

UC Irvine

UC Irvine Previously Published Works

Title

GTP energy dependence of endocytosis and autophagy in the aging brain and Alzheimer's disease

Permalink

<https://escholarship.org/uc/item/9780n3d6>

Journal

GeroScience, 45(2)

ISSN

2509-2715

Authors

Martínez, Ricardo A Santana
Pinky, Priyanka D
Harlan, Benjamin A
[et al.](#)

Publication Date

2023-04-01

DOI

10.1007/s11357-022-00717-x

Peer reviewed



GTP energy dependence of endocytosis and autophagy in the aging brain and Alzheimer's disease

Ricardo A. Santana Martínez ·
Priyanka D. Pinky · Benjamin A. Harlan ·
Gregory J. Brewer 

Received: 26 September 2022 / Accepted: 15 December 2022 / Published online: 9 January 2023
© The Author(s) 2023

Abstract Increased interest in the aging and Alzheimer's disease (AD)-related impairments in autophagy in the brain raise important questions about regulation and treatment. Since many steps in endocytosis and autophagy depend on GTPases, new measures of cellular GTP levels are needed to evaluate energy regulation in aging and AD. The recent development of ratiometric GTP sensors (GEVALS) and findings that GTP levels are not homogenous inside cells raise new issues of regulation of GTPases by the local availability of GTP. In this review, we highlight the metabolism of GTP in relation to the Rab GTPases involved in formation of early endosomes, late endosomes, and

lysosomal transport to execute the autophagic degradation of damaged cargo. Specific GTPases control macroautophagy (mitophagy), microautophagy, and chaperone-mediated autophagy (CMA). By inference, local GTP levels would control autophagy, if not in excess. Additional levels of control are imposed by the redox state of the cell, including thioredoxin involvement. Throughout this review, we emphasize the age-related changes that could contribute to deficits in GTP and AD. We conclude with prospects for boosting GTP levels and reversing age-related oxidative redox shift to restore autophagy. Therefore, GTP levels could regulate the numerous GTPases involved in endocytosis, autophagy, and vesicular trafficking. In aging, metabolic adaptation to a sedentary lifestyle could impair mitochondrial function generating less GTP and redox energy for healthy management of amyloid and tau proteostasis, synaptic function, and inflammation.

R. A. S. Martínez · P. D. Pinky · B. A. Harlan ·
G. J. Brewer (✉)
Department of Biomedical Engineering, University
of California Irvine, Irvine, CA 92697, USA
e-mail: gjbrewer@uci.edu

R. A. S. Martínez
e-mail: rasanta1@uci.edu

P. D. Pinky
e-mail: pdp0014@auburn.edu

B. A. Harlan
e-mail: jib5183@yahoo.com

G. J. Brewer
Center for Neurobiology of Learning and Memory,
University of California Irvine, Irvine, CA 92697, USA

G. J. Brewer
MIND Institute, University of California Irvine, Irvine,
CA 92697, USA

Keywords GTP · Energetics · Autophagy ·
Mitophagy · Endocytosis · Lysosomes · Alzheimer's ·
Aging

GTP, the other energy currency, in aging and AD

Synthesis of the high-energy molecules ATP and GTP both rely on the redox state of NAD⁺/NADH and their sufficient concentrations. ATP synthesis from glycolysis is primarily powered by the

oxidative power of NAD⁺ acting on reducing sugars. ATP synthesis by oxidative phosphorylation in mitochondria is powered by NADH generation in the TCA cycle for reduction of pyruvate, glycerol, or fatty acids with oxygen as the terminal electron acceptor in the electron transport chain. Our label-free studies in live rat or mouse hippocampal neurons indicate an age-related depletion of NAD and NADH, that could impair synthesis of ATP and GTP [1–4]. This depletion was further exacerbated in neurons from mice carrying transgenes for human beta amyloid and tau. Further, postmortem analysis of TCA cycle enzymes indicated up to 40% impairment in the TCA cycle enzymes from AD brains compared to age-matched controls [5]. Thus, age-related energetic depletion of NAD⁺ and NADH on top of AD-related mitochondrial impairment in the activity of TCA enzymes could locally affect ATP and GTP levels, even if their bulk concentrations were minimally affected.

The energy available from GTP hydrolysis is the same as ATP hydrolysis, but GTP is utilized for different purposes than ATP due to the selectivity of specific enzymes. GTP concentrations in cells are on average tenfold lower than the millimolar ATP. GTP is a major regulator of multiple energy-dependent cellular processes of protein synthesis and vesicular trafficking involving endocytosis and autophagy. The estimated total cellular GTP concentration in mammalian cells is in the range of 250–700 μM [6], but free GTP in cancer cell protrusions is closer to 30 μM [7]. Major proteins powered by GTP include dynamins for membrane fission and fusion, the small regulatory GTPases, and microtubules as detailed below. Beside de novo synthesis from inosine by IDMPH2 and xanthosine by GMPS, cellular sources of GTP are primarily localized nucleoside diphosphate kinases (NDPKs) from the non-metastatic genes (NME; sometimes called NM23) that convert ATP to GTP [8]. There are at least ten members of the NME gene family but only the first four have nucleoside diphosphate kinase activity. According to the Allen Brain Atlas, NME1 through 4 are strongly and selectively expressed in the mouse and human hippocampus where memory is impaired in AD. NME1–3 are cytoplasmic, while NME4 is mitochondrial. Since both distinct processes of endocytosis and autophagy are affected in aging and AD, we posit that alterations upstream affect both, possibly the age and

AD-related consequence of limiting GTP levels on these amyloid-critical functions.

Measurement of GTP levels

Although GTP regulates several crucial processes for cellular survival, measuring GTP levels in the live cell under physiological and pathological conditions is difficult because of high turnover and existence of both a protein-bound and free state. Bianchi-Smiraglia et al. [9] described a novel genetically encoded GTP sensor (GEVAL) based on yellow fluorescent protein that detects ratiometric changes in GTP-free and GTP-bound concentration in vitro and in vivo. Recently, the same group reported a mechanism for regulation of GTPase Rac1 in cell invasion by a human melanoma cell line driven by local GTP production [7] and reviewed their methodology [10]. As we will see in this review, GTP plays a crucial role during endocytosis and autophagy related to amyloid processing. Currently, we are studying the age-related GTP changes in processing of A β in primary cultures of hippocampal neurons from the triple transgenic 3xTg-AD mouse and its effect on alterations in autophagy. Figure 1 shows preliminary results of GEVAL530 transfection into primary adult neurons from this AD mouse model compared to non-transgenic mouse neurons. As observed by Bianchi-Smiraglia et al. [7], we see a non-uniform distribution of GTP in the processes and edges of the soma when measuring free GTP (Fig. 1Aa), with lower free GTP levels in the 3xTg-AD neurons (Fig. 1Ac). As detailed in Section 10, pretreatment of these neurons with an NAD⁺-precursor, nicotinamide, raised the free GTP levels (Fig. 1Ab, d). The 530- μM Kd for GEVAL530 used in these experiments suggests that the local free GTP concentration is within a few-fold of 530 μM . In Fig. 1B, we examined the bound GTP that appears to be localized in vesicles, and increased by nicotinamide (Fig. 1Bf, h), especially in the 3xTg-AD neurons. Preliminary evidence indicates that neurons treated with NME1 siRNA lowers their free GTP levels (Santana Martinez, unpublished). Further studies with adult neurons across the age-spectrum will determine the age-dependence of GTP deficits and the ability to remediate them with NAD⁺-precursors. Given the essential role of GTP in vesicular trafficking processes of endocytosis and autophagy and this

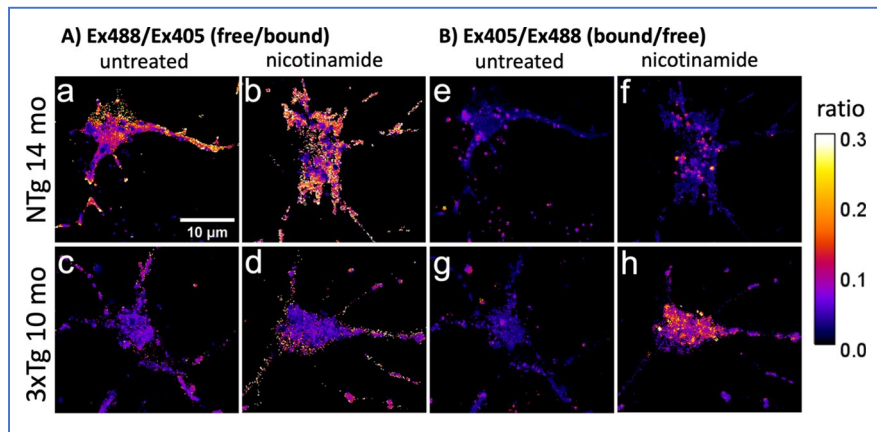


Fig. 1 Ratiometric GTP-measurements in primary mouse hippocampal neurons from middle-age mice transfected with GEVAL530 sensor. False-colored pixel-by-pixel ratiometric images of neurons show non-uniform distributions of GTP. (a) Excess free-GTP/bound-GTP of middle-age (14 mo.) non-transgenic (NTg) at edges and apical dendrite compared to

(c) middle-age (10 mo.) 3xTg-AD neuron. (b, d) Increase in free GTP in neurons treated with 2 mM nicotinamide by 24 h. (e, g) Untreated neurons exhibit vesicular bound GTP that is increased in (f, h) neurons treated with 2 mM nicotinamide by 24 h

preliminary evidence for changes in GTP with AD-like genetics, the following sections provide details of these processes and the effects of aging and Alzheimer's disease on them. The result of this review highlights the need for new methods like GEVAL probes to directly measure changes in bound and free GTP and total GTP concentrations.

Endocytosis: GTP dependence in aging and AD

In the endocytosis process, the cell internalizes macromolecules and ligand-bound receptors and surface proteins including the amyloid precursor protein (APP) [11] (Fig. 2A). Of the two main endocytic pathways, plasma membrane-embedded APP uptake occurs mostly through clathrin-mediated uptake (Fig. 2B.1) rather than caveola-mediated from cholesterol-rich patches (Fig. 2B.2). However, hydrophobic A β partitions into cholesterol-rich patches of lipid rafts in the plasma membrane with subsequent uptake by the caveola pathway [12] (Fig. 2B.2). In both cases, dynamin GTPase complexes assemble at the invagination to catalyze GTP-dependent membrane curvature for final fission and release of mature vesicles from the plasma membrane [13–15]. Local GTP fueling is catalyzed by NME1 and 2 nucleotide phosphate kinases that bind to dynamin [16].

Early endosome Rab5 GTPase decision toward exocytosis and receptor recycling or transformation to late endosome Rab7 GTPase for degradation

The Rab5 GTPase is considered a marker for the early endosome [17]. Rab5 protein is present in the plasma membrane, in clathrin-coated vesicles, and early endosomes (Fig. 2). In cell and fly models of Huntington's disease, Rab5 participates in autophagosome formation to regulate autophagy and eliminate toxic mutant huntingtin [18]. Inhibition of Rab5 reduces autophagy-regulating proteases Atg5-Atg12 conjugation, resulting in decreased autophagosome formation. The Atg12-Atg5 conjugate promotes lipidation of Atg8, and lipidated Atg8 facilitates autophagosome formation and selective cargo recognition during autophagy [19].

Another contributing factor to Rab5-mediated autophagy is phosphatidylinositol-3-kinase (PI3K). PI3-kinase complex inhibits autophagy by activating the Akt/mTOR-signaling pathway [20]. The P110 β subunit of the PI3K complex regulates the catalytic activity of the Vps34 complex to promote PI3P generation [21]. This step is essential for autophagosome formation. P110 β promotes the transition from Rab5-GDP to Rab5-GTP. P110 β overexpression mitigates autophagic deficiency following activation of the macromolecular complex composed of Rab5, Vps34,

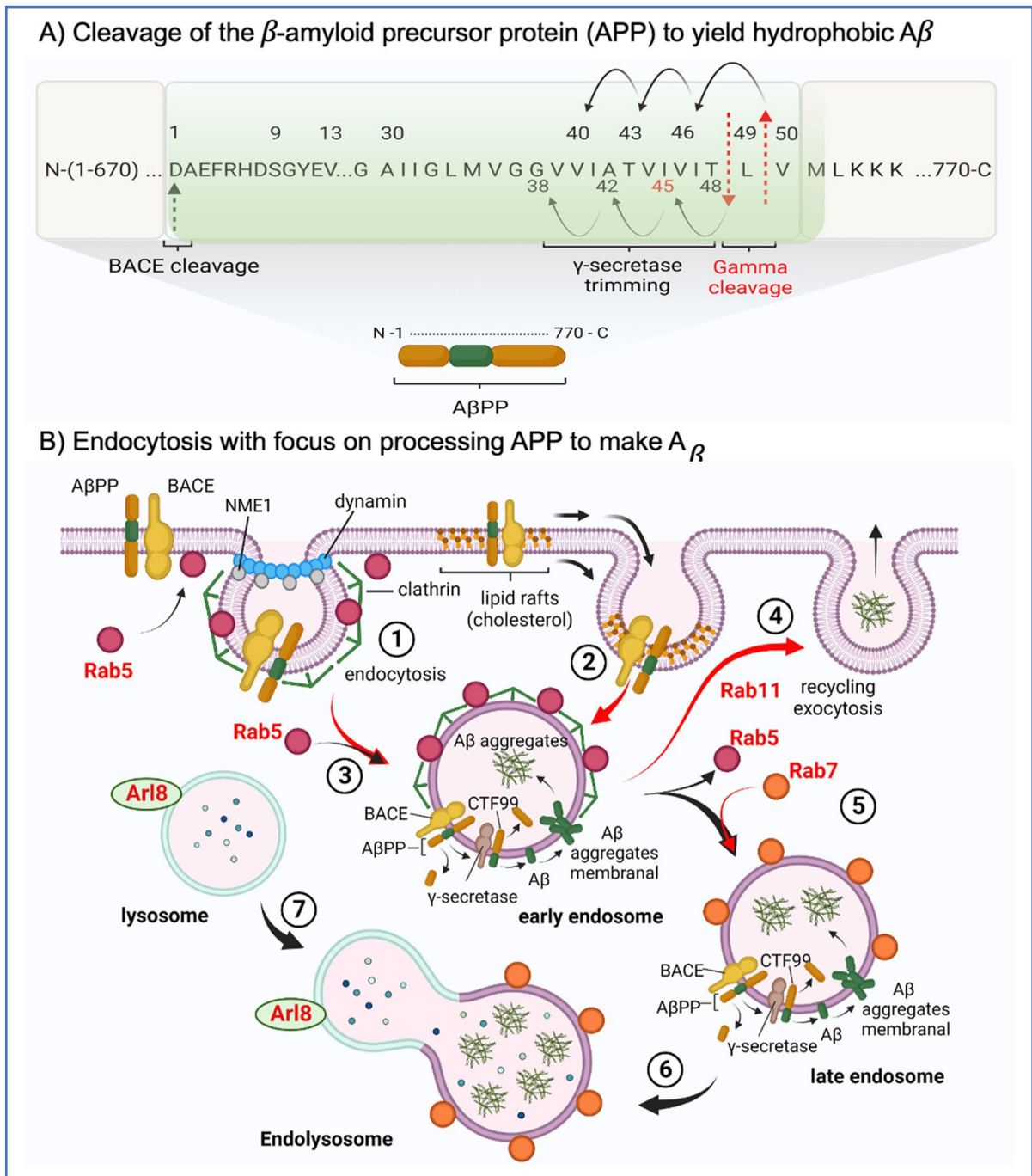


Fig. 2 Endocytosis of the amyloid precursor protein (APP). **A** The N-terminal portion of the 770 residue APP protein with green shading of the aggregation-prone $A\beta$ requiring cleavage by BACE and Gamma peptidases. **B** Endocytosis of APP. (1) Formation of the inward bud from the plasma membrane orchestrated by clathrin with endoproteases BACE and γ -secretase. Endocytosis supported by dynamin receives local GTP by NME1. (2) Microdomains enriched in cholesterol (lipid rafts) bind APP processed to $A\beta$ or $A\beta$ adsorbed from

the parenchyma. (3) Early endosome attracts the GTPase Rab5 which is recognized by another GTPase, Rab11, to mediate (4) endocytic recycling of receptors back to the plasma membrane. Alternatively, (5) Rab7 mediates segregation early into late endosomes allowing further processing of APP and accumulation of $A\beta$. (6) Late endosome fusion with (7) lysosome to degrade contents. Some aggregated $A\beta$ may be resistant to digestion, and with endosome disruption $A\beta$ may accumulate in the cytosol

and Beclin1, which in turn leads to autophagosome formation [22]. Targeting these specific proteins may elucidate therapeutic avenues to attenuate AD pathology.

Membrane-bound Rab5 is a key factor to directly promote Mon1-Ccz1 dependent Rab7 activation and Rab7-dependent membrane fusion [23]. Mon1-Ccz1 is a heterodimeric guanine nucleotide exchange factor GEF-complex which activates Rab5. Mon1-mediated displacement of the Rab5 GEF results in displacement of Rab5 by Rab7. C-Vps complex acts as a GEF for Rab7 and promotes Rab7 transiting from the GDP-bound to the GTP-bound state for Rab7 activation. Ultraviolet radiation resistance-associated gene (UVRAG) stimulates Rab7 activation by UVRAG-C-Vps interaction through GDP/GTP exchange of Rab7 [24]. Another regulatory point in the autophagic process is homotypic fusion and vacuole protein-sorting (HOPS) complex that activates the yeast vacuolar Ypt-Rab GTPase during membrane fusion [25]. One of the 6 subunits of HOPS complex is the Class C vacuolar protein sorting (C-Vps) complex which contains Vps11, Vps16, Vps18, and Vps33. Rab7 late endocytic vesicles subsequently fuse with lysosomes for cargo degradation (Fig. 2).

Changes in Rab 5 and Rab 7 endosomal trafficking in mouse models and in AD

In mouse models of AD with mutations in the amyloid precursor protein (APP) or A β -producing presenilin, the endosome–autophagosome–lysosome pathway appears dysregulated partly because of impaired acidification of lysosomes that fails to sufficiently activate protease and lipases [26]. In mouse models of AD, large perinuclear bodies containing A β aggregates label with the autophagosome marker LC3 colocalized with Rab7 and late endosomal and lysosomal components. This could be due to failed protective upregulation or pathologic impairment. In neurons cultured from 3xTg-AD mice across the age-span, we found aggregated vesicular A β to increase 30–50 fold with age, most prominently in Rab5-labeled early endosomes and mitochondria, but also within Rab7-labeled late endosomes and autophagosomes [27]. The snow-ball-like accumulation of A β 42 and A β 45 suggests that old neurons were unable to complete autophagic degradation of these longer aggregates of A β . We hypothesize that

impaired energy production in old neurons limits the energetic capacity for completion of autophagy. Compared to levels in young hippocampus and cortex, in 7–12-month-old APP/PS1 mouse hippocampus, Rab7 levels decreased along with reductions in Beclin1 and Rubicon activators and an increase in the Rubicon inhibitor [28]. In younger hippocampus and all ages in cortex, Beclin1 activates Rab7, while Rubicon inhibits the activation of Rab7 via suppression on UV radiation resistance-associated gene protein (UVRAG)-vacuolar protein sorting gene (Vps) interaction. Rab7 deficiency in yeast and fruit flies results in massive accumulation of autophagosomes [29]. Rab7 knockdown also inactivates mTORC1/S6K1, and localization of mTOR in late endosomes [30]. Interestingly, the inhibition of other stages of endocytic trafficking does not change the activity of mTORC1 suggesting that intact late endosomes are crucial for mTORC1 signaling in autophagy. This is an important reason why targeting mTOR may not slow aging or AD.

Regionally selective upregulation of Rab 5 and Rab7 proteins and mRNAs in AD, suggests selective contributions to disease pathology or an insufficient protective mechanism [31]. A β accumulates predominantly in Rab7-positive late endosomes and autophagic vacuoles of neuronal cells [32] in an age-related manner [27] causing internalization of A β and subsequent increase of Rab7 triggering neuronal degeneration [33]. Blocking the late endocytic pathway by Rab7 suppression induces A β -dependent amyloid fibril formation on the cell surface causing endosomal enlargement [34] resulting in accelerated recycling of Rab7 and endocytic trafficking of A β to lysosomes for degradation [35]. Inhibition of lysosomal proteolysis can also affect the axonal retrograde transport of autophagic organelles causing AD-like axonal dystrophy [36]. However, practical interventions to promote Rab7 mediated autophagy could delay or reverse AD progression. Examples include a shift from a sedentary state to exercise, Mediterranean diet, and energy boosting compounds like NAD precursors to create a redox shift [4] (Section 10). Another avenue is the strange discovery that guanosine monophosphate reductase 1 (NADPH-dependent GMPR1) levels are increased in AD brains that could itself lower GTP levels and raise AMP signaling [37]. Neurofibrillary tangles of tau were lowered when this activity was inhibited in AD-mice.

With progression from mild cognitive impairment to AD, both endosomal Rab5 and Rab7 expression were upregulated in hippocampal CA1 neurons as measured by transcriptional microarray [31]. Enlargement of Rab5-positive endosomes was associated with neurofibrillary tangles and amyloid deposition. Neurotrophic factors such as nerve growth factor (NGF) bind to their Trk receptors and internalize into Rab5-positive endosomes to initiate downstream signaling [38]. It is interesting to note that, the signaling endosome is retained as Rab5 early endosomes, and does not progress to Rab7 late endosomes, during their transit within the long axons [39]. This differentiates receptor-mediated signal delivery from endosome processing toward autophagy [40]. Following activation by NGF, TrkA recruits a Rab5-GAP to quickly convert GTP-Rab5 to GDP-Rab5 to keep the level of GTP-Rab5 in check, which prevents the Rab5 to Rab7 conversion resulting in inhibition of the NGF/TrkA signaling and premature degradation.

Rab5 function is compromised in early phases of AD [41]. Persistent hyperactivation of Rab5 promoted the endocytic pathway toward late endosomes, lysosome fusion, and autophagy resulting in premature degradation of the neurotrophic factor signaling and neuronal atrophy. Vps35 and Vps26, two key retromer proteins, were also reduced in AD brains [42]. Vps26 binds to SorLA which is a sorting receptor that controls APP trafficking from endosomes to the Golgi. A reduction in Vps retromer proteins leads to abnormal Vps-SorLA complex that hinders APP trafficking causing APP accumulation in the endosomes where it is subject to beta-secretase. However, mimicking APP phosphorylation at S655, within the APP 653YTSI656 basolateral motif, can enhance APP retrieval in a retromer-mediated process causing decreased APP lysosomal targeting, and decreased A β production [43]. An increase in full-length APP and APP fragment β -CTF can in turn elevate Rab5 GTPase activity inducing enlargement of early endosome [44]. Interestingly, neuronal atrophy induced by the APP β -CTFs could be rescued by a dominant-negative Rab5 mutant both in vitro [39] and in vivo [45].

The net result with aging is depletion of late endosomes, late endocytic dysfunction, and impaired lysosomal fusion. In AD, endocytosis of A β increases into enlarged Rab5 early endosomes. The role of Rab7 in AD is yet to be clearly elucidated as both

early upregulation and late downregulation of Rab7 have been observed affecting neuronal health in both directions. As emphasized by Bianchi-Smiraglia et al. [7], total cellular GTP that is measured in homogenates does not allow assessment of free GTP available locally for the Rab GTPases. Free GTP measures have not been reported yet, so their possible role in age and AD-related impairments in endocytosis is unexplored. Therefore, it will be important to measure free GTP in live cells and the effects of variations in free GTP on endocytosis as a function of age and in AD models.

Common GTPase signaling pathways in autophagy: macroautophagy (mitophagy), microautophagy, and CMA

The GTPase superfamily comprises a wide range of proteins that act as molecular switches by binding and hydrolyzing GTP molecules for tethering, docking, and fusion of vesicles to target membranes [46]. Changes in GTPase function are related to alterations in the trafficking of cargo [47]. The switch between an “active” state (GTP-bound GTPase) and an “inactive” state (GDP-bound GTPase) requires a guanine nucleotide exchange factor (GEF) and GTPase activating protein (GAP) [48] (Fig. 3), termed the Rab GEF/GAP cascade [22]. For many GTPases, the lifetime of the activated, GTP-bound state is believed to serve as a regulator in determining the activation time of a biological event such as membrane fusion and signal transduction. However, proper function of the GTPases could also be limited by a decrease in intracellular GTP levels.

Autophagosome formation involves Rab family members Rab1, Rab5, Rab7, Rab9A, Rab11, Rab23, Rab32, and Rab33B. Rab9 is required in non-canonical autophagy. Rab7, Rab8B, and Rab24 have a key role in autophagosome maturation. Rab8A and Rab25 are involved in unknown aspects of autophagy [22]. Rab11 is required for exocytosis [49]. Failing autophagy is associated with several unhealthy conditions such as metabolic stress and aggregation of proteins associated with neurodegenerative disorders including Alzheimer’s disease. Rab8b has a key role in orchestrating autophagic maturation although its downstream effector Tbk-1, which directly phosphorylates p62 at Ser-403, a crucial residue for

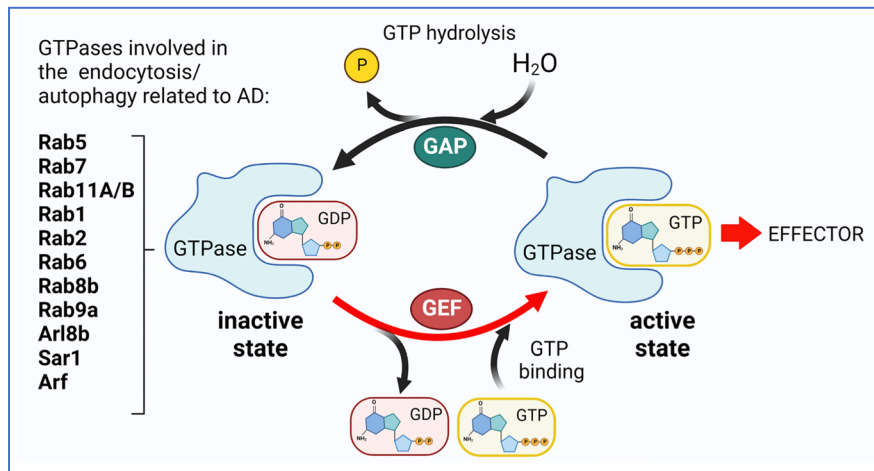


Fig. 3 Molecular switch between GTPase states. Small GTPase's active state (GTP-bound) is achieved by nucleotide exchange factor that catalyzes the exchange of GDP to GTP resulting in GTPase activation. Active GTPase interacts

with several kinds of downstream effectors to modulate their activity. GTPase activating protein (GAP) inactivates the GTP-bound proteins by boosting their activity for GTP hydrolysis. The GDP-bound form cannot bind effectors

autophagic function of p62 [50]. TBK-1 is also required for cytokinesis-induced autophagic elimination of bacteria, whereas Rab8 knockdown after induction of autophagy caused a decrease of phagosomes [50].

Since GDP is tightly bound by Rab GTPases and their intrinsic GTP hydrolysis rates are low despite their high affinity 10^{-1} – 10^{-5} μM kKm [51], the Rab GEFs catalyze the dissociation of GDP. Rab GAPs facilitate the hydrolysis of GTP. Both regulators are required to coordinate the temporal-spatial activity of Rab GTPases [52]. The activity of Rab GTPases, GEFs, and GAPs are crucial for transport and trafficking of autophagosomes for macroautophagy. Macroautophagy needs stringent control, and some Rab GAPs seem to function in overlapping pathways. For example, TBC domain containing proteins, TBC1D14 and TBC1D15, coordinate endosomal trafficking and autophagosome biogenesis. TBC1D5 is a Rab7 GAP that is recruited to mitochondria by protein FIS1 to handle Rab7 GTP hydrolysis, allowing also mitochondria to regulate the contact untethering with lysosomes [53]. Since mitochondria-lysosome contacts mark the sites of Drp1-positive mitochondrial fission events, alterations in the Rab7 GTP hydrolysis lead to both abnormal lysosomal morphology and markedly reduced the rate of mitochondrial motility [53]. TBC1D2, which affects the Rab7 GTPase and modulates autophagosome-lysosome fusion, was shown to

be activated by LRRK1 upon macroautophagy induction [54].

The protein family of small Rab GTPases control vesicle transport routes and ensures trafficking of vesicles to their appropriate target compartments. Rab GTPases interact with effector proteins such as cargo sorting complexes, motor proteins, and tethering factors, which are required for vesicle budding, transport, and fusion of various intracellular organelles. In mammalian cells, there are three primary types of autophagy: macroautophagy, microautophagy, and chaperone-mediated autophagy (CMA) (Fig. 4). Each subtype consists of different mechanisms of substrate delivery to the lysosome; however, the result is the same for all of them culminating in the delivery of cargo to the lysosome for degradation and recycling.

Macroautophagy of mitochondria is the most studied and is commonly referred as mitophagy. Mitophagy signaling is mainly controlled by the Target of Rapamycin protein complex 1 (TORC1). Following induction of autophagy, a sequestering membrane called a phagophore is formed (Fig. 4A). The phagophore encloses misfolded proteins and/or dysfunctional organelles until it is completed into an enveloped autophagosome. The autophagosome then fuses with the lysosome, transferring the cytoplasmic cargo for hydrolysis. The organelles inside the cargo are then degraded into amino acids and simple fatty acids and carbohydrates for release into

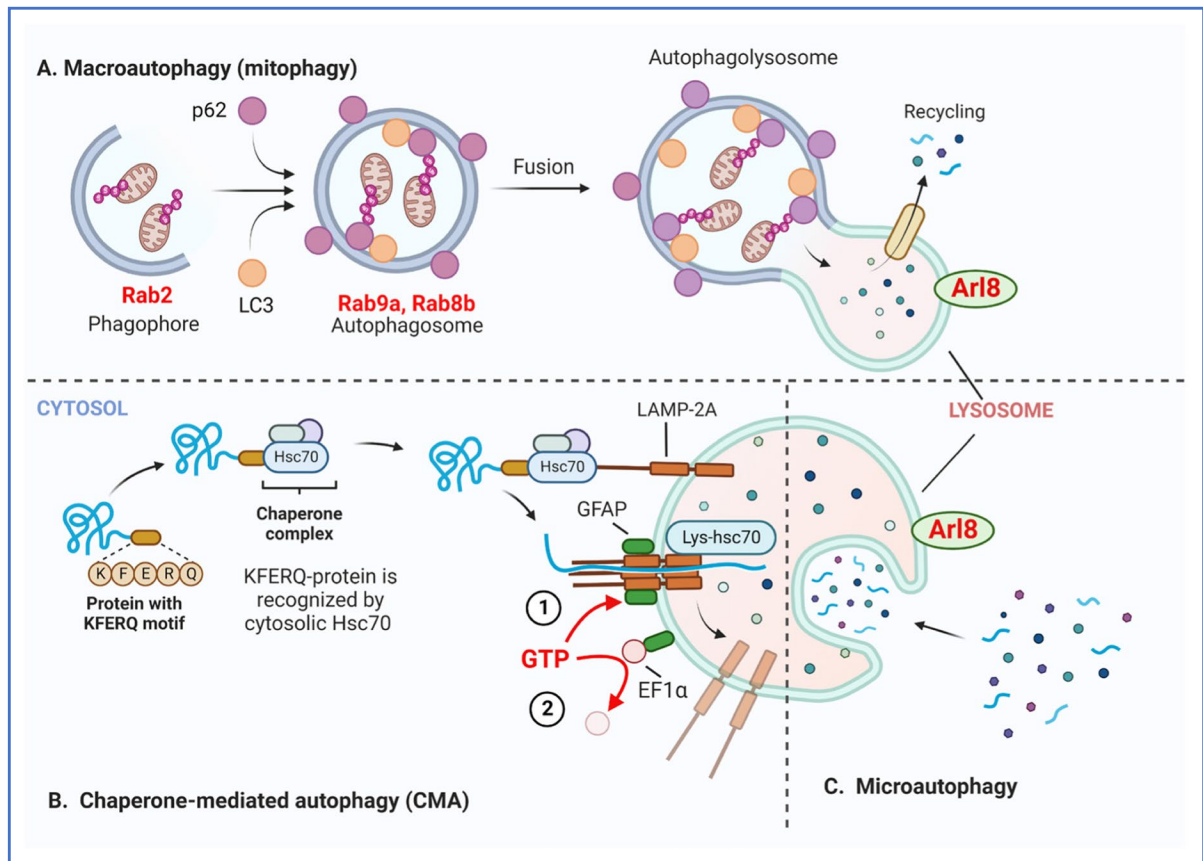


Fig. 4 Types of autophagy pathways regulated by GTP **A** In macroautophagy (mitophagy), ubiquitin (Ub)-labeled proteins recruit p62 to interact with LC3 to form autophagosomes. Rab2 participates in phagophore formation, whereas Rab8b and Rab9a participate in autophagosome maturation. Arl8 is located on the lysosome membrane and facilitates lysosomal trafficking. **B** In chaperone-mediated autophagy (CMA), substrate proteins bind to the monomeric form of LAMP-2A after recognition by a KFERQ motif by cytosolic Hsc70 chaperon complex. Unfolding of the complete substrate is required for its translocation into a multimeric complex with LAMP-2A.

There are two pools of GFAP in the membrane of lysosomes: 1, in conditions with high CMA activity, GFAP interacts with LAMP-2A to stabilize the complex required for the translocation of CMA cargo in a GTP-dependent manner; 2, GFAP interacts with EF1 α , while GTP induces the release of EF1 α from GFAP inhibiting CMA and promoting the disassembly of multimeric LAMP-2A. **C**. In microautophagy, autophagic cargo are engulfed by invaginations of the lysosomal membrane to capture content. In all of them, recycling occurs after lysosomal degradation

the cytoplasm by lysosomal/vacuolar membrane permeases and re-use in biosynthesis [55].

CMA (Fig. 4B) does not use membranous structures to sequester cargo, but instead uses chaperones to identify cargo proteins and translocate them to the lysosomal membrane, while microautophagy (Fig. 4C) utilizes invaginations or protrusions of lysosomal membrane to capture and deliver the autophagic cargo to the lysosomal membrane [56, 57].

Reactive oxygen species (ROS) partially controls autophagy. Starvation stimulates ROS (mainly H₂O₂) production in mitochondria, which appears to be necessary for autophagosome formation. In yeast, autophagy can be regulated by ROS via Atg4, through oxidation–reduction of a disulfide bond between residues Cys338 and Cys 394, which is required for a proper autophagosome biogenesis [58]. Atg4, a redox protease, acts as a conjugating enzyme cleavage C-terminus in immature Atg8 (mammalian

homologue LC3) to expose the conserved glycine residue for its subsequent linkage to phosphatidylethanolamine (PE). Further, Atg4 acts also as a deconjugation enzyme that cleaves the amide bond between Atg8 and PE, which releases it from the membrane for recycling, which is crucial for the conjugation systems essential for autophagy.

Autophagy: GTP regulation in aging and AD

Autophagic clearance of damaged cellular components or aberrant protein aggregates becomes increasingly important to handle the increased oxidative stress associated with aging and AD [59]. The bidirectional trafficking into and out of cells must be highly coordinated. Autophagy works in association with endocytosis and exocytosis, both of which contribute to turnover of damaged intracellular constituents employing lysosomal fusion and digestion. Autophagy involves tagging defective cellular organelles and protein aggregates for degradation, followed by the assembly of an autophagic structure to transport cargo for fusion with lysosomes to degrade damaged proteins and organelles for recycling amino acids and lipids or disposal. It is a highly dynamic process essential to maintain cellular homeostasis and functions. Dysregulation of autophagy has been linked to aging and pathological neurodegenerative diseases including Alzheimer's disease. Although it is well known that ATP levels decline with age under pathological conditions [60], age-related changes in GTP levels are poorly elucidated.

Autophagy induction is dependent on ADP/ATP balance. Decreased ATP production stimulates AMP-activated protein kinase (AMPK), and stimulation of AMPK inactivates mTOR. AMPK increases autophagy not only indirectly through inactivation of mTOR but also directly through phosphorylation of Unc-51-like kinase 1 (Ulk1) which is the molecular target of mTOR in the autophagic machinery [61].

GTP depletion may impact autophagy in aging and AD

Post-mortem analysis of AD patients indicates an accumulation of autophagosomes and other prelysosomal autophagic vacuoles in dystrophic neurites and synaptic terminals, which are neuropathological

hallmarks of AD [62]. Upregulation of autophagosomes in hippocampal CA1 pyramidal neurons is related to changes in expression of autophagy-related genes (ATG3, ATG5, ATG12, ULK1 and PIK3C3/VPS34) and proteins (LC3B-II and LC3B-I) at early AD stages [63]. These facts suggest that AD is associated with alterations in trafficking of autophagosomes.

Autophagy dysregulation occurs in both AD patients and animal models. Accumulation of large amounts of autophagic vacuoles in neuronal dendrites occurs in PS1/APP double transgenic mice appears even before A β plaques [64]. Similarly, immature autophagic vesicles in axons were observed in hippocampal neurons of AD mice, far before synaptic and neuronal loss (Cataldo et al., 2004 [44, 65, 66]. Tau aggregates are also degraded through the autophagy pathway [67, 68]. The wild type presenilin gene 1 (PS1) acts as a ligand of the v-ATPase V0a1 subunit regulating the distribution of v-ATPase subunits into lysosomes for acidification. Intracellular A β binds to v-ATPase and inhibits acidification [69]. Mutation in PS1 thus contributes to the dysregulation of the autophagy-lysosome degradation system [70]. The major genetic risk factor for sporadic AD, Apolipoprotein E4 (ApoE4), also contributes to induction of autophagy by lysosomal leakage [71] leading to impaired endolysosomal trafficking, disruption of synaptic homeostasis and reduced amyloid clearance. Altogether this suggests that the defective autophagy-lysosome proteolysis pathway might be responsible for the accumulation of pathogenic proteins such as A β and tau in AD. A conditional knock out of a gene needed for autophagosome formation, *Atg7^{fllox/fllox}*, crossed with APP23 transgenic mice, indicated that impaired autophagy deficiency promoted A β accumulation in CA1 and cortical pyramidal neurons, as well as a drastic reduction in the extracellular A β plaque burden and inhibition of A β secretion [72]. This observation suggests that alterations in autophagosome formation avoids proper A β processing that could lead an aberrant accumulation in the soma [27]. A β accumulation has been also observed in the cis- and trans-face of the Golgi vesicles in the late Golgi apparatus, indicating this organelle could also produce functional alterations that impair the proper formation of the phagophore [73]. Supporting this supposition, the small GTPase Rab2 connects the Golgi network to the autophagy pathway machinery [74]. Rab2 participates in the formation of phagophores by further recruiting

and activating Ulk1. Rab2 interacts with Rubcn1 and Stx17 (an autophagosomal SNARE protein) to further specify the recruitment of HOPS complex to facilitate autophagosome maturation and fusion with lysosomes [74].

Another member of Ras super family of small GTPases involved in vesicle formation is the ADP-ribosylation factor (Arf). Arf GTPase mainly participates in the budding process in the Golgi complex, recruitment of coat proteins during vesicle formation for membrane trafficking. Arf GTPase is involved in control of APP trafficking through MINTs proteins, crucial components for the fusion of synaptic vesicles. MINT proteins bind directly to Arf GTPases

and co-localize with APP-containing vesicles to regions of the Golgi/trans-Golgi network (TGN), with intracellular APP levels proportional to MINT levels [75]. The knockdown of Arf1 decreased secretion of amyloid peptides [76], suggesting the impact of failures in Arf1 function on trafficking of APP, which could converge in intracellular accumulation (Fig. 5).

Dysregulation of various Rab proteins also contributes to AD pathology. Rab1 dysregulation induces fragmentation of the Golgi apparatus, which triggers hyperphosphorylation of tau by activation of cdk5 and ERK1/2 [77, 78]. A defect in Rab6 can influence the secretion of APP into the

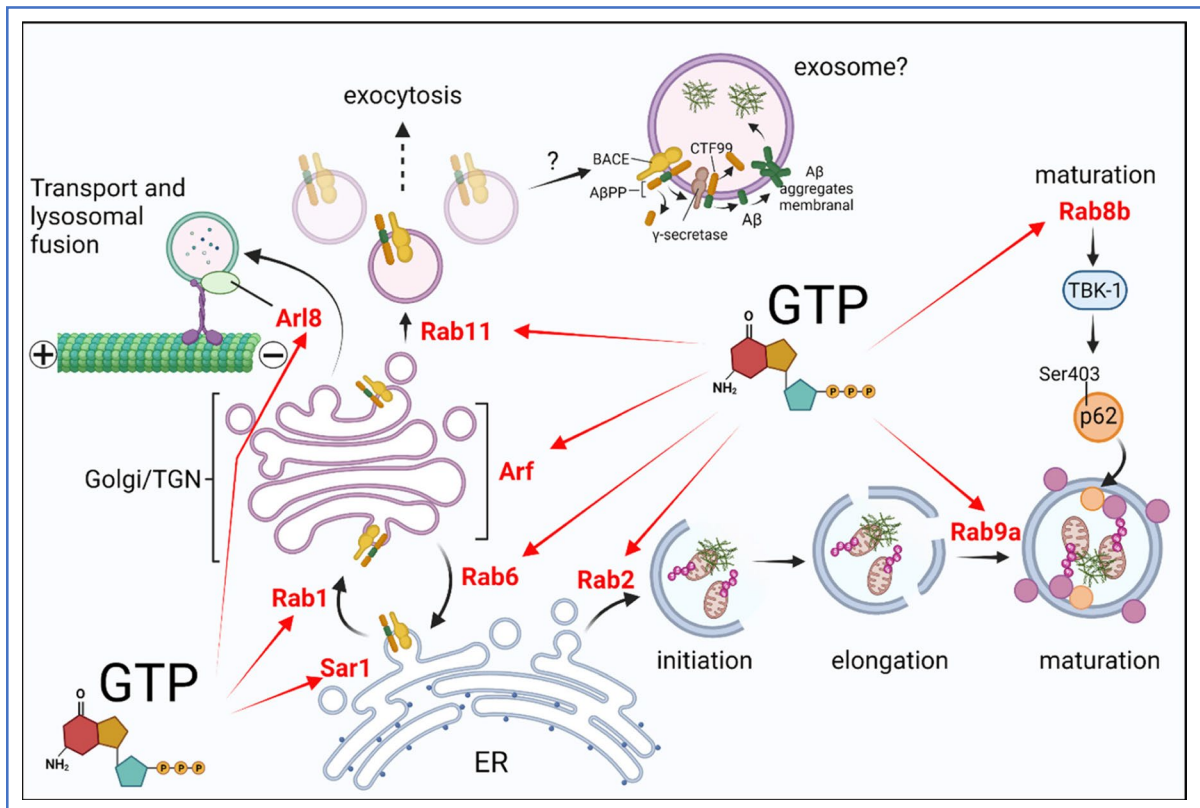


Fig. 5 Autophagy requires the participation of several small GTPases. In the initiation phase, Rab2 recruits and activates ULK1 in the phagophore formation, while Rab9a and Rab8b participate in autophagosome maturation. Arf takes part in budding in the Golgi complex, and recruitment of coat proteins during vesicle formation. Rab1 participates in the bidirectional vesicular transport route between the endoplasmic reticulum (ER) and Golgi apparatus. Sar1 GTPase is involved in the COPII-mediated transport as the exit route of the APP once its

final conformation has been reached. Rab6 resides in the TGN and participates in retrograde transport from Golgi to ER, and has been associated with the regulation of the vesicular transport and processing of APP. Rab11 controls the endosome recycling to the plasma membrane. Disruptions in vesicular export that contains APP-BACE could cause an accumulation of exosomes. Arl8 regulates the transport and lysosomal fusion via microtubules along the neuron

medium, leading to alterations in the secretion rate, which could promote changes in the anterograde trafficking of APP leading to intracellular accumulation. Supporting intracellular accumulation, an exome sequencing analysis identified Rab11A/B as a component of late-onset AD risk. Rab11 controls the endosome recycling to the plasma membrane. Silencing Rab11A/B in primary neurons isolated from APP transgenic mice reduced A β levels in the supernatants collected and analyzed by electrochemiluminescence (ECL) [79]. These data suggest that the accumulation of A β could also be generated by failures in the export of APP-containing vesicles, which could give way to a cleavage by the secretase in the vesicular membrane [27] (Fig. 1B). Since formation of the APP-containing vesicles and the formation of the phagophore involve a correct functioning of the ER-Golgi driven by small GTPases, deficits in GTP levels would cause disturbances in

the packaging of APP that could compromise its secretion and promote intracellular accumulation.

These relationships of autophagy to accumulation of intracellular A β are supported by an age-related colocalization signal between p62-directed formation of autophagosome and A β forms observed in primary cultured hippocampal neurons from adult 3xTg-AD mice [27] and in APP23-transgenic mice with Atg7 floxed mice [73]. Another interesting fact is that no signal of aggregates was observed in autophagolysosomes positive for cathepsin D [27]. These data may indicate dysfunction in earlier steps in autophagy or in lysosomal function in AD model neurons (Fig. 6) [26].

GTP regulation in CMA in AD

Instead of organelles, CMA degrades small molecules in eukaryotic cells induced by prolonged starvation

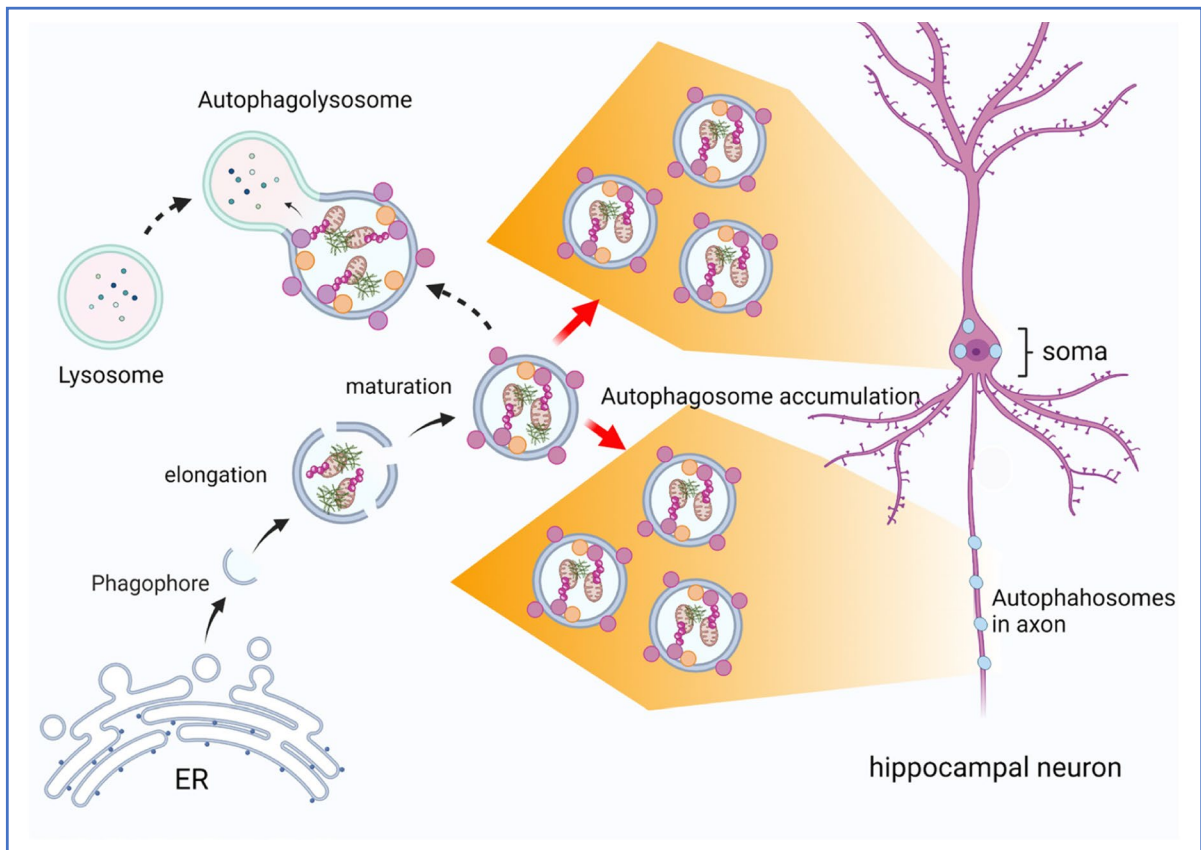


Fig. 6 Deficiency in autophagolysosome formation in AD. Mature autophagosomes (p62 and LC3 positive cells) containing A β aggregates are accumulated in the soma and axons leading to vesicular imbalance further associated with energy failure

or mild oxidative stress. CMA has been proposed to be regulated by GTP levels [80]. This regulation involves the participation of the intermediate filament glial fibrillary acidic protein (GFAP) and the elongation factor-1 alpha (EF1 α) [46, 81]. GFAP is present in two different pools at the lysosomal membrane, a portion bound to LAMP-2A and another unbound form that interacts with EF1 α . The three KFERQ motifs in the Hsc70 chaperon direct it to lysosomal membranes where it interacts with protein complex LAMP-2A and stabilizes the translocation of CMA-cargo into the lysosomal lumen. The portion of GFAP not bound to LAMP-2A contributes to GTP regulation. In the presence of GTP, EF1 α is released from GFAP at the lysosomal membrane which promotes the dissociation between GFAP and LAMP-2A, mobilizing LAMP-2A to the lipid microdomains for its degradation and subsequent CMA inhibition [81]. Failures in LAMP2A complex density on lysosomal membrane are also associated with aging, which also leads to a decrease in CMA function [82] (Fig. 4B).

Yet another form of mammalian autophagosome biogenesis operates through an enigmatic non-canonical VPS34-independent pathway. Phosphoinositides (PIs) define the membrane identity and control several membrane trafficking events. Phosphatidylinositol 5-kinase (PIKfyve) converts endosome-localized phosphatidylinositol-3-phosphate (PI(3)P) to PI(3,5)P₂, a key regulator of early to late endosome membrane trafficking [83]. Further, PIKfyve complex is also responsible for production of PI(5)P from PIs and regulates autophagosome formation. PI(5)P regulates autophagy via PI(3)P effectors (recruiting WIPI2 and DFCP1 proteins), which provide a mechanistic framework for this alternative autophagy pathway. PI(5)P is used by phosphatidylinositol 5-phosphate 4-kinase β (PI5P4K β) that regulates PI(5)P levels using GTP rather than ATP for PI(5)P phosphorylation to obtain PI(4,5)P₂ as a final product that regulates actin cytoskeleton remodeling [84]. They found PI(3,5)P₂ has low affinity for cofilin, which disassembles actin filaments and high affinity for N-WASP, which activates Arp2/3 complex to initiate actin nucleation to transport endocytic vesicles from the plasma membrane. PI5P4K β activity is proposed to reflect changes in direct proportion to physiological GTP concentration, acting as an intracellular GTP-sensor [85]. In cells lacking PI3P (low PI(3,5)P₂) with locked VPS34, the PIKfyve complex

sustains autophagy through the use of PI5P [85]. These data indicate PIKfyve has a pivotal role in the modulation of autophagy. Impaired PIKfyve function drives formation of swollen vacuoles, easily visible at low magnification in living cells [86]. The intracellular domain of APP binds the Vac14 subunit of PIKfyve complex, affecting PI(3,5)P₂ production [87]. PI(3,5)P₂ binds and activates the endolysosomal TRPML channel [88]. They found enlarged vacuoles in PI(3,5)P₂-deficient mouse fibroblasts that were suppressed by overexpression of healthy TRPML1 channel. TRPML conductivity and lysosomal acidification were impaired [89].

Dependence of mitochondrial fission and fusion on GTP

In addition to providing ATP through oxidative phosphorylation, mitochondria also provide GTP from NME4 and its nucleoside diphosphate kinase activity [8]. This mitochondrial supply of energy is essential for generation of synaptic vesicles for release at axon terminals as well as for vesicular recycling at synapses. Synaptic loss in AD could be caused by loss of bioenergetic capacity to maintain these essential processes. Therefore, the number and localization of mitochondria to synapses likely determines the energetic capacity for endocytosis and exocytosis. Mitochondrial dynamics delicately balance fission and fusion controlled by Drp1 and Fis1, Mf1, Mfn2 and Opa1 [90, 91] (Fig. 7). ATP conversion to GTP is locally controlled by localized nucleoside diphosphate kinases (NDPKs) from the NME genes 1–4 [51]. NME1 and 2 (sometimes called NM23 H1 and H2) are predominantly cytosolic, while NME3 and 4 (NM23 H3, H4) are mitochondrial. The mitochondrial NDPKs complex with specific dynamin GTPases to channel GTP directly from ATP hydrolysis [16]. The balance of mitochondria fission and fusion is sensitive to redox imbalance. Either endogenous or exogenous application of ROS activates mitochondrial fission, inducing mitochondrial fragmentation and subsequent mitochondrial dysfunction [90]. This leads to further ROS overproduction and a vicious cycle that amplifies oxidative stress and ultimately causes oxidative imbalance in AD [92]. Fission is regulated by the dynamin GTPases Drp1 and Fis1 with K_m 's around 100 μ M [51]. They polymerize and constrict tubular

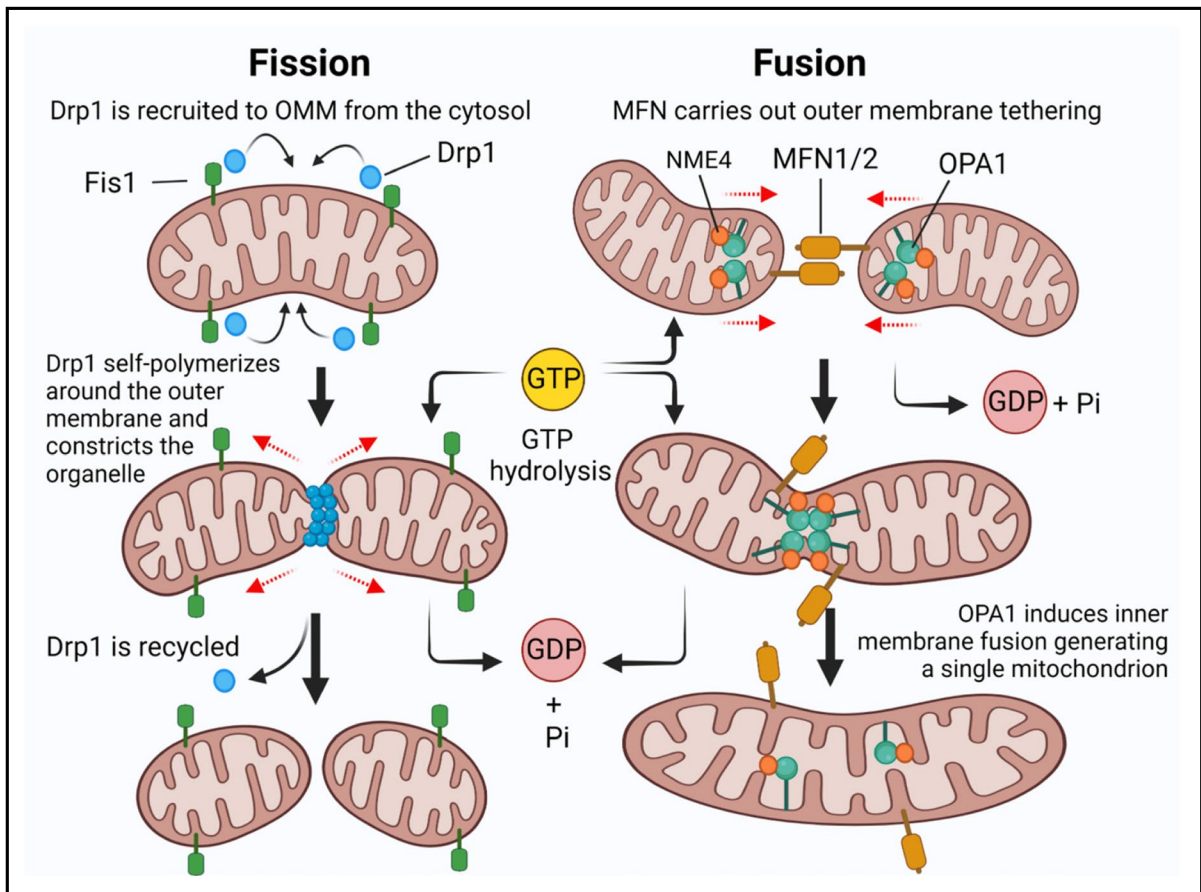


Fig. 7 GTP-dependent mitochondrial dynamic morphology of fission and fusion. Left fission panel: Dynamin-related protein-1 (Drp1) executes the mitochondrial fission by self-polymerizing around the outer mitochondrial membrane constricting and severing both membranes in a process dependent on GTP hydrolysis. Right fusion panel: Mitofusin-1 and -2

(MTF1/2) tethers the adjacent outer mitochondrial membranes in a process dependent on GTP hydrolysis. OPA1 enables inner mitochondrial membrane fusion using local GTP provided by NME4. Red dashed arrows indicate the displacement of the organelle

membranes much like endocytosis. Fis1 is localized in the outer mitochondrial membrane [90, 93, 94].

Fusion is controlled by three GTPase proteins: Opa1, located in the inner mitochondria membrane and Mfn1 and Mfn2, located in the outer mitochondrial membrane. The Opa1 GTPase with a K_m around $500 \mu\text{M}$ would require abundant levels of GTP to polymerize and mechanize mitochondrial membrane fusion into tubes [51].

Mitophagy can be activated by several stimuli such as hypoxia, energetic stress, and increased oxidized redox state. Increased oxidative stress and elevated ROS levels caused fragmentation of mitochondria and induction of DRP1 fission-dependent mitophagy

in mouse and HeLa cells. This did not result in cell death and autophagy because moderate levels of ROS were not sufficient to trigger non-selective autophagy [95]. Indeed, this mitophagy can be inhibited by N-acetyl-L-cysteine through fueling of the glutathione pool and possible action on Atg4. A decrease of the glutathione pool also induced mitophagy but not general autophagy. Conversely, the addition of a cell-permeable form of glutathione inhibited mitophagy [96]. Thus, an oxidative redox state promotes target-selective removal of dysfunctional mitochondria. This also suggests integration of redox balance and energy levels to maximize healthy mitochondrial function and control turnover of damaged mitochondria.

Impairment in fusion and fission has been implicated in AD. In a mouse model, A β interacts with fission protein Drp1, with a subsequent increase in free radical production, which further activates Drp1 and Fis1, causing excessive mitochondria fragmentation, defective transport of mitochondria to synapses, lowers synaptic ATP, and ultimately leads to synaptic dysfunction [97]. p-tau also interacts with Drp1 and enhances GTPase Drp1 enzymatic activity, leading to excessive fragmentation of mitochondria and mitochondrial dysfunction in AD [98]. A Drp1 S-nitrosylation adduct (SNO-Drp1), further stimulated its activity, and led to excessive mitochondrial fragmentation, and synapse loss [99].

Impaired lysosomal function in AD caused by age-related energy depletion

Lysosomal digestion of autophagic cargo is the last step for the completion of autophagy. Therefore, lysosomes must maintain its acidic milieu for pH-based degradation of cargo by acid-activated peptidases, lipases, nucleases, and glycosidases. For lysosomal acidification, the influx of protons is carried out by both the v-ATPase, an ATP-dependent proton pump and chloride proton antiporters, while cation efflux is mediated by transporters TPC and TRPML, which are also involved in the pH balance [100]. Presenilin-1 (PS1) regulates the distribution of v-ATPase subunits into lysosomes acting as a ligand of the v-ATPase V0a1 subunit and maintains lysosomal homeostasis via TRPML1 [69]. PS1 mutations have been linked to low lysosomal acidification, dysregulation of the autophagy-lysosome degradation system and the pathogenesis of early onset AD [26].

Arl8 (an Arf-like G protein) is a small GTPase located on lysosomes that acts as a linker between lysosomes and kinesin-1 to facilitate lysosomal trafficking along axons (Fig. 8B) [101]. Arl8b also acts as a switch to regulate the association of HOPS complex with the lysosomal membrane [102]. Disruption of Arl8b function causes abnormal accumulation of cholesterol in the membranes of lysosomes driving impaired axonal lysosome trafficking and leading to autophagic stress and axonal autophagosome accumulation [103]. Additionally, elevated Arl8b expression rescued lysosome transport into axons and autophagic stress. Overexpression of Arl8 also stimulated the

bidirectional motility of lysosomes on microtubules by binding to the kinesin-1 linker SKIP to link kinesin and power motility [104]. SKIP required Arl8 in its active GTP-bound state for binding. However, overexpression of Arl8b also caused a striking alkalization of lysosomes and movement toward the cell periphery versus a more uniform distribution of lysosomes throughout the cytoplasm [105]. A proteomic study in human tissue reported enrichment of Arl8b in amyloid plaques [106]. These data suggest that changes in the expression or functioning of Arl8 affects the fusion of the lysosome with the autophagosome or late endosome, which could have implications in autophagosome accumulation observed in AD. Since formation of secretory vesicles delivered to the plasma membrane is a GTP dependent process, requiring Arf and Rab GTPases, deficits in GTP levels may compromise proper vesicular trafficking.

Lysosome biogenesis is regulated by mTORC1 and the transcription factor EB (TFEB) creating a reversible signaling complex on the lysosome surface. mTORC1 phosphorylates TFEB on Ser211, driving thereby enabling TFEB transport to the nucleus to upregulate v-ATPase expression and other genes involved in lysosome biogenesis and autophagosome formation [107, 108]. Addition of A β to microglial cells lowers TFEB in the nucleus and impairs processing of A β [109]. This regulation by mTORC1 is in turn supported by small GTPases including Rheb and Rag, amino acid-sensing components included in the multiprotein signaling complex regulator which acts as an activator of mTORC1 [110]. Since lysosomal acidification requires ATP for vATPase function and GTP for GTPase-mediated regulation of TORC1, we hypothesize that energy depletion due to aging or AD-like pathological conditions directly impairs lysosomal function. Additionally, considering that lysosomal acidification involves a nutrient intake-mediated regulatory interaction between v-ATPase and TORC1 via TFEB, cycles of fasting and nutrient consumption could benefit lysosomal function in AD [111], while frequent sugar consumption may impair function. Impaired lysosomes could lead to accumulation of autophagosomes full of damaged mitochondria and enriched with A β -aggregates incapable of being degraded. Since autophagy carries out the replacement of damaged or aged organelles, its maintenance is affected by metabolic changes due to age. Age-related metabolic shifts and impaired lysosomes

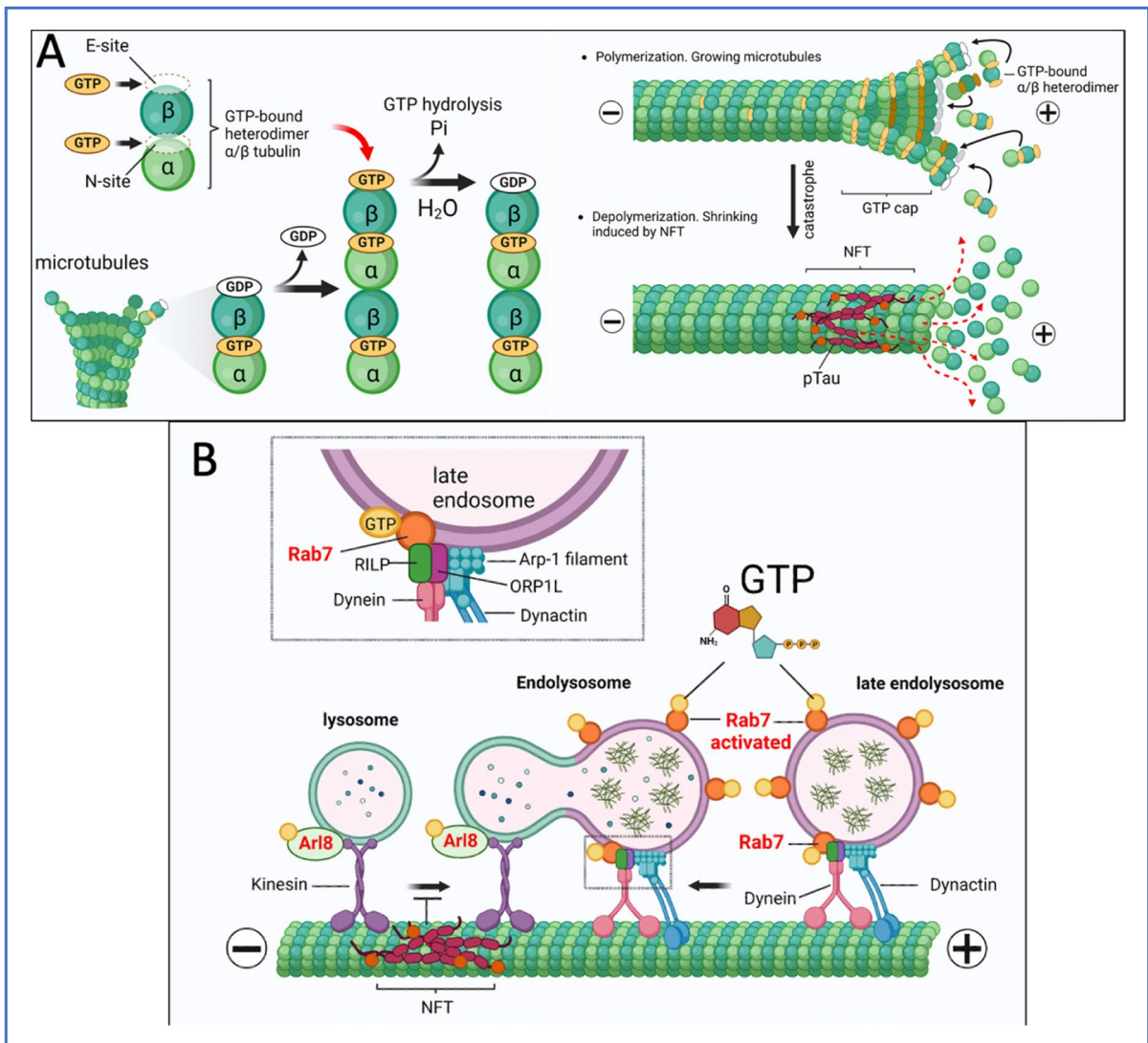


Fig. 8 Dynamic instability of microtubules associated with GTP. **A** GTP molecules bind to E-site and N-site on α/β heterodimer tubulins. GTP at the E-site is hydrolyzed to become GDP- β -tubulin and exchanged for a new GTP-bound heterodimer α/β to assemble subunits. Heterodimers are added to the growing microtubule lattice for polymerization forming a new layer of GTP-heterodimers known as a GTP-cap. Depolymerization occurs when heterodimers leave the shrinking microtubules lattice. Hyperphosphorylation of microtubule associated protein, tau promotes neurofibrillary tangle (NFT) formation and microtubule destabilization. The transition from a growing

state to a catastrophic shrinking state. **B** Endolysosome formation requires lysosomes move along microtubule tracks in the positive direction, while late endosomes move in the negative direction. The lysosomal multiprotein complex BORC (not shown) activates the small GTPase Arl8 to engage kinesin-driven plus-end transport. For minus-end transport, the Rab7-GTP-bound state recruits the Rab-interacting lysosomal protein (RILP) and the cytosolic oxysterol-binding protein-related protein 1 (ORP1L) forming the dynein-dynactin complex. NFT formation disrupts vesicular traffic along microtubules

could lead to accumulation of metabolic shifts such as energy depletion and/or oxidative redox shifts. Thus, GTP levels could be lower in low-activity sedentary lifestyles, which would lead to inefficient pathways

of GTP-dependent protein degradation with advancing age. These observations strongly suggest a relationship between alterations in the maturation process of the autophagosome and the metabolic deficiencies

that occur with the progression of the disease and ageing. Further, aberrant A β accumulation could be the consequence of upstream deficiencies in GTP that impair autophagic processing of A β and tau, possibly earlier than aggregate amyloid secretion, inflammation and plaque buildup.

GTP-dependence of tubulin assembly, vesicle transport and protein synthesis

Microtubules are key players in axonal growth and provide structural support to axo-dendritic vesicular trafficking. Microtubule ends undergo a repeated processes of polymerization/depolymerization, called dynamic instability. Microtubule polymerization requires the addition of GTP-bound α/β heterodimers to their ends (Fig. 8A). Tubulin heterodimers bind to two molecules of GTP at two separate sites. The N-site is located at the intradimer interface, between α - and β -tubulin at which GTP is not hydrolyzed and exchanges at a slow rate. The E-site is at the intradimer interface formed by the β -subunit of one heterodimer and the α -subunit of a neighboring heterodimer. GTP at the E-site is hydrolyzed to GDP and exchanged for a new GTP nucleotide. Since microtubules exhibit a high rate of dynamic equilibrium between polymerization states, cytoplasmic levels of GTP must be high enough to power the demand of the dynamic instability in the microtubules (Fig. 8A).

Microtubules form the cytoskeletal tracks on which lysosomes travel to their target endosomes and phagosomes (Fig. 8B). The kinesin motor powers transport of the lysosomes toward the positive-end of axonal microtubules with ATP, but the connection of the kinesin motor to the lysosome requires Arl8 GTPase with bound GTP [101]. Conversely, transport of late endosomes on microtubules toward lysosomes requires Rab7 GTPase and bound GTP to attach the dynein motor.

In AD, the microtubule-associated protein tau polymerizes with hyperphosphorylation into insoluble filaments of axo-dendritic neurofibrillary tangles (NFT) [112, 113]. Once hyperphosphorylated, tau loses its affinity for the microtubules. Wide-ranging studies have approached the alterations in microtubule dynamics, but have failed to identify a clear cause of failure in tau function. Regulation of tau involves GTPases. In AD brains, Rac1 GTPase protein levels

decline 50% [114]. In vitro, Rac1 GTPase activation caused hyperphosphorylation of tau₁₈₁, increased A β 42 production and decreased actin stability in spines [114]. This group also saw a biphasic rise in Rac1 GTPase in young 3xTg-AD hippocampus, followed by a later decline, as in late-stage human AD brains. In another study of tauopathy mice, increased farnesyl transferase activity to translocate the Rhes GTPase to autophagic vesicles was associated with low tau hyperphosphorylation, while inhibition of Rhes promoted tau hyperphosphorylation [115]. Thus, GTPase-dependent autophagy effectively clears pTau [116] and impaired autophagy from low GTP levels or oxidative redox state would impair clearance of pTau.

GTP is also essential for protein synthesis, the synthesis side of proteostasis, that requires the hydrolysis of two GTP molecules for each amino acid incorporated into a polypeptide. While ATP is used for charging aminoacyl-tRNA, for RNA helicase and recycling GDP to GTP, GTP itself is essential for the activity of elongation factors at initiation (IF-2), elongation (EF-Tu, EF-G) and ribosome release factor for termination (RF1) [117]. This group used a cell-free reconstituted system to measure an overall K_m for ATP of 27 μ M and 14 μ M for GTP. These high affinities suggest prioritization of protein synthesis during energy limitation to maintain high capacity, perhaps at the expense of endocytosis and autophagy which likely have higher K_m 's. Consequently, a large amount of GTP is required at synaptic plasticity where metabolic demands are the highest to maintain ionic homeostasis for synaptic function and protein turnover. Protein synthesis is crucial for presynaptic neurotransmitter release as well as consolidation of post synaptic plasticity [118]. Nevertheless, protein synthesis was decreased along with ribosomal RNA and tRNA levels, while RNA oxidation increased in the early phase of AD [119]. Altered protein synthesis leads to a constant accumulation of oxidized proteins leading to misfolding and aggregation. Protein aggregation in turn impairs the activity of cellular proteolytic systems resulting in further accumulation of oxidized proteins [120]. Thus, a vicious cycle results in excess protein ubiquitination and dysproteostasis. Several other genes that encode ribosomal proteins are abnormally regulated leading to altered protein levels of elongation factors eIF2 α , eIF3 η and eIF5 in AD [121]. Increased eIF2 α and decreased eIF3 η and

eIF5 levels were observed in the hippocampal CA1 region of AD brain. Persistent eIF2 α phosphorylation at Ser51 through prolonged overactivation of regulatory kinases inhibits the delivery of initiator methionyl-tRNA preventing the general translation initiation of a subset of mRNAs. Thus, proteostasis is could be impaired at severely limiting GTP concentrations.

Impaired GSH-Trx system as a trigger of autophagy failure in aging and AD

Age is the major risk factor in AD and is associated with an imbalance between a decrease in redox buffer protection and an oxidative shift that increases ROS production. However, the proposal that ROS damage is a causal factor in the pathogenesis of AD has not been validated by antioxidant therapy and an oxidative redox shift appears to be upstream of ROS damage [122]. An epigenetic oxidized redox shift (EORS) has been proposed to precede ROS-mediated oxidative damage to account for lasting metabolic alterations in aging and AD [123]. This manifests as a decrease in reductive intracellular redox ratios from the critical systems that maintain redox homeostasis: cysteine/cystine, GSH/GSSH and NAD(P)H/NAD(P) [3]. Across the age-span of non-transgenic mouse brains, an oxidative glutathione redox state precedes an Akt metabolic shift and old-age buildup of aggregated proteins which are greatly accelerated in the 3xTg-AD mouse model before intracellular hippocampal accumulation of A β or extracellular plaques [124]. Mutations in enzymes related to GSH metabolism have also been associated with the regulation of autophagy such as glutathione reductase gene (*grs-1*) in a *C. elegans* model that abolishes the nuclear translocation of HLH-30 transcription factor (orthologue of mammalian TFEB) and triggering reductions in the transcription of genes related to clearance of protein aggregates by autophagy [125]. Subcellular free NADH concentration decreases with age in mitochondria, nuclei and cytoplasm in live wild-type and AD-like neurons [2]. Could this redox shift stimulate the logarithmic increase in intracellular A β -aggregates with age [27]?

Although it seems evident that some combination of an increase in the rate of formation of intracellular A β or their rate of autophagic clearance decreases with the age (the main risk factor), the mechanisms

that trigger these alterations remain unclear. We propose that the redox state of cells and possibly a systemic redox shift are an essential driver of aging [123]. GSH and thioredoxin (Trx) are the most important thiol redox system in the cells against oxidative stress. However, their distribution in the cell is very different. The physiological GSH concentration is in the range of mM, ~1000-fold higher than Trx (in the μ M range). Despite this, the Trx system regulates a broader range of proteins than GSH system [126]. Proteomic analysis identified involvement of Trx in the regulation of pathways mainly linked to glycolysis/gluconeogenesis and cytoskeletal remodeling, while pathways affected by both Trx and GSH are related to insulin regulation of translation, lipid metabolism and cell adhesion [126]. In the same study, Trx was also associated with regulation of other proteins involved in handling GTP and autophagy, such as (i) the IQ motif contained in GTPase activating protein 1 (GAP), a scaffolding protein that integrates Rho GTPase and Ca²⁺/calmodulin signals in the maintenance of cytoskeletal integrity; (ii) GDP dissociation inhibitor-1 (GDI-1), a protein that maintains Rab proteins in the GDP-bound conformation; (iii) ubiquitin-activating enzyme E1(UBA1), a protein required for Atg7- and Atg3-independent autophagy [127]; and (iv) cofilin-1, an actin-depolymerizing factor whose function is crucial for maintaining synaptic spines and instigating mitochondrial fission and mitophagy [128]. Interestingly, cofilin-1 levels are elevated in AD model mice and human AD, indicating its unbalanced regulation [129]. These data suggest that Trx could have a possible role in maintaining and handling of GTP and autophagy.

The N-terminus of the Rab5 and Rab7 GTPases contain single or double cysteines that should be subject to oxidation–reduction to cystine or farnesylation to anchor in membranes or nitrosylation, perhaps as these Rabs aggregate. Other redox-sensitive proteins in endocytosis such as TXNL1 [130] could accelerate or impede transitions between steps in the pathway.

The low redox potential in the disulfide bond in Atg4 is efficiently reduced and activated by Trx, suggesting a role as a redox regulator of autophagy mediated by Atg4 [131]. Other components in the thioredoxin system are involved in autophagy regulation. Deletion of thioredoxin reductase-2 (TrxR2, mitochondrial isoform) caused mitochondrial degeneration accompanied by overexpression of LC3,

p62, LAMP1, and accumulation of autophagic bodies in cardiomyocytes [132]. But a TrxR1 deficiency enhanced oxidative stress, interrupted early autophagy and decreased protein-degradation in lysosomes [133]. These data suggest that several key proteins of the autophagy machinery, such as Atg in yeast, may be regulated in response to changes of intracellular redox conditions, in addition to potential regulation by GTP levels.

Boosting energy to rescue AD pathology

While stimulation of the overall autophagic flux is beneficial for cell and tissue fitness, without sufficient energy to execute autophagy, the system will be inhibited. Increasing evidence shows that a decrease in NAD⁺/NADH availability with age [2, 134] plays a critical role in age-related neurovascular and cerebrovascular dysfunction [135, 136]. Thus, rejuvenation of cellular oxidative and reductive energy may benefit the age-related increased demands of autophagy. Restoring cellular NAD⁺ and NADH levels by nicotinamide mononucleotide (NMN) supplement in aged mice rescued neurovascular function, increased cerebral blood flow, and improved performance on cognitive tasks [135, 136]. Nicotinamide riboside (NR), a precursor to NMN, improves mitochondrial and adult stem cell function in aged mice as well as extend overall lifespan [137]. These NAD precursors can also alter autophagy. NR can prevent the blockage of autophagic flux and reduce oxidative stress in doxorubicin-treated cardiomyocytes, leading to enhanced autolysosome clearance via the NAD⁺/SIRT1 signaling pathway [138]. NR treatment also increased autophagic function and restored mitochondrial health in animals models of Parkinson's disease, suggesting possible treatments in other neurodegenerative diseases [139]. Since the product of Sirtuins and PARP is nicotinamide, cells need the NAD salvage pathway to recycle nicotinamide back into NMN and ultimately NAD⁺. Two labs have treated 3xTg-AD mice with nicotinamide and observed beneficial improvement in memory, DNA repair, autophagy, accumulation of A β , and phosphorylated tau and improved bioenergetics [140, 141]. Similarly, an improvement in cognitive functionality on multiple behavioral tests in 3xTgAD mice treated with nicotinamide riboside suggests a pivotal role for cellular

NAD⁺ depletion upstream of neuronal degeneration in AD [142]. Therefore, we treated adult 3xTg-AD and non-transgenic neurons with nicotinamide to see that the bioenergetic improvement extended to improvement in GTP levels, both bound and free (Fig. 1). Overall, increased NAD⁺ levels may benefit numerous downstream functions such as NAD-dependent SIRT1 activated genes which rejuvenate mitochondria and inhibit inflammation and apoptosis.

Another class of molecules benefitting old neurons are Nrf2 activators [143, 144]. One example is the ester of epigallocatechin and gallic acid, (-)-Epigallocatechin-3-Gallate (EGCG), the most bioactive polyphenol found in green tea extract. EGCG significantly increased mRNA expression of the key autophagy adaptor proteins NDP52 and p62 and enhanced the clearance of AD-relevant phosphorylated tau species in primary neurons [145], as well as improving cerebrovascular tone [146]. EGCG also activated autophagic pathways by inducing Sirt1, exerting protective effects against human prion protein-induced neurotoxicity [69]. Similar effects were observed using other Nrf2 electrophiles sulforaphane [147], fisetin and urolithin A from pomegranate [148].

Conclusion and future studies

A surprising array of specific GTPase control macroautophagy (mitophagy), microautophagy, chaperone-mediated autophagy (CMA), and endocytosis and vesicle and mitochondrial trafficking. Since they all depend on local GTP levels, it will be important to determine their binding constants for GTP. Zala et al. [51] demonstrated functional significance for mitochondrial fission GTPases Drp1 and Fis1 with K_m around 100 μ M, while the Opa1 GTPase with a K_m around 500 μ M, would require higher GTP levels to promote mitochondrial fusion. In AD, the endocytic and exocytic processing of the synaptic adhesion protein beta-APP may be impaired by lower local GTP levels and GTPases as well as local oxidative redox states. More clear in AD is the accumulation of autophagic vesicles from either blockages in the long pathway or failure to upregulate a sufficiently robust response to internal cellular damage coupled with failure to sufficiently acidify lysosomes for activation of cargo degradation. Less

clear is how much of these deficits are caused by age-related degeneration, known to be associated with age-related oxidative redox shifts. Age-related changes in GTP levels or capacity to upregulate bioenergetic fluxes will be an important aspect of future investigation, especially if NAD⁺ precursors are able to boost capacity and promote more autophagic clearance. The metabolic adaptation to a sedentary lifestyle could downregulate mitochondrial function with less energy for healthy management of amyloid and tau proteostasis, synaptic function, and inflammation. In subjects with deficits, certain energy precursors and redox modulators may add to the benefits of exercise and a healthy diet to extend and promote maximum healthspan.

Acknowledgements Figures were created in Biorender.com.

Funding This work was supported in part by the National Institutes of Health grant RF1 AG058218 to GJB. RAS receives support from the Secretaría de Educación, Ciencia, Tecnología e Innovación (SECTEI) for the elaboration of the products derived from this postdoctoral work.

Data availability Raw image data for Figure 1 is available on reasonable request to the corresponding author.

Declarations

Conflict of interest The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Parihar MS, Kunz EA, Brewer GJ. Age-related decreases in NAD(P)H and glutathione cause redox declines before ATP loss during glutamate treatment of hippocampal neurons. *J Neurosci Res.* 2008;86:2339–52.
- Dong Y, Digman MA, Brewer GJ. Age- and AD-related redox state of NADH in subcellular compartments by fluorescence lifetime imaging microscopy. *Geroscience.* 2019;41:51–67.
- Dong Y, Harlan BA, Brewer GJ. When aging switches on Alzheimer's. *Aging.* 2021;13:13376–7.
- Dong Y, Sameni S, Digman MA, Brewer GJ. Reversibility of age-related oxidized free NADH redox states in Alzheimer's disease neurons by imposed external Cys/CySS redox shifts. *Sci Rep.* 2019;9:11274.
- Bubber P, Haroutunian V, Fisch G, Blass JP, Gibson GE. Mitochondrial abnormalities in Alzheimer brain: mechanistic implications. *Ann Neurol.* 2005;57:695–703.
- Traut TW. Physiological concentrations of purines and pyrimidines. *Mol Cell Biochem.* 1994;140:1–22.
- Bianchi-Smiraglia A, Wolff DW, Marston DJ, Deng Z, Han Z, Moparthy S, Wombacher RM, Mussell AL, Shen S, Chen J, Yun DH, O'Brien Cox A, Furdui CM, Hurley E, Feltri ML, Qu J, Hollis T, Kengne JBN, Fonggang B, Sousa RJ, Kandel ME, Kandel ES, Hahn KM, Nikiforov MA. Regulation of local GTP availability controls RAC1 activity and cell invasion. *Nat Commun.* 2021;12:6091.
- Schlattner U. The complex functions of the NME family—a matter of location and molecular activity. *Int J Mol Sci.* 2021;22:13083
- Bianchi-Smiraglia A, Rana MS, Foley CE, Paul LM, Lipchick BC, Moparthy S, Moparthy K, Fink EE, Bagati A, Hurley E, Affronti HC, Bakin AV, Kandel ES, Smiraglia DJ, Feltri ML, Sousa R, Nikiforov MA. Internally ratiometric fluorescent sensors for evaluation of intracellular GTP levels and distribution. *Nat Methods.* 2017;14:1003–9.
- Bianchi-Smiraglia A, Nikiforov MA. Assessment of intracellular GTP levels using genetically encoded fluorescent sensors. *Methods Mol Biol.* 2022;2394:163–9.
- Burrinha T, Martinsson I, Gomes R, Terrasso AP, Gouras GK, Almeida CG. Up-regulation of APP endocytosis by neuronal aging drives amyloid dependent-synapse loss. *J Cell Sci.* 2021;134:jcs255752
- Cho YY, Kwon OH, Chung S. Preferred endocytosis of amyloid precursor protein from cholesterol-enriched lipid raft microdomains. *Molecules.* 2020;25:5490.
- Ferguson SM, Raimondi A, Paradise S, Shen H, Mesaki K, Ferguson A, Destaing O, Ko G, Takasaki J, Cremona O, Toole EO, De Camilli P. Coordinated actions of actin and BAR proteins upstream of dynamin at endocytic clathrin-coated pits. *Dev cell.* 2009;17:811–22.
- Schmid SL, Frolov VA. Dynamin: functional design of a membrane fission catalyst. *Annu Rev Cell Dev Biol.* 2011;27:79–105.
- Elkin SR, Lakoduk AM, Schmid SL. Endocytic pathways and endosomal trafficking: a primer. *Wien Med Wochenschr.* 2016;166:196–204.
- Boissan M, Montagnac G, Shen Q, Griparic L, Guitton J, Romao M, Sauvonnet N, Lagache T, Lascu I, Raposo G, Desbourdes C, Schlattner U, Lacombe ML, Polo S, van der Bliek AM, Roux A, Chavrier P. Membrane trafficking. Nucleoside diphosphate kinases fuel dynamin superfamily proteins with GTP for membrane remodeling. *Science.* 2014;344:1510–5.

17. Stenmark H. Rab GTPases as coordinators of vesicle traffic. *Nat Rev Mol Cell Biol.* 2009;10:513–25.
18. Tanida I. Autophagosome formation and molecular mechanism of autophagy. *Antioxid Redox Signal.* 2011;14:2201–14.
19. Noda NN, Fujioka Y, Hanada T, Ohsumi Y, Inagaki F. Structure of the Atg12-Atg5 conjugate reveals a platform for stimulating Atg8-PE conjugation. *EMBO Rep.* 2013;14:206–11.
20. Levine B, Kroemer G. Autophagy in the pathogenesis of disease. *Cell.* 2008;132:27–42.
21. Dou Z, Chattopadhyay M, Pan JA, Guerriero JL, Jiang YP, Ballou LM, Yue Z, Lin RZ, Zong WX. The class IA phosphatidylinositol 3-kinase p110-beta subunit is a positive regulator of autophagy. *J Cell Biol.* 2010;191:827–43.
22. Ao X, Zou L, Wu Y. Regulation of autophagy by the Rab GTPase network. *Cell Death Differ.* 2014;21:348–58.
23. Langemeyer L, Borchers AC, Herrmann E, Fullbrunn N, Han Y, Perz A, Auffarth K, Kummel D, Ungermann C. A conserved and regulated mechanism drives endosomal Rab transition. *eLife.* 2020;9:e56090.
24. Liang C, Lee JS, Inn KS, Gack MU, Li Q, Roberts EA, Vergne I, Deretic V, Feng P, Akazawa C, Jung JU. Beclin1-binding UVRAG targets the class C Vps complex to coordinate autophagosome maturation and endocytic trafficking. *Nat Cell Biol.* 2008;10:776–87.
25. Collins KM, Thorngren NL, Fratti RA, Wickner WT. Sec17p and HOPS, in distinct SNARE complexes, mediate SNARE complex disruption or assembly for fusion. *EMBO J.* 2005;24:1775–86.
26. Lee JH, Yang DS, Goulbourne CN, Im E, Stavrides P, Pensalfini A, Chan H, Bouchet-Marquis C, Bleiwas C, Berg MJ, Huo C, Peddy J, Pawlik M, Levy E, Rao M, Staufenbiel M, Nixon RA. Faulty autolysosome acidification in Alzheimer's disease mouse models induces autophagic build-up of Abeta in neurons, yielding senile plaques. *Nat Neurosci.* 2022;25:688–701.
27. Brewer GJ, Herrera RA, Philipp S, Sosna J, Reyes-Ruiz JM, Glabe CG. Age-related intraneuronal aggregation of amyloid-beta in endosomes, mitochondria, autophagosomes, and lysosomes. *J Alzheimer's Dis : JAD.* 2020;73:229–46.
28. Ba L, Chen XH, Chen YL, Nie Q, Li ZJ, Ding FF, Zhang M. Distinct Rab7-related endosomal-autophagic-lysosomal dysregulation observed in cortex and hippocampus in APP^{swe}/PSEN1^{dE9} mouse model of Alzheimer's disease. *Chin Med J (Engl).* 2017;130:2941–50.
29. Kirisako T, Baba M, Ishihara N, Miyazawa K, Ohsumi M, Yoshimori T, Noda T, Ohsumi Y. Formation process of autophagosome is traced with Apg8/Aut7p in yeast. *J Cell Biol.* 1999;147:435–46.
30. Fujita N, Huang W, Lin TH, Groulx JF, Jean S, Nguyen J, Kuchitsu Y, Koyama-Honda I, Mizushima N, Fukuda M, Kiger AA. Genetic screen in *Drosophila* muscle identifies autophagy-mediated T-tubule remodeling and a Rab2 role in autophagy. *eLife.* 2017;6:e23367.
31. Ginsberg SD, Mufson EJ, Counts SE, Wu J, Alldred MJ, Nixon RA, Che S. Regional selectivity of rab5 and rab7 protein upregulation in mild cognitive impairment and Alzheimer's disease. *J Alzheimer's Dis: JAD.* 2010;22:631–9.
32. Muresan Z, Muresan V. Neuritic deposits of amyloid-beta peptide in a subpopulation of central nervous system-derived neuronal cells. *Mol Cell Biol.* 2006;26:4982–97.
33. Song MS, Baker GB, Todd KG, Kar S. Inhibition of beta-amyloid1-42 internalization attenuates neuronal death by stabilizing the endosomal-lysosomal system in rat cortical cultured neurons. *Neuroscience.* 2011;178:181–8.
34. Yuyama K, Yanagisawa K. Late endocytic dysfunction as a putative cause of amyloid fibril formation in Alzheimer's disease. *J Neurochem.* 2009;109:1250–60.
35. Lee CY, Tse W, Smith JD, Landreth GE. Apolipoprotein E promotes beta-amyloid trafficking and degradation by modulating microglial cholesterol levels. *J Biol Chem.* 2012;287:2032–44.
36. Lee S, Sato Y, Nixon RA. Lysosomal proteolysis inhibition selectively disrupts axonal transport of degradative organelles and causes an Alzheimer's-like axonal dystrophy. *J Neurosci.* 2011;31:7817–30.
37. Liu H, Luo K, Luo D. Guanosine monophosphate reductase 1 is a potential therapeutic target for Alzheimer's disease. *Sci Rep.* 2018;8:2759.
38. Valdez G, Philippidou P, Rosenbaum J, Akmentin W, Shao Y, Halegoua S. Trk-signaling endosomes are generated by Rac-dependent macroendocytosis. *Proc Natl Acad Sci USA.* 2007;104:12270–5.
39. Xu W, Weissmiller AM, White JA 2nd, Fang F, Wang X, Wu Y, Pearn ML, Zhao X, Sawa M, Chen S, Gunawardena S, Ding J, Mobley WC, Wu C. Amyloid precursor protein-mediated endocytic pathway disruption induces axonal dysfunction and neurodegeneration. *J Clin Investig.* 2016;126:1815–33.
40. Xu W, Fang F, Ding J, Wu C. Dysregulation of Rab5-mediated endocytic pathways in Alzheimer's disease. *Traffic.* 2018;19:253–62.
41. Kim S, Sato Y, Mohan PS, Peterhoff C, Pensalfini A, Rigoglioso A, Jiang Y, Nixon RA. Evidence that the rab5 effector APPL1 mediates APP-betaCTF-induced dysfunction of endosomes in Down syndrome and Alzheimer's disease. *Mol Psychiatry.* 2016;21:707–16.
42. Lane RF, Steele JW, Cai D, Ehrlich ME, Attie AD, Gandy S. Protein sorting motifs in the cytoplasmic tail of SorCS1 control generation of Alzheimer's amyloid-beta peptide. *J Neurosci.* 2013;33:7099–107.
43. Vieira SI, Rebelo S, Esselmann H, Wiltfang J, Lah J, Lane R, Small SA, Gandy S, da Cruz ESEF, da Cruz ESOA. Retrieval of the Alzheimer's amyloid precursor protein from the endosome to the TGN is S655 phosphorylation state-dependent and retromer-mediated. *Mol Neurodegener.* 2010;5:40.
44. Cataldo AM, Petanceska S, Terio NB, Peterhoff CM, Durham R, Mercken M, Mehta PD, Buxbaum J, Haroutunian V, Nixon RA. Abeta localization in abnormal endosomes: association with earliest Abeta elevations in AD and Down syndrome. *Neurobiol Aging.* 2004;25:1263–72.
45. Kim S, Choi KJ, Cho SJ, Yun SM, Jeon JP, Koh YH, Song J, Johnson GV, Jo C. Fisetin stimulates autophagic degradation of phosphorylated tau via the

- activation of TFEB and Nrf2 transcription factors. *Sci Rep.* 2016;6:24933.
46. Beau I, Codogno P. GTP: gatekeeper for autophagy. *Mol Cell.* 2010;39:485–6.
 47. Mitra S, Cheng KW, Mills GB. Rab GTPases implicated in inherited and acquired disorders. *Semin Cell Dev Biol.* 2011;22:57–68.
 48. Driscoll CA, Menotti-Raymond M, Roca AL, Hupe K, Johnson WE, Geffen E, Harley EH, Delibes M, Pontier D, Kitchener AC, Yamaguchi N, O'Brien SJ, Macdonald DW. The Near Eastern origin of cat domestication. *Science.* 2007;317:519–23.
 49. Jullie D, Choquet D, Perrais D. Recycling endosomes undergo rapid closure of a fusion pore on exocytosis in neuronal dendrites. *J Neurosci.* 2014;34:11106–18.
 50. Pilli M, Arko-Mensah J, Ponpuak M, Roberts E, Master S, Mandell MA, Dupont N, Ornatowski W, Jiang S, Bradfute SB, Bruun JA, Hansen TE, Johansen T, Deretic V. TBK-1 promotes autophagy-mediated antimicrobial defense by controlling autophagosome maturation. *Immunity.* 2012;37:223–34.
 51. Zala D, Schlattner U, Desvignes T, Bobe J, Roux A, Chavrier P, Boissan M. The advantage of channeling nucleotides for very processive functions. *F1000Res.* 2017;6:724.
 52. Kern A, Dikic I, Behl C. The integration of autophagy and cellular trafficking pathways via RAB GAPs. *Autophagy.* 2015;11:2393–7.
 53. Wong YC, Ysselstein D, Krainc D. Mitochondria-lysosome contacts regulate mitochondrial fission via RAB7 GTP hydrolysis. *Nature.* 2018;554:382–6.
 54. Toyofuku T, Morimoto K, Sasawatari S, Kumanogoh A. Leucine-rich repeat kinase 1 regulates autophagy through turning on TBC1D2-dependent Rab7 inactivation. *Mol Cell Biol.* 2015;35:3044–58.
 55. Nixon RA. The role of autophagy in neurodegenerative disease. *Nat Med.* 2013;19:983–97.
 56. Uddin MS, Stachowiak A, Mamun AA, Tzvetkov NT, Takeda S, Atanasov AG, Bergantin LB, Abdel-Daim MM, Stankiewicz AM. Autophagy and Alzheimer's disease: from molecular mechanisms to therapeutic implications. *Front Aging Neurosci.* 2018;10:04.
 57. Schuck S. Microautophagy - distinct molecular mechanisms handle cargoes of many sizes. *J Cell Sci.* 2020;133:jcs246322.
 58. Maruyama T, Noda NN. Autophagy-regulating protease Atg4: structure, function, regulation and inhibition. *J Antibiot (Tokyo).* 2017;71:72–78.
 59. Aman Y, Schmauck-Medina T, Hansen M, Morimoto RI, Simon AK, Bjedov I, Palikaras K, Simonsen A, Johansen T, Tavernarakis N, Rubinsztein DC, Partridge L, Kroemer G, Labbadia J, Fang EF. Autophagy in healthy aging and disease. *Nat Aging.* 2021;1:634–50.
 60. Miyoshi T, Tsuji T, Higashida C, Hertzog M, Fujita A, Narumiya S, Scita G, Watanabe N. Actin turnover-dependent fast dissociation of capping protein in the dendritic nucleation actin network: evidence of frequent filament severing. *J Cell Biol.* 2006;175:947–55.
 61. Kim J, Kundu M, Viollet B, Guan KL. AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat Cell Biol.* 2011;13:132–41.
 62. Nixon RA, Wegiel J, Kumar A, Yu WH, Peterhoff C, Cataldo A, Cuervo AM. Extensive involvement of autophagy in Alzheimer disease: an immuno-electron microscopy study. *J Neuropathol Exp Neurol.* 2005;64:113–22.
 63. Bordi M, Berg MJ, Mohan PS, Peterhoff CM, Alldred MJ, Che S, Ginsberg SD, Nixon RA. Autophagy flux in CA1 neurons of Alzheimer hippocampus: Increased induction overburdens failing lysosomes to propel neurotrophic dystrophy. *Autophagy.* 2016;12:2467–83.
 64. Yu WH, Cuervo AM, Kumar A, Peterhoff CM, Schmidt SD, Lee JH, Mohan PS, Mercken M, Farmery MR, Tjernberg LO, Jiang Y, Duff K, Uchiyama Y, Naslund J, Mathews PM, Cataldo AM, Nixon RA. Macroautophagy—a novel beta-amyloid peptide-generating pathway activated in Alzheimer's disease. *J Cell Biol.* 2005;171:87–98.
 65. Sanchez-Varo R, Trujillo-Estrada L, Sanchez-Mejias E, Torres M, Baglietto-Vargas D, Moreno-Gonzalez I, De Castro V, Jimenez S, Ruano D, Vizuete M, Davila JC, Garcia-Verdugo JM, Jimenez AJ, Vitorica J, Gutierrez A. Abnormal accumulation of autophagic vesicles correlates with axonal and synaptic pathology in young Alzheimer's mice hippocampus. *Acta Neuropathol.* 2012;123:53–70.
 66. Tomiyama T, Matsuyama S, Iso H, Umeda T, Takuma H, Ohnishi K, Ishibashi K, Teraoka R, Sakama N, Yamashita T, Nishitsuji K, Ito K, Shimada H, Lambert MP, Klein WL, Mori H. A mouse model of amyloid beta oligomers: their contribution to synaptic alteration, abnormal tau phosphorylation, glial activation, and neuronal loss in vivo. *J Neurosci.* 2010;30:4845–56.
 67. Ji C, Tang M, Johnson GVW. Assessing the degradation of tau in primary neurons: The role of autophagy. *Methods Cell Biol.* 2017;141:229–44.
 68. Wang Y, Mandelkow E. Tau in physiology and pathology. *Nat Rev Neurosci.* 2016;17:5–21.
 69. Lee JH, McBrayer MK, Wolfe DM, Haslett LJ, Kumar A, Sato Y, Lie PP, Mohan P, Coffey EE, Kompella U, Mitchell CH, Lloyd-Evans E, Nixon RA. Presenilin 1 Maintains Lysosomal Ca(2+) Homeostasis via TRPML1 by regulating vATPase-mediated lysosome acidification. *Cell Rep.* 2015;12:1430–44.
 70. Lee JH, Yu WH, Kumar A, Lee S, Mohan PS, Peterhoff CM, Wolfe DM, Martinez-Vicente M, Massey AC, Sovak G, Uchiyama Y, Westaway D, Cuervo AM, Nixon RA. Lysosomal proteolysis and autophagy require presenilin 1 and are disrupted by Alzheimer-related PS1 mutations. *Cell.* 2010;141:1146–58.
 71. Ji ZS, Mullendorff K, Cheng IH, Miranda RD, Huang Y, Mahley RW. Reactivity of apolipoprotein E4 and amyloid beta peptide: lysosomal stability and neurodegeneration. *J Biol Chem.* 2006;281:2683–92.
 72. Nilsson P, Loganathan K, Sekiguchi M, Matsuba Y, Hui K, Tsubuki S, Tanaka M, Iwata N, Saito T, Saido TC. Abeta secretion and plaque formation depend on autophagy. *Cell Rep.* 2013;5:61–9.
 73. Nilsson P, Sekiguchi M, Akagi T, Izumi S, Komori T, Hui K, Sorgjerd K, Tanaka M, Saito T, Iwata N, Saido TC. Autophagy-related protein 7 deficiency in amyloid beta (Abeta) precursor protein transgenic mice

- decreases Abeta in the multivesicular bodies and induces Abeta accumulation in the Golgi. *Am J Pathol.* 2015;185:305–13.
74. Ding X, Jiang X, Tian R, Zhao P, Li L, Wang X, Chen S, Zhu Y, Mei M, Bao S, Liu W, Tang Z, Sun Q. RAB2 regulates the formation of autophagosome and autolysosome in mammalian cells. *Autophagy.* 2019;15:1774–86.
 75. Hill K, Li Y, Bennett M, McKay M, Zhu X, Shern J, Torre E, Lah JJ, Levey AI, Kahn RA. Munc18 interacting proteins: ADP-ribosylation factor-dependent coat proteins that regulate the traffic of beta-Alzheimer's precursor protein. *J Biol Chem.* 2003;278:36032–40.
 76. Bansal A, Kirschner M, Zu L, Cai D, Zhang L. Coconut oil decreases expression of amyloid precursor protein (APP) and secretion of amyloid peptides through inhibition of ADP-ribosylation factor 1 (ARF1). *Brain Res.* 2019;1704:78–84.
 77. Jiang Q, Wang L, Guan Y, Xu H, Niu Y, Han L, Wei YP, Lin L, Chu J, Wang Q, Yang Y, Pei L, Wang JZ, Tian Q. Golgin-84-associated Golgi fragmentation triggers tau hyperphosphorylation by activation of cyclin-dependent kinase-5 and extracellular signal-regulated kinase. *Neurobiol Aging.* 2014;35:1352–63.
 78. Mohamed NV, Desjardins A, Leclerc N. Tau secretion is correlated to an increase of Golgi dynamics. *PLoS ONE.* 2017;12:e0178288.
 79. Udayar V, Buggia-Prevot V, Guerreiro RL, Siegel G, Rambabu N, Soohoo AL, Ponnusamy M, Siegenthaler B, Bali J, Aesg SM, Ries J, Puthenveedu MA, Hardy J, Thinnakaran G, Rajendran L. A paired RNAi and RabGAP overexpression screen identifies Rab11 as a regulator of beta-amyloid production. *Cell Reports.* 2013;5:1536–51.
 80. Kon M, Cuervo AM. Chaperone-mediated autophagy in health and disease. *FEBS Lett.* 2010;584:1399–404.
 81. Bandyopadhyay U, Sridhar S, Kaushik S, Kiffin R, Cuervo AM. Identification of regulators of chaperone-mediated autophagy. *Mol Cell.* 2010;39:535–47.
 82. Zhang C, Cuervo AM. Restoration of chaperone-mediated autophagy in aging liver improves cellular maintenance and hepatic function. *Nat Med.* 2008;14:959–65.
 83. Dove SK, Dong K, Kobayashi T, Williams FK, Michell RH. Phosphatidylinositol 3,5-bisphosphate and Fab1p/PIKfyve underpin endo-lysosome function. *Biochem J.* 2009;419:1–13.
 84. Senju Y, Lappalainen P. Regulation of actin dynamics by PI(4,5)P2 in cell migration and endocytosis. *Curr Opin Cell Biol.* 2019;56:7–13.
 85. Vicinanza M, Puri C, Rubinsztein DC. Coincidence detection of RAB11A and PI(3)P by WIP1 directs autophagosome formation. *Oncotarget.* 2019;10:2579–80.
 86. Ikononov OC, Sbrissa D, Shisheva A. Mammalian cell morphology and endocytic membrane homeostasis require enzymatically active phosphoinositide 5-kinase PIKfyve. *J Biol Chem.* 2001;276:26141–7.
 87. Balklava Z, Niehage C, Currinn H, Mellor L, Guscott B, Poulin G, Hoflack B, Wassmer T. The Amyloid Precursor Protein Controls PIKfyve Function. *PLoS One.* 2015;10:e0130485.
 88. Dong XP, Shen D, Wang X, Dawson T, Li X, Zhang Q, Cheng X, Zhang Y, Weisman LS, Delling M, Xu H. PI(3,5)P(2) controls membrane trafficking by direct activation of mucolipin Ca(2+) release channels in the endolysosome. *Nat Commun.* 2010;1:38.
 89. Soyombo AA, Tjon-Kon-Sang S, Rbaibi Y, Bashllari E, Bisceglia J, Muallem S, Kiselyov K. TRP-ML1 regulates lysosomal pH and acidic lysosomal lipid hydrolytic activity. *J Biol Chem.* 2006;281:7294–301.
 90. Reddy PH, Reddy TP, Manczak M, Calkins MJ, Shirendeb U, Mao P. Dynamin-related protein 1 and mitochondrial fragmentation in neurodegenerative diseases. *Brain Res Rev.* 2011;67:103–18.
 91. Zhu X, Perry G, Smith MA, Wang X. Abnormal mitochondrial dynamics in the pathogenesis of Alzheimer's disease. *J Alzheimer's Dis : JAD.* 2013;33(Suppl 1):S253–262.
 92. Wang X, Campos CR, Peart JC, Smith LK, Boni JL, Cannon RE, Miller DS. Nrf2 upregulates ATP binding cassette transporter expression and activity at the blood-brain and blood-spinal cord barriers. *J Neurosci.* 2014;34:8585–93.
 93. Chen H, Chan DC. Mitochondrial dynamics—fusion, fission, movement, and mitophagy—in neurodegenerative diseases. *Hum Mol Genet.* 2009;18:R169–176.
 94. Kandimalla R, Reddy PH. Multiple faces of dynamin-related protein 1 and its role in Alzheimer's disease pathogenesis. *Biochem Biophys Acta.* 2016;1862:814–28.
 95. Wang C, Chen K, Xia Y, Dai W, Wang F, Shen M, Cheng P, Wang J, Lu J, Zhang Y, Yang J, Zhu R, Zhang H, Li J, Zheng Y, Zhou Y, Guo C. N-acetylcysteine attenuates ischemia-reperfusion-induced apoptosis and autophagy in mouse liver via regulation of the ROS/JNK/Bcl-2 pathway. *PLoS One.* 2014;9:e108855.
 96. Deffieu M, Bhatia-Kissova I, Salin B, Galinier A, Manon S, Camougrand N. Glutathione participates in the regulation of mitophagy in yeast. *J Biol Chem.* 2009;284:14828–37.
 97. Calkins MJ, Manczak M, Mao P, Shirendeb U, Reddy PH. Impaired mitochondrial biogenesis, defective axonal transport of mitochondria, abnormal mitochondrial dynamics and synaptic degeneration in a mouse model of Alzheimer's disease. *Hum Mol Genet.* 2011;20:4515–29.
 98. Manczak M, Reddy PH. Abnormal interaction of oligomeric amyloid-beta with phosphorylated tau: implications to synaptic dysfunction and neuronal damage. *J Alzheimer's Dis : JAD.* 2013;36:285–95.
 99. Nakamura T, Oh CK, Zhang X, Tannenbaum SR, Lipton SA. Protein transnitrosylation signaling networks contribute to inflammation and neurodegenerative disorders. *Antioxid Redox Signal.* 2021;35:531–50.
 100. Krogsaeter E, Rosato AS, Grimm C. Metabolomic profiles predict individual multidisease outcomes: Targets for lysosomal storage and neurodegenerative disease therapy? *Cell Calcium.* 2022;103:102553.
 101. Farias GG, Guardia CM, De Pace R, Britt DJ, Bonifacino JS. BORC/kinesin-1 ensemble drives polarized transport of lysosomes into the axon. *Proc Natl Acad Sci USA.* 2017;114:E2955–64.
 102. Jongsma ML, Bakker J, Cabukusta B, Liv N, van Elsland D, Fermie J, Akkermans JL, Kuijl C, van der Zanden SY, Janssen L, Hoogzaad D, van der Kant R, Wijdeven RH,

- Klumperman J, Berlin I, Neeffjes J. SKIP-HOPS recruits TBC1D15 for a Rab7-to-Arl8b identity switch to control late endosome transport. *EMBO J*. 2020;39:e102301.
103. Roney JC, Li S, Farfel-Becker T, Huang N, Sun T, Xie Y, Cheng XT, Lin MY, Platt FM, Sheng ZH. Lipid-mediated impairment of axonal lysosome transport contributing to autophagic stress. *Autophagy*. 2021;17:1796–8.
 104. Rosa-Ferreira C, Munro S. Arl8 and SKIP act together to link lysosomes to kinesin-1. *Dev Cell*. 2011;21:1171–8.
 105. Ponsford AH, Ryan TA, Raimondi A, Cocucci E, Wycislo SA, Frohlich F, Swan LE, Stagi M. Live imaging of intra-lysosome pH in cell lines and primary neuronal culture using a novel genetically encoded biosensor. *Autophagy*. 2021;17:1500–18.
 106. Drummond E, Kavanagh T, Pires G, Marta-Ariza M, Kanshin E, Nayak S, Faustin A, Berdah V, Ueberheide B, Wisniewski T. The amyloid plaque proteome in early onset Alzheimer's disease and Down syndrome. *Acta Neuropathol Commun*. 2022;10:53.
 107. Pena-Llopis S, Vega-Rubin-de-Celis S, Schwartz JC, Wolff NC, Tran TA, Zou L, Xie XJ, Corey DR, Brugarolas J. Regulation of TFEB and V-ATPases by mTORC1. *EMBO J*. 2011;30:3242–58.
 108. Roczniaik-Ferguson A, Petit CS, Froehlich F, Qian S, Ky J, Angarola B, Walther TC, Ferguson SM. The transcription factor TFEB links mTORC1 signaling to transcriptional control of lysosome homeostasis. *Sci Signal*. 2012;5:ra42.
 109. Guo X, Tang P, Chen L, Liu P, Hou C, Zhang X, Liu Y, Chong L, Li X, Li R. Amyloid beta-induced redistribution of transcriptional factor EB and lysosomal Dysfunction in Primary Microglial Cells. *Front Aging Neurosci*. 2017;9:228.
 110. Inoki K, Li Y, Xu T, Guan KL. Rheb GTPase is a direct target of TSC2 GAP activity and regulates mTOR signaling. *Genes Dev*. 2003;17:1829–34.
 111. Mattson MP, Longo VD, Harvie M. Impact of intermittent fasting on health and disease processes. *Ageing Res Rev*. 2017;39:46–58.
 112. Singh TJ, Grundke-Iqbal I, Iqbal K. Differential phosphorylation of human tau isoforms containing three repeats by several protein kinases. *Arch Biochem Biophys*. 1996;328:43–50.
 113. Wang YP, Biernat J, Pickhardt M, Mandelkow E, Mandelkow EM. Stepwise proteolysis liberates tau fragments that nucleate the Alzheimer-like aggregation of full-length tau in a neuronal cell model. *Proc Natl Acad Sci USA*. 2007;104:10252–7.
 114. Borin M, Saraceno C, Catania M, Lorenzetto E, Pontelli V, Paterlini A, Fostinelli S, Avesani A, Di Fede G, Zanusso G, Benussi L, Binetti G, Zorzan S, Ghidoni R, Buffelli M, Bolognin S. Rac1 activation links tau hyperphosphorylation and Abeta dysmetabolism in Alzheimer's disease. *Acta Neuropathol Commun*. 2018;6:61.
 115. Hernandez I, Luna G, Rauch JN, Reis SA, Giroux M, Karch CM, Boctor D, Sibih YE, Storm NJ, Diaz A, Kaushik S, Zekanowski C, Kang AA, Hinman CR, Cerovac V, Guzman E, Zhou H, Haggarty SJ, Goate AM, Fisher SK, Cuervo AM, Kosik KS. A farnesyltransferase inhibitor activates lysosomes and reduces tau pathology in mice with tauopathy. *Sci Transl Med*. 2019;11:eaat3005.
 116. Lin H, Tang M, Ji C, Girardi P, Cvetojevic G, Chen D, Koren SA, Johnson GVW. BAG3 regulation of RAB35 mediates the endosomal sorting complexes required for transport/endolysosome pathway and tau clearance. *Biol Psychiat*. 2022;92:10–24.
 117. Jewett MC, Miller ML, Chen Y, Swartz JR. Continued protein synthesis at low [ATP] and [GTP] enables cell adaptation during energy limitation. *J Bacteriol*. 2009;191:1083–91.
 118. Younts TJ, Monday HR, Dudok B, Klein ME, Jordan BA, Katona I, Castillo PE. Presynaptic protein synthesis is required for long-term plasticity of GABA release. *Neuron*. 2016;92:479–92.
 119. Ding Q, Markesbery WR, Chen Q, Li F, Keller JN. Ribosome dysfunction is an early event in Alzheimer's disease. *J Neurosci*. 2005;25:9171–5.
 120. Reeg S, Grune T. Protein oxidation in aging: does it play a role in aging progression? *Antioxid Redox Signal*. 2015;23:239–55.
 121. Hernandez-Ortega K, Garcia-Esparcia P, Gil L, Lucas JJ, Ferrer I. Altered machinery of protein synthesis in Alzheimer's: from the nucleolus to the ribosome. *Brain Pathol*. 2016;26:593–605.
 122. Ghosh D, Levault KR, Brewer GJ. Relative importance of redox buffers GSH and NAD(P)H in age-related neurodegeneration and Alzheimer disease-like mouse neurons. *Aging Cell*. 2014;13:631–40.
 123. Brewer GJ. Epigenetic oxidative redox shift (EORS) theory of aging unifies the free radical and insulin signaling theories. *Exp Gerontol*. 2010;45:173–9.
 124. Pontrello CG, McWhirt JM, Glabe CG, Brewer GJ. Age-related oxidative redox and metabolic changes precede intraneuronal A β accumulation and plaque deposition in a transgenic Alzheimer's disease mouse model. *J Alz Dis*. 2022;909:1501–1521.
 125. Guerrero-Gomez D, Mora-Lorca JA, Saenz-Narciso B, Naranjo-Galindo FJ, Munoz-Lobato F, Parrado-Fernandez C, Goikolea J, Cedazo-Minguez A, Link CD, Neri C, Sequedo MD, Vazquez-Manrique RP, Fernandez-Suarez E, Goder V, Pane R, Cabisco E, Askjaer P, Cabello J, Miranda-Vizuete A. Loss of glutathione redox homeostasis impairs proteostasis by inhibiting autophagy-dependent protein degradation. *Cell Death Differ*. 2019;26:1545–65.
 126. Go YM, Roede JR, Walker DI, Duong DM, Seyfried NT, Orr M, Liang Y, Pennell KD, Jones DP. Selective targeting of the cysteine proteome by thioredoxin and glutathione redox systems. *Mol Cell Proteomics : MCP*. 2013;12:3285–96.
 127. Chang TK, Shrivage BV, Hayes SD, Powers CM, Simin RT, Wade Harper J, Baehrecke EH. Uba1 functions in Atg7- and Atg3-independent autophagy. *Nat Cell Biol*. 2013;15:1067–78.
 128. Li GB, Zhang HW, Fu RQ, Hu XY, Liu L, Li YN, Liu YX, Liu X, Hu JJ, Deng Q, Luo QS, Zhang R, Gao N. Mitochondrial fission and mitophagy depend on cofilin-mediated actin depolymerization activity at the mitochondrial fission site. *Oncogene*. 2018;37:1485–502.
 129. Rush T, Martinez-Hernandez J, Dollmeyer M, Frandemich ML, Borel E, Boisseau S, Jacquier-Sarlin M, Buisson A. Synaptotoxicity in Alzheimer's disease

- involved a dysregulation of actin cytoskeleton dynamics through Cofilin 1 phosphorylation. *J Neurosci*. 2018;38:10349–61.
130. Felberbaum-Corti M, Morel E, Cavalli V, Vilbois F, Gruenberg J. The redox sensor TXNL1 plays a regulatory role in fluid phase endocytosis. *PLoS One*. 2007;2:e1144.
 131. Perez-Perez ME, Zaffagnini M, Marchand CH, Crespo JL, Lemaire SD. The yeast autophagy protease Atg4 is regulated by thioredoxin. *Autophagy*. 2014;10:1953–64.
 132. Kiermayer C, Northrup E, Schrewe A, Walch A, de Angelis MH, Schoensiegel F, Zischka H, Prehn C, Adamski J, Bekeredjian R, Ivandic B, Kupatt C, Briemeier M. Heart-specific knockout of the mitochondrial thioredoxin reductase (Txnrd2) induces metabolic and contractile dysfunction in the aging myocardium. *J Am Heart Assoc*. 2015;4:e002153.
 133. Nagakannan P, Iqbal MA, Yeung A, Thliveris JA, Rastegar M, Ghavami S, Eftekharpour E. Perturbation of redox balance after thioredoxin reductase deficiency interrupts autophagy-lysosomal degradation pathway and enhances cell death in nutritionally stressed SH-SY5Y cells. *Free Radical Biol Med*. 2016;101:53–70.
 134. Zhu XH, Lu M, Lee BY, Ugurbil K, Chen W. In vivo NAD assay reveals the intracellular NAD contents and redox state in healthy human brain and their age dependences. *Proc Natl Acad Sci USA*. 2015;112:2876–81.
 135. Tarantini S, Valcarcel-Ares NM, Yabluchanskiy A, Fulop GA, Hertelendy P, Gautam T, Farkas E, Perz A, Rabinovitch PS, Sonntag WE, Csiszar A, Ungvari Z. Treatment with the mitochondrial-targeted antioxidant peptide SS-31 rescues neurovascular coupling responses and cerebrovascular endothelial function and improves cognition in aged mice. *Aging Cell*. 2018;70:e12731.
 136. Kiss T, Nyul-Toth A, Balasubramanian P, Tarantini S, Ahire C, Yabluchanskiy A, Csipo T, Farkas E, Wren JD, Garman L, Csiszar A, Ungvari Z. Nicotinamide mononucleotide (NMN) supplementation promotes neurovascular rejuvenation in aged mice: transcriptional footprint of SIRT1 activation, mitochondrial protection, anti-inflammatory, and anti-apoptotic effects. *Geroscience*. 2020;42:527–46.
 137. Zhang H, Ryu D, Wu Y, Gariani K, Wang X, Luan P, D'Amico D, Ropelle ER, Lutolf MP, Aebersold R, Schoonjans K, Menzies KJ, Auwerx J. NAD(+) replacement improves mitochondrial and stem cell function and enhances life span in mice. *Science*. 2016;352:1436–43.
 138. Zheng D, Zhang Y, Zheng M, Cao T, Wang G, Zhang L, Ni R, Brockman J, Zhong H, Fan GC, Peng T. Nicotinamide riboside promotes autolysosomal clearance in preventing doxorubicin-induced cardiotoxicity. *Clin Sci (Lond)*. 2019;133:1505–21.
 139. Schondorf DC, Ivanyuk D, Baden P, Sanchez-Martinez A, De Cicco S, Yu C, Giunta I, Schwarz LK, Di Napoli G, Panagiotakopoulou V, Nestel S, Keatinge M, Pruszk J, Bandmann O, Heimrich B, Gasser T, Whitworth AJ, Deleidi M. The NAD⁺ precursor nicotinamide riboside rescues mitochondrial defects and neuronal loss in iPSC and fly models of Parkinson's disease. *Cell Rep*. 2018;23:2976–88.
 140. Green KN, Steffan JS, Martinez-Coria H, Sun X, Schreiber SS, Thompson LM, LaFerla FM. Nicotinamide restores cognition in Alzheimer's disease transgenic mice via a mechanism involving sirtuin inhibition and selective reduction of Thr231-phosphotau. *J Neurosci*. 2008;28:11500–10.
 141. Liu D, Pitta M, Jiang H, Lee JH, Zhang G, Chen X, Kawamoto EM, Mattson MP. Nicotinamide forestalls pathology and cognitive decline in Alzheimer mice: evidence for improved neuronal bioenergetics and autophagy procession. *Neurobiol Aging*. 2013;34:1564–80.
 142. Hou Y, Lautrup S, Cordonnier S, Wang Y, Croteau DL, Zavala E, Zhang Y, Moritoh K, O'Connell JF, Baptiste BA, Stevnsner TV, Mattson MP, Bohr VA. NAD(+) supplementation normalizes key Alzheimer's features and DNA damage responses in a new AD mouse model with introduced DNA repair deficiency. *Proc Natl Acad Sci USA*. 2018;115:E1876–85.
 143. Esteras N, Dinkova-Kostova AT, Abramov AY. Nrf2 activation in the treatment of neurodegenerative diseases: a focus on its role in mitochondrial bioenergetics and function. *Biol Chem*. 2016;397:383–400.
 144. Goodfellow MJ, Borcar A, Proctor JL, Greco T, Rosenthal RE, Fiskum G. Transcriptional activation of antioxidant gene expression by Nrf2 protects against mitochondrial dysfunction and neuronal death associated with acute and chronic neurodegeneration. *Exp Neurol*. 2020;328:113247.
 145. Chesser AS, Ganeshan V, Yang J, Johnson GV. Epigallocatechin-3-gallate enhances clearance of phosphorylated tau in primary neurons. *Nutr Neurosci*. 2016;19:21–31.
 146. Redford KE, Rognant S, Jepps TA, Abbott GW. KCNQ5 potassium channel activation underlies vasodilation by tea. *Cell Physiol Biochem*. 2021;55:46–64.
 147. Jo C, Kim S, Cho SJ, Choi KJ, Yun SM, Koh YH, Johnson GV, Park SI. Sulforaphane induces autophagy through ERK activation in neuronal cells. *FEBS Lett*. 2014;588:3081–8.
 148. Kaushik S, Tasset I, Arias E, Pampliega O, Wong E, Martinez-Vicente M, Cuervo AM. Autophagy and the hallmarks of aging. *Ageing Res Rev*. 2021;72:101468.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.