Lawrence Berkeley National Laboratory

Joint Genome Institute

Title

Complete Genome Sequences of Evolved Arsenate-Resistant Metallosphaera sedula Strains

Permalink https://escholarship.org/uc/item/9hs5z07q

Journal Microbiology Resource Announcements, 3(5)

ISSN 2576-098X

Authors

Ai, Chenbing McCarthy, Samuel Schackwitz, Wendy <u>et al.</u>

Publication Date

2015-10-29

DOI

10.1128/genomea.01142-15

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <u>https://creativecommons.org/licenses/by/4.0/</u>

Peer reviewed





Complete Genome Sequences of Evolved Arsenate-Resistant Metallosphaera sedula Strains

Chenbing Ai,^{a,b} Samuel McCarthy,^a Wendy Schackwitz,^c Joel Martin,^c Anna Lipzen,^c Paul Blum^a

School of Biological Sciences, University of Nebraska-Lincoln, Lincoln, Nebraska, USA^a, School Of Minerals Processing and Bioengineering, Central South University, Hunan, China^b; U.S. Department of Energy-Joint Genome Institute, Walnut Creek, California, USA^c

Metallosphaera sedula is a thermoacidophilic crenarchaeote with a 2.19-Mb genome. Here, we report the genome sequences of several evolved derivatives of *M. sedula* generated through adaptive laboratory evolution for enhanced arsenate resistance.

Received 18 August 2015 Accepted 19 August 2015 Published 1 October 2015

Citation Ai C, McCarthy S, Schackwitz W, Martin J, Lipzen A, Blum P. 2015. Complete genome sequences of evolved arsenate-resistant *Metallosphaera sedula* strains. Genome Announc 3(5):e01142-15. doi:10.1128/genomeA.01142-15.

Copyright © 2015 Ai et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Paul Blum, pblum1@unl.edu

Metallosphaera sedula is a thermoacidophilic crenarchaeote that grows optimally at 75°C and pH 2.0 and is capable of lithoautotrophy (1, 2). Its ability to oxidize iron and sulfur (2) is used in biomining, and this organism has been used to extract copper from low-grade copper sulfide ores and tailings, especially by heap bioleaching (3). The oxidation of copper sulfide ores is an exothermic process that can elevate temperatures inside mineral heaps to 60 to ~80°C (4). These temperatures are lethal to mesophilic acidophiles and moderate the thermoacidophiles used in biomining. However, as an extreme thermoacidophile, *M. sedula* is able to withstand and continue bioleaching at these high temperatures.

Arsenic is very prevalent in low-grade copper sulfide ores and tailings, and during biomining, it is released from the ores along with the other metals. Extreme thermoacidophiles used in bioleaching, such as *M. sedula*, are very sensitive to arsenic (1, 5). Bioinformatics analysis has shown that previously studied arsenic resistance pathways in biomining mesophiles and moderate thermoacidophiles, such as the ars operon (6, 7), are not present in these organisms. Therefore, arsenate-resistant M. sedula strains would be beneficial for the bioprocessing of arsenic-bearing copper sulfide ores and tailings. Four arsenate-adapted derivatives of M. sedula (DSM 5348) and the copper-resistant M. sedula strain CuR1 (8) were isolated through adaptive laboratory evolution, involving extensive passage during selection for the biological trait of increased arsenate resistance (unpublished data). Here, we report the complete genome sequence of arsenate-resistant isolates derived from M. sedula DSM 5348, named ARS120-1 and ARS120-2, and those derived from *M. sedula* strain CuR1 (8), named ARS50-1 and ARS50-2.

High-molecular-weight genomic DNA was prepared from clonal cultures of the *M. sedula* strains, as described previously (2, 9). The integrity and purity of the DNA samples were verified by spectroscopic measurements at 260/280 and 260/230 nm and confirmed by agarose gel electrophoresis. DNA and RNA library preparation was conducted using the Joint Genome Institute (JGI)'s automated process with a Biomek FX robot. The samples were sheared using a Covaris E210 sonicator, followed by end repair and phosphorylation. Fragments ranging from 100 to 500 bp were selected for sequencing using an automated solid-phase reversible immobilization selection system. The addition of 3' terminal adenine was made to the fragments, followed by adaptor sequence ligation. Genome sequencing of the libraries was done using an Illumina HiSeq 2500, generating paired-end 100-bp reads. Samples were applied to a 25-Gb 2×100 channel that gave 1 Gb of sequence information per sample (500× coverage). The sequences were mapped to the *M. sedula* DSM 5348 reference genome (GenBank accession no. CP000682.1) using Bowtie2 (version 2.1.0) and SAMtools (version 1.0).

Nucleotide sequence accession numbers. The genome sequences of these evolved arsenate-resistant isolates of *M. sedula* DSM 5348, named ARS120-1 and ARS120-2, and *M. sedula* strain CuR1 (8), named ARS50-1 and ARS50-2, have been deposited in GenBank under accession numbers CP012174, CP012175, CP012172, and CP012173, respectively.

ACKNOWLEDGMENTS

This work was supported by the Department of Energy-Joint Genome Institute (DOE-JGI) under the Community Sequencing Program (CSP) (proposal ID 1515, project IDs 1036419, 1036422, 1036443, and 1036446), and the University of Nebraska Cell Development Facility.

REFERENCES

- Huber G, Spinnler C, Gambacorta A, Stetter KO. 1989. Metallosphaera sedula gen. and sp. nov. represents a new genus of aerobic, metalmobilizing, thermoacidophilic archaebacteria. Syst Appl Microbiol 12: 38–47. http://dx.doi.org/10.1016/S0723-2020(89)80038-4.
- Auernik KŠ, Maezato Y, Blum PH, Kelly RM. 2008. The genome sequence of the metal-mobilizing, extremely thermoacidophilic archaeon *Metallosphaera sedula* provides insights into bioleaching-associated metabolism. Appl Environ Microbiol 74:682–692. http://dx.doi.org/10.1128/ AEM.02019-07.
- Brierley CL, Brierley JA. 2013. Progress in bioleaching: part B: applications of microbial processes by the minerals industries. Appl Microbiol Biotechnol 97:7543–7552. http://dx.doi.org/10.1007/s00253-013-5095-3.
- Petersen J, Dixon DG. 2002. Thermophilic heap leaching of a chalcopyrite concentrate. Miner Eng 15:777–785. http://dx.doi.org/10.1016/S0892 -6875(02)00092-4.

- Watling HR, Watkinb EL, Ralphe DE. 2010. The resilience and versatility of acidophiles that contribute to the bio-assisted extraction of metals from mineral sulphides. Environ Technol 31:915–933. http://dx.doi.org/ 10.1080/09593331003646646.
- Kotze AA, Tuffin IM, Deane SM, Rawlings DE. 2006. Cloning and characterization of the chromosomal arsenic resistance genes from *Acidithiobacillus caldus* and enhanced arsenic resistance on conjugal transfer of *ars* genes located on transposon TnAtcArs. Microbiology 152:3551–3560. http://dx.doi.org/10.1099/mic.0.29247-0.
- 7. Li B, Lin J, Mi S, Lin J. 2010. Arsenic resistance operon structure in

Leptospirillum ferriphilum and proteomic response to arsenic stress. Bioresour Technol 101:9811-9814. http://dx.doi.org/10.1016/j.biortech.2010.07.043.

- Maezato Y, Johnson T, McCarthy S, Dana K, Blum P. 2012. Metal resistance and lithoautotrophy in the extreme thermoacidophile *Metallosphaera sedula*. J Bacteriol 194:6856–6863. http://dx.doi.org/10.1128/ JB.01413-12.
- 9. Maezato Y, Dana K, Blum P. 2011. Engineering thermoacidophilic archaea using linear DNA recombination. Methods Mol Biol 765:435–445. http://dx.doi.org/10.1007/978-1-61779-197-0_26.