# **UC Irvine**

# **UC Irvine Previously Published Works**

# **Title**

Response to Rougeron et al.: Leishmania population genetics: Clonality, selfing and aneuploidy

# **Permalink**

https://escholarship.org/uc/item/95x128nb

# **Journal**

Trends in Parasitology, 31(7)

# **ISSN**

1471-4922

### **Authors**

Tibayrenc, M Ayala, FJ

# **Publication Date**

2015-07-01

# DOI

10.1016/j.pt.2015.04.002

Peer reviewed

which should purge heterozygosity at each haploid cycle [9] and renders tests based on the hypothesis of diploidy invalid [2,3]. The view that microsatellite data do not fit the hypothesis of aneuploidy [1] is questionable [10]. Rougeron *et al.* [1] propose that aneuploidy could be transitory, which is not supported by a genomic analysis that deals with natural isolates and not experimental populations [11]. Even if it were true, heterozygosity purging at each haploid cycle [8] should remain.

#### Frequent recombination?

It is questionable to state that in some lineages (and possibly most lineages) of *Leishmania* sexual recombination is frequent [1], because the evidence against it is strong [2,3]. The studies cited to support sexual recombination [1], rather, deal with heterozygote deficit, which is considered as evidence for selfing. Selfing leads to lack of recombination and LD, not to sexual recombination [2].

The efforts by Rougeron et al. [1] to explore more finely the role played by selfing in Leishmania evolution are valuable. However, as recalled many times [2–4], selfing does not challenge the PCE model, since this model considers it as a particular case of clonality. Moreover, methodological difficulties (in particular the strong evidence for aneuploidy in Leishmania) make it tentative to evidence selfing. We consider that the PCE model by far fits the best Leishmania population genetic data, which do not show any evidence of frequent sexual recombination [1]. The development of whole-genome sequencing will certainly

help in clarifying parasite evolutionary patterns, as it has done in several major bacterial species [2].

#### References

- 1 Rougeron, V. et al. (2015) A primer for Leishmania population genetic studies. Trends Parasitol. 31, 52–59
- 2 Tibayrenc, M. and Ayala, F.J. (2012) Reproductive clonality of pathogens: a perspective on pathogenic viruses, bacteria, fungi, and parasitic protozoa. *Proc. Natl. Acad. Sci. U.S.A.* 109, E3305–E3313
- 3 Tibayrenc, M. and Ayala, F.J. (2013) How clonal are *Trypanosoma* and *Leishmania? Trends Parasitol.* 29, 264–269
- 4 Tibayrenc, M. et al. (1991) Are eukaryotic microorganisms clonal or sexual? A population genetics vantage. Proc. Natl. Acad. Sci. U.S.A. 88, 5129–5133
- 5 Alam, M.Z. et al. (2009) The paraphyletic composition of Leishmania donovani zymodeme MON-37 revealed by multilocus microsatellite typing. Microbes Infect. 11, 707-715
- 6 Gouzelou, E. et al. (2012) Multilocus microsatellite typing (MLMT) of strains from Turkey and Cyprus reveals a novel monophyletic L. donovani sensu lato group. PLoS Negl. Trop. Dis. 6, e1507
- 7 Ayala, F.J. and Escalante, A. (1996) The evolution of human populations: a molecular perspective. Mol. Phylogenet. Evol. 5, 188–201
- 8 Llewellyn, M.S. et al. (2009) Trypanosoma cruzi IIc: phylogenetic and phylogeographic insights from sequence and microsatellite analysis and potential impact on emergent Chagas disease. PLoS Negl. Trop. Dis. 3, e510
- 9 Sterkers, Y. et al. (2012) Novel insights into genome plasticity in eukaryotes: mosaic aneuploidy in Leishmania. Mol. Microbiol. 86, 15–23
- 10 Lachaud, L. et al. (2014) Constitutive mosaic aneuploidy is a unique genetic feature widespread in the Leishmania genus. Microbes Infect. 16, 61–66
- 11 Downing, T. et al. (2011) Whole genome sequencing of multiple Leishmania donovani clinical isolates provides insights into population structure and mechanisms of drug resistance. Genome Res. 21, 2143–2156

# Response to Tibayrenc *et al.*: can recombination in *Leishmania* parasites be so rare?

# Virginie Rougeron<sup>1,2\*</sup>, Thierry De Meeûs<sup>3,4\*</sup>, and Anne-Laure Bañuls<sup>1</sup>

The letter by Tibayrenc and Ayala [1] disagrees with several statements on the population genetics of *Leishmania* parasites that we recently published [2]. They consider that these parasites display a preponderant clonal evolution (PCE)

nation, which is thus supposed to represent the best model fitting *Leishmania* population genetics data.

model, suggesting no evidence of frequent sexual recombi-

Confusing selfing and clonality

The first argument appearing in Tibayrenc and Ayala's letter is that 'most scientists working on pathogens consider selfing as a particular case of clonality'. Tibayrenc and Ayala self-cite their own paper [3] where, if we retrieve self-citations and some other papers where authors never

Corresponding author: Rougeron, V. (rougeron.virginie@gmail.com). Keywords: Leishmania; selfing; population genetics; clonality.

\*These two authors contributed equally to this work.

1471-4922/

© 2015 Elsevier Ltd. All rights reserved. http://dx.doi.org/10.1016/j.pt.2015.05.006



<sup>&</sup>lt;sup>1</sup> Maladies Infectieuses et Vecteurs: Ecologie, Génétique, Evolution et Contrôle (MIVEGEC), Unité Mixte de Recherche 5290 (UMR), Centre National de la Recherche Scientifique (CNRS) – Institut de Recherche pour le Développement (IRD) 224 – Université de Montpellier, Montpellier, France

<sup>&</sup>lt;sup>2</sup> Centre International de Recherches Médicales de Franceville BP769 (CIRMF), Franceville, Gabon

<sup>&</sup>lt;sup>3</sup> IRD/Centre International de Recherche-Développement sur l'Elevage en zone Subhumide (CIRDES), UMR177, INTERTRYP IRD-Centre de Coopération Internationale en Recherche Agronomique pour le Développement (CIRAD), CIRDES 01, BP 454 Bobo-Dioulasso 01, Burkina Faso

<sup>&</sup>lt;sup>4</sup> IRD/Centre International de Recherche-Développement sur l'Elevage en zone Subhumide (CIRDES), UMR177, INTERTRYP IRD, TA A-17/G, Campus International de Baillarguet, 34398 Montpellier Cedex 5, France

wrote that selfing was a particular case of clonality (e.g., [4]) or even clearly made the distinction between clonality and selfing [5], we are left with only few remaining articles (most of which refer to Tibayrenc and Ayala's papers) asserting that selfing and clonality are the same thing. Then, Tibayrenc and Ayala's PCE model proposes that the evolutionary consequences of strict clonality and strong selfing are similar in that they lead to linkage disequilibrium (LD) and propagation of unchanged multilocus repeated genotypes (MLGs) [3]. In other words, the authors assimilate individual inbreeding as evidence of clonality [3]. The only common feature between clonality and selfing is that strongly inbred populations (i.e., small populations of extremely small sizes) will produce the MLGs that clonality classically generates [6]. It should be noted that genetic drift in small populations will also lead finally to LD as well as repeated MLGs over time, even in sexually reproducing populations that display no selfing (T. de Meeûs, personal communication). Thus, considering LD and repeated MLGs as key clues for defining clonality or selfing is not always appropriate. Moreover, it is relevant to distinguish between selfing and clonality because population genetic studies will use different models that will lead to different inferences. This is obvious in diploids, where clonality generates heterozygote excesses [7], while selfing leads to homozygosity excesses [8]. From an evolutionary point of view, selfing and clonality are different. Selfers undertake meiosis at each generation, which leads to higher mutation rates compared with mitotic propagation [9]. Thus, recurrent genome-wide reshuffling will recombine those new and more numerous mutations in selfers, while, for clonal propagators, the only way to restore damaged (mutated) important sites will be through localized mitotic recombination, such as gene conversion.

#### LD and inappropriate sampling strategies

Tibayrenc and Ayala state again that one of the criteria to be taken into account for the PCE model is LD. To avoid that Wahlund effect in LD, the authors recommend performing broad sampling across the whole ecogeographical range of species [1]. Computer simulations showed that LD does not provide reliable measurements to make the distinction between clonal and sexual reproductions [10]. Moreover, in our review, we specifically underlined that spatiotemporal effects, combined with inbreeding and/or clonality, produce unpredictable effects on LDs. By considering a broad range of sampling, how would the PCE model proposed by Tibayrenc and Ayala give any valuable information on the reproductive strategies used by Leishmania parasites based on this genetic parameter? We do not understand how the near clade and Russian doll criteria proposed would unravel such combined spatiotemporal Wahlund effects with selfing and/or clonal propagation and small effective local population sizes. In any case, such criteria would need to be validated by analytical modeling or simulations.

Aneuploidy and population genetics in Leishmania Tibayrenc and Ayala explain that, even if *Leishmania* are aneuploid organisms, heterozygosity purging at each

haploid stage would remain and, thus, the recurrent heterozygote deficits that we (and several others) obtained in population genetic studies are irrelevant. Nevertheless, they fail to explain why more than two alleles were never reported in any population genetics survey (and there are many), as expected in cases of hereditary aneuploidy, because aneuploidy is known to involve numerous cases of tri-, tetra-, and pentazomic chromosomes [11]. This is why we suggest that these aneuploidy states are more likely transient. Moreover, the life-stage specificity of aneuploidy deduced by Inbar *et al.* from their experimental studies strongly supports this assumption [12].

#### Recombination in Leishmania parasites

Given the above arguments, those illustrated in our review, and in the absence of any theoretical or experimentally controlled validation of Tibayrenc and Ayala's PCE, near clade, and Russian dolls criteria [1], we are left with the reasonable conclusion that most *Leishmania* populations recurrently experience sexual recombination, mainly endogamic and less frequently allogamic. These assessments are in agreement with the results obtained in most empirical population genetics studies with accurate sampling strategies, as well as experimental studies.

#### **Acknowledgments**

We are grateful to the IRD and the CNRS for financial support. This work was also supported by the French National Project ANR 06- SEST-20 IAEL. We would like to thank the anonymous referee for their comments that helped improve this letter.

#### References

- 1 Tibayrenc, M. and Ayala, F.J. (2015) Leishmania population genetics: clonality, selfing and aneuploidy. Trends Parasitol. 31, 279–280
- 2 Rougeron, V. et al. (2015) A primer for Leishmania population genetic studies. Trends Parasitol. 31, 52–59
- 3 Tibayrenc, M. and Ayala, F.J. (2012) Reproductive clonality of pathogens: a perspective on pathogenic viruses, bacteria, fungi, and parasitic protozoa. *Proc. Natl. Acad. Sci. U.S.A.* 109, E3305–E3313
- 4 Birky, C.W. (2010) Giardia sex? Yes, but how and how much? Trends Parasitol. 26, 70–74
- 5 Lehmann, T. et al. (2004) Variation in the structure of Toxoplasma gondii and the roles of selfing, drift, and epistatic selection in maintaining linkage disequilibria. Infect. Genet. Evol. 4, 107-114
- 6 Halkett, F. et al. (2005) Tackling the population genetics of clonal and partially clonal organisms. Trends Ecol. Evol. 20, 194–201
- 7 Balloux, F. et al. (2003) The population genetics of clonal and partially clonal diploids. Genetics 164, 1635–1644
- 8 De Meeûs, T. et al. (2007) Population genetics and molecular epidemiology or how to 'débusquer la bête.'. Infect. Genet. Evol. 7, 308–332
- 9 Séré, M. et al. (2014) Null allele, allelic dropouts or rare sex detection in clonal organisms: simulations and application to real data sets of pathogenic microbes. Parasit. Vectors 7, 331
- 10 De Meeûs, T. and Balloux, F. (2004) Clonal reproduction and linkage disequilibrium in diploids: a simulation study. *Infect. Genet. Evol.* 4, 345–351
- 11 Rogers, M.B. et al. (2014) Genomic confirmation of hybridisation and recent inbreeding in a vector-isolated *Leishmania* population. PLoS Genet. 10, e1004092
- 12 Inbar, E. et al. (2013) The mating competence of geographically diverse Leishmania major strains in their natural and unnatural sand fly vectors. PLoS Genet. 9, e1003672