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### Title

An experimental test of the effect of predation upon behaviour and trait correlations in the threespine stickleback

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1 **An experimental test of predation's effect upon behaviour and trait correlations**  
2 **in threespine stickleback**

3

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11 **Running title:** Stickleback behaviour correlates with armour

12

13 **Abstract:**

14 Benthic and limnetic threespine stickleback (*Gasterosteus aculeatus*) are a classic  
15 example of ecological speciation. Behavioural divergence between these species has been  
16 predicted to be the result of divergent selection driven in part by differential predation from  
17 cutthroat trout (*Oncorhynchus clarki*). To experimentally test this prediction, we reared  
18 split families of benthic-limnetic hybrids in the presence or absence of trout predation. Our  
19 results show that the presence of trout had little effect upon stickleback behaviour. We  
20 then compared performance in behavioural assays among stickleback that varied in armour.  
21 Our measurements also revealed trait correlations between several behaviours and  
22 components of armour morphology. The strength of the correlations between traits did not  
23 differ between predation treatments therefore differential predation between benthics and  
24 limnetics is unlikely to be the cause of these correlations. The presence of trait correlations  
25 in advanced generation hybrids suggests that pleiotropy or linkage between genes  
26 underlying behaviour and armour morphology may be greater than previously appreciated.

27

28 *Keywords: adaptation – armour – ecological speciation – Gasterosteus aculeatus –*

29 *pleiotropy - shoaling – selection experiment*

## 30 **Introduction**

31 Ecological speciation occurs when reproductive isolation evolves as a consequence of  
32 divergent natural selection between contrasting environments (Schluter, 2009; Nosil, 2012).  
33 While there are many examples of ecological speciation in nature, our understanding of the  
34 underlying mechanisms remains incomplete (Rundle & Nosil, 2005; Nosil, 2012). Divergent  
35 selection can occur in response to differences in resource availability and as a result of  
36 biotic interactions such as predation, competition, or intraguild predation (Schluter, 2000;  
37 2009; Miller, Metcalf, & Schluter, 2015). Experimental studies have shown that differential  
38 predation can lead to the evolution of divergent morphological traits (e.g. Jiggins et al.  
39 2001; Vamosi & Schluter, 2002; Rundle *et al.*, 2003; Nosil & Crespi, 2006; Langerhans,  
40 Gifford, & Joseph, 2007; Diabaté *et al.*, 2008; Marchinko, 2009; Svanbäck & Eklöv, 2011).  
41 However, less attention has been given to the role of divergent selection in the evolution of  
42 behavioural diversity.

43 Benthic and limnetic threespine stickleback (*Gasterosteus aculeatus sp.*) are a classic  
44 example of ecological speciation; the two species have evolved in sympatry in five lakes in  
45 coastal British Columbia (Schluter & McPhail, 1992). The species differ in many  
46 morphological and behavioural traits. Limnetics primarily eat zooplankton in the open  
47 water while benthics consume macroinvertebrates in the littoral zone (Schluter & McPhail,  
48 1992). In the open water, limnetics encounter cutthroat trout (*Oncorhynchus clarki*) more  
49 frequently (Reimchen, 1994). Consequently, many of the phenotypic differences between  
50 the species are thought to be the result of differential predation on limnetics by trout  
51 (Vamosi & Schluter, 2002). Relative to benthics, limnetics have longer spines and more  
52 lateral plates (Vamosi, 2002). Limnetics also have an increased shoaling preference

53 (Vamosi & Schluter, 2002; Wark *et al.*, 2011), and are generally found higher in the water  
54 column (Larson, 1976). In comparison, benthics are more often solitary (Vamosi &  
55 Schluter, 2002; Odling-Smee, Boughman, & Braithwaite, 2008; Wark *et al.*, 2011), and  
56 prefer to be lower in the water column (Larson, 1976).

57 To determine if a trait is the target of divergent selection, indirect evidence from  
58 observational or comparative studies is insufficient (Schluter, 2009). The presence of  
59 aquatic predators can co-vary with environmental factors (*e.g.* abiotic conditions, food  
60 resources) (Jackson, Peres-Neto, & Olden, 2001). Therefore it is necessary to use  
61 controlled experiments manipulating the presence/absence of predators to confirm that trait  
62 shifts are caused by divergent selection from predation. Comparing trait shifts between  
63 species is further problematic because species have fixed differences in many traits. As a  
64 result, it is difficult to separate the trait(s) that are the target of divergent selection from  
65 those traits that are genetically linked but not under direct selection. Predation may also  
66 lead to selection for correlations between advantageous combinations of behaviour and  
67 defence morphology (Sinervo & Svensson, 2002; Murren, 2012). Creating advanced  
68 generation crosses between species with divergent phenotypes can create trait  
69 combinations not normally seen in the wild. When such crosses are combined with  
70 predator manipulation, it is possible to test if predation is responsible for changes in traits  
71 and trait correlations.

72 We experimentally tested the hypothesis that differences in behaviour between  
73 benthic and limnetic stickleback are the result of divergent selection from cutthroat trout  
74 predation. Benthic-limnetic hybrid families were introduced into large, naturalistic  
75 experimental ponds in the presence/absence of trout predation. Experimental stickleback

76 reproduced annually in the ponds and underwent two generations of differential selection  
77 prior to measurement in behavioural assays. We measured two putative anti-predator  
78 behaviours, which have been previously shown to differ between the two species -  
79 preferred position in the water column and shoaling preference (Larson, 1976; Vamosi,  
80 2002; Kozak & Boughman, 2008; Wark *et al.*, 2011). Behaviours that differ consistently  
81 between control and predation ponds can be interpreted to arise in response to trout  
82 predation. We then tested for correlations between behaviour and defensive armour, and  
83 compared the strength of these correlations between treatments. If trout predation selects  
84 for combinations of behaviour and defensive armour, trait correlation will be greater in the  
85 predation treatment.

86

## 87 **Methods**

### 88 *Experimental Design*

89 In May 2011, four F1 crosses were made between wild-caught benthic females and  
90 limnetic males from Paxton Lake, Texada Island. These F1 crosses were reared in 300L  
91 tanks in the laboratory without predators for one year until adulthood. In May 2012, adult  
92 stickleback were collected from First Lake, an advanced generation hybrid population. First  
93 Lake is a small shallow lake on Texada Island that was founded in 1981 with Paxton Lake  
94 benthic x limnetic F1 stickleback (McPhail, 1993). We consider this population to be a  
95 single family of  $\sim$ F29 benthic-limnetic hybrids at the time of sampling. The First Lake  
96 population was included in the study because the greater number of recombination events  
97 this population has undergone affords us the opportunity to investigate the effect of linkage  
98 on adaptation.

99            In May 2012, the five hybrid families (Four F1s and one First Lake) were introduced  
100 in a split plot design to pairs of semi-natural ponds (n=21-31 individuals/pond; 10 ponds  
101 total) at the University of British Columbia's experimental pond facilities. Each paired pond  
102 contained a single family. Stickleback bred in all experimental ponds creating F2s or ~F30s  
103 (First Lake ponds) in the summer of 2012. In the summer of 2013, the F2/F30 stickleback  
104 bred to form a F3/F31 generation. All behavioural assays were conducted on adult  
105 stickleback from the 2013 (F3/F31) cohort.

106            The experimental ponds are 25m x 15m with a shallow littoral area and a 6m deep  
107 open water region. These ponds contain a natural assemblage of food resources and  
108 contain invertebrate and avian predators. For each set of paired ponds, one pond was  
109 randomly assigned to a predation treatment and the other pond to a control treatment.  
110 Adult Cutthroat trout were collected from Placid Lake in the Malcolm Knapp Research  
111 Forest. Two trout were added to each predation pond in September 2012. The trout died  
112 in the summer of 2013 and were replaced with three new trout in September 2013.

113

#### 114 *Behavioural Assays*

115            Behavioural assays were conducted from November 8-14, 2013, in tanks adjacent to  
116 the experimental ponds. Twelve randomly chosen stickleback were collected from each  
117 pond with unbaited minnow traps (n=120 total). Paired ponds were tested sequentially,  
118 alternating between treatments. Sticklebacks were transferred in a bucket from the pond  
119 to the behavioural assay area for a 15-minute acclimation period prior to the start of the  
120 behavioural trials. At that time, each stickleback was placed into an individual mesh basket  
121 inside a larger aquarium so that we could follow the behaviour of individuals across assays.

122 Behavioural tests were conducted in the following order: stickleback were tested in the  
123 novel tank test, returned to the holding basket for 15 minutes, and then tested in the  
124 shoaling assay.

125         The novel tank diving test measures stickleback movement and position in a new  
126 tank. Vertical position in the water column of a tank has been used as a proxy for habitat  
127 usage in guppies and stickleback (Larson, 1976; Torres-Dowdall *et al.*, 2012; Miller *et al.*,  
128 2015). It has also been found that anxiety in zebrafish (e.g. following exposure to alarm  
129 pheromones) leads to a reduction in exploration and a lower position in a tank (Egan *et al.*,  
130 2009; Cachat *et al.*, 2010; Stewart *et al.*, 2012). During the trial, a focal fish was gently  
131 introduced to the top centre of an empty unfamiliar 35.5 cm x 22 cm x 20 cm tank and  
132 allowed to move freely for 630 seconds. All assays were recorded with wireless D-Link  
133 DCS-930L webcams (DLink Corporation, Taiwan). We excluded the first 30 seconds of each  
134 assay as the introduction of a stickleback often resulted in erratic movement (Miller *et al.*,  
135 2015). Videos were subsampled to 0.5 frames per second using VirtualDub software  
136 ([www.virtualdub.org](http://www.virtualdub.org)). The MtrackJ plugin (Meijering, Dzyubachyk, & Smal, 2012) in  
137 ImageJ (Schneider, Rasband, & Eliceiri, 2012) was used to measure the x and y coordinates  
138 of the focal fish every 2 seconds. We calculated the mean vertical position of the focal fish,  
139 the latency to enter the upper half of the tank, and the distance that the focal fish travelled  
140 during the assay.

141         The second assay assesses shoaling preference by measuring the time that the focal  
142 stickleback spends near a stimulus shoal (Vamosi, 2002; Kozak & Boughman, 2008; Wark  
143 *et al.*, 2011). Assay tanks were 75 cm x 30 cm x 46 cm with two 10 cm end compartments  
144 on either side of the tank that were separated from a large centre arena with window

145 screen (Figure S1). Ten stimulus stickleback (shoal) were added to one end compartment  
146 and two stimulus stickleback (distractor) were added to the other end compartment (Wark  
147 et al., 2011). The stimulus sticklebacks were limnetic stickleback from Priest Lake reared at  
148 the experimental pond facility. This population was unrelated and unfamiliar to the  
149 experimental stickleback and was chosen because individuals have a high shoaling  
150 tendency (Wark et al., 2011) and were similar in size to the experimental stickleback. At  
151 the start of the shoaling assay, the focal stickleback was gently introduced into the centre  
152 arena and was allowed to move for 630 seconds. We measured the x and y coordinates of  
153 the focal fish every 2 seconds following the method used in the novel tank test. We used  
154 two metrics to assess shoaling behaviour: the mean horizontal position in the tank  
155 (shoaling position), and the time that the focal fish spends within one body length of the  
156 experimental shoal (shoaling preference).

157       As a result of camera error, two trials were not analysed. Following Wark *et al.*  
158 (2011), we excluded trials in which the focal fish did not move during the trial (novel tank  
159 n=10; shoaling n=12). In total, 110 novel tank trials and 108 shoaling trials were  
160 measured.

161

## 162 *Armour*

163       Immediately following the shoaling assay, stickleback were euthanized in MS-222  
164 and fixed in 10% formalin. Specimens were later stained with alizarin red to highlight bony  
165 structures following established protocols (Peichel et al., 2001). On the left side of each  
166 stained specimen we measured the length of length of the first and second dorsal spines,  
167 pelvic spine, pelvic girdle, the number of lateral plates and standard length. Specimens

168 lacking an armour component were assigned a value of zero. Lateral plate number and  
169 standard length were not significantly correlated. All other armour traits were positively  
170 correlated with standard length and were size corrected to the average length (43.82 mm)  
171 using the equation  $Y_i = X_i - \beta(L_i - \bar{L})$ . Where  $Y_i$  is the size-adjusted trait,  $X_i$  is the original  
172 trait,  $\beta$  is the regression coefficient of the original trait values on standard length,  $L_i$  is the  
173 standard length of the individual and  $\bar{L}$  is the average length (Vamosi, 2002). For second  
174 dorsal spine, pond had a significant effect on  $\beta$  and thus this trait was size corrected  
175 independently for each pond (pond did not have a significant effect for other traits). A  
176 principal component analysis (PCA) of the correlation matrix of size-corrected armour traits  
177 was used to visualize the overall defensive armour of each stickleback. The first principal  
178 component (PC1) accounted for 40.9% of the variation in stickleback armour and primarily  
179 describes the pelvic spine and pelvic girdle (Table S1). The second principal component  
180 (PC2) accounted for 25.8% of the variation and describes the length of the first and second  
181 dorsal spine.

182

### 183 *Statistical Analysis*

184 A linear mixed effects model was used to test if performance in behavioural assays  
185 differed between treatments and if armour traits affected these behaviours. Principal  
186 component score, treatment, and population (Paxton Lake or First Lake) were fixed factors.  
187 Pond and family were random factors. Population was not a significant covariate and was  
188 dropped from the final model.

189 All traits were not normally distributed. Therefore, Spearman's rank correlations  
190 were used to evaluate the correlations between armour and behavioural measurements.

191 Confidence intervals for trait correlations were calculated by bootstrapping (1000 replicates)  
192 with RVAideMemoire (Hervé, 2014). For traits with significant correlations, we compared  
193 the magnitude of the correlations between treatments using the Wilcoxon signed-rank test  
194 on Spearman rank correlations calculated separately for each pond. All statistical analysis  
195 were conducted in R (version 3.1) (R Core Team, 2014)

196

## 197 **Results**

198         The presence of trout did not have a measurable effect upon stickleback behaviour  
199 (Table 1; Figure S2). Predation and control ponds did not differ in vertical position in the  
200 water column, the latency to enter the upper half of the tank, or distance travelled during  
201 the novel tank assay. Fish from all ponds spent more time shoaling than the random  
202 expectation, regardless of treatment (one sample t-test: effect size = \_\_\_\_,  $t_{10}=9.29$ ,  
203  $P<0.0001$ ). In the shoaling assay, we observed a trend of increased time spent with the  
204 shoal (shoaling preference) in the control ponds for four of the five families (Treatment:  
205 effect size = \_\_\_\_,  $F_{1,4}=3.24$ ,  $P=0.15$ ), and focal fish from control ponds travelled more  
206 during the assay (Figure 1; Treatment: effect size = \_\_\_\_,  $F_{1,4}=5.69$ ,  $P=0.08$ ), although  
207 these results were not significant.

208         We observed variation in armour traits among experimental families (Table S2). PC1  
209 differentiated stickleback with robust pelvic armour (limnetic-like) and stickleback with  
210 reduced pelvic armour (benthic-like), while PC2 separated individuals with longer dorsal  
211 spines (limnetic-like) from those with reduced dorsal spines (benthic-like). Predation and  
212 control ponds did not differ in PC1 (Treatment:  $F_{1,4}=0.43$ ,  $P=0.55$ ), PC2 (Treatment:  
213  $F_{1,4}=2.5$ ,  $P=0.18$ ), or standard length (Treatment:  $F_{1,4}=0.19$ ,  $P=0.69$ ).

214           There was a positive correlation between PC1 score and mean vertical position  
215 during the novel tank test (Figure 1A; Spearman's rank correlation coefficient,  $\rho = 0.261$ ,  
216  $P = 0.006$ , 95% CI: 0.068-0.442). Individuals with increased pelvic armour preferred a  
217 higher vertical position in the water column (PC1:  $F_{4,97} = 4.10$ ,  $P = 0.045$ ). There was a  
218 negative correlation between PC2 and distance travelled during the novel tank test (Figure  
219 1C;  $\rho = -0.260$ ,  $P = 0.006$ , 95% CI: -0.428 - 0.071). PC2 and distance travelled during the  
220 shoaling assay were not correlated (Table S3), but there was a significant Treatment x PC2  
221 interaction ( $F_{1,95} = 4.52$ ,  $P = 0.04$ ). One individual had an extreme value for PC2. However,  
222 the correlation between these traits remained significant when this point was removed  
223 (without point,  $\rho = -0.245$ ,  $P = 0.01$ ). Behaviour was not correlated with standard length  
224 (Table S3). All other armour and behaviour correlations were non-significant (Table 1,  
225 Table S3).

226           Trout predation did not change the strength of the correlations between PC1 and  
227 water column position (Figure 1B; Wilcoxon signed-rank test,  $z = 9$ ,  $n = 5$ ,  $P = 0.812$ ), or PC2  
228 and distance travelled during the water column assay (Figure 1D;  $z = 5$ ,  $n = 5$ ,  $P = 0.625$ ).  
229

## 230 **Discussion**

231           Divergent selection from trout predation has been hypothesized to be an important  
232 driver of behavioural differences between benthic and limnetic stickleback (e.g. Larson,  
233 1976; Vamosi, 2002; Vamosi & Schluter, 2004; Wark *et al.*, 2011). We reared families of  
234 benthic-limnetic hybrids in naturalistic experimental ponds in the presence or absence of  
235 trout predation. Contrary to predictions, there was no significant difference in behaviour

236 between predation and control ponds. Instead, armour morphology was a stronger  
237 predictor of behaviour than trout predation.

238

239

#### 240 *Stickleback Behaviour*

241 Preferred position in the water column did not differ between predation and control  
242 ponds. Stickleback in predation ponds had a decreased shoaling preference, but this result  
243 was non-significant. If differences in benthic and limnetic behaviour are not the  
244 consequence of divergent selection from trout predation, then behavioural differences may  
245 be the result of selection from other factors that differ between the benthic and limnetic  
246 habitats. For example, benthics forage for invertebrates in the littoral zone, while limnetics  
247 eat zooplankton near the surface of the water (Larson, 1976; Odling-Smee et al., 2008).  
248 Therefore differences in water column preference may be due to divergence in diet and/or  
249 foraging behaviour between the two species. Similarly, limnetics are frequently observed in  
250 large aggregations (Larson, 1976) and have a stronger shoaling preference than benthics  
251 (Vamosi, 2002; Kozak & Boughman, 2008; Wark et al., 2011). The differences in shoaling  
252 behaviour in the lakes may be due to differences in the structural complexity and amount of  
253 open space between the two environments (Odling-Smee et al., 2008) rather than a  
254 consequence of increased trout predation. A shift in resource or habitat use could also  
255 have driven changes in shoaling preference. Compared to control ponds, predation ponds  
256 had a decrease in population density and a shift in diet towards benthic resources (S.  
257 Rudman, per. comm.). Selection for benthic-like trophic characteristics may have led to a  
258 decrease in shoaling preference. Trout predation may have also led to non-consumptive

259 changes in behaviour by reducing competition and increasing intimidation in the open water  
260 environment (Preisser, Bolnick, & Benard, 2005).

261         The experimental ponds provide an improvement over behavioural studies  
262 conducted in mesocosms or in the laboratory because we were able to manipulate  
263 experimental subjects in a natural environment. Our findings suggest that differential  
264 predation alone is unlikely to explain the large differences in shoaling behaviour and water  
265 column preference observed in the wild. However, the paired design limited the statistical  
266 power of this experiment to detect small differences in behaviour between treatments.  
267 Additionally, behaviours were assayed at a single end point, therefore if paired ponds did  
268 not start at the same trait value this would decrease our ability to detect a treatment effect.

269

#### 270 *Correlation between morphology and behaviour*

271         The likelihood that an individual escapes a predation event may be determined by  
272 an interaction between behavioural and morphological traits (e.g. Brodie, 1992; Dewitt,  
273 1999; Buskirk & McCollum, 2000; Relyea, 2001). We found a correlation between  
274 behavioural traits and bony armour. Armour PC1 (increased pelvic armour) was associated  
275 with a higher position in the water column and armour PC2 (longer dorsal spines) was  
276 associated with increased movement during the water column assay. Functionally these  
277 associations match the greater pelvic armour and preference for a higher water column  
278 position found in limnetics (Larson, 1976). A previous study by Grand (2000) found that  
279 within benthic stickleback that those individuals with reduced pelvic armour were less bold  
280 than individuals with increased pelvic armour. Behavioural traits have high variance and any  
281 measurement error can decrease the correlation between traits (Whitlock & Schluter,

282 2014), as a result, correlations between these traits in the wild are likely greater than  
283 reported in this study.

284         The correlations we found between armour morphology and behaviour could result  
285 from genetic linkage or pleiotropy (Schlosser & Wagner, 2004). Several inferences can be  
286 made regarding the possible genetic basis of the correlations. Recombination events in  
287 advanced generation hybrids should uncouple most traits that were genetically linked in  
288 limnetics and benthics, aside from regions of especially low recombination. Yet three  
289 generations of recombination were insufficient to break up the association between armour  
290 and behaviour in the F3 families and >30 generations of recombination in First Lake ponds  
291 did not decrease the correlation. The maintenance of these correlations in spite of  
292 genome-wide recombination indicates that tight genetic linkage or pleiotropy underlies  
293 these associations.

294         Prior studies in stickleback support a role for tight linkage or pleiotropy between  
295 behaviour and morphology. Lateral plate number and body orientation during schooling  
296 have been genetically mapped to the same chromosomal segment (Greenwood et al.  
297 2013). A single gene (*Ectodysplasin*) in this low recombination region has been previously  
298 shown to have pleiotropic effects upon lateral plate development, neuromast position,  
299 schooling behaviour, and salinity preference (Barrett et al. 2009; Wark & Peichel, 2009;  
300 Wark et al. 2012; Mills et al. 2014). A recent study has also uncovered a correlation  
301 between anti-predator behaviour and pigmentation in juvenile stickleback (Kim & Velando,  
302 2015), suggesting that these correlations may be more widespread than previously  
303 appreciated.

304           When certain trait combinations are preferentially favoured, natural selection may  
305 directly or indirectly lead to an increase in the correlation between these traits (Sinervo &  
306 Svensson, 2002; Murren, 2012). While we describe a correlation between multiple armour  
307 and behavioural traits, the strength of these correlations did not differ between treatments.  
308 Therefore we were unable to support the hypothesis that trout predation is the causal  
309 mechanism for the associations. However, the lack of change in correlation between  
310 treatments could be a consequence of the limited power of our experiment, or insufficient  
311 variation in correlation for selection to act upon. Trout may have also played an important  
312 role during the historical divergence between benthic and limnetic stickleback. Therefore,  
313 while trout predation may not be the proximate cause for the correlation between defence  
314 morphology and behaviour, it may be the ultimate cause for this association. Future work  
315 examining the genetic basis of these traits will be required to elucidate the role of  
316 pleiotropy and tight linkage in behaviour and armour morphology in the threespine  
317 stickleback.

318  
319

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<b>Novel Tank Test</b>	<b>Treatment</b>				<b>PC1</b>				<b>Treatment x PC1</b>			
	df	F	P	95% CI	df	F	P	95% CI	df	F	P	95% CI
<i>Mean Vertical Position</i>	1,4	1.42	0.30	-29.4-4.7	1,97	4.10	<b>0.05</b>	-0.8-17.3	1,97	0.14	0.71	-15.0-10.2
<i>Latency to upper tank</i>	1,4	1.12	0.35	-37.3-77.5	1,97	0.17	0.68	-6.5-35.6	1,97	2.69	0.10	-55.1-5.2
<i>Distance traveled</i>	1,4	0.98	0.38	-4026-1808	1,97	0.78	0.38	-886-1027	1,97	0.61	0.44	-820-1888

**Shoaling Assay**

<i>Mean horizontal position</i>	1,4	0.71	0.45	-1.5-0.77	1,95	0.04	0.83	-0.5-0.4	1,95	0.03	0.86	-0.7-0.6
<i>Shoaling preference</i>	1,4	3.24	0.15	-84.0-11.9	1,95	0.24	0.62	-23.7-16	1,95	0.00	0.98	-28.9-29.7
<i>Distance traveled</i>	1,4	5.69	0.08	-3266-252	1,95	0.00	0.94	-784-511	1,95	0.52	0.47	-571-1226

<b>Novel Tank Test</b>	<b>Treatment</b>				<b>PC2</b>				<b>Treatment x PC2</b>			
	df	F	P	95% CI	df	F	P	95% CI	df	F	P	95% CI
<i>Mean Vertical Position</i>	1,4	1.33	0.31	-34.2-15.7	1,97	0.49	0.49	-16.4-6.0	1,97	0.38	0.54	-11.0-20.8
<i>Latency to upper tank</i>	1,4	0.85	0.41	-44.8-86.6	1,97	0.02	0.88	-39.6-12.5	1,97	3.6	0.06	-1.9-75.6
<i>Distance traveled</i>	1,4	0.90	0.4	-1838-1585	1,97	1.69	0.20	-1696-609	1,97	0.02	0.88	-1838-1585

**Shoaling Assay**

<i>Mean horizontal position</i>	1,4	0.75	0.44	-1.2-0.6	1,95	1.78	0.19	-0.6-0.5	1,95	2.3	0.13	-1.4-0.2
<i>Shoaling preference</i>	1,4	3.27	0.15	-82.7-12.8	1,95	0.81	0.37	-38.0-11.1	1,95	0.39	0.54	-26.1-49.9
<i>Distance traveled</i>	1,4	5.88	0.07	-3269-189	1,95	0.09	0.77	-176-1378	1,95	4.52	<b>0.04</b>	-2380- -82

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