UC Irvine

UC Irvine Previously Published Works

Title

A targeted genetic association study of epithelial ovarian cancer susceptibility

Permalink https://escholarship.org/uc/item/9ww2j29f

Journal

Oncotarget, 7(7)

ISSN

1949-2553

Authors

Earp, Madalene Winham, Stacey J Larson, Nicholas <u>et al.</u>

Publication Date 2016-02-16

DOI 10.18632/oncotarget.7121

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at https://creativecommons.org/licenses/by/4.0/

Peer reviewed

A targeted genetic association study of epithelial ovarian cancer susceptibility

Madalene Earp¹, Stacey J. Winham², Nicholas Larson², Jennifer B. Permuth³, Hugues Sicotte², Jeremy Chien⁴, Hoda Anton-Culver⁵, Elisa V. Bandera⁶, Andrew Berchuck⁷, Linda S. Cook⁸, Daniel Cramer^{9,10}, Jennifer A. Doherty¹¹, Marc T. Goodman¹², Douglas A. Levine¹³, Alvaro N.A. Monteiro³, Roberta B. Ness¹⁴, Celeste L. Pearce¹⁵, Mary Anne Rossing^{16,17}, Shelley S. Tworoger^{10,18}, Nicolas Wentzensen¹⁹, Maria Bisogna¹³, Louise Brinton¹⁹, Angela Brooks-Wilson^{20,21}, Michael E. Carney²², Julie M. Cunningham²³, Robert P. Edwards²⁴, Zachary C. Fogarty², Edwin S. Iversen²⁵, Peter Kraft²⁶, Melissa C. Larson², Nhu D. Le²⁷, Hui-Yi Lin³, Jolanta Lissowska²⁸, Francesmary Modugno^{24,29,30}, Kirsten B. Moysich³¹, Sara H. Olson³², Malcolm C. Pike^{15,31}, Elizabeth M. Poole¹⁸, David N. Rider², Kathryn L. Terry^{9,10}, Pamela J. Thompson¹², David van den Berg¹⁵, Robert A. Vierkant², Allison F. Vitonis⁹, Lynne R. Wilkens³³, Anna H. Wu¹⁵, Hannah P. Yang¹⁹, Argyrios Ziogas³⁴, Catherine M. Phelan³, Joellen M. Schildkraut^{35,36}, Yian Ann Chen³, Thomas A. Sellers³, Brooke L. Fridley³⁷ and Ellen L. Goode¹

¹ Department of Health Sciences Research, Division of Epidemiology, Mayo Clinic, Rochester, MN, USA

² Department of Health Sciences Research, Division of Biomedical Statistics and Informatics, Mayo Clinic, Rochester, MN, USA

³ Department of Cancer Epidemiology, H. Lee Moffitt Cancer Center and Research Institute, Tampa, FL, USA

⁴ Department of Cancer Biology, University of Kansas Cancer Center, Kansas City, KS, USA

⁵ Department of Epidemiology, University of California Irvine, Irvine, CA, USA

⁶ Rutgers Cancer Institute of New Jersey and Robert Wood Johnson Medical School, New Brunswick, NJ, USA

⁷ Duke Cancer Institute, Duke University Medical Center, Durham, NC, USA

⁸ Division of Epidemiology and Biostatistics, University of New Mexico, Albuquerque, NM, USA

⁹ Obstetrics and Gynecology Epidemiology Center, Brigham and Women's Hospital and Harvard Medical School, Boston, MA, USA

¹⁰ Department of Epidemiology, Harvard School of Public Health, Boston, MA, USA

¹¹ Section of Biostatistics and Epidemiology, The Geisel School of Medicine at Dartmouth, Lebanon, NH, USA

¹² Samuel Oschin Comprehensive Cancer Institute, Cedars Sinai Medical Center, Los Angeles, CA, USA

¹³ Gynecology Service, Department of Surgery, Memorial Sloan-Kettering Cancer Center, New York, NY, USA

¹⁴ The University of Texas School of Public Health, Houston, TX, USA

¹⁵ Department of Preventive Medicine, Keck School of Medicine, University of Southern California, Los Angeles, CA, USA

¹⁶ Department of Epidemiology, University of Washington, Seattle, WA, USA

¹⁷ Program in Epidemiology, Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, WA, USA

¹⁸ Channing Division of Network Medicine, Harvard Medical School and Brigham and Women's Hospital, Boston, MA, USA

¹⁹ Division of Cancer Epidemiology and Genetics, National Cancer Institute, Bethesda, MD, USA

²⁰ Genome Sciences Centre, BC Cancer Agency, Vancouver, BC, Canada

²¹ Department of Biomedical Physiology and Kinesiology, Simon Fraser University, Burnaby, BC, Canada

²² Clinical and Translational Research Program, University of Hawaii Cancer Center, Honolulu, HI, USA

²³ Department of Laboratory Medicine and Pathology, Division of Experimental Pathology, Mayo Clinic, Rochester, MN, USA

²⁴ Department of Obstetrics, Gynecology and Reproductive Sciences, Division of Gynecologic Oncology, University of Pittsburgh School of Medicine, Pittsburgh, PA, USA

²⁵ Department of Statistical Science, Duke University, Durham, NC, USA

²⁶ Departments of Epidemiology and Biostatistics, Harvard School of Public Health, Boston, MA, USA

²⁷ Cancer Control Research, BC Cancer Agency, Vancouver, BC, Canada

²⁸ Department of Cancer Epidemiology and Prevention, M. Sklodowska-Curie Memorial Cancer Center & Institute of Oncology, Warsaw, Poland

²⁹ Department of Epidemiology, University of Pittsburgh Graduate School of Public Health, Pittsburgh, PA, USA

³⁰ Cancer Research Program, Magee-Women's Research Institute and University of Pittsburgh Cancer Institute, Pittsburgh, PA, USA

³¹ Department of Cancer Prevention and Control, Roswell Park Cancer Institute, Buffalo, NY, US

³² Department of Epidemiology and Biostatistics, Memorial Sloan-Kettering Cancer Center, New York, NY, USA

³³ Cancer Epidemiology Program, University of Hawaii Cancer Center, Honolulu, HI, USA

³⁴ Department of Epidemiology, Center for Cancer Genetics Research and Prevention, School of Medicine, University of California Irvine, Irvine, CA, USA

³⁵ Department of Community and Family Medicine, Duke University Medical Center, Durham, NC, USA

³⁶ Cancer Prevention, Detection and Control Research Program, Duke Cancer Institute, Durham, NC, USA

³⁷ Kansas IDeA Network of Biomedical Research Excellence Bioinformatics Core, University of Kansas Cancer Center, Kansas City, KS, USA

Correspondence to: Ellen L. Goode, email: egoode@mayo.edu

Keywords: ovarian cancer, high-grade serous carcinoma, genetic association, susceptibility loci, NF-кВ Received: October 16, 2015 Accepted: January 24, 2016 Published: February 01, 2016

ABSTRACT

Background: Genome-wide association studies have identified several common susceptibility alleles for epithelial ovarian cancer (EOC). To further understand EOC susceptibility, we examined previously ungenotyped candidate variants, including uncommon variants and those residing within known susceptibility loci.

Results: At nine of eleven previously published EOC susceptibility regions (2q31, 3q25, 5p15, 8q21, 8q24, 10p12, 17q12, 17q21.31, and 19p13), novel variants were identified that were more strongly associated with risk than previously reported variants. Beyond known susceptibility regions, no variants were found to be associated with EOC risk at genome-wide statistical significance ($p < 5x10^{-8}$), nor were any significant after Bonferroni correction for 17,000 variants (p < 3x10-6).

Methods: A customized genotyping array was used to assess over 17,000 variants in coding, non-coding, regulatory, and known susceptibility regions in 4,973 EOC cases and 5,640 controls from 13 independent studies. Susceptibility for EOC overall and for select histotypes was evaluated using logistic regression adjusted for age, study site, and population substructure.

Conclusion: Given the novel variants identified within the 2q31, 3q25, 5p15, 8q21, 8q24, 10p12, 17q12, 17q21.31, and 19p13 regions, larger follow-up genotyping studies, using imputation where necessary, are needed for fine-mapping and confirmation of low frequency variants that fall below statistical significance.

INTRODUCTION

Epithelial ovarian cancer (EOC) is the second most common gynecologic cancer in the US, but it leads in deaths owing to its tendency to be diagnosed in the late stages of disease [1]. EOC is composed of five major histologic types [2]: high-grade serous carcinoma (HGSC), accounting for most cases (~70%); and the rarer clear cell, endometrioid, mucinous, and low-grade serous carcinomas (LGSC). Known rare mutations in DNA repair and mismatch repair genes are thought to account for 10%-15% of all EOCs [3-9]. Common alleles identified by genome-wide association studies (GWAS) are thought to account for an additional 3%-4% of EOC risk [10-17]. Still, much of about the heritability of EOC remains unaccounted for. Here, we sought to identify additional EOC susceptibility variants through direct genotyping and analysis of EOC cases and controls from 13 independent studies. We targeted variants based on innovative pilot studies, hypothesizing that previously ungenotyped variants may be responsible for a proportion of the unexplained EOC susceptibility.

Known EOC susceptibility regions

One goal of this project was to compare the relative strength of the associations between known and novel variants within the first eleven published EOC risk loci[10-12, 14, 18] (Supplemental Table 1). The variants most strongly associated with EOC risk in this study (all histology or HGSC) are given in Table 1 and plotted regionally in Figure 1, Figure 2, and Supplemental Figure 1. Compared to published variants, novel variants were more strongly associated with susceptibility of all histologies of EOC at nine loci (2q31, 3q25, 5p15, 8q21, 8q24, 10p12, 17q12, 17q21.31, and 19p13) (Table 1); all but three (8q21, 17q21.31, and 19p13) are in moderate LD $(r^{2} > 0.4)$ with known variants (Supplemental Figure 1). At the 3q25 locus variant rs62273902 ($p_{all-histology} = 2 \times 10^{-8}$) was the most strongly associated variant (Figure 1), and at the 17q21.31 locus variant rs2532240 ($p_{all-histology} = 3 \times 10^{-7}$) was the most strongly associated variant (Figure 2). In the HGSC only analysis, novel variants were more strongly associated with susceptibility at seven loci (2q31, 3q25, 8q24, 10p12, 17q12, 17q21.31, and 19p13; Table 1); all but two (2q31, 17q21.31) are in moderate LD ($r^{2} > 0.4$) with known variants (Supplemental Figure 1). With two exceptions (noted below), novel variants were common (minor allele frequency (MAF)>0.05), in the intron of genes or intergenic, in moderate-to-strong LD with known variants, and conferred modest effects on susceptibility. One exception was the association of rare intergenic variant rs74955251 at 8q21 (MAF_{overall} = 0.00028, OR= 3.9×10^{-6} , 95% confidence interval [CI]: 3.4×10^{-118} -4.6 x 10^{106} , $p_{all-histology} = 4 \times 10^{-3}$). Given its rarity, rs74955251 requires assessment in a much larger sample of cases and controls. A second exception was the association of common (MAF_{overall} = 0.50) missense variant rs2363956 in the gene ANKLE1 at 19p13 (OR= 0.91, 95% CI: 0.87-0.97, $p_{all-histology} = 2 \times 10^{-3}$, protein change Leu184Trp).

Beyond known EOC susceptibility regions

We targeted 5,320 variants which showed suggestive association with susceptibility in a pilot-scale whole genome sequence analysis that compared germline sequence of EOC patients (N= 19) to 1000 GP participants (N= 174). No novel variants reached genome-wide significance for association with EOC risk overall or HGSC ($p \le 5x10^{-8}$), nor were significant after Bonferroni correction ($p \le 9x10^{-6}$). Nonetheless, the risk estimates generally were in the expected direction based on pilot data, and several variants merit investigation in larger case-control collections (Table 2). For example, among variants targeted because they were present only in EOC

germline sequence data (WGS EOC+ in Table 2), the most strongly associated risk variant was rs138643956 (OR= 3.68; p_{HGSC} = 2 x10⁻⁴). For variants selected because they were absent from whole genome sequenced EOC cases (WGS EOC- in Table 2), the most associated variant was rs9380516 (OR= 0.83; p_{HGSC} = 6 x10⁻⁵); in the current study this variant showed a case MAF of 0.15, suggesting that this was a missed variant in the pilot sequencing study. For variants targeted which were present in whole genome sequenced EOC cases and in 1000 GP data, but differed in MAF (WGS EOC \uparrow and WGS EOC \downarrow in Table 2), the current analyses were generally consistent, including rs117841616 on chromosome 20 ($p_{all histology} = 2 \times 10^{-4}$) and rs240783 ($p_{HGSC} = 8 \times 10^{-4}$) on chromosome 6. In general, very few variants targeted based on suggestive association in pilot sequence study had appreciable MAF differences (case vs. control) in the current genotyping study. As expected due to small sample size, we observed that case MAF estimates in our pilot whole genome sequencing study were both inflated and deflated compared to the current study.

Finally, among NF-kB-related variants and those hypothesized to associate with endometrioid EOC risk, the most suggestive results for variants which disrupt NF-kB binding [19, 20] were intergenic variants rs10143322 on chromosome 14 ($p_{all-histology} = 3 \times 10^{-5}$) and rs6092485 on chromosome 20 ($p_{HGSC} = 7 \times 10^{-4}$) (Table 2). If Bonferroni correction for the number NF-kB binding site variants tested is applied, the threshold for statistical significance is $p < 4 \times 10^{-5}$ (p= 0.05/1,302), and this single variant, rs10143322, is declared significant; using experiment-wide and certainly genome-wide multiple testing corrections, it is not significant. Among variants previously identified in a pilot GWAS of endometrioid EOC, the most significant variants were intronic variant rs2638653 ($p_{endometrioid} = 1 \times 10^{-4}$) on chromosome 8 (Table 2), and intergenic rs9264042 on chromosome 6 ($p_{all-histology} =$ 5 x10-4). These modest associations may also warrant follow-up in larger studies.

DISCUSSION

The objective of this study was to test whether novel variants identified through a combination of approaches were associated with EOC susceptibility. We took an innovative approach to the selection of variants, including the use of whole genome sequencing data to target novel variants correlated with known GWAS risk variants, comparison of sequencing EOC cases to 1000 GP participants beyond these regions, NF- κ B functional data, and GWAS analysis of EOC cases with endometrioid histology.

In nine of the eleven susceptibility regions investigated, novel variants were more highly associated with all histology EOC risk than previously reported variants, and, in the HGSC only analysis, novel variants were more strongly associated with susceptibility at seven loci. Further work on thesevariant may provide more biological insight. For example, at the 3q25 locus, the novel variant rs62273902 (all histology) coincides with a genomic sequence that appears functionally active in a range of cell lines and tissues relevant to EOC, including ovary, as assayed by the Roadmap Epigenomics Mapping Consortium (REMC), http://www.epigenomebrowser. org/. rs62273902 resides within a DNase peak, an active transcription start site (TSS), and multiple proteins across diverse tissues bind the sequence spanning this variant. rs62273902 is therefore a good functional candidate variant at this locus. As well, at 17q21.31, the novel variants (rs2532240 in all histologies and rs3785880 in HGSC-only analysis) are separated by 272 kb and not correlated with each other or the previously reported variant rs1294266. rs2532240 coincides with a chromatin region marked as a weak/poised enhancer in several tissues, including ovary (REMC[21]); however, it does not overlap transcription factor binding sites (TFBS) or DNase peaks. rs12942666 does not coincide with promoter or enhancer regions in tissues relevant to EOC in the REMC data, suggesting it is unlikely to be functionally relevant. The 17q21.31 variants are located in a large region of strong LD previously identified as the "17q21.31 inversion" (~900kb long), which exists either as a direct (H1) or inverted (H2) haplotype in the European population [14, 15, 22]. Further investigation of how these variants might impact EOC risk is needed.

Of critical note, a large EOC meta-GWAS with imputation to revised Phase I 1000 GP data was recently completed with over 23,000 cases and 35,000 controls of the Ovarian Cancer Association Consortium, including many of the participants in the current analysis. We inspected our most associated variants from the 11 known susceptibility regions in in an online look-up of results based on these data (http://apps.ccge.medschl.cam.ac.uk/ consortia/ocac/contact/contact.html). In general, the variants reported here were highly ranked in the EOC meta-GWAS data (i.e., in the top 50 most associated variants in the regions we defined). At 8q24, the novel directly genotyped variant presented here (rs1400482) was the most associated variant in the larger imputationbased study. At 3q25, 10p12, 17q21.31, and 17q21.32,

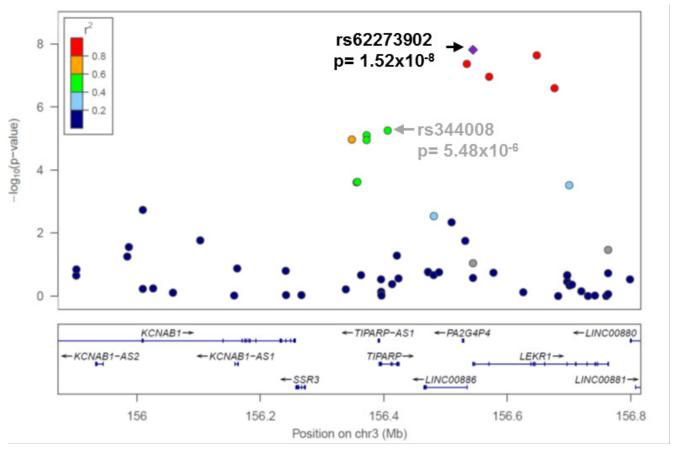


Figure 1: Novel variant rs62273902 in the 5'-untranslated region of LEKR1 has the strongest association signal at 3q25. Regional association plot for variants genotyped at 3q25 in all EOC histologies cases (N = 4,973) and controls (N = 5,640). Linkage disequilibrium between rs62273902 and each variant is estimated using data from 5,640 controls and indicated by the color scheme. The previously reported risk variant rs2665390 in this region {Goode, 2010 #23} was not genotyped; rs344008 ($p = 5x10^{-6}$) is indicated in its place to allow comparison of the novel (rs62273902) and known (rs2665390) most associated variants ($r^2 = 1$ for rs344008 and rs2665390 in 1000 Genomes Project phase 1 European data).

the most significant variant in the current study was not among the most significant in the imputation-based study. Nonetheless, novel variants at 3q25 and 17q21.32 remained highly significant (rs62273902 at 3q25 $p_{meta} =$ 2 x10⁻²⁸, and rs9303542 at 17q21.32 $p_{meta} =$ 3 x10⁻¹²). Although novel variants at 3q25 and 17q21.32 remained highly significant (rs62273902 at 3q25 $p_{meta} =$ 2 x10⁻²⁸, and rs9303542 at 17q21.32 $p_{meta} =$ 3 x10⁻¹²), at these two regions as well as at 10p12 and 17q21.31, the imputationbased study revealed stronger associations with other variants.

Among variants genotyped based on our pilot study comparing whole genomes of EOC cases and 1000 GP participants, none were significant after multiple testing correction for 5,320 variants. Despite our sample size (4,973 cases and 5,640 controls), power to detect associations with low MAF variants was limited. Variants in NF- κ B binding sites were also not associated with EOC risk at genome-wide significance. Noting the debate regarding the use of p<5 x10⁻⁸ as the threshold for statistical significance when evaluating potentially functional variants with presumed higher prior probability[23], Bonferroni correction for the number of NF- κ B binding site variants yields one statistically significant variant (rs10143322, p= 3x10⁻⁵). rs2638653, a variant selected based on an unpublished GWAS of endometrioid EOC, and found here to be the variant most associated with endometrioid EOC risk (p= 1 x10⁻⁴), coincides with chromatin marked as being an active promoter in ovary tissues (of *PSD3*), but not an enhancer or DNase site. Interestingly, loss of heterozygosity on chromosome 8p22, where this variant is located, is common in EOC tumors, and reduced expression of genes in this region has been found to negatively impact survival in EOC [24].

In summary, we developed a diverse panel of previously ungenotyped variants to directly test for association with EOC susceptibility in 4,973 EOC cases and 5,640 controls from 13 independent studies. Our innovative approach to variant selection included the first use of whole-genome sequencing data from EOC cases in novel variant discovery. At several EOC

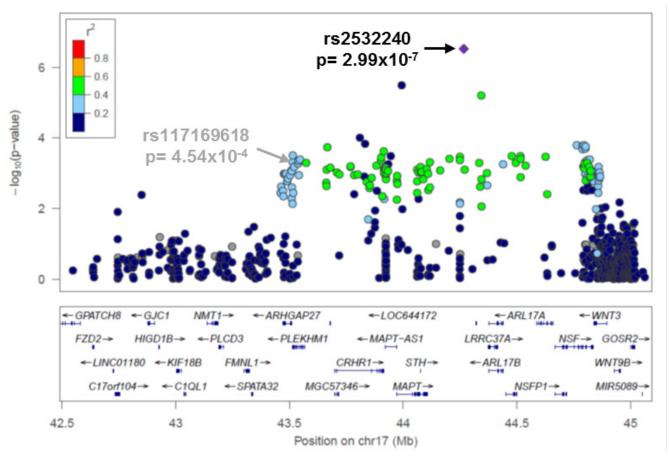


Figure 2: Novel variant rs2532240 has the strongest association signal at 17q21.31. Regional association plot for variants genotyped at 17q21.31 in all EOC histologies cases (N = 4,973) and controls (N = 5,640). The most associated variant was rs2532240 ($p = 3 \times 10^{-7}$). Linkage disequilibrium between rs2532240 and each variant is estimated using data from 5,640 controls and indicated by the color scheme. The previously reported risk variant rs12942666 in this region {Permuth-Wey, 2013 #28} was not genotyped, but rs117169618 ($p = 5 \times 10^{-4}$) is indicated in its place to allow comparison of the novel (rs2532240) and known (rs12942666) most variants ($r^2 = 0.8$ for rs117169618 and rs12942666 in 1000 Genomes Project phase 1 European data).

Table 1: Most significant associations within eleven known EOC susceptibility regions

	All histologies	s (4,973 cases	, 5,640 controls	5)		High grade serous (3,573 cases, 5,640 controls)						
Region	Variant	Position (hg19)	Nearest Gene	MAF overall	OR (95% CI)	P value	Variant	Position (hg19)	Nearest Gene	MAF overall	OR (95% CI)	P value
2q31	rs711830	177037311	3' UTR of HOXD3	0.33	1.12 (1.05-1.18)	2.05 x 10 ⁻⁴	rs1374325	177043971	Intron of HOXD1-AS	0.32	0.89 (0.83-0.96)	1.37 x 10 ⁻³
3q25	rs62273902	156544488	5' UTR of LEKR1	0.06	1.40 (1.25-1.57)	1.52 x 10 ⁻⁸	rs62275810	156647851	Intron of LEKR1	0.05	1.46 (1.27-1.68)	2.58 x 10 ⁻⁷
5p15	rs4975538	1280830	Intron of TERT	0.35	1.13 (1.06-1.20)	8.24 x 10 ⁻⁵	rs10069690	1279790	Intron of TERT	0.26	1.14 (1.06-1.22)	6.28 x 10 ⁻⁴
8q21	rs74955251	82230643	34kb 3' of <i>FABP5</i>	2.8 x 10 ⁻⁴	3.94 x 10 ⁻⁶ (3.4 x 10 ⁻¹¹⁸ -4.6 x 10 ¹⁰⁶)	5.71 x 10 ⁻³	rs11782652	82653644	Intron of CHMP4C	0.07	1.23 (1.09-1.39)	6.49 x 10 ⁻⁴
8q24	rs1400482	129541931	Intron of LINC00824	0.12	0.82 (0.75-0.89)	2.57 x 10 ⁻⁶	rs1400482	129541931	Intron of LINC00824	0.13	0.80 (0.73-0.89)	1.71 x 10⁵
9p22	rs3814113	16915021	44kb 5' of BNC2	0.32	0.81 (0.76-0.86)	7.96 x 10 ⁻¹³	rs3814113	16915021	44kb 5' of BNC2	0.32	0.75 (0.70-0.81)	3.20 x 10 ⁻⁵
10p12	rs4364959	22285371	Intron of DNAJC1	0.30	1.15 (1.08-1.22)	9.66 x 10 ⁻⁶	rs9971210	21879084	Intron of MLLT10	0.49	1.11 (1.04-1.19)	1.21 x 10 ⁻³
17q12	rs7405776	36093022	Intron of HNF1B	0.38	1.18 (1.11-1.24)	1.12 x 10 ⁻⁸	rs7405776	36093022	Inton of HNF1B	0.37	1.25 (1.17-1.34)	2.23 x 10 ⁻¹¹
17q21.31	rs2532240	44265839	Intron of KANSL1	0.41	1.16 (1.10-1.23)	2.99 x 10 ⁻⁷	rs3785880	43993376	Intron of MAPT	0.45	0.87 (0.82-0.93)	4.95 x 10 ⁻⁵
17q21.32	rs9303542	46411500	Intron of SKAP1	0.29	1.15 (1.08-1.23)	8.39 x 10 ⁻⁶	rs9303542	46411500	Intron of SKAP1	0.27	1.16 (1.08-1.25)	4.88 x 10 ⁻⁵
19p13	rs2363956	17394124	Missense mutation (L184W) in <i>ANKLE1</i>	0.50	0.91 (0.87-0.97)	1.57 x 10 ⁻³	rs56069439	17393925	Intron of ANKLE1	0.30	1.15 (1.07-1.23)	1.82 x 10 ⁻⁴

Bolded variants were previously reported as the most strongly associated variants in these susceptibility regions (Supplemental Table 1). MAF, minor allele frequency; OR, odds ratio; CI, confidence interval; UTR, untranslated region. Associations adjust for age, site, and three European principal components.

Table 2: Most significant EOC risk associations by selection criteria outside of eleven known susceptibility regions

	Variant	Chr.		Location	Pilot WGS Study		Case-control study data (13 sites)						
Selection Criteria			Position		EOC	1000 GP	Histology	Case	Control	N case,	OR (95%CI)	P value	
			(hg19)		MAF	MAF		MAF	MAF	N control			
WGS EOC+	rs138643956	10	79367857	Intron of KCNMA1	0.053	0.000	HGSC	0.004	0.001	3035, 5637	3.68 (1.79 - 7.55)	1.85 x 10 ⁻⁴	
WGS EOC†	rs117841616	20	57855211	20kb 5' of EDN3	0.079	0.006	All	0.008	0.005	4973, 5634	1.93 (1.36 - 2.74)	2.06 x 10 ⁻⁴	
WGS EOC↓	rs240783	6	100968737	Intron of ASCC3	0.184	0.494	HGSC	0.400	0.428	2956, 5518	0.89 (0.84 - 0.95)	7.90 x 10 ⁻⁴	
WGS EOC-	rs9380516	6	35502202	10kb 5' of RP3-340B19.3	0.000	0.155	HGSC	0.147	0.171	3027, 5633	0.83 (0.76 - 0.91)	6.44 x 10 ⁻⁵	
NF-κB	rs10143322	14	91556577	24kb 5' of C14orf159	n.a.	n.a.	All	0.220	0.246	4562, 5634	0.87 (0.82 - 0.93)	2.99 x 10 ⁻⁵	
NF-κB	rs6092485	20	56045014	26kb 3' of CTCFL	n.a.	n.a.	HGSC	0.335	0.311	3019, 5625	1.13 (1.05 – 1.21)	6.77 x 10 ⁻⁴	
Endometrioid GWAS	6 rs9264042	6	31196801	25kb 3' of HCG27	n.a.	n.a.	All	0.125	0.107	4440, 5505	1.17 (1.07 – 1.28)	4.71 x 10 ⁻⁴	
Endometrioid GWAS	6 rs2638653	8	18666210	Intron of PSD3	n.a.	n.a.	EC	0.409	0.362	832, 5619	1.23 (1.11 – 1.37)	1.28 x 10 ⁻⁴	

WGS, whole-genome sequencing; EOC, epithelial ovarian cancer; 1000 GP, 1000 Genomes Project; GWAS, genome wide association study; Chr, chromosome; MAF, minor allele frequency; OR, odds ratio; CI, confidence interval; HGSC, high grade serous carcinoma; n.a., not applicable. WGS EOC+ variant selection criteria: MAF> 0% in serous EOC cases, monomorphic in 1000 GP European individuals; WGS EOC↑ variant selection criteria: polymorphic in cases and 1000 GP, MAF greater in WGS patients; WGS EOC↓ variant selection criteria: polymorphic in cases and 1000 GP, MAF greater in 1000 GP. variant selection criteria: polymorphic in cases and 1000 GP; WGS 1000 EOC− variant selection criteria: monomorphic in cases, MAF> 0% in 1000 GP.

susceptibility regions, we report novel risk variants for further association and functional investigation. Beyond known regions, this first pass at using whole genome sequencing pilot analyses, although underpowered, also yielded variants of potential interest (rs138643956 and rs117841616). The key strength of this report is the use of direct genotyping of novel variants, some rare, while its key limitation is an inability to more comprehensively examine rare variation. Larger scale genotyping and/or improved genotype imputation accuracy will facilitate further scrutiny of the variants highlighted here.

MATERIALS AND METHODS

Study participants

Study participants were drawn from 13 independent EOC case-control studies of the Ovarian Cancer Association Consortium and were restricted to women of European ancestry. Characteristics of the contributing studies are given in Supplemental Table 2 and have been described previously [18]. Cases (N=4,973) consisted of women aged 18 and older with a pathologically confirmed primary invasive EOC, fallopian tube cancer, or primary peritoneal cancer; controls (N=5,640) were matched by age and region.

Genotyping array

A total of 17,439 germline DNA variants were genotyped using a customized Affymetrix Axiom Exome array (Affymetrix Corporation, Santa Clara, CA). These variants were drawn from four discovery categories: 1) from eleven known EOC susceptibility regions (N =6,948; Supplemental Table 3)[10-14, 18], identified by in silico fine-mapping and a small germline whole genome sequencing study of EOC cases, 2) variants outside these eleven regions which showed suggestive association in pilot whole genome sequencing of serous EOC cases (compared to 1000 Genomes Project [GP] data)(N = 7,189), 3) variants with a hypothesized role in disrupted binding of NF-kB transcription factors, which are known to have central roles in immune and inflammatory responses and cancer [19, 20, 25, 26] (N = 1,302), and 4) the top associated variants from a pilot GWAS of endometrioid EOC (N = 2,000). See the Supplemental Methods for more detail on the selection of these variants.

Quality control

Germline DNA was genotyped at the Affymetrix Research Services Laboratory (Santa Clara, CA) using default quality control (QC) and genotype calling criteria. Variants failed QC if: (1) the call rate was < 95%; (2) p-values of Hardy-Weinberg equilibrium in controls were $< 10^{-7}$; or (3) there was > 2% discordance in duplicate pairs. Further, monomorphic variants were removed. Of 6,948 variants genotyped within 11 known EOC risk regions, 4,919 (71%) met these QC criteria and were polymorphic. Outside of these regions, of 7,189 variants selected based on whole-genome sequencing data, 5,286 (74%) met QC criteria and were polymorphic. Of 1,302 variants associated with NF-kB binding, 980 (75%) met QC criteria and were polymorphic, and, of 2,000 variants selected from a GWAS of endometrioid EOC, 1,826 (91%) met QC criteria and were polymorphic. Most variants were excluded for being monomorphic. Thus, a total of 13,011 genotyped variants remained for analysis.

Association analysis

All cases were included in the overall EOC risk association analyses (N=4,973). Subset analyses were performed on histologic subsets based on *a priori* selection; HGSC (N=3,573) and endometrioid EOC (N=835). For each analysis, 5,640 controls were used. Associations were estimated using logistic regression assuming an additive genetic model, adjusting for age, study site, and population substructure by including the first three eigenvalues from principal components analysis[18]. All analyses were conducted in R version

3.0.2 (http://www.R-project.org/).

ACKNOWLEDGMENTS

The NHS/NHSII studies thank the following state cancer registries for their help: AL, AZ, AR, CA, CO, CT, DE, FL, GA, ID, IL, IN, IA, KY, LA, ME, MD, MA, MI, NE, NH, NJ, NY, NC, ND, OH, OK, OR, PA, RI, SC, TN, TX, VA, WA, and WY.

CONFLICTS OF INTEREST

The authors have no conflicts of interest to disclose.

GRANT SUPPORT

Funding support for the Follow-up of Ovarian Cancer Genetic Association and Interaction Studies (FOCI) was provided through the National Cancer Institute's Cancer Post-GWAS Initiative, Genetic Associations and Mechanisms in Oncology (GAME-ON) (U19-CA148112). In addition, we acknowledge the following: DOV: National Institutes of Health R01-CA112523 and R01-CA87538; HAW: National Institutes of Health (R01-CA58598, N01-CN-55424 and N01-PC-67001); HOP: DOD DAMD17-02-1-0669 and NCI K07-CA080668, R01-CA95023, P50-CA159981; NIH/National Center for Research Resources/General Clinical Research Center grant M01-RR000056; MAY: National Institutes of Health (R01-CA122443, P30-CA15083-41, P50-CA136393); NCO: National Institutes of Health (R01-CA76016) and the Department of Defense (DAMD17-02-1-0666); NEC: National Institutes of Health R01-CA54419 and P50-CA105009 and Department of Defense W81XWH-10-1-02802; NHS: NIH (P01-CA87696 and R01-CA49449): NJO: National Cancer Institute (K07-CA095666, R01-CA83918,K22-CA138563, and P30-CA072720) and the Cancer Institute of New Jersey; NCI CCSG award (P30-CA008748); OVA: This work was supported by Canadian Institutes of Health Research grant (MOP-86727); LEK is supported by a Canadian Institutes of Health Research New Investigator award (MSH-87734); POL: Intramural Research Program of the National Cancer Institute; UCI: R01-CA058860, R01-CA092044, US Public Health Service PSA-042205, and the Lon V Smith Foundation grant LVS-39420; USC: P01-CA17054, P30-CA14089, R01-CA61132, N01-PC67010, R03-CA113148, R03-CA115195, N01-CN025403, and California Cancer Research Program (00-01389V-20170, 2II0200). CMP is supported by National Institutes of Health R01-CA-149429.

REFERENCES

1. Center M, Siegel R and Jemal A. (2011). Global Cancer

Facts & Figures. (Atlanta: American Cancer Society).

- Kobel M, Kalloger SE, Lee S, Duggan MA, Kelemen LE, Prentice L, Kalli KR, Fridley BL, Visscher DW, Keeney GL, Vierkant RA, Cunningham JM, Chow C, Ness RB, Moysich K, Edwards R, et al. Biomarker-based ovarian carcinoma typing: a histologic investigation in the ovarian tumor tissue analysis consortium. Cancer Epidemiol Biomarkers Prev. 2013; 22:1677-1686.
- Hall JM, Lee MK, Newman B, Morrow JE, Anderson LA, Huey B and King MC. Linkage of early-onset familial breast cancer to chromosome 17q21. Science. 1990; 250:1684-1689.
- Lynch HT, Schuelke GS, Kimberling WJ, Albano WA, Lynch JF, Biscone KA, Lipkin ML, Deschner EE, Mikol YB, Sandberg AA and et al. Hereditary nonpolyposis colorectal cancer (Lynch syndromes I and II). II. Biomarker studies. Cancer. 1985; 56:939-951.
- Lynch HT, Conway T and Lynch J. Hereditary ovarian cancer. Pedigree studies, Part II. Cancer Genet Cytogenet. 1991; 53:161-183.
- Boyd J and Rubin SC. Hereditary ovarian cancer: molecular genetics and clinical implications. Gynecol Oncol. 1997; 64:196-206.
- Walsh T, Casadei S, Lee MK, Pennil CC, Nord AS, Thornton AM, Roeb W, Agnew KJ, Stray SM, Wickramanayake A, Norquist B, Pennington KP, Garcia RL, King MC and Swisher EM. Mutations in 12 genes for inherited ovarian, fallopian tube, and peritoneal carcinoma identified by massively parallel sequencing. Proc Natl Acad Sci U S A. 2011; 108:18032-18037.
- Narod SA, Madlensky L, Bradley L, Cole D, Tonin P, Rosen B and Risch HA. Hereditary and familial ovarian cancer in southern Ontario. Cancer. 1994; 74:2341-2346.
- Risch HA, McLaughlin JR, Cole DE, Rosen B, Bradley L, Kwan E, Jack E, Vesprini DJ, Kuperstein G, Abrahamson JL, Fan I, Wong B and Narod SA. Prevalence and penetrance of germline BRCA1 and BRCA2 mutations in a population series of 649 women with ovarian cancer. Am J Hum Genet. 2001; 68:700-710.
- Song H, Ramus SJ, Tyrer J, Bolton KL, Gentry-Maharaj A, Wozniak E, Anton-Culver H, Chang-Claude J, Cramer DW, DiCioccio R, Dork T, Goode EL, Goodman MT, Schildkraut JM, Sellers T, Baglietto L, et al. A genomewide association study identifies a new ovarian cancer susceptibility locus on 9p22.2. Nat Genet. 2009; 41:996-1000.
- Goode EL, Chenevix-Trench G, Song H, Ramus SJ, Notaridou M, Lawrenson K, Widschwendter M, Vierkant RA, Larson MC, Kjaer SK, Birrer MJ, Berchuck A, Schildkraut J, Tomlinson I, Kiemeney LA, Cook LS, et al. A genome-wide association study identifies susceptibility loci for ovarian cancer at 2q31 and 8q24. Nat Genet. 2010; 42:874-879.
- 12. Bolton KL, Tyrer J, Song H, Ramus SJ, Notaridou M,

Jones C, Sher T, Gentry-Maharaj A, Wozniak E, Tsai YY, Weidhaas J, Paik D, Van Den Berg DJ, Stram DO, Pearce CL, Wu AH, et al. Common variants at 19p13 are associated with susceptibility to ovarian cancer. Nat Genet. 2010; 42:880-884.

- Bojesen SE, Pooley KA, Johnatty SE, Beesley J, Michailidou K, Tyrer JP, Edwards SL, Pickett HA, Shen HC, Smart CE, Hillman KM, Mai PL, Lawrenson K, Stutz MD, Lu Y, Karevan R, et al. Multiple independent variants at the TERT locus are associated with telomere length and risks of breast and ovarian cancer. Nat Genet. 2013; 45:371-384, 384e371-372.
- 14. Permuth-Wey J, Lawrenson K, Shen HC, Velkova A, Tyrer JP, Chen Z, Lin HY, Chen YA, Tsai YY, Qu X, Ramus SJ, Karevan R, Lee J, Lee N, Larson MC, Aben KK, et al. Identification and molecular characterization of a new ovarian cancer susceptibility locus at 17q21.31. Nat Commun. 2013; 4:1627.
- 15. Couch FJ, Wang X, McGuffog L, Lee A, Olswold C, Kuchenbaecker KB, Soucy P, Fredericksen Z, Barrowdale D, Dennis J, Gaudet MM, Dicks E, Kosel M, Healey S, Sinilnikova OM, Bacot F, et al. Genome-wide association study in BRCA1 mutation carriers identifies novel loci associated with breast and ovarian cancer risk. PLoS Genet. 2013; 9:e1003212.
- 16. Chen K, Ma H, Li L, Zang R, Wang C, Song F, Shi T, Yu D, Yang M, Xue W, Dai J, Li S, Zheng H, Wu C, Zhang Y, Wu X, et al. Genome-wide association study identifies new susceptibility loci for epithelial ovarian cancer in Han Chinese women. Nat Commun. 2014; 5:4682.
- 17. Kuchenbaecker KB, Ramus SJ, Tyrer J, Lee A, Shen HC, Beesley J, Lawrenson K, McGuffog L, Healey S, Lee JM, Spindler TJ, Lin YG, Pejovic T, Bean Y, Li Q, Coetzee S, et al. Identification of six new susceptibility loci for invasive epithelial ovarian cancer. Nat Genet. 2015.
- Pharoah PD, Tsai YY, Ramus SJ, Phelan CM, Goode EL, Lawrenson K, Buckley M, Fridley BL, Tyrer JP, Shen H, Weber R, Karevan R, Larson MC, Song H, Tessier DC, Bacot F, et al. GWAS meta-analysis and replication identifies three new susceptibility loci for ovarian cancer. Nat Genet. 2013; 45:362-370.
- Kasowski M, Grubert F, Heffelfinger C, Hariharan M, Asabere A, Waszak SM, Habegger L, Rozowsky J, Shi M, Urban AE, Hong MY, Karczewski KJ, Huber W, Weissman SM, Gerstein MB, Korbel JO, et al. Variation in transcription factor binding among humans. Science. 2010; 328:232-235.
- Karczewski KJ, Tatonetti NP, Landt SG, Yang X, Slifer T, Altman RB and Snyder M. Cooperative transcription factor associations discovered using regulatory variation. Proc Natl Acad Sci U S A. 2011; 108:13353-13358.
- Ernst J and Kellis M. ChromHMM: automating chromatinstate discovery and characterization. Nat Methods. 2012; 9:215-216.
- 22. Steinberg KM, Antonacci F, Sudmant PH, Kidd JM,

Campbell CD, Vives L, Malig M, Scheinfeldt L, Beggs W, Ibrahim M, Lema G, Nyambo TB, Omar SA, Bodo JM, Froment A, Donnelly MP, et al. Structural diversity and African origin of the 17q21.31 inversion polymorphism. Nat Genet. 2012; 44:872-880.

- Broer L, Lill CM, Schuur M, Amin N, Roehr JT, Bertram L, Ioannidis JP and van Duijn CM. Distinguishing true from false positives in genomic studies: p values. Eur J Epidemiol. 2013; 28:131-138.
- 24. Pils D, Horak P, Gleiss A, Sax C, Fabjani G, Moebus VJ, Zielinski C, Reinthaller A, Zeillinger R and Krainer M. Five genes from chromosomal band 8p22 are significantly down-regulated in ovarian carcinoma: N33 and EFA6R have a potential impact on overall survival. Cancer. 2005; 104:2417-2429.
- 25. Charbonneau B, Block MS, Bamlet WR, Vierkant RA, Kalli KR, Fogarty Z, Rider DN, Sellers TA, Tworoger SS, Poole E, Risch HA, Salvesen HB, Kiemeney LA, Baglietto L, Giles GG, Severi G, et al. Risk of ovarian cancer and the NF-kappaB pathway: genetic association with IL1A and TNFSF10. Cancer Res. 2014; 74:852-861.
- 26. Block MS, Charbonneau B, Vierkant RA, Fogarty Z, Bamlet WR, Pharoah PD, Rossing MA, Cramer D, Pearce CL, Schildkraut J, Menon U, Kjaer SK, Levine DA, Gronwald J, Culver HA, Whittemore AS, et al. Variation in NF-kappaB signaling pathways and survival in invasive epithelial ovarian cancer. Cancer Epidemiol Biomarkers Prev. 2014; 23:1421-1427.