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Susceptibility of Several Citrus Relatives to Satsuma Dwarf Virus

T. Iwanami, M. Omura, and H. Ieki

ABSTRACT. The ability of satsuma dwarf virus (SDV) to infect several citrus relatives was investigated. The plants examined were: Fortunella polyandra, Clymenia polyandra, Microcitrus australis, Eremocitrus glauca, Atalantia monophylla, Severinia buxifolia, Feroniella lucida, Swinglea glutinosa, and Aegle marmelos. The scions of these plants were grafted on rough lemon rootstocks which were infected with SDV by graft-inoculation. ELISA and indexing on sesame showed that all inoculated plants were infected with SDV. However, when the plants were maintained at a low temperature (22 C day, 18 C night), SDV was not detected from S. glutinosa by ELISA. Furthermore, the titer of citrus tristeza virus was lower in S. glutinosa than in rough lemon at low and high (30 C day, 24 C night) temperatures. The low virus titer of citrus tristeza virus (CTV) in S. glutinosa under these conditions was confirmed also by dot-immunobinding assay. These results suggest that S. glutinosa is resistant to SDV at a low temperature and to CTV at both low and high temperatures.

Since the discovery of satsuma dwarf, efforts have been made to find a cultivar resistant to this formidable virus disease. It was shown in early studies that many citrus cultivars other than satsuma mandarin do not show typical satsuma dwarf symptoms (20). These cultivars are now recognized to be infected with satsuma dwarf virus (SDV) without showing typical symptoms, rather than being resistant to infection. Some strains of SDV induce severe symptoms on sweet orange (5). Miyakawa (13) reported that some citrus relatives were infected with SDV. Furthermore, China laurestine (Viburnum odoratissium Ker.), which is commonly planted as a windbreak in Japan, was shown to be infected with SDV (11) and this suggested a relatively wide host range of SDV among woody plants.

In this study, we investigated on the ability of SDV to infect several more citrus relatives which were previously unexamined, hoping that we could find some plants that are immune to SDV, just as *Poncirus trifoliata* is immune to citrus tristeza virus (CTV)(19).

MATERIALS AND METHODS

Plant materials and growing conditions. Tests were conducted in an air-conditioned greenhouse with ambient temperatures of 22-30 C/18-24 C

(day/night). Rough lemon rootstocks were used throughout the tests. Scions of citrus relatives (see Table 1) were obtained from the collection of the gene bank of MAFF, Japan. Two replicates were prepared per species. Plants were grown following the methods described by Koizumi and Kano (11).

Virus and inoculation. SDV and a closely related virus, citrus mosaic virus (CiMV, 18) were used as inoculum. The source of SDV, S-58 was isolated from satsuma mandarin showing typical satsuma dwarf symptoms. The source of CiMV, Ci-968 was isolated from satsuma mandarin with mosaic symptoms on fruit. We consider both to be strains of SDV. For comparative study, CTV isolate 1595, which causes severe stem pitting (7,8) was used. Blind buds of rough lemon which contains one of these viruses were side-grafted onto the rootstocks.

Virus indexing. Double sandwich enzyme-linked immunosorbent assay (DAS-ELISA) and dot-immunobinding assay (DIBA) using polyclonal antibodies were conducted for the detection of SDV and CTV. Virus was extracted from the young tender leaves and the midrib of the young leaves for SDV and CTV respectively. Polyclonal antibodies against S-58, Ci-968, and 1595 had been raised in rabbits previously in our laboratory. DAS-ELISA were conducted essentially as described earlier (1, 15). For DIBA, the

TABLE 1
ASSAY BY DAS-ELISA AND BY INOCULATION TO SESAME ON SEVERAL CITRUS RELATIVES GRAFT-INOCULATED WITH SATSUMA DWARF VIRUS

Plants	Inoculum	DAS-ELISA	Sesame indexing	
Atalantia monophylla-1	S-58	1.223		
Atalantia monophylla-2	S-58	1.262	25/25	
Fortunella polyandra-1	S-58	1.343	28/28	
Fortunella polyandra-2	S-58	1.299	20/21	
Clymenia polyandra-1	S-58	1.320	16/28	
Clymenia polyandra-2	S-58	1.311	18/21	
Microcitrus australis-1	Ci-968	0.960	19/28	
Microcitrus australis-2	Ci-968	0.982	21/28	
Severinia buxifolia-1	Ci-968	0.920	17/28	
Severinia buxifolia-2	Ci-968	0.954	20/18	
Eremocitrus glauca-2	Ci-968	0.952	12/21	
Eremocitrus glauca-2	Ci-968	0.960	14/21	
Swinglea glutinosa-1	S-58	0.668	2/21	
Swinglea glutinosa-2	Ci-968	0.748	2/21	
Feroniella lucida-1	Ci-968	0.962	10/21	
Feroniella lucida-2	Ci-968	0.986	15/21	
Aegle marmelos-1	S-58	1.176	10/21	
Aegle marmelos-2	S-58	1.124	12/21	
Rough lemon-1	S-58	1.324	25/25	
Rough lemon-2	Ci-968	0.980	15/21	
Healthy rough lemon		0.002	0/21	

²Absorbance at 415 nm after 2 hr of incubation in the presence of substrate. DAS-ELISA was conducted using polyclonal antibodies to SDV isolate S-58 for the detection of S-58 and polyclonal antibodies to CiMV isolate Ci-968 for the detection of Ci-968. Values are the average of four wells.

^yNumber of sesame plants which expressed symptoms.

*Number of sesame plants inoculated.

Immun-Blot Assay Kit (BIO-RAD, goat anti-rabbit IgG HRP conjugate) was used. Biological indexing on sesame (10) was also conducted for SDV. All these virus tests were conducted at least six months after graft-inoculation.

RESULTS

Screening of plants for resistance to SDV. The initial screening of the citrus relatives revealed that none of the plants tested was immune to SDV. The results of DAS-ELISA and sesame indexing are summarized in Table 1. Although it became clear that all the tested plants were infected with SDV, the relatively low ELISA values and lower number of positive plants in sesame indexing of Swinglea glutinosa were noteworthy. Further investigation was conducted to examine virus titer of SDV in S. glutinosa.

Virus titer of SDV in Swinglea glutinosa at different ambient temperatures. ELISA values of SDV in S. glutinosa were significantly lower than those in SDV-infected rough lemon and Poncirus trifoliata at three temperature conditions (30 C/24 C, 28 C/22 C, 22 C/18 C, day/night). The ELISA values of S. glutinosa were very low at a low temperature condition (22 C/18 C), while P. trifoliata and rough lemon showed high ELISA values (Table 2).

Virus titer of CTV in Swinglea glutinosa at different ambient temperatures. ELISA values of CTV in S. glutinosa were also significantly lower than those of CTV-infected rough lemon at the three examined temperature conditions. At low (22 C/18 C) and high (30 C/24 C) temperatures, the ELISA values of CTV-infected S. alutinosa were as low as those of healthy controls and of P. trifoliata, which is immune to CTV, while rough lemon showed high ELISA values (Table 2). The ELISA of S. glutinosa was significantly higher than those of the healthy control, but lower than those of rough

TABLE 2
DAS-ELISA OF SDV AND CTV-INOCULATED PLANTS KEPT AT DIFFERENT AMBIENT TEMPERATURES

Plants	SDV inoculation			CTV inoculation		
	30/24C	Ambient temperatures 28/22C	22/18C	30/24C	Ambient temperatures 28/22C	22/18C
Poncirus trifoliata	1.148(0.018)	1.173(0.037)	1.020(0.039)	0.010(0.009)	0.004(0.001)	0.058(0.005)
Roughlemon	1.112(0.001)	1.150(0.026)	0.846(0.069)	1.205(0.005)	1.246(0.006)	1.138(0.030)
Swinglea glutinosa	0.590(0.060)	0.688(0.019)	0.193(0.007)	0.037(0.009)	0.492(0.098)	0.043(0.020)
Healthyroughlemon	0.009(0.002)	0.007(0.002)	0.017(0.003)	0.001(0.001)	0.004(0.001)	0.018(0.006)

²Plants were graft-inoculated with either SDV (S-58) or CTV (1595) (see text) and grown in air-conditioned greenhouses with controlled ambient temperatures (day/night). DAS-ELISA were carried out using anti-S-58 or anti-CTV-1595 polyclonal antibodies. Values are the average of OD415 readings of four wells X three repetitions. Standard deviation is indicated in parentheses.

lemon at 28 C/22 C. The low virus titer shown by DAS-ELISA was further confirmed by DIBA (Fig. 1).

DISCUSSION

The initial screening of citrus relatives did not reveal plants which were immune to SDV or to the closely related CiMV. The plants tested in this study, in addition to the ones tested in an earlier study (13) include most of the citrus relatives which are graft-compatible with citrus. These results indicate that it seems unlikely that graft-compatible citrus relatives which are immune to SDV will be found.

The results of DAS-ELISA showed that *S. glutinosa* is resistant to SDV at a low temperature (22 C/18 C, day/

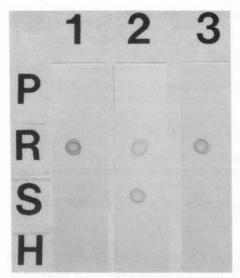


Fig. 1. Reaction of CTV-infected plants grown at different temperatures to CTV polyclonal antibodies in dot-immunobinding assay. Lane 1: 30 C/24 C, Lane 2: 28 C/22 C, Lane 3: 22 C/18 C (day/night). P: Poncirus trifoliata, R: rough lemon, S: Swinglea glutinosa, H: healthy rough lemon.

night). As shown previously, temperature is a significant factor which influences the expression of genes for virus resistance (3, 4, 9). It is probable that *S. glutinosa* has temperature-dependent resistant gene(s) to SDV.

S. glutinosa makes a good rootstock for citrus if planted in tropical and subtropical areas (2, 6, 16). This means that S. glutinosa is not suitable as a rootstock in Japan, where soil temperature in winter is relatively low. However, it now seems possible to obtain a somatic hybrid between S. glutinosa and cold-resistant citrus or Poncirus. Recently, the regeneration of intergeneric somatic hybrids between citrus and S. glutinosa utilizing electrical fusion was reported (17).

S. glutinosa was also resistant to CTV at low and high temperatures. Müller and Garnsey (14) reported that CTV was not detected in S. glutinosa by DAS-ELISA after graft-inoculation. We speculate that the tissue used for DAS-ELISA in their study was collected from S. glutinosa plants which were grown in the green-house with relative low or high ambient temperatures.

It is noteworthy that *S. glutinosa* was resistant to both SDV and CTV at low temperatures. It is probable that *S. glutinosa* is resistant to other citrus viruses, such as citrus tatter leaf virus and citrus vein enation virus, at a low temperature. Investigations are now in progress in our laboratory to confirm this hypothesis.

It is interesting that CTV does not replicate well both at low and high temperatures in *S. glutinosa*. The mechanism for CTV resistance in *S. glutinosa* might be different from that of SDV, where virus titer is very low only at a low temperature.

LITERATURE CITED

1. Clark, M. F. and A. N. Adams

1977. Characteristics of the microplate method of enzyme-like immunosorbent assay for the detection of plant viruses. J. Gen. Virol. 34: 475-483.

2. Frank D. V.

1957. Trials with Swinglea glutinosa (Blanco) Merr. as a rootstock for citrus. Proc. Fla. Hort. Soc. 70: 406-307.

3. Fraser, R. S. S. and S. A. R. Laughlin

1982. Effects of temperature on the *Tm-1* gene for resistance to tobacco mosaic virus in tomato. Physiol. Plant Path. 20: 109-117.

4. Iwai, H., Y. Horikawa, G. Figueirede, and C. Hiruki

1992. A recessive resistance gene of alfalfa to alfalfa mosaic virus and impact of high ambient temperature on virus resistance. Plant Pathology 41: 69-75.

5. Iwanami, T., T. Kano, M. Koizumi, and Y. Watanabe

1991. Leaf blotch and stem mosaic symptoms in sweet orange (Citrus siensis Osbeck) caused by a strain of satsuma dwarf virus (SDV). Bull. Fruit Trees Res. Stn. 21: 75-83.

Jones, D. T.

1990. A background for the utilization of citrus genetic resources in southeast Asia. 1: Classification of the Aurantioideae, p. 31-37. *In* Proc. 4th Int. Asia Pacific Conf. Citrus Rehab. FAO, Rome.

7. Kano, T., S. M. Garnsey, M. Koizumi, and T. A. Permar

1991. Serological diversity of field sources of citrus tristeza virus (CTV) in Japan, p. 51-55. *In*: Proc. 11th Conf. IOCV. IOCV, Riverside.

8. Kano, T. and M. Koizumi

1991. Separation of citrus tristeza virus (CTV) serotypes through aphid transmission, p. 82-85. *In*: Proc. 11th Conf. IOCV, IOCV, Riverside.

9. Kassanis B. and D. MacCarthy

1967. The quality of virus as affected by the ambient temperature. J. Gen. Virol. 1:425-440.

10. Kishi, K. and S. Tanaka

1964. Studies on the indicator plants for citrus viruses. 2. Mechanical transmission of the virus causing satsuma dwarf to sesame (Sesamum indicum L.). Ann. Phytopath. Soc. Japan 29: 142-148.

11. Koizumi, M. and T. Kano

1991. Improvement of citrus nursery systems used for indexing of viruses. Bull. Fruit Tree Res. Stn. 20: 79-91.

12. Koizumi, M., T. Kano, H. Ieki, and H. Mae

1988. China laurestine: a symptomless carrier of satsuma dwarf virus which accelerates natural transmission in the fields, p. 348-352. *In*: Proc. 10th Conf. IOCV. IOCV, Riverside.

13. Miyakawa, T.

1969. Susceptibility of *Citrus* spp. and other related plants to satsuma dwarf virus (SDV). Ann. Phytopath, Soc. Japan 35: 224-233.

14. Müller, G. W. and S. M. Garnsey

1984. Susceptibility of citrus varieties, species, citrus relatives, and non-rutaceous plants to slash-cut mechanical inoculation with citrus tristeza virus (CTV), p. 33-40. *In*: Proc. 9th Conf. IOCV. IOCV, Riverside.

15. Rocha-Pena, M. A., R. F. Lee, and C. L. Niblett

1991. Development of a dot-immunobinding assay for detection of citrus tristeza virus. J. Virol. Methods 34: 297-309.

6. Swingle, W. T. and P. C. Reece

1967. The botany of citrus and its wild relatives, Swinglea, p. 403-406. In: W. Rehther, H. J. Webber, and L. D. Bathelor (eds.). The Citrus Industry Vol. 1. Univ. Calif., Calif., Riverside.

17. Takayanagi, Rika, T. Hidaka, and M. Omura

1992. Regeneration of intergeneric somatic hybrids by electrical fusion between *Citrus* and its wild relatives: Mexican lime (*Citrus aurantifolia*) and Java feroniella (*Feroniella lucida*) or Tabog (*Swinglea glutinosa*). J. Japan. Soc. Hort. Sci. 60: 799-804.

18. Tanaka, H. and S. Yamada

1969. Evidence for a relationship among the viruses of satsuma dwarf, citrus mosaic, navel infectious mottling, natsudaidai dwarf, citrus variegation, and citrus crinkly leaf, p. 71-76. *In*: Proc. 5th Conf. IOCV. Univ. Florida, Press, Gainesville.

19. Tanaka, H., S. Yamada, and J. Nakanishi

1971 Approach to eliminating tristeza virus from citrus trees by using trifoliate orange seedlings. Bull. Hort. Res. Stn., Japan, Ser. B 11: 157-165.

20. Yamada, S. and K. Sawamura

1952. Studies on the dwarf disease of satsuma orange, Citrus unshiu Marcovitch. (preliminary report) Bull. Hort. Div. Tokai-Kinki Agr. Exp. Stn. 1: 61-71.