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Generation of T-cell-receptor-negative CD8 $\alpha\beta$ -positive CAR T cells from T cell-derived induced pluripotent stem cells

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Abstract

The production of autologous T cells expressing a chimaeric antigen receptor (CAR) is time-consuming, costly and occasionally unsuccessful. T-cell-derived induced pluripotent stem cells (TiPS) are a promising source for the generation of ‘off-the-shelf’ CAR T cells, but the in vitro differentiation of TiPS often yields T cells with suboptimal features. Here we show that the premature expression of the T-cell receptor (TCR) or a constitutively expressed CAR in TiPS promotes the acquisition of an innate phenotype, which can be averted by disabling the TCR and relying on the CAR to drive differentiation. Delaying CAR expression and calibrating its signalling strength in TiPS enabled the generation of human TCR– CD8 $\alpha\beta$ + CAR T cells that perform similarly to CD8 $\alpha\beta$ + CAR T cells from peripheral blood, achieving effective tumour control on systemic administration in a mouse model of leukaemia and without causing

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Author contributions.

S.J.C.v.d.S. designed the study, performed experiments, analysed and interpreted data, and wrote the manuscript. P.L.L. and R.M.P. performed experiments, analysed and interpreted data. H.X. performed RNA-sequencing analysis. M.P.D., V.A., Y.Z., M.H., J.M.S., J.E. and A.C. performed experiments. X.W. and I.R. generated and provided clinical experimental materials. R.A., T.L., R.C., and B.V. generated TiPS lines, developed iCD34 methodology. M.T. and I.R. contributed to experimental design and data analysis. M.S. designed the study, analysed and interpreted data and wrote the manuscript.

Competing interests.

R.A., T.L., R.C., B.V., are employees of Fate Therapeutics Inc. and have equity in the company. M.S. reports research funding from Takeda Pharmaceuticals, Atara Biotherapeutics and Fate Therapeutics. M.S. served on the scientific advisory board of St Jude Children’s Research Hospital.

graft-versus-host disease. Driving T-cell maturation in TiPS in the absence of a TCR by taking advantage of a CAR may facilitate the large-scale development of potent allogeneic CD8 $\alpha\beta$ + T cells for a broad range of immunotherapies.

T cells that are engineered to express a chimaeric antigen receptor (CAR) can direct potent therapeutic responses in patients with chemorefractory haematologic malignancies¹. CARs are synthetic receptors that redirect T cell specificity and augment T cell functions to overcome tumour resistance². CAR T cells are now under investigation in a range of diseases, including solid tumours, infectious disease, autoimmunity and senescence-associated pathologies³⁻⁵. CAR T cells are generally produced in autologous fashion, which is effective but presents some challenges. Manufacturing time is critical for patients with rapidly progressing disease; cell product variability is high because patient T cells may be reduced in number or functionality due to progressive disease or previous therapies; manufacturing processes and release testing are required on an individual basis and are costly^{6, 7}. Alternate T cell sources are under investigation to enable 'off-the-shelf' cellular therapy, including donor-derived lymphoid progenitors, T cells lacking alloreactive potential (virus-specific T cells, $\gamma\delta$ TCR-T cells, invariant natural killer T (iNKT) cells and TCR-edited T cells) and pluripotent stem cell-derived T cells^{8, 9}. The use of allogeneic T cells collected from healthy donors is the most explored alternative source, with some promising clinical results already obtained¹⁰. However, maintaining product consistency after genetic engineering and manufacturing large cell batches of sufficient cell purity to avert graft-versus host disease (GvHD) remain a challenge¹¹. Pluripotent stem cells provide an attractive solution to overcome challenges associated with the use of autologous or allogeneic blood cells¹². Their self-renewing capacity should facilitate the selection of clones of a desired genotype, eventually combining multiple edits to enhance anti-tumour functions and pre-empt alloreactivity and allorejection, and support large-scale production^{8, 13, 14}.

We have previously demonstrated that CAR T cells can be generated from reprogrammed T-cell-derived induced pluripotent stem cells (TiPS)¹². These original TiPS were retrovirally transduced to express a CD19-specific CAR (CAR-TiPS) and re-differentiated into T cells. The CAR T cells induced therefrom (CAR-iT) homogeneously expressed both the transduced CAR (1928z) and the endogenous $\alpha\beta$ TCR. The CAR-iT cells were specific for CD19, highly lytic and controlled tumour growth in an intraperitoneal lymphoma model in NSG mice¹². Despite expressing their endogenous $\alpha\beta$ TCR, these CAR-iT cells did not acquire a conventional CD4 or CD8 $\alpha\beta$ T cell phenotype, but rather an innate-like CD4⁻CD8 $\alpha\beta$ ⁻ double-negative (DN) or CD8 $\alpha\alpha$ single-positive (SP) phenotype. Transcriptional analyses confirmed that these CAR-iT cells more closely resembled $\gamma\delta$ TCR-T cells than $\alpha\beta$ TCR-T cells¹². This lineage divergence was recently observed in another model wherein the constitutive expression of an LMP2-specific CAR in TiPS yielded DN and CD8 α T cell phenotypes¹⁵. In another report, the lymphoid differentiation of induced pluripotent stem cells expressing a GPC3-specific CAR failed to induce characteristic T-cell markers such as CD5 and CD3 and rather produced natural killer (NK) or innate lymphoid cells (ILC)¹⁶. These reports raise the questions of why T lineage commitment is deferred towards innate phenotypes and what is required to induce CD8 $\alpha\beta$ CAR T cells.

Here we set out to analyse the impact of TCR and CAR expression on the commitment of TiPS-derived lymphoid progenitors to adaptive T-cell lineages. Physiological $\alpha\beta$ TCR-T-cell development is regulated by TCR gene recombination, Notch and preTCR/TCR signalling¹⁷. The TCR and Notch are central to the adoption of either $\gamma\delta$ TCR- or $\alpha\beta$ TCR-T-cell fates. The earlier rearrangement of the γ - and δ -chains results in maturation of $\gamma\delta$ TCR-T-cells with a DN or CD8 α SP phenotype¹⁸, whereas successful pre-TCR and $\alpha\beta$ TCR assembly drive progression from the DN to the CD4⁺CD8 $\alpha\beta$ ⁺ double positive (DP) stage before yielding SP T-cells^{19, 20}. We show that premature $\alpha\beta$ TCR or constitutive CAR expression interfere with DP formation, depending on the strength of Notch stimulation. Delaying CAR expression through *TRAC* promoter-controlled expression and calibration of CAR signalling through CD3 ζ immunoreceptor tyrosine activation motif (ITAM) mutations enable DP CAR⁺ induced T (iT) cell development. In absence of an $\alpha\beta$ TCR, the CAR drives T-cell maturation and yields CD8 $\alpha\beta$ ⁺ CAR⁺ iT cells that mediate durable remissions in a systemic leukaemia model.

Results

Delta-like ligand 4 (DLL4) stimulation facilitates CD4⁺CD8 $\alpha\beta$ ⁺ DP iT cell development from WT-TiPS but not CAR-TiPS.

We determined the yield of CD4⁺CD8 $\alpha\beta$ ⁺ DP $\alpha\beta$ TCR-T cell precursors from pluripotent stem cells using the OP9-mDLL1 stromal cell line in the differentiation protocol shown in Fig. 1a. The human embryonic stem (ES) cell line H1 and fibroblast-derived induced pluripotent stem cells (FiPS) cells consistently yielded a DP population, typically arising by day 35 and followed by the appearance of CD3⁺ cells by day 40 (Fig. 1b and Extended Data Fig. 1a,b). When differentiating unmodified, wild-type TiPS (WT-TiPS, Extended Data Fig. 1a) under the same conditions, very few DP cells were induced (Fig. 1b). Nonetheless, CD3⁺ $\alpha\beta$ TCR⁺ cells were generated, appearing much earlier, typically by day 25 (Fig. 1b and Extended Data Fig. 1b) and eschewing the DP intermediate state.

As the variable (V), diversity (D), and junctional (J) $\alpha\beta$ TCR genes are pre-rearranged in WT-TiPS, in contrast to their germline configuration in ES and FiPS cells, we reasoned that V gene transcription, which normally precedes VDJ recombination, would result in the premature expression of rearranged $\alpha\beta$ TCR genes in TiPS. We further hypothesised that the early formation of an $\alpha\beta$ TCR would mimic the earlier timing of a productive $\gamma\delta$ TCR rearrangement, and thus bypass DP cell formation and impart an innate phenotype¹⁸. To preclude early TCR assembly, we abolished TCR α chain expression by disrupting the *TRAC* locus (Extended Data Fig. 2a–c). Disrupting the *TRAC* locus (*TRAC*^{-/-}-TiPS) indeed allowed for increased DP cell formation from *TRAC*^{-/-}-TiPS compared with WT-TiPS (Fig. 1c and Extended Data Fig. 2d), supporting the notion that early TCR signalling prevents $\alpha\beta$ TCR-T cell lineage commitment²¹. The altered lineage commitment also corroborated that our differentiation protocol (Fig. 1a) supports DP cell development and pointed to the critical importance of the timing of TCR assembly in determining the fate of TiPS-derived T cells.

As Notch signalling also plays a critical role at the $\alpha\beta$ - versus $\gamma\delta$ -lineage commitment junction²², we investigated its role by first assessing different Notch ligands for their ability

to support T cell differentiation from TiPS. We engineered OP9 stromal cells to express either one of the four human Notch ligands, Delta-like ligand 1 (DLL1), Delta-like ligand 4 (DLL4), Jagged-1 (JAG1) or Jagged-2 (JAG2) (Extended Data Fig. 3a, b). These ligands displayed a gradation in their level of Notch signalling induction (Extended Data Fig. 3c) and, correspondingly, their ability to support T lineage commitment and DP formation from WT-TiPS (Fig. 1d and Extended Data Fig. 3d). DLL1 and JAG1 were unable to support DP T cell development, in contrast with JAG2 and DLL4, the latter showing the greatest efficiency in both T lineage commitment (CD7⁺CD5⁺ positive cells, Extended Data Fig. 3d) and DP formation (Fig. 1d, e). Interestingly, DLL4 has been previously shown to efficiently support *in vitro* T cell differentiation after TCR gene rearrangement²³ and induces the strongest signalling from Notch1²⁴ (Extended Data Fig. 3c).

These findings support a model wherein DP cell formation depends on an intricate interaction between TCR and Notch stimulation, in which a potent DLL4-mediated signal is required in WT-TiPS because of the earlier expression of a functional TCR, whereas DLL1 suffices for H1 and *TRAC*^{-/-}-TiPS. The requirement for stronger Notch engagement in the context of earlier TCR assembly suggests that the TCR interferes with Notch signalling, which can be over-ridden by more potent Notch ligands, and that a strong activation signal from the CAR^{25, 26} and the TCR would offset stronger Notch signalling. Consistent with this model, we found that TiPS that constitutively expressed the 1928z CAR (CAR-TiPS, Extended Data Fig. 3e), had increased levels of extracellular signal-regulated kinase (ERK)1/2 phosphorylation (Extended Data Fig. 3f), induced fewer CD7⁺CD5⁺ cells, and did not produce DP cells, even in the presence of DLL4, instead generating DN and CD8 α iT cells (Fig. 1f, g and Extended Data Fig. 3g). To assess whether premature CAR signalling induces apoptosis in emerging DP cells, we measured apoptotic cells at the different developmental stages (DN, DP, CD4 induced SP, CD8 α SP and CD8 α β SP) in WT-TiPS and CAR-TiPS from D27 – D35, when the induction of the DP population occurs in WT-TiPS (Extended Data Fig. 3h). Levels of apoptosis were uniformly low (<5%) in both WT-TiPS and CAR-TiPS, and similar in all different developmental stages, suggesting that the lack of DP establishment from CAR-TiPS is not due to global apoptosis of the DP population. To verify whether the development of CD8 α β iT cells is feasible in the presence of a CAR, we transduced the CAR into DP cells arising from WT-TiPS on D35. Delaying the onset of CAR expression in this manner resulted in the development of functional SP cells, including CD8 α β CAR⁺ iT cells (Extended Data Fig. 4a, b). This finding not only established that CD8 α β CAR⁺ iT cells can be generated from TiPS, but also confirmed that the lack of DP formation from CAR-TiPS was probably due to interference with DP commitment arising from the early CAR expression afforded by the constitutive Ubiquitin C promoter in CAR-TiPS.

Regulated CAR expression facilitates CD4⁺CD8 α β ⁺ DP iT cell development.

To restrict CAR expression, we placed it under the transcriptional control of the *TRAC* promoter (*TRAC*-1928z-TiPS, Extended Data Fig. 4c–e). CAR expression from the *TRAC* locus not only resulted in the expected absence of TCR expression throughout differentiation (Fig. 2a, top panel), but also showed the remarkable similarity in temporal cell-surface expression between *TRAC*-CAR and the α β TCR in both WT-TiPS and CAR-TiPS (Fig.

2a). The differentiation of *TRAC*-1928z-TiPS towards early T cell lineage commitment improved, as reflected in a greater CD7⁺CD5⁺ population (Fig. 2b) compared with CAR-TiPS (Extended Data Fig. 3g). DP induction, however, was still not enhanced (Fig. 2b, c and Extended Data Fig. 5a).

We hypothesized that despite its delayed onset of expression, the *TRAC*-encoded 1928z still interfered enough to prevent DP commitment. We therefore sought to attenuate CAR signalling strength by substituting the 1928z with 1928z-1XX, a CAR in which the second and third ITAM have been inactivated²⁷. In peripheral blood mononuclear cells (PBMC)-derived T cells, CAR expression through the endogenous *TRAC* promoter reduced signalling in the absence of antigen exposure²⁸, and the mutation of the second and third ITAMs pre-empted their phosphorylation²⁷. Phosphorylation of ITAM1 and ITAM3 was readily detected in retrovirally expressed CARs (γ RV-1928z) in the absence of antigen, increasingly in T cells with the highest CAR expression (Extended Data Fig. 6a–c). *TRAC*-encoded 1928z showed reduced phosphorylation of ITAM1 and ITAM3, while the latter was abolished in *TRAC*-1XX T cells (Extended Data Fig. 6a–c).

Upon their differentiation, *TRAC*-1XX-TiPS (Extended Data Fig. 4f, g) not only maintained the same heightened propensity to induce CD7 and CD5 expression as *TRAC*-1928z-TiPS, but additionally increased their progression to the DP stage (Fig. 2d, e and Extended Data Fig. 5a). By day 35, these DP cells expressed CD1a, CD2 and CD45RO, consistent with the phenotype of human DP thymocytes^{29–31} (Extended Data Fig. 5b). Intracellular CD3 confirmed their T lineage commitment despite the absence of CD3 and $\alpha\beta$ TCR expression at their cell surface (Extended Data Fig. 5c).

CAR expression affects Notch and TCR downstream target gene induction.

To further support our hypothesis that the induction to the DP stage is controlled by CAR and Notch interactions, we assessed the impact of CAR regulation on Notch target transcript levels during the T lineage commitment phase of our *in vitro* differentiation protocol. We assessed the expression level of several genes that have been reported to be associated with T lymphoid development and $\alpha\beta/\gamma\delta$ lineage commitment, including *NOTCH1*, *NOTCH3*, *ID3*, *TCF7*, *DTX1*, *GATA3* and *PTCRA* (Fig. 3a)^{32–34}. Constitutive CAR expression did indeed grossly perturb the expression pattern observed in WT-TiPS. CAR-TiPS showed an early increase in the expression of the (pre)TCR target *ID3* between D24 and D27 (Fig. 3b), with correspondingly reduced levels of *NOTCH1*, *NOTCH3* and Notch targets *TCF7*, *DTX1* and *PTCRA*. In the *TRAC*-1XX-TiPS, *ID3* expression was decreased while *NOTCH1*, *NOTCH3* and their downstream targets increased to levels nearing those found in WT-TiPS (Fig. 3b). *GATA3* expression, which is a direct target gene of not just Notch but also the pT α ³⁴, was noticeably upregulated in CAR-TiPS relative to WT-TiPS and reduced in *TRAC*-1XX-TiPS (Fig. 3b), suggesting that constitutive CAR expression results in stronger *GATA3* induction compared with the TCR or pT α .

Notably, one of the key Notch targets, *PTCRA*, which encodes the pT α , is repressed in CAR-TiPS, but induced in differentiating WT-TiPS as well as in *TRAC*-1XX-TiPS (Fig. 3b). In thymic development, pT α pairs with the rearranged β -chain to allow for α chain rearrangement and progression to the DP stage¹⁹. To assess whether *PTCRA* induction in

TRAC-1XX-TiPS was associated with pT α protein expression, we measured cell surface-protein levels by flow cytometry and found cell surface pT α expression in the induced-CD4 SP and DP populations (Fig. 3c, d), consistent with its physiological pattern³⁵.

CAR engagement facilitates maturation to CD8 $\alpha\beta$ single-positive iT cells.

Having established that CAR regulation allows for DP cell development in the absence of a TCR, we set out to determine whether the CAR could substitute for the TCR in further driving maturation of DP cells into CD8 $\alpha\beta$ SP iT cells. In the absence of a TCR-dependent positive selection process, we stimulated D35 DP iT cells on cells expressing the CAR target antigen (NIH/3T3 fibroblasts expressing CD19)³⁶. CAR engagement was required for iT cell survival as co-culture with parental fibroblasts lacking CD19 did not yield viable iT cells. Phenotypic analysis on CD19 stimulated *TRAC-1XX-iT* cells after a week (day 42) showed that CD4 and CD1a expression had waned and that a population of CD8 $\alpha\beta$ SP iT cells was induced, consistent with maturation from the DP to the SP stage (Fig. 4a and Extended Data Fig. 7a). Some DN cells and a small population of CD8 $\alpha\alpha$ iT cells still coexisted (Fig. 4b). The matured SP D42 *TRAC-1XX-iT* cells displayed a phenotype resembling activated T cells, including the upregulation of CD25, CD69, CD56 and transition from CD45RO to CD45RA (Extended Data Fig. 7a). However, the cells also downregulated CD5 and expanded poorly (Fig. 4c and Extended Data Fig. 7a).

Analysing the phenotype and induction of activation and co-stimulatory markers upon antigen exposure, we found that 4-1BB was induced transiently 8 h after antigen exposure (Fig. 4d) and hypothesized that engaging the 4-1BB pathway at this stage may promote iT cell expansion. We thus engineered the 3T3-CD19 cells to co-express 4-1BB ligand (3T3-CD19-41BBL). When D35 *TRAC-1XX-iT* cells were stimulated on 3T3-CD19-41BBL, they maintained the ability to form SP iT cells (Fig. 4e, f) and expanded 30-fold (Fig. 4g). To confirm that the resulting SP iT cells at D42 were indeed derived from D35 DP precursors, we sorted the DP cells at D35 and then exposed them to 3T3-CD19-41BBL. After 7 d, the DP cells had lost CD4 expression and matured to CD8 $\alpha\beta$ SP iT cells (Extended Data Fig. 7b). Phenotypically, 3T3-CD19-41BBL-matured cells retained CD5 and CD2 expression, more so than 3T3-CD19-matured cells, and showed higher expression of CD45RO, CD28 and CD56 (Extended Data Fig. 7c).

To determine whether CD19 levels may influence acquisition of an effector-like phenotype of the *TRAC-1XX-iT* cells, we matured iT cells on titrated levels of recombinant CD19 (Extended Data Fig. 7d). Increasing CD19 positively affected the expansion and CD8 $\alpha\beta$ SP iT-cell content but did not reduce the effector-like phenotype.

To determine whether exposure to 4-1BB co-stimulation qualitatively affected *TRAC-1XX-iT* maturation, we compared the function of the 3T3-CD19 and 3T3-CD19-41BBL-matured *TRAC-1XX-iT* cells. *In vitro* cytolytic function (Fig. 4i) and cytokine production (Fig. 4j) were similar between the two groups. However, whereas 3T3-CD19 matured cells failed to expand upon repeated exposure to antigen, 3T3-CD19-41BBL maturation improved their expansion and survival (Fig. 4k). We proceeded to compare these two populations in the NALM6 leukaemia model³⁷ (Fig. 4l), wherein iT cells matured on 3T3-CD19-41BBL showed improved tumour control and survival (Fig. 4m,n), which was associated with

an increased persistence of *TRAC*-1XX-iT cells (Fig. 4o). The cytolytic function of 3T3-CD19–41BBL-matured *TRAC*-1XX-iT cells was also demonstrated *in vitro* to be antigen specific (Extended Data Fig. 7e), and responsive not only to NALM6, but also to primary patient-derived CD19⁺ chronic lymphocytic leukaemia (CLL) cells (Fig. 4p and Extended Data Fig. 7f).

TRAC-1XX-iT cells overall resemble peripheral-blood derived CD8 $\alpha\beta$ T cells.

Having established that T-cell commitment, differentiation, maturation, and expansion of CD8 $\alpha\beta$ CAR T cells can be driven by CAR expression in the absence of a TCR, we sought out to compare the resulting CD8 $\alpha\beta$ *TRAC*-1XX-iT cells (CD8 $\alpha\beta$ iT) to naturally occurring peripheral blood lymphocytes. Analysis of cell-surface markers associated with $\alpha\beta$ TCR-T cells, NK cells or $\gamma\delta$ TCR-T cells, showed that CD8 $\alpha\beta$ iT cells express classical T-cell markers including CD45RO, CD25, CD27, CD28 and low levels of CD62L and CCR7 (Fig. 5a and Extended Data Fig. 8a, b). The stimulated cells expressed the T-cell activation/NK-cell marker CD56 but lacked canonical NK markers such as CD16 and KIR2D (Fig. 5a and Extended Data Fig. 8a, b). They also did not express $\gamma\delta$ TCR-T-cell-associated markers including the $\gamma\delta$ TCR (V δ 2) and CD161 (Fig. 5a and Extended Data Fig. 8a, b). To assess the nature of the cells more closely, we compared the transcriptomic profile of the CD8 $\alpha\beta$ iT cells to that of healthy donor PBMC-derived lymphocytes. To enhance comparability, CD4 $\alpha\beta$ TCR-T (CD4), CD8 $\alpha\beta$ TCR-T (CD8), $\gamma\delta$ TCR-T ($\gamma\delta$ T) and NK cells were engineered to express the 1928z-1XX CAR (either through *TRAC*-targeted integration in CD4 and CD8 or retroviral expression in $\gamma\delta$ T and NK cells) and purified for the CAR⁺ populations. Unsupervised hierarchical clustering analysis based on a dissimilarity matrix showed that, on the basis of overall gene expression, the CD8 $\alpha\beta$ iT cells are most closely related to the CD4 and CD8 T cells (Fig. 5b). Principal component (PC) analysis confirmed that within the first two PCs, CD8 $\alpha\beta$ iT cells cluster more closely with peripheral blood $\alpha\beta$ TCR-T cells than with $\gamma\delta$ TCR-T cells (Extended Data Fig. 8c). Pearson's correlation distinguished that CD8 $\alpha\beta$ iT cells are more closely related to the peripheral blood CD8 T cells ($r = 0.99$, Fig. 5c).

TRAC-1XX-iT cells achieve tumour control in a systemic *in vivo* leukaemia model.

To determine whether the $\alpha\beta$ TCR-T lineage commitment of *TRAC*-1XX-iT cells enhanced their functional capabilities, we compared their *in vitro* and *in vivo* functions to CAR-iT cells and healthy-donor PBMC-derived CD8⁺ *TRAC*-1XX $\alpha\beta$ TCR-T cells (CD8 *TRAC*-1XX) (Extended Data Fig. 9a). *In vitro* cytolytic activity was antigen-specific and similar among the three cell populations in an 18 h cytotoxicity assay (Fig. 6a and Extended Data Fig. 9b). However, Granzyme B and CD107a production was reduced in CAR-iT cells (Extended Data Fig. 9c). Importantly, *TRAC*-1XX-iT and CD8 *TRAC*-1XX cells were able to control repeated *in vitro* exposure to tumour cells, whereas CAR-iT cells failed after a third challenge (Fig. 6b). *In vitro* cytokine secretion was reduced in both iT populations compared with CD8 *TRAC*-1XX T cells. *TRAC*-1XX-iT cells were able to produce IFN γ and TNF α , whereas minimal secretion was detected in CAR-iT cells. Notably, both *TRAC*-1XX-iT and CAR-iT cells lacked the ability to produce IL-2 in response to antigen (Fig. 6c and Extended Data Fig. 9d). To determine whether the differences in *in vitro* function between the *TRAC*-1XX-iT and CAR-iT cells translated into

differences in *in vivo* tumour control, we compared them in the systemic NALM6 leukaemia model. We found that *TRAC-1XX-iT* cells showed improved tumour control (Fig. 6d, e), associated with increased iT cell persistence in the bone marrow, spleen and blood (Fig. 6f, g).

To provide a potency benchmark under these conditions, we administered diminishing doses of CD8 *TRAC-1XX* T cells and found that a single infusion of 2×10^6 *TRAC-1XX-iT* cells provided a survival response that was not significantly different from 4×10^5 healthy donor-derived CD8 *TRAC-1XX* T cells (Extended Data Fig. 9e, f). 4×10^6 cells *TRAC-1XX-iT* cells induced complete and durable responses (Fig. 7a–c). Notably, mice infused with 4×10^6 CD8 *TRAC-1XX* T cells eventually succumbed while presenting GvHD-like symptoms including weight loss, diarrhoea, and loss of fur, probably caused by the remaining small population of TCR⁺ cells¹¹, none of which occurred in *TRAC-1XX-iT* cell recipient mice (Fig. 7c). Enumeration of tumour cells in bone marrow, spleen, and blood at 6 and 12 d after infusion, showed complete absence of detectable tumour in the bone marrow for both treatment groups at either timepoint (Fig. 7d and Extended Data Fig. 10a, b). *TRAC-1XX-iT* cells showed similar persistence to CD8 *TRAC-1XX* T cells in bone marrow, spleen and blood by day 6 and in bone marrow on day 12 but were less abundant in spleen and blood by day 12 (Fig. 7d and Extended Data Fig. 10b). Phenotypically, both populations increased CD45RA expression *in vivo*, and diminished CD62L expression (Fig. 7e and Extended Data Fig. 10c). *TRAC-1XX-iT* cells downregulated CD27 and CD28 but did not display increased exhaustion markers compared with peripheral blood CD8 *TRAC-1XX* T cells.

Discussion

We have described the generation of therapeutic CD8 $\alpha\beta$ CAR iT cells from TiPS. We investigated how premature TCR or CAR expression interferes with adaptive T-cell maturation, and show that delayed expression and calibrated CAR signalling enable DP T-cell development and terminal CD8 $\alpha\beta$ iT cell expansion in the absence of a TCR.

In vitro T cell development from TiPS that constitutively express a CAR yields T cells with an innate-like CD8 $\alpha\alpha$ T cell phenotype^{12, 15} or NK-like features^{16, 26}. We show that premature CAR expression at the DN stage interferes with Notch signalling, skewing differentiation away from DP differentiation and towards the acquisition of an innate-like phenotype. The Notch ligand DLL1 was sufficient to induce DP cell formation during T lineage development from precursor cells that bear TCR VDJ genes in germline configuration, but not from WT-TiPS, which encode a rearranged $\alpha\beta$ TCR and required DLL4 to progress to the DP stage (Fig. 1d). In the presence of constitutive CAR expression however, DLL4 was no longer sufficient to evoke DP differentiation (Fig. 1f). Constitutive CAR expression diminished *NOTCH1* expression and deregulated downstream gene expression, including *DTX1*, *TCF7* and *PTCRA* (Fig. 3b). Consistent with *in vitro* models of lymphopoiesis, stronger TCR signals or interference with Notch signalling impairs the DN to DP transition in $\alpha\beta$ TCR T cells^{38, 39} and defaults the cells towards an innate/ $\gamma\delta$ TCR-like fate.

The combination of regulating CAR expression under the control of the *TRAC* locus²⁸ and inactivating the CAR's second and third ITAM (ref. ²⁷) brought down the potential for constitutive signalling (Extended Data Fig. 6) and rescued the induction of genes downstream of Notch and DP development (Figs. 2d and 3b). The partially restored induction of *PTCRA* in differentiating *TRAC*-1XX-TiPS is noteworthy (Fig. 3b–d), given the crucial role the preTCR-complex plays in the development of $\alpha\beta$ TCR-T cells (but not $\gamma\delta$ TCR-T cells¹⁹). PreTCR expression is required for β chain selection^{19, 40, 41} and its absence diverts T cell differentiation towards a $\gamma\delta$ TCR-T cell phenotype⁴². *PTCRA* expression is absent in the presence of a constitutively expressed CAR (Fig. 3b), consistent with Notch downregulation⁴³. Facilitating DP development through attenuation of CAR signalling is also consistent with the finding that attenuated $\gamma\delta$ TCR signalling can allow for DP maturation in the absence of an $\alpha\beta$ TCR⁴⁴.

TRAC-1XX-TiPS cannot assemble a functional $\alpha\beta$ TCR and therefore depend on the CAR to direct T cell maturation past the DP stage. Exposure to the CAR antigen resulted in the maturation of CD8 $\alpha\beta$ SP by day D42, but failed to support their expansion (Fig. 4a, c). Provision of 4–1BB co-stimulation together with antigen enabled the emerging SP *TRAC*-1XX-iT cells to expand. Upregulation of 4–1BB has been observed in murine DP cells undergoing positive selection *in vivo*⁴⁵. The benefit of providing co-stimulation is consistent with TCR/major histocompatibility complex interaction alone not sufficing to induce complete CD8 T cell maturation^{46, 47}. The emerging SP *TRAC*-1XX-iT cells exposed to 4–1BBL not only expanded but also acquired robust effector functions (Fig. 4i–m) and the ability to expand upon repeated antigen stimulation (Fig. 4k). However, these iT cells do not have a classical naïve phenotype, as they maintain CD5 and CD7 expression but do not homogeneously express CD45RA, CD62L and CCR7, as would be expected in naïve T cells and recent thymic emigrants⁴⁸. They rather express CD45RO, CD28, CD25 and CD56, which are hallmarks of recently activated T cells (Extended Data Fig. 7c). This effector-like phenotype is commonly observed following extrathymic differentiation of T cells, irrespective of CAR expression or maturation protocol^{49, 50}. While the *TRAC*-1XX-iT described here do not display a canonical naïve phenotype, it is noteworthy that they also do not express common exhaustion markers. Transcriptional studies confirmed that CAR-induced maturation produces CD8 $\alpha\beta$ *TRAC*-1XX-iT cells that are more similar to peripheral blood-derived CD8 $\alpha\beta$ $\alpha\beta$ TCR-T cells, than to CD4 $\alpha\beta$ TCR-T cells and yet are more distinct from $\gamma\delta$ TCR-T cells and NK cells (Fig. 5b, c).

When comparing *TRAC*-1XX-iT function to CAR-iT and peripheral blood-derived CD8 *TRAC*-1XX, we found that *TRAC*-1XX-iT had improved cytolytic capacity and cytokine secretion compared with CAR-iT (Fig. 6b,c), as well as improved anti-tumour activity *in vivo* (Fig. 6d,e). *TRAC*-1XX-iT cells still produced significantly lower levels of cytokines than CD8 *TRAC*-1XX cells, notably lacking IL-2 production, which was therefore provided exogenously (Fig. 6c). *TRAC*-1XX-iT cells nonetheless provide substantial anti-tumour activity in a systemic NALM6 model, which CAR-iT cannot achieve (Fig. 6d,e), while requiring a higher dosage than CD8 *TRAC*-1XX T cells (Extended Data Fig. 9e,f). Phenotypically, both *TRAC*-1XX-iT and CD8 *TRAC*-1XX cells differentiated towards an effector phenotype upon encounter with the tumour in the bone marrow. *TRAC*-1XX-iT cells downregulated CD62L, CD27 and CD28, but did not show accelerated acquisition

of exhaustion markers (Fig. 7e and Extended Data Fig. 10c). *TRAC*-1XX-iT cells showed reduced persistence in spleen and blood over time (Extended Data Fig. 10b). Despite these differences, *TRAC*-1XX-iT cells were able to induce long-term remission and survival following intravenous infusion of a single dose of 4×10^6 iT cells (Fig. 7a–c). Tumour control by CAR-iT cells has only been hitherto achieved in intraperitoneal models^{12, 15, 16}.

TiPS are a highly attractive resource for allogeneic, ‘off-the-shelf’ immunotherapy⁸. The self-renewing capacity of TiPS allows for the establishment of gene edited, clonally selected master cell banks^{51, 52} which can be utilized to mass produce genetically homogeneous T cell populations, eliminating donor-dependent T cell variability and thereby standardizing treatment. Combined with genome editing, the use of TiPS allows for careful selection of a desired genotype, screening for insertional mutagenesis⁵³, off-target editing and translocations^{54, 55}, and facilitates the complete elimination of TCR expression to prevent GvHD¹¹ or the accidental transduction of malignant cells in apheresis products⁵⁶. Genotype selection, including detection of off-target editing events, makes TiPS particularly attractive for multiplexed gene editing strategies, such as combining *TRAC* locus editing with the ablation of CD52, CD70, or PD1 (refs. ^{54, 55, 57}). The use of TiPS-derived T cells is not limited to targeting CD19 as described here and is applicable to other target tumour associated antigens, as well as applications beyond cancer immunotherapy⁵⁸. Nonetheless, CARs that produce stronger signalling or engage antigens expressed during T cell development, may require the same careful analysis as described here to avert interference with DP development.

In the present protocol, 1 million *TRAC*-1XX-iT cells are generated from a single TiPS in 42 days (Fig. 4h). Its engineering flexibility and expansion potential provide a process that can be easily scaled to generate clinically relevant iT cell numbers. This may allow for off-the-shelf CAR T cell therapy utilizing uniform and consistent iT cells produced from the same engineered master cell bank. The development of feeder-free systems that support the delicate signalling balance required for DP development and $\alpha\beta$ TCR-T cell lineage commitment and expansion is needed to facilitate large-scale manufacturing. A major advantage of the TiPS approach is to enable safe multiplexed T cell engineering to overcome histocompatibility and tumour microenvironmental barriers^{8, 9}. In summary, we have shown that synthetic receptors like CARs can substitute for the TCR in driving directed T cell differentiation and show that the induction of TCR^{-/-}, $\alpha\beta$ TCR-T cell-like CD8 $\alpha\beta$ CAR iT cells is feasible and holds great potential for the large scale production of potent T-cell-based immunotherapies.

Methods

Cell Lines

OP9-mDLL1.—OP9-mDLL1 cells were obtained from Isabelle Rivière and cultured as previously described¹².

OP9-DLL1, -DLL4, -JAG1, -JAG2.—Parental OP9 cells were obtained from ATCC. Plasmids encoding the Moloney murine leukaemia virus-based SFG γ retroviral vector⁵⁹ were used to clone bi-cistronic constructs to express one of the human Notch ligands (DLL1,

DLL4, JAG1 or JAG2) and green fluorescent protein (GFP). Plasmids containing sequences of hDLL1, hDLL4, hJAG1 and hJAG2 were obtained from GenScript (NM_005618, NM_019074, NM_000214 and NM_002226 respectively) and were cloned using standard molecular biology techniques by replacing the FFLuc element in the SFG-FFLuc-P2A-GFP retroviral vector with the desired Notch ligand. Vesicular stomatitis virus glycoprotein G (VSV-G) pseudotyped retroviral supernatants derived from transduced gpg29 fibroblasts (H29) was used to transduce OP9. Transduced cells were purified by flow cytometry based on Notch ligand and GFP expression. Antibodies used to detect Notch ligand expression were hDLL1 – PE (MHD1–314; BioLegend), hDLL4 – PE (MHD4–46; BioLegend), hJagged-1 – PE (MHJ1–152; BD) and hJagged-2 – PE (MHJ2–523; BioLegend) respectively. Purified OP9 cells were cultured in MEM α (Gibco) media with 20% Fetal Bovine Serum (FBS, Hyclone), 1X MEM Non-Essential Amino Acids (NEAA, Corning), 2mM GlutaMAX (Gibco), 100U/mL Penicillin (Pen) and 100 μ g/mL Streptomycin (Strep, Corning), 55 μ M 2-Mercaptoethanol (2-ME, Gibco) and 50mg/mL ascorbic acid (Sigma) as previously described¹².

NALM6.—NALM6 cells were obtained from ATCC. NALM6 CD19^{-/-} were generated as previously described⁶⁰. NALM6 were transduced to express GFP and firefly Luciferase (FFLuc) for *in vitro* and *in vivo* detection³⁷. Cells were cultured in RPMI 1640 (Corning) with 10% FBS (Hyclone) 1x NEAA, 2mM GlutaMAX, 100U/mL Pen, 100 μ g/mL Strep, 2mM HEPES (Corning) and 55 μ M 2-ME). For Incucyte-based analysis, NALM6 CD19⁺ and NALM6 CD19^{-/-} were transduced with Incucyte NuLight Red lentiviral reagent (NLR, Essen BioScience) and selected with puromycin according to manufacturer's instructions.

3T3-CD19.—NIH 3T3 cells expressing CD19 were used as artificial antigen presenting cells as previously described³⁷. 3T3-CD19–4-1BBL were generated utilizing a previously described SFG γ retroviral vector encoding the 4–1BBL transgene⁵⁹. VSV-G pseudotyped retroviral supernatant derived from transduced H29 was used to transduce 3T3-CD19. Transduced cells were purified by flow cytometry based on 4–1BBL (4–1BBL – PE, 5F4; BioLegend) expression. Cells were cultured in DMEM media (Corning) with 10% Cosmic Calf Serum (Hyclone).

K562-mbIL-21–4-1BBL.—K562 cells were transduced to express membrane-bound IL-21 and 4–1BBL as previously described⁶¹. Cells were cultured in RPMI 1640 (Corning) with 10% FBS (Hyclone), 2mM GlutaMAX, 100U/mL Pen 100 μ g/mL Strep, 55 μ M 2-ME.

H1.—The human embryonic stem cell line was obtained from the Studer Lab and cultured on MEF in hES media (DMEM-F12 (Corning) with 20% knock-out serum replacement (KSR), 1X NEAA, 2mM GlutaMAX, 100U/mL Pen 100 μ g/mL Strep, 55 μ M 2-ME) supplemented with 8ng/mL human basic fibroblast growth factor (hbFGF) (R&D systems).

Primary CLL cells: Apheresis product before CAR T cell infusion were obtained from patients that were consented and enrolled in phase I 1928z CAR T cell clinical trials approved by the MSKCC Institutional Review Board (IRB).

Generation of iPS

FiPS—Fibroblast-derived iPS (FiPS) were generated as previously described⁵¹, transfected with a transient plasmid-based reprogramming system to initiate cellular reprogramming. In brief, cells were transfected with reprogramming vector backbone (pCEP4, Life Technologies) containing *OCT4*, *NANOG*, and *SOX2* under regulation of the EF1 α promoter. Transfected cells were plated on Matrigel and selected with hygromycin until FiPS colonies were established.

WT-TiPS.—WT-TiPS were generated as previously described (Clone T-iPSC-1.10)¹². In brief, healthy-donor peripheral blood mononuclear cells (PBMCs) were activated with phytohemagglutinin (PHA, 2 μ g/mL) and transduced with two tri-cistronic SFG γ retroviral vectors, each vector encoding reprogramming factors and a different fluorescent marker (f-Citrine-P2A-cMYC-E2A-SOX2 and f-vexGFP-P2A-OCT4-T2A-KLF4). Transduced cells were seeded on MEF feeder cells and TiPS colonies were established.

CAR-TiPS.—CAR-TiPS were generated as previously described (1928z-T-iPSC)¹². In brief, clone T-iPSC-1.10 was stably transduced with a bi-cistronic lentiviral vector (mCherry-P2A-1928z) and purified for mCherry expression by flow cytometry.

TRAC^{-/-}-TiPS.—TRAC^{-/-}-TiPS were generated through CRISPR /Cas9-targeted integration of a EF1 α -GFP-P2A-Puromycin-bGHpA(G2AP) expression unit into the *TRAC* locus leading to knockout of the TCRA gene into the WT-TiPS (Clone T-iPSC-1.10). WT-TiPS were electroporated using Lonza Cell Line Nucleofector Kit V solution and Lonza Nucleofector-II. Five million cells were resuspended in 100 μ L nucleofection solution, with 2.6 μ g pBS-*TRAC* gRNA1, 2.6 μ g pBS-TRAC-HR-G2AP and 2.6 μ g hCas9 plasmid, and electroporated using program B-025. Transfected cells were plated on Matrigel-coated plates using TiPS complete medium containing 10 μ M ROCK inhibitor. After 48 h cells were selected with 0.8 μ g/ml Puromycin and sorted by flow cytometry for GFP expression. pBS-TRAC gRNA1 was generated by cloning the *TRAC* gRNA target sequence (5'-CAGGGTTCTGGATATCTGT) into pBS-gRNA MCS plasmid, which contains the human U6 promoter and the gRNA fold described in Mali et al., Science 2013. pBS-TRAC-HR-G2AP was generated by cloning the left (~0.9 kb) and right (1 kb) *TRAC* homology arms (HAs) into pBluescript II SK (+), followed by the insertion of the EF1 α -GFP-P2A-Puromycin-bGHpA expression unit in between of the HAs. hCas9 plasmid⁶² was obtained from Addgene (41815).

TRAC-1928z-TiPS.—TRAC-1928z T cells were generated as previously described^{28, 57}. In brief, $\alpha\beta$ TCR-T cells were purified from PBMCs with the Pan T cell Isolation kit (Miltenyi Biotec) on the AutoMACS Pro according to manufacturer instructions. Purified cells were activated with CD3/CD28 Dynabeads (1:1 beads:cell) in X-Vivo 15 media (Lonza) supplemented with 5% Human Serum (HS) (Gemini Bioproducts) with 5ng/mL rhIL-7 (R&D Systems) and 5ng/mL rhIL-15 (R&D Systems). 48 h after $\alpha\beta$ TCR-T cell activation, CD3/CD28 beads were magnetically removed, and T cells were transfected by electrotransfer of *TRAC* ribonucleoprotein using the Nucleofector II device (Lonza). Then 2 \times 10⁶ T cells were resuspended in P3 buffer (Lonza) and mixed with 60pmol *TRAC*

ribonucleoprotein in a total volume of 20 μ L. Following electroporation and considering 66.7% viability, cells were diluted into culture medium and 1 \times 10⁶/mL and incubated at 37°C, 5% CO₂. Recombinant AAV6 donor vector pAAV-*TRAC*-1928z²⁸ was added to the culture 30min after electroporation at a multiplicity of infection of 3 \times 10⁵ genome copies. Twenty-four h after targeting, T cells were reprogrammed as described above (WT-TiPS) and *TRAC*-1928z-TiPS colonies were established and cloned on MEF feeder cells. PCRs were performed to determine biallelic, specific target transgene integration into the *TRAC* locus.

***TRAC*-1XX-TiPS.**— α β TCR-T cells were isolated, activated and transfected as described above (*TRAC*-1928z-TiPS). Following electroporation cells were transduced with the previously described recombinant AAV6 donor vector pAAV-*TRAC*-1XX containing the 1928z-1XX CAR construct. The 1928z-1XX CAR contains tyrosine-to-phenylalanine point mutations within ITAM2 and 3 of the CD3 ζ domain rendering only ITAM1 functional²⁷ *TRAC*-1XX-T cells were reprogrammed as described above for the FiPS. Emerging iPSCs colonies were expanded and cloned by limiting dilution. PCRs were performed to determine biallelic, specific target transgene integration into the *TRAC* locus.

iPS culture.—iPS lines were maintained on MEF prior to EB-based differentiation, in serum-free hES medium supplemented with 8ng/mL hbFGF. Prior to monolayer-based iCD34 differentiation, iPS lines were cultured on Matrigel in hES media containing 0.4 μ M PD032590, 1 μ M CHIR99021, 5 μ M Thiazovivin, 2 μ M SB431542 (all Biovision), 10 μ M ROCK-inhibitor (Ascent) and 10ng/mL hbFGF (R&D Systems) as previously described⁶³.

Fresh media was provided every day and cells were passaged every 3–4 days as previously described^{12, 51}. iPS lines were tested for mycoplasma contamination every 2 months.

iPSC surface marker expression

iPS lines were assessed for cell surface pluripotency marker expression including SSEA4 – FITC (MC813–70; BD), TRA-1–81 – af647 (TRA-1–81; BD) and CD30 – PE (BerH8; BD).

Verification of transgene integration into the *TRAC* locus

Genomic DNA was isolated using QuickExtract™ DNA Extraction Solution (Lucigen) following manufacturer's protocols. PCRs were performed using KAPA 2X HiFi Hot Start Ready Mix following manufacturer's recommended conditions. PCR products were analysed using ethidium bromide-stained agarose gel electrophoresis and imaged using the Bio-Rad ChemiDoc.

***TRAC*^{-/-}-TiPS:** Successful disruption of the *TRAC* locus through insertion of the G2AP expression unit was verified through PCR of the region spanning between the left- and right homology arms. Primers: 5'-GATGATACGCGTCTCTTCTCCTTTCTCATTGAGC and 5'-TCGAGTAAACGGTAGTGCTG. Non-targeted alleles produce a 1603bp PCR product, targeted alleles a 4434bp product.

***TRAC*-1928z-TiPS and *TRAC*-1XX-TiPS** successful integration of the 1928z and 1928z-1XX CAR construct respectively were assessed as previously described^{27, 28}.

T cell differentiation from iPS and expansion of iT cells.—For the differentiation of iPS to haematopoietic precursors, we used optimized serum- and feeder-free *in vitro* differentiation protocols based on Embryoid Body (EB) formation¹² or monolayer-based (iCD34)⁵¹.

Haematopoietic precursor differentiation.—EB-based precursor differentiation was performed as previously described¹². Undifferentiated TRAC^{-/-}-TiPS or WT-TiPS colonies were transferred to ultra-low attachment plates to allow for EB formation in serum-free differentiation medium (StemPro-34 (Invitrogen), with 2mM GlutaMAX, 1X NEAA, 100U/mL Pen, 100µg/mL Strep, 55µM 2-ME, and 50mg/mL ascorbic acid). Mesoderm induction was facilitated through EB culture with 30ng/mL human bone morphogenetic protein 4 (hBMP-4) and 5ng/mL hbFGF until day 4. Next, haematopoietic specification and expansion was achieved in the presence of 20ng/mL human vascular endothelial growth factor (hVEGF) and a cocktail of haematopoietic cytokines (100ng/mL rhSCF, 20ng/mL rhFlt3L, 20ng/mL rhIL-3 and 5ng/mL hbFGF). Cells were transferred to fresh media with cytokines every 48 h. All cytokines were obtained from R&D systems. Day 10 EBs containing haematopoietic progenitor cells were dissociated with Accutase (StemCell Technologies) prior to culture on OP9 for T lymphoid commitment and expansion.

Monolayer-based iCD34 differentiation was performed as previously described⁶¹. H1, FiPS or TiPS were differentiated to mesoderm and subsequently to CD34⁺ haematopoietic progenitors on matrigel in StemPro34 differentiation media supplemented with a combination of 5ng/mL hBMP-4, 10ng/mL hbFGF, 10ng/mL rhVEGF, 50ng/mL hSCF, 10ng/mL hIL-6, 10ng/mL hIL-11 for 10 days. Media supplemented with fresh cytokines was added every 48 h. All cytokines were obtained from R&D systems. On day 10, CD34⁺ cells were enriched through positive selection with CD34 microbeads (Miltenyi Biotec) on the AutoMACS Pro according to manufacturer instructions, prior to T lymphoid differentiation on OP9.

T lymphoid commitment and expansion.—Day 10 single cells were seeded on OP9 monolayers in OP9 medium (MEMα with 20% FBS), supplemented with 10ng/mL rhTPO, 5ng/mL rhIL-3, 30ng/mL rhSCF, 10ng/mL rhIL-7 and 10ng/mL rhFlt3L to initiate lymphoid lineage commitment until day 20, and with 30ng/mL rhSCF, 10ng/mL rhIL-7 and 10ng/mL rhFlt3L to complete T lineage commitment until day 35. Differentiating T lymphoid cells were passaged onto fresh OP9 monolayers every four days, fresh media with cytokines was supplemented 48 h after each passage. For the stimulation and expansion of TRAC^{-1XX}-iT and CAR-iT cells, differentiated cells were harvested from OP9 monolayers on day 35 and seeded on a monolayer of irradiated 3T3-CD19±4-1BBL at a 3:1 E:T ratio in T cell expansion media (CTS Optimizer Media (Gibco) with 1x CTS T cell maturation/expansion supplement and 1x CTS Immune cell serum replacement, 5ng/mL rhIL-7 and 25ng/mL rhIL-21). Cells were fed with fresh expansion media every 48 h. For maturation of TRAC^{-1XX}-iT cells on CD19 recombinant protein, flat-bottom tissue culture plates were coated with recombinant human CD19-Fc chimeric protein (R&D Systems), in 100mM sodium-bicarbonate coating buffer, overnight at 4°C. Plate were blocked with PBS + 5% FBS for 30min at room temperature and washed twice with PBS. iT cells were resuspended

at 0.25×10^6 cells/mL in T cell expansion media with 5ng/mL rhIL-7, 25ng/mL rhIL-21 and 3 μ g/mL Urelumab (Creative Biolabs). Cells were passaged after 48 h and fresh media supplemented with cytokines was added every 2 days.

PBMC derived cell isolation, activation, culture, and transduction.—Buffy coats from healthy volunteer donors were obtained from the New York Blood Center. PBMCs were isolated by density gradient centrifugation.

$\alpha\beta$ TCR-T cells.— $\alpha\beta$ TCR-T cells were purified and engineered as described above (*TRAC*-1928z-TiPS). After AAV6 transduction with pAAV-*TRAC*-1XX, cells were cultured in media supplemented with 5ng/mL IL-7 and 5ng/mL IL-15 for 3–5 days. After expansion, CD4 and CD8 cells were purified using EasySep™ CD4⁺ or CD8⁺ negative selection T cell isolation kit (Stem Cell Technologies) and purified for CAR expression by flow cytometry.

$\gamma\delta$ TCR-T cells.—PBMCs were resuspended in lymphocyte media (RPMI 1640 media with 10% FBS) supplemented with 1 μ g/mL Zoledronic Acid (Stem Cell Technologies) 10ng/mL rhIL-15 (R&D systems) and 100U/mL IL-2 (Proleukin). After 72 h cells were fed with additional media and cytokines, after 6 days $\gamma\delta$ TCR-T cells were purified with the Miltenyi Biotec TCR γ/δ^+ T cell isolation kit. Purified $\gamma\delta$ TCR-T cells were transduced with SFG γ -1928z-1XX-P2A-LNGFR to induce CAR expression. Cells were expanded for 5–7 days in media supplemented with cytokines and purified for expression of the transduction reporter low-affinity nerve growth factor reporter (LNGFR) using magnetic isolation with LNGFR – PE (C401457; BD) and anti-PE microbeads (Miltenyi Biotec).

NK cells.—NK cells were purified from PBMCs with the NK Cell Isolation Kit (Miltenyi Biotec). NK cells were resuspended in lymphocyte medium and activated with K562-mbIL21–41BBL at a 1:1 E:T, supplemented with 1000U/mL IL-2. 48 h after activation NK cells were transduced with SFG γ -1928z-1XX-P2A-LNGFR to induce CAR expression. Cells were expanded for five days in media supplemented with cytokines and purified for LNGFR expression using magnetic isolation with LNGFR – PE (C401457; BD) and anti-PE microbeads (Miltenyi Biotec) on the AutoMACS Pro.

Retroviral vector constructs, retroviral production, and transduction.

Plasmids encoding the SFG γ -retroviral vector⁵⁹ were prepared as previously described^{36, 64}. VSV-G pseudotyped retroviral supernatants derived from transduced H29 were used to construct stable retroviral-producing cell lines as previously described⁶⁵. T and NK cells were transduced by centrifugation on Retronectin (Takara)-coated plates.

iT transduction.—Day 35 WT-TiPS iT cells were harvested and transduced by centrifugation on Retronectin-coated plates in the presence of 100U/mL IL-2. Cells were fed every 48 h with fresh lymphocyte media and cytokines.

Flow cytometry

Cell surface proteins.—The following conjugated antibodies were used to monitor T lymphocyte lineage development during differentiation. CD45 – BV605 (HI30; BioLegend),

CD3 – BUV737 (UCHT1; BD), TCR $\alpha\beta$ – PE-Cy7 (IP16; Invitrogen), CD4 – BV785 (SK3; BioLegend), CD8 α – BUV395 (HIT8a; BD), CD8 β – PE (SIDI8BEE; Invitrogen), CD8 $\alpha\beta$ – APC (2ST8.5H7; BD), CD7 – APC-H7 (M-T701; BD), CD5 – PerCP-Cy5.5 (UCHT2; BioLegend), CD56 – BV421 (HCD56; BioLegend), CD1a – PE-Cy7 (HI149; BioLegend), CD2 – BV711 (RPA-2.10; BD). The TCR-T and NK cell phenotypes were determined with CD45RA – BV605 (HI100; BioLegend), CD45RO – BV421 (UCHL1; BioLegend), CD62L – BV711 (DREG-56; BioLegend), CCR7 – PE-Cy7 (G043H7; BioLegend), CD25 – BB515 (2A3; BD), CD69 – PerCP-Cy5.5 (FN50; BioLegend), CD27 – BUV737 (M-T271; BD), CD28 – PE-Cy7 (CD28.2; BioLegend), CD16 – BUV737 (3G8; BD), NKG2C – PE (S19005E; BioLegend), KIR2D – FITC (NKVFS1; Miltenyi Biotec), NKp46 – FITC (9E2; BioLegend), NKp30 – PerCP-Cy5.5 (P30-15; BioLegend), TCRV $\delta 2$ – PerCP-Cy5.5 (B6; BioLegend) and CD161 – BV421 (HP-3G10; BioLegend), CCR2 – PE (K036C2; BioLegend) in addition to the aforementioned T lymphocyte lineage commitment markers. 4-1BB induction was measured with 4-1BB – BV605 (4B4-1; BioLegend). CAR expression was measured with biotin-conjugated goat anti-mouse F(ab')₂ antibody (GaM-biotin; Jackson ImmunoResearch), followed by a blocking incubation with 2% mouse serum (MP Biomedicals) and streptavidin-PE (BioLegend) or streptavidin-APC (BioLegend).

pT α stain.—Cells were incubated with anti-pT α antibody (2F1; BD), biotin-labelled anti-mouse IgG1 (RMG1-1; BioLegend) and Streptavidin-PE (BD). Staining for additional cell surface proteins was performed after completion of pT α staining.

Intracellular phospho-protein analyses.—SFG γ -1928z, *TRAC-1928z* and *TRAC-1XX* T cells were fixed with Phosflow Fix Buffer I (BD) and stained for the CAR with GaM-af647 (Jackson ImmunoResearch), followed by 2% mouse serum, and subsequently permeabilized with Phosflow Perm Buffer III (BD) following the manufacturer's procedure. Permeabilized samples were stained with antibodies detecting phosphorylated CD3 ζ ITAM1 – af488 (EP776(2)Y; Abcam) or phosphorylated CD3 ζ ITAM3 – PE (K25-407.69; BD).

All antibodies were titrated prior to use. Flow cytometric data were acquired on Fortessa X-20 (BD) or 5-laser Aurora (Cytek Biosciences) Flow cytometer voltages were calibrated with Ultra Rainbow Calibration Kit (SpheroTech, URCP-38-2K) prior to every acquisition. Analysis was performed using FCS Express 7 (De Novo Software). Negative and positive gates were set based on (un)stained PBMC and TiPS controls (Supplementary Fig. 1).

Apoptosis Analysis.—WT-TiPS and CAR-TiPS cells were harvested daily between D27 and D35 of the differentiation and stained for viability and Annexin-V – APC (Invitrogen) according to manufacturer's instructions, followed by cell surface staining for CD45, CD7, CD4, CD8 α and CD8 β as described above. Percentage of apoptotic cells within populations (CD45⁺CD7⁺, DN, DP, CD4, CD8 $\alpha\alpha$ or CD8 $\alpha\beta$) was calculated based on live Annexin-V⁺ stain.

Notch induction in differentiating TiPS cells.—WT-TiPS D20 lymphoid progenitor cells were co-cultured with parental OP9, OP9-hDLL1, OP9-hDLL4, OP9-hJAG1 or OP9-hJAG2. At 0, 4, 8, 12, 24, 48 and 72 h of co-culture, cells were harvested, cell pellets were

snap-frozen and stored at -80°C for until RNA extraction. Gene induction was measured by ddPCR as described below. Relative level of *DTX1* induction was normalized to 0h.

Notch/TCR target gene induction during iT differentiation.—TiPS (WT-TiPS, CAR-TiPS and *TRAC-1XX*-TiPS) were differentiated as described. During the T lymphoid commitment phase of the differentiation (D24, D27, D31 and D35) suspension cells were harvested, cell pellets were snap-frozen and stored at -80°C until RNA extraction. Gene induction was measured by ddPCR as described below.

dd PCR.—ddPCR gene expression assays for *Notch1* (dHsaCPE5050282), *Notch3* (dHsaCPE5046836), *TCF7* (dHsaCPE5031804), *DTX1* (dHsaCPE5192773), *GATA3* (dHsaCPE5034292), *ID3* (dHsaCPE5027720), *PTCRA* (dHsaCPE5031466) and *RPL13A* (dHsaCPE5037592) were obtained from Bio-Rad. ddPCR reactions were set up according to One-Step RT-ddPCR Advanced Kit for Probes protocol on a QX200 ddPCR system (Bio-Rad). Each sample was evaluated in technical triplicates. Reactions were partitioned into a median of $\sim 15,000$ droplets per well using the QX200 droplet generator. Emulsified reactions were amplified on a 96-well thermal cycler. Plates were read and analysed with the QuantaSoft software to assess the number of droplets positive for the target gene. The number of messenger RNA (mRNA) molecules per droplet relative to *RPL13A* was calculated assuming a Poisson distribution, as prescribed by Bio-Rad.

RNA extraction, library generation and sequencing.—Total RNA was isolated from $0.3 \times 10^6 - 0.5 \times 10^6$ cells using the RNeasy 96 Kit (Qiagen, 74181) according to the manufacturer's protocol. RNA quality was measured by High Sensitivity RNA ScreenTape (Agilent) on the Agilent 42000 TapeStation System. RNA quantity was measured using the Thermo Scientific™ Qubit™ Flex Fluorometer (Invitrogen). 200ng of total RNA was used per sample to generate mRNA library using NEBNext®Ultra™ II Directional RNA Library Kit for Illumina® (New England BioLabs) per sample. Final libraries were quantified using the Qubit™ 1X dsDNA high sensitivity Assay kit on the Qubit™ Flex Fluorometer. Library quality and size were measured using Agilent High Sensitivity D1000 ScreenTape. Libraries were calculated to nM, diluted to 4nM, and pooled evenly for high throughput sequencing. Sequencing was performed on Illumina NextSeq 500 Instrument (Illumina) with 2×76 pair-end reads targeting a minimum of 16 million pair-end reads per sample.

RNA sequencing analysis.—Sequencing data were trimmed using Trim Galore! 0.6.0 to remove Illumina adapters. Resulting reads were mapped to the human reference genome (assembly GRCh38.86) using Salmon v0.13.1 in quasi-mapping-based mode, with GC bias correction, selective alignment, and range factorization. The data was analysed using the statistical software R. The aggregated read counts were normalized for sequencing depth and RNA composition with DESeq2. Pseudogenes identified by the GENCODE project and lowly expressed genes were filtered out prior to downstream analysis. Principal Component Analysis (PCA) was performed with normalized read counts in R. Hierarchical Clustering Analysis was carried out with the unweighted pair group method with arithmetic mean (UPGMA) method on Euclidean distance matrix. Correlation matrix was generated using Pearson's statistics.

Cytotoxicity Assays.—The *in vitro* toxicity of TRAC-1XX T cells was determined by a standard firefly luciferase (FFLuc)-based assay⁶⁰ or by NLR⁺ imaging on the Incucyte Live Cell Analysis System (Sartorius). For FFLuc based cytotoxicity, FFLuc-expressing NALM6 served as target cells. The effector (E) and tumour target (T) cells were co-cultured in triplicates at the indicated E:T ratio using black-walled 96-well plates with 1×10^5 target cells in a total volume of 100 μ L per well in T cell expansion medium. Four hours later, 50 μ L luciferase substrate (Bright-Glo, Promega) was directly added to each well. Emitted light (RLU) was detected in a luminescence plate reader (Agilent BioTek S1L), and lysis was calculated using the formula $100 \times (1 - (RLU_{\text{sample}}) / (RLU_{\text{target alone}}))$. For the Incucyte cytotoxicity assay, flat-bottom 96-well plates were pre-coated with 5 μ g/mL Fibronectin (Sigma) at 4°C overnight. The E:T ratios were plated in with 3×10^4 target cells in a total volume of 200 μ L per well in T cell expansion medium, in technical triplicates. Hourly brightfield and fluorescence imaging was performed for a 72 h period. Cell survival was quantified based on NLR⁺ surface area by Incucyte S3 software (Essen BioScience) and normalized to the NLR⁺ surface area at 0 h. For the flow-cytometry based cytotoxic T lymphocyte (CTL) assay, CD19⁺ cells were purified from apheresis product (EasySep CD19 Positive selection kit, StemCell Technologies) and cultured overnight in RPMI media with 10% human serum, 1x NEAA, 2mM GlutaMAX, 100U/mL Pen, 100 μ g/mL Strep. TRAC-1XX-iT cells were counted and plated in triplicate at the indicated E:T ratios with 1×10^5 CD19⁺ CLL target cells in a total volume of 100 μ L per well in T cell expansion media. Six hours later, cells were stained with CD19 – PE-Cy7 (SJ25C1, BioLegend), CD45 – BV605 (HI30, BioLegend), CD7 – APC-H7 and Sytox Blue Dead Cell Stain (Invitrogen) and the number of remaining, target cells (live, CD7⁻CD19⁺ cells) were enumerated by flow using AccuCount beads (Spherotech). Percentage lysis was calculated using the formula $(\text{sample count} \times 100) / (\text{target alone count})$.

Antigen restimulation assay.—Restimulation assays were performed as previously described³⁷. In brief, 1×10^6 T cells were co-cultured with 3×10^5 3T3-CD19 in 1 mL T cell expansion media. Fresh media was supplied every 48 h. Cells were counted after seven days and restimulated on fresh 3T3-CD19 monolayers.

***In vitro* NALM6 rechallenge assay.**— 3×10^4 iT cells were co-cultured with 3×10^4 NLR⁺ NALM6 CD19⁺ tumour cells in 200 μ L T cell expansion media. Hourly brightfield and fluorescence imaging was performed for a 10 day period. At day 3 and day 6, plates were removed from the Incucyte, and 50 μ L media was replaced with 50 μ L media supplemented with 3×10^4 fresh NLR⁺ NALM6 cells and 4x cytokines. Cell survival was quantified based on NLR⁺ surface area by Incucyte S3 software (Essen BioScience) and normalized to the NLR⁺ surface area at 0, 72 and 144 h respectively.

Cytokine analyses.—To measure intracellular levels, iT cells were cultured for 4 h at 1×10^6 cells/mL together with NALM6 at a 1:1 ratio in the presence of Brefeldin A (BD) monensin (BioLegend) and CD107a – BV421 (H4A3; BD). Cells were stained with ef506 Fixable Viability dye (ThermoFisher) prior to fixation and permeabilization using BD Cytofix/Cytoperm Plus kit as per manufacturer's instructions, followed by staining with anti-cytokine and cell-surface antibodies GranzymeB – APC (GB12; Invitrogen), IFN γ –

PE-Cy7 (4S.B3; Invitrogen), IL-2 – BUV737 (MQ1–17H12; BD), TNF α – PE (Mab11; Invitrogen), IL-17 – af488 (BL168, BioLegend), CD45 – BV605 (HI30; BioLegend). Percentage of cytokine producing cells was determined by flow cytometry.

To measure secreted cytokine levels, 0.5×10^6 T cells were cultured together with NALM6 at a 1:1 ratio or without target cells for 24 h. Supernatants were collected and stored at -80°C . Secreted cytokines were quantified using BD Cytometric Bead Array kits (IL-2 – 558270, IFN γ – 560111, TNF α – 560112) and flow cytometry.

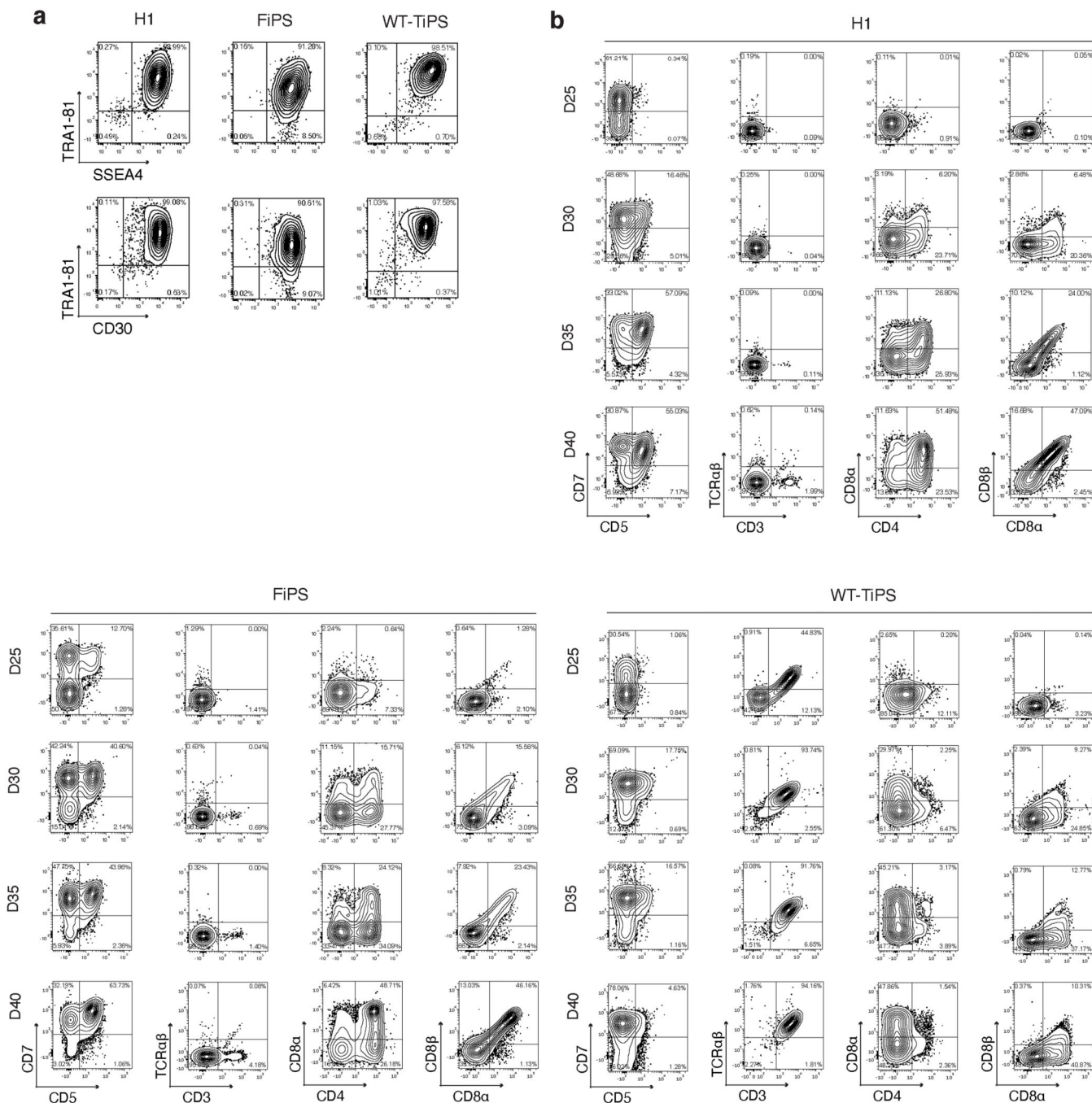
ERK1/2 phosphorylation analysis.—Phosphorylated-ERK1/2 was quantified in day 35 WT-TiPS and CAR-TiPS. Cells were lysed in 1x denaturation buffer supplemented with $10\mu\text{g/mL}$ aprotinin, leupeptin and pepstatin at 1mg/mL total protein content. Phosphorylated ERK1/2 was quantified using the BD Cell Signaling Master Buffer Kit (560005) and Phospho ERK1/2 (560012) according to manufacturer's instructions.

Mouse systemic tumour model.—6–8 week-old NOD/SCID/IL-2R γ -null (NSG) mice were obtained from Jackson Laboratory. A dose of 0.1×10^6 FFLuc-NALM6 was administered by tail vein injection and three days later the indicated dose of T cells were administered by tail vein injection per mouse. Mice received IL-2 (Proleukin 100KU/mouse) and rhIL-15 (150ng/mouse) in $200\mu\text{L}$ PBS intraperitoneally twice per week for three weeks post T cell injection. Tumour burden was measured by bioluminescence imaging using the Xenogen IVIS Imaging System (Xenogen). Living Image software (Xenogen) was used to analyse the acquired bioluminescence data. No blinding method was used. All animal experiments were conducted in accordance with protocols approved by MSKCC Institutional Animal Care and Use Committee (IACUC) and following National Institutes of Health (NIH) guidelines for animal welfare.

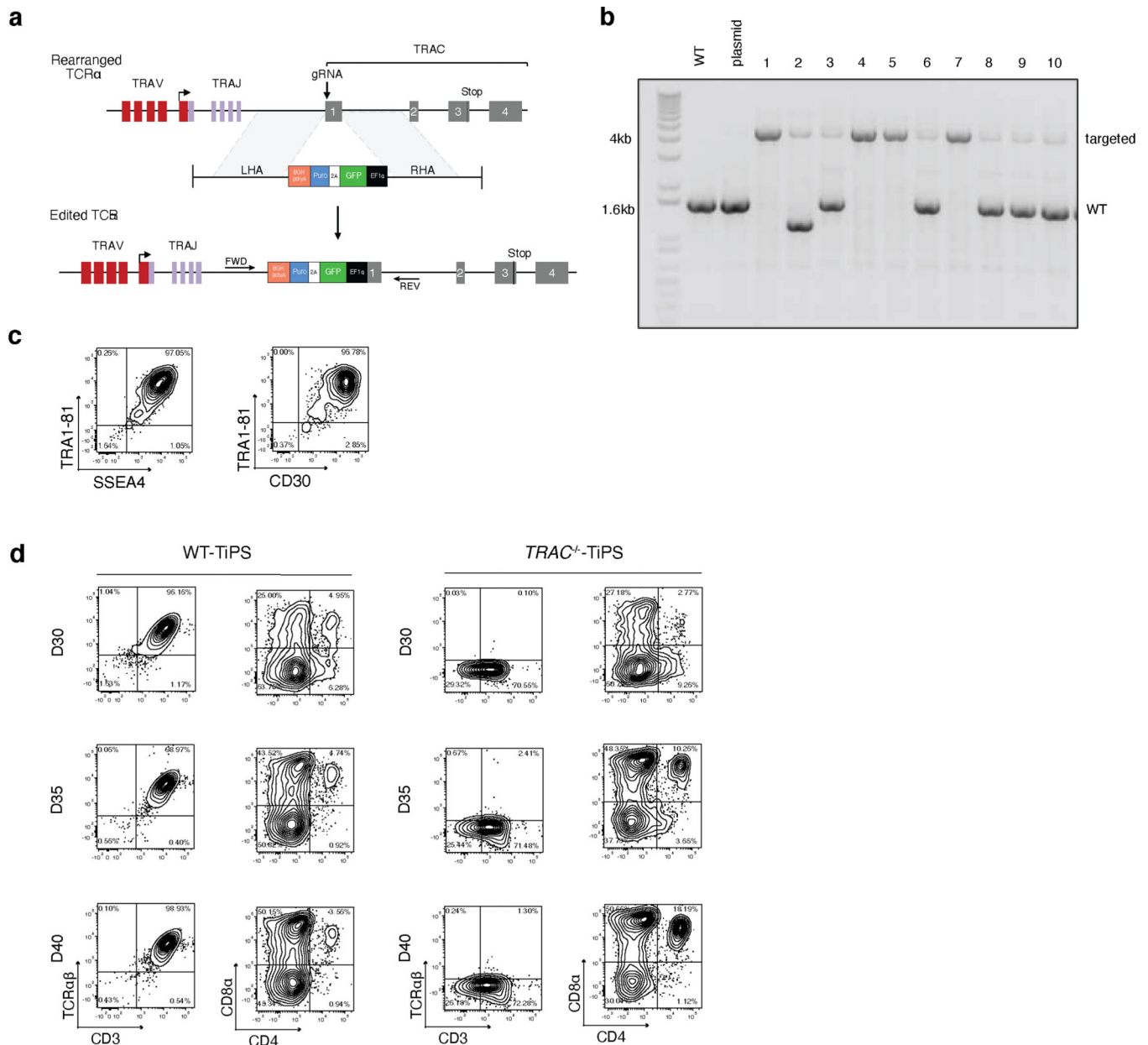
Cell enumeration.—Three mice per group were euthanised at day 6 or day 12 post T cell injection and cells were isolated from the blood, bone marrow and spleen as described³⁷. Cells were stained for viability (ef506), CD45, CAR, CD4, CD8 α , CD8 β , CD45RA, CD45RO, CD27, CD28, CD25, CD62L, CD56, CD19 (all as described above), mCD45 – BV421 (30-F11; BioLegend), TIGIT – BV605 (A15153G; BioLegend), LAG3 – PE-Cy7 (11C3c65; BioLegend), PD1 – BV711 (EH12.2H7; BioLegend) and GFP (tumour cells) and analysed by flow cytometry in the presence of counting beads (Countbright, Invitrogen).

Statistics.—All experimental data are presented as mean \pm S.D. No statistical methods were used to predetermine sample size. Appropriate statistical tests were used to analyse data, as described in the figure legends. Statistical analysis was performed on GraphPad Prism 7 software and R. Significance was set at $p < 0.05$.

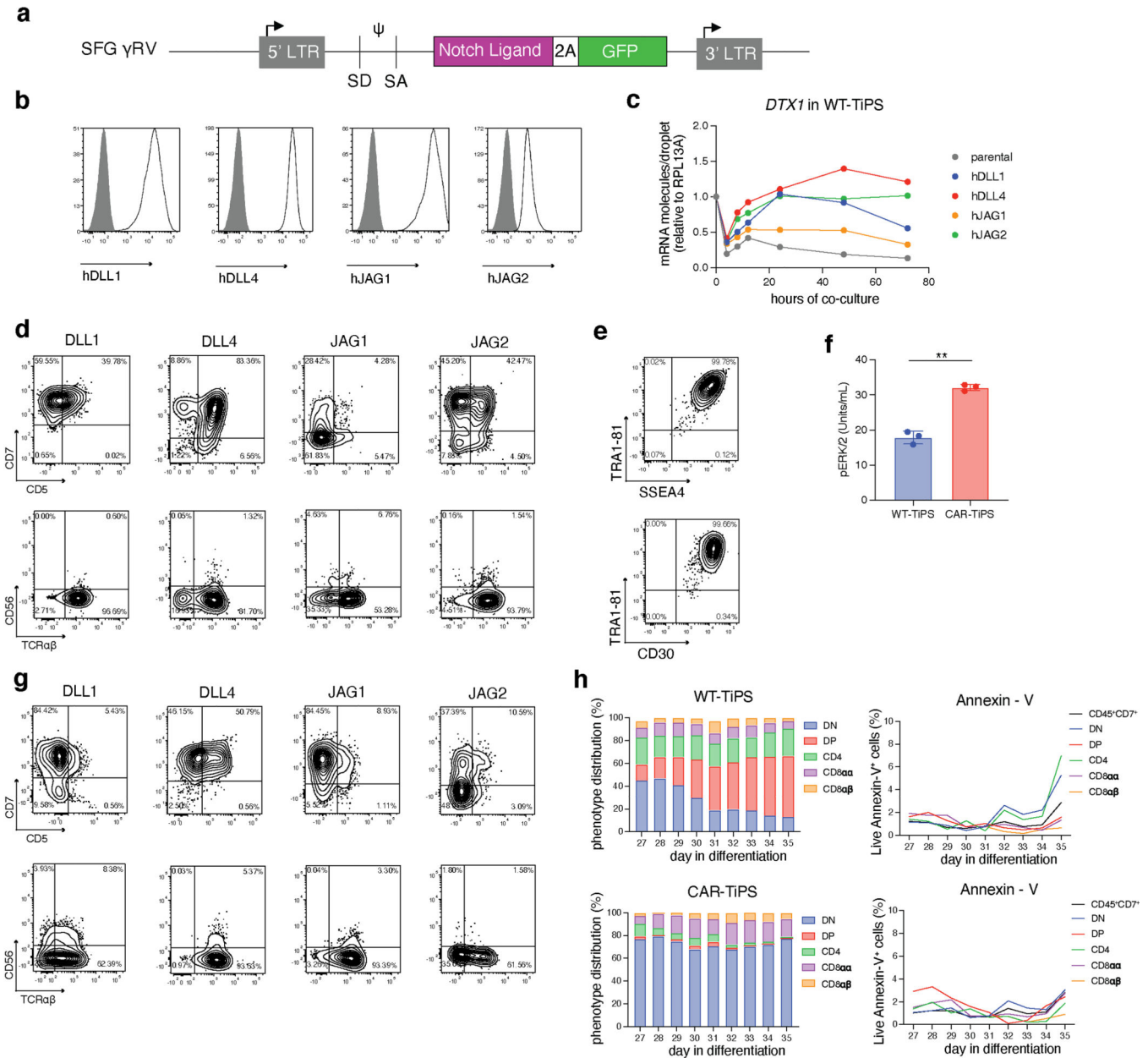
Extended Data



Extended Data Fig. 1. T lymphoid commitment of hES, FiPS and TiPS on OP9-mDLL1
a, Flow cytometric analysis of pluripotency marker expression on H1, FiPS and WT-TiPS
b, Flow cytometric analysis of T lymphoid markers of H1, FiPS and WT-TiPS during differentiation on OP9-mDLL1 at indicated timepoints. Plots depicting CD7/CD5 are gated on live CD45⁺ cells, plots depicting CD3/TCRαβ, CD4/CD8α and CD8α/CD8β are gated on live CD45⁺CD7⁺ cells. CD3/TCRαβ and CD4/CD8α at D40 are as presented in Fig. 1b.



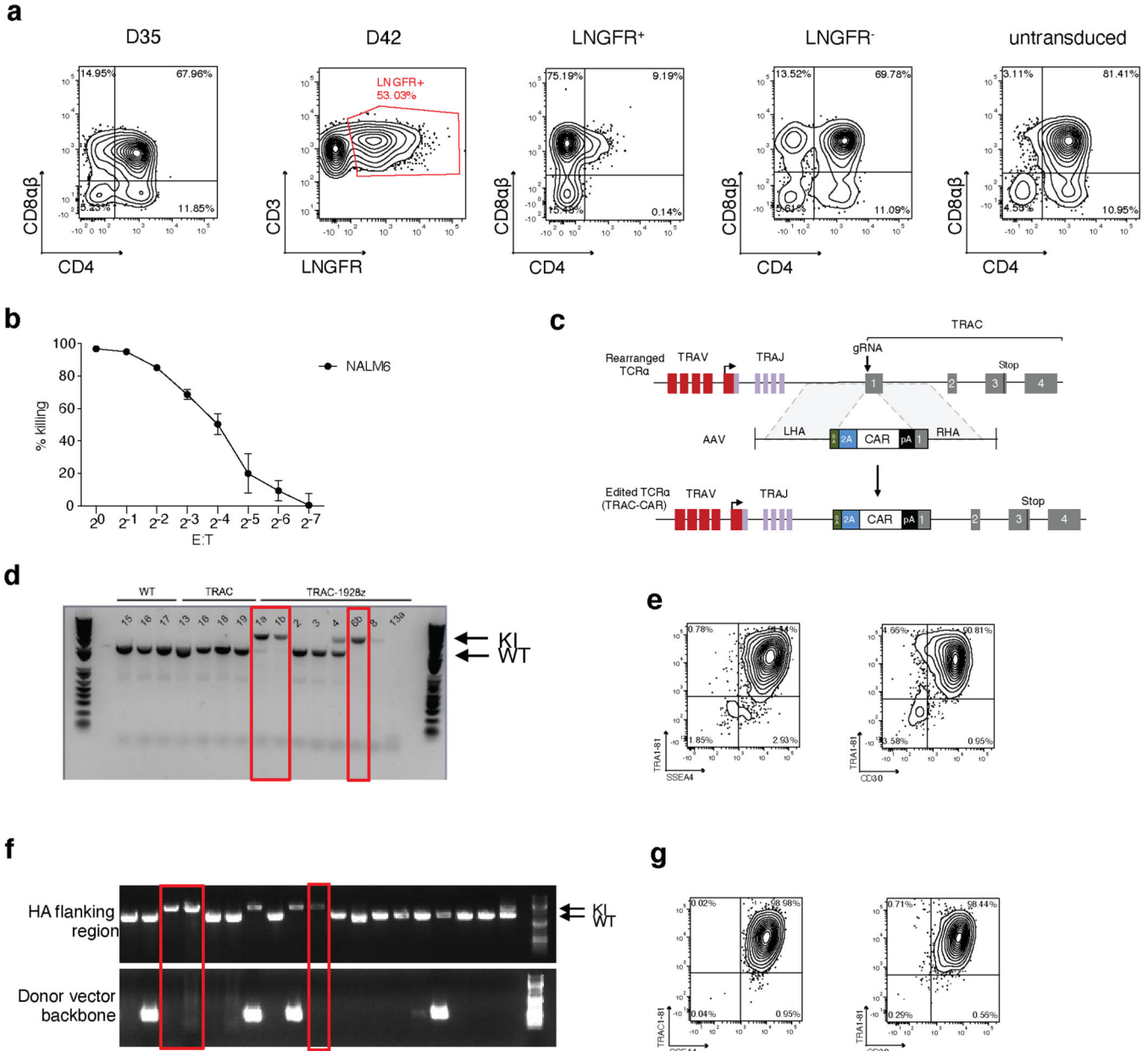
Extended Data Fig. 2. Generation, validation, and differentiation of *TRAC*^{-/-}-TiPS
a, CRISPR/Cas9-targeted integration of EF1a-GFP-P2A-Puromycin-bGHpA (G2AP) expression unit into the *TRAC* locus. Top, *TRAC* locus; middle, plasmid containing the G2AP expression unit flanked by homology arms; bottom, edited *TRAC* locus. ‘FWD’ and ‘REV’ indicate the location of the forward and reverse primers used in **b**. **b**, PCR validation of G2AP integration into the *TRAC* locus of TiPS clones. **c**, Flow cytometric analysis of pluripotency marker expression on *TRAC*^{-/-}-TiPS. Gated on live cells. **d**, T lymphoid makers of WT-TiPS and *TRAC*^{-/-}-TiPS during differentiation on OP9-mDLL1 at the indicated timepoints. Gated on live CD45⁺ cells. D40 is as presented in Fig. 1c.



Extended Data Fig. 3. Early T lymphoid commitment of WT-TiPS and CAR-TiPS on human Notch ligands

a. SFG γ RV plasmid design to transduce human Notch ligands (DLL1, DLL4, JAG1 or JAG2) into parental OP9 cells. **b.** Notch ligand expression on engineered OP9 lines. Filled grey histogram are stained parental OP9 cells, open black histogram are transduced OP9 cells. **c.** *DTX1* induction in WT-TiPS by OP9 expressing indicated Notch ligand. D20 differentiating WT-TiPS cells were co-cultured with indicated OP9. *DTX1* induction was measured by ddPCR, relative to endogenous *RPL13A*. The fold change was calculated relative to 0 h. Data shown is average of $n = 2$ technical replicates. **d, g.** Flow cytometric analysis of T lymphoid commitment marker expression (CD7, CD5, TCR $\alpha\beta$ and CD56) of WT-TiPS (**d**) and CAR-TiPS (**g**) differentiated on OP9 expressing indicated human Notch

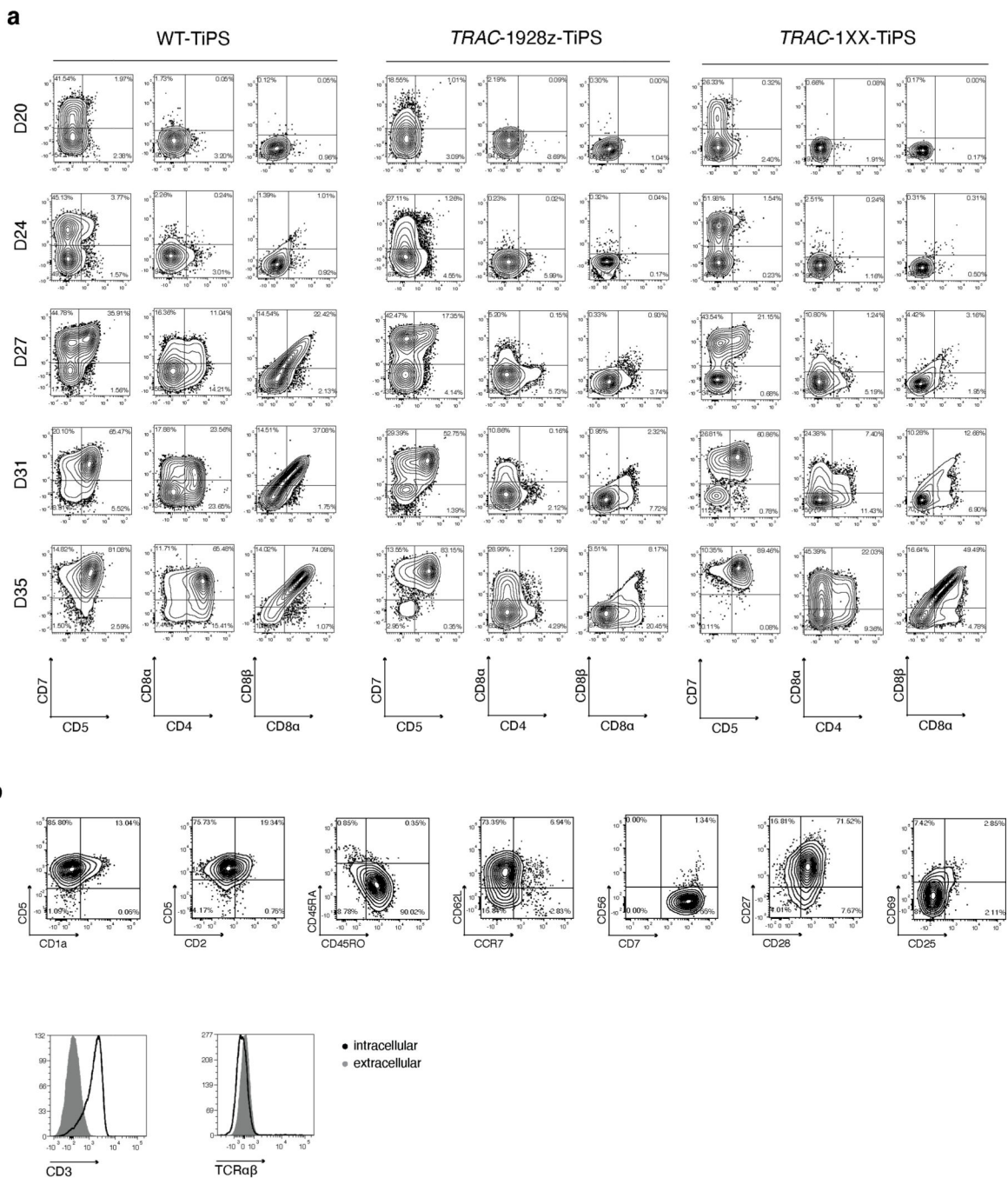
ligands. Gated on live CD45⁺ cells. **e**, Flow cytometric analysis of pluripotency marker expression on CAR-TiPS. Gated on live cells. **f**, Phosphorylated-ERK1/2 levels in WT-TiPS (blue) and CAR-TiPS (red) on D35 (n = 3 technical replicates). **h**, Phenotype (left panels) and apoptosis levels (right panels) of WT-TiPS (top) and CAR-TiPS (bottom) from D27 – D35 of differentiation on OP9-DLL4. Percentage of apoptotic cells in each T lineage developmental stage was based on percentage live Annexin-V⁺ cells. * P<0.05, ** P<0.001, *** P<0.001, Welch’s 2-sample t test, data are means ± s.d (**f**).



Extended Data Fig. 4. CD8αβ single positive CAR⁺ iT cell development

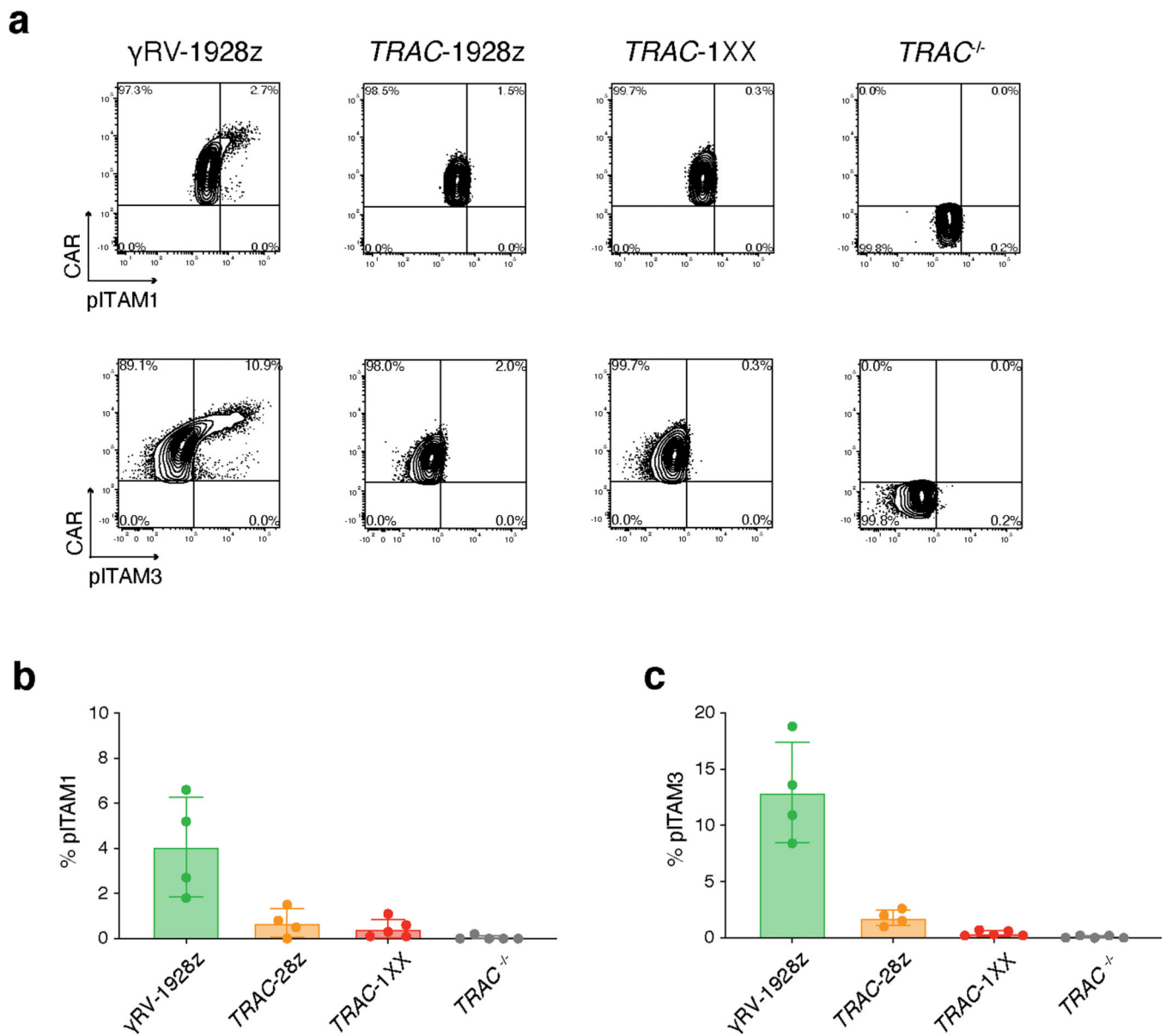
WT-TiPS were differentiated on OP9-DLL4 and transduced to express the 1928z CAR at D35 utilizing γRV SFG-1928z-P2A-LNGFR. Cells were expanded for 7 days in expansion

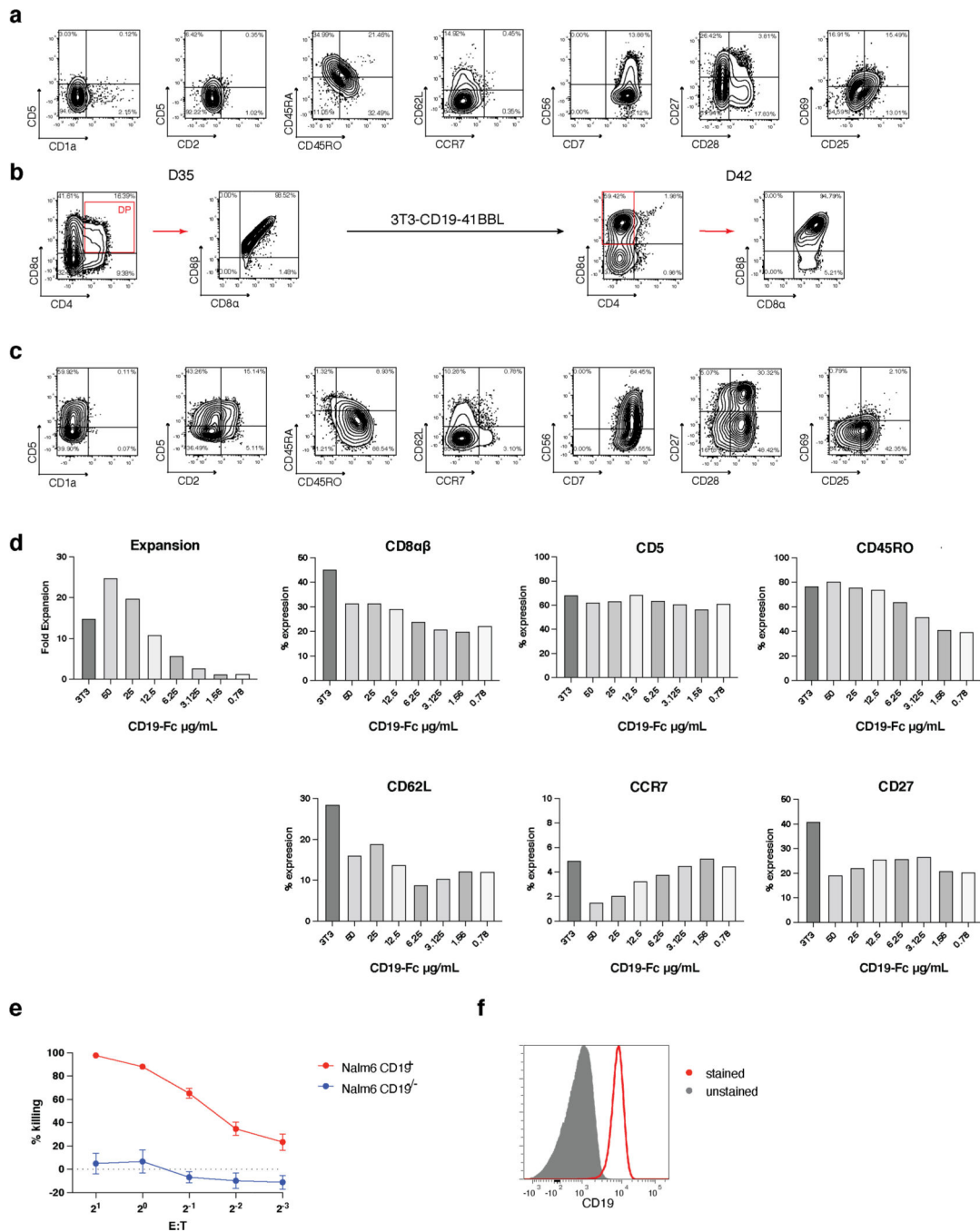
media supplemented with IL-2. **a**, CD4/CD8 $\alpha\beta$ expression prior to transduction (D35) and on D42 in LNGFR⁺ cells, LNGFR⁻ cells and untransduced control cells which remained in differentiation on OP9-DLL4. Gated on live CD45⁺ cells. **b**, Cytotoxic activity of CAR⁺ iT cells in a 18 h bioluminescence assay, using FFLuc- NALM6 as target cells (n = 3 technical replicates, data are mean \pm s.d). **c**, CRISPR/Cas9-targeted integration of CAR transgene into the *TRAC* locus. Top, *TRAC* locus; middle, plasmid containing the CAR transgene cassette flanked by homology arms; bottom, edited *TRAC* locus. **d, f**, PCR validation of CAR integration into the *TRAC* locus of *TRAC*-1928z-TiPS (**d**) and *TRAC*-1XX-TiPS (**f**) clones. **e, g**, Pluripotency marker expression on *TRAC*-1928z-TiPS (**e**) and *TRAC*-1XX-TiPS (**g**), gated on live cells.



Extended Data Fig. 5. T lineage commitment of TRAC-CAR-TiPS

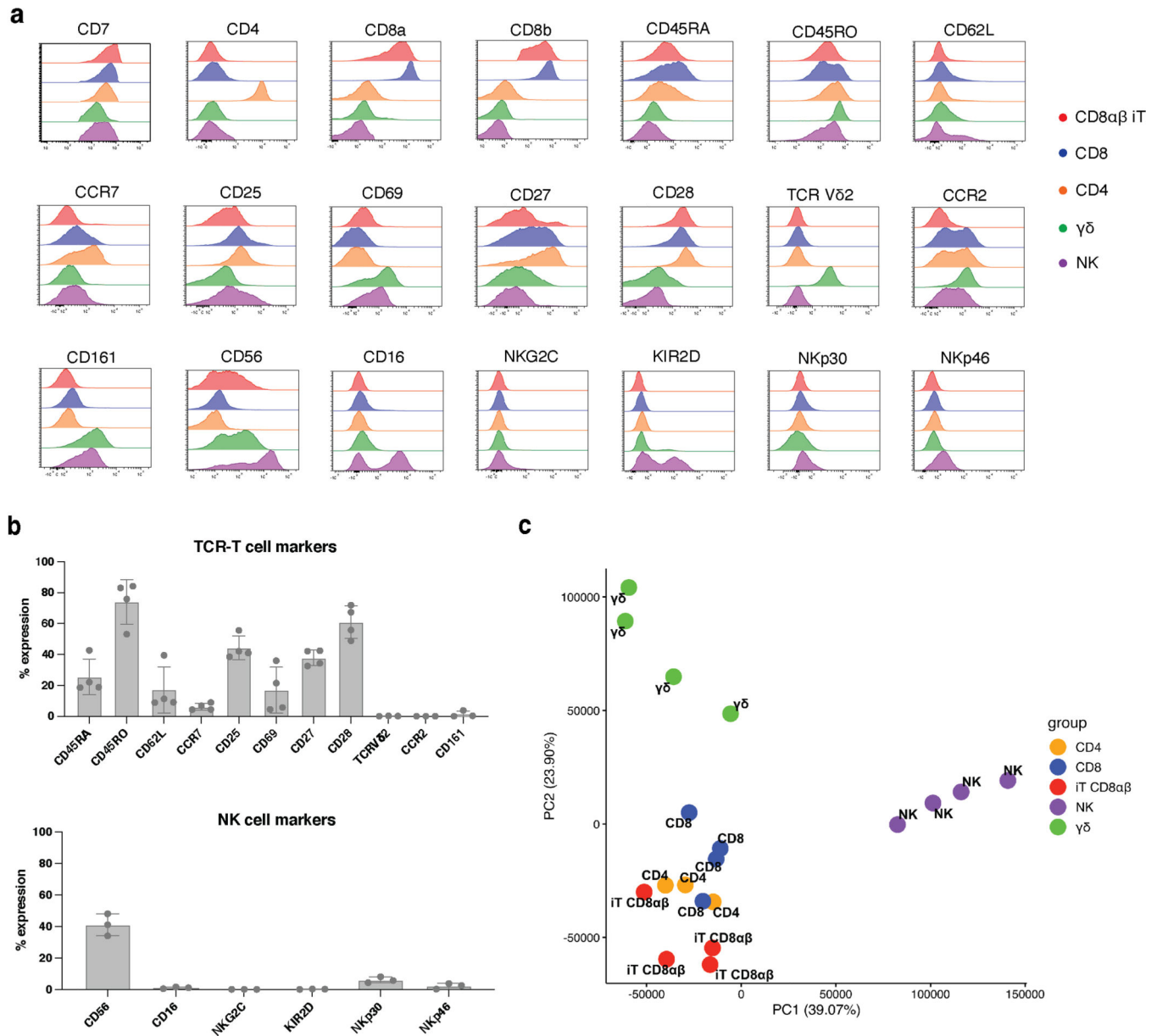
a, T lineage commitment marker expression (CD7/CD5, CD4/CD8 α , CD8 α /CD8 β) of WT-TiPS (left), *TRAC-1928z*-TiPS (middle) and *TRAC-1XX*-TiPS on OP9-DLL4 at the indicated timepoints. CD7/CD5 is gated on live CD45⁺ cells, others are gated on live CD45⁺CD7⁺ cells. **b**, Flow cytometric analysis of T cell phenotype markers of D35 DP *TRAC-1XX*-iT cells. Gated on live CD45⁺CD7⁺CD4⁺CD8 α β ⁺ cells. **c**, Intracellular and cell-surface expression of CD3 and TCR α β on D35 *TRAC-1XX*-iT cells.





Extended Data Fig. 7. DP *TRAC-1XX-iT* cell mature to *CD8αβ* SP *iT* cells on 3T3-CD19-41BBL
a, c, Flow cytometric analysis of D42 cells matured on 3T3-CD19 (**a**) or 3T3-CD19-41BBL (**c**). Gated on live $CD45^+CD7^+$ cells. **b**, Flow cytometric analysis of D35 and D42 phenotypes of stimulated DP *TRAC-1XX-iT* cells. D35 *TRAC-1XX-iT* cells were sorted for a $CD4^+CD8αβ^+$ DP phenotype, stimulated on 3T3-CD19-41BBL and expanded for seven days. Gated on live $CD45^+CD7^+$ cells. **d**, Fold Expansion and T cell phenotype marker expression of *TRAC-1XX-iT* cells matured on 3T3-CD19-41BBL (3T3) or recombinant CD19-Fc. **e**, 4 h cytotoxicity assay of 3T3-CD19-41BBL stimulated *TRAC-1XX-iT* cells

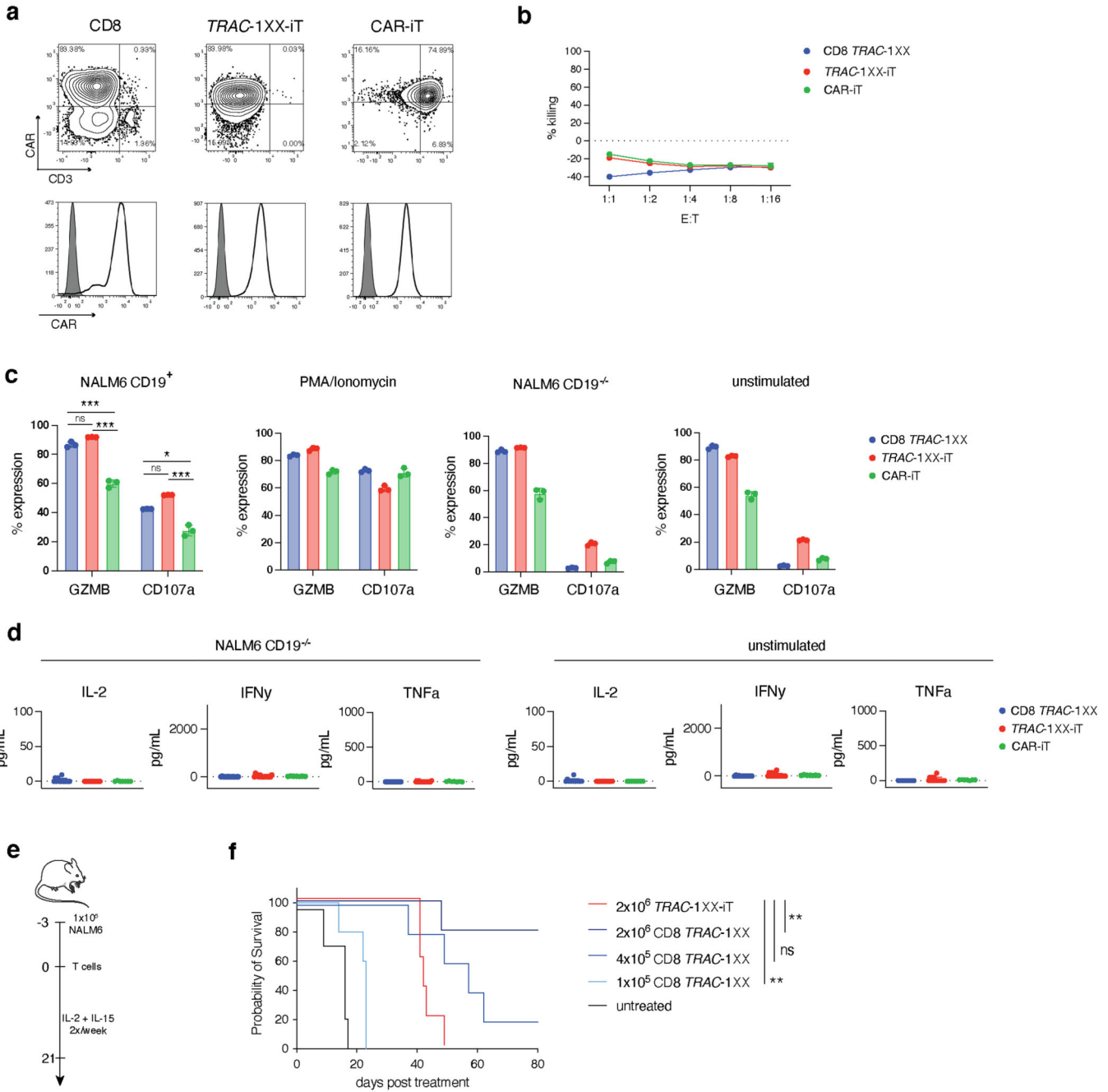
in response to NALM6-CD19⁺ an NALM6-CD19^{-/-} target cells (n = 3 technical replicates, data are means ± s.d.) **f** CD19 expression on primary CLL cells.



Extended Data Fig. 8. Comparison of CD8 $\alpha\beta$ TRAC-1XX-iT cells and peripheral blood lymphocytes

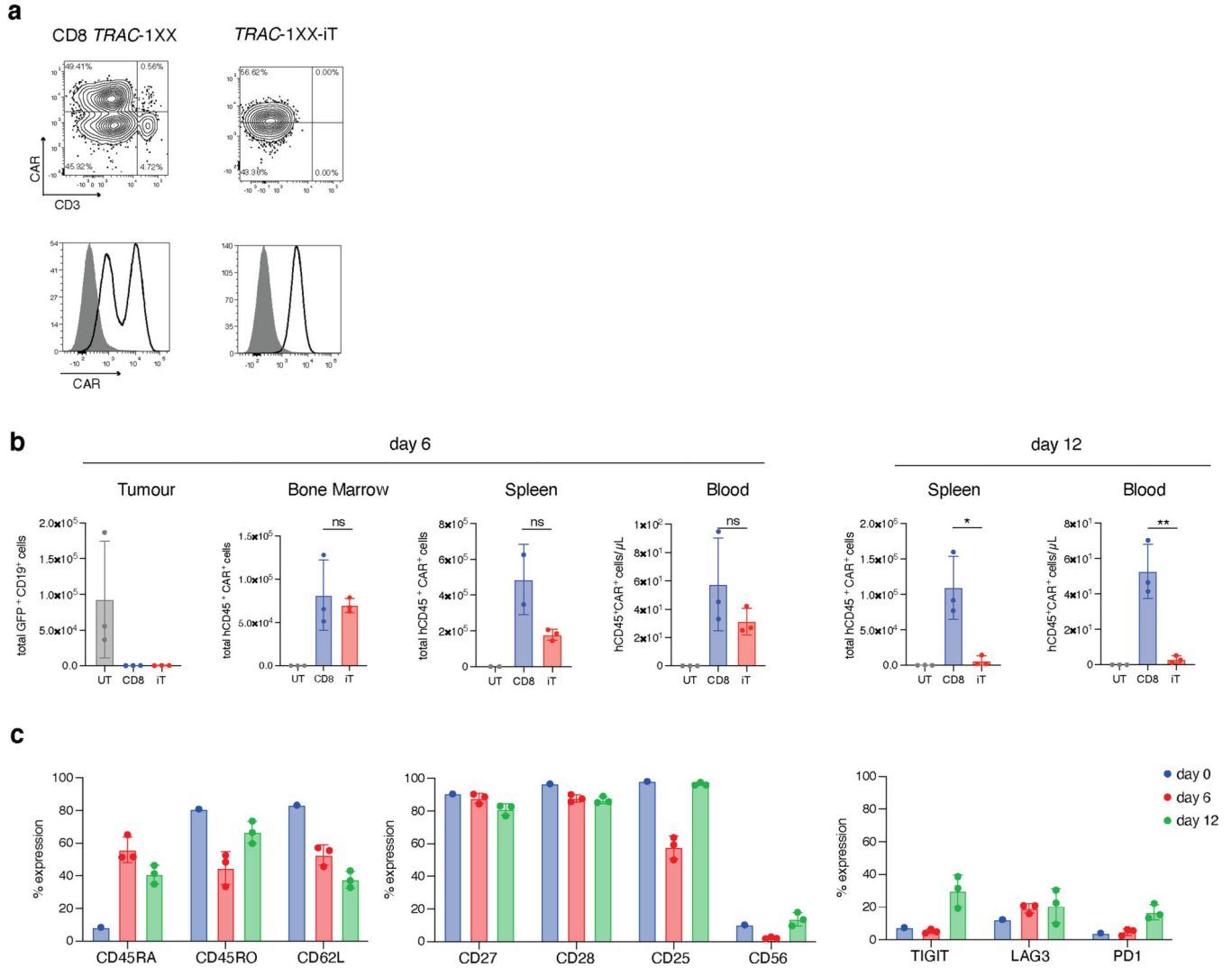
a, Representative examples of lymphoid phenotype marker expression in CD8 $\alpha\beta$ TRAC-1XX-iT (red), CD8 $\alpha\beta$ $\alpha\beta$ TCR-T (blue), CD4 $\alpha\beta$ TCR-T (orange), $\gamma\delta$ TCR-T (green) and NK cells (purple). CD8 $\alpha\beta$ TRAC-1XX-iT cells are the same as represented in Fig. 5a. **b**, Variability of lymphoid phenotype marker expression in CD8 $\alpha\beta$ TRAC-1XX-iT cells (n = 3–4 biological replicates, data are means ± s.d.). Biological replicates shown are samples utilized in RNA analysis (Fig. 5b, c). **c**, Principal Component Analysis comparing TRAC-1XX CD8 $\alpha\beta$ $\alpha\beta$ TCR-T cells (CD8, n = 4), TRAC-1XX CD4 $\alpha\beta$ TCR-T cells (CD4,

n = 3), γ RV-1XX $\gamma\delta$ TCR-T cells ($\gamma\delta$, n = 4), γ RV-1XX NK cells (NK, n = 4) and CD8 $\alpha\beta^+$ TRAC-1XX-iT cells (iT CD8 $\alpha\beta$, n = 4).



Extended Data Fig. 9. Functional comparison of TRAC-1XX-iT, CAR-iT and CD8 TRAC-1XX
 Functional comparison of healthy-donor peripheral blood TRAC-1XX CD8 $\alpha\beta^+$ $\alpha\beta$ TCR-T (CD8 TRAC-1XX), CAR-iT and TRAC-1XX-iT cells. CD8 TRAC-1XX cell doses represent number of CAR⁺ cells utilized in the assay. **a**, CAR and CD3 expression in CD8 TRAC-1XX, CAR-iT and TRAC-1XX-iT cells (black line) compared to unstained control (grey filled histogram). **b**, 18 h incucyte cytotoxicity assay with NLR⁺ CD19^{-/-}

NALM6 target cells (n = 3 technical replicates). **c**, 4 h intracellular cytokine detection in T cells stimulated with NALM6 CD19⁺ target cells (at a 1:1 E:T), PMA/Ionomycin, NALM6 CD19^{-/-} target cells (at a 1:1 E:T) unstimulated controls (n = 3 technical replicates). **d**, 24 h cytokine secretion using NALM6-CD19^{-/-} as target cells at a 1:1 E:T (n = 11–18 biological replicates, left panel) or unstimulated control (n = 11–18 biological replicates, right panel). **e**, Schematic representation of the NALM6 in vivo tumour model. **f**, Kaplan-Meier analysis of tumour free survival (2×10^6 TRAC-1XX-iT vs 2×10^6 CD8 TRAC-1XX p=0.0062, 2×10^6 TRAC-1XX-iT vs 1×10^5 CD8 TRAC-1XX p=0.0034). * P<0.05, ** P<0.01, *** P<0.001, Chi-Square test (**b**) log-rank Mantel-Cox test (**f**). All data are means \pm s.d.



Extended Data Fig. 10. TRAC-1XX-iT function compared to healthy donor peripheral blood-derived CD8 TRAC-1XX T cells.

In vivo functional comparison of healthy-donor peripheral blood TRAC-1XX CD8αβ αβTCR-T (CD8 TRAC-1XX), TRAC-1XX-iT cells. CD8 TRAC-1XX cell doses represent number of CAR⁺ cells utilized in the assay. **a**, CAR and CD3 expression in CD8 TRAC-1XX and TRAC-1XX-iT cells (black line) compared to unstained control (grey

filled histogram). **b**, Enumeration of tumour cells in the bone marrow and T cells in bone marrow, spleen and blood 6 or 12 days post T cell infusion (n = 2–3 mice, T cell in bone marrow day 12, CD8 *TRAC-1XX* vs *TRAC-1XX-iT* p=0.0161, T cells in spleen day 12 CD8 *TRAC-1XX* vs *TRAC-1XX-iT* p=0.0052). **c**, Phenotype of CD8 cells prior to infusion (day 0, n = 1) and of cells derived from the bone marrow on day 6 (n = 3 mice) and 12 (n = 3 mice). **d**, Kaplan-Meier analysis of overall survival. * P<0.05, ** P<0.01, *** P<0.001 Welch's 2-sample two-sided t test (**b**) All data are means ± s.d.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Data Availability

The RNA sequencing data have been deposited in the Gene Expression Omnibus under the accession number GSE210364. Source data for tumour growth are provided with this paper. Other data generated for this manuscript are available from the corresponding author upon reasonable request.

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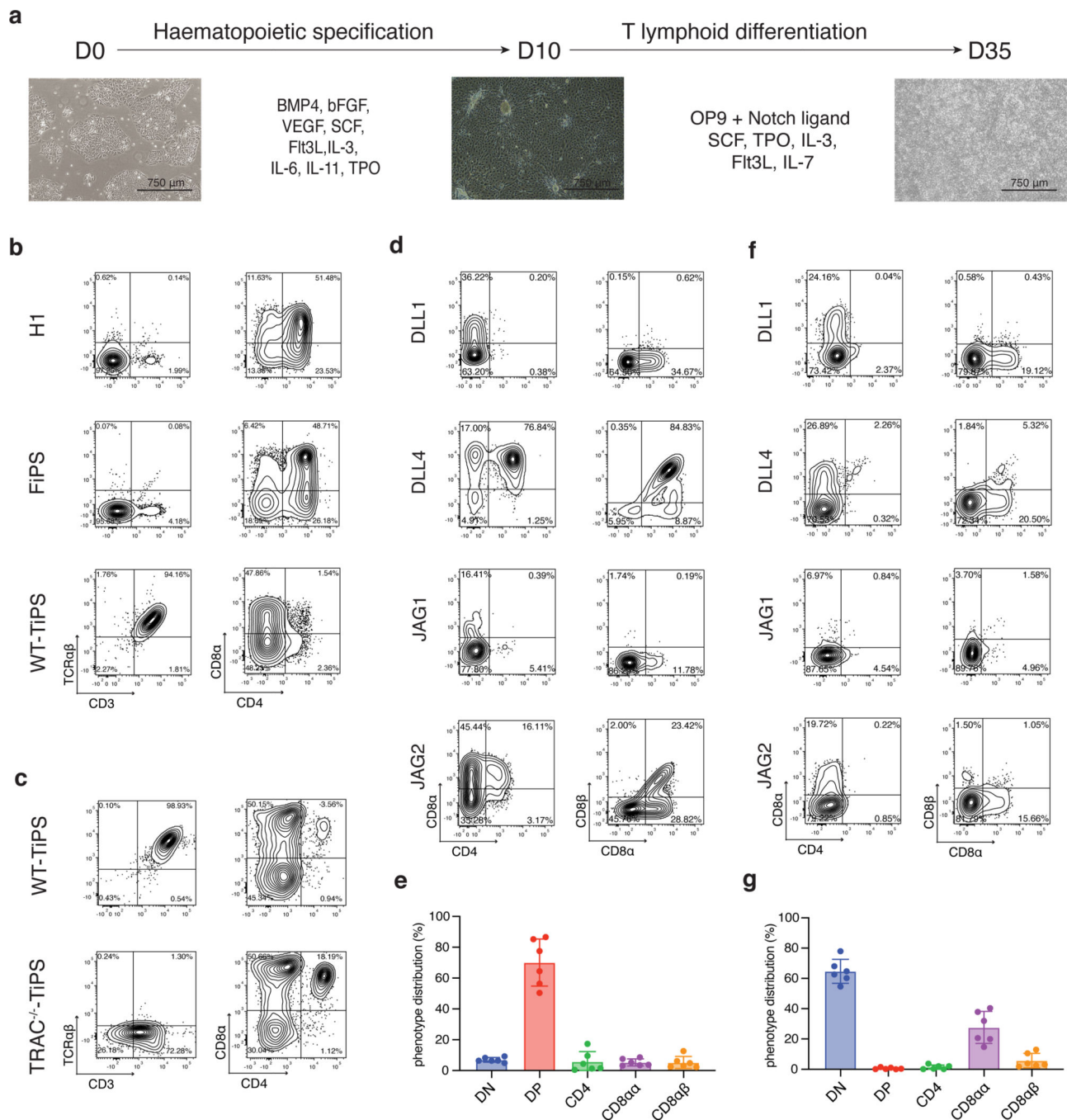


Fig. 1. DLL4 supports *in vitro* $\alpha\beta$ TCR-T cell development of WT-TiPS but not CAR-TiPS
a, Schematic representation of *in vitro* T cell differentiation protocol. Microscope images are at 4x magnification, scale bar represents 750 μ m. **b**, Flow cytometric analysis of T lineage commitment of H1, FiPS and WT-TiPS on OP9-mDLL1, gated on live CD45⁺CD7⁺ cells at day 40 (D40) in the differentiation. **c**, Flow cytometric analysis of T lineage commitment of WT-TiPS and *TRAC*^{-/-}-TiPS on OP9-mDLL1, gated on live CD45⁺ cells at D40 in the differentiation. **d**, **f**, Representative flow cytometric analysis of T lineage commitment of WT-TiPS (**d**) and CAR-TiPS (**f**) on D35 in differentiation on OP9 expressing the indicated

human Notch ligand, gated on live CD45⁺CD7⁺ cells. **e, g**, Phenotype distribution of WT-TiPS (**e**, n = 6 biological replicates) or CAR-TiPS (**g**, n = 6 biological replicates) on D35 of differentiation on OP9-DLL4, gated on live CD45⁺CD7⁺ cells. All data are means ± s.d.

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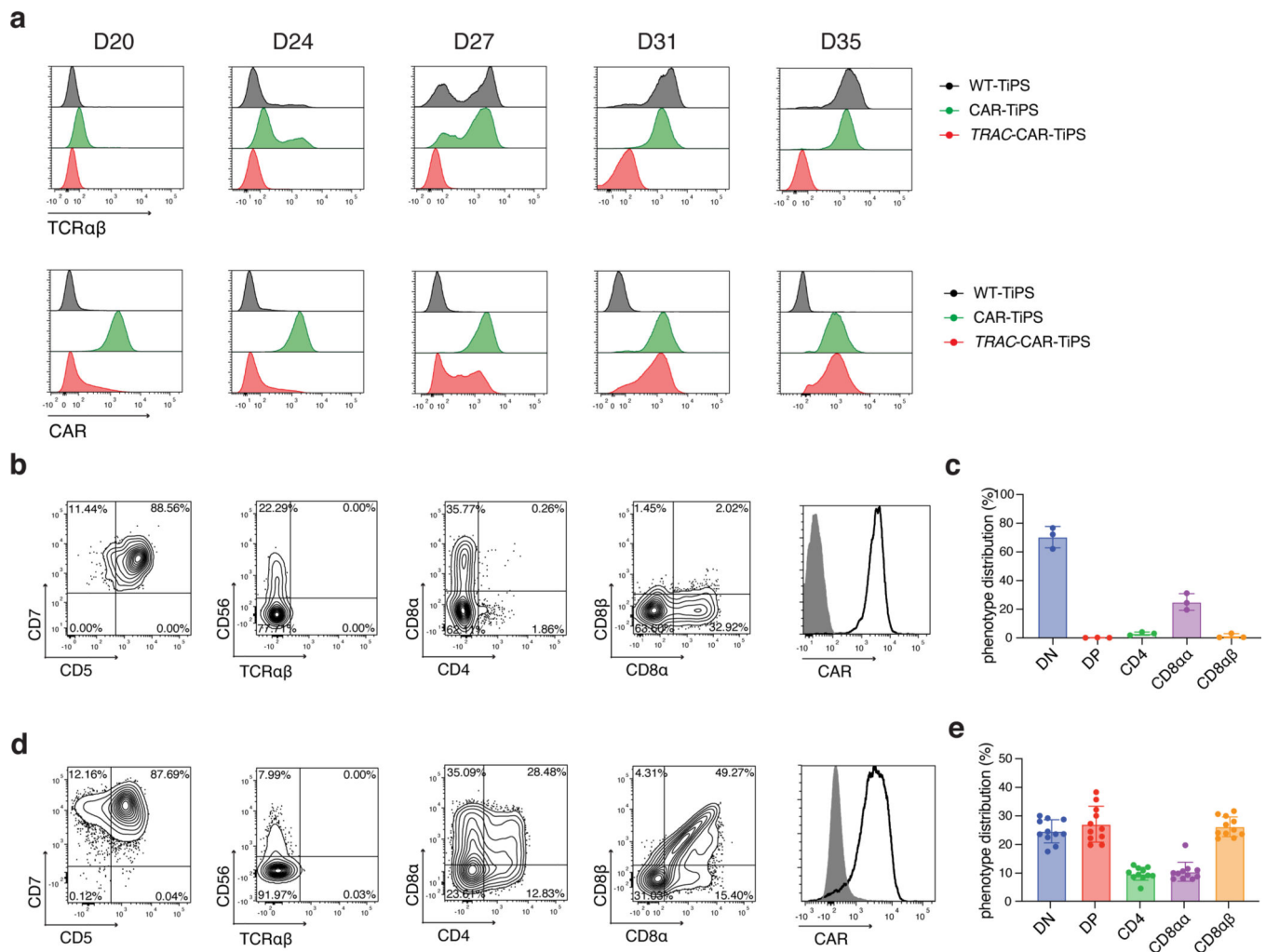


Fig. 2. TRAC-controlled 1928z-1XX CAR expression facilitates DP T cell development

a, Induction of $\alpha\beta$ TCR (upper panel) and CAR (lower panel) expression in WT-TiPS, CAR-TiPS and TRAC-CAR-TiPS throughout T lymphoid development on OP9-DLL4 at the indicated timepoints. Gated on live CD45⁺CD7⁺ cells. **b**, **d**, Representative flow cytometric analysis of T lineage commitment markers of TRAC-1928z-TiPS (**b**) and TRAC-1XX-TiPS (**d**) gated on live CD45⁺ cells at D35 in differentiation on OP9-DLL4. **c**, **e**, D35 phenotype distribution of TRAC-1928z-TiPS (**c**, n = 3 biological replicates) and TRAC-1XX-TiPS (**e**, n = 11 biological replicates) at D35 gated on live CD45⁺CD7⁺ cells. All data are means \pm s.d.

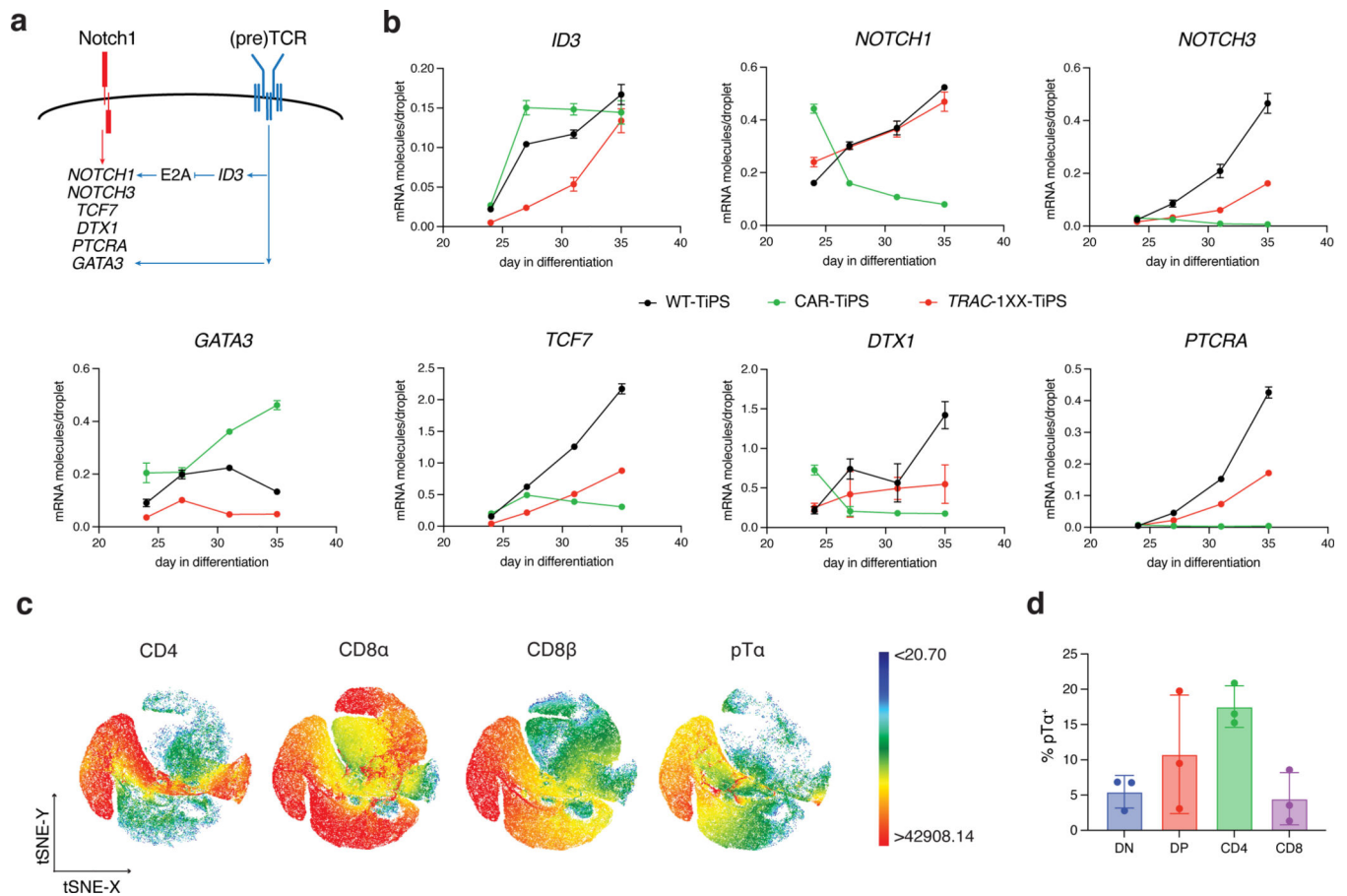


Fig. 3. CAR regulation influences Notch and TCR target gene induction

a, Schematic representation of Notch and (pre)TCR signalling interactions as reported in the literature. **b**, ddPCR analysis of Notch and (pre)TCR target genes at D24, 27, 31 and 35 of T cell differentiation (normalized to *RPL13A*) ($n = 3$ technical replicates). **c**, t-distributed Stochastic Neighbour embedding (tSNE) analysis of cell surface expression of CD4, CD8 α , CD8 β and pT α on D35 *TRAC-1XX-TiPS* iT cells. Colour scale represent level of marker expression. **d**, Distribution of pT α expression on D35 *TRAC-1XX-TiPS* ($n = 3$ biological replicates). All data are means \pm s.d.

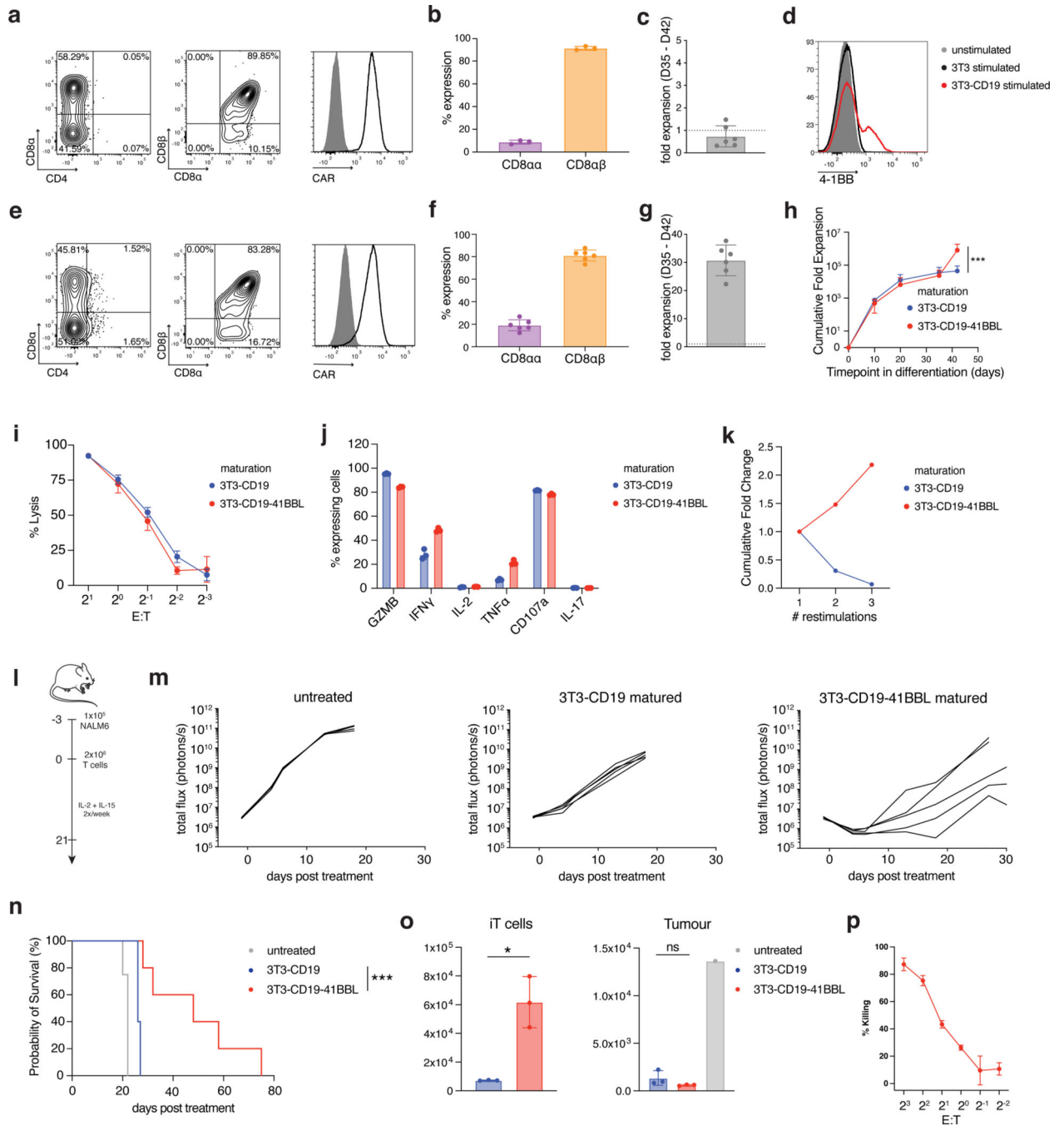


Fig. 4. 4-1BBL co-stimulation enhances CD8 $\alpha\beta$ TRAC-1XX-iT proliferation and function
a, Representative phenotype of TRAC-1XX-iT cells matured on 3T3-CD19 for 7 days (D35-D42), gated on live CD45⁺CD7⁺ (left and right) and CD45⁺CD7⁺CD8 α ⁺ (middle). **b**, Distribution of CD8 α α and CD8 α β phenotype in the CD8 α ⁺ compartment (n = 3 biological replicates). **c**, Expansion of TRAC-1XX-iT cells from D35-D42 after maturation on 3T3-CD19 (n = 6 biological replicates). **d**, 4-1BB cell-surface expression on TRAC-1XX-iT cells on D35 8 h after exposure to parental 3T3 (black), 3T3-CD19 (red) or left unstimulated (grey). **e**, Representative phenotype of TRAC-1XX-iT cells matured on 3T3-CD19-41BBL,

gated on live CD45⁺CD7⁺ (left and right) and CD45⁺CD7⁺CD8α⁺ (middle). **f**, Distribution of CD8αα and CD8αβ phenotype in the CD8α⁺ compartment (n = 6 biological replicates). **g**, Expansion of *TRAC*-1XX-iT cells from D35-D42 after maturation on 3T3-CD19-41BBL (n = 6 biological replicates). **h**, Total cell expansion from D0-D42 in iT differentiation with maturation on 3T3-CD19±41BBL (n = 6 biological replicates for each group, p=0.0008). **i**, Cytotoxic activity measured in an 18 h bioluminescence assay, using firefly luciferase (FFLuc)-expressing NALM6 at the indicated effector-to-target (E:T) ratios (n = 3; technical replicates). **j**, 4 h intracellular cytokine detection of 3T3-CD19±41BBL-matured cells in response to NALM6 (n = 3 technical replicates). **k**, Representative expansion of D42 cells matured on 3T3-CD19±41BBL upon repeated weekly antigen exposure on 3T3-CD19. **l**, Schematic representation of NALM6 *in vivo* tumour model. **m**, Tumour burden (total flux in photons per second) of NALM6-bearing mice treated with 2×10⁶ D42 *TRAC*-1XX-iT cells (n = 4–5, line = one mouse). **n**, Kaplan-Meier analysis of overall survival (p<0.0001). **o**, Flow cytometric quantification of iT cells (left panel, p=0.0339) and tumour cells (right panel) in bone marrow 6 days after T cell infusion (n = 3). **p**, Cytotoxic activity measured in a 6 h flow cytometry assay, using primary CD19⁺ CLL cells at the indicated E:Ts with 3T3-CD19-41BBL-matured *TRAC*-1XX-iT cells (n = 3 technical replicates) * P<0.05, *** P<0.001, Welch's 2-sample two-sided t test (**h**, **o**), log-rank Mantel-Cox test (**n**). All data are means ± s.d.

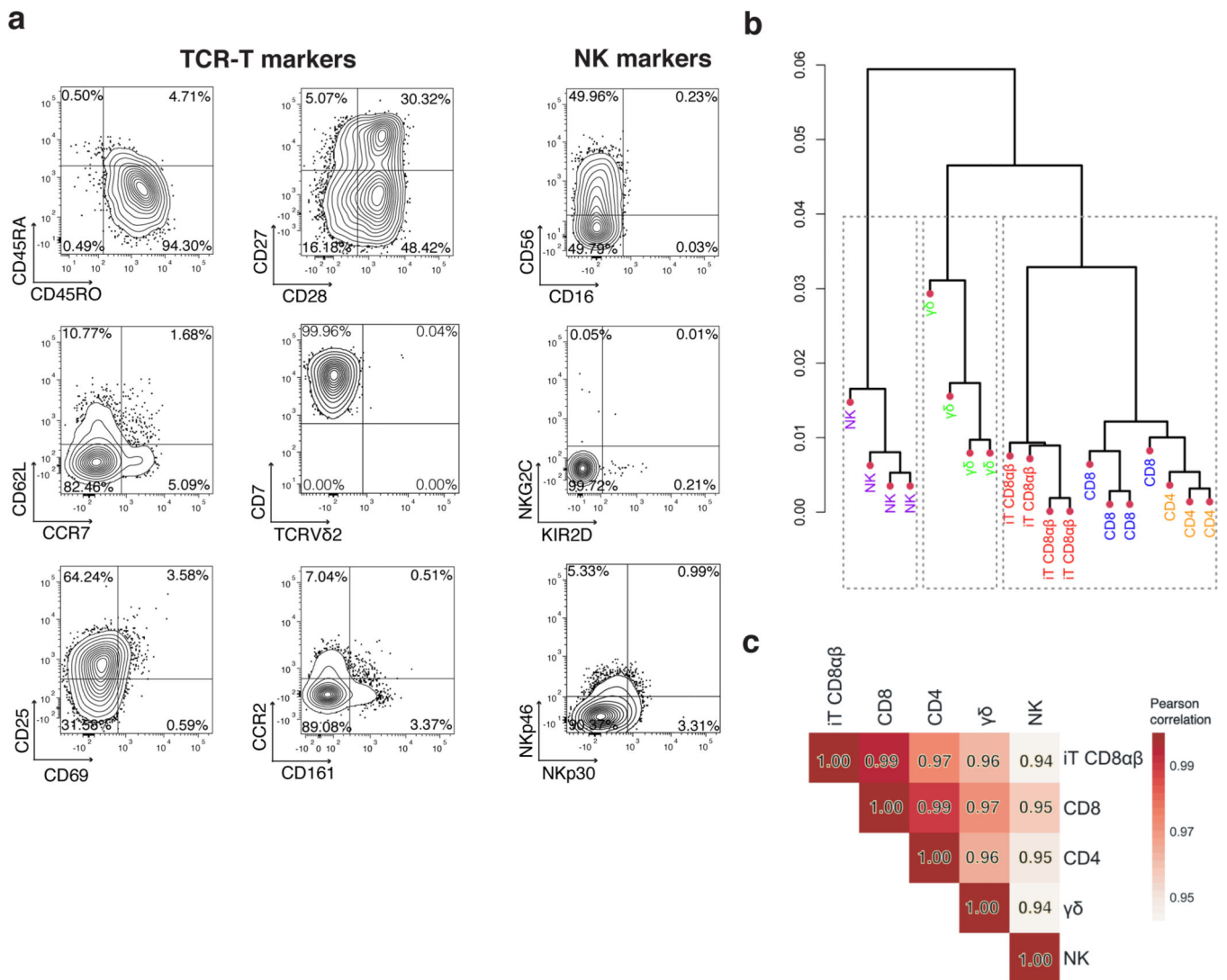


Fig. 5. CD8αβ TRAC-1XX-iT cells resemble peripheral-blood derived CD8αβ T cells

a, Phenotype analysis of 3T3-CD19–41BBL-matured D42 CD8αβ TRAC-1XX-iT cells for TCR-T cell markers (left and middle panel) and NK-cell markers (right panel). Data is representative of four independent experiments, gated on live CD45⁺CD7⁺CD8αβ⁺ cells.

b, Dendrogram of hierarchical clustering analysis based on Euclidian distance matrix comparing the transcriptome of TRAC-1XX CD8αβ αβTCR-T cells (CD8, blue, n = 4 biological replicates), TRAC-1XX CD4 αβTCR-T cells (CD4, orange, n = 3 biological replicates), γRV-1XX γδTCR-T cells (γδ, green, n = 4 biological replicates), γRV-1XX NK cells (NK, purple, n = 4 biological replicates) and CD8αβ⁺ TRAC-1XX-iT cells (iT CD8αβ, red, n = 4 biological replicates). **c**, Correlation matrix using Pearson’s statistics comparing same groups as in (b).

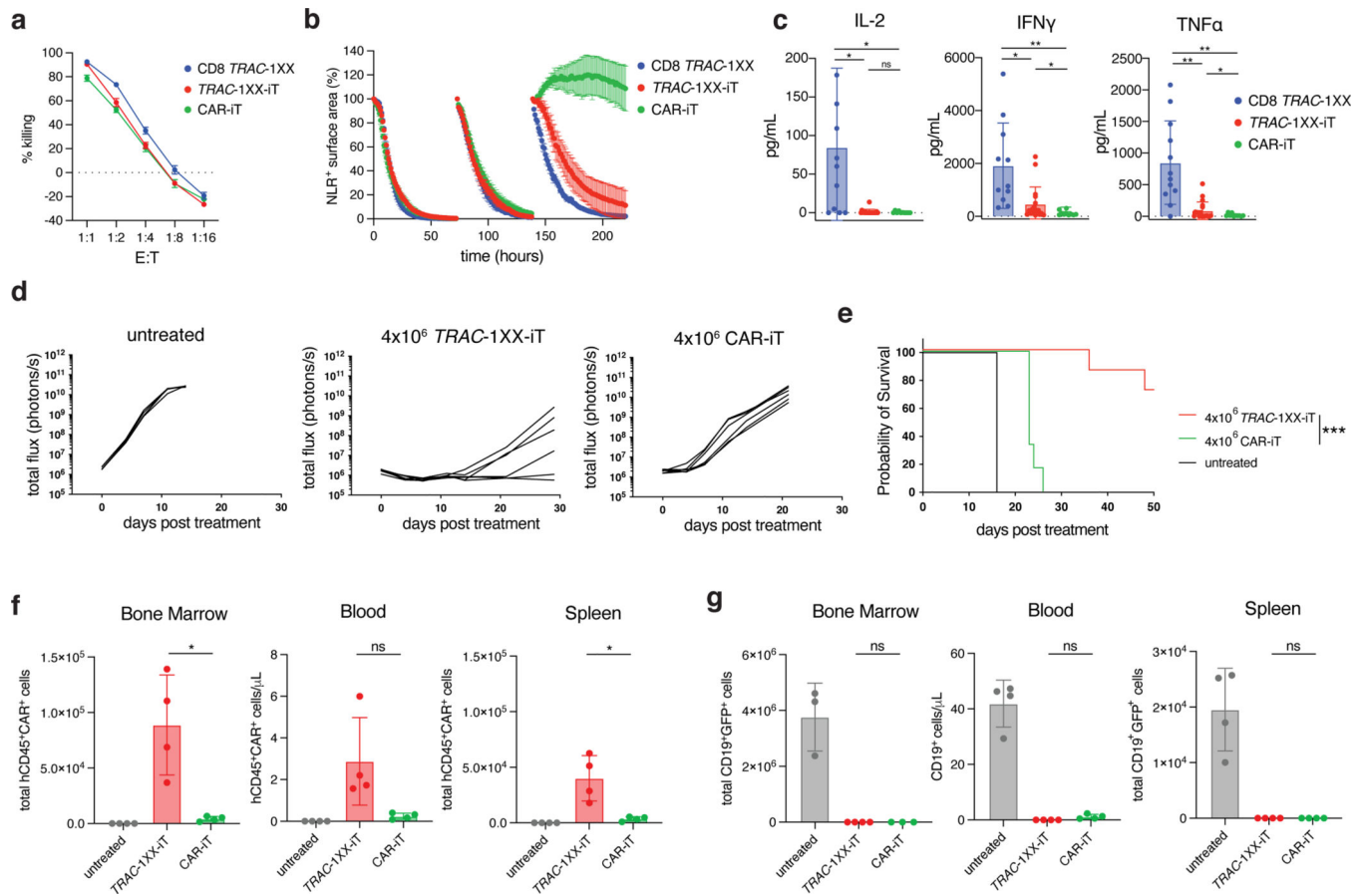


Fig. 6. TRAC-1XX-iT have improved persistence and function over CAR-iT cells

Functional comparison of healthy-donor peripheral blood TRAC-1XX CD8 $\alpha\beta$ $\alpha\beta$ TCR-T cells (CD8 TRAC-1XX), CAR-iT, and TRAC-1XX-iT cells (matured on 3T3-CD19–41BBL). CD8 TRAC-1XX doses reflect number of CAR⁺ T cells utilized in the assay. **a**, Cytotoxic activity using a 18 h Incucyte assay, using NLR-expressing NALM6 as target cells (n = 3 technical replicates). **b**, NALM6 rechallenge assay. NLR⁺ NALM6 and T cells were co-cultured at a 1:1 E:T. Every 72 h T cells were rechallenged with 1x NLR⁺NALM6 and cytokines. NALM6 clearance was measured in NLR⁺ surface area reduction compared to the timepoint of rechallenge (n = 3 technical replicates). **c**, Twenty-four h cytokine secretion using NALM6 as target cells at a 1:1 E:T ratio (CD8 TRAC-1XX n = 15, TRAC-1XX-iT n = 18, CAR-iT n = 11 biological replicates, IL-2 CD8 TRAC-1XX vs CAR-iT p=0.02, IL-2 CD8 TRAC-1XX vs TRAC-1XX-iT p=0.0222, IFN γ CD8 TRAC-1XX vs CAR-iT p=0.004, IFN γ CD8 TRAC-1XX vs TRAC-1XX-iT p=0.0293, IFN γ TRAC-1XX-iT vs CAR-iT p=0.0363, TNF α CD8 TRAC-1XX vs CAR-iT p=0.001, TNF α CD8 TRAC-1XX vs TRAC-1XX-iT p=0.0027, TNF α TRAC-1XX-iT vs CAR-iT p=0.0262). **d**, Tumour burden (total flux in photons per second) of NALM6-bearing, untreated mice (n = 4) or mice treated with 4x10⁶ TRAC-1XX-iT (middle) or CAR-iT (right) cells (n = 6, line = one mouse). **e**, Kaplan-Meier analysis of overall survival (p=0.002). **f,g**, Enumeration of iT cells (**f**) and tumour cells (**g**) in the bone marrow, spleen and blood 12 days post T cell infusion (n = 4 mice, iT in bone marrow TRAC-1XX-iT vs CAR-iT p=0.0329, iT in

spleen *TRAC-1XX-iT* vs *CAR-iT* $p=0.0369$). * $P<0.05$, ** $P<0.01$, *** $P<0.001$, Welch's 2-sample two-sided t test (**c, f, g**), log-rank Mantel-Cox test (**e**). All data are means \pm s.d.

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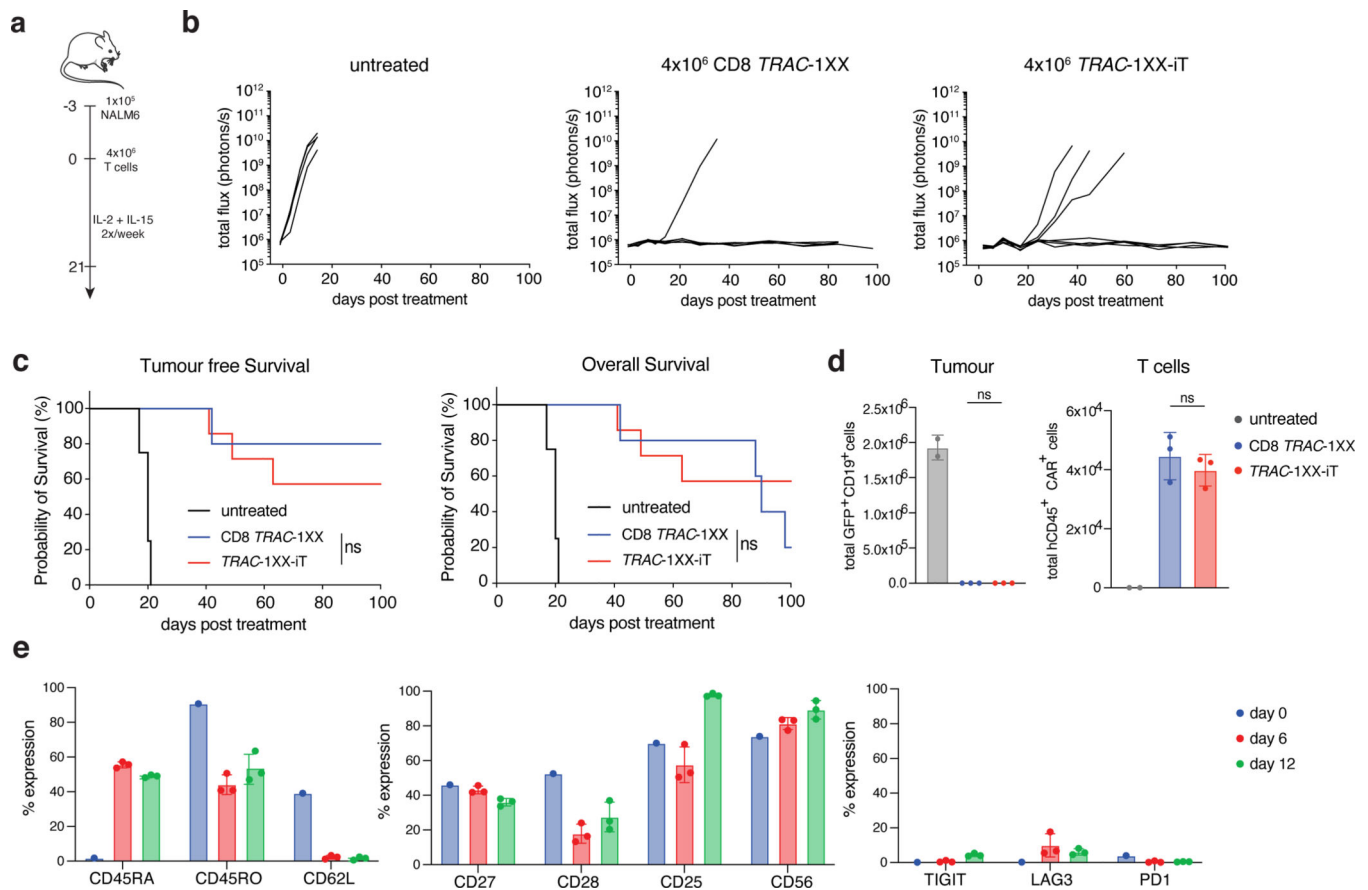


Fig. 7. TRAC-1XX-iT cells cure systemic NALM6 tumour model without inducing graft-versus-host disease

a, Schematic representation of systemic NALM6 tumour model. **b**, Tumour burden (total flux in photons per second) of NALM6-bearing untreated mice ($n = 4$), or mice treated with 4×10^6 CD8 TRAC-1XX ($n = 5$) or TRAC-1XX-iT cells ($n = 7$, line = one mouse). **c**, Kaplan-Meier analysis of tumour-free survival (left) and overall survival (right). **d**, Flow cytometric quantification of tumour cells (left) and T cells (right) in bone marrow 12 days after T cell infusion ($n = 2-3$ mice). **e**, Phenotype of persisting TRAC-1XX-iT cells prior to infusion (day 0, $n = 1$) and of cells derived from the bone marrow on day 6 and 12 days after TRAC-1XX-iT cell infusion ($n = 3$ mice). * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, log-rank Mantel-Cox test (c), Welch's 2-sample two-sided t test (d). All data are means \pm s.d.