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Article

Against the Odds: Hybrid Zones between Mangrove Killifish Species with Different Mating Systems

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Abstract: Different mating systems are expected to affect the extent and direction of hybridization. Due to the different levels of sexual conflict, the weak inbreeder/strong outbreeder (WISO) hypothesis predicts that gametes from self-incompatible (SI) species should outcompete gametes from self-compatible (SC) ones. However, other factors such as timing of selfing and unilateral incompatibilities may also play a role on the direction of hybridization. In addition, differential mating opportunities provided by different mating systems are also expected to affect the direction of introgression in hybrid zones involving outcrossers and selfers. Here, we explored these hypotheses with a unique case of recent hybridization between two mangrove killifish species with different mating systems, *Kryptolebias ocellatus* (obligately outcrossing) and *K. hermaphroditus* (predominantly self-fertilizing) in two hybrid zones in southeast Brazil. Hybridization rates were relatively high (~20%), representing the first example of natural hybridization between species with different mating systems in vertebrates. All F1 individuals were sired by the selfing species. Backcrossing was small, but mostly asymmetrical with the SI parental species, suggesting pattern commonly observed in plant hybrid zones with different mating systems. Our findings shed light on how contrasting mating systems may affect the direction and extent of gene flow between sympatric species, ultimately affecting the evolution and maintenance of hybrid zones.

Keywords: asymmetric introgression; *Kryptolebias*; mixed mating; reproductive isolation; self-fertilization

1. Introduction

Hybridization is a major source of evolutionary innovation, with important implications for phenotypic diversification, adaptation and ultimately speciation [1,2]. Prezygotic barriers, such as spatial, temporal, and behavioral differences play an important role in regulating the extent of hybridization amongst sympatric species in nature [3]. In addition, differences in mating systems (defined as the proportion of self-fertilization versus outcrossing [4]) can largely influence the extent of hybridization, as well as the direction of gene

flow and introgression [5,6]. For example, in monkeyflowers (genus *Mimulus*), differences in the mating system (predominantly selfing vs obligate outcrossing) have a strong effect as a prezygotic mechanism, leading to nearly complete reproductive isolation [6–8].

Natural crosses between self-compatible (SC) and self-incompatible (SI) plants generally produce viable embryos only when SI pollen (male parent) fertilizes SC styles (female parent), but not vice-versa. This pattern of unilateral incompatibility (UI) is known as the ‘SI × SC rule’ [9,10]. This rule could be explained by the weak inbreeder/strong outbreeder (WISO) hypothesis [11], according to which the higher magnitude of maternal-paternal conflict would result in outcrossers’ gametes outcompeting selfer’s gametes in crosses between species with different mating systems. In fact, sperm from outcrossers and/or species with higher levels of sperm competition are more successful at fertilization [12–14]. The WISO hypothesis could, therefore, explain the patterns of UI often observed in the ‘SI × SC rule’ that, despite some exceptions [9,15], is commonly observed in plants [11,16–20]. The ‘SI × SC rule’ has also been detected in some artificial interspecific crosses of *Caenorhabditis* nematodes, with males of outcrossing species having more competitive sperm than selfing males [14]. However, the opposite pattern was also observed in *Caenorhabditis* interspecific crosses involving other species [21], where more progeny was generated in crosses involving selfer males and outcrossing hermaphrodites than the ones in the opposite direction. These mixed results call for further research into the generality of the ‘SI × SC rule’ in animals, particularly under natural conditions.

In addition to the differential levels of genome conflict proposed by the WISO hypothesis, other factors can affect the direction of hybridization between species with different mating systems. SC organisms can self-fertilize before (prior selfing), during (competing selfing) or after (delayed selfing) outcrossing [22]. These differential selfing timings are likely to affect the opportunities for outcrossing, either by conspecific or heterospecific mates [5,23]. Prior selfing is known to represent a strong reproductive barrier for hybridization compared to the other types of selfing [24]. Contrasting mating systems can also influence the direction of introgression [5,25]. Given the possibility of both selfing and outcrossing in the F1s, the predictions about later hybrid generations in hybrid zones with SI and SC species will depend on the mating system of the hybrids themselves [19]. Although UI may emerge postzygotically according to the direction of introgression [26], in hybrid zones between SI and SC species, F1 individuals are expected to backcross more often with the SI than SC parent [5,19], creating an asymmetrical (from SC into SI) pattern of introgression. This prediction relies on the fact that highly selfing taxa usually have stronger reproductive isolation due to the low availability of gametes to outcross [22,27]. Therefore, male F1s are more likely to interact with ‘fertilizable’ eggs from the SI when compared to SC parental species [5].

Unlike plants, relatively little is known about the dynamics of hybridization between animals with different mating systems [28]. Most animals, in particular vertebrates, are dioecious (i.e., different individuals are either male or female) and therefore unable to self-fertilize [29]. *Kryptolebias hermaphroditus*, the mangrove killifish, is one of two known examples of self-fertilizing hermaphroditism in vertebrates [30]. Its populations mostly consists of selfing hermaphrodites and very rare males [31,32]. *Kryptolebias hermaphroditus* coexists with the outcrossing species *K. ocellatus* in the mangrove forests of southeast Brazil (Figures 1a and 2) [33]. Intermediate levels of individual heterozygosity in some populations suggest that outcrossing, likely to only happen between males and hermaphrodites in mangrove killifishes [34], sometimes occurs in *K. hermaphroditus*, but selfing is the major mode of reproduction in the species [35]. In mangrove killifishes, selfing occurs internally, with most of the eggs laid externally being already fertilized via selfing by the hermaphrodites (e.g., prior selfing) [36,37]. The frequency of prior selfing and, consequently the availability of unfertilized eggs for outcrossing, is influenced by environmental factors, such as temperature [38]. In contrast, *K. ocellatus*’ populations are composed by males and hermaphrodites in approximately equal ratio. Although selfing cannot be fully discarded in *K. ocellatus*, breeding experiments only generated offspring when males and

hermaphrodites were kept together [39]. In addition, population genetic studies using microsatellites across most of the species' distribution showed a general lack of deviation from Hardy-Weinberg equilibrium across loci, strongly suggesting that *K. ocellatus* is an exclusively outcrossing species [33,40]. The two species are genetically very divergent, approximately 10% at mtDNA [33], which is a divergence level higher than is commonly observed in 'well-established vertebrate species pairs [41,42]. *Kryptolebias ocellatus* is likely to be the sister species of the common ancestor between the selfing mangrove killifishes *K. hermaphroditus* and *K. marmoratus* [30,43].

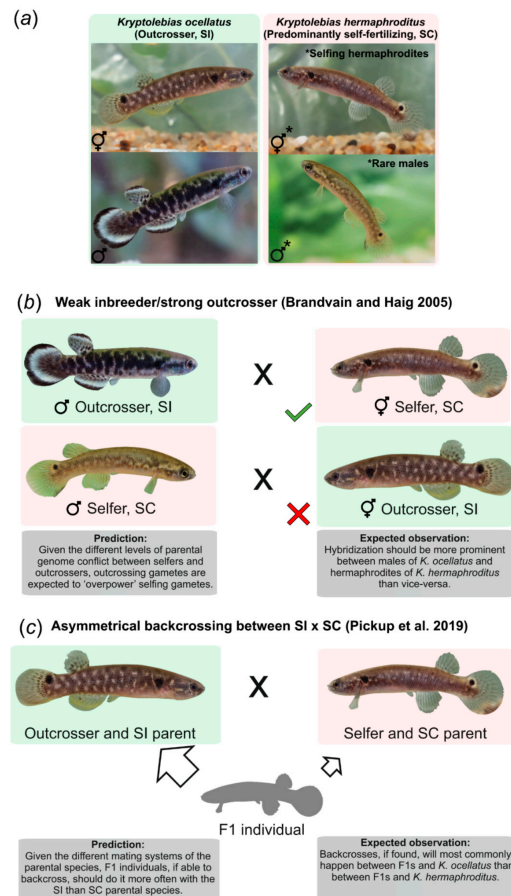


Figure 1. Mangrove killifish species and schematic representation of predictions and expected observations. (a) Mangrove killifish species with respective mating systems. Mating system classifications for *Kryptolebias ocellatus* and *K. hermaphroditus* mating were retrieved from [33,35], respectively. 'SI' refers to self-incompatible. 'SC' denotes self-compatible. (b) Representation for the weak inbreeder, strong outcrosser hypothesis (WISO) [11]. This hypothesis predicts that outcrossers have higher potential for genome conflict given the higher likelihood of crossing between divergent genomes when compared to selfers. This higher potential for genome conflict should reflect on the level of gametes competitiveness with outcrossers gametes being able 'overpower' selfers gametes [11]. Thus, the prediction is that hybridization between species with different mating systems should occur more often between males of the outcrossing species with hermaphrodites of selfers (as competitive sperm should overcome the potential reproductive barrier imposed by the less competitive selfers eggs). The opposite direction is less likely to happen as outcrossers' eggs should impose a stronger barrier for the less competitive sperm from selfers. (c) Representation for asymmetrical backcrossing between SI and SC hypothesis [5]. Given the differences between parental mating systems, if F1 individuals are viable and able to backcross, the direction of backcrossing should be biased towards the SI parent (higher gametes transmission) rather than the SC (gametes partially fertilized through selfing) parent.

Table 1. Sampling localities and sampling sizes for *Kryptolebias ocellatus* (Koce) and *Kryptolebias hermaphroditus* (Kher) in south and southeast Brazil. Microsatellites information for Guaratiba (GUA in Figure 1) was extracted from individuals sampled in two different sampling periods, 2017 and 2007. ‘Msats’ refers to sampling sizes for microsatellites. ‘SNPs’ refers to final sampling sizes using single-nucleotide polymorphisms. ‘Reference for Msats’ refer to references from which microsatellite data was extracted. RJ, Rio de Janeiro State; SC, Santa Catarina State. Asterisks denote sympatric populations.

| Sample ID | Location | Latitude | Longitude | Msats (Koce/Kher) | SNPs (Koce/Kher) | Reference for Msats |
|-----------|--|---------------|----------------|----------------------|---------------------|--|
| FUN 2017* | Fundão mangrove, Rio de Janeiro, RJ | 22°52′2.50″ S | 43°13′27.50″ W | 11/16 | 6/10 | Berbel-Filho et al. [33]/This study |
| GUA 2017* | Piracão mangrove, Guaratiba, RJ | 23°0′1.90″ S | 43°34′51.50″ W | 19/16 | 11/12 | Berbel-Filho et al. [33]/This study |
| GUA 2007* | Piracão mangrove, Guaratiba, RJ | 23°0′1.90″ S | 43°34′51.50″ W | 24/10 | -/- | Tatarenkov et al. [40] |
| SFR | Linguado channel, São Francisco do Sul, SC | 26°22′0.02″ S | 48°39′58.40″ W | 19/- | 7/- | Berbel-Filho et al. [33] |
| FLO | Rio Ratonés estuary, Florianópolis, SC | 27°28′3.84″ S | 48°29′33.76″ W | 30/- | 7/- | Berbel-Filho et al. [33] |
| Total | | | | 103/42 | 31/22 | |

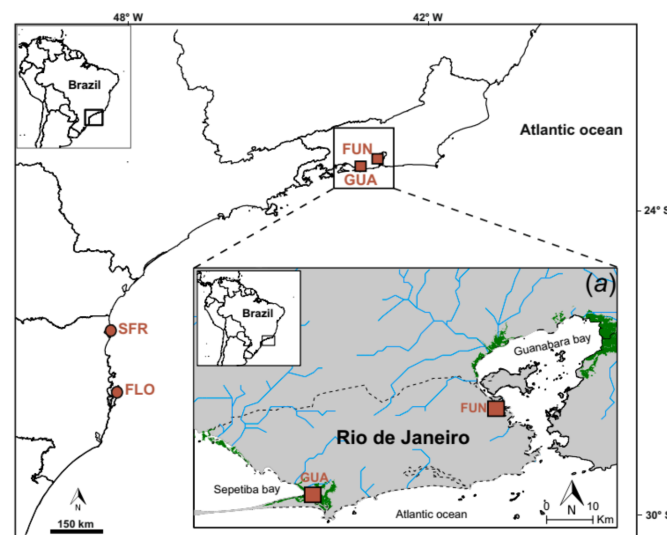


Figure 2. Sampling sites for the individuals included in the genetic analysis. Site names and details included in Table 1. Brown squares represent syntopic populations of *Kryptolebias ocellatus* and *K. hermaphroditus* and circles represent populations where only *K. ocellatus* was found. (a) Detailed map of the Rio de Janeiro municipality and its surrounding bays and mangroves.

The sympatry between the SI and obligately outcrossing *K. ocellatus* and the SC and predominantly selfing *K. hermaphroditus* [33] make these species a unique vertebrate system to understand the evolutionary dynamics of hybrid zones in animals with dissimilar mating systems. Here we take advantage of this system to explore, for the first time in vertebrates, some of the existing hypotheses regarding hybrid zones containing SC and SI species. First, given the low availability of gametes for outcrossing due to prior selfing in highly selfing organisms (such as *K. hermaphroditus*), hybridization rates between SC and SI taxa are expected to be generally low [5]. Second, although we cannot formally test WISO hypothesis without an experimental approach, based on its predictions, we expect that most of the natural hybrids to be sired by the outcrosser species (*K. ocellatus*), rather than by the selfer species (*K. hermaphroditus*) (Figure 1b). Alternatively, given the prior selfing nature of the SC *K. hermaphroditus*, substantial differences in mating opportunities in the wild could result in the opposite crossing direction (hereafter called the ‘mating

opportunity' hypothesis). Finally, given the differences in mating systems of the parental species (and therefore availability of fertilizable eggs), we expect that backcrossing, if existent, should be biased towards the SI parental species. In this case, we expect to find more backcross individuals between *K. ocellatus* and F1s than between *K. hermaphroditus* and F1s (Figure 1c).

2. Materials and Methods

2.1. Sampling Design and Genetic Markers

We analyzed two different genetic datasets. First, we analyzed microsatellite data for 103 *K. ocellatus* individuals (all from two previous datasets collected either in 2017 [33] or 2007 [40]) and 42 *K. hermaphroditus* individuals (32 sampled in 2017 and genotyped in this study; 10 sampled in 2007 [40]) from four localities in south and southeast Brazil (Figure 2; Table 1). The species are sympatric in two of those locations, Guaratiba and Fundão (GUA and FUN respectively in Figure 2), at the west and east limits, respectively, of the Rio de Janeiro municipality in Brazil [32,33]. The other two localities (SFR and GUA in Figure 2) represent an area only inhabited by *K. ocellatus*. Sampling was carried out under license ICMBio/SISBIO 57145-1/2017. Fish species were identified morphologically and confirmed by cytochrome oxidase subunit I (*cox1*) barcoding [33]. For the newly sampled material (*K. hermaphroditus* sampled in 2017; Table 1) we used a set of 16 microsatellites from Mackiewicz, et al. [44]. Micro-checker v. 2.2 (van Oosterhout et al. 2004) was used to check for errors or presence of null alleles. FSTAT v. 2.9.3.2 [45] was used to retrieve basic statistics from microsatellite data, which are provided in Supplementary Table S2.

Although microsatellites allowed us to genotype many individuals, the few loci analyzed could result in incomplete information about the genome-wide admixture proportions. Therefore, we expanded our sampling of genomic loci by obtaining new genomic data using a methylation sensitive genotype-by sequencing (msGBS) from pectoral-fin samples of 55 hermaphrodite individuals (33 *K. ocellatus* and 22 *K. hermaphroditus* sampled in 2017) (Table 1). Genomic DNA was extracted using Qiagen[®] DNeasy Blood and Tissue kit (Qiagen[®], Hilden, Germany) following the manufacturer's protocol. msGBS libraries were prepared as described in Kitimu, et al. [46]. In brief, extracted DNA was digested using restriction enzymes EcoRI and HpaII and ligated to sequencing adapters. A single library was produced by pooling 20 ng of processed DNA from each restriction/ligation product and amplified in eight separate PCR reactions which were pooled after amplification, size-selected (range 200–350 bp) and sequenced in a single lane of an Illumina NextSeq500 sequencer.

Paired-end reads were demultiplexed using GBSX v 1.3 [47]. We then filtered (-qtrim r; -minlength 25) and merged the reads by individual using BBmap tools [48] mapped to the *Kryptolebias marmoratus* reference genome [49] using Bowtie 2 v. 2.2.3 and generated filtered and indexed individual BAM files with SAMtools v. 1.9 [50]. Given the closer phylogenetic proximity between *K. hermaphroditus* and *K. marmoratus* (species from the reference genome) [43], a higher number of uniquely mapped reads was found between *K. hermaphroditus* and *K. marmoratus* (average 89%), than between *K. ocellatus* and *K. marmoratus* (average 80%) (Supplementary Table S1). To call genotypes across all samples, we used ANGSD v 0.9.2.9 [51]. Given the methylation sensitivity of HpaII and the differential mapping efficacy between species, we only allowed for a maximum of 5% of missing data per loci across all samples. Single- and double-tons were removed and, using SAMtools genotype likelihood model, we estimated posterior genotype probabilities assuming a uniform prior (-doPost 2). We also used the ANGSD (-SNP_pval 1×10^{-6}) to carry out a Likelihood Ratio Test to compare between the null (minimum allele frequency = 0) and alternative (estimated minimum allele frequency) hypotheses by using a Chi-squared distribution with one degree of freedom. More details about the library preparation and data processing are provided in the supplementary material.

These analyses produced two genomic datasets for 53 samples: dataset I with 597,733 sites, with average coverage per individual between 12.0X and 346.5X (mean

145.2X) and average missing data per individual ranging from 0% to 7.2% (mean 0.50%), and dataset II with 5477 SNPs, average coverage per individual between 12.4X and 382.6X (mean 152.9X) and average missing data per individual ranging from 0% to 4.9% (mean 0.34%) (Supplementary Table S1). A strong correlation ($R^2 = 0.93$, $p < 0.001$) between the size of the 3073 scaffolds of the *K. marmoratus* reference genome and the number of SNPs from each scaffold indicated that the SNPs were evenly distributed throughout the reference genome.

2.2. Population Genetics and Hybridization Analysis with Microsatellite and SNP Data

To investigate the structure and direction of the potential hybrid zones between *K. ocellatus* and *K. hermaphroditus*, STRUCTURE 2.3.4 [52] was used with microsatellite data with the following parameters: 10 iterations per K (K ranging 2–10), a total of 1,000,000 MCMC, 100,000 burn-in, admixture model and independent allele frequencies. Independent STRUCTURE runs were aligned and plotted using CLUMPAK [53]. To identify the uppermost hierarchical level of genetic structure, we chose the most likely K value using second-order rate of change of likelihood ΔK method [54], implemented in Structure Harvester [55]. For the microsatellites, genotypic associations among individuals were visualized using the factorial correspondence analysis (FCA) implemented in GENETIX v. 4.04 [56]. FCA is similar to principal component analysis but using categorical variables (in our case microsatellite genotypes) [57]. To estimate individual ancestries for the SNP data (dataset II), we used ngsAdmix v. 3.2 [58] with K ranging between 2–10 for 100 replicates using default parameters, except for tolerance for convergence ($-\text{tol } 1 \times 10^{-6}$), log likelihood difference in 50 iterations ($-\text{tolLike50 } 1 \times 10^{-3}$), and a maximum number of EM iterations ($-\text{maxiter } 10,000$). A pairwise genetic distance matrix was computed directly from the genotype likelihoods using ngsDist v.1.0.2 [59] and was then used for Multidimensional Scaling (MDS) using the R package *cmdscale*.

The proportion of heterozygote sites per individuals was calculated with ANGSD to compute the unfolded global estimate of the Site Frequency Spectrum (SFS) for dataset I. The observed fraction of heterozygous sites was calculated as the ratio between the number of heterozygotes and the total number of sites with information.

We used NEWHYBRIDS v. 1.1 [60] to estimate the posterior probability of each individual belonging to one of the parental species, F1 hybrids, F2 or backcrosses between F1 and each parental species, based on their allele frequencies. The analysis was run using the default genotype proportions, uniform prior option, burn-in period of 50,000 iteration and 300,000 MCMC sweeps. With the SNP data, we used individuals from FUN and GUA (sympatric populations) for both species (39 individuals: 17 *K. ocellatus*, 22 *K. hermaphroditus*). We called SNPs with the same parameters described above using ANGSD, but this time with no missing data allowed and selected those (3108 SNPs) with the highest pairwise F_{ST} values between species. Pairs of SNPs with significant LD were removed and randomly replaced with other SNPs to complete a dataset of 200 SNPs (the upper limit of NEWHYBRIDS). We then ran NEWHYBRIDS v.1.1 with the same parameters described for the microsatellites to investigate the posterior probability of each individual to belong to one of the six hybrid classes.

3. Results

Both microsatellite genotypes and SNPs confirmed the hybridization between the outcrossing and SI *K. ocellatus* and the predominantly selfing SC *K. hermaphroditus* in two hybrid zones (FUN and GUA in Figure 2) in Southeast Brazil (Figure 3).

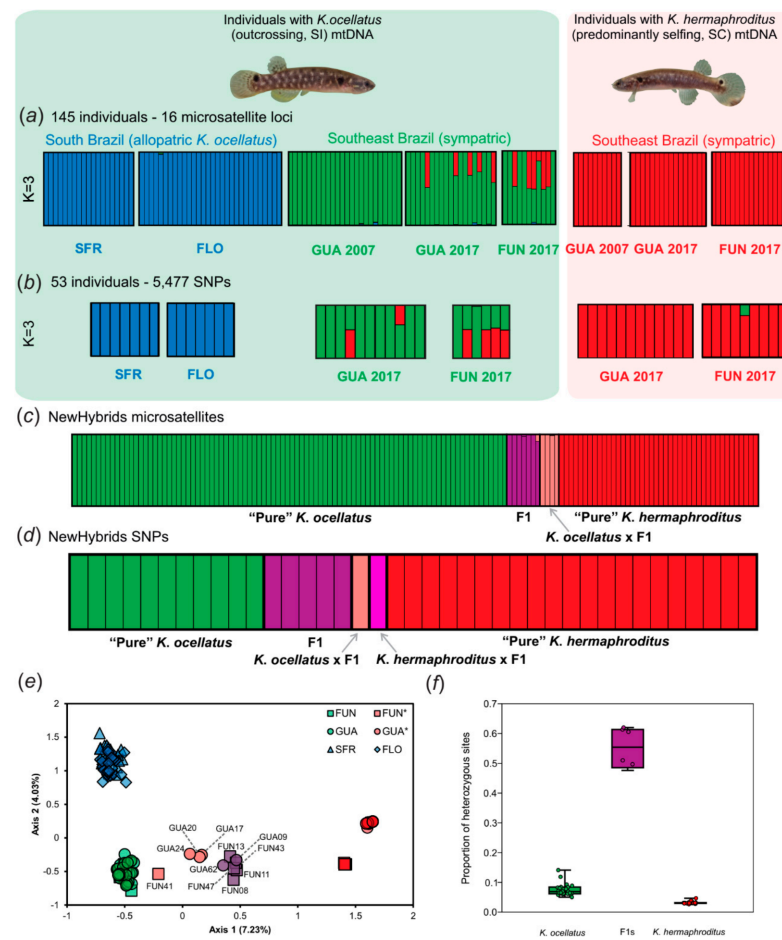


Figure 3. Genetic analyses for the hybrid zone between *Kryptolebias ocellatus* and *K. hermaphroditus* in Southeast Brazil. Admixture plots showing the genetic clusters ($K = 3$) for the (a) 16 microsatellites loci amplified in *K. ocellatus* and *K. hermaphroditus* according to STRUCTURE and (b) for the 5477 SNPs according to ngsAdmix (see Methods for details). Each individual is represented by a bar, and each color represents a genetic cluster. (c,d) NEWHYBRIDS individual classification into hybrid classes (*K. ocellatus*, *K. hermaphroditus*, F1, F2, *K. ocellatus* \times F1 and *K. hermaphroditus* \times F1) using microsatellite and SNPs data. (e) Factorial correspondence analysis using microsatellites data for all *K. ocellatus* and *K. hermaphroditus* individuals colored (green and blue for *K. ocellatus* from Southeast and South Brazil, respectively; red for *K. hermaphroditus*) and shaped according to sampling sites (squares, circles, triangles, and diamonds represent individuals from FUN, GUA, SFR and FLO, respectively). Hybrid individuals (see Results) are highlighted with their respective labels and colored (purple for F1s, pink for backcrosses) according to the hybrid class indicated by NEWHYBRIDS analysis. Sampling locations with asterisks represent *K. hermaphroditus* individuals. (f) Proportion of heterozygous sites between *K. ocellatus*, *K. hermaphroditus* and F1 hybrid individuals.

Altogether, microsatellites and SNPs identified 12 individuals with admixed genomes between *K. ocellatus* and *K. hermaphroditus*, seven of those from Fundão in Guanabara Bay (FUN 08, 11, 13, 26, 41, 43, 47) and five from Guaratiba in Sepetiba Bay (GUA 09, 17, 20, 24, 62) all of them sampled in 2017 (Figure 3a,b). The only disagreement between microsatellite and SNP data occurred with one individual from Fundão (FUN 26), that was classified as a pure *K. hermaphroditus* individual with the microsatellites but showed evidence of genetic admixture with SNPs data (see results below). Five individuals identified as hybrids by microsatellite data were not included in the SNPs analyses as they failed to produce our threshold of reads for the GBS library (cut-off ≥ 500 k reads) two from FUN (FUN 13, 41) and three from GUA (GUA 20, 24, 62) (Supplementary Table S1). The individual ancestry analyses (STRUCTURE for microsatellites and ngsAdmix for SNP data) supported the

identification of the hybrids (Figure 3a,b). At $K = 3$ (the most likely K according to Evanno's ΔK method) indicated that all *K. ocellatus* were assigned with nearly 100% probability to one cluster, while *K. hermaphroditus* individuals were assigned with nearly 100% probability to another cluster, apart from the subset of the divergent FUN and GUA fish, with admixed genetic backgrounds of both species. The third cluster consisted of the southernmost *K. ocellatus* individuals (from the allopatric populations SFR and FLO), reflecting the deep genetic structuring previously found for this species, with isolated populations occupying discontinuous patches of mangrove forests between Southeast and South in Brazil [33]. For both microsatellite and SNP data, the other K values tested indicated admixture between *K. ocellatus* and *K. hermaphroditus* genomes in the subset of FUN and GUA individuals (Supplementary Figures S1 and S2).

Overall, 12 out of 145 individuals analyzed (8.2%) had admixed ancestry of *K. ocellatus* and *K. hermaphroditus*. However, when considering only the sympatric populations FUN and GUA, the proportion of admixed individuals increases to 12.5%. Finally, when considering only contemporary sympatric populations from the most recent sampling effort (FUN and GUA from 2017, as no admixed individuals have been detected in the GUA microsatellite dataset from 2007) the proportion of admixed individuals in FUN and GUA in 2017 increases to 19.3%. Of these, eleven of the 30 individuals with *K. ocellatus* mtDNA (36.6%) had admixed nuclear genomes, while only one (3.1%) of the 32 with *K. hermaphroditus* mtDNA had evidence of admixture with *K. ocellatus* at the nuclear genome (Table 1; Figure 3a,b).

The WISO hypothesis predicts that hybridization should be most likely between males of the SI species *K. ocellatus* and hermaphrodites of the SC species *K. hermaphroditus*. Accordingly, F1 hybrids should predominately have *K. hermaphroditus* mtDNA haplotypes. Contrary to this prediction, NEWHYBRIDS analysis with both microsatellite and SNP data revealed seven F1 hybrids (FUN 08, 11, 13, 47, 48; GUA 09 and 62), all of them with mtDNA of the outcrossing species *K. ocellatus*. This result reveals that the most prominent direction of hybridization was between the outcrossing and SI *K. ocellatus* hermaphrodites and the SC *K. hermaphroditus* males. NEWHYBRIDS results also revealed four (FUN 41; GUA 17, 20, 24) backcrosses between *K. ocellatus* and a F1 hybrid, and one as a backcross between *K. hermaphroditus* and a F1 hybrid (FUN 26, only indicated with SNP data). This partial evidence of asymmetrical direction of backcrossing is consistent with the prediction that introgression is most common from the SC into the SI species. No individual, regardless of the genetic marker, was identified as a F2 (Figure 3c,d). All hybrids, except for FUN 26, had a mtDNA *cox1* haplotype typical of the outcrossing SI species *K. ocellatus*. In terms of overall genomic variation, both factorial correspondence and multidimensional scaling analyses positioned the F1s and backcrosses in between the parental genotypes along the first axes, while the variation along the second axis separated the *K. ocellatus* populations from south and southeast Brazil (Figure 3e and Supplementary Figure S3).

4. Discussion

Differences in mating systems are expected to influence the extent, direction, and the structure of hybrid zones involving SI and SC species [5,19,25]. Given the strong reproductive isolation of highly selfing species, such as *K. hermaphroditus* [30,35], theory predicts that there would be low hybridization rates in hybrid zones involving SI \times SC species [5]. Contrary to this prediction, we found relatively high hybridization rates between *K. ocellatus* (obligately outcrossing) and *K. hermaphroditus* (predominantly selfing), with admixed individuals representing approximately 20% of the individuals in the co-existing populations. To our knowledge, this represents the first case of hybridization between vertebrate species with different mating systems. Contrary to the expectations of the 'SI \times SC rule' [11] our data indicates that all seven F1 hybrids were sired by the rare males of predominantly selfing species *K. hermaphroditus*. Given the difficulties to detangle mating opportunities and hybrid viability in natural populations, we acknowledge that our results do not represent a formal test of the weak inbreeder/strong outbreeder

(WISO) hypothesis [11], which would require an experimental manipulation of mating opportunities, a task that has been proven particularly challenging for mangrove killifishes in laboratory conditions [36,44,61]. Although our sample size of backcrosses was small, those were mostly asymmetrical, showing a bias (four out of five) towards backcrosses between F1s and the SI parental species, *K. ocellatus*. A higher rate of backcrossing from the SC to SI species is consistent with the asymmetrical direction of introgression commonly observed in plant hybrid zones with different mating systems [5–7,19].

In most killifish species, egg fertilization occurs externally [62]. However, in the selfing mangrove killifishes, most of the eggs laid externally by the hermaphrodites are already fertilized internally via selfing [36,37]. This reproductive trait, similar to the ‘prior selfing’ in plants [22], leaves a limited window of opportunity for fertilization via outcrossing by either the (rare) conspecific or heterospecific males. Given that evidence suggest that outcrossing in the predominantly selfing mangrove killifishes only happens between male and hermaphrodites [34,44], outcrossing opportunities involving this species depend both on the density and mating activity of sexually active males as well as the availability of unfertilized eggs [36]. Despite extensive historical sampling, particularly in southeast Brazil, males of the SC *K. hermaphroditus* seem to be rare or even absent in some populations [35,63]. In fact, males of this species were only discovered recently [31,32]. The WISO hypothesis predicts that SI gametes should represent a strong barrier to fertilization for gametes of SC species. This prediction was not supported by our results, as all F1s were sired by males of the selfing *K. hermaphroditus*. With our data, we could not formally distinguish whether this pattern was caused by the potential differential reproductive barriers caused by gamete incompatibilities (WISO) or due to differential mating opportunities in the wild for crosses of opposite directions to happen. However, interspecific laboratory crosses of *Caenorhabditis* nematode species also revealed the opposite pattern to the SI × SC rule [21], suggesting that there are exceptions to this rule, as previously observed in plant hybrids [9,15]. In plants, it has been suggested that exceptions to the SI × SC rule occur in species where SC recently evolved from SI (Brandvain and Haig; 2005). Yet, the timing of the transition from SI to SC within *Kryptolebias* genus is unknown. Future research is also needed to investigate whether occasional outcrossing among divergent selfing lineages of *K. hermaphroditus* [35] may maintain competitiveness of the SC gametes (and therefore higher levels of sperm-egg conflict), possibly attenuating the effects of relaxed selection on sperm performance and other sexually selected traits due to prolonged selfing [a common effect of the selfing syndrome, 28].

Hybrids between highly divergent species (such as *K. ocellatus* and *K. hermaphroditus* [33,43]) are expected to harbor many genetic incompatibilities caused by the gradual accumulation of divergent alleles [64,65]. Although higher-resolution genomic data is still needed to investigate the possibility of ancestral introgression in this system, the absence of admixture in the one of the hybrids zones in recent collections (GUA in 2007) and the lack of further hybrid generations beyond F1s suggest that the hybrid zones found here are relatively recent, and the reproductive isolation between *K. ocellatus* and *K. hermaphroditus* is strong, despite being incomplete. The mangrove forests (FUN and GUA) where hybridization was found are situated within Guanabara and Sepetiba Bays, respectively. Those bays surround the Rio de Janeiro municipality and are amongst the most polluted Brazilian estuaries [66]. Water pollutants are known to disrupt physiology, mate choice and reproductive isolation between coexisting species [67,68]. The fact that we have not found evidence for hybridization in GUA samples from 2007, but found it in high rates in 2017, suggests that a recent environmental change (possibly human-induced) may have disrupted some of the reproductive barriers between syntopic *K. hermaphroditus* and *K. ocellatus* populations. Further sampling in the area is urgently needed to shed light the potential links between hybridization rates and environmental stressors across a larger number of sympatric populations.

The contrasting nature of genetic load between selfers and outcrossers [25] could provide a potential explanation for the observed structure and direction of backcrossing

found in our study. While outcrossers are expected to have many highly deleterious recessive alleles, selfers are expected to purge those and have many mildly deleterious codominant alleles [5]. Selfed progeny of F1s between selfing and outcrossing species are expected to have low fitness given the increased exposure of an outcrosser's recessive load in homozygosity. Although we cannot fully rule out the possibility that F1s between *K. ocellatus* and *K. hermaphroditus* are able to self-fertilize, our results indicate that they are at least able to outcross via backcrossing. Therefore, it might be possible that high genetic load (with many mildly deleterious codominant alleles) originating from selfers via introgression is quickly removed by natural selection from the outcrossing population. This would result in hybrid zones only containing early generation hybrids [5], with some individuals of *K. ocellatus* still bearing self-derived ancestry given recent backcrossing, but with no evidence of ancestral introgression. Further research is needed to investigate whether *K. ocellatus* individuals with self-derived ancestry show any reduction of fitness compared to 'pure' *K. ocellatus* individuals.

Another property of the hybrid zone between *K. ocellatus* and *K. hermaphroditus* is its asymmetry (unidirectional hybridization, and partial evidence for asymmetrical backcrossing). Asymmetrical hybridization suggests differential strengths of barriers to gene flow according to the direction of hybridization [5]. The influence of mating system differences vs selection and/or other reproductive barriers on hybridization zones are hard to disentangle in natural populations [6,69]. Although we cannot rule out the possibility of postzygotic unilateral incompatibility [also known as the 'Darwin's corollary to Haldane's rule, 26], the asymmetries in the direction of hybridization and the small but biased ratio of backcrossing from SC to SI suggest that differential mating, or more precisely, differential fertilization opportunities (the 'mating opportunity' hypothesis) may have also had a role in the structure of the hybrid zones. This hypothesis is substantiated by some life-history details of the species involved: (i) the rare males of the SC *K. hermaphroditus* species are more likely to find heterospecific than conspecific unfertilized eggs available for fertilization, given the 'prior selfing' nature of its conspecific hermaphrodites; (ii) backcrossing, although small, was bidirectional, revealing that at least some of the offspring between F1s and *K. hermaphroditus* are viable. Potential male F1s are more likely to find fertilizable eggs laid by the SI *K. ocellatus* than the SC *K. hermaphroditus*, while potential female F1 individuals would also be more likely to be fertilized by the more abundant males of the SI *K. ocellatus* than the rare males in SC *K. hermaphroditus*. Although we observe a slight bias of backcrosses from SC to SI (4:1 ratio), the overall small number of backcrosses calls for further research on the role of mating opportunities on backcrossing asymmetry. In addition to the increased fertilization opportunity, the propensity of males of the SC *K. hermaphroditus* to fertilize heterospecific eggs could be intensified by its patterns of disassortative mating. In the SC *K. marmoratus*, sister species of *K. hermaphroditus* [30,43], males tend to associate with genetically dissimilar hermaphrodites [70]. If this pattern of disassortative association holds true for *K. hermaphroditus* in a heterospecific context, both egg availability and behavioral preferences may have influenced the asymmetric direction of hybridization observed here. Further studies are needed to elucidate the patterns of assortative mating among *K. ocellatus*, *K. hermaphroditus*, and their hybrids.

5. Conclusions

Investigating the influence of different prezygotic barriers in hybrid zones is crucial for the understanding of the evolutionary consequences of hybridization, the evolution of reproductive isolation, and ultimately, speciation. Although the role of contrasting mating systems in hybrid zones has been largely explored in plants [5,7,11,20], empirical studies in animals are scarce. Our study contributed to this understanding by exploring, for the first time in a vertebrate system, the effect of different mating systems (predominantly self-fertilizing vs outcrossing) in natural hybrid zones. We observed hybridization between a predominantly self-fertilizing and an outcrossing fish species that differed in several aspects from what has been observed in other hybrid zones between SI and SC taxa,

particularly in plant systems. First, hybridization rates were relatively high for a highly selfing taxa with prior selfing such as *K. hermaphroditus*. All F1 hybrids were sired by the selfing species, providing a potential exception for the SC \times SI rule in a natural animal system. Although these findings partially challenge the predictions of the WISO hypothesis, further studies are necessary to detangle the role of mating opportunities and/or hybrid viability according to the crosses directions.

While hard to disentangle from other reproductive barriers, the asymmetry in hybridization and backcrossing found here suggest that fertilization opportunities may have had a strong role in shaping the direction of gene flow between the mangrove killifish species. In addition to influencing the degree of reproductive isolation between sympatric species, asymmetrical introgression can also affect the reproductive compatibility within species between allopatric and sympatric populations, if the latter are experiencing asymmetrical introgression [71]. Further studies are needed to investigate whether the asymmetric gene flow of *K. hermaphroditus* genome into *K. ocellatus* in the hybrid zones has impacted the reproductive isolation between allopatric vs sympatric *K. ocellatus* populations.

Supplementary Materials: The following are available online at <https://www.mdpi.com/article/10.3390/genes12101486/s1>, Figure S1: Individual ancestry plots with the highest likelihood runs for each K value ran in STRUCTURE with genotypes for 16 microsatellites for 103 *K. ocellatus* and 42 *K. hermaphroditus* individuals. Asterisk denotes locations for individuals with *K. hermaphroditus* mtDNA. Figure S2. Individual ancestry plots with each K value ran in nsgAdmix with 5477 SNPs from 31 *K. ocellatus* individuals and 22 *K. hermaphroditus* individuals. Asterisk denotes locations for individuals with *K. hermaphroditus* mtDNA. Figure S3. Multidimensional Scaling (MDS) based on the genetic distances from 5477 SNPs. Hybrid individuals (see results) are highlighted with their respective labels. Table S1. Summary of 58 samples sequenced in the genotype-by-sequencing (msGBS) library. Individuals in red failed to pass the filtering cutoff (≥ 500 kb reads). Superscript letters on ID denotes F1 and backcrosses. Parameters 'proportion of heterozygous sites' (for Dataset I), 'coverage and missing data' (for Dataset II) are described in Material and Methods. Table S2. Descriptive statistics of genetic variation at microsatellite loci in (a) 103 *Kryptolebias ocellatus* and (b) 42 *K. hermaphroditus* individuals. N = sample size; P99 = proportion of polymorphic loci (99% criterion); A = average number of alleles; HE = expected heterozygosity; HO = observed heterozygosity. Sampling locations are described in Table 1. Supplementary File 2 contains sample information, mtDNA sequences and microsatellite genotypes for all samples involved in the study.

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References

1. Janko, K.; Pačes, J.; Wilkinson-Herbots, H.; Costa, R.J.; Roslein, J.; Drozd, P.; Iakovenko, N.; Rídl, J.; Hroudová, M.; Kočí, J. Hybrid asexuality as a primary postzygotic barrier between nascent species: On the interconnection between asexuality, hybridization and speciation. *Mol. Ecol.* **2018**, *27*, 248–263. [[CrossRef](#)]
2. Mitchell, N.; Owens, G.L.; Hovick, S.M.; Rieseberg, L.H.; Whitney, K.D. Hybridization speeds adaptive evolution in an eight-year field experiment. *Sci. Rep.* **2019**, *9*, 6746. [[CrossRef](#)]
3. Chunco, A.J. Hybridization in a warmer world. *Ecol. Evol.* **2014**, *4*, 2019–2031. [[CrossRef](#)] [[PubMed](#)]
4. Barrett, S.C. Evolution of mating systems: Outcrossing versus selfing. In *The Princeton Guide of Evolution*; Losos, J.B., Ed.; Princeton University Press: Princeton, NJ, USA, 2014; pp. 356–362.
5. Pickup, M.; Brandvain, Y.; Fraïsse, C.; Yakimowski, S.; Barton, N.H.; Dixit, T.; Lexer, C.; Cereghetti, E.; Field, D.L. Mating system variation in hybrid zones: Facilitation, barriers and asymmetries to gene flow. *New Phytol.* **2019**, *224*, 1035–1047. [[CrossRef](#)] [[PubMed](#)]
6. Brandvain, Y.; Kenney, A.M.; Flagel, L.; Coop, G.; Sweigart, A.L. Speciation and introgression between *Mimulus nasutus* and *Mimulus guttatus*. *PLoS Genet.* **2014**, *10*, e1004410. [[CrossRef](#)] [[PubMed](#)]
7. Martin, N.H.; Willis, J.H. Ecological divergence associated with mating system causes nearly complete reproductive isolation between sympatric *Mimulus* species. *Evolution* **2007**, *61*, 68–82. [[CrossRef](#)]
8. Zuellig, M.P.; Sweigart, A.L. A two-locus hybrid incompatibility is widespread, polymorphic, and active in natural populations of *Mimulus*. *Evolution* **2018**, *72*, 2394–2405. [[CrossRef](#)] [[PubMed](#)]
9. Lewis, D.; Crowe, L.K. Unilateral interspecific incompatibility in flowering plants. *Heredity* **1958**, *12*, 233–256. [[CrossRef](#)]
10. Murfett, J.; Strabala, T.J.; Zurek, D.M.; Mou, B.; Beecher, B.; McClure, B.A. S RNase and interspecific pollen rejection in the genus *Nicotiana*: Multiple pollen-rejection pathways contribute to unilateral incompatibility between self-incompatible and self-compatible species. *Plant Cell* **1996**, *8*, 943–958. [[CrossRef](#)] [[PubMed](#)]
11. Brandvain, Y.; Haig, D. Divergent mating systems and parental conflict as a barrier to hybridization in flowering plants. *Am. Nat.* **2005**, *166*, 330–338. [[CrossRef](#)]
12. Palopoli, M.F.; Peden, C.; Woo, C.; Akiha, K.; Ary, M.; Cruze, L.; Anderson, J.L.; Phillips, P.C. Natural and experimental evolution of sexual conflict within *Caenorhabditis* nematodes. *BMC Evol. Biol.* **2015**, *15*, 93. [[CrossRef](#)] [[PubMed](#)]
13. Martín-Coello, J.; Benavent-Corai, J.; Roldan, E.R.; Gomendio, M. Sperm competition promotes asymmetries in reproductive barriers between closely related species. *Evol. Int. J. Org. Evol.* **2009**, *63*, 613–623. [[CrossRef](#)]
14. Ting, J.J.; Woodruff, G.C.; Leung, G.; Shin, N.-R.; Cutter, A.D.; Haag, E.S. Intense sperm-mediated sexual conflict promotes reproductive isolation in *Caenorhabditis* nematodes. *PLoS Biol.* **2014**, *12*, e1001915. [[CrossRef](#)]
15. Liedl, B.; McCormick, S.; Mutschler, M. Unilateral incongruity in crosses involving *Lycopersicon pennellii* and *L. esculentum* is distinct from self-incompatibility in expression, timing and location. *Sex. Plant Reprod.* **1996**, *9*, 299–308. [[CrossRef](#)]
16. Li, L.; Liu, B.; Deng, X.; Zhao, H.; Li, H.; Xing, S.; Fetzer, D.D.; Li, M.; Nasrallah, M.E.; Nasrallah, J.B. Evolution of interspecies unilateral incompatibility in the relatives of *Arabidopsis thaliana*. *Mol. Ecol.* **2018**, *27*, 2742–2753. [[CrossRef](#)]
17. Baek, Y.S.; Covey, P.A.; Petersen, J.J.; Chetelat, R.T.; McClure, B.; Bedinger, P.A. Testing the SI × SC rule: Pollen–pistil interactions in interspecific crosses between members of the tomato clade (*Solanum* section *Lycopersicon*, Solanaceae). *Am. J. Bot.* **2015**, *102*, 302–311. [[CrossRef](#)]
18. Li, W.; Chetelat, R.T. Unilateral incompatibility gene *ui1.1* encodes an S-locus F-box protein expressed in pollen of *Solanum* species. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 4417–4422. [[CrossRef](#)] [[PubMed](#)]
19. Ruhsam, M.; Hollingsworth, P.; Ennos, R. Early evolution in a hybrid swarm between outcrossing and selfing lineages in *Geum*. *Heredity* **2011**, *107*, 246–255. [[CrossRef](#)] [[PubMed](#)]
20. İltaş, Ö.; Svitok, M.; Cornille, A.; Schmickl, R.; Placette, C.L. Early evolution of reproductive isolation: A case of weak inbreeder/strong outbreeder leads to an intraspecific hybridization barrier in *Arabidopsis lyrata*. *Evolution* **2021**, *75*, 1466–1476. [[CrossRef](#)]
21. Kozłowska, J.L.; Ahmad, A.R.; Jahesh, E.; Cutter, A.D. Genetic variation for postzygotic reproductive isolation between *Caenorhabditis briggsae* and *Caenorhabditis* sp. 9. *Evol. Int. J. Org. Evol.* **2012**, *66*, 1180–1195. [[CrossRef](#)]
22. Lloyd, D.G. Some reproductive factors affecting the selection of self-fertilization in plants. *Am. Nat.* **1979**, *113*, 67–79. [[CrossRef](#)]
23. Goodwillie, C.; Weber, J.J. The best of both worlds? A review of delayed selfing in flowering plants. *Am. J. Bot.* **2018**, *105*, 641–655. [[CrossRef](#)]
24. Brys, R.; Van Cauwenberghe, J.; Jacquemyn, H. The importance of autonomous selfing in preventing hybridization in three closely related plant species. *J. Ecol.* **2016**, *104*, 601–610. [[CrossRef](#)]
25. Kim, B.Y.; Huber, C.D.; Lohmueller, K.E. Deleterious variation shapes the genomic landscape of introgression. *PLoS Genet.* **2018**, *14*, e1007741. [[CrossRef](#)] [[PubMed](#)]
26. Turelli, M.; Moyle, L.C. Asymmetric postmating isolation: Darwin’s corollary to Haldane’s rule. *Genetics* **2007**, *176*, 1059–1088. [[CrossRef](#)] [[PubMed](#)]

27. Brys, R.; Broeck, A.V.; Mergeay, J.; Jacquemyn, H. The contribution of mating system variation to reproductive isolation in two closely related *Centaureum* species (Gentianaceae) with a generalized flower morphology. *Evolution* **2014**, *68*, 1281–1293. [[CrossRef](#)] [[PubMed](#)]
28. Cutter, A.D. Reproductive transitions in plants and animals: Selfing syndrome, sexual selection and speciation. *New Phytol.* **2019**, *224*, 1080–1094. [[CrossRef](#)]
29. Jarne, P.; Auld, J.R. Animals mix it up too: The distribution of self-fertilization among hermaphroditic animals. *Evolution* **2006**, *60*, 1816–1824. [[CrossRef](#)] [[PubMed](#)]
30. Tatarenkov, A.; Lima, S.M.Q.; Earley, R.L.; Berbel, W.M.; Vermeulen, F.B.M.; Taylor, D.S.; Marson, K.; Turner, B.J.; Avise, J.C. Deep and concordant subdivisions in the self-fertilizing mangrove killifishes (*Kryptolebias*) revealed by nuclear and mtDNA markers. *Biol. J. Linn. Soc.* **2017**, *122*, 558–578. [[CrossRef](#)]
31. Berbel-Filho, W.M.; Espirito-Santo, H.M.V.; Lima, S.M.Q. First record of a male of *Kryptolebias hermaphroditus* Costa, 2011 (Cyprinodontiformes: Cynolebiidae). *Neotrop. Ichthyol.* **2016**, *14*, e160024. [[CrossRef](#)]
32. Costa, W.J.E.M. Colouration, taxonomy and geographical distribution of mangrove killifishes, the *Kryptolebias marmoratus* species group, in southern Atlantic coastal plains of Brazil (Cyprinodontiformes: Rivulidae). *Ichthyol. Explor. Freshw.* **2016**, *27*, 183–192.
33. Berbel-Filho, W.M.; Tatarenkov, A.; Espirito-Santo, H.M.; Lira, M.G.; De Leaniz, C.G.; Lima, S.M.; Consuegra, S. More than meets the eye: Syntopic and morphologically similar mangrove killifish species show different mating systems and patterns of genetic structure along the Brazilian coast. *Heredity* **2020**, *125*, 140–352. [[CrossRef](#)] [[PubMed](#)]
34. Furness, A.I.; Tatarenkov, A.; Avise, J.C. A genetic test for whether pairs of hermaphrodites can cross-fertilize in a selfing killifish. *J. Hered.* **2015**, *106*, 749–752. [[CrossRef](#)]
35. Berbel-Filho, W.M.; de Leaniz, C.G.; Morán, P.; Cable, J.; Lima, S.M.; Consuegra, S. Local parasite pressures and host genotype modulate epigenetic diversity in a mixed-mating fish. *Ecol. Evol.* **2019**, *9*, 8736–8748. [[CrossRef](#)] [[PubMed](#)]
36. Lomax, J.L.; Carlson, R.E.; Wells, J.W.; Crawford, P.M.; Earley, R.L. Factors affecting egg production in the selfing mangrove rivulus (*Kryptolebias marmoratus*). *Zoology* **2017**, *122*, 38–45. [[CrossRef](#)] [[PubMed](#)]
37. Harrington, R.W. How ecological and genetic factors interact to determine when self-fertilizing hermaphrodites of *Rivulus marmoratus* change into functional secondary males, with a reappraisal of the modes of intersexuality among fishes. *Copeia* **1971**, *1971*, 389–432. [[CrossRef](#)]
38. Park, C.-B.; Kim, Y.J.; Soyano, K. Effects of increasing temperature due to aquatic climate change on the self-fertility and the sexual development of the hermaphroditic fish, *Kryptolebias marmoratus*. *Environ. Sci. Pollut. Res.* **2017**, *24*, 1484–1494. [[CrossRef](#)]
39. Seegers, L. Zur revision der *Rivulus*-arten sudost-Brasilens, mit einer Neubeschreibung von *Rivulus luelingi* n. spp. und *Rivulus caudomarginatus* n. spp. (Pisces: Cyprinodontidae: Rivulinae). *Zool. Beitr.* **1984**, *28*, 271–320.
40. Tatarenkov, A.; Lima, S.M.Q.; Taylor, D.S.; Avise, J.C. Long-term retention of self-fertilization in a fish clade. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 14456–14459. [[CrossRef](#)] [[PubMed](#)]
41. Hebert, P.D.; Stoeckle, M.Y.; Zemlak, T.S.; Francis, C.M. Identification of birds through DNA barcodes. *PLoS Biol.* **2004**, *2*, e312. [[CrossRef](#)] [[PubMed](#)]
42. Ward, R.D.; Hanner, R.; Hebert, P.D. The campaign to DNA barcode all fishes, FISH-BOL. *J. Fish Biol.* **2009**, *74*, 329–356. [[CrossRef](#)]
43. Kanamori, A.; Sugita, Y.; Yuasa, Y.; Suzuki, T.; Kawamura, K.; Uno, Y.; Kamimura, K.; Matsuda, Y.; Wilson, C.A.; Amores, A.; et al. A genetic map for the only self-fertilizing vertebrate. *G3 Genes Genom. Genet.* **2016**, *6*, 1095–1106. [[CrossRef](#)] [[PubMed](#)]
44. Mackiewicz, M.; Tatarenkov, A.; Perry, A.; Martin, J.R.; Elder, J.F.; Bechler, D.L.; Avise, J.C. Microsatellite documentation of male-mediated outcrossing between inbred laboratory strains of the self-fertilizing mangrove killifish (*Kryptolebias marmoratus*). *J. Hered.* **2006**, *97*, 508–513. [[CrossRef](#)] [[PubMed](#)]
45. Goudet, J. FSTAT (version 1.2): A computer program to calculate F-statistics. *J. Hered.* **1995**, *86*, 485–486. [[CrossRef](#)]
46. Kitimu, S.R.; Taylor, J.; March, T.J.; Tairo, F.; Wilkinson, M.J.; Rodríguez López, C.M. Meristem micropropagation of cassava (*Manihot esculenta*) evokes genome-wide changes in DNA methylation. *Front. Plant Sci.* **2015**, *6*, 590. [[CrossRef](#)]
47. Herten, K.; Hestand, M.S.; Vermeesch, J.R.; Van Houdt, J.K. GBSX: A toolkit for experimental design and demultiplexing genotyping by sequencing experiments. *BMC Bioinform.* **2015**, *16*, 73. [[CrossRef](#)]
48. Bushnell, B. *BBMap: A Fast, Accurate, Splice-Aware Aligner*; Lawrence Berkeley National Lab. (LBNL): Berkeley, CA, USA, 2014.
49. Rhee, J.-S.; Choi, B.-S.; Kim, J.; Kim, B.-M.; Lee, Y.-M.; Kim, I.-C.; Kanamori, A.; Choi, I.-Y.; Scharl, M.; Lee, J.-S. Diversity, distribution, and significance of transposable elements in the genome of the only selfing hermaphroditic vertebrate *Kryptolebias marmoratus*. *Sci. Rep.* **2017**, *7*, 40121. [[CrossRef](#)] [[PubMed](#)]
50. Li, H.; Handsaker, B.; Wysoker, A.; Fennell, T.; Ruan, J.; Homer, N.; Marth, G.; Abecasis, G.; Durbin, R. Genome Project Data Processing, S. The sequence alignment/map format and SAMtools. *Bioinformatics* **2009**, *25*, 2078–2079. [[CrossRef](#)]
51. Korneliussen, T.S.; Albrechtsen, A.; Nielsen, R. ANGSD: Analysis of next generation sequencing data. *BMC Bioinform.* **2014**, *15*, 356. [[CrossRef](#)] [[PubMed](#)]
52. Pritchard, J.K.; Stephens, M.; Donnelly, P. Inference of population structure using multilocus genotype data. *Genetics* **2000**, *155*, 945–959. [[CrossRef](#)]
53. Kopelman, N.M.; Mayzel, J.; Jakobsson, M.; Rosenberg, N.A.; Mayrose, I. CLUMPAK: A program for identifying clustering modes and packaging population structure inferences across K. *Mol. Ecol. Resour.* **2015**, *15*, 1179–1191. [[CrossRef](#)]
54. Evanno, G.; Regnaut, S.; Goudet, J. Detecting the number of clusters of individuals using the software STRUCTURE: A simulation study. *Mol. Ecol.* **2005**, *14*, 2611–2620. [[CrossRef](#)]

55. Earl, D.A. STRUCTURE HARVESTER: A website and program for visualizing STRUCTURE output and implementing the Evanno method. *Conserv. Genet. Resour.* **2012**, *4*, 359–361. [[CrossRef](#)]
56. Belkhir, K. GENETIX 4.05, Logiciel Sous Windows TM pour la Génétique des Populations. 2004. Available online: <http://www.genetix.univ-montp2.fr/genetix/genetix.htm> (accessed on 29 May 2020).
57. Sourial, N.; Wolfson, C.; Zhu, B.; Quail, J.; Fletcher, J.; Karunanathan, S.; Bandeen-Roche, K.; Béland, F.; Bergman, H. Correspondence analysis is a useful tool to uncover the relationships among categorical variables. *J. Clin. Epidemiol.* **2010**, *63*, 638–646. [[CrossRef](#)] [[PubMed](#)]
58. Skotte, L.; Korneliussen, T.S.; Albrechtsen, A. Estimating individual admixture proportions from next generation sequencing data. *Genetics* **2013**, *195*, 693–702. [[CrossRef](#)] [[PubMed](#)]
59. Vieira, F.G.; Lassalle, F.; Korneliussen, T.S.; Fumagalli, M. Improving the estimation of genetic distances from Next-Generation sequencing data. *Biol. J. Linn. Soc.* **2015**, *117*, 139–149. [[CrossRef](#)]
60. Anderson, E.; Thompson, E. A model-based method for identifying species hybrids using multilocus genetic data. *Genetics* **2002**, *160*, 1217–1229. [[CrossRef](#)]
61. Nakamura, Y.; Suga, K.; Sakakura, Y.; Sakamoto, T.; Hagiwara, A. Genetic and growth differences in the outcrossings between two clonal strains of the self-fertilizing mangrove killifish. *Can. J. Zool.* **2008**, *86*, 976–982. [[CrossRef](#)]
62. Costa, W.J.; Amorim, P.F.; Mattos, J.L.O. Molecular phylogeny and evolution of internal fertilization in South American seasonal cynopoeciline killifishes. *Mol. Phylogenet. Evol.* **2016**, *95*, 94–99. [[CrossRef](#)] [[PubMed](#)]
63. Costa, W.J.E.M. Identity of *Rivulus ocellatus* and a new name for a hermaphroditic species of *Kryptolebias* from south-eastern Brazil (Cyprinodontiformes: Rivulidae). *Ichthyol. Explor. Fres.* **2011**, *22*, 185–192.
64. Coughlan, J.M.; Matute, D.R. The importance of intrinsic postzygotic barriers throughout the speciation process. *Philos. Trans. R. Soc. B* **2020**, *375*, 20190533. [[CrossRef](#)]
65. Coyne, J.A.; Orr, H.A. “Patterns of speciation in *Drosophila*” revisited. *Evolution* **1997**, 295–303.
66. Barletta, M.; Lima, A.R.A. Fish ecology and anthropogenic impacts in South American estuaries: Setting priorities for ecosystem conservation. *Front. Mar. Sci.* **2019**, *6*, 237. [[CrossRef](#)]
67. Kime, D.E. The effects of pollution on reproduction in fish. *Rev. Fish Biol. Fish.* **1995**, *5*, 52–95. [[CrossRef](#)]
68. Candolin, U.; Wong, B.B. Mate choice in a polluted world: Consequences for individuals, populations and communities. *Philos. Trans. R. Soc. B* **2019**, *374*, 20180055. [[CrossRef](#)]
69. Ruhsam, M.; Hollingsworth, P.M.; Ennos, R.A. Patterns of mating, generation of diversity, and fitness of offspring in a *Geum* hybrid swarm. *Evolution* **2013**, *67*, 2728–2740. [[CrossRef](#)]
70. Ellison, A.; Jones, J.; Inchley, C.; Consuegra, S. Choosy males could help explain androdioecy in a selfing fish. *Am. Nat.* **2013**, *181*, 855–862. [[CrossRef](#)]
71. Ostevik, K.L.; Rifkin, J.L.; Xia, H.; Rausher, M.D. Morning glory species co-occurrence is associated with asymmetrically decreased and cascading reproductive isolation. *Evol. Lett.* **2021**, *5*, 75–85. [[CrossRef](#)] [[PubMed](#)]