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Variability of p27 Gene from Several Isolates of Citrus Tristeza Virus as Analyzed by Single-Strand Conformation Polymorphism

S. Gago-Zachert, L. Semorile, N. Costa, and O. Grau

ABSTRACT. Hybridization analysis showed a region of the citrus tristeza virus (CTV) p27 gene which exhibits different hybridization patterns using probes derived from 10 biological distinct CTV isolates. In an attempt to determine if that gene region differs between mild and severe strains, 10 CTV isolates representing different biogroups were compared for nucleotide sequence variation by single strand conformation polymorphism (SSCP) analysis. The p27 gene was reverse transcribed, amplified by PCR and 30 clones of each isolate were obtained. From each clone, two gene fragments were amplified by PCR, (a) 459 bp and (b) 281 bp. Sequence variations in both gene fragments were studied by SSCP analysis. The p27 b gene region showed a higher rate of sequence variation than the p27 a region. On the other hand, the intrapopulation variability for the p27 gene showed no correlation with the biological characteristics of the viral isolates or the environmental conditions. The analysis of SSCP profiles of the majority of the clones suggests that the mild CTV isolates (biogroups I and II) are more homogeneous than the severe isolates (biogroups III, IV and V) for the p27 gene region.

The citrus tristeza virus (CTV) genome is composed of a single stranded RNA with two capsid proteins (CP) with the p25 CP encapsidating about 95% of the particle length, and a p27 diverged (d)CP present only on one end of the particle, forming a "rattlesnake" structure (5, 6). Moreover, the dCP may have a function in the transmission by aphid vectors as was suggested in beet yellows closterovirus (1).

CTV populations in citrus trees are unusually complex mixtures of viral genotypes and defective RNAs developed during long-term vegetative propagation and by additional mixing by aphid transmission (14). A variety of CTV strains, differing by symptoms induced on different host species, aphid transmissibility or capacity to interfere with other strains have been described (2, 10, 13). Several approaches for CTV isolate identification and differentiation have been tested in an attempt to develop a quick and specific procedure that could be applied to routine purposes.

Single-strand conformation polymorphism (SSCP) (9) is a method for detecting sequence differences of ssDNA by non-denaturing polyacrylamide gel electrophoresis (PAGE). Partially denatured DNA in the gel matrix has a sequence-specific three dimensional conformation having a characteristic mobility. Even a single nucleotide difference can produce a mobility shift. SSCP analysis has been shown to be an appropriate technique for the study of viral populations. Complex mixtures of the same size, such as viral guasispecies, can be separated into bands of different mobility (4). Rubio et al. (11) applied this technique to CTV isolates and compared them for variations in their CP gene.

From dsRNA isolated from field isolate C268-2 (from Concordia, Entre Ríos, Argentina), we obtained a cDNA library composed of 30 clones (12). Southern blot hybridizations of these clones with short copy cDNAs probes generated from the dsRNAs from 10 CTV isolates, representing biogroups I to V, allowed us to select 11 genomic regions, including the p27 gene, usable to discriminate among biologically different viral isolates (7).

With the aim of screening for variability of the p27 gene among CTV isolates, we compared 10 CTV isolates for sequence variations in the p27 gene region by SSCP analysis. Moreover, we used SSCP to analyze the variability of viral RNA populations in the CTV isolates studied. The origins and biological characteristics of the CTV isolates used in this study are shown in Table 1.

Bark from young shoots infected with CTV was pulverized in a mortar while frozen in liquid nitrogen. Nucleic acid preparations enriched in dsRNAs were obtained by extraction using buffered-saturated phenol and fractioned by column chromatography on non-ionic CF-11 cellulose (Whatman) as previously described (8).

The p27 gene was reverse transcribed with the antisense p27 ORF primer (5), amplified by PCR with the sense and antisense p27 ORF primers (5) and 30 clones of each isolate were obtained. SSCP analyses were performed on PCR products of the p27 gene split into two fragments of about 460 and 280 bp in size. Usually, PCR products were extracted twice, first with phenolchloroform and then with chloroform, precipitated and resuspended in 10 µl of double distilled water. A fraction of 1 μ l was mixed with 9 μ l of the denaturing solution (95% formamide, 20 mM EDTA and 500 mg/ liter bromophenol blue), heated for 10 min at 100°C, and chilled on ice. Denatured DNA was electrophoresed in a non-denaturing polyacrylamide 10×8 cm minigel (SE 250/SE 260, Hoefer Scientific), using $1\times$ TBE (90 mM Tris borate, 2 mM EDTA) as electrophoresis buffer, 200 V, 3.5 h. During the electrophoresis, temperature was maintained constant by water circulation. The gels were stained with silver nitrate (3).

From each p27 clone, two gene fragments were amplified: Fragment a, 459 bp long and including domains I and II, and Fragment b, 281 bp long and including domains II, III and IV. To obtain good discrimination of SSCP profiles of p27 gene Fragment a, we had to use a 12% polyacrylamide gel concentration and a lower temperature (20°C); whereas for Fragment b, we used 15% acrylamide and a temperature of 22°C. In most clones, only two intense bands, corresponding to only one stable conformation of each DNA strand, were obtained. More than one SSCP profile was observed in the 30 clones analyzed in each of the p27 gene fragments. These results indicate the heterogeneity of the population.

According to the results, the p27 Fragment b showed a higher rate of sequence variation than p27 Fragment a. These changes may be related eventually with the biological properties of the virus considering that p27 Fragment b, which contains domains II, III, and IV that are involved in protein folding, showed the highest number of variants. The number of SSCP patterns that could be distinguished for p27 Fragment a varied between four (for the biogroup V isolates) to 11 (for Pera GS of biogroup III). On the other hand, p27 Fragment b yielded a higher number of different SSCP patterns: 12 patterns in the most variable isolates (Pera GS, biogroup III; Baraõ B, biogroup IV and, T388, biogroup V), and three distinct patterns in the less variable isolate (C268-2, biogroup III). The clone number that showed variation in both p27 gene fragments varied between one (T32, biogroup I) to 12 (Pera GS, biogroup III). On the other hand, the quantity of clones for each isolate that show any variation in p27 gene varied from 21 (C268-2) to six (Pera GS).

SSCP has been shown to be a powerful alternative for detection of variation within a virus population, therefore, it was applied to study the intrapopulation variability for the p27 gene of CTV. With this purpose, the percentage of principal clones for the complete p27 gene without sequence variations was

| TABI ORIGIN AND BIOGROUP INFORMATION FOR THE CITR | ,Ε 1 US TRISTEZA V | VIRUS ISOL | ATES USED | IN THIS STI | UDY | |
|---|------------------------------------|--|---|-------------|----------|--------------------|
| Isolates | ML ^z (SP ^y) | SwO ^z /SO ^z (decline) | $\operatorname{Gpft}^{z}\operatorname{or}\operatorname{SO}_{(\operatorname{SY}^{Y})}$ | Gpft (SP) | SwO (SP) | Biogroup (type) |
| T385 (Valencia, Spain), T32 (Valencia, Spain) | + | 1 | I | 1 | 1 | |
| T300 (Valencia, Spain), T312 (Valencia, Spain) | + | + | | | | П |
| C268-2 (Argentina), Pera GS (Brazil, isolate used in cross-protection assays) | + | + | + | | | III |
| C269-6 (Star Ruby grapefruit, Argentina), Baraõ B (Brazil) | + | + | + | + | | IV |
| T388 (Valencia, Spain), Capaõ Bonito (Brazil) | + | + | + | + | + | Λ |
| | | | | | | |

*Indicator species: ML = Mexican lime; SwO = sweet orange; Gpft = grapefruit; SO = sour orange. *Symptoms: SP = stem pitting; SY = seedling yellows.

340

also estimated. C268-2 exhibited the higher percentage (66.7%) for the whole gene, being the most homogeneous isolate for this gene region. In contrast, the Pera GS isolate had the lowest percentage (20%) of clones without nucleotide changes and was the most heterogeneous isolate for the p27 gene region among the CTV isolates analyzed. The results obtained showed any correlation between biological characterof the viral isolates or istics environmental conditions and the variability of the viral population for this gene region.

The SSCP profiles of one representative majority clone for each CTV isolate were compared by PAGE. The p27 Fragment a of the isolates in biogroup I, and one isolate (T312) from biogroup II exhibited the same SSCP profile, but T300 (biogroup II) and the other six isolates compared exhibited different SSCP profiles among themselves and with the fore-mentioned isolates. For p27 Fragment b fragment, the four isolates of biogroup I and II shared the same SSCP pattern while all the other isolates showed a different SSCP profile among themselves and with biogroups I and II. These results suggest that if the p27 CTV gene region is considered, the mild isolates (biogroups I and II) are more homogeneous in nucleotide sequence. The more severe isolates (biogroups III, IV, and V) showed more nucleotide sequence heterogeneity in both intragroup and intergroup comparisons. Moreover, no correlation was found between geographical origin and SSCP pattern for the analyzed gene region.

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LITERATURE CITED

- Agranovsky, A. A., D. E. Lesemann, E. Maiss, R. Hull, and J. G. Atabekov 1995. Rattlesnake structure of a filamentous plant RNA virus built of two capsid proteins. Proc. Natl. Acad. Sci. 92: 2470-2473.
- Ballester-Olmos, J. F., J. A. Pina, E. Carbonell, P. Moreno, A. Hermoso de Mendoza, M. Cambra, and L. Navarro 1993. Biological diversity of citrus tristeza virus (CTV) isolates in Spain. Plant Pathol.
- 42: 219-229.
 Beidler, L. L., P. R. Hilliard, and R. L. Rill
- 1982. Ultrasensitive staining of nucleic acids with silver. Anal. Biochem. 120: 126: 374-380.
- Enomoto, N., M. Kurosaki, Y. Tanaka, F. Marumo, and C. Sato 1994. Fluctuation of hepatitis C virus quasispecies in persistent infection and interferon treatment revealed by single-strand conformation polymorphism analysis. J. Gen. Virol. 75: 1361-1369.
- Febres, V. J., H. R. Pappu, E. J. Anderson, S. S. Pappu, R. F. Lee, and C. L. Niblett 1994. The diverged copy of the citrus tristeza virus coat protein is expressed *in vivo*. Virology 201: 178-181.
- Febres, V. J., L. Ashoulin, M. Mawassi, A. Frank, M. Bar-Joseph, R. F. Lee, and C. L. Niblett 1996. The p27 protein is present at one end of citrus tristeza virus particles. Phytopathology 86: 13331-1335.
- Gago-Zachert, S., N. Costa, L. Semorile, and O. Grau 2000. Differential genomic regions among citrus tristeza virus isolates. In: Proc. 14th Conf. IOCV, 332-334. IOCV, Riverside, CA.

- 8. Moreno, P., J. Guerri, and N. Muñoz
 - 1990. Identification of Spanish strains of citrus tristeza virus (CTV) by analysis of double-stranded RNAs (dsRNAs). Phytopathology 80: 477-482.
- Orita, M., Y. Suzuki, T. Sekiya, and K. Hayashi 1989. Rapid and sensitive detection of point mutations and DNA polymorphisms. Genomics 5: 874-879.
- Raccah, B., M. Bar-Joseph, and G. Loebenstein 1980. Aphid transmissibility of citrus tristeza virus in infected trees. Phytopathology 70: 1173-1178.
- Rubio-Miguelez, L., M. A. Ayllón, J. Guerri, H. Pappu, C. L. Niblett, and P. Moreno 1996. Differentiation of citrus tristeza virus (CTV) isolates by single-strand conformation polymorphism analysis of the coat protein gene. Ann. Appl. Biol. 129: 479-489.
- Semorile, L., R. Dewey, M. L. García, E. Dal Bó, P. D. Ghiringhelli, V. Romanowski, and O. Grau
 1993. cDNA clones of CTV that discriminate severe and mild strains. In: Proc. 12th

1993. cDNA clones of CTV that discriminate severe and mild strains. In: *Proc. 12th Conf. IOCV*, 28-32. IOCV, Riverside, CA.

- Van Vuuren, S. P., R. P. Collins, and J. V. da Graça 1993. Evaluation of citrus tristeza virus isolates for cross-protection of grapefruit. Plant Dis. 77: 24-28.
- 14. Yang, Z. N., D. M. Mathews, J. A. Dodds, and T. E. Mirkov
 - 1999. Molecular characterization of an isolate of citrus tristeza virus that causes severe symptoms in sweet orange. Virus Genes 19: 131-142.