

UC Davis

UC Davis Previously Published Works

Title

Zika Virus Infection Results in Biochemical Changes Associated With RNA Editing, Inflammatory and Antiviral Responses in Aedes albopictus

Permalink

<https://escholarship.org/uc/item/8pt7864b>

Authors

Onyango, Maria G
Attardo, Geoffrey M
Kelly, Erin Taylor
et al.

Publication Date

2020

DOI

10.3389/fmicb.2020.559035

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <https://creativecommons.org/licenses/by/4.0/>

Peer reviewed



Zika Virus Infection Results in Biochemical Changes Associated With RNA Editing, Inflammatory and Antiviral Responses in *Aedes albopictus*

OPEN ACCESS

Edited by:

Akio Adachi,
Kansai Medical University, Japan

Reviewed by:

Rushika Perera,
Colorado State University,
United States
Jinbao Gu,
Southern Medical University, China
Carol D. Blair,
Colorado State University,
United States

*Correspondence:

Laura D. Kramer
laura.kramer@health.ny.gov

Specialty section:

This article was submitted to
Virology,
a section of the journal
Frontiers in Microbiology

Received: 04 May 2020

Accepted: 14 September 2020

Published: 02 October 2020

Citation:

Onyango MG, Attardo GM,
Kelly ET, Bialosuknia SM, Stout J,
Banker E, Kuo L, Ciota AT and
Kramer LD (2020) Zika Virus Infection
Results in Biochemical Changes
Associated With RNA Editing,
Inflammatory and Antiviral Responses
in *Aedes albopictus*.
Front. Microbiol. 11:559035.
doi: 10.3389/fmicb.2020.559035

Maria G. Onyango¹, Geoffrey M. Attardo², Erin Taylor Kelly², Sean M. Bialosuknia^{1,2,3},
Jessica Stout¹, Elyse Banker¹, Lili Kuo¹, Alexander T. Ciota^{1,3} and Laura D. Kramer^{1,3*}

¹ Wadsworth Center, New York State Department of Health, Slingerlands, NY, United States, ² Department of Entomology and Nematology, University of California, Davis, Davis, CA, United States, ³ School of Public Health, State University of New York, Albany, NY, United States

Rapid and significant range expansion of both the Zika virus (ZIKV) and its *Aedes* vector species has resulted in the declaration of ZIKV as a global health threat. Successful transmission of ZIKV by its vector requires a complex series of interactions between these entities including the establishment, replication and dissemination of the virus within the mosquito. The metabolic conditions within the mosquito tissues play a critical role in mediating the crucial processes of viral infection and replication and represent targets for prevention of virus transmission. In this study, we carried out a comprehensive metabolomic phenotyping of ZIKV infected and uninfected *Ae. albopictus* by untargeted analysis of primary metabolites, lipids and biogenic amines. We performed a comparative metabolomic study of infection state with the aim of understanding the biochemical changes resulting from the interaction between the ZIKV and its vector. We have demonstrated that ZIKV infection results in changes to the cellular metabolic environment including a significant enrichment of inosine and pseudo-uridine (Ψ) levels which may be associated with RNA editing activity. In addition, infected mosquitoes demonstrate a hypoglycemic phenotype and show significant increases in the abundance of metabolites such as prostaglandin H2, leukotriene D4 and protoporphyrinogen IX which are associated with antiviral activity. These provide a basis for understanding the biochemical response to ZIKV infection and pathology in the vector. Future mechanistic studies targeting these ZIKV infection responsive metabolites and their associated biosynthetic pathways can provide inroads to identification of mosquito antiviral responses with infection blocking potential.

Keywords: Zika virus, *Aedes albopictus*, primary metabolites, lipids, biogenic amines, metabolomic phenotyping

INTRODUCTION

Zika virus (ZIKV) was first isolated in Zika forest, Uganda from a sentinel rhesus monkey in 1947 as well as from *Aedes* (*Stegomyia*) *africana* (Dick, 1952; Dick et al., 1952; Hayes, 2009; Petersen et al., 2016) and is a member of the family Flaviviridae, genus *Flavivirus* with a 10,794 base positive-sense single stranded RNA genome (Faye et al., 2014), classified as a member of the Spondweni group (Cook and Holmes, 2006). The virus is primarily transmitted by mosquitoes of the genus *Aedes* (Wikan and Smith, 2016). Recently, ZIKV has become a focus of intense research due to its rapid geographic spread in the Americas as well as its association with birth defects in offspring from infected mothers (e.g., microcephaly) and neurological syndromes (Ventura et al., 2016; de Oliveira et al., 2017; World Health Organization [WHO], 2019). This has resulted in a concerted effort to understand the biology of ZIKV and the interactions it has with its vector host.

The main route of ZIKV transmission is by the bite of infected *Aedes* mosquitoes, in addition, sexual, transplacental, and blood transfusion have been documented (Petersen et al., 2016). At the moment, no FDA-approved medication or vaccine to treat or prevent ZIKV infection exists (Gulland, 2016). Symptomatic relief by medications are the only source of relief for infected individuals (Kazmi et al., 2020). Zika disease control is achieved primarily by prevention of mosquito bites, vector abatement and prevention of sexual transmission (Petersen et al., 2016). Despite the progress made in understanding Zika etiology and its transmission, there exists a significant knowledge gap in the understanding of interactions between ZIKV and its vector.

The biochemical interactions and chemical processes within a living organism is collectively known as its metabolism. Metabolites are the basic units of cellular function, involved in enzyme-catalyzed chemical processes and are important for cellular function (Schrompe-Rutledge et al., 2016). The cascade of biological information in a living organism flows from the genes, to the transcripts through proteins and subsequently to the metabolites. Hence, any perturbations observed at the metabolome level is a direct consequence of alterations in the genome, transcriptome and the proteome of a biological system. In addition, the metabolome of an organism is considered a reflection of the chemical phenotype of that organism. In fact, metabolomics is considered a promising link between the genotype and phenotype gap (Bujak et al., 2015). Because of the sensitive nature of metabolomics, any alterations in the biological pathways can result in the understanding of the mechanisms underlying physiological conditions and processes resulting from disease progression in an organism (Johnson et al., 2016).

The study of metabolomics perturbations due to viral infection is very important due to the potential identification of metabolites that distinguish disease states and outcomes. The identification of these metabolites could lead to diagnostic and therapeutic targets. In addition, it facilitates a deeper understanding of the vector pathogen interactions (Byers et al., 2019).

Using high-resolution mass spectrometry of mosquito cells, Perera et al. (2012) demonstrated that DENV infection resulted

in enrichment of lipids that modulates the curvature of membranes as well as those which influence the permeability of membranes. Further, the same study showed an increase in sphingolipids and other bioactive signaling molecules involved in control of membrane fusion, fission and trafficking. In addition, molecules associated with cytoskeletal reorganization were enriched in DENV infected cells. Melo et al. (2016), utilized a lipidomics approach on ZIKV-infected C6/36 mosquito cell lines in a bid to characterize new biomarkers of ZIKV infection as well as establish new targets for viral controls in vertebrates and invertebrate vectors. They identified 13 lipids as specific marker for ZIKV infection. Those lipids were associated with intracellular mechanism of viral replication and or cell recognition. These findings are similar to that of Chotiwan et al. (2018). They analyzed the temporal metabolic perturbations associated with DENV infection of the midgut tissue in *Ae. aegypti* and observed an increase in glycerophospholipids, sphingolipids and fatty acyls that accompanied DENV replication. Levels of acyl-carnitines were also enriched while points in the sphingolipid pathway influencing dihydroceramide to ceramide ratios were noted as very important for the life of the virus in that study.

Furthermore, Bottino-Rojas et al. (2019) utilized a whole cell lipidomics approach to interrogate lipid perturbations in West Nile virus strain Kunjin (WNV_{KUN}) infected Vero cells. This reveal elevated levels of phospholipase A2 activity in the infected cells resulting in increased production of lyso-phosphatidylcholine moieties associated with subcellular sites of viral replication.

To understand the role of oxidative stress in combating arboviral infections in mosquitoes, Shrinet et al. (2018) carried out a metabolomics analysis of *Aedes* mosquitoes infected and co-infected with Chikungunya (CHIKV) and DENV. Further, they obtained different -omics data from the public database including proteomics and transcriptomics. The data analysis demonstrated that the different pathways were harmonized in their response to infection in regulating oxidative stress during arboviral infection of *Aedes* mosquitoes.

The goal of this study is to provide a comprehensive analysis of dynamic changes in the global metabolic profile (primary metabolites, lipids and biogenic amines) in whole *Ae. albopictus* mosquitoes infected with ZIKV. We identified an array of metabolic biomarkers that change in abundance in response to ZIKV infection. These biomarkers provide insights into how the virus interacts with its vector and help to elucidate potential novel barriers to transmission.

MATERIALS AND METHODS

Mosquitoes Species and Maintenance

All experimental protocols in this study were approved by Wadsworth Institutional Review Board. *Aedes albopictus* were collected from Suffolk County, NYS in 2015 (kindly provided by I. Rochlin) and colonized at the New York State Department of Health (NYSDOH) Arbovirus Laboratory. The mosquito strain was continually maintained at 28°C with relative humidity of

60% and 16:8 h light: dark (LD) photoperiod under standard rearing conditions. To confirm the temperature and humidity of the chambers, both internal chamber log and independent HOBO data loggers (Onset, Cape Cod, MA, United States) were used. F15 *Ae. albopictus* were vacuum-hatched and the immature stages reared under the standard rearing conditions in the lab. The larvae were fed on Tetra Pond Koi growth feed, the adults were transferred to 3.8 L cardboard cartons and allowed to mate for 5 days while being provided with sugar and water *ad libitum*. Water and sugar were withdrawn 24 h before being offered 8.3 log₁₀ PFU/ml ZIKV HND (Ciota et al., 2017) blood meal. The control group were provided a non-infectious blood meal.

Blood Meal Preparations and Mosquito Viral Infections

We used frozen ZIKV HND (2016-19563, GenBank accession no. KX906952) strain. The infectious blood meal consisted of 1:1 dilution of defibrinated sheep blood plus 2.5% sucrose; sodium bicarbonate (to adjust pH to 8.0) and virus. Infectious blood meals contained a viral titer of 8.3 log₁₀ PFU/ml ZIKV HND (Ciota et al., 2017; Onyango et al., 2020). The non-infectious blood meal (negative control) contained a final concentration of 2.5% sucrose solution. The female mosquitoes were exposed to blood meals in a 37°C pre-heated Hemotek membrane feeding system (Discovery Workshops, Acrinton, United Kingdom) with a porcine sausage casing membrane. All methods were carried out in accordance with relevant guidelines and regulations. Both blood meals were offered to the female mosquitoes after withdrawal of sugar 24 h before blood feeding. After an hour, the mosquitoes were anesthetized with CO₂ and immobilized on a pre-chilled tray connected to 100% CO₂. Engorged females were separated and placed in three separate 0.6 L cardboard cartons (30 individuals per carton). In addition, 1 ml of each blood meal was transferred to a 1.5 ml Safe Seal microtube (Eppendorf, Hamburg Germany) and stored at −80°C. Blood fed females were maintained on 10% sucrose solution provided *ad libitum*.

Sample Collection for Metabolomics Analysis Mosquito Dissection

To reduce the amount of concentrated carbohydrates in the mosquitoes in order to avoid disruption to the mass spectrometer, sucrose solution was withdrawn from the mosquito cages at 4 days post infection (dpi) and replaced with plain water. At 7 dpi, the female mosquitoes were immobilized using triethylamine (Sigma Aldrich, St. Louis, MO, United States). To examine for ZIKV dissemination in the mosquito, the legs were removed from the mosquitoes and placed in individual tubes containing 500 µl mosquito diluent (20% heat-inactivated fetal bovine serum in Dulbecco phosphate-buffered saline plus 50 µg/ml penicillin/streptomycin, 50 µg/ml gentamicin and 2 µg/ml Fungizone [Sigma Aldrich, St. Louis, MO, United States]) and a bead. All samples were held at −80°C until assayed. The corresponding whole bodies were snap frozen and stored at

−80°C. To ensure uniformity during metabolite extraction, the legs of the individual female mosquitoes exposed to non-infectious blood meal were also removed and the whole bodies snap frozen and stored at −80°C.

Quantitative RT-PCR Screening of ZIKV in *Ae. albopictus*

A ZIKV-specific quantitative PCR assay that targets the NS1 region was utilized to obtain viral titer (Lanciotti et al., 1992) from the legs as described by Ciota et al. (2017). Individual female mosquitoes showing a disseminated virus infection (ZIKV-positive legs) were identified as “ZIKV-infected mosquitoes.” A threshold C_t value of 35 was used to discriminate disseminated from non-disseminated individuals. In order to perform metabolite extraction, both the individuals that were exposed infectious (and were disseminating ZIKV) as well as those exposed to the non-infectious blood meal were pooled. The pools consisted of five individuals and ten biological replicates per sample types. The samples were frozen at −80°C before submission to the NIH West Coast Metabolomics Center at UC Davis. Microfuge tubes handled by the different individuals involved in the sample preparations were also sent to the core for normalization of the variation introduced during sample collection and preparation.

To characterize the metabolic changes incurred by *Ae. albopictus* during ZIKV infection, we performed metabolites analysis on whole bodies of *Ae. albopictus* 7 dpi with ZIKV and analyzed primary metabolites, lipids and biogenic amines.

Metabolite Extraction

A total of 20 pools (*N* = 10 replicates per infection status; *n* = 5 mosquitoes per pool) of whole body *Ae. albopictus* females exposed to infectious and non-infectious ZIKV blood meals were analyzed using three untargeted metabolomic panels to identify metabolic perturbations associated with ZIKV infection. The three analyses utilized were designed to screen for relative differences in the abundance of primary metabolites, lipids and biogenic amines. Each replicate was partitioned into three fractions using the protocol described in Matyash et al. (2008). Briefly, each of the experimental treatments were homogenized and vortexed with a bead mill homogenizer (FastPrep-24 MP Biomedicals) in 975 µl of ice-cold methanol: methyl-tert-butyl ether (3:10) extraction solution. Phase separation of the homogenate was induced by the ether (3:10) extraction solution. Phase separation of the homogenate was induced by the addition of 188 µl of LC-MS grade water followed by 6 min of shaking at 4°C. tubes were then vortexed for 20 s and then centrifuged for 2 min at 14,000 × *g*. The organic phase was transferred to a separate tube for lipidomics analysis. The remaining aqueous fraction was split into two aliquots for the primary metabolite and biogenic amine analysis panels. Quality control samples were generated for each of the three metabolite panels by pooling 100 µl of each replicate from both experimental treatments. Blank controls consisting of empty tubes were also included in the analysis to account for contaminants associated with handling. The

aqueous phase aliquots were dried down by centrifugation under vacuum in a speedvac.

Primary Metabolite Analysis by GC TOF

Primary metabolites (including sugars, amino acids, hydroxyl acids and related biochemical pathways) were profiled using the Gas Chromatography time of flight (GC TOF) untargeted protocol as described in Fiehn (2017). Dried sample aliquots were resuspended in 450 μ l of nitrogen-degassed 50:50 (v/v) acetonitrile/water at room temperature. Resuspended samples were centrifuged for 2 min at $14,000 \times g$ at room temperature. Supernatant was transferred to a new tube and dried down in a speedvac evaporator.

Samples and controls were derivatized (chemical conversion of non-volatile compounds to volatile compounds) by treatment of dried extracts with 10 μ l of 20 mg/ml methoxamine hydrochloride solution in pyridine. Samples were then shaken at maximum speed on an orbital heating plate for 1.5 h at 30°C. Then 91 μ l of *N*-methyl-*N*-(trimethylsilyl)-trifluoroacetamide (MSTFA) was added to each sample and controls which were shaken at maximum speed for 30 min at 37°C. Derivatized samples and controls were then added to glass vials for GC-MS analysis and data acquisition.

Samples were injected via an Agilent 7693 autosampler (Santa Clara, CA, United States) into a 7890 Agilent GC fitted with a RtX-5Sil MS (30 m length \times 0.25 mm internal diameter with 0.25 μ m film made of 95% dimethyl/5% diphenylpolysiloxane) Restek corporation (Middelburg, ZEELAND, Netherlands) chromatographic column using helium as a mobile phase at 1 ml/min. Sample volumes of 0.5 μ l were injected at 50°C ramped to 250°C by 12°C/s. the oven temperature program was 50°C for 1 min then ramped at 20°C/min to 330°C, held constant for 5 min.

Data acquisition was performed with a Leco Pegasus IV time-of-flight MS instrument (Leco, St. Joseph/MI, United States). Mass spectrometry parameters were used as follows: the mass spectrometer is run with unit mass resolution at 17 spectra/s from 80 to 500 Da at 70 eV ionization energy and 1800 V detector voltage with a 230°C transfer line and a 250°C ion source. The multi baffled glass injection liners were changed every ten injections followed by reinjection of the pooled quality control sample throughout the course of the analytical batch.

Lipidomic Profiling by LC/Q-TOF CSH

Lipidomic analysis was performed by liquid chromatography (LC) coupled to a quadrupole time-of flight (QTOF) charged surface hybrid column (CSH) mass spectrometer (Waters MS Technologies, Manchester, United Kingdom). The lipid containing organic phase from the metabolite extractions (350 μ l) were dried in a Speed Vac (Thermo Fisher, Pittsburgh, United States). The remaining residue was redissolved in 110 μ l MeOH:Tol (9:1) including an internal standard mix (50 ng/ml) (see **Supplementary Table 2** for lipidomic internal standard composition). The resuspended samples were then vortexed for 10 s, sonicated for 5 min and centrifuged for 2 min at $16,100 \times g$. The supernatant was aliquoted into two tubes for analysis in positive and negative electrospray ionization modes.

The data was analyzed on an Agilent 6530 Q-TOF LC/MS UPLC mass spectrometer. Samples were separated on a C₁₈ column equipped with charged surface hybrid column.

Data acquisition was attained as described in Dong et al. (2017). Briefly, each sample was injected at 4°C into a Waters Acquity UPLC CSH C₁₈ column (100 mm length \times 2.1 mm internal diameter; 1.7 μ m particles) at 65°C with the flow rate set at 6 ml/min.

Each sample was subjected to electrospray ionization (ESI) in positive and negative modes with a capillary voltage of 3.2 kV, cone voltage of 30 kV and desolvation gas temperature of 400°C. positive and negative specific ionization settings are as follows:

Positive mode settings:

Spectrophotometer: Agilent 6530 Q-TOF LC/MS UPLC

Column: Waters Acquity UPLC CSH C₁₈ (100 mm length \times 2.1 mm internal diameter; 1.7 μ m particles)

Mobile phase (A): 60:40 v/v acetonitrile:water + 10 mM ammonium formate + 0.1% formic acid

Mobile phase (B): 90:10 v/v isopropanol:acetonitrile + 10 mM ammonium formate + 0.1% formic acid

Scan range positive mode: m/z 120–1200 Da

Injection volume: 1.7 μ l

ESI capillary voltage: ESI (+): + 3.5 kV

Negative mode settings:

Spectrophotometer: Agilent 6530 Q-TOF LC/MS UPLC

Column: Waters Acquity UPLC CSH C₁₈ (100 mm length \times 2.1 mm internal diameter; 1.7 μ m particles)

Mobile phase (A): 60:40 v/v acetonitrile: water + 10 mM ammonium acetate

Mobile phase (B): 90:10 v/v isopropanol: acetonitrile + 10 mM ammonium acetate

Scan range negative mode: m/z 60–1200 Da

Injection volume: 5 μ l

ESI capillary voltage: ESI (+): –3.5 kV

The elution gradient for both positive and negative modes was 0 min 15% (B), 0–2 min 30% (B), 2–2.5 min 48% (B), 2.5–11 min 82% (B), 11–11.5 min (B), 11.5–12 min 99% (B), 12–12.1 min 15% (B), 12.1–15 min 15% (B).

Biogenic Amine Analysis Using HILIC LC/Q-TOF

Biogenic amines are low molecular weight organic bases resulting from decarboxylation of free amino acids by animals, plants and microorganisms (Ito and Nakata, 2016). A rapid untargeted analysis of polar metabolites of Zika infected *Ae. albopictus* tissues was carried out by Hydrophilic Interaction Chromatography (HILIC) coupled to Quantum Time of Flight (Q-TOF) mass spectrometry. The analysis was undertaken as described in Chintapalli et al. (2013) with modifications. Dried aliquots for biogenic amine analysis were resuspended in 100 μ l acetonitrile (ACN): water (80:20) including internal standards (see **Supplementary Table 2** for biogenic amine internal standards) followed by 10 s of vortexing, 15 min of sonication

and centrifugation for 2 min at $16,000 \times g$. All processes were performed at 4°C . A pooled quality control sample was run after every 10 samples throughout the course of the analytical batch.

HILIC LC/Q-TOF settings:

Spectrophotometer: Agilent 6530 Q-TOF LC/MS UPLC

Pre-column: Waters Acquity UPLC BEH Amide VanGuard pre-column ($1.7\ \mu\text{m}$, $5\ \text{mm} \times 2.1\ \text{mm}$)

Column: Waters Acquity UPLC BEH Amide column, $1.7\ \mu\text{m}$, $2.1\ \text{mm} \times 150\ \text{mm}$)

Mobile phase (A): Solvent A was Ultrapure water with 10 mM ammonium formate + 0.125% formic acid, pH 3

Mobile phase (B): Solvent B was 95.5 v/v acetonitrile: ultrapure water with 10 mM ammonium formate + 0.125% formic acid, pH 3

Injection volume: $3\ \mu\text{l}$

Flow rate: $400\ \mu\text{l}/\text{min}$

ESI (+) capillary voltage: + 4.5 kV

ESI (+) collision energy: + 45 eV

Scan Range: m/z 60–1200 Da

Spectral acquisition speed: 2 spectra/s

Mass resolution: 10,000

Elution gradient 0 min 100% (B), 0–2 min 100% (B), 2–7 min 70% (B), 7.7–9 min 40% (B), 9.5–10.25 min 30% (B), 10.25–12.75 min 100% (B), 16.75 min 100% (B). The vial tray was kept at a constant temperature of 34°C while the column was set at ambient temperature.

Data Processing

Data for detected metabolites in all three panels were provided as peak heights for the quantification ion (m/z value) at the associated retention time (rt value). These values were obtained via the following processes for the respective panels.

Primary Metabolite Panel Data Processing

The raw GC-MS data from the primary metabolite panel was preprocessed in the Leco ChromaTOF version 2.32 software without smoothing using a 3 s peak width baseline subtraction just above the noise level and automatic mass spectral deconvolution/peak detection at signal/noise levels of 5:1. Apex masses were output for comparison against the BinBase metabolomics database (Skogerson et al., 2011). The BinBase algorithm (rtx5) used the following settings: validity of chromatogram (<10 peaks with intensity $> 10^7$ counts s^{-1}), unbiased retention index marker detection (MS similarity > 800 , validity of intensity range for high m/z marker ions), retention index calculation by 5th order polynomial regression. Spectra are cut to 5% base peak abundance and matched to database entries from most to least abundant spectra using the following matching filters: retention index window ± 2000 units (equivalent to about ± 2 s retention time), validation of unique ions and apex masses (unique ion must be included in apexing masses and present at $>3\%$ of base peak abundance), mass spectrum similarity must fit criteria dependent on peak purity and

signal/noise ratios and a final isomer filter. Failed spectra are automatically entered as new database entries if $s/n > 25$, purity < 1.0 and presence in the biological study design class was 80%. All thresholds reflect settings for ChromaTOF v 2.32. Quantification is reported as peak height using the unique ion as default, unless a different quantification ion is manually set in the BinBase administration software BinView. A quantification report table is produced for all database entries that are positively detected in more than 10% of the samples of a study design class (as defined in the miniX database) for unidentified metabolites.

Lipidomics Panel Data Processing

Raw LC/MS spectra were processed with MS-DIAL using the default parameters (Tsugawa et al., 2015). Data from blank samples were used to subtract background and contaminants from sample data based on the max peak height relative to blank average height, the average of all non-zero peak heights for samples, and if the feature is found in at least one sample. This is followed by sample cleanup using MS-FLO to eliminate potential duplicates and isotopes (Defelice et al., 2017). Internal standards served as retention time alignment markers and for quality control purposes. Peaks are annotated in manual comparison of MS/MS spectra and accurate masses of the precursor ion to spectra given in the Fiehn laboratory's LipidBlast spectral library (Kind et al., 2013). The confidence of annotation was guided by recommendations from the Metabolites Standard Initiative (Sumner et al., 2007). Unique chromatographic features that could not be identified are reported as "unknown."

Biogenic Amine Data Processing

The raw data from the biogenic amines panel was processed using mzMine 2.0 (Pluskal et al., 2010) for peak information. Internal standards served as retention time alignment markers and for quality control purposes. Alternatively, selected peaks were collated and constrained into Agilent's MassHunter quantification method on the accurate mass precursor ion level, using the MS/MS information and the NIST14/Metlin/MassBank libraries to identify metabolites with manual confirmation of adduct ions and spectral scoring accuracy.

Data Normalization and Analysis

Comparative analysis of the processed data from the ZIKV infected versus uninfected samples was performed using the MetaboAnalyst (V4.0) package (Chong et al., 2018). Raw data were submitted as peak intensity tables with samples in columns in unpaired format. Data were filtered to remove variables with low repeatability using Relative standard deviation (RSD). Samples were normalized by dividing the spectral profile of the sample by the median of all feature intensities of the sample. The data was log transformed and auto scaled (mean-centered and divided by the standard deviation of each variable).

Datasets were analyzed within Metaboanalyst using the following integrated tools and parameters. Significant differences in metabolite abundances between treatment groups were evaluated by Wilcoxon Rank Test assuming unequal variance with a threshold of an adjusted P -value (FDR- Benjamini and Hochberg method) of 0.05. Multivariate analysis of the datasets

was performed using Partial Least Squares Discriminant Analysis (PLS-DA). Top features from the PLS-DA analyses were signified by associated Variable Importance In Projection (VIP) scores. Heatmaps visualizing relative compound abundances in infected and uninfected samples were generated from top VIP scoring compounds. Distance measures between samples were calculated using Euclidean distance measure algorithm and clustered by the Ward algorithm. Features were standardized by autoscaling.

Pathway and Network Analysis of Metabolites

Further pathway-based Analysis were performed on the LC-MS data derived from the lipidomic and biogenic amines panel datasets using the Peaks to Pathways function in Metaboanalyst which is a web based iteration of the Mummichog analytical package (Li et al., 2013). Analysis of the primary metabolite dataset via Mummichog was not possible as it was derived from gas chromatography and lacks the required accuracy in mass/charge ratio and retention times required to make accurate metabolite predictions. The pathway analysis placing the experimental observations into relevant biological and disease context. Mummichog utilizes dta from defined metabolic network to predict functional activity directly from feature tables based on mass/charge ratio (mz) and retention time (rt). Data for this analysis were filtered, normalized and scaled using equivalent parameters to those in the section on dataset normalization and analysis. Following filtration and normalization, data were reformatted for submission to the peaks to pathways algorithm. All compounds (including unknowns) were represented by mz, rt, *p*-value, *T*-score and electro spray mode (positive or negative). Reformatted data files representing the lipidomic and biogenic amine panels were combined into a single dataset and submitted for analysis via version 2.0 of the peaks to pathways algorithm using the following parameters: molecular weight tolerance (ppm)- 10, analytical mode- mixed, compounds ranked by *P*-value and primary ions not enforced. Analysis algorithms utilized were Mummichog with a *P*-value cutoff of 0.05 and GSEA using the *Drosophila melanogaster* KEGG library.

Compound identifications resulting from the Peaks to Pathways analysis in the form of KEGG ids were mapped back to annotate unidentified compounds in the original datasets. Compounds with multiple potential annotations were matched with the annotation with the lowest mass difference relative to the compound. Network analysis of KEGG annotated compounds was performed in Cytoscape 3.8.0 (Shannon et al., 2003) using the Metscape (Karnovsky et al., 2012) plugin. Compounds displayed represent members of metabolic pathways identified as significantly enriched by Mummichog analysis.

Correlation Analysis

The correlation analysis of metabolite abundance and viral titer was performed in R version 4.0.2 (R Core Team, 2003) using the following packages: corrr correlation analysis (Kuhn et al., 2020), dplyr (Wickham et al., 2020) tidyr (Wickham and Henry, 2020), ggplot2 (Wickham, 2016), and viridis (Garnier, 2018).

Compounds included in the correlation analysis were those with significant shifts in abundance relative to ZIKV infection status (FDR < 0.05).

Inosine Assay

To confirm changes in inosine concentration upon viral infection a secondary biochemical assay was performed to verify the metabolomic predictions. The analysis was performed using a commercial inosine assay kit (Sigma-Aldrich, MO, United States). The assay utilizes a coupled enzyme reaction in which inosine is converted to hypoxanthine which reacts with the substrate mix and probe resulting in a fluorometric product ($\lambda_{\text{ex}} = 535 \text{ nm}/\lambda_{\text{em}} = 587 \text{ nm}$). The resulting absorbance is proportional to the inosine abundance. Mosquitoes were reared and offered a blood meal as described above and 7 days post infectious blood meal, the mosquito's legs were dissected and placed in a labeled vial and the whole body transferred to a separate vial. The legs of the mosquitoes were screened for ZIKV infection as described above to ascertain infection.

Six singleton female mosquitoes exposed to ZIKV infectious and non-infectious blood meals were homogenized in 0.7 ml of ice-cold 0.4 M perchloric acid and centrifuged at $13,000 \times g$ for 10 min in order to remove the insoluble material. The supernatant was transferred to a new tube and then neutralized by adding ice cold 70 μl of 4M potassium carbonate solution. The solution was incubated on ice for 7–10 min and then centrifuged at $13,000 \times g$ for 10 min.

Each sample was then diluted 1:10 and 1 μl of both diluted and undiluted sample were added to the wells of black plates with clear bottom. The samples were brought to a final volume of 50 μl with inosine assay buffer and 50 μl of the reaction mix was added to each well including the inosine standards and the sample blank. The fluorescence intensity was measured at $\lambda_{\text{ex}} = 535 \text{ nm}/\lambda_{\text{em}} = 587 \text{ nm}$. The amount of inosine present in the samples was determined from the standard curve. Differences in the concentration of inosine across the infection status was compared by *T*-test.

RESULTS

Global Metabolic Profile Analysis

A total of 209 *Ae. albopictus* individuals were exposed to $8.3 \log_{10}$ PFU/ml ZIKV HND strain infectious blood meal, the overall infection rate of exposed individuals was 26%, with an average of $\log_{10} 7.64$ copy number standard ZIKV and a standard deviation of ± 0.89 (Supplementary File S1).

Samples were subjected to untargeted metabolic analyses screening for primary metabolites (carbohydrates, proteins, amino acids, vitamins, acetone, and ethanol), lipids (fats, phospholipids, vitamins, and steroids) and biogenic amines (monoamine neurotransmitters and polyamines). Only features (molecular identities defined by a unique mass to charge ratio, m/z and retention time) detected in all the 20 samples (representing 10 infected and 10 uninfected) were considered 'present' in the samples. Comparison of ZIKV infected versus uninfected mosquitoes revealed clusters of metabolites showing

significant differences in abundance between infected and uninfected mosquitoes.

A total of 473 primary metabolites were detected, however, after filtering and normalization, 425 were analyzed. Of these, 29% were identified at the structural level, while the rest remain unidentified. Of the identified primary metabolites, 41 show differential abundances between treatment groups using a log fold change ≥ 2 and *T*-test/FDR *P*-values ≤ 0.05 being considered differential.

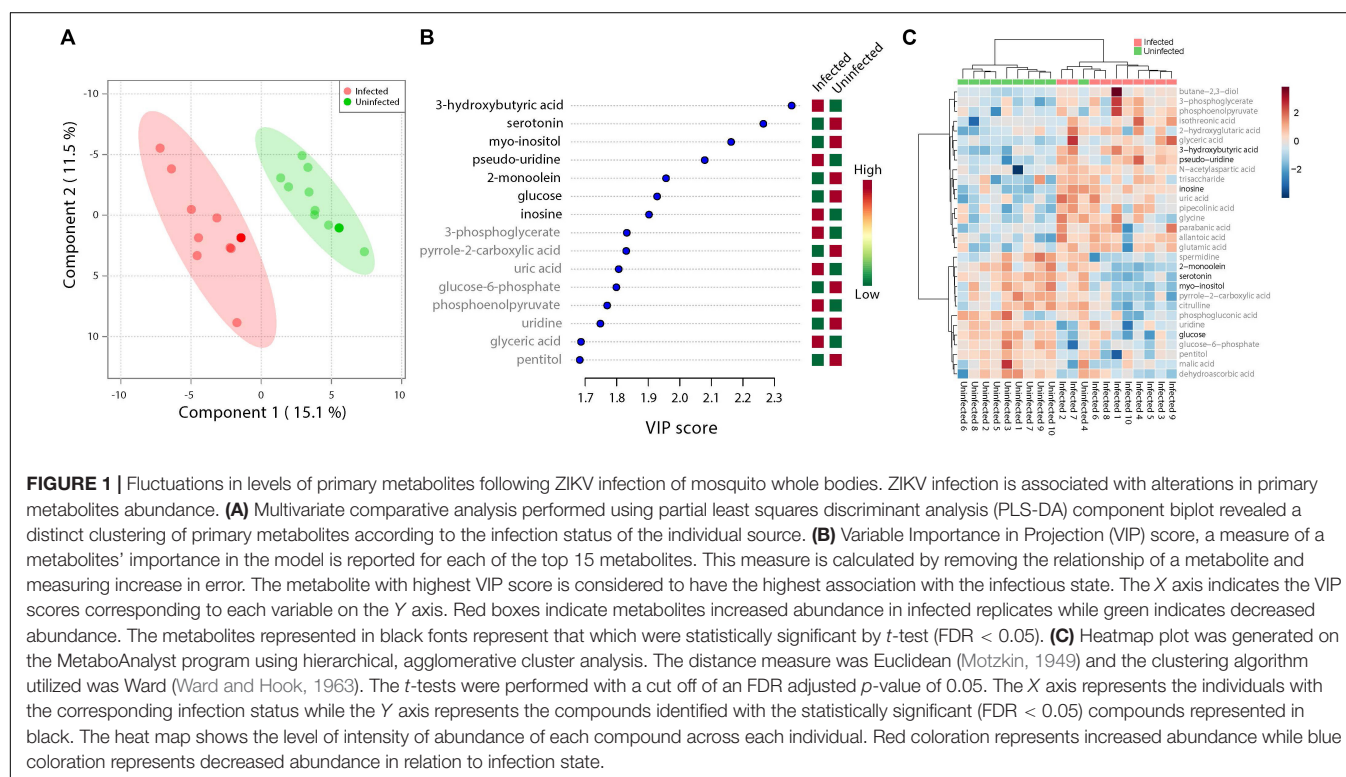
After normalization, 1809 lipids and lipid related compounds were detected and 409 were identified to the structural level. Of these, 167 showed differential abundances based on the same statistical parameters as used for the primary metabolites. Of the compounds detected in the biogenic amines panel, 896 were detected with 28% of those identified to structure. Of those, 26 met the threshold of differential abundance. Analysis of the detected compounds from the three panels by partial least squares – discriminant analysis (PLS-DA) revealed a clear separation between uninfected and infected samples and highlighted an array of compounds correlating with infection state. ZIKV-infection distinguished the primary metabolite, identified and unidentified lipids and biogenic amines when plotted together.

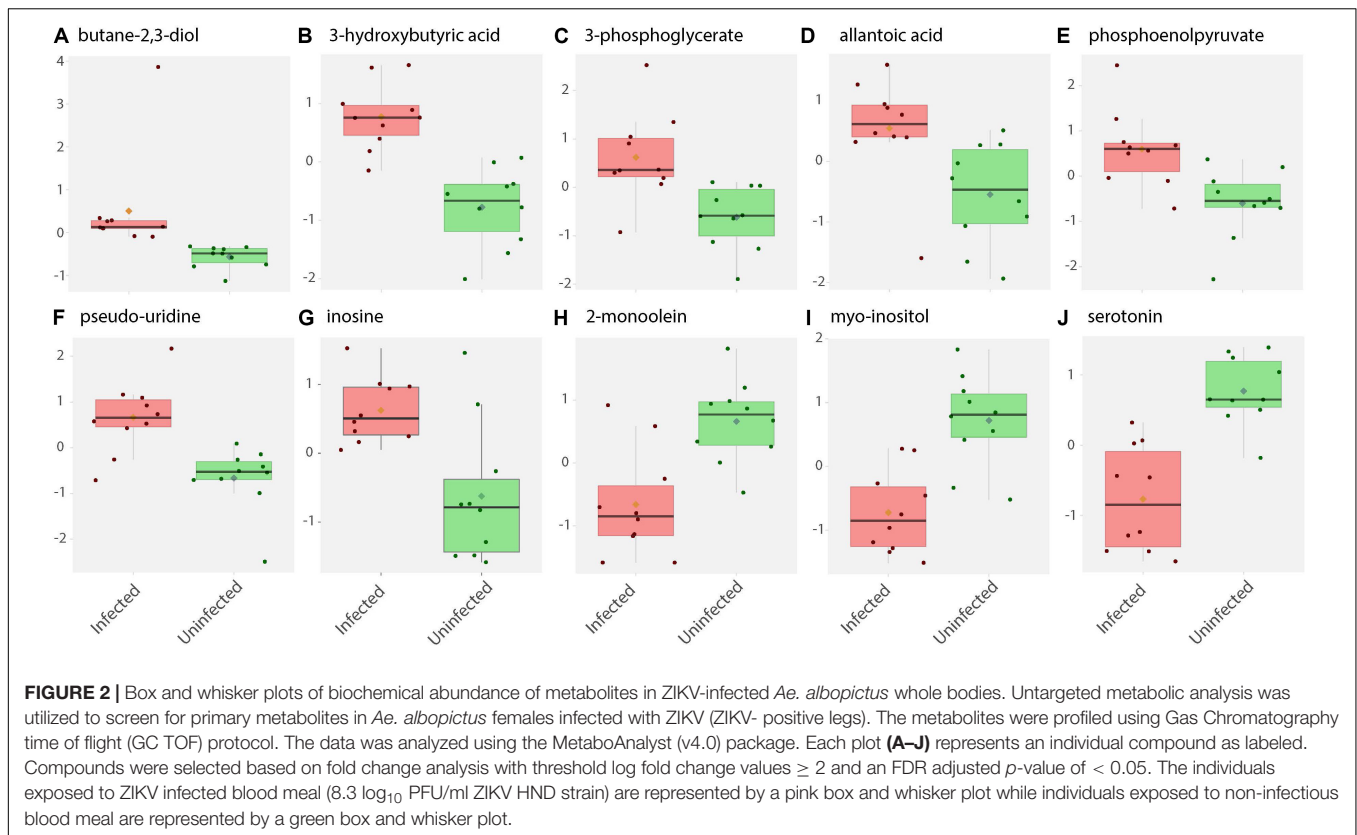
Metabolites Associated With RNA Editing and Glucose Levels Reduction Is Observed in ZIKV-Infected *Ae. albopictus*

ZIKV infection resulted in an enrichment of levels of metabolites associated with RNA editing as well as a perturbation of glucose

metabolism levels. RNA editing catalyzed by enzymes of the adenosine deaminase acting on RNA (ADAR) family converts adenosine to inosine in a double-stranded RNA (Zinshteyn and Nishikura, 2009). Additionally, pseudo-uridine, an isomer of the nucleoside uridine, occurs in rRNA, tRNA and small nuclear and nucleolar RNA but not in mRNA or viral genomic RNAs (Carlile et al., 2019). We measured a significant increase in pseudo-uridine abundance among the pools of *Ae. albopictus* exposed to infectious blood meals (*T* – stat 4.165; *P* = 0.0005; FDR = 0.019781) (Figures 1B,C, 2F). Inosine, a purine nucleotide is found at the wobble position of tRNA, playing a role in the proper translation of mRNA at the ribosome. Apart from its role in translation, it plays a role in the immune system exhibiting both inflammatory and anti-inflammatory effects. We measured the impact of ZIKV infection of *Ae. albopictus* on the levels of inosine. Further, increased abundance of inosine was also detected in the infected groups (*T* – stat 3.5434; *P* = 0.002; FDR = 0.005) (Figures 1B,C, 2G, 3). We carried out a separate inosine assay experiment to determine the impact of ZIKV infection on inosine levels of *Ae. albopictus*. We observed an enrichment of inosine among ZIKV in *Ae. albopictus* (*t*-test *P* = 0.0178; *t* = 2.832 df = 10) (Figure 3).

Specifically, levels of phosphoenolpyruvate, [responsible for generation of adenosine triphosphate (ATP), a major source of chemical energy within cells], were increased among infected samples relative to the uninfected (*T* – stat 3.5845; *P* = 0.002; FDR = 0.04) (Figure 2E). In addition, a significant increase of 3- phosphoglycerate metabolites among the infected individuals was noted (*T* – stat 3.9136; *P* = 0.001; FDR = 0.02) (Figures 1B,C, 2C), allantoinic acid, involved in





uric acid degradation was upregulated among ZIKV-infected individuals (Figure 2D), 3- phosphoglycerate is an important component of the glycolysis pathway. Levels of the ketone body 3- hydroxybutyrate, were also elevated (T – stat 5.5305; $P = 2.99E-05$; FDR = 0.0040654) (Figures 1B,C, 2B). This metabolite is generated during the metabolism of fatty acids (e.g., butyrate) and is usually synthesized to provide backup energy in the case of reduced glucose levels (Newman and Verdin, 2017). The infected groups appear to demonstrate increased energy expenditure as evidenced by reduced levels of glucose (T – stat -3.6259; $P = 0.0019$; FDR = 0.04) (Figures 1B,C) and glucose -6- phosphate (Figures 1B,C), a major intermediate in the mobilization of glucose as an energy source. This is a potential indicator that infected mosquitoes are undergoing increased rates of glycolysis in association with viral replication. Both myo-inositol and butane- 2,3- diol, products of glucose metabolism demonstrated significant change in abundance due to ZIKV infection (Figure 2I). Butane- 2,3 – diol, a volatile compound associated with certain strains of root-associated bacteria (Kong et al., 2018) and capable of being produced from glucose (Chu et al., 2015) was highly abundant ($P = 0.013$; Log₂⁵ fold change) in the infected samples (Figure 2A).

Serotonin as a neurotransmitter controls multiple processes related to feeding and nutrition in mosquitoes (Moffett and Moffett, 2005). Results of this study analysis revealed that infected mosquitoes show a significant decrease in serotonin levels (T – stat -5.0104; $P = 9.08E-05$; FDR = 0.0062) indicating that the infection may be having an effect on the serotonergic system

(Figures 1B,C, 2J). In addition, 2-monoolein, a monoglyceride linked to cell membrane fluidity, was downregulated in the infected tissues (T – stat -3.7195; $P = 0.002$; FDR = 0.04) (Figures 1B,C, 2H) (Singh et al., 2019). Apart from the above-mentioned samples, there were an array of metabolites whose abundance were differentiated but at an insignificant level. These includes metabolites such as uric acid, glyceric acid, pentitol, malic acid, and many others.

Enrichment of Arachidonic Acid Observed in ZIKV-Infected *Ae. albopictus*

Infection with ZIKV differentiated the lipids present in samples of *Ae. albopictus*. This was observed both when Multivariate comparative analysis was done using identified compounds as well as identified and non-identified compounds (Figures 4A,B). A large number of the lipids whose levels were altered due to ZIKV infection have not been previously identified, however, they demonstrated a clear profile of abundance in association with the infection status of the individuals (Figures 4B,D).

Lipids play critical roles in multiple stages in the virus replication cycle and the unique induction of a lipid profile upon viral infection is required for optimal virus replication (Diamond et al., 2010; Heaton and Randall, 2010; Peña and Harris, 2012; Perera et al., 2012; Martin-Acebes et al., 2014; Xu and Nagy, 2015; Melo et al., 2016; Chotiwan et al., 2018). Infected mosquitoes showed multiple perturbations in their lipid constitution. In particular, arachidonic acid, a precursor of

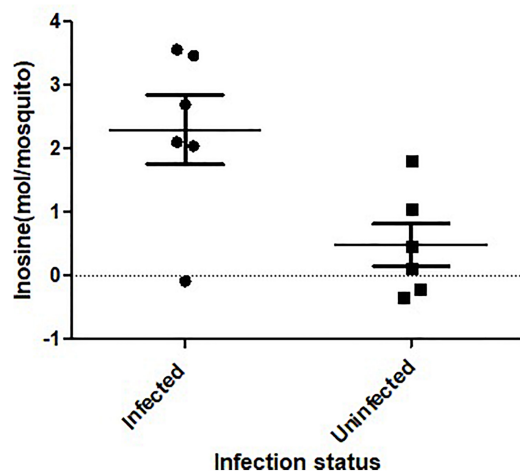


FIGURE 3 | Inosine quantification in ZIKV-infected *Ae. albopictus*.

A fluorometric assay was utilized to compare inosine levels in ZIKV infected (ZIKV- positive legs) and non-infected *Ae. albopictus* individuals. The individuals were offered 8.3 log₁₀ PFU/ml ZIKV HND strain blood meal or uninfected blood meal. At 7 dpi, the legs were dissected and screened for infection and the whole body of individual mosquito was used to determine the inosine levels. The background was corrected by subtracting the blank fluorescent reading value from all readings and the standard curve generated from the corrected values of the inosine standards. The sample blank value was subtracted from the sample readings and the amount of inosine present was determined from the standard curve using the corrected measurements. The X axis represents the infection status of the mosquitoes while the Y axis represents the concentration (moles) per mosquito.

eicosapentaenoic acid (Eijkelkamp et al., 2018) (Figures 4, 5) was differentially expressed due to ZIKV infection. When the unidentified compounds were included in the analysis, some show a highly significant correlation with ZIKV infection status. Both the identified (arachidonic acid and eicosapentaenoic acid) were plotted based on their mass/charge ratio against the Gas Chromatography Column Retention Time. The lipids within the top 200 VIP scores were annotated and they appeared to cluster in a similar space as the short chain lysophospholipids demonstrating a potential for shared properties in terms of M/Z and retention time (Figure 5).

ZIKV Infection Results in the Differential Abundance of Metabolites Associated With Immunomodulatory Roles

Analysis of metabolites identified among ZIKV-infected and uninfected *Ae. albopictus* individuals resulted in defined clusters of biogenic amines associated with either infected or uninfected *Ae. albopictus*. Biogenic amines are low molecular weight organic compounds formed by decarboxylation of amino acids. They are known to modulate energy metabolism in insects and regulate the secretion of other hormones (Hirashima et al., 2000). They also function as neurotransmitters and mediate cross talk between the nervous and immune systems (Sternberg, 1997). Acetylcholine, known for its role in neurotransmission (Picciotto et al., 2012) and immunity (Rajendran et al., 2015) is increased in abundance

among the infected samples (T – stat 7.9272; $P = 2.79E-07$; FDR = 2.44E-05) (Figure 6B). In addition, both 6-azuridine (T – stat 3.7079; $P = 0.002$; FDR = 0.03) and biliverdin (T – stat 3.8715; $P = 0.001$; FDR = 0.02) were upregulated among the infected individuals (Figures 6B,C). Stachydrine (T – stat -4.3757; $P = 0.0004$; FDR = 0.01) as well as 3-dehydrocarnitine (T – stat -4.5924; $P = 0.0002$; FDR = 0.008) (an intermediate in the production of carnitine) were significantly depleted in infected individuals (Figures 6B,C). Increased levels of *N*-methyl-D-aspartic acid were observed among the infected individuals (T – stat 4.6483; $P = 0.0002$; FDR = 0.008) (Figures 6B,C). *N*-alpha-acetyl-L-arginine levels were enriched among infected individuals (T – stat 3.8675; $P = 0.001$; FDR = 0.02). L-arginine may inhibit glycation and advanced glycosylated end product formation (Servetnick et al., 1996). Increased 5-aminolevulinic acid levels were measured among ZIKV-infected individuals (T – stat 4.6483; $P = 0.0002$; FDR = 0.008). It is a product of glycine and succinyl CoA and its synthesis is regulated by the amount of heme in the cell (Peng et al., 1997). ZIKV infection was associated with enrichment of Citrulline (T – stat 3.828; $P = 0.001$; FDR = 0.02). Citrulline is important for protein biosynthesis and normally, ornithine is converted to citrulline before conversion to arginine (Windmueller and Spaeth, 1981). In addition, propionyl-carnitine, a biogenic amine known to possess superoxide scavenging and antioxidant properties (Vanella et al., 2000) was enriched among the infected individuals (T – stat 3.8197; $P = 0.001$; FDR = 0.02) (Figures 6B,C).

Pathway and Network Analysis

Analysis of compounds present in the lipidomic and biogenic amine datasets for enrichment of specific biochemical pathways revealed statistically significant abundance shifts across multiple pathways. Identified pathways include: glycine, serine and threonine metabolism; valine, leucine and isoleucine biosynthesis; glycerophospholipid metabolism; arachidonic acid metabolism; retinol metabolism; pentose and glucuronate interconversion; sphingolipid metabolism; arginine and proline metabolism as well as porphyrin and chlorophyll metabolism (Figure 7). Metabolites from these pathways along with their relative abundances and statistical values were visualized to identify relationships between these pathways and to highlight large changes in metabolite abundance.

This analysis highlights multiple features of note in infected mosquitoes. The most prominent feature is a >300-fold increase in the compound protoporphyrinogen IX (a known antiviral that blocks viral cell invasion) (Paiva-silva et al., 2006) (Figure 8). This compound is synthesized from biliverdin which is a byproduct of heme metabolism in blood feeding insects and also shows a significant increase in abundance in infected mosquitoes.

Another observation is a 100-fold increase in abundance of prostaglandin H2, which is the precursor to all prostaglandin biosynthesis. In connection with this, compounds required for prostaglandin synthesis (including arachidonate, ethanolamine phosphate and sphinganine) were also significantly enriched among infected individuals (Figure 8). These findings suggest the arachidonic acid pathway is playing an important

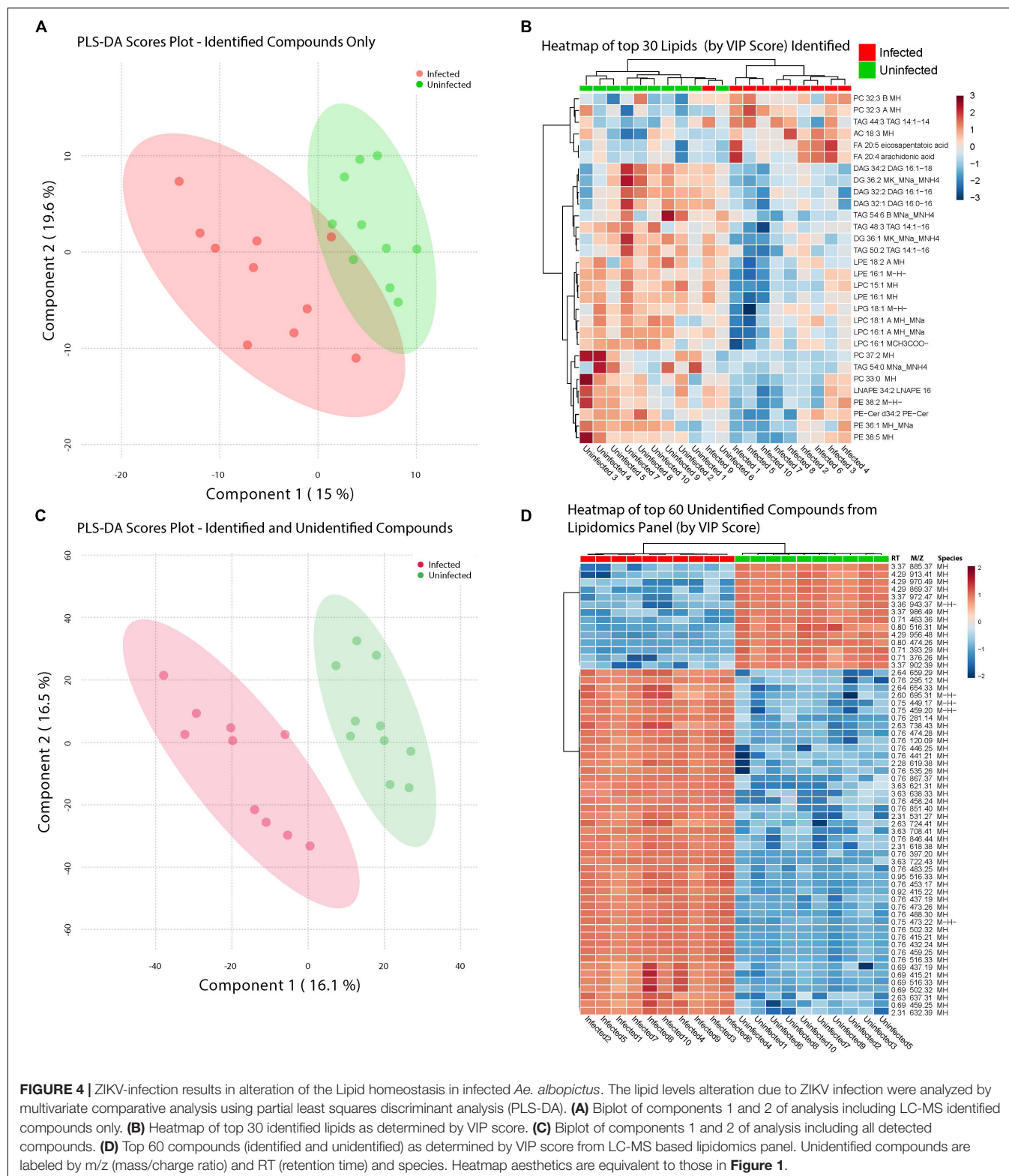
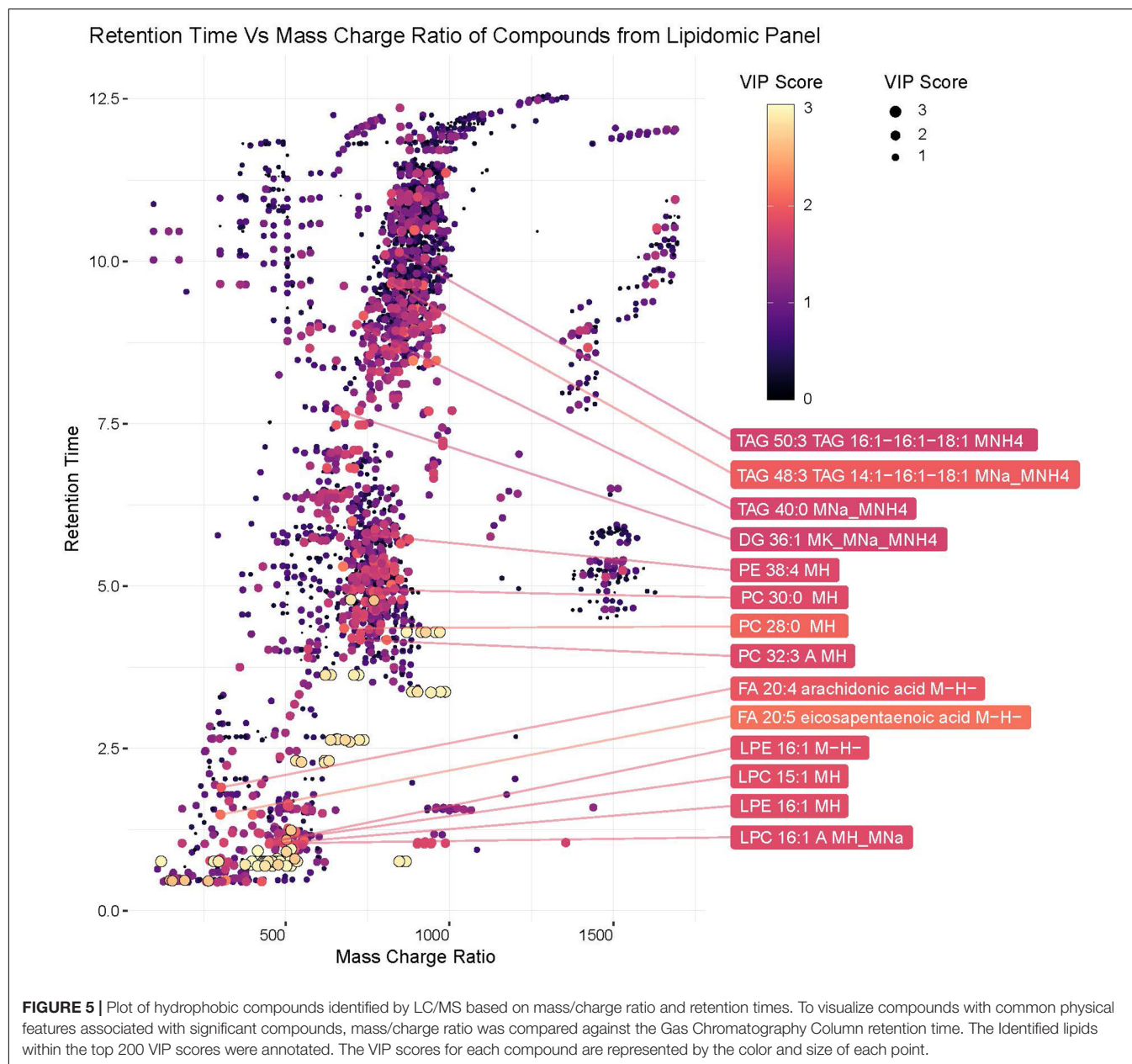


FIGURE 4 | ZIKV-infection results in alteration of the Lipid homeostasis in infected *Ae. albopictus*. The lipid levels alteration due to ZIKV infection were analyzed by multivariate comparative analysis using partial least squares discriminant analysis (PLS-DA). **(A)** Biplot of components 1 and 2 of analysis including LC-MS identified compounds only. **(B)** Heatmap of top 30 identified lipids as determined by VIP score. **(C)** Biplot of components 1 and 2 of analysis including all detected compounds. **(D)** Top 60 compounds (identified and unidentified) as determined by VIP score from LC-MS based lipidomics panel. Unidentified compounds are labeled by m/z (mass/charge ratio) and RT (retention time) and species. Heatmap aesthetics are equivalent to those in **Figure 1**.

role in the infection response and facilitating increased prostaglandin synthesis.

In addition, a large increase in the abundance of leukotriene D4 was observed in infected mosquitoes along with perturbations

in other compounds associated with its biosynthesis. As with prostaglandins, leukotrienes are also associated with inflammatory responses to infection and allergens in vertebrates (**Figures 7, 8**).

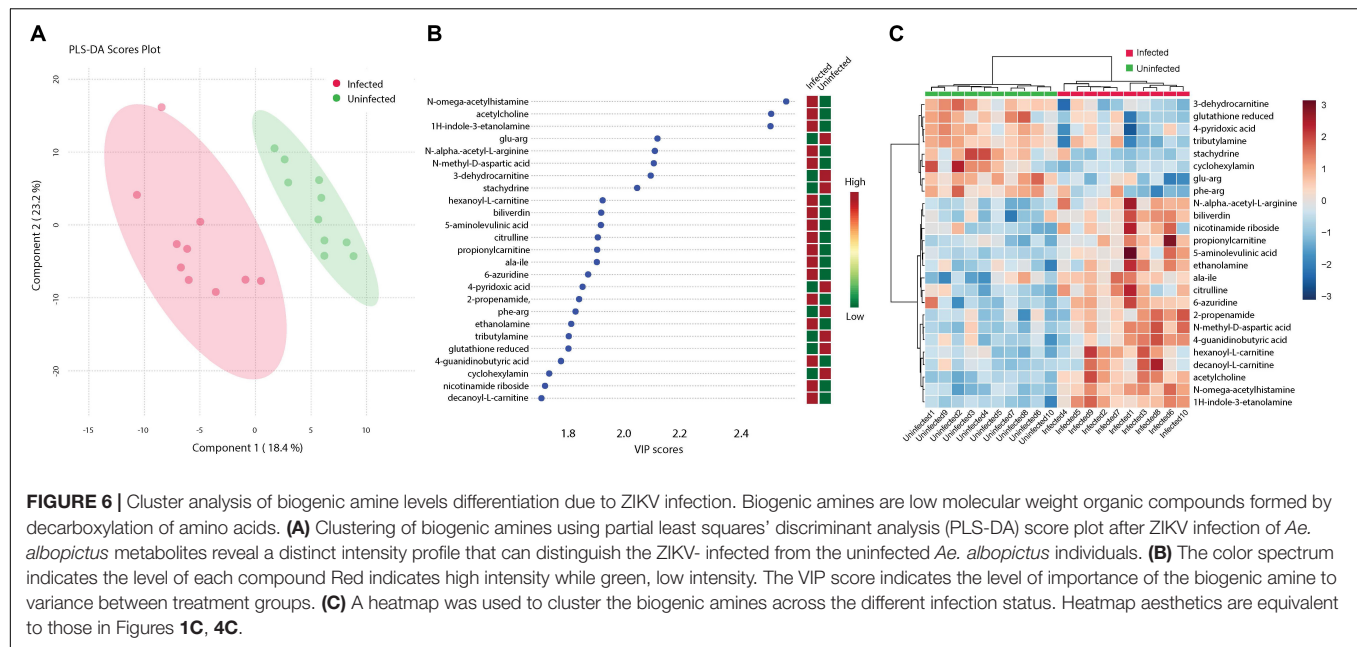


Arachidonic acid metabolism pathway was found to be significantly enriched among infected individuals as well as three prostaglandins (prostaglandin E2, prostaglandin H2 and (5Z,13E)- (15S)-9alpha,15-dihydroxy-11-oxoprost-5,13-dienoate, that showed very high abundance in the infected flies (Figures 7, 9). The prostaglandins are likely derived from the arachidonic acid as that feeds into the prostaglandin synthetic pathway.

DISCUSSION

This study observed significant differences in metabolites between uninfected and ZIKV infected mosquitoes. These

include differential abundances of pseudo-uridine and inosine metabolites which are associated with post transcriptional RNA modification activities. This was confirmed in a separate experimental assay showing significant enrichment in the levels of inosine among infected *Ae. albopictus*. RNA-editing enzymes, Adenosine deaminases that act on RNA (ADARs) deaminates double stranded RNA (dsRNA) converting adenosine (A) to inosine (I) in pre-mRNA hairpins (Bass and Weintraub, 1988). The conversion of A to I is read by the translation machinery as guanosine hence altering the protein sequence that's encoded by edited mRNA (Polson and Bass, 1994). *In silico* analysis using complete coding sequences of the ZIKV polyprotein have demonstrated that host-mediated RNA editing of adenosines (ADAR) play a role in molecular evolution of



ZIKV (Piontkivska et al., 2017). Further, most of the ADAR-edited transcripts are expressed in the central nervous system of *Drosophila* and hypothesized to result in more than one protein produced from a single gene (Palladino et al., 2000). In fact, a nuclease has been shown to cleave edited dsRNA in *Drosophila* demonstrating that ADARs may be involved in tagging viral RNA for degradation (Carpenter et al., 2009). Further, ADAR has been shown to result in hyper-editing in the sigma virus, a negative sense RNA virus that is a pathogen of *D. melanogaster* (Carpenter et al., 2009). ADAR is considered a part of the antiviral response, indeed the loss of *Adar* RNA editing induces the expression of immune-related genes in *Drosophila* (Carpenter et al., 2009). In order to establish causality between ZIKV infection and post transcriptional modification due to ADAR expression, future mechanistic studies are necessary.

In this study, ZIKV infection of *Ae. albopictus* resulted in hypoglycemia and an enrichment of phosphoenolpyruvate and ketone body 3- hydroxybutyrate which is an energy source produced when glucose is not readily available (Laffel, 1999). Energy depletion would be a consequence of a direct competition of resources between the mosquito and ZIKV (Maier et al., 1987) or the infected mosquito would require extra nutrients to restore damaged tissues or even to fuel the cost of mounting immune response (Ferdig et al., 1993; Ahmed et al., 2002). The results of our study corroborates the findings of Shrinet et al. (2018) which demonstrated that glycolysis/gluconeogenesis played an important role in both mono and co-infection of *Ae. aegypti* mosquitoes with CHIKV and DENV. In addition, using a proteomics differential approach with two-Dimensional differential in-Gel Electrophoresis (2D-DIGE), Tchankouo-Nguetcheu et al. (2010), studied the protein modulations in the midgut of *Ae. aegypti* after seven day oral infection with DENV-2 and CHIKV viruses. The study demonstrated that enzymes participating in the glycolytic pathway were upregulated

by CHIKV or DENV-2 suggesting extensive glucose utilization during midgut infection.

Serotonin levels were decreased among the infected individuals. Serotonin is a neurotransmitter and its immunoreactive neurons which innervate the chemosensory systems of mosquitoes suggest that serotonin is a neuromodulator in the chemosensory processes of disease vector mosquitoes (Siju et al., 2008). In fact, serotonin depletion in *Aedes triseriatus* resulted in low blood feeding levels but did not affect host seeking behavior (Novak and Rowley, 1994) and in *Ae. aegypti*, low serotonin levels resulted in prolonged probing as well as a lower blood feeding success rate (Novak et al., 1995). A decrease in serotonin levels may be indicative of the modulation of insect behavior and fitness effect of ZIKV infection.

Lipids in both the viral envelope and host cell membranes play important roles throughout the attachment process during viral infection (Lorizate and Kräusslich, 2011). In this study, a significant number of lipids that were significantly enriched due to ZIKV infection have not been previously determined. Our findings on lipid perturbations due to ZIKV infections corroborates the findings of Chotiwan et al. (2018), study which demonstrated that upon DENV infection of *Ae. aegypti*, a majority of lipids that showed greater than 2 -fold changes in intensity had higher abundances upon DENV infection.

Fatty acids play a very important role in virus protein synthesis as well as replication (Cordero et al., 2014). In this study, both eicosapentaenoic and arachidonic acid were upregulated among the infected individuals. The enzymatic oxidation of arachidonic acid results in the production of prostaglandin (Black et al., 1980). Our study demonstrated that infection of *Ae. albopictus* with ZIKV resulted in enriched levels of prostaglandins including prostaglandins H2. Prostaglandin A1 reduced Mayaro virus replication in *Ae. albopictus* mosquito cells

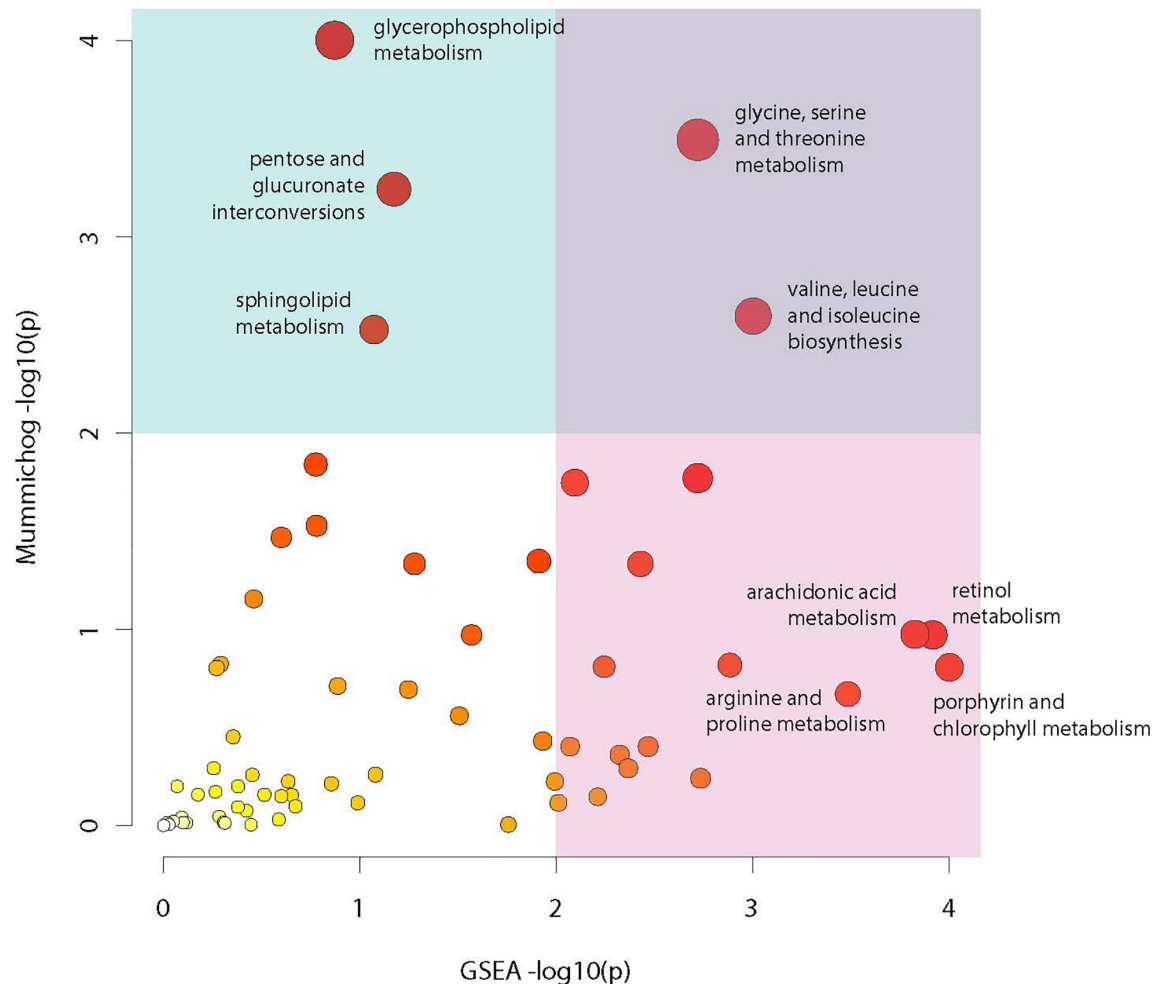


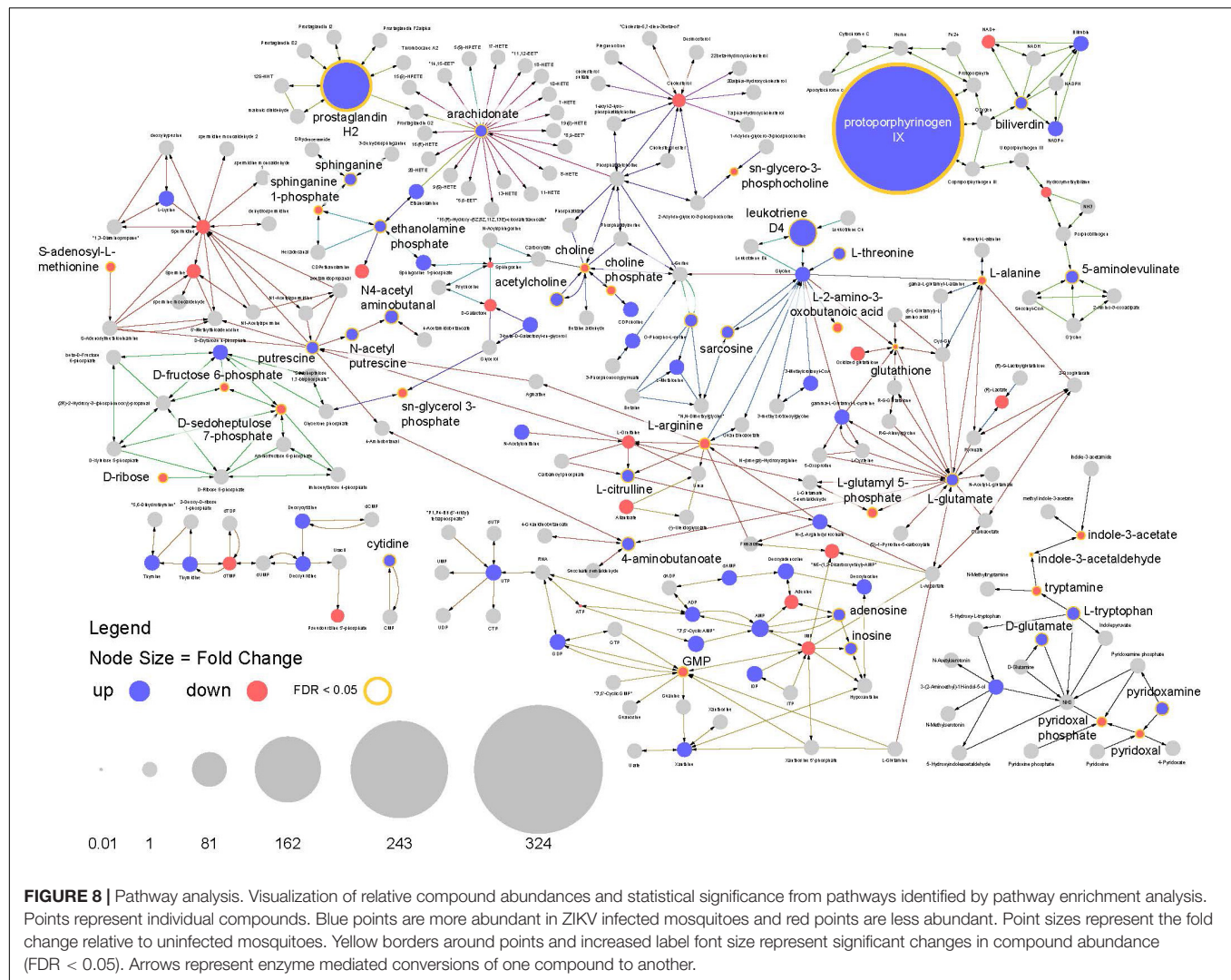
FIGURE 7 | Metabolic pathway enrichment. Visualization of enriched metabolic pathways as identified using the Mummichog pathway analysis package. Analysis utilized raw m/z and rt values for LC-MS profiles from the lipidomic and biogenic amine analysis panels in combination with p -values from statistical comparisons between treatment groups to identify pathways with perturbations in compound abundances in infected mosquitoes. The X axis represents $-\log_{10} p$ -value derived from the GSEA (Gene Set Enrichment Analysis) algorithm and the Y axis represents the $-\log_{10} p$ -value derived from the Mummichog enrichment analysis algorithm. Size and color of dots represents significance.

in culture by inhibiting virus-specific protein synthesis while in mock infected cells, the PGA1 stimulated the synthesis of several proteins (Barbosa and Rebello, 1995). Future mechanistic studies aimed at understanding the impact of prostaglandin on the immune priming of mosquitoes could result in novel transmission barrier mechanisms.

In our study, when we set the P cut-off value at $P \leq 0.1$, we measured a decrease of phosphatidylethanolamine (PE), important for the biophysical properties of membranes such as fluidity and curvature (Xu and Nagy, 2015). Indeed, lipid metabolites vital to virus assembly such as triacylglycerols (TAG) (Liefhebber et al., 2014) consisted the highest abundance relative to other compounds within the lipidome. Phosphatidylethanolamine (PE) and phosphatidylcholine (PC) were the second and third most abundant lipids respectively, phosphatidylcholine (PC) was increased in abundance among the infected individuals. The significance was lost when we decreased

the P -value threshold to ≤ 0.05 . The results of our studies differ from the results obtained from a study on lipid composition after DENV infection whereby PC, PE and phosphatidylserine levels were significantly enriched upon DENV infection of insect cell lines (Perera et al., 2012). In addition, glycerophospholipids, sphingolipids and fatty acyls were enriched in *Ae. aegypti* mosquitoes infected with DENV (Chotiwan et al., 2018). Further, our study findings differ from that of a study that utilized a whole cell lipidomics approach seeking to identify the cellular lipids important for WNV_{KUN} replication. This study observed an elevated level of phospholipase A2 as well as lyso phosphatidylcholine (Liebscher et al., 2018). We cannot rule out differences in pathogen vector host interactions as well as disease models utilized as causing the different results between these studies.

Further, biliverdin levels was upregulated among ZIKV-infected *Ae. albopictus*. Hemoglobin, the main blood protein is

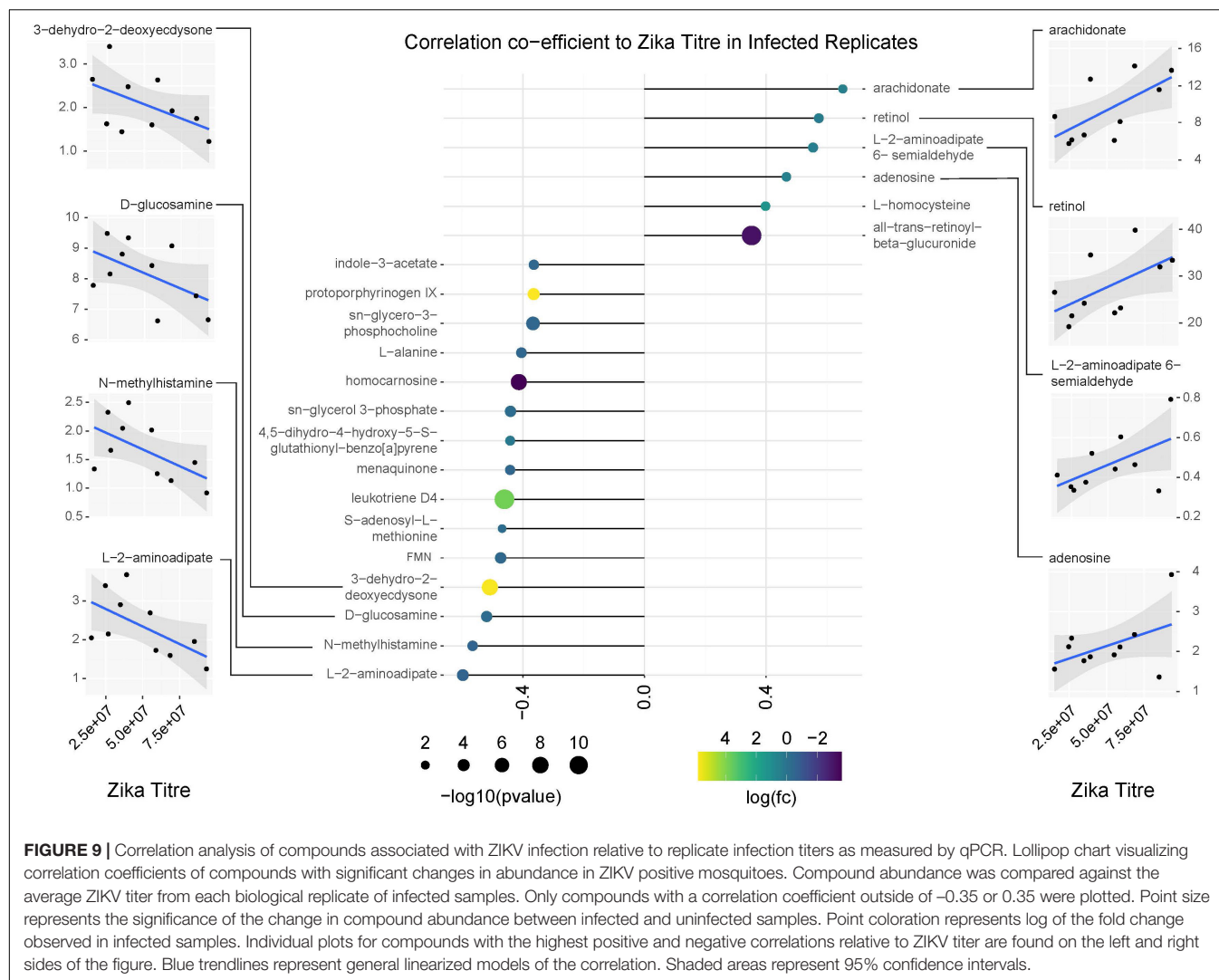


digested by mosquitoes to produce large amounts of heme which is further digested by heme oxygenase to produce biliverdin IX alpha, CO and free iron (Bottino-Rojas et al., 2019). Heme increases the microbial load in the mosquito midgut which has an antagonistic effect on DENV infection (Xi et al., 2008; Ramirez et al., 2012). In addition, heme also directly suppresses mosquito immune activity leading to a higher susceptibility of mosquitoes to DENV infection (Sim et al., 2013). Our study documents a > 300-fold increase in the compound protoporphyrinogen IX. This compound is synthesized from biliverdin. The antiviral properties of natural and synthetic porphyrins have been demonstrated (Staudinger et al., 1996; Vzorov et al., 2002; Chen-collins et al., 2003; Benati et al., 2009; Assuncao-Miranda et al., 2016). It has been hypothesized that the mechanism involved in viral inactivation by porphyrins to be dependent on its interaction with viral particles. In fact, DENV replication was inhibited only when porphyrins were in direct contact with newly formed particles or in infection with very low multiplicity of infection (MOI). Further porphyrins were shown to be capable of inhibiting the early stages of DENV

infection such as adsorption and penetration in susceptible cells (Assuncao-Miranda et al., 2016).

Though its primary role is to protect organisms from infection and injury, an overactive immune response can result in excessive inflammation and damaged organs. We measured an increase in levels of acetylcholine in ZIKV-infected individuals, suggesting it may have proviral function during ZIKV infection. Understanding how acetylcholine may modulate the immune pathways will be important for understanding viral tropism in mosquitoes.

In addition, in this study, histidine metabolism pathway was significantly perturbed by ZIKV infection with an enrichment of L-histidine and imidazole-4-acetaldehyde. Imidazoles are heterocycles with five-member ring structure. Imidazoles have gained importance due to their exceptional pharmacological activities (Kumar et al., 2017). In a search for new compounds with potential for clinical use as antiviral agents, high throughput screening of a compound library resulted in the identification of imidazole-4,5-dicarboxamide derivative and demonstrated it to have a high DENV inhibitory activity (Saudi et al., 2014).



Future mechanistic studies to clarify the role of imidazole-4-acetaldehyde in ZIKV infection is important.

In summary, elevation of pseudo-uridine and inosine in ZIKV-infected *Ae. albopictus* may be indicative of a post-transcriptional modification of RNAs as a result of ZIKV infection. However, mechanistic studies to establish causality is important. Future studies targeting select differentiated metabolites for targeted metabolic studies is very important in order to validate the observations made by this study. Understanding the interactions between proviral pseudo-uridine, inosine and ZIKV replication will impact our understanding of the process and pathology of ZIKV. Further, the results of our study suggest that infected mosquitoes may be impacted by negative fitness effects that increase transmission efficiency. The reduced levels of serotonin may affect salivary gland function resulting in decreased feeding efficiency and increased vector/host contact. The metabolites responsive to ZIKV infection identified in this study may represent an immune response by the mosquito and in future, upon validation may act as points to prevent ZIKV transmission. Overall, these findings

provide an array of insights into the biochemical interactions occurring during ZIKV infection in *Ae. albopictus* and provide potential targets that can inform the development of novel intervention strategies.

DATA AVAILABILITY STATEMENT

All datasets generated for this study are included in the article/**Supplementary Material**.

AUTHOR CONTRIBUTIONS

MO was involved in the study design, conducted the experiments, and drafted the manuscript. GA was involved in the study design, analyzed the results, and participated in the writing of the manuscript. SB, JS, and EB were involved in the mosquito rearing and assisted in mosquito manipulation exercises. EK was involved in data analysis and visualization. LK was generated and propagated

the ZIKV infectious clone utilized in this study. AC and LK were involved in the study design and participated in the writing of the manuscript. All the authors contributed to the article and approved the submitted version.

FUNDING

This publication was supported by the Cooperative Agreement Number U01CK000509 funded by the Center for Disease Control and Prevention. Its content is solely the responsibility of the authors and do not necessarily represent the official views of the Center for Disease Control and Prevention or the Department of Health and Human Services. We would like to extend our gratitude to Illia Rochlin of Suffolk County Health Department for providing us with *Ae. albopictus* mosquitoes and the Arbovirus laboratory insectary crew for mosquito maintenance. Data for these analyses were acquired at the University of California, Davis West Coast Metabolomics Center, U2C ES030158. We would like to express our gratitude to Dr. Alice Trimmer for her suggestions and edits.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmicb.2020.559035/full#supplementary-material>

REFERENCES

- Ahmed, A. M., Baggott, S. L., Maingon, R., and Hurd, H. (2002). The costs of mounting an immune response are reflected in the reproductive fitness of the mosquito *Anopheles gambiae*. *Oikos* 97, 371–377. doi: 10.1034/j.1600-0706.2002.970307.x
- Assuncao-Miranda, I., Cruz-Oliveira, C., Neris, R., Figueiredo, C., Pereira, L., Rodrigues, D., et al. (2016). Inactivation of Dengue and Yellow Fever viruses by heme, cobalt-protoporphyrin IX and tin-protoporphyrin IX. *J. Appl. Microbiol.* 120, 790–804. doi: 10.1111/jam.13038
- Barbosa, J., and Rebello, M. (1995). Prostaglandin A1 inhibits replication of Mayaro virus in *Aedes albopictus* cells. *Braz. J. Med. Biol. Res.* 28, 27–30.
- Bass, B., and Weintraub, H. (1988). An unwinding activity that covalently modifies its double-stranded RNA substrate. *Cell* 55, 1089–1098. doi: 10.1016/0092-8674(88)90253-x
- Benati, F. J., Lauretti, F., Faccin, L. C., Nodari, B., Ferri, D. V., Mantovani, M. S., et al. (2009). Effects of chlorophyllin on replication of poliovirus and bovine herpesvirus *in vitro*. *Lett. Appl. Microbiol.* 49, 791–795. doi: 10.1111/j.1472-765X.2009.02744.x
- Black, A. K., Fincham, N., Greaves, M. W., and Hensby, C. N. (1980). Time course changes in levels of arachidonic acid and prostaglandins D2 E2 F2a in human skin following ultraviolet irradiation. *Br. J. Clin. Pharmacol.* 10, 453–457. doi: 10.1111/j.1365-2125.1980.tb01788.x
- Bottino-Rojas, V., Pereira, L. O. R., Silva, G., Talyuli, O. A. C., Dunkov, B. C., Oliveira, P. L., et al. (2019). Non-canonical transcriptional regulation of heme oxygenase in *Aedes aegypti*. *Sci. Rep.* 9:13726. doi: 10.1038/s41598-019-49396-3
- Bujak, R., Struck-Lewicka, W., Markuszewski, M. J., and Kaliszan, R. (2015). Metabolomics for laboratory diagnostics. *J. Pharm. Biomed. Anal.* 113, 108–120. doi: 10.1016/j.jpba.2014.12.017
- Byers, N. M., Fleshman, A. C., Perera, R., and Molins, C. R. (2019). Metabolomic insights into human arboviral infections: Dengue, chikungunya, and zika viruses. *Viruses* 11:225. doi: 10.3390/v11030225
- Supplementary File 1** | An excel file showing the pooling of the *Ae. albopictus* samples used in the metabolomics study and demonstrating the ZIKV copy number standards in each individual associated with each of the pool.
- Supplementary File 2** | A table listing the internal standards included in the lipidomic and biogenic amine panels.
- Supplementary File 3** | Raw GC-MS peak values with annotations from primary metabolites panel.
- Supplementary File 4** | Raw LC-MS peak values with annotations from lipidomics panel.
- Supplementary File 5** | Raw LC-MS peak values with annotations from biogenic amines panel.
- Supplementary File 6** | Primary metabolite GC-MS peak data formatted for Metaboanalyst submission.
- Supplementary File 7** | Lipidomics LC-MS peak data formatted for Metaboanalyst submission.
- Supplementary File 8** | Biogenic amine LC-MS peak data formatted for Metaboanalyst submission.
- Supplementary File 9** | Combined Lipidomics and Biogenic amine LC-MS data formatted for submission to Peaks-to-Pathways (Mummichog) package on Metaboanalyst.
- Supplementary File 10** | Pathway enrichment data output from Peaks-to-Pathways.
- Supplementary File 11** | Compound KEGG identifications output from Peaks-to-Pathways.
- Supplementary File 12** | Excel workbook containing the normalized combined lipidomic and biogenic amine LC-MS datasets annotated with KEGG ids for correlation analysis and pathway analysis in Cytoscape.
- Carlile, T. M., Martinez, N. M., Schaening, C., Su, A., Tristan, A., Zinshteyn, B., et al. (2019). mRNA structure determines modification by pseudouridine synthase 1. *Nat. Chem. Biol.* 15, 966–974. doi: 10.1038/s41589-019-0353-z
- Carpenter, J. A., Keegan, L. P., Wilfert, L., O'Connell, M. A., and Jiggins, F. M. (2009). Evidence for ADAR-induced hypermutation of the *Drosophila* sigma virus (Rhabdoviridae). *BMC Genet.* 10:75. doi: 10.1186/1471-2156-10-75
- Chen-collins, A. R. M., Dixon, D. W., Vzorov, A. N., Marzilli, L. G., and Compans, R. W. (2003). Prevention of poxvirus infection by tetrapyrroles. *BMC Infect. Dis.* 3:9. doi: 10.1186/1471-2334-3-9
- Chintapalli, V. R., Al Bratty, M., Korzekwa, D., Watson, D. G., and Dow, J. A. T. (2013). Mapping an atlas of tissue-specific *Drosophila melanogaster* metabolomes by high resolution mass spectrometry. *PLoS One* 8:e78066. doi: 10.1371/journal.pone.0078066
- Chong, J., Soufan, O., Li, C., Caraus, I., Li, S., Bourque, G., et al. (2018). MetaboAnalyst 4.0: towards more transparent and integrative metabolomics analysis. *Nucleic Acids Res.* 46, W486–W494. doi: 10.1093/nar/gky310
- Chotiwan, N., Andre, B. G., Sanchez-Vargas, I., Islam, M. N., Grabowski, J. M., Hopf-Jannasch, A., et al. (2018). Dynamic remodeling of lipids coincides with dengue virus replication in the midgut of *Aedes aegypti* mosquitoes. *PLoS Pathog.* 14:e1006853. doi: 10.1371/journal.ppat.1006853
- Chu, H., Xin, B., Liu, P., Wang, Y., Li, L., Liu, X., et al. (2015). Metabolic engineering of *Escherichia coli* for production of (2S,3S)-butane-2,3-diol from glucose. *Biotechnol. Biofuels* 8:143. doi: 10.1186/s13068-015-0324-x
- Ciota, A., Bialosuknia, S., Zink, S., Brecher, M., Ehrbar, D., Morrisette, M., et al. (2017). Effects of Zika virus strain and *Aedes* mosquito species on vector competence. *Emerg. Infect. Dis.* 23, 1110–1117.
- Cook, S., and Holmes, E. C. (2006). A multigene analysis of the phylogenetic relationships among the flaviviruses (Family: *Flaviviridae*) and the evolution of vector transmission. *Arch. Virol.* 151, 309–325. doi: 10.1007/s00705-005-0626-6

- Cordero, J. G., Juárez, M. L., González-Y-Merchand, J. A., Barrón, L. C., and Castañeda, B. G. (2014). Caveolin-1 in lipid rafts interacts with dengue virus NS3 during polyprotein processing and replication in HMEC-1 cells. *PLoS One* 9:e90704. doi: 10.1371/journal.pone.0090704
- Defelice, B. C., Mehta, S. S., Samra, S., Cajka, T., Wancewicz, B., Fahrman, J. F., et al. (2017). Mass Spectral Feature List Optimizer (MS-FLO): a tool to minimize false positive peak reports in untargeted Liquid Chromatography-Mass Spectrometry (LC-MS) data processing. *Anal. Chem.* 89, 3250–3255. doi: 10.1021/acs.analchem.6b04372
- de Oliveira, W. K., Vinicius, G., de França, A., Carmo, E. H., Duncan, B. B., Kuchenbecker, R. D. S., et al. (2017). Infection-related microcephaly after the 2015 and 2016 Zika virus outbreaks in Brazil: a surveillance-based analysis. *Lancet* 390, 861–870. doi: 10.1016/S0140-6736(17)31368-5
- Diamond, D. L., Syder, A. J., Jacobs, J. M., Sorensen, C. M., Walters, K. A., Prohl, S. C., et al. (2010). Temporal proteome and lipidome profiles reveal hepatitis C virus-associated reprogramming of hepatocellular metabolism and bioenergetics. *PLoS Pathog.* 6:e1000719. doi: 10.1371/journal.ppat.1000719
- Dick, G. (1952). Zika virus. II. Pathogenicity and physical properties. *Trans. R. Soc. Trop. Med. Hyg.* 46, 521–534. doi: 10.1016/0035-9203(52)90043-6
- Dick, G. W., Kitchen, S., and Haddow, A. (1952). Zika virus. I. Isolations and serological specificity. *Trans. R. Soc. Trop. Med. Hyg.* 46, 509–520. doi: 10.1016/0035-9203(52)90042-4
- Dong, S., Zhang, R., Liang, Y., Shi, J., Li, J., Shang, F., et al. (2017). Changes of myocardial lipidomics profiling in a rat model of diabetic cardiomyopathy using UPLC/Q-TOF/MS analysis. *Diabetol. Metab. Syndr.* 9:56. doi: 10.1186/s13098-017-0249-6
- Eijkelkamp, B. A., Begg, S. L., Pederick, V. G., Trapetti, C., Gregory, M. K., Whittall, J. J., et al. (2018). Arachidonic acid stress impacts pneumococcal fatty acid homeostasis. *Front. Microbiol.* 9:813. doi: 10.3389/fmicb.2018.00813
- Faye, O., Freire, C. C. M., Iamarino, A., Faye, O., de Oliveira, J. V. C., Diallo, M., et al. (2014). Molecular evolution of Zika virus during its emergence in the 20th century. *PLoS Negl. Trop. Dis.* 8:e2636. doi: 10.1371/journal.pntd.0002636
- Ferdig, M., Beerntsen, B., Spray, F., Li, J., and Christensen, B. (1993). Reproductive costs associated with resistance in a mosquito-filarial worm system. *Am. J. Trop. Med. Hyg.* 49, 756–762. doi: 10.4269/ajtmh.1993.49.756
- Fiehn, O. (2017). Metabolomics by Gas Chromatography-Mass Spectrometry: the combination of targeted and untargeted profiling. *Curr. Protoc. Mol. Biol.* 114, 21.33.1–21.33.11. doi: 10.1002/0471142727.mb3004s114
- Garnier, S. (2018). *viridisLite: Default Color Maps from 'matplotlib' (Lite Version)*. Available online at: <https://cran.r-project.org/web/packages/viridisLite/viridisLite.pdf> (accessed June 14, 2020).
- Gulland, A. (2016). Zika virus is a global public health emergency, declares WHO. *BMJ* 352:i657. doi: 10.1136/bmj.i657
- Hayes, E. B. (2009). Zika virus outside Africa. *Emerg. Infect. Dis.* 15, 2007–2010. doi: 10.3201/eid1509.090442
- Heaton, N. S., and Randall, G. (2010). Dengue virus-induced autophagy regulates lipid metabolism. *Cell Host Microbe* 8, 422–432. doi: 10.1016/j.chom.2010.10.006
- Hirashima, A., Sukhanova, M. J., and Rauschenbach, I. Y. (2000). Biogenic amines in *Drosophila virilis* under stress conditions. *Biosci. Biotechnol. Biochem.* 64, 2625–2630. doi: 10.1271/bbb.64.2625
- Ito, S., and Nakata, F. (2016). *Direct Analysis of Biogenic Amines in Food by HILIC MS*. Available online at: <https://separations.eutoshbioscience.com/OpenPDF.aspx?path=/File%20Library/TBG/Products%20Download//Application%20Note/a15148a.pdf> (accessed June 14, 2020).
- Johnson, C. H., Ivanisevic, J., and Siuzdak, G. (2016). Metabolomics: beyond biomarkers and towards mechanisms. *Nat. Rev. Mol. Cell Biol.* 17, 451–459. doi: 10.1038/nrm.2016.25
- Karnovsky, A., Weymouth, T., Hull, T., Glenn Tarcea, V., Scardoni, G., Laudanna, C., et al. (2012). Metscape 2 bioinformatics tool for the analysis and visualization of metabolomics and gene expression data. *Bioinformatics* 28, 373–380. doi: 10.1093/bioinformatics/btr661
- Kazmi, S. S., Ali, W., Bibi, N., and Nouroz, F. (2020). A review on Zika virus outbreak, epidemiology, transmission and infection dynamics. *J. Biol. Res.* 27:5. doi: 10.1186/s40709-020-00115-4
- Kind, T., Liu, K.-H., Lee, D. Y., DeFelice, B., Meissen, J. K., and Fiehn, O. (2013). LipidBlast - in-silico tandem mass spectrometry database for lipid identification. *Nat. Methods* 10, 755–758. doi: 10.1016/j.physbeh.2017.03.040
- Kong, H. G., Shin, T. S., Kim, T. H., and Ryu, C. M. (2018). Stereoisomers of the bacterial volatile compound 2,3-butanediol differentially elicit systemic defense responses of pepper against multiple viruses in the field. *Front. Plant Sci.* 9:90. doi: 10.3389/fpls.2018.00090
- Kuhn, M., Jackson, S., and Cimentada, J. (2020). *corr: Correlations in R*. Available online at: <https://CRAN.R-project.org/package=corr> (accessed June 14, 2020).
- Kumar, M., Kumar, D., and Raj, V. (2017). Studies on imidazole and its derivatives with particular emphasis on their chemical/biological applications as bioactive molecules/intermediated to bioactive molecule. *Curr. Synth. Syst. Biol.* 5, 135–145. doi: 10.4172/2332-0737.1000135
- Laffel, L. (1999). Ketone bodies: a review of physiology, pathophysiology and application of monitoring to diabetes. *Diabetes Metab. Res. Rev.* 15, 412–426. doi: 10.1002/(sici)1520-7560(199911/12)15:6<412::aid-dmrr72>3.0.co;2-8
- Lancioti, R. S., Calisher, C. H., Gubler, D. J., Chang, G. J., and Vorndam, A. V. (1992). Rapid detection and typing of dengue viruses from clinical samples by using reverse transcriptase-polymerase chain reaction. *J. Clin. Microbiol.* 30, 545–551. doi: 10.1128/jcm.30.3.545-551.1992
- Li, S., Park, Y., Duraisingham, S., Strobel, F. H., Khan, N., Soltow, Q. A., et al. (2013). Predicting network activity from high throughput metabolomics. *PLoS Comput. Biol.* 9:e1003123. doi: 10.1371/journal.pcbi.1003123
- Liebscher, S., Ambrose, R. L., Aktepe, T. E., Mikulasova, A., Prier, J. E., Gillespie, L. K., et al. (2018). Phospholipase A2 activity during the replication cycle of the flavivirus West Nile virus. *PLoS Pathog.* 14:e1007029. doi: 10.1371/journal.ppat.1007029
- Liefhebber, J. M. P., Hague, C. V., Zhang, Q., Wakelam, M. J. O., and McLauchlan, J. (2014). Modulation of triglyceride and cholesterol ester synthesis impairs assembly of infectious hepatitis C virus. *J. Biol. Chem.* 289, 21276–21288. doi: 10.1074/jbc.M114.582999
- Lorizate, M., and Kräusslich, H.-G. (2011). Role of lipids in virus replication. *Cold Spring Harb. Perspect. Biol.* 3:a004820. doi: 10.1101/cshperspect.a004820
- Maier, W., Becker-Feldman, H., and Seitz, H. (1987). Pathology of malaria-infected mosquitoes. *Parasitol. Today* 3, 216–218. doi: 10.1016/0169-4758(87)90063-9
- Martin-Acebes, M. A., Merino-Ramos, T., Blazquez, A.-B., Casas, J., Escribano-Romero, E., Sobrino, F., et al. (2014). The composition of West Nile virus lipid envelope unveils a role of sphingolipid metabolism in flavivirus biogenesis. *J. Virol.* 88, 12041–12054. doi: 10.1128/jvi.02061-14
- Matyash, V., Liebisch, G., Kurzchalia, T. V., Shevchenko, A., and Schwudke, D. (2008). Lipid extraction by methyl-terf-butyl ether for high-throughput lipidomics. *J. Lipid Res.* 49, 1137–1146. doi: 10.1194/jlr.D700041-JLR200
- Melo, C. F. O. R., De Oliveira, D. N., De Oliveira Lima, E., Guerreiro, T. M., Esteves, C. Z., Beck, R. M., et al. (2016). A lipidomics approach in the characterization of zika-infected mosquito cells: potential targets for breaking the transmission cycle. *PLoS One* 11:e0164377. doi: 10.1371/journal.pone.0164377
- Moffett, S. B., and Moffett, D. F. (2005). Comparison of immunoreactivity to serotonin, FMRFamide and SCPb in the gut and visceral nervous system of larvae, pupae and adults of the yellow fever mosquito *Aedes aegypti*. *J. Insect Sci.* 5:20. doi: 10.1673/031.005.2001
- Motzkin, T. (1949). The euclidean algorithm. *Bull. Am. Soc.* 55, 1142–1146.
- Newman, J. C., and Verdin, E. (2017). β -Hydroxybutyrate: a signaling metabolite. *Annu. Rev. Nutr.* 37, 51–76. doi: 10.1146/annurev-nutr-071816-064916
- Novak, M., and Rowley, W. (1994). Serotonin depletion affects blood-feeding but not host-seeking ability in *Aedes triseriatus* (Diptera: Culicidae). *J. Med. Entomol.* 31, 600–606. doi: 10.1093/jmedent/31.4.600
- Novak, M. G., Ribeiro, J. M. C., and Hildebrand, J. G. (1995). 5-Hydroxytryptamine in the salivary glands of adult female *Aedes aegypti* and its role in regulation of salivation. *J. Exp. Biol.* 198, 167–174.
- Onyango, M. G., Bialosuknia, S. M., Payne, A. F., Mathias, N., Kuo, L., Vigneron, A., et al. (2020). Increased temperatures reduce the vectorial capacity of *Aedes* mosquitoes for Zika virus. *Emerg. Microbes Infect.* 9, 67–77. doi: 10.1080/22221751.2019.1707125
- Paiva-silva, G. O., Cruz-oliveira, C., Nakayasu, E. S., Maya-monteiro, C. M., Dunkov, B. C., Masuda, H., et al. (2006). A heme-degradation pathway in a blood-sucking insect. *Proc. Natl. Acad. Sci. U.S.A.* 103, 8030–8035. doi: 10.1073/pnas.0602224103

- Palladino, M. J., Keegan, L. P., O'Connell, M. A., and Reenan, R. A. (2000). A-to-I pre-mRNA editing in *Drosophila* is primarily involved in adult nervous system function and integrity. *Cell* 102, 437–449. doi: 10.1016/S0092-8674(00)00049-0
- Peña, J., and Harris, E. (2012). Early dengue virus protein synthesis induces extensive rearrangement of the endoplasmic reticulum independent of the UPR and SREBP-2 pathway. *PLoS One* 7:e38202. doi: 10.1371/journal.pone.0038202
- Peng, Q., Berg, K., Moan, J., Kongshaug, M., and Nesland, J. M. (1997). 5-Aminolevulinic acid-based photodynamic therapy: principles and experimental research. *Photochem. Photobiol.* 65, 235–251. doi: 10.1111/j.1751-1097.1997.tb08549.x
- Perera, R., Riley, C., Isaac, G., Hopf-Jannasch, A. S., Moore, R. J., Weitz, K. W., et al. (2012). Dengue virus infection perturbs lipid homeostasis in infected mosquito cells. *PLoS Pathog.* 8:e1002584. doi: 10.1371/journal.ppat.1002584
- Petersen, L. R., Jamieson, D. J., Powers, A. M., and Honein, M. A. (2016). Zika Virus. *N. Engl. J. Med.* 374, 1552–1563. doi: 10.1056/NEJMra1602113
- Picciotto, M. R., Higley, M., and Mineur, Y. (2012). Acetylcholine as a neuromodulator: cholinergic signaling shapes nervous system function and behavior. *Neuron* 76, 116–129. doi: 10.1016/j.neuron.2012.08.036
- Piontkivska, H., Frederick, M., Miyamoto, M. M., and Wayne, M. L. (2017). RNA editing by the host ADAR system affects the molecular evolution of the Zika virus. *Ecol. Evol.* 7, 4475–4485. doi: 10.1002/ece3.3033
- Pluskal, T., Castillo, S., Villar-Briones, A., and Orešić, M. (2010). MZmine 2: modular framework for processing, visualizing, and analyzing mass spectrometry-based molecular profile data. *BMC Bioinformatics* 11:395. doi: 10.1186/1471-2105-11-395
- Polson, A. G., and Bass, B. L. (1994). Preferential selection of adenosines for modification by double-stranded RNA adenosine deaminase. *EMBO J.* 13, 5701–5711. doi: 10.1002/j.1460-2075.1994.tb06908.x
- R Core Team (2003). *R: A Language and Environment for Statistical Computing*. Vienna: R Foundation for Statistical Computing.
- Rajendran, R., Borghi, E., Falleni, M., Perdoni, F., Tosi, D., Lappin, D. F., et al. (2015). Acetylcholine protects against *Candida albicans* infection by inhibiting biofilm formation and promoting hemocyte function in a *Galleria mellonella* infection model. *Am. Soc. Microbiol.* 14, 834–844. doi: 10.1128/EC.00067-15
- Ramirez, J. L., Souza-Neto, J., Cosme, R. T., Rovira, J., Ortiz, A., Pascale, J. M., et al. (2012). Reciprocal tripartite interactions between the *Aedes aegypti* midgut microbiota, innate immune system and dengue virus influences vector competence. *PLoS Negl. Trop. Dis.* 6:e1561. doi: 10.1371/journal.pntd.0001561
- Saudi, M., Zmurko, J., Kaptein, S., Rozenski, J., Neyts, J., and Van Aerschot, A. (2014). Synthesis and evaluation of imidazole-4,5-and pyrazine-2,3-dicarboxamides targeting dengue and yellow fever virus. *Eur. J. Med. Chem.* 87, 529–539. doi: 10.1016/j.ejmech.2014.09.062
- Schrompe-Rutledge, A., Codreanu, S., Sherrod, S., and McLean, J. (2016). Untargeted metabolomics strategies-challenges and emerging directions. *J. Am. Soc. Mass Spectrom.* 27, 1897–1905. doi: 10.1016/j.physbeh.2017.03.040
- Servetnick, D., Bryant, D., Wells-Knecht, K., and Wiesenfeld, P. (1996). L-Arginine inhibits in vitro nonenzymatic glycation and advanced glycosylated end product formation of human serum albumin. *Amino Acids* 11, 69–81. doi: 10.1007/BF00805722
- Shannon, P., Markiel, A., Ozier, O., Baliga, N. S., Wang, J. T., Ramage, D., et al. (2003). Cytoscape: a software environment for integrated models of biomolecular interaction networks. *Genome Res.* 13, 2498–2504. doi: 10.1101/gr.1239303.metabolite
- Shrinet, J., Bhavesh, N. S., and Sunil, S. (2018). Understanding oxidative stress in *Aedes* during Chikungunya and Dengue virus infections using integrative analysis. *Viruses* 10:314. doi: 10.3390/v10060314
- Siju, K., Hansson, B., and Ignell, R. (2008). Immunocytochemical localization of serotonin in the central and peripheral chemosensory system of mosquitoes. *Arthropod Struct. Dev.* 37, 248–259. doi: 10.1016/j.asd.2007.12.001
- Sim, S., Jupatanakul, N., Ramirez, J. L., Kang, S., Romero-Vivas, C. M., Mohammed, H., et al. (2013). Transcriptomic profiling of diverse *Aedes aegypti* strains reveals increased basal-level immune activation in dengue virus-refractory populations and identifies novel virus-vector molecular interactions. *PLoS Negl. Trop. Dis.* 7:e2295. doi: 10.1371/journal.pntd.0002295
- Singh, P., Sharma, V., Singha, S., Sakai, V., and Mukhopadhyay, R. (2019). Unraveling the role of monolein in fluidity and dynamical response of a mixed cationic lipid bilayer. *Langmuir* 35, 4682–4692. doi: 10.1021/acs.langmuir.9b00043
- Skogerson, K., Wohlgemuth, G., Barupal, D. K., and Fiehn, O. (2011). The volatile compound BinBase mass spectral database. *BMC Bioinformatics* 12:321. doi: 10.1186/1471-2105-12-321
- Staudinger, R., Abraham, N., Levere, R., and Kappas, A. (1996). Inhibition of human immunodeficiency virus-1 reverse transcriptase by heme and synthetic heme analogs. *Proc. Assoc. Am. Physicians* 108, 47–54.
- Sternberg, E. M. (1997). Neural-immune interactions in health and disease. *J. Clin. Invest.* 100, 2641–2647. doi: 10.1172/jci119807
- Sumner, L. W., Amberg, A., Barrett, D., Beale, M. H., Beger, R., Daykin, C. A., et al. (2007). Proposed minimum reporting standards for chemical analysis. *Metabolomics* 3, 211–221. doi: 10.1007/s11306-007-0082-2
- Tchankouo-Nguetcheu, S., Khun, H., Pincet, L., Roux, P., Bahut, M., Huerre, M., et al. (2010). Differential protein modulation in midguts of *Aedes aegypti* infected with chikungunya and dengue 2 viruses. *PLoS One* 5:e13149. doi: 10.1371/journal.pone.0013149
- Tsugawa, H., Cajka, T., Kind, T., Ma, Y., Higgins, B., Ikeda, K., et al. (2015). MS-DIAL: data independent MS/MS deconvolution for comprehensive metabolome analysis. *Nat. Methods* 12, 523–526. doi: 10.1016/j.physbeh.2017.03.040
- Vanella, A., Russo, A., Acquaviva, R., Campisi, A., Di Giacomo, C., Sorrenti, V., et al. (2000). L-Propionyl-carnitine as superoxide scavenger, antioxidant, and DNA cleavage protector. *Cell Biol. Toxicol.* 16, 99–104.
- Ventura, C. V., Maia, M., Bravo-Filho, V., Góis, A. L., and Belfort, R. (2016). Zika virus in Brazil and macular atrophy in a child with microcephaly. *Lancet* 387:228. doi: 10.1016/S0140-6736(16)00006-4
- Vzorov, A. N., Dixon, D. W., Trommel, J. S., Marzilli, L. G., and Compans, R. W. (2002). Inactivation of human immunodeficiency virus Type 1 by porphyrins. *Antimicrob. Agents Chemother.* 46, 3917–3925. doi: 10.1128/AAC.46.12.3917
- Ward, J., and Hook, M. (1963). Application of an hierarchical grouping procedure to a problem of grouping profiles. *Educ. Psychol. Meas.* 23, 69–81. doi: 10.1177/001316446302300107
- Wickham, H. (2016). *ggplot2: Elegant Graphics for Data Analysis*. New York, NY: Springer-Verlag.
- Wickham, H., François, R., Henry, L., and Müller, K. (2020). *dplyr: A Grammar of Data Manipulation 2020*. Vienna: R Foundation for Statistical Computing.
- Wickham, H., and Henry, L. (2020). *tidyr: Tidy Messy Data*. Available online at: <https://cran.r-project.org/web/packages/tidyr/index.html> (accessed June 14, 2020).
- Wikan, N., and Smith, D. R. (2016). Zika virus: history of a newly emerging arbovirus. *Lancet Infect. Dis.* 16, E119–E126. doi: 10.1016/S1473-3099(16)30010-X
- Windmueller, H., and Spaeth, A. (1981). Source and fate of circulating citrulline. *Am. J. Physiol.* 241, E473–E480.
- World Health Organization [WHO] (2019). *ZIKA EPIDEMIOLOGY UPDATE*. Geneva: WHO.
- Xi, Z., Ramirez, J. L., and Dimopoulos, G. (2008). The *Aedes aegypti* toll pathway controls dengue virus infection. *PLoS Pathog.* 4:e1000098. doi: 10.1371/journal.ppat.1000098
- Xu, K., and Nagy, P. D. (2015). RNA virus replication depends on enrichment of phosphatidylethanolamine at replication sites in subcellular membranes. *Proc. Natl. Acad. Sci. U.S.A.* 112, E1782–E1791. doi: 10.1073/pnas.1418971112
- Zinshteyn, B., and Nishikura, K. (2009). Adenosine-to-inosine RNA editing. *Wiley Interdiscip. Rev. Syst. Biol. Med.* 1, 202–209. doi: 10.1002/wsbm.10

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Onyango, Attardo, Kelly, Bialosuknia, Stout, Banker, Kuo, Ciota and Kramer. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.