

UCLA

UCLA Previously Published Works

Title

Fabrication of antibiotic-loaded dissolvable calcium sulfate beads: an in vitro mixing lab utilizing various antibiotic mixing formulas.

Permalink

<https://escholarship.org/uc/item/89g6m0dp>

Journal

Journal of Bone and Joint Infection, 6(9)

ISSN

2206-3552

Authors

McPherson, Edward

Dipane, Matthew

Chowdhry, Madhav

et al.

Publication Date

2021

DOI

10.5194/jbji-6-405-2021

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <https://creativecommons.org/licenses/by/4.0/>

Peer reviewed



Fabrication of antibiotic-loaded dissolvable calcium sulfate beads: an in vitro mixing lab utilizing various antibiotic mixing formulas

Edward J. McPherson¹, Matthew V. Dipane¹, Madhav Chowdhry², and Andrew J. Wassef³

¹Department of Orthopaedic Surgery, David Geffen School of Medicine at UCLA, Santa Monica, 90404, USA

²Shanti Hospital and Nursing Home, Aligarh, 202001, India

³Long Beach Lakewood Orthopedic Institute, Long Beach, 90808, USA

Correspondence: Matthew V. Dipane (mdipane@mednet.ucla.edu)

Received: 9 September 2021 – Revised: 23 October 2021 – Accepted: 25 October 2021 – Published: 10 November 2021

Abstract. Chronic periprosthetic joint infection (PJI) is a devastating complication that requires an aggressive eradication protocol. Local antimicrobial delivery via dissolvable calcium sulfate (CaSO_4) using small-sized beads (3–8 mm) has been utilized as an adjunctive treatment combined with implant exchange, radical debridement, and antimicrobial loaded acrylic spacers. The non-exothermic setting of CaSO_4 theoretically allows for any antimicrobial agent to be used, so long as mixing methods provide a consistent fabrication within a reasonable set time. This study performed the first in vitro mixing study, in which various antimicrobial agents, used singularly and in combination, were mixed with a synthetic CaSO_4 product to observe and document their interactions. The study was performed in a simulated operating room environment. We report a standard mix formula with set times, testing 22 different antimicrobial agents, combinations, and doses. For some antimicrobials and combinations, set times using the standard formula were either too fast or exceedingly slow. For these 14 antimicrobial agents and combinations, we were able to arrive at individualized mixing methods. We present all mixing formulas and set times. In all, we were able to establish mixing methods that incorporate all antimicrobial agents and combinations that we have seen utilized via surgeon-directed use.

1 Introduction

Joint replacement surgery for degenerative arthrosis provides improved function and pain relief in the majority of cases where this procedure is utilized (Merx et al., 2003; Lohmander et al., 2006; Lützner et al., 2011; Maradit Kremers et al., 2015). However, adverse responses do occur at relatively low rates (McPherson et al., 2020). Such risks include fixation failure, mechanical dysfunction, fracture, reactive wear debris phenomenon, and infection. Of these complications, that which stands out as the most problematic is periprosthetic joint infection (PJI). This is due to the difficulty in eradicating the infection, as well as the enormous impact placed upon the patient and healthcare system (Kurtz et al., 2008, 2012; Costerton et al., 1999, 2005). Further complicating matters is the extended use of parenteral antimicrobial agents, which disrupts numerous human physiologic systems, confers hu-

man biome resistance, and affects future treatment options (Levast et al., 2021).

Alternatives to parenteral antimicrobial treatment for PJI are limited. Local antimicrobial delivery by various carriers has been utilized with increasing frequency in the last 3 decades. The mainstay agents include polymethyl methacrylate (PMMA) in bead form and PMMA in bulk form. In bulk form, the antimicrobial-loaded acrylic cement (ALAC) serves to maintain the joint space. PROSTALAC (PROSThesis + ALAC) implants provide additional stability and function to the resected joint. The other utilized agent for local intra-articular antimicrobial delivery is calcium sulfate (CaSO_4), which dissolves and releases antimicrobials. These agents are utilized only for physician-directed use but are considered acceptable methods by those orthopaedic surgeons focusing on PJI treatment (Dacquet et al., 1992; Dahners and Funderburk, 1987; Evrard et al., 1990).

Newer forms of synthetic CaSO_4 have been developed that are considered to be improved products. Legacy CaSO_4 products were “mined and refined” with process techniques that allow implantation into human hosts. Impurities with these products were thought to cause adverse reactions affecting treatment (Lee et al., 2002). Subsequent second- and third-generation products are synthetic, as they are produced from pure elemental form. These agents are better able to accommodate antibiotic impregnation and provide a more consistent performance in terms of mixing, dissolution, and clinical application, although wound drainage still remains a concern for surgeons, with reported rates in the literature varying considerably (Kelly et al., 2001; Ziran et al., 2007; Ferguson et al., 2014; Kallala et al., 2018). Newer antimicrobial delivery agents are being developed for the specific property of antimicrobial compatibility but are not yet in clinical trials (Ambrose et al., 2014). Thus, for the foreseeable future, antimicrobial-loaded CaSO_4 beads, combined with ALAC or PROSTALAC spacers, are the common treatment methods for local antimicrobial delivery.

With the advent of microbial next-generation DNA sequencing combined with specified PJI culture methods, the number of identified microbiota associated with PJI is growing considerably (Aul et al., 1998; Tarabichi et al., 2018). This requires the employment of a wider variety of antimicrobial agents to be used in isolation or in concert. This paper reports our work with a mixing study whereby, in a laboratory setting, we mixed antimicrobial agents into a synthetic, commercially pure CaSO_4 product, creating a dissolvable antimicrobial bead. In cursory review, this concept seems simple, but in reality, the process can be immensely frustrating and difficult. Our purpose is to report our baseline standardized mixing techniques, developing formulas, and doses of antimicrobials added to CaSO_4 . Furthermore, when encountering difficulty in mixing specific antimicrobial agents, we developed modified mixing techniques to create an antimicrobial-loaded bead that previously was problematic with our standard technique.

2 Methods

The CaSO_4 product used for this study was Synthecure® (Austin Medical Ventures, Memphis, TN). We used the 10 cc bead kits that consisted of CaSO_4 hemihydrate powder that is mixed with 5 cc of sterile saline. When mixed, the unloaded CaSO_4 product makes a paste volume of 10 cc. The mixed product is spread into a sterile silicone bead mat, allowing for the fabrication of 3, 4.8, or 6 mm beads. For the purposes of this study, we selected the 4.8 mm beads. The bead mat and bead product formed in this laboratory study are shown in Fig. 1. The antimicrobial formulae selected for study were based on clinical and academic observations of formulae currently utilized in the orthopaedic community.

In the majority of added antimicrobial formulae, the sterile powdered form of the antimicrobial was premixed into the CaSO_4 powder, after which the sterile saline liquid was added. Mixing was performed in a 6 cm diameter mixing bowl with a spatula for 15–45 s, until a paste consistency devoid of antimicrobial clumps was achieved. The paste product was transferred via the spatula to the bead mat. For this study, we measured the time from initial mix until the product became a consistently smooth product devoid of clumps. Once the product was spread onto the bead mat, the product was left to set on the laboratory table in standard conditions of 70°F and 40 % humidity. We tested the beads for setting by gently bending the mat at 1 min intervals to assess separation from the silicone mat. In addition, we examined the bead itself. If the bead showed “fissuring” as the mat was gently bent, the curing process was not complete, and the mat was placed back onto the laboratory table. We defined the product as “set” when we could bend the bead mat as it was turned upside down, and the beads would fall onto the table.

A second confirmatory test included a digital bead compression test when the bead was on the table. In some cases, the CaSO_4 product would fall out of the bead mat but was still soft. In this scenario, the beads were tested by compressing the bead with the tip of the index finger. If the bead was not fully cured, the bead would collapse. When “fully set,” the bead would not collapse. Thus, the measured “set time” was defined as the time from initial mix until the bead dropped out of the bead mat and was confirmed as fully set via the digital compression test.

Two exceptions to using the antimicrobial powder mix method for this study were Tobramycin and Gentamicin. In many centers, both Tobramycin and Gentamicin come conveniently premixed in 2 mL vials and can be used as a substitute for the sterile saline that is included in the mix kit. Liquid Tobramycin (or Gentamicin) has two advantages. First, as noted above, it is conveniently provided in 2 mL vials (80 mg of antibiotic per vial). Secondly, as Tobramycin (or Gentamicin) is provided in liquid form, additional antimicrobial powders can be added to the CaSO_4 powder, thus allowing combination formulas to be tested. We elected to use 6 mL of liquid Tobramycin and Gentamicin in our formulae, rather than the manufacturer recommended 5 mL, to make matters simple and efficient for operating room staff, rather than requiring the exact measurement of one-half vial of liquid to achieve 5 mL. Preliminary testing between formulae of 6 mL and those of 5 mL demonstrated no significant difference in set time or quality.

During the standardized mix tests, we found difficulty mixing numerous antimicrobial agents. In many cases, we encountered extremely long set times, ranging from 25 min to over 3 h. Additionally, we experienced antimicrobial– CaSO_4 combinations that set too quickly to effectively mat and fabricate a quality bead product. For these irregular set times, we experimented with alternative mixing methods. We arrived at several mixing techniques to provide reasonable

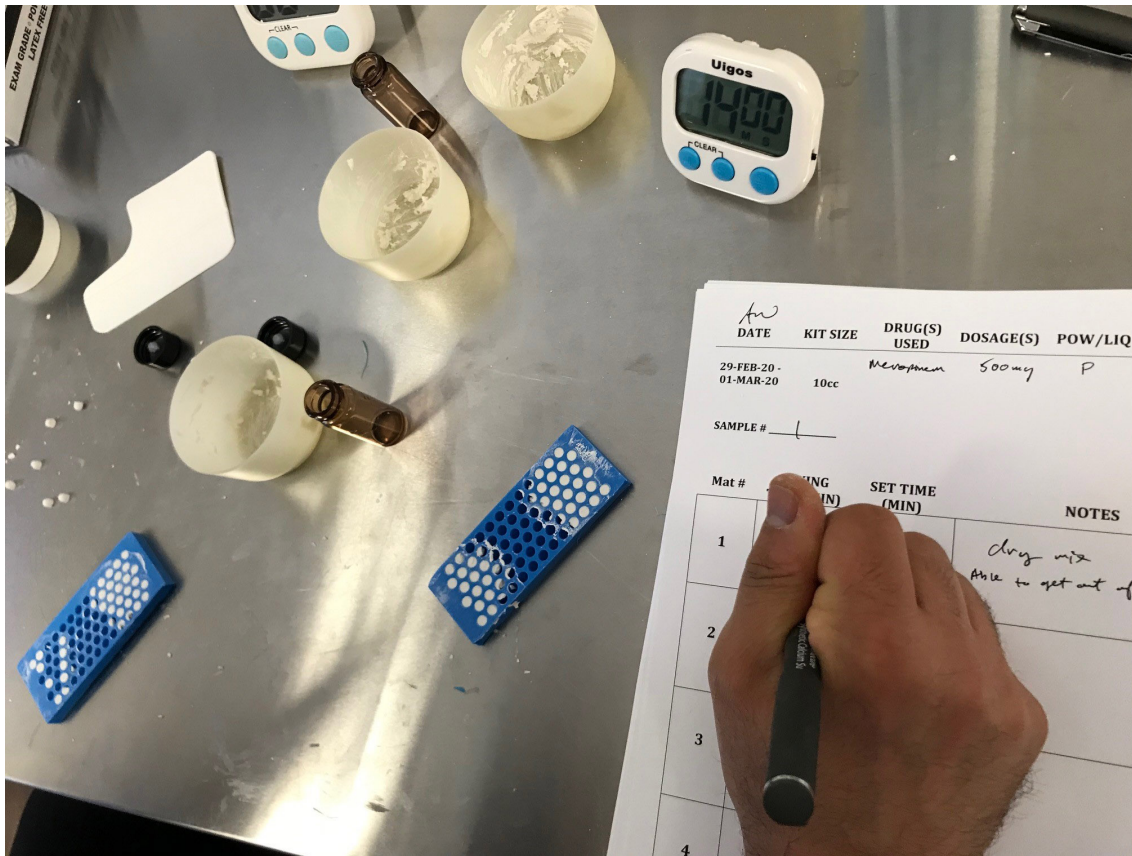


Figure 1. Photograph of 4.8 mm bead mat. The antimicrobial-loaded CaSO_4 paste is shown after spreading the paste into the molds. Once applied, the bead mats were left on the countertop to set, in order to imitate operating room procedure. Time measurements were recorded to establish a set time for each formulation.

set times. These separate methods are annotated in a separate section where the modified mixing instructions are described in each particular instance. The set time was recorded for each modified technique. The group of modified mixing instructions are grouped into the category of “post-mix protocols.”

For each antimicrobial agent or combination tested, we performed the test a minimum of three times (range three–four tests) between the two authors (Edward J. McPherson, Andrew J. Wassef). The mix time and set time for each antimicrobial agent and combination was then averaged for the reported result.

3 Results

The standard mix times are reported in Table 1. The two measured times include the mix time and set time. In the cases where liquid Tobramycin and Gentamicin were substituted for saline, we also report mix time and set time in a similar fashion. For the standard mix method, all tested antimicrobial(s) were set within 4 to 15 min. The recorded mix times

and set times tested between the two testing authors varied no more than 35 s in all agents tested.

The post-mix protocols and described methods are reported in Table 2. For each unique mixing method, we describe the modified mixing technique. The set times were variable, dependent upon the antimicrobial agent(s) added. We will note that we were able to get the CaSO_4 beads to set within a reasonable time with each modified technique. All the recorded mix times and set times between the two testing authors varied no more than 100 s in all agents tested.

4 Discussion

Surgeons face a myriad of obstacles in effectively treating PJI. Amongst the most daunting are microbiota reserves hidden within the interstices of bone known as the cortico-canalicular reserve and the retention of “biofilm islands” attached to avascular soft tissue, bone, and residual small pieces of foreign material(s) (Zoller et al., 2020; de Mesy Bentley et al., 2017; Sindeldecker et al., 2020). Parenteral antibiotics have been a traditionally used adjuvant treatment, combined with implant removal, for PJI but have garnered

Table 1. Set times of Synthecure with various antimicrobials and combinations.

Antibiotic(s)/antifungal	Dose (mg)	Powder (P)/liquid (L)	Avg mix time (min:s)	Avg set time (min:s)	Observations
Control	n/a	n/a	0:30	4:10	
Amikacin	500	P	0:27	5:35	Easy, smooth mix.
Amphotericin B	250	P	0:52	5:58	Longer to mix into a smooth paste. Mats ok.
Cefazolin	1000	P	0:51	4:41	Thicker paste, doughy. Spreads ok into mat.
Clarithromycin	200	P	0:38	4:30	Smooth mix and mat.
Clindamycin	900	P	0:30	7:03	Slightly runny. Wait 2 min to mat.
Fluconazole	400	P	0:36	4:20	Chalky. Spread into mat quickly.
Gentamicin	240	L	0:25	6:15	Runny. Wait 2 min to mat.
Gentamicin	500	P	0:45	5:48	Easy mix, smooth paste.
Meropenem	500	P	0:43	14:07	Dry mix, comes out of mat at 10 min. Sets on table top.
Rifampin	600	P	0:35	4:23	Need to mix and plate fast.
Tobramycin	240	L	0:22	9:30	Runny. Wait 3 min to mat.
Vancomycin	1000	P	0:40	3:58	Coarse, doughy. Be firm with spatula when matting.
Cefazolin/Gentamicin	1000/240	P/L	0:25	6:57	Easy to mix. Slightly runny. Easy to mat.
Cefazolin/Tobramycin	1000/240	P/L	0:25	9:25	Easy mix.
Clindamycin/Tobramycin	900/240	P/L	0:23	13:32	Runny. Wait 2 min to mat.
Vancomycin/Fluconazole	1000/400	P/P	0:38	4:40	Smooth paste. Easy to mat.
Vancomycin/Gentamicin	1000/240	P/L	0:45	7:44	Runny. Wait 2 min to mat.
Vancomycin/Tobramycin	1000/240	P/L	0:45	7:19	Runny. Wait 2 min to mat.
Clindamycin/Fluconazole/ Tobramycin	900/400/240	P/P/L	0:30	14:30	Sets well. Easy to mix.
Vancomycin/Amphotericin B/ Tobramycin	1000/250/240	P/P/L	0:35	8:13	Runny to start, but do not wait to mat.
Vancomycin/Fluconazole/ Tobramycin	1000/400/240	P/P/L	0:32	8:36	Runny to start. Wait 4 min to mat.
Vancomycin/Rifampin/ Tobramycin	1000/600/240	P/P/L	0:45	10:40	Runny to start. Mild increase in volume. Do not wait to mat.

n/a: not applicable.

criticism for their harmful effect to host physiology (Levast et al., 2021). Local antimicrobial delivery into the PJI region, on the other hand, is advantageous for many reasons. First, local delivery can provide substantial antimicrobial levels, high enough to diffuse through the biofilm and enact microbiota kill (Brooks et al., 2021; Moore et al., 2021). Similarly,

high local concentrations allow for diffusion into the small corticocanalicular systems of bone, thus killing microbiota reserves (Maale, 2009). Even in the event of regional devascularization of bone via debridement, microbiota kill can still occur via antimicrobial diffusion. This is salutary in that suspect bone can be treated with antimicrobial agents. Similarly,

Table 2. Synthecure set times that required modified mixing techniques.

Antibiotic(s)/antifungal	Dose (mg)	Powder (P)/liquid (L)	Avg mix time (min:s)	Avg set time (min:s)	Observations
Control	n/a	n/a	0:30	4:10	
Ciprofloxacin	1000	P	1:30	7:25	PMP. Add powder at 60 s. Mix for an additional 30 s.
Ciprofloxacin	500	P	1:30	14:30	PMP. Add powder at 60 s. Mix for an additional 30 s.
Cefepime	500	P	1:30	10:20	PMP. Add powder at 60 s. Mix an additional 30 s.
Cefepime	1000	P	1:50	9:10	PMP. Add powder at 60 s. Mix an additional 50 s.
Ceftriaxone	500	P	0:25	19:20	Add an additional 5 cc of saline at beginning of mix.
Daptomycin	500	P	3:00	12:30	PMP. Add powder at 120 s (paste will be almost set). Mix an additional 60 s.
Ertapenem	500	P	2:15	13:15	PMP. Add powder at 120 s (paste will be almost set). Mix an additional 15 s.
Nafcillin	500	P	1:25	12:20	PMP. Add powder at 60 s. Mix an additional 25 s.
Nafcillin	1000	P	1:25	16:30	PMP. Add powder at 60 s. Mix an additional 25 s.
Unasyn	1500	P	2:20	10:10	PMP. Add powder at 120 s (paste will be almost set). Mix an additional 20 s.
Vancomycin/Ciprofloxacin	1000/1000	P/P	1:25	6:30	PMP. Mix Vanc. and CaSO ₄ . Wait 60 s before adding Cipro. Mix an additional 25 s.
Vancomycin/Tobramycin	1000/1200	P/P	1:45	17:50	PMP. Mix Vanc. and CaSO ₄ . Wait 45 s before adding Tobra. Mix an additional 60 s.
Vancomycin/Zosyn	1000/2250	P/P	2:00	16:15	PMP. Mix Vanc. and CaSO ₄ . Wait 90 s before adding Zosyn. Mix an additional 30 s.
Voriconazole/Vancomycin/Tobramycin	200/1000/240	P/P/L	0:50	22:00	Inject 6 cc of liquid Tobramycin into the vial of Voriconazole. Shake bottle vigorously for 60 s, until all Voriconazole has dissolved. Aspirate 6 cc of liquid from the vial, add to Synthecure + Vancomycin powder, and mix until a smooth paste is achieved.

PMP: post-mix protocol. Mix CaSO₄ with liquid until a smooth paste is achieved. Wait the prescribed time, then add desired antibiotic powder. n/a: not applicable.

the periprosthetic soft tissue envelope partly devascularized by radical debridement can be treated with high levels of antimicrobial agents (Maale et al., 2020). Lastly, local delivery avoids systemic side effects associated with parenteral antibiotics (Levast et al., 2021).

The use of dissolvable antimicrobial-loaded CaSO₄ beads has been utilized for over 3 decades. Its cleared use stems from a process permitting the use of CaSO₄ in the presence of active bone infection dating back to 1997 (Wright Medical Technology, 1997). First-generation products are “mined and refined.” These products are derived from mined gypsum that is subsequently processed to create a medical-grade product that is cleared for insertion in human subjects. Despite their safe use, these products still contain minute amounts of impurities. These minerals include feldspars, dolomite, silica, and corundum (Liu, 2016). These first-generation products have reported variable results and the mineral processing has been thought to be related to side effects, including wound drainage and inflammatory synovitis (Lee et al., 2002). Many companies have since used the above CaSO₄ clearance to create second- and third-generation products that we see today. These products are commercially pure, having undergone manufacturing utilizing pharmaceutical base reagents. The enhanced purity is thought to provide consistency in mixing, dissolution, and application. At present, CaSO₄ is the most common carrier agent that is utilized in local antimicrobial delivery to bone or joint that does not necessitate a second surgical procedure for removal. It is unlikely that new potential carrier systems will enter the market in the near future due to the complicated approval process(es) currently in place.

Our experience in the mixing lab was humbling. First, we encountered antimicrobial–CaSO₄ combinations that set quickly and could not be spread onto the mat fast enough to fabricate quality beads. In cases such as these, we suspect a “hydrophilic steal” phenomenon. Specifically, these antimicrobial powders absorbed H₂O from the saline additive, depriving the H₂O required to convert calcium sulfate hemihydrate to calcium sulfate dihydrate, which is the final product form. To mitigate fast set times, the antimicrobial powder was added later into the mix, such that the dihydrate was allowed to form. In the extreme case of Ceftriaxone, it was necessary to add additional saline in order to lengthen the set time.

On the other hand, we also encountered prolonged set times, some of which required hours to set with the standard mixing method. In these cases, we suspect “stoichiometric interference.” It is our belief that these antimicrobial powders interfered with the seeding and proliferation of calcium sulfate dihydrate. To mitigate such prolonged set times, we again employed a delayed introduction of the antimicrobial powder to achieve calcium sulfate dihydrate crystal formation. In all instances where we encountered initial difficulties, we were able to create a modified mixing technique to produce a quality bead in a timely fashion.

This compendium of mixing formulas is by no means a recommended treatment algorithm(s) for PJI. Local periprosthetic delivery with antimicrobial-loaded CaSO₄ is being used considerably throughout the world. Up to this point, antibiotic and antifungal formulas used with CaSO₄ in the form of insertable beads have been within the purview of surgeon-to-surgeon or surgeon-to-industry representative sidebar discussions. We wanted to bring these discussions to the forefront. We want to educate those physicians treating bone and joint infections that these antimicrobial formulas exist. Those uninformed in this area of treatment could potentially over-treat with adjuvant parenteral antibiotics, thus causing unwanted side effects. In contrast, studies are slowly emerging that show local delivery via CaSO₄ may mitigate the need for adjuvant parenteral antimicrobial therapy (McPherson et al., 2013). The description of these mixing formulas is to serve as a starting basis for discussion while providing a reference point for set times. We note that it is important to leave the beads undisturbed in the bead mat so that the curing process can take place. Disruption of the curing process could cause incomplete curing, which may lead to more rapid resorption in the body. In the future we hope to establish an open-access site such that all physician and surgeons worldwide can report upon mixing formulas that can help other physicians with difficult infection cases. Additionally, we hope to one day see the expansion of mixing formulas that extend beyond the current realm of antibiotic and antifungal chemicals and transition to other non-specific antimicrobial agents.

Ethical statement. This research did not involve human or animal subjects and therefore was exempt from any IRB approval.

Data availability. All of our data are presented in Tables 1 and 2.

Author contributions. EJM, MC, and AJW designed the mixing lab. EJM and AJW carried out the mixing lab. MVD prepared the manuscript with contributions from all co-authors.

Competing interests. The contact author has declared that neither they nor their co-authors have any competing interests.

Disclaimer. Publisher’s note: Copernicus Publications remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Review statement. This paper was edited by Bryan Springer and reviewed by two anonymous referees.

References

- Ambrose, C. G., Clyburn, T. A., Mika, J., Gogola, G. R., Kaplan, H. B., Wanger, A., and Mikos, A. G.: Evaluation of antibiotic-impregnated microspheres for the prevention of implant-associated orthopaedic infections, *J. Bone Joint Surg. Am.*, 96, 128–134, <https://doi.org/10.2106/JBJS.L.01750>, 2014.
- Aul, J. J., Anderson, K. W., Wadowsky, R. M., Doyle, W. J., Kingsley, L. A., Post, J. C., and Ehrlich, G. D.: Comparative evaluation of culture and PCR for the detection and determination of persistence of bacterial strains and DNAs in the Chinchilla laniger model of otitis media, *Ann. Otol. Rhinol. Laryngol.*, 107, 508–513, <https://doi.org/10.1177/000348949810700609>, 1998.
- Brooks, J. R., Dusane, D. H., Moore, K., Gupta, T., Delury, C., Aiken, S. S., Laycock, P. A., Sullivan, A. C., Granger, J. F., Dipane, M. V., McPherson, E. J., and Stoodley, P.: *Pseudomonas aeruginosa* biofilm killing beyond the spacer by antibiotic-loaded calcium sulfate beads: an in vitro study, *J. Bone Joint Infect.*, 6, 119–129, <https://doi.org/10.5194/jbji-6-119-2021>, 2021.
- Costerton, J. W., Stewart, P. S., and Greenberg, E. P.: Bacterial biofilms: a common cause of persistent infections, *Science*, 284, 1318–1322, <https://doi.org/10.1126/science.284.5418.1318>, 1999.
- Costerton, J. W., Montanaro, L., and Arciola, C. R.: Biofilm in implant infections: its production and regulation, *Int. J. Artif. Organs*, 28, 1062–1068, <https://doi.org/10.1177/039139880502801103>, 2005.
- Dacquet, V., Varlet, A., Tandogan, R. N., Tahon, M. M., Fournier, L., Jehl, F., Monteil, H., and Bascoulergue, G.: Antibiotic-impregnated plaster of Paris beads. Trials with teicoplanin, *Clin. Orthop. Relat. Res.*, 282, 241–249, 1992.
- Dahners, L. E. and Funderburk, C. H.: Gentamicin-loaded plaster of Paris as a treatment of experimental osteomyelitis in rabbits, *Clin. Orthop. Relat. Res.*, 219, 278–282, 1987.
- de Mesy Bentley, K. L., Trombetta, R., Nishitani, K., Bello-Irizarry, S. N., Ninomiya, M., Zhang, L., Chung, H. L., McGrath, J. L., Daiss, J. L., Awad, H. A., Kates, S. L., and Schwarz, E. M.: Evidence of Staphylococcus Aureus Deformation, Proliferation, and Migration in Canaliculi of Live Cortical Bone in Murine Models of Osteomyelitis, *J. Bone Miner. Res.*, 32, 985–990, <https://doi.org/10.1002/jbmr.3055>, 2017.
- Evrard, J., Kerri, O., Martini, M., and Conort, O.: Treatment of bone infection by plaster of Paris pellets impregnated with antibiotics, *Pathol. Biol. (Paris)*, 38, 543–547, 1990.
- Ferguson, J. Y., Dudareva, M., Riley, N., D., Stubbs, D., Atkins, B. L., and McNally, M. A.: The use of a biodegradable antibiotic-loaded calcium sulphate carrier containing tobramycin for the treatment of chronic osteomyelitis: a series of 195 cases, *Bone Joint J.*, 96-B, 829–836, <https://doi.org/10.1302/0301-620X.96B6.32756>, 2014.
- Kallala, R., Harris, W. E., Ibrahim, M., Dipane, M., and McPherson, E.: Use of Stimulan absorbable calcium sulphate beads in revision lower limb arthroplasty: Safety profile and complication rates, *Bone Joint Res.*, 7, 570–579, <https://doi.org/10.1302/2046-3758.710.BJR-2017-0319.R1>, 2018.
- Kelly, C. M., Wilkins, R. M., Gitelis, S., Hartjen, C., Watson, J. T., and Kim, P. T.: The use of a surgical grade calcium sulfate as a bone graft substitute: results of a multicenter trial, *Clin. Orthop. Relat. Res.*, 382, 42–50, <https://doi.org/10.1097/00003086-200101000-00008>, 2001.
- Kurtz, S. M., Lau, E., Schmier, J., Ong, K. L., Zhao, K., and Parvizi, J.: Infection burden for hip and knee arthroplasty in the United States, *J. Arthroplasty*, 23, 984–991, <https://doi.org/10.1016/j.arth.2007.10.017>, 2008.
- Kurtz, S. M., Lau, E., Watson, H., Schmier, J. K., and Parvizi, J.: Economic burden of periprosthetic joint infection in the United States, *J. Arthroplasty*, 27, 61–65.e1, <https://doi.org/10.1016/j.arth.2012.02.022>, 2012.
- Lee, G. H., Khoury, J. G., Bell, J.-E., and Buckwalter, J. A.: Adverse reactions to OsteoSet bone graft substitute, the incidence in a consecutive series, *Iowa Orthop. J.*, 22, 35–38, 2002.
- Levast, B., Benech, N., Gasc, C., Batailler, C., Senneville, E., Lustig, S., Poudroux, C., Boutoille, D., Boucinha, L., Dauchy, F.-A., Zeller, V., Maynard, M., Cazanave, C., Thi, T.-T. L., Josse, J., Doré, J., Laurent, F., and Ferry, T.: Impact on the Gut Microbiota of Intensive and Prolonged Antimicrobial Therapy in Patients With Bone and Joint Infection, *Front. Med.*, 8, 586875, <https://doi.org/10.3389/fmed.2021.586875>, 2021.
- Liu, H. (Ed.): *Nanocomposites for Musculoskeletal Tissue Regeneration*, Elsevier, Philadelphia, PA, 2016.
- Lohmander, L. S., Engesaeter, L. B., Herberts, P., Ingvarsson, T., Lucht, U., and Puolakka, T. J. S.: Standardized incidence rates of total hip replacement for primary hip osteoarthritis in the 5 Nordic countries: similarities and differences, *Acta Orthop.*, 77, 733–740, <https://doi.org/10.1080/17453670610012917>, 2006.
- Lütznér, J., Hübel, U., Kirschner, S., Günther, K.-P., and Krummenauer, F.: Long-term results in total knee arthroplasty. A meta-analysis of revision rates and functional outcome, *Chir. Z. Alle Geb. Oper. Medizin*, 82, 618–624, <https://doi.org/10.1007/s00104-010-2001-8>, 2011.
- Maale, G.: The use of antibiotic loaded synthesized calcium sulfate pellets in the one stage treatment for Osteomyelitis, Annual Open Scientific Meeting of the Musculoskeletal Infection Society 2009, 7–8 August 2009, San Diego, CA, 2009.
- Maale, G. E., Eager, J. J., Mohammadi, D. K., and Calderon, F. A.: Elution Profiles of Synthetic CaSO₄ Hemihydrate Beads Loaded with Vancomycin and Tobramycin, *Eur. J. Drug Metab. Pharmacokinet.*, 45, 547–555, <https://doi.org/10.1007/s13318-020-00622-8>, 2020.
- Maradit Kremers, H., Larson, D. R., Crowson, C. S., Kremers, W. K., Washington, R. E., Steiner, C. A., Jiranek, W. A., and Berry, D. J.: Prevalence of Total Hip and Knee Replacement in the United States, *J. Bone Joint Surg. Am.*, 97, 1386–1397, <https://doi.org/10.2106/JBJS.N.01141>, 2015.
- McPherson, E. J., Dipane, M. V., and Sherif, S. M.: Dissolvable Antibiotic Beads in Treatment of Periprosthetic Joint Infection and Revision Arthroplasty: The Use of Synthetic Pure Calcium Sulfate (Stimulan) Impregnated with Vancomycin and Tobramycin, *Reconstructive Rev.*, 3, 32–44, 2013.
- McPherson, E. J., Werner, B. C., Gill, G. K., and Browne, J. A.: Adult Reconstruction, in: *Review of Orthopaedics*, 8th edition, edited by: Miller, M. D. and Thompson, S. R., Elsevier, Philadelphia, PA, 356–437, ISBN-13 978-0323609784, 2020.
- Merx, H., Dreinhöfer, K., Schröder, P., Stürmer, T., Puhl, W., Günther, K.-P., and Brenner, H.: International variation in hip replacement rates, *Ann. Rheum. Dis.*, 62, 222–226, <https://doi.org/10.1136/ard.62.3.222>, 2003.

- Moore, K., Wilson-van Os, R., Dusane, D. H., Brooks, J. R., Delury, C., Aiken, S. S., Laycock, P. A., Sullivan, A. C., Granger, J. F., Dipane, M. V., McPherson, E. J., and Stoodley, P.: Elution Kinetics from Antibiotic-Loaded Calcium Sulfate Beads, Antibiotic-Loaded Polymethacrylate Spacers, and a Powdered Antibiotic Bolus for Surgical Site Infections in a Novel In Vitro Draining Knee Model, *Antibiot. Basel Switz.*, 10, 270, <https://doi.org/10.3390/antibiotics10030270>, 2021.
- Sindeldecker, D., Moore, K., Li, A., Wozniak, D. J., Anderson, M., Dusane, D. H., and Stoodley, P.: Novel Aminoglycoside-Tolerant Phoenix Colony Variants of *Pseudomonas aeruginosa*, *Antimicrob. Agents Chemother.*, 64, e00623-20, <https://doi.org/10.1128/AAC.00623-20>, 2020.
- Tarabichi, M., Shohat, N., Goswami, K., and Parvizi, J.: Can next generation sequencing play a role in detecting pathogens in synovial fluid?, *Bone Jt. J.*, 100-B, 127–133, <https://doi.org/10.1302/0301-620X.100B2.BJJ-2017-0531.R2>, 2018.
- Wright Medical Technology, Inc.: 510(k) Premarket Notification – Wright Plaster of Paris Pellets, Food and Drug Administration, available at: <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfpmn/pmn.cfm?ID=K963562> (last access: 30 August 2021), 1997.
- Ziran, B. H., Smith, W. R., and Morgan, S. J.: Use of calcium-based demineralized bone matrix/allograft for nonunions and posttraumatic reconstruction of the appendicular skeleton: preliminary results and complications, *J. Trauma*, 63, 1324–1328, <https://doi.org/10.1097/01.ta.0000240452.64138.b0>, 2007.
- Zoller, S. D., Hegde, V., Burke, Z. D., C., Park, H. Y., Ishmael, C. R., Blumstein, G. W., Sheppard, W., Hamad, C., Loftin, A. H., Johansen, D. O., Smith, R. A., Sprague, M. M., Hori, K. R., Clarkson, S. J., Borthwell, R., Simon, S. I., Miller, J. F., Nelson, S. D., and Bernthal, N. M.: Evading the host response: *Staphylococcus* “hiding” in cortical bone canalicular system causes increased bacterial burden, *Bone Res.*, 8, 43, <https://doi.org/10.1038/s41413-020-00118-w>, 2020.