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Nickel-Catalyzed Cross-Coupling Reactions:
Stereospecific Arylations, Formation of 2-PyridylZinc Reagents
& Directed Hydroarylation of Alkynes

DISSERTATION

submitted in partial satisfaction of the requirements
for the degree of

DOCTOR OF PHILOSOPHY

in Chemistry

by

Luke Edward Hanna

Dissertation Committee:
Professor Elizabeth R. Jarvo, Chair
Professor Vy M. Dong
Professor Christopher D. Vanderwal

2016

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DEDICATION

To

my parents, family and friends

in recognition of their inspiring character, love and support

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that Professor Jarvo shows her students are what make the Jarvo group an excellent environment to do and learn organic chemistry.

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Tollefson E. J.; Hanna L. E.; Jarvo, E. R.; “Stereospecific Nickel-Catalyzed Cross-Coupling Reactions of Benzylic Ethers and Esters” *Acc. Chem. Res.* **2015**, *48*, 2344.**

Hanna, L. E.; Jarvo E. R. “Selective Cross-Electrophile Coupling By Dual Catalysis” *Angew. Chem. Int. Ed.* **2015**, *54*, 15618.**

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ABSTRACT OF THE DISSERTATION

Nickel-Catalyzed Cross-Coupling Reactions:
Stereospecific Arylations, Formation of 2-PyridylZinc Reagents
& Directed Hydroarylation of Alkynes

by

Luke E. Hanna

Doctor of Philosophy in Chemistry

University of California, Irvine, 2016

Professor Elizabeth R. Jarvo, Chair

Cross-coupling technology has become an indispensable tool for the rapid and efficient synthesis of complex molecules. Over the past few decades a foundational understanding of organometallic chemistry has been laid using palladium and other precious metals. Recent research on first row base metal catalysts such as nickel, cobalt and iron has uncovered new and complementary modes of reactivity compared to their more well-studied precious metal counterparts. While nickel sits one row above palladium on the periodic table, ongoing research has illustrated that nickel possesses a unique reactivity profile. Thus, while nickel is commonly thought of as a cheaper alternative to palladium, research in the field of nickel catalysis has demonstrated far more potential than this. The unique propensity of nickel to undergo single electron chemistry as well as its ability to break strong carbon oxygen bonds make research into nickel reactivity an immensely beneficial endeavor to the fields of inorganic, organometallic and synthetic organic chemistry.

Chapter 1 describes the development of a stereospecific Suzuki coupling of benzylic carbamates and pivalates with aryl- and heteroarylboronic esters. The reaction proceeds with selective inversion or retention at the electrophilic carbon, depending on the identity of the ligand used. Tricyclohexylphosphine ligand provides products with retention of configuration at the electrophilic carbon, while an N-heterocyclic carbene ligand SIMes provides products with inversion.

Chapter 2 discusses the development of a regio- and stereoselective nickel-catalyzed hydroarylation of alkynes using propargylic carbamates as directing groups. The reaction proceeds under mild reaction conditions using arylboronic acids in the absence of base. A range of heterocycles and functional groups are tolerated under the reaction conditions. Additionally, the method is applied to the synthesis of tamoxifen.

Chapter 3 details a nickel-catalyzed cross-electrophile coupling reaction of benzylic esters and aryl halides. Both inter- and intramolecular variants proceed under mild reaction conditions. A range of heterocycles and functional groups are tolerated under the reaction conditions. Additionally, the first example of a stereospecific cross-electrophile coupling of a secondary benzylic ester is described.

Chapter 4 presents secondary benzylzinc reagents generated from 2-pyridylcarbinols using a nickel catalyst and diethylzinc. Substrates are activated in situ using a chlorophosphate reagent. Quenching the organozinc reagents allows for facile deoxygenation of 2-pyridylcarbinols in a one-pot reaction with straightforward incorporation of a deuterium label from deuteromethanol. An intramolecular conjugate addition of a secondary benzylzinc reagent with an α,β -unsaturated ester is also demonstrated.

Chapter 1

Nickel-Catalyzed Stereospecific Suzuki Cross-Coupling:

A Novel Approach to Optically Enriched Triarylmethanes

1.1 Introduction

Triarylmethanes are a diverse class of compounds, from their early uses as precursors to dyes, their utility has since expanded to include a range of chemical and biological applications (Figure 1.1).¹ For example, triarylmethanes have been used as compounds for live cell imaging, selective sensors for metal ions, anticancer and antitubercular agents, potassium ion channel blockers, and even have applications to material science.^{1,2} Hence, the synthesis of triarylmethanes is a rich and growing area of research. Given the wide ranging applications of triarylmethanes, many methods have emerged to access racemic mixtures;³ however, methods that address their asymmetric synthesis have notably been limited.⁴ In this Chapter, an account of the development of a stereospecific nickel-catalyzed Suzuki cross-coupling that can access a library of enantioenriched triarylmethanes is described.

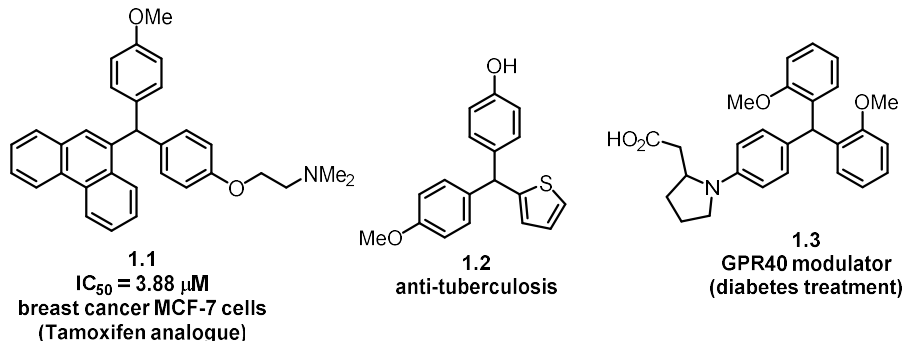
¹ (a) Nambo M.; Crudden C. M. *ACS Catal.* **2015**, *5*, 4734. (b) Mondal, S.; Panda, G. *RSC Adv.*, **2014**, *4*, 28317. Biological activity: (c) Palchauthuri, R.; Nesterenko, V.; Hergenrother, P. J. *J. Am. Chem. Soc.* **2008**, *130*, 10274. (d) Shagufta; Srivastava, A. K.; Sharma, R.; Mishra, R.; Balapure, A. K.; Murthy, P. S. R.; Panda, G. *Bioorg. Med. Chem.* **2006**, *14*, 1497. (e) Parai, M. K.; Panda, G.; Chaturvedi, V.; Manju, Y. K.; Sinha, S. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 289. (f) Ellsworth, B. A.; Ewing, W. R.; Jurica, E. U.S. Patent Application 2011/0082165A1, Apr 7, **2011**. (g) Baba, K.; Maeda, K.; Tabata, Y.; Doi, M.; Kozawa, M. *Chem. Pharm. Bull.* **1988**, *36*, 2977. Materials: (h) Herron, N.; Johansson, G. A.; Radu, N. S. US Patent Application 2005/0187364, Aug 25, **2005**. (i) Xu, Y.-Q.; Lu, J.-M.; Li, N.-J.; Yan, F.; Xia, X.; Xu, Q. *Eur. Polym. J.* **2008**, *44*, 2404.

² Physical properties of triarylmethanes: (a) Breslow, R.; Chu, W. *J. Am. Chem. Soc.* **1973**, *95*, 411. (b) Finocchiaro, P.; Gust, D.; Mislow, K. *J. Am. Chem. Soc.* **1974**, *96*, 3198. (c) Duxbury, D. F. *Chem. Rev.* **1993**, *93*, 381.

³ (a) Zhang, J.; Bellomo, A.; Creamer, A. D.; Dreher, S. D.; Walsh, P. J. *J. Am. Chem. Soc.* **2012**, *134*, 13765. (b) Yu, J.-Y.; Kuwano, R. *Org. Lett.* **2008**, *10*, 973. (c) Molander, G.A.; Elia, M. D. *J. Org. Chem.* **2006**, *71*, 9198. (d) Li, Y.-Z.; Li, B.-J.; Lu, X.-Y.; Lin, S.; Shi, Z.-J. *Angew. Chem. Int. Ed.* **2009**, *48*, 3817. (e) Iovel, I.; Mertins, K.; Kischel, J.; Zapf, A.; Beller, M. *Angew. Chem. Int. Ed.* **2005**, *44*, 3913. (f) For a representative Freidel–Crafts strategy, see: Esquivias, J.; Arrayás, R. G.; Carretero, J. C. *Angew. Chem. Int. Ed.* **2006**, *45*, 629.

⁴ (a) Shi, B.-F.; Mangel, N.; Zhang, Y.-H.; Yu, J.-Q. *Angew. Chem. Int. Ed.* **2008**, *47*, 4882. (b) Sun, F.-L.; Zheng, X.-J.; Gu, Q.; He, Q.-L.; You, S.-L. *Eur. J. Org. Chem.* **2010**, 47.

Figure 1.1. Examples of biologically active triarylmethanes



The mechanisms of alkyl cross-coupling reactions are hardwired with implications for the stereochemical outcome at the reactive centers.⁵ Simple changes to the reaction conditions do not typically perturb the inherent bias for racemization, retention, or inversion at the reactive centers. For example, palladium-catalyzed reactions of alkyl electrophiles are typically stereospecific and proceed with inversion at the stereogenic center,^{6,7} while nickel-catalyzed reactions of alkyl halides proceed with racemization at the electrophilic carbon⁸ and judicious use of a chiral catalyst permits stereoconvergent reactions.⁹ Overcoming the intrinsic preference, such that a reaction that typically proceeds with inversion at the stereogenic center can proceed with retention is quite unusual, and requires a significant change to the mechanism of the transformation. For stereospecific reactions, special cases using α -chiral *transmetallating agents* have been reported where modification of reaction conditions or substrate structure can effect a switch in the sense of

⁵ Jana, R.; Pathak, T. P.; Sigman, M. S. *Chem. Rev.* **2011**, *111*, 1417.

⁶ (a) Lau, K. S. Y.; Fries, R. W.; Stille, J. K. *J. Am. Chem. Soc.* **1974**, *96*, 4983. (b) Netherton, M. R.; Fu, G. C. *Angew. Chem. Int. Ed.* **2002**, *41*, 3910. (c) Legros, J.-Y.; Toffano, M.; Fiaud, J.-C. *Tetrahedron* **1995**, *51*, 3235. (d) Rodriguez, N.; de Arellano, C. R.; Asensio, G.; Medio-Simon, M. *Chem. Eur. J.* **2007**, *13*, 4223. (e) Lopez-Perez, A.; Adrio, J.; Carretero, J. C. *Org. Lett.* **2009**, *11*, 5514. (f) He, A.; Falck, J. R. *J. Am. Chem. Soc.* **2010**, *132*, 2524. (g) Rudolph, A.; Rackelmann, N.; Lautens, M. *Angew. Chem. Int. Ed.* **2007**, *46*, 1485.

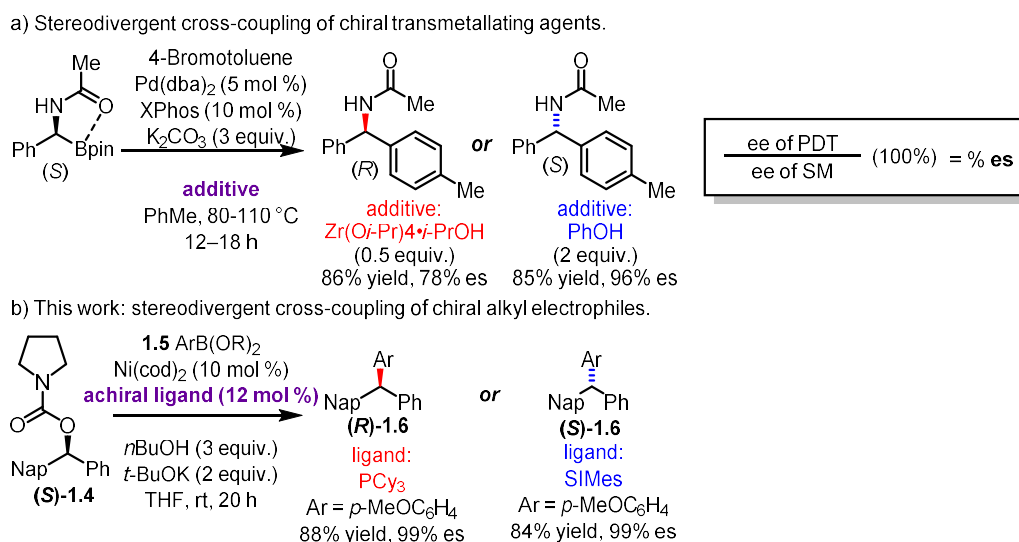
⁷ Pd-catalyzed *allylic* substitutions can occur with inversion or retention, depending on the nucleophile. See: Trost, B. M.; VanVranken, D. L. *Chem. Rev.* **1996**, *96*, 395.

⁸ Stille, J. K.; Cowell, A. B. *J. Organomet. Chem.* **1977**, *124*, 253.

⁹ (a) Saito, B.; Fu, G. C. *J. Am. Chem. Soc.* **2008**, *130*, 6694. (b) Wilsily, A.; Tramutola, F.; Owston, N. A.; Fu, G. C. *J. Am. Chem. Soc.* **2012**, *134*, 5794, and cited therein. (c) Glorius, F. *Angew. Chem. Int. Ed.* **2008**, *47*, 8347.

absolute configuration.¹⁰ Transmetalation typically occurs with retention at the stereogenic center,^{11,12} select examples that proceed with inversion have been reported.¹³ In seminal contributions, Hiyama demonstrated that palladium-catalyzed couplings of alkylsilanes could proceed with retention or inversion, depending on the reaction conditions.^{14,15} Recently, the Suginome group has developed stereodivergent reactions of α -(acetylamino)benzylboronic esters that are controlled by choice of additive to afford, selectively, either retention or inversion (Scheme 1.1a).

Scheme 1.1. Control of product stereochemistry in stereospecific reactions



¹⁰ For a discussion, see: Molander, G. A.; Wisniewski, S. R. *J. Am. Chem. Soc.* **2012**, *134*, 16856.

¹¹ For labeling studies, see: (a) Bock, P. L.; Boschetto, D. J.; Rasmussen, J. R.; Demers, J. P.; Whitesides, G. M. *J. Am. Chem. Soc.* **1974**, *96*, 2814. (b) Ridgway, B. H.; Woerpel, K. A. *J. Org. Chem.* **1998**, *63*, 458. (c) Matos, K.; Soderquist, J. A. *J. Org. Chem.* **1998**, *63*, 461. (d) Taylor, B. L. H.; Jarvo, E. R. *J. Org. Chem.* **2011**, *76*, 7573.

¹² (a) Ye, J.; Bhatt, R. K.; Falck, J. R. *J. Am. Chem. Soc.* **1994**, *116*, 1. (b) Hölzer, B.; Hoffmann, R. W. *Chem. Comm.* **2003**, 732. (c) Campos, K. R.; Klapars, A.; Waldman, J. H.; Dormer, P. G.; Chen, C.-Y. *J. Am. Chem. Soc.* **2006**, *128*, 3538. (d) Lange, H.; Fröhlich, R.; Hoppe, D. *Tetrahedron* **2008**, *64*, 9123. (e) Imao, D.; Glasspoole, B. W.; Laberge, S. V.; Crudden, C. M. *J. Am. Chem. Soc.* **2009**, *131*, 5024. (f) Li, H.; He, A.; Falck, J. R.; Liebeskind, L. S. *Org. Lett.* **2011**, *13*, 3682. (g) See reference 5.

¹³ (a) LaBadie, J. W.; Stille, J. K. *J. Am. Chem. Soc.* **1983**, *105*, 669. (b) Kells, K. W.; Chong, J. M. *J. Am. Chem. Soc.* **2004**, *126*, 15666. (c) Sandrock, D. L.; Jean-Gérard, L.; Chen, C.-Y.; Dreher, S. D.; Molander, G. A. *J. Am. Chem. Soc.* **2010**, *132*, 17108. (d) Ohmura, T.; Awano, T.; Suginome, M. *J. Am. Chem. Soc.* **2010**, *132*, 13191. (e) Lee, J. C. H.; McDonald, R.; Hall, D. G. *Nat. Chem.* **2011**, *3*, 894.

¹⁴ (a) Hatanaka, Y.; Hiyama, T. *J. Am. Chem. Soc.* **1990**, *112*, 7793. (b) Hiyama, T. *J. Organomet. Chem.* **2002**, 653, 58.

¹⁵ For enantiodivergent reactions of alkyl lithium reagents: Stymiest, J. L.; Bagutski, V.; French, R. M.; Aggarwal, V. M. *Nature*, **2008**, 456, 778.

In this Chapter, we demonstrate catalyst control of the stereochemical course with respect to the *electrophilic* partner in a cross-coupling reaction. Stereospecific nickel-catalyzed cross-coupling reactions of benzylic alcohol derivatives typically proceed with inversion at the electrophilic carbon.^{16,17} Nickel-catalyzed cross-coupling of benzylic esters where the achiral ligand structure dictates if the reaction proceeds with retention or inversion (Scheme 1.1b). Use of SIMes, an N-heterocyclic carbene (NHC) ligand, affords inversion, while PCy₃ gives retention. To the best of our knowledge, these results constitute the first cross-coupling reactions of alkyl electrophiles that proceed via two distinct stereospecific mechanistic pathways to provide either retention or inversion at the electrophilic carbon.

In previous work, we established the synthesis of enantioenriched triarylmethanes by stereospecific nickel-catalyzed cross-coupling of ethers with aryl Grignard reagents.^{16b} As part of our ongoing interest in developing nickel-catalyzed stereospecific reactions of alkyl electrophiles, we chose to examine cross-coupling reactions of arylboronic esters for triarylmethane synthesis. The functional group tolerance and availability of a wide range of boronic esters makes them attractive coupling partners.

1.2 Development of a Stereospecific Cross-Coupling Reaction of Arylboronic Esters

We began by examining a range of benzylic alcohol derivatives (Table 1.1). Our initial reaction conditions resulted in a modest conversion of carbonate (**S**)-**1.4** and low enantiospecificity (es) (entry 1).¹⁸ To our surprise, in contrast to the Kumada coupling, the product, (**R**)-**1.6**, results

¹⁶ (a) Taylor, B. L. H.; Swift, E. C.; Waetzig, J. D.; Jarvo, E. R. *J. Am. Chem. Soc.* **2011**, *133*, 389. (b) Taylor, B. L. H.; Harris, M. R.; Jarvo, E. R. *Angew. Chem. Int. Ed.* **2012**, *51*, 7790. (c) Greene, M. A.; Yonova, I. M.; Williams, F. J.; Jarvo, E. R. *Org. Lett.* **2012**, *14*, 4293. (d) For a review, see: Taylor, B. L. H.; Jarvo, E. R. *Synlett*, **2011**, *19*, 2761.

¹⁷ For recent studies of the stereochemical course of nickel-catalyzed reactions of epoxides and aziridines, see: (a) Beaver, M. G.; Jamison, T. F. *Org. Lett.* **2011**, *13*, 4140. (b) Sylvester, K. T.; Wu, K.; Doyle, A. G. *J. Am. Chem. Soc.* **2012**, *134*, 9541. (c) Lin, B. L.; Clough, C. R.; Hillhouse, G. L. *J. Am. Chem. Soc.* **2002**, *124*, 2890.

¹⁸ es: Denmark, S. E.; Vogler, T. *Chem.–Eur. J.* **2009**, *15*, 11737.

from *retention* at the electrophilic carbon. An improvement to 43% es was observed when the solvent was changed from toluene to THF (entry 2). Alcohol additives further improved the yield and stereochemical fidelity of the reaction, with *n*-BuOH providing the highest es, 87% (entry 4). More sterically encumbered alcohols provided more modest improvements, while water and the electron-deficient alcohol trifluoroethanol proved detrimental to the reaction (entries 3, 5, and 7). The enantiospecificity of the reaction showed a marked dependence on the identity of the leaving group. While the use of pivalate (**(S)**-1.8 in the cross-coupling reaction resulted in lower enantiomeric excess of the product (entry 8), the benzoate and carbamate derivatives (**(S)**-1.9 and (**(S)**-1.4 showed a significant increase in product ee, providing 91 and 95% es, respectively (entries 8, 10, and 12). An additional small improvement in yield and es resulted from using a 1:1 mixture of THF/toluene as the solvent (c.f. entries 12 and 15).

Table 1.1. Optimization of Stereospecific Suzuki Cross-coupling

Entry	R	ligand ^a	solvent	additive	% yield ^b	es ^c	retention/ inversion	Entry	R	ligand ^a	solvent	additive	% yield ^b	es ^c	retention/ inversion
1		PCy ₃	PhMe	none	46	7	retention	10		PCy ₃	THF	<i>n</i> -BuOH	57	91	retention
2		PCy ₃	THF	none	53	43	retention	11		SIMes	THF	<i>n</i> -BuOH	83	>99	inversion
3		PCy ₃	THF	H ₂ O	74	10	retention	12		PCy ₃	THF	<i>n</i> -BuOH	62	95	retention
4		PCy ₃	THF	<i>n</i> -BuOH	76	87	retention	13		PCy ₃	THF/PhMe	none	67	35	retention
5		PCy ₃	THF	<i>i</i> -PrOH	46	78	retention	14		SIMes	THF/PhMe	none	82	92	inversion
6		PCy ₃	THF	<i>t</i> -BuOH	55	43	retention	15		PCy ₃	THF/PhMe	<i>n</i> -BuOH	88	99	retention
7		PCy ₃	THF	F ₃ CCH ₂ OH	< 5	na	retention	16		SIMes	THF/PhMe	<i>n</i> -BuOH	84	99	inversion
8		PCy ₃	THF	<i>n</i> -BuOH	53	76	retention	^a PCy ₃ (20 mol %), SIMes (11 mol %). ^b Isolated yield after column chromatography. ^c Enantiospecificity (es) = ee _{product} /ee _{starting material} × 100%.							
9		SIMes	THF	<i>n</i> -BuOH	60	77	inversion								

We examined other ligands¹⁹ under the reaction conditions and found that the NHC ligand SIMes²⁰ afforded comparable yields and enantiospecificity of **1.6**, however, the major product was the (*S*)-enantiomer, resulting from *inversion* at the electrophilic carbon.²¹ Catalyst-control of the stereochemical outcome of the reaction was consistent across the range of esters and carbamates that we examined: PCy₃ and SIMes reliably afforded opposite enantiomers of product (entries 8–11, 15 and 16).²² Under the optimal reaction conditions, addition of *n*-BuOH was found to improve stereochemical fidelity when using either ligand (c.f. entries 13–16).

1.3 Investigation of Scope in Aryl Boronic Ester

Having optimized reaction conditions for the stereospecific synthesis of either enantiomer of product, we turned our attention to the scope of the reaction with respect to the boronic ester (Table 1.2). Electron-donating and withdrawing substituents on the arylboronic ester are well tolerated under the reaction conditions (entries 1–6). Reaction conditions are mild and allow for broad functional group tolerance. Boronic esters containing ketone, free alcohol and carbamate functional groups all undergo cross-coupling in good yield and es (entries 9–13). Heterocyclic boronic esters including pyrimidine, furan, and indole undergo smooth cross-coupling (entries 14–19). The reaction conditions developed for the formation of either enantiomer of **1.6** are general across the range of boronic esters that we examined: of 20 examples, 18 provide high es. Therefore, by choosing the appropriate ligand, PCy₃ or SIMes, either enantiomer of a given product can be obtained from the same enantiomer of starting material.

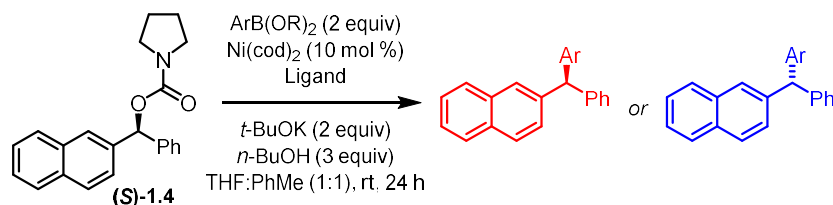
¹⁹ For results with other ligands, see the Experimental Details (table 1.4)

²⁰ SIMes = 1,3-Bis(2,6-diisopropylphenyl)-4,5-di-hydroimidazoliumtetrafluoroborate

²¹ Comparison of NHC to PR₃: (a)Clavier, H.; Nolan, S. P. *Chem. Commun.* **2010**, *46*, 841. (b)Dorta, R., Stevens, E. D., Scott, N. M., Costabile, C., Cavallo, L., Hoff, C. D., Nolan, S. P. *J. Am. Chem. Soc.* **2005**, *127*,2485. (c)Kelly, R. A., III; Clavier, H.; Giudice, S.; Scott, N. M.; Stevens, E. D.; Bordner, J.; Samardjiev, I.; Hoff, C. D.; Cavallo, L.; Nolan, S. P. *Organometallics* **2008**, *27*, 202.

²² Changing PCy₃ loading from 20 mol % to 11 mol % does not affect the stereochemical outcome; see the Experimental Details.

Table 1.2. Investigation of Scope with respect to Aryl Boronic Ester^a



Entry	Ar	ligand ^b	yield (%) ^c	SM ee (%) ^d	product ee (%) ^d	es (%)	retention/inversion	Entry	Ar	ligand ^b	yield (%) ^c	SM ee (%) ^d	product ee (%) ^d	es (%)	retention/inversion		
1	R' = OMe	PCy ₃	88	93	92	98	retention	11		R' = CH ₂ OH	PCy ₃	67	93	82	88	retention	
2		OMe	SIMes	84	93	93	>99	inversion	12		CH ₂ NHBoc	PCy ₃	84	93	91	98	retention
3		NMe ₂	PCy ₃	86	93	92	99	retention	13		CH ₂ NHBoc	SIMes	84	98	95	97	inversion
4		NMe ₂	SIMes	71	93	92	98	inversion	14			PCy ₃	86	93	89	96	retention
5		F	PCy ₃	82	93	90	97	retention	15 ^d			SIMes	75	98	92	94	inversion
6		F	SIMes	80	97	88	91	inversion	16			PCy ₃	79	93	94	>99	retention
7		CF ₃	PCy ₃	88	97	57	59	retention	17			SIMes	65	98	82	84	inversion
8		CF ₃	SIMes	70	93	91	98	inversion	18			PCy ₃	90	93	93	99	retention
9		COMe	PCy ₃	76	93	89	96	retention	19			SIMes	71	93	92	98	inversion
10		COMe	SIMes	99	98	97	99	inversion									

^aAll data are average of two experiments unless otherwise indicated. ^bPCy₃ (20 mol %), SIMes (11 mol %).

^cIsolated yield after column chromatography. ^dDetermined by chiral SFC chromatography. ^eData obtained from a single experiment.

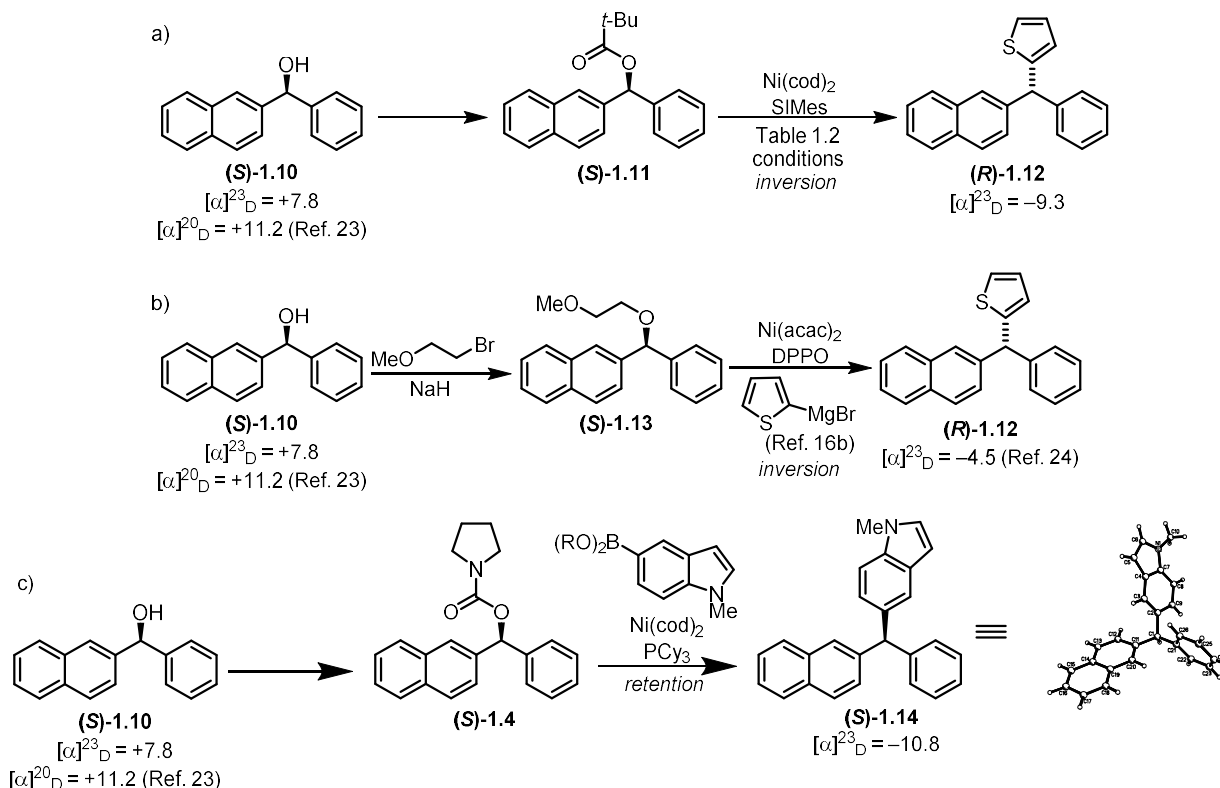
1.4 Stereochemical Course of the Cross-Coupling Reaction

To demonstrate the stereochemical course of the reaction with either catalyst, we utilized X-ray crystallographic analysis. We first showed that the reaction proceeds with inversion when SIMes is used as the ligand by comparison of the optical rotation of the triarylmethane (**R**)-**1.12** with the literature value (Scheme 1.2a).^{23,24} This product corresponds to net inversion at the benzylic carbon during the Suzuki–Miyaura cross-coupling reaction. Previous work in our group demonstrated that cross-coupling of aryl Grignard reagents and benzylic ethers also results in inversion at the benzylic carbon and assigned the absolute configuration of (**R**)-**1.12** based on X-ray crystallographic analysis. The stereochemical course of the Suzuki–Miyaura cross-coupling

²³ For optical rotation data for **1.10**, see: (a) Yamamoto, Y.; Kurihara, K.; Miyaura, N. *Angew. Chem. Int. Ed.* **2009**, *48*, 4414. (b) Shannon, J.; Bernier, D.; Rawson, D.; Woodward, S. *Chem. Commun.* **2007**, 3945. (c) Tjosaas, F.; Anthonsen, T.; Jacobsen, E. E. *ARKIVOC* **2008**, (6), 8190.

²⁴ For characterization data for **1.12** including optical rotation and X-ray crystallographic data, see: Taylor, B. L. H.; Harris, M. R.; Jarvo, E. R. *Angew. Chem., Int. Ed.* **2012**, *51*, 7790.

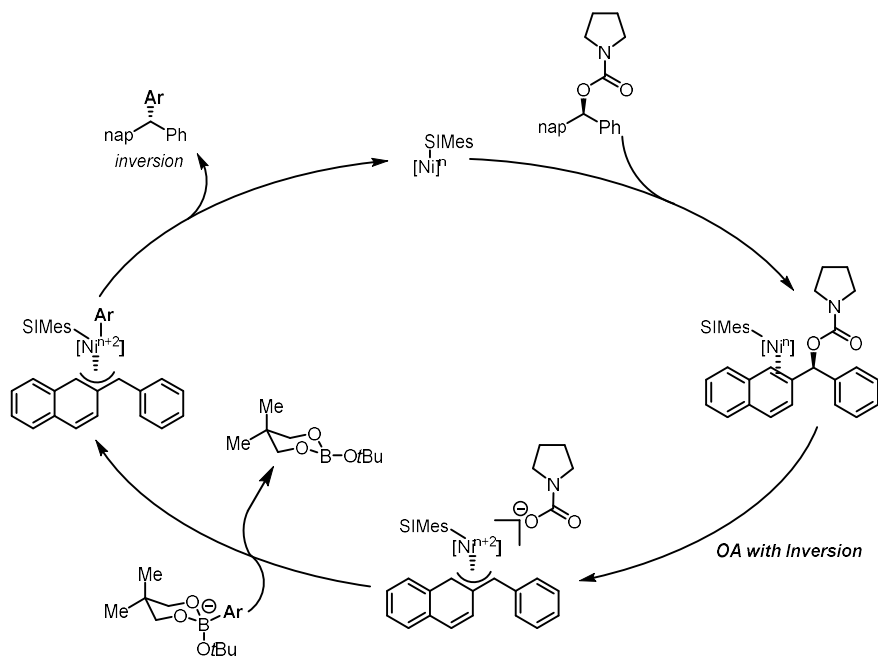
reaction when PCy₃ is used as the ligand provides the opposite enantiomer. Based on the absolute configuration of **(S)**-1.14, the reaction proceeds with retention at the benzylic carbon (Scheme 1.2c). **Scheme 1.2.** Demonstration of stereochemical course of cross-coupling reaction



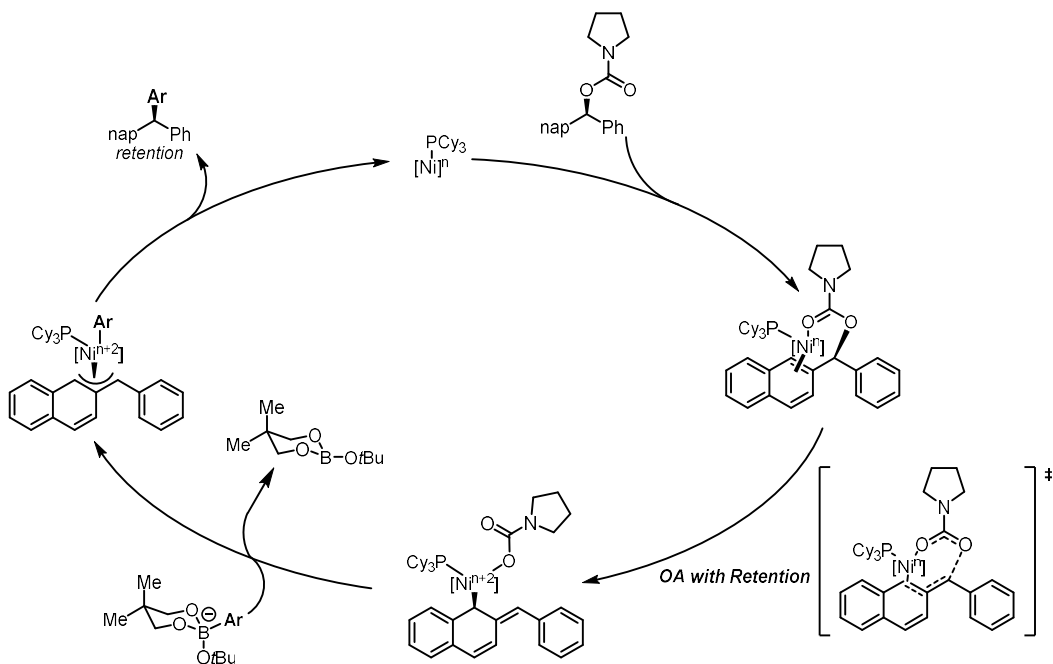
A mechanistic model for obtaining either product of retention or inversion is shown in Scheme 1.3b. We propose that when PCy₃ ligand is employed, the carbamate leaving group coordinates to the nickel catalyst. This coordination event directs the oxidative addition of the catalyst to the substrate, resulting in oxidative addition with retention. Transmetalation followed by reductive elimination with retention leads to formation of product with overall retention. In contrast, when SIMes is used as the ligand, coordination of the carbamate to the catalyst prior to oxidative addition does not occur. Without precoordination of the nickel catalyst to the leaving group, oxidative addition proceeds with inversion, ultimately affording triarylmethane products with overall inversion.

Scheme 1.3. Current mechanistic model for stereodivergent pathways using PCy₃ or SiMes

a) Catalytic cycle for Ni-SiMes catalyst; oxidative addition with *inversion* of configuration



b) Catalytic cycle for Ni-PCy₃ catalyst; oxidative addition with *retention* of configuration



1.5 Scope of the Oxidative Addition Partner

We set as our goal the cross-coupling of oxidative addition partners that do not include a naphthalene moiety. Electrophiles that only have simple benzene rings are typically less reactive in cross-coupling reactions,^{16c} and were not competent for triarylmethane synthesis via Kumada coupling.^{16b} Indeed, neither the corresponding carbamates nor the use of PCy₃ as ligand provide acceptable yields of product. However, benzhydryl pivalates undergo smooth cross-coupling under our optimized reaction conditions when SIMes is used as the ligand (Table 1.3). Efficient cross-coupling was achieved for pivalates with a range of arylboronic esters, including an indoleboronic ester (entries 1–3). Heterocyclic functionality was also tolerated on the electrophile: furan- and benzodioxane- substituted pivalates couple in good yield and excellent es (entries 4 and 5).

Table 1.3. Scope of Substrates Bearing Non-extended Arenes^a

Entry	R'	Ar	yield (%) ^b	SM ee (%) ^c	product ee (%) ^c	es (%)
1	Ph	<i>p</i> -MeOC ₆ H ₄	85	96	84	88
2	Ph	<i>p</i> -(Me ₂ N)C ₆ H ₄	75	82	79	96

3	Ph		66	96	96	>99
4			80	93	87	94
5			60	93	93	99

^aAll data are average of two experiments. ^bIsolated yield after column chromatography. ^cDetermined by chiral SFC chromatography.

1.6 Conclusions

In summary, we have developed a nickel-catalyzed Suzuki-Miyaura cross-coupling reaction for the synthesis of enantioenriched triarylmethanes. Reactions proceed with high stereochemical fidelity. Achiral ligand identity controls if the reaction proceeds with inversion or retention at the electrophilic carbon; therefore either enantiomer of product can be formed from a single enantiomer of starting material. This method expands the range of triarylmethanes that may be prepared in enantioenriched form, as simple benzhydryl pivalates and a variety of functionalized arylboronic esters, including heterocyclic compounds, can be used in the reaction.

1.7 Experimental Details

General Procedures

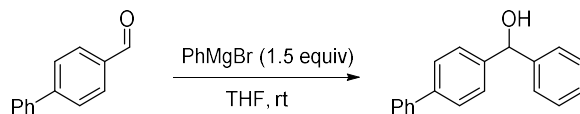
All reactions were carried out under an atmosphere of N₂, or Ar when noted. All glassware was oven- or flame-dried prior to use. Tetrahydrofuran (THF), diethyl ether (Et₂O), dichloromethane (CH₂Cl₂), and toluene (PhMe) were degassed with Ar and then passed through two 4 x 36 inch columns of anhydrous neutral A-2 alumina (8 x 14 mesh; LaRoche Chemicals; activated under a flow of argon at 350 °C for 12 h) to remove H₂O. All other solvents utilized were purchased “anhydrous” commercially, or purified as described. ¹H NMR spectra were recorded on Bruker DRX-400 (400 MHz ¹H, 100 MHz ¹³C, 376.5 MHz ¹⁹F), GN-500 (500 MHz ¹H, 125.7 MHz ¹³C), or CRYO-500 (500 MHz ¹H, 125.7 MHz ¹³C) spectrometers. Proton chemical shifts are reported in ppm (δ) relative to internal tetramethylsilane (TMS, δ 0.00). Data are reported as follows: chemical shift (multiplicity [singlet (s), broad singlet (br s), doublet (d), doublet of doublets (dd), triplet (t), doublet of triplets (dt), doublet of doublet of triplets (ddt), triplet of triplets (tt), quartet (q), multiplet (m)], coupling constants [Hz], integration). Carbon chemical shifts are reported in ppm (δ) relative to TMS with the respective solvent resonance as the internal standard (CDCl₃, δ 77.16 ppm). Unless otherwise indicated, NMR data were collected at 25 °C. Infrared (IR) spectra were obtained on a Perkin-Elmer Spectrum 1000 FT-IR Systems and are reported in terms of frequency of absorption (cm⁻¹). Analytical thin-layer chromatography (TLC) was performed using Silica Gel 60 F₂₅₄ precoated plates (0.25 mm thickness). Visualization was accomplished by irradiation with a UV lamp and/or staining with KMnO₄, ceric ammonium molybdate (CAM), or *p*-anisaldehyde (PAA) solutions. Flash chromatography was performed using Silica Gel 60 (170-400 mesh) from Fisher Scientific. Melting points (m.p.) were obtained using a Mel-Temp melting point apparatus and are uncorrected. Optical rotations were measured on a Rudolph Research

Analytical Autopol IV Automatic Polarimeter. SFC determinations of enantiopurity were performed on a Berger Analytical instrument using a Daicel™ Chiralpak® column (OD-H, OJ-H, or AD-H; 100 bar, 50 °C, 215 nm). High resolution mass spectrometry was performed by the University of California, Irvine Mass Spectrometry Center. Boronic esters were prepared from the corresponding boronic acids and 2,2-dimethylpropane-1,3-diol.²⁵ Boronic acids were generously donated from Frontier, stored at 4 °C, and used as received. 1,8-bis(1,5-cyclooctadiene)nickel was purchased from Strem, stored in a glovebox freezer (−20 °C) under an atmosphere of N₂, and used as received. Tricyclohexylphosphine (PCy₃), (1,3-Bis(2,6-diisopropylphenyl)-4,5-dihydroimidazoliumtetrafluoroborate (SIMes), and tris(di-benzylideneacetone)dipalladium (Pd₂(dba)₃) were purchased from Strem, stored in a glovebox, and used as received. All other reagents were purchased commercially and used as received.

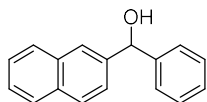
²⁵ Tivola, B. P.; Deagostino, A.; Prandi, C.; Venturello, P. *Org. Lett.* **2002**, *4*, 1275.

Synthesis and Characterization of Substrates

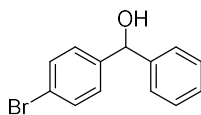
A. Representative procedure for synthesis of diarylmethyl alcohols.



Rac-1.15 In a flame-dried round-bottom flask, to a solution of biphenyl-4-carboxaldehyde (1.04 g, 5.68 mmol, 1.00 equiv) in THF (10 mL) was added phenylmagnesium bromide (0.71 M in THF, 12 mL, 8.5 mmol, 1.5 equiv). After stirring at room temperature for 4 h, saturated ammonium chloride (10 mL) was added and the reaction was extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (3 x 5 mL), dried over MgSO₄, and concentrated in vacuo to afford **rac-1.15** as a white solid (1.2 g, 4.7 mmol, 55%). Analytical data are consistent with the values listed for (**S**)-**1.15** (vide infra).

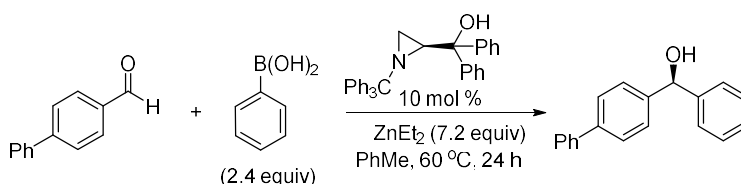


Rac-1.10 Using the representative procedure A outlined above, the following amounts of reagents were used: 2-naphthaldehyde (6.24 g, 40.0 mmol, 1.00 equiv), phenylmagnesium bromide (58 mL, 0.83 M in THF, 48 mmol, 1.2 equiv), and THF (25 mL). The reaction mixture was purified by silica gel flash column chromatography (5–20% EtOAc/hexanes) to afford the product as a white solid (6.74 g, 28.7 mmol, 72%). Analytical data are consistent with the values listed for (**S**)-**1.10** (vide infra).



Rac-1.16 Using the representative procedure A outlined above, the following amounts of reagents were used: 4-bromobenzaldehyde (1.85 g, 10.0 mmol, 1.00 equiv), phenylmagnesium bromide (7.0 mL, 1.7 M in THF, 12 mmol, 1.2 equiv), and THF (10 mL). The crude reaction mixture was purified by flash chromatography (5–20% EtOAc/hexanes) to afford the product as a white solid (1.92 g, 7.29 mmol, 73%). Analytical data is consistent with the values listed below for **(S)-1.16**.

B. Representative procedure for enantioselective synthesis of diarylmethyl alcohols by asymmetric arylation.

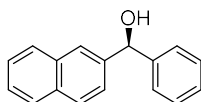


Enantioenriched alcohols were prepared according to a modified procedure of Braga and co-workers.²⁶

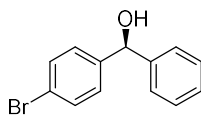
(S)-1.15 To a solution of phenylboronic acid (0.732 g, 6.00 mmol, 2.40 equiv) in toluene (10 mL) was added diethylzinc (18 mL, 18 mmol, 1.0 M in toluene, 7.2 equiv), and the solution was allowed to stir at 60 °C for 12 h. Upon cooling to room temperature, (*S*)-(1-tritylaziridin-2-yl)diphenylmethanol (0.084 g, 0.06 mmol, 0.01 equiv) was added as a solution in toluene (5 mL) and the reaction mixture was allowed to stir for 10 minutes before the addition of a solution of biphenyl-4-carboxaldehyde (0.456 g, 2.50 mmol, 1.00 equiv) in toluene (5 mL). After stirring 12 h at room temperature, 1 N hydrochloric acid (10 mL) was added and the product was extracted

²⁶ Braga, A. R.; Paixao, M. W.; Westeman, B.; Schneider, P. H.; Wessjohan, L.A. *J. Org. Chem.* **2008**, *73*, 2879.

with EtOAc (3 x 10 mL). The combined organics were washed with brine (10 mL), dried over MgSO₄, and concentrated in vacuo. The product was purified by flash column chromatography (0–1% EtOAc/benzene) and then recrystallized from hexanes and EtOAc to upgrade the ee (0.488 g, 1.85 mmol, 75% yield, 96% ee). **TLC** *R_f* = 0.2 (benzene); **m.p.** = 90–92 °C; **¹H NMR** (400 MHz, CDCl₃) δ 7.55 (m, 4H), 7.40 (m, 6H), 7.32 (m, 4H), 5.81 (s, 1H), 2.32 (d, *J* = 2.8, 1H); **¹³C NMR** δ (100 MHz, CDCl₃) δ 143.8, 142.9, 140.9, 140.6, 128.9, 128.7, 127.8, 127.4, 127.38, 127.2, 127.1, 126.7, 76.1; **IR** (neat) 3361, 3029, 1408, 1006, 763 cm⁻¹; **HRMS** (TOF MS ES+) *m/z* calcd for C₁₉H₁₆O (M + Na)⁺ 283.1099, found 283.1110; **[α]²³_D** +4.72 (*c* 1.10, CHCl₃); **SFC** analysis (AD-H, 15% IPA, 3 mL/min) indicated 96% ee: *t_R* (major) = 18.9 minutes, *t_R* (minor) = 20.5 minutes.

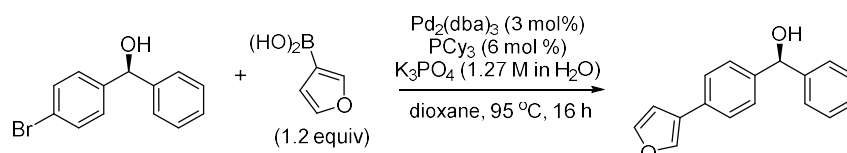


(S)-1.10 Using the representative procedure B outlined above, the following amounts of reagents were used: phenylboronic acid (0.732 g, 6.00 mmol, 2.4 equiv), diethylzinc (18 mL, 18 mmol, 1.0 M in toluene), (*S*)-diphenyl(1-tritylaziridin-2-yl)methanol (116 mg, 0.250 mmol, 0.100 equiv), and 2-naphthaldehyde (0.390 g, 2.50 mmol, 1.00 equiv). The product was purified by flash chromatography (10–20% EtOAc/hexanes) to afford the product as a white solid (0.608 g, 2.59 mmol, 93%, 89% ee). The product was then recrystallized from hexanes to upgrade the ee (99% ee). Analytical data is consistent with literature values. **Error! Bookmark not defined.** **¹H NMR** (400 MHz, CDCl₃) δ 7.91 (s, 1H), 7.82 (dt, *J* = 9.2, 2.6 Hz, 2H), 7.80 (d, *J* = 8.2 Hz, 1H), 7.49–7.40 (m, 5H), 7.35 (t, *J* = 7.0 Hz, 2H), 7.29 (dt, *J* = 7.4, 1.5 Hz, 1H), 6.02 (d, *J* = 3.5 Hz, 1H), 2.29 (d, *J* = 3.5 Hz, 1H); **[α]²³_D** +7.8 (*c* 0.92, CHCl₃), literature **[α]²⁰_D** +11.2 (*c* 0.83, CHCl₃); **SFC** analysis (OD-H, 20% 2-propanol, 3 mL/min) indicated >99% ee: *t_R* (major) = 6.4 min, *t_R* (minor) = 7.3 min.



(S)-1.16 Using the representative procedure B outlined above, the following amounts of reagents were used: phenylboronic acid (0.732 g, 6.00 mmol, 2.4 equiv), diethylzinc (18 mL, 18 mmol, 1.0 M in toluene), (*S*)-(1-tritylaziridin-2-yl)diphenylmethanol (116 mg, 0.250 mmol, 0.100 equiv), and 4-bromobenzaldehyde (0.463 g, 2.50 mmol, 1.00 equiv). The product was purified by flash chromatography (10–20% EtOAc/hexane) to afford the product as a white solid (0.608 g, 2.31 mmol, 93%, 92% ee). The product was then recrystallized from hexanes to yield higher enantiopurity (96% ee). Analytical data is consistent with literature values.²⁷ **¹H NMR** (500 MHz, CDCl₃) δ 7.44 (d, *J* = 8.4 Hz, 2H), 7.36–7.30 (m, 4H), 7.29–7.25 (m, 1H), 7.23 (d, *J* = 8.4 Hz, 2H), 5.76 (d, *J* = 3.3 Hz, 1H), 2.34 (d, *J* = 3.3 Hz, 1H); **¹³C NMR** (125 MHz, CDCl₃) δ 143.5, 142.8, 131.7, 128.8, 128.3, 128.0, 126.6, 121.5, 75.8; [α]_D²³ +17.5 (*c* 1.65, CHCl₃); **SFC** analysis (AD-H, 10% IPA, 2.5 mL/min) indicated 96% ee: *t*_R (major) = 10.4 minutes, *t*_R (minor) = 9.8 minutes.

C. Representative procedure for the Suzuki cross-coupling of aryl bromide (S)-1.17 with aryl boronic acids.

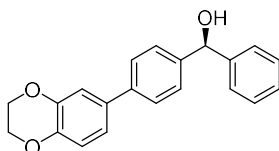


(S)-1.17 The product was prepared according to a modified procedure by Fu and co-workers.²⁸ Tris(dibenzylideneacetone)dipalladium (55 mg, 0.060 mmol, 0.030 equiv) and tricyclohexylphosphine (39 mg, 0.14 mmol, 0.070 equiv) were weighed out into a flame dried two neck, round bottom flask inside a glovebox. The flask was fitted with septa, removed from the

²⁷ Wu, X.; Liu, X.; Zhao, G. *Tetrahedron: Asymmetry* **2005**, *16*, 2299.

²⁸ Kudo, N.; Perseghini, M.; Fu, G. C. *Angew. Chem. Int. Ed.* **2006**, *45*, 1282.

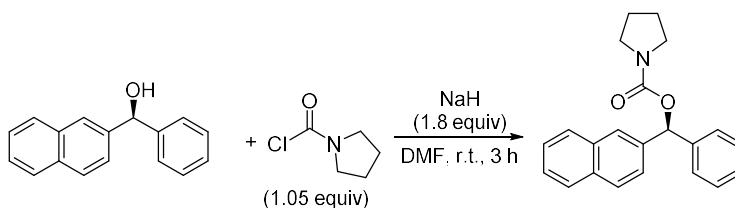
glovebox, and 3-furanboronic acid (0.262 g, 2.20 mmol, 1.10 equiv), **(S)**-**1.16** (0.526 g, 2.00 mmol, 1.00 equiv), aqueous potassium phosphate (2.7 mL, 3.4 mmol, 1.3 M in H₂O, 1.7 equiv) and dioxane (6 mL) were added. The reaction flask was fitted with a reflux condenser and heated to 95 °C for 16 h. After cooling, the solvent was removed under reduced pressure. The resultant residue was purified by flash column chromatography (10–20% EtOAc/hexane) to afford **(S)**-**1.17** as a yellow solid (0.437 g, 1.75 mmol, 87%, 97% ee). **TLC R_f** = 0.2 (4:1 hexane/EtOAc); **m.p.** = 97–99 °C; **¹H NMR** (500 MHz, CDCl₃) δ 7.70 (s, 1H), 7.45 (s, 2H), 7.43 (s, 1H), 7.37 (t, *J* = 8.2 Hz, 3H), 7.33 (t, *J* = 7.3 Hz, 3H), 7.27 (d, *J* = 7.4 Hz, 1H), 6.67 (s, 1H), 5.82 (s, 1H), 2.32 (s, 1H); **¹³C NMR** (125 MHz, CDCl₃) δ 143.9, 143.8, 142.7, 138.6, 131.8, 128.7, 127.8, 127.1, 126.7, 126.2, 126.1, 108.9, 76.1; **IR** (neat) 3279, 1160, 1012, 780, 699 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₁₇H₁₄O₂ (M + Na)⁺ 273.0891, found 273.0883; **[α]_D²⁹** –37.3 (*c* 1.00, CHCl₃); **SFC** analysis (OD-H, 13% IPA, 2.5 mL/min) indicated 97% ee: *t_R* (major) = 12.9 minutes, *t_R* (minor) = 14.7 minutes.



(S)-**1.18** Using representative procedure C outlined above, the following amounts of reagents were used: tris(dibenzylideneacetone)dipalladium (28 mg, 0.030 mmol, 0.030 equiv), tricyclohexylphosphine (20 mg, 0.07 mmol, 0.070 equiv), 1,4-benzodioxane-6-boronic acid (0.198 g, 1.10 mmol, 1.10 equiv), **(S)**-**1.16** (0.263 g, 1.00 mmol, 1.00 equiv), aqueous potassium phosphate (1.4 mL, 1.7 mmol, 1.3 M in H₂O, 1.7 equiv) and dioxane (3 mL). The product was purified by flash column chromatography (10–30% EtOAc/hexane) to afford **(S)**-**1.18** as a brown solid (0.296 g, 0.929 mmol, 93%, 96% ee). **TLC R_f** = 0.2 (4:1 hexane/EtOAc); **TLC R_f** = 0.3 (30% EtOAc/hexanes); **m.p.** = 108–110 °C; **¹H NMR** (500 MHz, CDCl₃) δ 7.48 (d, *J* = 7.7 Hz,

2H), 7.40 (q, $J = 7.8$ Hz, 4H), 7.34 (t, $J = 7.2$ Hz, 2H), 7.27 (d, $J = 7.3$ Hz, 1H), 7.08 (d, $J = 1.9$ Hz, 1H), 7.04 (dd, $J = 8.5, 2.1$ Hz, 1H), 6.90 (d, $J = 8.2$ Hz, 1H), 5.85 (s, 1H), 4.36 (s, 4H), 2.35 (s, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 143.9, 143.8, 143.3, 142.5, 140.0, 134.5, 128.7, 127.7, 127.1, 127.0, 126.6, 120.2, 117.7, 115.9, 76.2, 64.6, 64.5; IR (neat) 3550, 1494, 1304, 1284, 1070 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{21}\text{H}_{18}\text{O}_3$ ($\text{M} + \text{Na}$) $^+$ 341.1154, found 341.1147; $[\alpha]^{29}_{\text{D}} +3.1$ (c 1.04, CHCl_3); SFC analysis (AD-H, 14% IPA, 2.5 mL/min) indicated 96%: t_{R} (major) = 6.9 minutes, t_{R} (minor) = 8.8 minutes.

Preparation of protected carbinols.

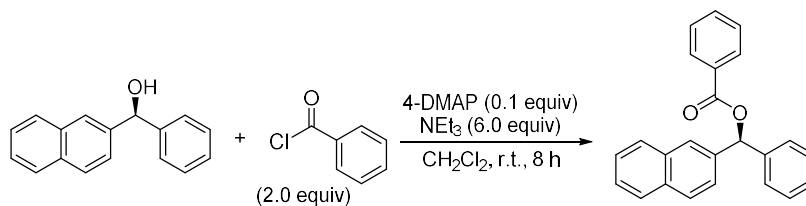


(S)-1.4 The product was prepared according to a modified procedure by Zhang and co-workers.²⁹

To a suspension of NaH (0.153 g, 6.37 mmol, 1.80 equiv) in DMF (3 mL) was added a solution of **(S)-1.10** (0.823 g, 3.54 mmol, 1.00 equiv) in DMF (2 mL) at 0 °C. The mixture was stirred for 1 h before addition of neat 1-pyrrolidinecarbonyl chloride (0.41 mL, 3.7 mmol, 1.1 equiv) at room temperature. After stirring for 3 h, the reaction was quenched with saturated aqueous ammonium chloride (6 mL), and the product was extracted with methylene chloride (3 x 10 mL). The combined organics were washed with brine (5 mL), dried over Na_2SO_4 , and concentrated in vacuo. The product was purified by flash column chromatography (20% EtOAc/hexane) to afford **(S)-1.4** as a white solid (0.963 g, 2.91 mmol, 83%, 94% ee): TLC $R_f = 0.2$ (20% EtOAc/hexanes); m.p. = 151–153 °C; ^1H NMR (500 MHz, CDCl_3) δ 7.84 (s, 1H), 7.82–7.77 (m, 3H), 7.47–7.44 (m, 3H),

²⁹ DeKorver, K. A.; Johnson, W. L.; Zhang, Y.; Hsung, R. P.; Dai, H.; Deng, J.; Lohse, A. G.; Zhang, Y. S. *J. Org. Chem.* **2011**, *76*, 5092.

7.41 (d, $J = 7.3$ Hz, 2H), 7.33 (t, $J = 7.3$ Hz, 2H), 7.27 (d, $J = 7.4$ Hz, 1H), 7.00 (s, 1H), 3.55 (t, $J = 6.7$ Hz, 2H), 3.40 (t, $J = 6.7$ Hz, 2H), 1.90 (dt, $J = 13.3, 6.7$ Hz, 2H), 1.84 (dt, $J = 13.3, 6.7$ Hz, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 154.2, 141.4 138.7, 133.2, 133.0, 128.5, 128.4 128.3, 127.8, 127.7, 127.2, 126.3, 126.2, 126.1, 125.2, 77.4, 46.4, 46.0, 25.9, 25.0; IR (neat) 1690, 1412, 1102, 828, 765 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{22}\text{H}_{21}\text{NO}_2$ ($\text{M} + \text{Na}$) $^+$ 354.1470, found 354.1463; $[\alpha]^{29}_{\text{D}}$ +45.9 (c 1.15, CHCl_3); SFC analysis (OD-H, 18% IPA, 2.5 mL/min) indicated 93% ee: t_{R} (major) = 7.1 minutes, t_{R} (minor) = 6.6 minutes.

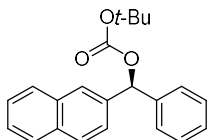


(S)-1.9 The product was prepared according to a modified procedure by Hassner and co-workers.³⁰

To a 25 mL round bottom flask was added alcohol **(S)-1.10** (0.175 g, 0.750 mmol, 1.00 equiv), and 4-(dimethylamino)pyridine (9.0 mg, 0.075 mmol, 0.10 equiv). The flask was evacuated and backfilled with nitrogen before addition of methylene chloride (6 mL), triethylamine (0.48 mL, 4.5 mmol, 6.0 equiv), and benzoyl chloride (0.18 mL, 1.5 mmol, 2.0 equiv). After stirring for 8 h, the reaction was quenched with 1 M HCl (6 mL), and the product was extracted with methylene chloride (3 x 10 mL). The combined organics were washed with brine (10 mL), dried over MgSO_4 , and concentrated in vacuo. The product was purified by flash column chromatography (5–10% EtOAc/hexane) to afford **(S)-1.9** as a white solid (0.177 g, 0.523 mmol, 70%, 89% ee): TLC R_f = 0.4 (10% EtOAc/hexanes); m.p. = 91–93 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.26 (d, $J = 7.9$ Hz, 2H), 8.00 (s, 1H), 7.91–7.90 (m, 3H), 7.66 (t, $J = 7.2$ Hz, 1H), 7.61–7.53 (m, 6H), 7.45 (t, $J = 7.1$

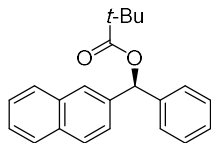
³⁰ Basel, Y.; Hassner, A. *J. Org. Chem.* **2000**, *65*, 6368.

Hz, 2H), 7.39 (d, $J = 7.2$ Hz, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 165.7, 140.3, 137.7, 133.3, 133.2, 133.1, 130.3, 130.0, 128.7, 128.6, 128.3, 128.2, 127.8, 127.4, 126.44, 126.40, 126.3, 125.1, 77.7; IR (neat) 1712, 1259, 1108, 732, 700 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{24}\text{H}_{18}\text{O}_2$ ($\text{M} + \text{Na}$) $^+$ 361.1205, found 361.1201; $[\alpha]_D^{29} +10.0$ (c 0.99, CHCl_3); SFC analysis (OD-H, 10.0% IPA, 2.5 mL/min) indicated 89% ee: t_R (major) = 6.5 minutes, t_R (minor) = 6.3 minutes.



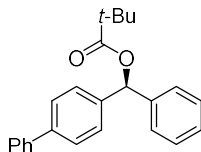
(S)-1.7 The product was prepared according to a modified procedure by Hassner and co-workers.³⁰ To a 25 mL round bottom flask was added alcohol **(S)-1.10** (0.234 g, 1.00 mmol, 1.00 equiv), and 4-(dimethylamino)pyridine (12 mg, 0.010 mmol, 0.10 equiv). The flask was evacuated and backfilled with nitrogen before addition of methylene chloride (8 mL), triethylamine (0.10 mL, 1.2 mmol, 1.2 equiv), and di-*tert*-butyl dicarbonate (0.228 g, 1.05 mmol, 1.05 equiv). After stirring for 8 h, the reaction was quenched with 1 M HCl (6 mL), and the product was extracted with methylene chloride (3 x 10 mL). The combined organics were washed with brine (10 mL), dried over MgSO_4 , and concentrated in vacuo. The product was purified by flash column chromatography (5–10% EtOAc/hexane) to afford **(S)-1.7** as a white solid (0.284 g, 0.849 mmol, 85%, 88% ee): TLC $R_f = 0.4$ (9:1 hexane/EtOAc); m.p. = 90–92 °C; ^1H NMR (500 MHz, CDCl_3) δ 7.87 (s, 1H), 7.83 (d, $J = 7.0$ Hz, 1H), 7.79 (d, $J = 8.2$ Hz, 2H), 7.48–7.45 (m, 2H), 7.43 (s, 1H), 7.41 (d, $J = 8.3$ Hz, 2H), 7.33 (t, $J = 7.3$ Hz, 2H), 7.27 (d, $J = 7.7$ Hz, 1H), 1.47 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 153.1, 140.1, 137.6, 133.2, 133.1, 128.7, 128.5, 128.3, 128.1, 127.8, 127.2, 126.4, 126.3, 126.0, 125.0, 82.7, 80.0, 27.9; IR (neat) 1742, 1270, 1251, 1150, 1081 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{22}\text{H}_{22}\text{O}_3$ ($\text{M} + \text{Na}$) $^+$ 357.1467, found 357.1467; $[\alpha]_D^{29} -19.3$ (c 0.90, CHCl_3); SFC analysis (AD-H, 5% IPA, 3.0 mL/min) indicated 88% ee: t_R (major) = 5.6

minutes, t_R (minor) = 6.1 minutes.

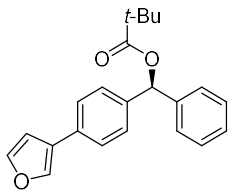


(S)-1.8 The product was prepared according to a modified procedure by Hassner and co-workers.³⁰

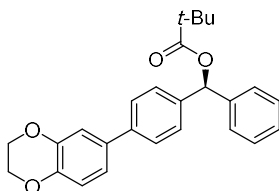
To a 25 mL round bottom flask was added alcohol **(S)-1.10** (0.281 g, 1.20 mmol, 1.20 equiv), and 4-(dimethylamino)pyridine (15 mg, 0.012 mmol, 0.10 equiv). The flask was evacuated and backfilled with nitrogen before addition of methylene chloride (8 mL), triethylamine (0.19 mL, 2.6 mmol, 2.2 equiv), and trimethylacetyl chloride (0.160 mL, 1.26 mmol, 1.05 equiv). After stirring for 8 h, the reaction was quenched with 1M HCl (6 mL), and the product was extracted with methylene chloride (3 x 10 mL). The combined organics were washed with brine (10 mL), dried over MgSO₄, and concentrated in vacuo. The product was purified by flash column chromatography (5–10% EtOAc/hexane) to afford **(S)-1.8** as a white solid (0.334 g, 1.05 mmol, 88%, 82% ee): **TLC** R_f = 0.5 (10% EtOAc/hexanes); **m.p.** = 80–83 °C; **¹H NMR** (500 MHz, CDCl₃) δ 7.82 (s, 2H), 7.79 (d, J = 8.8 Hz, 2H), 7.48–7.45 (m, 2H), 7.42 (d, J = 8.3 Hz, 1H), 7.38 (d, J = 7.6 Hz, 2H), 7.33 (t, J = 7.1 Hz, 2H), 7.28 (d, J = 7.6 Hz, 1H), 1.27 (s, 9H); **¹³C NMR** (125 MHz, CDCl₃) δ 177.4, 140.6, 138.0, 133.2, 133.0, 128.6, 128.5, 128.3, 127.9, 127.8, 127.1, 126.4, 126.3, 126.1, 125.0, 76.8, 39.1, 27.3; **IR** (neat) 1721, 1276, 1148, 1123, 823 cm⁻¹; **HRMS** (TOF MS ES+) m/z calcd for C₂₂H₂₂O₂ (M + Na)⁺ 341.1518, found 341.1526; **$[\alpha]_D^{29}$** –37.4 (c 1.18, CHCl₃); **SFC analysis** (AD-H, 5.0% IPA, 3.0 mL/min) indicated 82% ee: t_R (major) = 6.8 minutes, t_R (minor) = 7.1 minutes.



(S)-1.19 The product was prepared according to a modified procedure by Zhang and co-workers.²⁹ NaH (500 mg, 20.8 mmol, 4.00 equiv) was suspended in 40 mL of dry DMF and cooled to 0 °C. To this solution, alcohol **(S)-1.15** (1.28 g, 4.92 mmol, 1.00 equiv) in dry DMF (10 mL) was added dropwise. The mixture was allowed to stir at 0 °C for 30 minutes after which pivaloyl chloride (4.3 mL, 35 mmol, 7.0 equiv) was added dropwise. The reaction was stirred at 0 °C for 1.5 hours then warmed to room temperature and stirred for 22 hours. The reaction was quenched by consecutive addition of water (5 x 2 mL) and stirring for 3 minutes. The reaction was diluted with more water (10 mL) and the organics were extracted with EtOAc (3 x 30 mL). The combined organic layers were dried over MgSO₄ and the solvent was removed under reduced pressure. The crude was purified by flash chromatography (0–1% Et₂O/petroleum ether) yielding **(S)-1.19** as a white solid (1.56 g, 4.53 mmol, 92%). **TLC** *R_f* = 0.4 (10% Et₂O:petroleum ether); **m.p.** = 109–110 °C; **¹H NMR** (400 MHz, CDCl₃) δ 7.54 (dd, *J* = 6.1, 1.9 Hz, 4H), 7.35 (m, 10H), 6.87 (s, 1H), 1.27 (s, 9H); **¹³C NMR** (100 MHz, CDCl₃) δ 177.4, 140.8, 140.6, 139.8, 128.9, 128.7, 127.9, 127.8, 127.5, 127.40, 127.38, 127.2, 127.0, 76.5, 39.1, 27.3; **IR** (neat) 3029, 2974, 1722, 1275, 1138 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₄H₂₄O₂ (M + Na)⁺ 367.1674, found 367.1681; **[α]_D²³** –23.6 (*c* 1.09, CHCl₃); **SFC** analysis (AD-H, 10% IPA, 3 mL/min) indicated 96% ee: *t_R* (minor) = 4.0 minutes, *t_R* (major) = 6.4 minutes.



(S)-1.20 The product was prepared according to a modified procedure by Zhang and co-workers.²⁹ To a suspension of NaH (72 mg, 3.0 mmol, 2.0 equiv) in DMF (3 mL) was added a solution of **(S)-1.17** (0.374 g, 1.50 mmol, 1.00 equiv) in DMF (2 mL) at 0 °C. The mixture was stirred for 1 h before addition of neat trimethylacetyl chloride (0.200 mL, 1.60 mmol, 1.05 equiv) at room temperature. After stirring for 3 h, the reaction was quenched with saturated aqueous ammonium chloride (6 mL), and the product was extracted with methylene chloride (3 x 10 mL). The combined organics were washed with brine (5 mL), dried over Na₂SO₄, and concentrated in vacuo. The product was purified by flash column chromatography (30% Et₂O/hexane) to afford **(S)-1.20** as a pale yellow solid (0.427 g, 1.28 mmol, 85%, 93% ee): **TLC** *R_f* = 0.2 (4:1 hexane/Et₂O); **m.p.** = 105–108 °C; **¹H NMR** (500 MHz, CDCl₃) δ 7.70 (s, 1H), 7.45 (s, 2H), 7.43 (s, 1H), 7.35 (s, 4H), 7.33 (s, 2H), 7.29–7.26 (m, 1H), 1.26 (s, 9H); **¹³C NMR** (125 MHz, CDCl₃) δ 177.4, 143.8, 140.6, 139.5, 138.7, 132.1, 128.6, 127.9, 127.6, 127.0, 126.2, 126.1, 108.9, 76.5, 39.0, 27.3; **IR** (neat) 1724, 1159, 1138, 757, 699 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₂H₂₂O₃ (M + Na)⁺ 357.1467, found 357.1475; **[α]_D²⁹** -26.0 (c 1.25, CHCl₃); **SFC** analysis (OJ-H, 8% IPA, 2.0 mL/min) indicated 93% ee: *t_R* (major) = 3.6 minutes, *t_R* (minor) = 4.2 minutes.

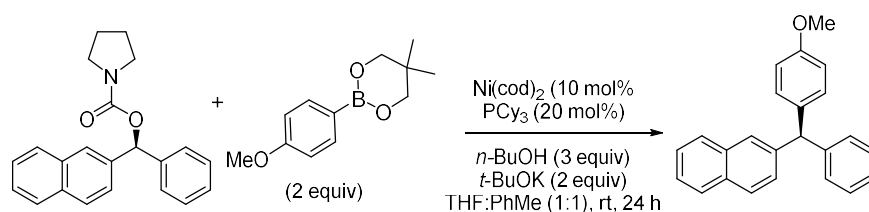


(S)-1.21 The product was prepared according to a modified procedure by Zhang and co-workers.²⁹ To a suspension of NaH (35 mg, 1.4 mmol, 1.8 equiv) in DMF (3 mL) was added a solution of

(S)-1.18 (0.254 g, 0.800 mmol, 1.00 equiv) in DMF (2 mL) at 0 °C. The mixture was stirred for 1 h before addition of neat trimethylacetyl chloride (0.103 mL, 0.840 mmol, 1.05 equiv) at room temperature. After stirring for 3 h, the reaction was quenched with saturated aqueous ammonium chloride (6 mL), and the product was extracted with methylene chloride (3 x 10 mL). The combined organics were washed with brine (5 mL), dried over Na₂SO₄, and concentrated in vacuo. The product was purified by flash column chromatography (30% Et₂O/hexanes) to afford **(S)-1.21** as a tan solid (0.232 g, 0.576 mmol, 73%, 94% ee): **TLC** *R_f* = 0.1 (20% Et₂O/hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.48 (d, *J* = 8.2 Hz, 2H), 7.37–7.32 (m, 6H), 7.28 (d, *J* = 7.2 Hz, 1H), 7.08 (s, 1H), 7.04 (d, *J* = 8.4 Hz, 1H), 6.90 (d, *J* = 8.6 Hz, 1H), 6.84 (s, 1H), 4.27 (s, 4H), 1.27 (s, 9H); **¹³C NMR** (125 MHz, CDCl₃) δ 177.4, 143.8, 143.4, 140.7, 140.2, 139.3, 134.4, 128.7, 127.9, 127.4, 127.0, 126.9, 120.2, 117.7, 115.9, 76.5, 64.6, 64.5, 39.0, 27.3; **IR** (neat) 1723, 1494, 1309, 1147, 1068 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₆H₂₆O₄ (M + Na)⁺ 425.1729, found 425.1715; **[α]_D²⁹** -20.3 (c 0.96, CHCl₃); **SFC** analysis (OD-H, 30% MeOH, 2.5 mL/min) indicated 94% ee: *t_R* (major) = 6.9 minutes, *t_R* (minor) = 8.8 minutes.

Procedures for Cross-Coupling Reactions

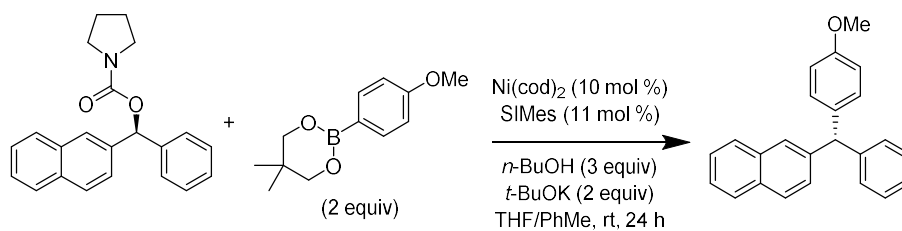
A. Procedure for the synthesis of products with retention (Table 2).



(R)-1.6 To a flame dried vial in a glovebox was added 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), tricyclohexylphosphine (11 mg, 0.040 mmol, 0.20 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μL, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(4-methoxyphenyl)-1,3,2-dioxaborinane (88 mg, 0.40 mmol, 2.0 equiv), **(S)-1.4** (66

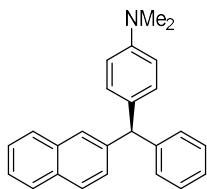
mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The reaction was stirred for 24 hours before removing the vial from the glovebox, opening to atmosphere, and running through a silica gel plug (1:1 Et₂O:hexane). The combined organics were concentrated in vacuo, internal standard (PhTMS, 0.20 mmol) was added and ¹H NMR yield was collected. The product was purified by flash chromatography (1–3% Et₂O/pentane) to afford (**R**)-**1.6** as a colorless oil. First run: (56.0 mg, 0.173 mmol, 86%, 93% ee). Second run: (56.4 mg, 0.174 mmol, 87%, 93% ee). Analytical data is consistent with literature values:^{13b} ¹H NMR (500 MHz, CDCl₃) δ 7.81–7.76 (m, 1H), 7.74 (d, *J* = 8.6 Hz, 1H), 7.72–7.67 (m, 1H), 7.46 (s, 1H), 7.42 (dt, *J* = 9.5, 3.2 Hz, 2H), 7.29 (t, *J* = 7.2 Hz, 3H), 7.24–7.19 (m, 1H), 7.15 (d, *J* = 7.2 Hz, 2H), 7.07 (d, *J* = 8.4 Hz, 2H), 6.83 (d, *J* = 8.8 Hz, 2H), 5.65 (s, 1H), 3.78 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 158.2, 144.1, 142.0, 136.0, 133.5, 132.2, 130.6, 129.6, 128.4, 128.2, 128.0, 127.9, 127.8, 127.6, 126.4, 126.1, 125.7, 113.8, 56.2, 55.3; [α]²³_D –0.77 (*c* 2.70, CHCl₃); SFC analysis (AD-H, 15% IPA, 2.5 mL/min) indicated 93% ee: t_R (major) = 13.9 minutes, t_R (minor) = 13.2 minutes.

B. Procedure for the synthesis of products with inversion of configuration.



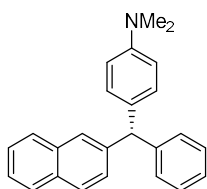
(S)-1.6 To a flame dried vial in a glovebox was added 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-Bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.27 mg, 0.0210 mmol, 0.105 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(4-methoxyphenyl)-1,3,2-dioxaborinane (88 mg, 0.40 mmol, 2.0 equiv), **(S)-1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The reaction was stirred for 24 hours before removing the vial from the glovebox, opening to atmosphere, and running through a silica gel plug (1:1 Et₂O:hexane). The combined organics were concentrated in vacuo, internal standard (PhTMS, 0.20 mmol) was added and ¹H NMR yield was collected. The product was purified by flash chromatography (1–3% Et₂O/pentane) to afford **(S)-1.6** as a colorless oil. First run: (53.2 mg, 0.164 mmol, 82%, 93% ee). Second run: (56.0 mg, 0.173 mmol, 86%, 93% ee). Analytical data is consistent with the values listed above for **(R)-1.6** [α]_D²³ +2.1 (*c* 2.70, CHCl₃); SFC analysis (AD-H, 15% IPA, 2.5 mL/min) indicated 90% ee: t_R (major) = 13.2 minutes, t_R (minor) = 13.9 minutes.

Characterization Data for Products



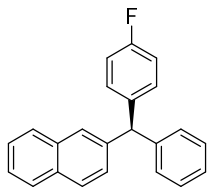
(R)-1.22 Using representative procedure A outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), tricyclohexylphosphine (11 mg, 0.040 mmol, 0.20 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-

butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(4-dimethylaminophenyl)-1,3,2-dioxaborinane (83 mg, 0.40 mmol, 2.0 equiv), (**S**)-**1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash chromatography (1–3% Et₂O/pentane) to afford the product as a colorless oil. First run: (49.8 mg, 0.148 mmol, 80%, 90% ee). Second run: (52.6 mg, 0.167 mmol, 84%, 90% ee). Analytical data is consistent with literature values:^{13b} **¹H NMR** (500 MHz, CDCl₃) δ 7.80–7.75 (m, 1H), 7.73 (d, J = 8.7 Hz, 1H), 7.71–7.67 (m, 1H), 7.48 (s, 1H), 7.40 (dt, J = 9.4, 3.3 Hz, 2H), 7.31 (d, J = 8.4 Hz, 1H), 7.27 (t, J = 7.6 Hz, 2H), 7.19 (t, J = 7.5 Hz, 1H), 7.16 (d, J = 7.5 Hz, 2H), 7.01 (d, J = 8.8 Hz, 2H), 6.67 (d, J = 8.8 Hz, 2H), 5.61 (s, 1H), 2.90 (s, 6H); **¹³C NMR** (125 MHz, CDCl₃) δ 149.2, 144.6, 142.5, 133.6, 132.2, 131.8, 130.3, 129.6, 128.4 (2C), 128.0, 127.8, 127.7, 127.6, 126.3, 126.0, 125.6, 112.6, 56.2, 40.8; **IR** (neat) 3054, 3023, 2879, 1612, 1350 cm⁻¹; [α]²³_D -9.43 (c 2.28, CHCl₃); **SFC** analysis (AD-H, 20% MeOH, 3 mL/min) indicated 92% ee: t_R (major) = 4.2 min, t_R (minor) = 4.8 min.

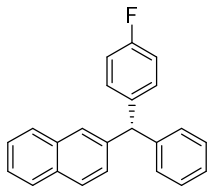


(**S**)-**1.22** Using representative procedure B outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-Bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.27 mg, 0.0210 mmol, 0.105 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(4-dimethylaminophenyl)-1,3,2-dioxaborinane (83 mg, 0.40 mmol, 2.0 equiv), (**S**)-**1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product

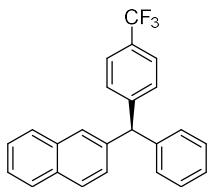
was purified by flash chromatography (1–3% Et₂O/pentane) to afford the product as a colorless oil. First run: (42.1 mg, 0.125 mmol 62%, 92% ee). Second run: (53.6 mg, 79%, 0.159 mmol, 92% ee). Analytical data is consistent with the values listed above for **(R)**-1.22 [α]²³_D +8.0 (*c* 1.00, CHCl₃); **SFC** analysis (AD-H, 20% MeOH, 3 mL/min) indicated 92% ee: *t*_R (major) = 3.9 min, *t*_R (minor) = 4.6 min.



(R)-1.23 Using representative procedure A outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), tricyclohexylphosphine (11 mg, 0.040 mmol, 0.2 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(4-fluorophenyl)-1,3,2-dioxaborinane (83 mg, 0.40 mmol, 2.0 equiv), **(S)**-1.4 (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash chromatography (1–3% Et₂O/pentane) to afford the product as a colorless oil. First run: (49.8 mg, 0.159 mmol, 80%, 90% ee). Second run: (52.6 mg, 0.168 mmol, 84%, 90% ee). Analytical data is consistent with literature values: ¹³_b **¹H NMR** (500 MHz, CDCl₃) δ 7.81–7.77 (m, 1H), 7.75 (d, *J* = 8.6 Hz, 1H), 7.71–7.67 (m, 1H), 7.45–7.39 (m, 3H), 7.31–7.21 (m, 4H), 7.15–7.07 (m, 4H), 6.97 (t, *J* = 8.8 Hz, 2H), 5.70 (s, 1H); **¹³C NMR** (125 MHz, CDCl₃) δ 161.6 (d, *J* = 245 Hz), 143.6, 141.4, 139.5 (d, *J* = 3 Hz), 133.5, 132.3, 131.1 (d, *J* = 8 Hz), 129.6, 128.6, 128.1, 128.05, 127.99, 127.8, 127.7, 126.7, 126.2, 125.9, 115.3 (d, *J* = 21 Hz), 56.3; [α]²³_D +4.5 (*c* 4.47, CHCl₃); **SFC** analysis (OJ-H, 12% IPA, 2.5 mL/min) indicated 90% ee: *t*_R (major) = 9.4 minutes, *t*_R (minor) = 8.7 minutes.

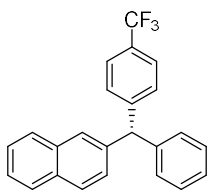


(S)-1.23 Using representative procedure B outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-Bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.27 mg, 0.0210 mmol, 0.105 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(4-fluorophenyl)-1,3,2-dioxaborinane (83 mg, 0.40 mmol, 2.0 equiv), **(S)-1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash chromatography (1–3% Et₂O/pentane) to afford the product as a colorless oil. First run: (49.8 mg, 0.159 mmol, 80%, 88% ee). Second run: (50.0 mg, 0.168 mmol, 84%, 88% ee). Analytical data is consistent with literature values:^{13b} **¹H NMR** (500 MHz, CDCl₃) δ 7.81–7.77 (m, 1H), 7.75 (d, J = 8.6 Hz, 1H), 7.71–7.67 (m, 1H), 7.45–7.39 (m, 3H), 7.31–7.21 (m, 4H), 7.15–7.07 (m, 4H), 6.97 (t, J = 8.8 Hz, 2H), 5.70 (s, 1H); **¹³C NMR** (125 MHz, CDCl₃) δ 161.6 (d, J = 245 Hz), 143.6, 141.4, 139.5 (d, J = 3 Hz), 133.5, 132.3, 131.1 (d, J = 8 Hz), 129.6, 128.6, 128.1, 128.05, 127.99, 127.8, 127.7, 126.7, 126.2, 125.9, 115.3 (d, J = 21 Hz), 56.3; [α]²³_D –3.6 (*c* 4.10, CHCl₃); **SFC** analysis (OJ-H, 12% IPA, 2.5 mL/min) indicated 88% ee: t_R (major) = 9.7 minutes, t_R (minor) = 10.6 minutes.



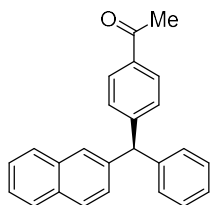
(R)-1.24 Using representative procedure A outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv),

tricyclohexylphosphine (11 mg, 0.040 mmol, 0.2 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.00 equiv), 5,5-dimethyl-2-(4-trifluoromethylphenyl)-1,3,2-dioxaborinane (103 mg, 0.400 mmol, 2.00 equiv), (**S**)-**1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash column chromatography (1% Et₂O/pentane) to afford the product as a colorless oil. First run: (64.4 mg, 0.178 mmol, 89%, 57% ee). Second run: (62.1 mg, 0.172 mmol, 86%, 57% ee). **TLC** R_f = 0.4 (pentane); **¹H NMR** (500 MHz, CDCl₃) δ 7.85–7.79 (m, 1H), 7.77 (d, *J* = 8.6 Hz, 1H), 7.74–7.68 (m, 1H), 7.55 (d, *J* = 8.1 Hz, 2H), 7.49–7.41 (m, 3H), 7.32 (t, *J* = 7.4 Hz, 2H), 7.29–7.22 (m, 4H), 7.14 (d, *J* = 7.6 Hz, 2H), 5.75 (s, 1H); **¹³C NMR** (125 MHz, CDCl₃) δ 148.0, 142.9, 140.6, 133.5, 132.4, 130.0, 129.6, 128.9 (q, *J* = 32.4 Hz), 128.7, 128.3, 128.02, 128.00, 127.9, 127.7, 126.9, 126.4, 126.1, 125.5 (q, *J* = 3.7 Hz), 124.4 (q, *J* = 271.9 Hz), 56.9; **IR** (neat) 3057, 1600, 1323, 1119 cm⁻¹; **HRMS** (TOF MS CI⁺) *m/z* calcd for C₁₅H₁₆O (M)⁺ 362.1282, found 362.1273; [α]_D²³ +4.84 (*c* 0.915, CHCl₃); **SFC** analysis (AD-H, 5% IPA, 2.5 mL/min) indicated 57% ee: t_R (major) = 7.7 minutes, t_R (minor) = 7.0 minutes.



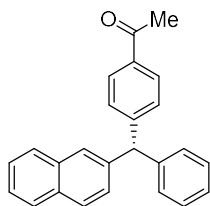
(**S**)-**1.24** Using representative procedure B outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.27 mg, 0.0210 mmol, 0.105 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.00 equiv), 5,5-dimethyl-2-(4-trifluoromethylphenyl)-1,3,2-dioxaborinane (103 mg, 0.400 mmol, 2.00 equiv), (**S**)-**1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product

was purified by flash column chromatography (1% Et₂O/pentane) to afford the product as a colorless oil. First run: (52.8 mg, 0.146 mmol, 73%, 91% ee). Second run: (48.6 mg, 0.134 mmol, 67%, 90% ee). **TLC** R_f = 0.4 (pentane); **¹H NMR** (500 MHz, CDCl₃) δ 7.85–7.79 (m, 1H), 7.77 (d, J = 8.6 Hz, 1H), 7.74–7.68 (m, 1H), 7.55 (d, J = 8.1 Hz, 2H), 7.49–7.41 (m, 3H), 7.32 (t, J = 7.4 Hz, 2H), 7.29–7.22 (m, 4H), 7.14 (d, J = 7.6 Hz, 2H), 5.75 (s, 1H); **¹³C NMR** (125 MHz, CDCl₃) δ 148.0, 142.9, 140.6, 133.5, 132.4, 130.0, 129.6, 128.9 (q, J = 32.4 Hz), 128.7, 128.3, 128.02, 128.00, 127.9, 127.7, 126.9, 126.4, 126.1, 125.5 (q, J = 3.7 Hz), 124.4 (q, J = 271.9 Hz), 56.9; **IR** (neat) 3057, 1600, 1323, 1119 cm⁻¹; **HRMS** (TOF MS CI⁺) m/z calcd for C₁₅H₁₆O (M)⁺ 362.1282, found 362.1273; **[α]_D²³** -16.5 (c 1.00, CHCl₃); **SFC** analysis (AD-H, 5% IPA, 2.5 mL/min) indicated 89% ee: t_R (major) = 6.6 minutes, t_R (minor) = 7.3 minutes.



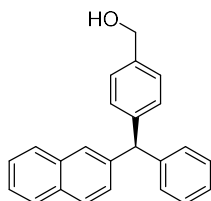
(R)-1.25 Using representative procedure A outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), tricyclohexylphosphine (11 mg, 0.040 mmol, 0.20 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μL, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(4-acetylphenyl)-1,3,2-dioxaborinane (93 mg, 0.40 mmol, 2.0 equiv), **(S)-1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash column chromatography (10% EtOAc/hexanes) to afford the product as an amorphous white solid. First run: (50.8 mg, 0.151 mmol, 76%, 89% ee). Second run: (51.0 mg, 0.152 mmol, 76%, 89% ee). **TLC** R_f = 0.4 (20% EtOAc/hexanes); **¹H**

NMR (500 MHz, CDCl₃) δ 7.80 (d, J = 8.4 Hz, 2H), 7.73–7.69 (m, 1H), 7.67 (d, J = 8.6 Hz, 1H), 7.63–7.57 (m, 1H), 7.40–7.30 (m, 3H), 7.21 (q, J = 7.7 Hz, 2H), 7.19–7.10 (m, 4H), 7.05 (d, J = 7.5 Hz, 2H), 5.65 (s, 1H), 2.47 (s, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 197.9, 149.4, 142.9, 140.7, 135.6, 133.5, 132.3, 129.9, 129.6, 128.7, 128.6, 128.3, 127.97, 127.95, 127.9, 127.7, 126.9, 126.3, 126.0, 57.0, 26.7; **IR** (neat) 3055, 2923, 1679, 1600, 1506 cm⁻¹; **HRMS** (TOF MS CI⁺) m/z calcd for C₂₅H₂₀O (M)⁺ 336.1514, found 3316.1514; **[α]²³_D** -17.2 (c 2.3, CHCl₃); **SFC** analysis (OD-H, 20% IPA, 3.0 mL/min) indicated 89% ee: t_R (major) = 6.3 minutes, t_R (minor) = 5.9 minutes.

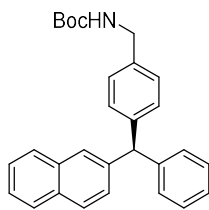


(S)-1.25 Using representative procedure B outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-Bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.67 mg, 0.0220 mmol, 0.11 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(4-acetylphenyl)-1,3,2-dioxaborinane (93 mg, 0.40 mmol, 2.0 equiv), **(S)-1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash column chromatography (10% EtOAc/hexanes) to afford the product as an amorphous white solid. First run: (66.5 mg, 0.198 mmol, 99%, 97% ee). Second run: (66.0 mg, 0.196 mmol, 98%, 97% ee). **TLC** R_f = 0.4 (20% EtOAc/hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.80 (d, J = 8.4 Hz, 2H), 7.73–7.69 (m, 1H), 7.67 (d, J = 8.6 Hz, 1H), 7.63–7.57 (m, 1H), 7.40–7.30 (m, 3H), 7.21 (q, J = 7.7 Hz, 2H), 7.19–7.10 (m, 4H), 7.05 (d, J = 7.5 Hz, 2H), 5.65 (s, 1H), 2.47 (s, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 197.9, 149.4, 142.9, 140.7, 135.6, 133.5, 132.3, 129.9, 129.6, 128.7, 128.6, 128.3, 127.97, 127.95, 127.9, 127.7, 126.9, 126.3, 126.0, 57.0, 26.7; **IR** (neat) 3055,

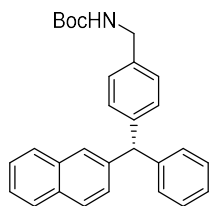
2923, 1679, 1600, 1506 cm^{-1} ; **HRMS** (TOF MS CI+) m/z calcd for $\text{C}_{25}\text{H}_{20}\text{O}$ (M)⁺ 336.1514, found 3316.1514; $[\alpha]_{\text{D}}^{29} +5.05$ (c 1.01, CHCl_3); **SFC** analysis (OD-H, 20% IPA, 3.0 mL/min) indicated 97% ee: t_{R} (major) = 5.9 minutes, t_{R} (minor) = 6.5 minutes.



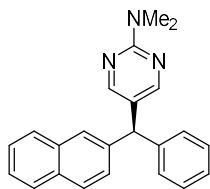
(R)-1.26 Using representative procedure A outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), tricyclohexylphosphine (11 mg, 0.040 mmol, 0.20 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μL , 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(4-hydroxymethylphenyl)-1,3,2-dioxaborinane (88 mg, 0.40 mmol, 2.0 equiv), **(S)-1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash column chromatography (10% EtOAc/hexane) to afford the product as an oil. First run: (51.0 mg, 0.157 mmol, 79%, 82% ee). Second run: (50.0 mg, 0.154 mmol, 77%, 81% ee). **TLC** R_{f} = 0.2 (20% EtOAc/hexanes); **¹H NMR** (500 MHz, CDCl_3) δ 7.82–7.76 (m, 1H), 7.74 (d, J = 8.7 Hz, 1H), 7.71–7.66 (m, 1H), 7.46 (s, 1H), 7.41 (dt, J = 9.5, 4.4 Hz, 2H), 7.34–7.24 (m, 5H), 7.24–7.19 (m, 1H), 7.15 (d, J = 8.4 Hz, 4H), 5.69 (s, 1H), 4.64 (d, J = 4 Hz, 2H), 1.77 (t, J = 4.5 Hz, 1H); **¹³C NMR** (125 MHz, CDCl_3) δ 143.7, 143.3, 141.5, 139.1, 133.5, 132.3, 129.9, 129.6, 128.5, 128.1, 128.02, 127.96, 127.9, 127.7, 127.3, 126.6, 126.1, 125.8, 65.2, 56.8; **IR** (neat) 3330 (br), 2953, 1600, 1506 cm^{-1} ; **HRMS** (TOF MS CI+) m/z calcd for $\text{C}_{24}\text{H}_{18}\text{O}$ ($\text{M} - 2\text{H}$)⁺ 322.1358, found 322.1364; $[\alpha]_{\text{D}}^{23} -18.3$ (c 1.66, CHCl_3); **SFC** analysis (AD-H, 30% MeOH, 2.5 mL/min) indicated 89% ee: t_{R} (major) = 4.3 minutes, t_{R} (minor) = 6.1 minutes.



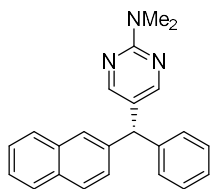
(R)-1.27 Using representative procedure A outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), tricyclohexylphosphine (11 mg, 0.040 mmol, 0.2 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(4-[(*tert*-butoxycarbonyl)amino]methyl}phenyl)-1,3,2-dioxaborinane (128 mg, 0.400 mmol, 2.00 equiv), **(S)-1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash column chromatography (15–25% Et₂O/hexanes) to afford the product as a white solid. First run: (71.0 mg, 0.168 mmol, 84%, 92% ee). Second run: (70.5 mg, 0.166 mmol, 83%, 89% ee). **TLC** R_f = 0.3 (20% EtOAc/hexanes); **m.p.** = 57 °C; **¹H NMR** (500 MHz, CDCl₃) δ 7.81–7.76 (m, 1H), 7.74 (d, J = 8.6 Hz, 1H), 7.71–7.66 (m, 1H), 7.45 (s, 1H), 7.41 (dt, J = 9.5, 1.0 Hz, 2H), 7.28 (t, J = 7.4 Hz, 3H), 7.22 (d, J = 7.6 Hz, 1H), 7.19 (d, J = 8.0 Hz, 2H), 7.14 (d, J = 7.5 Hz, 2H), 7.11 (d, J = 8.0 Hz, 2H), 5.67 (s, 1H), 4.83 (br s, 1H), 4.29 (d, J = 5.1 Hz, 2H), 1.45 (s, 9H); **¹³C NMR** (125 MHz, CDCl₃) δ 156.0, 143.7, 142.9, 141.5, 137.1, 133.5, 132.2, 129.9, 129.6, 128.5, 128.1, 128.01, 127.95, 127.8, 127.64, 127.59, 126.5, 126.1, 125.8, 79.5, 56.7, 44.4, 28.5; **IR** (neat) 3346, 2876, 1698, 1600, 1365 cm⁻¹; **HRMS** (TOF MS ES⁺) m/z calcd for C₂₉H₂₉O₂N (M + Na)⁺ 446.2096, found 446.2078; **$[\alpha]^{23}_D$** –14.3 (c 4.4, CHCl₃); **SFC** analysis (AS-H, 20% MeOH, 2.5 mL/min) indicated 92% ee: t_R (major) = 4.3 minutes, t_R (minor) = 4.7 minutes.



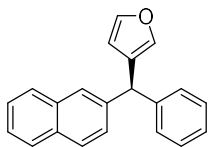
(S)-1.27 Using representative procedure B outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-Bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.27 mg, 0.0210 mmol, 0.105 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(4-{{(tert-butoxycarbonyl)amino}methyl}phenyl)-1,3,2-dioxaborinane (128 mg, 0.400 mmol, 2.00 equiv), **(S)-1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash column chromatography (15–25% Et₂O/hexanes) to afford the product as a white solid. First run: (84.0 mg, 0.198 mmol, 99%, 96% ee). Second run: (75.4 mg, 0.178 mmol, 89%, 94% ee). **TLC** R_f = 0.3 (20% EtOAc/hexanes); **m.p.** = 57 $^{\circ}$ C; **¹H NMR** (500 MHz, CDCl₃) δ 7.81–7.76 (m, 1H), 7.74 (d, J = 8.6 Hz, 1H), 7.71–7.66 (m, 1H), 7.45 (s, 1H), 7.41 (dt, J = 9.5, 1.0 Hz, 2H), 7.28 (t, J = 7.4 Hz, 3H), 7.22 (d, J = 7.6 Hz, 1H), 7.19 (d, J = 8.0 Hz, 2H), 7.14 (d, J = 7.5 Hz, 2H), 7.11 (d, J = 8.0 Hz, 2H), 5.67 (s, 1H), 4.83 (br s, 1H), 4.29 (d, J = 5.1 Hz, 2H), 1.45 (s, 9H); **¹³C NMR** (125 MHz, CDCl₃) δ 156.0, 143.7, 142.9, 141.5, 137.1, 133.5, 132.2, 129.9, 129.6, 128.5, 128.1, 128.01, 127.95, 127.8, 127.64, 127.59, 126.5, 126.1, 125.8, 79.5, 56.7, 44.4, 28.5; **IR** (neat) 3346, 2876, 1698, 1600, 1365 cm^{-1} ; **HRMS** (TOF MS ES⁺) m/z calcd for C₂₉H₂₉O₂N (M + Na)⁺ 446.2096, found 446.2078; **$[\alpha]_D^{29}$** +22.1 (c 1.01, CHCl₃); **SFC** analysis (AS-H, 20% MeOH, 2.5 mL/min) indicated 96% ee: t_R (major) = 4.5 minutes, t_R (minor) = 4.3 minutes.



(S)-1.28 Using representative procedure A outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), tricyclohexylphosphine (11 mg, 0.040 mmol, 0.20 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-((dimethylamino)-5-pyrimidinylphenyl)-1,3,2-dioxaborinane (94 mg, 0.40 mmol, 2.0 equiv), **(S)-1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash column chromatography (5% EtOAc/benzene) to afford the product as a white solid. First run: (58.2 mg, 0.171 mmol, 86%, 89% ee). Second run: (58.6 mg, 0.173 mmol, 86%, 89% ee). **TLC** R_f = 0.5 (5% EtOAc/benzene); **m.p.** = 45–47 $^{\circ}$ C; **1 H NMR** (500 MHz, CDCl_3) δ 8.04 (s, 2H), 7.73–7.67 (m, 1H), 7.66 (d, J = 8.6 Hz, 1H), 7.64–7.59 (m, 1H), 7.40 (s, 1H), 7.33 (dt, J = 9.5, 3.5 Hz, 2H), 7.23–7.16 (m, 3H), 7.15–7.10 (m, 1H), 7.06 (d, J = 7.6 Hz, 2H), 2.39 (s, 1H), 3.08 (s, 6H); **13 C NMR** (125 MHz, CDCl_3) δ 161.3, 158.5, 142.9, 140.7, 133.5, 132.3, 129.8, 128.7, 128.3, 127.9, 127.67, 127.65, 127.6, 126.8, 126.3, 125.9, 123.6, 51.7, 37.2; **IR** (neat) 3054, 3023, 2861, 1599, 1531 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{23}\text{H}_{21}\text{N}_3$ ($\text{M} + \text{H}$) $^+$ 340.1814, found 340.1819; **$[\alpha]^{23}_{\text{D}}$** +15.7 (c 2.51, CHCl_3); **SFC** analysis (AD-H, 30% MeOH, 2.5 mL/min) indicated 89% ee: t_R (major) = 4.6 minutes, t_R (minor) = 6.4 minutes.

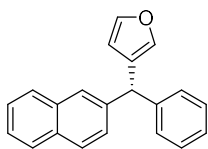


(S)-1.28 Using representative procedure B outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-Bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.27 mg, 0.0210 mmol, 0.105 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-((dimethylamino)-5-pyrimidinylphenyl)-1,3,2-dioxaborinane (94 mg, 0.40 mmol, 2.0 equiv), **(S)-1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash column chromatography (5% EtOAc/benzene) to afford the product as a white solid (50.8 mg, 0.150 mmol, 75%, 92% ee). **TLC** R_f = 0.5 (5% EtOAc/benzene); **m.p.** = 45–47 °C; **¹H NMR** (500 MHz, CDCl₃) δ 8.04 (s, 2H), 7.73–7.67 (m, 1H), 7.66 (d, J = 8.6 Hz, 1H), 7.64–7.59 (m, 1H), 7.40 (s, 1H), 7.33 (dt, J = 9.5, 3.5 Hz, 2H), 7.23–7.16 (m, 3H), 7.15–7.10 (m, 1H), 7.06 (d, J = 7.6 Hz, 2H), 2.39 (s, 1H), 3.08 (s, 6H); **¹³C NMR** (125 MHz, CDCl₃) δ 161.3, 158.5, 142.9, 140.7, 133.5, 132.3, 129.8, 128.7, 128.3, 127.9, 127.67, 127.65, 127.6, 126.8, 126.3, 125.9, 123.6, 51.7, 37.2; **IR** (neat) 3054, 3023, 2861, 1599, 1531 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for C₂₃H₂₁N₃ (M + H)⁺ 340.1814, found 340.1819; **$[\alpha]_D^{29}$** –13.2 (c 0.675, CHCl₃); **SFC** analysis (AD-H, 30% MeOH, 2.5 mL/min) indicated 92% ee: t_R (major) = 6.1 minutes, t_R (minor) = 4.5 minutes.



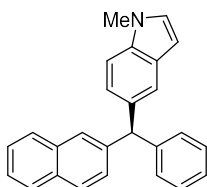
(S)-1.29 Using representative procedure A outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), tricyclohexylphosphine (11 mg, 0.040 mmol, 0.20 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(3-furanyl)-1,3,2-dioxaborinane (72 mg, 0.40 mmol, 2.0 equiv), **(S)-1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene

(1 mL). The product was purified by flash column chromatography (0.5–1% Et₂O/pentane) to afford the product as a white solid. First run: (45.8 mg, 0.161 mmol, 80%, 94% ee). Second run: (44.0 mg, 0.155 mmol, 78%, 94% ee). **TLC** *R_f* = 0.5 (1% Et₂O/pentane); **m.p.** = 65–67 °C; **¹H NMR** (500 MHz, CDCl₃) δ 7.83–7.69 (m, 3H), 7.60 (s, 1H), 7.47–7.37 (m, 3H), 7.34 (dd, *J* = 8.6, 1 Hz, 1H), 7.31–7.16 (m, 5H), 6.97 (s, 1H), 6.26 (s, 1H), 5.42 (s, 1H); **¹³C NMR** (125 MHz, CDCl₃) δ 143.4, 143.3, 141.3, 141.1, 133.5, 132.4, 129.0, 128.6, 128.3, 128.1, 128.0, 127.7, 127.6, 127.1, 126.7, 126.2, 125.8, 111.6, 48.3; **IR** (neat) 3145, 3024, 1599, 1492 cm⁻¹; **HRMS** (TOF MS CI⁺) *m/z* calcd for C₂₁H₁₆O (M)⁺ 284.1201, found 284.1203; **[α]_D²³** +22.3 (*c* 1.67, CHCl₃); **SFC** analysis (AD-H, 5% IPA, 2.5 mL/min) indicated 94% ee: *t_R* (major) = 12.2 minutes, *t_R* (minor) = 11.3 minutes.



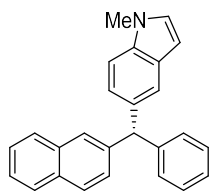
(R)-1.29 Using representative procedure B outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-Bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.27 mg, 0.0210 mmol, 0.105 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μL, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(3-furanyl)-1,3,2-dioxaborinane (72 mg, 0.40 mmol, 2.0 equiv), **(S)-1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash column chromatography (0.5–1% Et₂O/pentane) to afford the product as a white solid. First run: (35.5 mg, 0.125 mmol, 62.5 %, 82% ee). Second run: (38.7 mg, 0.136 mmol, 68%, 84% ee). **TLC** *R_f* = 0.5 (1% Et₂O/pentane); **m.p.** = 65–67 °C; **¹H NMR** (500 MHz, CDCl₃) δ 7.83–7.69 (m, 3H), 7.60 (s, 1H), 7.47–7.37 (m, 3H), 7.34 (dd, *J* = 8.6, 1 Hz, 1H), 7.31–7.16 (m, 5H), 6.97 (s, 1H), 6.26 (s, 1H), 5.42 (s, 1H); **¹³C NMR** (125 MHz, CDCl₃) δ 143.4, 143.3, 141.3, 141.1, 133.5,

132.4, 129.0, 128.6, 128.3, 128.1, 128.0, 127.7, 127.6, 127.1, 126.7, 126.2, 125.8, 111.6, 48.3; **IR** (neat) 3145, 3024, 1599, 1492 cm^{-1} ; **HRMS** (TOF MS CI^+) m/z calcd for $\text{C}_{21}\text{H}_{16}\text{O}$ (M) $^+$ 284.1201, found 284.1203; $[\alpha]^{29}_{\text{D}}$ -22.0 (c 1.00, CHCl_3); **SFC** analysis (AD-H, 5% IPA, 2.5 mL/min) indicated 84% ee: t_{R} (major) = 12.2 minutes, t_{R} (minor) = 13.4 minutes.

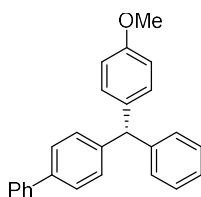


(S)-1.14 Using representative procedure A outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), tricyclohexylphosphine (11 mg, 0.040 mmol, 0.20 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μL , 0.60 mmol, 3.0 equiv), 5-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)-1-methyl-1*H*-indole (97 mg, 0.40 mmol, 2.0 equiv), **(S)-1.4** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash column chromatography (5–20% Et_2O /hexane, 0.5% TEA) to afford the product as a white solid. First run: (63.4 mg, 0.182 mmol, 91%, 92% ee). Second run: (61.4 mg, 0.178 mmol, 89%, 93% ee). **TLC** R_f = 0.3 (20% Et_2O /hexane); **m.p.** = 49–52 $^\circ\text{C}$; **^1H NMR** (500 MHz, CDCl_3) δ 7.80–7.75 (m, 1H), 7.73 (d, J = 8.6 Hz, 1H), 7.70–7.64 (m, 1H), 7.50 (s, 1H), 7.40 (dt, J = 9.3, 4.9 Hz, 2H), 7.37–7.31 (m, 2H), 7.27 (t, J = 7.3 Hz, 2H), 7.25–7.16 (m, 4H), 7.07 (dd, J = 8.7, 1.0 Hz, 1H), 6.98 (d, J = 2.9 Hz, 1H), 6.36 (d, J = 2.9 Hz, 1H), 5.83 (s, 1H), 3.72 (s, 3H); **^{13}C NMR** (125 MHz, CDCl_3) δ 144.7, 142.6, 135.6, 134.9, 133.5, 132.2, 129.8, 129.2, 128.55, 128.52, 128.4, 128.0, 127.9, 127.8, 127.6, 126.3, 126.0, 125.6, 123.9, 121.7, 109.2, 101.1, 57.1, 33.0; **IR** (neat) 3022, 2884, 1599, 1489 cm^{-1} ; **HRMS** (TOF MS ES^+) m/z calcd for $\text{C}_{26}\text{H}_{21}\text{N}$ ($\text{M} + \text{Na}$) $^+$ 370.1572, found 370.1576; $[\alpha]^{23}_{\text{D}}$ -10.8 (c 1.00, CHCl_3); **SFC** analysis (AD-H, 20% MeOH, 2.5 mL/min) indicated 93% ee: t_{R} (major)

= 8.1 minutes, t_R (minor) = 9.0 minutes. Single crystals suitable for X-ray crystallographic analysis were grown by slow diffusion of hexane into a solution of **(S)-1.14** in benzene at 4 °C.

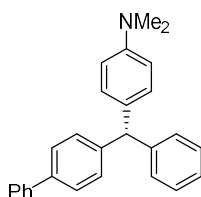


(R)-1.14 Using representative procedure B outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.3 mg, 0.021 mmol, 0.11 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)-1-methyl-1*H*-indole (97 mg, 0.40 mmol, 2.0 equiv), **(S)-2.1** (66 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash column chromatography (5–20% Et₂O/hexanes, 0.5% TEA) to afford the product as a white solid. First run: (41.5 mg, 0.119 mmol, 57%, 92% ee). Second run: (57.0 mg, 0.164 mmol, 82%, 92% ee). Analytical data is consistent with the values listed above for **(S)-1.4** [α]²³_D +6.0 (*c* 0.9, CHCl₃); **SFC** analysis (AD-H, 20% MeOH, 2.5 mL/min) indicated 93% ee: *t*_R (major) = 8.9 minutes, *t*_R (minor) = 8.1 minutes.



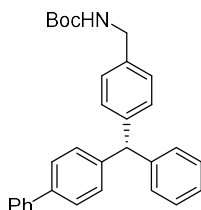
(R)-1.30 Using representative procedure B above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.3 mg, 0.0210 mmol, 0.11 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5,5-dimethyl-2-(4-methoxyphenyl)-1,3,2-dioxaborinane (88 mg, 0.40 mmol, 2.0 equiv), **(S)-1.19** (69 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was

purified by flash column chromatography (1–3% Et₂O/hexanes) to afford the product as a colorless oil. First run: (54.8 mg, 0.156 mmol, 78%, 81% ee). Second run: (55.8 mg, 0.159 mmol, 80%, 81% ee). **TLC** R_f = 0.4 (10% Et₂O/Hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.56 (d, J = 7.2, 2H), 7.51 (d, J = 7.1, 2H), 7.42 (t, J = 7.1, 2H), 7.29 (m, 3H), 7.19 (m, 5H), 7.06 (d, J = 7.8, 2H), 6.83 (d, J = 8.2, 2H) 5.53 (s, 1H), 3.78 (s, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 158.2, 144.3, 143.5, 141.0, 139.2, 136.2, 130.5, 129.9, 129.5, 128.9, 128.5, 128.3, 127.15, 127.14, 126.4, 113.9, 55.9, 55.4; **IR** (neat) 3020, 2996, 1508, 1244, 1030 cm⁻¹; **HRMS** (TOF MS ES⁺) m/z calcd for C₂₆H₂₂O (M + Na)⁺ 350.1671, found 367.1679; **[α]²³_D** +1.2 (c 1.01, CHCl₃), **SFC** analysis (AD-H, 10% MeOH, 2.5 mL/min) indicated 84% ee: t_R (minor) = 21.5 minutes, t_R (major) = 19.8 minutes.

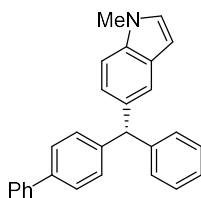


(R)-1.31 Using representative procedure B above, the following amounts and reagents: 1,8-bis(1,5-cyclooctadiene)nickel (8.3 mg, 0.030 mmol, 0.10 equiv), 1,3-bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (12 mg, 0.030 mmol, 0.10 equiv), potassium *tert*-butoxide (64 mg, 0.60 mmol, 2.0 equiv), 4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)-*N,N*-dimethylaniline (134 mg, 0.600 mmol, 2.00 equiv), **(S)-1.19** (103 mg, 0.300 mmol, 1.00 equiv) and 1-butanol (54 μ L, 0.90 mmol, 3.00 equiv). Purified by flash column chromatography (0–10% Et₂O/hexanes) to afford **(R)-1.32**, as a light yellow oil (55 mg, 0.15 mmol, 75%). **TLC** R_f = 0.3 (10% Et₂O/Hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.57 (d, J = 7.2, 2H), 7.49 (d, J = 8.2, 2H), 7.41 (t, J = 7.7, 2H), 7.29 (m, 3H), 7.27 (s, 1H) 7.21 (m, 5H), 7.02 (dd, J = 8.6, 2H) 6.67 (d, J = 8.9, 2H), 6.4 (d, J = 2.8, 1H), 5.5 (s, 1H), 2.9 (s, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 149.2, 144.7, 143.9, 141.1, 139.0, 131.9, 130.2, 129.9, 129.5, 128.8, 128.4, 127.20, 127.15, 127.1, 126.3, 112.7,

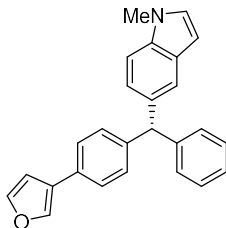
55.7, 40.8; **IR** (neat) 3024, 2841, 2360, 1613, 1485, 1347, 763 cm^{-1} ; **HRMS** (TOF MS EI+) m/z calcd for $\text{C}_{27}\text{H}_{25}\text{O}$ ($\text{M} + \text{Na}$)⁺ 364.2065, found 364.2061; $[\alpha]_{\text{D}}^{23}$ -2.9 (c 1.07, CHCl_3); **SFC** analysis (AD-H, 16% MeOH, 3.0 mL/min) indicated 79% ee: t_{R} (minor) = 26.3 minutes, t_{R} (major) = 11.6 minutes.



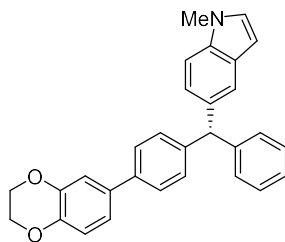
(S)-1.32 Using representative procedure B outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.67 mg, 0.022 mmol, 0.11 equiv), potassium *tert*-butoxide (45 mg, 0.4 mmol, 2.0 equiv), *tert*-butyl 4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)benzylcarbamate (127.3 mg, 0.200 mmol, 2.00 equiv), **(S)-1.19** (68.9 mg, 0.200 mmol, 1.00 equiv) and 1-butanol (54 μL , 0.60 mmol, 3.0 equiv). Purified by flash column chromatography (0–15 % EtOAc/Hexane) to afford the desired triarylmethane as a clear colorless oil (48.5 mg, 54%); **TLC** R_{f} = 0.1 (9:1 Hexanes:EtOAc); **^1H NMR** (500 MHz, CDCl_3) δ 7.57 (dd, J = 1.5, J = 8.4, 2H), 7.51 (d, J = 8.3, 2H), 7.39 (t, J = 7.5, 2H), 7.29 (m, 3H), 7.25 (m, 3H), 7.15 (m, 6H), 5.56 (s, 1H), 4.82 (s, 1H), 4.30 (d, J = 5.3, 1H); **^{13}C NMR** (125 MHz, CDCl_3) δ 156.0, 143.9, 143.13, 143.05, 140.9, 139.3, 137.1, 129.9, 129.8, 129.5, 128.9, 128.5, 127.6, 127.3, 127.16, 127.14, 126.5, 56.3, 44.3, 28.5; **IR** (neat) 3294, 3028, 1695, 1486, 1316, 1016, 757 cm^{-1} ; **HRMS** (TOF MS EI+) m/z calcd for $\text{C}_{31}\text{H}_{31}\text{NO}_2$ [$\text{M} + \text{Na}$]⁺ 472.2253, found 472.2261. $[\alpha]_{\text{D}}^{29}$ -8.2 **SFC** analysis (AS-H, 20% MeOH, 2.5 mL/min) indicated 92% ee: t_{R} (major) = 6.86 minutes, t_{R} (minor) = 7.47 minutes.



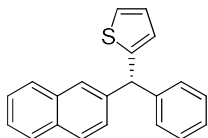
(R)-1.33 Using representative procedure B outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.27 mg, 0.0210 mmol, 0.105 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)-1-methyl-1*H*-indole (97 mg, 0.40 mmol, 2.0 equiv), **(S)-1.19** (69 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified by flash column chromatography (5–20% Et₂O/hexane, 0.5% TEA) to afford the product as a white solid. First run: (61.0 mg, 0.163mmol, 82%, 96% ee). Second run: (37.8 mg, 0.101 mmol, 51%, 96% ee). **TLC** *R_f* = 0.3 (10% EtOAc/hexanes); **m.p.** = 54–55 °C; **¹H NMR** (400 MHz, CDCl₃) δ 7.56 (d, *J* = 7.4, 2H), 7.49 (d, *J* = 8.2, 2H), 7.37 (t, *J* = 7.4, 2H), 7.27 (m, 3H), 7.26 (s, 1H) 7.22 (m, 9H), 7.07 (dd, *J* = 1.3, 8.6, 1H) 6.87 (d, *J* = 3.0, 1H), 6.37 (d, *J* = 2.8, 1H), 5.70 (s, 1H), 3.73 (s, 3H); **¹³C NMR** (100 MHz, CDCl₃) δ 144.9, 144.2, 141.1, 138.9, 135.6, 135.1, 130.1, 129.7, 129.2, 128.8, 128.6, 128.4, 127.17, 127.15, 127.0, 126.2, 123.8, 121.6, 109.2, 101.1, 56.7, 32.9; **IR** (neat) 3025, 2360, 1486, 1449, 1246, 1006, 760 cm⁻¹; **HRMS** submitted; **[α]²³_D** – 5.2; **SFC** analysis (AD-H, 25% MeOH, 2.5 mL/min) indicated 96% ee: *t_R* (major) = 11.3 minutes, *t_R* (minor) = 16.1 minutes.



(R)-1.34 Using representative procedure B outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-Bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.27 mg, 0.0210 mmol, 0.105 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)-1-methyl-1*H*-indole (97 mg, 0.40 mmol, 2.0 equiv), **(S)-1.20** (67 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified twice by flash column chromatography (6% Et₂O/hexane and then 60% benzene/pentane) to afford the desired triarylmethane as a white solid. First run: (58.5 mg, 0.161 mmol, 80%, 87% ee). Second run: (58.5 mg, 0.161 mmol, 80%, 87% ee). **TLC** R_f = 0.6 (40% pentane/benzene); **m.p.** = 149–151 °C; **¹H NMR** (500 MHz, CDCl₃) δ 7.59 (s, 1H), 7.34 (s, 1H), 7.29 (d, J = 8.3 Hz, 2H), 7.23 (s, 1H), 7.18 (t, J = 7.4 Hz, 2H), 7.15–7.09 (m, 2H), 7.07 (t, J = 7.4 Hz, 4H), 6.95 (d, J = 8.6 Hz, 1H), 6.90 (d, J = 2.8 Hz, 1H), 6.57 (s, 1H), 6.29 (d, J = 2.8 Hz, 1H), 5.58 (s, 1H), 3.63 (s, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 144.9, 143.9, 141.7, 138.5, 135.6, 135.0, 130.3, 130.1, 129.7, 129.2, 128.5, 128.3, 126.4, 126.2, 125.8, 123.7, 121.5, 109.2, 109.0, 101.1, 56.7, 33.0; **IR** (neat) 3145, 3024, 1599, 1492 cm⁻¹; **[α]²³_D** –3.1 (*c* 2.24, CHCl₃); **HRMS** submitted; **SFC** analysis (OJ-H, 30% MeOH, 3.0 mL/min) indicated 87% ee: t_R (major) = 23.6 minutes, t_R (minor) = 26.2 minutes.



(R)-1.35 Using representative procedure B outlined above, the following amounts of reagents were used: 1,8-bis(1,5-cyclooctadiene)nickel (5.5 mg, 0.020 mmol, 0.10 equiv), 1,3-bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (8.27 mg, 0.0210 mmol, 0.105 equiv), potassium *tert*-butoxide (45 mg, 0.40 mmol, 2.0 equiv), 1-butanol (54 μ L, 0.60 mmol, 3.0 equiv), 5-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)-1-methyl-1*H*-indole (97 mg, 0.40 mmol, 2.0 equiv), **(S)-1.21** (67 mg, 0.20 mmol, 1.0 equiv), tetrahydrofuran (1 mL) and toluene (1 mL). The product was purified twice by flash column chromatography (6% EtOAc/hexanes and then 70% benzene/pentane) to afford the desired triarylmethane as a white solid. First run: (51.4 mg, 0.119 mmol, 60%, 93% ee). Second run: (52.0 mg, 0.121 mmol, 60%, 93% ee). **TLC** R_f = 0.3 (40% pentane/benzene); **m.p.** = 81–84 $^{\circ}$ C; **1 H NMR** (500 MHz, CDCl_3) δ 7.43 (s, J = 8.0 2H), 7.33 (s, 1H), 7.29 (t, J = 7.4 Hz, 2H), 7.24 (s, 1H), 7.22–7.14 (m, 5H), 7.10 (s, 1H), 7.06 (dt, J = 8.2, 2.0 Hz, 2H), 7.01 (d, J = 2.7 Hz, 1H), 6.90 (d, J = 8.2 Hz, 1H), 6.39 (d, J = 2.5 Hz, 1H), 5.69 (s, 1H), 4.27 (s, 4H), 3.76 (s, 3H); **13 C NMR** (125 MHz, CDCl_3) δ 144.9, 143.8, 143.7, 143.1, 138.4, 135.6, 135.1, 134.8, 130.0, 129.7, 129.2, 128.6, 128.4, 126.6, 126.2, 123.8, 121.6, 120.2, 117.6, 115.8, 109.2, 101.1, 64.58, 64.56, 56.7, 33.0; **IR** (neat) 2916, 1586, 1513, 1449 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{30}\text{H}_{25}\text{O}_2\text{N}$ ($\text{M} + \text{Na}$) $^+$ 454.1783, found 454.1772; **$[\alpha]_D^{23}$** –8.2 (c 2.36, CHCl_3); **SFC** analysis (OD-H, 25% IPA, 2.5 mL/min) indicated 93% ee: t_R (major) = 23.6 minutes, t_R (minor) = 26.2 minutes.



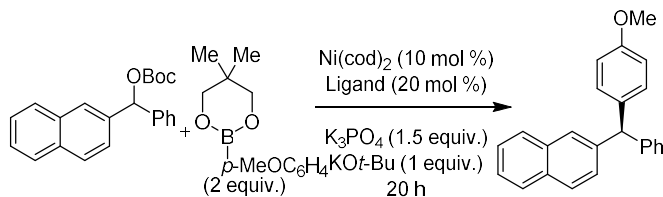
(R)-1.12 Prepared according to general procedure B using the following amounts and reagents: Ni(cod)₂ (2.8 mg, 0.010 mmol, 0.10 equiv), 1,3-bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate (3.9 mg, 0.010 mmol, 0.10 equiv), potassium *tert*-butoxide (22 mg, 0.20 mmol, 2.0 equiv), 5,5-dimethyl-2-(thiophen-2-yl)-1,3,2-dioxaborinane (39.2 mg, 0.200 mmol, 2.00 equiv), **(S)-1.4** (33.1 mg, 0.100 mmol, 1.00 equiv) and 1-butanol (27 μ L, 0.30 mmol, 3.0 equiv). Purified by flash column chromatography (0–5% Et₂O/hexanes) to afford the desired triarylmethane as a yellow solid (11.4 mg, 0.0379 mmol, 38%). Analytical data is consistent with literature values.^{13b} **¹H NMR** (400 MHz, CDCl₃) δ 7.84–7.72 (m, 3H), 7.61 (s, 1H), 7.37–7.41 (m, 2H), 7.37 (dd, J = 1.6, 8.5), 7.34–7.20 (m, 6H), 6.95 (t, J = 4.3), 6.73 (d, J = 3.3), 5.84 (s, 1H); **¹³C NMR** δ (125 MHz, CDCl₃) δ 147.8, 143.7, 141.4, 133.5, 132.4, 129.1, 128.6, 128.2, 128.1, 127.7, 127.5, 127.4, 126.9, 126.8, 126.7, 126.2, 125.9, 124.8, 52.3 [α]²⁵_D –9.3 (c 0.57, CHCl₃).

Tables of results using alternative ligands and bases.

Other ligands and bases were tested under reaction conditions similar to Table 1.1.

Representative examples are shown below.

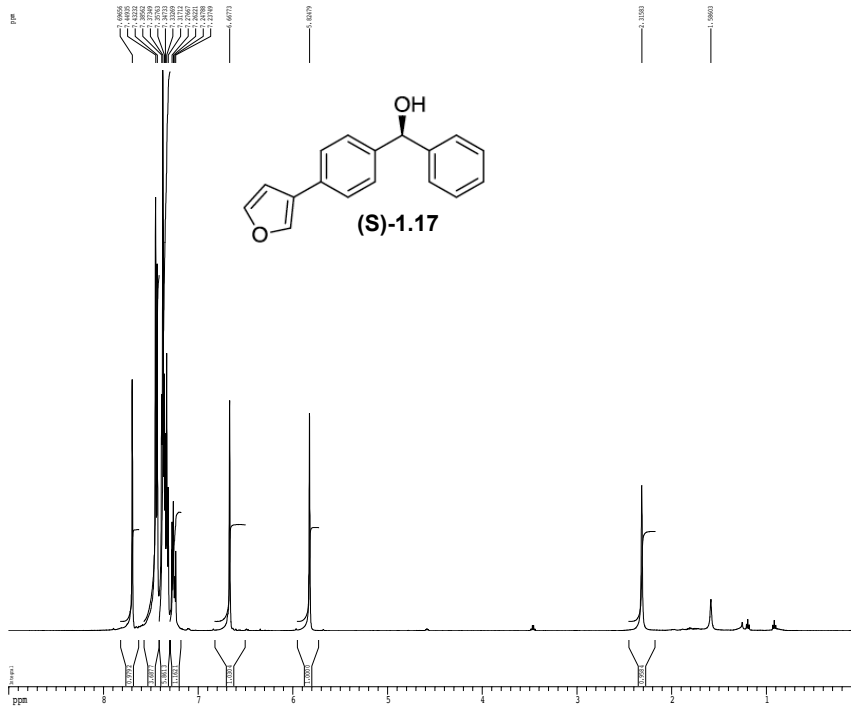
Table 1.4. Examination of additional ligands in the cross-coupling reaction



Entry	ligand	yield ^a
1	DPEphos (Bis[(2-diphenylphosphino)phenyl] ether)	< 5%
2	Cy-DPEphos (Bis[(2-dicyclohexylphosphino)phenyl] ether)	< 5%
3	DPPO (1,8-bis(diphenylphosphino)octane)	13%
4	PPh_3 (triphenylphosphine)	22%
5	$\text{P}(t\text{-Bu})_3$ tri- <i>tert</i> -butylphosphine	< 5%
6	XPhos (2-Dicyclohexylphosphino-2',4',6'-triisopropylbiphenyl)	< 5%
7	SPhos (2-Dicyclohexylphosphino-2',6'-dimethoxybiphenyl)	< 5%
8	SIPr-HBF ₄ (1,3-Bis(2,6-diisopropylphenyl)-4,5-dihydroimidazolium tetrafluoroborate)	31%
9	tricyclohexylphosphine	86%
10	PCy_3 tricyclohexylphosphine (11 mol %)	83%
11	SIMes-HBF ₄ 1,3-bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazolium tetrafluoroborate	84%
12	None	< 5%

^aDetermined by ¹H NMR analysis using an internal standard (PhSiMe_3).

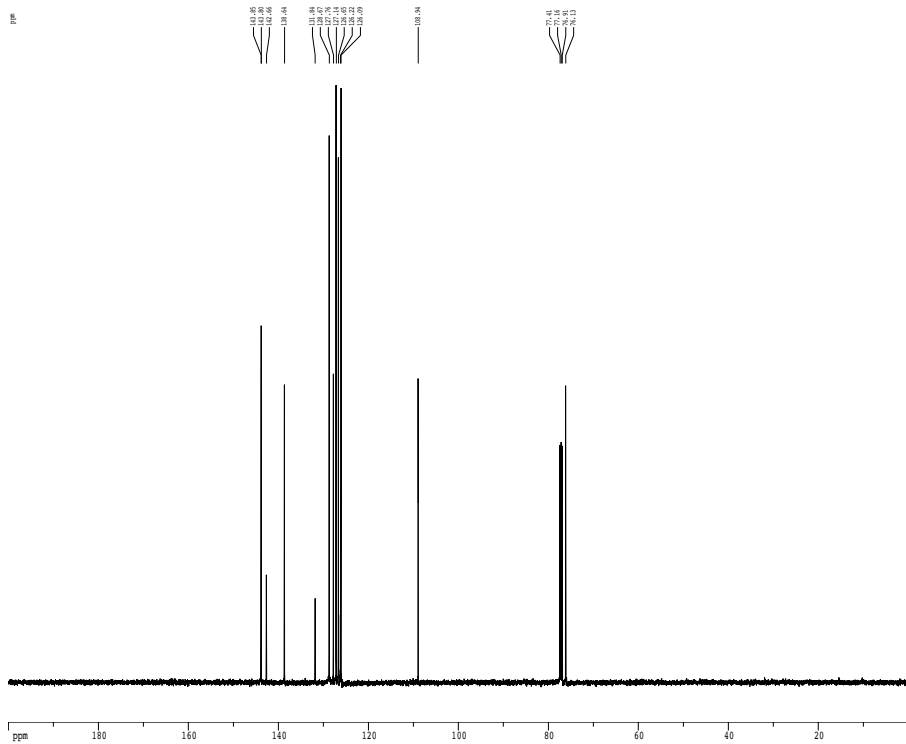
1H spectrum



```

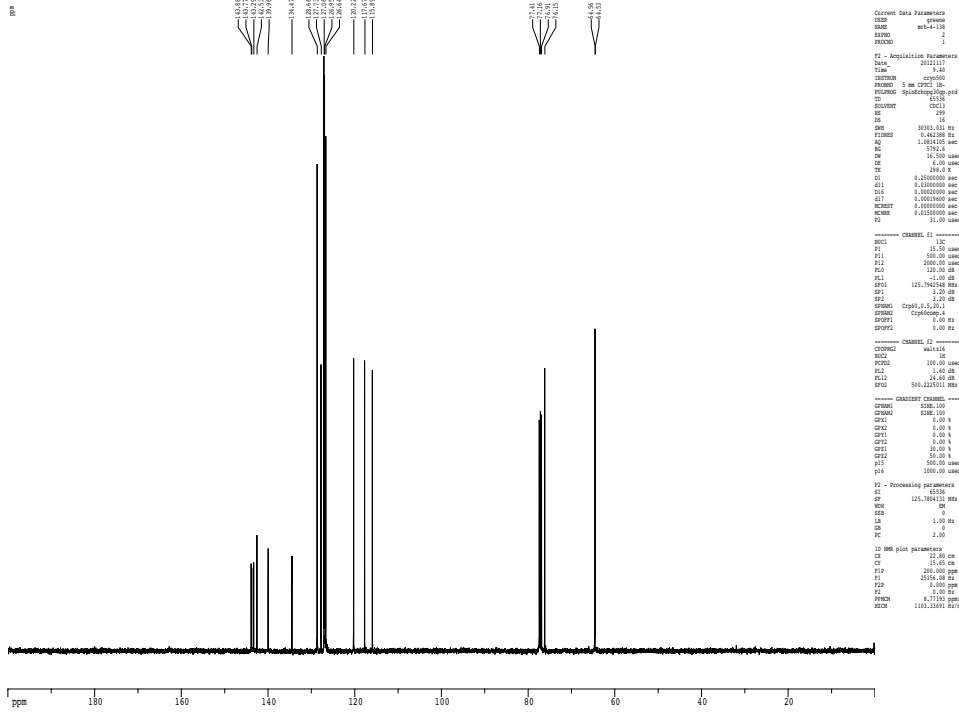
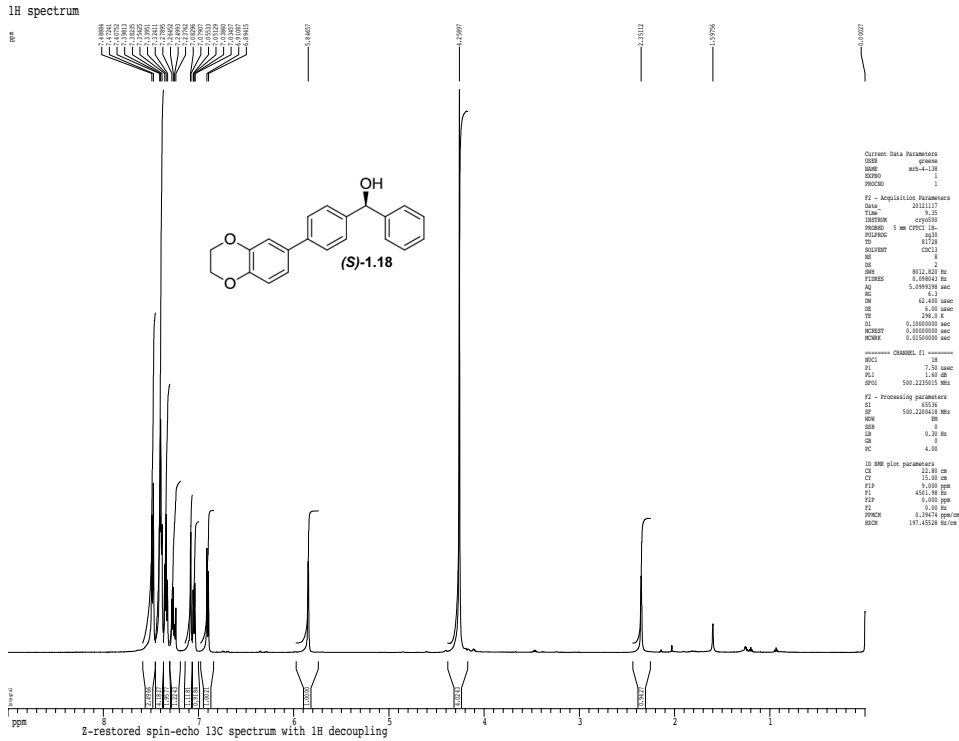
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PROCNO        1
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Date_         20121111
Time         15.00
INSTRUM       crypm00
PROBHD 5 mm CPXI 1H-
PULPROG       zgpg30
TD            65536
SOLVENT       CDCl3
NS            8
DS            4
SWH           6032.620 Hz
FIDRES       0.00041 Hz
AQ           5.0999300 sec
RG           67.000000
DE           42.000000
TE           300.2 K
NUC1          13C
D1           0.10000000 sec
DECT1        0.00000000 sec
PCNMR1       0.21500000 sec
===== CHANNEL f1 =====
NUC1          13C
P1           7.50000000 sec
PL1          190.000000 dB
SFO1         101.6261200 MHz
F2 - Processing parameters
SI           65536
SF           101.6261200 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB           0
PC           4.00
IS MRB plot parameters
CI           22.00 cm
CT           15.00 cm
F1P         9.0000000 ppm
F2P         400.1400000 MHz
F3P         0.000000000 Hz
F4P         0.00 Hz
F5P         0.39474 ppm/cm
PCNMR2       101.6261200 MHz
  
```

Z-restored spin-echo 13C spectrum with 1H decoupling

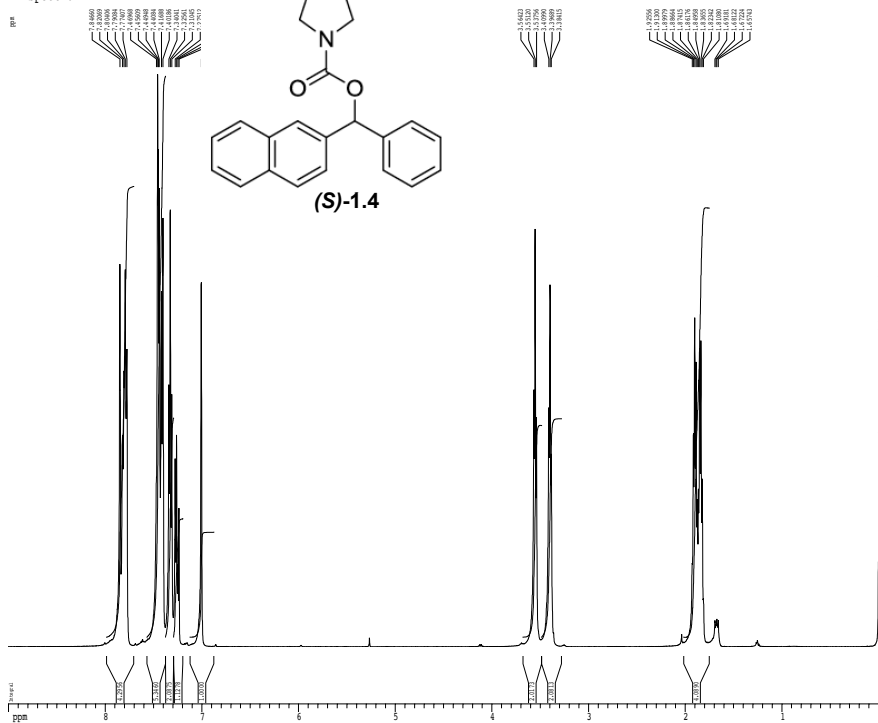


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Current Data Parameters
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PROCNO        1
F2 - Acquisition Parameters
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Time         15.00
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PULPROG       zgpg30
TD            65536
SOLVENT       CDCl3
NS            8
DS            4
SWH           30363.821 Hz
FIDRES       0.00041 Hz
AQ           1.0814105 sec
RG           1305.0
DE           16.500000
TE           300.2 K
NUC1          13C
D1           0.20000000 sec
DECT1        0.00000000 sec
D14          0.00020000 sec
d17          0.00190000 sec
DECT17       0.00000000 sec
PCNMR1       0.21500000 sec
F2           31.000000 sec
===== CHANNEL f1 =====
NUC1          13C
P1           15.50000000 sec
PL1          190.000000 dB
SFO1         101.6261200 MHz
F2 - Processing parameters
SI           65536
SF           101.6261200 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           2.00
IS MRB plot parameters
CI           22.00 cm
CT           15.00 cm
F1P         9.0000000 ppm
F2P         400.1400000 MHz
F3P         0.000000000 Hz
F4P         0.00 Hz
F5P         0.39474 ppm/cm
PCNMR2       101.6261200 MHz
  
```



1H spectrum



Current Data Parameters
NAME spwars
NAME mb-3-219
EXPNO 1
PROCNO 1

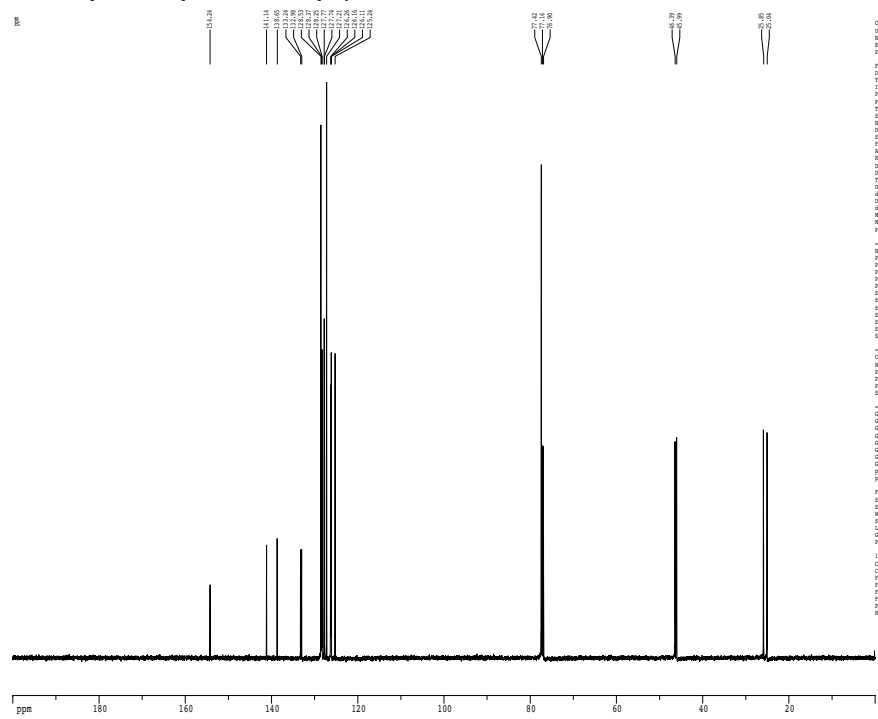
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PROBHD 1 mm CPYC 1H-
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DS 2
SWH 8012.822 Hz
FIDRES 0.090461 Hz
AQ 0.3993386 sec
RG 3
GB 0
PC 42.400 usec
DC 0.00 usec
TE 298.2 K
NUC1 13C
NUC2 1H
ACQRESPT 0.0000000 sec
ACQRES 0.0000000 sec
SOLVENT CDCl3

===== CHANNEL f1 =====
NUC1 13C
P1 15.00 usec
PL1 0.00 dB
SFO1 500.2253511 MHz

F1 - Processing parameters
SI 65536
SF 500.2253511 MHz
WDW EM
SSB 0
LB 0.300 Hz
GB 0
PC 4.00

1D NMR plot parameters
SI 65536
SF 500.2253511 MHz
WDW EM
SSB 0
LB 0.300 Hz
GB 0
PC 4.00

2-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
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NAME mb-3-219
EXPNO 1
PROCNO 1

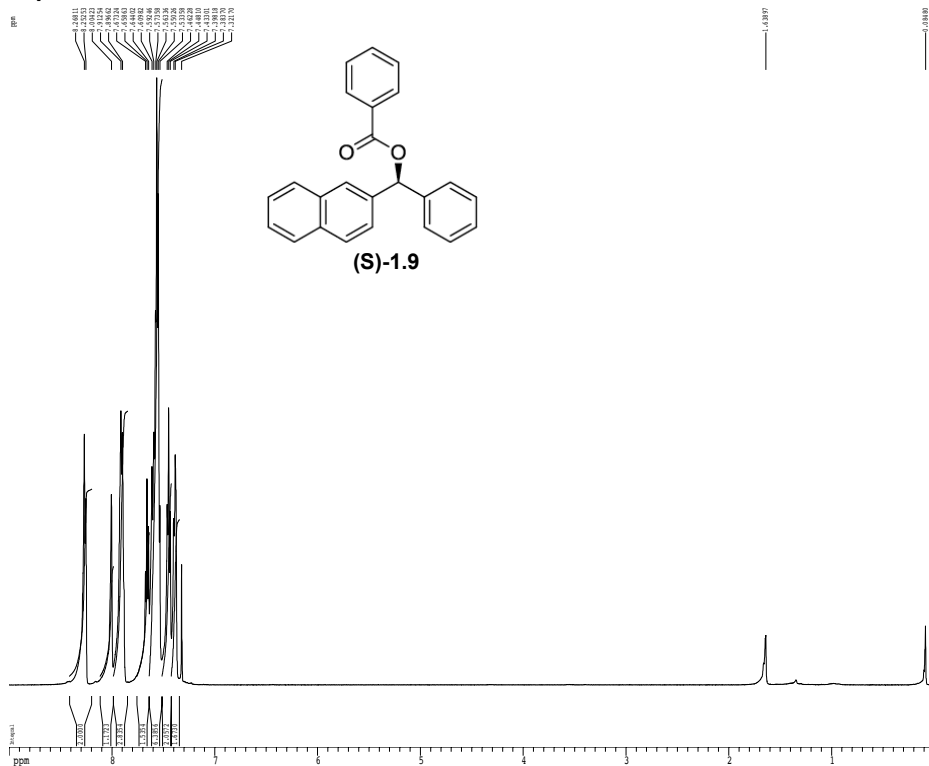
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DS 2
SWH 30561.031 Hz
FIDRES 0.642328 Hz
AQ 1.081440 sec
RG 3
GB 0
PC 15.500 usec
DC 0.00 usec
TE 298.2 K
NUC1 13C
NUC2 1H
ACQRESPT 0.0000000 sec
ACQRES 0.0000000 sec
SOLVENT CDCl3

===== CHANNEL f1 =====
NUC1 13C
P1 15.00 usec
PL1 0.00 dB
SFO1 500.2253511 MHz

F1 - Processing parameters
SI 65536
SF 500.2253511 MHz
WDW EM
SSB 0
LB 0.300 Hz
GB 0
PC 4.00

1D NMR plot parameters
SI 65536
SF 500.2253511 MHz
WDW EM
SSB 0
LB 0.300 Hz
GB 0
PC 4.00

1H spectrum



```

Current Data Parameters
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NAME          MB-3-144
EXPNO        2
PROCNO       1

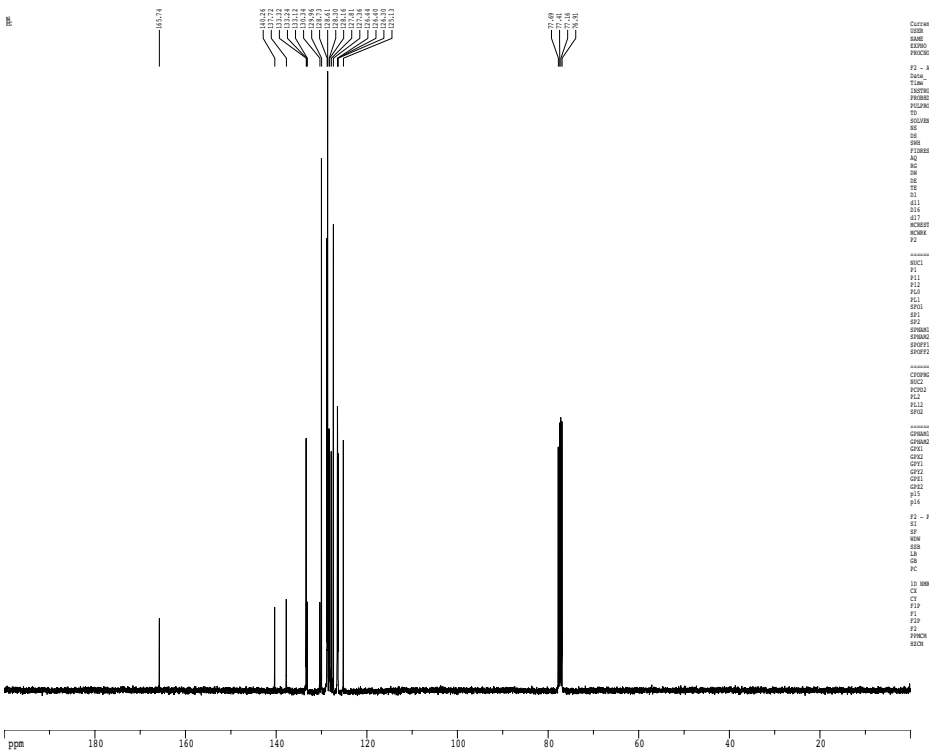
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PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           2
DS           4
SWH          8012.820 Hz
FIDRES      0.088414 Hz
AQ          1.8998774 sec
RG          14
SR          42.420 usec
SE          1.00 usec
TE          300.2 K
SI          0.10000000 sec
MCHST       0.00000000 sec
MCHSX       0.01500000 sec

===== CHANNEL f1 =====
NUC1         13C
P1           15.00 usec
PL1         -1.00 dB
PC1         500.2201011 MHz

F2 - Processing parameters
SI          500.220000 MHz
SF          500.220000 MHz
WDW         no
SSB         0
LB          0.00 Hz
GB          0
PC          4.00

ID NMR plot parameters
CX          22.80 cm
CT          15.00 cm
CF          9.400 ppm
F1          4501.38 Hz
F2          0.000 ppm
P2          0.00 Hz
PRCHA      0.28474 ppm/cm
SICH       197.45129 Hz/cm
    
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Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER          gpcwae
NAME          MB-3-144
EXPNO        2
PROCNO       1

F2 - Acquisition Parameters
Date_         20121118
Time          9.18
INSTRUM      cryo500
PROBHD      5 mm CPXI 13-
PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           2
DS           4
SWH          30300.012 Hz
FIDRES      0.461208 Hz
AQ          1.0814515 sec
RG          1158.2
SR          26.268 usec
SE          4.00 usec
TE          300.2 K
SI          0.20000000 sec
MCHST       0.00000000 sec
MCHSX       0.00010000 sec
MCHSY       0.00000000 sec
MCHSZ       0.01500000 sec
F2          0.00 usec

===== CHANNEL f1 =====
NUC1         13C
P1           15.00 usec
PL1         -1.00 dB
PC1         500.2201011 MHz

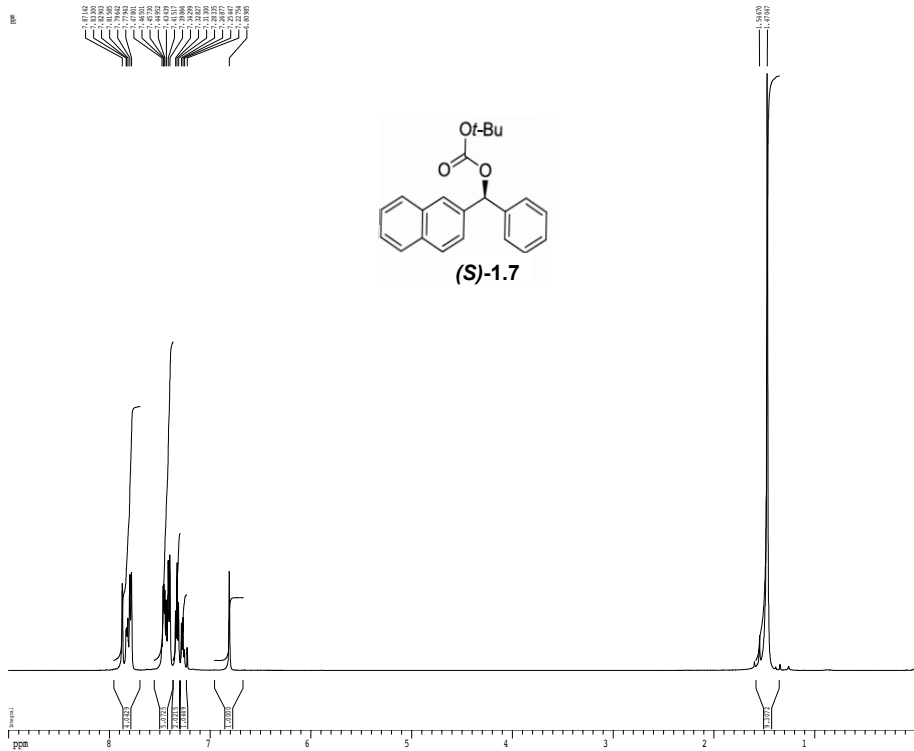
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NUC2         1H
P2           100.00 usec
PL2         1.00 dB
PC2         24.40 dB
SFO2        500.2201011 MHz

===== CHANNEL f3 =====
CPDPRG3     waltz16
NUC3         13C
P3           15.00 usec
PL3         -1.00 dB
PC3         500.2201011 MHz

F2 - Processing parameters
SI          500.220000 MHz
SF          500.220000 MHz
WDW         no
SSB         0
LB          1.00 Hz
GB          0
PC          2.00

ID NMR plot parameters
CX          22.80 cm
CT          15.00 cm
CF          200.000 ppm
F1          2514.08 Hz
F2          0.000 ppm
P2          0.00 Hz
PRCHA      8.71715 ppm/cm
SICH       100.33691 Hz/cm
    
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1H spectrum



Current Data Parameters
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NAME mh-3-123a
EXPNO 1
PROCNO 1

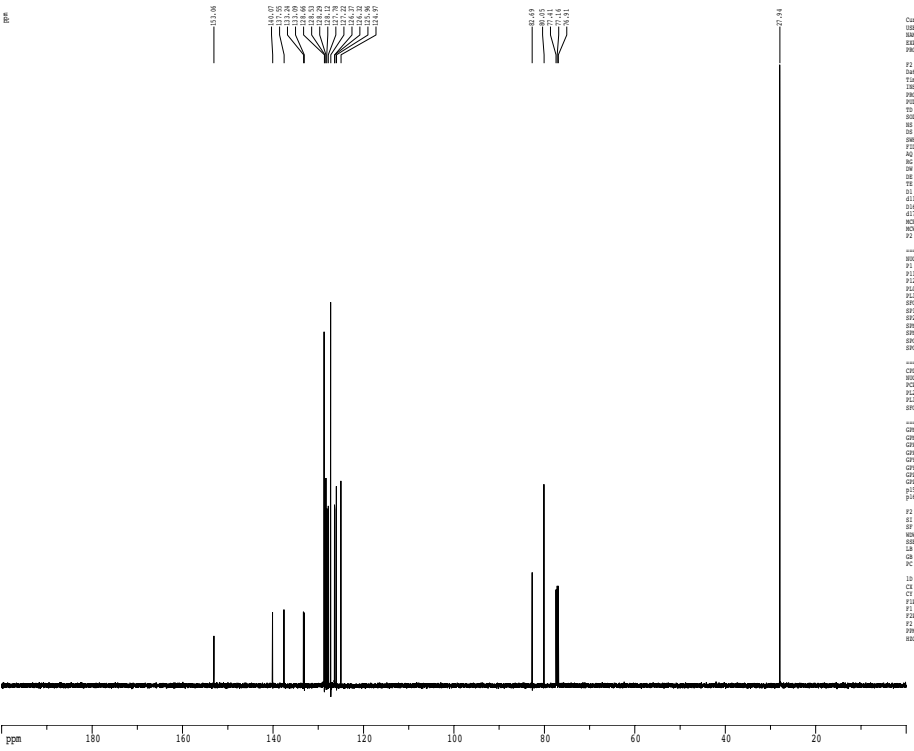
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PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 2
DS 4
SWH 8012.840 Hz
FIDRES 0.208043 Hz
AQ 0.0309281 sec
RG 128
RW 42.450 usec
DE 6.00 usec
TE 298.15 K
D1 0.1000000 sec
dCHIEP 0.0000000 sec
dCMR 0.0150000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 7.00 usec
PL1 1.00 dB
SFO1 100.225013 MHz

F2 - Processing parameters
SI 65536
SF 500.225013 MHz
WDW EM
SS 0
LA 0.20 Hz
GB 4.00

ID NMR plot parameters
CX 22.80 cm
CY 15.00 cm
FID 0.000 ppm
F1 451.58 Hz
F2 0.000 ppm
F3 0.00 Hz
F4 0.000 ppm/cm
F5 197.45328 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
USER greene
NAME mh-3-123a
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20131117
Time 8:57
INSTRUM cryo500
PROBHD 5 mm CPYC 1H-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 2
DS 4
SWH 30203.051 Hz
FIDRES 0.462088 Hz
AQ 1.0184452 sec
RG 4296
RW 18.200 usec
DE 6.00 usec
TE 298.15 K
D1 0.2000000 sec
d11 0.0000000 sec
d12 0.0000000 sec
d13 0.0000000 sec
d14 0.0000000 sec
d15 0.0000000 sec
d16 0.0000000 sec
d17 0.0000000 sec
d18 0.0000000 sec
d19 0.0000000 sec
d20 0.0000000 sec
d21 0.0000000 sec
d22 0.0000000 sec
d23 0.0000000 sec
d24 0.0000000 sec
d25 0.0000000 sec
d26 0.0000000 sec
d27 0.0000000 sec
d28 0.0000000 sec
d29 0.0000000 sec
d30 0.0000000 sec
d31 0.0000000 sec
d32 0.0000000 sec
d33 0.0000000 sec
d34 0.0000000 sec
d35 0.0000000 sec
d36 0.0000000 sec
d37 0.0000000 sec
d38 0.0000000 sec
d39 0.0000000 sec
d40 0.0000000 sec
d41 0.0000000 sec
d42 0.0000000 sec
d43 0.0000000 sec
d44 0.0000000 sec
d45 0.0000000 sec
d46 0.0000000 sec
d47 0.0000000 sec
d48 0.0000000 sec
d49 0.0000000 sec
d50 0.0000000 sec
d51 0.0000000 sec
d52 0.0000000 sec
d53 0.0000000 sec
d54 0.0000000 sec
d55 0.0000000 sec
d56 0.0000000 sec
d57 0.0000000 sec
d58 0.0000000 sec
d59 0.0000000 sec
d60 0.0000000 sec
d61 0.0000000 sec
d62 0.0000000 sec
d63 0.0000000 sec
d64 0.0000000 sec
d65 0.0000000 sec
d66 0.0000000 sec
d67 0.0000000 sec
d68 0.0000000 sec
d69 0.0000000 sec
d70 0.0000000 sec
d71 0.0000000 sec
d72 0.0000000 sec
d73 0.0000000 sec
d74 0.0000000 sec
d75 0.0000000 sec
d76 0.0000000 sec
d77 0.0000000 sec
d78 0.0000000 sec
d79 0.0000000 sec
d80 0.0000000 sec
d81 0.0000000 sec
d82 0.0000000 sec
d83 0.0000000 sec
d84 0.0000000 sec
d85 0.0000000 sec
d86 0.0000000 sec
d87 0.0000000 sec
d88 0.0000000 sec
d89 0.0000000 sec
d90 0.0000000 sec
d91 0.0000000 sec
d92 0.0000000 sec
d93 0.0000000 sec
d94 0.0000000 sec
d95 0.0000000 sec
d96 0.0000000 sec
d97 0.0000000 sec
d98 0.0000000 sec
d99 0.0000000 sec
d100 0.0000000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 15.50 usec
PL1 0.00 usec
PL2 0.00 usec
PL3 0.00 usec
PL4 0.00 usec
PL5 0.00 usec
PL6 0.00 usec
PL7 0.00 usec
PL8 0.00 usec
PL9 0.00 usec
PL10 0.00 usec
PL11 0.00 usec
PL12 0.00 usec
PL13 0.00 usec
PL14 0.00 usec
PL15 0.00 usec
PL16 0.00 usec
PL17 0.00 usec
PL18 0.00 usec
PL19 0.00 usec
PL20 0.00 usec
PL21 0.00 usec
PL22 0.00 usec
PL23 0.00 usec
PL24 0.00 usec
PL25 0.00 usec
PL26 0.00 usec
PL27 0.00 usec
PL28 0.00 usec
PL29 0.00 usec
PL30 0.00 usec
PL31 0.00 usec
PL32 0.00 usec
PL33 0.00 usec
PL34 0.00 usec
PL35 0.00 usec
PL36 0.00 usec
PL37 0.00 usec
PL38 0.00 usec
PL39 0.00 usec
PL40 0.00 usec
PL41 0.00 usec
PL42 0.00 usec
PL43 0.00 usec
PL44 0.00 usec
PL45 0.00 usec
PL46 0.00 usec
PL47 0.00 usec
PL48 0.00 usec
PL49 0.00 usec
PL50 0.00 usec
PL51 0.00 usec
PL52 0.00 usec
PL53 0.00 usec
PL54 0.00 usec
PL55 0.00 usec
PL56 0.00 usec
PL57 0.00 usec
PL58 0.00 usec
PL59 0.00 usec
PL60 0.00 usec
PL61 0.00 usec
PL62 0.00 usec
PL63 0.00 usec
PL64 0.00 usec
PL65 0.00 usec
PL66 0.00 usec
PL67 0.00 usec
PL68 0.00 usec
PL69 0.00 usec
PL70 0.00 usec
PL71 0.00 usec
PL72 0.00 usec
PL73 0.00 usec
PL74 0.00 usec
PL75 0.00 usec
PL76 0.00 usec
PL77 0.00 usec
PL78 0.00 usec
PL79 0.00 usec
PL80 0.00 usec
PL81 0.00 usec
PL82 0.00 usec
PL83 0.00 usec
PL84 0.00 usec
PL85 0.00 usec
PL86 0.00 usec
PL87 0.00 usec
PL88 0.00 usec
PL89 0.00 usec
PL90 0.00 usec
PL91 0.00 usec
PL92 0.00 usec
PL93 0.00 usec
PL94 0.00 usec
PL95 0.00 usec
PL96 0.00 usec
PL97 0.00 usec
PL98 0.00 usec
PL99 0.00 usec
PL100 0.00 usec

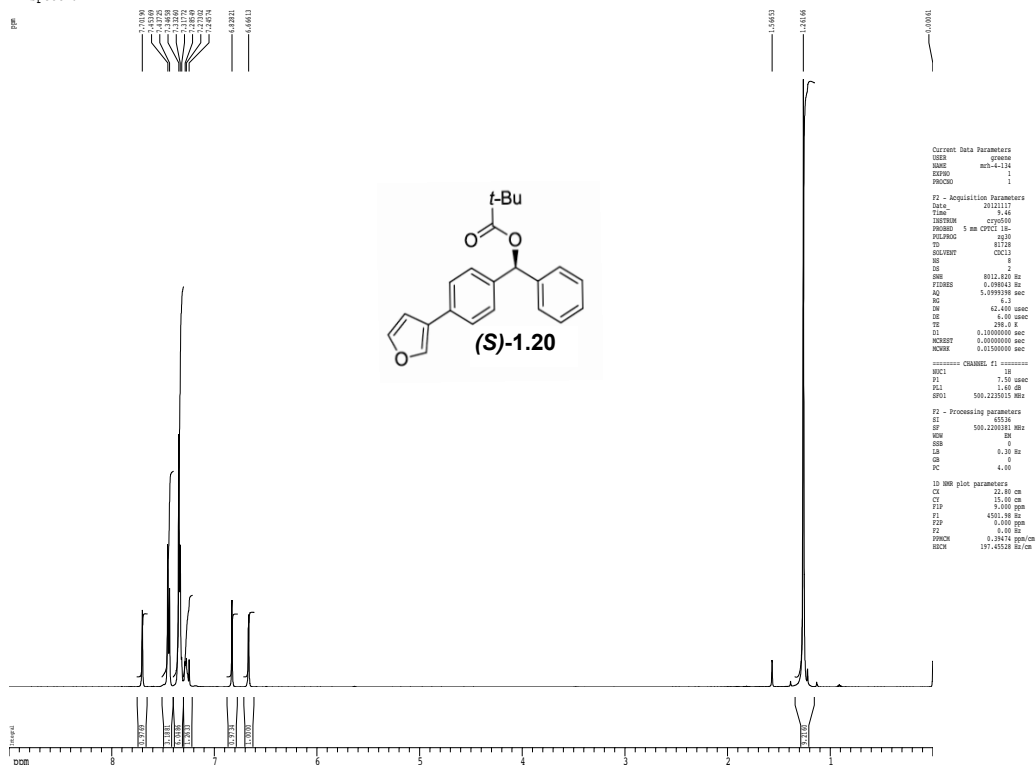
===== CHANNEL f2 =====
CPROG2 waltz16
NUC2 1H
P2P2 100.00 usec
PL2 1.00 dB
PL12 24.00 dB
SFO2 500.225013 MHz

===== GRABBY CHANNEL =====
GRAB1 5188.100
GRAB2 5188.100
GRAB3 5188.100
GRAB4 5188.100
GRAB5 5188.100
GRAB6 5188.100
GRAB7 5188.100
GRAB8 5188.100
GRAB9 5188.100
GRAB10 5188.100
GRAB11 5188.100
GRAB12 5188.100
GRAB13 5188.100
GRAB14 5188.100
GRAB15 5188.100
GRAB16 5188.100
GRAB17 5188.100
GRAB18 5188.100
GRAB19 5188.100
GRAB20 5188.100
GRAB21 5188.100
GRAB22 5188.100
GRAB23 5188.100
GRAB24 5188.100
GRAB25 5188.100
GRAB26 5188.100
GRAB27 5188.100
GRAB28 5188.100
GRAB29 5188.100
GRAB30 5188.100
GRAB31 5188.100
GRAB32 5188.100
GRAB33 5188.100
GRAB34 5188.100
GRAB35 5188.100
GRAB36 5188.100
GRAB37 5188.100
GRAB38 5188.100
GRAB39 5188.100
GRAB40 5188.100
GRAB41 5188.100
GRAB42 5188.100
GRAB43 5188.100
GRAB44 5188.100
GRAB45 5188.100
GRAB46 5188.100
GRAB47 5188.100
GRAB48 5188.100
GRAB49 5188.100
GRAB50 5188.100
GRAB51 5188.100
GRAB52 5188.100
GRAB53 5188.100
GRAB54 5188.100
GRAB55 5188.100
GRAB56 5188.100
GRAB57 5188.100
GRAB58 5188.100
GRAB59 5188.100
GRAB60 5188.100
GRAB61 5188.100
GRAB62 5188.100
GRAB63 5188.100
GRAB64 5188.100
GRAB65 5188.100
GRAB66 5188.100
GRAB67 5188.100
GRAB68 5188.100
GRAB69 5188.100
GRAB70 5188.100
GRAB71 5188.100
GRAB72 5188.100
GRAB73 5188.100
GRAB74 5188.100
GRAB75 5188.100
GRAB76 5188.100
GRAB77 5188.100
GRAB78 5188.100
GRAB79 5188.100
GRAB80 5188.100
GRAB81 5188.100
GRAB82 5188.100
GRAB83 5188.100
GRAB84 5188.100
GRAB85 5188.100
GRAB86 5188.100
GRAB87 5188.100
GRAB88 5188.100
GRAB89 5188.100
GRAB90 5188.100
GRAB91 5188.100
GRAB92 5188.100
GRAB93 5188.100
GRAB94 5188.100
GRAB95 5188.100
GRAB96 5188.100
GRAB97 5188.100
GRAB98 5188.100
GRAB99 5188.100
GRAB100 5188.100

F2 - Processing parameters
SI 65536
SF 125.761440 MHz
WDW EM
SS 0
LA 0.00 Hz
GB 0
PC 2.00

ID NMR plot parameters
CX 22.80 cm
CY 15.00 cm
FID 200.000 ppm
F1 2519.208 Hz
F2 0.000 ppm
F3 0.000 Hz
F4 0.000 ppm/cm
F5 1163.23601 Hz/cm

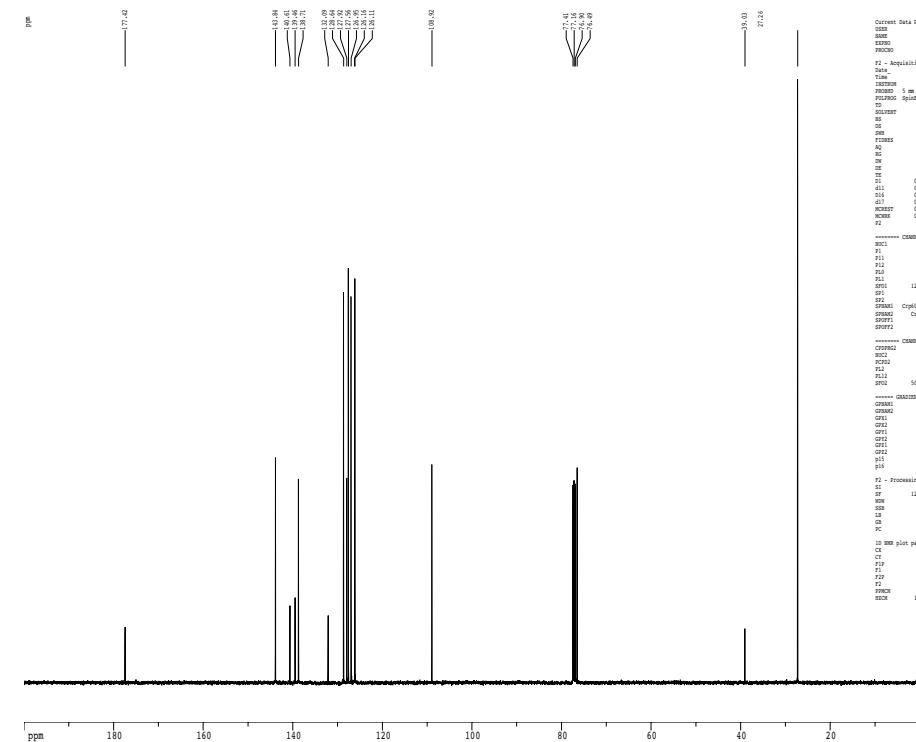
1H spectrum



```

Current Data Parameters
NAME          gsmse
EXPNO         1
PROCNO       1
F2 - Acquisition Parameters
Date_         201111
Time         9.45
INSTRUM      crys60
PROBHD       5 mm CPXI 1H-
PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           2
DS           2
SWH          8012.420 Hz
FREQZ       0.099643 Hz
AQ          5.099338 sec
RG           6.3
DM          42.400 usec
DE          6.00 usec
TE          298.2 K
SI          0.1000000 sec
SFO         0.0000000 sec
MCHSET      0.0000000 sec
MCHW        0.0000000 sec
===== CHANNEL f1 =====
NUC1         13C
P1          7.00 usec
PL1         1.00 dB
SFO1        500.1325115 MHz
F2 - Processing parameters
SI          65536
SF          500.1325115 MHz
WDW         EM
SSB         0
LA          0.30 Hz
GB          0
PC          4.00
IS MRB plot parameters
CX          22.80 cm
CY          15.00 cm
F1P         9.400 ppm
F2P         4501.88 Hz
F3P         0.000 ppm
F4          0.00 Hz
F5          0.00 Hz
F6          0.100 Hz
F7          0.100 Hz
F8          197.45528 Hz/cm
  
```

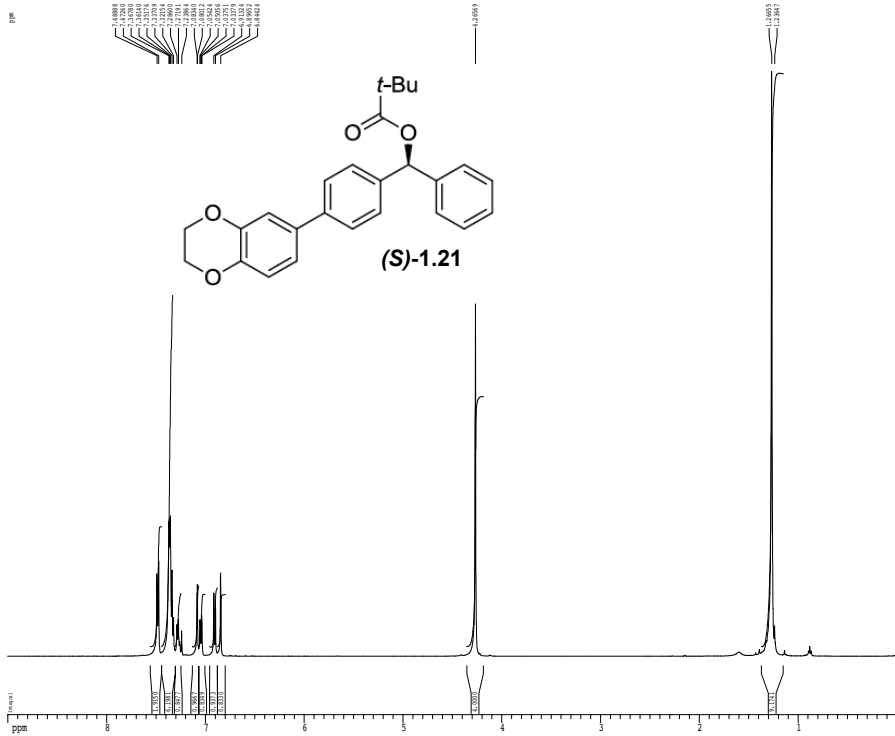
Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
NAME          gsmse
EXPNO         1
PROCNO       1
F2 - Acquisition Parameters
Date_         201111
Time         9.54
INSTRUM      crys60
PROBHD       5 mm CPXI 1H-
PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           2
DS           2
SWH          3089.020 Hz
FREQZ       0.041200 Hz
AQ          1.041100 sec
RG           7.0
DM          14.500 usec
DE          6.00 usec
TE          298.2 K
SI          0.2000000 sec
SFO         0.0000000 sec
MCHSET      0.0000000 sec
MCHW        0.0000000 sec
===== CHANNEL f1 =====
NUC1         13C
P1          1.00 usec
PL1         1.00 dB
SFO1        500.1325115 MHz
PL2         2000.00 dB
PL3         10.00 dB
PL4         10.00 dB
PL5         10.00 dB
PL6         10.00 dB
PL7         10.00 dB
PL8         10.00 dB
PL9         10.00 dB
PL10        10.00 dB
PL11        10.00 dB
PL12        10.00 dB
PL13        10.00 dB
PL14        10.00 dB
PL15        10.00 dB
PL16        10.00 dB
PL17        10.00 dB
PL18        10.00 dB
PL19        10.00 dB
PL20        10.00 dB
===== CHANNEL f2 =====
CPDPRG2     waltz16
NUC2         13C
P2          1.00 usec
PL2         1.00 dB
SFO2        100.6261500 MHz
PL3         20.00 dB
PL4         10.00 dB
PL5         10.00 dB
===== CHANNEL CHANDEL =====
CPDPRG3     zgpg30
NUC3         13C
P3          1.00 usec
PL3         1.00 dB
SFO3        100.6261500 MHz
PL4         10.00 dB
PL5         10.00 dB
PL6         10.00 dB
PL7         10.00 dB
PL8         10.00 dB
PL9         10.00 dB
PL10        10.00 dB
PL11        10.00 dB
PL12        10.00 dB
PL13        10.00 dB
PL14        10.00 dB
PL15        10.00 dB
PL16        10.00 dB
PL17        10.00 dB
PL18        10.00 dB
PL19        10.00 dB
PL20        10.00 dB
F2 - Processing parameters
SI          65536
SF          125.7604128 MHz
WDW         EM
SSB         0
LA          1.00 Hz
GB          0
PC          2.00
IS MRB plot parameters
CX          22.80 cm
CY          15.00 cm
F1P         250.000 ppm
F2P         250.000 ppm
F3P         0.000 ppm
F4          0.00 Hz
F5          0.00 Hz
F6          0.100 Hz
F7          0.100 Hz
F8          100.6261500 Hz/cm
F9          100.6261500 Hz/cm
  
```

1H spectrum



Current Data Parameters
USER greeno
NAME mH-4-128
EXPNO 1
PROCNO 1

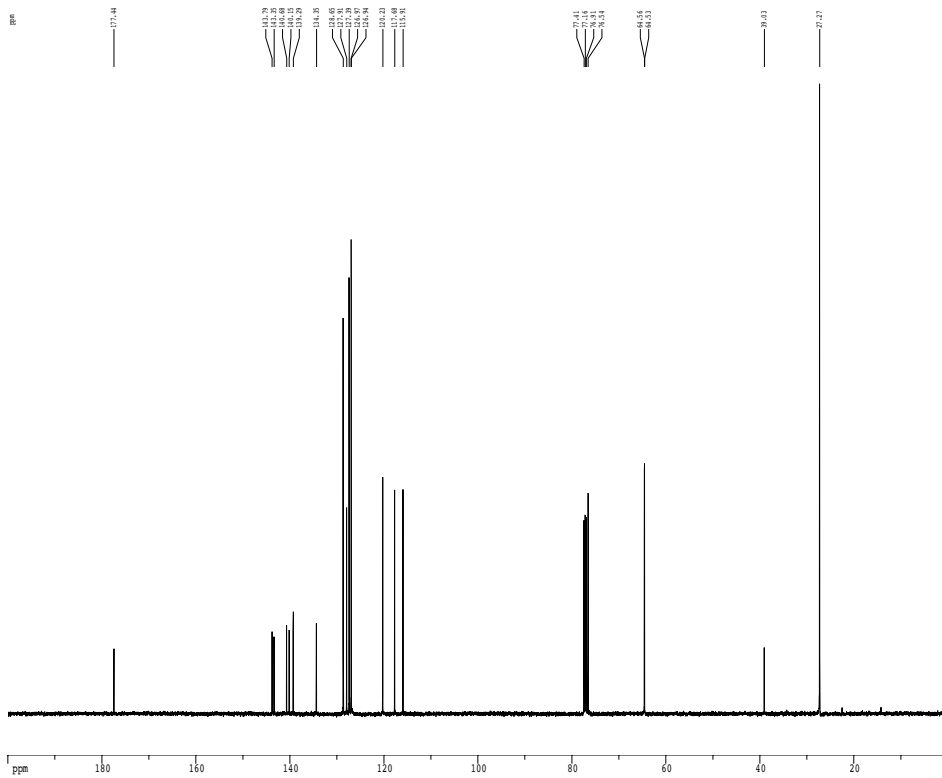
F2 - Acquisition Parameters
Date_ 20111117
Time 9.14
INSTRUM cryo50
PROBHD 5 mm CPMCI 1H-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 2
DS 2
SWH 8012.820 Hz
FIDRES 0.00040 Hz
AQ 5.000000 sec
RG 65
RG 62.400 usec
DE 6.00 usec
TE 298.2 K
SI 0.1000000 sec
IC 0.0000000 sec
MCHSET 0.0000000 sec
MCHRX 0.0000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 7.50 usec
PL1 1.00 dB
SFO1 500.200000 MHz

F2 - Processing parameters
SI 65536
SF 500.200000 MHz
WDW EM
SSB 0
LA 0.20 Hz
GB 0
PC 4.00

D0 DMR plot parameters
CX 22.00 cm
CY 22.00 cm
FLP 0.000 ppm
FL 4001.000 Hz
FZP 0.000 ppm
FZ 0.00 Hz
PPMCH 0.00174 ppm/cm
SFOCH 197.60328 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
USER greeno
NAME mH-4-128
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111117
Time 9.20
INSTRUM cryo50
PROBHD 5 mm CPMCI 1H-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 2
DS 2
SWH 30302.010 Hz
FIDRES 0.442208 Hz
AQ 1.0844100 sec
RG 655
RG 16.500 usec
DE 6.00 usec
TE 298.2 K
SI 0.2500000 sec
IC 0.0000000 sec
MCHSET 0.0000000 sec
MCHRX 0.0000000 sec
F2 13.00 usec

===== CHANNEL f1 =====
NUC1 13C
P1 15.00 usec
PL1 500.00 usec
PL2 2000.00 usec
PL3 120.00 dB
PL4 -1.00 dB
SFO1 125.764104 MHz
SFO2 77.000 MHz
SFO3 101.626100 MHz
SFO4 Cpu60, 6.5, 101.7
SFO5 Cpu60, 6.5, 101.7
SFO6 0.00 Hz
SFO7 0.00 Hz
SFO8 0.00 Hz

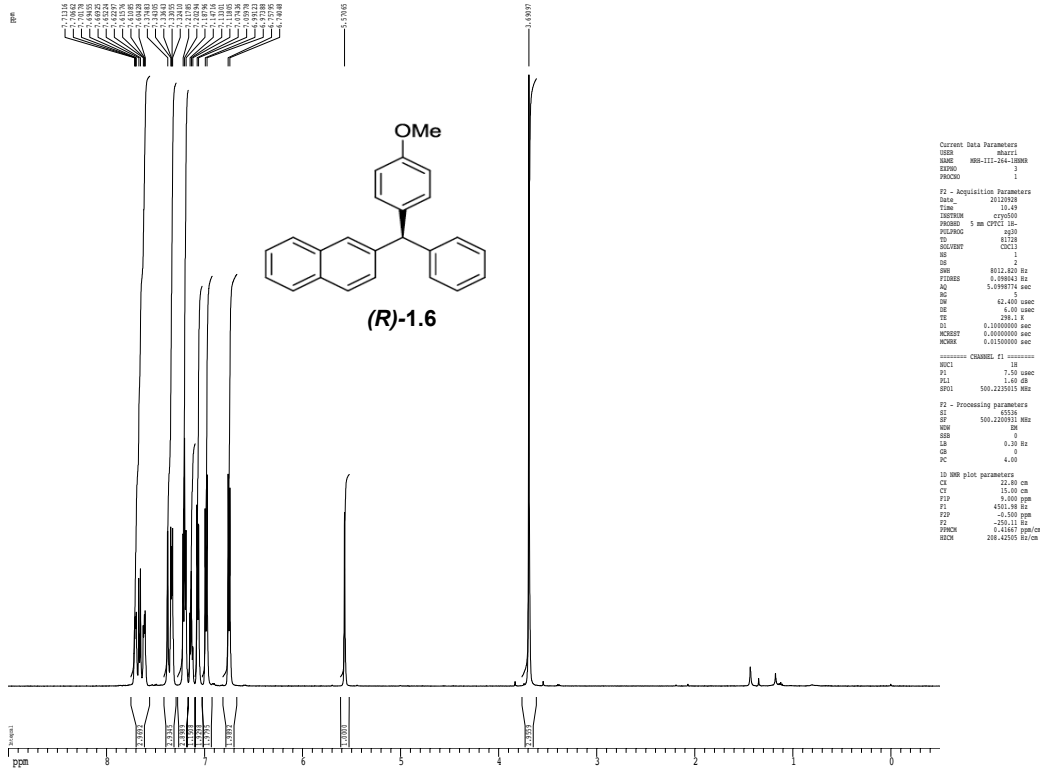
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 180.00 usec
PL2 1.00 dB
PL12 24.00 dB
SFO2 500.200000 MHz

===== CHANNEL CHANNE1 =====
CPDPRG1 BIRD-100
CPDPRG2 BIRD-100
SFO1 0.00 Hz
SFO2 0.00 Hz
SFO3 0.00 Hz
SFO4 0.00 Hz
SFO5 0.00 Hz
SFO6 0.00 Hz
SFO7 0.00 Hz
SFO8 0.00 Hz

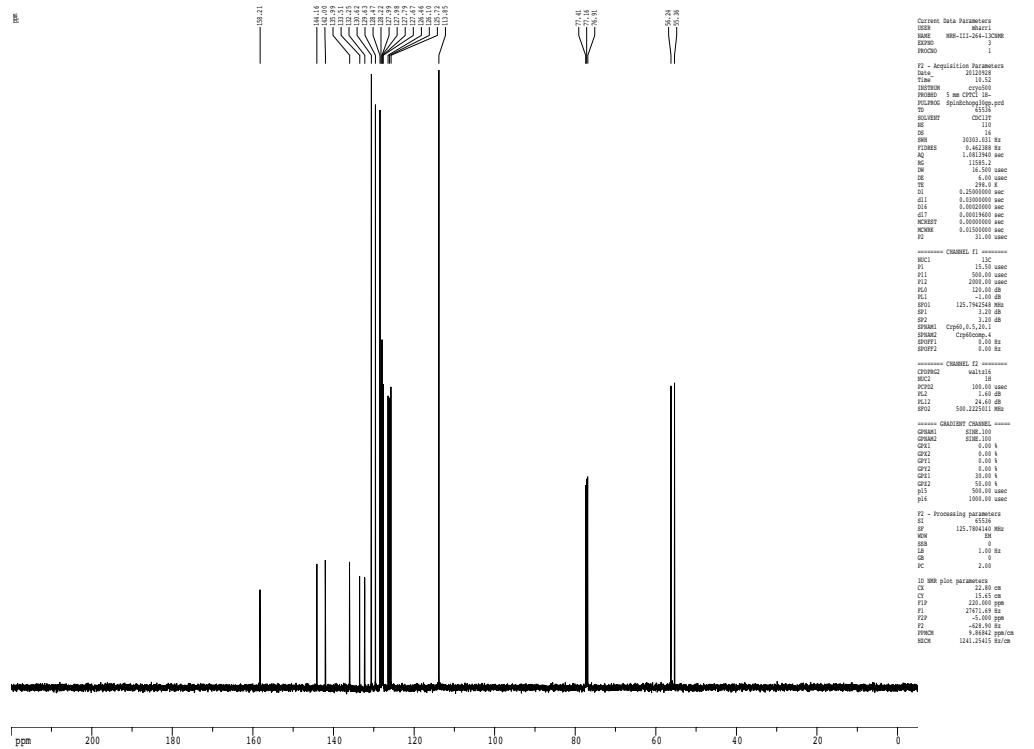
F2 - Processing parameters
SI 65536
SF 125.764104 MHz
WDW EM
SSB 0
LA 1.00 Hz
GB 0
PC 2.00

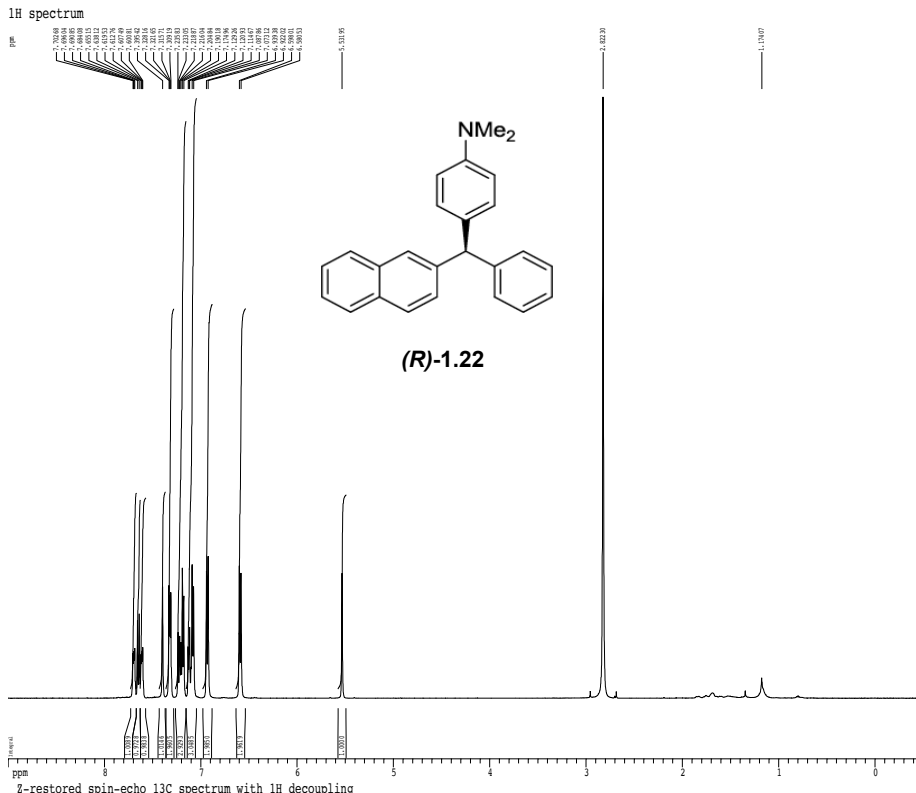
D0 DMR plot parameters
CX 22.00 cm
CY 22.00 cm
FLP 200.000 ppm
FL 2016.000 Hz
FZP 0.000 ppm
FZ 0.00 Hz
PPMCH 8.77192 ppm/cm
SFOCH 100.626100 Hz/cm

1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling





```

Current Data Parameters
USER      MARI1
NAME      M88-112-168-120M
EXPNO    1
PROCNO   1

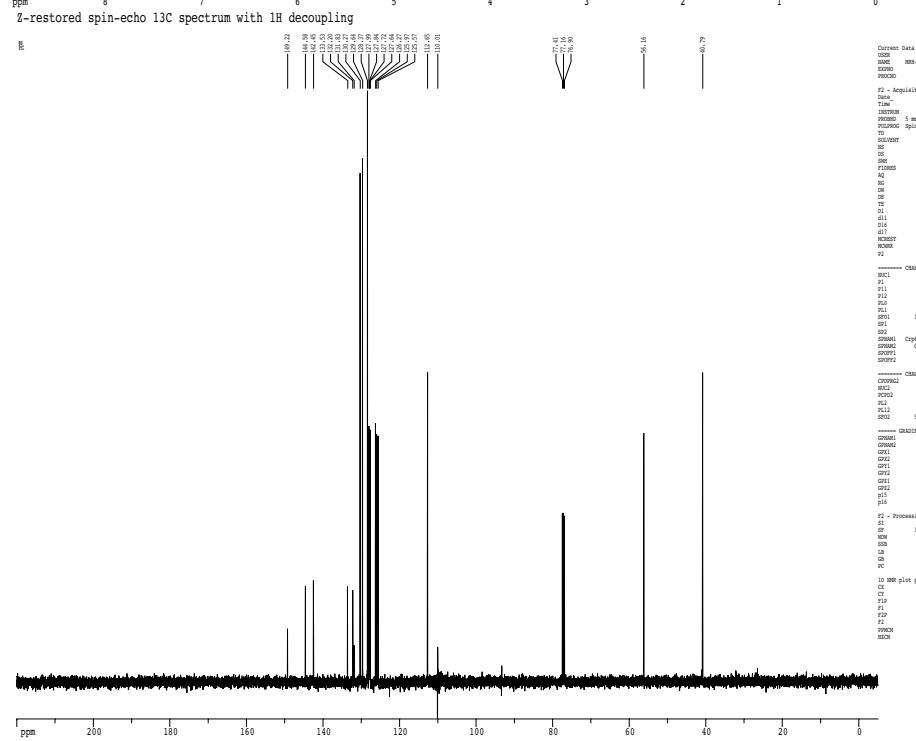
F2 - Acquisition Parameters
Date_    20120128
Time     15:10
INSTRUM  spect
PROBHD   5 mm CPDQ 1H-
PULPROG  zgpg30
SFO      500
SOLVENT  CDCl3
NS       2
DS       1
SWH      8012.820 Hz
FIDRES   0.09842 Hz
AQ       5.8997714 sec
RG        0
WDW      EM
SSB      0
LB      61.400 usec
GB       0
TE       298.2 K
C1       0.10000000 sec
MCHSET   0.00000000 sec
MCHWD    0.01000000 sec

===== CHANNEL f1 =====
NUC1      13C
P1        12.00 usec
PL1       0.00 dB
SFO1      125.761144 MHz

F2 - Processing parameters
SI        65536
SF        500.136092 MHz
WDW      EM
SSB      0
LB      0.00 Hz
GB       0
TE       298.2 K
PC       4.00

1D 1H NMR parameters
SI        65536
SF        500.136092 MHz
WDW      EM
SSB      0
LB      0.00 Hz
GB       0
TE       298.2 K
PC       4.00

1D 13C NMR parameters
SI        65536
SF        125.761144 MHz
WDW      EM
SSB      0
LB      0.00 Hz
GB       0
TE       298.2 K
PC       4.00
  
```



```

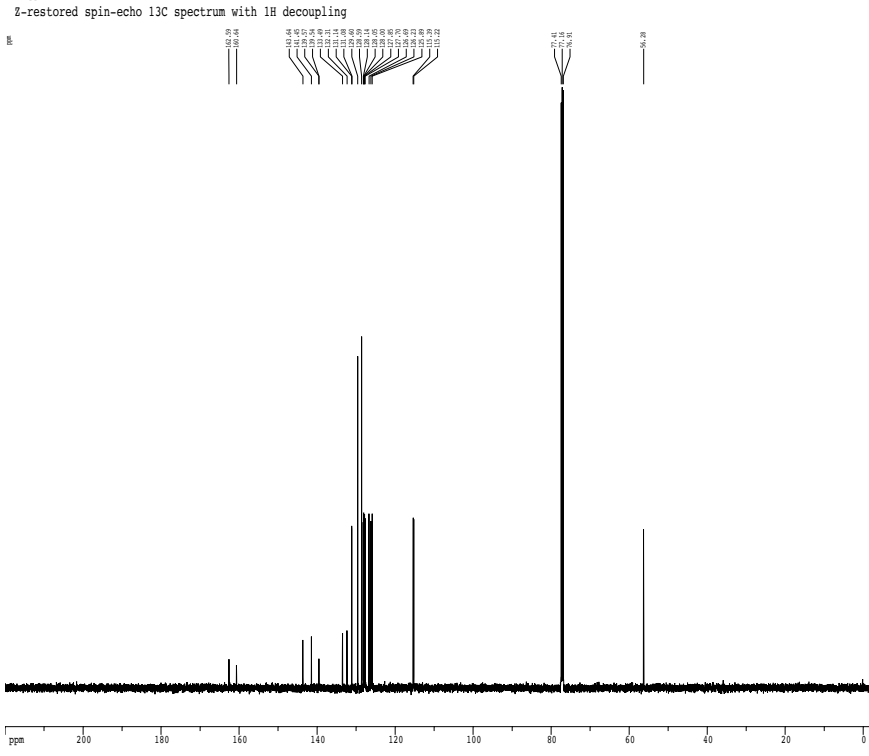
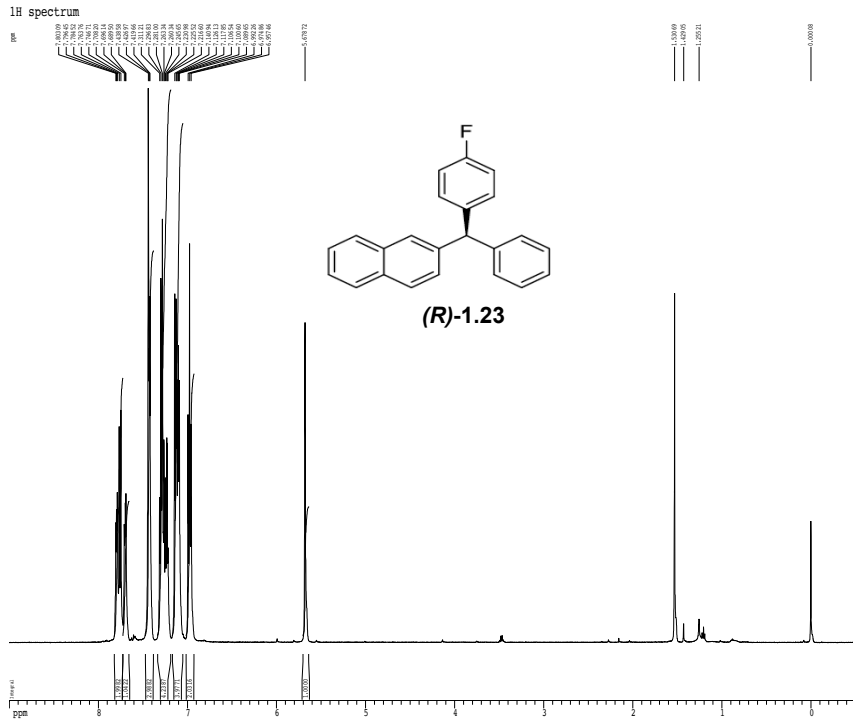
Current Data Parameters
USER      MARI1
NAME      M88-112-168-120M
EXPNO    1
PROCNO   1

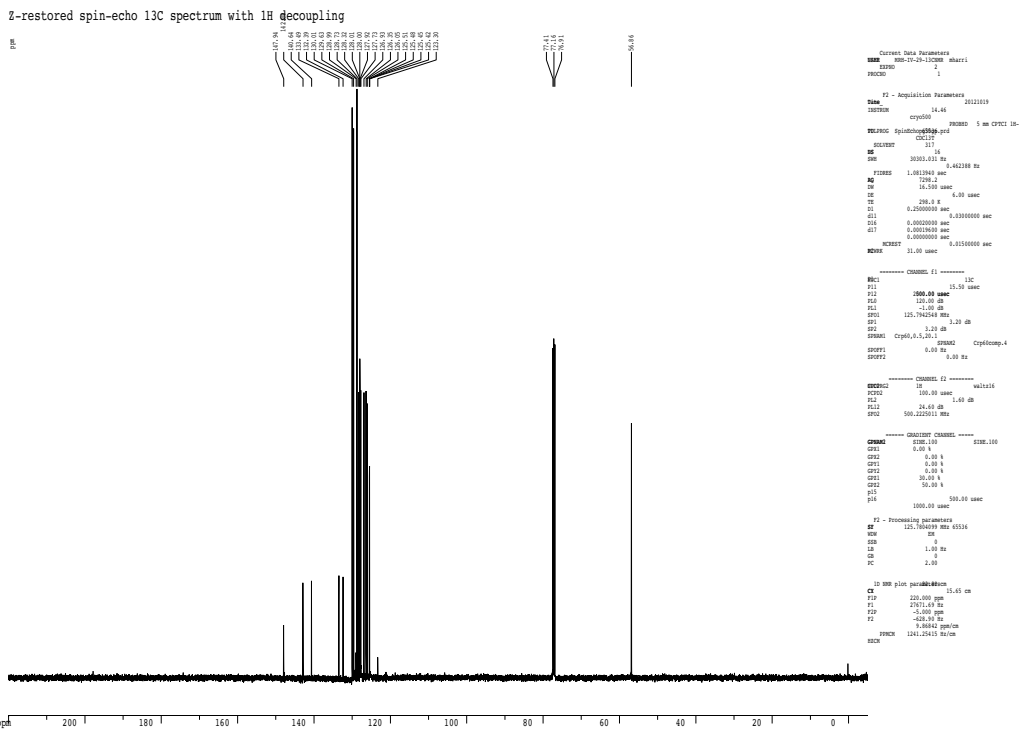
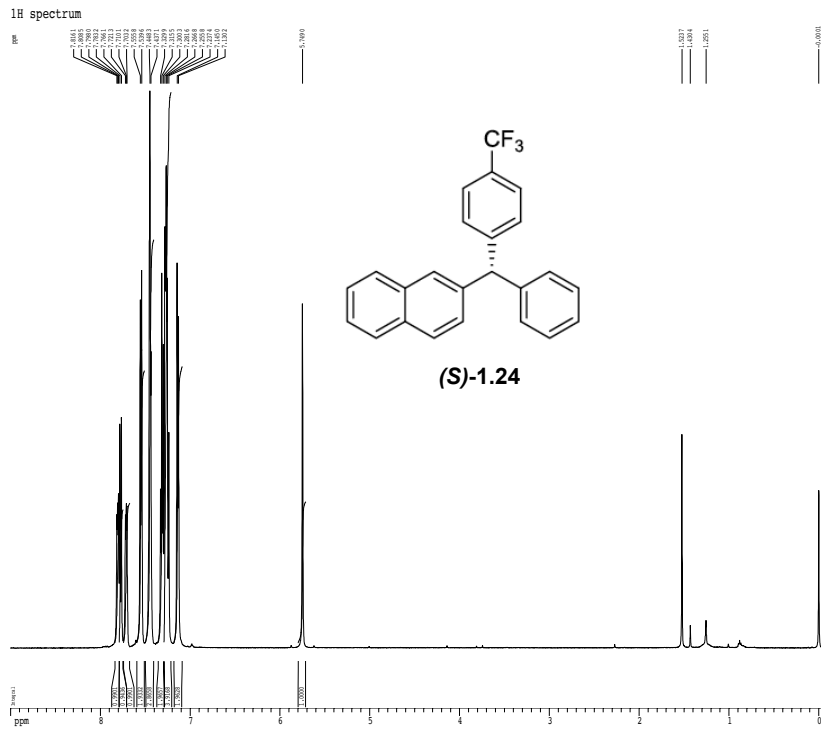
F2 - Acquisition Parameters
Date_    20120128
Time     15:17
INSTRUM  spect
PROBHD   5 mm CPDQ 1H-
PULPROG  zgpg30
SFO      500
SOLVENT  CDCl3
NS       2
DS       1
SWH      8012.820 Hz
FIDRES   0.09842 Hz
AQ       5.8997714 sec
RG        0
WDW      EM
SSB      0
LB      61.400 usec
GB       0
TE       298.2 K
C1       0.10000000 sec
MCHSET   0.00000000 sec
MCHWD    0.01000000 sec

===== CHANNEL f1 =====
NUC1      13C
P1        12.00 usec
PL1       0.00 dB
SFO1      125.761144 MHz

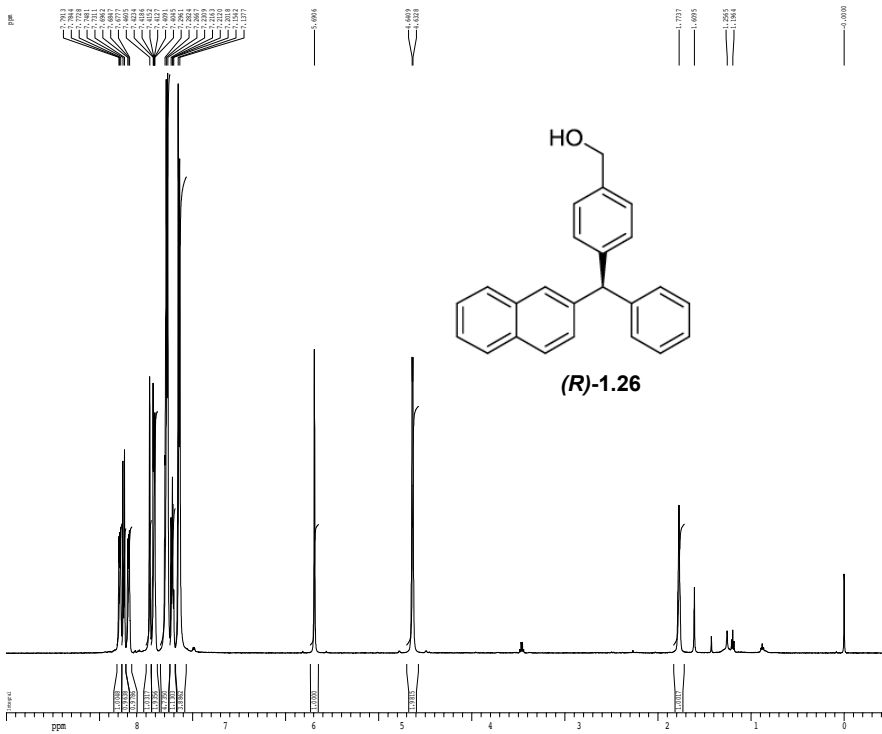
F2 - Processing parameters
SI        65536
SF        500.136092 MHz
WDW      EM
SSB      0
LB      0.00 Hz
GB       0
TE       298.2 K
PC       4.00

1D 13C NMR parameters
SI        65536
SF        125.761144 MHz
WDW      EM
SSB      0
LB      0.00 Hz
GB       0
TE       298.2 K
PC       4.00
  
```





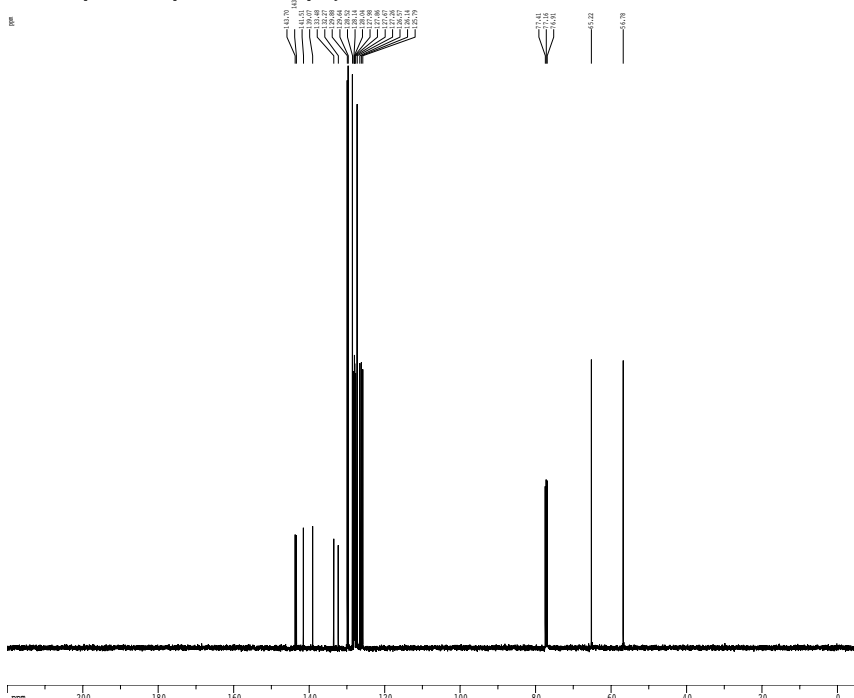
¹H spectrum



```

Current Data Parameters
NAME: 08-19-201008
EXPNO: 2
PROCNO: 1
F2 - Acquisition Parameters
Date_ : 20121013
Time : 14.34
INSTRUM : spect
PROBHD : 5 mm CPXI 13
PULPROG : zgpg30
TD : 65536
SOLVENT : DMS
NS : 6128
DS : 4
AQ : 802.620 Hz
RG : 0.000000 Hz
AQ : 5.099874 sec
RG : 3.2
DE : 62.400 usec
TE : 6.20 usec
TE : 298.2 K
SI : 0.1000000 sec
MCHSET : 0.0000000 sec
MCHSX : 0.0000000 sec
===== CHANNEL f1 =====
NUC1 : 13C
P1 : 1.50 usec
PL1 : 0.00 dB
SFO1 : 500.225011 MHz
F2 - Processing parameters
SI : 65536
SF : 500.225011 MHz
WDW : EM
SSB : 0
LA : 0.20 Hz
GB : 0
PC : 4.00
1D 13C NMR plot parameters
CX : 22.80 cm
CY : 10.00 cm
F1P : 0.000 ppm
F1 : 4501.90 Hz
F2P : -0.500 ppm
F2 : 4501.10 Hz
PRNC : 0.4164 ppm/cm
SDCN : 208.4200 Hz/cm
    
```

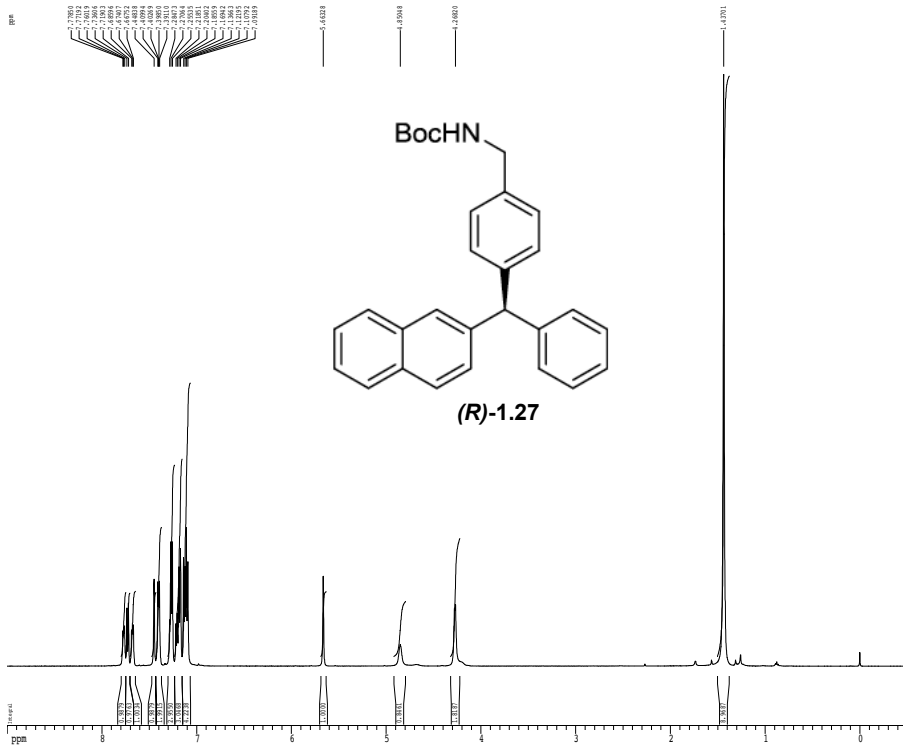
¹³C-restored spin-echo 13C spectrum with ¹H decoupling



```

Current Data Parameters
NAME: 08-19-201008
EXPNO: 1
PROCNO: 1
F2 - Acquisition Parameters
Date_ : 20121013
Time : 14.34
INSTRUM : spect
PROBHD : 5 mm CPXI 13
PULPROG : zgpg30
TD : 65536
SOLVENT : DMS
NS : 6128
DS : 4
AQ : 802.620 Hz
RG : 0.000000 Hz
AQ : 5.099874 sec
RG : 3.2
DE : 62.400 usec
TE : 6.20 usec
TE : 298.2 K
SI : 0.1000000 sec
MCHSET : 0.0000000 sec
MCHSX : 0.0000000 sec
===== CHANNEL f1 =====
NUC1 : 13C
P1 : 1.50 usec
PL1 : 0.00 dB
SFO1 : 500.225011 MHz
F2 - Processing parameters
SI : 65536
SF : 500.225011 MHz
WDW : EM
SSB : 0
LA : 0.20 Hz
GB : 0
PC : 4.00
1D 13C NMR plot parameters
CX : 22.80 cm
CY : 10.00 cm
F1P : 0.000 ppm
F1 : 4501.90 Hz
F2P : -0.500 ppm
F2 : 4501.10 Hz
PRNC : 0.4164 ppm/cm
SDCN : 208.4200 Hz/cm
    
```

1H spectrum



```

Current Data Parameters
NAME      M0811
NAME      M08-111-294-12000
EXPNO     1
PROCNO    1

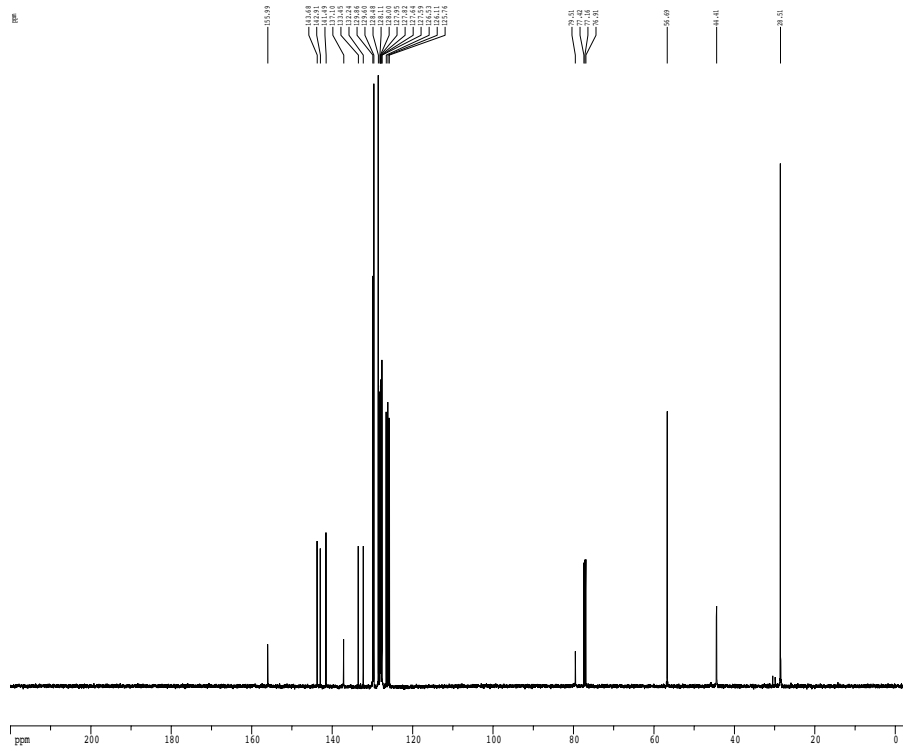
F2 - Acquisition Parameters
Date_     20121015
Time      15:33
INSTRUM   cryo500
PROBHD    5 mm CPXI 1H-
PULPROG   zgpg30
TD         65536
SFO       500.136261 MHz
AQ         0.18128
RG         682
AQ         0.18128
RG         682
SI         1
SF         500.136261 MHz
NUC1       13C
NUC2       13C
PC         1
PC2        1
PC3        1
PC4        1
PC5        1
PC6        1
PC7        1
PC8        1
PC9        1
PC10       1
PC11       1
PC12       1
PC13       1
PC14       1
PC15       1
PC16       1
PC17       1
PC18       1
PC19       1
PC20       1
PC21       1
PC22       1
PC23       1
PC24       1
PC25       1
PC26       1
PC27       1
PC28       1
PC29       1
PC30       1
PC31       1
PC32       1
PC33       1
PC34       1
PC35       1
PC36       1
PC37       1
PC38       1
PC39       1
PC40       1
PC41       1
PC42       1
PC43       1
PC44       1
PC45       1
PC46       1
PC47       1
PC48       1
PC49       1
PC50       1
PC51       1
PC52       1
PC53       1
PC54       1
PC55       1
PC56       1
PC57       1
PC58       1
PC59       1
PC60       1
PC61       1
PC62       1
PC63       1
PC64       1
PC65       1
PC66       1
PC67       1
PC68       1
PC69       1
PC70       1
PC71       1
PC72       1
PC73       1
PC74       1
PC75       1
PC76       1
PC77       1
PC78       1
PC79       1
PC80       1
PC81       1
PC82       1
PC83       1
PC84       1
PC85       1
PC86       1
PC87       1
PC88       1
PC89       1
PC90       1
PC91       1
PC92       1
PC93       1
PC94       1
PC95       1
PC96       1
PC97       1
PC98       1
PC99       1
PC100      1

===== CHANNEL f1 =====
NUC1       13C
P1         15.00 usec
PL1        0.00 dB
SFO1       125.764249 MHz
===== CHANNEL f2 =====
NUC1       13C
P1         15.00 usec
PL1        0.00 dB
SFO1       125.764249 MHz
===== CHANNEL f3 =====
NUC1       13C
P1         15.00 usec
PL1        0.00 dB
SFO1       125.764249 MHz

F2 - Processing parameters
SI         65536
SF         500.136261 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00

ID NMR plot parameters
SI         65536
SF         500.136261 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00
  
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
NAME      M0811
NAME      M08-111-294-12000
EXPNO     1
PROCNO    1

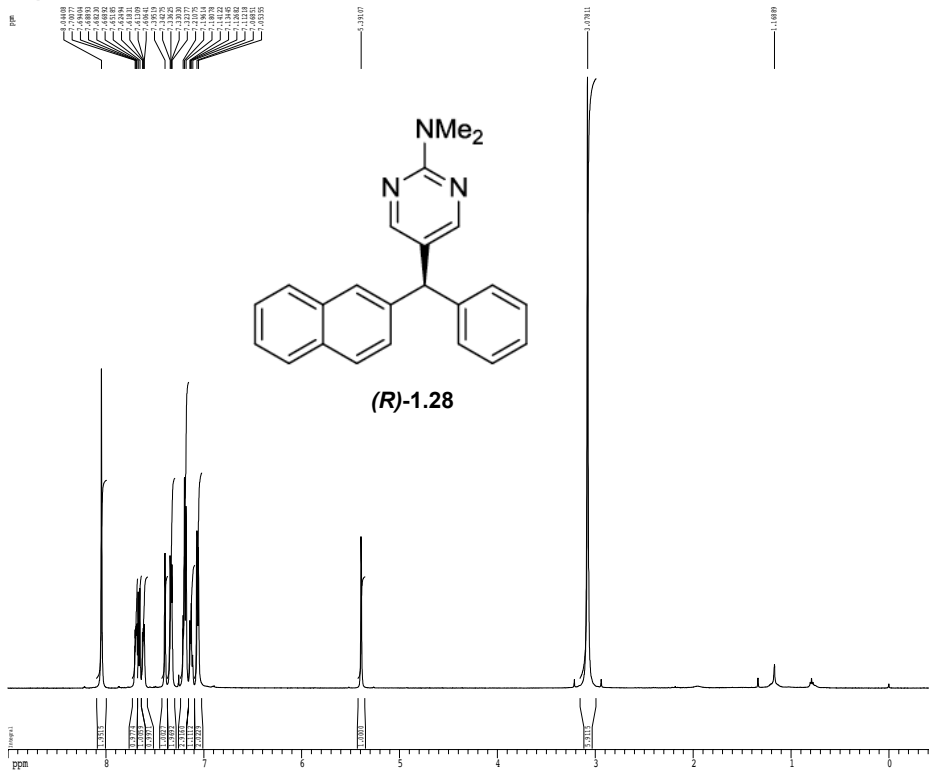
F2 - Acquisition Parameters
Date_     20121015
Time      15:35
INSTRUM   cryo500
PROBHD    5 mm CPXI 1H-
PULPROG   zgpg30
TD         65536
SFO       500.136261 MHz
AQ         0.18128
RG         682
AQ         0.18128
RG         682
SI         1
SF         500.136261 MHz
NUC1       13C
NUC2       13C
PC         1
PC2        1
PC3        1
PC4        1
PC5        1
PC6        1
PC7        1
PC8        1
PC9        1
PC10       1
PC11       1
PC12       1
PC13       1
PC14       1
PC15       1
PC16       1
PC17       1
PC18       1
PC19       1
PC20       1
PC21       1
PC22       1
PC23       1
PC24       1
PC25       1
PC26       1
PC27       1
PC28       1
PC29       1
PC30       1
PC31       1
PC32       1
PC33       1
PC34       1
PC35       1
PC36       1
PC37       1
PC38       1
PC39       1
PC40       1
PC41       1
PC42       1
PC43       1
PC44       1
PC45       1
PC46       1
PC47       1
PC48       1
PC49       1
PC50       1
PC51       1
PC52       1
PC53       1
PC54       1
PC55       1
PC56       1
PC57       1
PC58       1
PC59       1
PC60       1
PC61       1
PC62       1
PC63       1
PC64       1
PC65       1
PC66       1
PC67       1
PC68       1
PC69       1
PC70       1
PC71       1
PC72       1
PC73       1
PC74       1
PC75       1
PC76       1
PC77       1
PC78       1
PC79       1
PC80       1
PC81       1
PC82       1
PC83       1
PC84       1
PC85       1
PC86       1
PC87       1
PC88       1
PC89       1
PC90       1
PC91       1
PC92       1
PC93       1
PC94       1
PC95       1
PC96       1
PC97       1
PC98       1
PC99       1
PC100      1

===== CHANNEL f1 =====
NUC1       13C
P1         15.00 usec
PL1        0.00 dB
SFO1       125.764249 MHz
===== CHANNEL f2 =====
NUC1       13C
P1         15.00 usec
PL1        0.00 dB
SFO1       125.764249 MHz
===== CHANNEL f3 =====
NUC1       13C
P1         15.00 usec
PL1        0.00 dB
SFO1       125.764249 MHz

F2 - Processing parameters
SI         65536
SF         500.136261 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00

ID NMR plot parameters
SI         65536
SF         500.136261 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00
  
```

1H spectrum



```

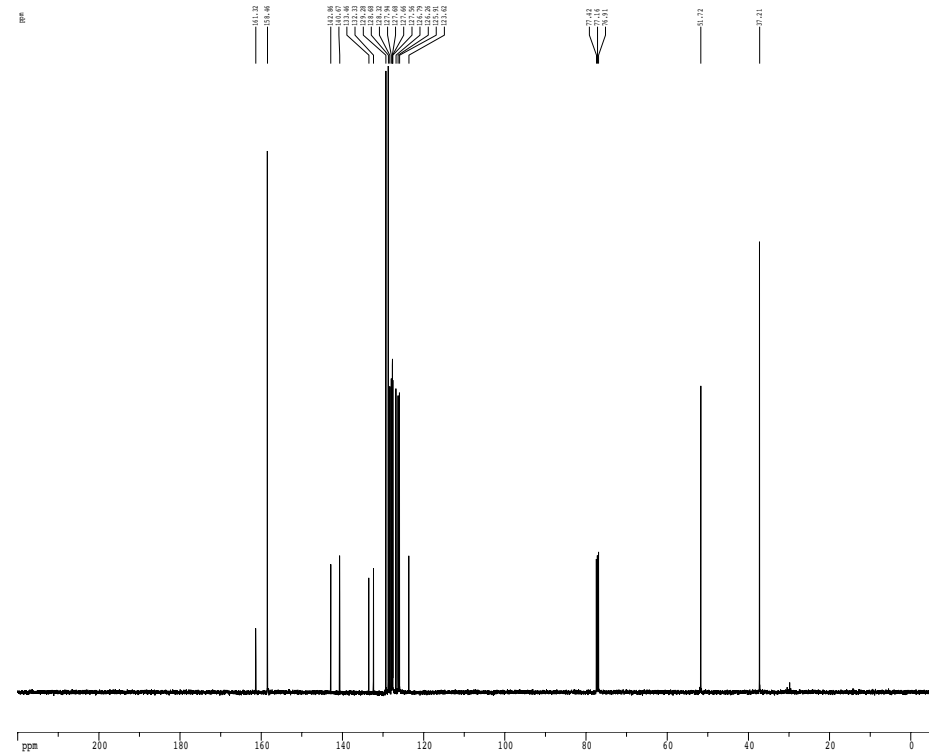
Current Data Parameters
NAME      HMR11
EXPNO    1
PROCNO   1
----- Acquisition Parameters
Date_     20120928
Time      11:19
INSTRUM   cryo500
PROBHD    5 mm CPXI 1h-
PULPROG   zgpg30
PC        40.729
SOLVENT   CDCl3
NS         2
DS         2
SFO       501.821 MHz
FIDRES    0.988643 Hz
AQ        0.5098734 sec
RG         4.5
OF        43.409 umsec
DE        6.00 umsec
TE        298.2 K
D1         0.10000000 sec
d11        0.00000000 sec
NOREST    0.00000000 sec
NORMX     0.15000000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         7.50 umsec
PL1        0.00 dB
SFO1       500.1255015 MHz

F2 - Processing parameters
SI         32768
SF         500.1255015 MHz
WDW        EM
SSB        0
LA         0.00 Hz
GB         0
PC        4.00

1D NMR plot parameters
SI         32768
SF         500.1255015 MHz
WDW        EM
SSB        0
LA         0.00 Hz
GB         0
PC        4.00
  
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
NAME      HMR11
EXPNO    1
PROCNO   1
----- Acquisition Parameters
Date_     20120918
Time      11:19
INSTRUM   cryo500
PROBHD    5 mm CPXI 1h-
PULPROG   zgpg30
PC        40.729
SOLVENT   CDCl3
NS         2
DS         2
SFO       501.821 MHz
FIDRES    0.988643 Hz
AQ        0.5098734 sec
RG         4.5
OF        43.409 umsec
DE        6.00 umsec
TE        298.2 K
D1         0.10000000 sec
d11        0.00000000 sec
NOREST    0.00000000 sec
NORMX     0.15000000 sec

===== CHANNEL f1 =====
NUC1       13C
P1         15.00 umsec
PL1        0.00 dB
SFO1       125.7603117 MHz

===== CHANNEL f2 =====
NAME      Multis
NUC2       13C
P2         100.00 umsec
PL2        0.00 dB
SFO2       125.7603117 MHz

===== CHANNEL f3 =====
NAME      Cryo6h-1.5.10-1
SFO3       1.5 MHz
SFO3A     1.5000000 MHz
SFO3B     1.5000000 MHz

===== CHANNEL f4 =====
NAME      Cryo6homp-4
SFO4       0.40 MHz
SFO4A     0.4000000 MHz
SFO4B     0.4000000 MHz

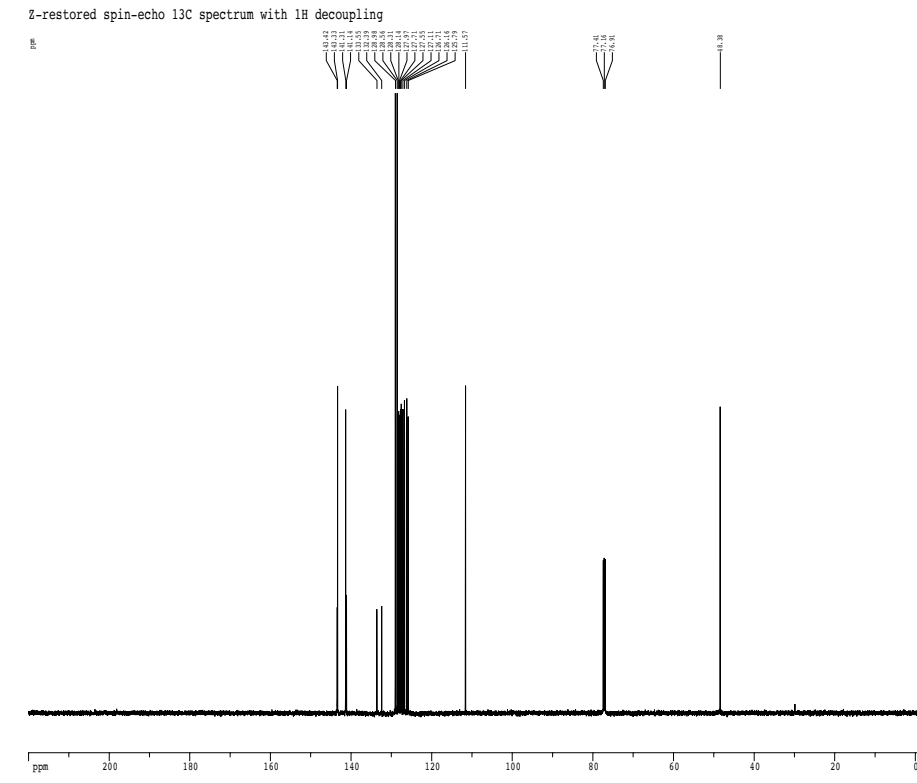
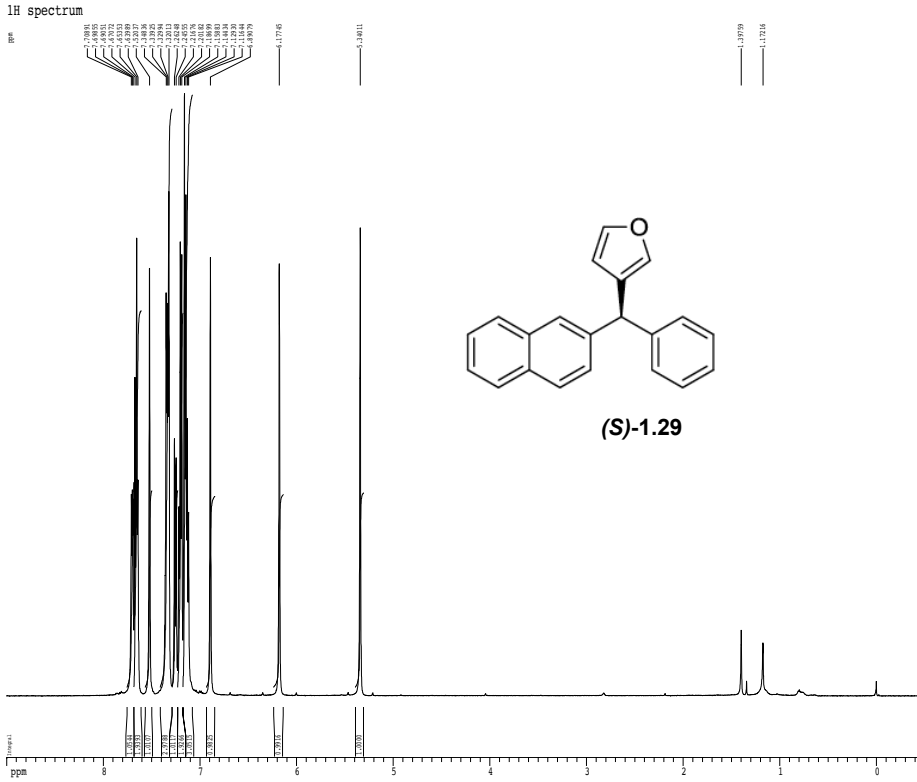
===== CHANNEL f5 =====
NAME      Multis
NUC5       13C
P5         100.00 umsec
PL5        0.00 dB
SFO5       125.7603117 MHz

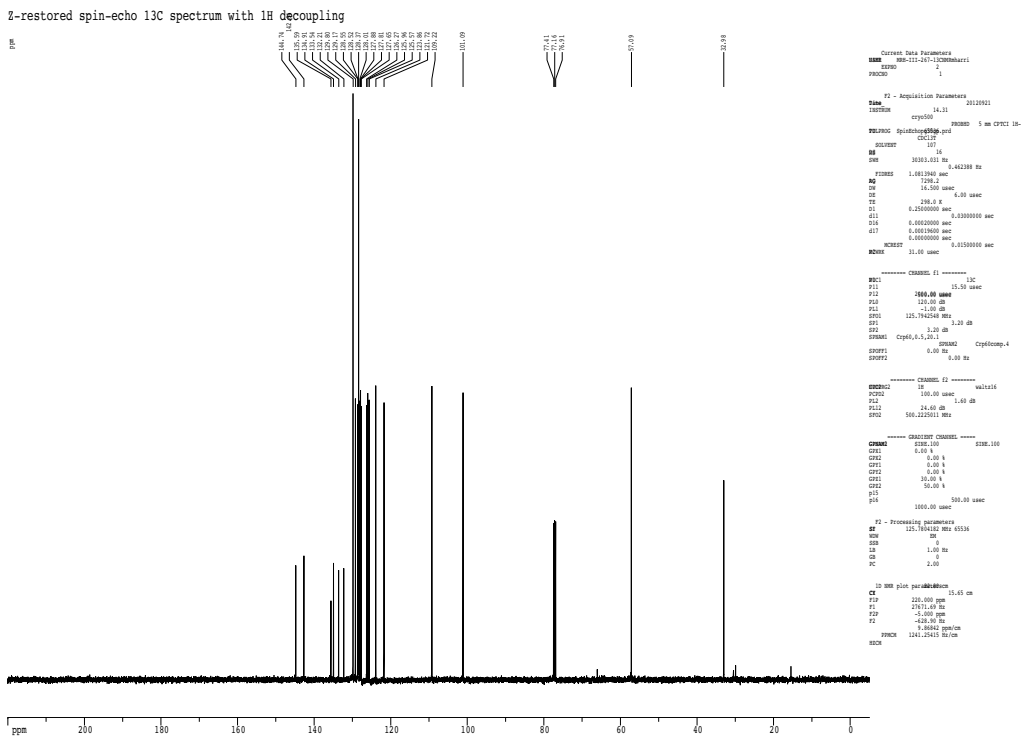
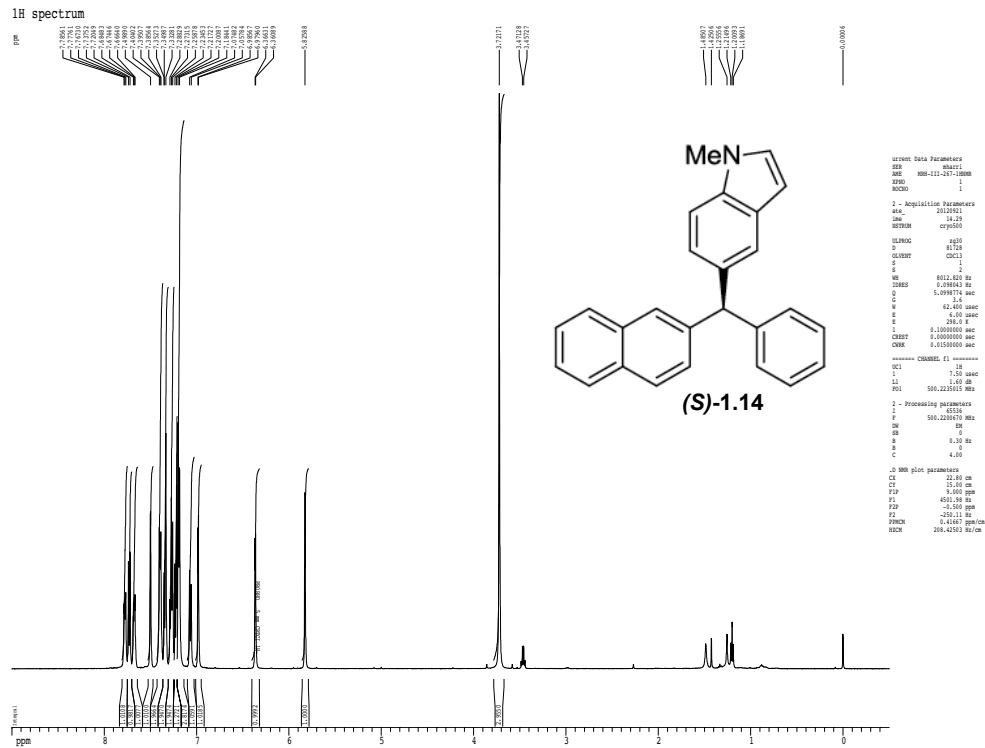
===== CHANNEL f6 =====
NAME      Cryo6h-1.5.10-1
SFO6       1.5 MHz
SFO6A     1.5000000 MHz
SFO6B     1.5000000 MHz

===== CHANNEL f7 =====
NAME      Cryo6homp-4
SFO7       0.40 MHz
SFO7A     0.4000000 MHz
SFO7B     0.4000000 MHz

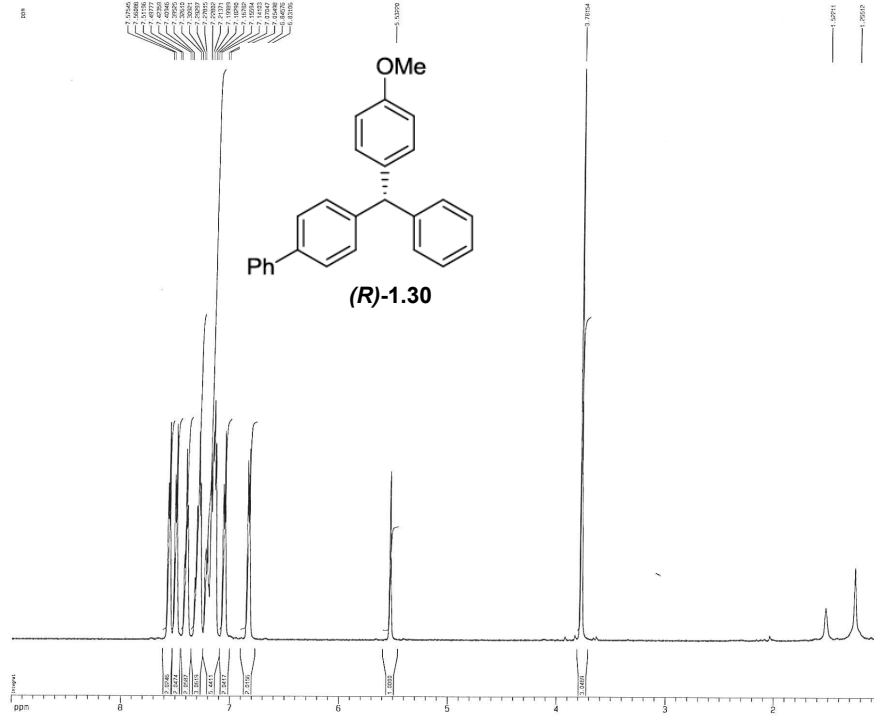
F2 - Processing parameters
SI         65536
SF         125.7603117 MHz
WDW        EM
SSB        0
LA         1.00 Hz
GB         0
PC        2.00

1D NMR plot parameters
SI         65536
SF         125.7603117 MHz
WDW        EM
SSB        0
LA         1.00 Hz
GB         0
PC        2.00
  
```





1H spectrum



```

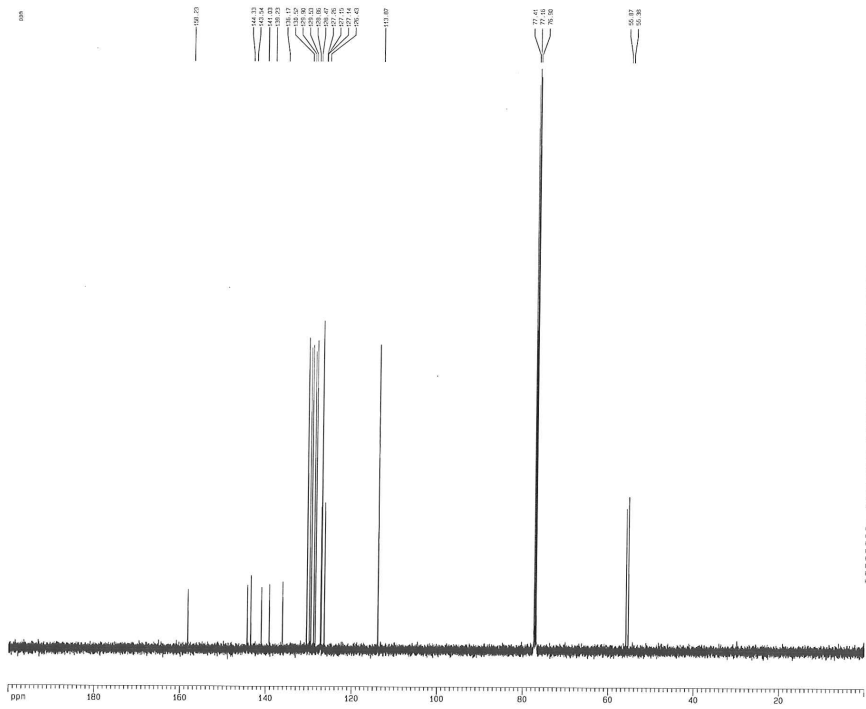
Current Data Parameters
=====
NAME      13C-1-118-1P
EXPNO    1
PROCNO   1
F2 - Acquisition Parameters
=====
Date_    20120313
Time     16:44
INSTRUM  spect
PROBHD   5 mm 1H/13
PULPROG  zgpg30
TD        65536
SOLVENT  CDCl3
NS        8
DS        4
SWH       8000.000 Hz
FIDRES    0.0001000 Hz
AQ         0.0001000 sec
RG         655.37
RM         0.0200000 sec
DE         0.0000000 sec
TE         300.2 K
SI         0.0000000 sec
RGHRT      0.0000000 sec
MCKW      0.0000000 sec
MCKK      0.0000000 sec

***** CHANNEL f1 *****
NUC1      13C
P1         12.00 usec
PL1        -1.00 dB
SFO1      101.62539 MHz

F2 - Processing parameters
=====
SI         65536
SF         101.62539 MHz
WDW        EM
SSB        0
LB         0.00 Hz
GB         0
PC         1.00

13C NMR list parameters
=====
CA         100.00 usec
CH1        15.00 usec
F1P        0.0000000 sec
F2         400.0000000 Hz
F2P        1.0000000 sec
F2         101.62539 MHz
PCMCW     0.0000000 sec
HDCR     170.0000000 Hz
    
```

13C spectrum with 1H decoupling



```

Current Data Parameters
=====
NAME      13C-1-118-1P
EXPNO    1
PROCNO   1
F2 - Acquisition Parameters
=====
Date_    20120313
Time     16:47
INSTRUM  spect
PROBHD   5 mm 1H/13
PULPROG  zgpg30
TD        65536
SOLVENT  CDCl3
NS        8
DS        4
SWH       8000.000 Hz
FIDRES    0.0001000 Hz
AQ         0.0001000 sec
RG         655.37
RM         0.0200000 sec
DE         0.0000000 sec
TE         300.2 K
SI         0.0000000 sec
RGHRT      0.0000000 sec
MCKW      0.0000000 sec
MCKK      0.0000000 sec

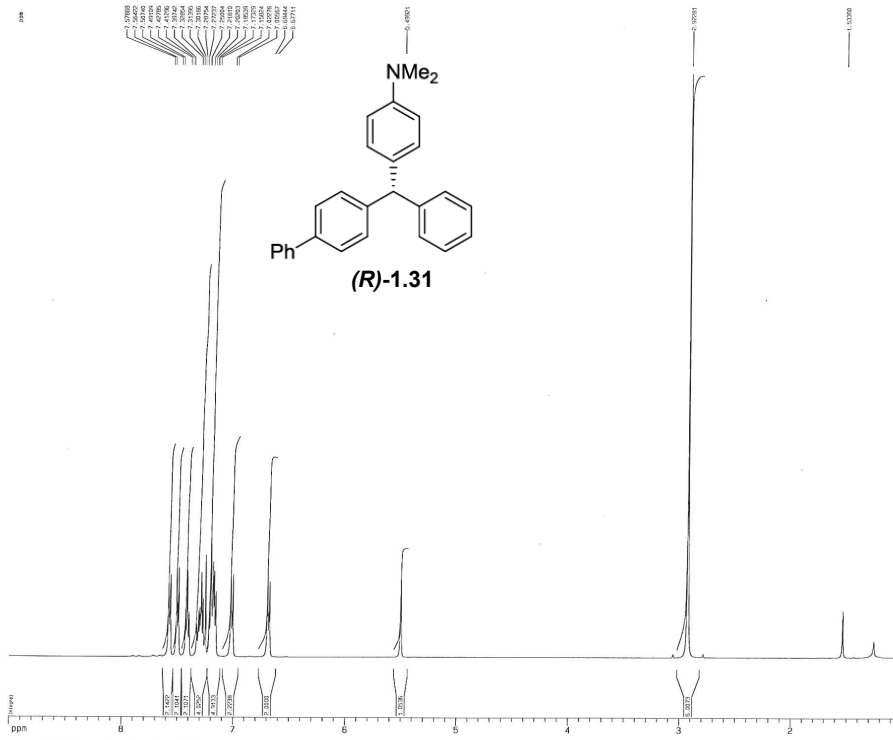
***** CHANNEL f1 *****
NUC1      13C
P1         12.00 usec
PL1        -1.00 dB
SFO1      101.62539 MHz

***** CHANNEL f2 *****
NUC2      1H
P2         12.00 usec
PL2        -1.00 dB
SFO2      400.14639 MHz

F2 - Processing parameters
=====
SI         65536
SF         101.62539 MHz
WDW        EM
SSB        0
LB         0.00 Hz
GB         0
PC         1.00

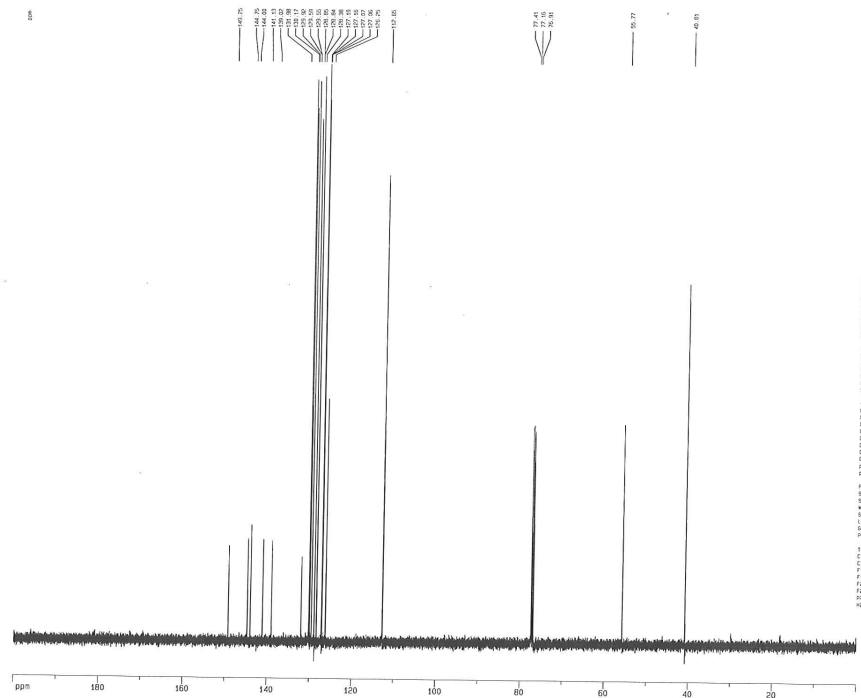
13C NMR list parameters
=====
CA         100.00 usec
CH1        15.00 usec
F1P        0.0000000 sec
F2         400.14639 MHz
F2P        1.0000000 sec
F2         101.62539 MHz
PCMCW     0.0000000 sec
HDCR     170.0000000 Hz
    
```


1H spectrum

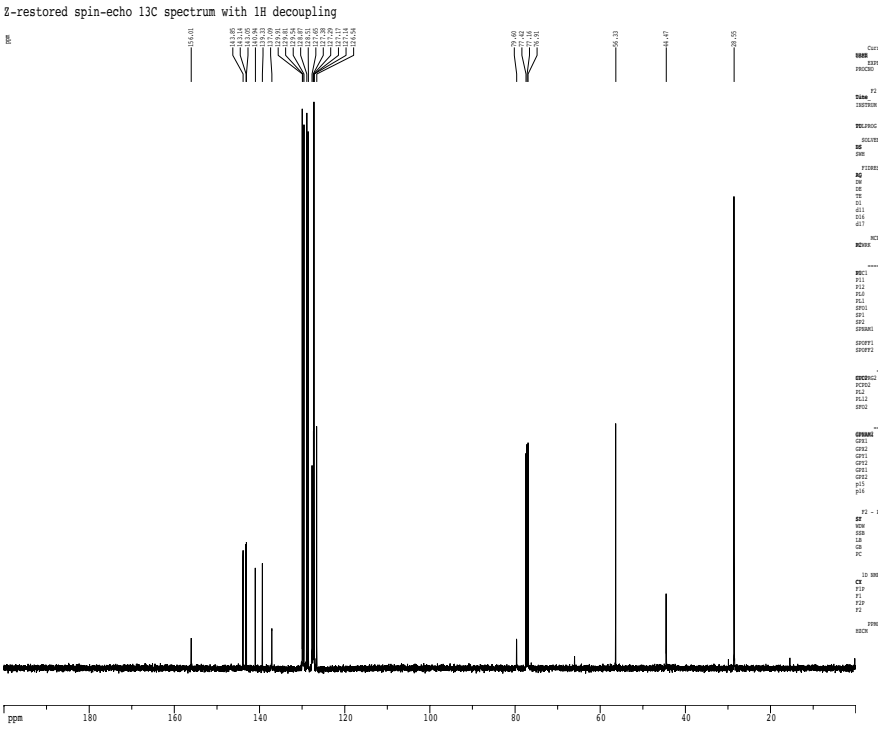
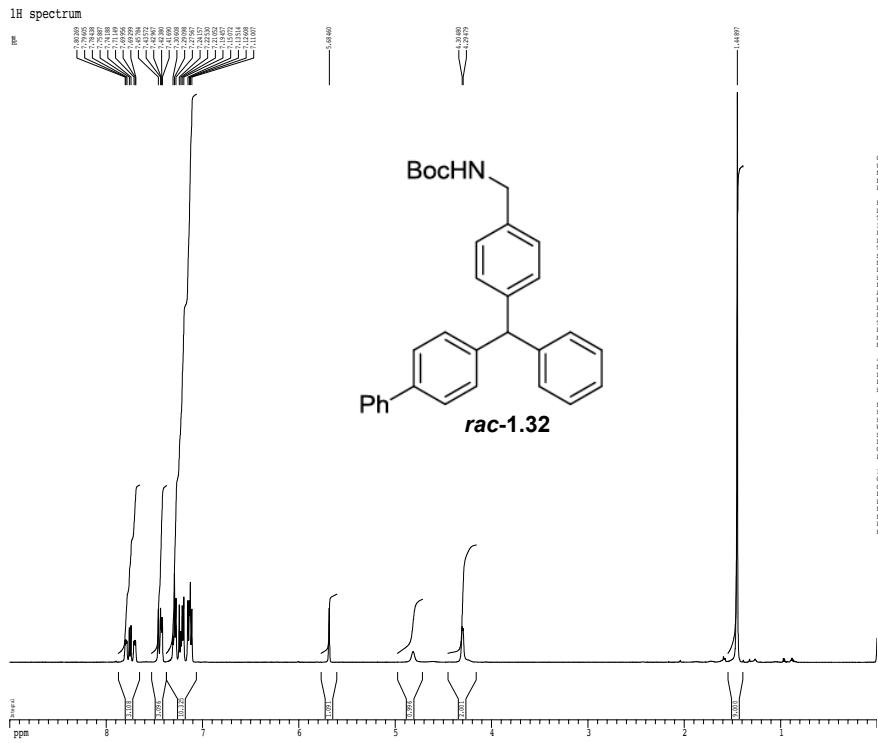


Current Data Parameters
 QEXP 1000000
 NAME LHM-1-148-13-125-2
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 DATE_ 20100909
 TIME 14:35
 INSTRUM spect
 PULPROG zgpg30
 NUC1 13C
 NUC2 1H
 SOLVENT CDCl3
 NS 2
 DS 2
 SWH 8019.850 Hz
 FIDRES 0.00049 Hz
 AQ 0.0000000 sec
 RG 327
 EQ 9.7
 DE 8.00 uMm
 TE 300.2 K
 D1 0.0000000 sec
 MCHST 0.0000000 sec
 MCKK 0.0000000 sec
 ***** CHANNEL f1 *****
 NUC1 1H
 P1 15.00 uMm
 PL1 0.00 dB
 SFO1 500.1360500 MHz
 F2 - Processing parameters
 SI 65536
 SF 500.1360500 MHz
 DS 4
 SWH 8019.850 Hz
 EQ 9.7
 DE 8.00 uMm
 TE 300.2 K
 D1 0.0000000 sec
 MCHST 0.0000000 sec
 MCKK 0.0000000 sec
 F1 4000.000 Hz
 F2 100.626126 Hz
 F3 100.626126 Hz
 F4 100.626126 Hz
 F5 100.626126 Hz
 F6 100.626126 Hz
 F7 100.626126 Hz
 F8 100.626126 Hz
 F9 100.626126 Hz
 F10 100.626126 Hz
 F11 100.626126 Hz
 F12 100.626126 Hz
 F13 100.626126 Hz
 F14 100.626126 Hz
 F15 100.626126 Hz
 F16 100.626126 Hz
 F17 100.626126 Hz
 F18 100.626126 Hz
 F19 100.626126 Hz
 F20 100.626126 Hz
 F21 100.626126 Hz
 F22 100.626126 Hz
 F23 100.626126 Hz
 F24 100.626126 Hz
 F25 100.626126 Hz
 F26 100.626126 Hz
 F27 100.626126 Hz
 F28 100.626126 Hz
 F29 100.626126 Hz
 F30 100.626126 Hz
 F31 100.626126 Hz
 F32 100.626126 Hz
 F33 100.626126 Hz
 F34 100.626126 Hz
 F35 100.626126 Hz
 F36 100.626126 Hz
 F37 100.626126 Hz
 F38 100.626126 Hz
 F39 100.626126 Hz
 F40 100.626126 Hz
 F41 100.626126 Hz
 F42 100.626126 Hz
 F43 100.626126 Hz
 F44 100.626126 Hz
 F45 100.626126 Hz
 F46 100.626126 Hz
 F47 100.626126 Hz
 F48 100.626126 Hz
 F49 100.626126 Hz
 F50 100.626126 Hz
 F51 100.626126 Hz
 F52 100.626126 Hz
 F53 100.626126 Hz
 F54 100.626126 Hz
 F55 100.626126 Hz
 F56 100.626126 Hz
 F57 100.626126 Hz
 F58 100.626126 Hz
 F59 100.626126 Hz
 F60 100.626126 Hz
 F61 100.626126 Hz
 F62 100.626126 Hz
 F63 100.626126 Hz
 F64 100.626126 Hz
 F65 100.626126 Hz
 F66 100.626126 Hz
 F67 100.626126 Hz
 F68 100.626126 Hz
 F69 100.626126 Hz
 F70 100.626126 Hz
 F71 100.626126 Hz
 F72 100.626126 Hz
 F73 100.626126 Hz
 F74 100.626126 Hz
 F75 100.626126 Hz
 F76 100.626126 Hz
 F77 100.626126 Hz
 F78 100.626126 Hz
 F79 100.626126 Hz
 F80 100.626126 Hz
 F81 100.626126 Hz
 F82 100.626126 Hz
 F83 100.626126 Hz
 F84 100.626126 Hz
 F85 100.626126 Hz
 F86 100.626126 Hz
 F87 100.626126 Hz
 F88 100.626126 Hz
 F89 100.626126 Hz
 F90 100.626126 Hz
 F91 100.626126 Hz
 F92 100.626126 Hz
 F93 100.626126 Hz
 F94 100.626126 Hz
 F95 100.626126 Hz
 F96 100.626126 Hz
 F97 100.626126 Hz
 F98 100.626126 Hz
 F99 100.626126 Hz
 F100 100.626126 Hz

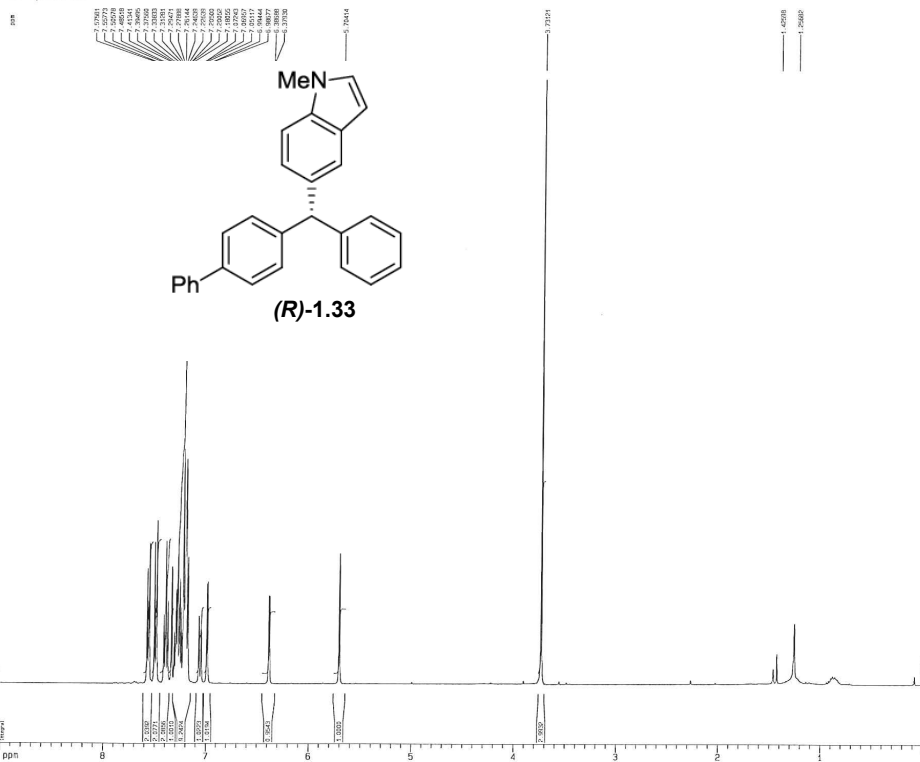
Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
 QEXP 1000000
 NAME LHM-1-148-13-125-2
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 DATE_ 20100909
 TIME 14:35
 INSTRUM spect
 PULPROG zgpg30
 NUC1 13C
 NUC2 1H
 SOLVENT CDCl3
 NS 2
 DS 2
 SWH 8019.850 Hz
 FIDRES 0.00049 Hz
 AQ 0.0000000 sec
 RG 327
 EQ 9.7
 DE 8.00 uMm
 TE 300.2 K
 D1 0.0000000 sec
 MCHST 0.0000000 sec
 MCKK 0.0000000 sec
 F1 4000.000 Hz
 F2 100.626126 Hz
 F3 100.626126 Hz
 F4 100.626126 Hz
 F5 100.626126 Hz
 F6 100.626126 Hz
 F7 100.626126 Hz
 F8 100.626126 Hz
 F9 100.626126 Hz
 F10 100.626126 Hz
 F11 100.626126 Hz
 F12 100.626126 Hz
 F13 100.626126 Hz
 F14 100.626126 Hz
 F15 100.626126 Hz
 F16 100.626126 Hz
 F17 100.626126 Hz
 F18 100.626126 Hz
 F19 100.626126 Hz
 F20 100.626126 Hz
 F21 100.626126 Hz
 F22 100.626126 Hz
 F23 100.626126 Hz
 F24 100.626126 Hz
 F25 100.626126 Hz
 F26 100.626126 Hz
 F27 100.626126 Hz
 F28 100.626126 Hz
 F29 100.626126 Hz
 F30 100.626126 Hz
 F31 100.626126 Hz
 F32 100.626126 Hz
 F33 100.626126 Hz
 F34 100.626126 Hz
 F35 100.626126 Hz
 F36 100.626126 Hz
 F37 100.626126 Hz
 F38 100.626126 Hz
 F39 100.626126 Hz
 F40 100.626126 Hz
 F41 100.626126 Hz
 F42 100.626126 Hz
 F43 100.626126 Hz
 F44 100.626126 Hz
 F45 100.626126 Hz
 F46 100.626126 Hz
 F47 100.626126 Hz
 F48 100.626126 Hz
 F49 100.626126 Hz
 F50 100.626126 Hz
 F51 100.626126 Hz
 F52 100.626126 Hz
 F53 100.626126 Hz
 F54 100.626126 Hz
 F55 100.626126 Hz
 F56 100.626126 Hz
 F57 100.626126 Hz
 F58 100.626126 Hz
 F59 100.626126 Hz
 F60 100.626126 Hz
 F61 100.626126 Hz
 F62 100.626126 Hz
 F63 100.626126 Hz
 F64 100.626126 Hz
 F65 100.626126 Hz
 F66 100.626126 Hz
 F67 100.626126 Hz
 F68 100.626126 Hz
 F69 100.626126 Hz
 F70 100.626126 Hz
 F71 100.626126 Hz
 F72 100.626126 Hz
 F73 100.626126 Hz
 F74 100.626126 Hz
 F75 100.626126 Hz
 F76 100.626126 Hz
 F77 100.626126 Hz
 F78 100.626126 Hz
 F79 100.626126 Hz
 F80 100.626126 Hz
 F81 100.626126 Hz
 F82 100.626126 Hz
 F83 100.626126 Hz
 F84 100.626126 Hz
 F85 100.626126 Hz
 F86 100.626126 Hz
 F87 100.626126 Hz
 F88 100.626126 Hz
 F89 100.626126 Hz
 F90 100.626126 Hz
 F91 100.626126 Hz
 F92 100.626126 Hz
 F93 100.626126 Hz
 F94 100.626126 Hz
 F95 100.626126 Hz
 F96 100.626126 Hz
 F97 100.626126 Hz
 F98 100.626126 Hz
 F99 100.626126 Hz
 F100 100.626126 Hz



1H spectrum



```

Current Data Parameters
NAME: LFN-133-01
EXPNO: 1
PROCNO: 1

F2 - Acquisition Parameters
Date_ : 20120305
Time : 11:55
INSTRUM : spect
PROBHD : 5 mm QNP 1H/1
PULPROG : zgpg30
TD : 65536
SOLVENT : CDCl3
NS : 4
DS : 2
SWH : 8410.258 Hz
FIDRES : 0.330113 Hz
AQ : 5.1181119 sec
RG : 321.6
CW : 736.000 MHz
DE : 4.50 MHz
TE : 298.1 K
D1 : 0.30000000 sec
d11 : 0.00000000 sec
MORPH : 0.00000000 sec
MORPH : 0.00000000 sec

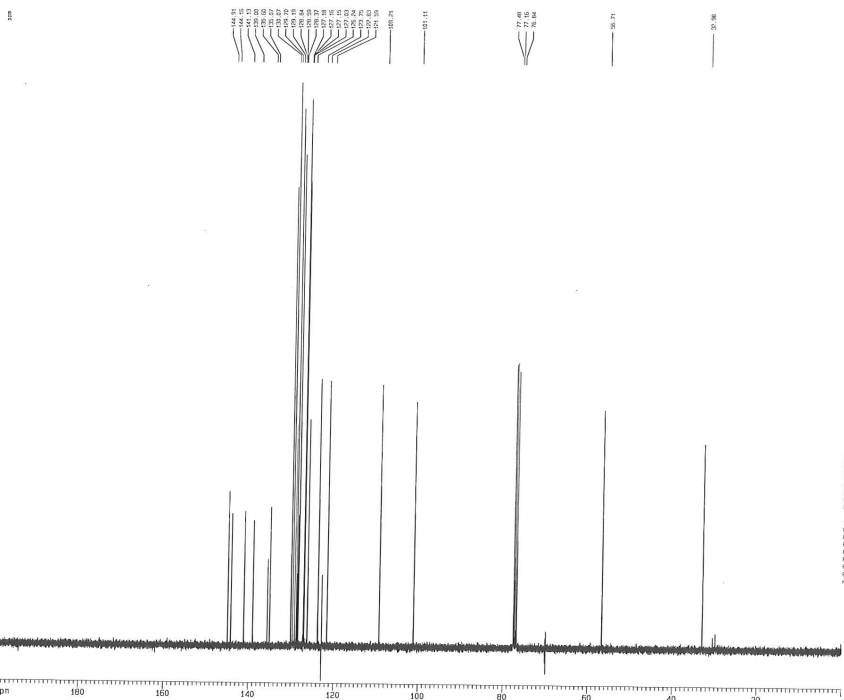
----- CHANNEL f1 -----
NUC1 : 1H
P1 : 12.00 MHz
PL1 : -2.50 dB
SFO1 : 400.1463000 MHz

F2 - Processing parameters
SI : 80000
SF : 400.1463000 MHz
WDW : HANNING
SSB : 0
LB : 0.00 Hz
GB : 0
PC : 2.00

ID NMR file parameters
EX : 20.00 cm
CY : 15.00 cm
FID : 8.00 cm
F1 : 3001.47 Hz
F2 : 0.00 cm
F3 : 0.00 Hz
P1 : 0.00 cm
P2 : 0.00 cm
P3 : 0.00 cm
P4 : 0.00 cm
P5 : 0.00 cm
P6 : 0.00 cm
P7 : 0.00 cm
P8 : 0.00 cm
P9 : 0.00 cm
P10 : 0.00 cm
P11 : 0.00 cm
P12 : 0.00 cm
P13 : 0.00 cm
P14 : 0.00 cm
P15 : 0.00 cm
P16 : 0.00 cm
P17 : 0.00 cm
P18 : 0.00 cm
P19 : 0.00 cm
P20 : 0.00 cm
P21 : 0.00 cm
P22 : 0.00 cm
P23 : 0.00 cm
P24 : 0.00 cm
P25 : 0.00 cm
P26 : 0.00 cm
P27 : 0.00 cm
P28 : 0.00 cm
P29 : 0.00 cm
P30 : 0.00 cm
P31 : 0.00 cm
P32 : 0.00 cm
P33 : 0.00 cm
P34 : 0.00 cm
P35 : 0.00 cm
P36 : 0.00 cm
P37 : 0.00 cm
P38 : 0.00 cm
P39 : 0.00 cm
P40 : 0.00 cm
P41 : 0.00 cm
P42 : 0.00 cm
P43 : 0.00 cm
P44 : 0.00 cm
P45 : 0.00 cm
P46 : 0.00 cm
P47 : 0.00 cm
P48 : 0.00 cm
P49 : 0.00 cm
P50 : 0.00 cm
P51 : 0.00 cm
P52 : 0.00 cm
P53 : 0.00 cm
P54 : 0.00 cm
P55 : 0.00 cm
P56 : 0.00 cm
P57 : 0.00 cm
P58 : 0.00 cm
P59 : 0.00 cm
P60 : 0.00 cm
P61 : 0.00 cm
P62 : 0.00 cm
P63 : 0.00 cm
P64 : 0.00 cm
P65 : 0.00 cm
P66 : 0.00 cm
P67 : 0.00 cm
P68 : 0.00 cm
P69 : 0.00 cm
P70 : 0.00 cm
P71 : 0.00 cm
P72 : 0.00 cm
P73 : 0.00 cm
P74 : 0.00 cm
P75 : 0.00 cm
P76 : 0.00 cm
P77 : 0.00 cm
P78 : 0.00 cm
P79 : 0.00 cm
P80 : 0.00 cm
P81 : 0.00 cm
P82 : 0.00 cm
P83 : 0.00 cm
P84 : 0.00 cm
P85 : 0.00 cm
P86 : 0.00 cm
P87 : 0.00 cm
P88 : 0.00 cm
P89 : 0.00 cm
P90 : 0.00 cm
P91 : 0.00 cm
P92 : 0.00 cm
P93 : 0.00 cm
P94 : 0.00 cm
P95 : 0.00 cm
P96 : 0.00 cm
P97 : 0.00 cm
P98 : 0.00 cm
P99 : 0.00 cm
P100 : 0.00 cm

```

13C spectrum with 1H decoupling



```

Current Data Parameters
NAME: LFN-133-01
EXPNO: 1
PROCNO: 1

F2 - Acquisition Parameters
Date_ : 20120305
Time : 12:00
INSTRUM : spect
PROBHD : 5 mm QNP 1H/1
PULPROG : zgpg30
TD : 65536
SOLVENT : CDCl3
NS : 4
DS : 2
SWH : 8410.258 Hz
FIDRES : 0.330113 Hz
AQ : 5.1181119 sec
RG : 321.6
CW : 736.000 MHz
DE : 4.50 MHz
TE : 298.1 K
D1 : 0.30000000 sec
d11 : 0.00000000 sec
MORPH : 0.00000000 sec
MORPH : 0.00000000 sec

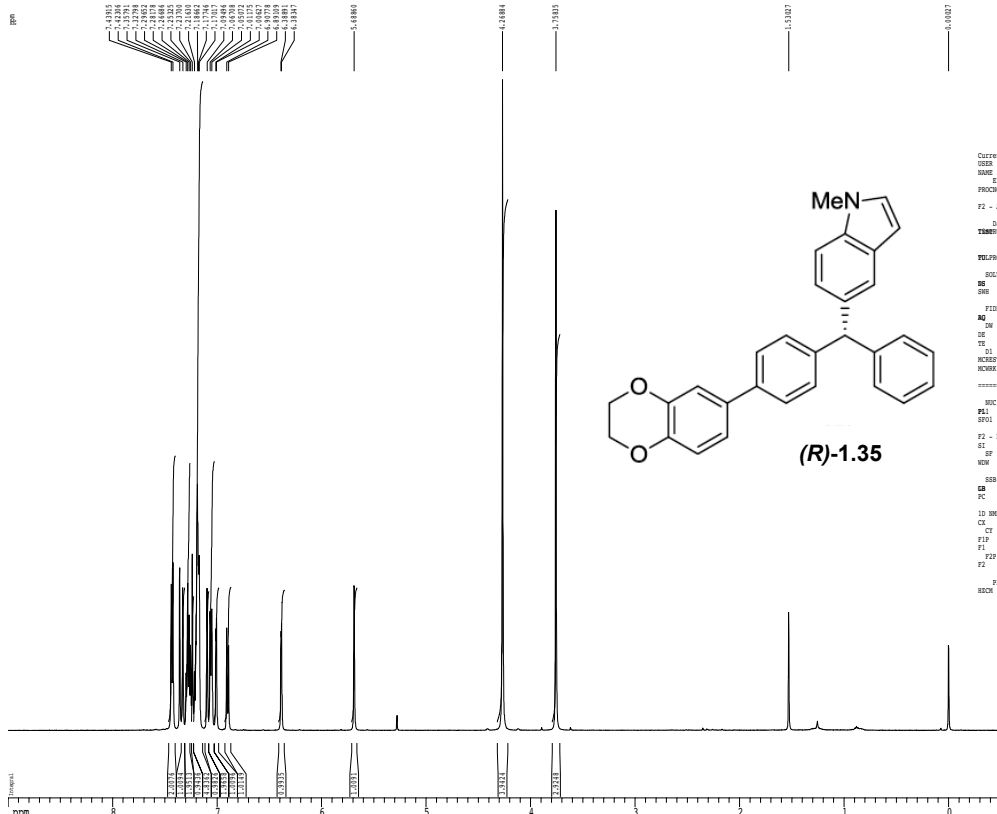
----- CHANNEL f1 -----
NUC1 : 13C
P1 : 12.00 MHz
PL1 : -2.50 dB
SFO1 : 100.6261200 MHz

F2 - Processing parameters
SI : 80000
SF : 100.6261200 MHz
WDW : HANNING
SSB : 0
LB : 0.00 Hz
GB : 0
PC : 2.00

ID NMR file parameters
EX : 20.00 cm
CY : 15.00 cm
FID : 8.00 cm
F1 : 3001.47 Hz
F2 : 0.00 cm
F3 : 0.00 Hz
P1 : 0.00 cm
P2 : 0.00 cm
P3 : 0.00 cm
P4 : 0.00 cm
P5 : 0.00 cm
P6 : 0.00 cm
P7 : 0.00 cm
P8 : 0.00 cm
P9 : 0.00 cm
P10 : 0.00 cm
P11 : 0.00 cm
P12 : 0.00 cm
P13 : 0.00 cm
P14 : 0.00 cm
P15 : 0.00 cm
P16 : 0.00 cm
P17 : 0.00 cm
P18 : 0.00 cm
P19 : 0.00 cm
P20 : 0.00 cm
P21 : 0.00 cm
P22 : 0.00 cm
P23 : 0.00 cm
P24 : 0.00 cm
P25 : 0.00 cm
P26 : 0.00 cm
P27 : 0.00 cm
P28 : 0.00 cm
P29 : 0.00 cm
P30 : 0.00 cm
P31 : 0.00 cm
P32 : 0.00 cm
P33 : 0.00 cm
P34 : 0.00 cm
P35 : 0.00 cm
P36 : 0.00 cm
P37 : 0.00 cm
P38 : 0.00 cm
P39 : 0.00 cm
P40 : 0.00 cm
P41 : 0.00 cm
P42 : 0.00 cm
P43 : 0.00 cm
P44 : 0.00 cm
P45 : 0.00 cm
P46 : 0.00 cm
P47 : 0.00 cm
P48 : 0.00 cm
P49 : 0.00 cm
P50 : 0.00 cm
P51 : 0.00 cm
P52 : 0.00 cm
P53 : 0.00 cm
P54 : 0.00 cm
P55 : 0.00 cm
P56 : 0.00 cm
P57 : 0.00 cm
P58 : 0.00 cm
P59 : 0.00 cm
P60 : 0.00 cm
P61 : 0.00 cm
P62 : 0.00 cm
P63 : 0.00 cm
P64 : 0.00 cm
P65 : 0.00 cm
P66 : 0.00 cm
P67 : 0.00 cm
P68 : 0.00 cm
P69 : 0.00 cm
P70 : 0.00 cm
P71 : 0.00 cm
P72 : 0.00 cm
P73 : 0.00 cm
P74 : 0.00 cm
P75 : 0.00 cm
P76 : 0.00 cm
P77 : 0.00 cm
P78 : 0.00 cm
P79 : 0.00 cm
P80 : 0.00 cm
P81 : 0.00 cm
P82 : 0.00 cm
P83 : 0.00 cm
P84 : 0.00 cm
P85 : 0.00 cm
P86 : 0.00 cm
P87 : 0.00 cm
P88 : 0.00 cm
P89 : 0.00 cm
P90 : 0.00 cm
P91 : 0.00 cm
P92 : 0.00 cm
P93 : 0.00 cm
P94 : 0.00 cm
P95 : 0.00 cm
P96 : 0.00 cm
P97 : 0.00 cm
P98 : 0.00 cm
P99 : 0.00 cm
P100 : 0.00 cm

```


1H spectrum



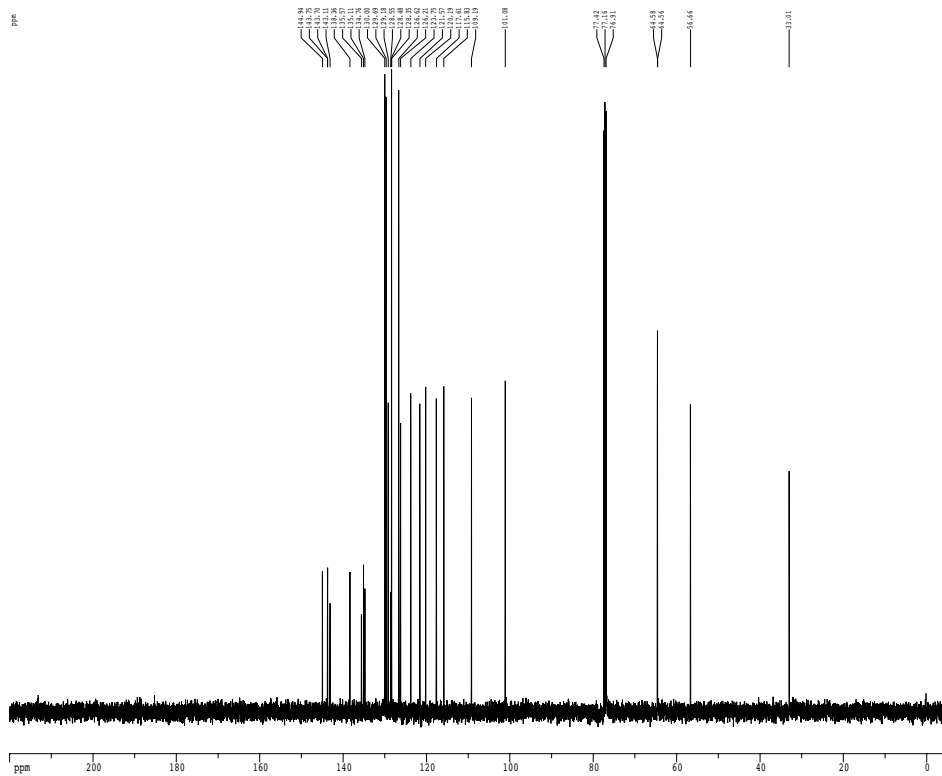
Current Data Parameters
USER mhart
NAME MM-27-142-12008
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20121119
Time 12.04
INSTRUM cryo500
PROBHD 5 mm CPXI 1H-
PULPROG zgpg30
SOLVENT 4
NS 2
DS 8012.820 Hz
SWH 0.898043 Hz
FIDRES 5.999874 sec
AQ 0
RG 62.400 usec
IN 62.400 usec
TE 298.2 K
SI 0.1000000 sec
MCX27 0.0000000 sec
MCW4 0.0100000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 7.50 usec
SFO1 500.2215040 MHz
F2 - Processing parameters
SI 65156
SF 500.220423 MHz
WDW EM
SSB 0
GB 0.00 Hz
PC 4.00

ID NH plot parameters
CX 21.80 cm
CY 15.40 cm
F1P 9.000 ppm
F2P 4561.98 Hz
FZ -250.11 Hz
PPHCH 0.416647 ppm/cm
SFOH 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
USER mhart
NAME MM-27-142-12008
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20121119
Time 12.04
INSTRUM cryo500
PROBHD 5 mm CPXI 1H-
PULPROG SpinEchoHsg-pod
SI 65156
SOLVENT 4
NS 80
DS 16
SWH 30362.020 Hz
FIDRES 0.482388 Hz
AQ 1.481294 sec
RG 7219.2
IN 156.200 usec
DE 6.00 usec
TE 298.2 K
SI 0.2000000 sec
D14 0.0002000 sec
d11 0.0002000 sec
d12 0.0002000 sec
MCX27 0.0000000 sec
MCW4 0.0100000 sec
F2 31.00 usec

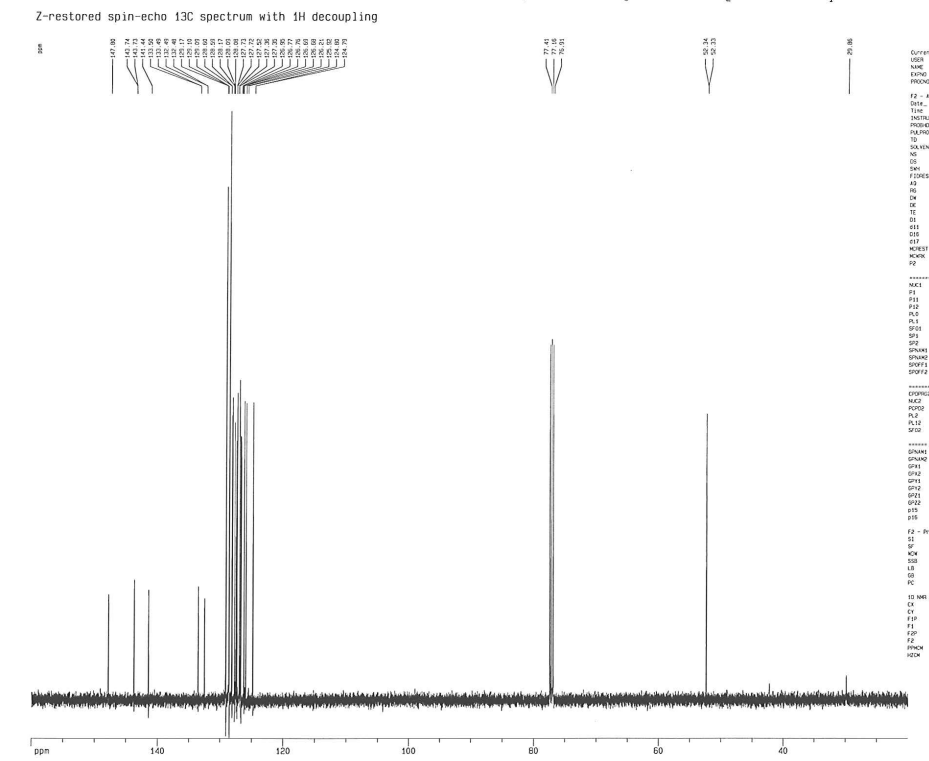
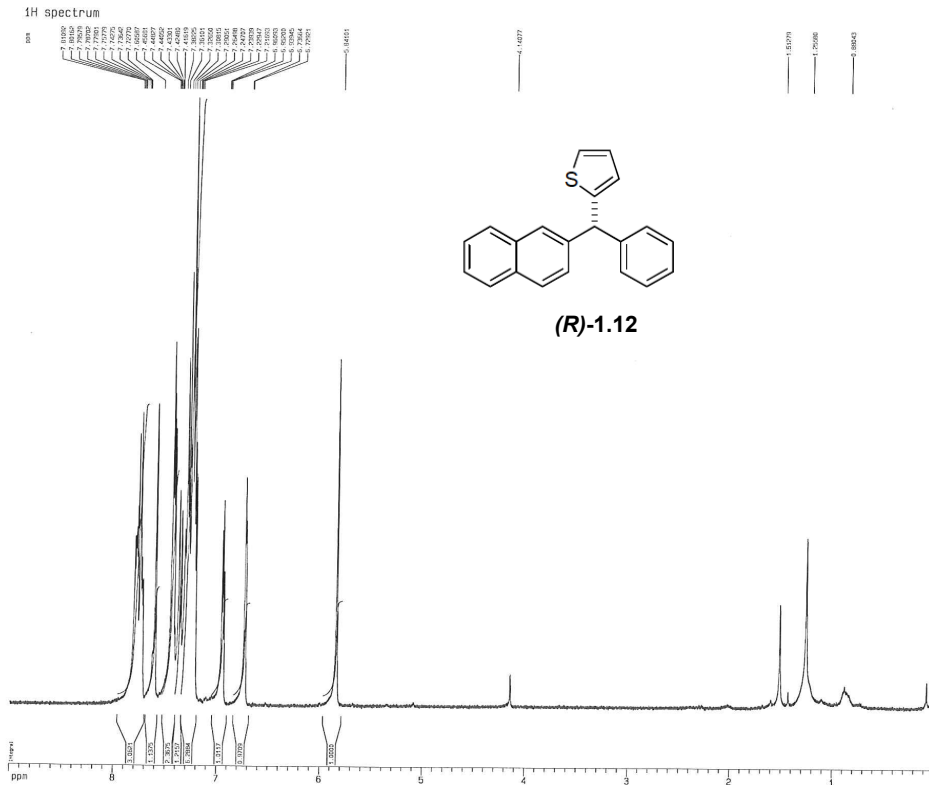
***** CHANNEL f1 *****
NUC1 13C
P1 15.50 usec
SFO1 500.00 usec
P12 2000.00 usec
PL2 120.00 dB
SFO2 125.7642150 MHz
SFO1 5.20 dB
SFO2 5.20 dB
SFOH1 Cryo5, 0.251261
SFOH2 CryoComp, 4
SFOFF1 0.00 Hz
SFOFF2 0.00 Hz

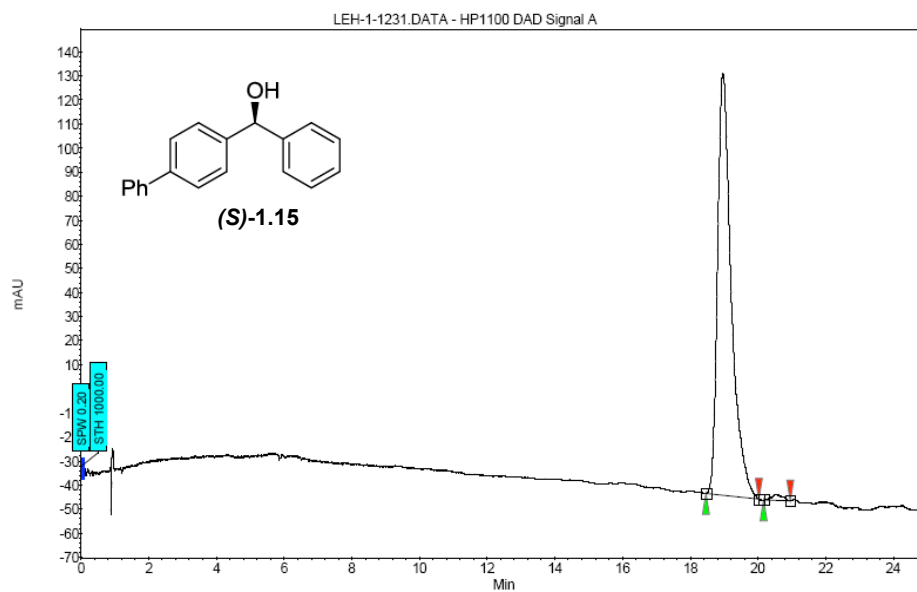
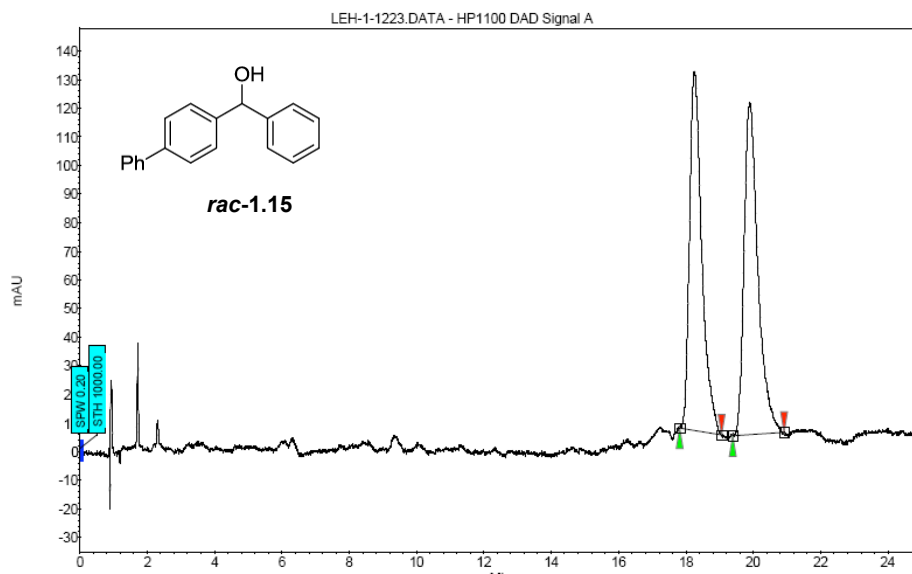
***** CHANNEL f2 *****
CPDPRG2 waltz16
NUC2 13C
PCPD2 180.00 usec
PL2 1.60 dB
PL12 24.40 dB
SFO2 500.2225611 MHz

***** QUALITY CHANNEL *****
CHS0H1 8700.100
CHS0H2 8700.100
CHS1 0.00 %
CHS2 0.00 %
CHS3 0.00 %
CHS4 0.00 %
CHS5 0.00 %
CHS6 0.00 %
CHS7 0.00 %
CHS8 0.00 %
CHS9 0.00 %
CHS10 0.00 %
CHS11 0.00 %
CHS12 0.00 %
CHS13 0.00 %
CHS14 0.00 %
CHS15 0.00 %
CHS16 0.00 %

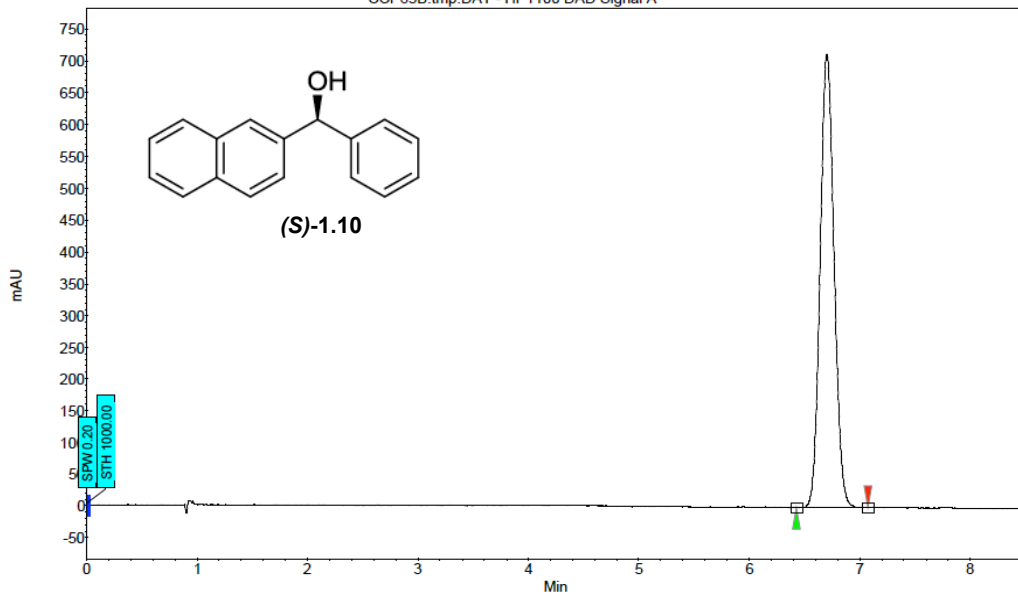
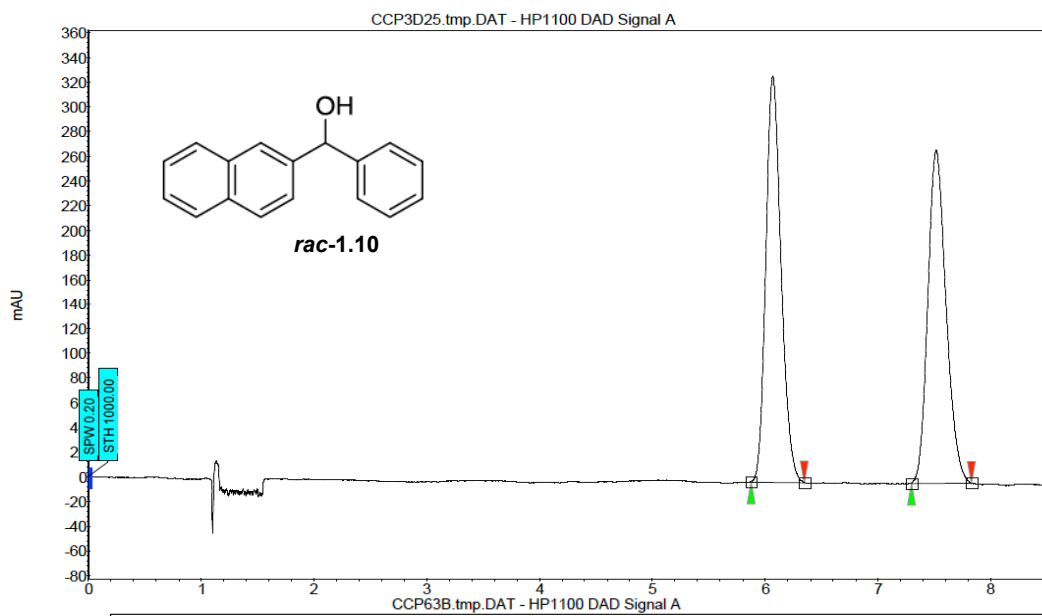
F2 - Processing parameters
SI 65156
SF 125.7642150 MHz
WDW EM
SSB 0
GB 1.00 Hz
PC 2.00

ID NH plot parameters
CX 21.80 cm
CY 15.40 cm
F1P 220.400 ppm
F2P 2747.60 Hz
FZ -250.11 Hz
PPHCH 1.481647 ppm/cm
SFOH 1247.25415 Hz/cm

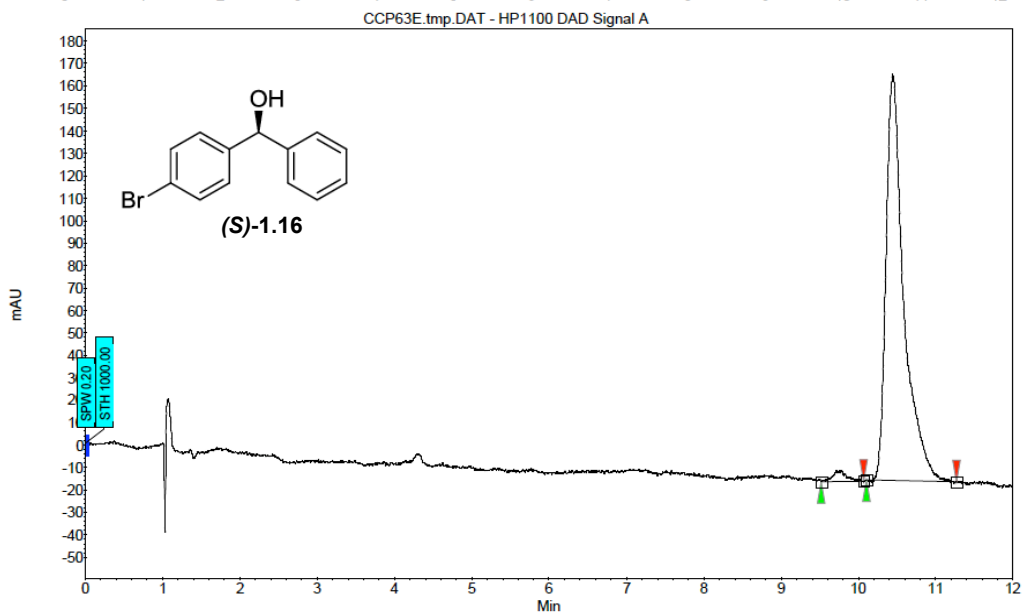
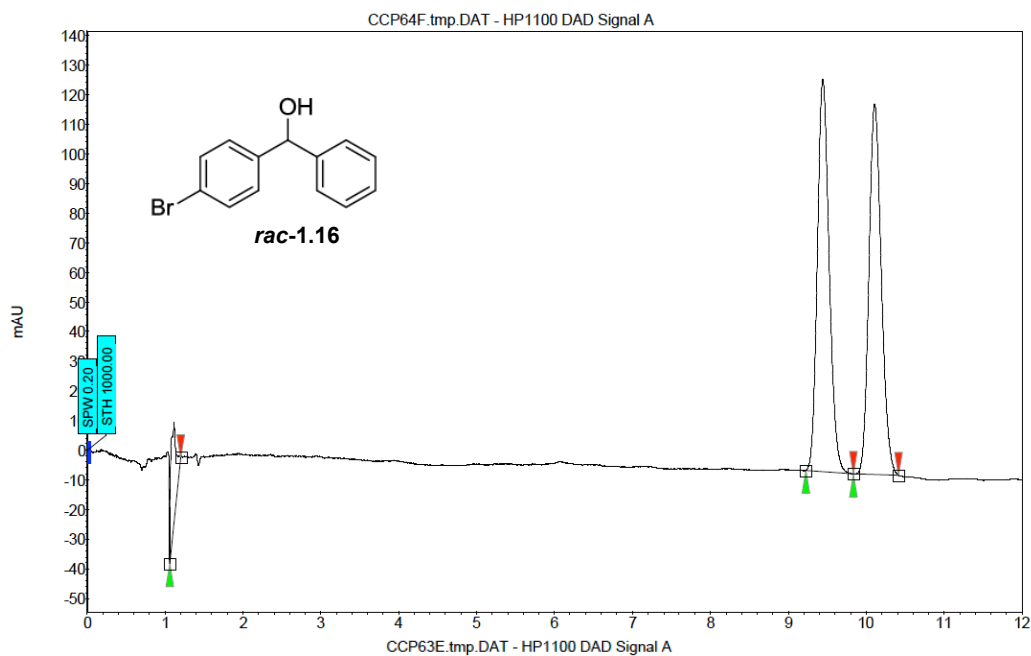




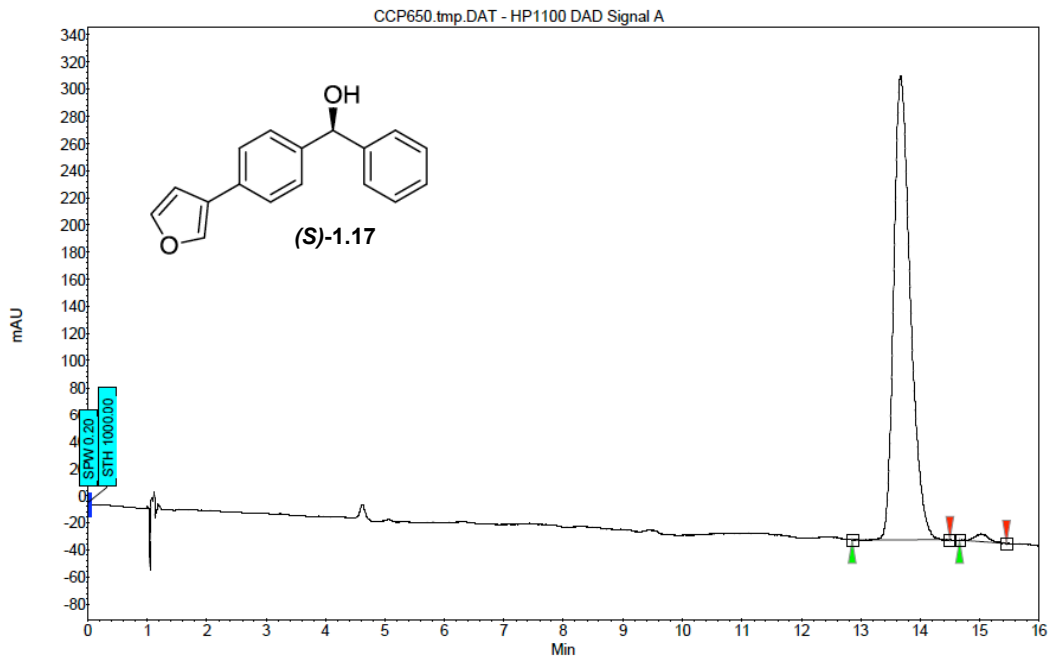
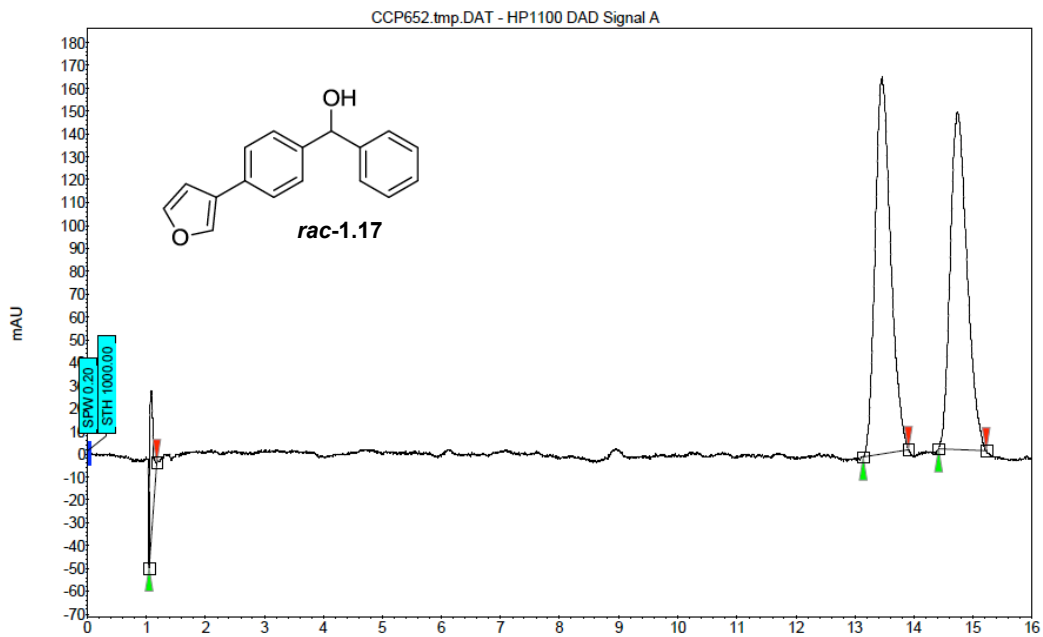
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	18.46	18.97	20.02	0.00	98.91	175.3	82.1	98.914
2	UNKNOWN	20.17	20.55	20.95	0.00	1.09	2.4	0.9	1.086
Total						100.00	177.7	83.0	100.000



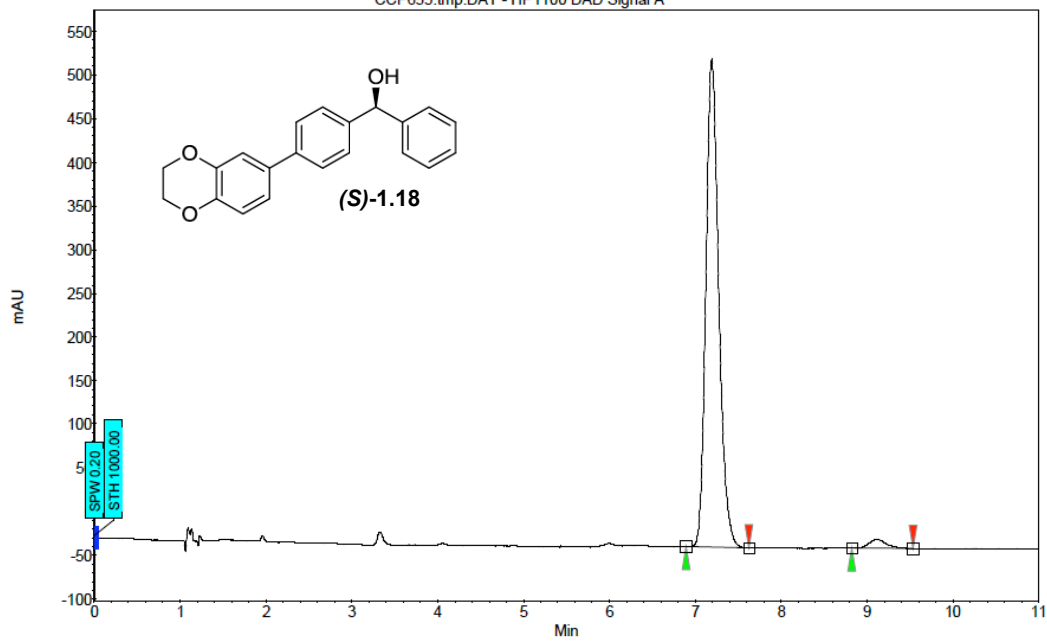
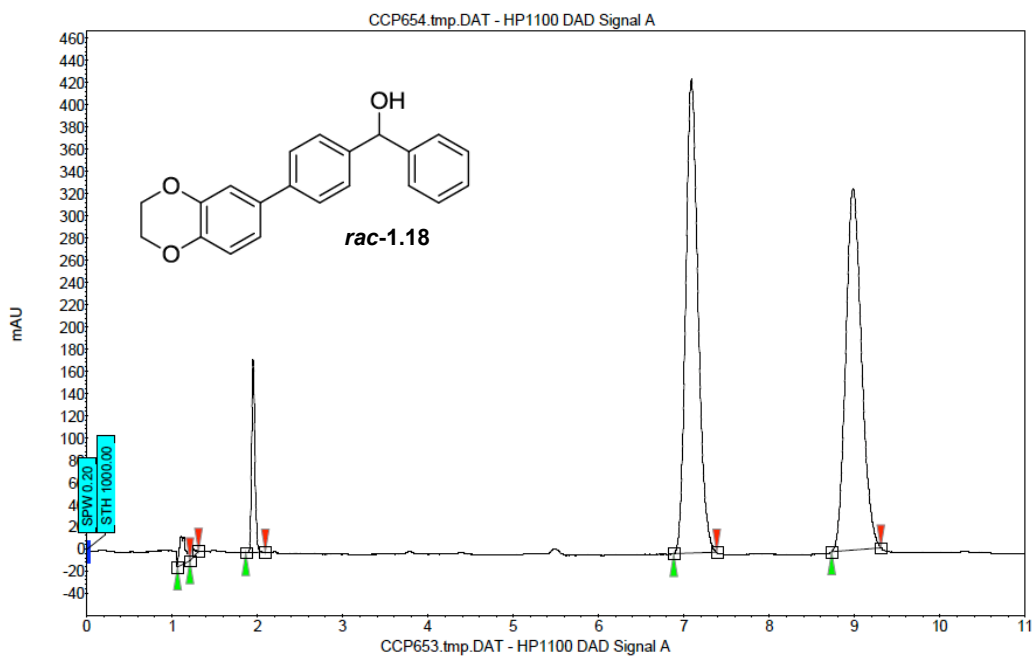
Index	Name	Start Time [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	6.43	6.70	7.08	0.00	100.00	712.9	104.3	100.000
Total						100.00	712.9	104.3	100.000



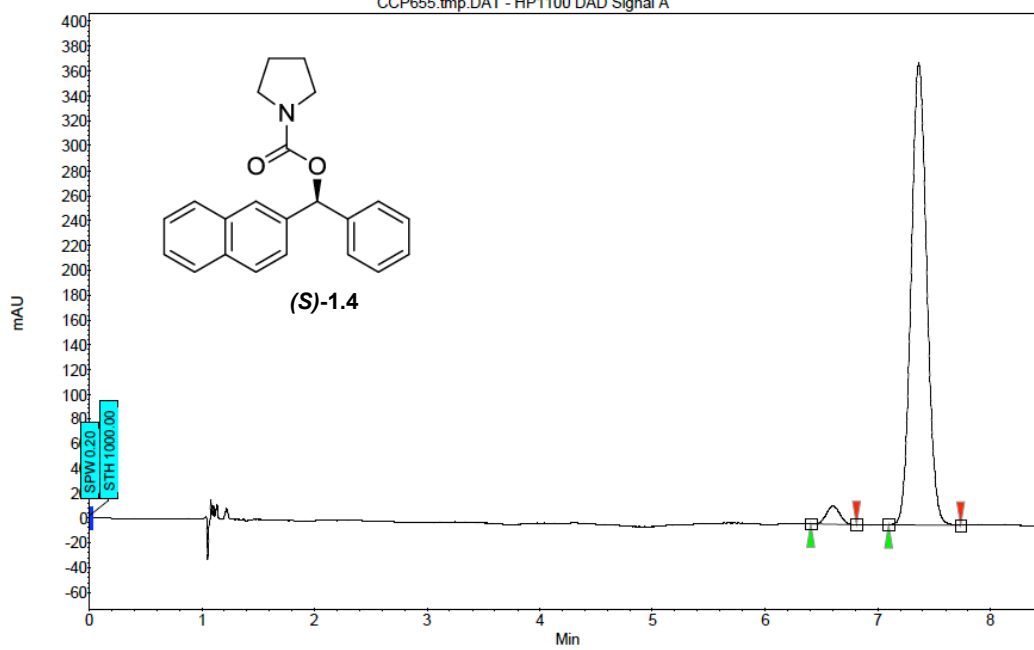
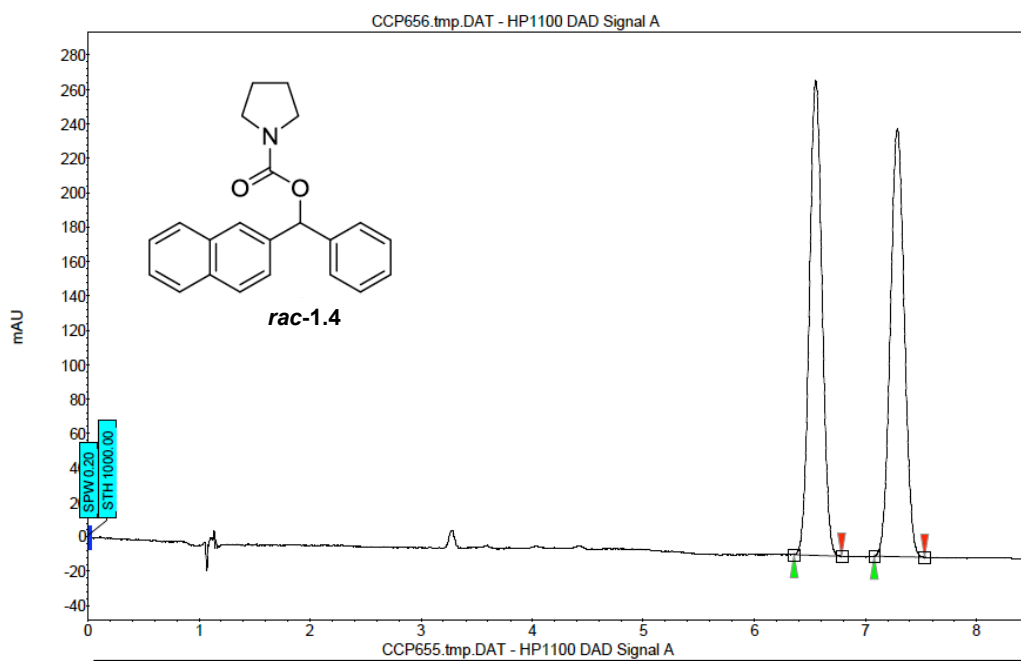
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [µV]	Area [µV.Min]	Area [%]
2	UNKNOWN	9.52	9.76	10.07	0.00	2.16	4.9	1.1	2.164
1	UNKNOWN	10.10	10.44	11.27	0.00	97.84	181.0	47.8	97.836
Total						100.00	185.8	48.9	100.000



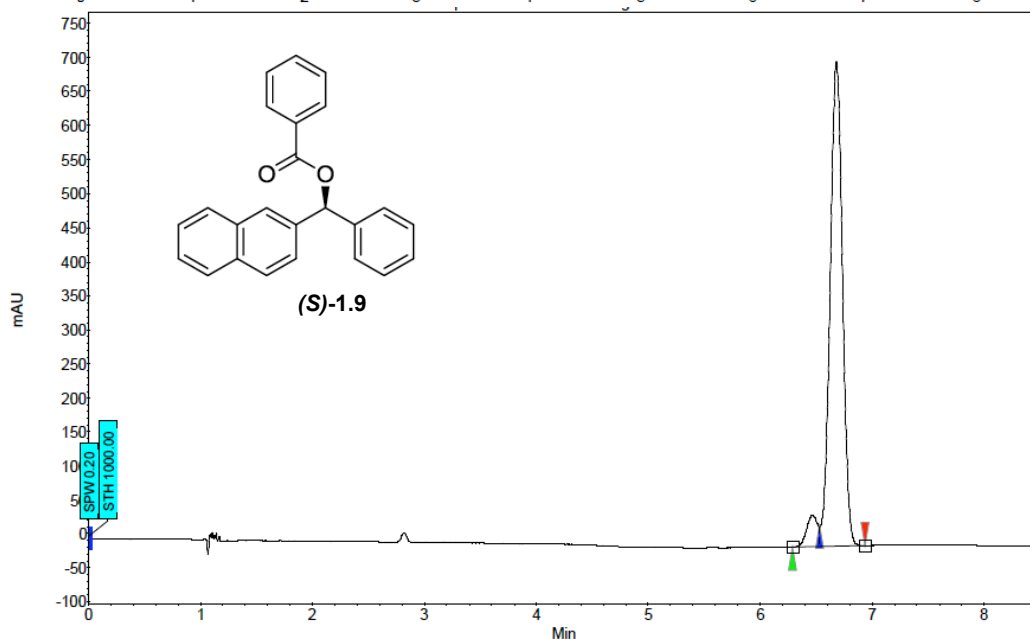
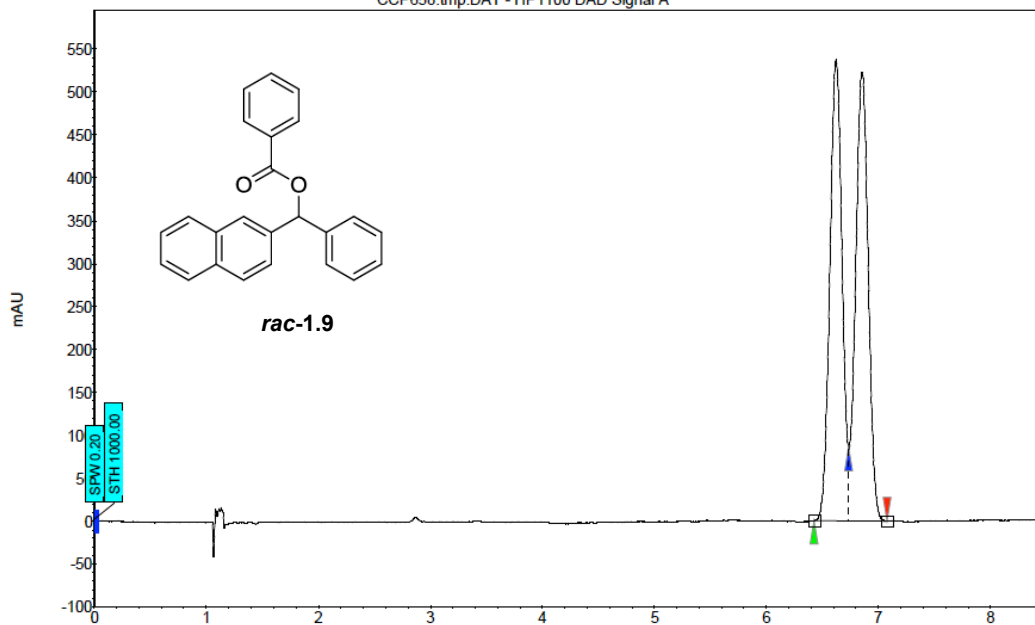
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [µV]	Area [µV.Min]	Area [%]
1	UNKNOWN	12.86	13.67	14.50	0.00	98.57	341.7	111.0	98.571
2	UNKNOWN	14.66	15.02	15.45	0.00	1.43	5.9	1.6	1.429
Total						100.00	347.6	112.6	100.000



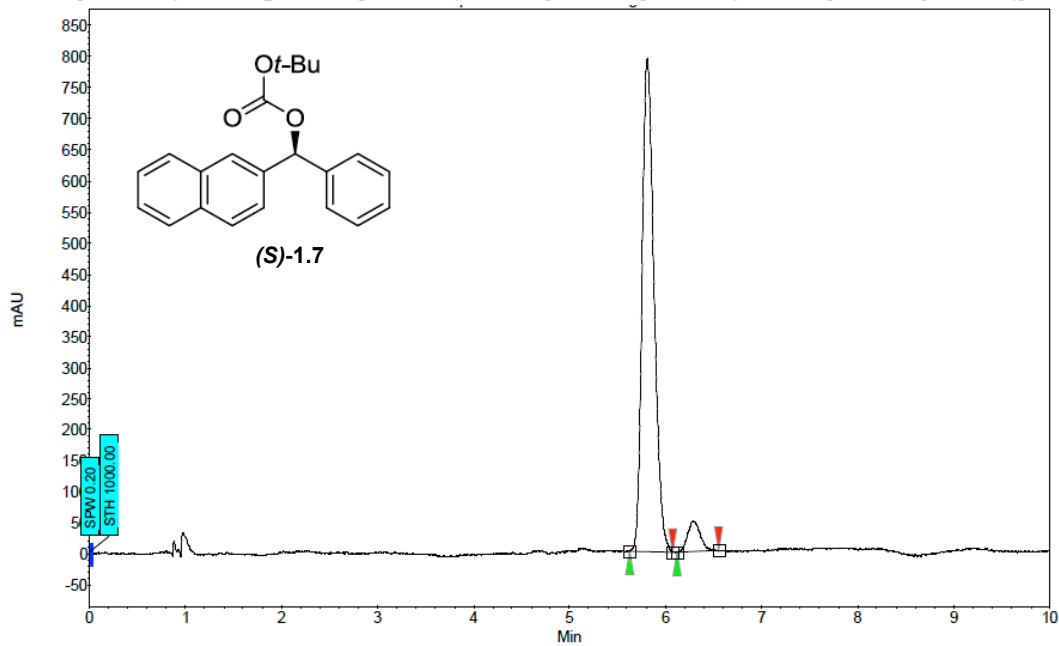
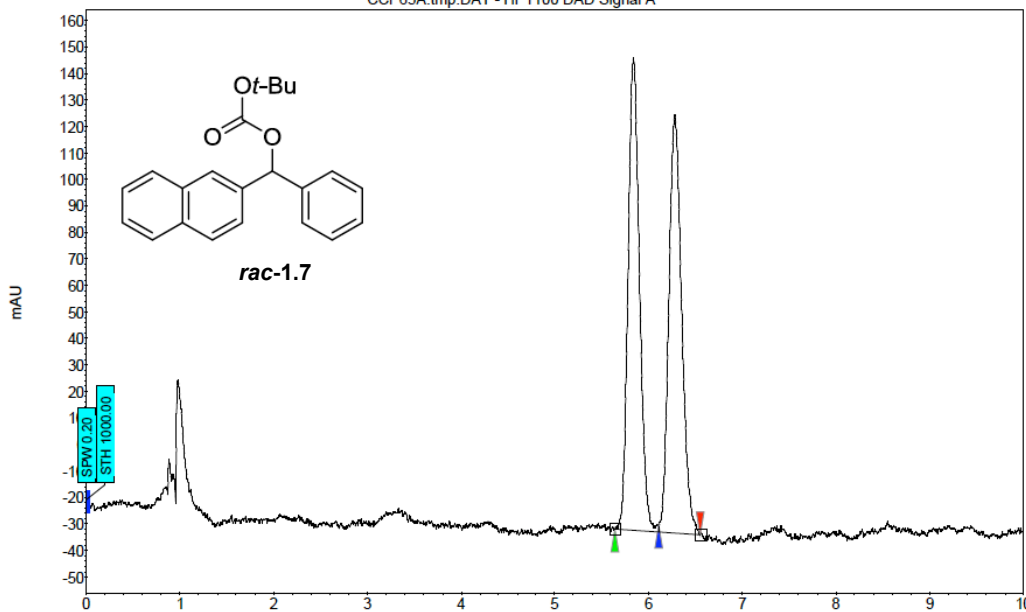
Index	Name	Time			RT Offset	Quantity	Height	Area	
		Start [Min]	End [Min]	Time [Min]				[μ V]	[μ V.Min]
1	UNKNOWN	6.89	7.19	7.62	0.00	97.79	558.2	96.4	97.790
2	UNKNOWN	8.82	9.11	9.53	0.00	2.21	9.8	2.2	2.210
Total						100.00	568.0	98.6	100.000



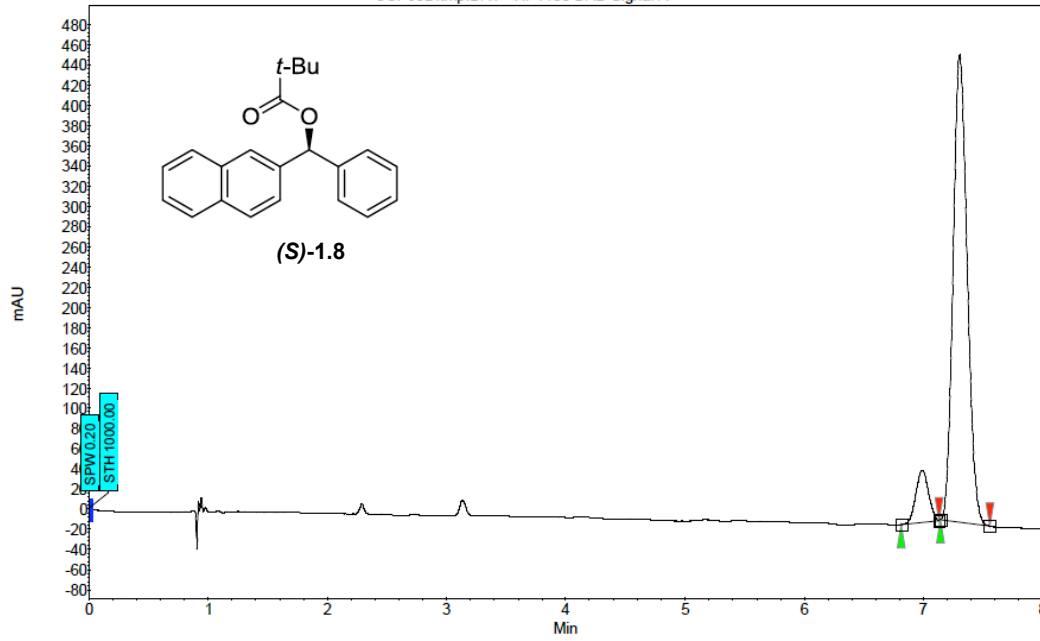
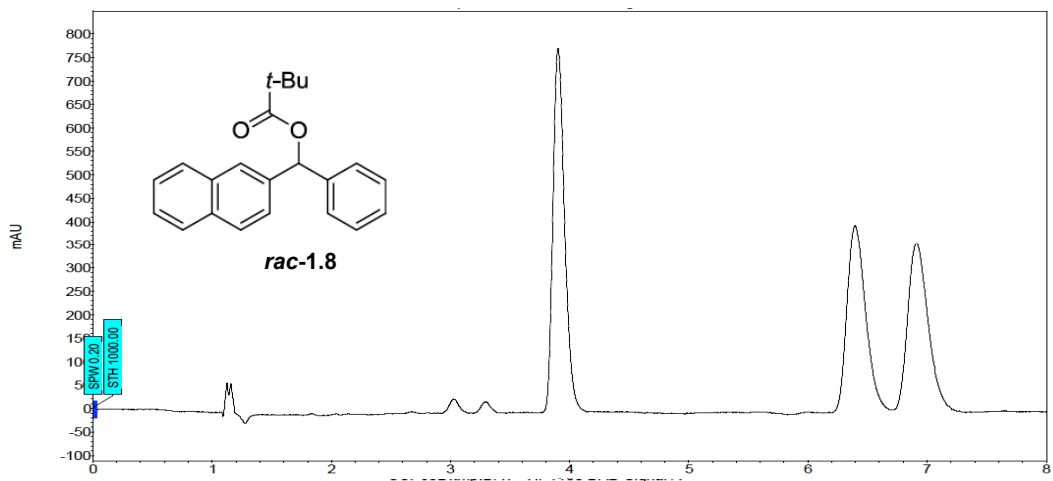
Index	Name	Time			RT Offset	Quantity	Height	Area	
		[Min]	[Min]	[Min]				[% Area]	[μ V]
1	UNKNOWN	6.41	6.60	6.81	0.00	3.40	14.9	2.1	3.400
2	UNKNOWN	7.10	7.37	7.74	0.00	96.60	372.0	59.5	96.600
Total						100.00	386.9	61.6	100.000



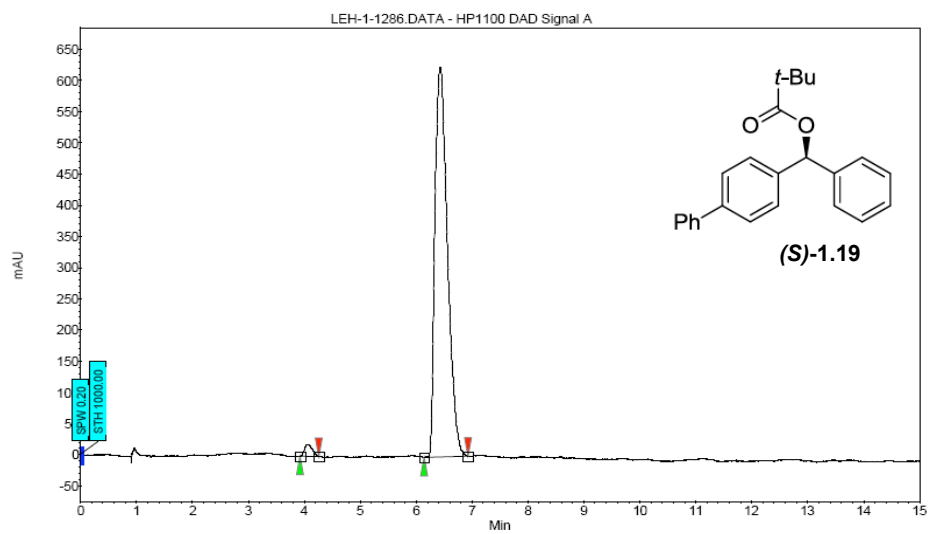
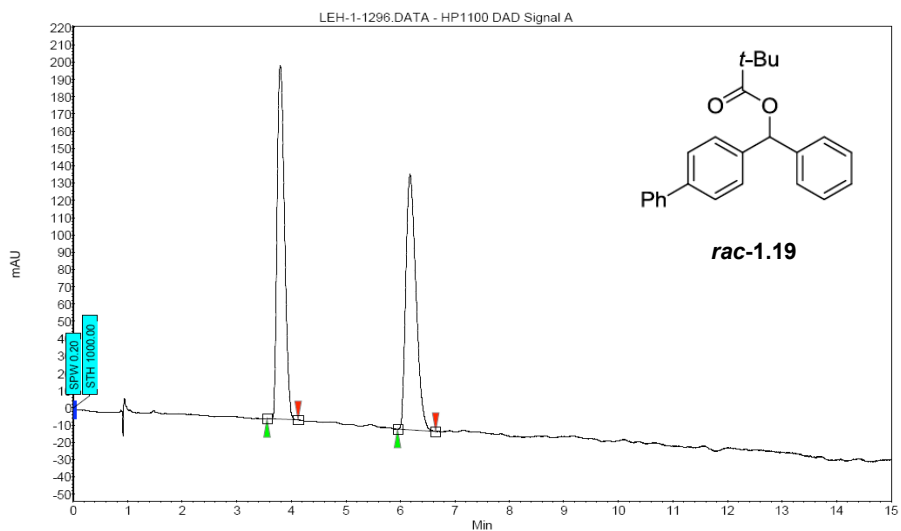
Index	Name	Start Time [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	6.29	6.47	6.53	0.00	5.45	46.2	5.2	5.451
2	UNKNOWN	6.53	6.68	6.94	0.00	94.55	711.7	90.9	94.549
Total						100.00	757.9	96.2	100.000



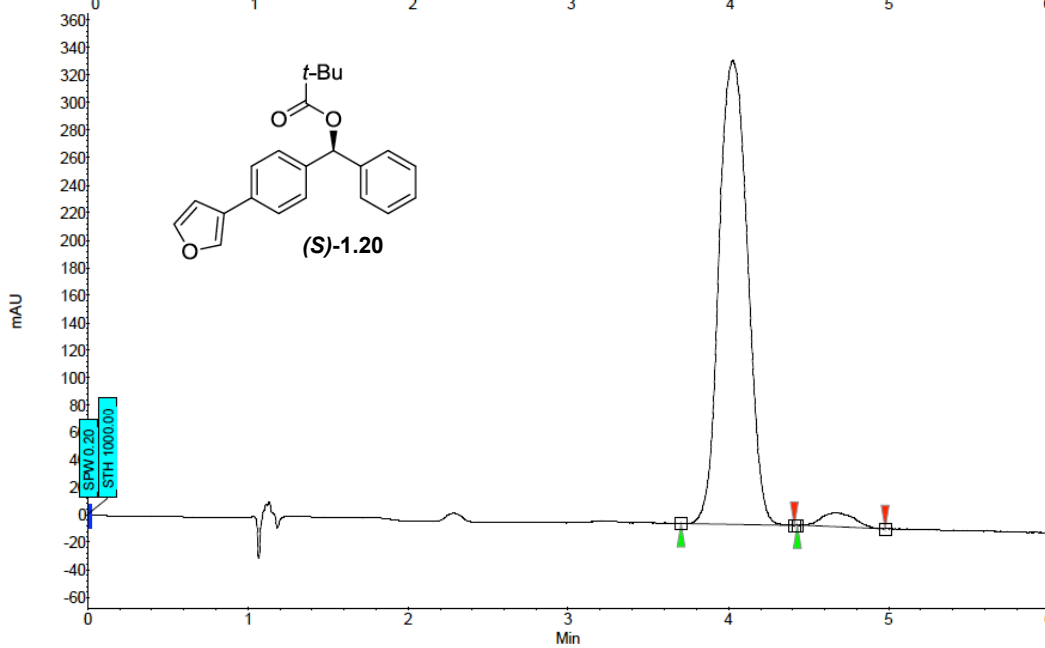
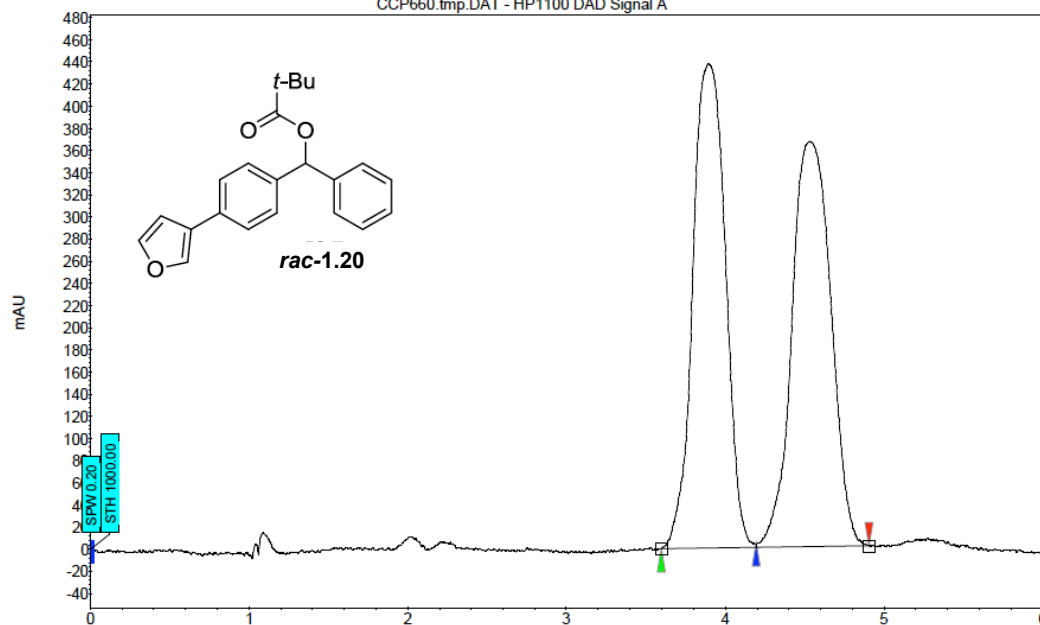
Index	Name	Start Time [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	5.63	5.81	6.08	0.00	93.79	791.8	113.4	93.788
2	UNKNOWN	6.12	6.29	6.55	0.00	6.21	50.0	7.5	6.212
Total						100.00	841.8	120.9	100.000



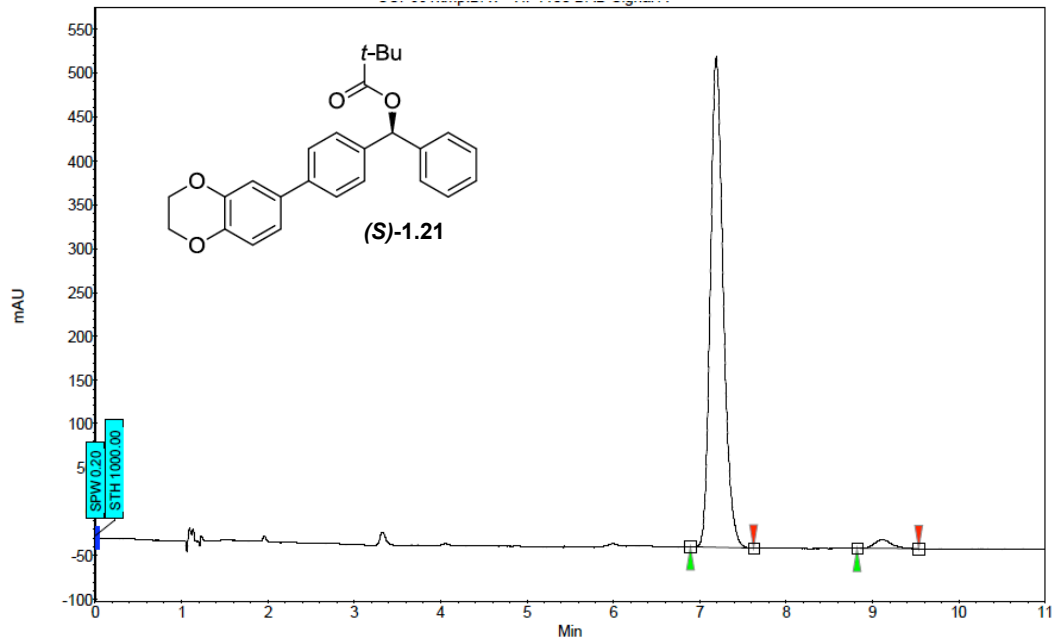
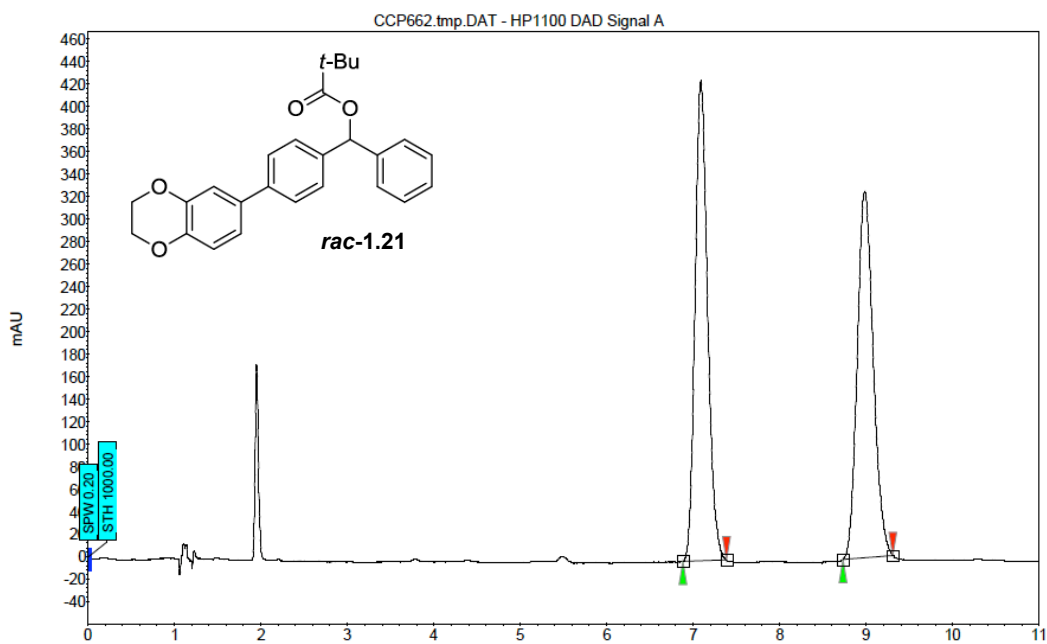
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	6.82	6.99	7.13	0.00	9.06	51.7	6.3	9.058
2	UNKNOWN	7.14	7.30	7.56	0.00	90.94	463.0	63.5	90.942
Total						100.00	514.7	69.9	100.000



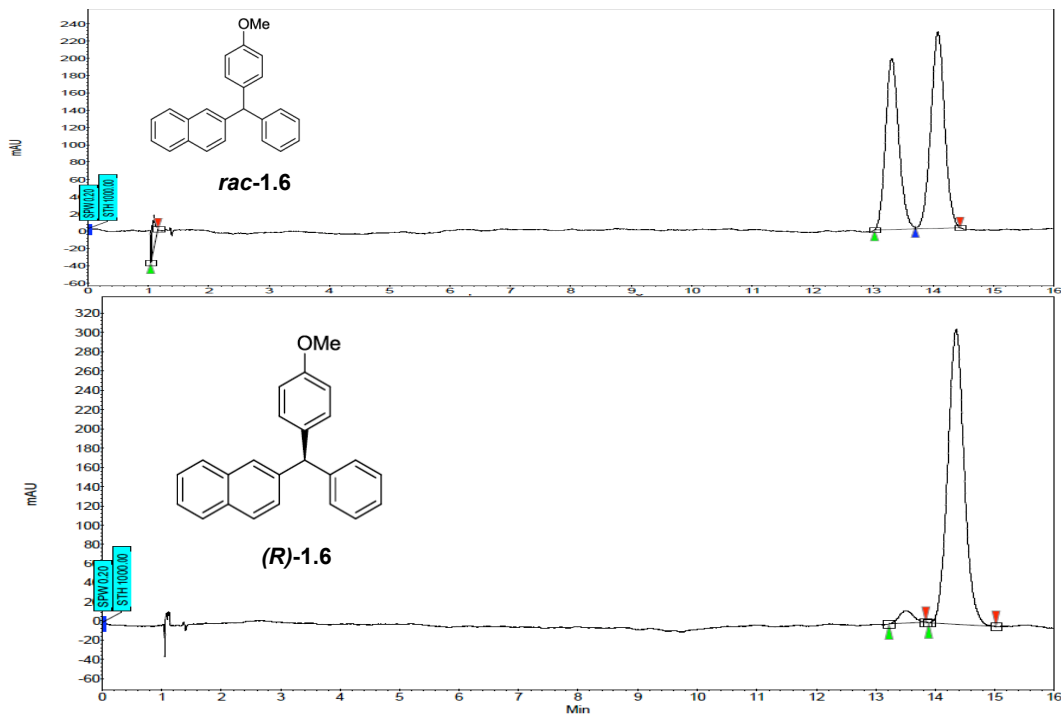
Index	Name	Start Time			End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]						
2	UNKNOWN	3.93	4.06	4.26	0.00	1.88	19.0	3.1	1.877	
1	UNKNOWN	6.14	6.43	6.93	0.00	98.12	625.4	161.3	98.123	
Total						100.00	644.3	164.4	100.000	



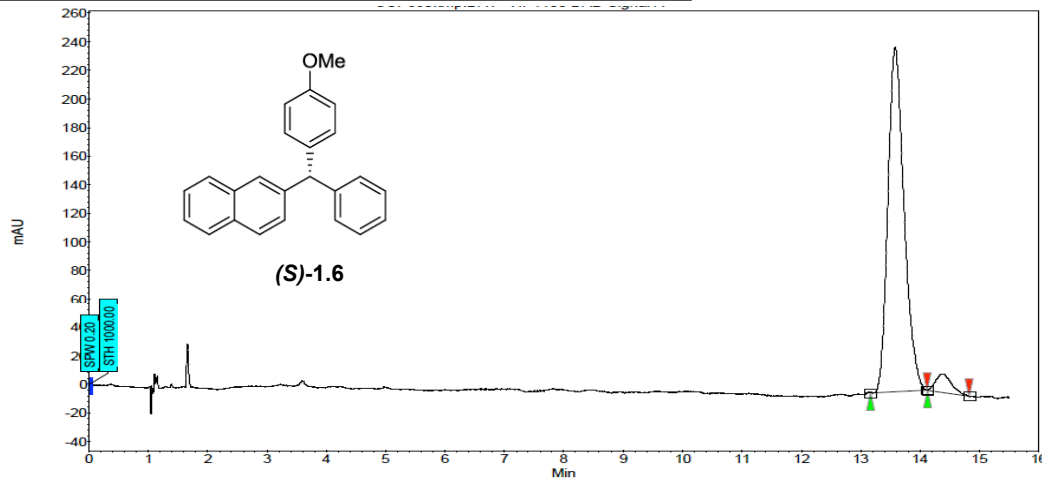
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	3.70	4.03	4.41	0.00	96.61	337.3	69.5	96.611
2	UNKNOWN	4.43	4.66	4.98	0.00	3.39	10.1	2.4	3.389
Total						100.00	347.4	71.9	100.000



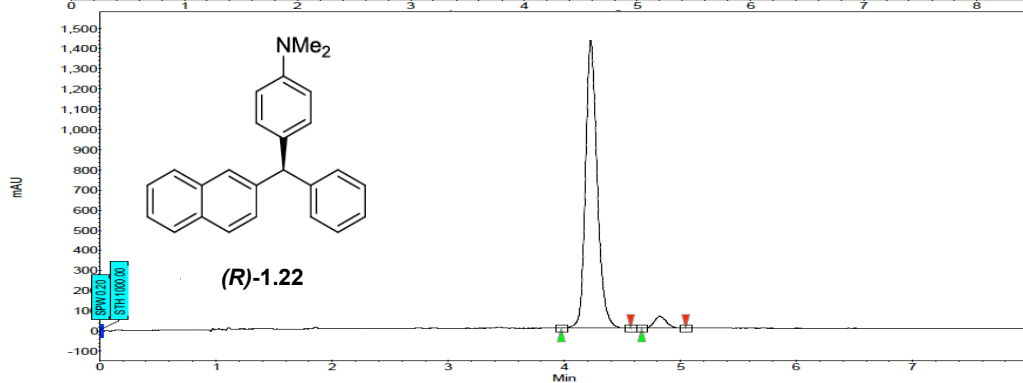
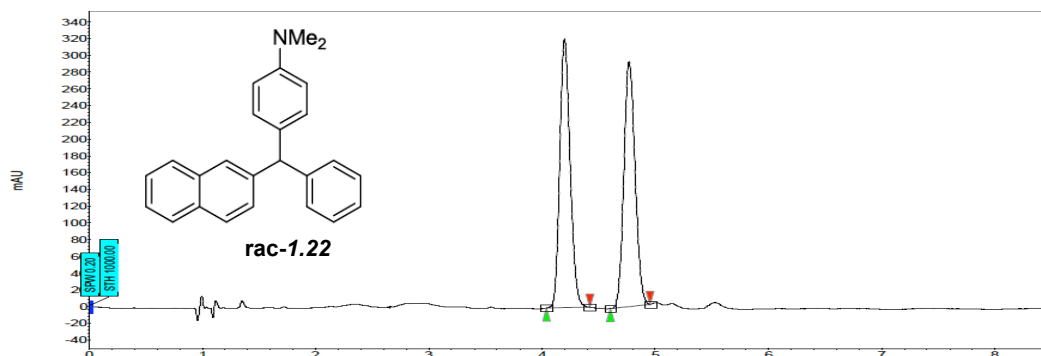
Index	Name	Start Time [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	6.89	7.19	7.62	0.00	97.79	558.2	96.4	97.790
2	UNKNOWN	8.82	9.11	9.53	0.00	2.21	9.8	2.2	2.210
Total						100.00	568.0	98.6	100.000



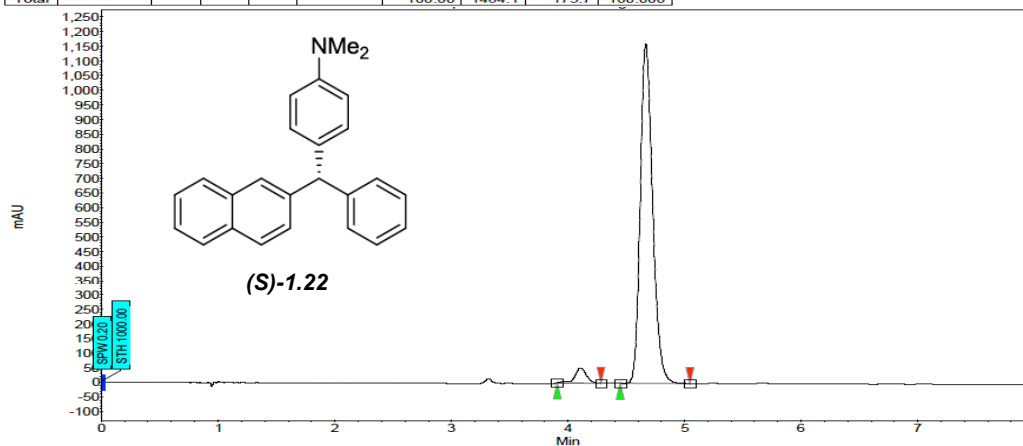
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	13.22	13.52	13.84	0.00	3.51	13.0	3.6	3.505
2	UNKNOWN	13.89	14.35	15.02	0.00	96.49	306.8	98.0	96.495
Total						100.00	319.8	101.6	100.000



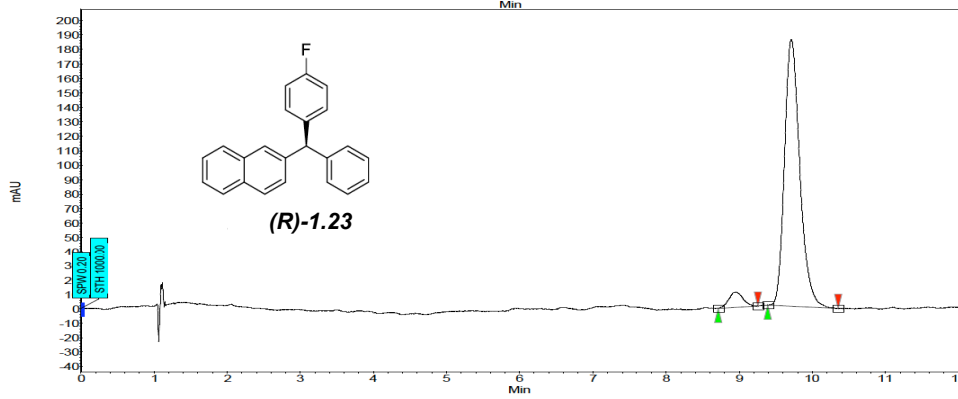
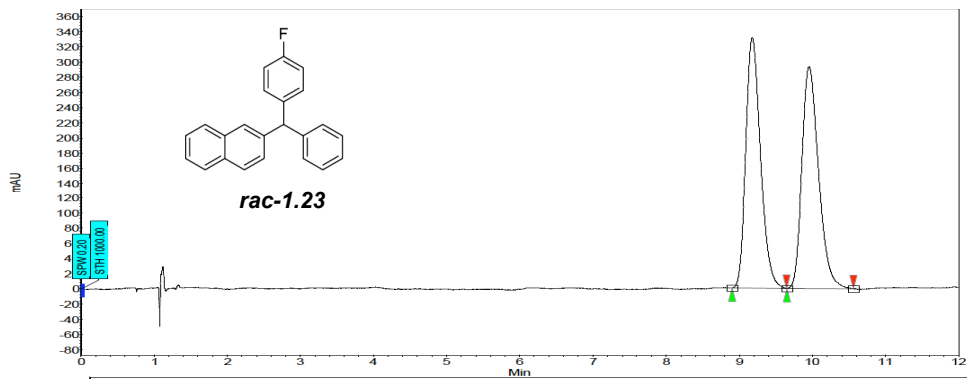
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	13.16	13.58	14.11	0.00	95.00	241.1	76.1	94.998
2	UNKNOWN	14.12	14.40	14.83	0.00	5.00	12.9	4.0	5.002
Total						100.00	254.0	80.1	100.000



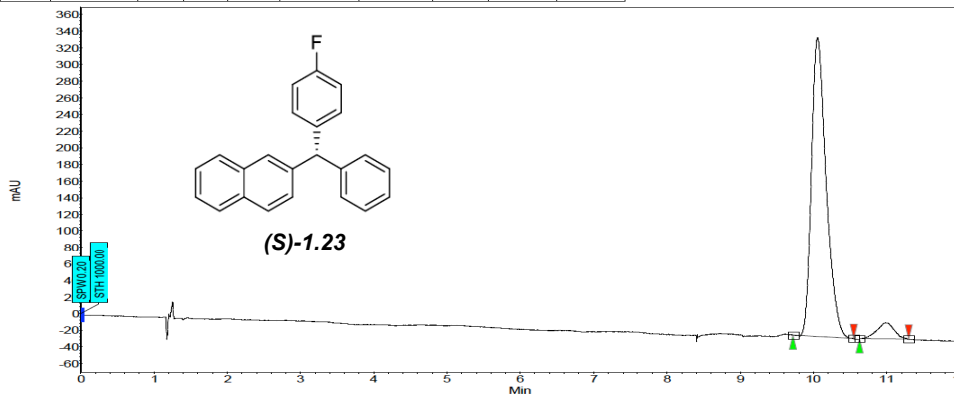
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	3.95	4.23	4.57	0.00	95.82	1425.1	166.4	95.819
2	UNKNOWN	4.67	4.83	5.05	0.00	4.18	59.1	7.3	4.181
Total						100.00	1484.1	173.7	100.000



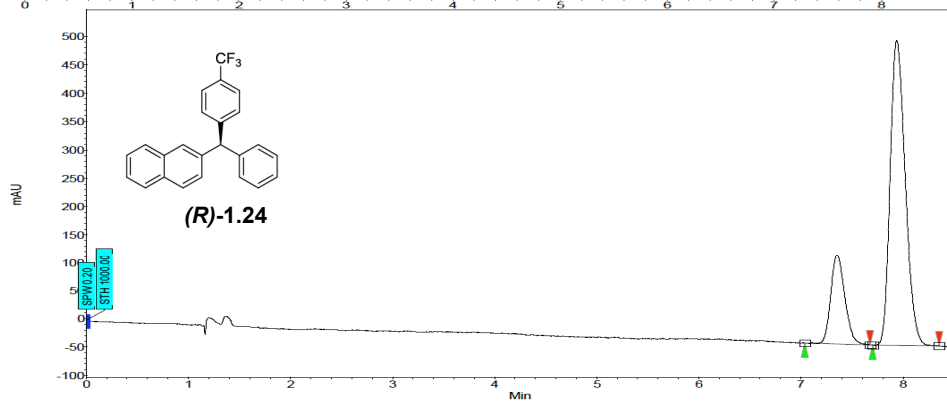
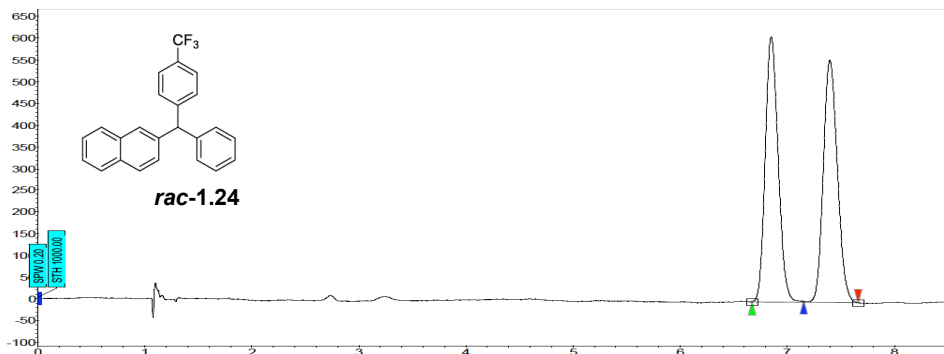
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2	UNKNOWN	3.91	4.11	4.29	0.00	4.01	52.1	6.0	4.006
1	UNKNOWN	4.45	4.67	5.05	0.00	95.99	1160.9	143.0	95.994
Total						100.00	1213.0	148.9	100.000



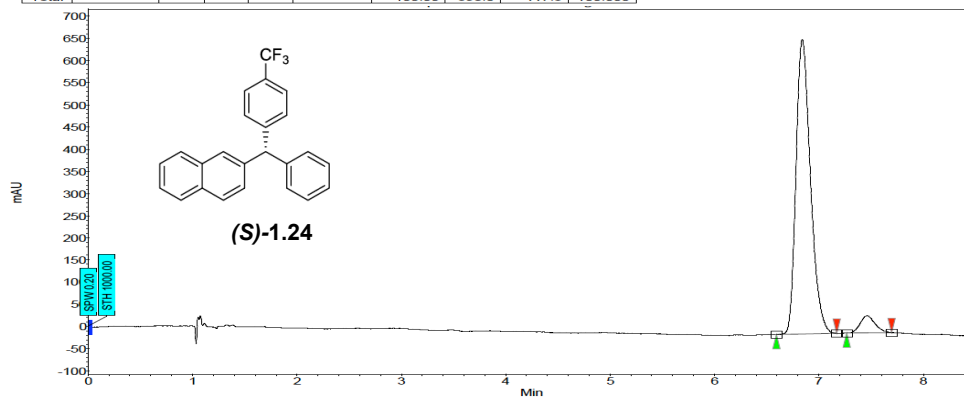
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1	UNKNOWN	8.71	8.96	9.26	0.00	4.97	10.6	2.4	4.971
2	UNKNOWN	9.39	9.71	10.35	0.00	95.03	185.0	45.1	95.029
Total						100.00	195.6	47.5	100.000



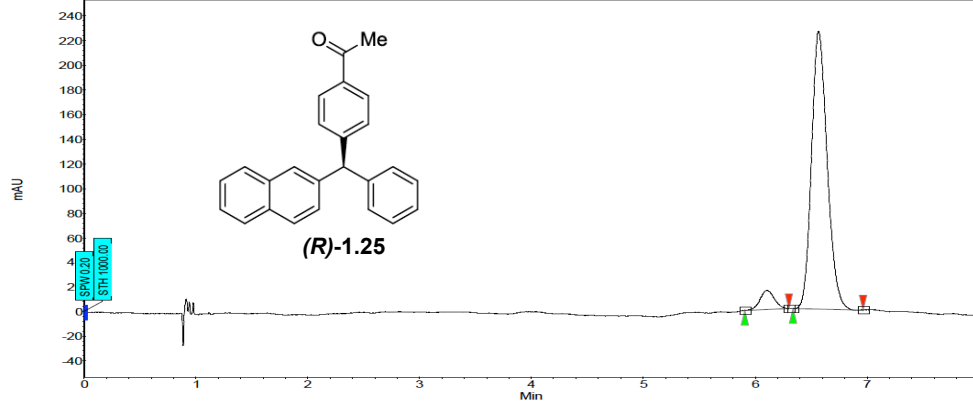
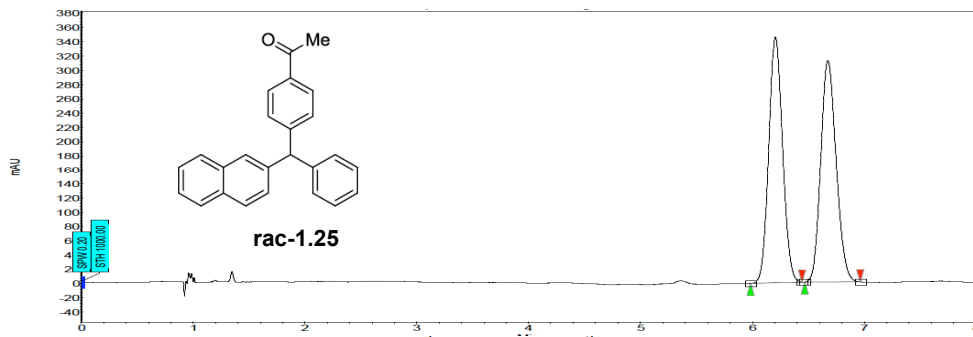
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1	UNKNOWN	9.72	10.06	10.55	0.00	94.00	359.5	82.9	94.005
2	UNKNOWN	10.63	10.98	11.30	0.00	6.00	19.5	5.3	5.995
Total						100.00	379.0	88.2	100.000



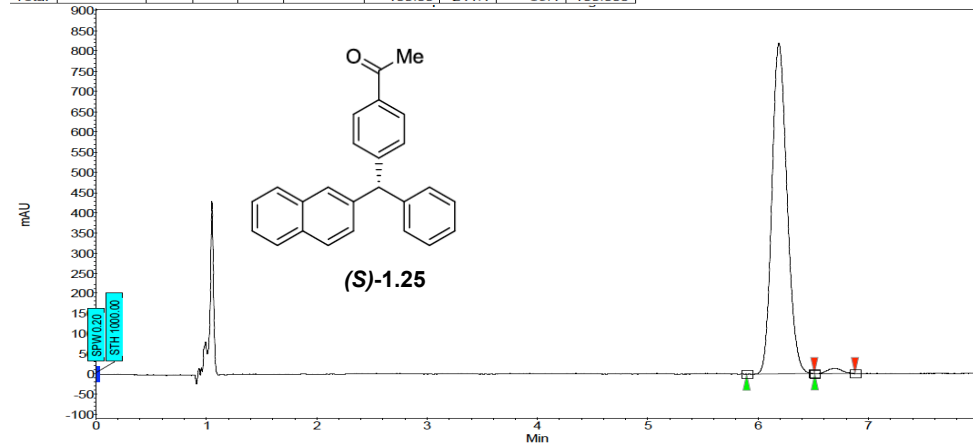
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1	UNKNOWN	7.04	7.35	7.67	0.00	21.30	157.7	25.0	21.299
2	UNKNOWN	7.70	7.94	8.35	0.00	78.70	540.3	92.5	78.701
Total						100.00	698.0	117.6	100.000



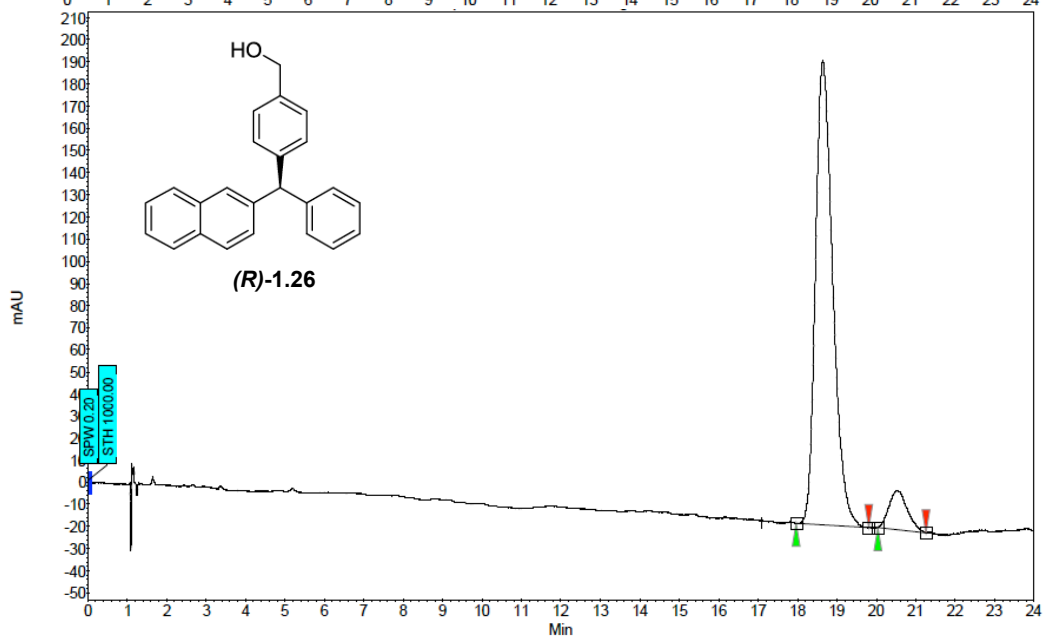
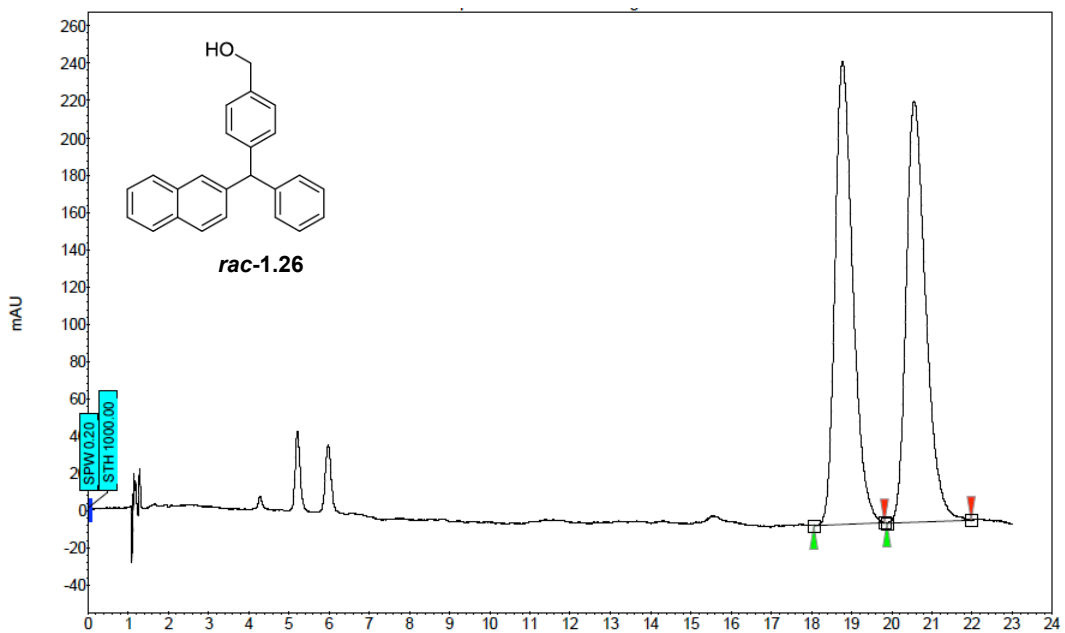
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1	UNKNOWN	6.59	6.84	7.17	0.00	94.37	663.8	106.4	94.372
2	UNKNOWN	7.27	7.46	7.70	0.00	5.63	38.2	6.3	5.628
Total						100.00	702.1	112.7	100.000



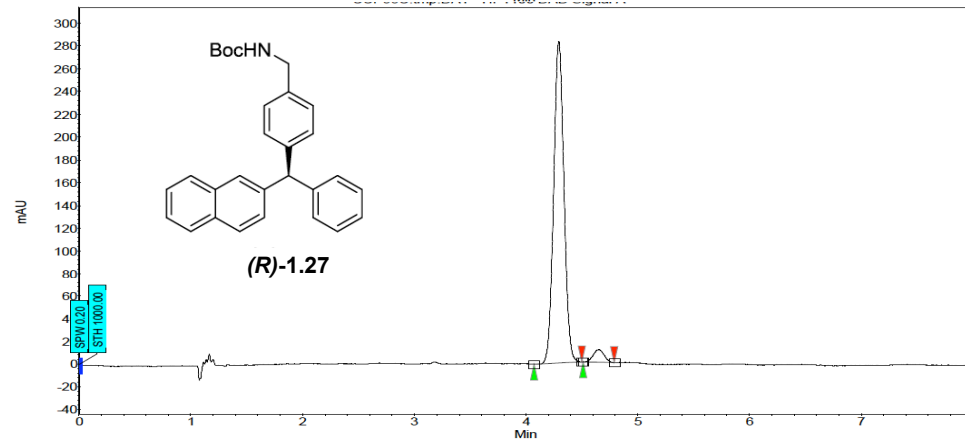
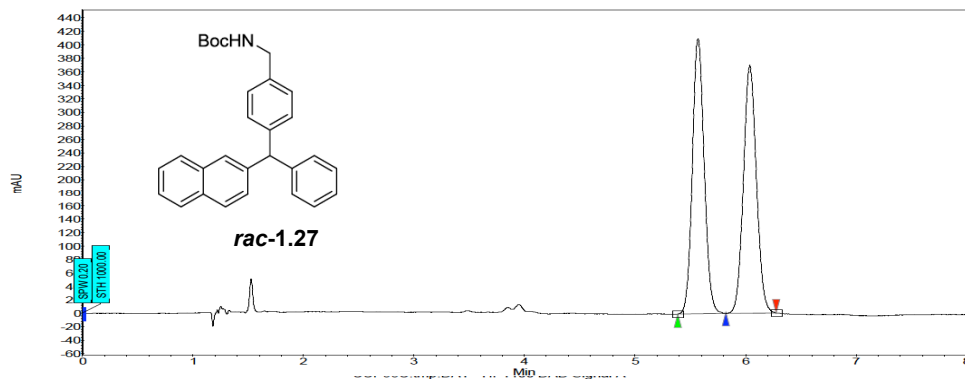
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
2	UNKNOWN	5.91	6.11	6.30	0.00	5.73	15.4	2.3	5.725
1	UNKNOWN	6.34	6.57	6.96	0.00	94.27	225.7	37.1	94.275
Total						100.00	241.1	39.4	100.000



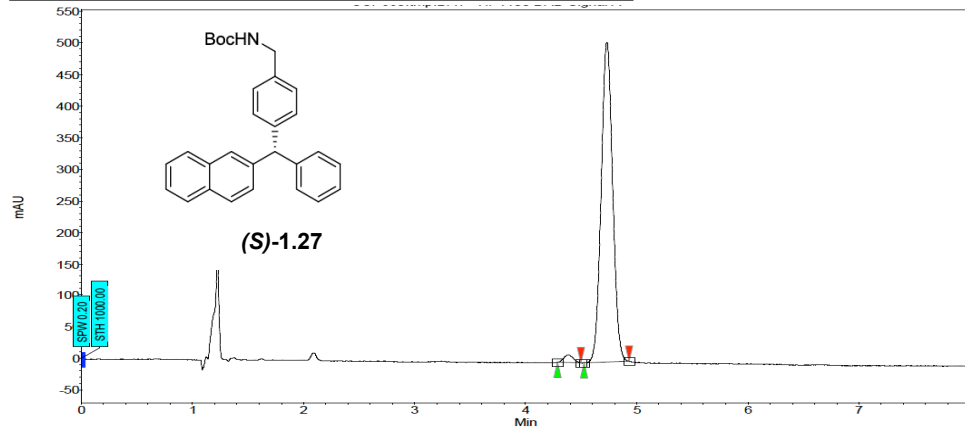
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	5.90	6.19	6.51	0.00	98.37	818.9	129.1	98.371
2	UNKNOWN	6.51	6.69	6.88	0.00	1.63	13.7	2.1	1.629
Total						100.00	832.6	131.2	100.000



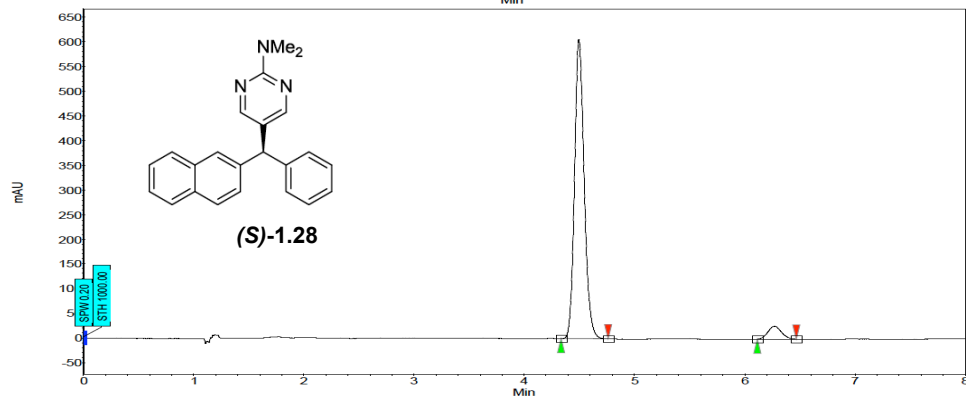
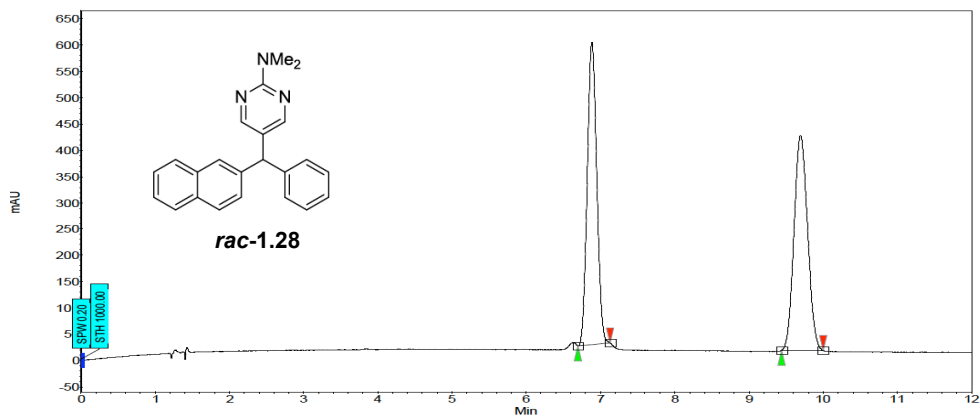
Index	Name	Start Time [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	17.97	18.64	19.81	0.00	91.88	209.7	106.3	91.876
2	UNKNOWN	20.04	20.53	21.26	0.00	8.12	17.7	9.4	8.124
Total						100.00	227.3	115.7	100.000



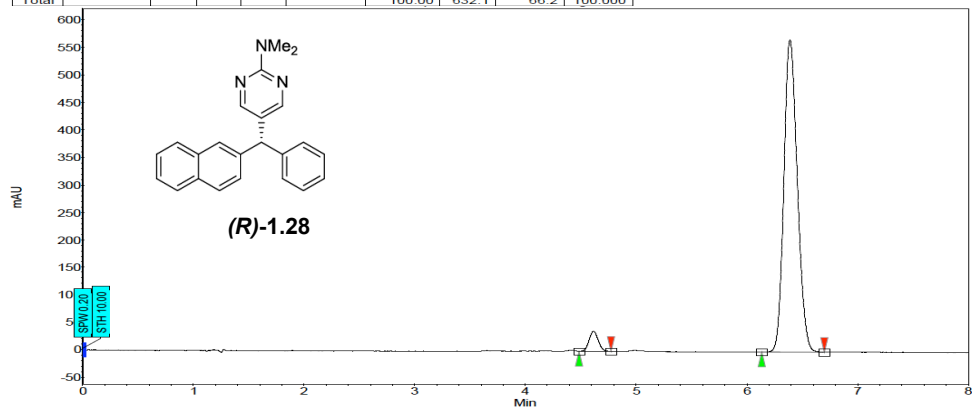
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1	UNKNOWN	4.07	4.29	4.50	0.00	96.08	283.0	30.0	96.085
2	UNKNOWN	4.51	4.66	4.79	0.00	3.92	11.0	1.2	3.915
Total						100.00	294.0	31.2	100.000



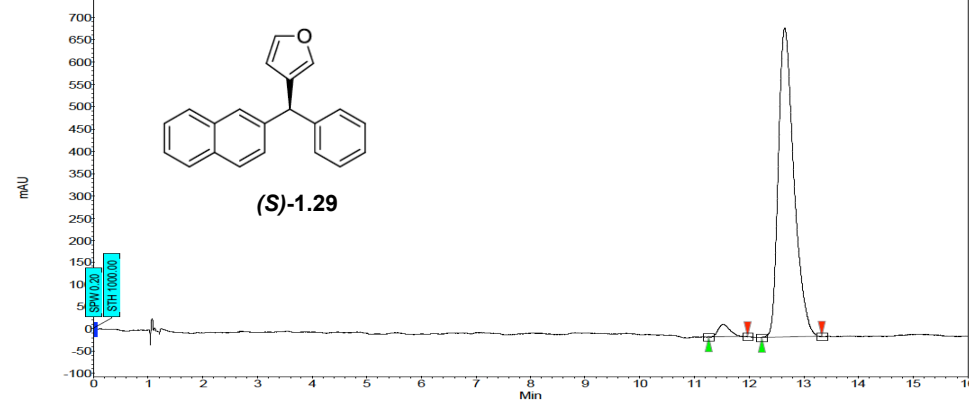
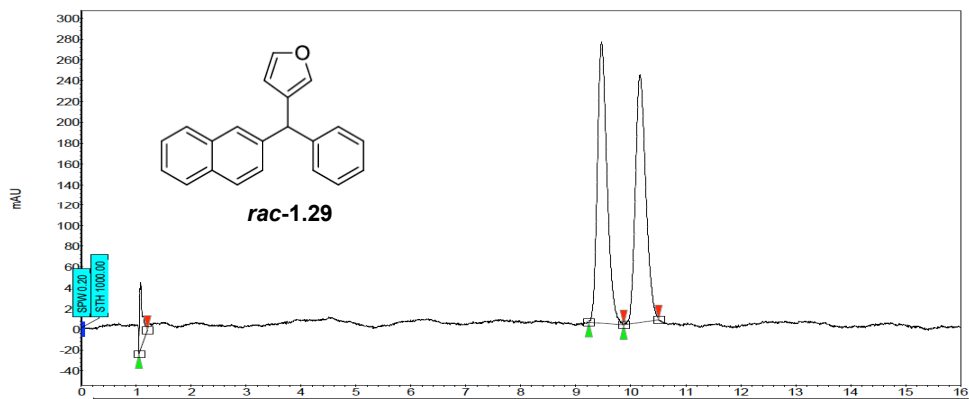
Index	Name	Start Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV Min]	Area [%]	
1	UNKNOWN	4.29	4.39	4.50	0.00	1.92	11.9	1.2	1.924
2	UNKNOWN	4.53	4.73	4.93	0.00	98.08	506.5	60.4	98.076
Total						100.00	518.5	61.6	100.000



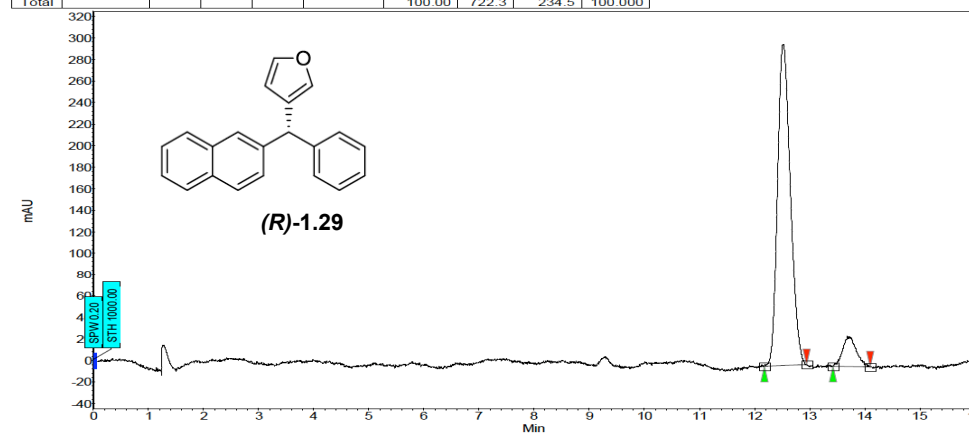
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1	UNKNOWN	4.33	4.50	4.76	0.00	94.50	606.4	62.5	94.501
2	UNKNOWN	6.12	6.27	6.47	0.00	5.50	25.7	3.6	5.499
Total						100.00	632.1	66.2	100.000



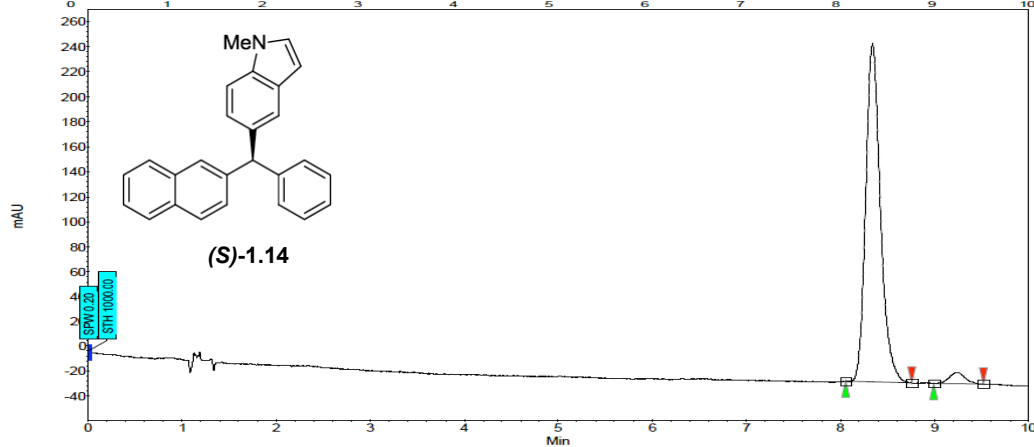
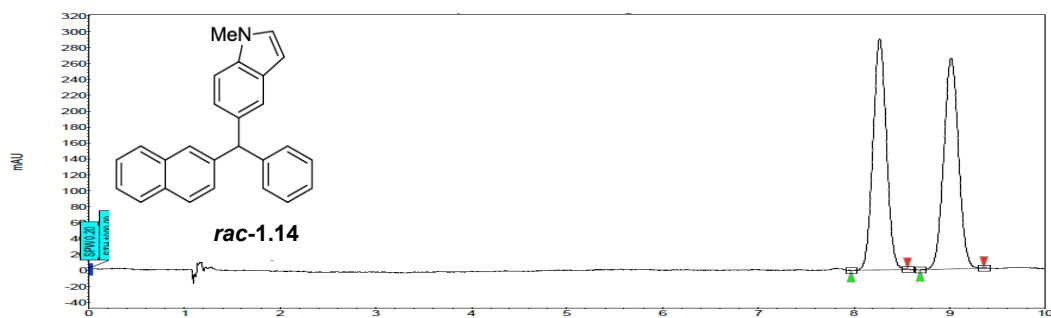
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV Min]	Area [%]
1	UNKNOWN	4.48	4.62	4.77	0.00	4.20	36.6	3.5	4.201
2	UNKNOWN	6.13	6.39	6.70	0.00	95.80	567.9	79.6	95.799
Total						100.00	604.4	83.1	100.000



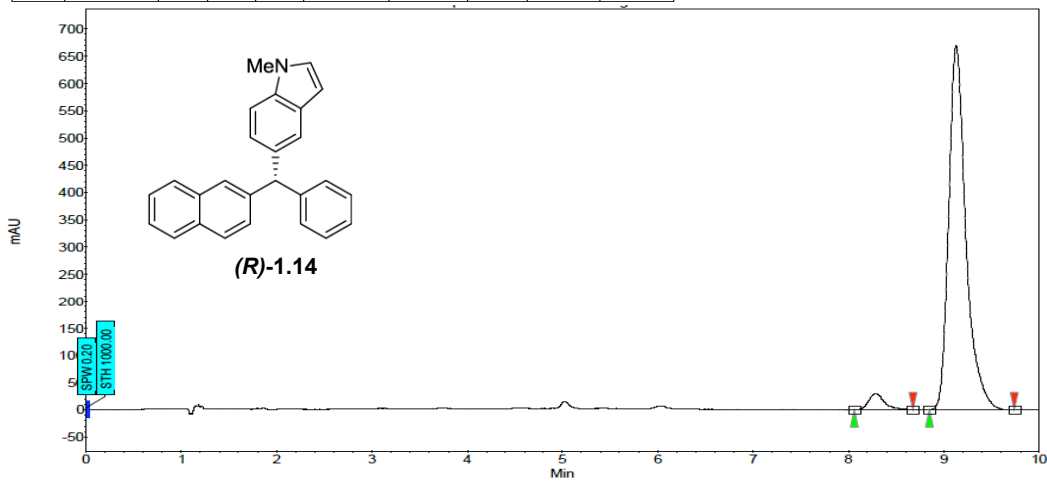
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [µV]	Area [µV Min]	Area [%]
2	UNKNOWN	11.26	11.53	11.97	0.00	2.94	28.1	6.9	2.937
1	UNKNOWN	12.23	12.65	13.32	0.00	97.06	694.1	227.6	97.063
Total						100.00	722.3	234.5	100.000



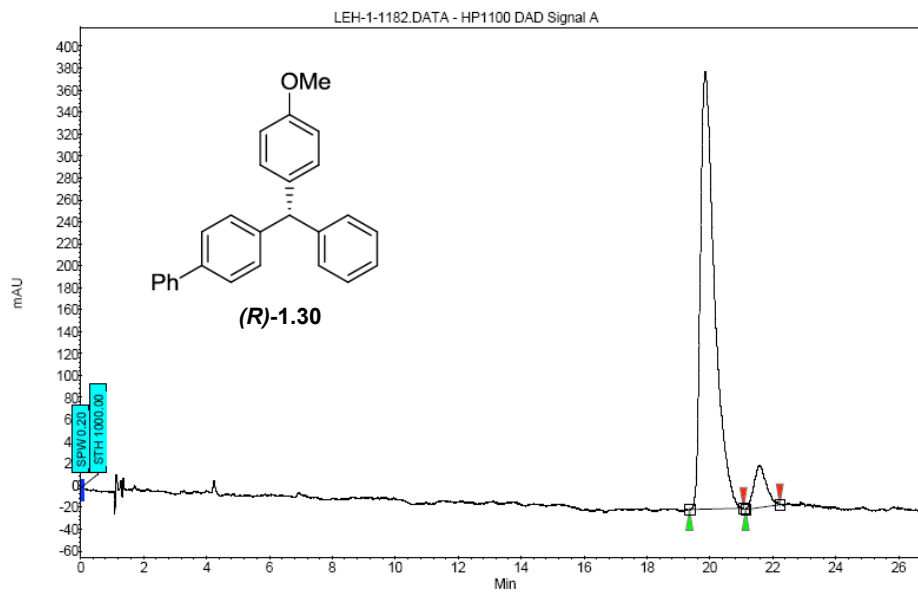
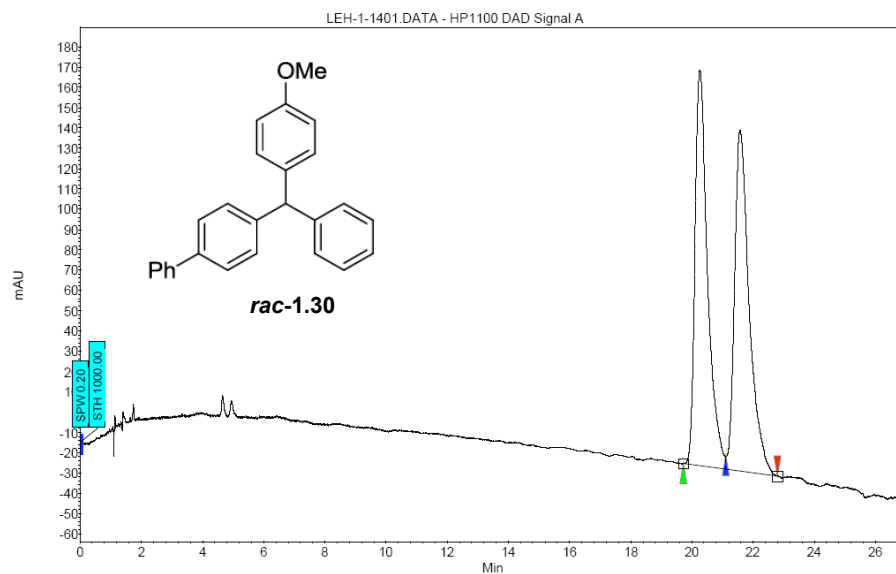
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [µV]	Area [µV Min]	Area [%]
1	UNKNOWN	12.18	12.52	12.95	0.00	90.99	298.5	80.3	90.991
2	UNKNOWN	13.42	13.69	14.10	0.00	9.01	27.8	7.9	9.009
Total						100.00	326.3	88.2	100.000



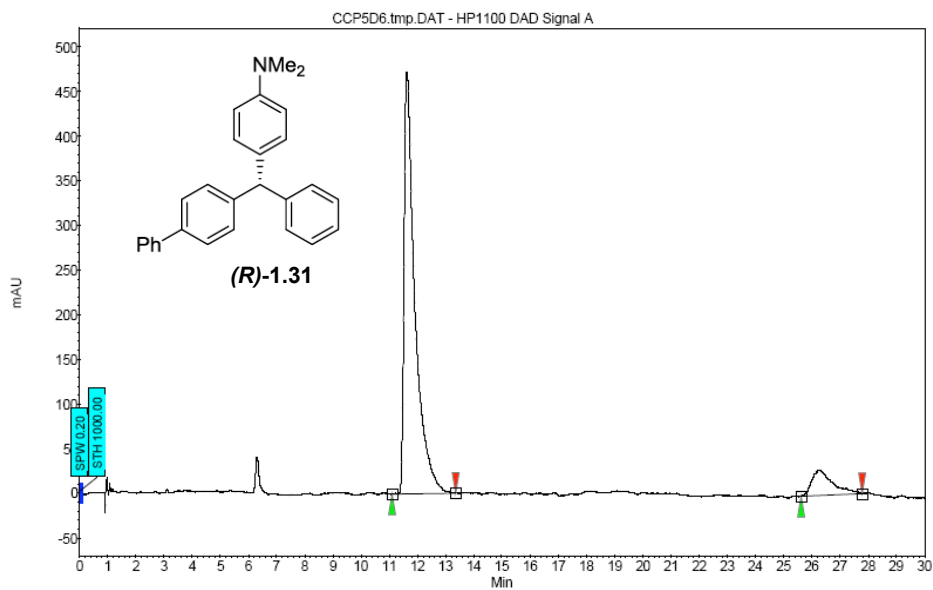
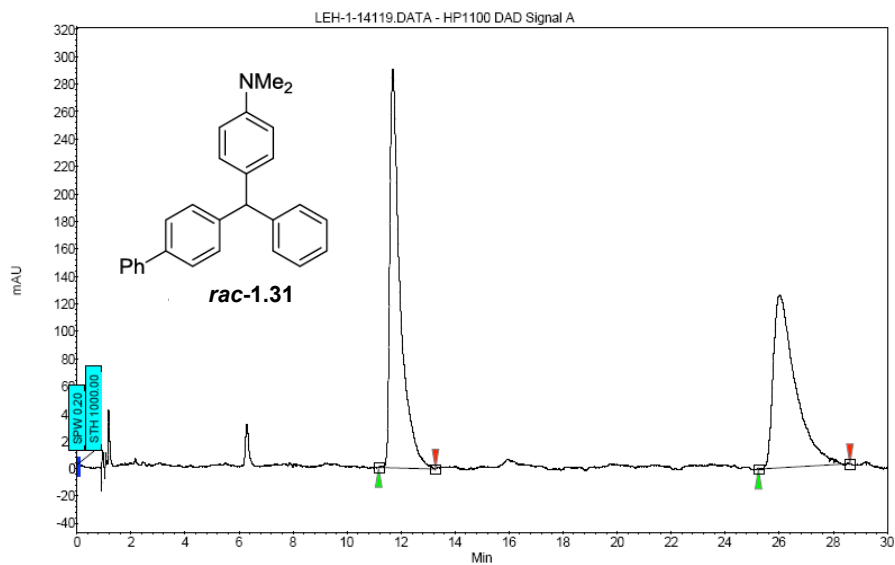
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	8.05	8.34	8.76	0.00	96.61	270.9	49.0	96.606
2	UNKNOWN	8.99	9.24	9.52	0.00	3.39	8.9	1.7	3.394
Total						100.00	279.8	50.7	100.000



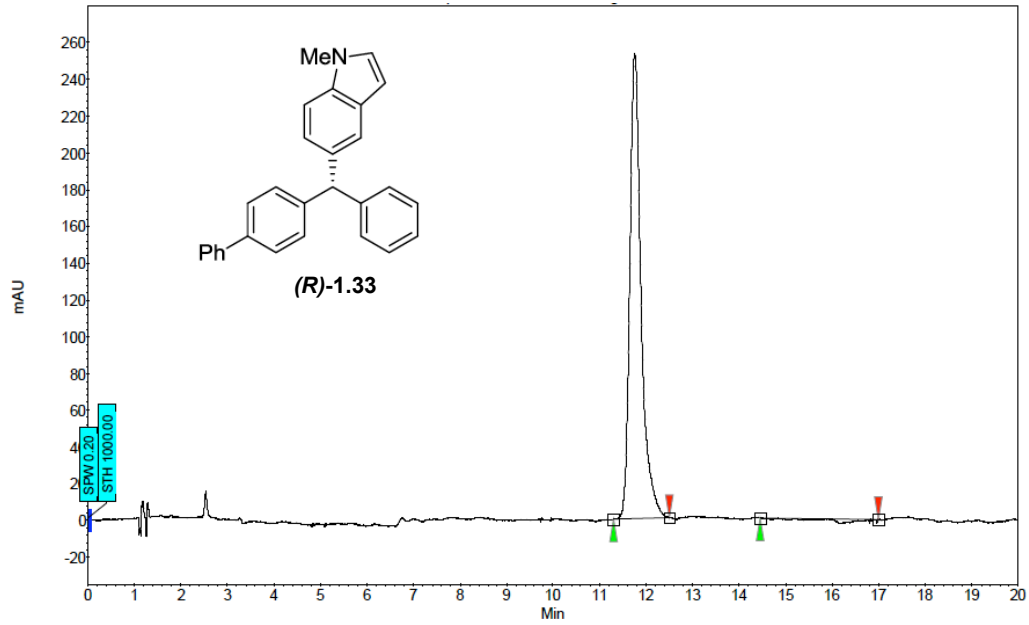
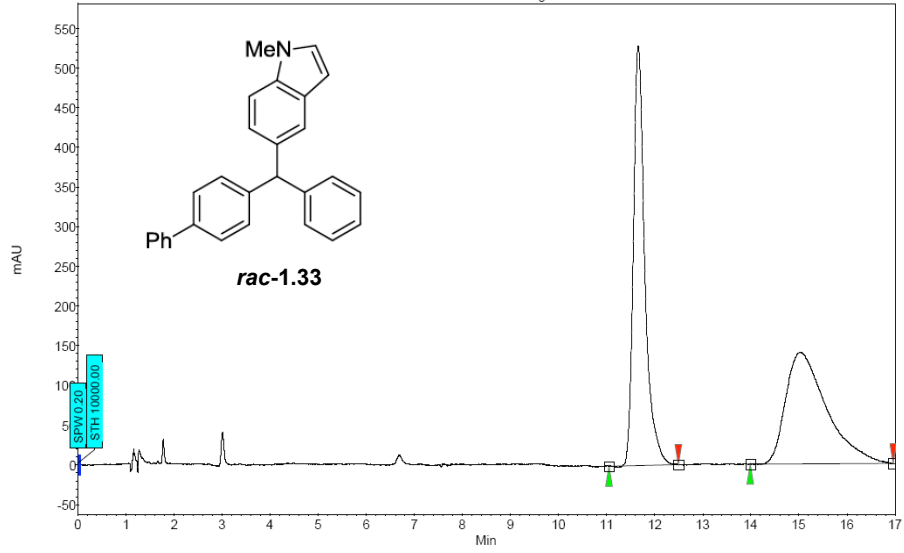
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1	UNKNOWN	8.06	8.28	8.67	0.00	3.91	29.8	5.7	3.910
2	UNKNOWN	8.85	9.13	9.73	0.00	96.09	668.3	139.6	96.090
Total						100.00	698.2	145.3	100.000



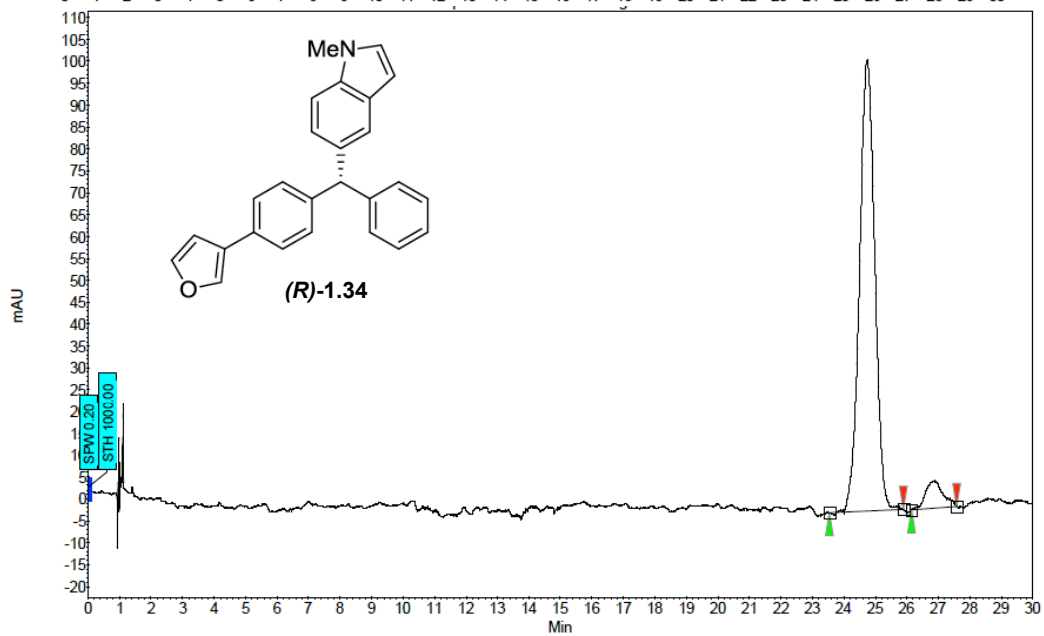
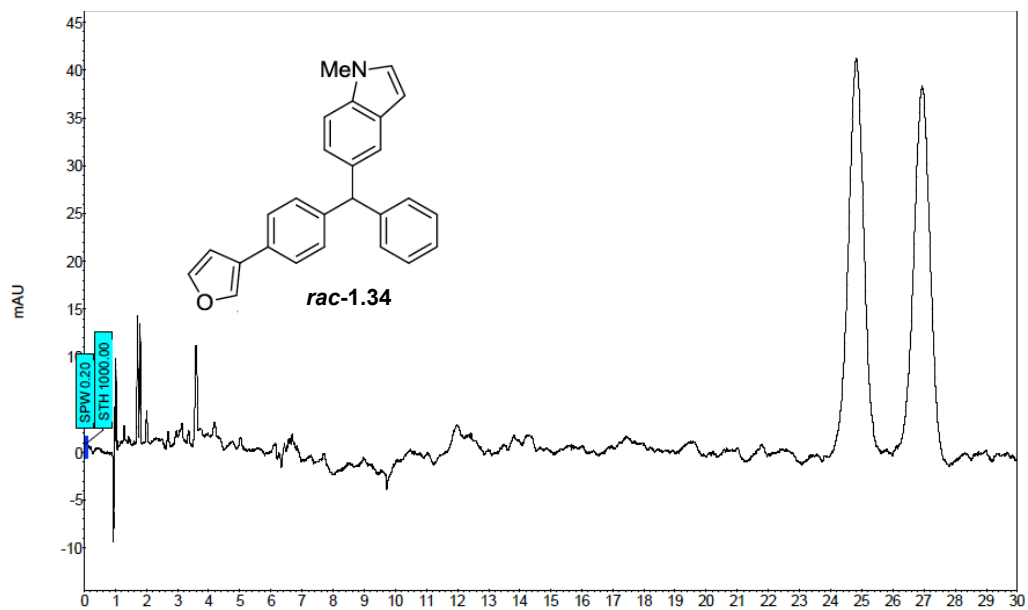
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1	UNKNOWN	19.35	19.85	21.07	0.00	92.25	398.9	207.1	92.249
2	UNKNOWN	21.13	21.57	22.22	0.00	7.75	38.6	17.4	7.751
Total						100.00	437.5	224.5	100.000



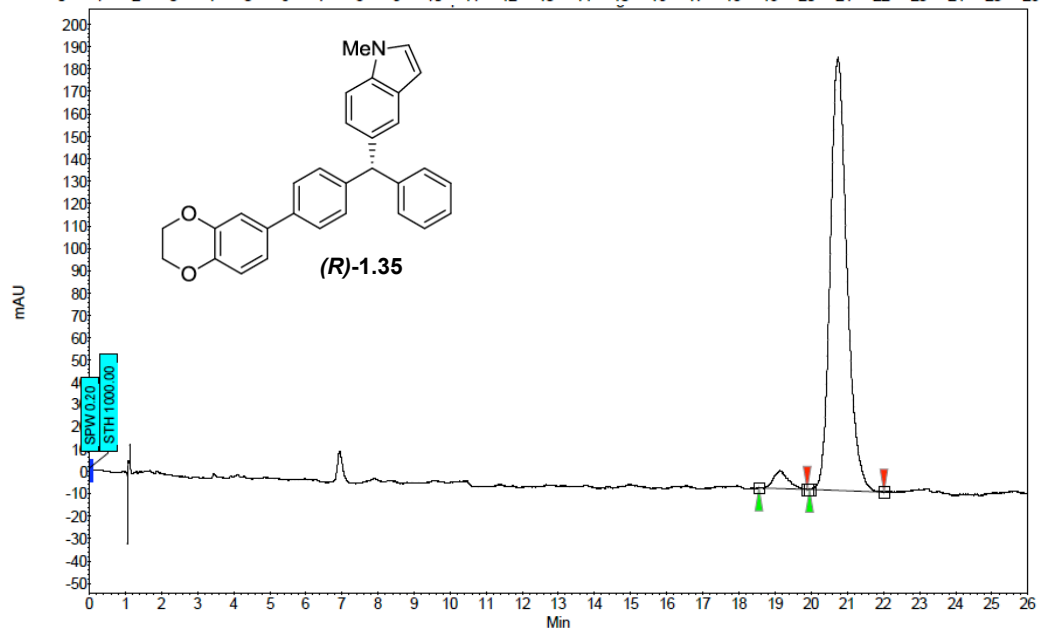
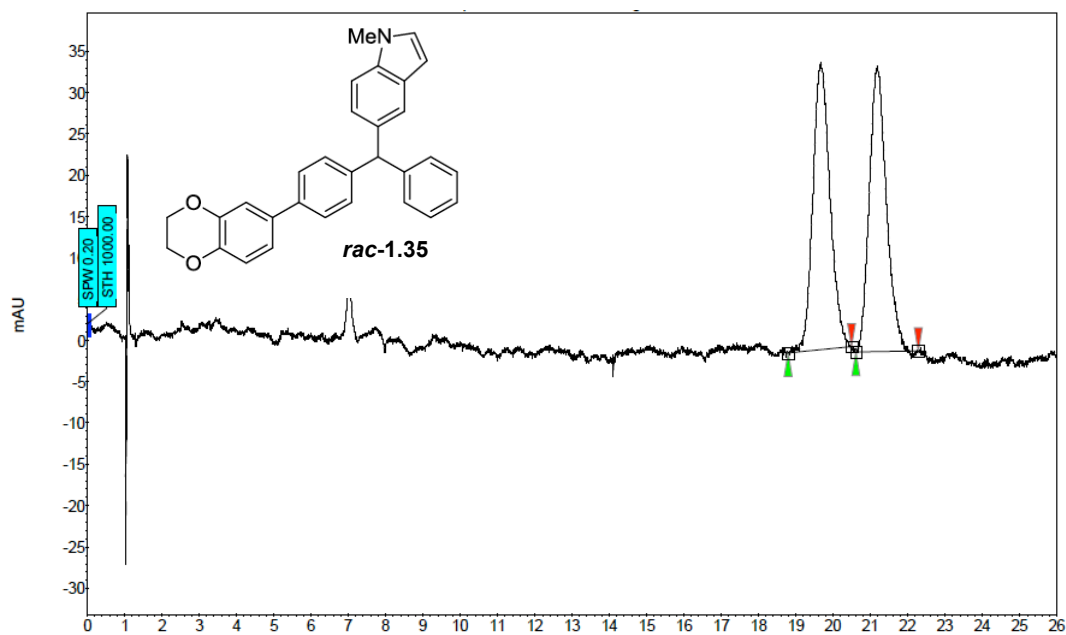
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1	UNKNOWN	11.09	11.62	13.35	0.00	89.58	472.3	210.6	89.580
2	UNKNOWN	25.61	26.26	27.77	0.00	10.42	28.2	24.5	10.420
Total						100.00	500.5	235.1	100.000



Index	Name	Start Time [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	11.31	11.76	12.51	0.00	97.87	252.3	70.7	97.873
2	UNKNOWN	14.45	16.15	17.00	0.00	2.13	2.3	1.5	2.127
Total						100.00	254.6	72.2	100.000

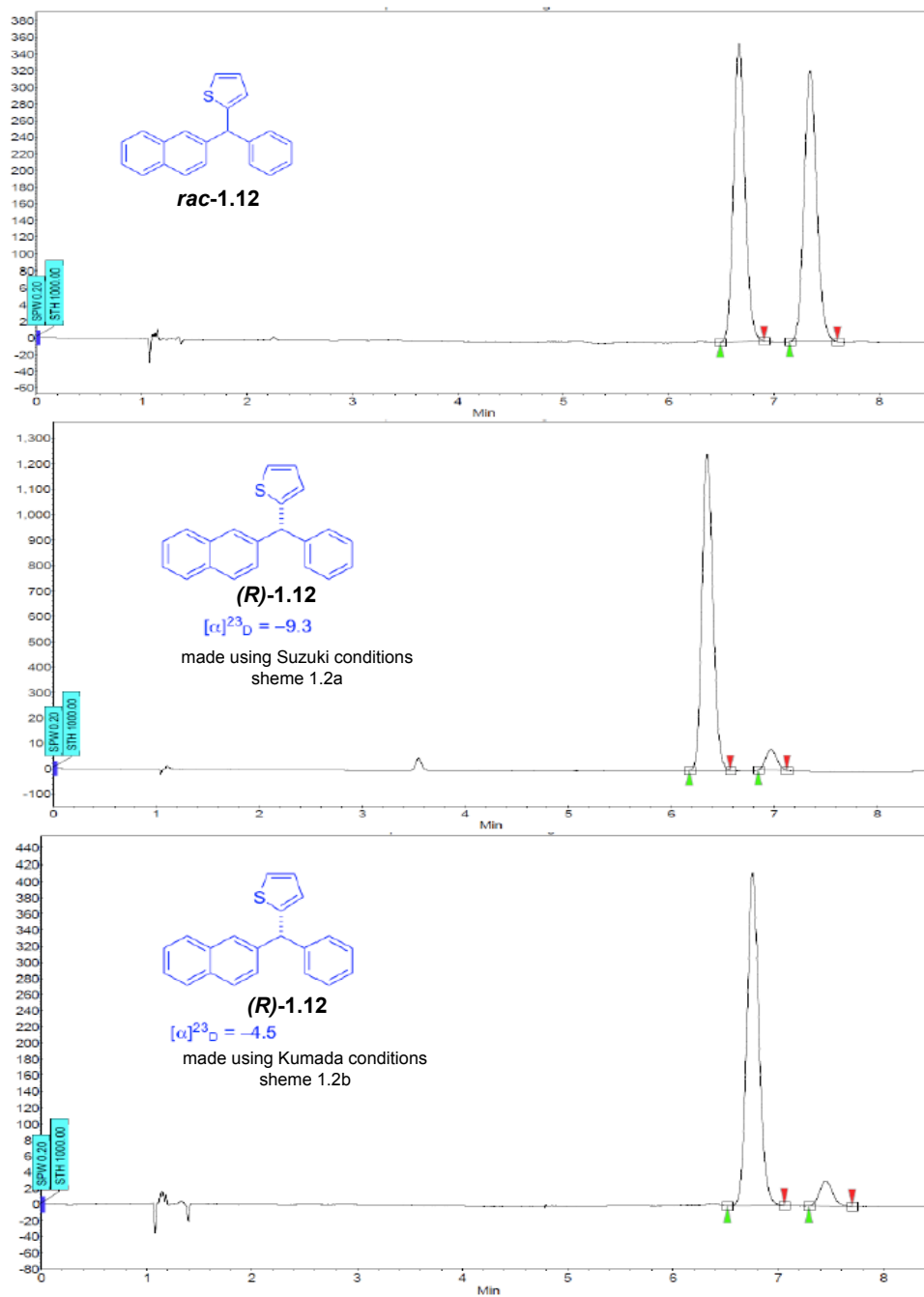


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μV]	[μV.Min]	[%]
1	UNKNOWN	23.55	24.74	25.88	0.00	93.55	103.0	57.7	93.545
2	UNKNOWN	26.15	26.89	27.58	0.00	6.45	6.2	4.0	6.455
Total						100.00	109.2	61.7	100.000



Index	Name	Start Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	18.56	19.13	19.88	0.00	3.46	7.8	3.7
2	UNKNOWN	19.96	20.74	22.01	0.00	96.54	193.6	103.4
Total					100.00	201.4	107.1	100.000

SFC traces (complete SFC data can be found in section X):



Chapter 2

Propargylic Carbamates Facilitate Regio- and Stereoselective

Nickel-Catalyzed Hydroarylation of Alkynes

2.1 Introduction

Functionalized alkenes are present in pharmaceuticals¹ and natural products, and are commonly used as building blocks in synthesis² and polymer chemistry³ (Figure 2.1).⁴ Hydroarylation reactions of alkynes offer an alternative manifold for the facile synthesis of substituted alkenes. The major challenges associated with hydroarylation of alkynes has primarily been control of regioisomeric ratios and control of *E/Z* stereochemistry of the resulting olefin without relying on steric or electronic biases in the substrate.⁵ For example, early work from Hayashi demonstrated that Rh(I) catalysts cleanly furnished hydroarylated products using symmetrical alkynes and arylboronic acids, however, using unsymmetrical starting materials that lacked strong electronic or steric bias resulted in mixtures of regioisomers (Scheme 2.1a).⁶ The Lautens group judiciously addressed this problem with the use of pyridine or alcohol directing

¹ (a) Liu, X.; Shimizu, M.; Hiyama, T. *Angew. Chem. Int. Ed.* **2004**, *43*, 879. (b) Levenson, A. S.; Jordan, V. C. *Eur. J. Cancer*, **1999**, *35*, 1628. (c) Prasit, P.; Wang, Z.; Brideau, C.; Chan, C. C.; Charleson, S.; Cromlish, W.; Ethier, D.; Evans, J. F.; Ford-Hutchinson, A. W.; Gauthier, J. Y.; Gordon, R.; Guay, J.; Gresser, M.; Kargman, S.; Kennedy, B.; Leblanc, Y.; Leger, S.; Mancini, J.; O'Neill, G. P.; Ouellet, M.; Percival, M. D.; Perrier, H.; Riendeau, D.; Rodger, I.; Tagari, P.; Therien, M.; Vickers, P.; Wong, E.; Xu, L. J.; Young, R. N.; Zamboni, R.; Boyce, S.; Rupniak, N.; Forrest, M.; Visco, D.; Patrick, D. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 1773. (d) Fallis, A. G.; Forgiione, P. *Tetrahedron*, **2001**, *28*, 5899.

²For examples of alkene utility in synthesis: (a) Hoffmann, N. *Chem. Rev.* **2008**, *108*, 1052. (b) Sato, F.; Urabe, H.; Okamoto, S. *Chem. Rev.* **2000**, *100*, 2835. (c) Nicolaou, K.C.; Snyder, S. A.; Montagnon, T.; Vassilikogiannakis, G. *Angew. Chem. Int. Ed.* **2002**, *41*, 1668.

³ Hall, H. K. *Angew. Chem., Int. Ed.* **1983**, *22*, 440.

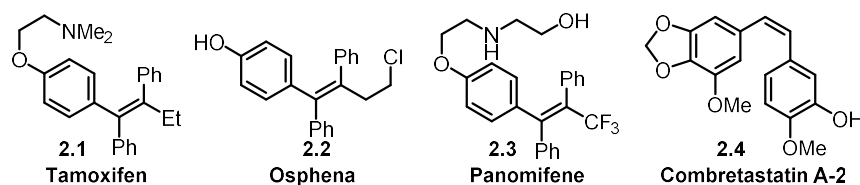
⁴ (a) Example of natural products: Williams, R.B.; Norris, A.; Slebodnick, C.; Merola, J.; Miller, J. S.; Andriantsiferana, R.; Rasamison, V. E.; Kingston, D. G. I. *J. Nat. Prod.* **2005**, *68*, 1371. (b) Lin, C. M.; Ho, H. H.; Pettit, G. R.; Hamel, E. *Biochemistry* **1989**, *28*, 6984.

⁵ for examples using electronic biased substrates (a) Lin, P.-S.; Jeganmohan, M.; Cheng, C.-H. *Chem. - Eur. J.* **2008**, *14*, 11296. (b) Bai, Y.; Yin, J.; Kong, W.; Mao, M.; Zhu G. *Chem. Commun.*, **2013**, *49*, 7650. (c) Cui, W.; Yin, J.; Zheng, R.; Cheng, C.; Bai, Y.; Zhu, G. *J. Org. Chem.* **2014**, *79*, 3487. Also see reference 6 for examples of steric and electronic bias

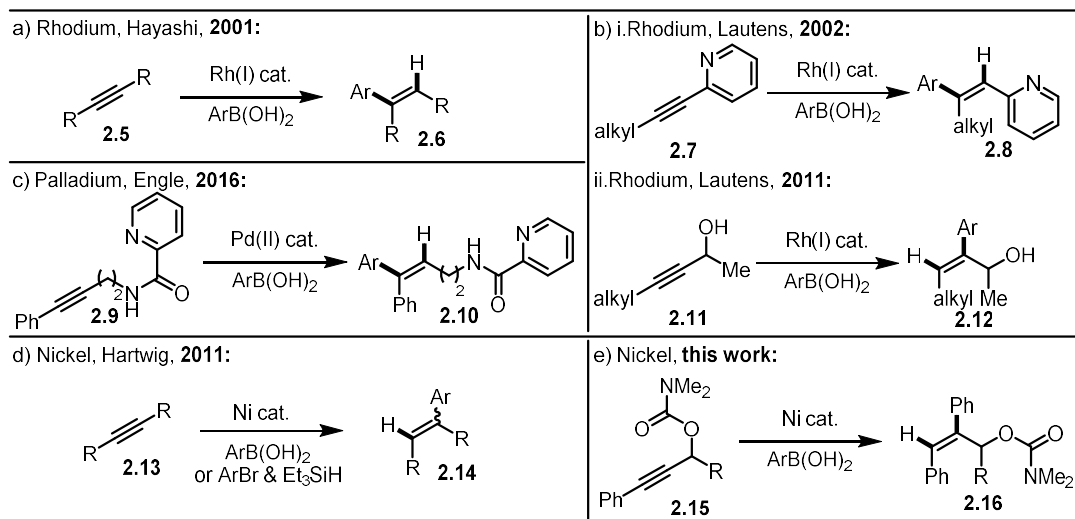
⁶(a) Hayashi, T.; Inoue, K.; Taniguchi, N.; Ogasawara, M. *J. Am. Chem. Soc.* **2001**, *123*, 9918. (b) Zhang, W.; Liu, M.; Wu, H.; Ding, J.; Cheng, J. *Tetrahedron Lett.* **2008**, *49*, 5214. (c) Genin, E.; Michelet, V.; Genêt, J.-P. *Tetrahedron Lett.* **2004**, *45*, 4157.

groups that could deliver hydroarylated products with excellent regio- and stereocontrol (Scheme 2.1b).⁷ Efforts to employ group 10 metals such as palladium and nickel in hydroarylation reactions have been explored with low regio- and stereocontrol (Scheme 2.1d).⁸ However, Engle and co-workers have recently reported the use of pendant directing groups that address these obstacles with great success in palladium-catalyzed reactions (Scheme 2.1c).⁹ To date, hydroarylation reactions that use nickel catalysts suffer from both regio- and stereocontrol of the resulting olefin. (Scheme 2d).

Figure 2.1. Examples of pharmaceutical agents and natural products bearing alkenes



Scheme 2.1. Summary of transition metal catalyzed hydroarylations using boronic acids



⁷ (a) Pantelev, J.; Huang, R. Y.; Lui, E. K. J.; Lautens, M. *Org. Lett.* **2011**, *13*, 5314. (b) Lautens, M.; Yoshida, M. *Org. Lett.* **2002**, *4*, 123.

⁸(a) Shirakawa, E.; Takahashi, G.; Tsuchimoto, T.; Kawakami, Y. *Chem. Commun.* **2001**, 2688 (b) Hartwig, J. F. *Science* **2011**, *333*, 1423. (c) Oh, C. H.; Jung, H. H.; Kim, K. S.; Kim, N. *Angew. Chem., Int. Ed.* **2003**, *42*, 805. (d) Xu, X.; Chen, J.; Gao, W.; Wu, H.; Ding, J.; Su, W. *Tetrahedron* **2010**, *66*, 2433

⁹ Liu, Z.; Derosa, J.; Engle, K. M. *J. Am. Chem. Soc.*, **2016**, *138*, 13076.

Moving forward, efforts in our group have focused on the use of base metals and propargylic directing groups to address this challenge. Previous work in our group has demonstrated that carbamates can serve as directing groups for nickel catalysts in Suzuki coupling reactions. Thus, we hypothesized that propargylic carbamates could also serve as directing groups for hydroarylation reactions as well. Herein, we report a regio- and stereoselective hydroarylation of alkynes under mild conditions (Scheme 2.1e).

2.2 Development of Nickel-Catalyzed Directed Hydroarylation

We began our investigation by investigating a range of propargylic directing groups including pivalates, carbonates and carbamates. Using pivalates and carbonates as directing groups led to undesired formation of allene **2.19** while still forming small amounts of desired product. A mixture of byproducts and only 10% hydroarylated product was observed when free secondary propargyl alcohols were used. Interestingly, nearly identical results were observed without the alcohol or directing groups present (Table 2.1, entries 4 and 5). Employing bulky carbamates such as diisopropyl carbamate led to a severe decrease in product formation. However, moving to sterically smaller alkyl groups such as pyrrolidine carbamate led to an increase in product formation, with a dimethyl carbamate giving the highest yields. A range of ligands were investigated during the optimization. The reaction performs well with monodentate phosphine ligands. During further studies of ligand effects, it was uncovered that catalysts with sterically bulky Buchwald type ligands such as TrixiePhos were uniquely capable of suppressing the formation of allene, while still furnishing the desired product in good yields.

Interestingly, the reaction performs much better in the absence of bases such as *t*-BuOK and K₃PO₄, commonly used bases for transmetalation of Suzuki reagents. Furthermore, the addition of just one equivalent of LiCl to the reaction mixture led to a complete shutdown of reactivity and

recovery of only starting material (Table 2.1, entry 12). Additionally, during an investigation of solvents, it was found that the use of strongly coordinating solvents such as DMF also shut down reactivity (Table 2.1, entry 11).

Table 2.1. Optimization of hydroarylation reaction conditions

entry	variation from standard conditions	recovered 2.17 (%) ^a	2.18 (%) ^a	2.19 (%) ^a
1	none	<2	70	<2
2	2.17a instead of 2.17	57	16	14
3	2.17b instead of 2.17	81	8	<2
4	2.17c instead of 2.17	73	20	<2
5	2.20 instead of 2.17	57	19	<2
6	no nickel	100	0	0
7	no ligand	8	55	15
8	SPhos instead of TrixiePhos	16	63	2
9	BrettPhos instead of TrixiePhos	11	50	9
10	CPhos instead of TrixiePhos	12	50	12
11	DMF instead of THF	86	13	7
12	1 equivalent of LiCl	100	0	0
13	1 equivalent of K ₃ PO ₄	10	40	6
14	PhBpin instead of PhB(OH) ₂	78	8	4
15	(PhBO) ₃ instead of PhB(OH) ₂	65	19	6

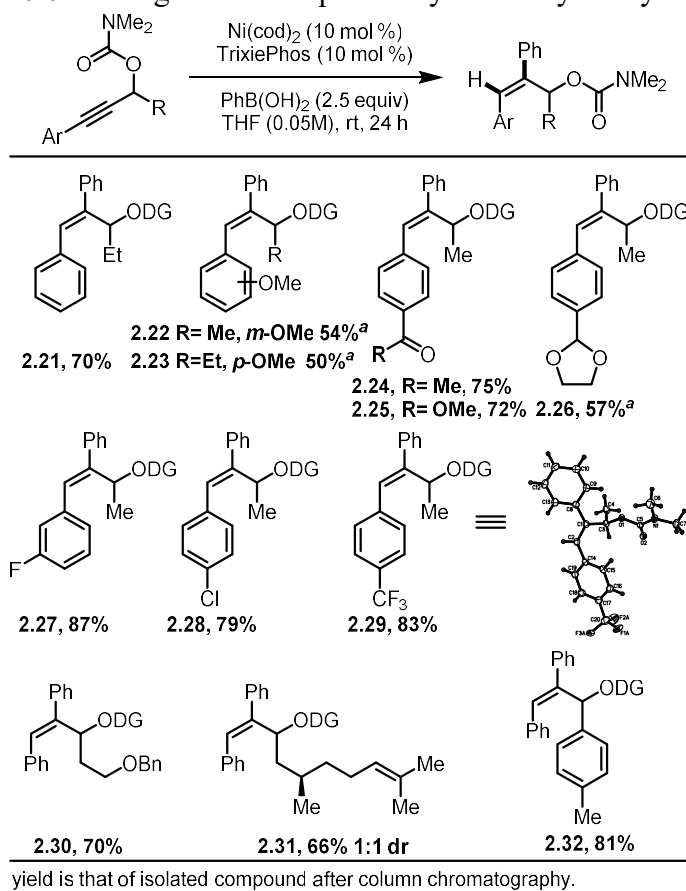
^aYield determined by ¹H NMR spectroscopy using PhTMS as internal standard.

2.3 Scope of Substrates for Hydroarylation

Having identified suitable reaction conditions for model substrate **2.17**, we turned our attention to interrogating a broad scope of internal alkynes (Table 2.2). Substrates bearing electron-deficient arenes provided the highest yields of hydroarylated product at room temperature while electron-rich arenes required heating (Table 2.2, entry **2.22** and **2.23**). Ketone and ester substituents were well tolerated, however, free aldehydes were not. Gratifyingly, this oxidation state can still be incorporated as an acetal, although heating is required. The presence of aryl fluorides and chlorides did not produce the corresponding Suzuki-coupled products, allowing for

further orthogonal functionalization (Table 2.2, entry **2.27** and **2.28**). An X-ray crystallographic structure of trifluoromethyl-substituted **2.29** was obtained, supporting the completely regioselective *cis*-hydroarylation. Benzyl-protected alcohols on the alkyl moiety of the starting materials were carried through the reaction untouched. Citronellal-derived substrate **2.31** shows that alkenes do not shut down reactivity and can be manipulated in downstream reactions. Surprisingly, sp^2 -hybridized side chains yielding both allylic and benzylic-activated carbamates did not undergo Tsuji-Trost type cross-coupling and provided products in high yields (entry **2.32**).

Table 2.2. Investigation of scope in alkynes for hydroarylation

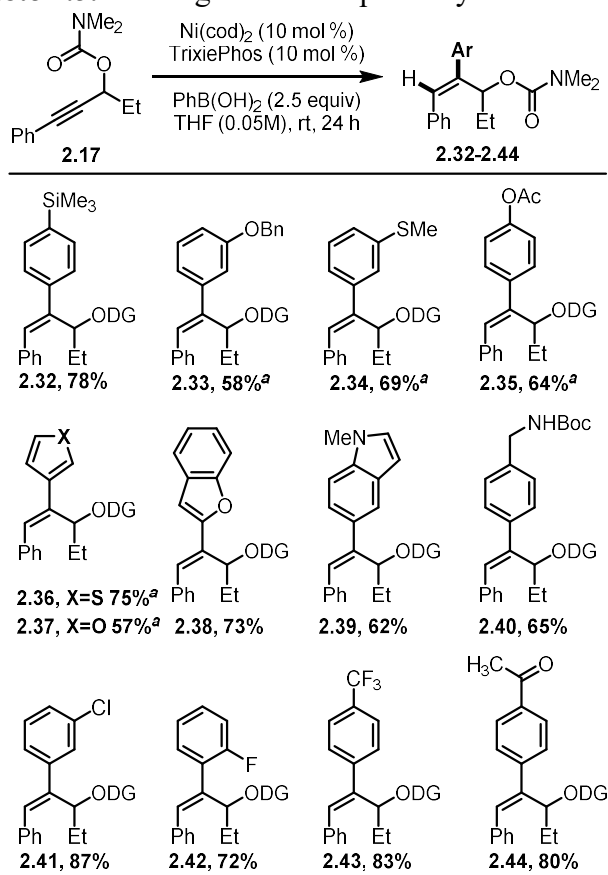


2.4 Scope of Arylboronic Acid for Hydroarylation

Next, we turned our attention to investigating the scope of arylboronic acids (Table 2.3). Again, a similar trend in sensitivity to electronics was observed and electron-deficient arylboronic acids provided the highest yields at room temperature while electron-rich arylboronic acids

required heating. A variety of functional groups were well-tolerated, allowing for the incorporation of phenyl TMS, acetoxy, ether, thioether, and trifluoromethyl moieties (Table 2.3, entries **2.32-2.34** & **2.43**). Ketones and Boc-protected amines were also well tolerated and provided hydroarylated product in good yields. Chloro- and fluorophenyl products, such as **2.41** and **2.42** did not participate in either hydrodehalogenation or cross-coupling. Furthermore, boronic acids containing heterocycles such as furan and thiophene underwent hydroarylation smoothly at elevated temperatures while benzofuran and indoleboronic acids could be coupled at room temperature. (Table 2.3, entries **2.38** and **2.39**).

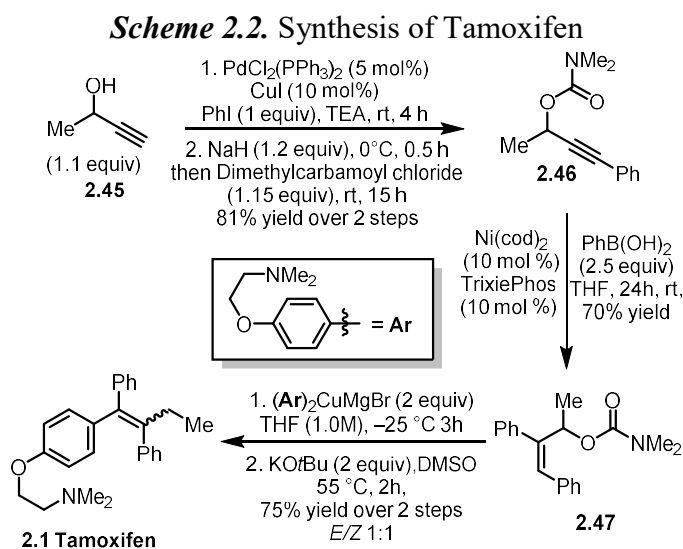
Table 2.3. Investigation of scope in aryl boronic acid



2.5 Synthesis of Tamoxifen

Functionalized allylic alcohols are common synthetic building blocks en route to more complex molecular scaffolds. Allylic carbamates can be used in Tsuji-Trost chemistry as well as

copper-mediated substitutions allowing for a range of back-to-back transition-metal-catalyzed reactions following hydroarylation. Scheme 2.2 demonstrates one of the many synthetic uses of these products. **2.47** can be converted to tamoxifen in two steps via a cuprate-mediated allylic substitution of the allylic carbamate followed by isomerization of the double bond to form the conjugated target **2.13**. As previously stated, these alkenes are known to be biologically active, and have been described to have anti-cancer activity. Hence, the herein described method can serve as a useful platform to generate a library of tamoxifen-like structures for SAR studies.

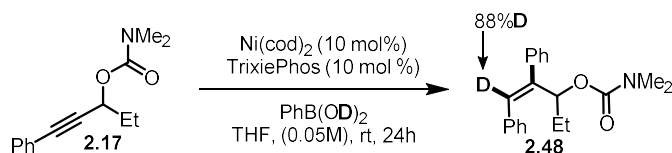


2.6 Mechanistic Studies and Deuterium Labeling

Lastly, we were interested gaining insight into the mechanism of the reaction by determining the origin of the hydrogen atom in our hydroarylation reaction. Initial attempts to replace phenylboronic acid with phenyl boroxine and deuterium oxide resulted in low yields of product (17-30%) with 99% deuterium incorporation. However, using $\text{PhB}(\text{OD})_2$ led to a 68% yield of product **2.48** with 88% deuterium incorporation (Equation 2.3). These results led us to definitively conclude that the acidic protons in the boronic acid were the source of hydrogen in the hydroarylation. These results are consistent with a mechanism that involves a directed hydrometalation to afford a vinylnickelintermediate, which can then undergo transmetallation and

reductive elimination to furnish the desired product. This is also consistent with the previously proposed hydroarylation reactions that do not require base.^{8c}

Equation 2.3. Deuterium Labeled Boronic Acid Reveals Origin of Hydrogen in Reaction



2.7 Conclusion

In conclusion, we have developed a regio- and stereoselective nickel-catalyzed hydroarylation of alkynes using a propargyl carbamate as a directing group. The reaction is tolerant of a range of functional groups and heterocycles. Furthermore, the synthesis of tamoxifen can be completed in two, operationally simple, steps in good overall yield.

2.8 Experimental Details

General Procedures

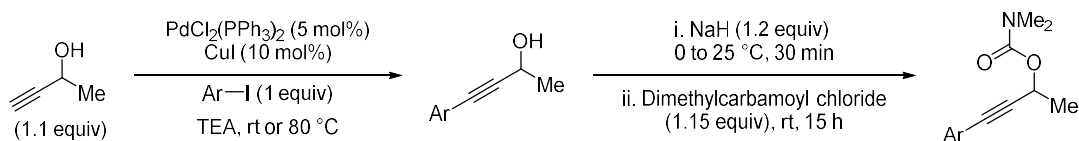
All hydroarylations were carried out under an atmosphere of N₂, or Ar when noted. All glassware was oven- or flame-dried prior to use. Tetrahydrofuran (THF), diethyl ether (Et₂O), dichloromethane (CH₂Cl₂), and dimethylacetamide (DMA) were degassed with Ar and then passed through two 4 x 36 inch columns of anhydrous neutral A-2 alumina (8 x 14 mesh; LaRoche Chemicals; activated under a flow of argon at 350 °C for 12 h) to remove H₂O. All other solvents utilized were purchased “anhydrous” commercially, or purified as described. ¹H NMR spectra were recorded on Bruker DRX-400 (400 MHz ¹H, 100 MHz ¹³C, 376.5 MHz ¹⁹F) or CRYO-500 (500 MHz ¹H, 125.7 MHz ¹³C) spectrometers. Proton chemical shifts are reported in ppm (δ) relative to internal tetramethylsilane (TMS, δ 0.00). Data are reported as follows: chemical shift (multiplicity [singlet (s), broad singlet (br s), doublet (d), doublet of doublets (dd), triplet (t), doublet of triplets (dt), doublet of doublet of triplets (ddt), triplet of triplets (tt), quartet (q), quintet (quin), apparent doublet (ad), apparent triplet (at), multiplet (m)], coupling constants [Hz], integration). Carbon chemical shifts are reported in ppm (δ) relative to TMS with the respective solvent resonance as the internal standard (CDCl₃, δ 77.16 ppm). Unless otherwise indicated, NMR data were collected at 25 °C. Infrared (IR) spectra were obtained on a Thermo Scientific Nicolet iS5 spectrometer with an iD5 ATR tip (neat) and are reported in terms of frequency of absorption (cm⁻¹). Analytical thin-layer chromatography (TLC) was performed using Silica Gel 60 F₂₅₄ precoated plates (0.25 mm thickness). Visualization was accomplished by irradiation with a UV lamp and/or staining with KMnO₄, ceric ammonium molybdate (CAM), or *p*-anisaldehyde (PAA) solutions. Flash chromatography was performed using Silica Gel 60 (170-400 mesh) from Fisher Scientific. Melting points (m.p.) were obtained using a Mel-Temp melting point apparatus and are uncorrected. Optical rotations were measured on a Rudolph Research Analytical Autopol IV Automatic Polarimeter. High resolution mass spectrometry was performed by the University

of California, Irvine Mass Spectrometry Center. Bis(cyclooctadiene)nickel Ni(cod)₂ complex was purchased from Strem, stored in a glovebox under an atmosphere of N₂, and used as received. All other reagents were purchased commercially and used as received.

GENERAL METHODS FOR STARTING MATERIAL SYNTHESIS

The general methods for starting material synthesis will be used throughout the rest of the SI. In each instance a general method is used, it is specified by letter (A, B, etc.) and the exact amounts of reagents used for each reaction are listed for the specific compounds synthesized.

METHOD A: SONOGASHIRA COUPLING, CARBAMATE INSTALLATION



SONOGASHIRA COUPLING:

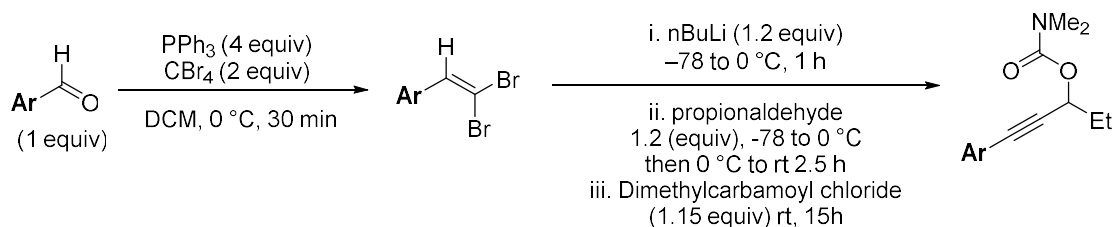
The products were prepared according to a modified procedure reported by Lautens.¹⁰ To 100 mL round bottom flask equipped with a stir bar was added PdCl₂(PPh₃)₂ (5 mol%), CuI (10 mol%), 1-butyne-3-ol (1.1 equiv), aryl iodide (1.0 equiv), and triethyl amine (25 mL). The reaction was stirred at room temperature for 4 hours. The reaction mixture was then filtered through silica to remove palladium and copper, and the silica was washed with diethyl ether. The solvents were removed under reduced pressure to provide the crude product as a dark yellow-brown oil. The crude mixtures were carried forward to carbamate installation without further purification.

CARBAMATE INSTALLATION:

¹⁰ Panteleev, J.; Huang, R. Y.; Lui, E. K. J.; Lautens, M. *Org. Lett.* **2011**, *13*, 5314

To a 100 mL round bottom flask was added NaH (1.1 equiv) and THF (30 mL). The suspension was cooled to 0 °C in an ice bath and a solution of the alcohol (1 M in THF) was added dropwise over 15 minutes. The ice bath was then removed and the mixture was allowed to warm to room temperature before dimethylcarbamoyl chloride (1.15 equiv) was added. The reaction was stirred overnight at room temperature for 15 hours after which the mixture was quenched with saturated aqueous ammonium chloride, extracted with ethyl ether, dried over MgSO₄, and concentrated under reduced pressure. The product was purified by flash column chromatography.

METHOD B: COREY-FUCHS, 1, 2 ADDITION, CARBAMATE INSTALLATION



COREY-FUCHS:

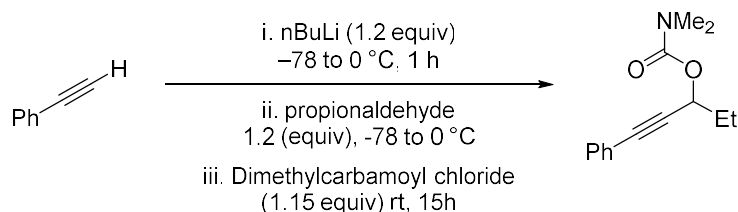
The product was prepared according to a modified procedure reported by Hoppe.¹¹ To a 250 mL round bottom flask was added PPh₃ (40 mmol, 4 equiv) and dichloromethane (35 mL) and was cooled to 0 °C, after which CBr₄ (20 mmol, 2 equiv) was added as a solution in dichloromethane (15 mL). The aryl aldehyde (10 mmol, 1 equiv) was then added as a solution in dichloromethane (10 mL) dropwise over the course of 10 minutes. The reaction was stirred for 30 min at 0 °C before the solvent was removed and the crude yellow oil was purified by column chromatography.

1, 2 ADDITION, CARBAMATE INSTALLATION:

¹¹ Oestreich, M; Fröhlich, R.; Hoppe, D. *J. Org. Chem.* **1999**, *64*, 8616

To a 250 mL round bottom flask was added *gem*-dibromo styrene and THF (50 mL). The solution was cooled to $-78\text{ }^{\circ}\text{C}$ in a dry ice and acetone bath after which *n*-BuLi (1.2 equiv) was added dropwise over the course for 20 min. The bath was then removed and the reaction was allowed to warm to room temperature and stirred for 30 minutes before it was cooled back down to $-78\text{ }^{\circ}\text{C}$, and a solution of propionaldehyde (1M in THF) was added dropwise over the course of 10 minutes. The reaction was allowed to warm to room temperature and stirred for 4 hours before being quenched with dimethylcarbamoyl chloride (1.15 equiv). The resulting mixture was stirred overnight at room temperature for 15 hours before being quenched with saturated aqueous ammonium chloride and extracted with ethyl ether, dried over MgSO_4 , and concentrated under reduced pressure. The product was purified by flash column chromatography.

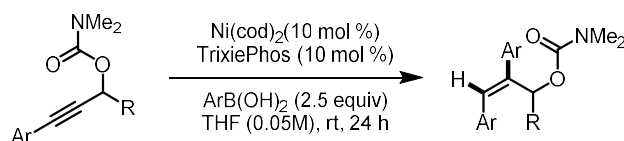
METHOD C: 1, 2 ADDITION, CARBAMATE INSTALLATION



To a 250 mL round bottom flask was added phenyl acetylene (1 equiv) and THF (50 mL). The solution was cooled to $-78\text{ }^{\circ}\text{C}$ in a dry ice and acetone bath after which *n*-BuLi (1.2 equiv) was added dropwise over the course for 20 min. The bath was then removed and the reaction was allowed to warm to room temperature and stirred for 30 minutes before it was cooled back down to $-78\text{ }^{\circ}\text{C}$, and a solution of propionaldehyde (1M in THF) was added dropwise over the course of 10 minutes. The reaction was allowed to warm to room temperature and stirred for 4 hours before being quenched with dimethylcarbamoyl chloride (1.15 equiv). The resulting mixture was stirred overnight at room temperature for 15 hours before being quenched with saturated aqueous

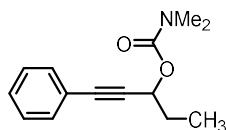
ammonium chloride and extracted with ethyl ether, dried over MgSO₄, and concentrated under reduced pressure. The product was purified by flash column chromatography.

METHOD D: NICKEL CATALYZED HYDROARYLATION



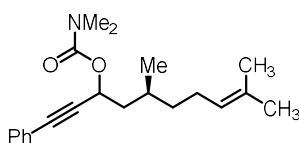
In a glove box, a flame dried 7 mL dram vial was charged with Ni(cod)₂ (5.5 mg, 0.02 mmol, 0.10 equiv), TrixiePhos (8.0 mg, 0.02 mg, 0.10 equiv), arylboronic acid (0.5 mmol, 2.5 equiv), and the alkyne (0.2 mmol, 1.0 equiv) and dissolved in THF (4 mL). The reaction was then stirred for 24 hours at 24 °C or 85 °C depending on the substrate and arylboronic acid used. The crude reaction mixture was then filtered through a pad of silica and eluted with Et₂O to remove the catalyst and then concentrated under reduced pressure. The crude mixture was purified by flash column chromatography to isolate pure product.

CHARACTERIZATION FOR STARTING MATERIALS

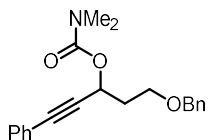


2.49. The product was prepared according to general method C using propionaldehyde (1.9 mL, 26.8 mmol), phenylacetylene (2.9 mL, 26.4 mmol) *n*-BuLi (11.4 mL, 28.5 mmol) and dimethylcarbamoyl chloride (2.5 mL, 27.5 mmol). The product was purified by flash column chromatography (20% Et₂O/pentane) to afford the title compound as a colorless oil (4.4 g, 19 mmol, 72% yield). **TLC** R_f = 0.4 (12% EtOA/hexane); **¹H NMR** (500 MHz, CDCl₃) δ 7.49–7.41

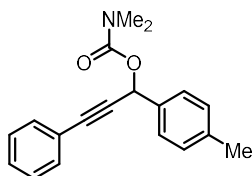
(m, 2H), 7.33–7.28 (m, 3H), 5.5 (t, $J = 6.48$ Hz, 1 H), 2.94 (s, 6H), 1.89 (quintet $J = 7.5$ Hz, 2H) 1.08 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 156.0, 132.2, 128.7, 128.5, 122.9, 87.6, 85.2, 66.9, 28.9, 9.8; IR (neat) 2970, 2935, 1702, 1489, 1128, 1392 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{14}\text{H}_{17}\text{NO}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 254.1157, found 254.1151



2.50. The product was prepared according to general method C using citronellal (2.0 mL, 11.0 mmol), phenylacetylene (1.1 mL, 10 mmol,) *n*-BuLi (4.4 mL, 11 mmol) and dimethylcarbamoyl chloride (1.0 ml, 11 mmol). The product was purified by flash column chromatography (20% Et_2O /pentane) to afford the title compound as a colorless oil (3.3 g, 10 mmol, 92% yield). TLC $R_f = 0.4$ (20% Et_2O /pentane); ^1H NMR (500 MHz, CDCl_3) δ 7.46–7.40 (m, 2H), 7.31–7.27 (m, 3H), 5.65–5.58 (m, 1H), 5.14–5.07 (m, 1H), 2.94 (s, 6H) 2.08–1.84, (m, 3H), 1.83–1.65 (m, 5H), 1.60 (s, 3H), 1.47–1.37 (m, 1H), 1.29–1.18 (m, 1H), 1.00–0.96 (m, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 156.1, 156.0, 132.21, 132.19, 131.66, 131.65, 128.68, 128.66, 128.49, 128.48, 124.91, 124.90, 123.0, 88.3, 88.0, 85.3, 85.0, 64.9, 64.4, 42.8, 42.5, 37.4, 37.2, 29.7, 29.4, 26.0, 25.66, 25.62, 19.97, 19.89, 18.0; IR (neat) 2955, 2924, 1705, 1489, 1392, 1178 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{21}\text{H}_{29}\text{NO}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 350.2096, found 350.2083

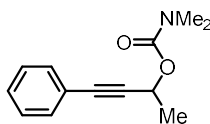


2.51. The product was prepared according to general method C using phenyl acetylene (0.45 mL, 4.1 mmol), *n*-BuLi (1.9 mL, 4.6 mmol), 3-(benzyloxy)propanal (660 mg, 4.0 mmol), and dimethyl carbamoyl chloride (1.1 mL, 11 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a colorless oil (1.1 g, 3.4 mmol, 85% yield). **TLC** R_f = 0.3 (15% EtOAc/hexanes); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 7.43–7.38 (m, 2H), 7.36–7.23 (m, 8H), 5.73 (t, J = 6.7 Hz, 1H), 4.53 (s, 2H), 3.74–3.62 (m 2H), 2.93 (s, 3H), 2.89 (s, 3H), 2.28–2.12 (m, 2H); **$^{13}\text{C NMR}$** (125 MHz, CDCl_3) δ 155.8, 138.6, 132.2, 128.8, 128.7, 128.5, 128.0, 127.8, 122.8, 87.5, 85.4, 73.3, 66.4, 63.4, 35.9; **IR** (neat) 2931, 2863, 1699, 1490, 1395, 1184 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{21}\text{H}_{23}\text{NO}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 360.1576, found 360.1564

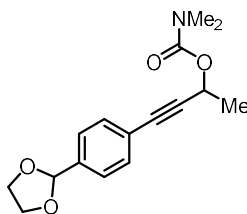


2.52. The product was prepared according to general method C using phenyl acetylene (1.1 mL 10 mmol) *n*-BuLi (4.8 mL, 12 mmol), *p*-Tolualdehyde (1.2 mL, 2.0 mmol), and dimethyl carbamoyl chloride (0.41mL, 4.4 mmol). The product was purified by flash column chromatography (20% Et_2O /pentanes) to afford the title compound as a colorless oil (1.14 g, 3.38mmol, 85% yield). **TLC** R_f = 0.3 (20% Et_2O /pentane); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 7.52–7.46, (m, 4H), 7.34–7.27 (m, 3H), 7.22–7.18 (m, 2H), 6.63 (s, 1H), 2.94 (s, 6H), 2.36 (s, 3H); **$^{13}\text{C NMR}$** (125 MHz, CDCl_3) δ 155.8, 138.9, 135.5, 132.2, 129.6, 128.9, 128.5, 127.9, 122.8, 87.0, 86.8, 67.2, 21.6; **IR** (neat)

2929, 1697, 1490, 1394, 1175 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{19}\text{H}_{19}\text{NO}_2\text{Na}$ ($\text{M} + \text{Na}$)⁺ 316.1313, found 316.1313

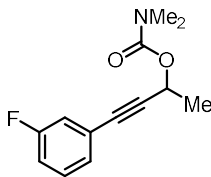


2.53. The product was prepared according to general method A using $\text{PdCl}_2(\text{PPh}_3)_2$ (90 mg), CuI (45 mg) 1-butyne-3-ol (0.54 mL, 7.0 mmol), aryl iodide (0.78 mL, 7.0 mmol), triethyl amine (30 mL), dimethylcarbamoyl chloride (0.71 mL, 7.7 mmol), NaH (200 mg, 8.4 mmol), THF (25 mL) The product was purified by flash column chromatography (20% Et_2O /pentanes) to afford the title compound as a colorless oil (1.3 g, 6.0 mmol, 86% yield). **TLC** $R_f = 0.4$ (12% EtOAc /hexane); **^1H NMR** (500 MHz, CDCl_3) δ 7.46–7.42 (m, 2H), 7.32–7.25 (m, 3H), 5.65 (q, $J = 6.6$ Hz, 1H), 2.93 (s, 6H), 1.59 (d, $J = 6.8$ Hz, 3H); **^{13}C NMR** (125 MHz, CDCl_3) δ 155.8, 132.1, 128.7, 128.5, 122.8, 88.6, 84.4, 61.9, 20.2; **IR** (neat) 2986, 2935, 1699, 1489, 1392, 1177 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{13}\text{H}_{15}\text{NO}_2\text{Na}$ ($\text{M} + \text{Na}$)⁺ 240.1001, found 240.0989

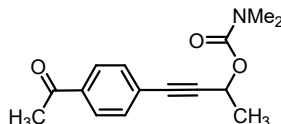


2.54 The product was prepared according to general method A using $\text{PdCl}_2(\text{PPh}_3)_2$ (350 mg, 0.5 mmol), CuI (190 mg, 1.0 mmol) 1-butyne-3-ol (0.83 mL, 11 mmol), aryl bromide (2.3g, 10 mmol), triethyl amine (30 mL), dimethylcarbamoyl chloride (1.0 mL, 11 mmol), NaH (290 mg, 12 mmol), THF (25 mL) The product was purified by flash column chromatography (12% EtOAc /hexanes) to afford the title compound as a pale yellow oil (2.3 g, 8.0 mmol, 80% yield). **TLC** $R_f = 0.2$ (12%

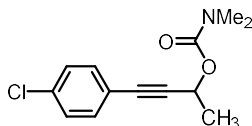
EtOAc/Hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.47–7.39 (m, 4H), 5.79 (s, 1H), 5.64 (q, *J* = 6.7 Hz, 1H), 4.14–3.99 (m, 4H), 2.94 (s, 6H), 1.58 (d, 3, *J* = 6.7 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 155.9, 138.4, 132.2, 126.7, 123.7, 103.6, 89.2, 84.1, 65.6, 62.0, 22.3; **IR** (neat) 2985, 2886, 1699, 1391, 1176, 1079 cm⁻¹; **HRMS** (TOF MS ES+) *m/z* calcd for C₁₆H₁₉NO₄Na (M + Na)⁺ 312.1212, found 312.1197



2.55 The product was prepared according to general method A using PdCl₂(PPh₃)₂ (250 mg, 0.35 mmol), CuI (130 mg, 0.70 mmol) 1-butyne-3-ol (0.58 mL, 7.7 mmol), aryl iodide (0.82 mL, 7.0 mmol), triethyl amine (30 mL), dimethylcarbamoyl chloride (0.71 mL, 7.7 mmol), NaH (200 mg, 8.2 mmol), THF (25 mL) The product was purified by flash column chromatography (12% EtOAc/hexanes) to afford the title compound as a pale yellow oil (1.9 g, 8.2 mmol, 82% yield). **TLC** R_f = 0.4 (12% EtOAc/Hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.29–7.20 (m, 2H), 7.16–7.12 (m, 1H), 7.05–6.98 (m, 1H), 5.63 (q, *J* = 6.8 Hz, 1H), 2.94 (s, 6H), 1.58 (d, *J* = 6.8 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 162.5 (d, *J* = 246.0 Hz) 155.8, 130.1 (d, *J* = 8.8 Hz) 128.1 (d, *J* = 3.2 Hz) 124.7 (d, *J* = 9.7 Hz), 119.0 (d, *J* = 22.7 Hz), 116.1 (d, *J* = 21.4 Hz), 89.7, 83.2, 61.9, 22.2; **IR** (neat) 2936, 1701, 1579, 1486, 1392, 1170 cm⁻¹; **HRMS** (TOF MS ES+) *m/z* calcd for C₁₃H₁₄FNO₂Na (M + Na)⁺ 258.0906, found 258.0905

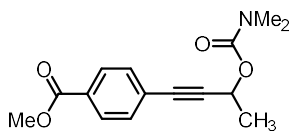


2.56. The product was prepared according to general method A using $\text{PdCl}_2(\text{PPh}_3)_2$ (180 mg, 0.25 mmol), CuI (95 mg, 0.50 mmol) 1-butyne-3-ol (0.43 mL, 5.5 mmol), aryl iodide (1.2 g, 5.0 mmol), triethyl amine (30 mL), dimethylcarbamoyl chloride (0.51 mL, 5.5 mmol), NaH (140 mg, 6 mmol), THF (25 mL) The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (0.86 g, 3.3 mmol, 66% yield). **TLC** R_f = 0.4 (12% EtOAc/Hexanes); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 7.89 (d, J = 8.4 Hz, 2H), 7.52 (d, J = 8.4 Hz, 2H), 5.65 (q, J = 6.7 Hz, 1H), 2.95 (s, 6H), 2.60 (s, 3H), 1.60 (d, J = 6.9 Hz, 3H); **$^{13}\text{C NMR}$** (125 MHz, CDCl_3) δ 197.7, 155.8, 136.7, 132.3, 128.4, 127.8, 92.0, 83.6, 61.8, 27.0, 22.1; **IR** (neat) 2987, 2936, 1701, 1682, 1392, 1260, 1176 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{15}\text{H}_{17}\text{NO}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 282.1106, found 282.1102

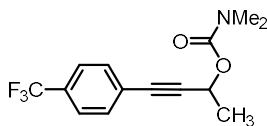


2.57. The product was prepared according to general method A using $\text{PdCl}_2(\text{PPh}_3)_2$ (180 mg, 0.25 mmol), CuI (95 mg, 0.50 mmol) 1-butyne-3-ol (0.43 mL, 5.5 mmol), aryl iodide (1.2 g, 5.0 mmol), triethyl amine (30 mL), dimethylcarbamoyl chloride (0.51 mL, 5.5 mmol), NaH (140 mg, 6 mmol), THF (25 mL) The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (0.89 g, 3.6 mmol, 71% yield). **TLC** R_f = 0.4 (12% EtOAc/Hexanes); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 7.37 (d, J = 8.6 Hz, 2H), 7.27 (d, J = 8.4 Hz, 2H), 5.62 (q, J = 6.7 Hz, 1H), 2.94 (s, 6H), 1.58 (d, J = 6.6 Hz, 3H); **$^{13}\text{C NMR}$** (125 MHz, CDCl_3) δ 155.8, 134.8, 133.4, 128.9, 121.4, 89.8, 83.3, 61.9, 22.2; **IR** (neat) 2987,

2934, 1701, 1488, 1391, 1177 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{13}\text{H}_{14}\text{ClNO}_2\text{Na}$ ($\text{M} + \text{Na}$)⁺ 274.0611, found 274.0617

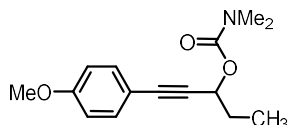


2.58. The product was prepared according to general method A using $\text{PdCl}_2(\text{PPh}_3)_2$ (180 mg, 0.25 mmol), CuI (95 mg, 0.50 mmol) 1-butyne-3-ol (0.43 mL, 5.5 mmol), aryl iodide (1.3 g, 5.0 mmol), triethyl amine (30 mL), dimethylcarbamoyl chloride (0.51 mL, 5.5 mmol), NaH (140 mg, 6 mmol), THF (25 mL) The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (1.0 g, 3.8 mmol, 75% yield). **TLC** R_f = 0.3 (15% EtOAc/Hexanes); **^1H NMR** (500 MHz, CDCl_3) δ 7.97 (d, J = 8.6 Hz, 2H), 7.50 (d, J 8.6 = Hz, 2H), 5.64 (q, J = 6.7 Hz, 1H), 3.92 (s, 3H), 2.95 (s, 6H) 1.59 (d, J = 6.7 Hz, 3H); **^{13}C NMR** (125 MHz, CDCl_3) δ 166.8, 155.8, 132.1, 130.0, 129.7, 127.6, 91.7, 83.7, 61.9, 52.6, 22.1; **IR** (neat) 2988, 2936, 1701, 1393, 1272, 1174, 1085 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{15}\text{H}_{17}\text{NO}_4\text{Na}$ ($\text{M} + \text{Na}$)⁺298.1055, found 298.1047

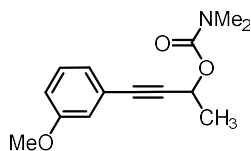


2.59. The product was prepared according to general method A using $\text{PdCl}_2(\text{PPh}_3)_2$ (180 mg, 0.25 mmol), CuI (95 mg, 0.50 mmol) 1-butyne-3-ol (0.43 mL, 5.5 mmol), aryl iodide (0.73 mL, 5.0 mmol), triethyl amine (30 mL), dimethylcarbamoyl chloride (0.51 mL, 5.5 mmol), NaH (140 mg, 6 mmol), THF (25 mL) The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (1.0 g, 3.7 mmol, 73% yield). **TLC** R_f = 0.4 (15% EtOAc/Hexanes); **^1H NMR** (500 MHz, CDCl_3) δ 7.58–7.52 (m, 4H), 5.64 (q,

J 7.0 = Hz, 1H), 2.95 (s, 6H), 1.59 (d, J = 6.7 Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 155.8, 132.4, 130.5 (q, J = 32.8 Hz), 126.7 (q, J = 1.8 Hz), 124.1 (q, J = 271.9 Hz) 125.5 (q, J = 3.7 Hz), 91.2, 83.1, 61.8, 22.0; IR (neat) 2938, 1703, 1394, 1319, 1165, 1064 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{14}\text{H}_{14}\text{F}_3\text{NO}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 308.0874, found 308.0863



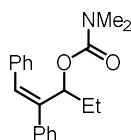
2.60. The product was prepared according to general method B using *p*-methoxy benzaldehyde (1.2 mL 10 mmol) CBr_4 (6.6 g, 20 mmol), PPh_3 (10.5 g 40.0 mmol), *n*-BuLi (8.4 mL, 21 mmol) propionaldehyde (0.72 mL, 10 mmol) and dimethyl carbamoyl chloride (1.0 mL, 11 mmol). The product was purified by flash column chromatography (12 % EtOAc/Hex) to afford the title compound as a pale yellow oil (1.2 g, 4.5 mmol, 45% yield). TLC R_f = 0.3 (12% EtOAc/Hexanes); ^1H NMR (500 MHz, CDCl_3) δ 7.39 (d, J = 8.8 Hz, 2H), 6.82 (d, J = 8.7, Hz, 2H), 5.50 (t, J = 6.5 Hz, 1H), 3.80 (s, 3H), 2.94 (s, 6H), 1.87 (quintet J = 7.2 Hz, 2H), 1.07 (t, J = 7.3 Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 160.0, 156.1, 133.7, 115.1, 114.1, 86.3, 85.1, 67.0, 55.6, 29.0, 9.8; IR (neat) 2962, 2930, 1698, 1509, 1248, 1176 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{15}\text{H}_{19}\text{NO}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 284.1263, found 284.1265



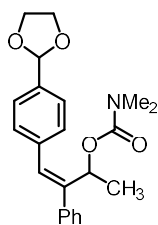
2.61. The product was prepared according to general method A using $\text{PdCl}_2(\text{PPh}_3)_2$ (180 mg, 0.25 mmol), CuI (95 mg, 0.50 mmol) 1-butyne-3-ol (0.43 mL, 5.5 mmol), aryl iodide (0.73 mL, 5.0 mmol), triethyl amine (30 mL), dimethylcarbamoyl chloride (0.51 mL, 5.5 mmol), NaH (140 mg,

6 mmol), THF (25 mL) The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (1.0 g, 3.7 mmol, 73% yield). **TLC** R_f = 0.3 (12% EtOAc/Hexanes); **^1H NMR** (500 MHz, CDCl_3) δ 7.20 (t, J = 7.7 Hz, 1H), 7.06–7.02 (m, 1H), 6.99–6.95 (m, 1H), 6.89–6.84 (m, 1H), 5.64 (q, J = 6.9 Hz, 1H), 3.79 (s, 3H), 2.94 (s, 6H), 1.58 (d, J = 6.7 Hz, 3H); **^{13}C NMR** (125 MHz, CDCl_3) δ 159.5, 155.9, 129.6, 124.7, 123.8, 116.9, 115.5, 88.5, 84.4, 62.0, 55.6, 22.3; **IR** (neat) 2936, 1700, 1574, 1392, 1173 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{13}\text{H}_{17}\text{NO}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 270.1106, found 270.1097

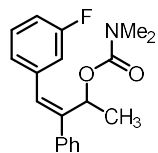
CHARACTERIZATION OF PRODUCTS



2.21 The product was prepared according to general method D using alkyne **2.49** (46.3 mg, 0.2 mmol), phenyl boronic acid (60 mg, 0.5 mmol), $\text{Ni}(\text{cod})_2$ (5.5 mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et_2O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (43.3 mg, 70% yield). **TLC** R_f = 0.4 (15% EtOAc/hexanes); **^1H NMR** (500 MHz, CDCl_3) δ 7.51–7.46 (m, 2H), 7.43–7.23 (m, 8H), 6.71 (s, 1H), 5.84 (t, J = 6.85 Hz, 1H), 2.88 (s, 3H), 2.7 (s, 3H), 1.85 (m, 1H), 1.65 (m, 3H), 0.82 (t, J = 7.83 Hz, 3H); **^{13}C NMR** (125 MHz, CDCl_3) 156.3, 141.9, 141.4, 137.3, 132.6, 129.2, 128.9, 128.6, 128.2, 127.5, 127.3, 75.6, 27.5, 10.5; **IR** (neat) 2968, 2934, 1699, 1598, 1443 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{20}\text{H}_{23}\text{NO}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 332.1627, found 332.1611

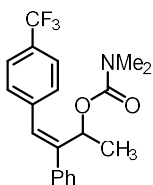


2.26. The product was prepared according to general method D using alkyne **2.54** (57.9 mg, 0.2 mmol), phenyl boronic acid (61 mg, 0.5 mmol), Ni(cod)₂ (5.5 mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (41.9 mg, 57% yield). **TLC** R_f = 0.3 (25% EtOAc/hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.51–7.45 (m, 4H), 7.41–7.28 (m, 5H), 6.64 (s, 1H), 6.02 (q, *J* = 6.9 Hz, 1H), 5.83 (s, 1H), 4.16–4.01 (m, 4H), 2.86 (s, 3H), 2.70 (s, 3H), 1.39 (d, *J* = 6.8 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) 156.1, 143.4, 141.0, 138.1, 137.0, 131.0, 129.2, 128.9, 128.2, 127.6, 126.8, 103.9, 70.6, 65.7, 65.6, 30.1, 20.8; **IR** (neat) 2926, 1698, 1442, 1390, 1183, 1080 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₂H₂₅NO₄Na (M + Na)⁺ 390.1681, found 390.1668

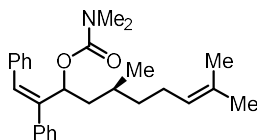


2.27 The product was prepared according to general method D using alkyne **2.55** (47.1 mg, 0.2 mmol), phenyl boronic acid (61 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (54.5 mg, 87% yield). **TLC** R_f = 0.3 (25% EtOAc/hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.48–7.44 (m, 2H), 7.39–7.29 (m, 4H), 7.17–7.13 (m, 1H), 7.10–7.05

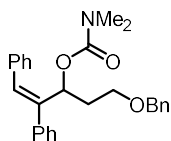
(m, 1H), 6.93 (dt, $J = 8.4$, $J = 2.4$ Hz, 1H), 6.58 (s, 1H), 5.99 (q, $J = 6.8$ Hz, 1H), 2.87 (s, 3H), 2.71 (s, 3H), 1.41 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 162.9 (d, $J = 245.5$ Hz) 156.0, 144.2, 140.8, 139.4 (d, $J = 7.9$ Hz), 130.1 (d, $J = 8.3$ Hz), 130.0 (d, $J = 1.8$ Hz), 128.9, 128.3, 127.7, 124.9, (d, $J = 3.2$ Hz), 116.1 (d, $J = 21.7$ Hz), 114.3 (d, $J = 21.3$ Hz), 70.4, 20.8; **IR** (neat) 2985, 2933, 1698, 1489, 1391, 1180 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{19}\text{H}_{20}\text{FNO}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 336.1376, found 336.1362



2.29. The product was prepared according to general method D using alkyne **2.59** (57.1 mg, 0.2 mmol), phenyl boronic acid (61 mg, 0.5 mmol), $\text{Ni}(\text{cod})_2$ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et_2O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (60.3mg, 83% yield). **TLC** $R_f = 0.4$ (10% EtOAc /hexanes); **^1H NMR** (500 MHz, CDCl_3) δ 7.65–7.61 (m, 2H), 7.51–7.46 (m, 4H), 7.40–7.31 (m, 3H), 6.63 (s, 1H), 5.95 (q, $J = 6.7$ Hz, 1H), 2.87 (s, 3H), 2.68 (s, 3H), 1.40 (d, $J = 6.7$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 156.0, 144.9, 140.9 (q, $J = 1.4$ Hz), 140.6, 129.7, 129.4, 129.3 (q, $J = 132.4$ Hz), 128.8, 128.4, 127.9, 125.6 (q, $J = 3.7$ Hz), 124.5 (q, $J = 271.9$), 70.4, 20.8; **IR** (neat) 2980, 2934, 1700, 1615, 1393, 1323, 1121 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{20}\text{H}_{20}\text{F}_3\text{NO}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 386.1344, found 386.1335

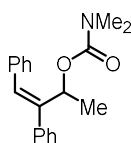


2.31. The product was prepared according to general method D using alkyne **2.50** (65.5 mg, 0.2 mmol), phenyl boronic acid (61 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (53.5 mg, 66% yield). **TLC** R_f = 0.6 (15% EtOAc/hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.50–7.44 (m, 2H), 7.44–7.40 (m, 2H), 7.38–7.32 (m, 4H), 7.31–7.27 (m, 1H), 7.27–7.22 (m, 1H), major diastereomer 6.69 (s, 0.52H), minor diastereomer 6.64 (s, 0.46H), 6.06–5.99 (m, 1H), 4.99–4.91 (m, 1H), major diastereomer 2.92– 2.80 (m, 3H) minor diastereomer 2.77-2.58 (m, 3H), 1.97–1.72 (m, 2H), 1.69–1.59 (m, 4H), 1.54–1.49 (m, 3H), 1.49–1.33 (m, 2H), 1.22–0.90 (m, 2H), minor diastereomer 0.73 (d, *J* = 6.6 Hz, 1.3H) major diastereomer 0.68 (d, *J* = 6.7 Hz, 1.59H); **¹³C NMR** (125 MHz, CDCl₃) δ 156.3, 156.2, 143.1, 142.4, 141.28, 141.24, 137.4, 137.3, 132.3, 131.33, 131.28, 131.26, 129.10, 129.09, 128.90, 128.89, 128.6, 128.5, 128.19, 128.16, 127.48, 127.45, 127.3, 127.2, 125.03, 125.01, 73.2, 72.7, 41.9, 41.3, 37.5, 37.0, 29.6, 29.4, 26.01, 26.00, 25.7, 25.4, 19.9, 19.4, 17.9; **IR** (neat) 3022, 2959, 1700, 1493, 1392, 1186 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₇H₃₅NO₂Na (M + Na)⁺ 428.2566, found 428.2574

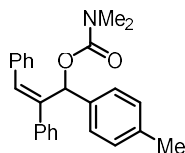


2.30. The product was prepared according to general method D using alkyne **2.51** (67.5 mg, 0.2 mmol), phenyl boronic acid (61 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and

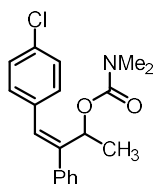
concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (58.2 mg, 70% yield). **TLC** R_f = 0.6 (20% EtOAc/hexanes); **^1H NMR** (500 MHz, CDCl_3) δ 7.48–7.40 (m, 4H), 7.36–7.19 (m, 11H), 6.67 (s, 1H), 6.16 (dd, $J=8.9$ Hz, $J=5.3$ Hz 1H), 4.35 (s, 2H), 3.50–3.40 (m, 2H), 2.84 (s, 3H), 2.64 (s, 3H), 2.22–2.12 (m, 1H), 2.06, 1.97 (m, 1H); **^{13}C NMR** (125 MHz, CDCl_3) δ 156.1, 141.9, 141.2, 138.7, 137.1, 132.3, 129.3, 128.9, 128.6, 128.5, 128.2, 127.9, 127.7, 127.5, 127.3, 73.1, 71.6, 67.0, 35.0; **IR** (neat) 3056, 2927, 1699, 1494, 1391, 1179 cm^{-1} ; **HRMS** (TOF MS ES^+) m/z calcd for $\text{C}_{27}\text{H}_{29}\text{NO}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 438.2045, found 438.2048



2.47 The product was prepared according to general method D using alkyne **2.53** (43.5 mg, 0.2 mmol), phenyl boronic acid (61 mg, 0.5 mmol), $\text{Ni}(\text{cod})_2$ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et_2O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (41.9 mg, 71% yield). **TLC** R_f = 0.4 (15% EtOAc/hexanes); **^1H NMR** (500 MHz, CDCl_3) δ 7.51–7.47 (m, 2H), 7.40–7.24 (m, 7H), 6.65 (s, 1H), 6.05 (q, $J=6.6$ Hz, 1H), 2.87 (s, 3H), 2.72 (s, 3H), 1.40 (d, $J=6.7$ Hz, 3H); **^{13}C NMR** (125 MHz, CDCl_3) δ 156.1, 142.9, 141.2, 137.1, 131.5, 129.2, 129.0, 128.7, 128.2, 127.5, 127.4, 70.5, 20.8; **IR** (neat) 3022, 2932, 1698, 1491, 1392, 1185 cm^{-1} ; **HRMS** (TOF MS ES^+) m/z calcd for $\text{C}_{19}\text{H}_{21}\text{NO}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 318.1470, found 318.1469

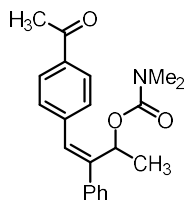


2.32 The product was prepared according to general method D using alkyne **2.52** (58.7 mg, 0.2 mmol), phenyl boronic acid (61 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (60.2 mg, 81% yield). **TLC** *R_f* = 0.4 (15% EtOAc/hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.52 (d, *J* = 8.1 Hz, 2H), 7.45–7.38 (m, 4H), 7.34–7.28 (m, 6H), 7.22 (s, 1H), 7.19 (d, *J* = 7.7 Hz, 2H), 7.05, (s, 1H), 2.92 (s, 3H), 2.68 (s, 3H), 2.40 (s, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 156.1, 140.5, 140.4, 137.4, 137.03, 136.94, 132.4, 129.4, 129.2, 128.78, 128.76, 128.1, 127.6, 127.5, 126.7, 74.6, 21.5; **IR** (neat) 3023, 2923, 1703, 1492, 1389, 1173 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₅H₂₅NO₂Na (M + Na)⁺ 394.1783, found 394.1784

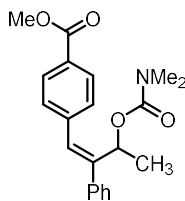


2.28 The product was prepared according to general method D using alkyne **2.57** (50.3 mg, 0.2 mmol), phenyl boronic acid (61 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (52.1 mg, 79% yield). **TLC** *R_f* = 0.5 (15% EtOAc/hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.48–7.44 (m, 2H), 7.38–7.28 (m, 7H), 6.57 (s, 1H), 5.99 (q, *J* = 6.7 Hz, 1H) 2.87 (s, 3H), 2.72 (s, 3H), 1.38 (d, *J* = 6.7 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 156.0, 143.7, 140.8, 135.6, 133.3, 130.5, 130.1, 128.85, 128.82, 128.3, 127.7, 70.3, 20.7; **IR** (neat) 2977,

2930, 1698, 1490, 1391, 1183 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{19}\text{H}_{20}\text{ClNO}_2\text{Na}$ ($\text{M} + \text{Na}$)⁺ 352.1080, found 352.1077

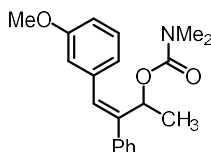


2.24. The product was prepared according to general method D using alkyne **2.56** (51.9 mg, 0.2 mmol), phenyl boronic acid (61 mg, 0.5 mmol), $\text{Ni}(\text{cod})_2$ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et_2O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (50.6mg, 75% yield). **TLC** R_f = 0.3 (25% EtOAc /hexanes); **^1H NMR** (500 MHz, CDCl_3) δ 8.00–7.95 (m, 2H), 7.51–7.45 (m, 4H), 7.40–7.31 (m, 3H), 6.66 (s, 1H), 6.01 (q, J = 6.7 Hz, 1H), 2.87 (s, 3H), 2.72 (s, 3H), 2.61 (s, 3H), 1.40 (d, J = 6.7 Hz, 3H); **^{13}C NMR** (125 MHz, CDCl_3) δ 156.0, 144.8, 142.1, 140.6, 135.9, 130.3, 129.4, 128.83, 128.76, 128.3, 127.8, 70.3, 27.0, 20.7; **IR** (neat) 2979, 2931, 1699, 1682, 1267, 1182 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{21}\text{H}_{23}\text{NO}_3\text{Na}$ ($\text{M} + \text{Na}$)⁺ 360.1576, found 360.1572

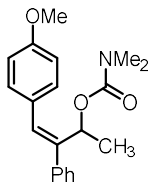


2.25 The product was prepared according to general method D using alkyne **2.58** (55.1 mg, 0.2 mmol), phenyl boronic acid (61 mg, 0.5 mmol), $\text{Ni}(\text{cod})_2$ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et_2O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title

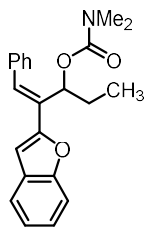
compound as a pale yellow oil (50.9mg, 72% yield. **TLC** R_f = 0.3 (15% EtOAc/hexanes); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 8.06–8.03 (m, 2H), 7.50–7.46 (m, 2H), 7.46–7.42 (m, 2H), 7.39–7.31 (M, 3H), 6.65 (s, 1H), 5.99 (q, J = 6.7 Hz, 1H), 3.92 (s, 3H), 2.86 (s, 3H), 2.69 (s, 3H), 1.43 (d, J = 6.7 Hz, 3H); **$^{13}\text{C NMR}$** (125 MHz, CDCl_3) δ 167.2, 156.0, 144.7, 142.0, 140.7, 130.3, 129.9, 129.2, 128.9, 128.8, 128.3, 127.8, 70.4, 52.4, 20.7; **IR** (neat) 2929, 1718, 1698, 1391, 1275, 1178 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{21}\text{H}_{23}\text{NO}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 376.1525, found 376.1522



2.22 The product was prepared according to general method D using alkyne **2.61** (49.5 mg, 0.2 mmol), phenyl boronic acid (61 mg, 0.5 mmol), $\text{Ni}(\text{cod})_2$ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et_2O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (35.1 mg, 54% yield). **TLC** R_f = 0.3 (12% EtOAc/hexanes); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 7.50–7.46 (m, 2H), 7.39–7.26 (m, 4H), 6.98–6.94 (m, 1H), 6.93–6.90 (m, 1H), 6.85–6.80 (m, 1H), 6.63 (s, 1H), 6.06 (q, J = 6.4 Hz 1H), 3.83 (s, 3H), 2.87 (s, 3H) 2.74 (s, 3H), 1.40 (d, J = 6.6 Hz 3H); **$^{13}\text{C NMR}$** (125 MHz, CDCl_3) δ 159.8, 143.1, 141.1, 138.5, 131.5, 129.7, 128.9, 128.2, 127.6, 121.6, 114.4, 113.4, 70.6, 55.5, 20.8; **IR** (neat) 2928, 1697, 1488, 1391, 1268, 1182 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{20}\text{H}_{23}\text{NO}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 348.1576, found 348.1577

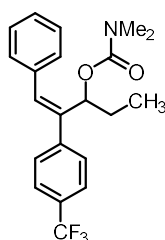


2.23 The product was prepared according to general method D using alkyne **2.61** (49.5 mg, 0.2 mmol), phenyl boronic acid (61 mg, 0.5 mmol), Ni(cod)₂ (5.5 mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (32.5 mg, 50% yield). **TLC** *R_f* = 0.3 (12% EtOAc/hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.50–7.46 (m, 2H), 7.39–7.26 (m, 5H), 6.94–6.88 (m, 2H), 6.65 (s, 1H), 5.89 (t, *J* = 7.34 Hz, 1H), 3.82 (s, 3H), 2.89 (s, 3H), 2.78 (s, 3H), 1.84 (m, 1H), 1.64 (m, 1H), 0.82 (t, *J* = 7.58 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 158.9, 141.5, 140.5, 132.5, 130.5, 129.7, 128.9, 128.2, 127.3, 114.1, 75.5, 55.6, 27.4, 10.5; **IR** (neat) 2929, 1698, 1573, 1442, 1298, 1033 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₁H₂₅NO₃Na (M + Na)⁺ 362.1732, found 362.1730

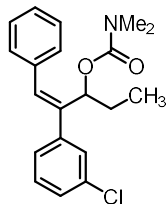


2.38 The product was prepared according to general method D using alkyne **2.17** (46.3 mg, 0.2 mmol), 2-benzofuranylboronic acid (81 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (51.0 mg, 73% yield). **TLC** *R_f* = 0.3 (15% EtOAc/hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.57–7.50 (m, 4H), 7.47 (d, *J* = 8.2 Hz, 1H), 7.40 (t, *J* = 7.7 Hz, 2H),

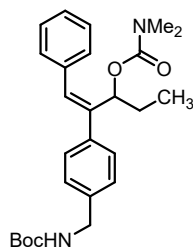
7.31–7.23 (m, 2H), 7.20 (t, $J = 7.6$ Hz, 1H), 6.96 (s, 1H), 6.03 (dd, $J = 8.6$, $J = 6.4$ Hz, 1H), 2.93 (s, 3H), 2.88 (s, 3H), 2.18–2.08 (m, 1H), 1.96–1.86 (m, 1H), 0.90 (t, $J = 7.5$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 156.3, 154.4, 154.3, 136.6, 130.8, 130.5, 129.5, 129.4, 128.8, 127.8, 124.8, 123.1, 121.2, 111.2, 105.3, 74.0, 28.1, 10.8; **IR** (neat) 3057, 2967, 1699, 1451, 1390, 1181 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{22}\text{H}_{23}\text{NO}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 372.1576, found 372.1582



2.43 The product was prepared according to general method D using alkyne **2.17** (46.3 mg, 0.2 mmol), 4-(Trifluoromethyl) phenyl boronic acid (95.0 mg, 0.5 mmol), $\text{Ni}(\text{cod})_2$ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et_2O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (62.6 mg, 83% yield). **TLC** $R_f = 0.4$ (15% EtOAc/Hex); ^1H NMR (500 MHz, CDCl_3) δ 7.63–7.58 (m, 4H), 7.43–7.36, (m, 4H), 7.31–7.26 (m, 1H), 6.73 (s, 1H), 5.85 (t, $J = 7.3$ Hz, 1H), 2.89 (s, 3H), 2.73 (s, 3H), 1.88–1.80 (m, 1H), 1.68–1.59 (m, 1H), 0.83 (t, $J = 7.5$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 156.2, 145.1 (q, $J = 1.4$ Hz), 140.9, 136.7, 133.7, 129.6 (q, $J = 32.4$ Hz) 129.23, 129.16, 128.7, 127.7, 125.2 (q, $J = 3.7$ Hz), 124.5 (q, $J = 127.4$ Hz) 75.4, 27.6, 10.5; **IR** (neat) 2969, 2934, 1699, 1322, 1164, 1122 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{21}\text{H}_{22}\text{F}_3\text{NO}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 400.1500, found 400.1489

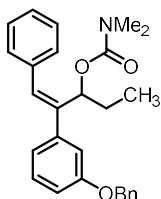


2.41 The product was prepared according to general method D using alkyne **2.17** (46.3 mg, 0.2 mmol), 3-chlorophenylboronic acid (78 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (59.8 mg, 87% yield). **TLC** R_f = 0.4 (15% EtOAc/Hex); **¹H NMR** (500 MHz, CDCl₃) δ 7.51–7.49 (m, 1H), 7.42–7.34 (m, 5H), 7.31–7.26 (m, 3H), 6.71 (s, 1H), 5.83 (dd, *J* = 8.1, *J* = 6.7, Hz, 1H), 2.89 (s, 3H), 2.76 (s, 3H), 1.88–1.78 (m, 1H), 1.67–1.58 (m, 1H), 0.82 (t, *J* = 7.3 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 156.2, 143.1, 140.7, 136.8, 134.0, 133.3, 129.5, 129.2, 129.1, 128.7, 127.59, 127.58, 127.1, 75.3, 27.5, 10.5; **IR** (neat) 2968, 2933, 1699, 1472, 1390, 1182 cm⁻¹; **HRMS** (TOF MS ES+) *m/z* calcd for C₂₀H₂₂ClNO₂Na (M + Na)⁺, 366.1237 found 366.1227

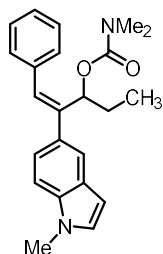


2.40 The product was prepared according to general method D using alkyne **2.17** (46.3 mg, 0.2 mmol), 4-(tert-butoxycarbonylamino)methylphenylboronic acid (126 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a

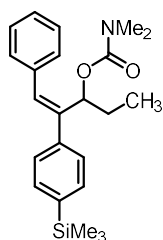
silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (57.0 mg, 65% yield). **TLC** *R_f* = 0.3 (25% EtOAc/Hex); **¹H NMR** (500 MHz, CDCl₃) δ 7.46 (d, *J* = 8.2 Hz, 2H), 7.42–7.33 (m, 4H), 7.29–7.23 (m, 3H), 6.69 (s, 1H), 5.83 (t, *J* = 7.2 Hz, 1H), 4.88 (s, 1H), 4.34 (d, *J* = 5.5 Hz, 2H), 2.88 (s, 3H), 2.74 (s, 3H), 1.88–1.78 (m, 1H), 1.66–1.58 (m, 1H), 1.48 (s, 9H), 0.80 (t, *J* = 7.5 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 156.3, 141.5, 140.4, 137.2, 132.6, 129.18, 129.11, 128.6, 127.3, 75.6, 44.7, 28.8, 27.5, 10.5; **IR** (neat) 3339, 2970, 2932, 1699, 1507, 1175 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₆H₃₄N₂O₄Na (M + Na)⁺, 461.2416 found 461.2411



2.33 The product was prepared according to general method D using alkyne **2.17** (46.3 mg, 0.2 mmol), 3-benzyloxyphenylboronic acid (114 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (48.2 mg, 58% yield). **TLC** *R_f* = 0.3 (25% EtOAc/Hex); **¹H NMR** (500 MHz, CDCl₃) δ 7.47–7.43 (m, 2H), 7.41–7.31 (m, 6H), 7.28–7.23 (m, 2H), 7.15–7.13 (m, 1H), 7.12–7.09 (m, 1H), 6.93 (ddd, *J* = 8.3, *J* = 2.6, *J* = 0.7 Hz, 1H), 6.72 (s, 1H), 5.82 (dd, *J* = 7.8, *J* = 6.8 Hz, 1H), 5.09 (s, 2H), 2.89 (s, 3H), 2.73 (s, 3H), 1.89–1.79 (m, 1H), 1.70–1.60 (m, 1H), 0.81 (t, *J* = 7.5, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 158.7, 156.3, 142.8, 141.7, 137.4, 137.2, 132.7, 129.20, 129.18, 128.9, 128.6, 128.3, 127.8, 127.4, 121.7, 115.7, 113.8, 75.6, 70.3, 27.5, 10.6; **IR** (neat) 2967, 2930, 1698, 1574, 1391, 1182 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₇H₂₉NO₃Na (M + Na)⁺, 438.2045 found 438.2029

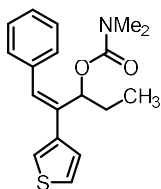


2.39 The product was prepared according to general method D using alkyne **2.17** (46.3 mg, 0.2 mmol), 1-Methyl-5-indolylboronic acid (87 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (44.9 mg, 62% yield). **TLC** R_f = 0.2 (15% EtOAc/Hex); **¹H NMR** (500 MHz, CDCl₃) δ 7.75–7.74 (m, 1H), 7.44–7.34 (m, 5H), 7.31–7.28 (m, 1H), 7.26–7.22 (m, 1H), 7.06 (d, *J* = 3.1 Hz, 1H), 6.74 (s, 1H), 6.49 (dd, *J* = 3.2 *J* = 0.7 Hz, 1H), 5.88 (t, *J* = 7.3 Hz, 1H), 3.80 (s, 3H), 2.89 (s, 3H), 2.74 (s, 3H), 1.91–1.81 (m, 1H), 1.73–1.64 (m, 1H), 0.81 (t, *J* = 7.5 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 156.5, 142.7, 137.9, 136.4, 132.7, 132.0, 129.5, 129.2, 128.5, 127.0, 123.0, 121.2, 108.8, 101.4, 76.2, 33.2, 27.6, 10.6; **IR** (neat) 2964, 2931, 2695, 1488, 1391, 1186 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₃H₂₆N₂O₂Na (M + Na)⁺, 385.1892 found 385.1900

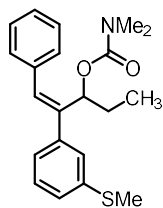


2.32 The product was prepared according to general method D using alkyne **2.17** (46.3 mg, 0.2 mmol), 4-trimethylsilylphenylboronic acid (97 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure

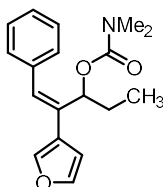
Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (59.5 mg, 78% yield). **TLC** *R_f* = 0.5 (15% EtOAc/Hex); **¹H NMR** (500 MHz, CDCl₃) δ 7.52–7.46 (m, 4H), 7.41–7.34 (m, 4H), 7.28–7.23 (m, 1H), 6.73 (s, 1H), 5.86 (dd, *J* = 7.7 *J* = 6.7 Hz, 1H), 2.89 (s, 3H), 2.75 (s, 3H), 1.89–1.80 (m, 1H), 1.70–1.60 (m, 1H), 0.81 (t, *J* = 7.5 Hz, 3H), 0.29 (s, 9H); **¹³C NMR** (125 MHz, CDCl₃) δ 156.4, 141.76, 141.74, 139.5, 137.4, 133.3, 132.8, 129.2, 128.6, 128.2, 127.3, 75.6, 27.5, 10.5, –0.7; **IR** (neat) 2955, 1703, 1460, 1392, 1248, 1183 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₃H₃₁NO₂SiNa (M + Na)⁺, 404.2022 found 404.2018



2.36 The product was prepared according to general method D using alkyne **2.17** (46.3 mg, 0.2 mmol), 3-thienylboronic acid (64 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (47.3mg, 75% yield). **TLC** *R_f* = 0.4 (15% EtOAc/Hex); **¹H NMR** (500 MHz, CDCl₃) δ 7.45–7.34 (m, 4H), 7.33–7.28 (m, 2H), 7.28–7.24 (m, 2H), 6.95 (s, 1H), 5.89 (dd, *J* = 8.1, *J* = 6.9 Hz, 1H), 2.88 (s, 3H), 2.85 (s, 3H), 1.98–1.87 (m, 1H), 1.77–1.68 (m, 1H), 0.83 (t, *J* = 7.5 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 156.3, 140.8, 137.2, 136.3, 131.1, 129.2, 128.7, 128.0, 127.4, 125.0, 122.5, 75.3, 27.7, 10.6; **IR** (neat) 2969, 1702, 1491, 1390, 1183 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₁₈H₂₁NO₂SNa (M + Na)⁺, 338.1191 found 338.1184

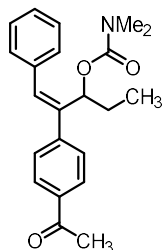


2.34 The product was prepared according to general method D using alkyne **2.17** (46.3 mg, 0.2 mmol), 3-methylthiophenylboronic acid (84 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (49.1mg, 69% yield). **TLC** *R_f* = 0.4 (15% EtOAc/Hex); **¹H NMR** (500 MHz, CDCl₃) δ 7.42–7.35 (m, 5H), 7.30–7.24 (m, 3H), 7.20 (dt, *J* = 7.1, *J* = 1.8 Hz, 1H) 6.70 (s, 1H), 5.83 (dd, *J* = 8.0, *J* = 6.7 Hz, 1H), 2.89 (s, 3H), 2.75 (s, 3H), 2.51 (s, 3H), 1.88–1.79 (m, 1H), 1.69–1.60 (m, 1H), 0.82 (t, *J* = 7.5 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 156.3, 142.0, 141.5, 138.2, 137.1, 132.9, 129.2, 128.7, 128.6, 127.4, 127.2, 125.8, 125.6, 75.5, 27.5, 16.2, 10.5; **IR** (neat) 2966, 1698, 1584, 1443, 1390, 1182 cm⁻¹; **HRMS** (TOF MS ES+) *m/z* calcd for C₂₁H₂₅NO₂SNa (M + Na)⁺, 378.1504 found 378.1510

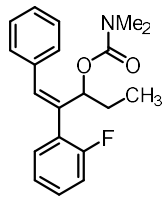


2.37 The product was prepared according to general method D using alkyne **2.17** (46.3 mg, 0.2 mmol), 3-furanboronic acid (56 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (34.1 mg, 57% yield). **TLC** *R_f* = 0.3 (15% EtOAc/Hex); **¹H NMR** (500 MHz, CDCl₃) δ 7.67 (s, 1H), 7.43–7.40 (m, 2H), 7.39–7.34 (m, 2H), 7.27–7.23 (m, 2H), 6.88

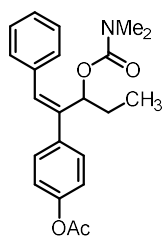
(s, 1H), 6.65–6.63 (m, 1H), 5.87 (dd, $J=8.3$, $J=6.6$ Hz, 1H), 2.90 (s, 6H), 2.0–1.89 (m, 1H), 1.78–1.69 (m, 1H), 0.84 (t, $J=7.6$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 156.3, 142.8, 140.5, 137.1, 132.6, 129.2, 129.1, 128.7, 127.3, 124.2, 109.8, 75.2, 27.7, 10.6; IR (neat) 2971, 1698, 1491, 1394, 1264, 1188 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{18}\text{H}_{21}\text{NO}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$, 322.1419 found 322.1426



2.44 The product was prepared according to general method D using alkyne **2.17** (46.3 mg, 0.2 mmol), 4-acetylphenylboronic acid (82 mg, 0.5 mmol), $\text{Ni}(\text{cod})_2$ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et_2O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (56.2 mg, 80% yield). TLC $R_f=0.2$ (12% EtOAc/Hex); ^1H NMR (500 MHz, CDCl_3) δ 7.95 (d, $J=8.2$ Hz, 2H) 7.60 (d, $J=8.3$ Hz, 2H), 7.43–7.35 (m, 4H), 7.31–7.27 (m, 1H), 6.77 (s, 1H), 5.87 (dd, $J=7.7$, $J=6.8$ Hz, 1H), 2.88 (s, 3H), 2.74 (s, 3H), 2.63 (s, 3H), 1.93–1.80 (m, 1H) 1.71–1.59 (m, 1H) 0.82 (t, $J=7.4$ Hz, 3H); ^{13}C NMR 198.2, 156.2, 146.4, 141.2, 136.8, 136.2, 133.7, 129.2, 129.1, 128.7, 128.4, 127.7, 75.4, 27.7, 27.0, 10.5; IR (neat) 2963, 2925, 1698, 1681, 1602, 1265, 1180 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{22}\text{H}_{25}\text{NO}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$, 374.1732 found 374.1729



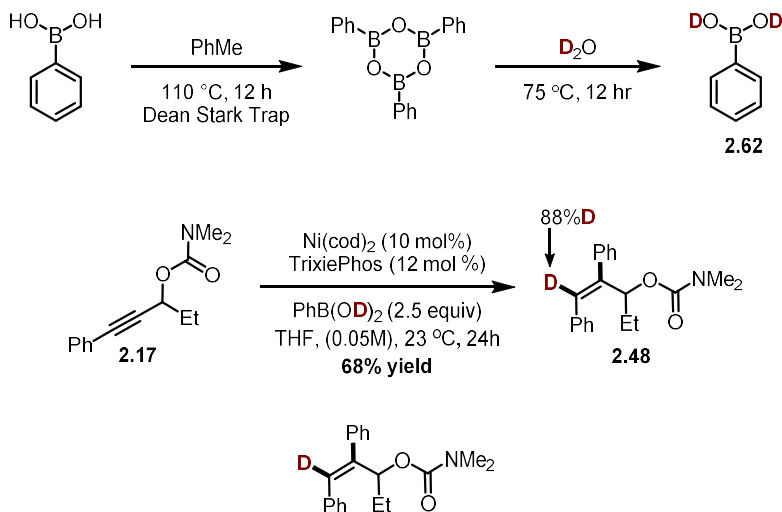
2.44 The product was prepared according to general method D using alkyne **2.17** (46.3 mg, 0.2 mmol), 2-fluorophenylboronic acid (70 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (47.1 mg, 72% yield). **TLC** R_f = 0.4 (12% EtOAc/Hex); **¹H NMR** (500 MHz, CDCl₃) δ 7.41–7.43 (m, 2H), 7.40–7.35 (m, 2H), 7.33–7.26 (m, 3H), 7.14–7.06 (m, 2H), 6.60 (s, 1H), 5.87 (dd, *J* = 8.4, *J* = 6.0 Hz, 1H), 2.89 (s, 3H), 2.68 (s, 3H), 1.81–1.58 (m, 2H), 0.85 (t, *J* = 7.4 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 160.6 (d, *J* = 245.5 Hz), 156.4, 134.1, (*J* = 1.4 Hz), 132.3 (*J* = 3.7 Hz), 131.9, 129.3, 129.21, 129.18, 128.7, 127.5, 123.6 (d, *J* = 3.7), 115.7 (d, *J* = 23.1 Hz), 75.3, 27.3, 10.5; **IR** (neat) 2931, 1698, 1487, 1391, 1182 cm⁻¹; **HRMS** (TOF MS CI+) *m/z* calcd for C₂₀H₂₂FNO₂ (M)⁺, 327.1635 found 327.1630



2.35 The product was prepared according to general method D using alkyne **2.17** (46.3 mg, 0.2 mmol), 4-acetoxyphenylboronic acid (90 mg, 0.5 mmol), Ni(cod)₂ (5.5mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et₂O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title

compound as a pale yellow oil (47.0 mg, 64% yield). **TLC** R_f = 0.3 (12% EtOAc/Hex); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 7.51–7.47 (m, 2H), 7.41–7.33 (m, 4H), 7.28–7.23 (m, 1H), 7.10–7.05 (m, 2H), 6.70 (s, 1H), 5.83 (dd, $J = 8.0$, $J = 6.6$ Hz, 1H), 2.87 (s, 3H), 2.72 (s, 3H), 2.32 (s, 3H), 1.91–1.79 (m, 1H), 1.71–1.60 (m, 1H), 0.82 (t, $J = 7.4$ Hz, 3H); **$^{13}\text{C NMR}$** (125 MHz, CDCl_3) δ 169.8, 156.3, 150.2, 141.1, 139.0, 137.2, 132.8, 129.9, 129.2, 128.6, 127.4, 121.3, 75.6, 27.5, 21.5, 10.6; **IR** (neat) 2926, 1760, 1698, 1503, 1186, 1166 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{22}\text{H}_{25}\text{NO}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$, 390.1681 found 390.1684

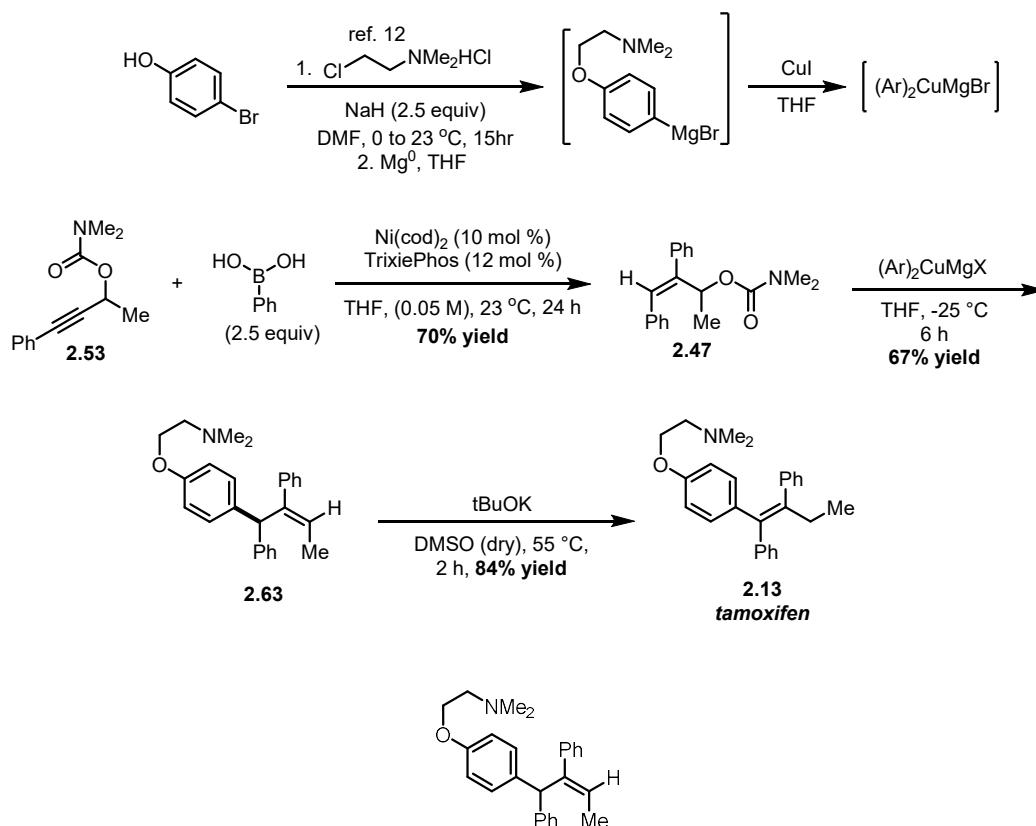
DEUTERIUM LABELING EXPERIMENTS



2.48 The product was prepared according to general method D using alkyne **2.17** (43.5 mg, 0.2 mmol), phenyl boronic acid- D_2 **2.62** (62 mg, 0.5 mmol), $\text{Ni}(\text{cod})_2$ (5.5 mg, 0.02 mmol), TrixiePhos (8.0 mg, 0.02 mmol). The reaction mixture was eluted through a silica plug (with pure Et_2O) and concentrated under reduced pressure. Purified by flash column chromatography to afford the title compound as a pale yellow oil (42.2 mg, 68% yield). **TLC** R_f = 0.4 (15% EtOAc/hexanes); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 7.51–7.46 (m, 2H), 7.43–7.23 (m, 8H), 6.71 (s, (12% H incorporation,

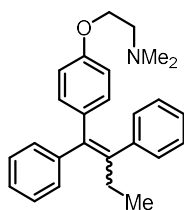
1H), 5.84 (t, $J = 6.85$ Hz, 1H), 2.88 (s, 3H), 2.7 (s, 3H), 1.85 (m, 1H), 1.65 (m, 3H), 0.82 (t, $J = 7.83$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) 156.3, 141.9, 141.4, 137.3, 132.6, 129.2, 128.9, 128.6, 128.2, 127.5, 127.3, 75.6, 27.5, 10.5; IR (neat) 2966, 2932, 1698, 1493, 1390, 1179 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{20}\text{H}_{22}\text{DNO}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 333.1689, found 333.1677.

SYNTHESIS OF TAMOXIFEN



2.63. The Grignard reagent was prepared by adding to Taillefer's method 25 mL round bottom flask: aryl bromide (0.80 g, 3.3 mmol), magnesium turnings (160 mg, 6.6 mmol, 2 equiv), THF (3 mL), and 1,2-dibromoethane (70 μL). The mixture was stirred at room temperature for 3 hours, resulting in a solution of ~ 0.55 M of Grignard reagent. To a 7 dram vial was added CuI (76 mg, 0.40 mmol) and Grignard reagent (1.5 mL, 0.80 mmol) at 0 °C, and the mixture was stirred for 30 minutes. The mixture was then cooled to -25 °C and a solution of allylic carbamate (30 mg, 0.10

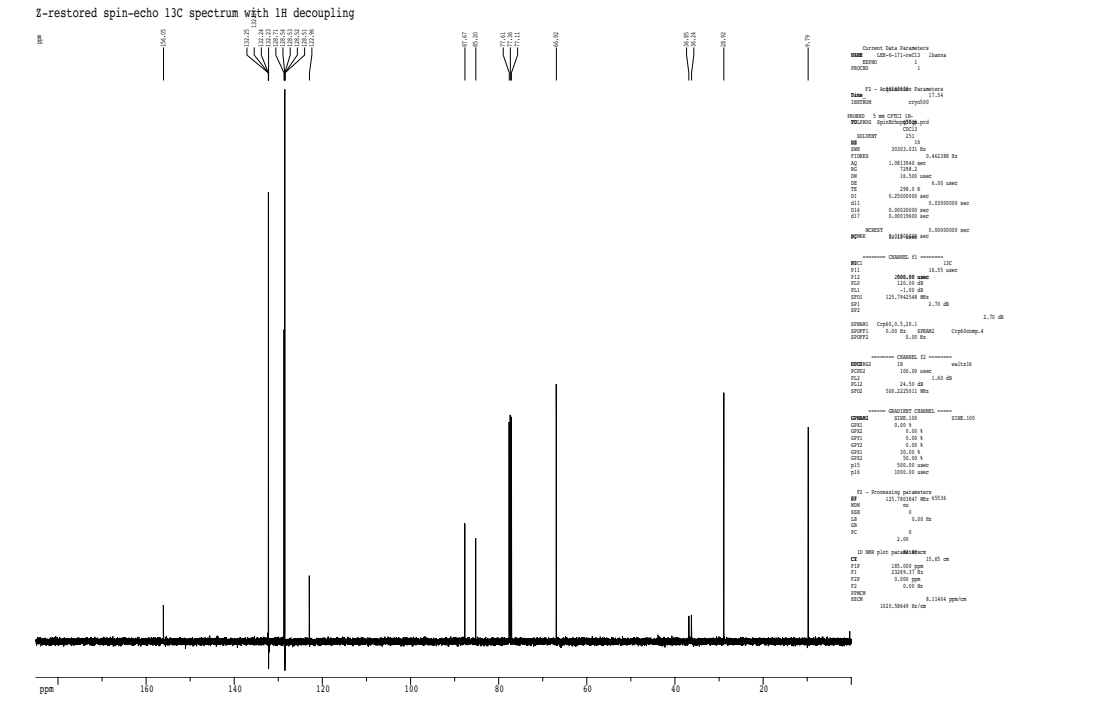
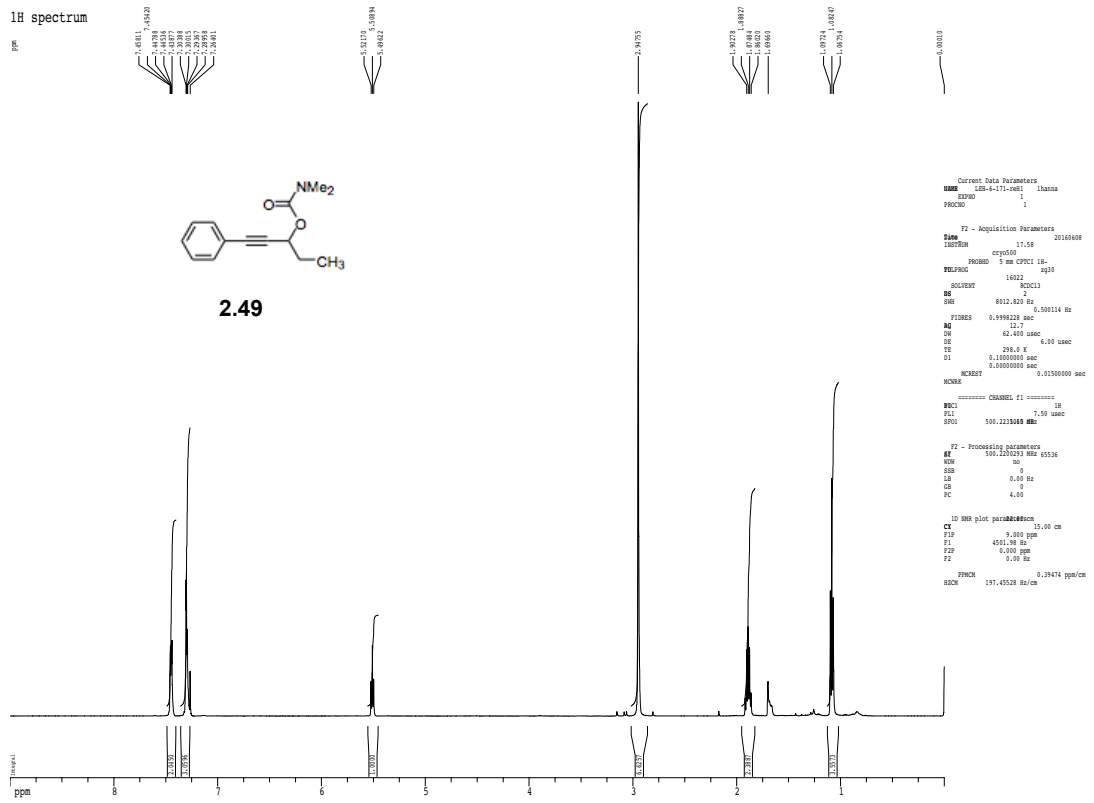
mmol, 0.1 M in THF) was added dropwise over 40 minutes. The mixture was stirred for 6 hours at $-25\text{ }^{\circ}\text{C}$ then quenched with saturated aqueous ammonium chloride, extracted with EtOAc, dried over Mg_2SO_4 and concentrated under reduced pressure. The product was purified by reverse-phase HPLC with $\text{H}_2\text{O}/\text{CH}_3\text{CN}$ (ZORBAX SB-C18 column, gradient elution of 40–70% CH_3CN w/ 0.1% TFA) to afford the compound as a colorless oil (24.1 mg, 65% yield). **TLC** $R_f = 0.3$ (15% EtOAc/hexanes w/ 5% NEt_3); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 7.37–7.28 (m, 5H), 7.27–7.16 (m, 7H), 6.78 (d, $J = 8.7$ Hz, 2H), 5.24 (q, $J = 7.0$ Hz, 1H), 5.08 (s, 1H), 4.30 (t, $J = 3.8$ Hz, 2H), 3.53 (t, $J = 3.9$ Hz, 2H), 2.99 (s, 6H), 1.58 (d, $J = 6.9$ Hz, 3H); **$^{13}\text{C NMR}$** (125 MHz, CDCl_3) δ 155.7, 144.2, 142.9, 141.8, 136.8, 131.2, 129.8, 129.1, 128.5, 128.3, 126.8, 126.6, 126.5, 114.3, 62.6, 58.9, 57.2, 44.3, 15.3; **IR** (neat) 3027, 1609, 1509, 1238, 1175^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{26}\text{H}_{29}\text{NOH}$ ($\text{M} + \text{H}$) $^+$ 372.2327, found 372.2328

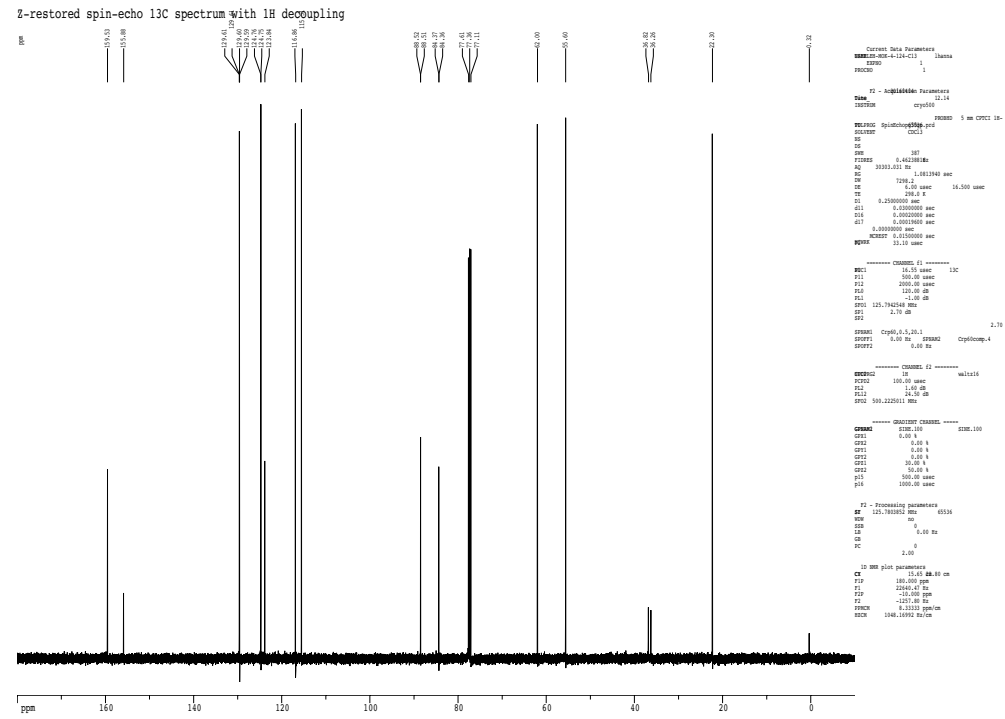
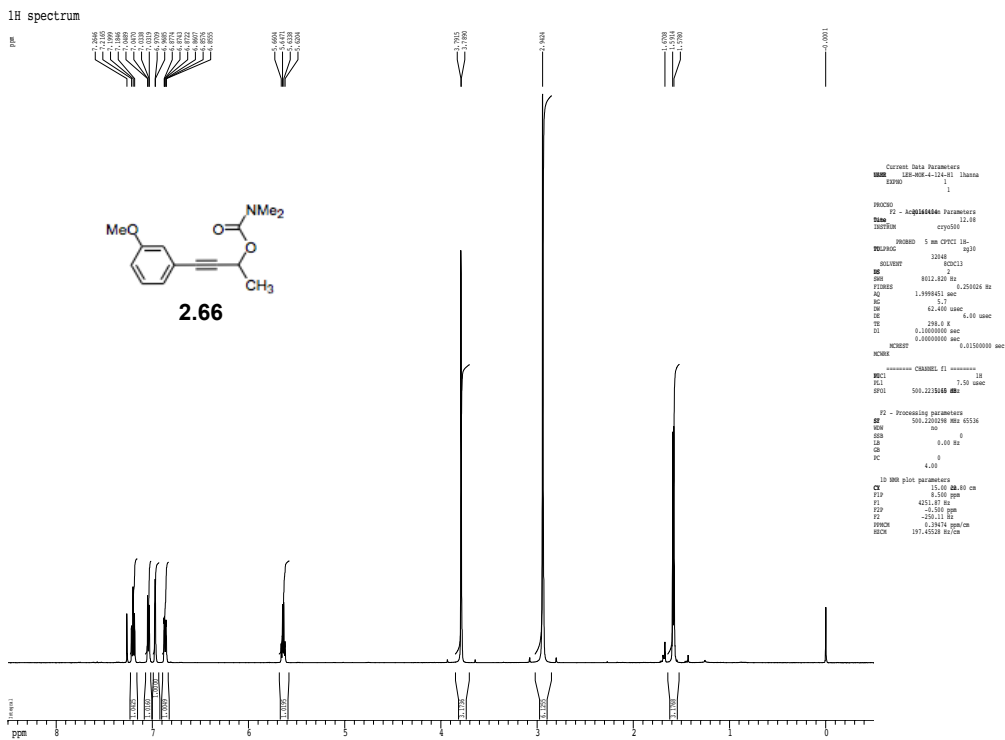


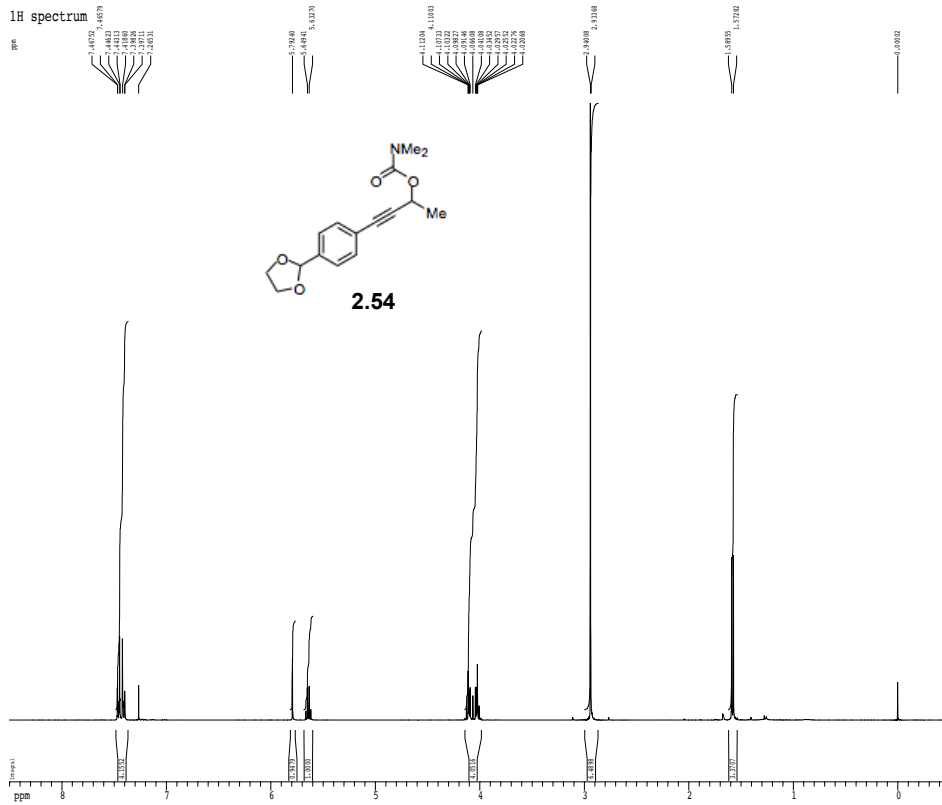
2.13 The product was prepared according to a modified procedure reported by Taillefer.¹² **2.63** (9.1 mg, 0.024 mmol) was dissolved in dry DMSO (0.8 mL) and $\text{KO}t\text{Bu}$ (14 mg, 0.12 mmol, 5 equiv) was added. The mixture was then heated at $50\text{ }^{\circ}\text{C}$ for 5 h. The crude reaction mixture was then filtered through a pad of silica and eluted with Et_2O then concentrated under reduced pressure. The product was purified by flash column chromatography to afford the title compound as a colorless oil (7.6 mg, 84% yield, 1:1 *E:Z*). **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 7.38–7.33 (m, 2H),

¹² Danoun, G.; Tlili, A.; Monnier, F.; Taillefer, M. *Angew. Chem. Int. Ed.* **2012**, *51*, 12815

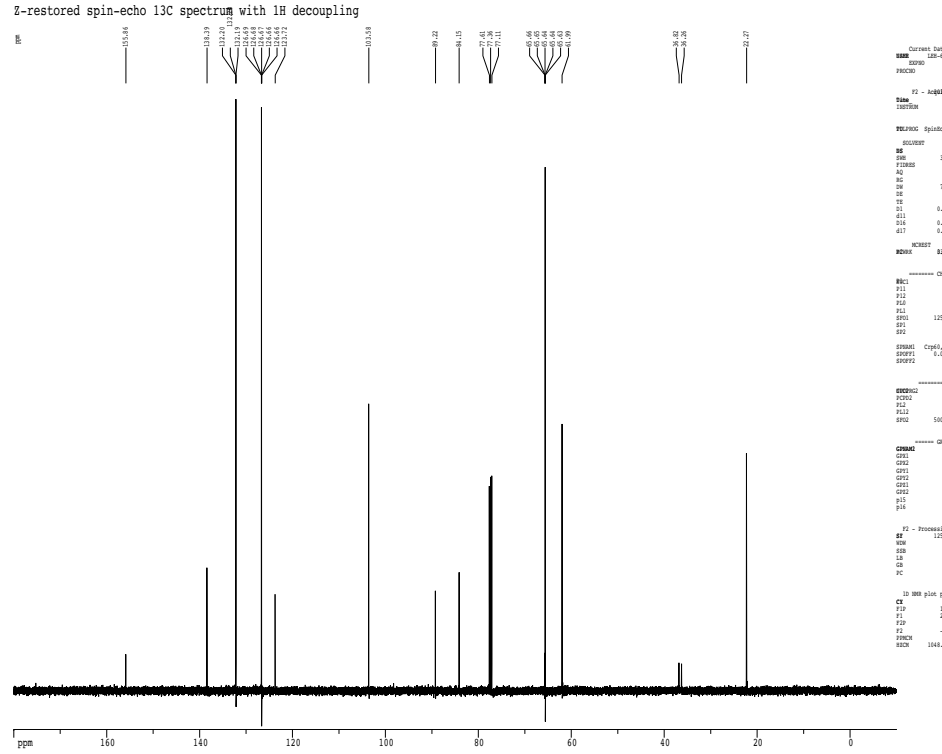
7.30–7.23 (m, 4H), 7.21–7.08 (m, 12H), 7.02–6.97 (m, 2H), 6.93–6.87 (m, 4H), 6.80–6.75 (m, 2H), 6.59–6.55 (m, 2H), 4.10 (t, $J = 6.1$ Hz, 1H), 3.94 (t, $J = 6.0$ Hz, 1H), 2.76 (t, $J = 5.8$ Hz, 1H), 2.65 (t, $J = 5.8$ Hz, 1H), 2.52 (q, $J = 7.4$ Hz, 1H), 2.47 (q, $J = 7.4$ Hz, 1H), 2.36 (s, 6H), 2.29 (s, 6H), 0.94 (q, $J = 8.6$ Hz, 6H).





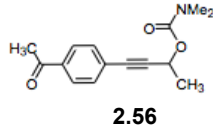
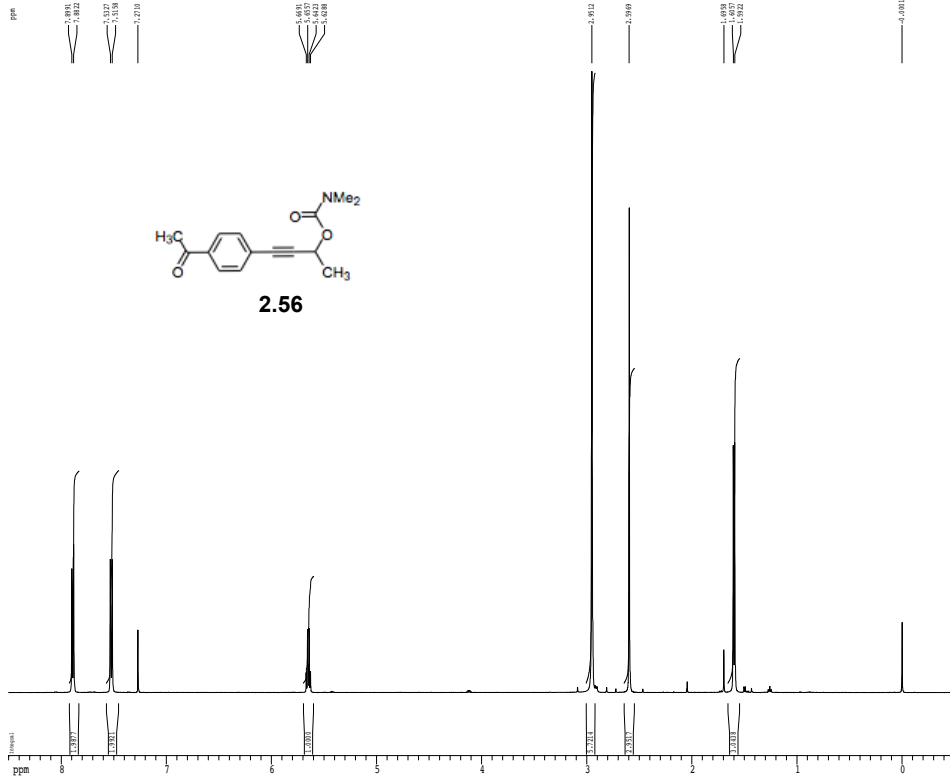


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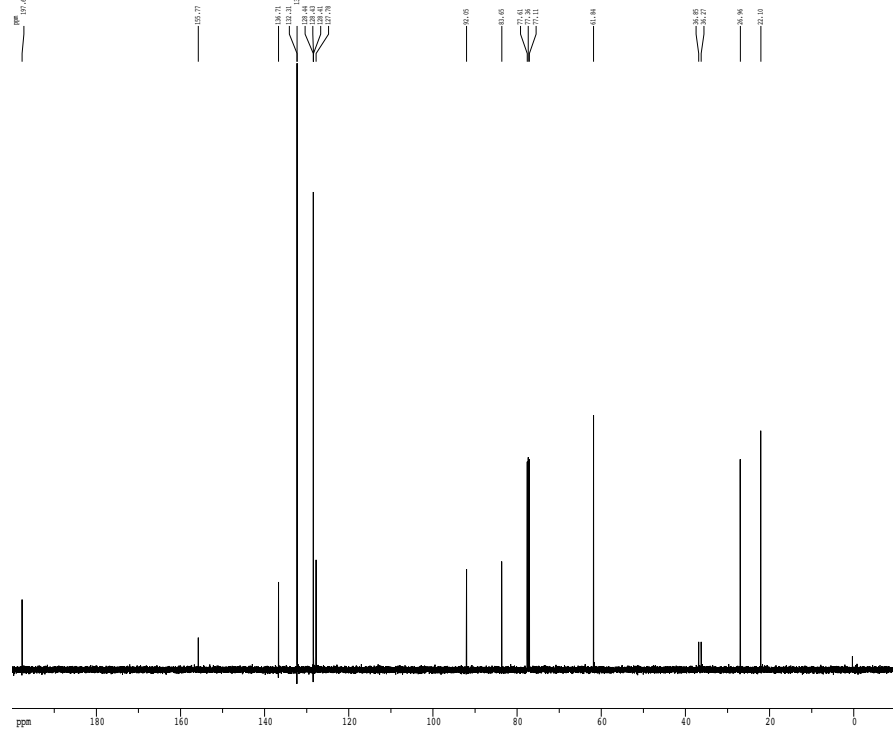
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1H spectrum



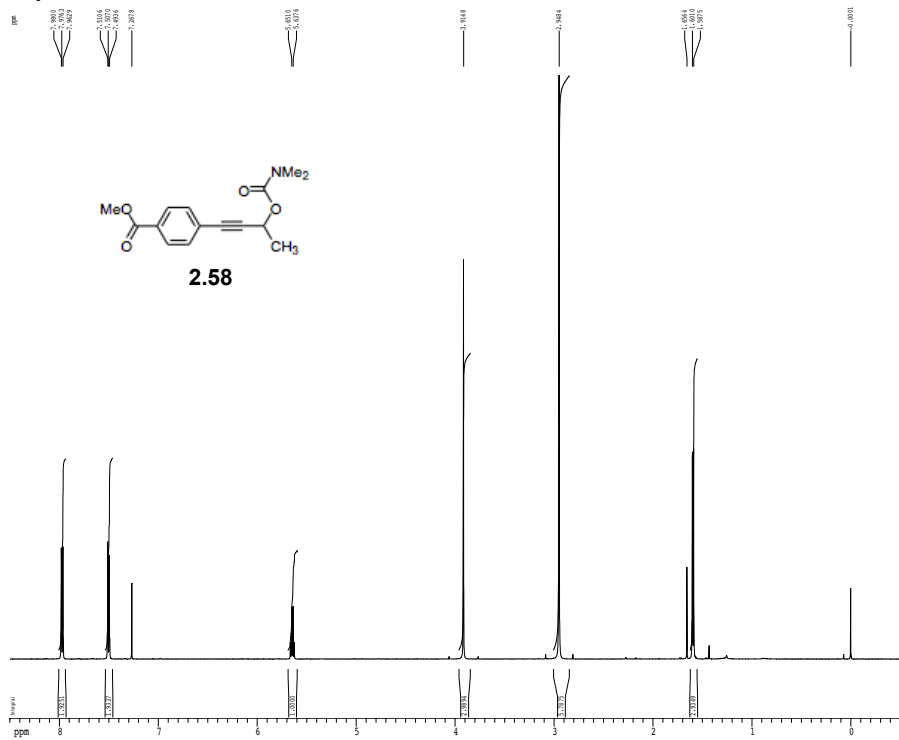
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Z-restored spin-echo 13C spectrum with 1H decoupling



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1H spectrum



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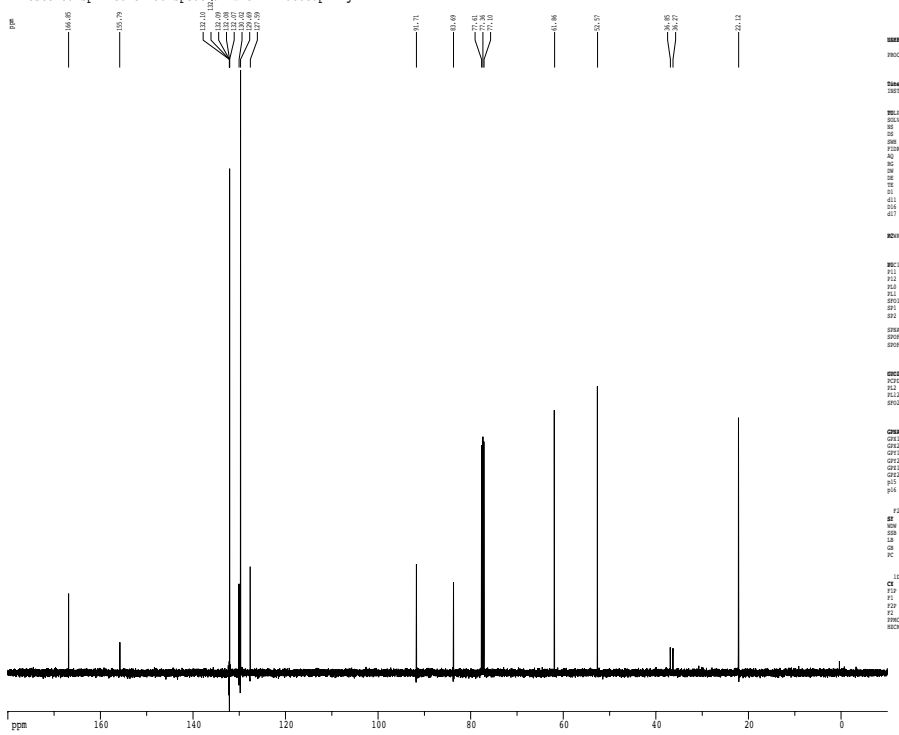
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SD 0
LB 0.00 Hz
GB 0
PC 4.00

1D NMR plot parameters
SI 15.00 dB 80 cm
F1 4210.47 Hz
F2 -20.000 ppm
F3 -299.15 Hz
FREQ 0.225015 MHz/cm
SFO 197.49518 Hz/cm
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```
Current Data Parameters
NAME=2.58-4-19-0-01 1 hanna
EXPNO 1
PROCNO 1

Date_ Time F2 - Acq040404 Parameters 15:31
INSTRUM cryo000
PROBHD 5 mm CPXI 1H-
PULPROG zgpg30
SOLVENT 30448 930
NS 316
DS 16
SFO 300.00130 MHz
AQ 0.442388 Hz
RG 512
SF 512.82400 usac
DE 6.00 usac 16.500 usac
TE 299.2 K
D1 0.25000000 sec
d11 0.00000000 sec
D15 0.00200000 sec
d17 0.00018000 sec
MCHW1 0.00000000 sec

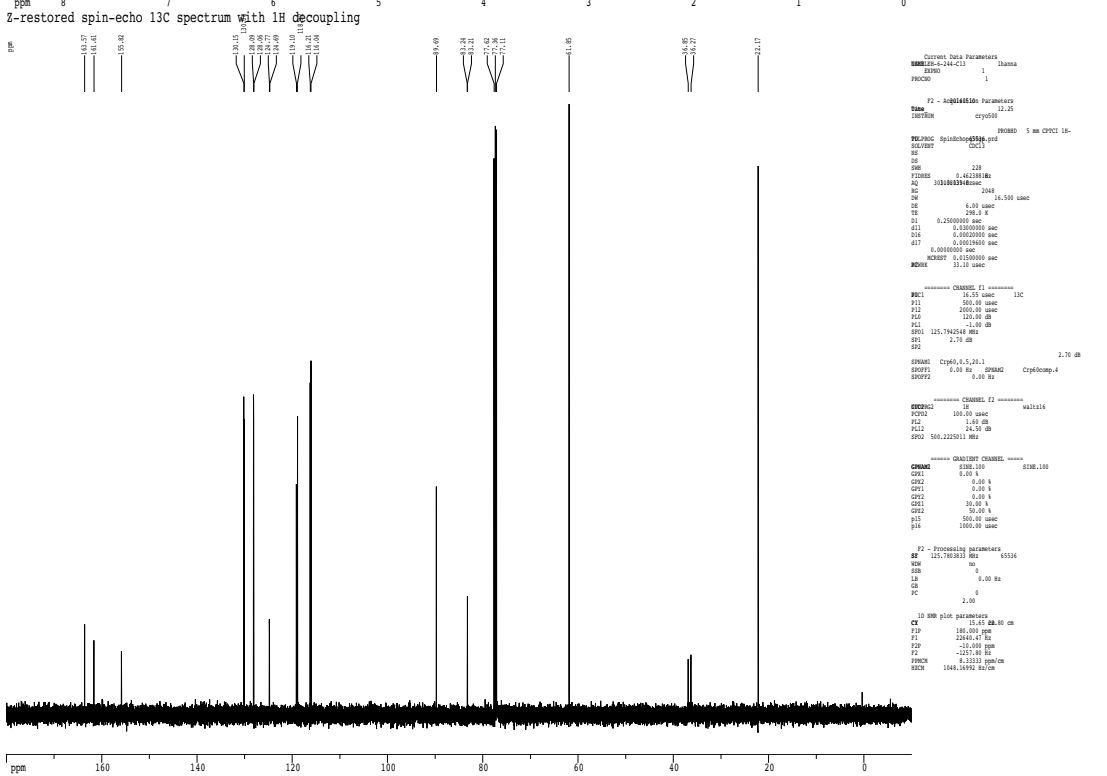
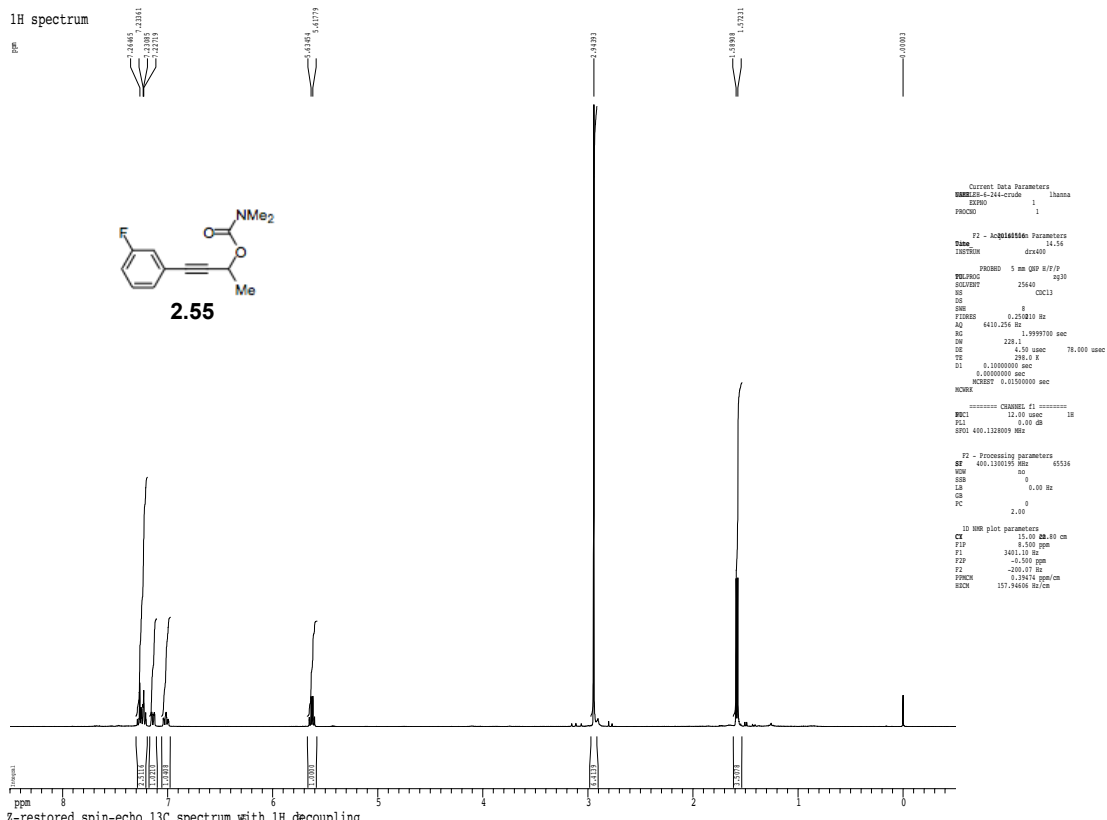
===== CHANNEL f1 =====
NUC1 13.10 usac 13C
P1 1.00 dB
SFO1 500.00 usac

===== CHANNEL f2 =====
PULPROG 16.00 usac 13C
PCPD 1.00 dB
P1 1.00 dB
SFO2 500.225015 MHz

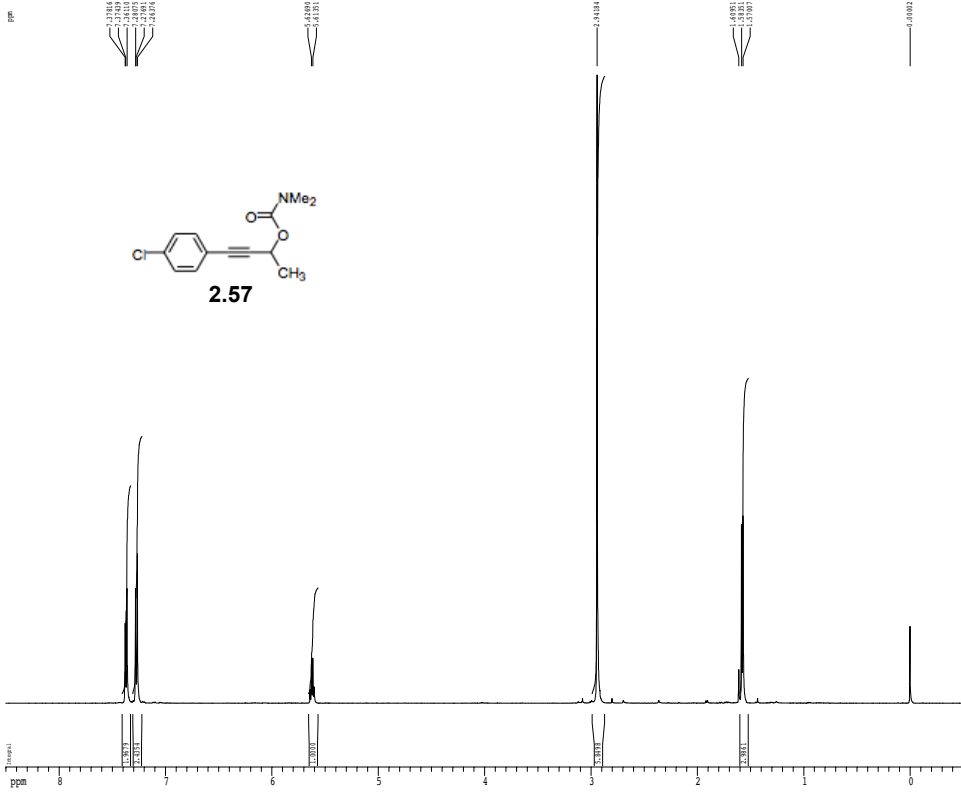
===== GRABFREQ CHANNEL =====
SI 100.00
GR1 0.00 %
GR2 0.00 %
GR3 0.00 %
GR4 0.00 %
GR5 30.00 %
GR6 50.00 %
P1 500.00 usac
P4 1000.00 usac

F2 - Processing parameters
SI 125.762382 MHz 65116
RG 0
SD 0
LB 0.00 Hz
GB 0
PC 2.00

1D NMR plot parameters
SI 15.00 dB 80 cm
F1 4210.47 Hz
F2 -20.000 ppm
F3 -299.15 Hz
FREQ 0.225015 MHz/cm
SFO 148.16972 Hz/cm
```

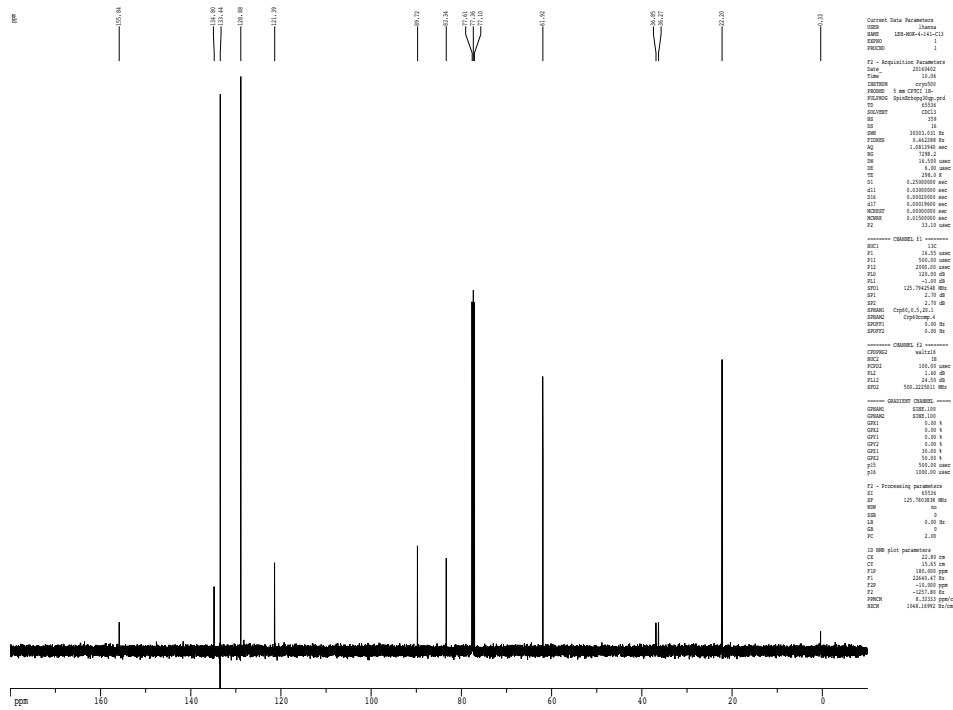


1H spectrum



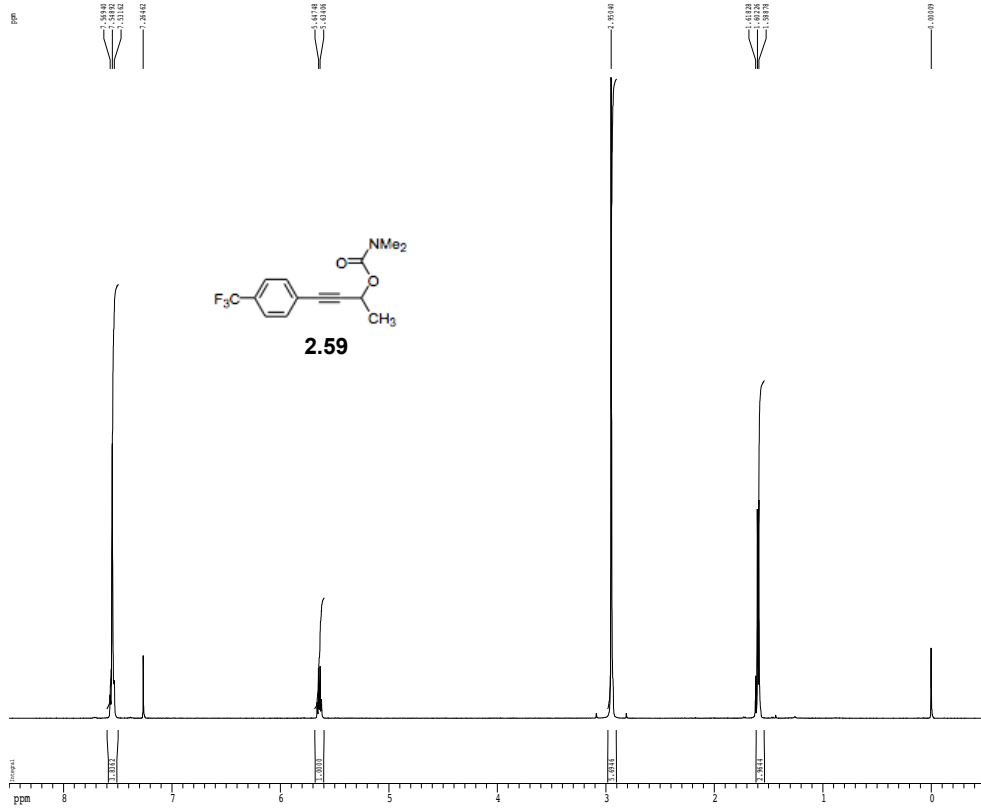
```
Current Data Parameters
USER ihama
NAME 128-MNH-4-14-03
EXPNO 1
PROCNO 1
F2 - Acquisition Parameters
Date_ 20160412
Time 9.30
INSTRUM cryo500
PROBHD 5 mm CPXI 1H
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 6
DS 2
SWH 9013.861 Hz
FIDRES 0.250026 Hz
AQ 1.999831 sec
RG 8
DM 62.400 usec
DE 6.00 usec
TE 300.2 K
D1 0.1000000 sec
d11 0.0000000 sec
d12 0.0000000 sec
d13 0.0000000 sec
d14 0.0000000 sec
===== CHANNEL f1 =====
NUC1 1H
P1 7.00 usec
PL1 0.00 dB
SFO1 500.225013 MHz
F2 - Processing parameters
SI 6526
SF 500.225013 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 4.00
ID NMR plot parameters
CX 22.80 cm
CY 15.00 cm
FID 8.500 ppm
F1 4231.87 Hz
F2 -0.000 ppm
F3 -256.11 Hz
PRN1 171.1231 Hz/cm
SDCH 191.45228 Hz/cm
```

Z-restored spin-echo 13C spectrum with 1H decoupling



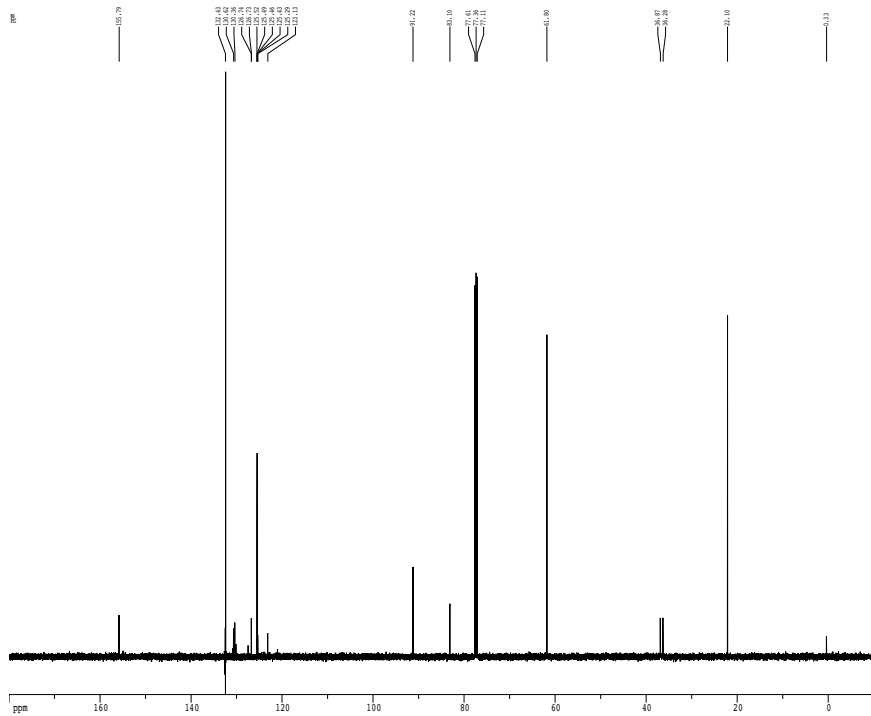
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Current Data Parameters
USER ihama
NAME 128-MNH-4-14-03
EXPNO 1
PROCNO 1
F2 - Acquisition Parameters
Date_ 20160412
Time 10.00
INSTRUM cryo500
PROBHD 5 mm CPXI 1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 6
DS 2
SWH 16013.47 Hz
FIDRES 0.46208 Hz
AQ 1.999831 sec
RG 8
DM 62.400 usec
DE 6.00 usec
TE 300.2 K
D1 0.2000000 sec
d11 0.0000000 sec
d12 0.0000000 sec
d13 0.0000000 sec
d14 0.0000000 sec
d15 0.0000000 sec
d16 0.0000000 sec
d17 0.0000000 sec
d18 0.0000000 sec
d19 0.0000000 sec
d20 0.0000000 sec
===== CHANNEL f1 =====
NUC1 13C
P1 16.00 usec
PL1 0.00 dB
PL2 190.00 usec
PL3 190.00 usec
PL4 190.00 usec
PL5 190.00 usec
PL6 190.00 usec
PL7 190.00 usec
PL8 190.00 usec
PL9 190.00 usec
PL10 190.00 usec
PL11 190.00 usec
PL12 190.00 usec
PL13 190.00 usec
PL14 190.00 usec
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
P2 16.00 usec
PL2 0.00 dB
PL3 190.00 usec
PL4 190.00 usec
PL5 190.00 usec
PL6 190.00 usec
PL7 190.00 usec
PL8 190.00 usec
PL9 190.00 usec
PL10 190.00 usec
PL11 190.00 usec
PL12 190.00 usec
PL13 190.00 usec
PL14 190.00 usec
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f3 =====
CPDPRG3 waltz16
NUC3 13C
P3 16.00 usec
PL3 0.00 dB
PL4 190.00 usec
PL5 190.00 usec
PL6 190.00 usec
PL7 190.00 usec
PL8 190.00 usec
PL9 190.00 usec
PL10 190.00 usec
PL11 190.00 usec
PL12 190.00 usec
PL13 190.00 usec
PL14 190.00 usec
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f4 =====
CPDPRG4 waltz16
NUC4 1H
P4 16.00 usec
PL4 0.00 dB
PL5 190.00 usec
PL6 190.00 usec
PL7 190.00 usec
PL8 190.00 usec
PL9 190.00 usec
PL10 190.00 usec
PL11 190.00 usec
PL12 190.00 usec
PL13 190.00 usec
PL14 190.00 usec
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f5 =====
CPDPRG5 waltz16
NUC5 13C
P5 16.00 usec
PL5 0.00 dB
PL6 190.00 usec
PL7 190.00 usec
PL8 190.00 usec
PL9 190.00 usec
PL10 190.00 usec
PL11 190.00 usec
PL12 190.00 usec
PL13 190.00 usec
PL14 190.00 usec
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f6 =====
CPDPRG6 waltz16
NUC6 1H
P6 16.00 usec
PL6 0.00 dB
PL7 190.00 usec
PL8 190.00 usec
PL9 190.00 usec
PL10 190.00 usec
PL11 190.00 usec
PL12 190.00 usec
PL13 190.00 usec
PL14 190.00 usec
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f7 =====
CPDPRG7 waltz16
NUC7 13C
P7 16.00 usec
PL7 0.00 dB
PL8 190.00 usec
PL9 190.00 usec
PL10 190.00 usec
PL11 190.00 usec
PL12 190.00 usec
PL13 190.00 usec
PL14 190.00 usec
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f8 =====
CPDPRG8 waltz16
NUC8 1H
P8 16.00 usec
PL8 0.00 dB
PL9 190.00 usec
PL10 190.00 usec
PL11 190.00 usec
PL12 190.00 usec
PL13 190.00 usec
PL14 190.00 usec
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f9 =====
CPDPRG9 waltz16
NUC9 13C
P9 16.00 usec
PL9 0.00 dB
PL10 190.00 usec
PL11 190.00 usec
PL12 190.00 usec
PL13 190.00 usec
PL14 190.00 usec
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f10 =====
CPDPRG10 waltz16
NUC10 1H
P10 16.00 usec
PL10 0.00 dB
PL11 190.00 usec
PL12 190.00 usec
PL13 190.00 usec
PL14 190.00 usec
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f11 =====
CPDPRG11 waltz16
NUC11 13C
P11 16.00 usec
PL11 0.00 dB
PL12 190.00 usec
PL13 190.00 usec
PL14 190.00 usec
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f12 =====
CPDPRG12 waltz16
NUC12 1H
P12 16.00 usec
PL12 0.00 dB
PL13 190.00 usec
PL14 190.00 usec
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f13 =====
CPDPRG13 waltz16
NUC13 13C
P13 16.00 usec
PL13 0.00 dB
PL14 190.00 usec
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f14 =====
CPDPRG14 waltz16
NUC14 1H
P14 16.00 usec
PL14 0.00 dB
PL15 190.00 usec
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f15 =====
CPDPRG15 waltz16
NUC15 13C
P15 16.00 usec
PL15 0.00 dB
PL16 190.00 usec
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f16 =====
CPDPRG16 waltz16
NUC16 1H
P16 16.00 usec
PL16 0.00 dB
PL17 190.00 usec
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f17 =====
CPDPRG17 waltz16
NUC17 13C
P17 16.00 usec
PL17 0.00 dB
PL18 190.00 usec
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f18 =====
CPDPRG18 waltz16
NUC18 1H
P18 16.00 usec
PL18 0.00 dB
PL19 190.00 usec
PL20 190.00 usec
===== CHANNEL f19 =====
CPDPRG19 waltz16
NUC19 13C
P19 16.00 usec
PL19 0.00 dB
PL20 190.00 usec
===== CHANNEL f20 =====
CPDPRG20 waltz16
NUC20 1H
P20 16.00 usec
PL20 0.00 dB
===== CHANNEL f21 =====
CPDPRG21 waltz16
NUC21 13C
P21 16.00 usec
PL21 0.00 dB
===== CHANNEL f22 =====
CPDPRG22 waltz16
NUC22 1H
P22 16.00 usec
PL22 0.00 dB
===== CHANNEL f23 =====
CPDPRG23 waltz16
NUC23 13C
P23 16.00 usec
PL23 0.00 dB
===== CHANNEL f24 =====
CPDPRG24 waltz16
NUC24 1H
P24 16.00 usec
PL24 0.00 dB
===== CHANNEL f25 =====
CPDPRG25 waltz16
NUC25 13C
P25 16.00 usec
PL25 0.00 dB
===== CHANNEL f26 =====
CPDPRG26 waltz16
NUC26 1H
P26 16.00 usec
PL26 0.00 dB
===== CHANNEL f27 =====
CPDPRG27 waltz16
NUC27 13C
P27 16.00 usec
PL27 0.00 dB
===== CHANNEL f28 =====
CPDPRG28 waltz16
NUC28 1H
P28 16.00 usec
PL28 0.00 dB
===== CHANNEL f29 =====
CPDPRG29 waltz16
NUC29 13C
P29 16.00 usec
PL29 0.00 dB
===== CHANNEL f30 =====
CPDPRG30 waltz16
NUC30 1H
P30 16.00 usec
PL30 0.00 dB
```

1H spectrum



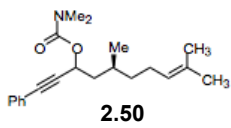
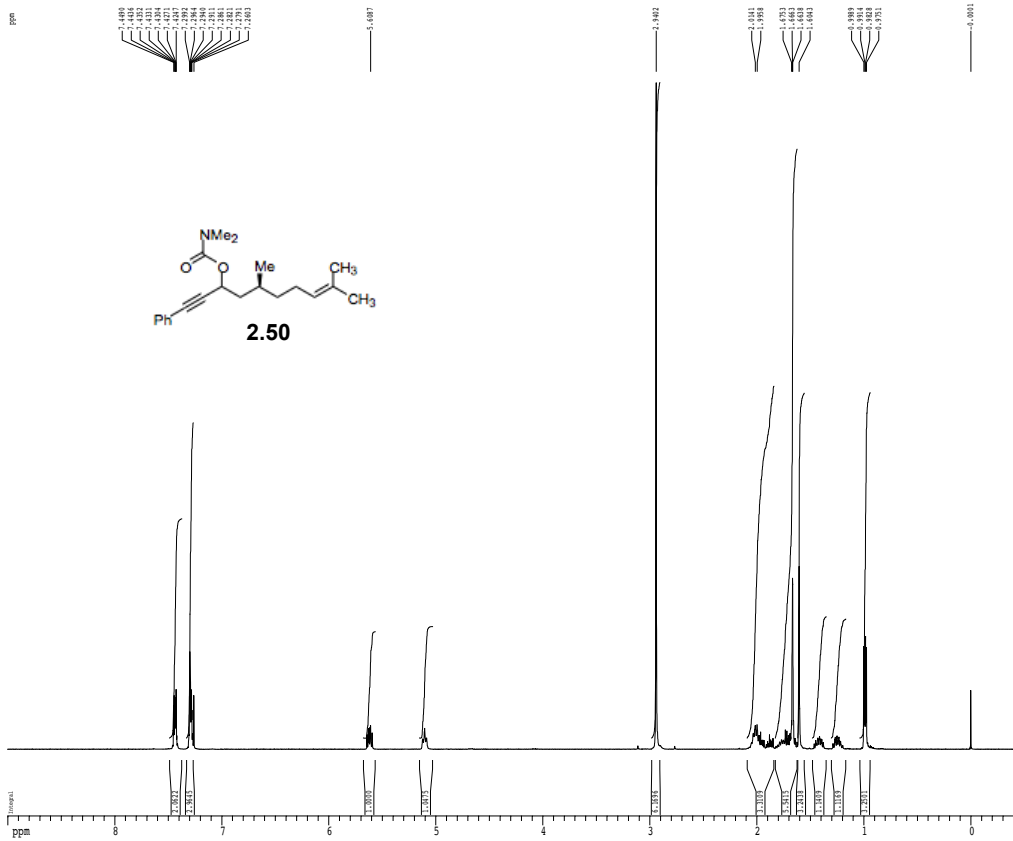
Current Data Parameters
NAME 128-MNH-4-112-81
PROCNO 1
F2 - Acquisition Parameters
Date_ 20160402
Time 9.30
INSTRUM cryo100
PROBHD 5 mm CPY1 16-
PULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 8
DS 2
SWH 8012.828 Hz
FIDRES 0.230026 Hz
AQ 1.999811 sec
RG 6.2
DE 62.400 umsec
TE 298.2 K
D1 0.1000000 sec
MCHSET 0.0000000 sec
MCHW 0.0150000 sec
===== CHANNEL f1 =====
NUC1 13C
P1 7.50 umsec
PL1 0.00 dB
SFO1 500.2215015 MHz
F2 - Processing parameters
SI 6526
SF 500.2200294 MHz
WDW no
SSB no
LB 0.00 Hz
GB 0
PC 4.00
1D NMR plot parameters
CX 21.80 cm
CY 15.00 cm
P1P 61.500 ppm
F1 4211.87 Hz
F2P -41.500 ppm
F2 -210.11 Hz
PRNCH 0.12474 ppm/cm
SICH 197.45528 Hz/cm

2-restored spin-echo 13C spectrum with 1H decoupling

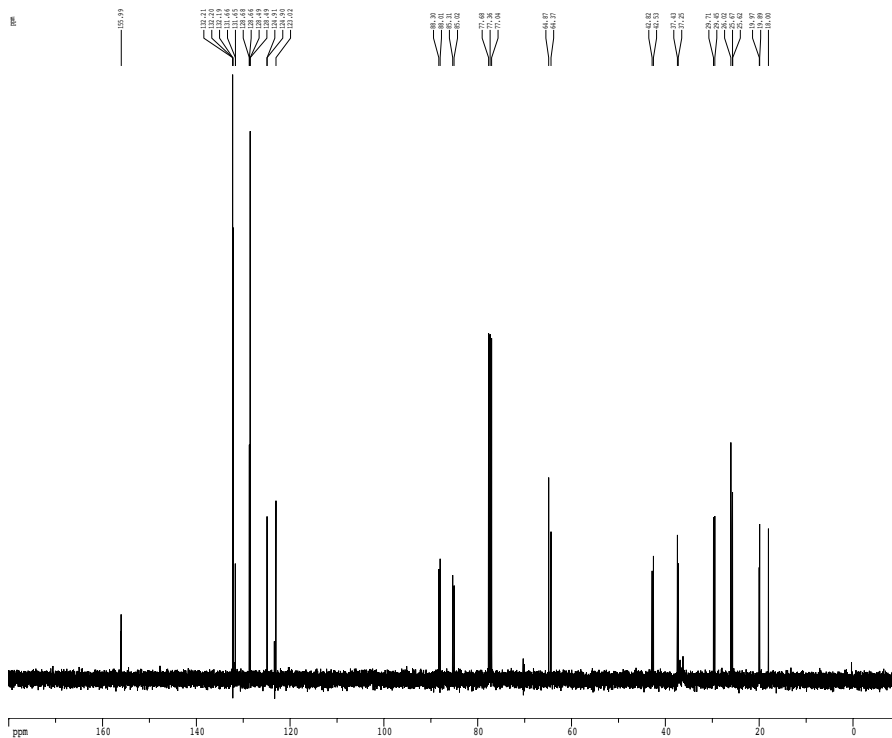


Current Data Parameters
NAME 128-MNH-4-112-81
PROCNO 1
F2 - Acquisition Parameters
Date_ 20160402
Time 9.34
INSTRUM cryo100
PROBHD 5 mm CPY1 16-
PULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 8
DS 2
SWH 8012.828 Hz
FIDRES 0.230026 Hz
AQ 1.999811 sec
RG 6.2
DE 62.400 umsec
TE 298.2 K
D1 0.1000000 sec
MCHSET 0.0000000 sec
MCHW 0.0150000 sec
===== CHANNEL f1 =====
NUC1 13C
P1 7.50 umsec
PL1 0.00 dB
SFO1 500.2215015 MHz
F2 - Processing parameters
SI 6526
SF 500.2200294 MHz
WDW no
SSB no
LB 0.00 Hz
GB 0
PC 4.00
1D NMR plot parameters
CX 21.80 cm
CY 15.00 cm
P1P 61.500 ppm
F1 4211.87 Hz
F2P -41.500 ppm
F2 -210.11 Hz
PRNCH 0.12474 ppm/cm
SICH 197.45528 Hz/cm

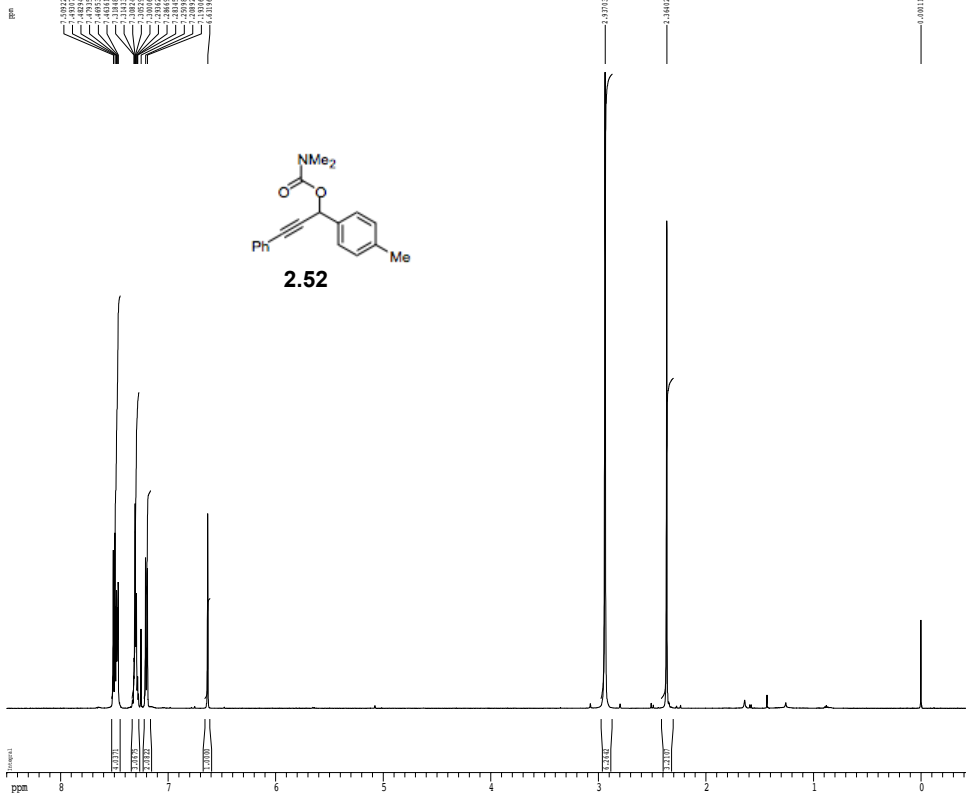
1H spectrum



13C spectrum with 1H decoupling

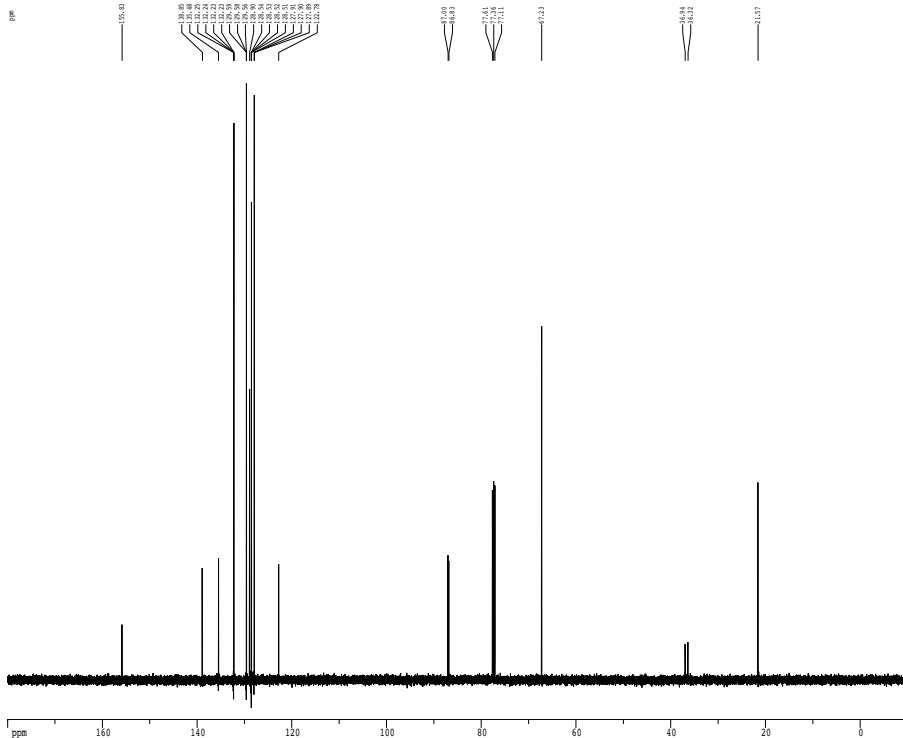


1H spectrum



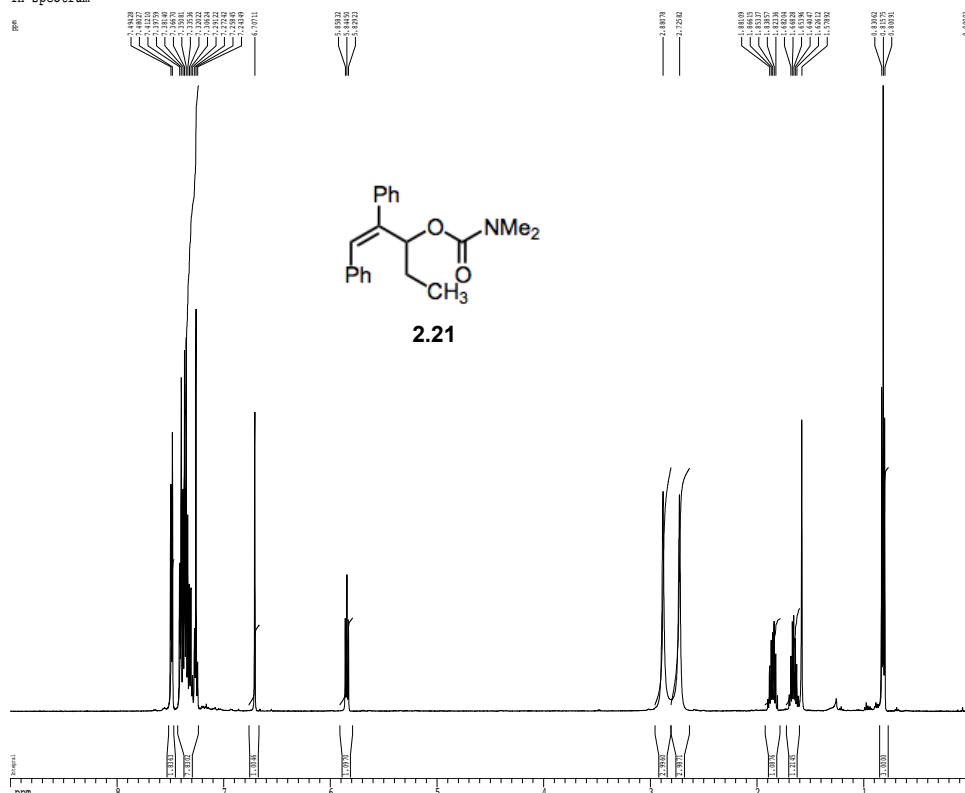
```
Current Data Parameters
USER: M00000
NAME: M04-IV-123char
EXPNO: 1
PROCNO: 1
F2 - Acquisition Parameters
Date_: 20160517
Time: 16:03
INSTRUM: cryo500
PROBHD: 5 mm CRY1 1H-
PULPROG: zgpg
TD: 327384
SOLVENT: CDCl3
NS: 2
DS: 2
SWH: 8012.828 Hz
FIDRES: 0.250026 Hz
AQ: 1.1896451 sec
RG: 6
RG2: 8
AQ2: 12.460 usec
DE: 4.00 usec
TE: 296.2 K
D1: 0.1000000 sec
MCHRG1: 0.0000000 sec
MCHRG2: 0.0150000 sec
***** CHANNEL f1 *****
NUC1: 13C
P1: 7.50 usec
P11: 1.00 dB
SFO1: 500.225013 MHz
F2 - Processing parameters
SI: 6552
SF: 500.225013 MHz
WDW: no
SSB: 0
LB: 0.00 Hz
GB: 0
PC: 4.00
1D NMR plot parameters
CP: 12.80 cm
CT: 15.00 cm
FIP: 8.500 ppm
FZ: 4251.87 Hz
F2P: -13.000 ppm
FZ2: -2561.10 Hz
FNUC1: 6.26458 ppm/cm
FNUC2: 197.45528 Hz/cm
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```
Current Data Parameters
USER: M00000
NAME: M04-IV-123char
EXPNO: 1
PROCNO: 1
F2 - Acquisition Parameters
Date_: 20160517
Time: 16:03
INSTRUM: cryo500
PROBHD: 5 mm CRY1 1H-
PULPROG: zgpg
TD: 327384
SOLVENT: CDCl3
NS: 2
DS: 2
SWH: 30323.216 Hz
FIDRES: 0.462289 Hz
AQ: 1.4812908 sec
RG: 5792.6
RG2: 8
AQ2: 6.00 usec
DE: 296.2 K
D1: 0.1000000 sec
MCHRG1: 0.0000000 sec
MCHRG2: 0.0000000 sec
MCHRG3: 0.0150000 sec
F1: 125.76120 MHz
***** CHANNEL f1 *****
NUC1: 13C
P1: 16.55 usec
P11: 500.00 dB
P12: 2000.00 usec
SFO1: 125.76120 MHz
P13: -1.00 dB
SFO2: 125.76120 MHz
SFO3: 125.76120 MHz
SFO4: 125.76120 MHz
SFO5: 125.76120 MHz
SFO6: 125.76120 MHz
SFO7: 125.76120 MHz
SFO8: 125.76120 MHz
SFO9: 125.76120 MHz
SFO10: 125.76120 MHz
***** CHANNEL f2 *****
CPROG2: waltz16
NUC2: 13C
P2: 100.00 usec
P21: 1.00 dB
P22: 1.00 dB
SFO1: 125.76120 MHz
SFO2: 500.225013 MHz
***** CHANNEL f3 *****
CPROG3: waltz16
NUC3: 13C
P3: 100.00 usec
P31: 1.00 dB
P32: 1.00 dB
SFO1: 125.76120 MHz
SFO2: 500.225013 MHz
***** CHANNEL f4 *****
CPROG4: waltz16
NUC4: 13C
P4: 100.00 usec
P41: 1.00 dB
P42: 1.00 dB
SFO1: 125.76120 MHz
SFO2: 500.225013 MHz
P4 - Processing parameters
SI: 6552
SF: 125.76120 MHz
WDW: no
SSB: 0
LB: 0.00 Hz
GB: 0
PC: 2.00
1D NMR plot parameters
CP: 12.80 cm
CT: 15.00 cm
FIP: 161.000 ppm
FZ: 2284.67 Hz
F2P: -13.000 ppm
FZ2: -2561.10 Hz
FNUC1: 6.26458 ppm/cm
FNUC2: 197.45528 Hz/cm
```


¹H spectrum



Current Data Parameters
 USER: llauna
 NAME: LES-4-284-R1
 EXPNO: 1
 PROCNO: 1

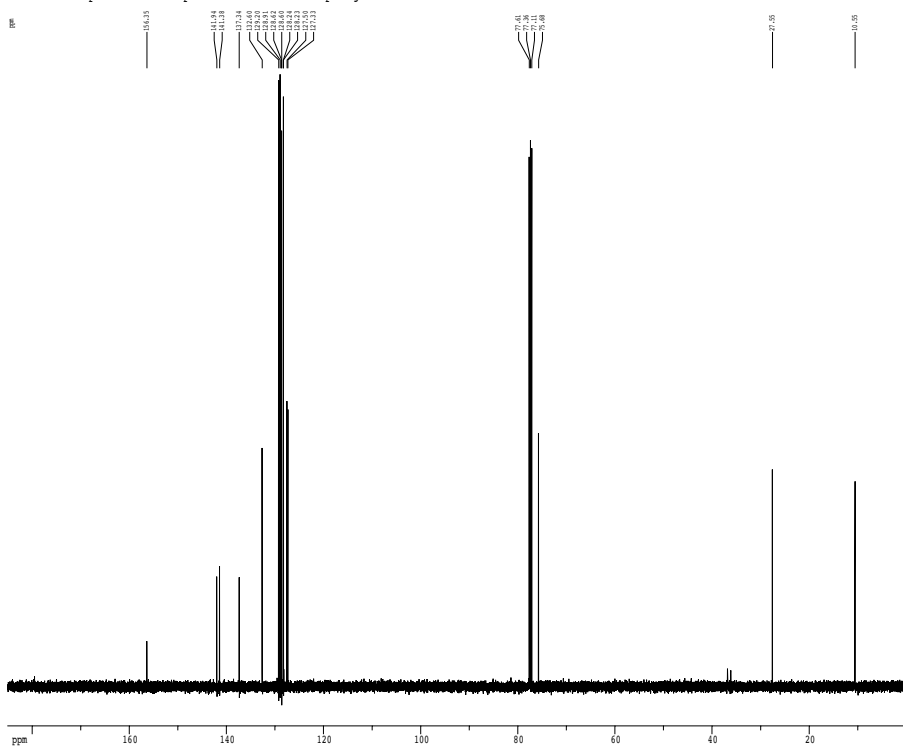
F2 - Acquisition Parameters
 DATE_: 20160119
 TIME: 11.01
 INSTRUM: cryo500
 PROBHD: 5 mm CPXC13
 PULPROG: zgpg30
 TD: 32768
 SFO1: 500.131360
 NS: 2
 DS: 4
 SWH: 8012.820 Hz
 FIDRES: 0.260114 Hz
 AQ: 0.9992228 sec
 RG: 71.1
 DW: 62.400 usec
 DE: 4.00 usec
 TE: 298.2 K
 D1: 0.2000000 sec
 MCHRG1: 0.0000000 sec
 MCHRG2: 0.0150000 sec

===== CHANNEL f1 =====
 NUCL1: 13C
 P1: 7.50 usec
 PL1: 0.00 dB
 SFO1: 500.225015 MHz

F2 - Processing parameters
 SI: 65536
 SF: 500.225015 MHz
 NS: 2
 DS: 4
 SSB: 0
 GB: 0.00 Hz
 PC: 0
 MC: 4.00

1D NMR plot parameters
 CX: 22.80 cm
 CY: 15.00 cm
 P1P: 9.000 ppm
 P1: 4500.00 Hz
 P2P: 0.000 ppm
 P2: 0.000 Hz
 PPM2M: 0.28474 ppm/cm
 SFO2: 137.4528 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
 USER: llauna
 NAME: LES-4-284-R1
 EXPNO: 1
 PROCNO: 1

F2 - Acquisition Parameters
 DATE_: 20160119
 TIME: 11.05
 INSTRUM: cryo500
 PROBHD: 5 mm CPXC13
 PULPROG: zgpg30
 TD: 65536
 SFO1: 500.131360
 NS: 2
 DS: 4
 SWH: 30382.031 Hz
 FIDRES: 0.462388 Hz
 AQ: 1.0000000 sec
 RG: 5161.6
 DW: 18.500 usec
 DE: 4.00 usec
 TE: 298.2 K
 D1: 0.2500000 sec
 d11: 0.0300000 sec
 D15: 0.0020000 sec
 d17: 0.0020000 sec
 MCHRG1: 0.0000000 sec
 MCHRG2: 0.0150000 sec
 F2: 33.10 usec

===== CHANNEL f1 =====
 NUCL1: 13C
 P1: 18.50 usec
 PL1: 580.00 usec
 PL2: 3000.00 usec
 PL3: 120.00 dB
 PL4: -2.00 dB
 SFO1: 125.7642548 MHz
 SFO2: 2.70 MHz
 SFO3: 2.70 MHz
 SFO4: CryoLoop, 4
 SFO5: 0.00 Hz
 SFO6: 0.00 Hz

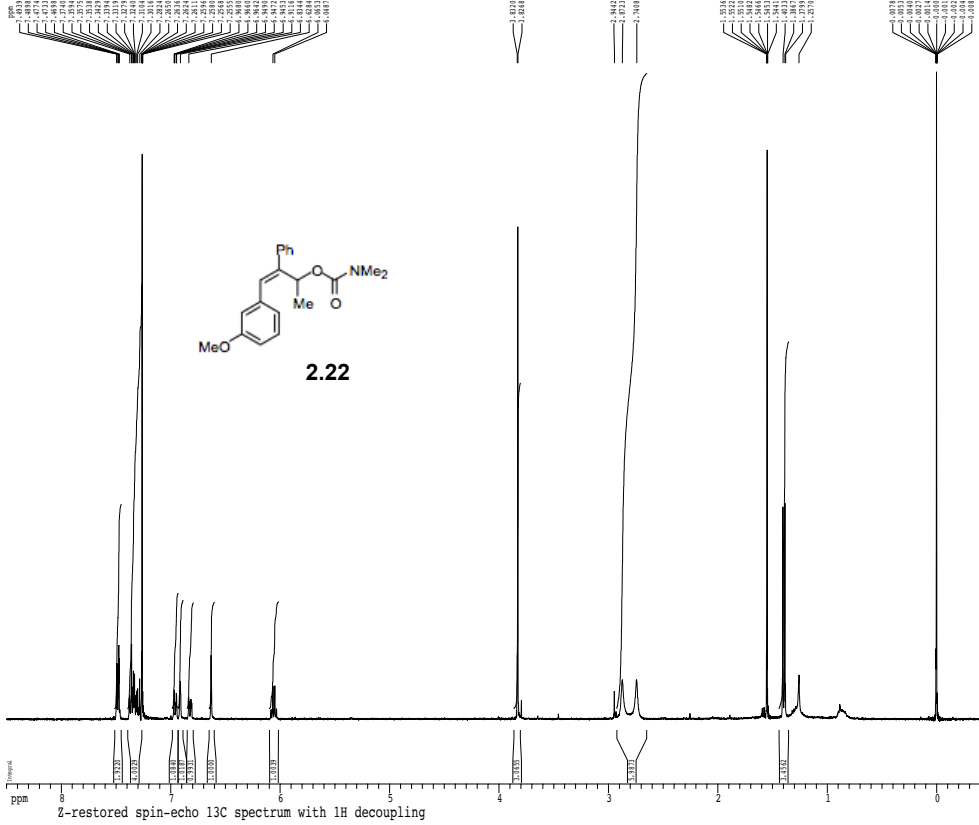
===== CHANNEL f2 =====
 CPDPRG2: waltz16
 NUCL2: 1H
 P2P2: 180.00 usec
 PL2: 1.00 dB
 PL12: 24.50 dB
 SFO2: 500.225015 MHz

===== CHANNEL f3 =====
 CHRG1: 5180.100
 CHRG2: 5180.100
 CP1: 0.00 %
 CP2: 0.00 %
 CP3: 0.00 %
 CP4: 0.00 %
 CP5: 0.00 %
 CP6: 50.00 %
 SFO3: 500.00 usec
 P15: 1000.00 usec

F2 - Processing parameters
 SI: 65536
 SF: 125.7693833 MHz
 NS: 2
 DS: 4
 SSB: 0
 GB: 0.00 Hz
 PC: 2.00

1D NMR plot parameters
 CX: 22.80 cm
 CY: 15.00 cm
 P1P: 181.000 ppm
 P1: 22887.37 Hz
 P2P: 0.000 ppm
 P2: 0.000 Hz
 PPM2M: 8.1484 ppm/cm
 SFO2: 1020.58463 Hz/cm

1H spectrum



```

Current Data Parameters
USER          jhans
NAME          LBN-6-22b-c13-14
EXPNO        1
PROCNO       1

F2 - Acquisition Parameters
Data_        2010429
Time         13:27
INSTRUM      cryo50
PROBHD       5 mm QNP 675 P9
PULPROG      zgpg
TD           65536
SOLVENT      CDCl3
NS           8
DS           2
SWH           6410.266 Hz
FIDRES       0.250000 Hz
AQ           1.989970 sec
RG           1149.4
MS           18.000 usec
WDW           EM
SS           4.50 usec
LB           298.0 K
GB           0.1000000 sec
PC           0.0000000 sec
MCHRG1       0.0000000 sec
MCHRG2       0.0000000 sec

===== CHANNEL f1 =====
NUC1          13
P1            12.00 usec
PL1           0.00 dB
SFO1         400.1218000 MHz

F2 - Processing parameters
SI            65536
SF           400.1218013 MHz
WDW          EM
SS           4.50
LB           0.00 Hz
GB           0
PC           2.00

ID NHN plot parameters
CN            22.80 cm
CT            15.00 cm
F1D           8.50 ppm
F1            3401.11 Hz
F2D           -2.50 ppm
F2            -201.77 Hz
F3D           0.0000000 sec
F3            0.19474 ppm/cm
F4D           0.0000000 sec
F4            157.34608 Hz/cm
    
```

```

Current Data Parameters
USER          jhans
NAME          LBN-6-22b-c13-14
EXPNO        1
PROCNO       1

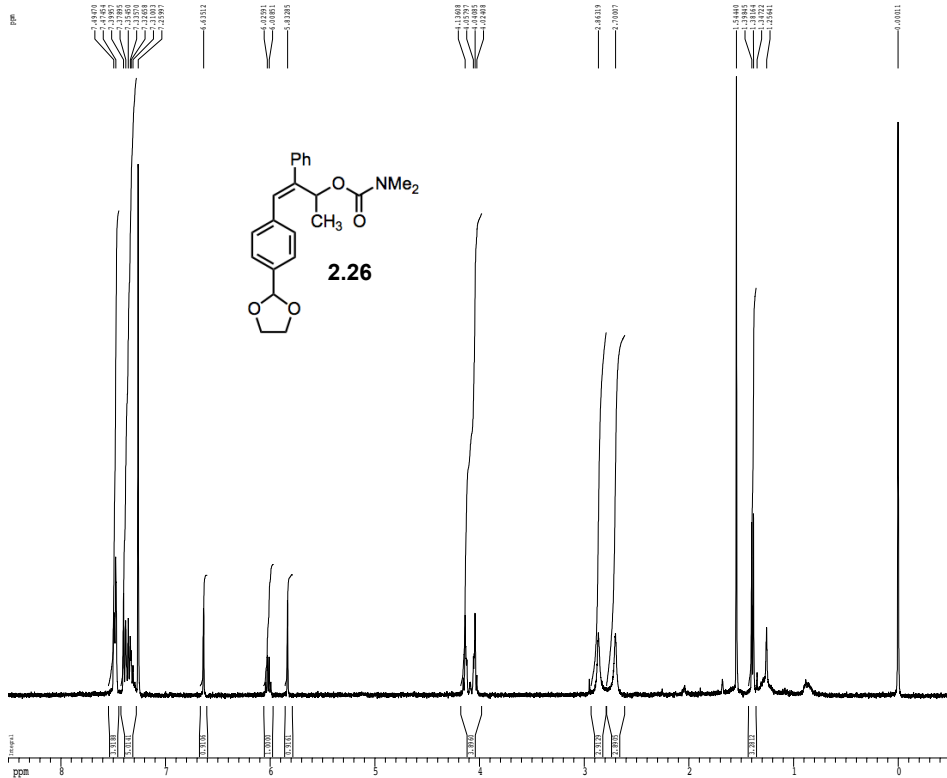
F2 - Acquisition Parameters
Data_        2010429
Time         13:27
INSTRUM      cryo50
PROBHD       5 mm QNP 675 P9
PULPROG      zgpg
TD           65536
SOLVENT      CDCl3
NS           8
DS           2
SWH           6410.266 Hz
FIDRES       0.250000 Hz
AQ           1.989970 sec
RG           1149.4
MS           18.000 usec
WDW           EM
SS           4.50 usec
LB           298.0 K
GB           0.1000000 sec
PC           0.0000000 sec
MCHRG1       0.0000000 sec
MCHRG2       0.0000000 sec

===== CHANNEL f1 =====
NUC1          13
P1            12.00 usec
PL1           0.00 dB
SFO1         400.1218000 MHz

F2 - Processing parameters
SI            65536
SF           400.1218013 MHz
WDW          EM
SS           4.50
LB           0.00 Hz
GB           0
PC           2.00

ID NHN plot parameters
CN            22.80 cm
CT            15.00 cm
F1D           8.50 ppm
F1            3401.11 Hz
F2D           -2.50 ppm
F2            -201.77 Hz
F3D           0.0000000 sec
F3            0.19474 ppm/cm
F4D           0.0000000 sec
F4            157.34608 Hz/cm
    
```


1H spectrum



```

Current Data Parameters
USER          : shana
NAME         : IES-4-255-21-050
EXPNO       : 1
PROCNO      : 1

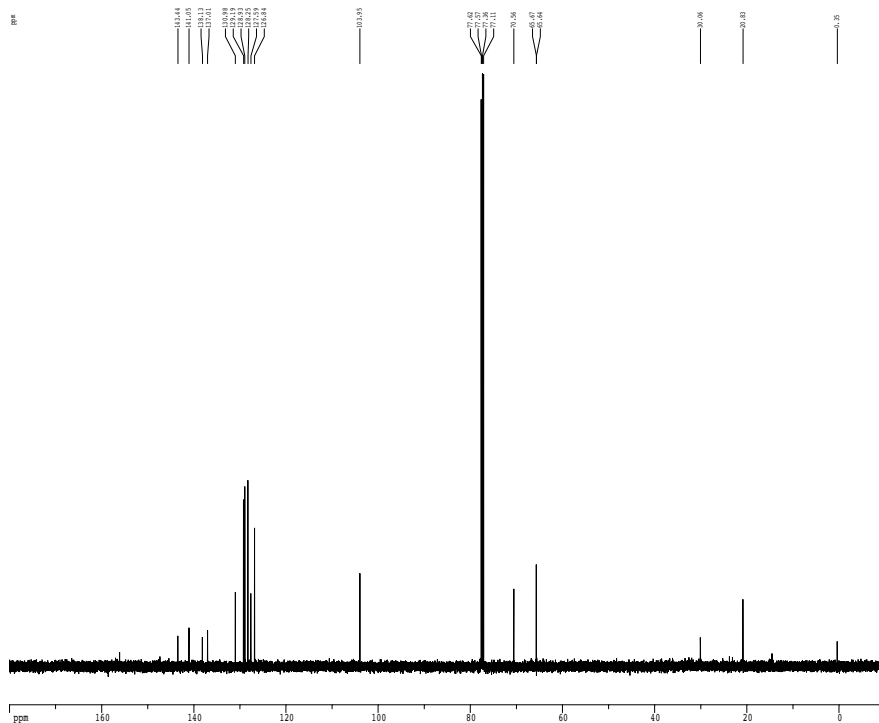
F2 - Acquisition Parameters
Date_       : 20160320
Time       : 12.44
INSTRUM    : draco
PROBHD     : 5 mm QNP 82TP
PULPROG    : zgpg
TD         : 26240
SOLVENT    : CDCl3
NS         : 2
DS         : 4
SFO        : 640.256 Hz
FIDRES     : 0.250010 sec
AQ         : 1.995701 sec
RG         : 1149.4
SQ         : 78.000 usec
DE         : 4.50 usec
TE         : 298.2 K
SI         : 0.1000000 sec
MCHRG     : 0.0000000 sec
MCHRG     : 0.0100000 sec

===== CHANNEL f1 =====
NUC1       : 13C
P1         : 12.00 usec
PL1        : 0.00 dB
SFO1      : 400.126009 MHz

F2 - Processing parameters
SI         : 65536
SF         : 400.1260114 MHz
WDW        : EM
SSB        : 0
LB         : 0.00 Hz
GB         : 0
PC         : 2.00

ID NMR plot parameters
CX         : 22.80 cm
CY         : 15.00 cm
FIDF      : 8.000 ppm
F1         : 3401.11 Hz
F2P       : -16.000 ppm
F3        : -240.00 Hz
FREQ      : 0.336174 ppm/cm
SFO      : 150.94688 MHz
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER          : shana
NAME         : IES-4-255-21-050
EXPNO       : 1
PROCNO      : 1

F2 - Acquisition Parameters
Date_       : 20160320
Time       : 14.44
INSTRUM    : draco
PROBHD     : 5 mm QNP 82TP
PULPROG    : zgpg
TD         : 26240
SOLVENT    : CDCl3
NS         : 2
DS         : 4
SFO        : 640.256 Hz
FIDRES     : 0.250010 sec
AQ         : 1.995701 sec
RG         : 1149.4
SQ         : 78.000 usec
DE         : 4.50 usec
TE         : 298.2 K
SI         : 0.2500000 sec
MCHRG     : 0.0000000 sec
MCHRG     : 0.0100000 sec
F2        : 33.10 usec

===== CHANNEL f1 =====
NUC1       : 13C
P1         : 16.00 usec
PL1        : 0.00 dB
SFO1      : 125.762448 MHz
SFO2      : 125.762448 MHz
SFO3      : 125.762448 MHz
SFO4      : 125.762448 MHz
SFO5      : 125.762448 MHz
SFO6      : 125.762448 MHz
SFO7      : 125.762448 MHz
SFO8      : 125.762448 MHz
SFO9      : 125.762448 MHz
SFO10     : 125.762448 MHz
SFO11     : 125.762448 MHz
SFO12     : 125.762448 MHz
SFO13     : 125.762448 MHz
SFO14     : 125.762448 MHz
SFO15     : 125.762448 MHz
SFO16     : 125.762448 MHz
SFO17     : 125.762448 MHz
SFO18     : 125.762448 MHz
SFO19     : 125.762448 MHz
SFO20     : 125.762448 MHz
SFO21     : 125.762448 MHz
SFO22     : 125.762448 MHz
SFO23     : 125.762448 MHz
SFO24     : 125.762448 MHz
SFO25     : 125.762448 MHz
SFO26     : 125.762448 MHz
SFO27     : 125.762448 MHz
SFO28     : 125.762448 MHz
SFO29     : 125.762448 MHz
SFO30     : 125.762448 MHz
SFO31     : 125.762448 MHz
SFO32     : 125.762448 MHz
SFO33     : 125.762448 MHz
SFO34     : 125.762448 MHz
SFO35     : 125.762448 MHz
SFO36     : 125.762448 MHz
SFO37     : 125.762448 MHz
SFO38     : 125.762448 MHz
SFO39     : 125.762448 MHz
SFO40     : 125.762448 MHz
SFO41     : 125.762448 MHz
SFO42     : 125.762448 MHz
SFO43     : 125.762448 MHz
SFO44     : 125.762448 MHz
SFO45     : 125.762448 MHz
SFO46     : 125.762448 MHz
SFO47     : 125.762448 MHz
SFO48     : 125.762448 MHz
SFO49     : 125.762448 MHz
SFO50     : 125.762448 MHz
SFO51     : 125.762448 MHz
SFO52     : 125.762448 MHz
SFO53     : 125.762448 MHz
SFO54     : 125.762448 MHz
SFO55     : 125.762448 MHz
SFO56     : 125.762448 MHz
SFO57     : 125.762448 MHz
SFO58     : 125.762448 MHz
SFO59     : 125.762448 MHz
SFO60     : 125.762448 MHz
SFO61     : 125.762448 MHz
SFO62     : 125.762448 MHz
SFO63     : 125.762448 MHz
SFO64     : 125.762448 MHz
SFO65     : 125.762448 MHz
SFO66     : 125.762448 MHz
SFO67     : 125.762448 MHz
SFO68     : 125.762448 MHz
SFO69     : 125.762448 MHz
SFO70     : 125.762448 MHz
SFO71     : 125.762448 MHz
SFO72     : 125.762448 MHz
SFO73     : 125.762448 MHz
SFO74     : 125.762448 MHz
SFO75     : 125.762448 MHz
SFO76     : 125.762448 MHz
SFO77     : 125.762448 MHz
SFO78     : 125.762448 MHz
SFO79     : 125.762448 MHz
SFO80     : 125.762448 MHz
SFO81     : 125.762448 MHz
SFO82     : 125.762448 MHz
SFO83     : 125.762448 MHz
SFO84     : 125.762448 MHz
SFO85     : 125.762448 MHz
SFO86     : 125.762448 MHz
SFO87     : 125.762448 MHz
SFO88     : 125.762448 MHz
SFO89     : 125.762448 MHz
SFO90     : 125.762448 MHz
SFO91     : 125.762448 MHz
SFO92     : 125.762448 MHz
SFO93     : 125.762448 MHz
SFO94     : 125.762448 MHz
SFO95     : 125.762448 MHz
SFO96     : 125.762448 MHz
SFO97     : 125.762448 MHz
SFO98     : 125.762448 MHz
SFO99     : 125.762448 MHz
SFO100    : 125.762448 MHz

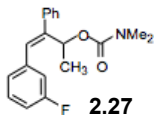
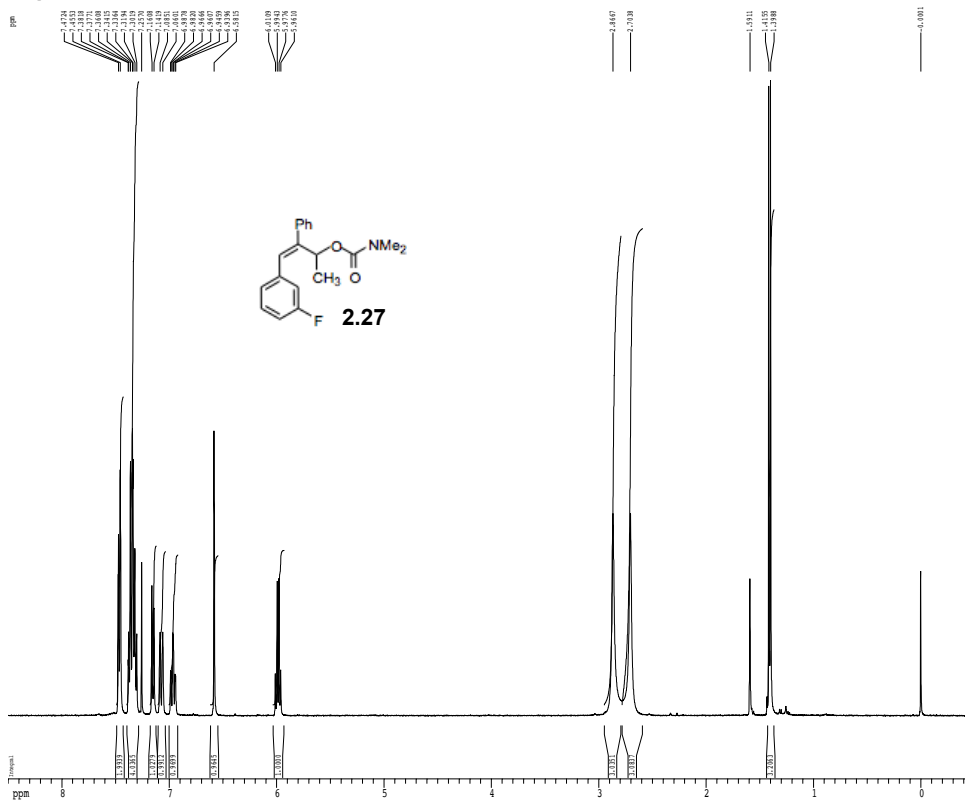
===== CHANNEL f2 =====
CPDPRG2   : waltz16
NUC2       : 1H
PCPD2     : 160.00 usec
PL2       : 0.00 dB
SFO2      : 500.136491 MHz
SFO3      : 500.136491 MHz

===== GRADIENT CHANNEL =====
GDMRG1    : 5198.100
DPMRG1    : 5198.100
GDMRG2    : 0.00 V
DPMRG2    : 0.00 V
GDMRG3    : 0.00 V
DPMRG3    : 0.00 V
GDMRG4    : 30.00 V
DPMRG4    : 30.00 V
GDMRG5    : 30.00 V
DPMRG5    : 30.00 V
GDMRG6    : 500.00 usec
DPMRG6    : 100.00 usec

F2 - Processing parameters
SI         : 125.762448 MHz
SF         : 125.762448 MHz
WDW        : EM
SSB        : 0
LB         : 0.00 Hz
GB         : 0
PC         : 2.00

ID NMR plot parameters
CX         : 22.80 cm
CY         : 15.00 cm
FIDF      : 160.000 ppm
F1         : 3444.47 Hz
F2P       : -16.000 ppm
F3        : -240.00 Hz
FREQ      : 0.336174 ppm/cm
SFO      : 124.4892 Hz/cm
    
```

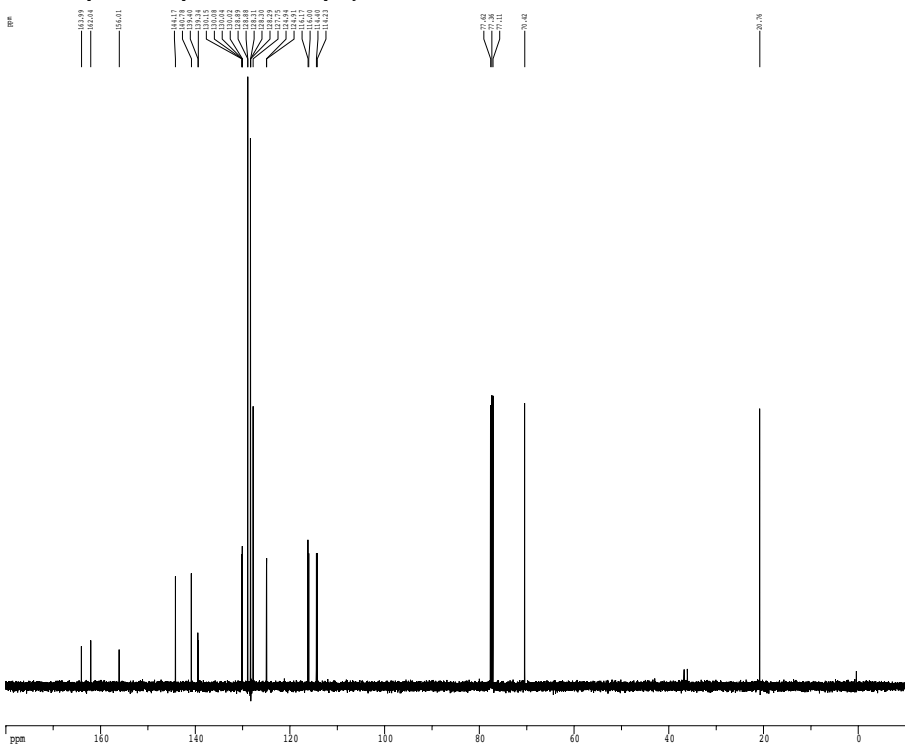

1H spectrum



Current Data Parameters
 USER thama
 NAME LHM-6-23-1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160223
 Time 13.40
 INSTRUM spect
 PROSD 5 mm QNP 817P
 PULPROG zgpg30
 TD 25640
 SFO 400.136225
 DS 4
 SW 4120.250 Hz
 FIDRES 0.205010 Hz
 AQ 1.9999700 sec
 RG 327.1
 DW 70.100 usec
 DE 6.50 usec
 TE 298.2 K
 D1 0.10000000 sec
 d11 0.00000000 sec
 d12 0.00000000 sec
 d13 0.00000000 sec
 d14 0.00000000 sec
 d15 0.00000000 sec
 d16 0.00000000 sec
 d17 0.00000000 sec
 d18 0.00000000 sec
 d19 0.00000000 sec
 d20 0.00000000 sec
 d21 0.00000000 sec
 d22 0.00000000 sec
 d23 0.00000000 sec
 d24 0.00000000 sec
 d25 0.00000000 sec
 d26 0.00000000 sec
 d27 0.00000000 sec
 d28 0.00000000 sec
 d29 0.00000000 sec
 d30 0.00000000 sec
 d31 0.00000000 sec
 d32 0.00000000 sec
 d33 0.00000000 sec
 d34 0.00000000 sec
 d35 0.00000000 sec
 d36 0.00000000 sec
 d37 0.00000000 sec
 d38 0.00000000 sec
 d39 0.00000000 sec
 d40 0.00000000 sec
 d41 0.00000000 sec
 d42 0.00000000 sec
 d43 0.00000000 sec
 d44 0.00000000 sec
 d45 0.00000000 sec
 d46 0.00000000 sec
 d47 0.00000000 sec
 d48 0.00000000 sec
 d49 0.00000000 sec
 d50 0.00000000 sec
 d51 0.00000000 sec
 d52 0.00000000 sec
 d53 0.00000000 sec
 d54 0.00000000 sec
 d55 0.00000000 sec
 d56 0.00000000 sec
 d57 0.00000000 sec
 d58 0.00000000 sec
 d59 0.00000000 sec
 d60 0.00000000 sec
 d61 0.00000000 sec
 d62 0.00000000 sec
 d63 0.00000000 sec
 d64 0.00000000 sec
 d65 0.00000000 sec
 d66 0.00000000 sec
 d67 0.00000000 sec
 d68 0.00000000 sec
 d69 0.00000000 sec
 d70 0.00000000 sec
 d71 0.00000000 sec
 d72 0.00000000 sec
 d73 0.00000000 sec
 d74 0.00000000 sec
 d75 0.00000000 sec
 d76 0.00000000 sec
 d77 0.00000000 sec
 d78 0.00000000 sec
 d79 0.00000000 sec
 d80 0.00000000 sec
 d81 0.00000000 sec
 d82 0.00000000 sec
 d83 0.00000000 sec
 d84 0.00000000 sec
 d85 0.00000000 sec
 d86 0.00000000 sec
 d87 0.00000000 sec
 d88 0.00000000 sec
 d89 0.00000000 sec
 d90 0.00000000 sec
 d91 0.00000000 sec
 d92 0.00000000 sec
 d93 0.00000000 sec
 d94 0.00000000 sec
 d95 0.00000000 sec
 d96 0.00000000 sec
 d97 0.00000000 sec
 d98 0.00000000 sec
 d99 0.00000000 sec
 d100 0.00000000 sec

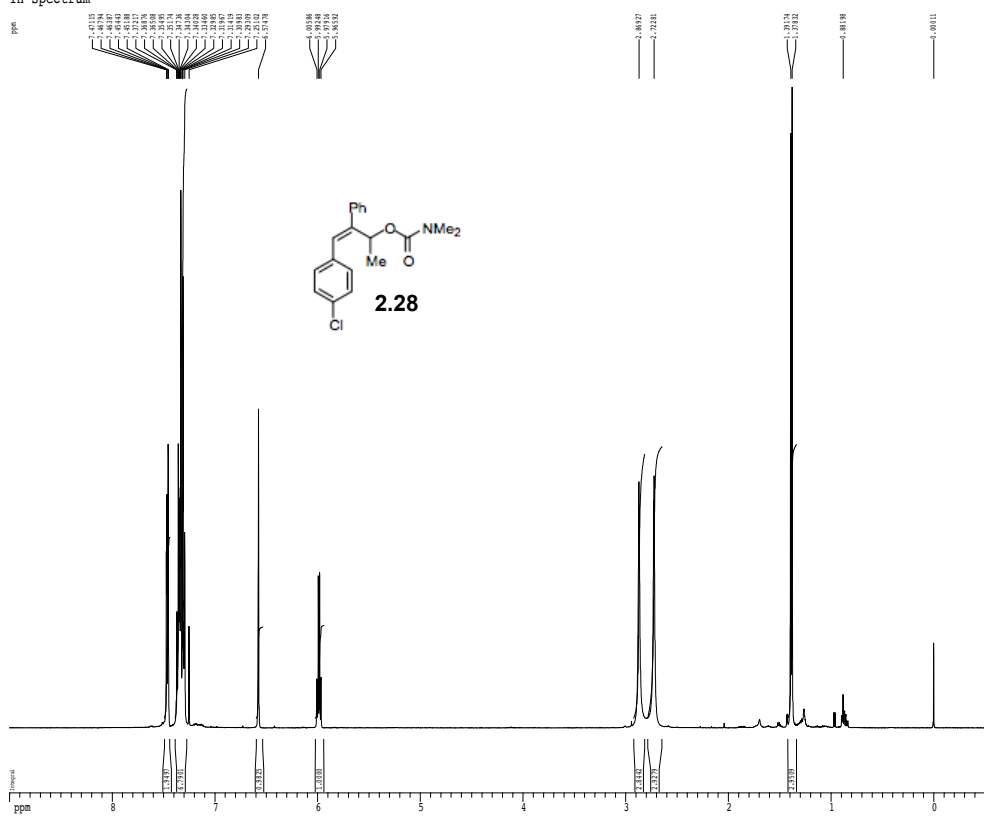
Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
 USER thama
 NAME LHM-6-23-1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160223
 Time 13.40
 INSTRUM spect
 PROSD 5 mm QNP 817P
 PULPROG zgpg30
 TD 25640
 SFO 400.136225
 DS 4
 SW 4120.250 Hz
 FIDRES 0.205010 Hz
 AQ 1.9999700 sec
 RG 327.1
 DW 70.100 usec
 DE 6.50 usec
 TE 298.2 K
 D1 0.10000000 sec
 d11 0.00000000 sec
 d12 0.00000000 sec
 d13 0.00000000 sec
 d14 0.00000000 sec
 d15 0.00000000 sec
 d16 0.00000000 sec
 d17 0.00000000 sec
 d18 0.00000000 sec
 d19 0.00000000 sec
 d20 0.00000000 sec
 d21 0.00000000 sec
 d22 0.00000000 sec
 d23 0.00000000 sec
 d24 0.00000000 sec
 d25 0.00000000 sec
 d26 0.00000000 sec
 d27 0.00000000 sec
 d28 0.00000000 sec
 d29 0.00000000 sec
 d30 0.00000000 sec
 d31 0.00000000 sec
 d32 0.00000000 sec
 d33 0.00000000 sec
 d34 0.00000000 sec
 d35 0.00000000 sec
 d36 0.00000000 sec
 d37 0.00000000 sec
 d38 0.00000000 sec
 d39 0.00000000 sec
 d40 0.00000000 sec
 d41 0.00000000 sec
 d42 0.00000000 sec
 d43 0.00000000 sec
 d44 0.00000000 sec
 d45 0.00000000 sec
 d46 0.00000000 sec
 d47 0.00000000 sec
 d48 0.00000000 sec
 d49 0.00000000 sec
 d50 0.00000000 sec
 d51 0.00000000 sec
 d52 0.00000000 sec
 d53 0.00000000 sec
 d54 0.00000000 sec
 d55 0.00000000 sec
 d56 0.00000000 sec
 d57 0.00000000 sec
 d58 0.00000000 sec
 d59 0.00000000 sec
 d60 0.00000000 sec
 d61 0.00000000 sec
 d62 0.00000000 sec
 d63 0.00000000 sec
 d64 0.00000000 sec
 d65 0.00000000 sec
 d66 0.00000000 sec
 d67 0.00000000 sec
 d68 0.00000000 sec
 d69 0.00000000 sec
 d70 0.00000000 sec
 d71 0.00000000 sec
 d72 0.00000000 sec
 d73 0.00000000 sec
 d74 0.00000000 sec
 d75 0.00000000 sec
 d76 0.00000000 sec
 d77 0.00000000 sec
 d78 0.00000000 sec
 d79 0.00000000 sec
 d80 0.00000000 sec
 d81 0.00000000 sec
 d82 0.00000000 sec
 d83 0.00000000 sec
 d84 0.00000000 sec
 d85 0.00000000 sec
 d86 0.00000000 sec
 d87 0.00000000 sec
 d88 0.00000000 sec
 d89 0.00000000 sec
 d90 0.00000000 sec
 d91 0.00000000 sec
 d92 0.00000000 sec
 d93 0.00000000 sec
 d94 0.00000000 sec
 d95 0.00000000 sec
 d96 0.00000000 sec
 d97 0.00000000 sec
 d98 0.00000000 sec
 d99 0.00000000 sec
 d100 0.00000000 sec

1H spectrum



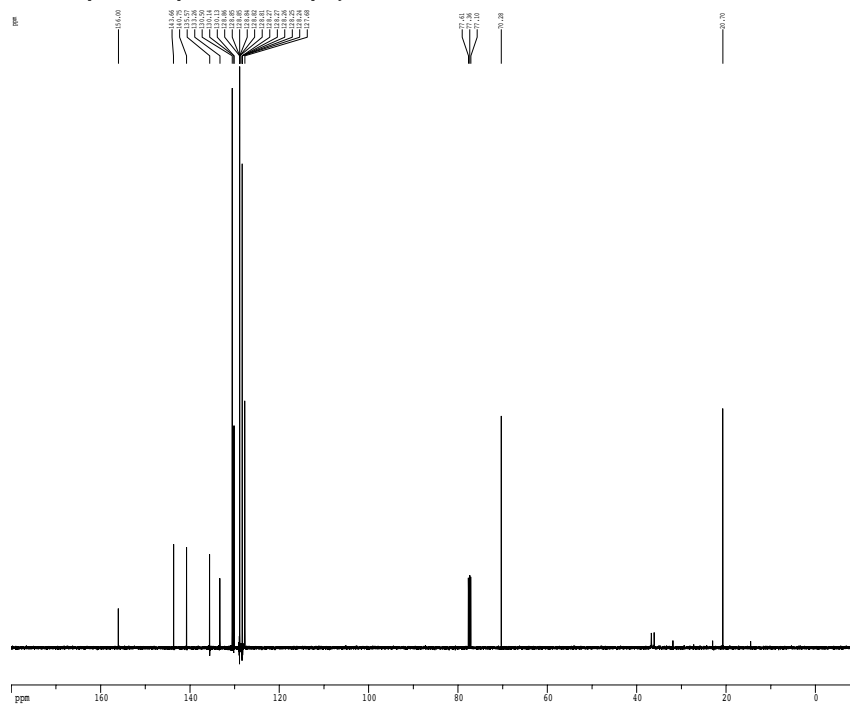
Current Data Parameters
 Date: 200412
 Name: NMR-19-157bhar
 ExpNO: 1
 PROCNO: 1

F2 - Acquisition Parameters
 Date_ : 200412
 Time : 14.51
 INSTRUM : cryo500
 PROBRW : 5 mm CPMG 1H-
 PULPROG : zgpg30
 TD : 65536
 SFO : 500.136385 MHz
 SOLVENT : CDCl3
 NS : 8
 DS : 2
 SWH : 8012.572 Hz
 FIDRES : 0.166677 Hz
 AQ : 2.3958877 sec
 RG : 5
 DB : 62.400 usac
 DE : 4.00 usac
 TE : 298.0 K
 D1 : 0.1000000 sec
 DELTAT : 0.0000000 sec
 NUC1 : 13C
 NU11 : 13C
 P1 : 7.50 usac
 PL1 : 1.60 dB
 SFO1 : 500.1258143 MHz

F2 - Processing parameters
 SI : 65536
 SF : 500.136385 MHz
 MDW : no
 CDR : 0
 LB : 0.00 Hz
 GB : 0
 PC : 4.00

ID NMR plot parameters
 CN : 22.00 cm
 CT : 15.00 cm
 FID : 8.000 ppm
 P1 : 4501.98 Hz
 F2P : -2.500 ppm
 FZ : -7501.11 Hz
 SFO : 0.41667 ppm/cm
 NUCN : 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



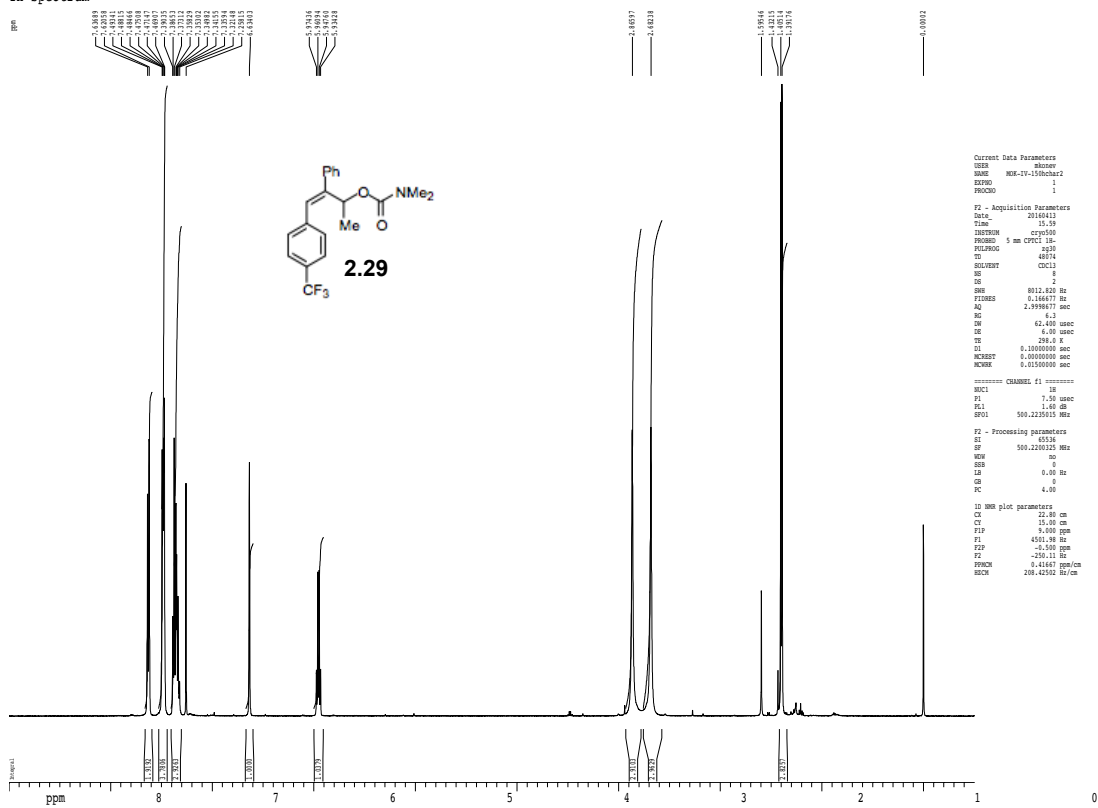
Current Data Parameters
 Date: 200412
 Name: NMR-19-157bhar
 ExpNO: 1
 PROCNO: 1

F2 - Acquisition Parameters
 Date: 200412
 Time: 14.54
 INSTRUM: cryo500
 PROBRW: 5 mm CPMG 1H-
 PULPROG: zgpg30decoupl-2
 TD: 65536
 SFO: 500.136385 MHz
 SOLVENT: CDCl3
 NS: 8
 DS: 2
 SWH: 50012.572 Hz
 FIDRES: 1.46288 Hz
 AQ: 1.08280 sec
 RG: 5
 DB: 14.400 usac
 DE: 4.00 usac
 TE: 298.0 K
 D1: 0.1000000 sec
 DELTAT: 0.0000000 sec
 NUC1: 13C
 NU11: 13C
 P1: 7.50 usac
 PL1: 1.60 dB
 SFO1: 500.1258143 MHz

F2 - Processing parameters
 SI: 65536
 SF: 125.7631925 MHz
 MDW: no
 CDR: 0
 LB: 0.00 Hz
 GB: 0
 PC: 2.40

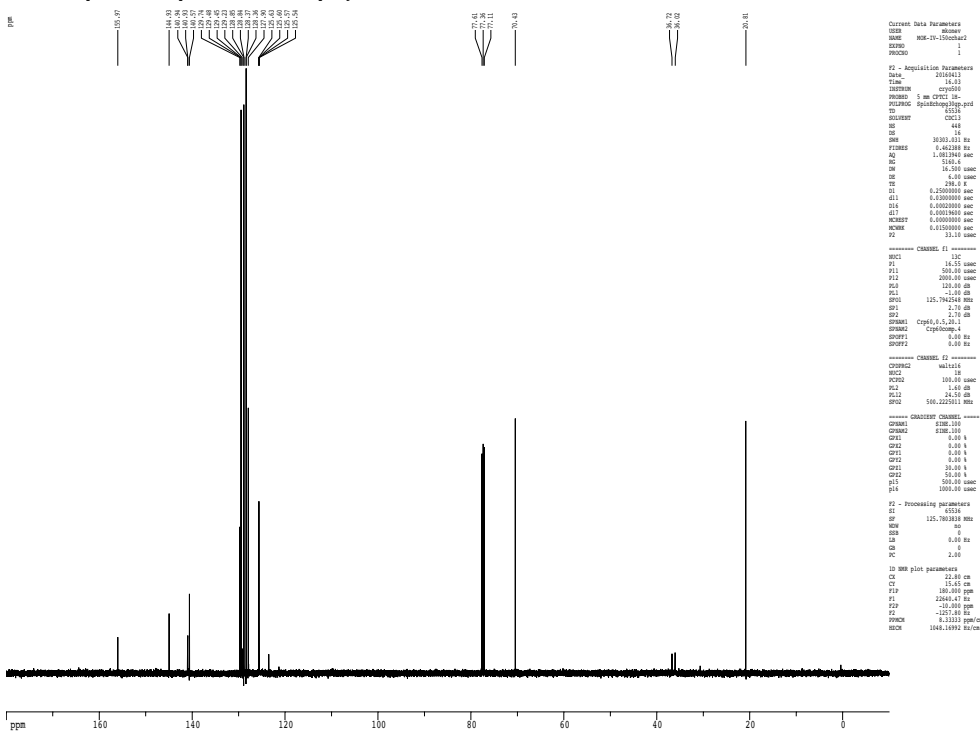
ID NMR plot parameters
 CN: 22.00 cm
 CT: 15.00 cm
 FID: 100.000 ppm
 P1: 2500.00 Hz
 F2P: -125.00 ppm
 FZ: -125.00 Hz
 SFO: 0.41667 ppm/cm
 NUCN: 1048.16992 Hz/cm

1H spectrum



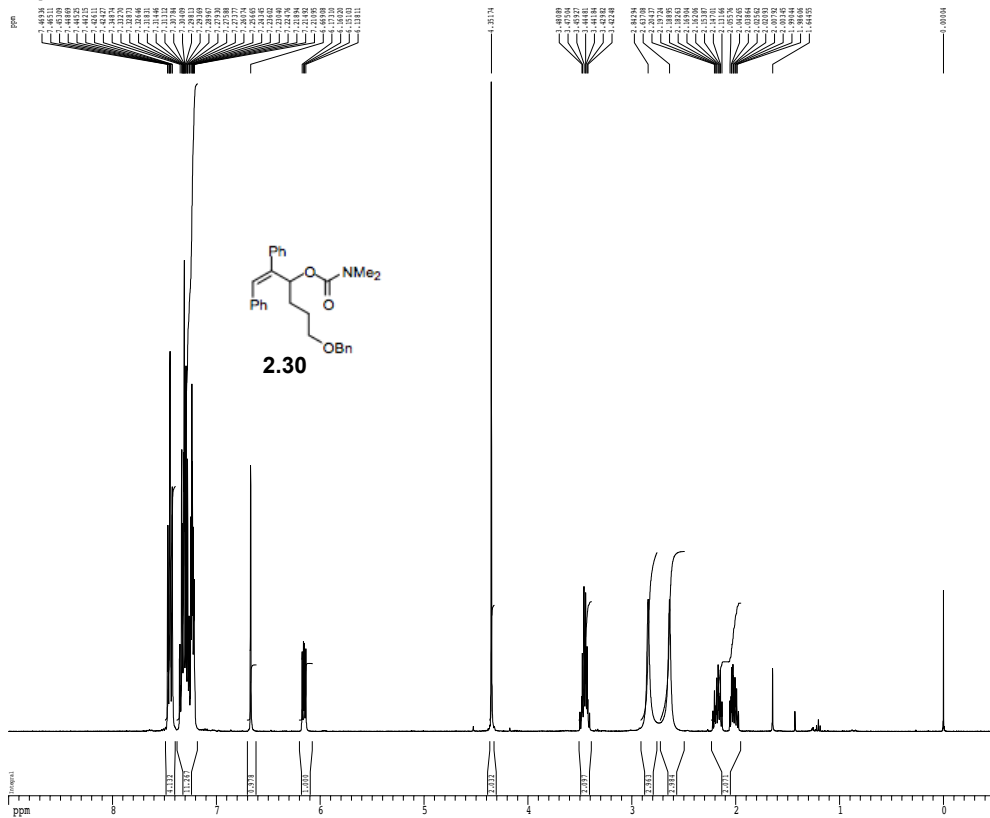
Current Data Parameters
 USER: mhseer
 NAME: MGE-TV-150char2
 EXPNO: 1
 PROCNO: 1
 F2 - Acquisition Parameters
 Date_: 20161011
 Time: 15:39
 INSTRUM: cryso50
 PROBMG: 5 mm CPPLC 1h-
 PULPROG: zgpg30
 TD: 65536
 SOLVENT: ccd13
 NS: 8
 DS: 2
 SFO: 801.220 Hz
 FIDRES: 0.168471 Hz
 AQ: 2.9398177 sec
 RG: 320
 SI: 62.400 umsec
 DE: 4.00 umsec
 TE: 298.2 K
 DI: 0.1000000 sec
 DELT: 0.0000000 sec
 MCHRG1: 0.0000000 sec
 MCHRG2: 0.0000000 sec
 CHANNEL F1: 150MHz
 MCH1: 1H
 P1: 7.50 umsec
 PL1: 2.00 dB
 SFO1: 500.235015 MHz
 F2 - Processing parameters
 SI: 65536
 SF: 500.235015 MHz
 DS: 8
 SSB: 0
 LB: 0.00 Hz
 GB: 0
 PC: 4.00
 ID: 150MHz plot parameters
 CX: 22.80 cm
 CY: 15.65 cm
 F1P: 9.000 ppm
 F2: 4501.98 Hz
 F3P: -2.500 ppm
 F4: -258.11 Hz
 FREQM: 6.45483 ppm/cm
 RSCN: 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
 USER: mhseer
 NAME: MGE-TV-150char2
 EXPNO: 1
 PROCNO: 1
 F2 - Acquisition Parameters
 Date_: 20161011
 Time: 16:02
 INSTRUM: cryso50
 PROBMG: 5 mm CPPLC 1h-
 PULPROG: zgpg30
 TD: 65536
 SOLVENT: ccd13
 NS: 8
 DS: 2
 SFO: 801.220 Hz
 FIDRES: 0.462388 Hz
 AQ: 1.0017614 sec
 RG: 320
 SI: 62.400 umsec
 DE: 4.00 umsec
 TE: 298.2 K
 DI: 0.1000000 sec
 DELT: 0.0000000 sec
 MCHRG1: 0.0000000 sec
 MCHRG2: 0.0000000 sec
 CHANNEL F1: 150MHz
 MCH1: 13C
 P1: 16.55 umsec
 PL1: 200.00 umsec
 PL2: 120.00 dB
 SFO1: 125.760548 MHz
 SFO2: 125.760548 MHz
 SF1: 2.70 dB
 SF2: 2.70 dB
 SPANM1: Cpplc, 5, 26, 1
 SPANM2: cpplcplcplc, 4
 SFOFF1: 0.00 Hz
 SFOFF2: 0.00 Hz
 CHANNEL F2: 500MHz
 MCH2: 13C
 SFO2: 125.760548 MHz
 MCH1: 150.00 umsec
 PC1: 1.40 dB
 PL2: 26.50 dB
 SFO3: 500.235015 MHz
 CHANNEL F3: 500MHz
 MCH3: 13C
 SFO3: 125.760548 MHz
 MCH2: 500.00 umsec
 MCH3: 500.00 umsec
 ID: 500MHz plot parameters
 CX: 22.80 cm
 CY: 15.65 cm
 F1P: 100.000 ppm
 F2: 24963.47 Hz
 F3P: -20.000 ppm
 F4: -1227.80 Hz
 FREQM: 8.33333 ppm/cm
 RSCN: 1648.16992 Hz/cm

1H spectrum



```

Current Data Parameters
USER          MROVER
NAME         MW-IV-150char
EXPNO        1
PROCNO       1

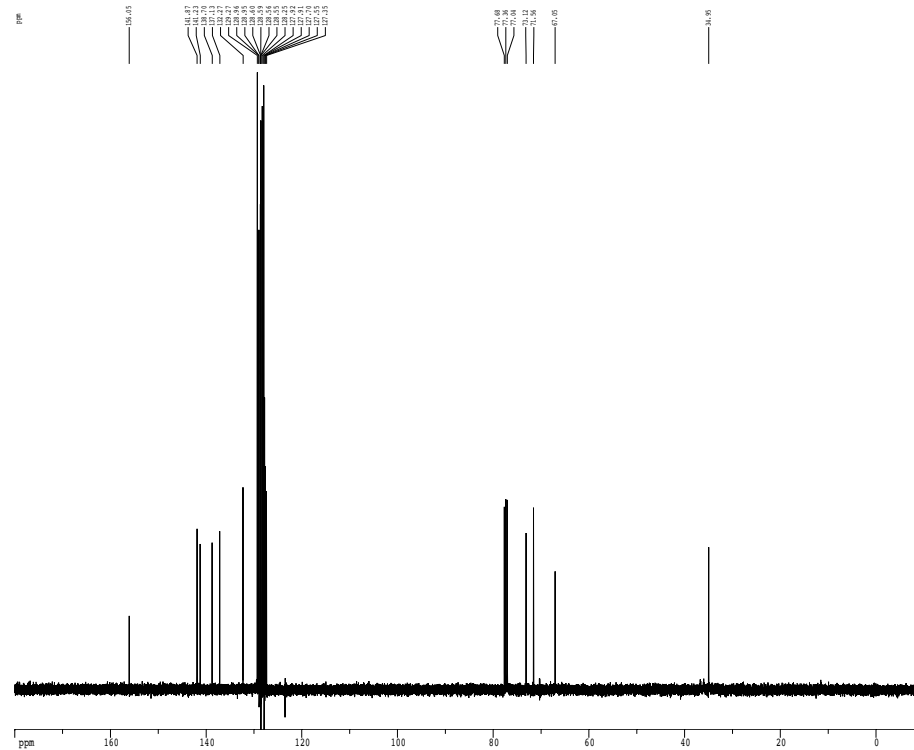
F2 - Acquisition Parameters
Date_        20160410
Time         17.27
INSTRUM     dr400
PROBHD      5 mm QNP 877 P
PULPROG     zgpg
TD           65536
SOLVENT     CDCl3
NS           4
DS           4
SWH          6419.256 Hz
F2RES        0.166879 Hz
AQ           2.9992299 sec
RG           101.6
DM           78.000 usec
DE           4.50 usec
TE           298.2 K
SI           0.1000000 sec
RG1          0.0000000 sec
RG2          0.0000000 sec
RG3          0.0100000 sec
===== CHANNEL f1 =====
NUC1         13C
P1           12.00 usec
PL1          0.00 dB
SFO1         400.128009 MHz

F2 - Processing parameters
SI           65536
SF           400.1280399 MHz
WDW          EM
SSB          0
LB           0.60 Hz
GB           0
PC           1.00

1D NMR plot parameters
CX           22.80 cm
CY           15.00 cm
FIDP         9.000 ppm
F1           1601.17 Hz
F2P          -200.06 ppm
F2           -200.06 Hz
FREQM        8.42487 ppm/cm
SFO          100.626019 MHz

```

13C spectrum with 1H decoupling



```

Current Data Parameters
USER          MROVER
NAME         MW-IV-150char
EXPNO        1
PROCNO       1

F2 - Acquisition Parameters
Date_        20160410
Time         17.30
INSTRUM     dr400
PROBHD      5 mm QNP 877 P
PULPROG     zgpg
TD           65536
SOLVENT     CDCl3
NS           4
DS           4
SWH          24154.290 Hz
F2RES        0.36979 Hz
AQ           1.1666452 sec
RG           101.6
DM           78.700 usec
DE           20.39 usec
TE           298.2 K
SI           0.1000000 sec
RG1          0.0000000 sec
RG2          0.0000000 sec
RG3          0.0100000 sec
===== CHANNEL f1 =====
NUC1         13C
P1           7.75 usec
PL1          -1.00 dB
SFO1         100.627944 MHz

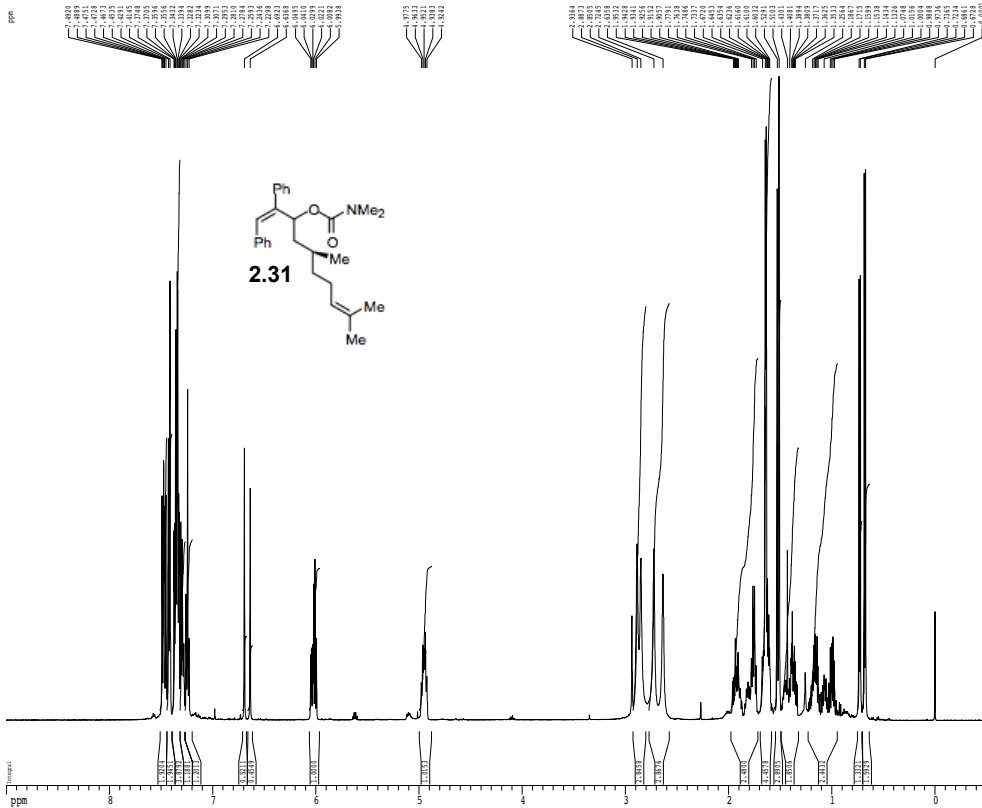
===== CHANNEL f2 =====
CPROG2      Mvpr4
NUC2         1H
P2           90.00 usec
PL2          0.00 dB
PL12         17.70 dB
SFO2         400.128009 MHz

F2 - Processing parameters
SI           65536
SF           100.6127433 MHz
WDW          EM
SSB          0
LB           0.60 Hz
GB           0
PC           1.00

1D NMR plot parameters
CX           22.80 cm
CY           15.00 cm
FIDP         180.000 ppm
F1           1610.19 Hz
F2P          -13.000 ppm
F2           -13.000 Hz
FREQM        8.33333 ppm/cm
SFO          638.43951 Hz/cm

```

1H spectrum



```

Current Data Parameters
USER          rhocver
NAME          MW-17-15304ar
EXPNO        1
PROCNO       1

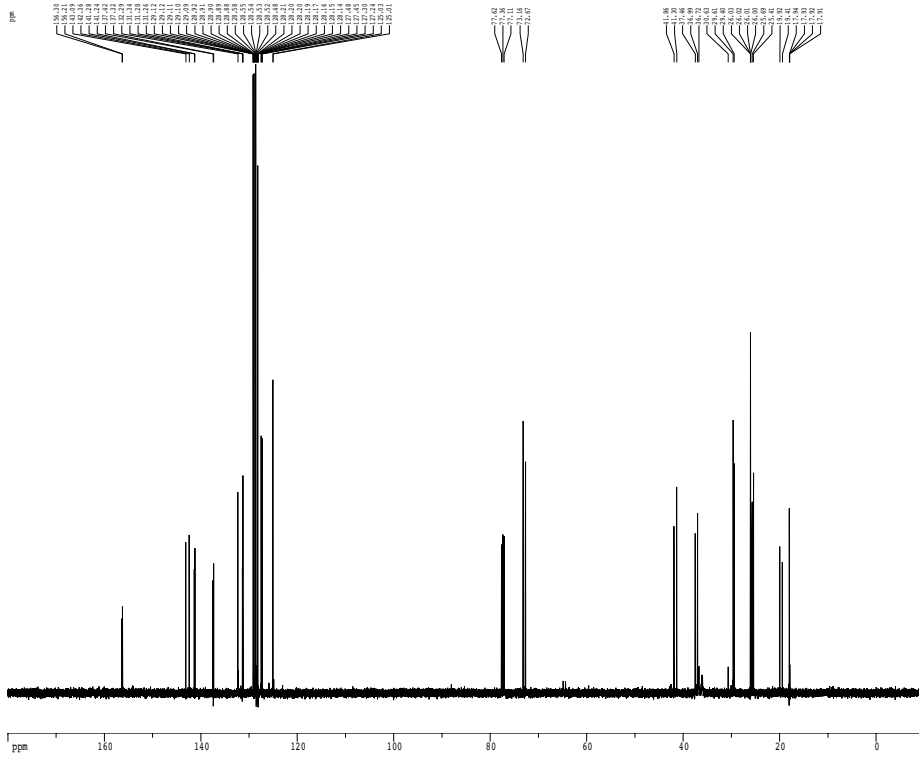
F2 - Acquisition Parameters
Date_        20160408
TIME        16.39
INSTRUM     cryo500
PROBHD      5 mm CPYC1 1H-
PULPROG     zgpg30
TD          65536
FIDRES      0.0000000
AQ          0.0000000
RG          655.36
SFO1        500.225015 MHz
SF          500.225015 MHz
WDW          EM
SSB          0
LB          0.00 Hz
GB          0
PC          4.00

===== CHANNEL f1 =====
NUC1         13C
P1           7.50 usec
PL1          0.00 dB
SFO1        500.225015 MHz

F1 - Processing parameters
SI           65536
SF          500.225015 MHz
WDW          EM
SSB          0
LB          0.00 Hz
GB          0
PC          4.00

ID 90M plot parameters
CX           22.80 cm
CY           15.00 cm
FIDP         9.000 ppm
FI           4501.50 Hz
FZP          -250.11 ppm
F2           -250.11 Hz
FREQM        8.442487 MHz/cm
HCHX        208.42162 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER          rhocver
NAME          MW-17-15304ar
EXPNO        1
PROCNO       1

F2 - Acquisition Parameters
Date_        20160408
TIME        16.43
INSTRUM     cryo500
PROBHD      5 mm CPYC1 1H-
PULPROG     zgpg30
TD          65536
FIDRES      0.0000000
AQ          0.0000000
RG          655.36
SFO1        500.225015 MHz
SF          500.225015 MHz
WDW          EM
SSB          0
LB          0.00 Hz
GB          0
PC          4.00

===== CHANNEL f1 =====
NUC1         13C
P1           16.35 usec
PL1          0.00 dB
PL2          200.00 usec
PL3          100.00 dB
PL4          -1.00 dB
SFO1        125.760248 MHz
SF1         125.760248 MHz
WDW          EM
SSB          0
LB          0.00 Hz
GB          0
PC          2.00

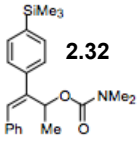
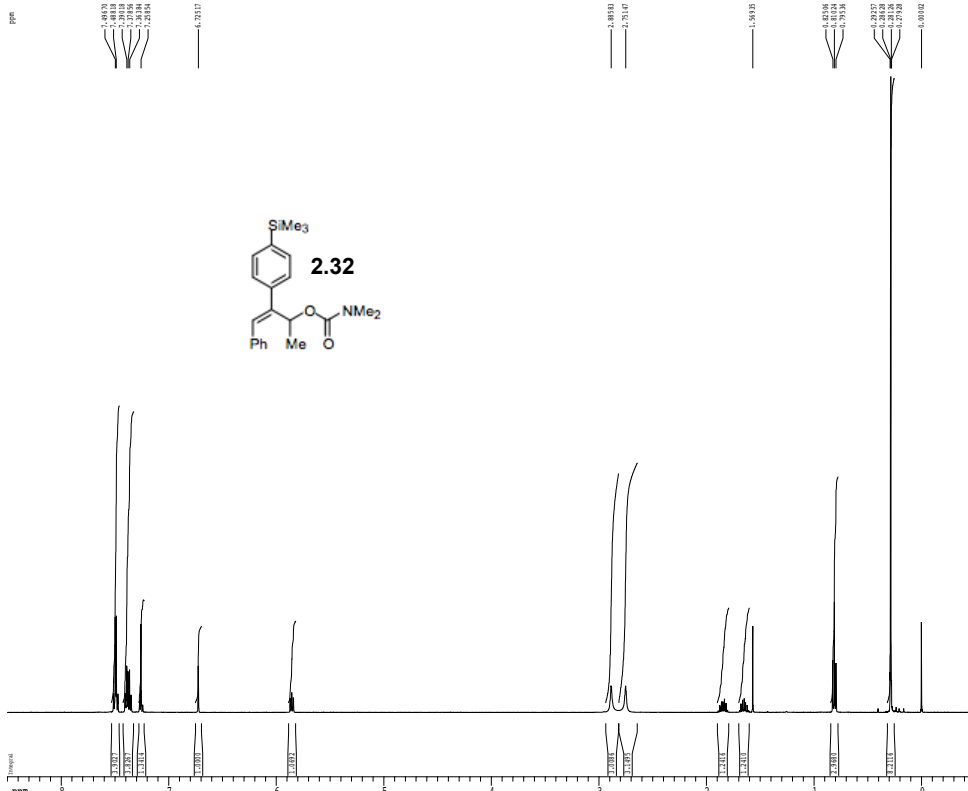
===== CHANNEL f2 =====
CPROG2      waltz16
NUC2         13C
P2           100.00 usec
PL2          0.00 dB
PL3          14.10 dB
SFO2        500.225015 MHz

===== CHANNEL CHANCE =====
CPHASE      0.00 usec
CPHASE2     0.00 usec
CPHASE3     0.00 usec
CPHASE4     0.00 usec
CPHASE5     0.00 usec
CPHASE6     0.00 usec
CPHASE7     0.00 usec
CPHASE8     0.00 usec
CPHASE9     0.00 usec
CPHASE10    0.00 usec
CPHASE11    0.00 usec
CPHASE12    0.00 usec
CPHASE13    0.00 usec
CPHASE14    0.00 usec
CPHASE15    0.00 usec
CPHASE16    0.00 usec
CPHASE17    0.00 usec
CPHASE18    0.00 usec
CPHASE19    0.00 usec
CPHASE20    0.00 usec
CPHASE21    0.00 usec
CPHASE22    0.00 usec
CPHASE23    0.00 usec
CPHASE24    0.00 usec
CPHASE25    0.00 usec
CPHASE26    0.00 usec
CPHASE27    0.00 usec
CPHASE28    0.00 usec
CPHASE29    0.00 usec
CPHASE30    0.00 usec
CPHASE31    0.00 usec
CPHASE32    0.00 usec
CPHASE33    0.00 usec
CPHASE34    0.00 usec
CPHASE35    0.00 usec
CPHASE36    0.00 usec
CPHASE37    0.00 usec
CPHASE38    0.00 usec
CPHASE39    0.00 usec
CPHASE40    0.00 usec
CPHASE41    0.00 usec
CPHASE42    0.00 usec
CPHASE43    0.00 usec
CPHASE44    0.00 usec
CPHASE45    0.00 usec
CPHASE46    0.00 usec
CPHASE47    0.00 usec
CPHASE48    0.00 usec
CPHASE49    0.00 usec
CPHASE50    0.00 usec
CPHASE51    0.00 usec
CPHASE52    0.00 usec
CPHASE53    0.00 usec
CPHASE54    0.00 usec
CPHASE55    0.00 usec
CPHASE56    0.00 usec
CPHASE57    0.00 usec
CPHASE58    0.00 usec
CPHASE59    0.00 usec
CPHASE60    0.00 usec
CPHASE61    0.00 usec
CPHASE62    0.00 usec
CPHASE63    0.00 usec
CPHASE64    0.00 usec
CPHASE65    0.00 usec
CPHASE66    0.00 usec
CPHASE67    0.00 usec
CPHASE68    0.00 usec
CPHASE69    0.00 usec
CPHASE70    0.00 usec
CPHASE71    0.00 usec
CPHASE72    0.00 usec
CPHASE73    0.00 usec
CPHASE74    0.00 usec
CPHASE75    0.00 usec
CPHASE76    0.00 usec
CPHASE77    0.00 usec
CPHASE78    0.00 usec
CPHASE79    0.00 usec
CPHASE80    0.00 usec
CPHASE81    0.00 usec
CPHASE82    0.00 usec
CPHASE83    0.00 usec
CPHASE84    0.00 usec
CPHASE85    0.00 usec
CPHASE86    0.00 usec
CPHASE87    0.00 usec
CPHASE88    0.00 usec
CPHASE89    0.00 usec
CPHASE90    0.00 usec
CPHASE91    0.00 usec
CPHASE92    0.00 usec
CPHASE93    0.00 usec
CPHASE94    0.00 usec
CPHASE95    0.00 usec
CPHASE96    0.00 usec
CPHASE97    0.00 usec
CPHASE98    0.00 usec
CPHASE99    0.00 usec
CPHASE100   0.00 usec

F1 - Processing parameters
SI           65536
SF          125.760248 MHz
WDW          EM
SSB          0
LB          0.00 Hz
GB          0
PC          2.00

ID 90M plot parameters
CX           22.80 cm
CY           15.00 cm
FIDP         9.000 ppm
FI           2246.47 Hz
FZP          -125.760248 ppm
F2           -125.760248 Hz
FREQM        8.442487 MHz/cm
HCHX        1648.16992 Hz/cm
    
```


1H spectrum



Current Data Parameters
 USER: sboner
 NAME: 128-4-2362car
 EXPNO: 1
 PROCNO: 1

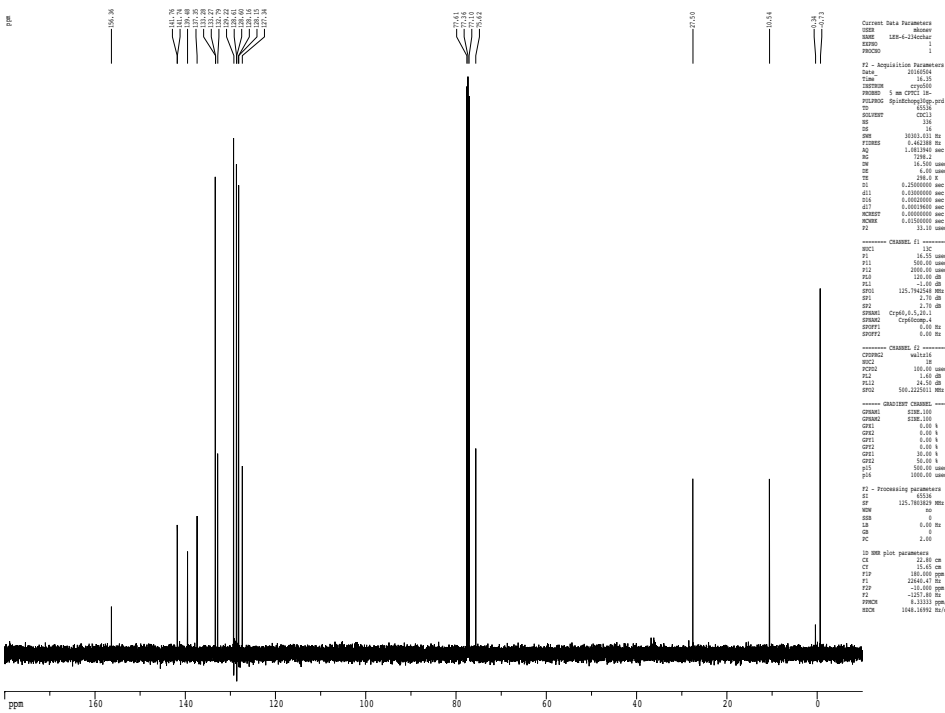
F2 - Acquisition Parameters
 Date_: 20160504
 TIME: 14.22
 INSTRUM: cryo500
 PROBHD: 5 mm CPMG1 1H-1
 PULPROG: zgpg30
 TD: 32768
 SFOF2: 500.1362773 MHz
 ACQRES: 6.250000 Hz
 AQ: 1.9998451 sec
 RG: 7.1
 CW: 62.400 usec
 DE: 6.00 usec
 TE: 298.2 K
 SI: 0.1000000 sec
 MCHWST: 0.0000000 sec
 MCHWEL: 0.0100000 sec

===== CHANNEL f1 =====
 NUC1: 1H
 P1: 7.50 usec
 PL1: 1.60 dB
 SFO1: 500.1362773 MHz

F2 - Processing parameters
 SI: 500.1362773 MHz
 SF: 500.1362773 MHz
 WDW: 0
 SSB: 0
 LB: 0.00 Hz
 GB: 0
 PC: 4.00

1D 1H NMR parameters
 CX: 22.80 cm
 CY: 15.00 cm
 FIP: 0.500 ppm
 F1: 4251.87 Hz
 F2: -0.500 ppm
 FZ: +256.21 Hz
 FWHM: 0.28474 ppm/cm
 SSCX: 197.45528 Hz/cm

2-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
 USER: sboner
 NAME: 128-4-2362car
 EXPNO: 1
 PROCNO: 1

F2 - Acquisition Parameters
 Date_: 20160504
 TIME: 14.35
 INSTRUM: cryo500
 PROBHD: 5 mm CP131 1H-13
 PULPROG: zgpg30
 TD: 65536
 SFOF2: 125.7603500 MHz
 ACQRES: 6.250000 Hz
 AQ: 1.9998451 sec
 RG: 7.1
 CW: 16.500 usec
 DE: 6.00 usec
 TE: 298.2 K
 SI: 0.2500000 sec
 MCHWST: 0.0000000 sec
 MCHWEL: 0.0000000 sec
 MCHWEL: 0.0100000 sec
 PC: 4.00

===== CHANNEL f1 =====
 NUC1: 13C
 P1: 16.50 usec
 PL1: 1.60 dB
 SFO1: 125.7603500 MHz

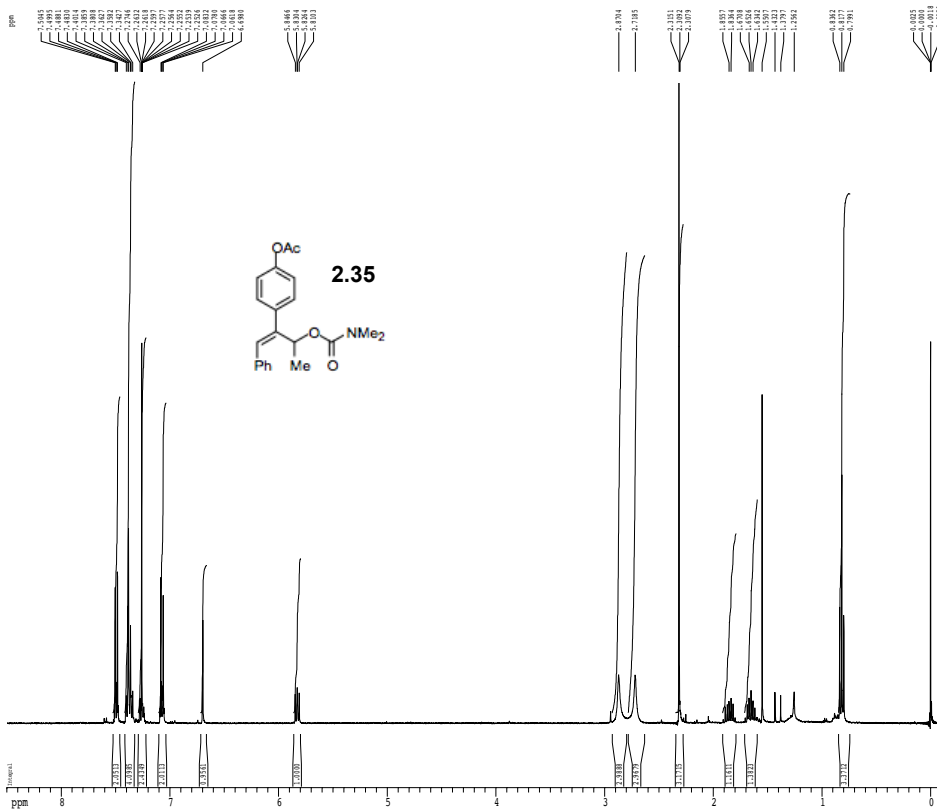
===== CHANNEL f2 =====
 NUC2: 1H
 P2: 7.50 usec
 PL2: 1.60 dB
 SFO2: 500.1362773 MHz

===== GRABBER CHANNEL =====
 GRAB1: 5188.100
 GRAB2: 5188.100
 GRF1: 0.00 A
 GRF2: 0.00 A
 GRF3: 0.00 A
 GRF4: 0.00 A
 GRF5: 0.00 A
 GRF6: 0.00 A
 GRF7: 0.00 A
 GRF8: 0.00 A
 GRF9: 0.00 A
 GRF10: 0.00 A

F2 - Processing parameters
 SI: 125.7603500 MHz
 SF: 125.7603500 MHz
 WDW: 0
 SSB: 0
 LB: 0.00 Hz
 GB: 0
 PC: 1.00

1D 13C NMR parameters
 CX: 22.80 cm
 CY: 15.00 cm
 FIP: 0.000 ppm
 F1: 2646.07 Hz
 F2: -0.500 ppm
 FZ: +256.21 Hz
 FWHM: 0.13132 ppm/cm
 SSCX: 1048.16992 Hz/cm

1H spectrum



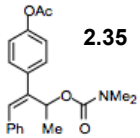
Current Data Parameters
USER lhamia
NAME IER-4-214-c3-113
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20160310
Time 19.10
INSTRUM dr400
PROBHD 5 mm QNP 1H/1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
DS 0
DE 0
SF 600.1326 MHz
F2RES 0.250010 Hz
AQ 1.999700 sec
RG 720.1
WE 4.00 usec
DE 4.00 usec
TE 298.0 K
D1 0.1000000 sec
MCHSEF 0.0000000 sec
MCHRX 0.0100000 sec

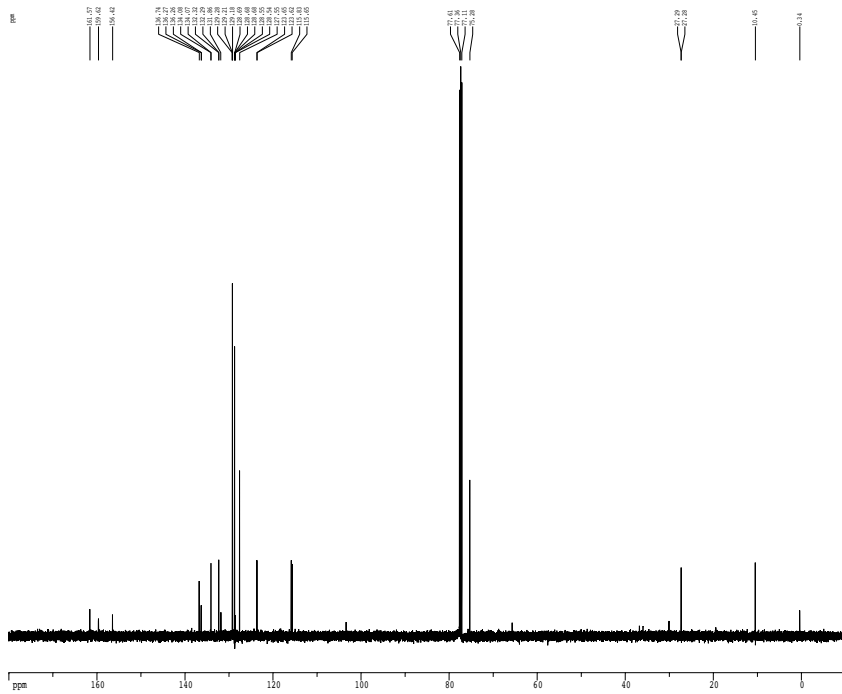
----- CHANNEL f1 -----
NUC1 1H
P1 12.00 usec
PL1 0.00 dB
SFO1 400.129999 MHz

F2 - Processing parameters
SI 65536
SF 400.126610 MHz
WDW EM
SSB 0
L8 0.00 Hz
GB 0
PC 2.00

ID HMR plot parameters
CX 22.80 cm
CY 12.00 cm
P1P 8.500 ppm
P1 1840.110 Hz
P2P -5.500 ppm
P2 -298.07 Hz
PPHWC 0.194714 ppm/cm
HSCX 157.94658 Hz/cm



Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
USER lhamia
NAME IER-4-225-111
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20160310
Time 19.10
INSTRUM cryo-500
PROBHD 5 mm QNP 1H/13
PULPROG zgpg30sp1
TD 65536
SOLVENT CDCl3
DS 0
DE 0
SF 125.761320 MHz
F2RES 0.250010 Hz
AQ 1.661300 sec
RG 300.0
WE 4.00 usec
DE 4.00 usec
TE 298.0 K
D1 0.2000000 sec
d11 0.0000000 sec
d12 0.0000000 sec
d13 0.0000000 sec
MCHSEF 0.0000000 sec
MCHRX 0.0100000 sec
PC 1.10 usec

----- CHANNEL f1 -----
NUC1 13C
P1 16.00 usec
PL1 0.00 dB
PL2 2000.00 usec
PL3 12.00 dB
PL4 -1.00 dB
SFO1 125.761324 MHz
SFO2 101.626126 MHz
SFO3 2.70 MHz
SFO4 100.626126 MHz
SFO5 Cyp6-A-1-11.1
SFO6C 9.70 Hz
SFO7 0.00 Hz
SFO8 0.00 Hz

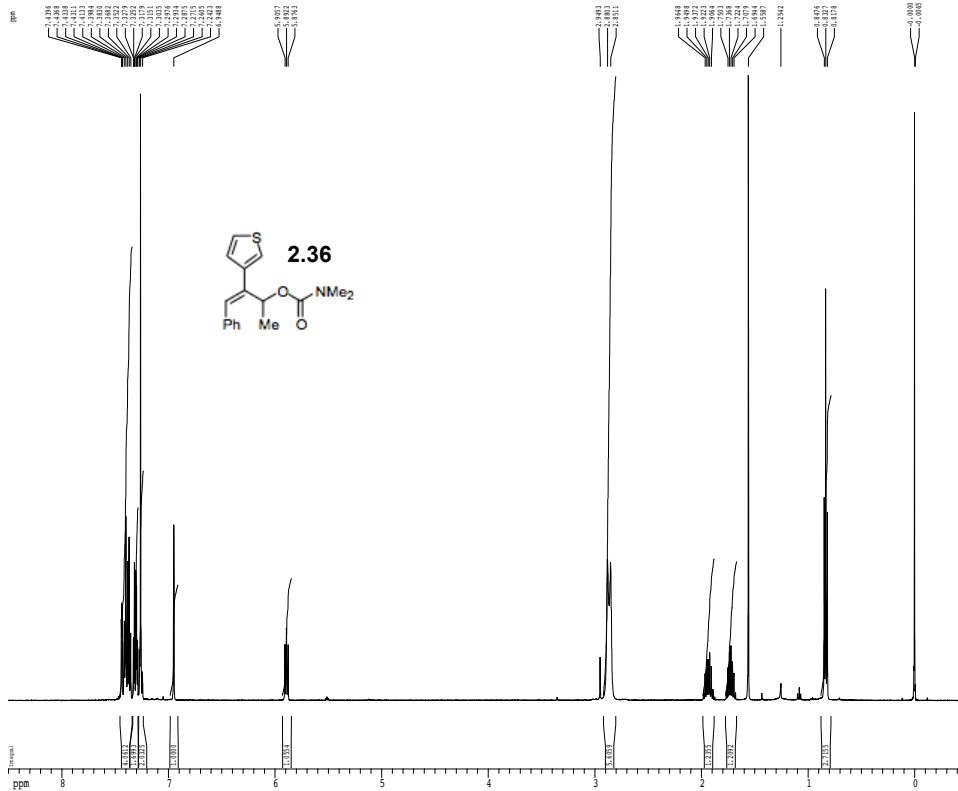
----- CHANNEL f2 -----
CPDPRG2 waltz16
NUC2 1H
P2 100.00 usec
PL2 1.00 dB
PL3 24.00 dB
SFO2 500.225101 MHz

----- CHANNEL CHANNEL -----
CHANI1 6786.100
CHANI2 8786.100 V
CHAM1 0.00 V
CHAM2 0.00 V
CHM1 0.00 V
CHM2 0.00 V
CHM3 50.00 Hz
CHM4 50.00 usec
PC 100.00 usec
P1 100.00 usec

F2 - Processing parameters
SI 65536
SF 125.761324 MHz
WDW EM
SSB 0
L8 0.00 Hz
GB 0
PC 1.00

ID HMR plot parameters
CX 22.80 cm
CY 12.00 cm
P1P 1840.110 ppm
P1 2160.47 Hz
P2P -15.500 ppm
P2 -201.80 Hz
PPHWC 4.12120 ppm/cm
HSCX 164.16992 Hz/cm

1H spectrum



```
Current Data Parameters
=====
USER      Mowse
NAME      L84-4-218char2
EXPNO     1
PROCNO    1

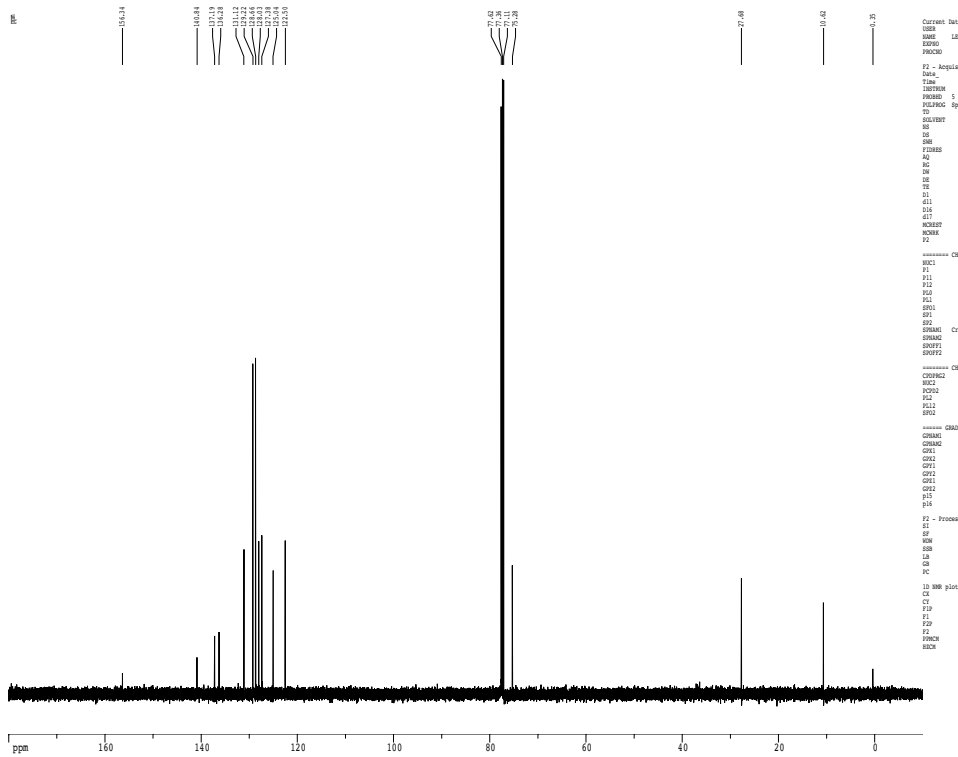
F2 - Acquisition Parameters
=====
Date_     20160505
Time      16.32
INSTRUM   cryo000
PROBHD    5 mm CPY 1H-
PULPROG   zgpg
TD         32768
SOLVENT   CDCl3
NS         8
DS         8
SWH        8012.820 Hz
FIDRES    0.250000 Hz
AQ         1.1998451 sec
RG         9
WDW         42.450 usec
DE         6.00 usec
TE         298.2 K
D1         0.1000000 sec
MCHOP     0.0000000 sec
MCMX      0.0150000 sec

===== CHANNEL f1 =====
NUC1      1H
P1         7.50 usec
PL1       0.00 dB
SFO1      500.2250151 MHz

F2 - Processing parameters
=====
SI         6554
SF         500.2250151 MHz
WDW        0
SSB        0
LB         0.00 Hz
GB         0
PC         4.00

LV 90H plot parameters
=====
CX         22.80 cm
CY         15.00 cm
FZ         8.200 ppm
P1         4251.87 Hz
P2         -12.500 ppm
P3         -256.11 Hz
PRNCH      4.13133 ppm/cm
RICHN      197.4528 Hz/cm
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```
Current Data Parameters
=====
USER      Mowse
NAME      L84-4-218char2
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
=====
Date_     20160505
Time      16.47
INSTRUM   cryo000
PROBHD    5 mm CPY 1H-
PULPROG   zgpg
TD         32768
SOLVENT   CDCl3
NS         8
DS         8
SWH        30363.621 Hz
FIDRES    0.462388 Hz
AQ         1.0813940 sec
RG         13520.1
WDW         6.200 usec
DE         16.500 usec
TE         298.2 K
D1         0.2500000 sec
d11        0.0200000 sec
D15        0.0200000 sec
d151       0.0010000 sec
MCHOP     0.0000000 sec
MCMX      0.0150000 sec
F2         31.10 usec

===== CHANNEL f1 =====
NUC1      13C
P1         16.35 usec
PL1       0.00 dB
PL2       2000.00 usec
PL3       130.00 dB
PC1       -1.00 dB
SFO1      125.7614264 MHz
SP1       2.70 dB
SP2       2.70 dB
SFO2      Cp40,0,5,26,1
SFO3      Cp40,0,5,26,1
SFO4      Cp40,0,5,26,1
SFO5      Cp40,0,5,26,1
SFO6      Cp40,0,5,26,1
SFO7      0.00 Hz
SFO8      0.00 Hz

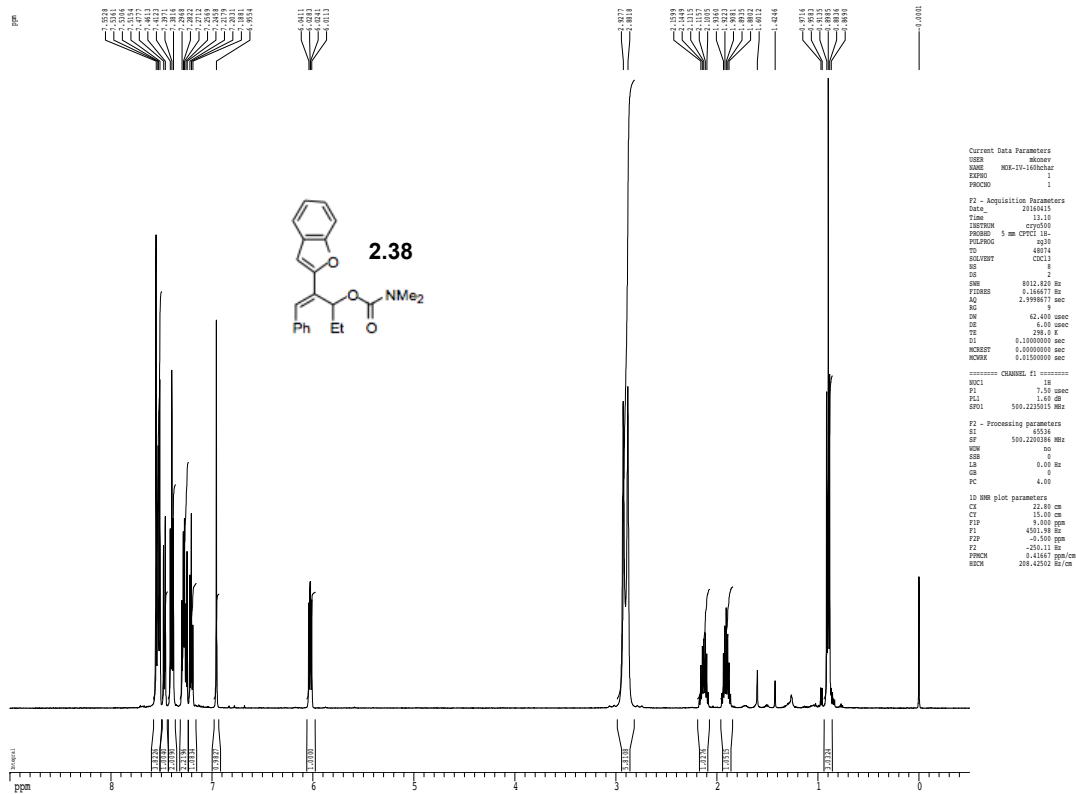
===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     100.00 usec
PL2       1.00 dB
PL3       34.16 dB
SFO2      500.2250151 MHz

===== CHANNEL CHANNEL =====
CHN1      128H.150
CHN2      819H.150
CHN3      0.00 V
CHN4      0.00 V
CHN5      0.00 V
CHN6      0.00 V
CHN7      30.00 V
CHN8      0.00 V
CHN9      500.00 usec
P30       1300.00 usec

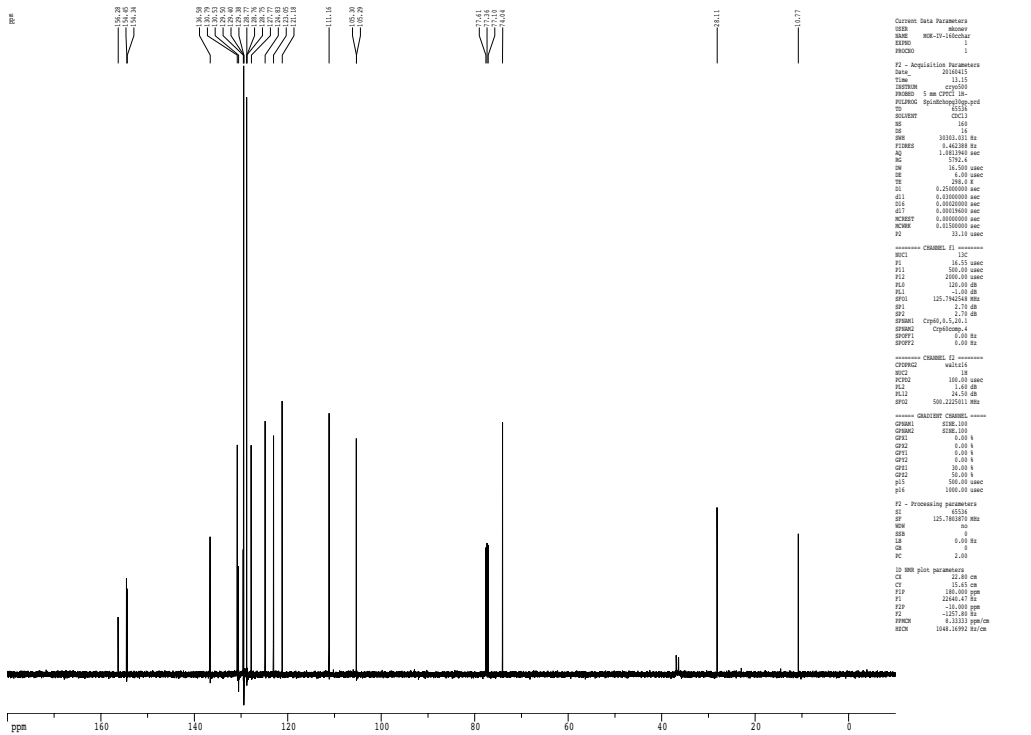
F2 - Processing parameters
=====
SI         6554
SF         125.7614264 MHz
WDW        0
SSB        0
LB         0.00 Hz
GB         0
PC         2.00

LV 90H plot parameters
=====
CX         22.80 cm
CY         15.00 cm
FZ         160.000 ppm
P1         22440.47 Hz
P2         -12.500 ppm
P3         -12.500 ppm
PRNCH      4.13133 ppm/cm
RICHN      104.16992 Hz/cm
```

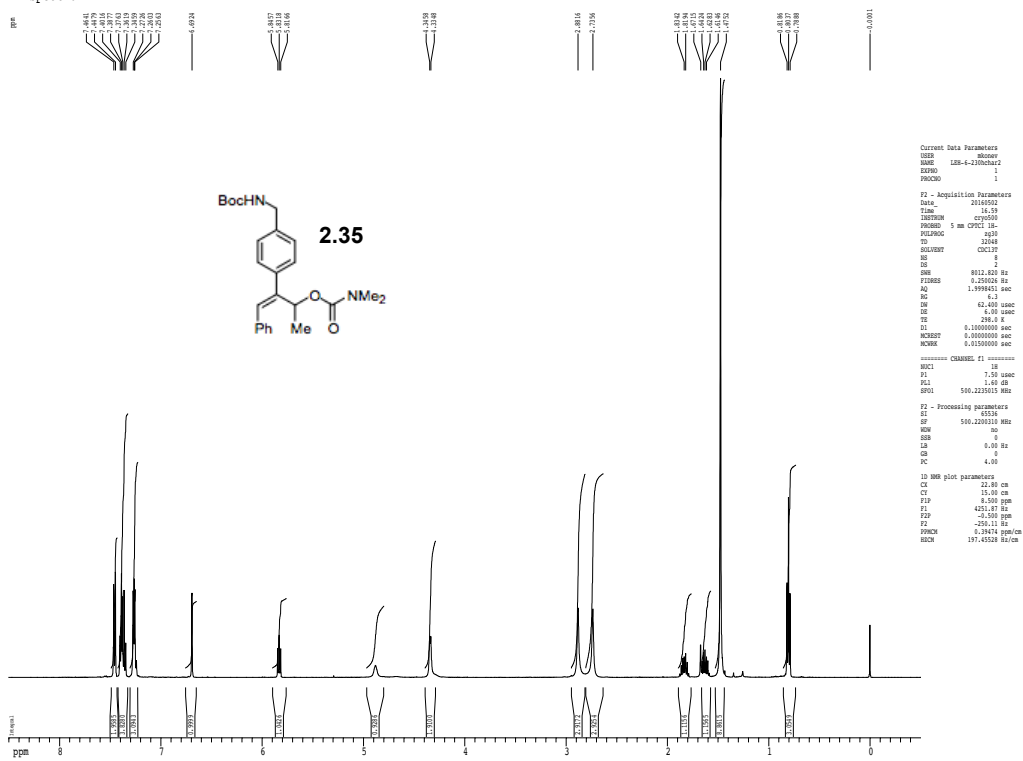
1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling

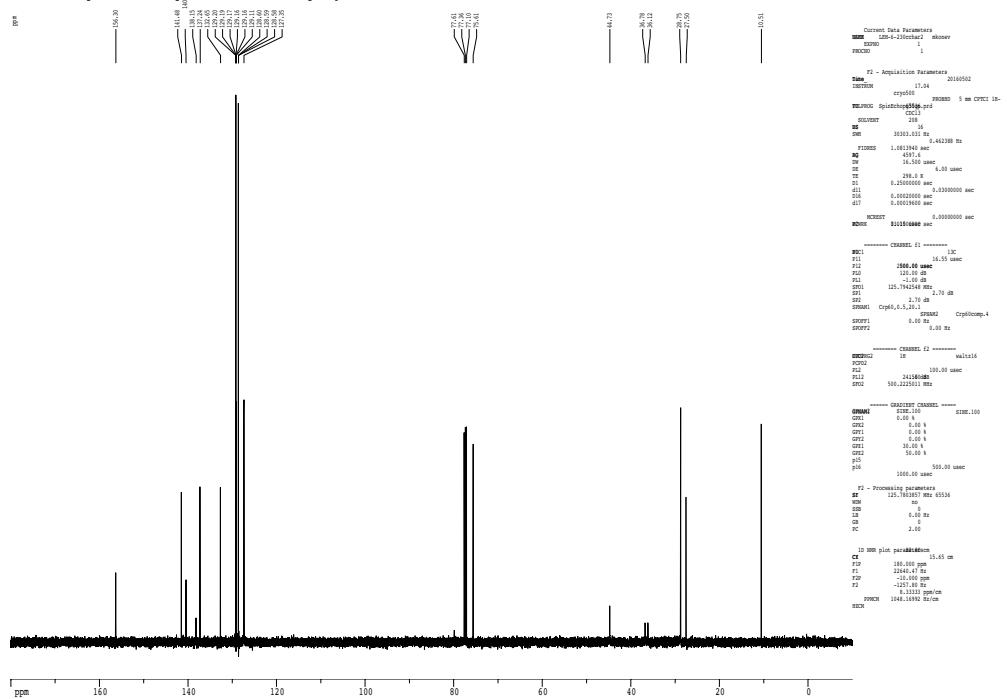


¹H spectrum



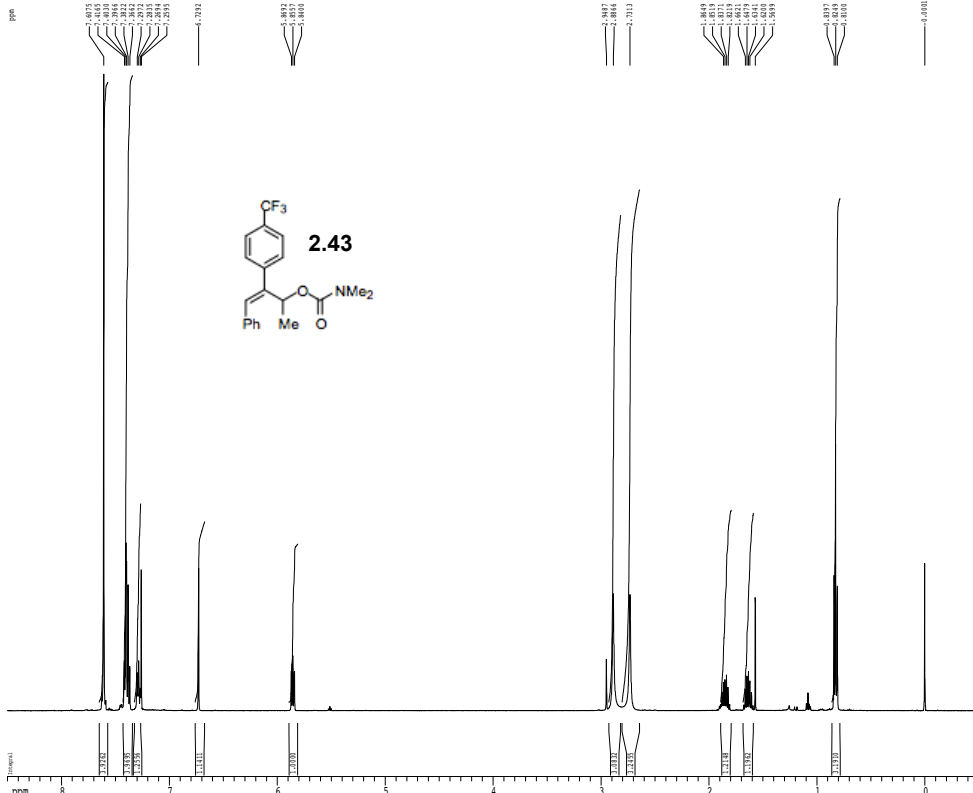
Current Data Parameters
NAME 128-4-210a2a2
PROCNO 1
F2 - Acquisition Parameters
Date_ 20160502
Time 16.19
INSTRUM crysol
PROBHD 5 mm CPYC 1H
PULPROG zgpg30
TD 32768
SOLVENT cdcl3
DS 2
SFO 801.829 MHz
FIDRES 0.20000 MHz
AQ 1.998431 sec
RG 61.3
DM 62.500 usec
DE 9.00 usec
TE 296.0 K
SI 0.1500000 sec
NUC1 1
NCHET 0.0000000 sec
NCHKA 0.1500000 sec
===== CHANNEL f1 =====
NUC1 1H
P1 16.55 usec
PL1 -1.80 dB
SFO1 500.125511 MHz
F2 - Processing parameters
SI 65236
SF 500.125511 MHz
WDW no
SSB 0
LA 0.00 Hz
GB 0
PC 4.00
ID NMR plot parameters
CX 22.81 cm
CY 15.00 cm
FID 8.450 ppm
F1 4201.87 Hz
F2 -12.000 ppm
PC 250.11 Hz
PUNCH 8.12814 mm/cm
HCN 197.45528 Hz/cm

Z-restored spin-echo 13C spectrum with ¹H decoupling



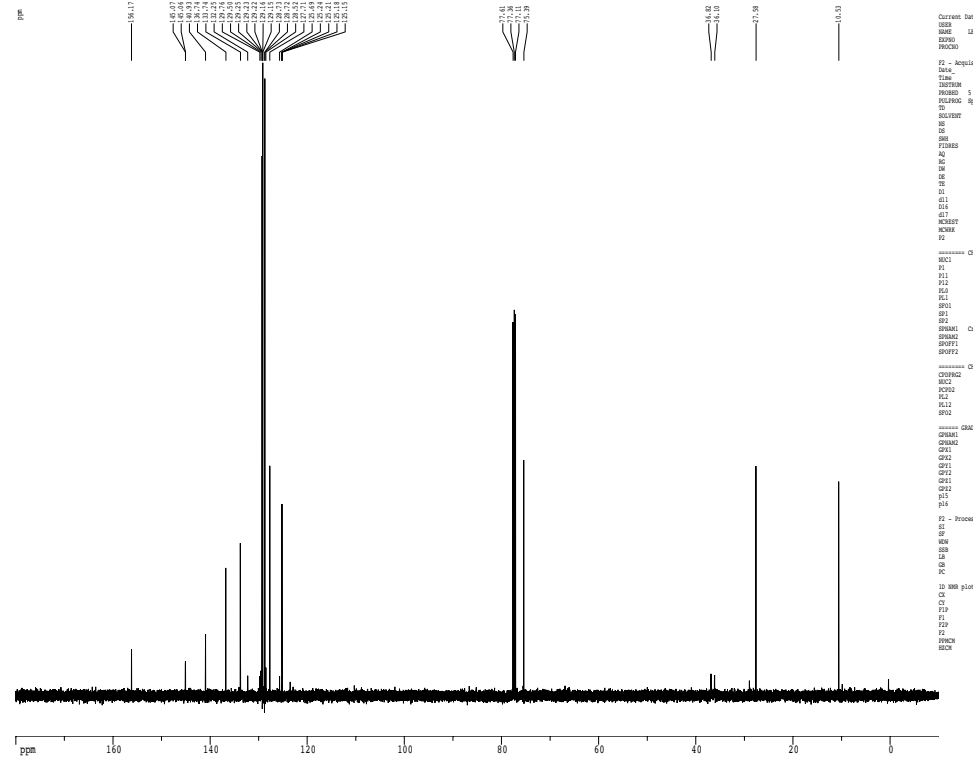
Current Data Parameters
NAME 128-4-210a2a2 aknow
PROCNO 1
F2 - Acquisition Parameters
Date_ 20160502
Time 17.04
INSTRUM crysol
PROBHD 5 mm CPYC 1H
PULPROG sphtchycp13c
SOLVENT cdcl3
DS 2
SFO 125.76187 MHz
FIDRES 1.081049 MHz
AQ 4.0713 sec
RG 34.333
DM 16.100 usec
DE 9.00 usec
TE 296.0 K
SI 0.1500000 sec
NUC1 13C
NCHET 0.0000000 sec
NCHKA 0.1500000 sec
===== CHANNEL f1 =====
NUC1 13C
P1 16.55 usec
PL1 18.00 dB
SFO1 125.76187 MHz
===== CHANNEL f2 =====
NUC2 1H
P2 9.19 usec
PL2 -1.80 dB
SFO2 500.125511 MHz
===== CHANNEL f3 =====
NUC3 1H
P3 9.19 usec
PL3 18.00 dB
SFO3 500.125511 MHz
===== CHANNEL f4 =====
NUC4 13C
P4 16.55 usec
PL4 -1.80 dB
SFO4 125.76187 MHz
===== CHANNEL f5 =====
NUC5 13C
P5 16.55 usec
PL5 -1.80 dB
SFO5 125.76187 MHz
===== CHANNEL f6 =====
NUC6 13C
P6 16.55 usec
PL6 -1.80 dB
SFO6 125.76187 MHz
===== CHANNEL f7 =====
NUC7 13C
P7 16.55 usec
PL7 -1.80 dB
SFO7 125.76187 MHz
===== CHANNEL f8 =====
NUC8 13C
P8 16.55 usec
PL8 -1.80 dB
SFO8 125.76187 MHz
===== CHANNEL f9 =====
NUC9 13C
P9 16.55 usec
PL9 -1.80 dB
SFO9 125.76187 MHz
===== CHANNEL f10 =====
NUC10 13C
P10 16.55 usec
PL10 -1.80 dB
SFO10 125.76187 MHz
===== CHANNEL f11 =====
NUC11 13C
P11 16.55 usec
PL11 -1.80 dB
SFO11 125.76187 MHz
===== CHANNEL f12 =====
NUC12 13C
P12 16.55 usec
PL12 -1.80 dB
SFO12 125.76187 MHz
===== CHANNEL f13 =====
NUC13 13C
P13 16.55 usec
PL13 -1.80 dB
SFO13 125.76187 MHz
===== CHANNEL f14 =====
NUC14 13C
P14 16.55 usec
PL14 -1.80 dB
SFO14 125.76187 MHz
===== CHANNEL f15 =====
NUC15 13C
P15 16.55 usec
PL15 -1.80 dB
SFO15 125.76187 MHz
===== CHANNEL f16 =====
NUC16 13C
P16 16.55 usec
PL16 -1.80 dB
SFO16 125.76187 MHz
===== CHANNEL f17 =====
NUC17 13C
P17 16.55 usec
PL17 -1.80 dB
SFO17 125.76187 MHz
===== CHANNEL f18 =====
NUC18 13C
P18 16.55 usec
PL18 -1.80 dB
SFO18 125.76187 MHz
===== CHANNEL f19 =====
NUC19 13C
P19 16.55 usec
PL19 -1.80 dB
SFO19 125.76187 MHz
===== CHANNEL f20 =====
NUC20 13C
P20 16.55 usec
PL20 -1.80 dB
SFO20 125.76187 MHz
===== CHANNEL f21 =====
NUC21 13C
P21 16.55 usec
PL21 -1.80 dB
SFO21 125.76187 MHz
===== CHANNEL f22 =====
NUC22 13C
P22 16.55 usec
PL22 -1.80 dB
SFO22 125.76187 MHz
===== CHANNEL f23 =====
NUC23 13C
P23 16.55 usec
PL23 -1.80 dB
SFO23 125.76187 MHz
===== CHANNEL f24 =====
NUC24 13C
P24 16.55 usec
PL24 -1.80 dB
SFO24 125.76187 MHz
===== CHANNEL f25 =====
NUC25 13C
P25 16.55 usec
PL25 -1.80 dB
SFO25 125.76187 MHz
===== CHANNEL f26 =====
NUC26 13C
P26 16.55 usec
PL26 -1.80 dB
SFO26 125.76187 MHz
===== CHANNEL f27 =====
NUC27 13C
P27 16.55 usec
PL27 -1.80 dB
SFO27 125.76187 MHz
===== CHANNEL f28 =====
NUC28 13C
P28 16.55 usec
PL28 -1.80 dB
SFO28 125.76187 MHz
===== CHANNEL f29 =====
NUC29 13C
P29 16.55 usec
PL29 -1.80 dB
SFO29 125.76187 MHz
===== CHANNEL f30 =====
NUC30 13C
P30 16.55 usec
PL30 -1.80 dB
SFO30 125.76187 MHz
===== CHANNEL f31 =====
NUC31 13C
P31 16.55 usec
PL31 -1.80 dB
SFO31 125.76187 MHz
===== CHANNEL f32 =====
NUC32 13C
P32 16.55 usec
PL32 -1.80 dB
SFO32 125.76187 MHz
===== CHANNEL f33 =====
NUC33 13C
P33 16.55 usec
PL33 -1.80 dB
SFO33 125.76187 MHz
===== CHANNEL f34 =====
NUC34 13C
P34 16.55 usec
PL34 -1.80 dB
SFO34 125.76187 MHz
===== CHANNEL f35 =====
NUC35 13C
P35 16.55 usec
PL35 -1.80 dB
SFO35 125.76187 MHz
===== CHANNEL f36 =====
NUC36 13C
P36 16.55 usec
PL36 -1.80 dB
SFO36 125.76187 MHz
===== CHANNEL f37 =====
NUC37 13C
P37 16.55 usec
PL37 -1.80 dB
SFO37 125.76187 MHz
===== CHANNEL f38 =====
NUC38 13C
P38 16.55 usec
PL38 -1.80 dB
SFO38 125.76187 MHz
===== CHANNEL f39 =====
NUC39 13C
P39 16.55 usec
PL39 -1.80 dB
SFO39 125.76187 MHz
===== CHANNEL f40 =====
NUC40 13C
P40 16.55 usec
PL40 -1.80 dB
SFO40 125.76187 MHz
===== CHANNEL f41 =====
NUC41 13C
P41 16.55 usec
PL41 -1.80 dB
SFO41 125.76187 MHz
===== CHANNEL f42 =====
NUC42 13C
P42 16.55 usec
PL42 -1.80 dB
SFO42 125.76187 MHz
===== CHANNEL f43 =====
NUC43 13C
P43 16.55 usec
PL43 -1.80 dB
SFO43 125.76187 MHz
===== CHANNEL f44 =====
NUC44 13C
P44 16.55 usec
PL44 -1.80 dB
SFO44 125.76187 MHz
===== CHANNEL f45 =====
NUC45 13C
P45 16.55 usec
PL45 -1.80 dB
SFO45 125.76187 MHz
===== CHANNEL f46 =====
NUC46 13C
P46 16.55 usec
PL46 -1.80 dB
SFO46 125.76187 MHz
===== CHANNEL f47 =====
NUC47 13C
P47 16.55 usec
PL47 -1.80 dB
SFO47 125.76187 MHz
===== CHANNEL f48 =====
NUC48 13C
P48 16.55 usec
PL48 -1.80 dB
SFO48 125.76187 MHz
===== CHANNEL f49 =====
NUC49 13C
P49 16.55 usec
PL49 -1.80 dB
SFO49 125.76187 MHz
===== CHANNEL f50 =====
NUC50 13C
P50 16.55 usec
PL50 -1.80 dB
SFO50 125.76187 MHz
===== CHANNEL f51 =====
NUC51 13C
P51 16.55 usec
PL51 -1.80 dB
SFO51 125.76187 MHz
===== CHANNEL f52 =====
NUC52 13C
P52 16.55 usec
PL52 -1.80 dB
SFO52 125.76187 MHz
===== CHANNEL f53 =====
NUC53 13C
P53 16.55 usec
PL53 -1.80 dB
SFO53 125.76187 MHz
===== CHANNEL f54 =====
NUC54 13C
P54 16.55 usec
PL54 -1.80 dB
SFO54 125.76187 MHz
===== CHANNEL f55 =====
NUC55 13C
P55 16.55 usec
PL55 -1.80 dB
SFO55 125.76187 MHz
===== CHANNEL f56 =====
NUC56 13C
P56 16.55 usec
PL56 -1.80 dB
SFO56 125.76187 MHz
===== CHANNEL f57 =====
NUC57 13C
P57 16.55 usec
PL57 -1.80 dB
SFO57 125.76187 MHz
===== CHANNEL f58 =====
NUC58 13C
P58 16.55 usec
PL58 -1.80 dB
SFO58 125.76187 MHz
===== CHANNEL f59 =====
NUC59 13C
P59 16.55 usec
PL59 -1.80 dB
SFO59 125.76187 MHz
===== CHANNEL f60 =====
NUC60 13C
P60 16.55 usec
PL60 -1.80 dB
SFO60 125.76187 MHz
===== CHANNEL f61 =====
NUC61 13C
P61 16.55 usec
PL61 -1.80 dB
SFO61 125.76187 MHz
===== CHANNEL f62 =====
NUC62 13C
P62 16.55 usec
PL62 -1.80 dB
SFO62 125.76187 MHz
===== CHANNEL f63 =====
NUC63 13C
P63 16.55 usec
PL63 -1.80 dB
SFO63 125.76187 MHz
===== CHANNEL f64 =====
NUC64 13C
P64 16.55 usec
PL64 -1.80 dB
SFO64 125.76187 MHz
===== CHANNEL f65 =====
NUC65 13C
P65 16.55 usec
PL65 -1.80 dB
SFO65 125.76187 MHz
===== CHANNEL f66 =====
NUC66 13C
P66 16.55 usec
PL66 -1.80 dB
SFO66 125.76187 MHz
===== CHANNEL f67 =====
NUC67 13C
P67 16.55 usec
PL67 -1.80 dB
SFO67 125.76187 MHz
===== CHANNEL f68 =====
NUC68 13C
P68 16.55 usec
PL68 -1.80 dB
SFO68 125.76187 MHz
===== CHANNEL f69 =====
NUC69 13C
P69 16.55 usec
PL69 -1.80 dB
SFO69 125.76187 MHz
===== CHANNEL f70 =====
NUC70 13C
P70 16.55 usec
PL70 -1.80 dB
SFO70 125.76187 MHz
===== CHANNEL f71 =====
NUC71 13C
P71 16.55 usec
PL71 -1.80 dB
SFO71 125.76187 MHz
===== CHANNEL f72 =====
NUC72 13C
P72 16.55 usec
PL72 -1.80 dB
SFO72 125.76187 MHz
===== CHANNEL f73 =====
NUC73 13C
P73 16.55 usec
PL73 -1.80 dB
SFO73 125.76187 MHz
===== CHANNEL f74 =====
NUC74 13C
P74 16.55 usec
PL74 -1.80 dB
SFO74 125.76187 MHz
===== CHANNEL f75 =====
NUC75 13C
P75 16.55 usec
PL75 -1.80 dB
SFO75 125.76187 MHz
===== CHANNEL f76 =====
NUC76 13C
P76 16.55 usec
PL76 -1.80 dB
SFO76 125.76187 MHz
===== CHANNEL f77 =====
NUC77 13C
P77 16.55 usec
PL77 -1.80 dB
SFO77 125.76187 MHz
===== CHANNEL f78 =====
NUC78 13C
P78 16.55 usec
PL78 -1.80 dB
SFO78 125.76187 MHz
===== CHANNEL f79 =====
NUC79 13C
P79 16.55 usec
PL79 -1.80 dB
SFO79 125.76187 MHz
===== CHANNEL f80 =====
NUC80 13C
P80 16.55 usec
PL80 -1.80 dB
SFO80 125.76187 MHz
===== CHANNEL f81 =====
NUC81 13C
P81 16.55 usec
PL81 -1.80 dB
SFO81 125.76187 MHz
===== CHANNEL f82 =====
NUC82 13C
P82 16.55 usec
PL82 -1.80 dB
SFO82 125.76187 MHz
===== CHANNEL f83 =====
NUC83 13C
P83 16.55 usec
PL83 -1.80 dB
SFO83 125.76187 MHz
===== CHANNEL f84 =====
NUC84 13C
P84 16.55 usec
PL84 -1.80 dB
SFO84 125.76187 MHz
===== CHANNEL f85 =====
NUC85 13C
P85 16.55 usec
PL85 -1.80 dB
SFO85 125.76187 MHz
===== CHANNEL f86 =====
NUC86 13C
P86 16.55 usec
PL86 -1.80 dB
SFO86 125.76187 MHz
===== CHANNEL f87 =====
NUC87 13C
P87 16.55 usec
PL87 -1.80 dB
SFO87 125.76187 MHz
===== CHANNEL f88 =====
NUC88 13C
P88 16.55 usec
PL88 -1.80 dB
SFO88 125.76187 MHz
===== CHANNEL f89 =====
NUC89 13C
P89 16.55 usec
PL89 -1.80 dB
SFO89 125.76187 MHz
===== CHANNEL f90 =====
NUC90 13C
P90 16.55 usec
PL90 -1.80 dB
SFO90 125.76187 MHz
===== CHANNEL f91 =====
NUC91 13C
P91 16.55 usec
PL91 -1.80 dB
SFO91 125.76187 MHz
===== CHANNEL f92 =====
NUC92 13C
P92 16.55 usec
PL92 -1.80 dB
SFO92 125.76187 MHz
===== CHANNEL f93 =====
NUC93 13C
P93 16.55 usec
PL93 -1.80 dB
SFO93 125.76187 MHz
===== CHANNEL f94 =====
NUC94 13C
P94 16.55 usec
PL94 -1.80 dB
SFO94 125.76187 MHz
===== CHANNEL f95 =====
NUC95 13C
P95 16.55 usec
PL95 -1.80 dB
SFO95 125.76187 MHz
===== CHANNEL f96 =====
NUC96 13C
P96 16.55 usec
PL96 -1.80 dB
SFO96 125.76187 MHz
===== CHANNEL f97 =====
NUC97 13C
P97 16.55 usec
PL97 -1.80 dB
SFO97 125.76187 MHz
===== CHANNEL f98 =====
NUC98 13C
P98 16.55 usec
PL98 -1.80 dB
SFO98 125.76187 MHz
===== CHANNEL f99 =====
NUC99 13C
P99 16.55 usec
PL99 -1.80 dB
SFO99 125.76187 MHz
===== CHANNEL f100 =====
NUC100 13C
P100 16.55 usec
PL100 -1.80 dB
SFO100 125.76187 MHz
ID NMR plot parameters
CX 15.45 cm
CY 15.00 cm
FID 8.450 ppm
F1 4201.87 Hz
F2 -12.000 ppm
PC 250.11 Hz
PUNCH 8.13312 mm/cm
HCN 197.45528 Hz/cm

1H spectrum



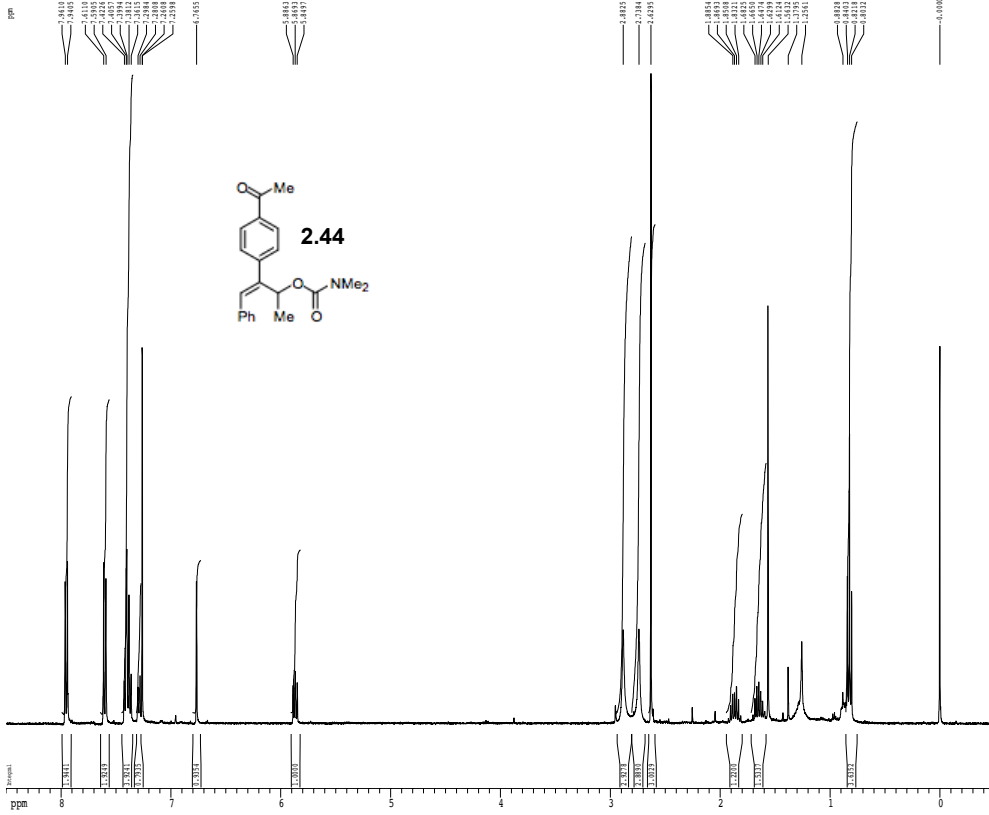
Current Data Parameters
USER: hmoser
NAME: LER-4-277084r
EXPNO: 1
PROCNO: 1
F2 - Acquisition Parameters
Date_: 20160429
Time: 6:17
INSTRUM: cryo500
PROBHD: 5 mm CPYC116-
PULPROG: zgpg30
TD: 32768
SOLVENT: CDCl3
AQ: 0
RG: 2
SI: 32768
SF: 500.136260 Hz
FIDRES: 0.250024 Hz
AQ: 1.998945 sec
RG: 1.1
SI: 64.000 usec
DE: 6.00 usec
TE: 298.2 K
SI: 0.1000000 sec
MCHET: 0.0000000 sec
MCHEX: 0.0100000 sec
===== CHANNEL f1 =====
NUC1: 13C
P1: 1.50 usec
PL1: 1.40 dB
SFO1: 500.2220101 MHz
F2 - Processing parameters
SI: 5534
SF: 500.1280100 MHz
WDW: no
SSB: 0
LB: 0.00 Hz
GB: 0
PC: 4.00
ID: 000 plot parameters
CX: 21.80 cm
CY: 11.00 cm
FID: 0.500 ppm
F1: 4211.67 Hz
F2: -120.11 Hz
F3: -120.11 Hz
F4: 0.28474 ppm/cm
F5: 137.45228 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling

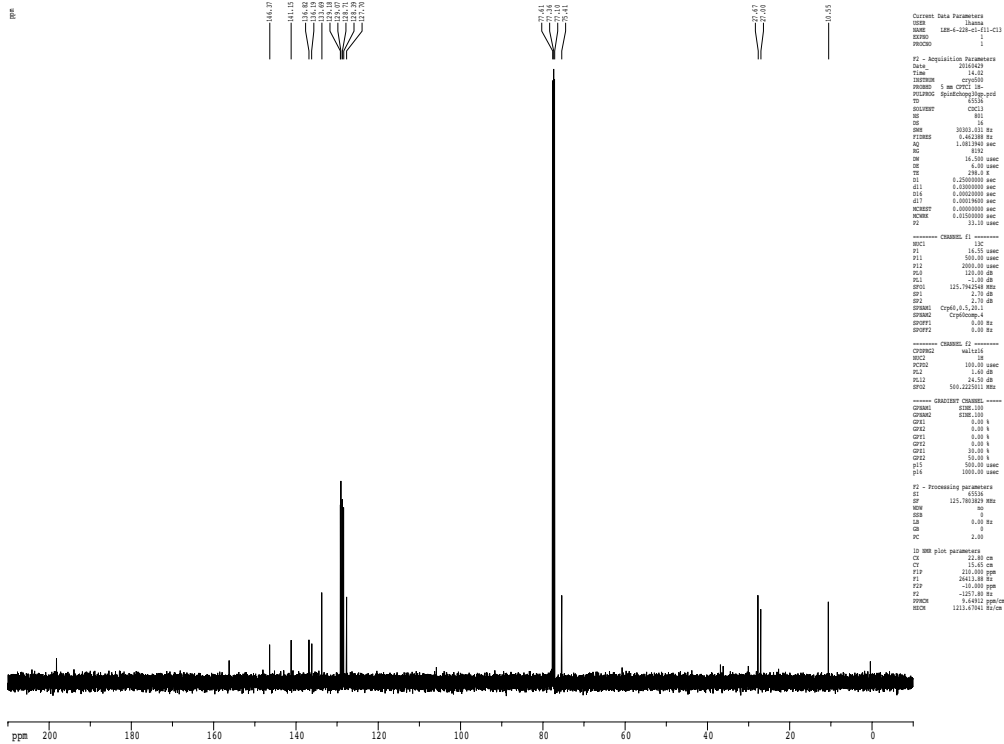


Current Data Parameters
USER: hmoser
NAME: LER-4-277084r
EXPNO: 1
PROCNO: 1
F2 - Acquisition Parameters
Date_: 20160429
Time: 6:20
INSTRUM: cryo500
PROBHD: 5 mm CPYC116-
PULPROG: spinlockgpg30-prd
TD: 65536
SOLVENT: CDCl3
AQ: 0
RG: 64
SI: 32768
SF: 125.761400 MHz
FIDRES: 0.462208 Hz
AQ: 1.0013940 sec
RG: 1.1
SI: 16.500 usec
DE: 6.00 usec
TE: 298.2 K
SI: 0.2500000 sec
MCH1: 0.1000000 sec
MCH2: 0.0500000 sec
MCH3: 0.0001000 sec
MCH4: 0.0000000 sec
MCH5: 0.1100000 sec
F2: 10.10 usec
===== CHANNEL f1 =====
NUC1: 13C
P1: 1.50 usec
PL1: 1.40 dB
SFO1: 125.761400 MHz
SFO2: 125.761400 MHz
SFO3: 125.761400 MHz
SFO4: 125.761400 MHz
SFO5: 125.761400 MHz
SFO6: 125.761400 MHz
SFO7: 125.761400 MHz
SFO8: 125.761400 MHz
SFO9: 125.761400 MHz
SFO10: 125.761400 MHz
SFO11: 125.761400 MHz
SFO12: 125.761400 MHz
SFO13: 125.761400 MHz
SFO14: 125.761400 MHz
SFO15: 125.761400 MHz
SFO16: 125.761400 MHz
SFO17: 125.761400 MHz
SFO18: 125.761400 MHz
SFO19: 125.761400 MHz
SFO20: 125.761400 MHz
SFO21: 125.761400 MHz
SFO22: 125.761400 MHz
SFO23: 125.761400 MHz
SFO24: 125.761400 MHz
SFO25: 125.761400 MHz
SFO26: 125.761400 MHz
SFO27: 125.761400 MHz
SFO28: 125.761400 MHz
SFO29: 125.761400 MHz
SFO30: 125.761400 MHz
SFO31: 125.761400 MHz
SFO32: 125.761400 MHz
SFO33: 125.761400 MHz
SFO34: 125.761400 MHz
SFO35: 125.761400 MHz
SFO36: 125.761400 MHz
SFO37: 125.761400 MHz
SFO38: 125.761400 MHz
SFO39: 125.761400 MHz
SFO40: 125.761400 MHz
SFO41: 125.761400 MHz
SFO42: 125.761400 MHz
SFO43: 125.761400 MHz
SFO44: 125.761400 MHz
SFO45: 125.761400 MHz
SFO46: 125.761400 MHz
SFO47: 125.761400 MHz
SFO48: 125.761400 MHz
SFO49: 125.761400 MHz
SFO50: 125.761400 MHz
SFO51: 125.761400 MHz
SFO52: 125.761400 MHz
SFO53: 125.761400 MHz
SFO54: 125.761400 MHz
SFO55: 125.761400 MHz
SFO56: 125.761400 MHz
SFO57: 125.761400 MHz
SFO58: 125.761400 MHz
SFO59: 125.761400 MHz
SFO60: 125.761400 MHz
SFO61: 125.761400 MHz
SFO62: 125.761400 MHz
SFO63: 125.761400 MHz
SFO64: 125.761400 MHz
SFO65: 125.761400 MHz
SFO66: 125.761400 MHz
SFO67: 125.761400 MHz
SFO68: 125.761400 MHz
SFO69: 125.761400 MHz
SFO70: 125.761400 MHz
SFO71: 125.761400 MHz
SFO72: 125.761400 MHz
SFO73: 125.761400 MHz
SFO74: 125.761400 MHz
SFO75: 125.761400 MHz
SFO76: 125.761400 MHz
SFO77: 125.761400 MHz
SFO78: 125.761400 MHz
SFO79: 125.761400 MHz
SFO80: 125.761400 MHz
SFO81: 125.761400 MHz
SFO82: 125.761400 MHz
SFO83: 125.761400 MHz
SFO84: 125.761400 MHz
SFO85: 125.761400 MHz
SFO86: 125.761400 MHz
SFO87: 125.761400 MHz
SFO88: 125.761400 MHz
SFO89: 125.761400 MHz
SFO90: 125.761400 MHz
SFO91: 125.761400 MHz
SFO92: 125.761400 MHz
SFO93: 125.761400 MHz
SFO94: 125.761400 MHz
SFO95: 125.761400 MHz
SFO96: 125.761400 MHz
SFO97: 125.761400 MHz
SFO98: 125.761400 MHz
SFO99: 125.761400 MHz
SFO100: 125.761400 MHz
===== CHANNEL f2 =====
CPUPROG: waltz16
NUC2: 13C
PCPD2: 100.00 usec
PL12: 1.40 dB
PL13: 24.50 dB
SFO2: 500.2220101 MHz
===== CHANNEL f3 =====
SFO3: 125.761400 MHz
SFO4: 125.761400 MHz
SFO5: 125.761400 MHz
SFO6: 125.761400 MHz
SFO7: 125.761400 MHz
SFO8: 125.761400 MHz
SFO9: 125.761400 MHz
SFO10: 125.761400 MHz
SFO11: 125.761400 MHz
SFO12: 125.761400 MHz
SFO13: 125.761400 MHz
SFO14: 125.761400 MHz
SFO15: 125.761400 MHz
SFO16: 125.761400 MHz
SFO17: 125.761400 MHz
SFO18: 125.761400 MHz
SFO19: 125.761400 MHz
SFO20: 125.761400 MHz
SFO21: 125.761400 MHz
SFO22: 125.761400 MHz
SFO23: 125.761400 MHz
SFO24: 125.761400 MHz
SFO25: 125.761400 MHz
SFO26: 125.761400 MHz
SFO27: 125.761400 MHz
SFO28: 125.761400 MHz
SFO29: 125.761400 MHz
SFO30: 125.761400 MHz
SFO31: 125.761400 MHz
SFO32: 125.761400 MHz
SFO33: 125.761400 MHz
SFO34: 125.761400 MHz
SFO35: 125.761400 MHz
SFO36: 125.761400 MHz
SFO37: 125.761400 MHz
SFO38: 125.761400 MHz
SFO39: 125.761400 MHz
SFO40: 125.761400 MHz
SFO41: 125.761400 MHz
SFO42: 125.761400 MHz
SFO43: 125.761400 MHz
SFO44: 125.761400 MHz
SFO45: 125.761400 MHz
SFO46: 125.761400 MHz
SFO47: 125.761400 MHz
SFO48: 125.761400 MHz
SFO49: 125.761400 MHz
SFO50: 125.761400 MHz
SFO51: 125.761400 MHz
SFO52: 125.761400 MHz
SFO53: 125.761400 MHz
SFO54: 125.761400 MHz
SFO55: 125.761400 MHz
SFO56: 125.761400 MHz
SFO57: 125.761400 MHz
SFO58: 125.761400 MHz
SFO59: 125.761400 MHz
SFO60: 125.761400 MHz
SFO61: 125.761400 MHz
SFO62: 125.761400 MHz
SFO63: 125.761400 MHz
SFO64: 125.761400 MHz
SFO65: 125.761400 MHz
SFO66: 125.761400 MHz
SFO67: 125.761400 MHz
SFO68: 125.761400 MHz
SFO69: 125.761400 MHz
SFO70: 125.761400 MHz
SFO71: 125.761400 MHz
SFO72: 125.761400 MHz
SFO73: 125.761400 MHz
SFO74: 125.761400 MHz
SFO75: 125.761400 MHz
SFO76: 125.761400 MHz
SFO77: 125.761400 MHz
SFO78: 125.761400 MHz
SFO79: 125.761400 MHz
SFO80: 125.761400 MHz
SFO81: 125.761400 MHz
SFO82: 125.761400 MHz
SFO83: 125.761400 MHz
SFO84: 125.761400 MHz
SFO85: 125.761400 MHz
SFO86: 125.761400 MHz
SFO87: 125.761400 MHz
SFO88: 125.761400 MHz
SFO89: 125.761400 MHz
SFO90: 125.761400 MHz
SFO91: 125.761400 MHz
SFO92: 125.761400 MHz
SFO93: 125.761400 MHz
SFO94: 125.761400 MHz
SFO95: 125.761400 MHz
SFO96: 125.761400 MHz
SFO97: 125.761400 MHz
SFO98: 125.761400 MHz
SFO99: 125.761400 MHz
SFO100: 125.761400 MHz
F2 - Processing parameters
SI: 65536
SF: 125.7618200 MHz
WDW: no
SSB: 0
LB: 0.00 Hz
GB: 0
PC: 2.00
ID: 000 plot parameters
CX: 21.80 cm
CY: 11.00 cm
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F4: -120.188 Hz
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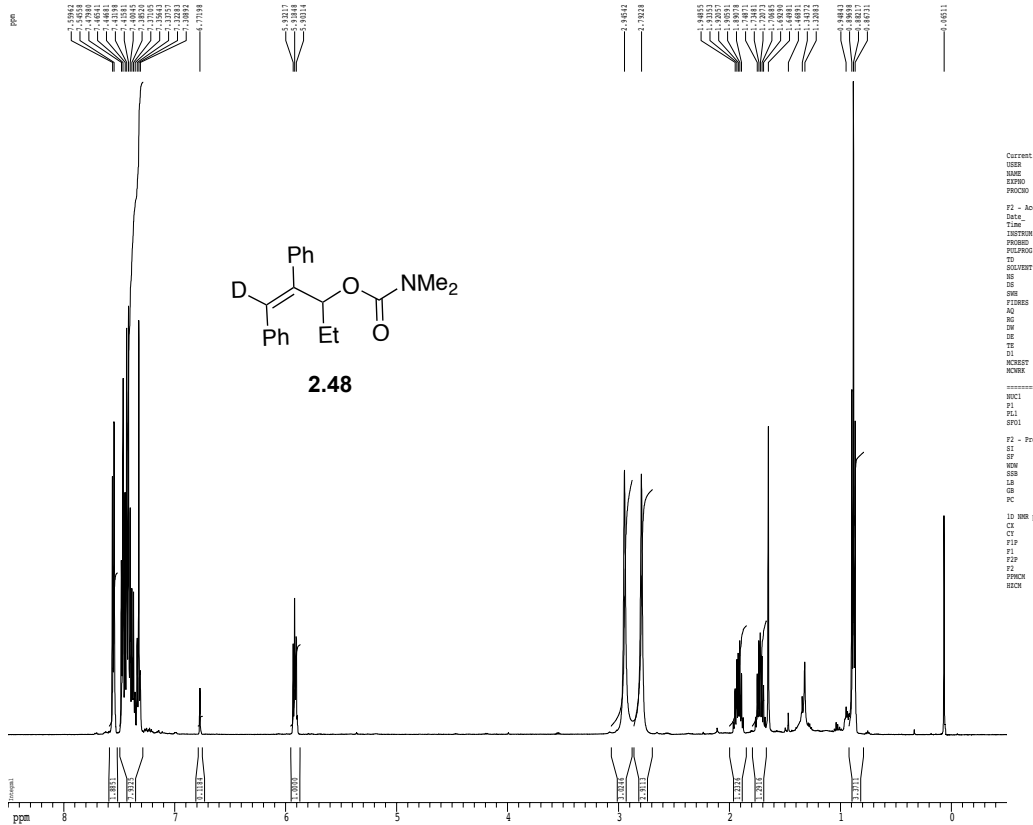
1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling

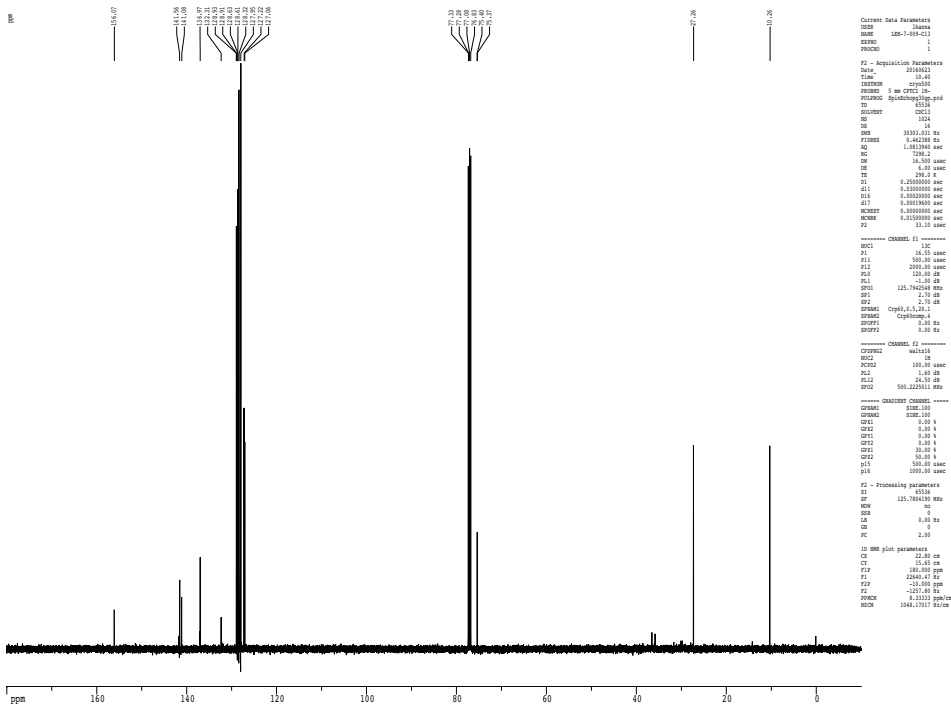


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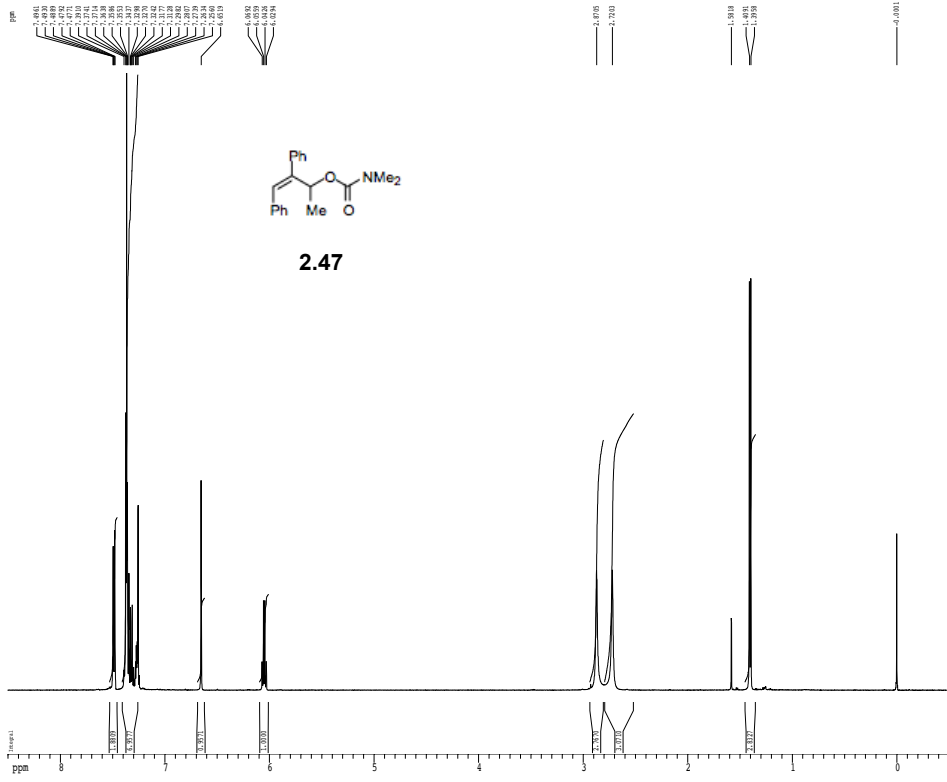
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SOLVENT CDCl3
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DS 4
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FIDRES 0.580114 Hz
AQ 0.0996226 sec
RG 12.1
WE 62.400 usec
DE 4.00 usec
TE 298.2 K
D1 0.1000000 sec
MCRET 0.0000000 sec
MCRM 0.0100000 sec
----- CHANNEL f1 -----
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PL1 1.67 dB
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SSB 0
LB 0.00 Hz
GB 0
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CY 15.00 cm
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PI 4251.81 Hz
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PC -250.15 Hz
PPHVA 0.29476 ppm/cm
RHVA 197.45326 Hz/cm

2-restored spin-echo 13C spectrum with 1H decoupling

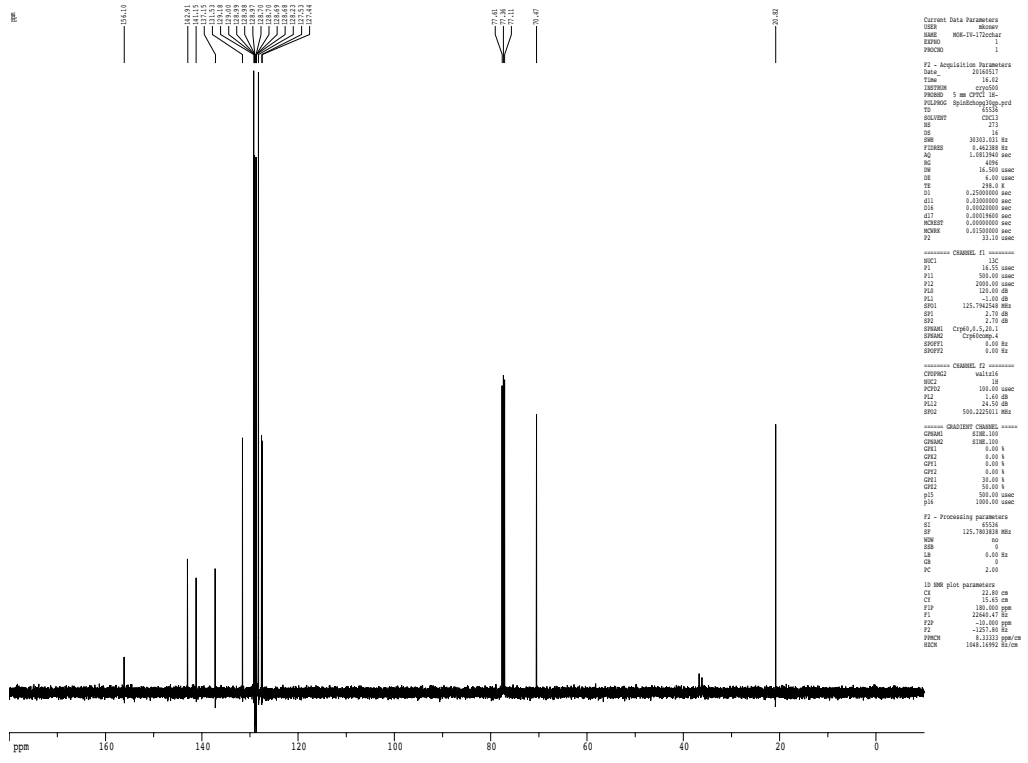


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TD 65536
SOLVENT CDCl3
NS 104
DS 4
SFO 101.6253152 Hz
FIDRES 0.462268 Hz
AQ 0.0122967 sec
RG 12.1
WE 16.50 usec
DE 4.00 usec
TE 298.2 K
D1 0.2500000 sec
d11 0.0300000 sec
d12 0.0000000 sec
d13 0.0000000 sec
MCRET 0.0000000 sec
MCRM 0.0100000 sec
F2 ----- CHANNEL f1 -----
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PL1 180.00 dB
SFO1 125.7642548 MHz
SFO2 125.7642548 MHz
SFO3 125.7642548 MHz
SFO4 125.7642548 MHz
SFO5 125.7642548 MHz
SFO6 125.7642548 MHz
SFO7 125.7642548 MHz
SFO8 125.7642548 MHz
SFO9 125.7642548 MHz
SFO10 125.7642548 MHz
SFO11 125.7642548 MHz
SFO12 125.7642548 MHz
SFO13 125.7642548 MHz
SFO14 125.7642548 MHz
SFO15 125.7642548 MHz
SFO16 125.7642548 MHz
SFO17 125.7642548 MHz
SFO18 125.7642548 MHz
SFO19 125.7642548 MHz
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GB 0
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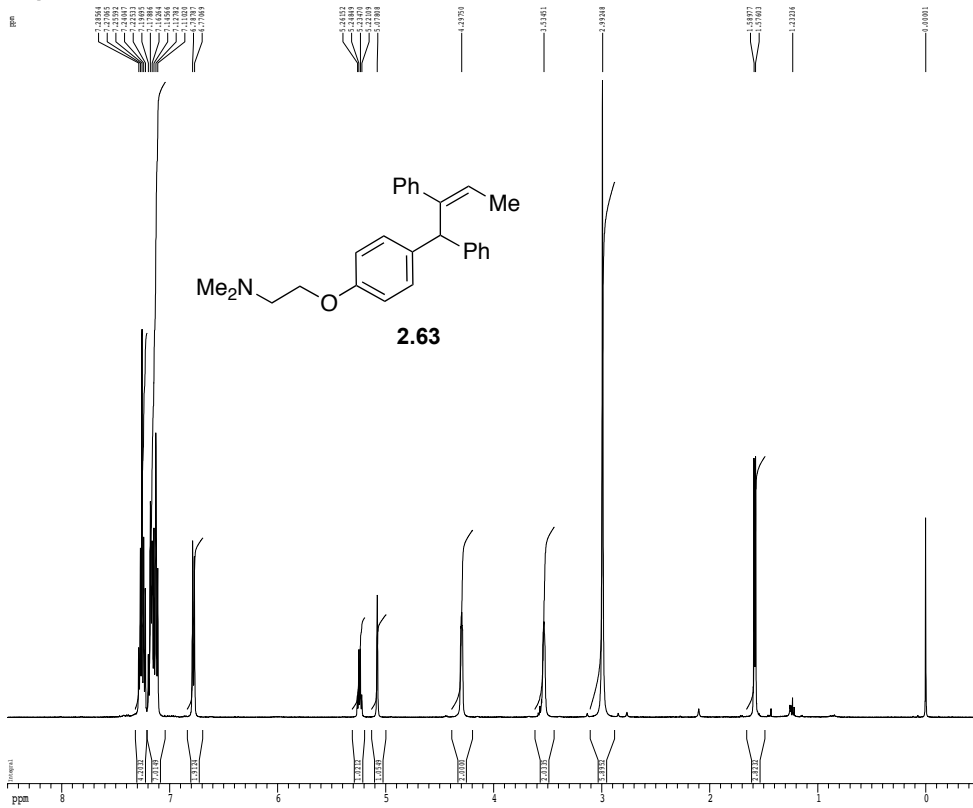
1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



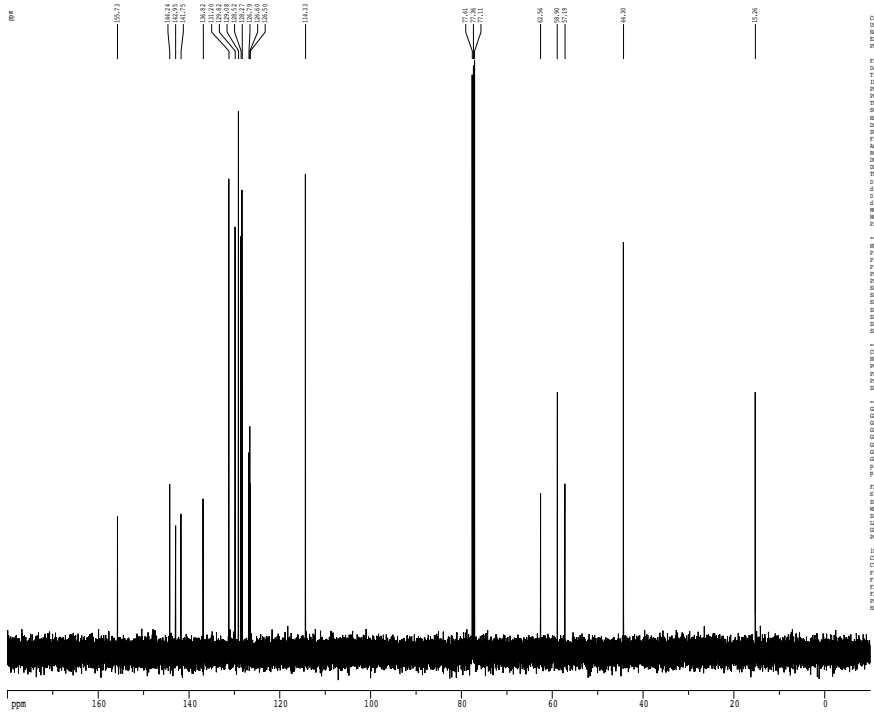
1H spectrum



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SCHEM1 0.0000000 sec
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CZ 25.00 cm
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F6 197.45528 Hz/cm
  
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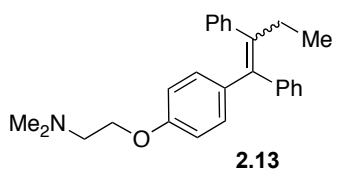
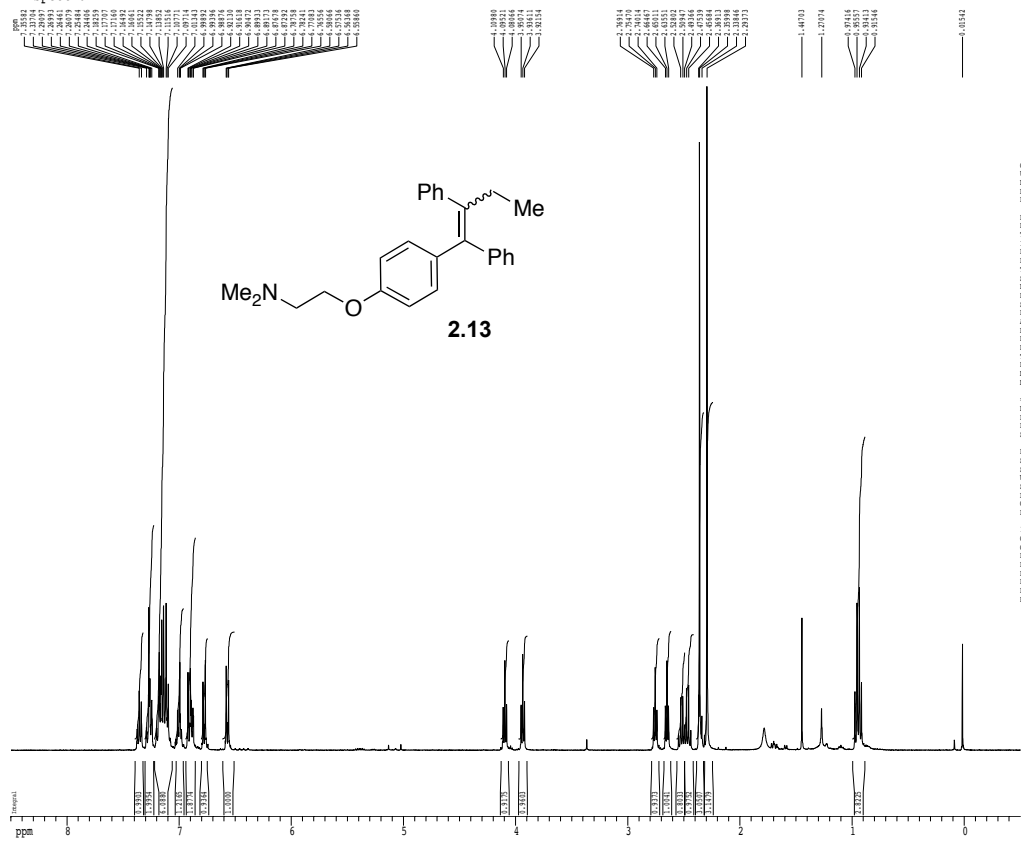
Z-restored spin-echo 13C spectrum with 1H decoupling



```

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PROCNO 1
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SOLVENT CDCl3
NS 8
DS 4
SWH 10383.015 Hz
FIDRES 0.00040 Hz
AQ 1.0813940 sec
RG 337.0
DE 0.00 usec
TE 298.2 K
SI 0.2500000 sec
SCHEM1 0.0000000 sec
MCHW1 0.0000000 sec
MCHW2 0.0100000 sec
F2 - Processing parameters
SI 65536
SF 125.7618183 MHz
WDW EM
SSB 0
GB 0
PC 2.00
IS MRB plot parameters
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CY 25.00 cm
CZ 25.00 cm
F1 2440.47 Hz
F2 2440.47 Hz
F3 -120.00 ppm
F4 -120.00 Hz
F5 6.3333 ppm/cm
F6 168.18992 Hz/cm
  
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1H spectrum



```

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NAME          MGE-TV1300cc
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PROCNO       1

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DS           2
SWH          6410.256 Hz
FIDRES      0.200000 Hz
AQ          2.4998439 sec
RG           201.1
DM           78.000 usec
DE           4.50 usec
TE           298.2 K
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MCHRGPT     0.0000000 sec
MCMWKE      0.01500000 sec

===== CHANNEL f1 =====
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PC1         0.00 dB
SFO1        400.1328099 MHz

F2 - Processing parameters
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SF          400.1308175 MHz
WDW          no
SSB          0
LA           0.00 Hz
GB           0
PC           2.00

ID NON plot parameters
CN           20.00 cm
CY           15.00 cm
FIDP        8.500 ppm
F1          3401.10 Hz
F2P         -8.500 ppm
FZ          -2461.07 Hz
PPHMC       0.29474 ppm/cm
SFCW       137.84506 Hz/cm
    
```

***Intramolecular Nickel-Catalyzed Reductive Cross-Electrophile Coupling Reactions of
Benzylic Esters with Aryl Halides***

3.1 Introduction

Nickel-catalyzed reductive cross-electrophile coupling reactions have recently undergone rapid advances in synthetic utility and mechanistic understanding.¹ Their mild reaction conditions can provide advantages to traditional cross-coupling reactions. For example, reductive coupling reactions offer an attractive strategy toward intramolecular cyclization reactions because they do not require installation of both electrophilic and organometallic functional groups into the starting material.² We envisioned a cross-electrophile reductive coupling reaction would provide straightforward synthesis of 1-arylidanes and tetralins, common motifs in natural products and pharmaceutical agents (Scheme 3.1).^{3,4} While intermolecular reductive coupling reactions have undergone rapid development in recent years,^{1,5} few intramolecular variants have been reported. The Peng group disclosed a stoichiometric nickel-catalyzed reductive coupling reaction to access nitrogen- and oxygen-containing heterocycles (Scheme 3.2a).⁶ In 2014, Gong and coworkers

¹(a) Knappke, E. I. C.; Grupe, G.; Gartner, D.; Corpet, M.; Gosmini, C.; Wangelin, A. J. V. *Chem. Eur. J.* **2014**, *20*, 6828. (b) Weix, D. *Acc. Chem. Res.* **2015**, *48*, 1767. (c) Moragas, T.; Correa, A.; Martin, R. *Chem. Eur. J.* **2014**, *20*, 8242. (d) Gu, J.; Wang, X.; Xue, W.; Gong, H. *Org. Chem. Front.* **2015**, *2*, 1141.

² Traditional cross-coupling reactions have had a transformative impact on synthesis. For reviews, see: (a) Bulger, P. G.; Sarlah, D.; Nicolaou, K. C. *Angew. Chem. Int. Ed.* **2005**, *44*, 4442. (b) Chemler, S. R.; Trauner, D.; Danishefsky, S. J. *Angew. Chem. Int. Ed.* **2001**, *40*, 4544.

³For lead references in alternative methods for synthesis of indanes and tetralins, see: (a) Parham, W. E.; Bradsher, C. K.; Reames, D. C. *J. Org. Chem.* **1981**, *46*, 4804. (b) Miyaura, N.; Ishiyama, T.; Sasaki, H.; Ishikawa, M.; Satoh, M.; Suzuki, A. *J. Am. Chem. Soc.* **1989**, *111*, 314. (c) Bailey, W. F.; Longstaff, S. C. *J. Org. Chem.* **1998**, *63*, 432. (d) Segura, J. L.; Martin, N. *Chem. Rev.* **1999**, *99*, 3199. (f) Begouin, J.-M.; Capitta, F.; Wu, X.; Niggemann, M. *Org. Lett.* **2013**, *15*, 1370. (g) Wang, Y.-M.; Bruno, N. C.; Placeres, A. L.; Zhu, S.; Buchwald, S. L. *J. Am. Chem. Soc.* **2015**, *137*, 10524.

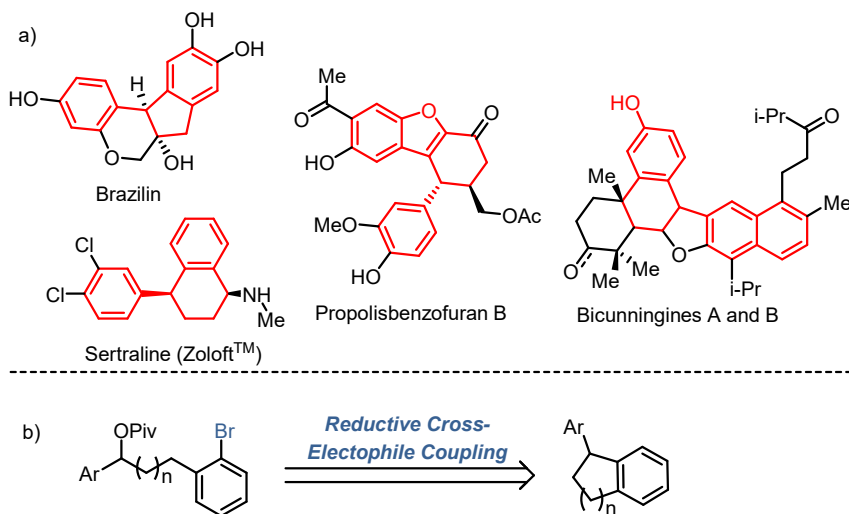
⁴For representative nickel-catalyzed reactions that generate indanes, see: (a) Deng, R.; Sun, L.; Li, Z. *Org. Lett.*, **2007**, *9*, 5207. (b) Watson, M. P.; Jacobsen, E. N. *J. Am. Chem. Soc.* **2008**, *130*, 12594. (c) Arp, F. O.; Fu, G. C. *J. Am. Chem. Soc.* **2005**, *127*, 10482.

⁵For lead references, see: (a) Kadunce, N. T.; Reisman, S. E. *J. Am. Chem. Soc.* **2015**, *137*, 10480. (b) Ackerman, L. K. G.; Lovell, M. M.; Weix, D. J. *Nature* **2015**, *524*, 454.

⁶ Yan, C. S.; Peng, Y.; Xu, X. B.; Wang, Y. W. *Chem. Eur. J.* **2012**, *18*, 6039.

reported a catalytic intramolecular cyclization of dihaloalkanes to access 5- and 6-membered rings (Scheme 3.2b).⁷ Recently, much interest has focused on the use of C–O electrophiles in reductive coupling reactions.^{8,9} Our laboratory has reported a reductive ring-contraction of 4-chlorotetrahydropyrans to generate cyclopropanes (Scheme 3.2c).¹⁰ To further expand the scope of intramolecular cross-electrophile coupling reactions, we targeted cyclization reactions of benzylic esters with aryl halides to afford valuable indanes and tetralins (Scheme 3.2d). These reactions would also complement recent efforts to develop intermolecular reductive coupling reactions to include alcohol derivatives.¹¹ In this chapter we report the intramolecular reductive cross-electrophile coupling reactions of benzylic pivalates with aryl halides and provide evidence for a stereospecific cyclization reaction.

Scheme 3.1. Intramolecular Reductive Cross-Electrophile Coupling Reaction for The Synthesis of Indanes and Tetralins.



⁷ Xue, W.; Xu, H.; Liang, Z.; Qian, Q.; Gong, H. *Org. Lett.* **2014**, *16*, 4984.

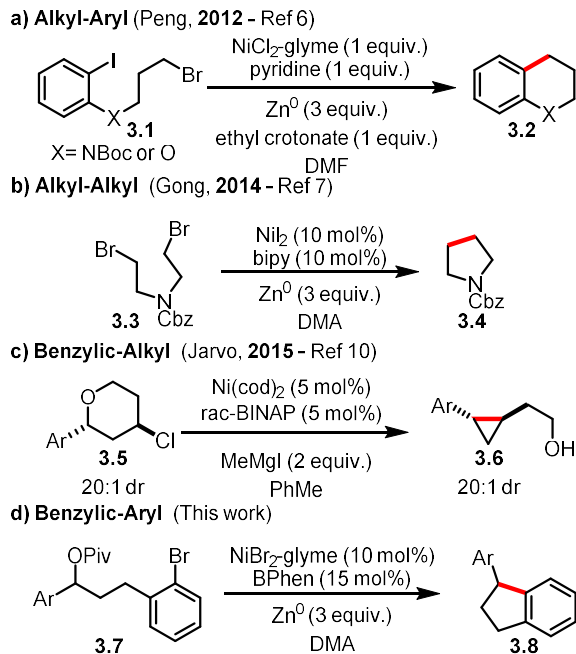
⁸ (a) Weix, D. *Acc. Chem. Res.* **2015**, *48*, 1767. (b) Moragas, T.; Correa, A.; Martin, R. *Chem. Eur. J.* **2014**, *20*, 8242. (c) Gu, J.; Wang, X.; Xue, W.; Gong, H. *Org. Chem. Front.* **2015**, *2*, 1141.

⁹For intermolecular reductive coupling reactions of benzylic alcohol derivatives, see: (a) Correa, A.; Leon, T.; Martin, R. *J. Am. Chem. Soc.* **2014**, *136*, 1062. (b) Ackerman, L. K. G.; Anka-Lufford, L. L.; Naodovic, M.; Weix, D. J. *Chem. Sci.* **2015**, *6*, 1115.

¹⁰Tollefson, E. J.; Erickson, L. W.; Jarvo, E. R. *J. Am. Chem. Soc.* **2015**, *137*, 9760.

¹¹ For intermolecular reductive coupling of allylic esters with halides, see: Anka-Lufford, L. L.; Prinsell, M. R.; Weix, D. J. *J. Org. Chem.* **2012**, *77*, 9989. (b) Cui, X.; Wang, S.; Zhang, Y.; Deng, W.; Qian, Q.; Gong, H. *Org. Biomol. Chem.* **2013**, *11*, 3094.

Scheme 3.2. Nickel-Catalyzed Intramolecular Reductive Cross-Electrophile Coupling Reactions



3.2 Development of Nickel-Catalyzed Reductive Cross-Electrophile Coupling

We designed secondary benzylic pivalate **3.9** as a model substrate, based on our previous work in cross-coupling reactions of benzylic electrophiles.¹² These substrates are easily prepared by lithiation of an arene and addition into the corresponding bromophenyl aldehyde. Additionally, benzylic pivalate esters are less reactive than their halide counterparts. Reaction of **3.9** in the presence of catalytic quantities of NiBr₂-glyme, bathophenanthroline (BPhen), and Zn⁰ provided desired product **3.10** in excellent yield with negligible yields of hydrodehalogenation (Table 3.1, entry 1). Less sterically encumbered esters, such as acetate, provided lower conversion under the reaction conditions (entry 2). Alternative reducing agents such as Mn⁰ also led to lower yields (entry 3). Utilizing phosphine and other aromatic nitrogen-containing ligands, such as bipy or pybox, resulted in a dramatic decrease in product formation (entries 4–7). In the absence of a ligand

¹² Tollefson, E. J.; Hanna, L. E.; Jarvo, E. R. *Acc. Chem. Res.* **2015**, *48*, 2344. For a related transformation, see: Zhou, Q.; Srinivas, H. D.; Dasgupta, S.; Watson, M. P. *J. Am. Chem. Soc.* **2013**, *135*, 3307.

or a nickel catalyst, the desired cyclization does not occur (entries 8 and 9). Additives known to promote reactivity in other reductive cross-electrophile coupling reactions were also examined.¹³ The addition of either pyridine or NaI favored hydrodehalogenation (entries 10 and 11).

Table 3.1. Optimization of reductive coupling reaction conditions

Entry	Variation From Standard Conditions	Recovered 3.9 (%) ^[a]	3.10 (%) ^[a]	3.11 (%) ^[a]
1	none	<2	90	<2
2	Ac instead of Piv	59	9	13
3	Mn instead of Zn	80	<2	<2
4	dppf instead of BPhen	26	25	49
5	bipy instead of BPhen	20	42	25
6	terpyridine instead of BPhen	86	<2	3
7	pybox instead of BPhen	98	<2	<2
8	no ligand	100	<2	<2
9	no NiBr ₂ -glyme	93	<2	<2
10	pyridine (40 mol%)	25	4	61
11	NaI (25 mol%)	<2	25	68
12	NiCl ₂ -glyme instead of NiBr ₂ -glyme	12	79	4
13	DMF instead of DMA	<2	44	35

^[a]Determined by ¹H NMR spectroscopy using PhTMS as internal standard.

3.3 Scope of Intramolecular Reductive Coupling

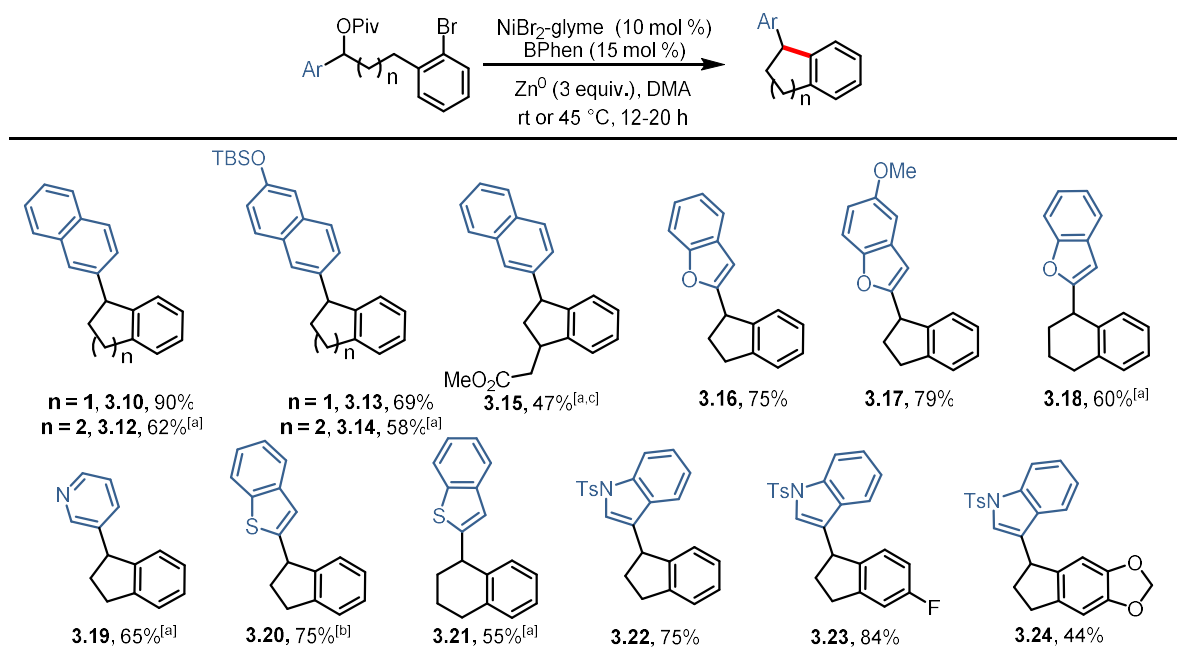
Having established conditions for the cyclization of model substrate **3.9**, we set out to investigate the scope of the transformation (Table 3.2). Cyclization of a series of naphthyl esters provides tetralins **3.12** and **3.14**, which correspond to the core of bicunningines A and B,¹⁴ and tetracyclic indanes **3.10**, **3.13**, and **3.15**. Benzofuran and benzothiophene moieties were well-tolerated, providing good yields for both indanes and tetralins (**3.16–18**, **3.20–21**). Additionally, substrates containing N-heterocycles were found to undergo the desired cyclization. Pyridine-

¹³ (a) Everson, D. A.; Shrestha, R.; Weix, D. J. *J. Am. Chem. Soc.* **2010**, *132*, 920. (b) Cherney, A. H.; Reisman, S. E. *J. Am. Chem. Soc.* **2014**, *136*, 14365.

¹⁴ Hou, X.-F.; Yao, S.; Mandi, A.; Kurtan, T.; Tang, C.-P.; Ke, C.-Q.; Li, X.-Q.; Ye, Y. *Org. Lett.* **2012**, *14*, 460.

substituted indane **3.19** can be synthesized in 65% yield and *N*-tosylindoles **3.22–24** were obtained in good yields. Substrates containing methoxy-, fluoro-, silyl- and ester substituents were also well-tolerated under the reaction conditions. Notably, these cyclization reactions proceed smoothly without the aid of a Thorpe-Ingold effect.¹⁵

Table 3.2. Scope of intramolecular cross-electrophile coupling



^[a]Reaction run with 15 mol % NiBr₂-glyme and 45 °C. ^[b]Reaction run at 45 °C. ^[c]Both sm and product are 1:1 dr.

3.4 Scope of Enantiospecific Intramolecular Reductive Coupling

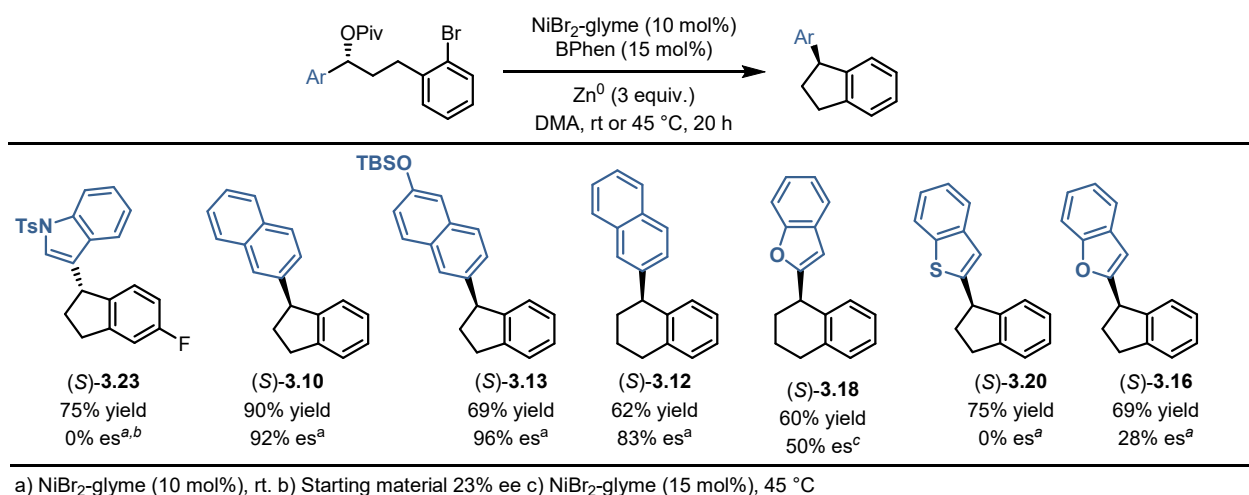
Finally, we sought to determine whether the intramolecular cyclization could proceed in an enantiospecific fashion. While several examples of stereoconvergent reductive coupling reactions have been reported,¹⁶ to the best of our knowledge, there is only one example of an enantiospecific reductive coupling reaction.⁸ Subjecting (*R*)-**3.9** to the optimized conditions afforded (*S*)-**3.10** in 90% yield in 88% enantiomeric excess with 92% enantiospecificity (Table

¹⁵ Beesley, R. M.; Ingold, C. K.; Thorpe, J. F. *J. Chem. Soc. Trans.* **1915**, 107, 1080.

¹⁶ Cui, X.; Wang, S.; Zhang, Y.; Deng, W.; Qian, Q.; Gong, H. *Org. Biomol. Chem.* **2013**, *11*, 3094. Cherney, A. H.; Kadunce, N. T.; Reisman, S. E. *J. Am. Chem. Soc.* **2013**, *135*, 7442. Cherney, A. H.; Reisman, S. E. *J. Am. Chem. Soc.* **2014**, *136*, 14365. Kadunce, N. T.; Reisman, S. E. *J. Am. Chem. Soc.* **2015**, *137*, 10480. Ackerman, L. K. G.; Anka-Lufford, L. L.; Naodovic, M.; Weix, D. J. *Chem. Sci.* **2015**, *6*, 1115.

3.3). Based on comparison of (**S**)-**3.10** to literature values,^{17,18} the reductive cross-electrophile coupling reaction proceeds with inversion at the benzylic center. Indane **3.13** was also formed with high enantiospecificity, as was tetralin **3.12**. Notably, all three of these substrates contain the naphthyl ether moiety which we hypothesize is prone to rapid and stereospecific oxidative addition reactions.^[12] Substrates wherein the ester is activated by a heterocycle such as benzofuran, benzothiophene, or indole provided lower enantiospecificity.¹⁹ This change in stereoselectivity likely correlates to a change in the reaction mechanism. Investigation of the mechanistic details is ongoing.

Table 3.3. Nickel-catalyzed stereospecific reductive cyclization



3.5 Conclusion

In summary, the intramolecular reductive cyclization for the synthesis of indanes and tetralins has been developed. The reactions are tolerant of a variety of heterocycles and functional groups. We have also demonstrated stereospecific cross-electrophile coupling reactions of benzylic esters for synthesis of enantioenriched 1-arylidanes and tetralins.

¹⁷ (a) Yu, N-U.; Xu, M-H. *J. Org. Chem.* **2013**, *78*, 2736. (b) Yue, G.; Lei, K.; Hirao, H.; Zhou, J. *Angew. Chem. Int. Ed.* **2015**, *54*, 6531.

¹⁸ For complete details, see the experimental details.

¹⁹ When enantioenriched starting materials were employed, products **3.16**, **3.18**, **3.20**, and **3.23** were formed with <50% es. For details, see the experimental details.

3.6 Experimental Details

General Procedures

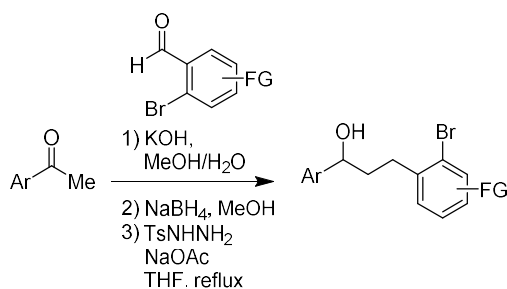
All reactions were carried out under an atmosphere of N₂, or Ar when noted. All glassware was oven- or flame-dried prior to use. Tetrahydrofuran (THF), diethyl ether (Et₂O), dichloromethane (CH₂Cl₂), and dimethylacetamide (DMA) were degassed with Ar and then passed through two 4 x 36 inch columns of anhydrous neutral A-2 alumina (8 x 14 mesh; LaRoche Chemicals; activated under a flow of argon at 350 °C for 12 h) to remove H₂O. All other solvents utilized were purchased “anhydrous” commercially, or purified as described. ¹H NMR spectra were recorded on Bruker DRX-400 (400 MHz ¹H, 100 MHz ¹³C, 376.5 MHz ¹⁹F) or CRYO-500 (500 MHz ¹H, 125.7 MHz ¹³C) spectrometers. Proton chemical shifts are reported in ppm (δ) relative to internal tetramethylsilane (TMS, δ 0.00). Data are reported as follows: chemical shift (multiplicity [singlet (s), broad singlet (br s), doublet (d), doublet of doublets (dd), triplet (t), doublet of triplets (dt), doublet of doublet of triplets (ddt), triplet of triplets (tt), quartet (q), quintet (quin), apparent doublet (ad), apparent triplet (at), multiplet (m)], coupling constants [Hz], integration). Carbon chemical shifts are reported in ppm (δ) relative to TMS with the respective solvent resonance as the internal standard (CDCl₃, δ 77.16 ppm). Unless otherwise indicated, NMR data were collected at 25 °C. Infrared (IR) spectra were obtained on a Thermo Scientific Nicolet iS5 spectrometer with an iD5 ATR tip (neat) and are reported in terms of frequency of absorption (cm⁻¹). Analytical thin-layer chromatography (TLC) was performed using Silica Gel 60 F₂₅₄ precoated plates (0.25 mm thickness). Visualization was accomplished by irradiation with a UV lamp and/or staining with KMnO₄, ceric ammonium molybdate (CAM), or *p*-anisaldehyde (PAA) solutions. Flash chromatography was performed using Silica Gel 60 (170-400 mesh) from Fisher Scientific. Melting points (m.p.) were obtained using a Mel-Temp melting point apparatus and are

uncorrected. Optical rotations were measured on a Rudolph Research Analytical Autopol IV Automatic Polarimeter. SFC determinations of enantiopurity were performed on a Berger Analytical instrument using a Daicel™ Chiralpak® column (OD-H, OJ-H, or AD-H; 100 bar, 50 °C, 215 nm). High resolution mass spectrometry was performed by the University of California, Irvine Mass Spectrometry Center. Nickel(II) bromide ethylene glycol dimethyl ether complex was purchased from Aldrich, stored in a glovebox under an atmosphere of N₂, and used as received. Zinc powder (100 Mesh) was purchased from Alfa Aesar and used as received. All other reagents were purchased commercially and used as received.

General methods for starting material synthesis

The general methods for starting material synthesis will be used throughout the rest of the SI. In each instance a general method is used, it is specified by letter (A, B, etc.) and the exact amounts of reagents used for each reaction are listed for the specific compounds synthesized.

METHOD A: ALDOL CONDENSATION, SODIUM BOROHYDRIDE REDUCTION, AND DIIMIDE REDUCTION

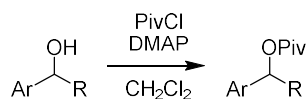


The products were prepared according to a modified procedure reported by Franzen.²⁰ To a 250 mL round bottom flask equipped with a stir bar was added ketone (1.0 equiv), aldehyde (1.0 equiv), KOH (2.0 equiv), and MeOH (30 mL). The reaction was stirred overnight at room temperature. The resulting solid was filtered, washed with water, dried by vacuum filtration, and taken on to

²⁰ Wang, Y.; Franzen, R. *Synlett.*, **2012**, 23, 925.

the next step without further purification. The unpurified chalcone was taken up in MeOH (50 mL) and NaBH₄ (1.2 equiv) was added in two portions over 10 minutes. Upon complete reaction of starting material, as judged by TLC, the mixture was quenched with careful addition of saturated NH₄Cl (15 mL). EtOAc (50 mL) was added, the layers were separated, and the aqueous layer was extracted with EtOAc (3 x 30 mL). The organic extract was washed with brine, dried over MgSO₄, and concentrated under reduced pressure and the unpurified allylic alcohol was taken on to the next step without further purification. The unpurified residue was dissolved in THF (100 mL) and tosylhydrazide (4.0 equiv) and NaOAc•3H₂O (4.0 equiv) were added. The reaction vessel was equipped with a reflux condenser and the mixture was heated at reflux for 24 hours. The mixture was cooled to room temperature and quenched with H₂O (30 mL), and the aqueous layer was extracted with EtOAc (3 x 40 mL). The organic extract was washed with brine, dried over MgSO₄, and concentrated under reduced pressure. The product was then purified by flash column chromatography.

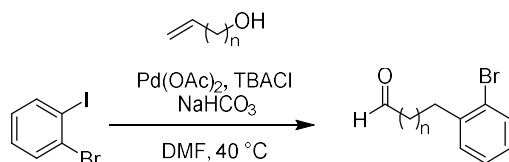
METHOD B: PIVALATION OF BENZYLIC ALCOHOLS



The product was prepared according to a modified procedure reported by Martin.²¹ To a solution of benzylic alcohol (1.0 equiv) in CH₂Cl₂ (20 mL) was added pivaloyl chloride (1.1 equiv) and dimethylaminopyridine (1.1 equiv). The reaction was allowed to stir overnight at room temperature. The reaction was quenched with water (10 mL) and extracted with CH₂Cl₂ (3 x 10 mL). The organic extract was washed with brine, dried over MgSO₄, and concentrated under reduced pressure. The product was purified by flash column chromatography.

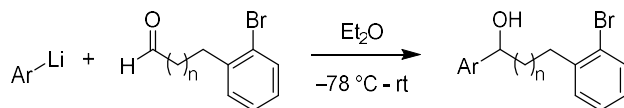
²¹ Zarate, C.; Martin, R. *J. Am. Chem. Soc.*, **2014**, *136*, 2236

METHOD C: HECK REACTION FOR THE PREPARATION OF 2-BROMOPHENYLALDEHYDES



The product was prepared according to a modified procedure reported by Tietze.²² A flame-dried 100 mL round bottom flask equipped with a stir bar and septum was charged with TBACl (1.0 equiv), NaHCO_3 (2.5 equiv), $\text{Pd}(\text{OAc})_2$ (0.05 equiv), and dry DMF (30 mL). The mixture was stirred for 5 minutes followed by simultaneous addition of 1,2-iodobromobenzene (1.0 equiv), and alkenol (1.5 equiv). The reaction mixture was heated to $40\text{ }^\circ\text{C}$ and stirred for 30 hours. The reaction was slowly quenched with saturated NH_4Cl (25 mL) and extracted with EtOAc (3 x 30 mL). The organic extract was washed with brine, dried over MgSO_4 , and concentrated under reduced pressure. The product was purified by flash column chromatography.

METHOD D: ARENE LITHIATION/LITHIUM-HALOGEN EXCHANGE, THEN ADDITION INTO ALDEHYDE



The product was prepared according to a modified procedure reported by O'Doherty.²³ A flame-dried 25 mL round bottom flask equipped with a stir bar and septum was charged with Et_2O (6 mL) and arene (1.3 equiv) and cooled to $-78\text{ }^\circ\text{C}$. *n*-BuLi (2.5 M in hexanes, 1.4 equiv) was added dropwise over 10 minutes. The mixture was stirred cold for 30 minutes and then allowed to warm to room temperature and stir for an additional 20 minutes. A separate flask of requisite aldehyde

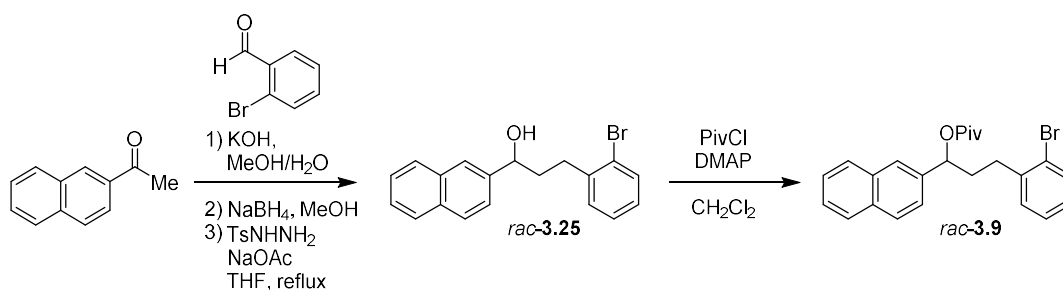
²² Tietze, L. F.; Hungerland, T.; Dufert, A.; Objartel, I.; Stalke, D. *Chem. Eur. J.*, **2012**, *18*, 3286

²³ Guo, H.; O'Doherty, G.A. *Org. Lett.* **2006**, *8*, 1609

(1.0 equiv) dissolved in Et₂O (10 mL) was cooled to -78 °C. The solution of aryl lithium was added dropwise over 30 minutes to the aldehyde. The reaction was warmed to room temperature overnight. The mixture was quenched with saturated NH₄Cl (15 mL). Layers separated and the aqueous layer was extracted with EtOAc (3 x 25 mL). The organic extract was washed with brine, dried over MgSO₄, and concentrated under reduced pressure. The product was purified by flash column chromatography.

SYNTHESIS AND CHARACTERIZATION OF STARTING MATERIALS FOR TABLE 3.2

Scheme 3.3: Synthesis of starting material for formation of *rac*-3.10 (Table 3.2)



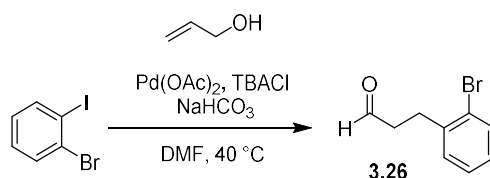
rac-3.25 The product was prepared according to general method A using 2-acetonaphthone (3.4 g, 20 mmol), 2-bromobenzaldehyde (2.3 mL, 20 mmol), and KOH (2.2 g, 40 mmol). Then NaBH₄ (1.0 g, 25 mmol), tosylhydrazide (15 g, 80 mmol) and NaOAc•3H₂O (11 g, 80 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (6.2 g, 18 mmol, 91% over 3 steps). **TLC** R_f = 0.6 (10% EtOAc/hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.86–7.78 (m, 4H), 7.54–7.43 (m, 4H), 7.24–7.18 (m, 2H), 7.04 (ddd, *J* = 8.1, 6.2, 3.3 Hz, 1H), 4.89 (t, *J* = 6.4 Hz, 1H), 2.97–2.76 (m, 2H), 2.22–2.09 (m, 2H), 2.05 (s, 1H); **¹³C NMR** (100 MHz, CDCl₃) δ 141.9, 141.3, 133.5, 133.3, 133.1, 130.7, 128.6, 128.2, 127.92, 127.90, 127.7, 126.4, 126.1, 124.9, 124.7, 124.3, 74.3, 39.0, 32.8; **IR** (neat) 3365, 3054,

2929, 1601, 1470, 1021 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{19}\text{H}_{17}\text{BrONa}$ ($\text{M} + \text{Na}$)⁺ 363.0360, found 363.0368.

rac-3.9. The product was prepared according to general method B using **rac-3.25** (1.7 g, 5.0 mmol), pivaloyl chloride (0.68 mL, 5.5 mmol, 1.1 equiv) and dimethylaminopyridine (0.67 g, 5.5 mmol, 1.1 equiv). The product was purified by flash column chromatography (5% EtOAc/hexanes) to afford the title compound as a colorless oil (1.7 g, 4.1 mmol, 82%). **TLC** R_f = 0.7 (5% EtOAc/hexanes); **^1H NMR** (400 MHz, CDCl_3) δ 7.86–7.74 (m, 4H), δ 7.52–7.39 (m, 4H), δ 7.22–7.11 (m, 2H), 7.00 (t, J = 7.0 Hz, 1H), 5.95 (t, J = 6.5 Hz, 1H), 2.94–2.68 (m, 2H), 2.36–2.11 (m, 2H), 1.26 (s, 9H); **^{13}C NMR** (100 MHz, CDCl_3) δ 177.9, 140.9, 138.2, 133.5, 133.3, 133.2, 130.6, 128.7, 128.3, 128.1, 128.0, 127.8, 126.5, 126.3, 125.8, 124.6, 124.3, 75.5, 39.2, 36.9, 32.8, 27.5; **IR** (neat) 3055, 2969, 1725, 1471, 1279, 1147, 1025 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{24}\text{H}_{25}\text{BrO}_2\text{Na}$ ($\text{M} + \text{Na}$)⁺ 447.0936, found 447.0926.

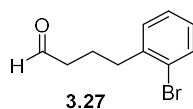
Synthesis of building blocks 3.26 and 3.27.

Aldehydes **3.26** and **3.27** were employed as building blocks in the synthesis of the starting materials for compounds **3.12** and **3.13**, **3.18-21** (Table 3.2).



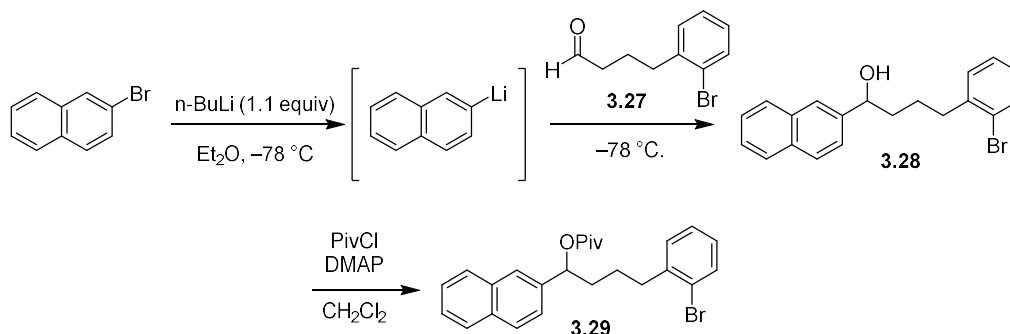
3.26 The product was prepared according to general method C using TBACl (5.6 g, 20.0 mmol), NaHCO_3 (4.2 g, 50 mmol), $\text{Pd}(\text{OAc})_2$ (0.11 g, 0.5 mmol), iodobromobenzene (2.6 mL, 20 mmol, 1.0 equiv), and allyl alcohol (2.1 mL, 30 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a yellow oil (2.4 g, 10 mmol, 52%). Analytical data is consistent with literature values.³ **^1H NMR** (400 MHz, CDCl_3) δ

9.84 (s, 1H), 7.54 (d, $J = 7.9$ Hz, 1H), 7.28–7.20 (m, 2H), 7.08 (q, $J = 4.1$ Hz, 1H), 3.07 (t, $J = 7.4$ Hz, 2H), 2.81 (t, $J = 7.4$ Hz, 2H).



3.27 The product was prepared according to general method C using TBABr (6.5 g, 20.0 mmol), NaHCO₃ (4.2 g, 50 mmol), Pd(OAc)₂ (0.11 g, 0.5 mmol), iodobromobenzene (2.6 mL, 20 mmol, 1.0 equiv), and 3-buten-1-ol (2.6 mL, 30 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a yellow oil (2.4 g, 10 mmol, 52%). Analytical data is consistent with literature values.²⁴ ¹H NMR (400 MHz, CDCl₃) δ 9.79 (s, 1H), 7.53 (d, $J = 7.9$ Hz, 1H), 7.25–7.18 (m, 2H), 7.07 (dd, $J = 7.9, 2.2$ Hz, 1H), 2.79 (dt, $J = 7.5$ Hz, 2H), 2.50 (td, $J = 7.4, 1.5$ Hz, 2H), 1.97 (d, $J = 7.5$ Hz, 2H).

Scheme 3.4: Synthesis of starting material for formation of 3.12 (Table 3.2)

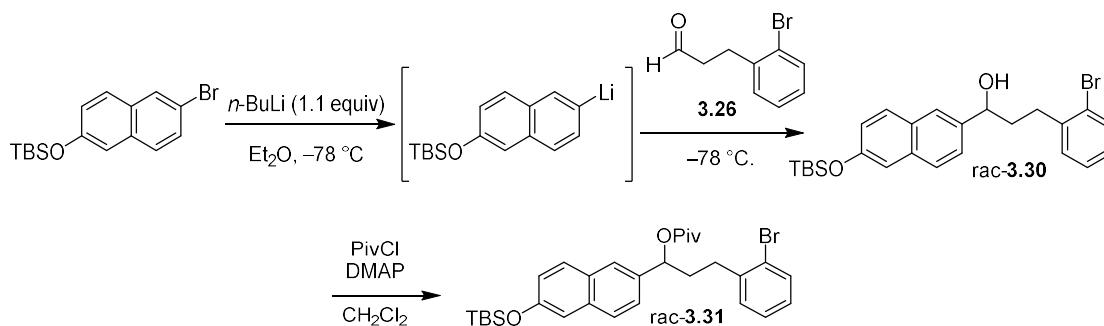


3.28 The product was prepared according to general method D using 2-bromonaphthalene (1.25 g, 6.00 mmol), *n*-BuLi (2.6 mL, 2.5 M in hexane, 6.6 mmol), and **3.27** (1.36 g, 6.00 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a yellow oil (1.89 g, 5.31 mmol, 85%).

²⁴ Gibson, S. E.; Jones, J. O.; McCague R.; Tozer, M. J.; Whitcombe, N. J. *Synlett*, **1999**, 954

3.29 The product was prepared according to general method B using *rac*-**3.28** (0.18 g, 0.50 mmol), pivaloyl chloride (0.080 mL, 0.60 mmol) and dimethylaminopyridine (0.80 g, 0.60 mmol). The product was purified by flash column chromatography (5% EtOAc/hexanes) to afford the title compound as a colorless oil (0.18 g, 0.40 mmol, 80%). **TLC** R_f = 0.4 (12% Et₂O/hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.85–7.78 (m, 3H), 7.76 (s, 1H), 7.52–7.41 (m, 4H), 7.19 (td, J = 7.2, 1.2 Hz, 1H), 7.14 (dd, J = 7.6, 1.27 Hz, 1H), 7.03 (td, J = 7.6, 1.9 Hz, 1H), 5.91 (dd, J = 8.0, 5.6 Hz, 1H), 2.76 (t, J = 7.9 Hz, 2H), 2.07 (dddd, J = 13.4, 10.2, 8.0, 5.1 Hz, 1H), 1.93 (ddt, J = 13.8, 11.2, 5.6 Hz, 1H), 1.78–1.58 (m, 2H), 1.22 (s, 9H). **¹³C NMR** (125 MHz, CDCl₃) δ 177.8, 141.4, 138.5, 133.3, 133.2, 133.0, 130.5, 128.5, 128.2, 127.8, 127.8, 127.6, 126.3, 126.1, 125.6, 124.6, 124.3, 75.6, 39.0, 36.3, 36.0, 27.4, 26.0; **IR** (neat) 2971, 2866, 1726, 1500, 1279, 1148 cm⁻¹; **HRMS** (TOF MS CI⁺) m/z calcd for C₂₅H₂₇BrO₂ (M)⁺ 438.1194, found 438.1187.

Scheme 3.5: Synthesis of starting material for formation of 3.13 (Table 3.2)

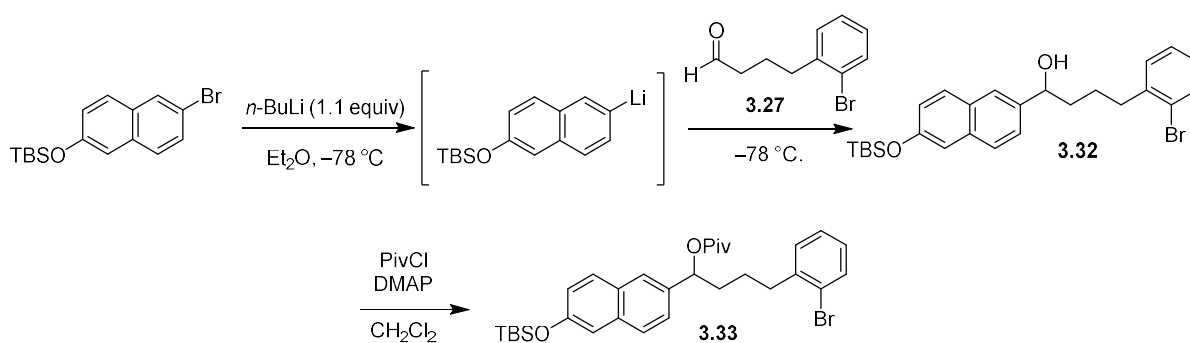


rac-**3.30** The product was prepared according to general method D using 2-bromo-6-(*tert*-butyldimethylsilyloxy) naphthalene (2.02 g, 6.00 mmol), *n*-BuLi (2.6 mL, 2.5 M in hexane, 6.6 mmol), and **3.26** (1.4 g, 6.0 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a yellow oil (1.9 g, 4.0 mmol, 67%).

rac-**3.31** The product was prepared according to general method B using *rac*-**3.30** (0.47 g, 1.0 mmol), pivaloyl chloride (0.13 mL, 1.1 mmol) and dimethylaminopyridine (0.13 g, 1.1 mmol). The product was purified by flash column chromatography (5% EtOAc/hexanes) to afford the title

compound as a colorless oil (0.45 g, 0.81 mmol, 81%). **TLC** R_f = 0.5 (5% EtOAc/hexanes); **^1H NMR** (400 MHz, CDCl_3) δ 7.73–7.66 (m, 3H), 7.50 (dd, J = 8.0, 1.1 Hz, 1H), 7.42 (dd, J = 8.5, 1.7 Hz, 1H), 7.22–7.13 (m, 3H), 7.07 (dd, J = 8.8, 2.4 Hz, 1H), 7.02 (ddd, J = 8.0, 6.9, 2.3 Hz, 1H), 5.91 (dd, J = 8.3, 5.3 Hz, 1H), 2.86 (ddd, J = 14.0, 11.2, 5.0 Hz, 2H), 2.73 (ddd, J = 13.5, 10.7, 5.7 Hz, 2H), 2.35–2.10 (m, 2H), 1.25 (s, 9H), 1.01 (s, 9H), 0.24 (s, 6H); **^{13}C NMR** (100 MHz, CDCl_3) δ 177.8, 153.9, 140.9, 136.1, 134.5, 133.0, 130.5, 129.6, 129.1, 127.9, 127.7, 127.3, 125.5, 124.54, 124.50, 122.5, 114.9, 75.5, 39.1, 36.7, 32.7, 27.4, 25.9, 18.4, -4.2; **IR** (neat) 2956, 2930, 1728, 1605, 1505, 1261, 1152 cm^{-1} ; **HRMS** (TOF MS CI^+) m/z calcd for $\text{C}_{30}\text{H}_{39}\text{BrO}_3\text{Si}$ ($\text{M} + \text{Na}$) $^+$ 554.1852, found 554.1850.

Scheme 3.6: Synthesis of starting material for formation of 3.14 (Table 3.2)

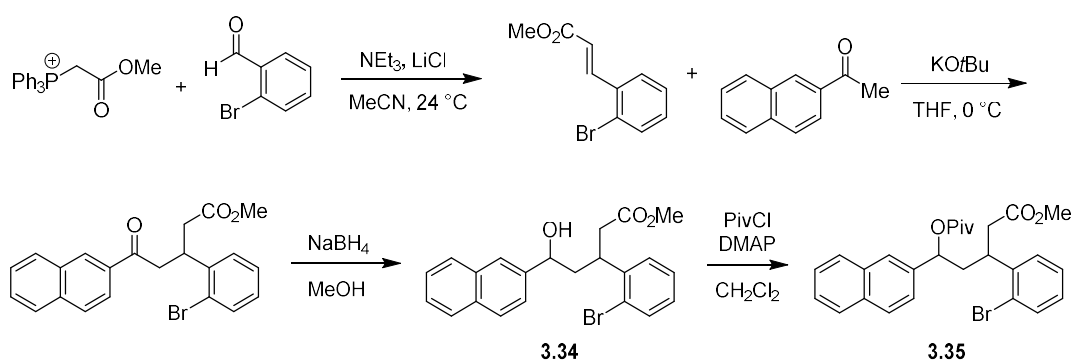


3.32. The product was prepared according to general method D using 2-bromo-6-(*tert*-butyldimethylsilyloxy) naphthalene (1.01 g, 3.00 mmol), *n*-BuLi (1.3 mL, 2.5 M in hexane, 3.3 mmol), and **3.27** (0.68 g, 3.0 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a yellow oil (1.4 g, 2.9 mmol, 98%).

3.33 The product was prepared according to general method B using **3.32** (0.75 g, 1.5 mmol), pivaloyl chloride (0.22 mL, 1.8 mmol) and dimethylaminopyridine (0.22 g, 1.8 mmol). The product was purified by flash column chromatography (5% EtOAc/hexanes) to afford the title

compound as a colorless oil (0.33 g, 0.77 mmol, 96%). **TLC** R_f = 0.5 (5% EtOAc/hexanes); **^1H NMR** (400 MHz, CDCl_3) δ 7.70–7.64 (m, 3H), 7.49 (dd, J = 8.0, 1.3 Hz, 1H), 7.38 (dd, J = 8.6, 1.8 Hz, 1H), 7.19–7.11 (m, 3H), 7.06 (dd, J = 8.9, 2.5 Hz, 1H), 7.00 (ddd, J = 8.0, 7.0, 2.1 Hz, 1H), 5.88 (dd, J = 7.8, 5.6 Hz, 1H), 2.75 (t, J = 7.9 Hz, 2H), 2.11–1.85 (m, 2H), 1.80–1.56 (m, 2H), 1.21 (s, 9H), 1.01 (s, 9H), 0.23 (s, 6H); **^{13}C NMR** (100 MHz, CDCl_3) δ 177.9, 153.9, 141.5, 136.5, 134.5, 133.1, 130.6, 129.7, 129.2, 127.8, 127.7, 127.3, 125.5, 124.7, 122.6, 115.1, 75.8, 39.1, 36.4, 36.1, 27.5, 26.1, 26.0, 18.5, -4.04; **IR** (neat) 2954, 2929, 1726, 1605, 1479, 1260, 1150 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{31}\text{H}_{41}\text{BrO}_3\text{SiNa}$ ($\text{M} + \text{Na}$)⁺ 591.1906, found 591.1910.

Scheme 3.7: Synthesis of starting material for formation of 3.15 (Table 3.2)



3.34 The product was prepared according to a modified procedure reported by Liu.²⁵ A 100 mL round bottom flask charged with LiCl (0.50 g, 10 mmol, 1.0 equiv) and a Teflon stir bar was flame-dried under vacuum. Once cooled to room temperature, the flask was removed from vacuum and charged with MeCN (50 mL), 2-bromobenzaldehyde (1.2 mL, 10 mmol, 1.0 equiv), trimethyl phosphonoacetate (2.00 mL, 12.5 mmol, 1.25 equiv), and triethylamine (1.4 mL, 10 mmol, 1.0 equiv), then sealed and stirred at 24 °C for 12 hr. The reaction mixture was passed through a cake

²⁵ Pan, X.; Liu, Z. *Tetrahedron*, **2014**, 70, 4602

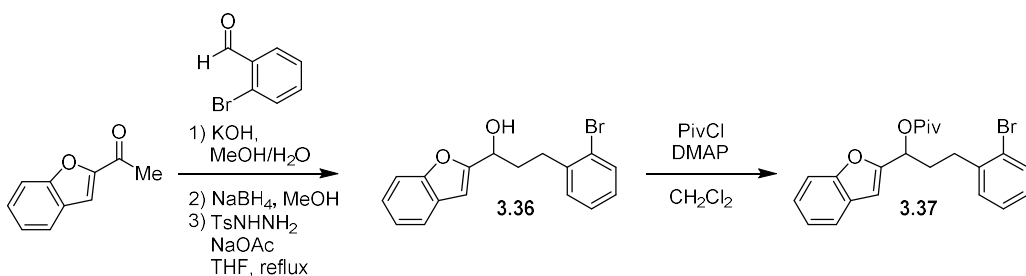
of silica and the solvent was removed under reduced pressure affording a clear, colorless oil that was taken directly to the next step in the synthetic sequence.

A 100 mL round bottom flask containing methyl (*E*)-3-(2-bromophenyl)-acrylate was charged with 2-acetonaphthone (1.7 g, 10 mmol, 1.0 equiv), dissolved in THF (40 mL) and cooled to -5 °C in a brine-ice bath. Following this, potassium *tert*-butoxide (1.2 g, 10 mmol, 1.0 equiv) was added and the reaction was stirred at -5 °C to 0 °C for 3 hrs. The reaction was then quenched with saturated NH₄Cl (20 mL) and extracted with Et₂O (45 mL). The ether layer was then washed with brine and dried over Na₂SO₄, filtered and concentrated under vacuum. The unpurified product was purified by flash column chromatography (gradient: hexanes to 15% Et₂O/hexanes). The fractions containing product were concentrated in a 100 mL round bottom flask, dissolved in MeOH (45 mL), cooled to 0 °C and NaBH₄ (0.45 g, 12 mmol 1.2 equiv) was added. The reaction was allowed to warm to room temperature over the course of 30 min and stirred for an additional 2 hr. The unpurified mixture was then concentrated under reduced pressure and the unpurified oil was passed through a cake of silica with ethyl ether to afford the title compound as a pale yellow oil (1.4 g, 3.3 mmol, 33% over 3 steps).

3.35 The product was prepared according to general method B using **3.34** (1.3 g, 3.3 mmol), pivaloyl chloride (0.45 mL, 3.7 mmol) and dimethylaminopyridine (0.46 g, 3.8 mmol). The product was purified by flash column chromatography (10% Et₂O/hexanes) to afford the title compound as a colorless oil (1.6 g, 3.2 mmol, 99%, 1:1 dr). **TLC** *R_f* = 0.35 (12% EtOAc/hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.84–7.76 (m, 3H), 7.63 (s, 1H), 7.55 (d, *J* = 8.3 Hz, 1H), 7.49–7.43 (m, 2H), 7.40 (dd, *J* = 8.4, 1.8 Hz, 1H), 7.35–7.28 (m, 2H), 7.11–7.06 (m, 1H), 5.79 (t, *J* = 7.3 Hz, 1H), 3.68 (quint, *J* = 7.2 Hz, 1H), 3.56 (s, 3H), 2.68 (dd, *J* = 15.4, 7.2 Hz, 1H), 2.61 (dd, *J*

= 15.5, 7.2 Hz, 1H), 2.49 (dt, $J = 15.4, 8.2$ Hz, 1H), 2.26–2.17 (m, 1H), 1.20 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 177.8, 172.0, 141.9, 137.5, 133.4, 133.27, 133.24, 128.6, 128.4, 128.2, 128.0, 127.8, 126.3, 126.2, 126.1, 124.8, 124.1, 74.0, 51.8, 40.8, 39.7, 38.9, 37.2, 27.2; IR (neat) 3056, 2972, 1728 (b), 1600, 1471, 1278, 1148, 1021 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{27}\text{H}_{29}\text{BrO}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 519.1147, found 519.1132.

Scheme 3.8: Synthesis of starting material for formation of 3.16 (Table 3.2)

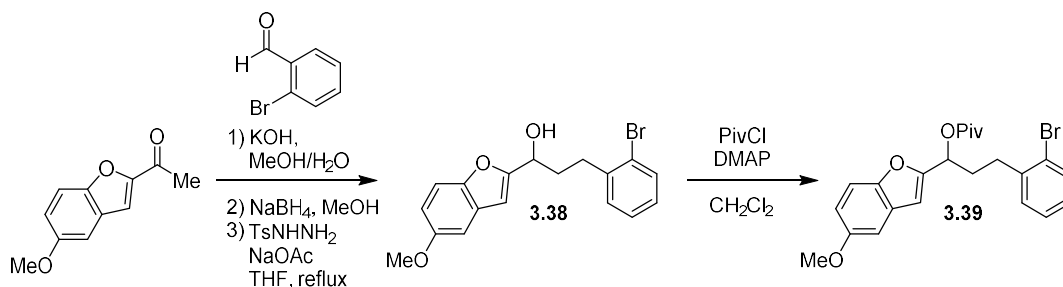


3.36 The product was prepared according to general method A using 2-benzofuran methyl ketone (2.9 g, 18 mmol), 2-bromobenzaldehyde (2.1 mL, 18 mmol), and KOH (2.0 g, 36 mmol). Then NaBH₄ (0.76 g, 20 mmol), tosylhydrazide (7.5 g, 40 mmol) and NaOAc•3H₂O (5.4 g, 40 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a yellow oil (2.1 g, 6.3 mmol, 35% over 3 steps).

3.37 The product was prepared according to general method B using **3.36** (0.66 g, 2.0 mmol), pivaloyl chloride (0.27 mL, 2.2 mmol) and dimethylaminopyridine (0.27 g, 2.2 mmol). The product was purified by flash column chromatography (5% EtOAc/hexanes) to afford the title compound as a colorless oil (0.59 g, 1.4 mmol, 70%). TLC $R_f = 0.5$ (5% EtOAc/hexanes); ^1H NMR (400 MHz, CDCl_3) δ 7.51 (d, $J = 8.3$ Hz, 1H), 7.47 (d, $J = 8.3$ Hz, 1H), 7.44 (d, $J = 8.2$ Hz, 1H), 7.27–7.12 (m, 4H), 7.02–6.94 (m, 1H), 6.67 (s, 1H), 6.04 (t, $J = 6.8$ Hz, 1H), 2.90–2.72 (m, 2H), 2.41–2.27 (m, 2H), 1.25 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 177.8, 155.6, 155.2, 140.6,

133.2, 130.7, 129.8, 128.3, 127.9, 124.74, 124.68, 123.2, 121.5, 111.6, 104.9, 68.8, 39.3, 33.3, 32.4, 27.5; **IR** (neat) 2970, 1729, 1453, 1278, 1142 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{22}\text{H}_{23}\text{BrO}_3\text{Na}$ ($\text{M} + \text{Na}$)⁺ 437.0728, found 437.0725.

Scheme 3.9: Synthesis of starting material for formation of 3.17 (Table 3.2)

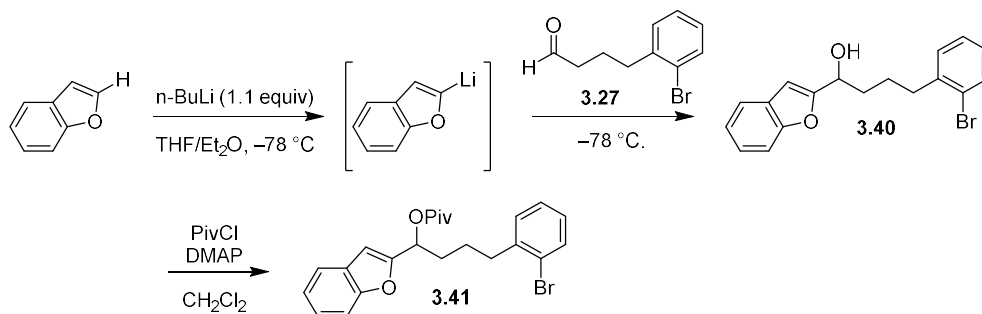


3.38 The product was prepared according to general method A using 2-acyl-5-methoxybenzofuran (2.0 g, 10 mmol), 2-bromobenzaldehyde (1.2 mL, 10 mmol), and KOH (1.2 g, 21 mmol). Then NaBH₄ (0.95 g, 25 mmol), tosylhydrazide (5.4 g, 29 mmol) and NaOAc•3H₂O (3.9 g, 29 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a yellow oil (2.0 g, 5.5 mmol, 55% over 3 steps).

3.39 The product was prepared according to general method B using **3.38** (0.50 g, 1.6 mmol), pivaloyl chloride (0.19 mL, 1.6 mmol) and dimethylaminopyridine (0.19 g, 1.6 mmol). The product was purified by flash column chromatography (12% EtOAc/hexanes) to afford the title compound as a colorless oil (0.52 g, 1.2 mmol, 75%). **TLC** R_f = 0.4 (12% EtOAc/hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.49 (d, J = 8.1 Hz, 1H), 7.33 (d, J = 8.9 Hz, 1H), 7.20–7.16 (m, 2H), 7.01 (ddd, J = 7.9, 6.0, 3.2 Hz, 1H), 6.99 (d, J = 2.6 Hz, 1H), 6.87 (dd, J = 8.9, 2.6 Hz, 1H), 6.62 (t, J = 0.6 Hz, 1H), 6.00 (t, J = 6.6 Hz, 1H), 3.78 (s, 3H), 2.90–2.72 (m, 2H), 2.41–2.26 (m, 2H), 1.25 (s, 9H); **¹³C NMR** (100 MHz, CDCl₃) δ 177.8, 156.34, 156.32, 150.1, 140.6, 133.2, 130.7,

128.8, 128.2, 127.9, 124.6, 113.5, 112.0, 105.0, 103.9, 68.8, 56.1, 39.2, 33.2, 32.3, 27.4; **IR** (neat) 2968, 1729, 1617, 1476, 1204, 1192 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{22}\text{H}_{25}\text{BrO}_4\text{Na}$ ($\text{M} + \text{Na}$)⁺ 467.0834, found 467.0832.

Scheme 3.10: Synthesis of starting material for formation of 3.18 (Table 3.2)

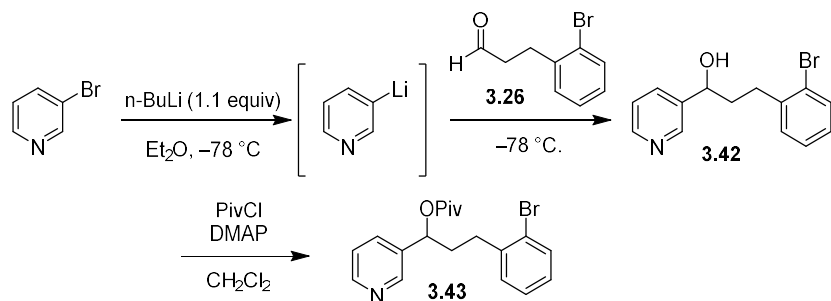


3.40 The product was prepared according to general method D using benzofuran (0.66 mL, 6.0 mmol), *n*-BuLi (2.6 mL, 2.5 M in hexane, 6.6 mmol), and **3.27** (1.0 g, 4.5 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a yellow oil (1.4 g, 3.8 mmol, 84%).

3.41 The product was prepared according to general method B using **3.40** (0.28 g, 0.80 mmol), pivaloyl chloride (0.11 mL, 0.90 mmol) and dimethylaminopyridine (0.11 g, 0.90 mmol). The product was purified by flash column chromatography (5% EtOAc/hexanes) to afford the title compound as a colorless oil (0.33 g, 0.77 mmol, 96%). **TLC** R_f = 0.5 (5% EtOAc/hexanes); **¹H NMR** (400 MHz, CDCl_3) δ 7.51 (d, J = 7.3 Hz, 1H), 7.48 (d, J = 8.1 Hz, 1H), 7.44 (d, J = 8.2 Hz, 1H), 7.24 (t, J = 8.0 Hz, 1H), 7.21–7.11 (m, 3H), 7.00 (td, J = 7.8, 1.8 Hz, 1H), 6.63 (s, 1H), 6.01 (t, J = 6.8 Hz, 1H), 2.77 (t, J = 7.8 Hz, 2H), 2.18–2.03 (m, 2H), 1.80–1.60 (m, 2H), 1.21 (s, 9H); **¹³C NMR** (100 MHz, CDCl_3) δ 177.8, 155.9, 155.1, 141.4, 133.1, 130.6, 128.2, 128.0, 127.7, 124.7, 124.6, 123.1, 121.4, 111.6, 104.6, 69.0, 39.2, 36.0, 32.7, 27.4, 25.7; **IR** (neat) 3057, 2969,

2869, 1728, 1471, 1454, 1278, 1254, 1143 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{23}\text{H}_{25}\text{BrO}_3\text{Na}$ ($\text{M} + \text{Na}$)⁺ 451.0885, found 451.0891.

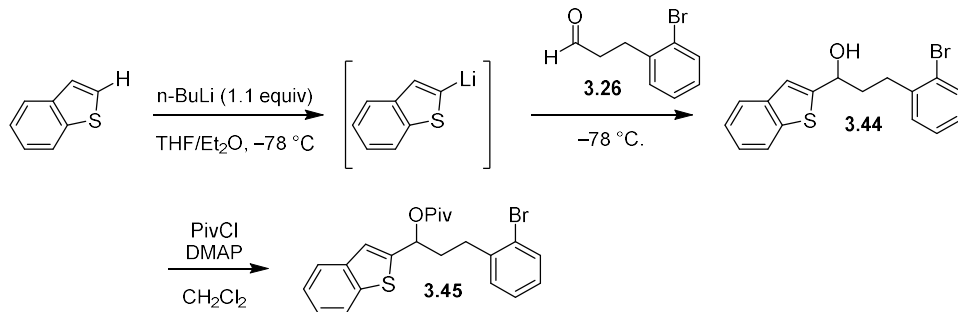
Scheme 3.11: Synthesis of starting material for formation of 3.19 (Table 3.2)



3.42 The product was prepared according to general method D using 3-bromopyridine (0.39 g, 4.0 mmol), *n*-BuLi (1.7 mL, 2.5 M in hexane, 4.4 mmol), and **3.26** (0.85 g, 4.0 mmol) and -78 °C. The product was purified by flash column chromatography (1% NEt₃ in EtOAc) to afford the title compound as a yellow oil (0.72 g, 2.5 mmol, 62%).

3.43 The product was prepared according to general method B using **3.42** (0.58 g, 1.5 mmol), pivaloyl chloride (0.22 mL, 1.8 mmol) and dimethylaminopyridine (0.22 g, 1.8 mmol). The product was purified by flash column chromatography (50% EtOAc/hexanes) to afford the title compound as a colorless oil (0.40 g, 1.1 mmol, 70%). **TLC** R_f = 0.2 (12% EtOAc/hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 8.63 (d, J = 1.4 Hz, 1H), 8.54 (dd, J = 4.9, 1.2 Hz, 1H), 7.65 (dt, J = 8.2, 1.8 Hz, 1H), 7.51 (d, J = 8.0 Hz, 1H), 7.27 (dd, J = 7.3, 4.8 Hz, 1H), 7.22 (t, J = 7.4 Hz, 1H), 7.16 (dd, J = 7.5, 1.5 Hz, 1H), 7.05 (td, J = 7.8, 1.5 Hz, 1H), 5.80 (dd, J = 8.2, 5.1 Hz, 1H), 2.89–2.68 (m, 2H), 2.30–2.05 (m, 2H), 1.25 (s, 9H); **¹³C NMR** (100 MHz, CDCl₃) δ 177.7, 149.5, 148.3, 140.4, 136.4, 134.1, 133.2, 130.5, 128.2, 127.9, 124.5, 123.6, 73.2, 39.1, 36.6, 32.5, 27.4; **IR** (neat) 3055, 2970, 1728, 1471, 1279, 1146 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{19}\text{H}_{22}\text{BrNO}_2\text{Na}$ ($\text{M} + \text{Na}$)⁺ 398.0732, found 398.0746.

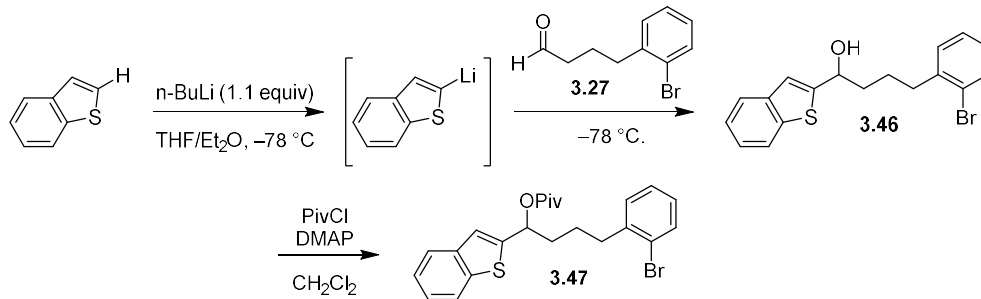
Scheme 3.12: Synthesis of starting material for formation of 3.20 (Table 3.2)



3.44 The product was prepared according to general method D using benzothiophene (0.54 g, 4.0 mmol), *n*-BuLi (1.7 mL, 2.5 M in hexane, 4.4 mmol), and **3.26** (0.85 g, 4.0 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a yellow oil (1.1 g, 3.2 mmol, 79%).

3.45 The product was prepared according to general method B using **3.44** (0.21 g, 0.60 mmol), pivaloyl chloride (0.08 mL, 0.7 mmol) and dimethylaminopyridine (0.080 g, 0.70 mmol). The product was purified by flash column chromatography (5% EtOAc/hexanes) to afford the title compound as a colorless oil (0.21 g, 0.48 mmol, 79%). **TLC** *R_f* = 0.5 (5% EtOAc/hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.84 (d, *J* = 7.7 Hz, 1H), 7.77 (d, *J* = 7.4 Hz, 1H), 7.57 (d, *J* = 8.0 Hz, 1H), 7.40–7.30 (m, 3H), 7.28–7.20 (m, 2H), 7.01 (dd, *J* = 8.1, 2.2 Hz, 1H), 6.21 (dd, *J* = 7.6, 5.7 Hz, 1H), 2.99–2.79 (m, 2H), 2.46–2.27 (m, 2H), 1.32 (s, 9H); **¹³C NMR** (100 MHz, CDCl₃) δ 177.7, 144.4, 140.5, 139.7, 139.5, 133.2, 130.6, 129.8, 128.2, 127.9, 124.7, 124.6, 124.0, 122.7, 122.3, 71.4, 39.2, 36.6, 32.6, 27.4; **IR** (neat) 3058, 2970, 1728, 1477, 1457, 1277, 1140, 1028 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₂H₂₃BrO₂SNa (M + Na)⁺ 453.0500, found 453.0512.

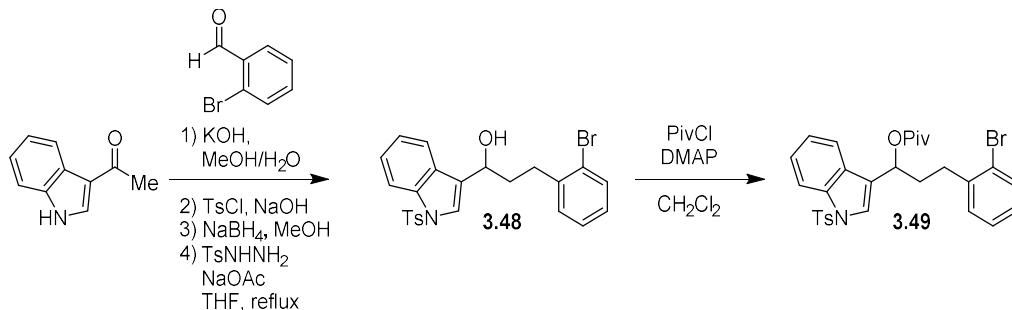
Scheme 3.13: Synthesis of starting material for formation of 2.21 (Table 3.2)



3.46 The product was prepared according to general method D using benzothiophene (0.34 g, 2.5 mmol), *n*-BuLi (1.1 mL, 2.5 M in hexane, 2.2 mmol), and **3.27** (0.45 g, 2.0 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a yellow oil (0.72 g, 2.0 mmol, 99%).

3.47 The product was prepared according to general method B using **3.46** (0.72 g, 2.0 mmol), pivaloyl chloride (0.27 mL, 2.3 mmol) and dimethylaminopyridine (0.27 g, 2.3 mmol). The product was purified by flash column chromatography (12% Et₂O/hexanes) to afford the title compound as a colorless oil (0.46 g, 1.0 mmol, 50%). **TLC** *R_f* = 0.6 (12% Et₂O/hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.81–7.77 (m, 1H), 7.73–7.69 (m, 1H), 7.51 (dd, *J* = 7.9, 1.2 Hz, 1H), 7.35–7.27 (m, 2H), 7.23–7.15 (m, 3H), 7.04 (ddd, *J* = 7.9, 7.0, 2.1 Hz, 1H), 6.12 (dd, *J* = 7.6, 6.0 Hz, 1H), 2.78 (t, *J* = 7.8 Hz, 2H), 2.17–1.96 (m, 2H), 1.82–1.61 (m, 2H), 1.22 (s, 9H); **¹³C NMR** (125 MHz, CDCl₃) δ 177.6, 144.5, 141.1, 139.4, 139.3, 132.9, 130.4, 127.7, 127.5, 124.5, 124.40, 124.35, 123.7, 122.4, 121.8, 71.4, 38.9, 35.9, 35.8, 27.2, 25.7; **IR** (neat) 3054, 2972, 1725, 1438, 1264, 1146 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₃H₂₅BrO₂SNa (M + Na)⁺ 467.0656, found 467.0652.

Scheme 3.14: Synthesis of starting material for formation of 3.22 (Table 3.2)



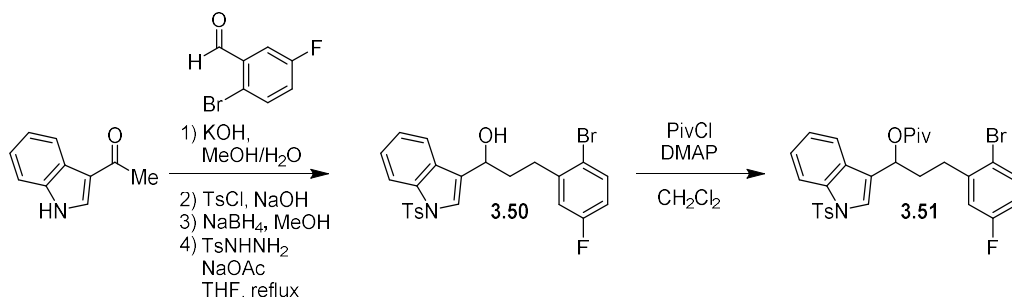
3.48 The product was prepared according to general method A using 3-acetylidole (1.6 g, 10 mmol), 2-bromobenzaldehyde (1.9 g, 10 mmol), and KOH (1.2 g, 22 mmol). The indolyl chalcone was then tosylated according to a modified procedure reported by Carreira. A using tosyl chloride (2.2 g, 12 mmol), TBABr (0.34 g, 0.12 mmol), and NaOH (30% w/v) and benzene.²⁶ The chalcone was then reduced by NaBH₄ (0.38 g, 10 mmol), and tosylhydrazide (7.5 g, 40 mmol) and NaOAc•3H₂O (5.5 g, 40 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a yellow oil (3.6 g, 7.4 mmol, 74% over 4 steps).

3.49 The product was prepared according to general method B using **3.48** (0.78 g, 1.6 mmol), pivaloyl chloride (0.25 mL, 2.0 mmol) and dimethylaminopyridine (0.25 g, 2.0 mmol). The product was purified by flash column chromatography (5% EtOAc/hexanes) to afford the title product as a colorless oil (0.86 g, 1.5 mmol, 94%). **TLC** R_f = 0.6 (15% EtOAc/hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 8.00 (d, *J* = 8.4 Hz, 1H), 7.71 (d, *J* = 8.2 Hz, 2H), 7.62 (d, *J* = 6.5 Hz, 2H), 7.43 (d, *J* = 7.8 Hz, 1H), 7.29 (t, *J* = 7.9 Hz, 1H), 7.20 (t, *J* = 7.7 Hz, 1H), 7.17–7.06 (m, 4H), 6.96 (td, *J* = 7.7, 1.7 Hz, 1H), 6.14 (t, *J* = 6.6 Hz, 1H), 2.84–2.64 (m, 2H), 2.39–2.19 (m, 2H), 2.17 (s, 3H), 1.21 (s, 9H); **¹³C NMR** (100 MHz, CDCl₃) δ 177.8, 145.3, 140.6, 135.7, 135.3, 133.1, 130.6,

²⁶ Waser, J.; Gaspar, B.; Nambu, H.; Carreira, E. M. *J. Am. Chem. Soc.*, **2006**, *128*, 11693.

130.2, 129.1, 128.2, 127.9, 127.0, 125.3, 124.6, 124.3, 123.7, 122.3, 120.6, 114.2, 68.9, 39.2, 35.1, 32.7, 27.5, 21.7; **IR** (neat) 3053, 2972, 1725, 1596, 1446, 1370, 1174, 1152, 1122 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{29}\text{H}_{30}\text{BrNO}_4\text{SNa}$ ($M + \text{Na}$)⁺ 590.0977, found 590.0964.

Scheme 3.15: Synthesis of starting material for formation of 3.23 (Table 3.2)

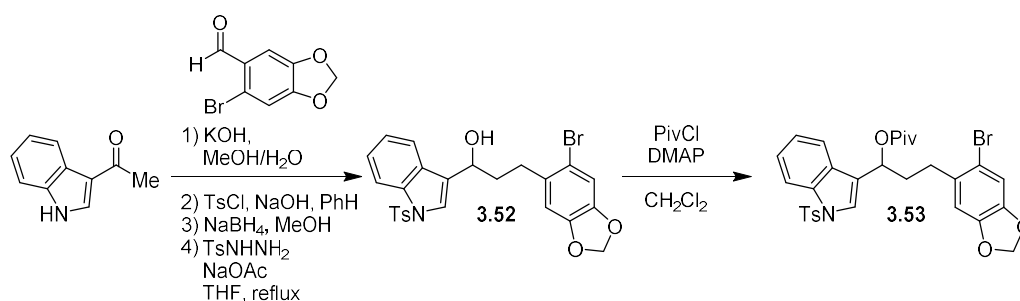


3.50 The product was prepared according to general method A using 3-acetylindole (1.6 g, 10 mmol), 2-bromo-5-fluorobenzaldehyde (2.0 g, 10 mmol), and KOH (1.4 g, 23 mmol). The indolyl chalcone was then tosylated according to a modified procedure reported by Carreira using tosyl chloride (2.2 g, 12 mmol), TBABr (0.34 g, 0.12 mmol), and NaOH (30% w/v) and benzene. The chalcone was then reduced by NaBH₄ (0.40 g, 10 mmol), and tosylhydrazide (7.5 g, 40 mmol) and NaOAc•3H₂O (5.5 g, 40 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a yellow oil (3.6 g, 7.4 mmol, 64% over 4 steps).

3.51 The product was prepared according to general method B using **3.50** (0.75 g, 1.5 mmol), pivaloyl chloride (0.25 mL, 2.0 mmol) and dimethylaminopyridine (0.24 g, 2.0 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title product as a yellow oil (0.66 g, 1.1 mmol, 73%). **TLC** R_f = 0.6 (15% EtOAc/hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.98 (d, J = 8.3 Hz, 1H), 7.74 (d, J = 8.3 Hz, 1H), 7.61 (d, J = 8.0 Hz, 1H), 7.69 (s, 1H), 7.42 (dd, J = 8.7, 6.3 Hz, 1H), 7.32 (t, J = 7.7 Hz, 1H), 7.23 (t, J = 7.6 Hz, 1H), 7.18 (d, J = 8.1 Hz, 2H), 6.85 (dd, J = 9.3, 3.0 Hz, 1H), 6.77 (td, J = 8.3, 3.0 Hz, 1H), 6.11 (t, J = 6.6

Hz, 1H), 2.80–2.61 (m, 2H), 2.36–2.17 (m, 5H), 1.21 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 177.8, 162.1 (d, $J = 247$ Hz), 145.3, 142.7 (d, $J = 7$ Hz), 135.7, 135.3, 134.2 (d, $J = 8$ Hz), 130.1, 129.0, 127.0, 125.2, 124.2, 123.6, 122.0, 120.5, 118.6 (d, $J = 3$ Hz), 117.3 (d, $J = 22$ Hz), 115.3 (d, $J = 22$ Hz), 114.1, 68.8, 39.2, 34.7, 32.7, 27.4, 21.8; IR (neat) 3068, 2972, 1726, 1598, 1370, 1174, 1121 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{29}\text{H}_{29}\text{BrFNO}_4\text{SNa}$ ($\text{M} + \text{Na}$) $^+$ 608.0883, found 608.0860.

Scheme 3.16: Synthesis of starting material for formation of 3.24 (Table 3.2)



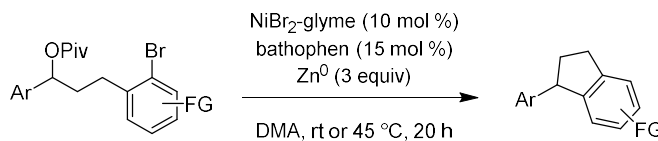
3.52 The product was prepared according to general method A using 3-acetylindole (1.6 g, 10 mmol), 2-bromo-5-fluorobenzaldehyde (2.3 g, 10 mmol), and KOH (1.4 g, 23 mmol). The indolyl chalcone was then tosylated according to a modified procedure reported by Carreira using tosyl chloride (2.2 g, 12 mmol), TBABr (0.34 g, 0.12 mmol), and NaOH (30% w/v) and benzene. The chalcone was then reduced by NaBH_4 (0.40 g, 10 mmol), and tosylhydrazide (7.5 g, 40 mmol) and $\text{NaOAc}\cdot 3\text{H}_2\text{O}$ (5.5 g, 40 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a yellow oil (3.6 g, 7.4 mmol, 11% over 4 steps).

3.53 The product was prepared according to general method B using **3.52** (0.60 g, 1.1 mmol), pivaloyl chloride (0.16 mL, 1.3 mmol) and dimethylaminopyridine (0.16 g, 1.3 mmol). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title product as a yellow oil (0.25 g, 0.41 mmol, 36%). TLC $R_f = 0.5$ (15% EtOAc/hexanes); ^1H NMR

(400 MHz, CDCl₃) δ 7.97 (d, J = 8.2 Hz, 1H), 7.73 (d, J = 8.4 Hz, 2H), 7.61 (d, J = 7.6 Hz, 1H), 7.58 (s, 1H), 7.30 (td, J = 8.4, 1.3 Hz, 1H), 7.22 (td, J = 8.1, 1.2 Hz, 1H), 7.17 (d, J = 8.0 Hz, 2H), 6.93 (s, 1H), 6.59 (s, 1H), 6.08 (t, J = 6.6 Hz, 1H), 5.89 (s, 2H), 2.73–2.54 (m, 2H), 2.32–2.11 (m, 5H), 1.20 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 177.8, 147.7, 147.1, 145.3, 135.7, 135.3, 133.5, 130.1, 129.0, 127.0, 125.2, 124.2, 123.6, 122.2, 120.6, 114.4, 114.1, 113.0, 110.1, 101.9, 68.8, 39.2, 35.2, 32.6, 27.4, 21.8; IR (neat) 2971, 1724, 1476, 1369, 1174, 1121 cm⁻¹; HRMS (TOF MS ES+) m/z calcd for C₃₀H₃₀BrNO₆SNa (M + Na)⁺ 634.0875, found 634.0860.

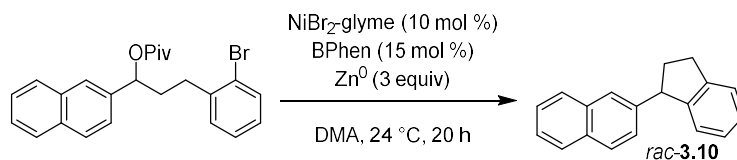
GENERAL PROCEDURES FOR REDUCTIVE CROSS-ELECTROPHILE COUPLING REACTIONS

METHOD E: INTRAMOLECULAR REDUCTIVE COUPLING REACTIONS

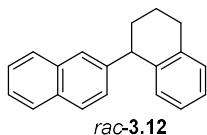


In a glovebox, a flame-dried 7 mL vial equipped with a stir bar was charged with NiBr₂•glyme (10 or 15 mol %), bathophenanthroline (15 mol %), Zn⁰ (3 equiv), DMA (0.60 mL), and substrate (1.00 equiv). The reaction was stirred for 20 h before removing the vial from the glovebox. The reaction mixture was eluted through a silica plug (with Et₂O) and concentrated under reduced pressure. The product was purified by flash column chromatography.

CHARACTERIZATION OF PRODUCTS IN TABLE 3.2

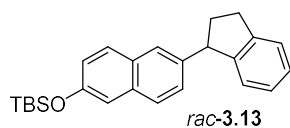


rac-3.10. The product was prepared according to general method E using NiBr₂•glyme (6.2 mg, 0.020 mmol, 10 mol %), bathophenanthroline (10.0 mg, 0.030 mmol, 15 mol %), Zn⁰ (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and *rac*-3.9 (85.1 mg, 0.200 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et₂O) and concentrated under reduced pressure. The product was purified by flash column chromatography (100% hexanes) to afford the title compound as a colorless oil (44.0 mg, 0.180 mmol, 90%). **TLC** R_f = 0.3 (100% hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.80–7.71 (m, 3H), 7.63 (s, 1H), 7.44–7.36 (m, 2H), 7.30 (d, *J* = 7.5 Hz, 1H), 7.26 (dd, *J* = 8.4, 1.6 Hz, 1H), 7.19 (t, *J* = 7.3 Hz, 1H), 7.11 (t, *J* = 8.1 Hz, 1H), 6.95 (d, *J* = 7.6 Hz, 1H), 4.48 (t, *J* = 8.3 Hz, 1H), 3.12–2.92 (m, 2H), 2.65–2.55 (m, 1H), 2.19–2.07 (m, 1H); **¹³C NMR** (100 MHz, CDCl₃) δ 147.1, 144.7, 143.1, 133.9, 132.7, 128.6, 127.98, 127.96, 126.99, 126.95, 126.82, 126.78, 126.3, 125.7, 125.4, 124.8, 52.1, 36.8, 32.3; **IR** (neat) 3050, 3018, 2937, 1599, 1506, 1477, 1457 cm⁻¹; **HRMS** (TOF MS CI⁺) *m/z* calcd for C₁₉H₁₆ (M)⁺ 224.1252, found 224.1257.



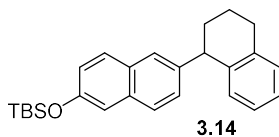
rac-3.12. The product was prepared according to general method E using NiBr₂•glyme (3.1 mg, 0.010 mmol, 10 mol %), bathophenanthroline (5.0 mg, 0.015 mmol, 15 mol %), Zn⁰ (18.8 mg, 0.300 mmol, 3 equiv), DMA (0.40 mL), and 3.29 (44.0 mg, 0.100 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et₂O) and concentrated under reduced pressure. The

product was purified by flash column chromatography (10% EtOAc/hexanes) to afford the title compound as a colorless oil (16 mg, 0.062 mmol, 62%). **TLC** R_f = 0.3 (hexanes); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 7.84–7.73 (m, 3H), 7.53 (s, 1H), 7.49–7.41 (m, 2H), 7.29–7.23 (m, 1H), 7.21–7.11 (m, 2H), 7.02 (t, J = 7.4 Hz, 1H), 6.86 (d, J = 7.8 Hz, 1H), 4.28 (t, J = 7.0 Hz, 1H), 3.02–2.84 (m, 2H), 2.27–2.18 (m, 1H), 2.03–1.90 (m, 2H), 1.86–1.75 (m, 1H); **$^{13}\text{C NMR}$** (125 MHz, CDCl_3) δ 145.1, 139.4, 137.8, 133.6, 132.3, 130.5, 129.2, 128.1, 127.79, 127.77, 127.5, 127.4, 126.2, 126.1, 125.9, 125.5, 46.0, 33.3, 30.0, 21.4; **IR** (neat) 3053, 2925, 2854, 1599, 1448 cm^{-1} ; **HRMS** (TOF MS CI^+) m/z calcd for $\text{C}_{20}\text{H}_{18}$ (M) $^+$ 258.1408, found 258.1415.

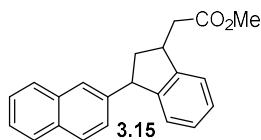


rac-3.13. The product was prepared according to general method E using $\text{NiBr}_2 \cdot \text{glyme}$ (6.2 mg, 0.020 mmol, 10 mol %), bathophenanthroline (10.0 mg, 0.030 mmol, 15 mol %), Zn^0 (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and **3.31** (111 mg, 0.200 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et_2O) and concentrated under reduced pressure. The product was purified by flash column chromatography (10% EtOAc/hexanes) to afford the title compound as a colorless oil (51.7 mg, 0.138 mmol, 69%). **TLC** R_f = 0.3 (hexanes); **$^1\text{H NMR}$** (400 MHz, CDCl_3) δ 7.64 (dd, J = 8.6, 6.8 Hz, 2H), 7.57 (s, 1H), 7.31 (d, J = 7.5 Hz, 1H), 7.23–7.10 (m, 4H), 7.05 (dd, J = 8.8, 2.5 Hz, 1H), 6.97 (d, J = 7.5 Hz, 1H), 4.47 (t, J = 8.4 Hz, 1H), 3.13–2.93 (m, 2H), 2.66–2.57 (m, 1H), 2.13 (dq, J = 12.7, 8.8 Hz, 1H), 1.02 (s, 9H), 0.23 (s, 6H); **$^{13}\text{C NMR}$** (125 MHz, CDCl_3) δ 153.3, 147.1, 144.6, 140.8, 133.6, 129.5, 129.2, 127.2, 127.1, 126.8, 126.6, 126.4, 125.2, 124.6, 122.4, 115.0, 51.8, 36.7, 32.1, 25.9, 18.5, -4.1; **IR** (neat) 2954, 2928,

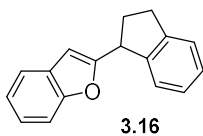
1602, 1478, 1258 cm^{-1} ; **HRMS** (TOF MS CI^+) m/z calcd for $\text{C}_{25}\text{H}_{30}\text{OSi}$ (M)⁺ 374.2066, found 374.2054.



3.14. The product was prepared according to general method E using $\text{NiBr}_2\cdot\text{glyme}$ (3.1 mg, 0.010 mmol, 5.0 mol %), bathophenanthroline (5.0 mg, 0.015 mmol, 7.5 mol %), Zn^0 (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and **3.33** (113.9 mg, 0.200 mmol, 1.00 equiv). After 8 hours, the reaction vial was taken out of the oil bath. In the glovebox, an additional amount of $\text{NiBr}_2\cdot\text{glyme}$ (3.1 mg, 0.010 mmol, 5 mol %) and bathophenanthroline (5.0 mg, 0.015 mmol, 7.5 mol %) was added to the reaction mixture. The reaction vial was put back into the oil bath for the remainder of the reaction time. The reaction mixture was eluted through a silica plug (with Et_2O) and concentrated under reduced pressure. The product was purified by flash column chromatography (100% pentane) to afford the title compound as a colorless oil (29.8 mg, 0.120 mmol, 58%). **TLC** R_f = 0.2 (pentane); **^1H NMR** (400 MHz, CDCl_3) δ 7.61 (dd, J = 8.6, 6.9 Hz, 2H), 7.45 (s, 1H), 7.18 (dd, J = 8.6, 1.8 Hz, 1H), 7.16–7.10 (m, 3H), 7.04 (dd, J = 8.8, 2.4 Hz, 1H), 7.04–6.99 (m, 1H), 6.87 (d, J = 7.7 Hz, 1H), 4.23 (t, J = 6.7 Hz, 1H), 3.02–2.82 (m, 2H), 2.26–2.15 (m, 1H), 2.00–1.88 (m, 2H), 1.84–1.73 (m, 1H), 1.01 (s, 9H), 0.23 (s, 6H); **^{13}C NMR** (100 MHz, CDCl_3) δ 153.2, 142.8, 139.5, 137.7, 133.3, 130.4, 129.3, 129.04, 129.02, 127.6, 127.2, 126.8, 126.0, 125.7, 122.1, 114.8, 45.7, 33.2, 29.9, 25.8, 21.2, 18.3, -4.3; **IR** (neat) 3053, 2928, 1633, 1602, 1478, 1259 cm^{-1} ; **HRMS** (TOF MS CI^+) m/z calcd for $\text{C}_{26}\text{H}_{32}\text{OSi}$ (M)⁺ 388.2222, found 388.2227.

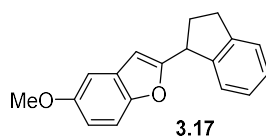


3.15. The product was prepared according to general method E using NiBr₂•glyme (6.2 mg, 0.020 mmol, 10 mol %), bathophenanthroline (10.0 mg, 0.030 mmol, 15 mol %), Zn⁰ (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and **3.35** (99.5 mg, 0.200 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et₂O) and concentrated under reduced pressure. The product was purified by flash column chromatography (10% EtOAc/hexanes) to afford the title compound as a colorless oil (29.7 mg, 0.094 mmol, 47%, 1:1 dr). **TLC** R_f = 0.6 (10% EtOAc in hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.82–7.73 (m, 3H), 7.60–7.57 (m, 1H), 7.47–7.40 (m, 2H), 7.30 (d, *J* = 7.5 Hz, 1H), 7.27–7.22 (m, 2H), 7.18 (tdd, *J* = 7.5, 1.5, 0.5 Hz, 1H), 7.01 (d, *J* = 7.5 Hz, 1H), 4.62 (t, *J* = 7.7 Hz, 1H), 3.87–3.78 (m, 1H), 3.72 (s, 3H), 2.76 (dd, *J* = 15.5, 6.1 Hz, 1H), 2.57 (dd, *J* = 15.4, 8.9 Hz, 1H), 2.51–2.36 (m, 2H); **¹³C NMR** (100 MHz, CDCl₃) δ 173.3, 146.4, 146.3, 142.6, 133.7, 132.6, 128.5, 127.86, 127.85, 127.6, 127.4, 126.7, 126.5, 126.2, 125.7, 124.1, 123.1, 51.9, 49.9, 42.5, 40.6, 40.2; **IR** (neat) 3051, 2949, 1734, 1599, 1435, 1250, 1164 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₂H₂₀O₂Na (M + Na)⁺ 339.1361, found 339.1367.

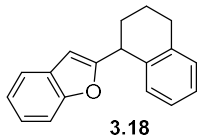


3.16. The product was prepared according to general method E using NiBr₂•glyme (6.2 mg, 0.020 mmol, 10 mol %), bathophenanthroline (10.0 mg, 0.030 mmol, 15 mol %), Zn⁰ (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and **3.37** (83.1 mg, 0.200 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (neat Et₂O) and concentrated under reduced pressure. The product was purified by flash column chromatography (100% hexanes) to afford the title

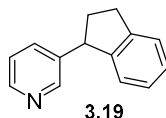
compound as a colorless oil (35.1 mg, 0.150 mmol, 75%). **TLC** R_f = 0.7 (5% EtOAc/hexanes); **^1H NMR** (400 MHz, CDCl_3) δ 7.54–7.50 (m, 1H), 7.48–7.44 (m, 1H), 7.36–7.29 (m, 2H), 7.27–7.19 (m, 4H), 6.43 (s, 1H), 4.61 (t, J = 7.7 Hz, 1H), 3.19–2.98 (m, 2H), 2.66–2.37 (m, 2H); **^{13}C NMR** (100 MHz, CDCl_3) δ 161.1, 155.2, 144.2, 143.5, 129.0, 127.5, 126.7, 125.1, 124.9, 123.6, 122.7, 120.7, 111.2, 102.4, 45.1, 32.4, 31.8; **IR** (neat) 3066, 3022, 2942, 1597, 1583, 1453, 1252 cm^{-1} ; **HRMS** (TOF MS CI^+) m/z calcd for $\text{C}_{17}\text{H}_{14}\text{ONH}_4$ ($\text{M} + \text{NH}_4$) $^+$ 252.1388, found 252.1386.



3.17. The product was prepared according to general method E using $\text{NiBr}_2 \cdot \text{glyme}$ (6.2 mg, 0.020 mmol, 10 mol %), bathophenanthroline (10.0 mg, 0.030 mmol, 15 mol %), Zn^0 (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and **3.39** (89.0 mg, 0.200 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et_2O) and concentrated under reduced pressure. The product was purified by flash column chromatography (100% hexanes) to afford the title compound as a colorless oil (41.2 mg, 0.156 mmol, 79%). **TLC** R_f = 0.3 (6% Et_2O /hexanes); **^1H NMR** (400 MHz, CDCl_3) δ 7.31–7.26 (m, 2H), 7.25–7.14 (m, 3H), 6.95 (d, J = 2.6 Hz, 1H), 6.81 (dd, J = 8.9, 2.6 Hz, 1H), 6.34–6.32 (m, 1H), 4.53 (t, J = 7.7 Hz, 1H), 3.82 (s, 3H), 3.14–2.93 (m, 2H), 2.61–2.51 (m, 1H) 2.41–2.30 (m, 1H); **^{13}C NMR** (100 MHz, CDCl_3) δ 162.0, 156.0, 150.2, 144.2, 143.5, 129.5, 127.5, 126.7, 125.0, 124.9, 112.0, 111.5, 103.5, 102.6, 56.2, 45.1, 32.3, 31.8; **IR** (neat) 3068, 2929, 1615, 1599, 1474, 1202, 1030 cm^{-1} ; **HRMS** (TOF MS CI^+) m/z calcd for $\text{C}_{18}\text{H}_{16}\text{O}_2\text{NH}_4$ ($\text{M} + \text{NH}_4$) $^+$ 282.1494, found 282.1494.

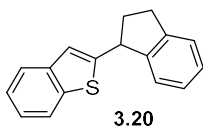


3.18. The product was prepared according to general method E using NiBr₂•glyme (6.2 mg, 0.020 mmol, 10 mol %), bathophenanthroline (10.0 mg, 0.030 mmol, 15 mol %), Zn⁰ (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and **3.41** (85.9 mg, 0.200 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et₂O) and concentrated under reduced pressure. The product was purified by flash column chromatography (100% hexanes) to afford the title compound as a colorless oil (29.8 mg, 0.120 mmol, 60%). **TLC** R_f = 0.8 (5% EtOAc/hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.44 (t, *J* = 8.3 Hz, 2H), 7.23 (td, *J* = 7.7, 1.4 Hz, 1H), 7.20–7.09 (m, 5H), 6.19 (s, 1H), 4.33 (t, *J* = 5.7 Hz, 1H), 2.91–2.78 (m, 2H), 2.35–2.08 (m, 2H), 1.95–1.75 (m, 2H); **¹³C NMR** (100 MHz, CDCl₃) δ 162.6, 155.5, 137.6, 136.0, 130.1, 129.6, 128.9, 126.9, 125.9, 123.5, 122.7, 120.6, 111.2, 104.3, 39.9, 29.6, 28.7, 20.5; **IR** (neat) 3064, 2926, 2858, 1453, 1253 cm⁻¹; **HRMS** (TOF MS CI⁺) *m/z* calcd for C₁₈H₁₆OH (M + H)⁺ 249.1279, found 249.1276.

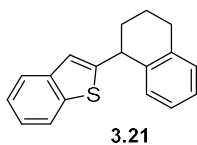


3.19. The product was prepared according to general method E using NiBr₂•glyme (9.2 mg, 0.030 mmol, 15 mol %), bathophenanthroline (10.0 mg, 0.030 mmol, 15 mol %), Zn⁰ (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and **3.43** (75.3 mg, 0.200 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et₂O) and concentrated under reduced pressure. The product was purified by flash column chromatography (100% hexanes) to afford the title compound as a colorless oil (25.4 mg, 0.130 mmol, 65%). **TLC** R_f = 0.1 (12% EtOAc: hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 8.52 (d, *J* = 1.8 Hz, 1H), 8.50 (dd, *J* = 4.9, 1.7 Hz, 1H), 7.45 (dt, *J* = 7.9, 2.0 Hz, 1H), 7.32 (d, *J* = 7.4 Hz, 1H), 7.25–7.20 (m, 2H), 7.16 (t, *J* = 7.4 Hz, 1H), 6.94 (d,

$J = 7.6$ Hz, 1H), 4.37 (t, $J = 8.2$ Hz, 1H), 3.13–2.95 (m, 2H), 2.63 (dtd, $J = 12.8, 7.9, 7.9, 3.9$ Hz, 1H), 2.08 (dq, $J = 12.7, 8.7$ Hz, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 150.0, 148.0, 145.7, 144.3, 140.8, 135.3, 127.0, 126.6, 124.7, 124.6, 123.6, 49.0, 36.5, 31.9; IR (neat) 3020, 2925, 1574, 1478, 1423 cm^{-1} ; HRMS (TOF MS CI^+) m/z calcd for $\text{C}_{14}\text{H}_{13}\text{NH}$ ($\text{M} + \text{H}$) $^+$ 196.1126, found 196.1119.

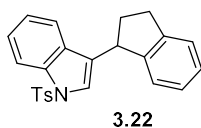


3.20. The product was prepared according to general method E using $\text{NiBr}_2 \cdot \text{glyme}$ (6.2 mg, 0.020 mmol, 10 mol %), bathophenanthroline (10.0 mg, 0.030 mmol, 15 mol %), Zn^0 (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and **3.45** (86.3 mg, 0.200 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et_2O) and concentrated under reduced pressure. The product was purified by flash column chromatography (100% pentane) to afford the title compound as a colorless oil (37.6 mg, 0.150 mmol, 75%). TLC $R_f = 0.5$ (pentane); ^1H NMR (400 MHz, CDCl_3) δ 7.74 (d, $J = 7.8$ Hz, 1H), 7.65 (d, $J = 8.0$ Hz, 1H), 7.32–7.14 (m, 6H), 7.06 (s, 1H), 4.68 (t, $J = 7.8$ Hz, 1H), 3.14–2.90 (m, 2H), 2.66 (dtd, $J = 12.6, 7.9, 4.6$ Hz, 1H), 2.25 (dq, $J = 12.7, 7.8$ Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 150.2, 145.6, 143.9, 140.2, 139.7, 127.4, 126.8, 125.2, 124.8, 124.4, 123.9, 123.2, 122.5, 120.9, 47.4, 36.6, 31.9; IR (neat) 3060, 2922, 1567, 1475, 1456, 1133 cm^{-1} ; HRMS (TOF MS CI^+) m/z calcd for $\text{C}_{17}\text{H}_{14}\text{OH}$ ($\text{M} + \text{H}$) $^+$ 251.0894, found 251.0894.

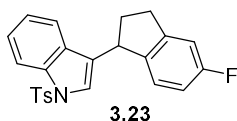


3.21. The product was prepared according to general method E using $\text{NiBr}_2 \cdot \text{glyme}$ (9.2 mg, 0.030 mmol, 15 mol %), bathophenanthroline (10.0 mg, 0.030 mmol, 15 mol %), Zn^0 (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and **3.47** (89.0 mg, 0.200 mmol, 1.00 equiv). The reaction

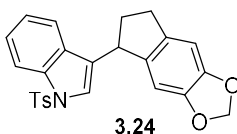
mixture was eluted through a silica plug (with Et₂O) and concentrated under reduced pressure. The product was purified by flash column chromatography (100% hexanes) to afford the title compound as a colorless oil (29.1 mg, 0.110 mmol, 55%). **TLC** *R_f* = 0.5 (hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.74 (d, *J* = 7.8 Hz, 1H), 7.63 (d, *J* = 7.6 Hz, 1H), 7.32–7.27 (m, 1H), 7.25–7.08 (m, 5H), 6.87 (s, 1H), 4.46 (t, *J* = 5.9 Hz, 1H), 2.96–2.79 (m, 2H), 2.29–2.07 (m, 2H), 2.00–1.75 (m, 2H); **¹³C NMR** (125 MHz, CDCl₃) δ 152.0, 139.8, 139.6, 137.9, 137.1, 130.2, 129.3, 126.6, 125.8, 124.1, 123.6, 123.0, 122.3, 122.1, 41.2, 32.8, 29.4, 20.4; **IR** (neat) 3060, 2915, 1456, 1435, 1308, 1129 cm⁻¹; **HRMS** (TOF MS CI⁺) *m/z* calcd for C₁₈H₁₆S (M)⁺ 264.0973, found 264.0966.



3.22. The product was prepared according to general method E using NiBr₂•glyme (6.2 mg, 0.020 mmol, 10 mol %), bathophenanthroline (10.0 mg, 0.030 mmol, 15 mol %), Zn⁰ (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and **3.49** (114 mg, 0.200 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et₂O) and concentrated under reduced pressure. The product was purified by flash column chromatography (5% EtOAc/hexanes) to afford the title compound as a colorless oil (59.4 mg, 0.153 mmol, 75%). **TLC** *R_f* = 0.4 (5% EtOAc/hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.98 (d, *J* = 8.4 Hz, 1H), 7.72 (d, *J* = 8.4 Hz, 2H), 7.34–7.26 (m, 3H), 7.25–7.09 (m, 6H), 7.01 (d, *J* = 7.6 Hz, 1H), 4.53 (t, *J* = 8.1 Hz, 1H), 3.08–2.92 (m, 2H), 2.55 (dtd, *J* = 12.4, 7.8, 4.6 Hz, 1H), 2.32 (s, 3H), 2.12 (dq, *J* = 12.6, 8.4 Hz, 1H); **¹³C NMR** (100 MHz, CDCl₃) δ 144.99, 144.98, 144.2, 136.0, 135.5, 130.6, 130.0, 127.2, 127.0, 126.7, 126.4, 124.90, 124.86, 124.8, 123.3, 123.2, 120.4, 114.1, 42.3, 34.2, 31.9, 21.8; **IR** (neat) 3065, 2926, 2850, 1596, 1446, 1368, 1171 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₄H₂₁NO₂SNa (M + Na)⁺ 410.1191, found 410.1193.



3.23. The product was prepared according to general method E using NiBr₂•glyme (6.2 mg, 0.020 mmol, 10 mol %), bathophenanthroline (10.0 mg, 0.030 mmol, 15 mol %), Zn⁰ (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and **3.51** (117.3 mg, 0.200 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et₂O) and concentrated under reduced pressure. The product was purified by flash column chromatography (5% EtOAc/hexanes) to afford the title compound as a colorless oil (68.1 mg, 0.168 mmol, 84%). **TLC** R_f = 0.4 (5% EtOAc/hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.98 (d, *J* = 8.6 Hz, 1H), 7.73 (d, *J* = 8.4 Hz, 2H), 7.32–7.27 (m, 2H), 7.23–7.20 (m, 3H), 7.16 (td, *J* = 7.7, 0.9 Hz, 1H), 7.01–6.97 (m, 1H), 6.92 (dd, *J* = 8.3, 5.4 Hz, 1H), 6.81 (td, *J* = 9.0, 2.3 Hz, 1H), 4.49 (t, *J* = 8.1 Hz, 1H), 3.05–2.91 (m, 2H), 2.57 (dtd, *J* = 12.6, 8.0, 4.7 Hz, 1H), 2.35 (s, 3H), 2.17 (dq, *J* = 12.7, 8.3 Hz, 1H); **¹³C NMR** (100 MHz, CDCl₃) δ 162.3 (d, *J* = 243 Hz), 146.2 (d, *J* = 8 Hz), 144.8, 140.2 (d, *J* = 2 Hz), 135.8, 135.3, 130.2, 129.9, 126.8, 125.5 (d, *J* = 9 Hz), 126.0, 124.7, 123.1, 123.0, 120.1 113.9, 113.4 (d, *J* = 22 Hz), 111.7 (d, *J* = 22 Hz), 41.4, 34.4, 31.7, 21.6; **IR** (neat) 2923, 1595, 1482, 1368, 1172, 1121 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₄H₂₀FNO₂SNa (M + Na)⁺ 428.1096, found 428.1081.

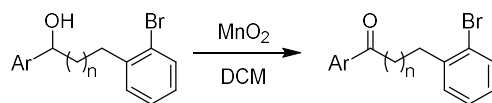


3.24. The product was prepared according to general method E using NiBr₂•glyme (6.2 mg, 0.020 mmol, 10 mol %), bathophenanthroline (10.0 mg, 0.030 mmol, 15 mol %), Zn⁰ (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and **3.53** (122.5 mg, 0.200 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et₂O) and concentrated under reduced pressure. The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title

compound as a colorless oil (38.0 mg, 0.088 mmol, 44%). **TLC** R_f = 0.5 (15% EtOAc/hexanes); **$^1\text{H NMR}$** (400 MHz, CDCl_3) δ 7.98 (d, J = 8.3 Hz, 1H), 7.72 (d, J = 8.3 Hz, 2H), 7.35–7.26 (m, 2H), 7.24–7.12 (m, 4H), 6.76 (s, 1H), 6.43 (s, 1H), 5.92 (d, J = 8.3 Hz, 2H), 4.43 (t, J = 7.7 Hz, 1H), 2.97–2.81 (m, 2H), 2.60–2.48 (m, 1H), 2.33 (s, 3H), 2.13 (dq, J = 12.7, 7.9 Hz, 1H); **$^{13}\text{C NMR}$** (100 MHz, CDCl_3) δ 147.3, 146.9, 145.0, 137.7, 137.0, 136.1, 135.5, 130.5, 130.0, 127.0, 126.6, 124.9, 123.3, 123.2, 120.4, 114.1, 105.33, 105.31, 101.2, 42.2, 34.6, 31.7, 21.8; **IR** (neat) 2926, 1596, 1474, 1367, 1172, 1120, 1021 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{25}\text{H}_{21}\text{NO}_4\text{SNa}$ ($M + \text{Na}$) $^+$ 454.1089, found 454.1085.

VIII. SYNTHESIS AND CHARACTERIZATION OF ENANTIOENRICHED ESTERS FOR TABLE 4 AND STEREOSPECIFIC INTRAMOLECULAR REDUCTIVE CROSS-ELECTROPHILE COUPLING REACTION

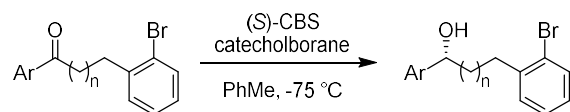
METHOD G: MANGANESE DIOXIDE OXIDATION OF BENZYLIC ALCOHOLS



The product was prepared according to a modified procedure reported by Wipf.²⁷ To a solution of *rac*-benzylic alcohol (1.0 equiv) in CH_2Cl_2 (30 mL) was added in a single portion MnO_2 (8 equiv). The reaction was allowed to stir overnight at room temperature. The resulting slurry was filtered through celite, and the celite was washed with CH_2Cl_2 . Solvent was removed under reduced pressure to afford the pure title compound.

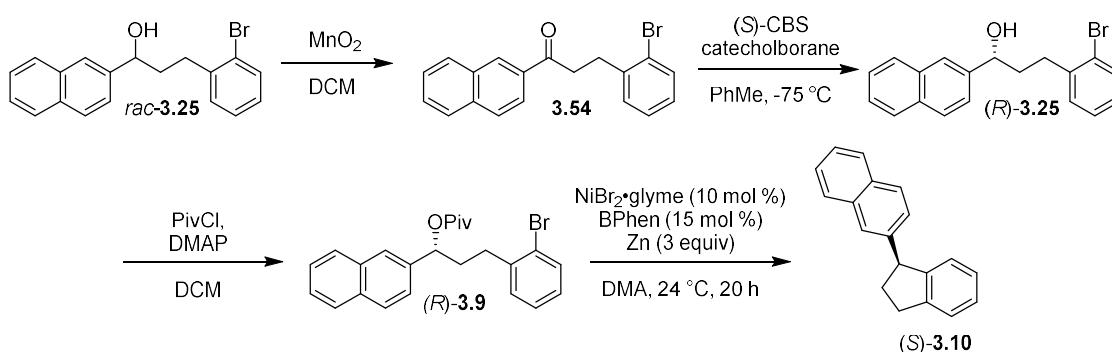
²⁷ Wipf, P.; Xu, W. *J. Org. Chem.* **1996**, *61*, 6556

METHOD H: CBS REDUCTION OF BENZYLIC KETONES



The product was prepared according to a modified procedure reported by Okamura.²⁸ In a glovebox, (*S*)-Me-CBS (0.100 equiv) was added to a 50 mL flame-dried round bottom flask equipped with a stir bar. The flask was capped with a septum and removed from the box. Benzylic ketone (1.0 equiv) was added to the flask as a solution in PhMe (20 mL). The reaction was then cooled to $-78\text{ }^{\circ}\text{C}$ and catecholborane (2.0 equiv) was added dropwise. After stirring for 24 h at $-78\text{ }^{\circ}\text{C}$, the reaction was warmed to ambient temperature and quenched with water. Saturated NaHCO_3 (15 mL) was added to the reaction flask and the mixture was extracted with EtOAc (3 x 30 mL). The combined organics were washed with brine, dried over MgSO_4 , and concentrated under reduced pressure. The product was purified by flash column chromatography to afford the title compound.

Scheme 3.17: Synthesis of starting material and formation of (*S*)- 3.10 (Table 3.3)



²⁸ Okamoto, K.; Hayashi, T. *Org. Lett.* **2007**, *9*, 5067

3.54. The product was prepared according to general procedure G using *rac*-**3.25** (1.32 g, 3.86 mmol, 1.0 equiv), MnO₂ (2.08 g, 24.0 mmol, 6.22 equiv). The product was purified by flash column chromatography to afford the pure title compound as a yellow oil (0.962 g, 2.84 mmol, 74%). **TLC** R_f = 0.6 (10% EtOAc/hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 8.47 (s, 1H), 8.04 (dd, *J* = 8.7, 1.8 Hz, 1H), 7.92 (d, *J* = 8.0 Hz, 1H), 7.86 (t, *J* = 8.6 Hz, 2H), 7.61–7.50 (m, 3H), 7.34 (dd, *J* = 7.6, 1.6 Hz, 1H), 7.27–7.21 (m, 1H), 7.08 (td, *J* = 7.8, 1.8 Hz, 1H), 3.44 (t, *J* = 8.3 Hz, 2H), 3.24 (t, *J* = 8.2 Hz, 2H); **¹³C NMR** (100 MHz, CDCl₃) δ 199.1, 140.9, 135.8, 134.3, 133.2, 132.8, 131.1, 130.0, 129.8, 128.7 (2C), 128.3, 128.0, 127.9, 127.0, 124.6, 124.1, 39.0, 31.2; **IR** (neat) 3057, 2928, 1677, 1626, 1469, 1182, 1122 cm⁻¹; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₁₉H₁₅BrONa (M + Na)⁺ 361.0204, found 361.0218.

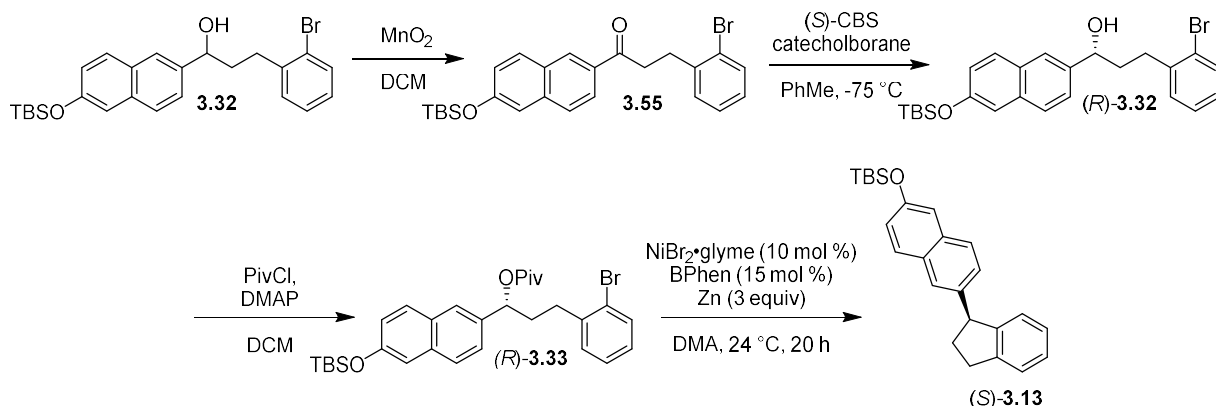
(R)-**3.25.** The product was prepared according to a general procedure H using *(S)*-Me-CBS (98 mg, 0.28 mmol, 0.100 equiv), **3.54** (0.96 g, 2.8 mmol, 1.0 equiv), and catecholborane (0.59 mL, 5.5 mmol, 2.0 equiv). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (0.91 g, 2.6 mmol, 94%, 96% ee). Analytical data is consistent with the values listed for **3.25** (vide supra).

(R)- **3.9.** The product was prepared according to general method B using *(R)*-**3.25** (0.60 g, 1.7 mmol, 1.0 equiv), pivaloyl chloride (0.24 mL, 1.9 mmol, 1.1 equiv) and dimethylaminopyridine (0.24 g, 1.9 mmol, 1.1 equiv). The product was purified by flash column chromatography (5% EtOAc/hexanes) to afford the title compound as a colorless oil (0.49 g, 1.1 mmol, 66%). Analytical data is consistent with the values listed for *rac*-**9** (vide supra). [α]_D²⁸ +39 (c 1.7, CHCl₃); **SFC**

analysis (OD-H, 6% IPA, 2.5 mL/min) indicated 96% ee: t_R (major) = 13.6 minutes, t_R (minor) = 14.9 minutes.

(*S*)- **3.10**. The product was prepared according to general method E using NiBr₂•glyme (6.2 mg, 0.020 mmol, 10 mol %), bathophenanthroline (10.0 mg, 0.030 mmol, 15 mol %), Zn⁰ (39.6 mg, 0.600 mmol, 3 equiv), DMA (0.60 mL), and (*R*)- **3.9** (85.1 mg, 0.200 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et₂O) and concentrated under reduced pressure. The product was purified by flash column chromatography (100% hexanes) to afford the title compound as a colorless oil (44.0 mg, 0.180 mmol, 90%, 88% ee). Analytical data is consistent with the values listed for *rac*-**3.10** (vide supra); [α]_D²⁸ +10 (c 0.9, CHCl₃); SFC analysis (OD-H, 6.0% IPA, 2.5 mL/min) indicated 88% ee: t_R (major) = 11.3 minutes, t_R (minor) = 10.5 minutes.

Scheme 3.18: Synthesis of starting material and formation of (*S*)- **3.13 (Table 3.3)**



3.55. The product was prepared according to general procedure G using *rac*-**3.32** (0.94 g, 2.0 mmol, 1.0 equiv), MnO₂ (1.8 g, 20.0 mmol, 10 equiv). The product was purified by flash column chromatography to afford the pure title compound as a yellow oil (0.56 g, 1.2 mmol, 60%). TLC R_f = 0.4 (5% EtOAc/hexanes); ¹H NMR (400 MHz, CDCl₃) δ 8.38 (s, 1H), 7.98 (dd, J = 8.6, 1.6 Hz, 1H), 7.78 (d, J = 9.0 Hz, 1H), 7.68 (d, J = 8.8 Hz, 1H), 7.51 (dd, J = 8.1, 1.0 Hz, 1H), 7.30

(dd, $J = 7.7, 1.6$ Hz, 1H), 7.22–7.17 (m, 2H), 7.10 (dd, $J = 8.9, 2.6$ Hz, 1H), 7.02 (td, $J = 7.6, 1.6$ Hz, 1H), 3.37 (t, $J = 7.9$ Hz, 2H), 3.20 (t, $J = 7.8$ Hz, 2H), 1.00 (s, 9H), 0.25 (s, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 198.5, 156.0, 140.8, 137.3, 133.0, 132.4, 131.4, 130.9, 129.8, 128.2, 128.1, 127.8, 127.2, 124.5, 124.4, 123.1, 115.0, 38.6, 31.1, 25.8, 18.4, -4.1; **IR** (neat) 3056, 2954, 1680, 1598, 1467, 1256 cm^{-1} ; **HRMS** (TOF MS CI^+) m/z calcd for $\text{C}_{25}\text{H}_{29}\text{BrO}_2\text{Si}$ (M) $^+$ 468.1120, found 468.1132.

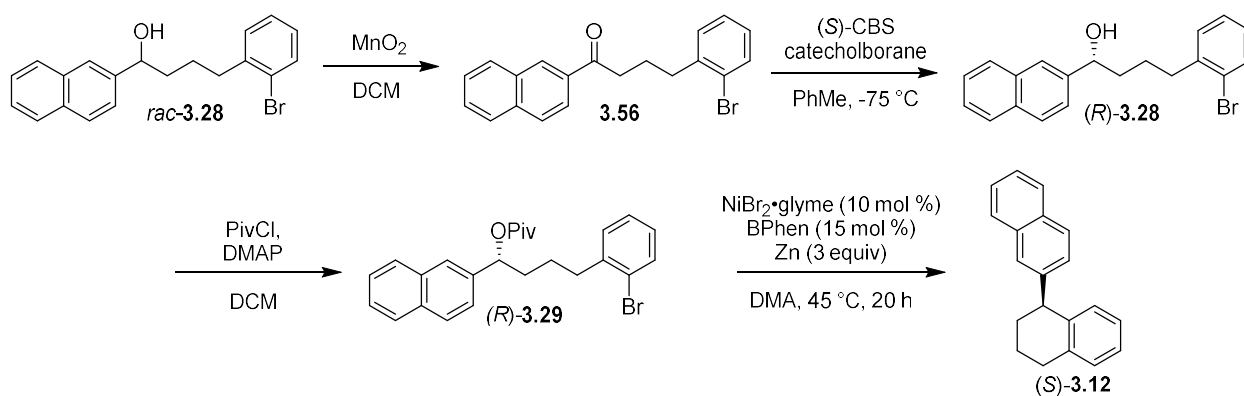
(*R*)-3.32. The product was prepared according to a general procedure H using (*S*)-Me-CBS (66 mg, 0.24 mmol, 0.200 equiv), **3.55** (0.56 g, 1.2 mmol, 1.0 equiv), and catecholborane (0.26 mL, 2.4 mmol, 2.0 equiv). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (0.42 g, 0.90 mmol, 75%). Analytical data is consistent with the values listed for **3.32** (vide supra).

(*R*)-3.33 The product was prepared according to general method B using (*R*)-**3.32** (0.40 g, 0.85 mmol, 1.0 equiv), pivaloyl chloride (0.12 mL, 0.98 mmol, 1.1 equiv) and dimethylaminopyridine (0.14 g, 1.0 mmol, 1.1 equiv). The product was purified by flash column chromatography (5% EtOAc/hexanes) to afford the title compound as a colorless oil (0.23 g, 0.41 mmol, 52%). Analytical data is consistent with the values listed for *rac*-**3.33** (vide supra). $[\alpha]^{27.5}_{\text{D}} +67$ (c 3.7, CHCl_3); **SFC** analysis (OD-H, 8% IPA, 2.5 mL/min) indicated 96% ee: t_{R} (major) = 8.1 minutes, t_{R} (minor) = 8.9 minutes.

(*S*)- 3.13. The product was prepared according to general method E using $\text{NiBr}_2 \cdot \text{glyme}$ (3.1 mg, 0.010 mmol, 10 mol %), bathophenanthroline (5.0 mg, 0.015 mmol, 15 mol %), Zn^0 (18.6 mg,

0.300 mmol, 3 equiv), DMA (0.40 mL), and (*R*)-**3.33** (55.5 mg, 0.100 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et₂O) and concentrated under reduced pressure. The product was purified by flash column chromatography (100% hexanes) to afford the title compound as a colorless oil (26 mg, 0.069 mmol, 69%, 92% ee). Analytical data is consistent with the values listed for *rac*-**3.13** (vide supra); [α]^{26.6}_D +91 (c 0.55, CHCl₃); SFC analysis (AD-H, 7.0% IPA, 2.5 mL/min) indicated 92% ee: t_R (major) = 7.1 minutes, t_R (minor) = 8.6 minutes.

Scheme 3.19: Synthesis of starting material and formation of (*S*)-3.12** (Table 3.3)**



3.56. The product was prepared according to general procedure G using *rac*-**3.28** (1.79 g, 5.03 mmol, 1.0 equiv), MnO₂ (4.4 g, 51 mmol, 10 equiv). The product was purified by flash column chromatography to afford the pure title compound as a yellow oil (1.22 g, 3.69 mmol, 73%). TLC R_f = 0.5 (12% Et₂O/hexanes); ¹H NMR (400 MHz, CDCl₃) δ 8.45 (s, 1H), 8.03 (dd, *J* = 8.6, 1.6 Hz, 1H), 7.97–7.93 (m, 1H), 7.91–7.85 (m, 1H), 7.58 (dddd, *J* = 18.5, 7.7, 6.6, 1.3 Hz, 3H), 7.29–7.22 (m, 3H), 7.07 (ddd, *J* = 8.0, 7.0, 2.1 Hz, 1H), 3.17 (t, *J* = 7.3 Hz, 2H), 2.90 (t, *J* = 7.7 Hz, 2H), 2.20–2.11 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 200.0, 141.2, 135.7, 134.4, 133.0, 132.7, 130.7, 129.86, 129.74, 128.62, 128.58, 128.0, 127.9, 127.7, 126.9, 124.8, 124.1, 38.0, 35.6, 24.7; IR

(neat) 2939, 2893, 1679, 1620, 1438, 1165 cm^{-1} ; **HRMS** (TOF MS CI^+) m/z calcd for $\text{C}_{20}\text{H}_{17}\text{BrOH}$ ($\text{M} + \text{H}$) $^+$ 353.0541, found 353.0442.

(R)-3.28. The product was prepared according to a general procedure H using (*S*)-Me-CBS (67 mg, 0.29 mmol, 0.100 equiv), **3.56** (1.0 g, 2.9 mmol, 1.0 equiv), and catecholborane (0.62 mL, 5.8 mmol, 2.0 equiv). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (0.83 g, 2.3 mmol, 80%). Analytical data is consistent with the values listed for **3.28** (vide supra).

(R)-3.29. The product was prepared according to general method B using (*R*)-**3.28** (0.728 g, 2.05 mmol, 1.0 equiv), pivaloyl chloride (0.30 mL, 2.4 mmol, 1.2 equiv) and dimethylaminopyridine (0.33 g, 2.7 mmol, 1.3 equiv). The product was purified by flash column chromatography (5% EtOAc/hexanes) to afford the title compound as a colorless oil (0.23 g, 0.41 mmol, 52%). Analytical data is consistent with the values listed for *rac*-**3.29** (vide supra). $[\alpha]_{\text{D}}^{27.6} +84$ (c 2.1, CHCl_3); **SFC** analysis (OD-H, 6% IPA, 2.5 mL/min) indicated 94% ee: t_{R} (major) = 18.2 minutes, t_{R} (minor) = 21.6 minutes.

(S)- 3.12. The product was prepared according to general method E using $\text{NiBr}_2 \cdot \text{glyme}$ (3.1 mg, 0.010 mmol, 10 mol %), bathophenanthroline (5.0 mg, 0.015 mmol, 15 mol %), Zn^0 (18.6 mg, 0.300 mmol, 3 equiv), DMA (0.40 mL), and (*R*)-**3.29** (44.0 mg, 0.100 mmol, 1.00 equiv). The reaction mixture was eluted through a silica plug (with Et_2O) and concentrated under reduced pressure. The product was purified by flash column chromatography (100% hexanes) to afford the title compound as a colorless oil (16 mg, 0.062 mmol, 62%, 78% ee). Analytical data is consistent

with the values listed for *rac*-**3.12** (vide supra); $[\alpha]^{27.6}_D +43$ (c 0.66, CHCl₃); SFC analysis (OD-H, 6.0% IPA, 2.5 mL/min) indicated 78% ee: t_R (major) = 12.5 minutes, t_R (minor) = 16.8 minutes.

STEREOCHEMICAL PROOF

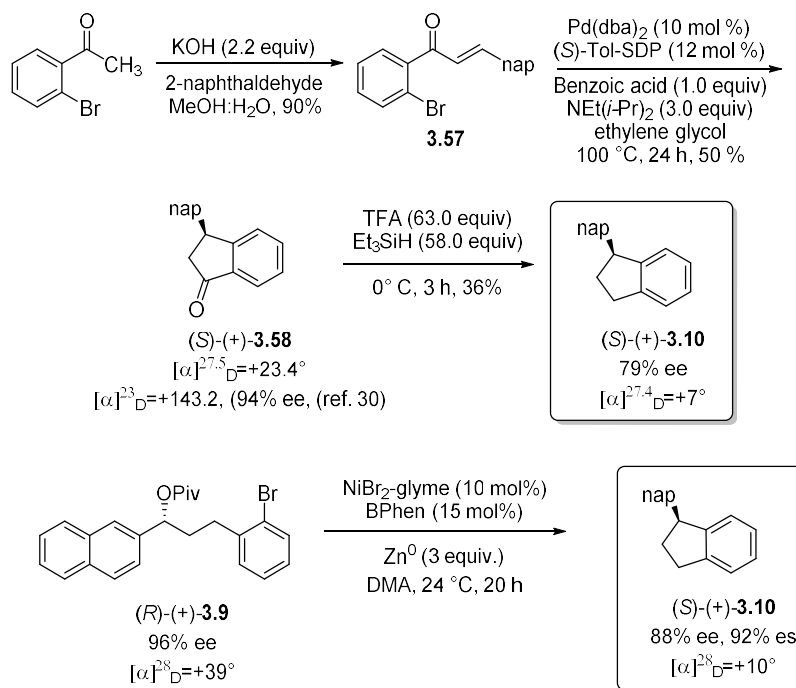
Enantioenriched alcohol (*R*)-**3.25** was prepared by enantioselective CBS reduction (vide supra). Absolute configuration of (*R*)-**3.25** and (*R*)-**3.9** were assigned based on the accepted model for selectivity in CBS reductions.²⁹

The absolute configuration of the enantioenriched indane **3.10** was assigned by derivatization of known enantioenriched indanone (*S*)-**3.58** to indane (*S*)-**3.10**. Enantioenriched (*S*)-**3.58** was prepared by an asymmetric reductive Heck reaction as reported by Zhou. The stereochemistry was verified by comparison of the optical rotation to the literature value. Reduction to indane (*S*)-**3.10** and subsequent comparison of the optical rotation and SFC data matched that of (*S*)-**3.10** synthesized by the reductive cross-electrophile coupling reaction. This product corresponds to net inversion at the benzylic center in the reductive cross-electrophile coupling reaction.

²⁹ Corey, E. J.; Helal, C. J. *Angew. Chem., Int. Ed.* **1998**, *37*, 1986

Scheme 3.20: Stereochemical course of the reductive cross-electrophile coupling reaction

(Table 3.3).



3.57. The product was prepared according to a modified procedure reported by Zhou.³⁰ To a 100 mL round bottom flask equipped with a stir bar was added 2'-bromoacetophenone (0.67 mL, 5.0 mmol, 1.0 equiv), naphthaldehyde (0.78 g, 5.0 mmol, 1.0 equiv), KOH (0.62 g, 11 mmol, 2.2 equiv), and MeOH/H₂O (15:15 mL). The reaction was stirred overnight at room temperature. The resulting solid was filtered, washed with MeOH/H₂O, and dried by vacuum filtration to afford the title compound as a solid (1.3 g, 4.5 mmol, 90 % yield).

(S)-3.58. The product was prepared according to a modified procedure reported by Zhou.¹⁵ In a glovebox, a flame-dried 7 mL vial equipped with a stir bar was charged with Pd(dba)₂ (14 mg,

³⁰ Yue, G.; Lei, K.; Hirao, H.; Zhou, J. *Angew. Chem. Int. Ed.* **2015**, *54*, 6531

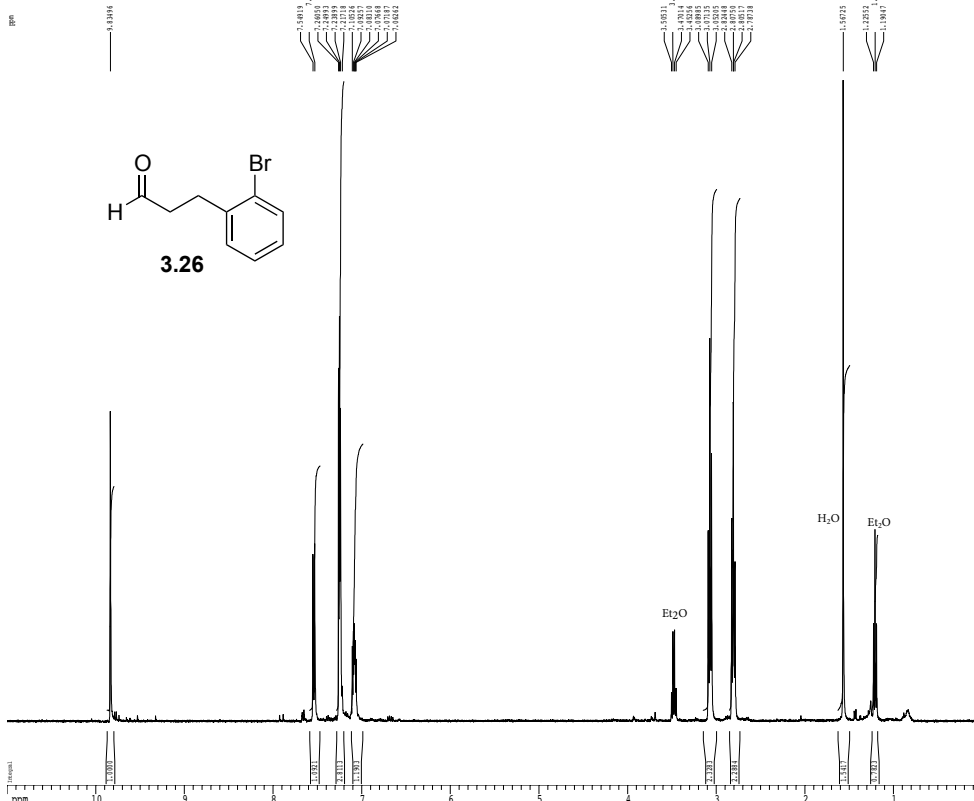
0.025 mmol, 10 mol %), (*S*)- Tol-SDP (18 mg, 0.03 mmol, 12 mol %), benzoic acid (30 mg, 0.25 mmol, 1.0 equiv) and degassed ethylene glycol (1.25 mL). After stirring for 10 min, *N*-diisopropylethylamine (130 μ L, 1.5 mmol, 3.0 equiv) and **3.57** (85 mg, 0.25 mmol, 1.0 equiv) were added and the mixture was taken out of the glove box. The reaction was stirred for 24 h in a pre-warmed oil bath at 100 °C then allowed to cool to room temperature. The reaction mixture was eluted through a silica plug (with Et₂O) and concentrated under reduced pressure. The product was purified by flash column chromatography (0%-10% Et₂O/hexanes) to afford the title compound as a colorless oil (27 mg, 0.13 mmol, 50% yield). Analytical data is consistent with literature values.³¹

¹H NMR (400 MHz, CDCl₃) δ 7.87 (d, *J* = 7.9 Hz, 1H), 7.84–7.77 (m, 3H), 7.67 (s, 1H), 7.08 (t, *J* = 7.5 Hz, 1H), 7.50–7.43 (m, 3H), 7.30 (d, *J* = 7.8 Hz, 1H), 7.15 (dd, *J* = 8.4, 1.6 Hz, 1H), 4.77 (dd, *J* = 8.2, 3.7 Hz, 1H), 3.31 (dd, *J* = 19.3, 8.1 Hz, 1H), 2.79 (dd, *J* = 19.3, 3.9 Hz, 1H).

(*S*)- **3.10**. A 20 ml scintillation vial was charged with **3.58** (27 mg, 0.13 mmol, 1.0 equiv), dissolved in TFA (0.5 ml) and cooled to 0 °C in an ice bath. Et₃SiH (1.0 ml) was added dropwise over the course of 20 min and the reaction was allowed to stir for 3 h at 0 °C. The reaction was then allowed to warm to room temperature, was concentrated under reduced pressure, and the residue was taken up and purified by flash column chromatography (100% hexanes) to afford the title compound as a colorless oil (9.2 mg, 0.038 mmol, 36% yield, 79% ee). Analytical data is consistent with the values listed for (*S*)-**10** (vide supra). $[\alpha]^{27.4}_{\text{D}} +7$ (c 0.46, CHCl₃); SFC analysis (OD-H, 6.0% IPA, 2.5 mL/min) indicated 79% ee: *t*_R (major) = 11.4 minutes, *t*_R (minor) = 10.6 minutes.

³¹ Yu, N-U.; Xu, M-H. *J. Org. Chem.*, **2013**, 78, 2736.

1H spectrum



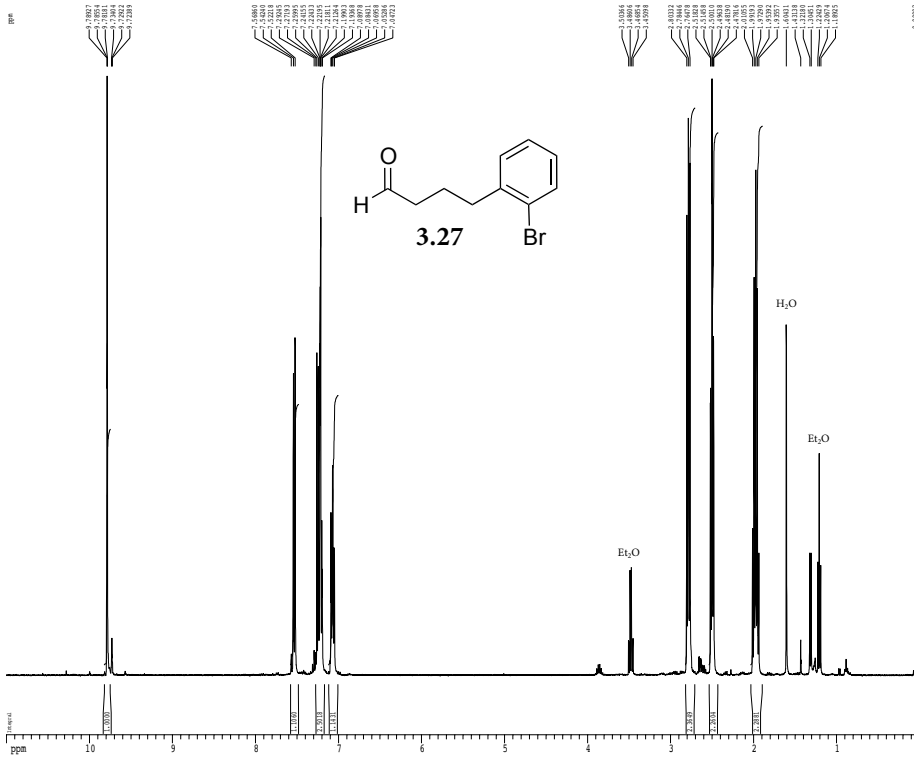
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1H spectrum



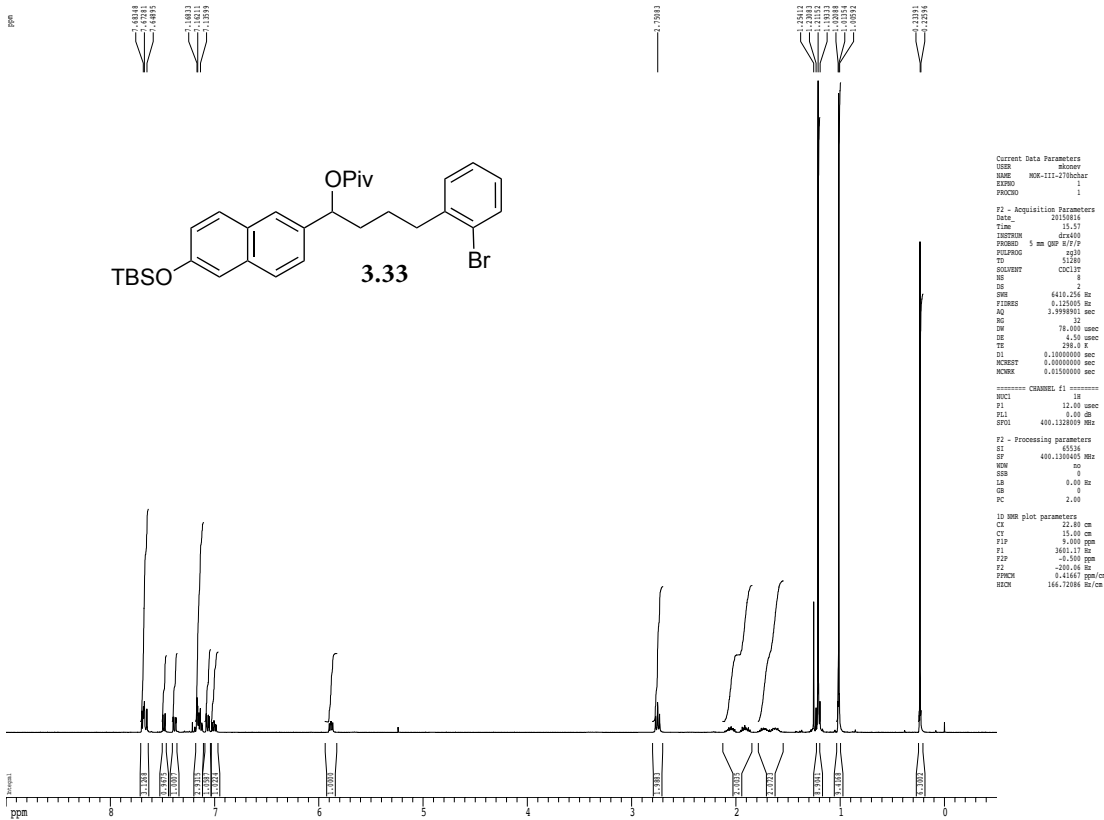
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1H spectrum



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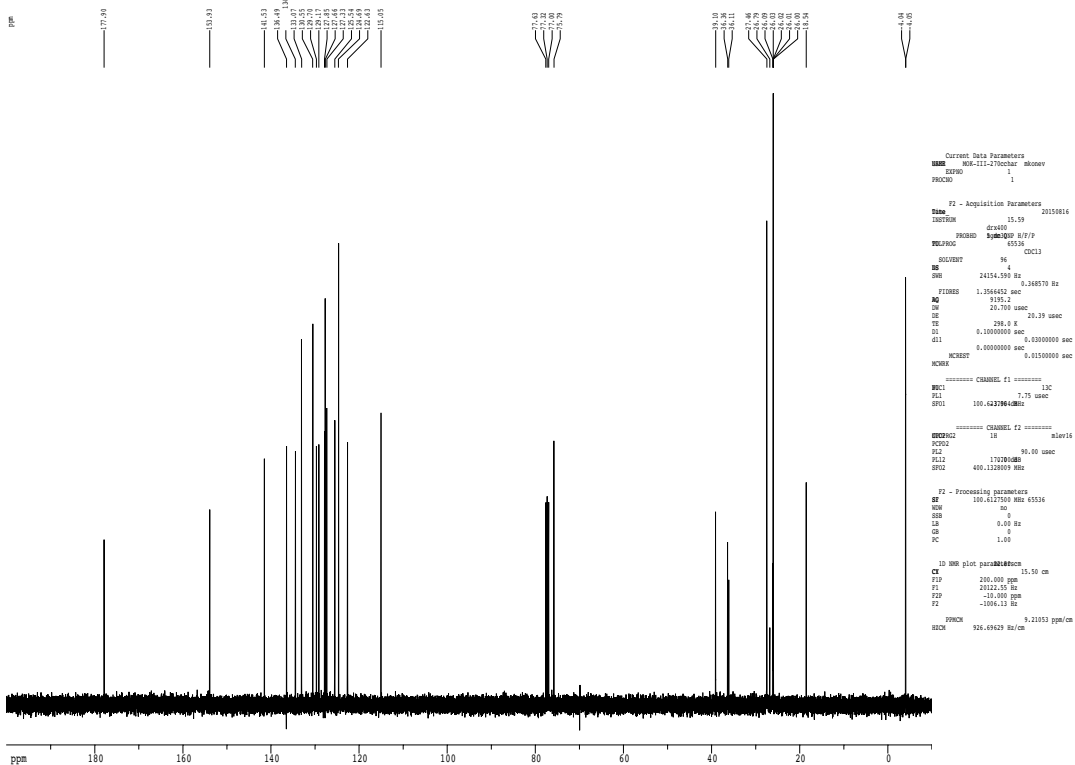
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13C spectrum with 1H decoupling



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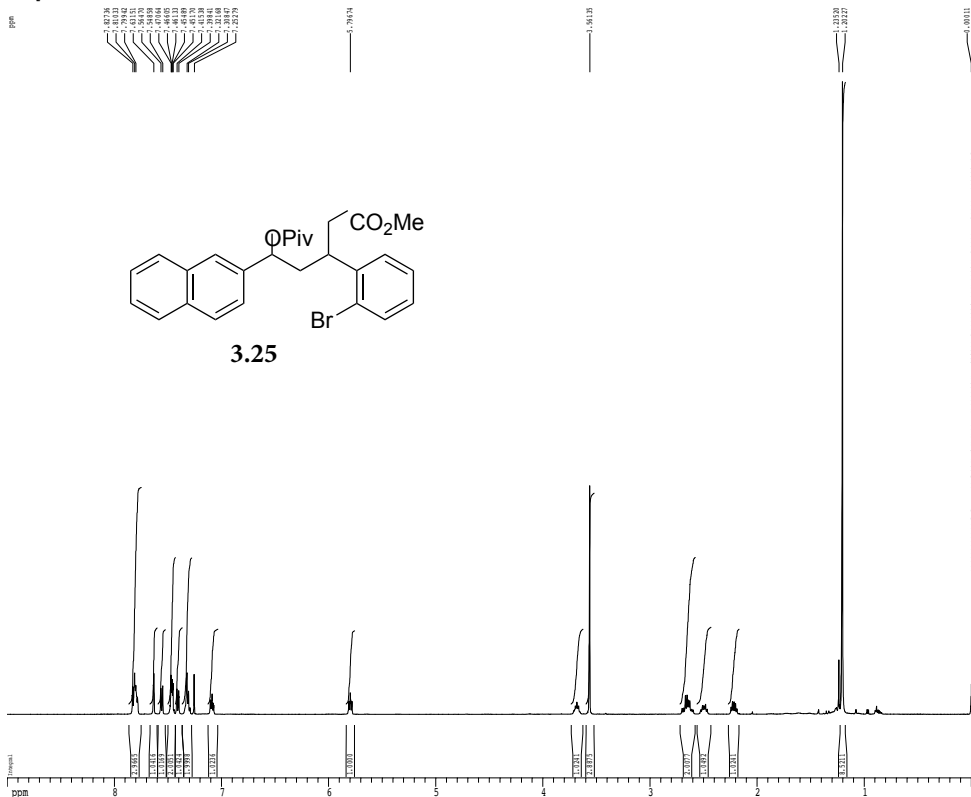
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1H spectrum



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EXPNO 1
PROCNO 1

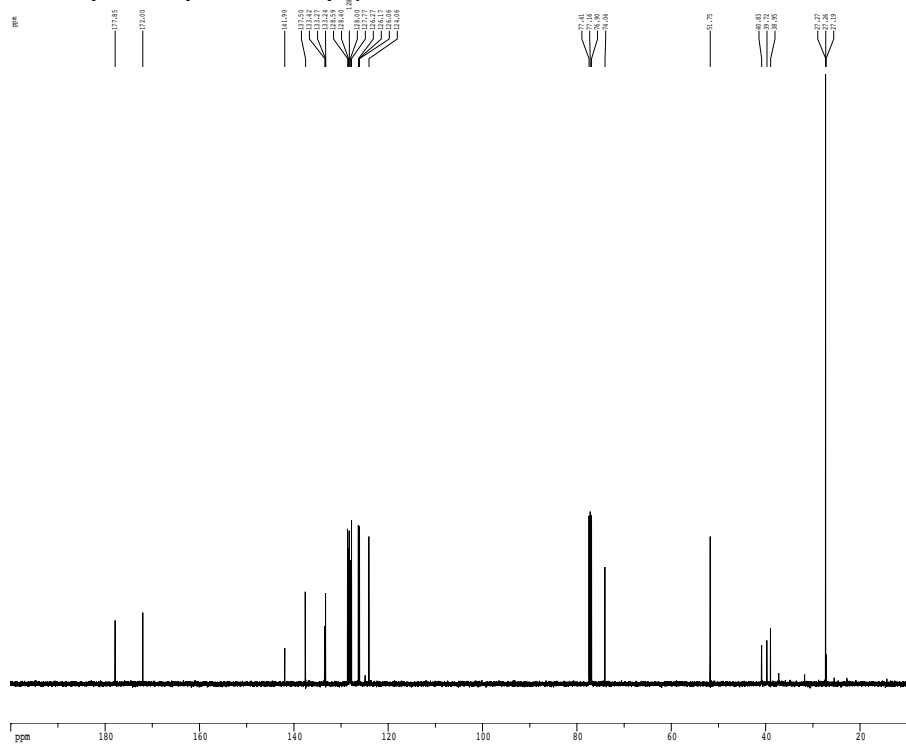
F2 - Acquisition Parameters
Date_ 20150731
TIME 14.40
INSTRUM cryo000
PROBHD 5 mm CPXI 1H-
PULPROG zgpg
TD 32768
SOLVENT CDCl3
DS 2
SWH 8012.000 Hz
FIDRES 0.250026 Hz
AQ 1.9998451 sec
RG 16
DM 62.400 usec
DE 6.00 usec
TE 298.0
SI 0.10000000 sec
MCHRESF 0.00000000 sec
MCHSR 0.01000000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 7.00 usec
PL1 1.60 dB
SFO1 500.222915 MHz

F2 - Processing parameters
SI 6536
SF 500.222915 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 4.00

ID NMR plot parameters
CX 22.80 cm
CY 15.00 cm
FIDP 0.000 ppm
F1 4501.58 Hz
F2P 0.000 ppm
F2 0.00 Hz
PPMCH 0.39474 ppm/cm
WCH 197.45028 ppm/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
NAME 188-6-017-0-111 188a
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150731
TIME 14.29
INSTRUM cryo000
PROBHD 5 mm CPXI 1H-
PULPROG zgpg
TD 32768
SOLVENT CDCl3
DS 2
SWH 30303.031 Hz
FIDRES 1.0813940 sec
AQ 16.500 usec
RG 16
DM 16.500 usec
DE 6.00 usec
TE 298.0
SI 0.25000000 sec
MCHRESF 0.03000000 sec
MCHSR 0.00000000 sec
MCHSR 0.00000000 sec
MCHSR 0.00000000 sec
MCHSR 0.01500000 sec

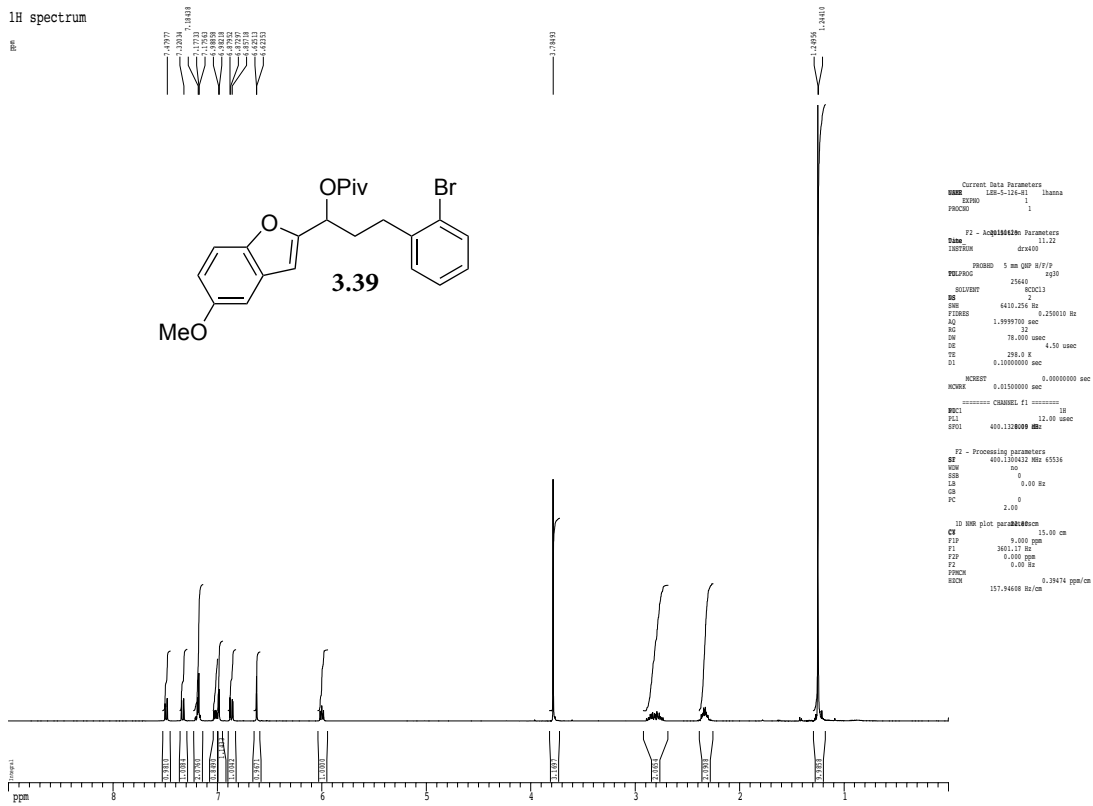
***** CHANNEL f1 *****
NUC1 13C
P1 16.50 usec
PL1 190.00 dB
PL2 120.00 dB
SFO1 125.760448 MHz
SFO2 101.626126 MHz
SFO3 77.000000 MHz
SFO4 50.761200 MHz
SFO5 27.500000 MHz
SFO6 15.625000 MHz
SFO7 7.812500 MHz
SFO8 3.906250 MHz
SFO9 1.953125 MHz
SFO10 0.976562 MHz
SFO11 0.488281 MHz
SFO12 0.244141 MHz
SFO13 0.122070 MHz
SFO14 0.061035 MHz
SFO15 0.030518 MHz
SFO16 0.015259 MHz
SFO17 0.007629 MHz
SFO18 0.003814 MHz
SFO19 0.001907 MHz
SFO20 0.000954 MHz
SFO21 0.000477 MHz
SFO22 0.000238 MHz
SFO23 0.000119 MHz
SFO24 0.000059 MHz
SFO25 0.000029 MHz
SFO26 0.000015 MHz
SFO27 0.000007 MHz
SFO28 0.000004 MHz
SFO29 0.000002 MHz
SFO30 0.000001 MHz
SFO31 0.000000 MHz
SFO32 0.000000 MHz
SFO33 0.000000 MHz
SFO34 0.000000 MHz
SFO35 0.000000 MHz
SFO36 0.000000 MHz
SFO37 0.000000 MHz
SFO38 0.000000 MHz
SFO39 0.000000 MHz
SFO40 0.000000 MHz
SFO41 0.000000 MHz
SFO42 0.000000 MHz
SFO43 0.000000 MHz
SFO44 0.000000 MHz
SFO45 0.000000 MHz
SFO46 0.000000 MHz
SFO47 0.000000 MHz
SFO48 0.000000 MHz
SFO49 0.000000 MHz
SFO50 0.000000 MHz
SFO51 0.000000 MHz
SFO52 0.000000 MHz
SFO53 0.000000 MHz
SFO54 0.000000 MHz
SFO55 0.000000 MHz
SFO56 0.000000 MHz
SFO57 0.000000 MHz
SFO58 0.000000 MHz
SFO59 0.000000 MHz
SFO60 0.000000 MHz
SFO61 0.000000 MHz
SFO62 0.000000 MHz
SFO63 0.000000 MHz
SFO64 0.000000 MHz
SFO65 0.000000 MHz
SFO66 0.000000 MHz
SFO67 0.000000 MHz
SFO68 0.000000 MHz
SFO69 0.000000 MHz
SFO70 0.000000 MHz
SFO71 0.000000 MHz
SFO72 0.000000 MHz
SFO73 0.000000 MHz
SFO74 0.000000 MHz
SFO75 0.000000 MHz
SFO76 0.000000 MHz
SFO77 0.000000 MHz
SFO78 0.000000 MHz
SFO79 0.000000 MHz
SFO80 0.000000 MHz
SFO81 0.000000 MHz
SFO82 0.000000 MHz
SFO83 0.000000 MHz
SFO84 0.000000 MHz
SFO85 0.000000 MHz
SFO86 0.000000 MHz
SFO87 0.000000 MHz
SFO88 0.000000 MHz
SFO89 0.000000 MHz
SFO90 0.000000 MHz
SFO91 0.000000 MHz
SFO92 0.000000 MHz
SFO93 0.000000 MHz
SFO94 0.000000 MHz
SFO95 0.000000 MHz
SFO96 0.000000 MHz
SFO97 0.000000 MHz
SFO98 0.000000 MHz
SFO99 0.000000 MHz
SFO100 0.000000 MHz

***** CHANNEL f2 *****
NUC12 13C
P12 16.50 usec
PL12 190.00 dB
PL22 120.00 dB
SFO12 125.760448 MHz
SFO22 101.626126 MHz
SFO32 77.000000 MHz
SFO42 50.761200 MHz
SFO52 27.500000 MHz
SFO62 15.625000 MHz
SFO72 7.812500 MHz
SFO82 3.906250 MHz
SFO92 1.953125 MHz
SFO102 0.976562 MHz
SFO112 0.488281 MHz
SFO122 0.244141 MHz
SFO132 0.122070 MHz
SFO142 0.061035 MHz
SFO152 0.030518 MHz
SFO162 0.015259 MHz
SFO172 0.007629 MHz
SFO182 0.003814 MHz
SFO192 0.001907 MHz
SFO202 0.000954 MHz
SFO212 0.000477 MHz
SFO222 0.000238 MHz
SFO232 0.000119 MHz
SFO242 0.000059 MHz
SFO252 0.000029 MHz
SFO262 0.000015 MHz
SFO272 0.000007 MHz
SFO282 0.000004 MHz
SFO292 0.000002 MHz
SFO302 0.000001 MHz
SFO312 0.000000 MHz
SFO322 0.000000 MHz
SFO332 0.000000 MHz
SFO342 0.000000 MHz
SFO352 0.000000 MHz
SFO362 0.000000 MHz
SFO372 0.000000 MHz
SFO382 0.000000 MHz
SFO392 0.000000 MHz
SFO402 0.000000 MHz
SFO412 0.000000 MHz
SFO422 0.000000 MHz
SFO432 0.000000 MHz
SFO442 0.000000 MHz
SFO452 0.000000 MHz
SFO462 0.000000 MHz
SFO472 0.000000 MHz
SFO482 0.000000 MHz
SFO492 0.000000 MHz
SFO502 0.000000 MHz
SFO512 0.000000 MHz
SFO522 0.000000 MHz
SFO532 0.000000 MHz
SFO542 0.000000 MHz
SFO552 0.000000 MHz
SFO562 0.000000 MHz
SFO572 0.000000 MHz
SFO582 0.000000 MHz
SFO592 0.000000 MHz
SFO602 0.000000 MHz
SFO612 0.000000 MHz
SFO622 0.000000 MHz
SFO632 0.000000 MHz
SFO642 0.000000 MHz
SFO652 0.000000 MHz
SFO662 0.000000 MHz
SFO672 0.000000 MHz
SFO682 0.000000 MHz
SFO692 0.000000 MHz
SFO702 0.000000 MHz
SFO712 0.000000 MHz
SFO722 0.000000 MHz
SFO732 0.000000 MHz
SFO742 0.000000 MHz
SFO752 0.000000 MHz
SFO762 0.000000 MHz
SFO772 0.000000 MHz
SFO782 0.000000 MHz
SFO792 0.000000 MHz
SFO802 0.000000 MHz
SFO812 0.000000 MHz
SFO822 0.000000 MHz
SFO832 0.000000 MHz
SFO842 0.000000 MHz
SFO852 0.000000 MHz
SFO862 0.000000 MHz
SFO872 0.000000 MHz
SFO882 0.000000 MHz
SFO892 0.000000 MHz
SFO902 0.000000 MHz
SFO912 0.000000 MHz
SFO922 0.000000 MHz
SFO932 0.000000 MHz
SFO942 0.000000 MHz
SFO952 0.000000 MHz
SFO962 0.000000 MHz
SFO972 0.000000 MHz
SFO982 0.000000 MHz
SFO992 0.000000 MHz
SFO1002 0.000000 MHz

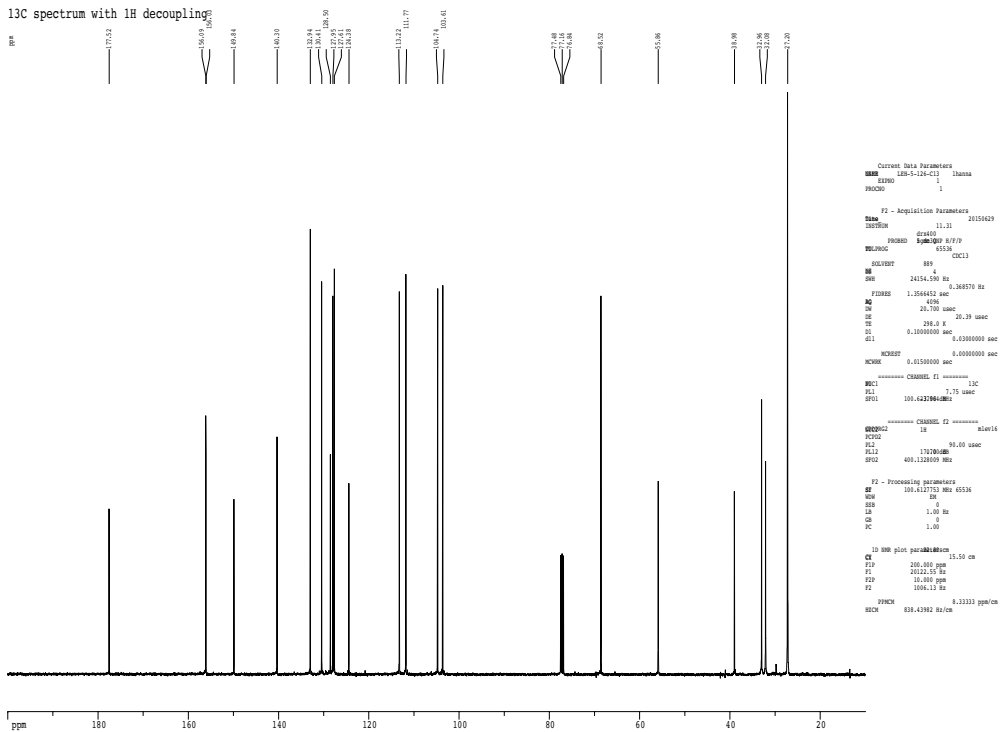
***** GRABBER CHANNEL *****
NAME 188-6-017-0-111 188a
SI 6536
SF 500.222915 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 4.00

ID NMR plot parameters
CX 22.80 cm
CY 15.00 cm
FIDP 0.000 ppm
F1 4501.58 Hz
F2P 0.000 ppm
F2 0.00 Hz
PPMCH 0.39474 ppm/cm
WCH 197.45028 ppm/cm
    
```

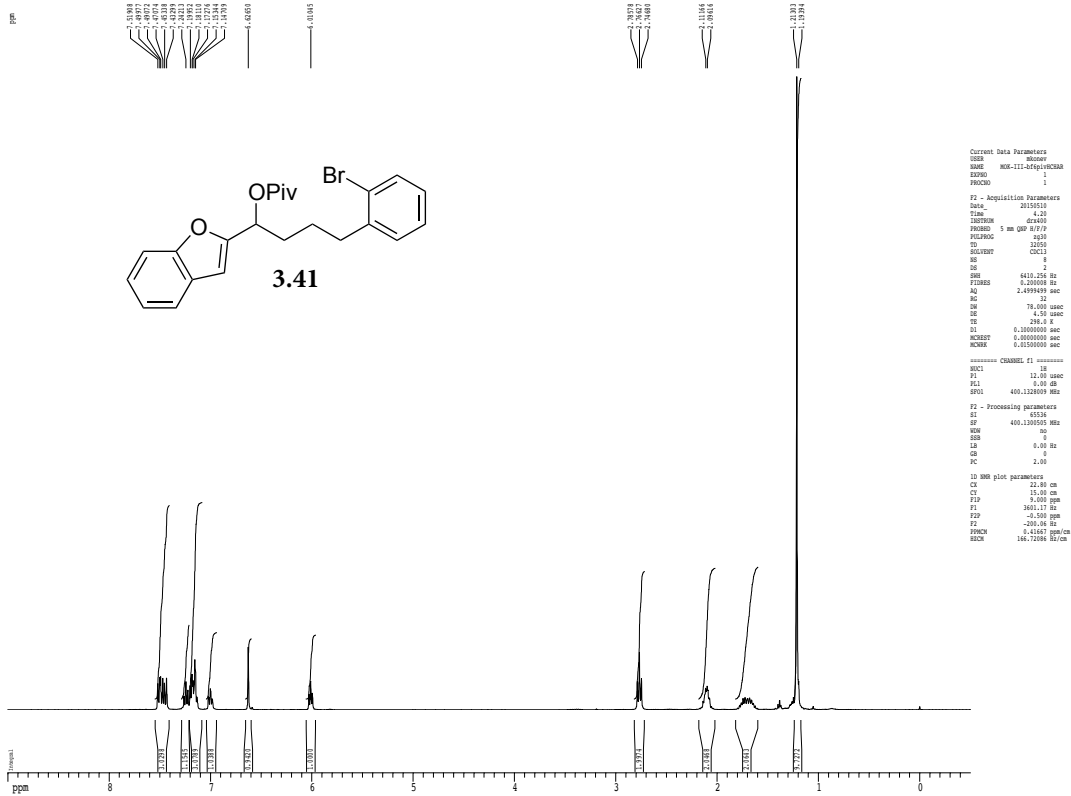

1H spectrum



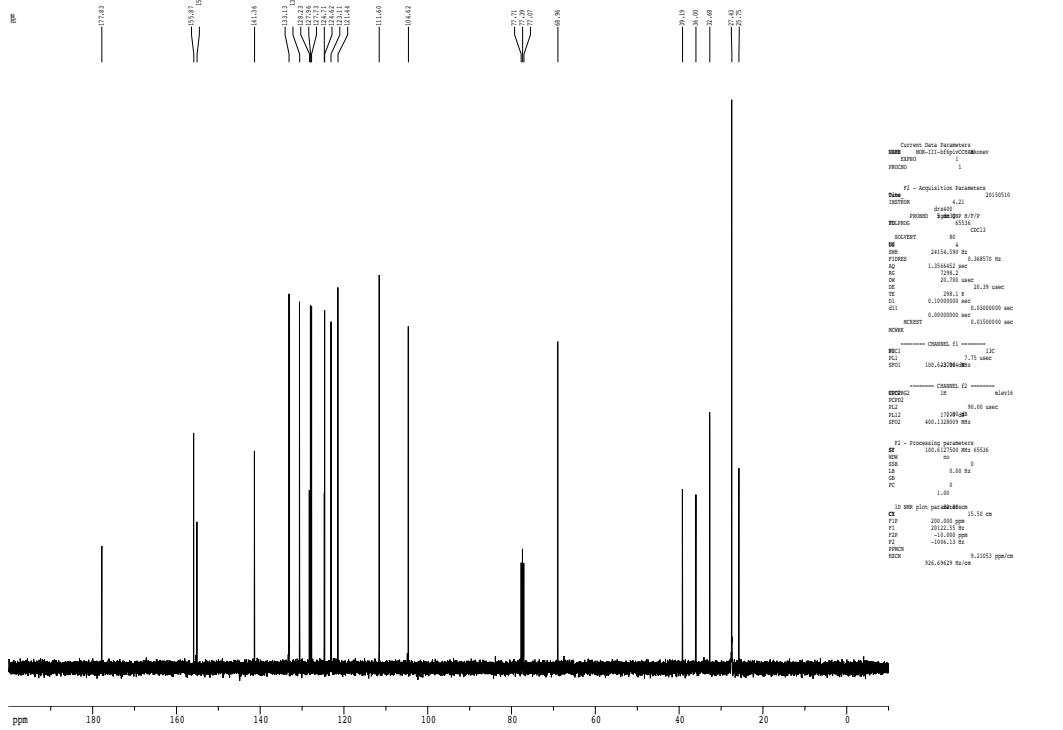
13C spectrum with 1H decoupling



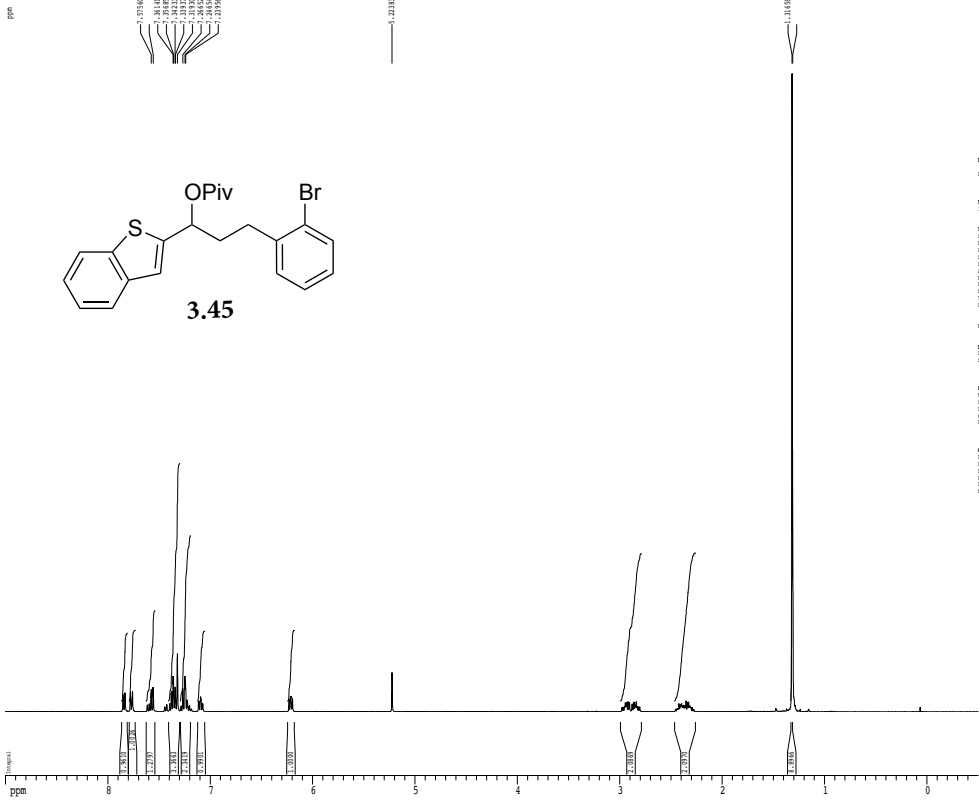
1H spectrum



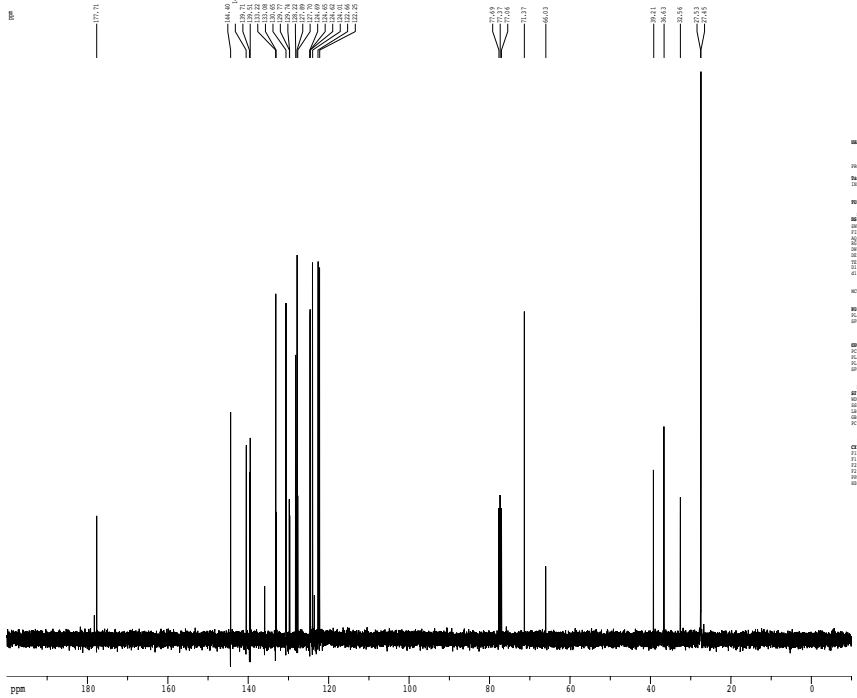
13C spectrum with 1H decoupling



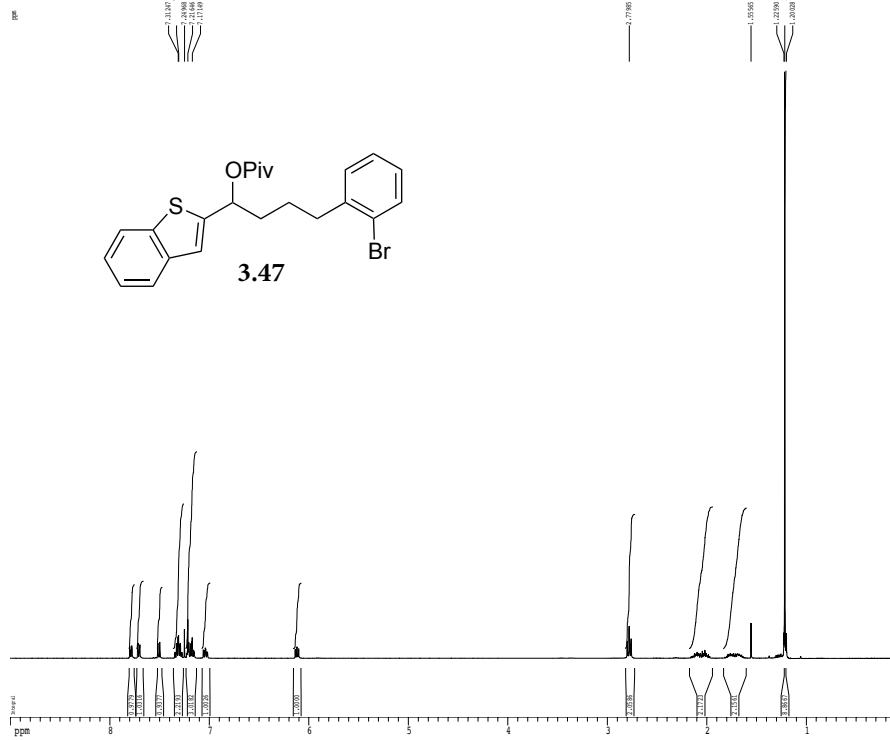
1H spectrum



13C spectrum with 1H decoupling

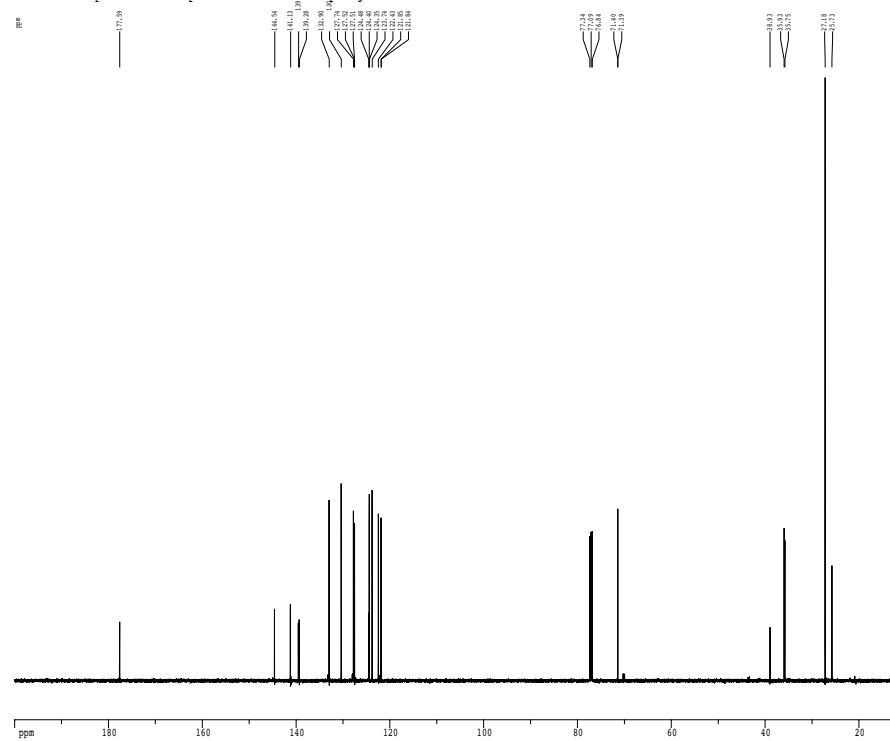


1H spectrum



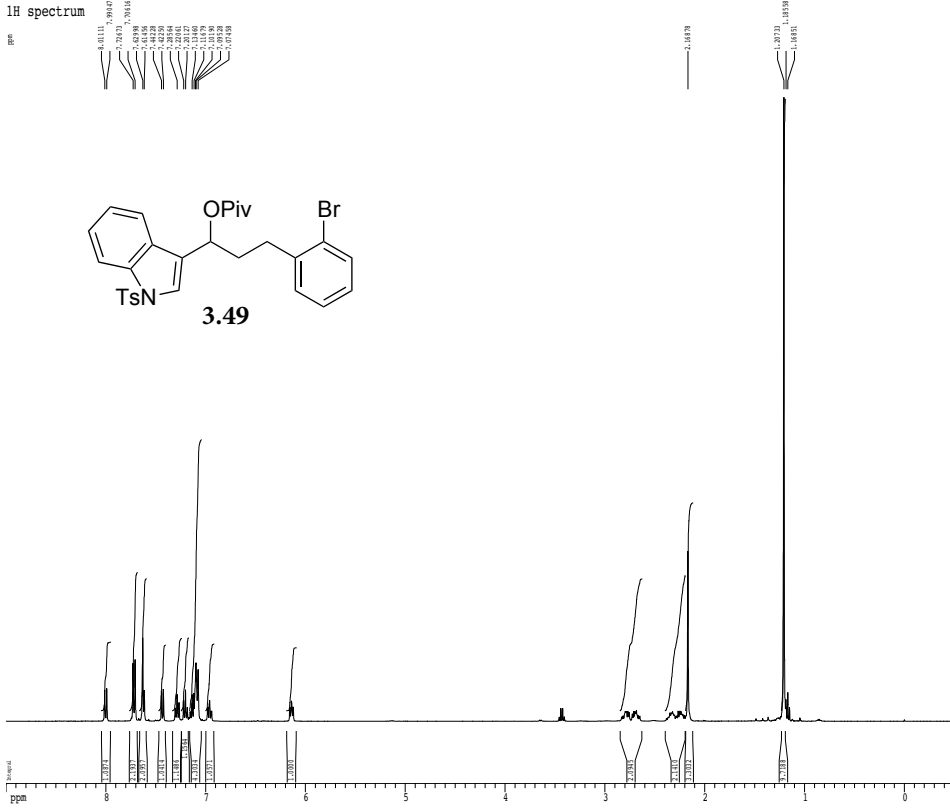
Current Data Parameters
 NAME 188-025-2-232 Thana
 EXPNO 1
 PROCNO 1
 F2 - Acq015410 Parameters
 Date_ 20.15
 INSTRUM dr400
 PROCES2 5 mm QNP 5/17/7
 PULPROG zgpg30
 SOLVENT 256d3
 NS 2564
 DS 8
 SFO 810.252 MHz
 FIDRES 1.999700 sec 0.250010 Hz
 AQ 1.999700 sec
 RG 256
 SD 78.000 usec
 DE 288.0 K
 TE 300.2 K
 D1 0.10000000 sec
 D11 0.00000000 sec
 ACQRES2 0.01500000 sec
 ===== CHANNEL f1 =====
 NUC1 13
 P1 12.00 usec
 SFO1 400.118000 MHz
 F2 - Processing parameters
 SF 400.118000 MHz 9536
 NH 2
 SWH 0.00 Hz
 LB 0.00 Hz
 GB 0
 PC 2.00
 1D 100 plot parameters
 CT 15.00 Hz 0.00 cm
 PTP 0.000 ppm
 FI 3601.17 Hz
 FZ 9.000 ppm
 FQ 2.00 Hz
 FWHM 0.29474 ppm/cm
 SFOH 157.74469 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



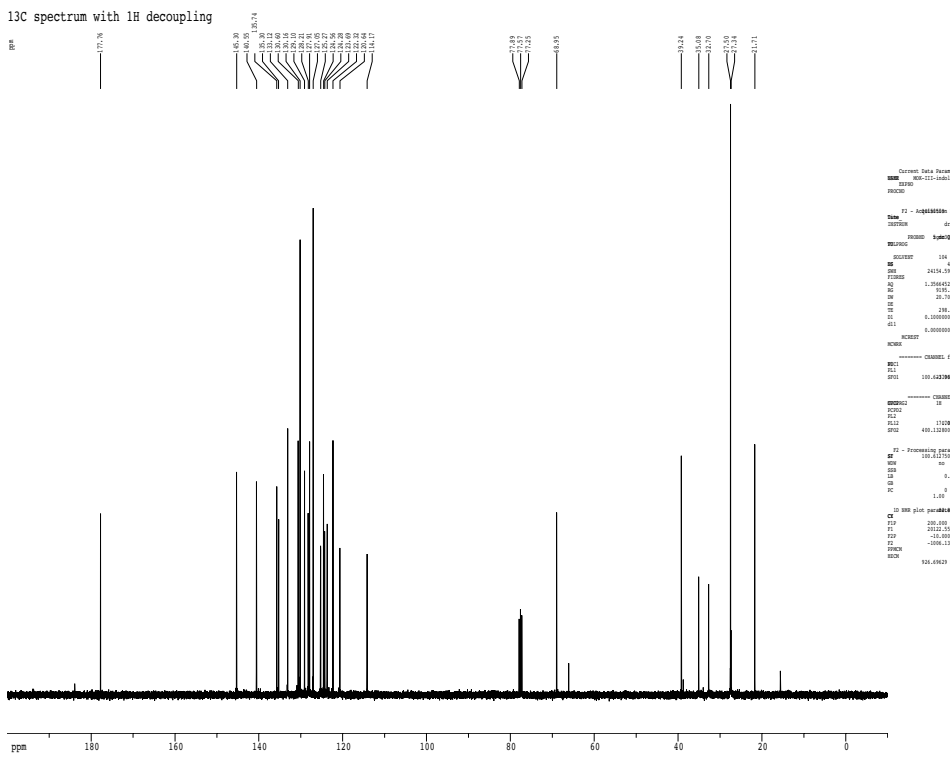
Current Data Parameters
 NAME 188-025-2-134 Thana
 EXPNO 1
 PROCNO 1
 F2 - Acq015410 Parameters
 Date_ cryo516
 INSTRUM cryo516
 PROCES2 Spinalchoy034p
 PROCNO 5 mm CPXI 13-
 SOLVENT gsd
 NS 3331.011 Hz
 DS 1
 SFO 303.1011 MHz
 FIDRES 1.0913840 sec 0.462388 Hz
 AQ 1.0913840 sec
 RG 7296.2
 SD 86.000 usec
 DE 288.0 K
 TE 300.2 K
 D1 0.25000000 sec
 D11 0.03000000 sec
 AC 0.00100000 sec
 AC17 0.00100000 sec
 AC18 0.00000000 sec
 ACRES2 0.01500000 sec
 ===== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 SFO1 125.7642540 MHz
 SFO2 125.7642540 MHz
 SFX 2.70 dB
 SFX2 2.70 dB
 SFOH1 Cryo5, 0.5, 20.1
 SFOH2 0.00 Hz 0.000000 CryoComp. 4
 SFOH3 0.00 Hz
 ===== CHANNEL f2 =====
 NUC2 13
 P2 100.00 usec
 SFO2 100.625000 MHz
 SFO3 24.25 MHz 1.60 dB
 SFO4 580.222501 MHz
 ===== GRABBER CHANNEL =====
 CP1MAG 0.000 MHz 0.000 MHz
 CP1 0.00 A
 CP2 0.00 A
 CP3 0.00 A
 CP4 0.00 A
 CP5 0.00 A
 CP6 0.00 A
 CP7 30.00 A
 CP8 30.00 A
 P15 500.00 usec
 P16 500.00 usec
 F2 - Processing parameters
 SF 125.764250 MHz 9536
 NH 2
 SWH 0.00 Hz
 LB 0.00 Hz
 GB 0
 PC 2.00
 1D 100 plot parameters
 CT 15.00 Hz 0.00 cm
 PTP 200.000 ppm
 FI 2156.00 Hz
 FZ 10.000 ppm
 FQ 10.00 Hz
 FWHM 0.33333 ppm/cm
 SFOH 1048.17017 Hz/cm

¹H spectrum



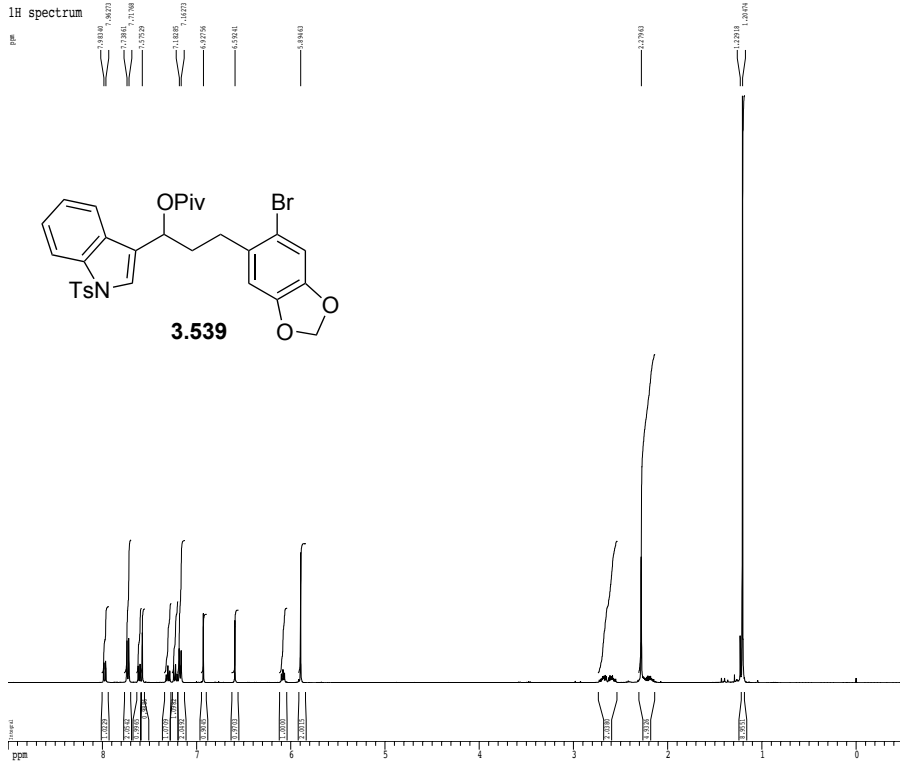
Current Data Parameters
 NAME: 001-11-040919-000000000
 EXPNO: 1
 PROCNO: 1
 Date_ Time: 21.09.09
 PROBRG: 5 mm QNP 61/70
 PULPROG: zgpg30
 SOLVENT: DMSO
 NS: 640
 DS: 4
 F2: 601.365 MHz
 F1: 150.916 MHz
 AQ: 0.200000 sec
 RG: 327.5
 IC: 12.00
 DE: 2.000000 sec
 TE: 300.2 K
 D1: 0.200000 sec
 DELTA: 7.000000 sec
 ACHNAME: 0.13150000 sec
 ACHVAL: 1.00
 CHANNEL: f1
 NUCL1: 13C
 P1: 12.00 usec
 PL1: 0.00 dB
 SFO1: 101.625000 MHz
 Processing parameters
 SF: 400.130427 MHz 6516
 DSF: 6516
 EQ: 0
 LQ: 0.00 Hz
 GQ: 0
 PC: 1.00
 1D 90B plot parameters
 CH: 15.00 dB, 90.00 cm
 FI: 3.000000 Hz
 PD: 3491.27 Hz
 ZF: -25.000000 ppm
 PR: -200.00 Hz
 FREQC: 0.143841 ppm/cm
 SFOC: 101.625000 MHz/cm

¹³C spectrum with ¹H decoupling



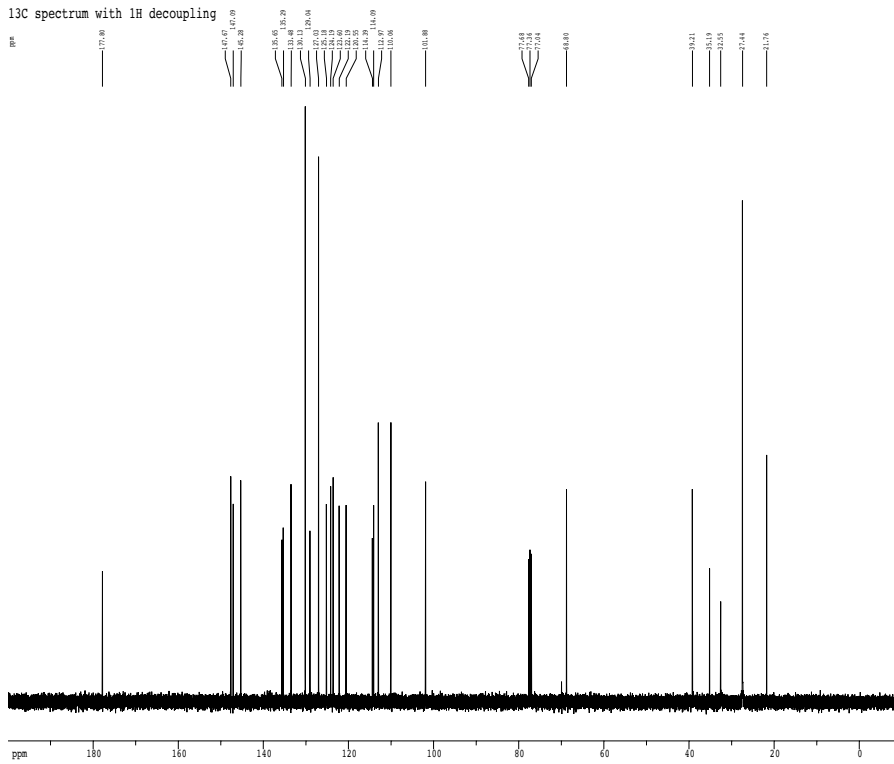
Current Data Parameters
 NAME: 001-11-040919-000000000
 EXPNO: 1
 PROCNO: 1
 Date_ Time: 21.09.09
 PROBRG: 5 mm QNP 61/70
 PULPROG: zgpg30
 SOLVENT: DMSO
 NS: 640
 DS: 4
 F2: 601.365 MHz
 F1: 150.916 MHz
 AQ: 0.200000 sec
 RG: 327.5
 IC: 12.00
 DE: 2.000000 sec
 TE: 300.2 K
 D1: 0.200000 sec
 DELTA: 7.000000 sec
 ACHNAME: 0.13150000 sec
 ACHVAL: 1.00
 CHANNEL: f1
 NUCL1: 13C
 P1: 12.00 usec
 PL1: 0.00 dB
 SFO1: 101.625000 MHz
 Processing parameters
 SF: 400.130427 MHz 6516
 DSF: 6516
 EQ: 0
 LQ: 0.00 Hz
 GQ: 0
 PC: 1.00
 1D 90B plot parameters
 CH: 15.00 dB, 90.00 cm
 FI: 3.000000 Hz
 PD: 3491.27 Hz
 ZF: -25.000000 ppm
 PR: -200.00 Hz
 FREQC: 0.143841 ppm/cm
 SFOC: 101.625000 MHz/cm

1H spectrum



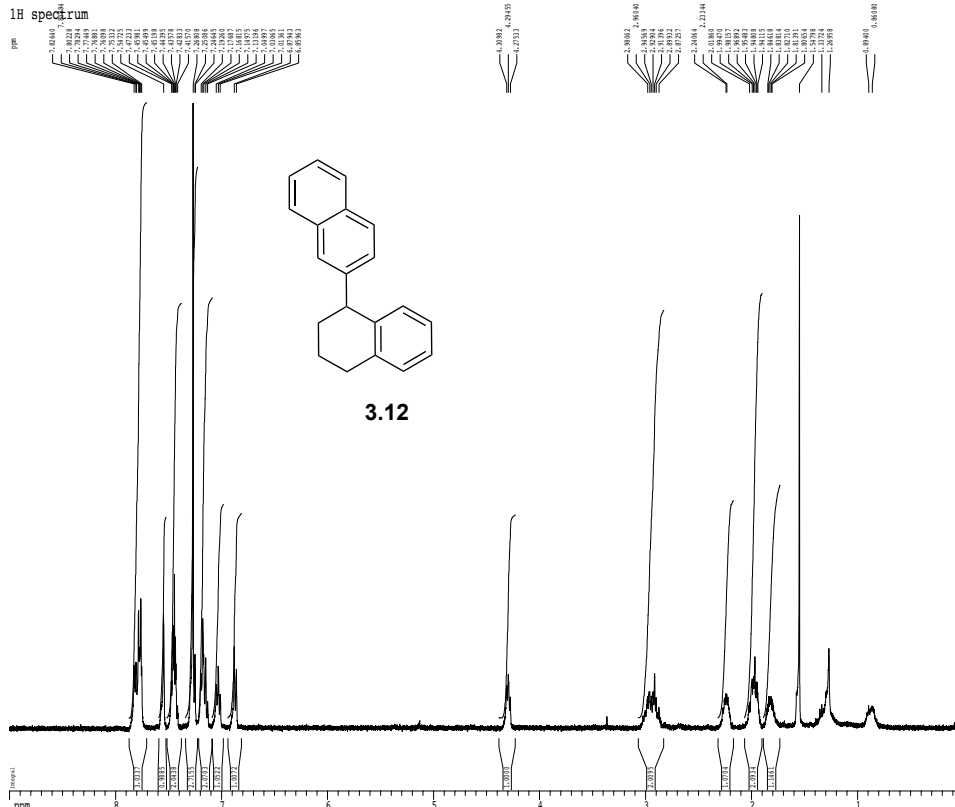
Current Data Parameters
NAME: 3.539-11-020917-C-H2O
PROCNO: 1
F2 - Acquisition Parameters
Date_: 21.47
Time: 20150709
INSTRUM: spect
PROBHD: 5 mm QNP 1H/1
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 2
DS: 4
SWH: 6433.234 Hz
FIDRES: 3.998891 sec
AQ: 0.1100000 sec
RG: 655.36
SR: 78.000 MHz
TE: 298.2 K
DE: 0.1000000 sec
TE1: 0.1000000 sec
TE2: 0.1000000 sec
TE3: 0.1000000 sec
TE4: 0.1000000 sec
TE5: 0.1000000 sec
TE6: 0.1000000 sec
TE7: 0.1000000 sec
TE8: 0.1000000 sec
TE9: 0.1000000 sec
TE10: 0.1000000 sec
===== CHANNEL f1 =====
NUC1: 13C
PUL1: zgpg30
SFO1: 400.12609 MHz
F2 - Processing parameters
SI: 32768
SF: 400.12609 MHz
WDW: EM
SSB: 0
LB: 0.40 Hz
GB: 0
PC: 2.00
===== 1D 13C NMR plot parameters =====
SI: 32768
SF: 400.12609 MHz
WDW: EM
SSB: 0
LB: 0.40 Hz
GB: 0
PC: 2.00
===== 2D 13C NMR plot parameters =====
SI: 32768
SF: 400.12609 MHz
WDW: EM
SSB: 0
LB: 0.40 Hz
GB: 0
PC: 2.00
===== 3D 13C NMR plot parameters =====
SI: 32768
SF: 400.12609 MHz
WDW: EM
SSB: 0
LB: 0.40 Hz
GB: 0
PC: 2.00

13C spectrum with 1H decoupling



Current Data Parameters
NAME: 3.539-11-020917-C-H2O
PROCNO: 1
F2 - Acquisition Parameters
Date_: 21.47
Time: 20150709
INSTRUM: spect
PROBHD: 5 mm QNP 1H/1
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 2
DS: 4
SWH: 24154.350 Hz
FIDRES: 0.348570 Hz
AQ: 1.106442 sec
RG: 655.36
SR: 20.700 MHz
TE: 298.2 K
DE: 0.1000000 sec
TE1: 0.1000000 sec
TE2: 0.1000000 sec
TE3: 0.1000000 sec
TE4: 0.1000000 sec
TE5: 0.1000000 sec
TE6: 0.1000000 sec
TE7: 0.1000000 sec
TE8: 0.1000000 sec
TE9: 0.1000000 sec
TE10: 0.1000000 sec
===== CHANNEL f1 =====
NUC1: 13C
PUL1: zgpg30
SFO1: 100.62786 MHz
F2 - Processing parameters
SI: 32768
SF: 100.62786 MHz
WDW: EM
SSB: 0
LB: 0.40 Hz
GB: 0
PC: 2.00
===== 1D 13C NMR plot parameters =====
SI: 32768
SF: 100.62786 MHz
WDW: EM
SSB: 0
LB: 0.40 Hz
GB: 0
PC: 2.00
===== 2D 13C NMR plot parameters =====
SI: 32768
SF: 100.62786 MHz
WDW: EM
SSB: 0
LB: 0.40 Hz
GB: 0
PC: 2.00
===== 3D 13C NMR plot parameters =====
SI: 32768
SF: 100.62786 MHz
WDW: EM
SSB: 0
LB: 0.40 Hz
GB: 0
PC: 2.00

1H spectrum



```

Current Data Parameters
NAME 2,8,4-85-cl-12D 1hanna
EXPNO 1
PROCNO 1

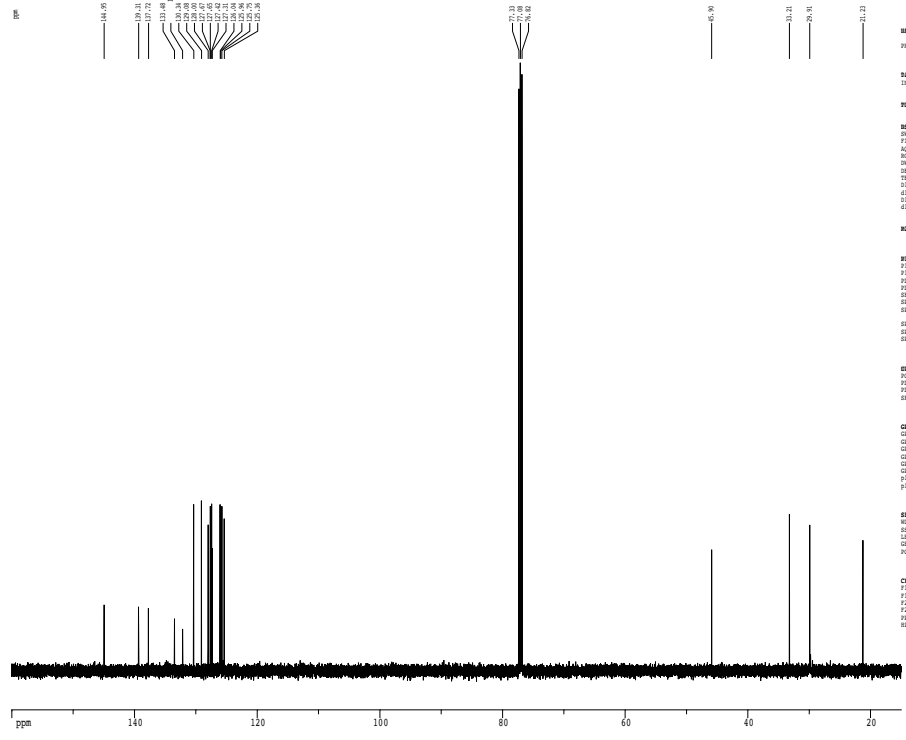
F2 - Acquisition Parameters
Date_   14.11
Time     09:10
INSTRUM crys16
PROBHD  5 mm QNP 1H/1
PULPROG zgpg30
SOLVENT  crys16
NS       2
DS       4
SWH      6410.256 Hz
FIDRES   0.097813 Hz
AQ       5.111579 sec
RG       741.4
SH       78.000 usec
TE       298.2 K
SI       0.1000000 sec
RGHRES   0.0100000 sec

===== CHANNEL f1 =====
NUC1     13C
P1      12.00 usec
SFO1    400.1328000 MHz

F2 - Processing parameters
SI       0
SF       400.130070 MHz 45534
WDW      EM
SSB      0
LB       0.00 Hz
GB       0
PC       2.00

1D NMR plot parameters
SI       0
F1      3601.17 Hz
F2      0.000 ppm
F3      0.00 Hz
P1      0.00 Hz
P2      0.00 Hz
P3      0.29474 ppm/cm
SFO1    157.94466 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
NAME 2,8,4-85-cl-12D 1hanna
EXPNO 1
PROCNO 1

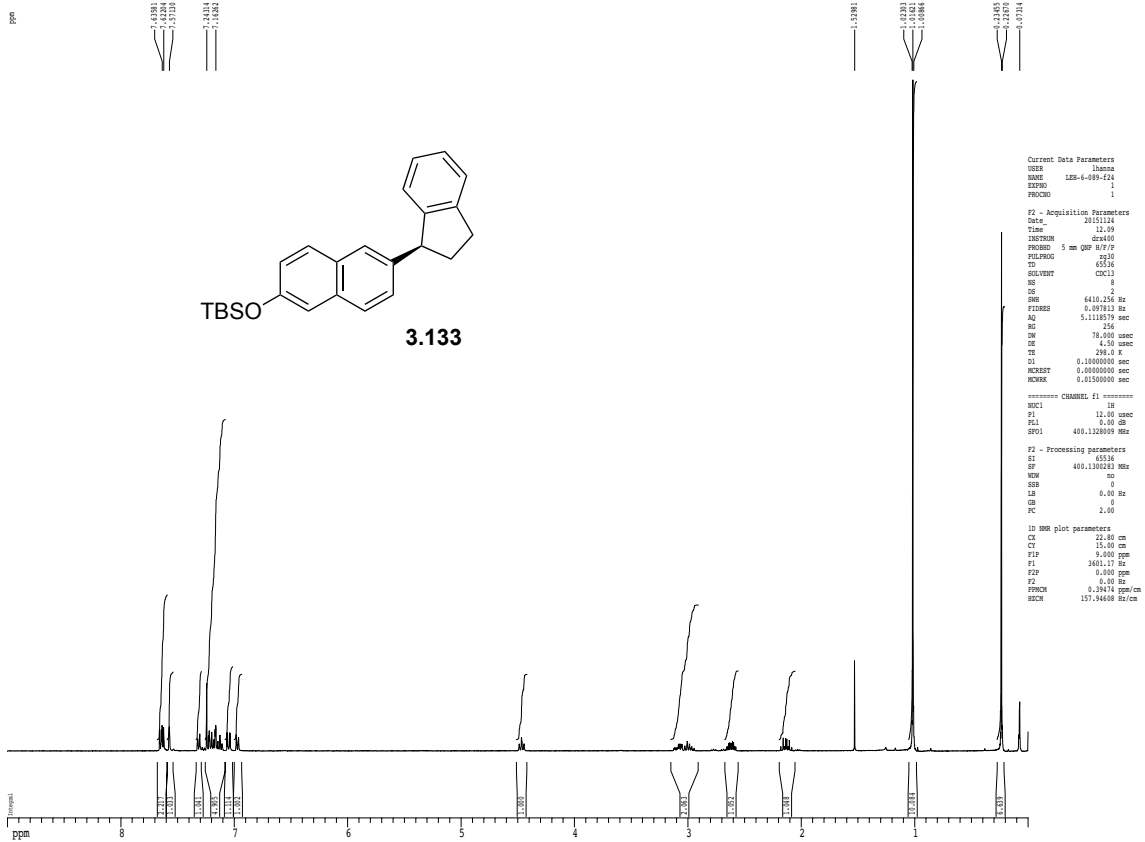
F2 - Acquisition Parameters
Date_   14.11
Time     09:10
INSTRUM crys16
PROBHD  5 mm QNP 1H-
PULPROG zgpg30
SOLVENT  crys16
NS       2
DS       4
SWH      6410.256 Hz
FIDRES   0.097813 Hz
AQ       5.111579 sec
RG       741.4
SH       78.000 usec
TE       298.2 K
SI       0.1000000 sec
RGHRES   0.0100000 sec

===== CHANNEL f1 =====
NUC1     13C
P1      12.00 usec
SFO1    400.1328000 MHz

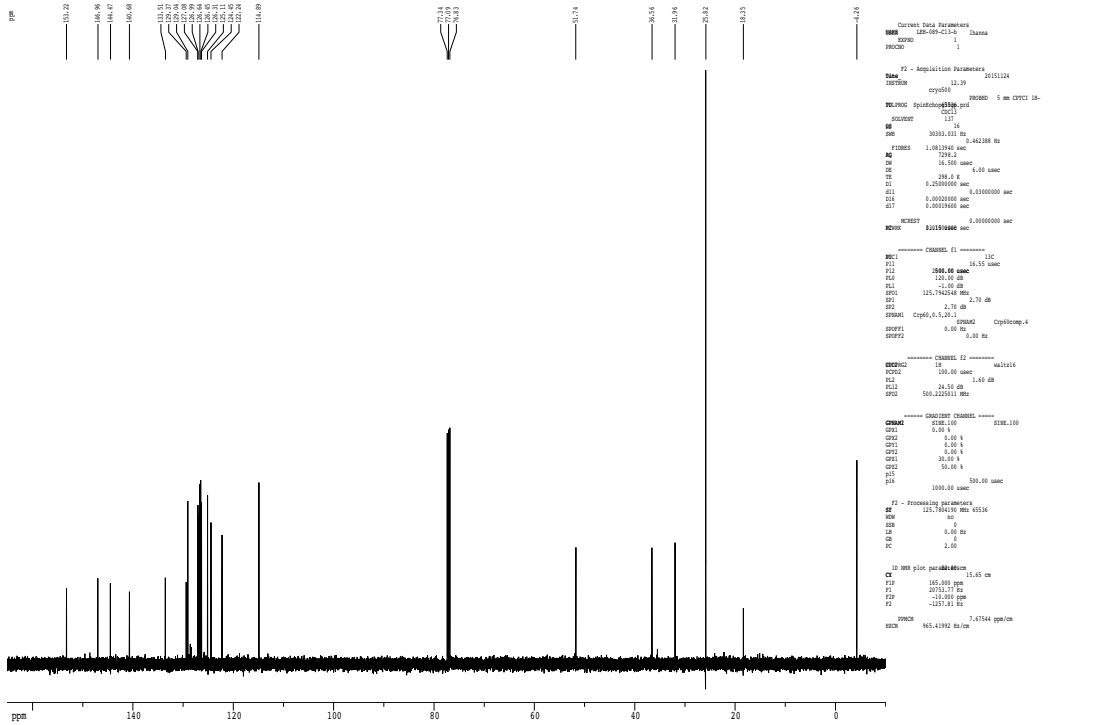
F2 - Processing parameters
SI       0
SF       400.130070 MHz 45534
WDW      EM
SSB      0
LB       0.00 Hz
GB       0
PC       2.00

1D NMR plot parameters
SI       0
F1      3601.17 Hz
F2      0.000 ppm
F3      0.00 Hz
P1      0.00 Hz
P2      0.00 Hz
P3      0.29474 ppm/cm
SFO1    157.94466 Hz/cm
    
```

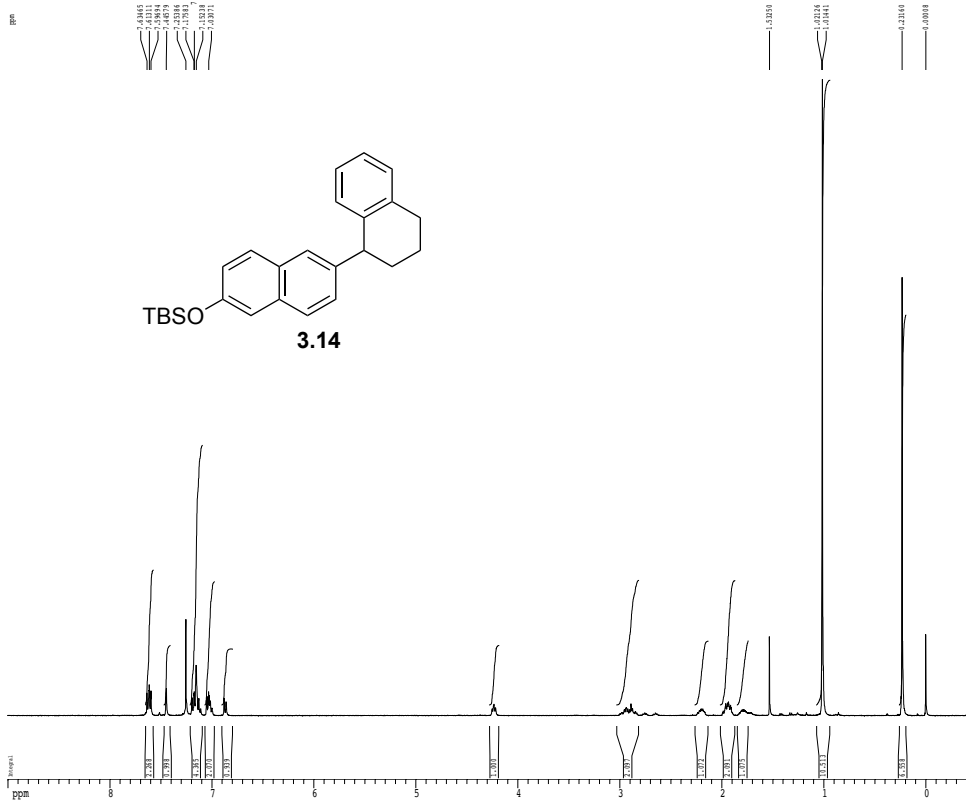
1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



Current Data Parameters
 NAME 3.14-11-2720 080609
 EXPNO 1
 PROCNO 1

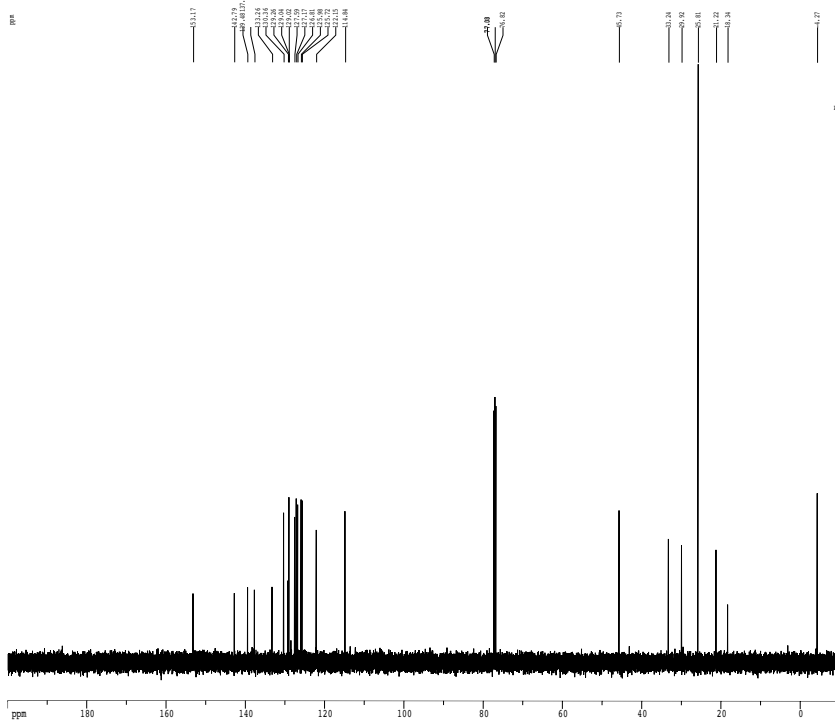
F2 - Acquisition Parameters
 20150817
 INSTRUM draco
 PROBHD 5 mm QNP R77/9
 PULPROG zgpg30
 SOLVENT CDCl3
 NS 2
 DS 4
 SWH 4410.256 Hz
 FIDRES 1.9999700 sec
 AQ 512
 RM 93.000 usec
 DE 390.0 u
 DI 0.1000000 sec
 MCNMR 0.0100000 sec
 MCHRES 0.0000000 sec

===== CHANNEL f1 =====
 NU1 18
 PL1 400.1128000 MHz
 SFO1 400.1128000 MHz

F2 - Processing parameters
 SI 400.1128000 MHz
 RF 400.1128000 MHz
 NS 2
 DS 4
 SW 0.00 Hz
 GB 0
 PC 2.00

10 MHz plot parameters
 CE 15.00 cm
 F2 300.137 Hz
 F1 251.137 Hz
 F2 -251.137 Hz
 F1 -251.137 Hz
 F2 251.137 Hz
 FWHM 0.41667 ppm/cm
 HCN 166.72086 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



===== CHANNEL f1 =====
 NAME 3.14-11-2720 080609
 EXPNO 1
 PROCNO 1

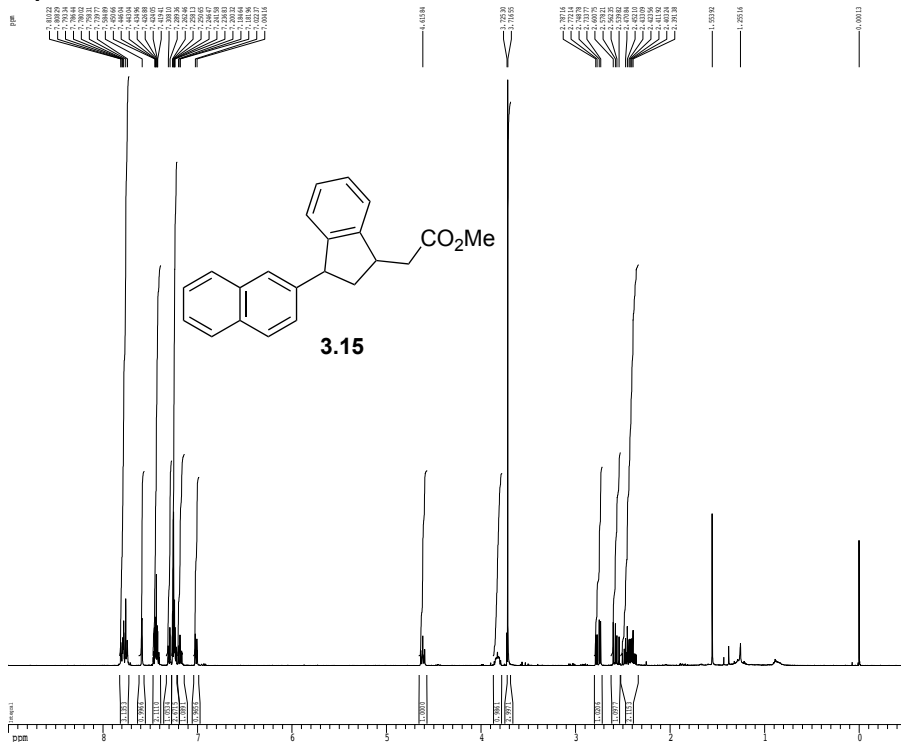
F2 - Acquisition Parameters
 20150817
 INSTRUM draco
 PROBHD 5 mm QNP R77/9
 PULPROG zgpg30
 SOLVENT CDCl3
 NS 2
 DS 4
 SWH 4410.256 Hz
 FIDRES 1.9999700 sec
 AQ 512
 RM 93.000 usec
 DE 390.0 u
 DI 0.1000000 sec
 MCNMR 0.0100000 sec
 MCHRES 0.0000000 sec

===== CHANNEL f2 =====
 NU2 18
 PL2 101.6261250 MHz
 SFO2 101.6261250 MHz

F2 - Processing parameters
 SI 101.6261250 MHz
 RF 101.6261250 MHz
 NS 2
 DS 4
 SW 0.00 Hz
 GB 0
 PC 2.00

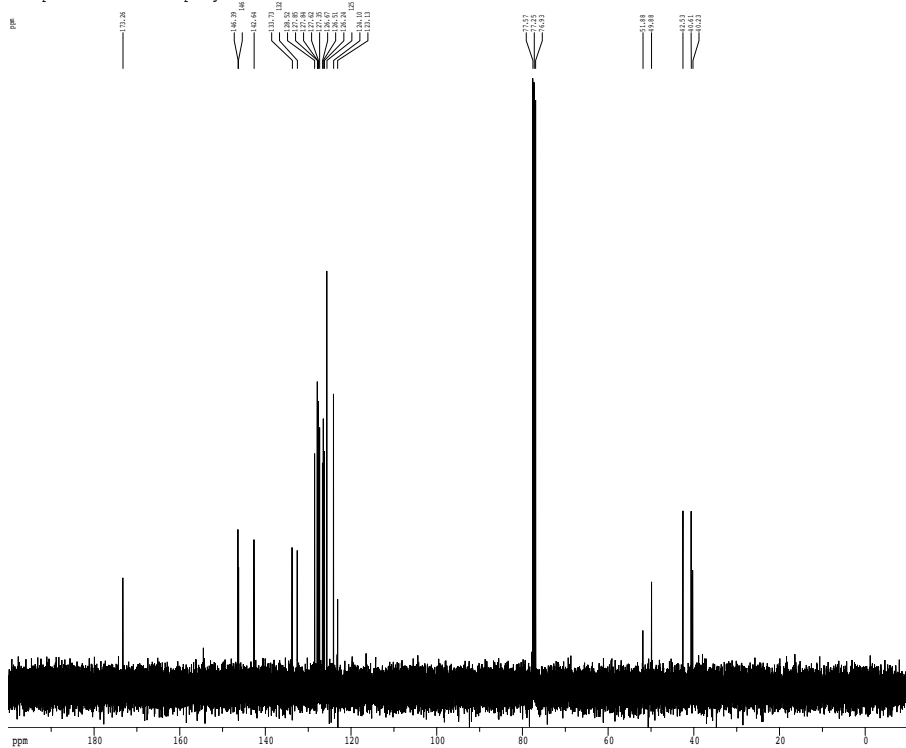
10 MHz plot parameters
 CE 15.00 cm
 F2 251.137 Hz
 F1 251.137 Hz
 F2 -251.137 Hz
 F1 -251.137 Hz
 F2 251.137 Hz
 FWHM 0.41667 ppm/cm
 HCN 166.72086 Hz/cm

1H spectrum



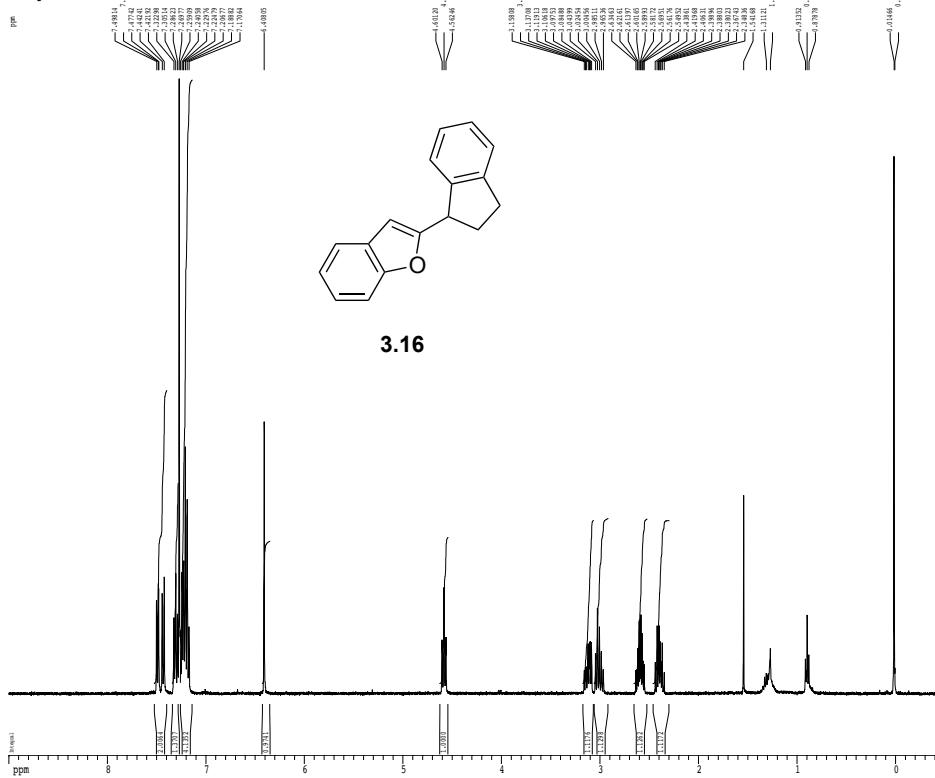
```
Current Data Parameters
NAME 3.15
EXPNO 1
PROCNO 1
F2 - Acquisition Parameters
Date_ 20150709
Time 14:28
INSTRUM spect
PROBHD 5 mm QNP 1H/1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 4
DS 4
SWH 6433.254 Hz
FIDRES 0.16673 Hz
AQ 2.79292 sec
RG 2072
AQ 78.300 usec
DE 6.20 usec
TE 298.2 K
D1 0.10000000 sec
d11 0.10000000 sec
dCHAPT 0.00000000 sec
dCHRG 0.02000000 sec
===== CHANNEL f1 =====
NUC1 13C
P1 12.00 usec
PL1 0.00 dB
SFO1 400.126000 MHz
F2 - Processing parameters
SI 65536
SF 400.1260000 MHz
WDW no
SSB 0
GB 0
PC 2.00
ID 800 plot parameters
SI 22.00 cm
CF 10.00 cm
F1P 0.000 ppm
F1 3601.71 Hz
F2P -0.500 ppm
F2 -600.11 Hz
PPHCA 9.43447 ppm/cm
PPHCB 94.70888 Hz/cm
```

13C spectrum with 1H decoupling



```
Current Data Parameters
NAME 3.15
EXPNO 1
PROCNO 1
F2 - Acquisition Parameters
Date_ 20150709
Time 14:10
INSTRUM spect
PROBHD 5 mm QNP 1H/1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 4
DS 4
SWH 24534.595 Hz
FIDRES 0.386570 Hz
AQ 1.9544652 sec
RG 939.2
AQ 20.790 usec
DE 20.29 usec
TE 298.2 K
D1 0.10000000 sec
d11 0.10000000 sec
dCHAPT 0.00000000 sec
dCHRG 0.01000000 sec
===== CHANNEL f1 =====
NUC1 13C
P1 12.00 usec
PL1 0.00 dB
SFO1 100.621796 MHz
===== CHANNEL f2 =====
NAME 13C
PCPD2 13
PL2 90.00 usec
PL12 170.000 dB
SFO2 400.126000 MHz
F2 - Processing parameters
SI 65536
SF 100.621796 MHz
WDW no
SSB 0
GB 0
PC 1.00
ID 800 plot parameters
SI 15.50 cm
CF 100.000 ppm
F1 30123.05 Hz
F2 -10000.11 Hz
PPHCA 9.21053 ppm/cm
PPHCB 932.69629 Hz/cm
```

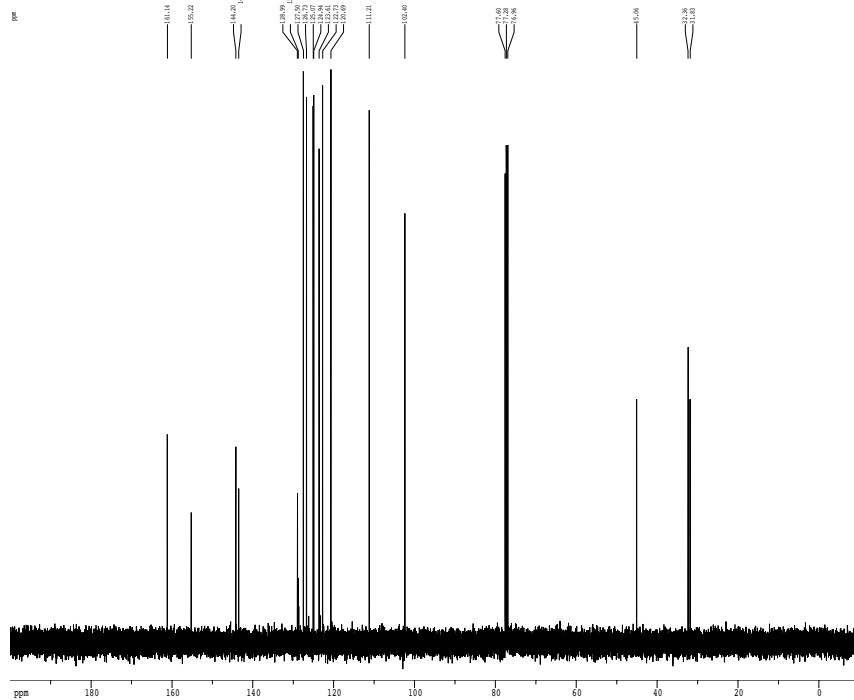
1H spectrum



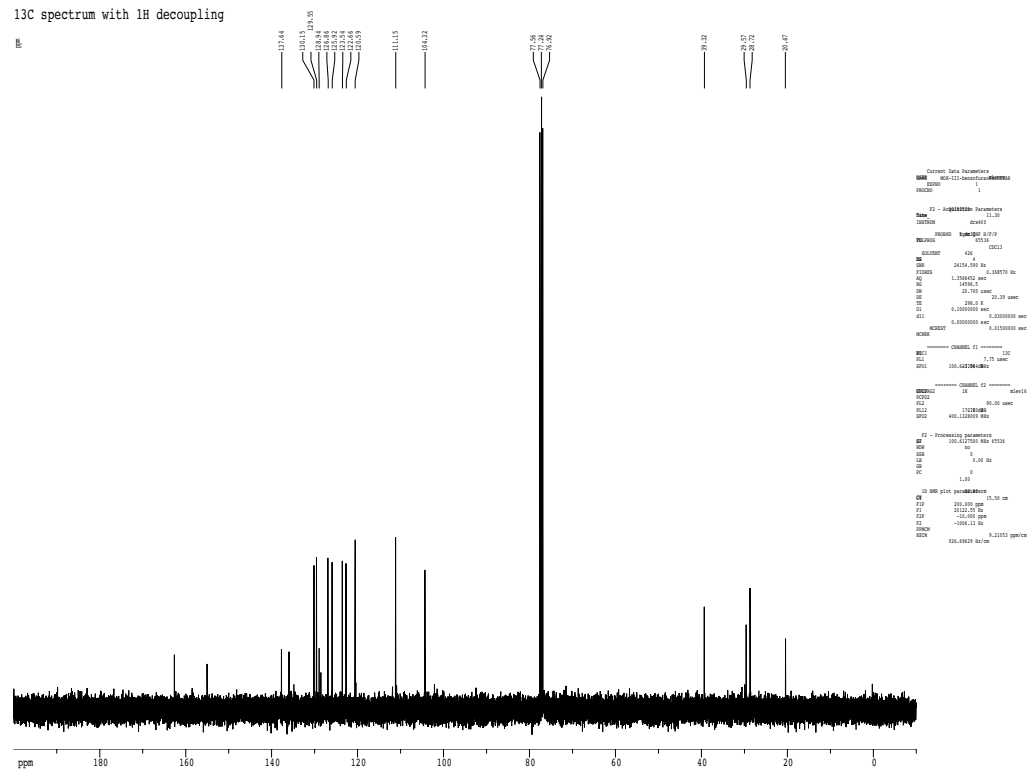
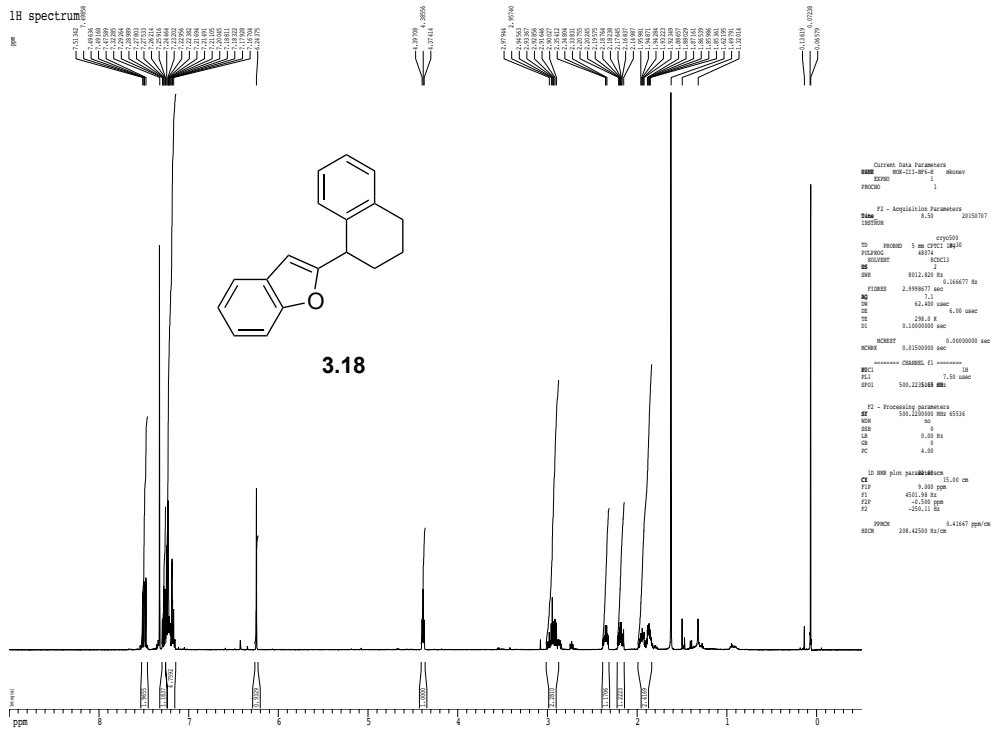
Current Data Parameters
 NAME: 400-120-0000-00000000
 EXPNO: 1
 PROCNO: 1
 F2 - Acquisition Parameters
 Date_ Time: 20151030 14:03
 INSTRUM: spect
 PROBRD: 5 mm QNP 8/197
 PULPROG: zgpg30
 SOLVENT: CDCl3
 NS: 640
 DS: 4
 FIDRES: 0.250000 Hz
 AQ: 0.12777777777777778 sec
 TM: 1.0000000000000000 sec
 DE: 4.50 umol
 TE: 297.5 K
 D1: 0.10000000000000000 sec
 DELTAD: 0.00000000000000000 sec
 NUC1: 13C
 NUC2: 1H
 P1: 12.00 umol
 SFO1: 400.120000 MHz
 F2 - Processing parameters
 SI: 32768
 SF: 400.1200000 MHz
 WDW: EM
 SSB: 0
 LB: 0.00 Hz
 GB: 0
 PC: 2.00
 MD NMR plot parameters
 CT: 15.00 cm
 F1: 9.1000 ppm
 F2: 3632.12 Hz
 F3: -0.500 ppm
 F4: -200.00 Hz
 PUNCH: 0.41667 ppm/cm
 HXCN: 166.7286 Hz/cm

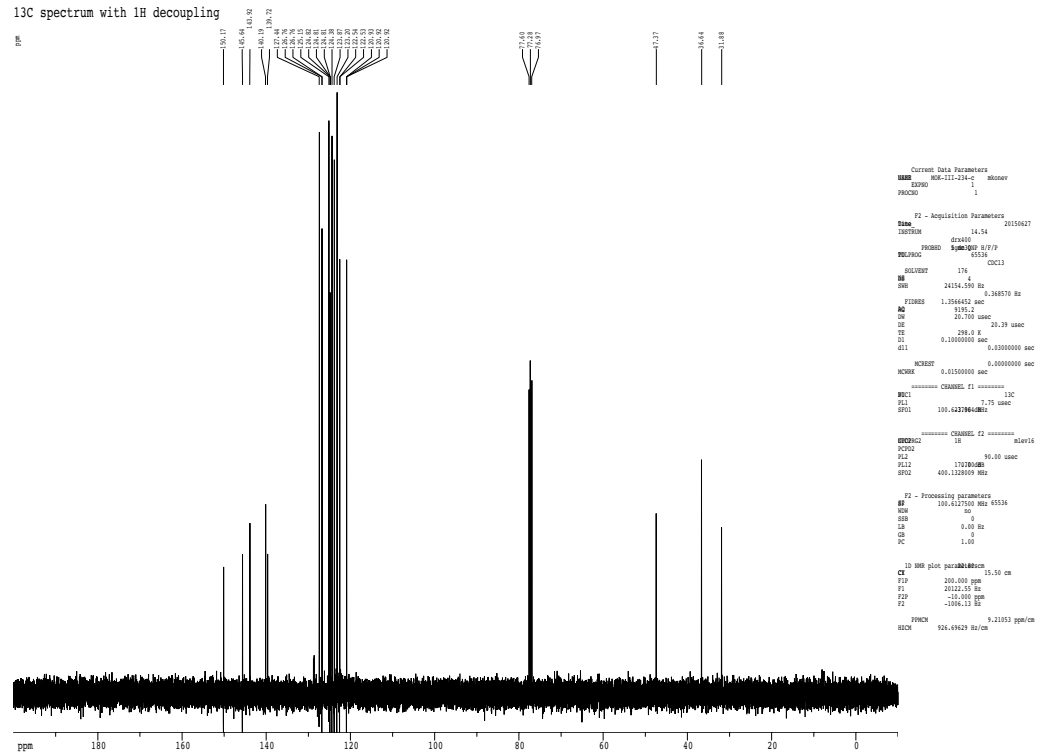
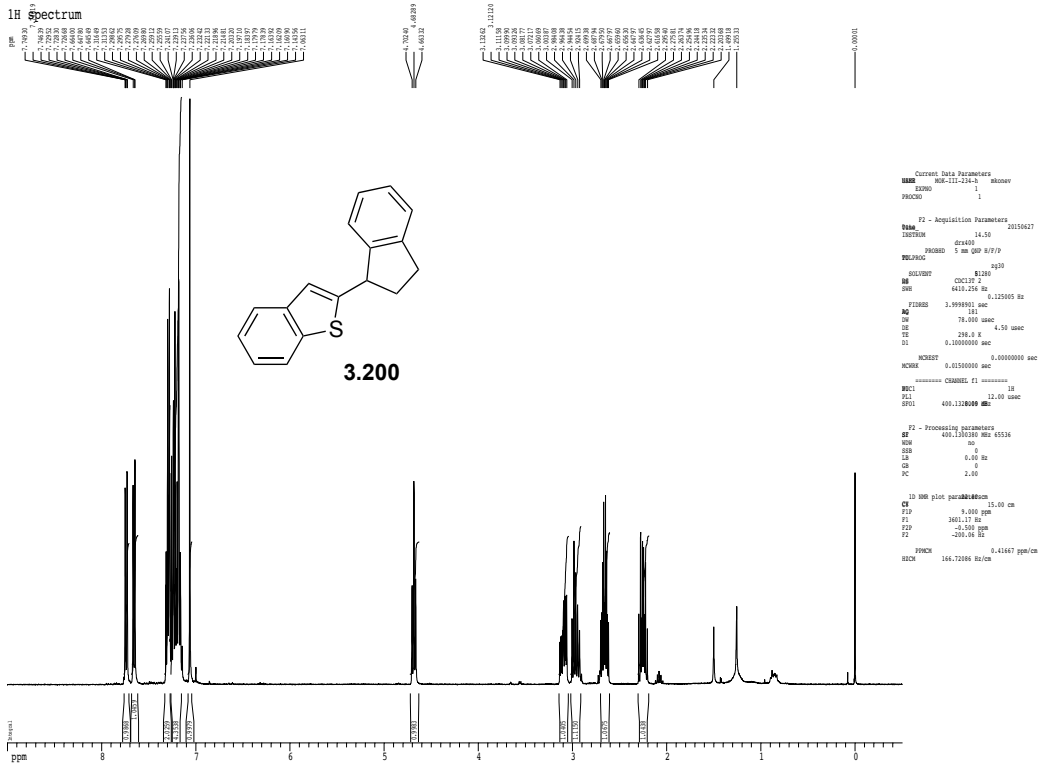
S68

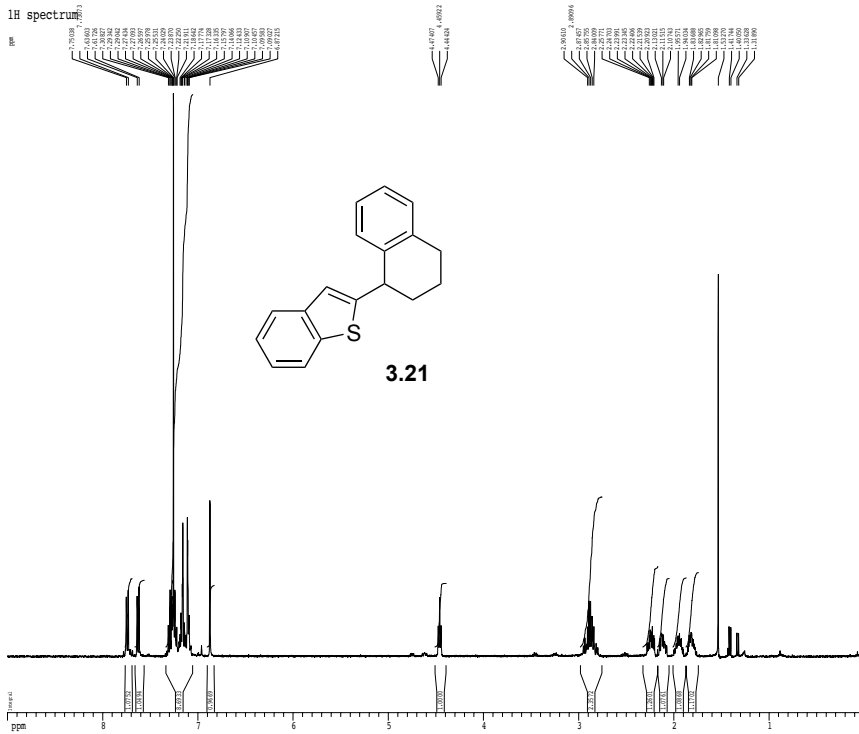
13C spectrum with 1H decoupling



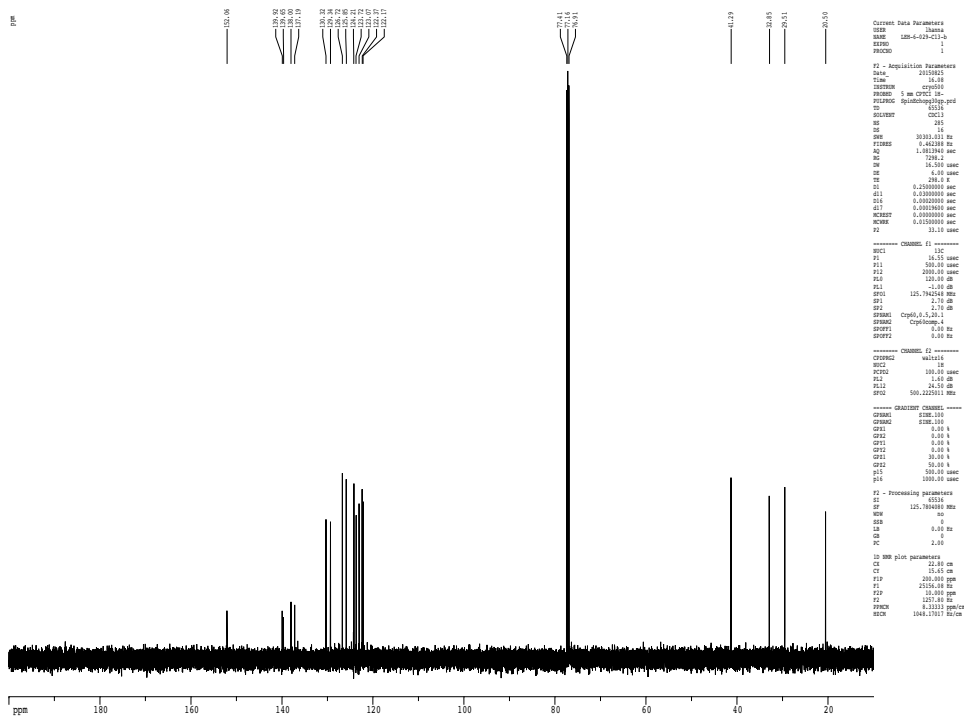
Current Data Parameters
 NAME: 400-120-0000-00000000
 EXPNO: 1
 PROCNO: 1
 F2 - Acquisition Parameters
 Date_ Time: 20151030 14:03
 INSTRUM: spect
 PROBRD: 5 mm QNP 8/197
 PULPROG: zgpg30
 SOLVENT: CDCl3
 NS: 640
 DS: 4
 FIDRES: 0.250000 Hz
 AQ: 0.12777777777777778 sec
 TM: 1.0000000000000000 sec
 DE: 4.50 umol
 TE: 297.5 K
 D1: 0.10000000000000000 sec
 DELTAD: 0.00000000000000000 sec
 NUC1: 13C
 NUC2: 1H
 P1: 12.00 umol
 SFO1: 400.120000 MHz
 F2 - Processing parameters
 SI: 32768
 SF: 100.6200000 MHz
 WDW: EM
 SSB: 0
 LB: 0.00 Hz
 GB: 0
 PC: 1.00
 MD NMR plot parameters
 CT: 15.00 cm
 F1: 9.1000 ppm
 F2: 3632.12 Hz
 F3: -0.500 ppm
 F4: -200.00 Hz
 PUNCH: 0.41667 ppm/cm
 HXCN: 166.7286 Hz/cm



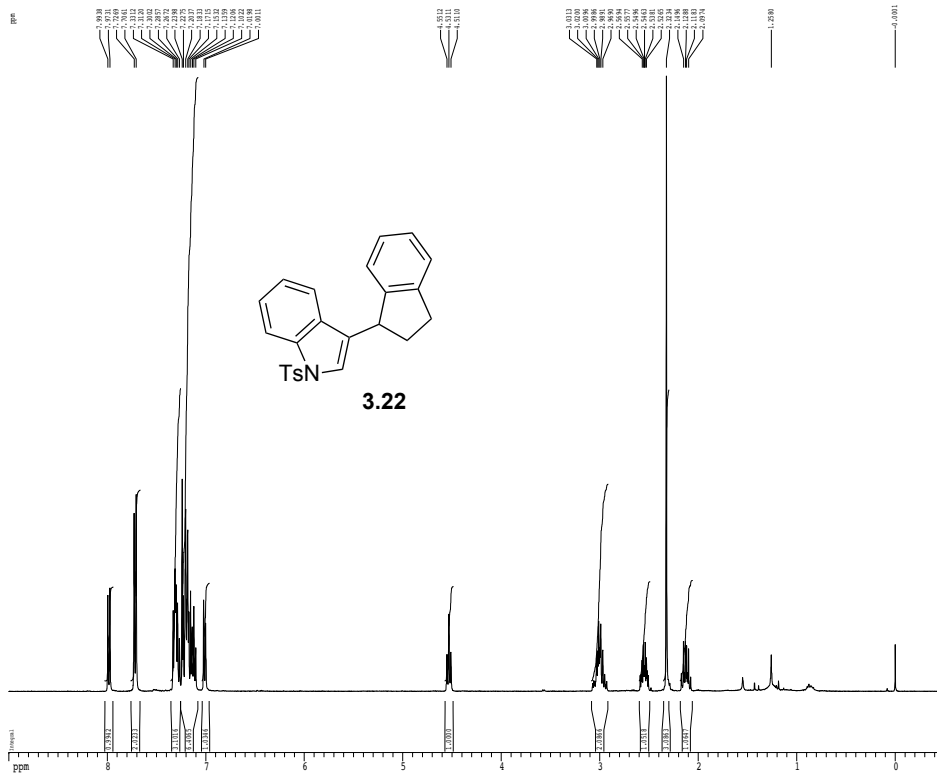




2-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum

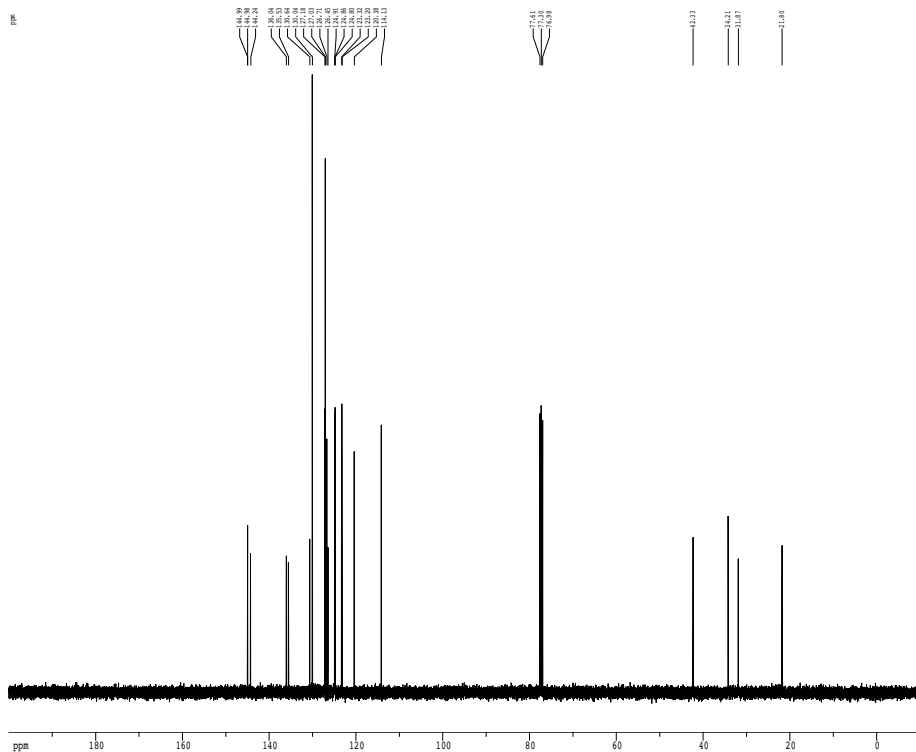


Current Data Parameters
NAME NMR-111-IndoleAcCCMuu
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
DATE_ 20150509
INSTRUM spect
PROBHD 5 mm QNP 1H/13
PULPROG zgpg30
DELTA 3055
SOLVENT DMSO-d6
NS 640
DS 4
SWH 6410.256 Hz
FIDRES 0.20000 Hz
AQ 2.99999 sec
RG 327.5
DW 78.000 usec
DE 4.50 usec
TE 298.2 K
D1 0.10000000 sec
DELTA2 0.15000000 sec
DELTA3 0.00000000 sec
DELTA4 0.00000000 sec
CHANNEL f1
NUC1 1H
NUC2 13C
NUC3 15N
RF1 400.142000 MHz
RF2 101.626126 MHz

F2 - Processing parameters
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00
PD 0.00000000
CD 0.00000000
CF 400.142000 MHz
CPDPRG zgpg30
FT 32768.000
FT2 101.626126
F2 101.626126 MHz
F1 400.142000 MHz
PROCNO 1
PRNCHN 146.72086 Hz/cm
RECN 146.72086 Hz/cm

13C spectrum with 1H decoupling

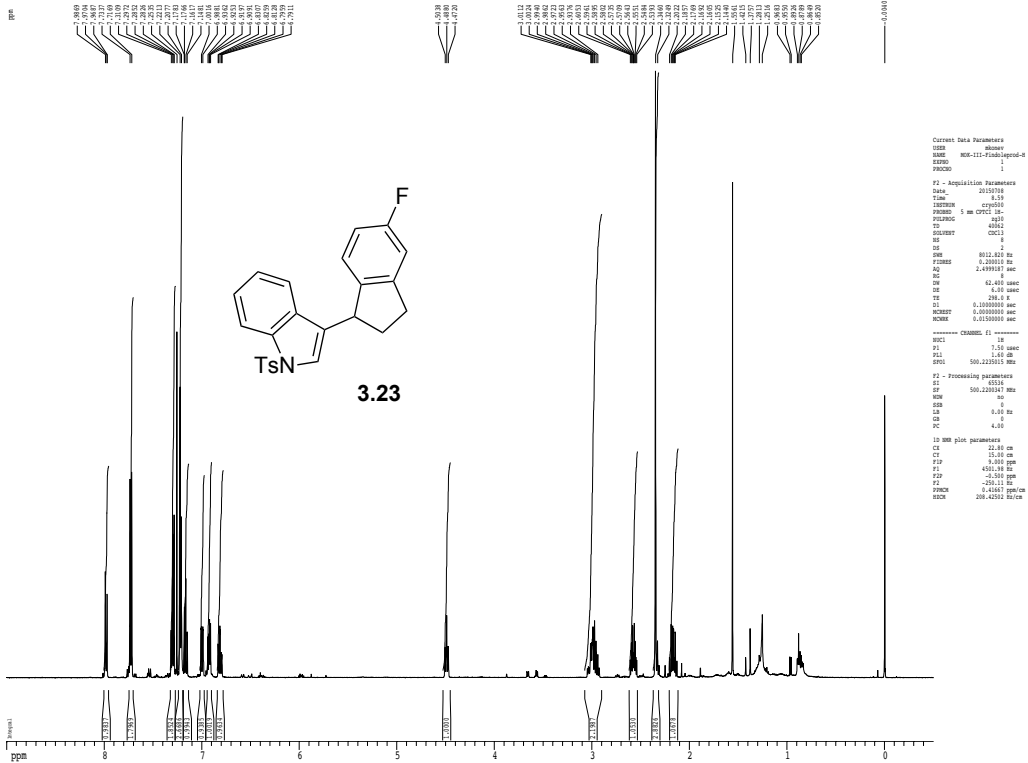


Current Data Parameters
NAME NMR-111-IndoleAcCCMuu
EXPNO 1
PROCNO 1

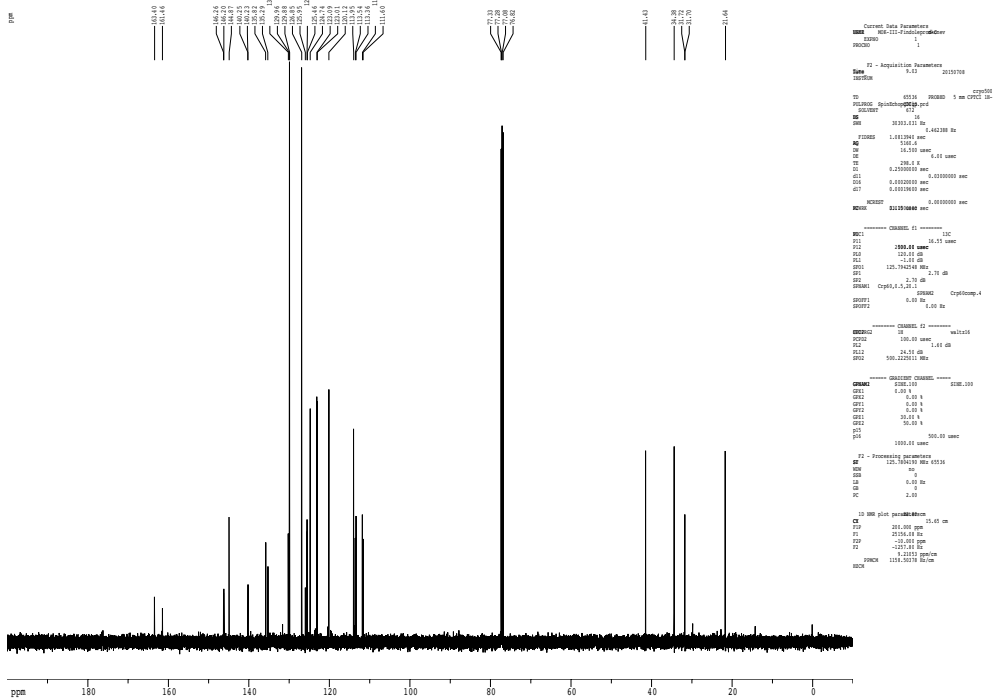
F2 - Acquisition Parameters
DATE_ 20150509
INSTRUM spect
PROBHD 5 mm QNP 1H/13
PULPROG zgpg30
DELTA 3055
SOLVENT DMSO-d6
NS 640
DS 4
SWH 24154.936 Hz
FIDRES 0.20000 Hz
AQ 2.99999 sec
RG 327.5
DW 78.000 usec
DE 4.50 usec
TE 298.2 K
D1 0.10000000 sec
DELTA2 0.15000000 sec
DELTA3 0.00000000 sec
DELTA4 0.00000000 sec
CHANNEL f1
NUC1 13C
NUC2 1H
RF1 101.626126 MHz
RF2 400.142000 MHz

F2 - Processing parameters
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00
PD 0.00000000
CD 0.00000000
CF 101.626126 MHz
CPDPRG zgpg30
FT 32768.000
FT2 400.142000
F2 400.142000 MHz
F1 101.626126 MHz
PROCNO 1
PRNCHN 926.69419 Hz/cm
RECN 926.69419 Hz/cm

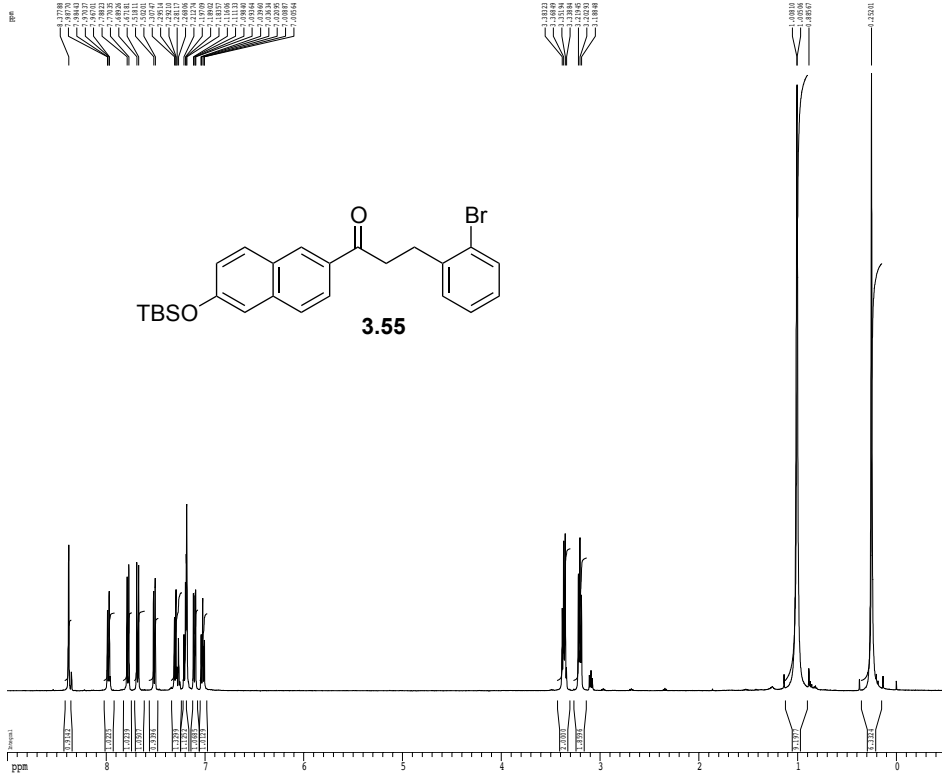
1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



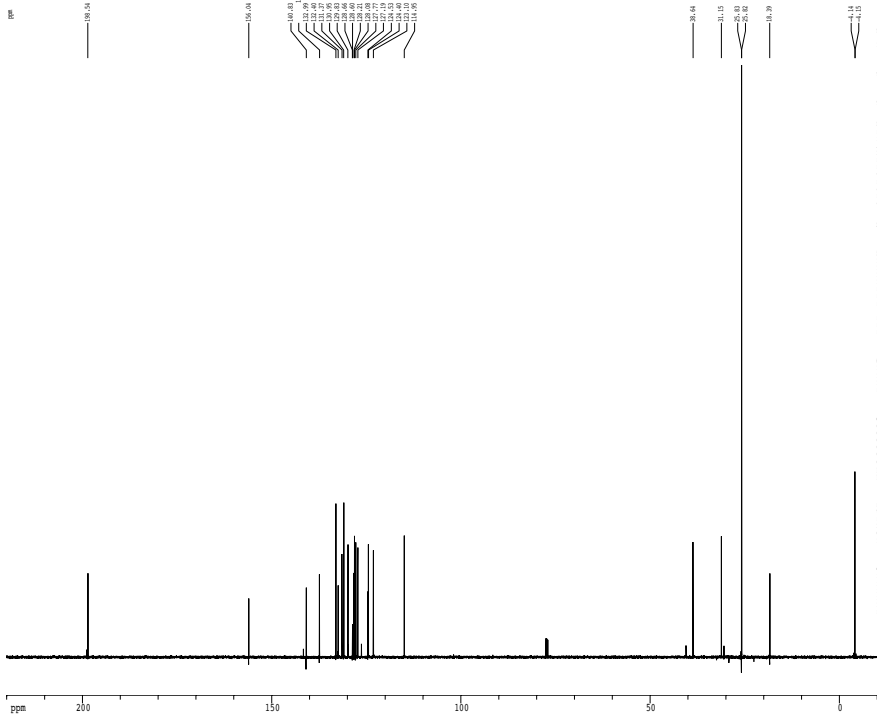
Current Data Parameters
USER: Mosew
NAME: M06-174-622chbr
EXPNO: 1
PROCNO: 1

F2 - Acquisition Parameters
Date_: 2011117
Time: 1.48
INSTRUM: cryo600
PROBHD: 5 mm CPXI 1H-
PULPROG: zgpg30
TD: 65536
SFO: 500.136099 MHz
AQ: 5.0998774 sec
RG: 1.00
WDW: EM
SSB: 0.000000 sec
LB: 0.00 Hz
GB: 0
PC: 4.00

F2 - Processing parameters
SI: 65536
SF: 500.136099 MHz
WDW: EM
SSB: 0
LB: 0.00 Hz
GB: 0
PC: 4.00

1D 1H NMR plot parameters
CH: 22.00 cm
CI: 15.00 cm
PI1: 8.000 ppm
PI2: 4501.38 Hz
F2: -3.500 ppm
F1: -226.11 Hz
PPHMC: 0.41667 ppm/cm
HSCX: 200.42000 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling

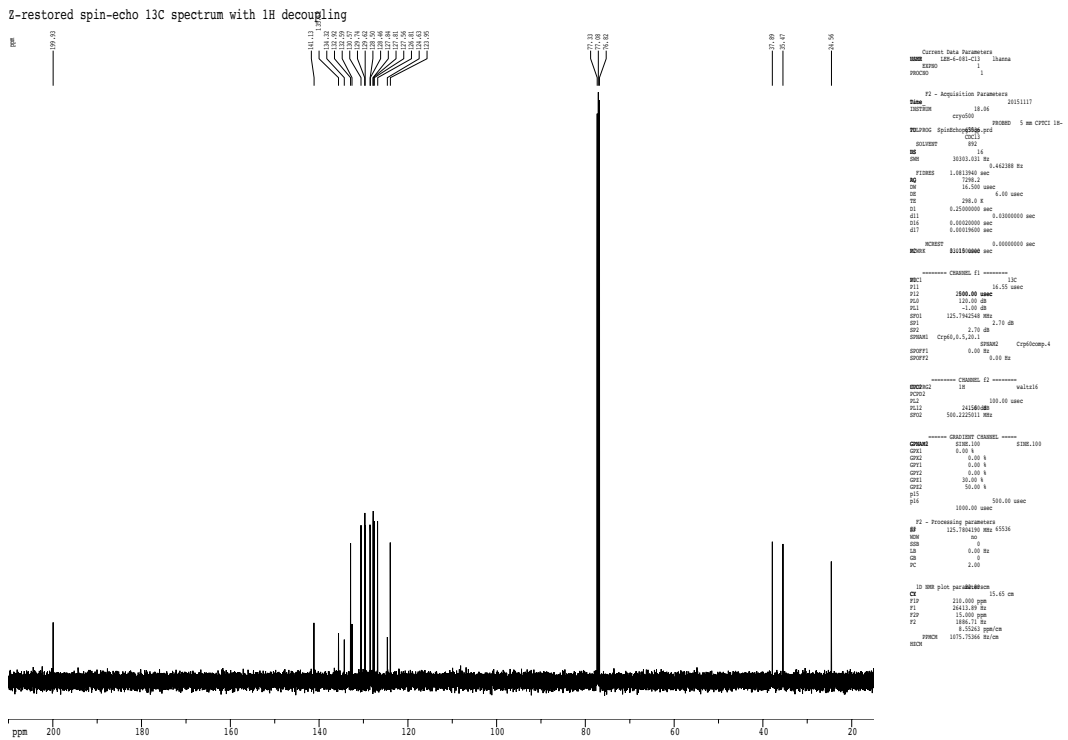
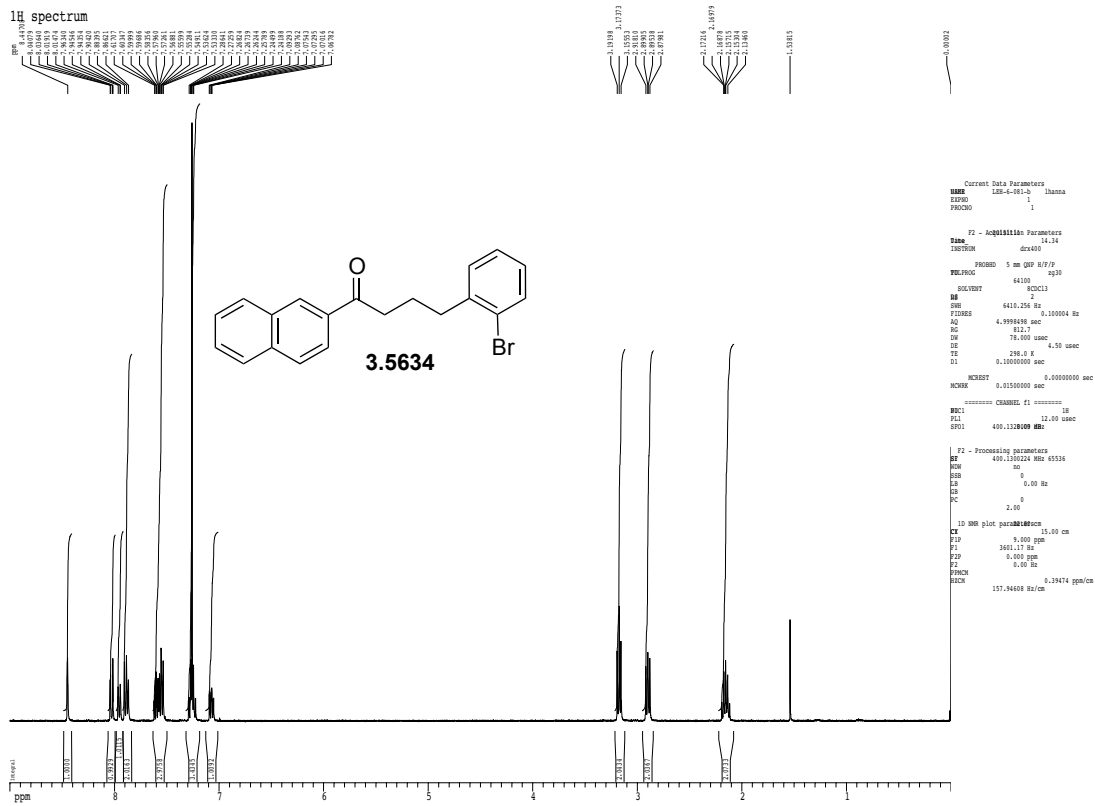


Current Data Parameters
USER: Mosew
NAME: M06-174-622chbr3
EXPNO: 1
PROCNO: 1

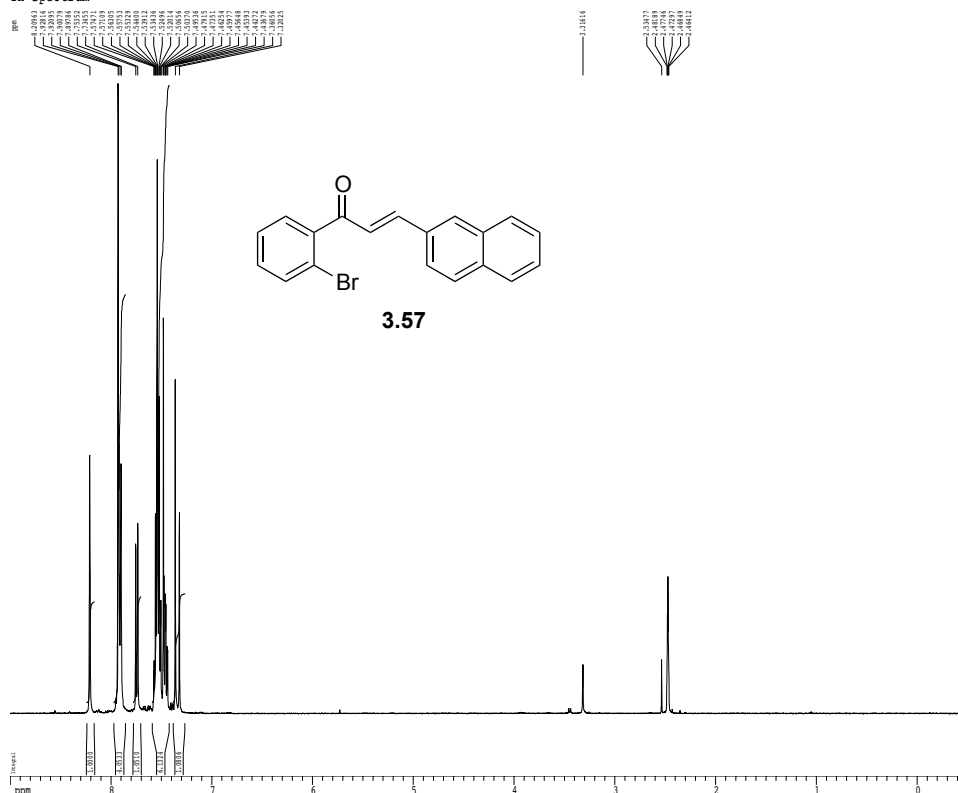
F2 - Acquisition Parameters
Date_: 2011117
Time: 1.54
INSTRUM: cryo600
PROBHD: 5 mm CPXI 1H-
PULPROG: zgpg30
TD: 65536
SFO: 500.136099 MHz
AQ: 5.0998774 sec
RG: 1.00
WDW: EM
SSB: 0.000000 sec
LB: 0.00 Hz
GB: 0
PC: 4.00

F2 - Processing parameters
SI: 65536
SF: 500.136099 MHz
WDW: EM
SSB: 0
LB: 0.00 Hz
GB: 0
PC: 4.00

1D 13C NMR plot parameters
CH: 15.00 cm
CI: 15.00 cm
PI1: 220.000 ppm
PI2: 27471.49 Hz
F2: -13.000 ppm
F1: -1257.09 Hz
PPHMC: 10.00773 ppm/cm
HSCX: 1200.87563 Hz/cm



1H spectrum

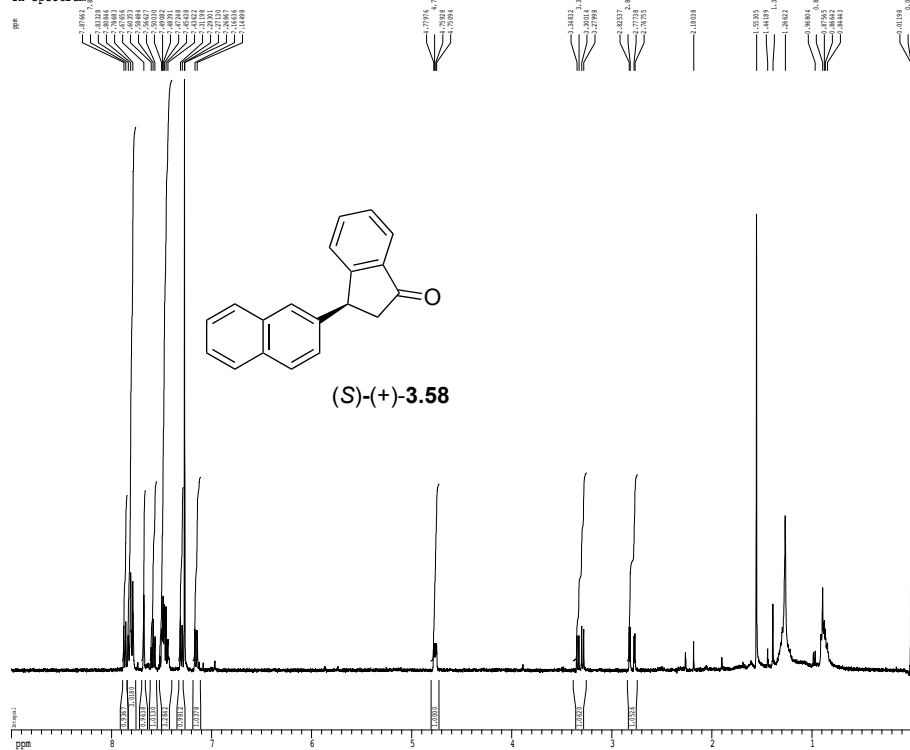


```

Current Data Parameters
NAME      H00000000
EXPNO     1
PROCNO    1
F2 - Acquisition Parameters
Date_     20151112
Time      17.24
INSTRUM   dr4000
PROBHD    5 mm QNP 8/7/P
PULPROG   zgpg30
TD         65536
SOLVENT   DMSO
NS         2
DS         2
SWH        6413.256 Hz
FIDRES     0.097818 Hz
AQ         5.1118179 sec
RG         261.1
NUC1       13
NUC2       13
DE         78.000 usec
TE         300.2 K
D1         0.10000000 sec
DELTA      0.10000000 sec
RG2        6.00000000 sec
RG3        6.00000000 sec
RG4        6.00000000 sec
RG5        6.00000000 sec
RG6        6.00000000 sec
RG7        6.00000000 sec
RG8        6.00000000 sec
RG9        6.00000000 sec
RG10       6.00000000 sec
RG11       6.00000000 sec
RG12       6.00000000 sec
RG13       6.00000000 sec
RG14       6.00000000 sec
RG15       6.00000000 sec
RG16       6.00000000 sec
RG17       6.00000000 sec
RG18       6.00000000 sec
RG19       6.00000000 sec
RG20       6.00000000 sec
===== CHANNEL f1 =====
NUC1       13
P1         12.00 usec
PC1        0.00 dB
SFO1       400.126009 MHz
F2 - Processing parameters
SI         65536
SF         400.1260175 MHz
WDW        EM
SSB        0
LA         0.00 Hz
GB         0
PC         2.00
1D NMR plot parameters
CX         22.00 cm
CT         12.00 cm
FT         9.000 ppm
F1         3601.17 Hz
F2         61.000 ppm
F3         0.00 Hz
PPMCH     0.41441 ppm/cm
SFOCH     164.72086 Hz/cm
    
```

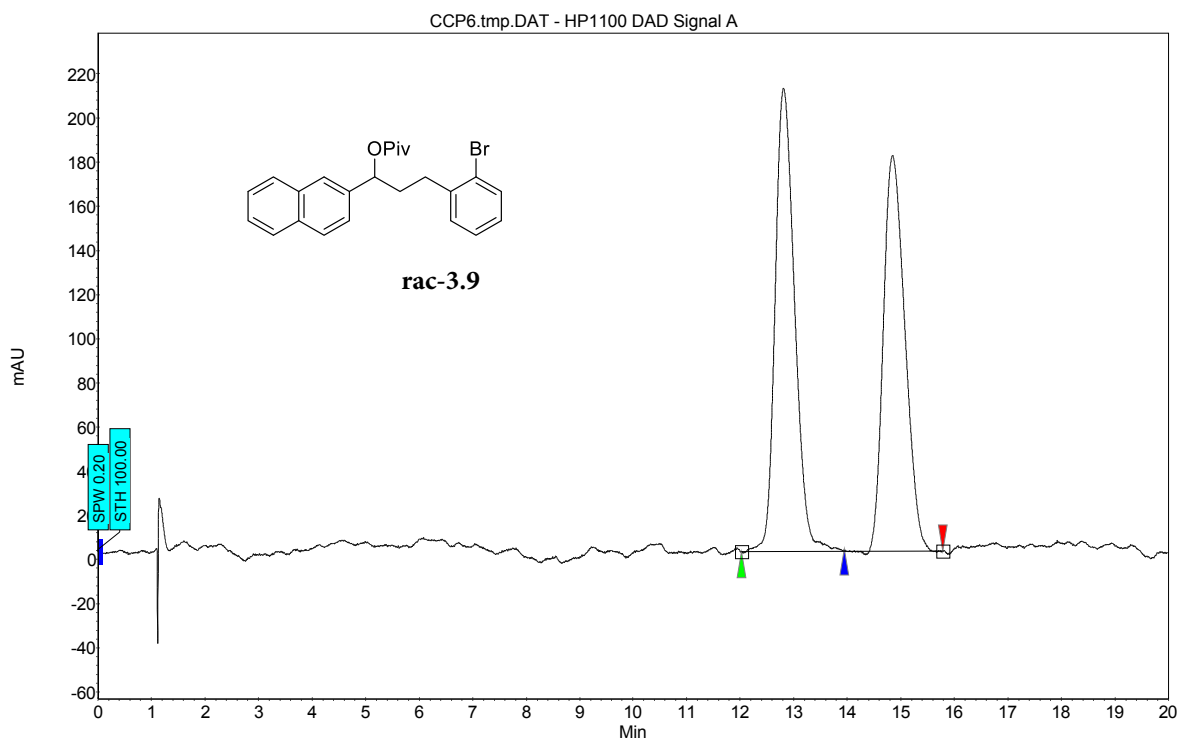
S96

1H spectrum

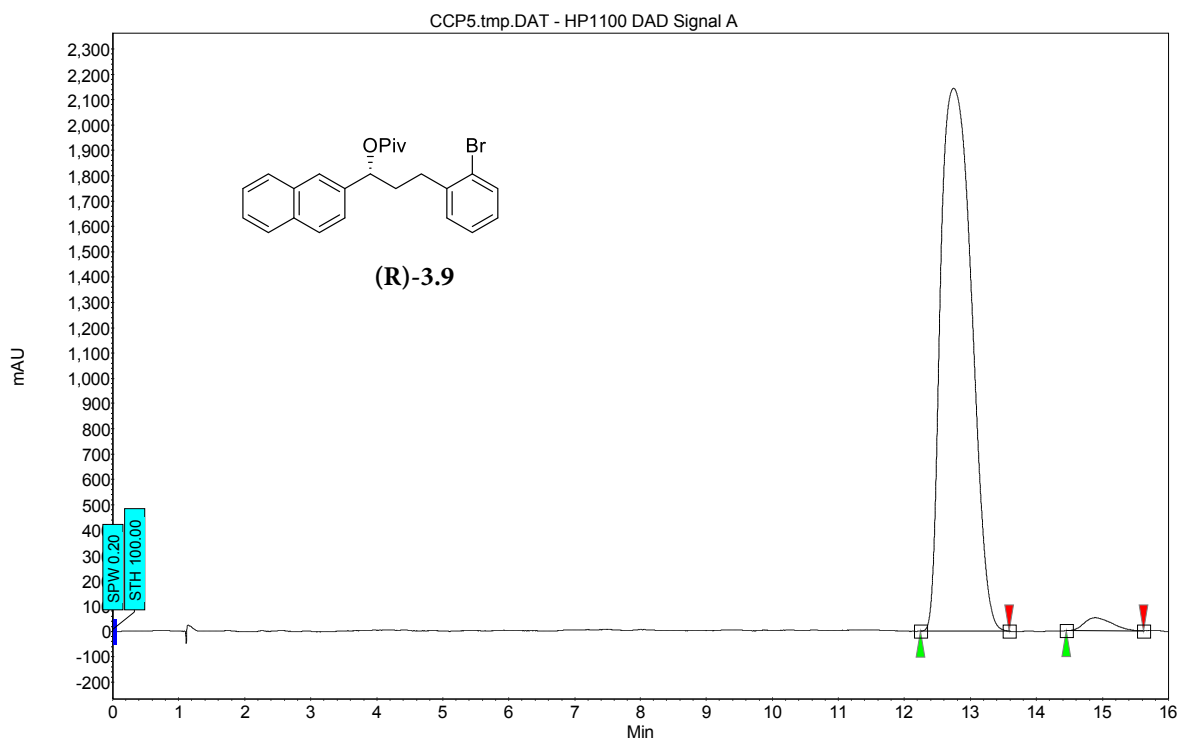


```

Current Data Parameters
NAME      1284-435-c1-c10 1hanna
EXPNO     1
PROCNO    1
F2 - Acquisition Parameters
Date_     20150831
Time      13.54
INSTRUM   dr4000
PROBHD    5 mm QNP 8/7/P
PULPROG   zgpg30
TD         26448
SOLVENT   d4-DMSO
NS         2
DS         2
SWH        6413.256 Hz
FIDRES     0.1250010 Hz
AQ         1.9997001 sec
RG         100
NUC1       13
NUC2       13
DE         78.000 usec
TE         300.2 K
D1         0.10000000 sec
DELTA      0.10000000 sec
RG2        6.00000000 sec
RG3        6.00000000 sec
RG4        6.00000000 sec
RG5        6.00000000 sec
RG6        6.00000000 sec
RG7        6.00000000 sec
RG8        6.00000000 sec
RG9        6.00000000 sec
RG10       6.00000000 sec
RG11       6.00000000 sec
RG12       6.00000000 sec
RG13       6.00000000 sec
RG14       6.00000000 sec
RG15       6.00000000 sec
RG16       6.00000000 sec
RG17       6.00000000 sec
RG18       6.00000000 sec
RG19       6.00000000 sec
RG20       6.00000000 sec
===== CHANNEL f1 =====
NUC1       13
P1         12.00 usec
PC1        0.00 dB
SFO1       400.126009 MHz
F2 - Processing parameters
SI         65536
SF         400.1260175 MHz
WDW        EM
SSB        0
LA         0.00 Hz
GB         0
PC         2.00
1D NMR plot parameters
CX         22.00 cm
CT         12.00 cm
FT         9.000 ppm
F1         3601.17 Hz
F2         61.000 ppm
F3         0.00 Hz
PPMCH     0.39476 ppm/cm
SFOCH     157.34406 Hz/cm
    
```



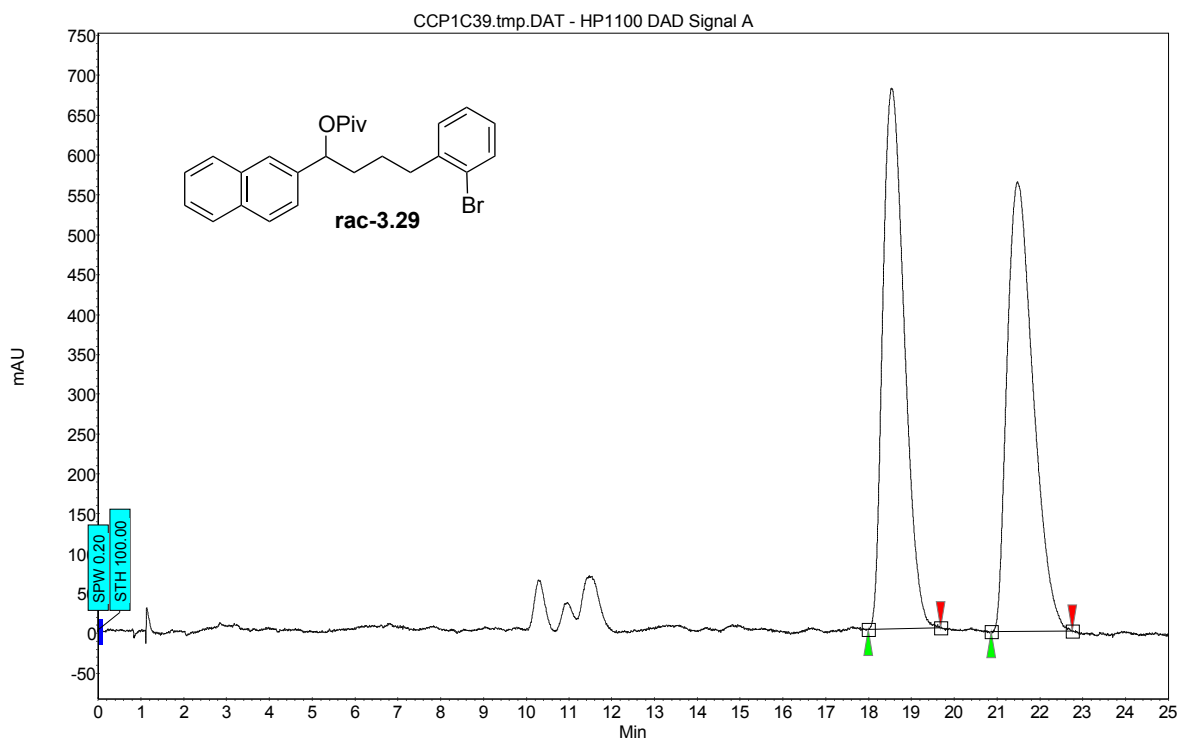
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	12.02	12.81	13.95	0.00	51.51	209.7	88.4	51.507
2	UNKNOWN	13.95	14.86	15.79	0.00	48.49	179.2	83.2	48.493
Total						100.00	388.8	171.5	100.000



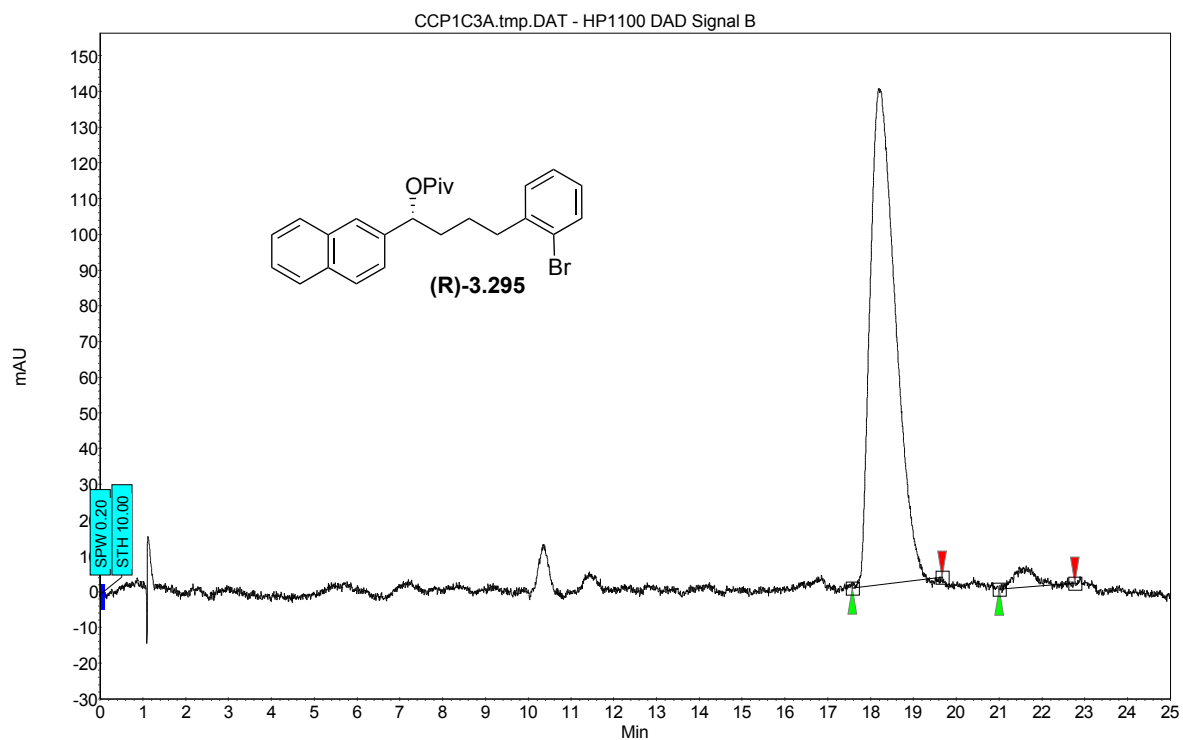
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μV]	[μV.Min]	[%]
1	UNKNOWN	12.25	12.74	13.59	0.00	97.90	2143.7	1166.3	97.901
2	UNKNOWN	14.45	14.90	15.62	0.00	2.10	51.7	25.0	2.099
Total						100.00	2195.5	1191.3	100.000

Method Name: MOK-II-266rac
Run Name: LEH-6-080-D3

Date: 11/24/2015
Time: 7:09:09 PM



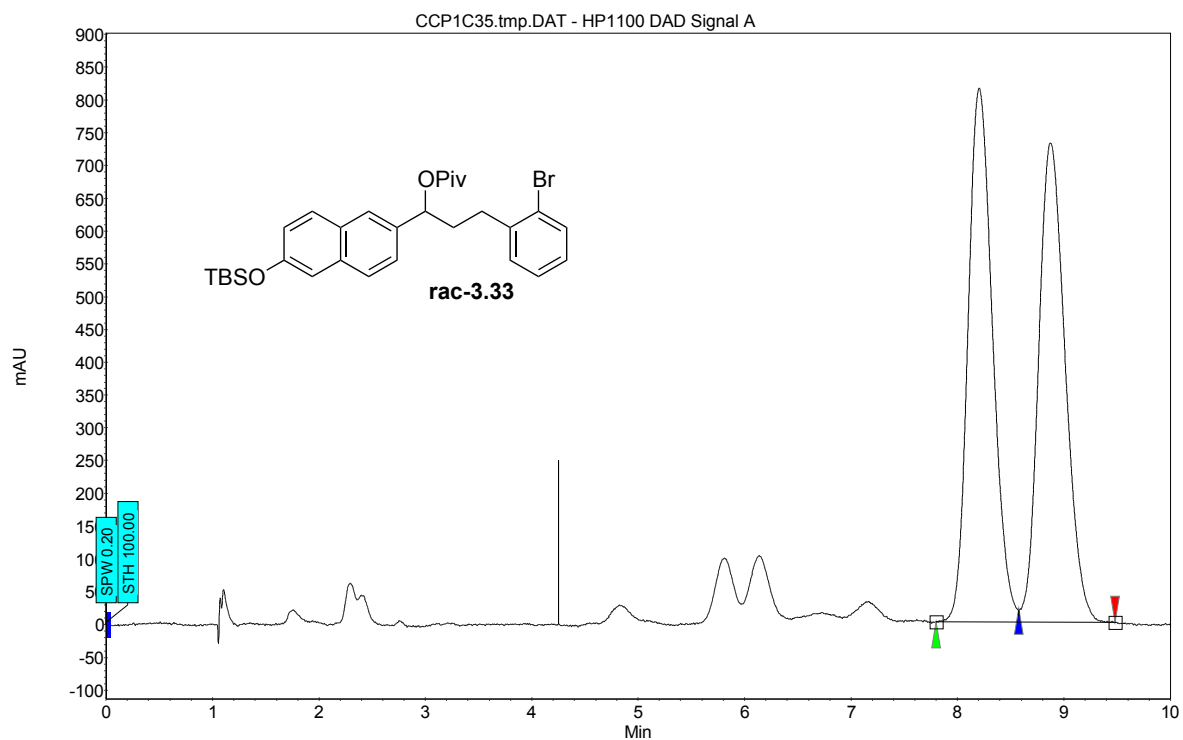
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	17.99	18.53	19.68	0.00	49.85	678.1	395.5	49.846
2	UNKNOWN	20.86	21.48	22.75	0.00	50.15	564.1	397.9	50.154
Total						100.00	1242.2	793.3	100.000



Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	17.57	18.20	19.67	0.00	96.84	138.8	94.2	96.842
2	UNKNOWN	21.01	21.63	22.76	0.00	3.16	5.9	3.1	3.158
Total						100.00	144.7	97.3	100.000

Method Name: MOK-IV-5otbspiv
 Run Name: MOK-IV-racopiv5tbs3

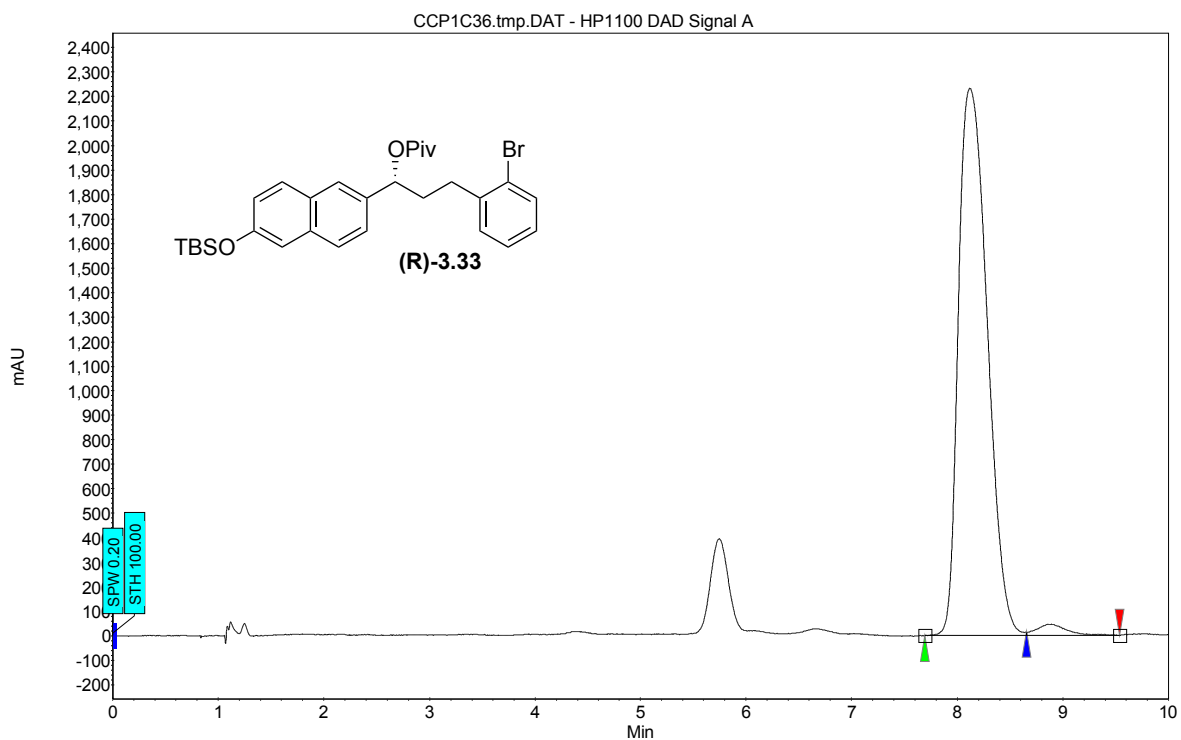
Date: 11/24/2015
 Time: 7:00:17 PM



Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	7.80	8.20	8.58	0.00	50.65	813.6	216.6	50.651
2	UNKNOWN	8.58	8.87	9.48	0.00	49.35	730.6	211.1	49.349
Total						100.00	1544.2	427.7	100.000

Method Name:MOK-IV-5otbspiv
 Run Name:MOK-IV-eeopiv5tbs2

Date:11/24/2015
 Time:7:01:31 PM

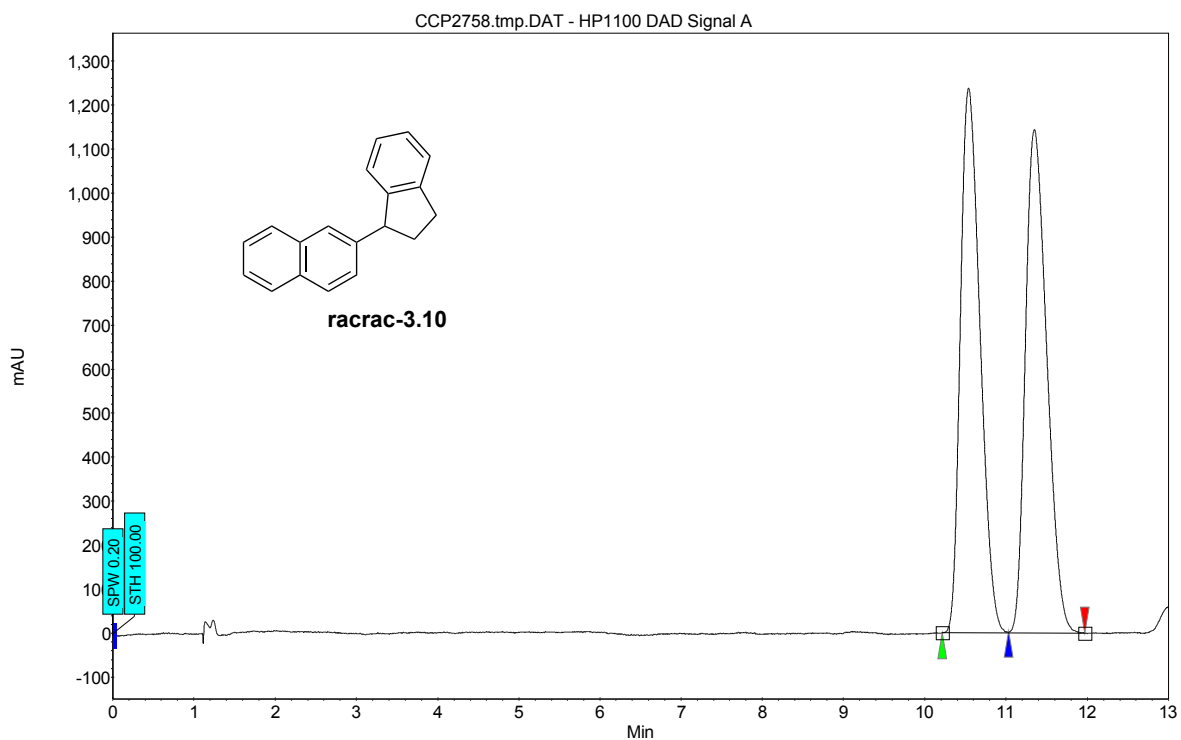


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	7.69	8.12	8.65	0.00	97.92	2231.6	705.5	97.918
2	UNKNOWN	8.65	8.86	9.54	0.00	2.08	45.6	15.0	2.082
Total						100.00	2277.2	720.5	100.000

S121

Method Name:MOK-II-266rac
Run Name:MOK-2-266rac3

Date:9/7/2015
Time:3:04:42 PM

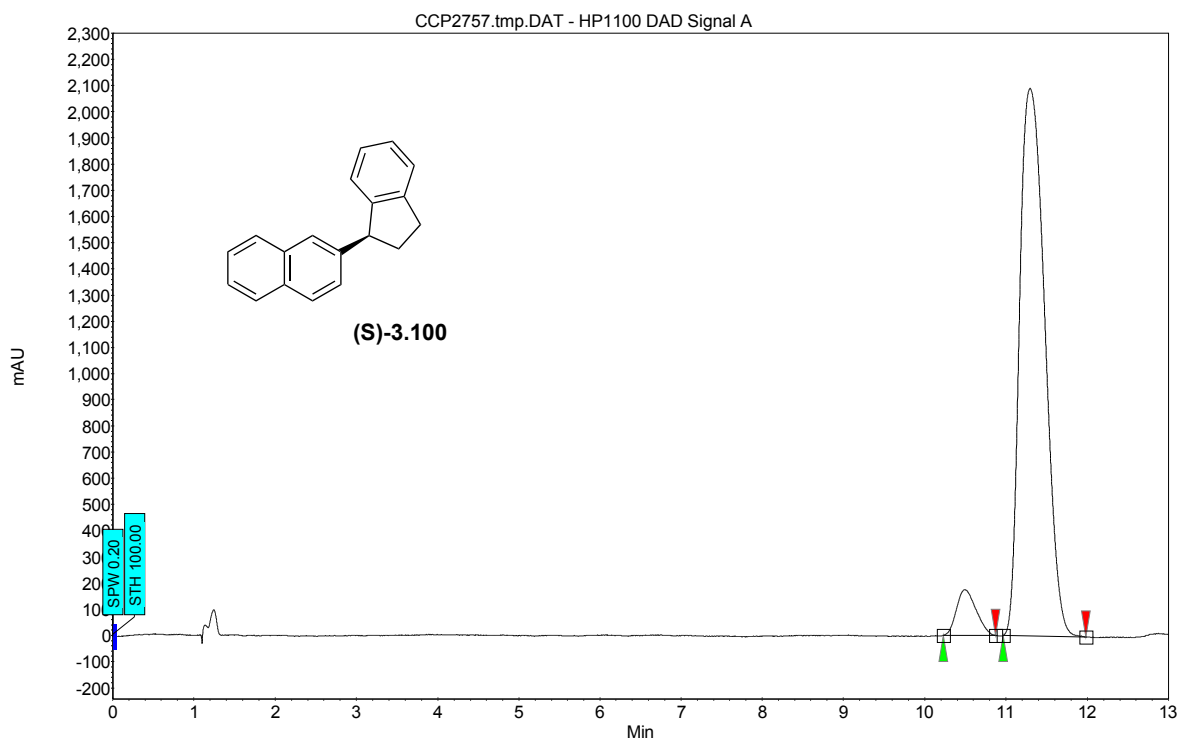


Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	10.21	10.54	11.04	0.00	49.76	1237.1	347.4	49.764
2	UNKNOWN	11.04	11.35	11.97	0.00	50.24	1143.6	350.7	50.236
Total						100.00	2380.8	698.2	100.000

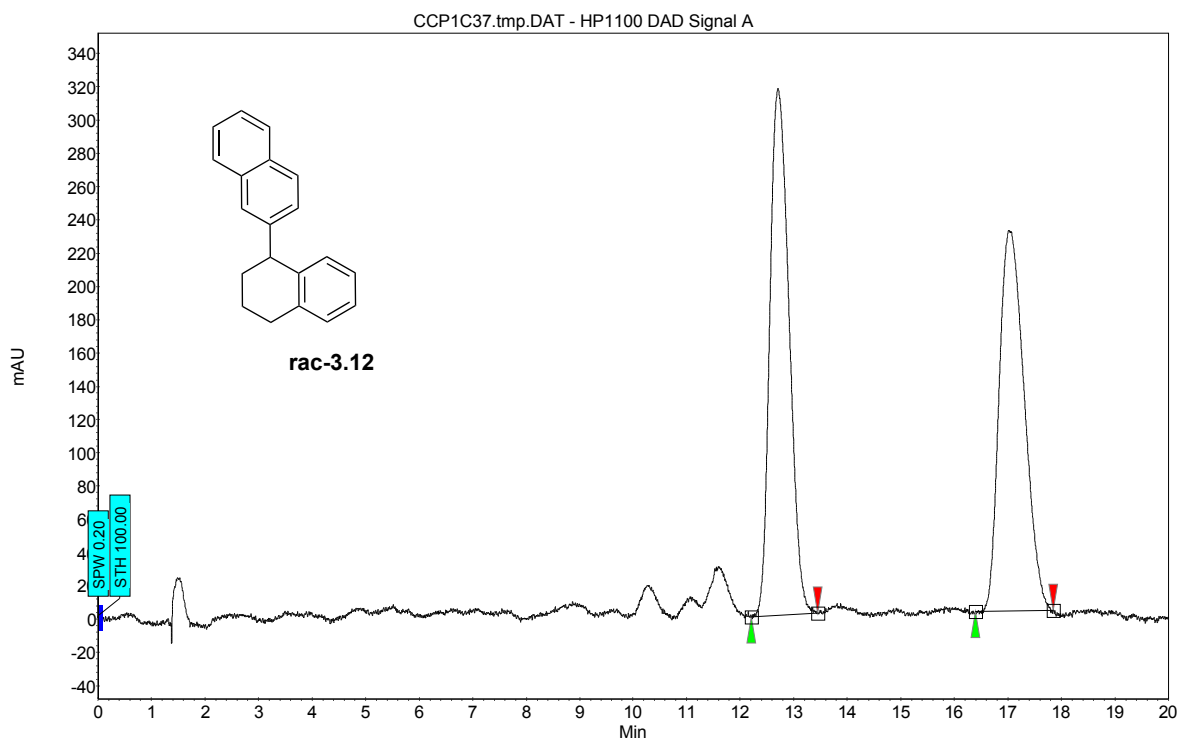
S122

Method Name: MOK-II-266rac
 Run Name: LEH-5-151-pub3

Date: 9/7/2015
 Time: 3:03:27 PM



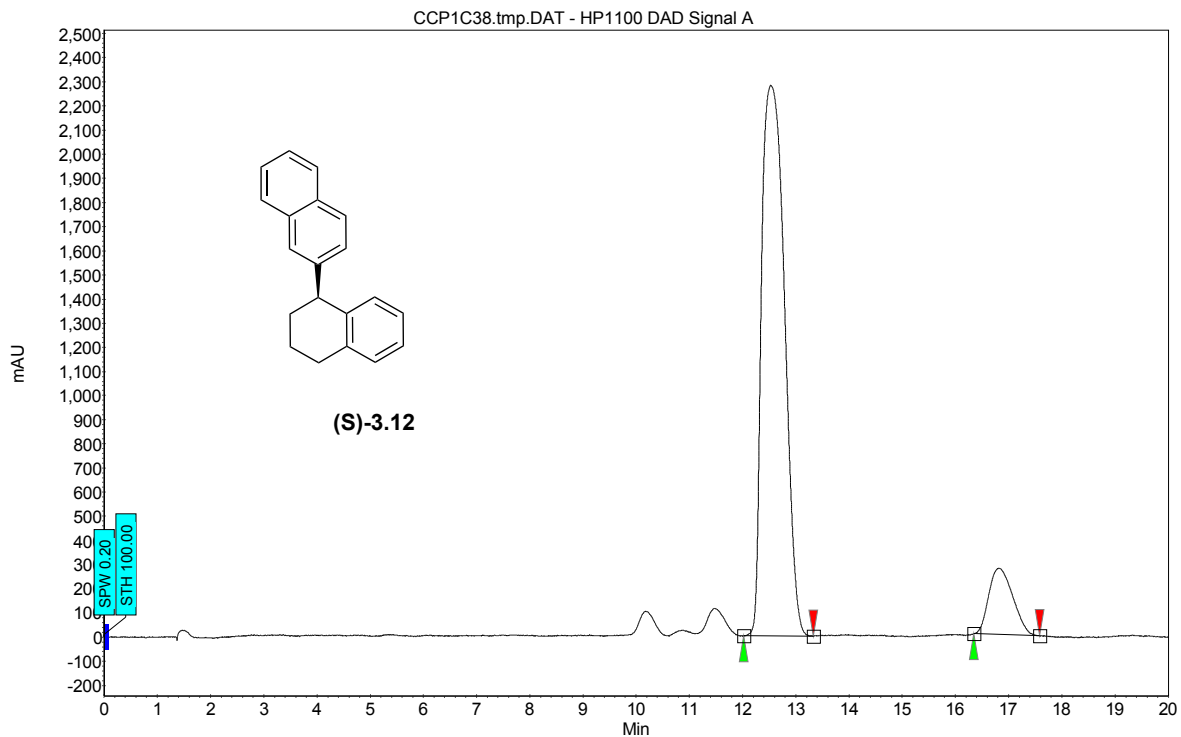
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
2	UNKNOWN	10.23	10.50	10.87	0.00	6.12	174.1	48.8	6.121
1	UNKNOWN	10.96	11.30	11.98	0.00	93.88	2091.5	747.8	93.879
Total						100.00	2265.6	796.6	100.000



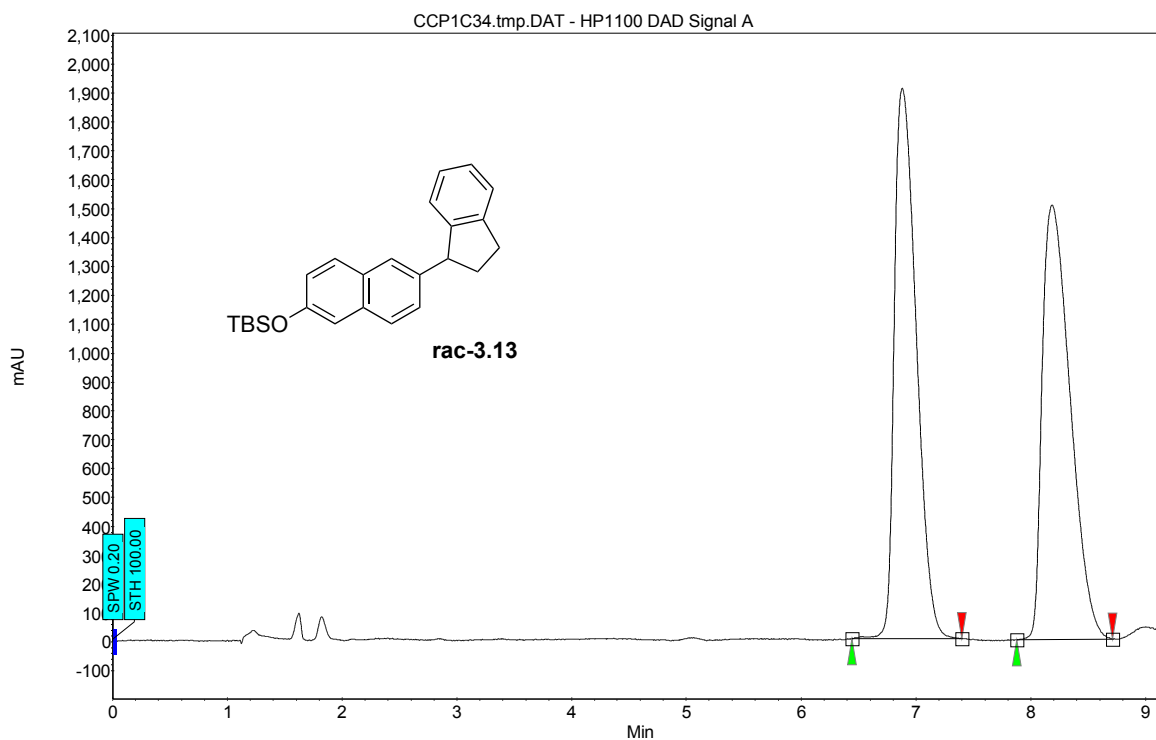
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
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1	UNKNOWN	12.21	12.71	13.45	0.00	51.53	316.7	131.5	51.529
2	UNKNOWN	16.40	17.02	17.84	0.00	48.47	228.9	123.7	48.471
Total						100.00	545.7	255.3	100.000

Method Name: MOK-266-NAP-6-Pdt
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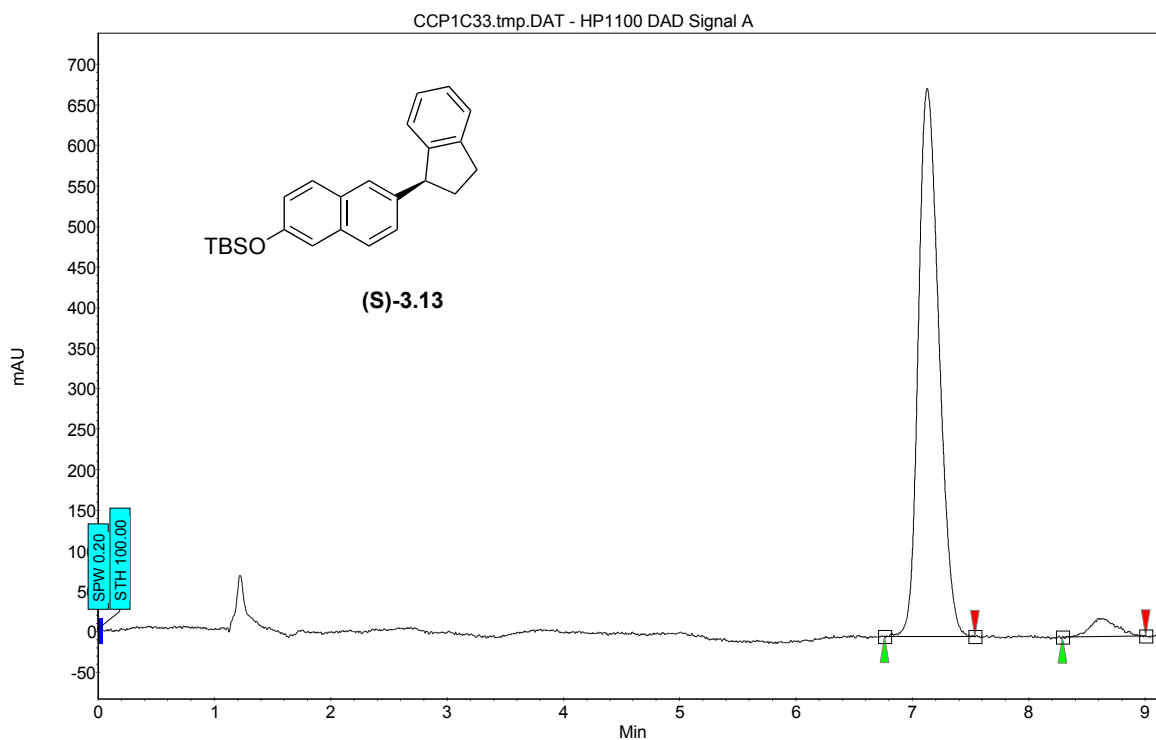
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1	UNKNOWN	12.02	12.53	13.33	0.00	88.75	2280.2	1130.7	88.750
2	UNKNOWN	16.35	16.81	17.57	0.00	11.25	273.6	143.3	11.250
Total						100.00	2553.9	1274.1	100.000



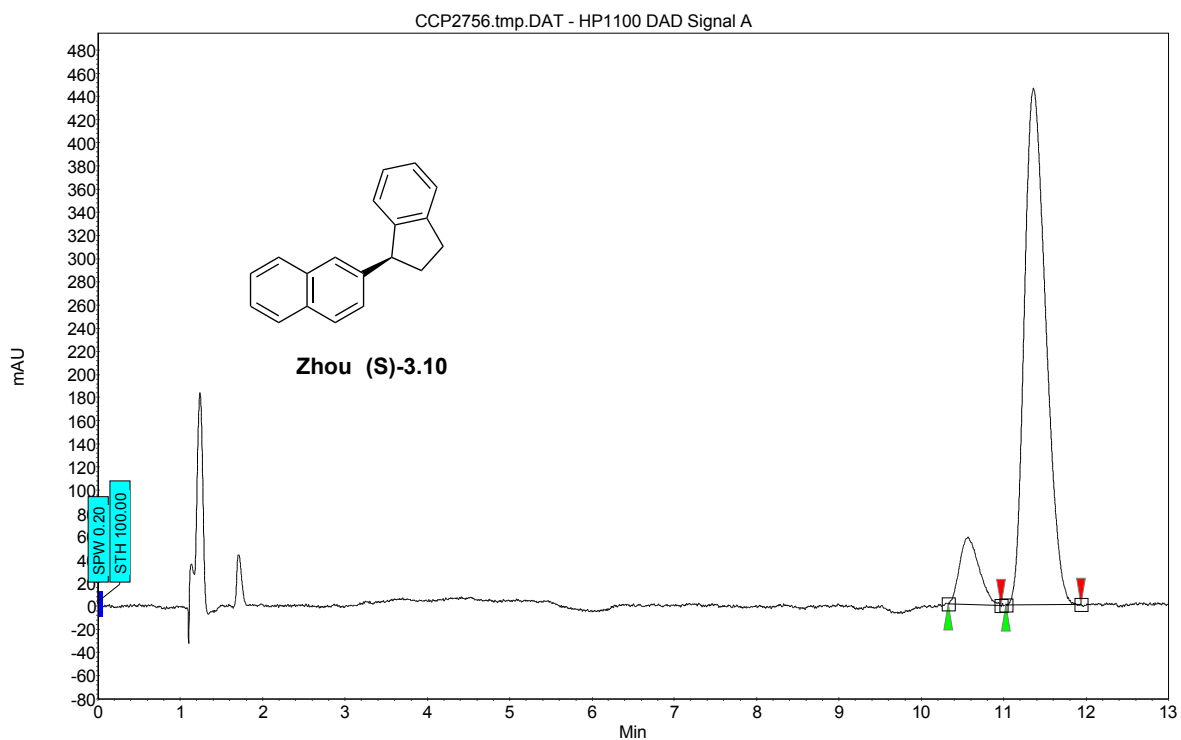
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		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	6.44	6.88	7.40	0.00	49.85	1904.4	421.7	49.849
2	UNKNOWN	7.88	8.19	8.71	0.00	50.15	1503.7	424.3	50.151
Total						100.00	3408.1	846.0	100.000



Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	6.76	7.13	7.53	0.00	95.85	676.0	138.3	95.849
2	UNKNOWN	8.29	8.61	9.00	0.00	4.15	22.6	6.0	4.151
Total						100.00	698.6	144.3	100.000

Method Name: MOK-II-266rac
 Run Name: LEH-5-036-realeezhou3

Date: 9/7/2015
 Time: 2:49:52 PM



Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
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1	UNKNOWN	10.33	10.57	10.96	0.00	10.54	58.2	15.8	10.536
2	UNKNOWN	11.03	11.36	11.93	0.00	89.46	446.0	134.4	89.464
Total						100.00	504.2	150.3	100.000

*Nickel-Catalyzed Generation of 2-PyridylZinc Reagents:**Applications to Deoxygenation and Intramolecular Conjugate Additions***4.1 Introduction**

Functionalized organozinc reagents are critical for the construction of natural products and other complex organic molecules.^{1,2} Their tendency to undergo selective reactions with transition metals enables the construction of new carbon–carbon bonds under mild reaction conditions. Methods to prepare functionalized organozincs are diverse and are an important topic of continuing research.³ However, the preparation of benzylzinc reagents present particular challenges. While benzylzinc reagents can be prepared by insertion of elemental zinc with primary benzylic halides (Scheme 4.1a), secondary benzylic halides frequently provide low yields due to formation of stable radicals that undergo competitive Wurtz-type coupling. Exchange of alkylboron reagents with a zinc (II) species provides an alternative method for the preparation of secondary benzylzinc reagents, but relies on synthesis of the requisite alkylboron reagent (Scheme 4.1b).⁴ Fragmentation of bulky homobenzylic zinc alkoxides is a creative approach that circumvents formation of benzylic radicals (Scheme 4.1c).⁵

¹ (a) G.S. Silverman, in: P.E. Rakita (Ed.), *Handbook of Grignard reagents*, Marcel Dekker, New York, 1996. (b) Knochel, P.; Leuser, H.; Gong, L.-Z.; Perrone, S.; Kneisel, F. F. *Handbook of Functionalized Organometallics: Applications in Synthesis*; Knochel, P., Ed.; Wiley: Weinheim, 2005; p 251.

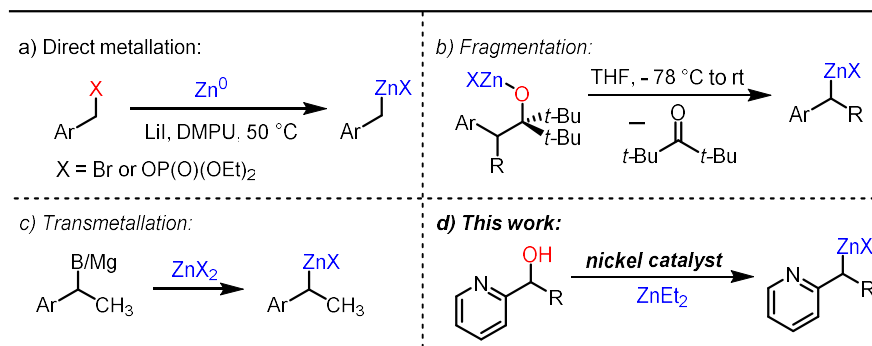
² (a) Knochel, P.; Leuser, H.; Gong, L.-Z.; Perrone, S.; Kneisel, F. F. *Polyfunctional Zinc Organometallics for Organic Synthesis*. In *Handbook of Functionalized Organometallics: Applications in Synthesis*; Knochel, P., Ed.; Wiley: Weinheim, 2005; p 251. For specific examples in total synthesis, see: (b) Fürstner, A.; Weintritt, H. *J. Am. Chem. Soc.* **1998**, *120*, 2817. (c) Masaki, H.; Maeyama, J.; Kamada, K.; Esumi, T.; Iwabuchi, Y.; Hatakeyama, S. *J. Am. Chem. Soc.* **2000**, *122*, 5216. (d) Aoyagi, S.; Hirashima, S.; Saito, K.; Kibayashi, C. *J. Org. Chem.* **2002**, *67*, 5517.

³ (a) Klatt, T.; Markiewicz, J. T.; Sämann, C.; Knochel, P. *J. Org. Chem.* **2014**, *79*, 4253. (b) Stathakis, C. I.; Manolikakes, S. M.; Knochel, P. *Org. Lett.* **2013**, *15*, 1302. (c) Colombe, J. R.; Bernhardt, S.; Stathakis, C.; Buchwald, S. L.; Knochel, P. **2013**, *15*, 5754. (d) Quinio, P.; François, C.; Cuesta, A. E.; Steib, A. K.; Achraimer, F.; Zipse, H.; Karaghiosoff, K.; Knochel, P. *Org. Lett.* **2015**, *17*, 1010.

⁴ (a) Langer, F.; Knochel, P. *Tetrahedron Lett.* **1995**, *36*, 4591. (b) Hupe, E.; Calaza, M. I.; Knochel, P. *J. Organomet. Chem.* **2003**, *680*, 136.

⁵ Piazza, C.; Millot, N.; Knochel, P. *J. Organomet. Chem.* **2001**, *624*, 88.

Scheme 4.1 Summary of methods for preparation of benzylic zinc reagents



Transition-metal catalysis offers alternative strategies for formation of organometallic reagents.^{6,7,8} Formation of benzylnickel complexes can be accomplished by oxidative addition of nickel catalysts with benzylic electrophiles.⁹ We reasoned that, with catalytic quantities of a nickel catalyst and in the presence of a super-stoichiometric organozinc reagent such as diethylzinc, transmetalation would generate the desired organozinc reagent. The Shi group has reported a related synthesis of primary benzylic Grignard reagents from benzylic alcohols.¹⁰ In this chapter, we report nickel-catalyzed formation of secondary benzylic zinc reagents from readily accessible 2-pyridylcarbinols (Scheme 4.1d). Activation of the alcohol is achieved using diethylchlorophosphate;¹¹ quenching with methanol provides the products of deoxygenation in a one-pot reaction. We propose that the pyridyl substituent serves as a directing group and stabilizes

⁶ (a) Ishiyama, T.; Murata, M.; Miyaura, N. *J. Org. Chem.* **1995**, *60*, 7508. (b) Rosen, B. M.; Huang, C.; Percec, V. *Org. Lett.* **2008**, *10*, 2597. (c) Ishiyama, T.; Takagi, J.; Ishida, K.; Miyaura, N.; Anastasi, N. R.; Hartwig, J. F. *J. Am. Chem. Soc.* **2002**, *124*, 390.

⁷ Gao, F.; Hoveyda, A. H. *J. Am. Chem. Soc.* **2010**, *132*, 10961.

⁸ Dao, H. T.; Baran, P. S. *Angew. Chem. Int. Ed.* **2014**, *53*, 14382.

⁹ (a) Wisniewska, H. M.; Swift, E. C.; Jarvo, E. R. *J. Am. Chem. Soc.* **2013**, *135*, 9083. (b) Do, H-Q.; Chandrashekar, E. R. R.; Fu, G. C. *J. Am. Chem. Soc.* **2013**, *135*, 16288

¹⁰ Yu, D-G.; Wang, X.; Zhu, R-Y.; Luo, S.; Zhang, X-B.; Wang, B-Q.; Wang, L.; Shi, Z-J. *J. Am. Chem. Soc.* **2012**, *134*, 14638.

¹¹ Phosphate esters generated in situ are effective electrophiles in nickel-catalyzed transformations, see: (a) Ackerman, L. K. G.; Anka-Lufford, L. L.; Naodovic, M.; Weix, D. J. *Chem. Sci.* **2015**, *6*, 1115.

the benzylzinc reagent.^{12,13} This method provides reduction of hydroxymethylpyridines with incorporation of deuterium in the benzylic position, which can be useful probing metabolic pathways.

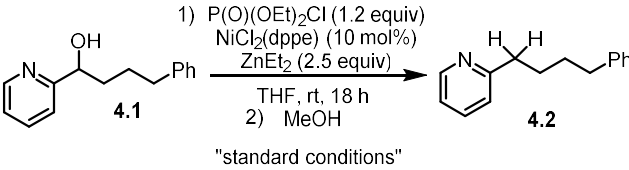
4.2 Development of Nickel-Catalyzed Formation of 2-Pyridyl Benzylzinc Reagents

We examined 2-pyridyl carbinol **4.1** as our model substrate. Reactions were performed employing diethylchlorophosphate to activate the alcohol, a series of nickel catalysts, and diethylzinc as transmetallating agent. We found that at room temperature, Ni(dppe)Cl₂ furnishes the desired deoxygenated product **4.2** in high yield (Table 4.1, entry1). The data in Table 4.1 illustrate how changes in the reaction parameters affect the transformation. When the nickel catalyst or diethylchlorophosphate are omitted, the reaction does not afford desired product (entries 2–4). These results are consistent with a mechanism of benzylic alcohol activation prior to reaction with a nickel catalyst. The use of a non-polar solvent or ZnMe₂ instead of ZnEt₂ does not significantly affect the efficiency of the reaction (entries 5 and 6). A decreased yield of **4.2** was observed when catalysts ligated by other bidentate or monodentate phosphine ligands were used in the reaction (entries 7-10). Under optimized conditions, the reaction scales well, providing desired product in high yield when run on gram scale (entry 11).

¹² General pyridyl directing groups: (a) Reviews on removable pyridine directing groups (a) Itami, K.; Mitsudo, K.; Nokami, T.; Kamei, T.; Koike, T.; Yoshida, J-I. *J. Organomet. Chem.* **2002**, *653*, 105. (b) Itami, K.; Yoshida, J-I *Synlett.* **2006**, *2*, 157. Directed reactions by Chatani/Murai (a) Chatani, N.; Inoue, S.; Yokata, K.; Tatamidani, H.; Fukumoto, Y. *Pure. Appl. Chem.* **2010**, *82*, 1443. (b) Chatani, N.; Ie, Y.; Kakiuchi, F.; Murai, S. *J. Am. Chem. Soc.* **1999**, *121*, 8645. (c) Tatamidani, H.; Yokata, K.; Kakiuchi, F.; Chatani, N. *J. Org. Chem.* **2004**, *69*, 5615. More recent publications: (a) Wang, J.; Chen, W.; Zuo, S.; Liu, L.; Zhang, X.; Wang, J. *Angew. Chem., Int. Ed.* **2012**, *51*, 12334. (b) Lei, Z-Q.; Pan, F.; Li, H.; Li, Y.; Zhang, X-S.; Chen, K.; Wang, X.; Li, Y-X.; Sun, J.; Shi, Z-J. *J. Am. Chem. Soc.* **2015**, *137*, 5012. Pyridine is a privileged directing group in C–H bond activation, see: (a) Suggs, J. W. *J. Am. Chem. Soc.* **1979**, *101*, 489. (b) Lim, Y-G.; Kang, J-B.; Kim, Y. H. *J. Chem. Soc., Perkin Trans.* **1996**, *1*, 2201. (c) Jun, C-H.; Hwang, D-C.; Na, S-J. *Chem. Commun.* **1998**, 1405. (d) Chatani, N.; Ishii, Y.; Ie, Y.; Kakiuchi F.; Murai, S. *J. Org. Chem.* **1998**, *63*, 5129. (e) Chatani, N.; Asaumi, T.; Ikeda, T.; Yorimitsu, S.; Ishii, Y.; Kakiuchi, F.; Murai, S. *J. Am. Chem. Soc.* **2000**, *122*, 12882.

¹³ Bioactive references: (a) Shi, N.; Lu, C.; Ho, C. C.; Shen, Y. *Rec. Nat. Prod.* **2013**, *7*, 1. (b) Fürstner, A.; Leitner, A.; Méndez, M.; Krause, H. *J. Am. Chem. Soc.* **2002**, *124*, 13856. (c) Kubota, T.; Ishiguro Y.; Yamamoto, S.; Fromont, J.; Kobayashi, J. *Heterocycles* **2010**, *80*, 1407. (d) Fischer, D. F.; Sarpong, R. *J. Am. Chem. Soc.* **2010**, *132*, 5926. Actos (Takeda). Ligand references: (a) Chelucci, G.; Thummel, R. P. *Chem. Rev.* **2002**, *102*, 3129.

Table 4.1. Effect of Reaction Parameters on the Efficiency of Deoxygenation of **4.1**

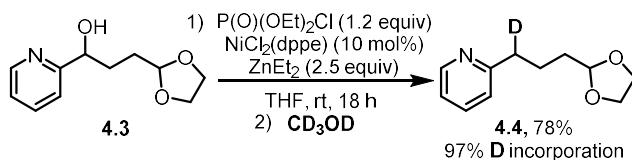


entry	change from standard conditions	yield (%) ^a
1	none	77
2	dppe instead of NiCl ₂ (dppe)	< 5
3	no NiCl ₂ (dppe)	< 5
4	no P(O)(OEt) ₂ Cl	< 5
5	PhMe instead of THF	72
6	ZnMe ₂ instead of ZnEt ₂	74
7	NiCl ₂ ·DME / DPEphos instead of NiCl ₂ (dppe)	42
8	NiCl ₂ ·DME / dpfp instead of NiCl ₂ (dppe)	55
9	NiCl ₂ ·DME / PCy ₃ instead of NiCl ₂ (dppe)	21
10	NiCl ₂ ·DME / PPh ₃ instead of NiCl ₂ (dppe)	50
11	Run on 1.0 g scale of 1	72 ^b

4.3 Exploration of Scope of Deoxygenation of 2-pyridyl Alcohols

Having developed robust conditions for the deoxygenation of **4.1**, we turned our attention to investigating the source of “H” found in the product. We proposed that **4.2** is formed from the methanol quench of the benzylzinc reagent generated in situ. To test our hypothesis, we performed an isotopic labeling study with methanol-d₄. Upon quenching the deoxygenation reaction of **4.3** with CD₃OD, we observed high yield and high deuterium incorporation of the corresponding product **4.4** (eq 4.1). The result of the isotopic labeling experiment provides strong support for our proposed hypothesis that the reaction proceeds by formation of a benzylzinc reagent.

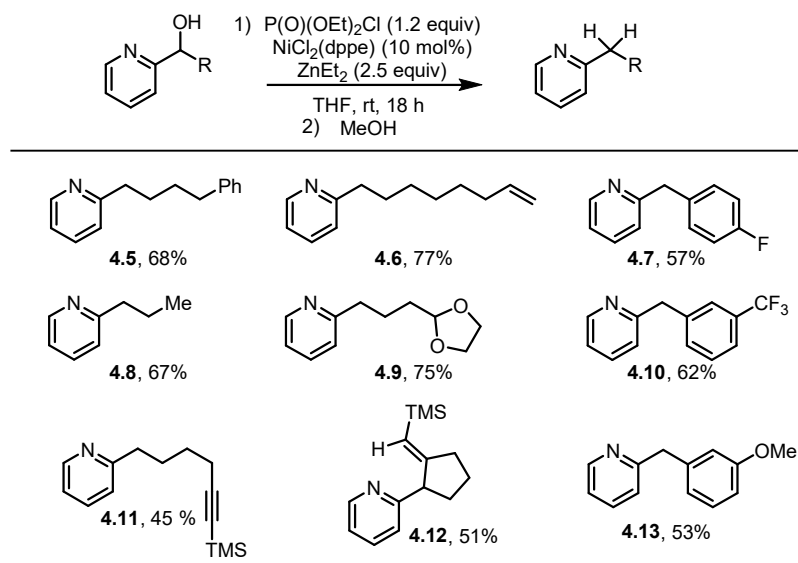
Equation 4.1. Deuterium incorporation via quenching with deuteromethanol



Next, we examined the formation of alkylzinc reagents from a range of benzylic alcohols (Table 4.2). Secondary benzylic alcohols bearing alkyl chains are well tolerated under the reaction conditions (entries **4.5**, and **4.8**). Substrates containing acetal and alkene functionalities provide

deoxygenated products in good yield (**4.6** and **4.11**). The high yield of **4.6** demonstrates excellent orthogonality to hydroboration transmetalation procedures often used to prepare secondary benzylic zinc reagents.¹⁴ Of particular note, is the formation of byproduct **4.12**, isolated from reactions desired to produce only **4.11**. We hypothesize this byproduct is the result of a metalloene reaction resulting from the proposed benzylic reagent which has been previously described.¹⁵

Table 4.2 Scope of Deoxygenation



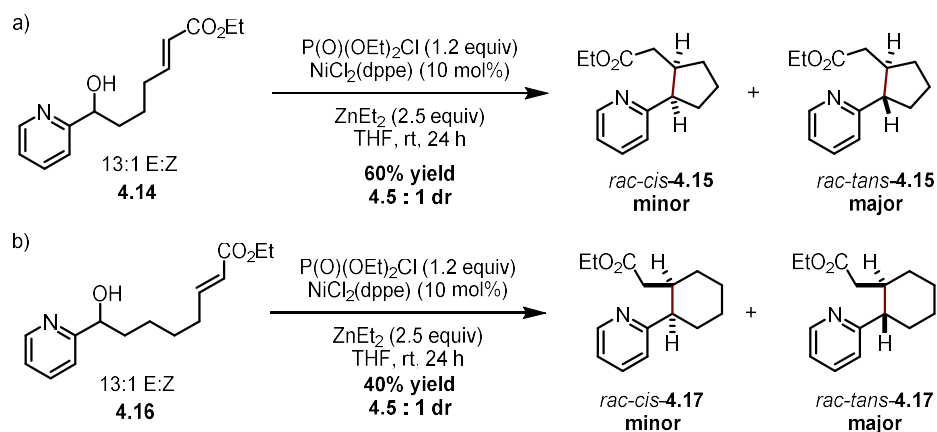
A mechanism consistent with the formation of the proposed benzylic zinc reagents is outlined in Figure 1. The active catalyst is formed by reduction of $\text{Ni}(\text{dppe})\text{Cl}_2$ with ZnEt_2 to give the reduced $\text{Ni}(0)(\text{dppe})$ species. The $\text{Ni}(0)$ catalyst then undergoes oxidative addition into the phosphorylated alcohol to form a benzylic $\text{Ni}(\text{II})$ intermediate. Transmetalation results in formation of the benzylic zinc reagent and an ethylnickel complex. β -Hydride elimination followed by loss of ethylene yields a nickel hydride intermediate. This intermediate undergoes a

¹⁴ Hupe, E.; Calaza, M. I.; Knochel, P. *J. Organomet. Chem.* **2003**, *680*, 136.

¹⁵ Meyer, C.; Marek, I.; Courtemanche, G.; Normant, J.-F. *J. Org. Chem.* **1995**, *60*, 863.

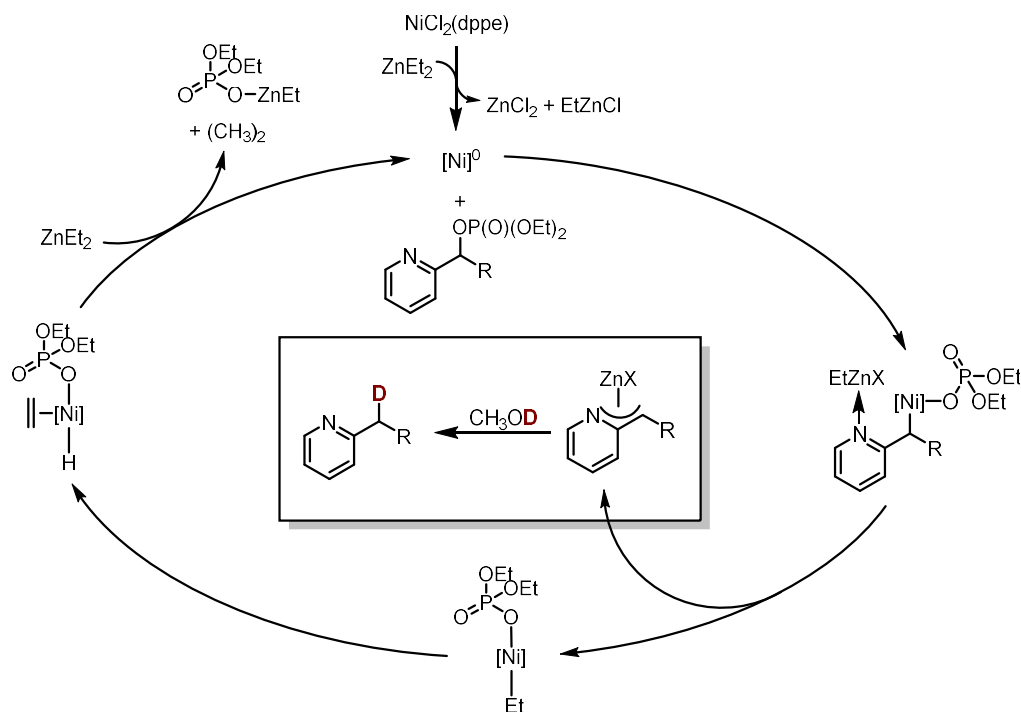
final transmetalation of an ethyl group from zinc followed by reductive elimination to form ethane and regenerate the Ni(0) catalyst.

Scheme 4.2. Intramolecular 1,4-addition



To further demonstrate the utility of this methodology, we coupled the formation of a benzylic zinc reagent to an intramolecular conjugate addition (Scheme 4.2). We anticipated that upon forming the corresponding benzylic zinc reagent from **4.14** or **4.16**, cyclization should occur to afford cyclopentane **4.15** or cyclohexane **4.17** (Scheme 4.2 a and b respectively). 2-Pyridyl carbinol **4.14** and **4.16** bearing a pendant α,β -unsaturated ester underwent smooth cyclization to produce the desired product in modest yield and 4.5:1 diastereomeric ratio. Upon investigation of smaller ring sized such as 3-membered rings yielded no cyclization product, this was also the case when we attempted to produce 7-membered rings using this methodology.

Figure 4.1. Proposed Mechanism for formation of 2-pyridyl zinc reagent



4.4 Conclusion

In summary, we have developed a concise route to secondary organozinc reagents directly from benzylic alcohols. We have successfully applied this methodology to the formal hydrogenolysis of a range of 2-pyridyl carbinols. The reaction proceeds in high yield and with straightforward incorporation of deuterium from deuteromethanol. The reaction has been applied to an intramolecular 1,4-addition. Further exploration of the reactivity of these compounds and elucidation of mechanistic details are underway.

4.5 Experimental Details

GENERAL PROCEDURES

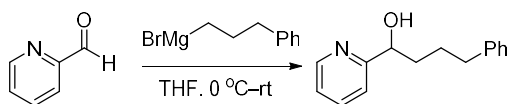
All reactions were carried out under an atmosphere of N₂, or Ar when noted. All glassware was oven- or flame-dried prior to use. Tetrahydrofuran (THF), diethyl ether (Et₂O), dichloromethane (CH₂Cl₂), and toluene (PhMe) were degassed with Ar and then passed through two 4 x 36 inch columns of anhydrous neutral A-2 alumina (8 x 14 mesh; LaRoche Chemicals; activated under a flow of argon at 350 °C for 12 h) to remove H₂O. All other solvents utilized were purchased “anhydrous” commercially, or purified as described. ¹H NMR spectra were recorded on Bruker DRX-400 (400 MHz ¹H, 100 MHz ¹³C, 376.5 MHz ¹⁹F), GN-500 (500 MHz ¹H, 125.7 MHz ¹³C), or CRYO-500 (500 MHz ¹H, 125.7 MHz ¹³C) spectrometers. Proton chemical shifts are reported in ppm (δ) relative to internal tetramethylsilane (TMS, δ 0.00). Data are reported as follows: chemical shift (multiplicity [singlet (s), broad singlet (br s), doublet (d), doublet of doublets (dd), triplet (t), doublet of triplets (dt), doublet of doublet of triplets (ddt), triplet of triplets (tt), quartet (q), quintet (quin), apparent doublet (ad), apparent triplet (at), multiplet (m)], coupling constants [Hz], integration). Carbon chemical shifts are reported in ppm (δ) relative to TMS with the respective solvent resonance as the internal standard (CDCl₃, δ 77.16 ppm). Unless otherwise indicated, NMR data were collected at 25 °C. Infrared (IR) spectra were obtained on a Thermo Scientific Nicolet iS5 spectrometer with an iD5 ATR tip (neat) and are reported in terms of frequency of absorption (cm⁻¹). Analytical thin-layer chromatography (TLC) was performed using Silica Gel 60 F₂₅₄ precoated plates (0.25 mm thickness). Visualization was accomplished by irradiation with a UV lamp and/or staining with KMnO₄, ceric ammonium molybdate (CAM), or *p*-anisaldehyde (PAA) solutions. Flash chromatography was performed using Silica Gel 60 (170-400 mesh) from Fisher Scientific. Melting points (m.p.) were obtained using a Mel-Temp melting

point apparatus and are uncorrected. High resolution mass spectrometry was performed by the University of California, Irvine Mass Spectrometry Center. [1,2-Bis(diphenylphosphino)ethane]dichloronickel(II) was purchased from Strem, stored in a glovebox under an atmosphere of N₂, and used as received. Diethylzinc (ZnEt₂) and diethyl chlorophosphate were purchased from Sigma and used as received. 1-isoquinolinecarboxaldehyde was prepared from selenium (IV) oxide oxidation of 1-methylisoquinoline by a procedure reported by Long.ⁱ All other reagents were purchased commercially and used as received.

SYNTHESIS AND CHARACTERIZATION OF ALL SUBSTRATES

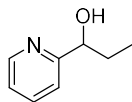
Synthesis of Benzylic Alcohols

General Procedure A. Grignard addition to aldehydes.

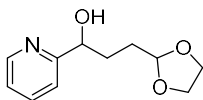


4.18. In a flame-dried round-bottom flask, to a solution of 2-pyridinecarboxaldehyde (1.43 mL, 15.0 mmol, 1.00 equiv) in THF (30 mL) was added at 0 °C (3-phenylpropyl)magnesium bromide (1.6 M in THF, 10 mL, 16 mmol, 1.1 equiv). After stirring at room temperature for 1 h, saturated ammonium chloride (25 mL) was added at 0 °C and the reaction was extracted with EtOAc (3 x 25 mL). The combined organic layers were washed with brine (1 x 40 mL), dried over Na₂SO₄, and concentrated in vacuo. The product was purified by column flash chromatography (25–40% EtOAc/hexanes) to afford the title compound as a yellow solid (1.70 g, 7.50 mmol, 50%). **TLC R_f** = 0.16 (20% EtOAc/hexanes, UV active); **m.p.** = 58–60 °C; **¹H NMR** (500 MHz, CDCl₃) δ 8.53 (d, *J* = 4.8 Hz, 1H), 7.66 (td, *J* = 7.8, 1.8 Hz, 1H), 7.29–7.23 (m, 2H), 7.22–7.13 (m, 5H), 4.75 (s, 1H), 4.19 (s, 1H), 2.71–2.59 (m, 2H), 1.91–1.82 (m, 1H), 1.80–1.67 (m, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 161.9, 148.2, 142.4, 136.7, 128.5, 128.3, 125.8, 122.3, 120.4, 72.5, 38.2, 35.8, 26.9; **IR**

(neat) 3192, 2925, 2861, 1594, 1492, 1452 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{15}\text{H}_{17}\text{NONa}$ ($\text{M} + \text{Na}$)⁺ 227.1310, found 227.1315.



4.19 Prepared according to a procedure reported by Braga¹⁶, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.38 mL, 4.00 mmol, 1.00 equiv), ethylmagnesium bromide (2.4 M in Et_2O , 2.0 mL, 4.8 mmol, 1.2 equiv), and THF (15 mL). Analytical data is consistent with literature values.¹⁷ **¹H NMR** (400 MHz, CDCl_3) δ 8.55 (dt, $J = 4.9, 1.3$ Hz, 1H), 7.68 (td, $J = 7.6, 1.8$ Hz, 1H), 7.25 (dd, $J = 7.8$ Hz, 0.5 Hz, 1H), 7.20 (ddd, $J = 7.6, 4.9, 0.5$ Hz, 1H), 4.73–4.66 (m, 1H), 4.17 (d, $J = 5.5$ Hz, 1H), 1.95–1.84 (m,



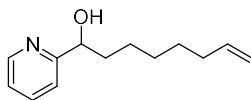
4.20 Using representative procedure A outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.95 mL, 10 mmol, 1.0 equiv), hept-6-en-1-ylmagnesium bromide (1.2 M in THF, 9.2 mL, 11 mmol, 1.1 equiv), and THF (10 mL). The product was purified by column flash chromatography (25–40% EtOAc/hexanes) to afford the title compound as a yellow oil (1.31 g, 6.38 mmol, 64%). Analytical data is consistent with literature values.¹⁸ **TLC** $R_f = 0.1$ (50% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl_3) δ 8.55–8.50 (m, 1H), 7.67 (td, $J = 7.8, 1.8$ Hz, 1H), 7.29 (d, $J = 7.8$ Hz, 1H), 7.21–7.15 (m, 1H), 4.93–4.88 (m, 1H), 4.83–4.75 (m, 1H), 4.33 (d, $J = 5.4$ Hz, 1H), 3.99–3.80 (m, 4H), 2.07–1.95 (m, 1H), 1.87–1.74 (m,

¹⁶ Braga, A. L.; Paixao, M. W.; Ludtke, D. S.; Silveira, C. C.; Rodrigues, O. E. D. *Org. Lett.* **2003**, 5, 2365.

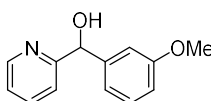
¹⁷ Moody, C. J.; Morfitt, C. N. *Synthesis* **1998**, 7, 1039.

¹⁸ Gebert, A.; Barth, M.; Linden, A.; Widmer, U.; Heimgartner, H. *Helv. Chim. Acta* **2012**, 95, 737.

3H); ^{13}C NMR (125 MHz, CDCl_3) δ 161.9, 148.3, 136.7, 122.3, 120.4, 104.4, 72.38, 64.9, 32.5, 29.5.

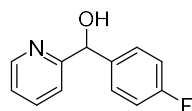


4.21 Using representative procedure A outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.95 mL, 10 mmol, 1.0 equiv), hept-6-en-1-ylmagnesium bromide (1.2 M in THF, 9.2 mL, 11 mmol, 1.1 equiv), and THF (10 mL). The product was purified by column flash chromatography (25–40% EtOAc/hexanes) to afford the title compound as a red oil (1.31 g, 6.38 mmol, 64%). **TLC** R_f = 0.4 (40% EtOAc/hexanes, UV active); ^1H NMR (500 MHz, CDCl_3) δ 8.53 (d, J = 4.7 Hz, 1H), 7.67 (t, J = 7.6 Hz, 1H), 7.26 (d, J = 7.6 Hz, 1H), 7.19 (t, J = 5.5 Hz, 1H), 5.79 (ddt, J = 17.2, 10.2, 6.9 Hz, 1H), 4.98 (d, J = 17.2 Hz, 1H), 4.92 (d, J = 10.2 Hz, 1H), 4.73 (dd, J = 7.7, 4.4 Hz, 1H), 4.25 (br s, 1H), 2.03 (dd, J = 10.2, 6.9 Hz, 2H), 1.87–1.77 (m, 1H) 1.74–1.65 (m, 1H), 1.50–1.24 (m, 6H); ^{13}C NMR (125 MHz, CDCl_3) δ 162.4, 148.3, 139.2, 136.7, 122.3, 120.4, 114.3, 72.9, 38.7, 33.8, 29.2, 28.9, 25.2; **IR** (neat) 3260, 2926, 1594 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{13}\text{H}_{19}\text{ONNa}$ ($\text{M} + \text{Na}$) $^+$ 228.1364, found 228.1364.

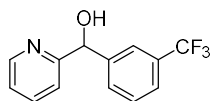


4.22 Using representative procedure A outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.95 mL, 10 mmol, 1.0 equiv) (3-methoxyphenyl)magnesium bromide (0.92 M in THF, 13 ml, 12 mmol, 1.2 equiv), and THF (20 mL). The product was purified by column flash chromatography (20–40% EtOAc/hexanes) to afford the title compound as a yellow oil (0.62 g, 3.1 mmol, 62%). Analytical data is consistent with literature values.⁶ **TLC** R_f

= 0.2 (20% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl₃) δ 8.56 (d, *J* = 4.4 Hz, 1H), 7.62 (td, *J* = 7.5, 1.3 Hz, 1H), 7.29–7.14 (m, 3H), 6.96 (d, *J* = 6.7 Hz, 1H), 6.93 (s, 1H), 6.81 (d, *J* = 7.2 Hz, 1H), 5.27 (d, *J* = 3.6 Hz, 1H), 5.29 (d, *J* = 4.3 Hz, 1H); **¹³C NMR** (125 MHz, CDCl₃) δ 160.6, 159.8, 147.8, 144.8, 136.9, 129.6, 122.5, 121.4, 119.5, 113.5, 112.5, 74.9, 55.3.



4.23 Using representative procedure A outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.36 mL, 3.8 mmol, 1.1 equiv) (4-fluorophenyl)magnesium bromide (0.27 M in THF, 15 ml, 4.0 mmol, 1.2 equiv), and THF (20 mL). The product was purified by column flash chromatography (20–40% EtOAc/hexanes) to afford the title compound as a yellow oil. (0.60 g, 3.0 mmol, 78%). Analytical data is consistent with literature values.¹⁹ **TLC** *R_f* = 0.2 (20% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl₃) δ 8.56 (d, *J* = 4.7 Hz, 1H), 7.62 (dt, *J* = 7.7, 1.8 Hz, 1H), 7.36–7.30 (m, 2H), 7.23–7.17 (m, 1H), 7.11 (d, *J* = 7.9 Hz, 1H), 7.05–6.97 (m, 2H), 5.73 (s, 1H), 5.31 (s, 1H); **¹³C NMR** (125 MHz, CDCl₃) δ 163.4, 161.5, 160.6, 147.9, 139.1 (d, *J_{CF}* = 2.77 Hz), 137.0, 128.9 (d, *J_{CF}* = 8.3 Hz), 122.6 (d, *J_{CF}* = 161.8 Hz), 115.6 (d, *J_{CF}* = 21.5 Hz), 74.31.

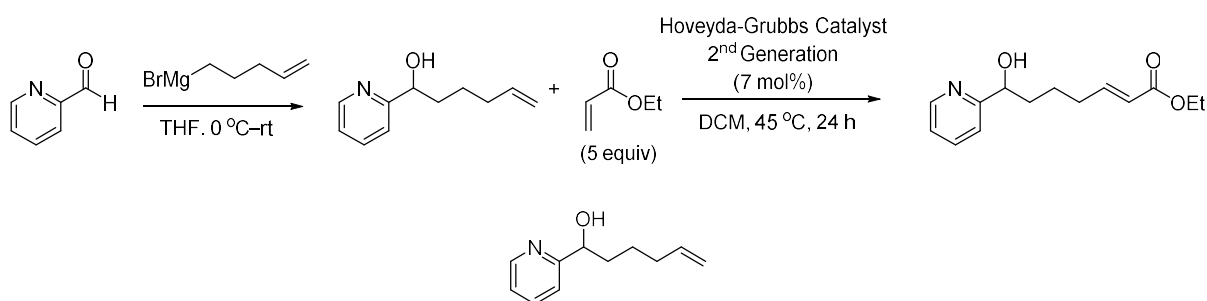


4.24 Using representative procedure A outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.95 mL, 10 mmol, 1.0 equiv) (3-(trifluoromethyl)phenyl)magnesium bromide (0.50 M in THF, 20 ml, 10 mmol, 1.2 equiv), and

¹⁹ Kamitani, M.; Ito, M.; Itazaki, M.; Nakazawa, H. *Chem. Commun.* **2014**, 50, 7941.

THF (20 mL). The product was purified by column flash chromatography (20–40% EtOAc/hexanes) to afford the title compound as a yellow oil (0.60 g, 3.0 mmol, 78%). Analytical data is consistent with literature values.²⁰ **TLC** R_f = 0.2 (20% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl₃) δ 8.57 (d, J = 5.0 Hz, 1H), 7.69–7.62 (m, 2H), 7.57 (d, J = 7.8 Hz, 1H), 7.53 (d, J = 7.6 Hz, 1H), 7.45 (t, J = 8.0 Hz, 1H), 7.25–7.20 (m, 1H), 7.14 (d, J = 8.0 Hz, 1H), 5.80 (s, 1H), 5.44 (s, 1H); **¹³C NMR** (125 MHz, CDCl₃) δ 159.9, 148.0, 144.2, 137.1, 130.8 (q J = 32.4 Hz), 130.4, 129.0, 124.8, 124.7 (q, J = 4.16 Hz), 123.8 (q, J = 3.70 Hz), 122.8, 121.2, 74.4.

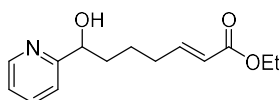
Synthesis of Substrate for Scheme 4.2



4.25 Using representative procedure A outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.95 mL, 10 mmol, 1.0 equiv), pent-4-en-1-ylmagnesium bromide (1.1 M in THF, 10 mL, 11 mmol, 1.1 equiv), and THF (15 mL). The product was purified by column flash chromatography (20–40% EtOAc/hexanes) to afford the title compound as a yellow oil (0.784 g, 4.40 mmol, 44%). **TLC** R_f = 0.4 (40% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl₃) δ 8.53 (d, J = 4.5 Hz, 1H), 7.68 (td, J = 7.6, 1.5 Hz, 1H), 7.25 (d, J = 8.1 Hz, 1H), 7.19 (dd, J = 6.9, 5.1 Hz, 1H), 5.79 (ddt, J = 17.1, 10.3, 6.7 Hz, 1H), 4.99 (dd, J = 17.1, 1.6 Hz, 1H), 4.94 (d, J = 10.3 Hz, 1H), 4.80–4.78 (m, 1H), 4.25 (s, 1H), 2.16–2.02 (m, 2H), 1.89–1.78

²⁰ Agai, B.; Proszenyak, A.; Tarkanyi, G.; Vida, L.; Faigl, F. *Eur. J. Org. Chem.* **2004**, 3623.

(m, 1H), 1.74–1.62 (m, 1H), 1.58–1.47 (m, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 162.3, 148.3, 138.8, 136.8, 122.4, 120.4, 114.7, 72.7, 38.1, 33.8, 24.6; IR (neat) 3260, 2936, 1594 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{11}\text{H}_{15}\text{ONNa}$ ($\text{M} + \text{Na}$) $^+$ 200.1051, found 200.1056.



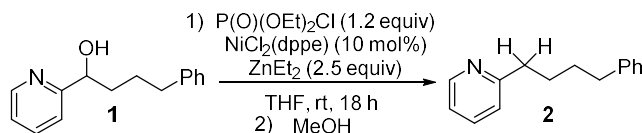
4.14 The title compound was prepared according to a modified procedure reported by Grubbs.²¹ In a glovebox, a flame-dried bomb flask was charged with a stir bar, **SI-10** (0.71 g, 4.0 mmol, 1.0 equiv), and Hoveyda-Grubbs Catalyst 2nd Generation (178 mg, 0.280 mmol, 0.0700 equiv). The flask was removed from the glovebox, and anhydrous CH_2Cl_2 (50 mL) and ethyl acrylate (2.2 mL, 20 mmol, 5.0 equiv) were added. The flask was sealed and heated to reflux over 24 h. The flask was then cooled to ambient temperature, and the solvent was removed in vacuo. The residue was purified by flash column chromatography to afford the title compound as a pale yellow oil (125 mg, 0.501 mmol, 13%, 13:1 E:Z). TLC R_f = 0.1 (40% EtOAc/hexanes, UV active); ^1H NMR (500 MHz, CDCl_3) δ 8.53 (d, J = 4.4 Hz, 1H), 7.68 (td, J = 7.6, 1.5 Hz, 1H), 7.26 (d, J = 7.7 Hz, 1H), 7.20 (dd, J = 6.9, 5. Hz, 1H), 6.93 (dt, J = 15.5, 7.1 Hz, 1H), 5.80 (dt, J = 15.5, 1.5 Hz, 1H), 4.75 (dd, J = 7.2, 4.3 Hz, 1H), 4.33 (br s, 1H), 4.16 (q, J = 7.2 Hz, 2H), 2.28–2.18 (m, 2H), 1.90–1.81 (m, 1H), 1.75–1.66 (m, 1H), 1.64–1.55 (m, 2H), 1.28 (t, J = 7.2 Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 166.8, 162.0, 148.9, 148.3, 136.8, 122.4, 121.7, 120.4, 72.5, 60.2, 38.0, 32.1, 23.7, 14.3; IR (neat) 2938, 1730 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{14}\text{H}_{19}\text{NO}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 272.1263, found 272.1271.

²¹ Chatterjee, A. K; Choi, T-L.; Sanders, D. P.; Grubbs, R. H. *J. Am. Chem. Soc.* **2003**, *125*, 11360.

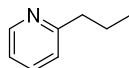
NICKEL CATALYZED FORMATION OF ALKYL ZINC REAGENTS AND DEOXYGENATION OF BENZYLIC CARBINOLS

Synthesis of Products for table 4.2

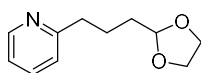
General Procedure A. Deoxygenation of benzylic carbinols



In a glovebox, a flame-dried 7 mL vial equipped with a stir bar was charged with NiCl₂(dppe) (11 mg, 0.020 mmol, 0.10 equiv), alcohol **1** (45 mg, 0.20 mmol, 1.0 equiv) and diethyl chlorophosphate (35 μ L, 0.24 mmol, 1.2 equiv) and suspended in THF (1.6 mL). The reaction vial was capped with a screw-cap, fitted with a septum and removed from the glove box. The reaction was placed under N₂, and stirred at which point diethyl zinc (1 M in PhMe, 0.5 mL, 0.5 mmol, 2.5 equiv) was added at room temperature resulting in an immediate color change from slightly orange to transparent yellow. The reaction was then sealed with parafilm and allowed to stir for 18 hours at 24 °C after which the reaction typically is dark brown or black. The reaction was quenched with MeOH and filtered through a plug of silica gel (100% EtOAc). The solvent was removed under reduced vacuum and the crude purified by flash column chromatography (12% EtOAc, 1% triethyl amine in hexanes) to afford the title compound as a pale yellow oil (29 mg, 0.13 mmol, 69%). **TLC** R_f = 0.3 (20% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl₃) δ 8.52 (ddd, *J* = 4.9, 1.9, 1.0 Hz, 1H), 7.56 (td, *J* = 2.0, 7.7 Hz, 1H), 7.29–7.23 (m, 2H), 7.19–7.14 (m, 3H), 7.13–7.06 (m, 2H), 2.81 (t, *J* = 7.3 Hz, 2H), 2.65 (t, *J* = 7.7 Hz, 2H), 1.83–1.74 (m, 2H) 1.74–1.64 (m, 2H); **¹³C NMR** (125 MHz, CDCl₃) δ 162.2, 149.3, 142.6, 136.3, 128.5, 128.3, 125.7, 122.7, 120.9, 38.3, 35.8, 31.3, 29.6; **IR** (neat) 3025, 2926, 2855, 2359, 2341, 1598 cm⁻¹; **HRMS** (TOF MS CI⁺) *m/z* calcd for C₁₆H₁₈ONH₄ (M + NH₄)⁺ 211.1361, found 211.1351.



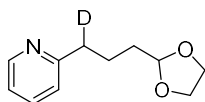
4.8 Using representative procedure B outlined above, the following amounts of reagents were used: NiCl₂(dppe) (11 mg, 0.020 mmol, 0.10 equiv), alcohol **SI-1** (28 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μ l, 0.24 mmol, 1.2 equiv), THF (1.6 mL) and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (16 mg, 0.13 mmol, 67%). Analytical data is consistent with literature values.ⁱⁱ **TLC R_f** = 0.3 (20% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl₃) δ 8.53 (d, J = 4.4 Hz, 1H), 7.58 (t, J = 7.7 Hz, 1H), 7.14 (d, J = 7.7 Hz, 1H), 7.09 (t, J = 6.5 Hz, 1H), 2.77 (t, J = 7.8 Hz, 2H), 1.76 (sex, J = 7.7 Hz, 2H), 0.97 (t, J = 7.7 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 162.4, 149.3, 136.3, 122.9, 121.0, 40.5, 23.2, 14.0.



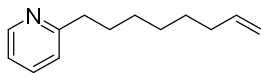
4.9 Using representative procedure B outlined above, the following amounts of reagents were used: NiCl₂(dppe) (11 mg, 0.020 mmol, 0.10 equiv), alcohol **3** (42 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μ l, 0.24 mmol, 1.2 equiv), THF (1.6 mL) and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (12% EtOAc/hexanes) to afford the title compound as a pale yellow oil (31 mg, 0.15 mmol, 74 %). Analytical data are consistent with literature values.²² **TLC R_f** = 0.1 (20% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl₃) δ 8.52 (d, J = 4.9 Hz, 1H), 7.57 (td, J = 7.6, 1.6 Hz, 1H),

²² Kitbunnadaj, R.; Zuiderveld, O.P.; Christophe, B.; Hulscher, S.; Menge, W.M.P.B.; Gelens, E.; Snip, E.; Bakker, R.A.; Celanire, S.; Gillard, M.; Talaga, P.; Timmerman, H.; Leurs, R. *J. Med. Chem.* **2004**, *47*, 2414.

7.14 (d, $J = 7.8$ Hz, 1H), 7.10 (dd, $J = 7.6, 5.3$ Hz, 1H), 4.89 (t, $J = 6$ Hz, 1H), 4.00–3.79 (m, 4H), 2.84 (t, $J = 7.7$ Hz, 2H), 1.92–1.82 (m, 2H), 1.76–1.68 (m, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 161.8, 149.3, 136.3, 122.8, 121.1, 104.5, 64.9, 38.1, 33.4, 24.2.

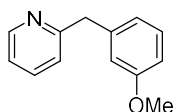


4.4 Using representative procedure B outlined above, the following amounts of reagents were used: $\text{NiCl}_2(\text{dppe})$ (11 mg, 0.020 mmol, 0.10 equiv), alcohol **3** (42 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μl , 0.24 mmol, 1.2 equiv), THF (1.6 mL) and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The reaction was quenched with CD_3OD from a sealed ampoule (1 mL) and the product was purified by flash column chromatography (12 % EtOAc, 1% triethyl amine, in hexane) to afford the title compound as a pale yellow oil (30 mg, 0.15 mmol, 78%). TLC $R_f = 0.1$ (20% EtOAc/hexanes, UV active); ^1H NMR (500 MHz, CDCl_3) δ 8.52 (d, $J = 4.6$ Hz, 1H), 7.58 (td, $J = 7.7, 1.9$ Hz, 1H), 7.15 (d, $J = 7.7$ Hz, 1H), 7.11–7.08 (m, 1H), 4.89 (t, $J = 4.7$ Hz, 1H), 4.01–3.79 (m, 4H), 2.87–2.79 (m, 1H), 1.91–1.83 (m, 2H), 1.76–1.68 (m, 2H); ^2H NMR (500 MHz, CHCl_3) 3.10 (s, 1D) ^{13}C NMR (125 MHz, CDCl_3) δ 161.8, 149.3, 136.3, 122.8, 121.1, 104.5, 64.9, 38.1, 33.4, 24.2; IR (neat) 2922, 2874, 1591, 1567, 1473, 1410 cm^{-1} ; HRMS (TOF MS Cl^+) m/z calcd for $\text{C}_{16}\text{H}_{18}\text{ONH}_4$ ($\text{M} + \text{NH}_4$) $^+$ 195.1244, found 195.1251.



4.6 Using representative procedure B outlined above, the following amounts of reagents were used: $\text{NiCl}_2(\text{dppe})$ (11 mg, 0.020 mmol, 0.10 equiv), alcohol **SI-3** (41 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μl , 0.24 mmol, 1.2 equiv), THF (1.6 mL) and diethyl zinc (1.0 M in PhMe,

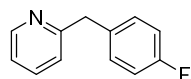
0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (29 mg, 0.15 mmol, 77%). **TLC** R_f = 0.2 (20% EtOAc/hexanes, UV active); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 8.52 (d, J = 4.5 Hz, 1H), 7.58 (td, J = 7.6, 1.5 Hz, 1H), 7.14 (d, J = 7.6 Hz, 1H), 7.09 (dd, J = 7.3, 5.5 Hz, 1H), 5.80 (ddt, J = 17.1, 10.4, 3.3 Hz, 1H), 4.98 (dd, J = 17.1, 1.1 Hz, 1H), 4.92 (d, J = 10.4 Hz, 1H), 2.78 (t, J = 7.8 Hz, 2H), 2.03 (dd, J = 13.5, 6.5 Hz, 2H), 1.72 (quint, J = 7.6 Hz, 2H), 1.44–1.30 (m, 6H); **$^{13}\text{C NMR}$** (125 MHz, CDCl_3) δ 162.2, 149.3, 139.3, 136.3, 122.8, 121.0, 114.3, 38.6, 33.9, 30.0, 29.6, 29.1, 28.9; **IR** (neat) 3075, 2925, 1589 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{13}\text{H}_{19}\text{NNa}$ ($\text{M} + \text{Na}$) $^+$ 212.1415, found 212.1406.



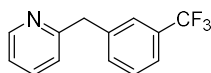
4.13 Using representative procedure B outlined above, the following amounts of reagents were used: $\text{NiCl}_2(\text{dppe})$ (11 mg, 0.020 mmol, 0.10 equiv), alcohol **SI-7** (43 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μl , 0.24 mmol, 1.2 equiv), THF (1.6 mL) and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (20 % EtOAc, 10% DCM, in hexanes) to afford the title compound as a pale yellow oil (21.5 mg, 0.120 mmol, 54%). Analytical data are consistent with literature values.²³ **TLC** R_f = 0.3 (20% EtOAc/hexanes, UV active); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 8.54 (d, J = 4.3 Hz, 1H), 7.57 (td, J = 7.6, 1.7, 1H), 7.21 (t, J = 7.9 Hz, 1H), 7.12 (s, 1H), 7.10 (t, J = 3.2 Hz, 1H), 6.85 (d, J = 7.4 Hz, 1H), 6.81 (s, 1H), 6.76 (dd, J = 8.4, 2.4 Hz, 1H), 4.13 (s, 2H), 3.77 (s, 3H),

²³ Chen, X.; Zhou, L.; Li, Y.; Xie, T.; Zhou, S. *J. Org. Chem.* **2014**, *79*, 230.

^{13}C NMR (125 MHz, CDCl_3) δ 160.9, 159.8, 149.4, 141.1, 136.6, 129.6, 123.2, 121.5, 121.3, 114.9, 111.9, 55.2, 44.8.



4.7 Using representative procedure B outlined above, the following amounts of reagents were used: $\text{NiCl}_2(\text{dppe})$ (11 mg, 0.020 mmol, 0.10 equiv), alcohol **SI-8** (43 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μl , 0.24 mmol, 1.2 equiv), THF (1.6 mL) and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The crude product was pre-absorbed to a minimal amount of silica and purified by flash column chromatography (12 % EtOAc, 1% triethyl amine, in hexane) to afford the title compound as a pale yellow oil (20.3 mg, 0.120 mmol, 60%). Analytical data are consistent with literature values.²⁴ **TLC** R_f = 0.3 (20% EtOAc/hexanes, UV active); **^1H NMR** (500 MHz, CDCl_3) δ 8.55 (ddd, J = 4.9, 1.9, 1.0 Hz, 1H), 7.58 (td, J = 7.6, 1.9 Hz, 1H), 7.24–7.19 (m, 2H), 7.14–7.07 (m, 2H), 7.01–6.95 (m, 2H), 4.12 (s, 2H); **^{13}C NMR** (125 MHz, CDCl_3) δ 162.5 (d, J_{CF} = 244.14 Hz), 160.6, 149.4, 136.6, 135.1 (d, J_{CF} = 3.24 Hz), 130.5 (d, J_{CF} = 7.87 Hz), 123.0, 121.3, 115.3 (d, J_{CF} = 21.2 Hz), 43.8.

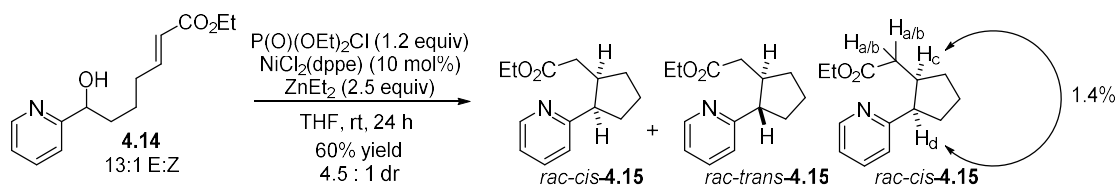


4.10 Using representative procedure B outlined above, the following amounts of reagents were used: $\text{NiCl}_2(\text{dppe})$ (11 mg, 0.020 mmol, 0.10 equiv), alcohol **SI-9** (51 mg, 0.20 mmol, 1.0 equiv),

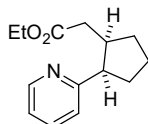
²⁴ De Houwer, J.; Tehrani, K. A.; Maes, B. U. W. *Angew. Chem., Int. Ed.* **2012**, *51*, 2745.

diethyl chlorophosphate (35 μ l, 0.24 mmol, 1.2 equiv), THF (1.6 ML) and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (30–50% EtOAc/hexanes) to afford the title compound as a pale yellow oil (29.4 mg, 0.124 mmol, 62%). Analytical data are consistent with literature values¹⁰ **TLC** R_f = 0.2 (10% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl₃) δ 8.56 (d, J = 4.9 Hz, 1H), 7.61 (dt, J = 2.0, 7.6 Hz, 1H), 7.53 (s, 1H), 7.50–7.38 (m, 3H), 7.17–7.09 (m, 2H), 4.21 (s, 2H); **¹³C NMR** (125 MHz, CDCl₃) δ 159.8, 149.6, 140.4, 136.8, 132.5, 128.9, 125.8, 125.7, 123.4, 123.3, 123.2, 121.6, 44.3.

Intramolecular 1,4-addition (Scheme 4.2)



4.15 Using representative procedure B outlined above, the following amounts of reagents were used: $NiCl_2(dppe)$ (11 mg, 0.020 mmol, 0.10 equiv), alcohol **16** (50 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μ l, 0.24 mmol, 1.2 equiv), THF (1.6 mL) and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (20% EtOAc/hexanes) to afford the title compound as a mixture 2.5:1 mixture of diastereomers (37 mg, 0.16 mmol, 80%). H_c was integrated to determine the ratio of diastereomers. Flash column chromatography (5–10% EtOAc/hexanes) was performed a second time to isolate analytically pure *cis* diastereomer (19 mg, 0.081 mmol, 40%) and a 1:1 mixture of diastereomers (16 mg, 0.07 mmol, 34%). The *cis* diastereomer was assigned based on the nOe correlation shown.

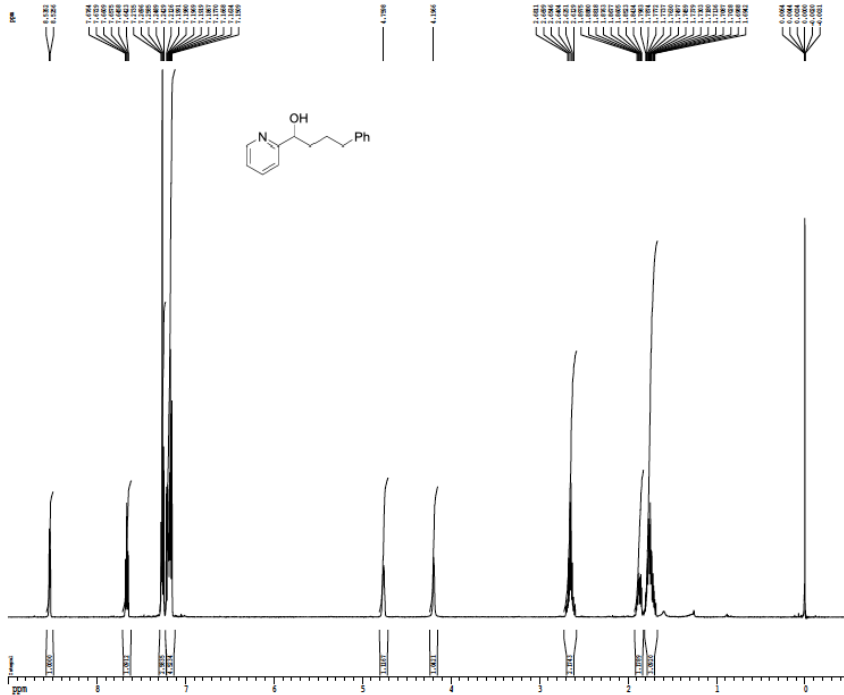


TLC R_f = 0.1 (10% EtOAc/hexanes, UV active); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 8.55 (d, J = 4.7 Hz, 1H), 7.56 (td, J = 7.7, 2.0 Hz, 1H), 7.12–7.07 (m, 2H), 4.00 (q, J = 7.2 Hz, 2H), 3.43 (q, J = 8.0 Hz, 1H), 2.79–2.71 (m, 1H), 2.14–2.04 (m, 2H), 2.02–1.90 (m, 4H), 1.75–1.67 (m, 1H), 1.63–1.54 (m, 1H), 1.17 (t, J = 7.2 Hz, 3H); **$^{13}\text{C NMR}$** (125 MHz, CDCl_3) δ 173.6, 163.2, 149.2, 136.0, 123.5, 121.2, 60.2, 49.8, 40.8, 36.2, 31.9, 30.2, 24.2, 14.3; **IR** (neat) 2935, 1716 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{14}\text{H}_{19}\text{NO}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 256.1313, found 256.1316.

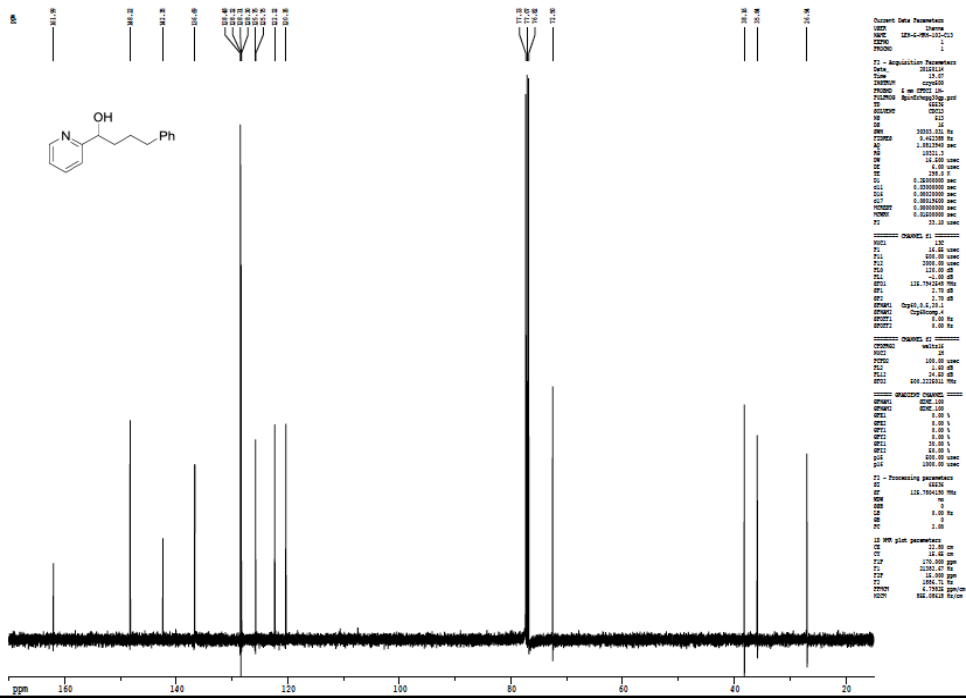
ⁱ Cao, B.; Wang, Y.; Ding, K.; Neamati, N.; Long, Y-Q. *Org. Biomol. Chem.* **2012**, *10*, 1239.

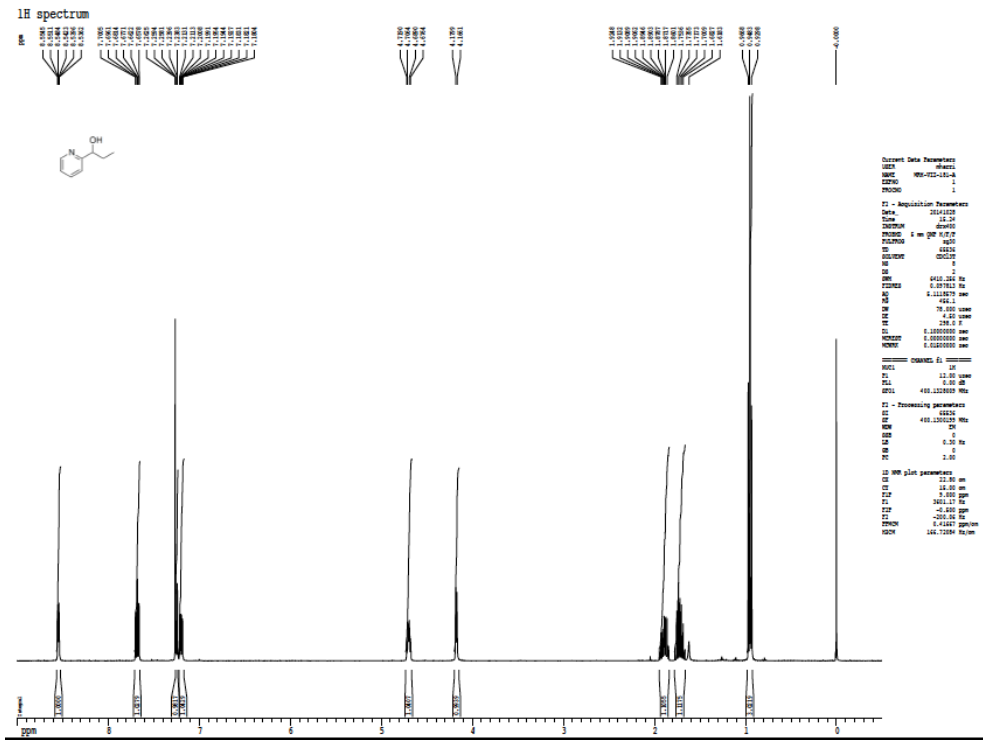
ⁱⁱ Groenhagen, U.; Maczka, M.; Dickschat, J. S.; Schulz, S. *Beilstein J. Org. Chem.* **2014**, *10*, 1421.

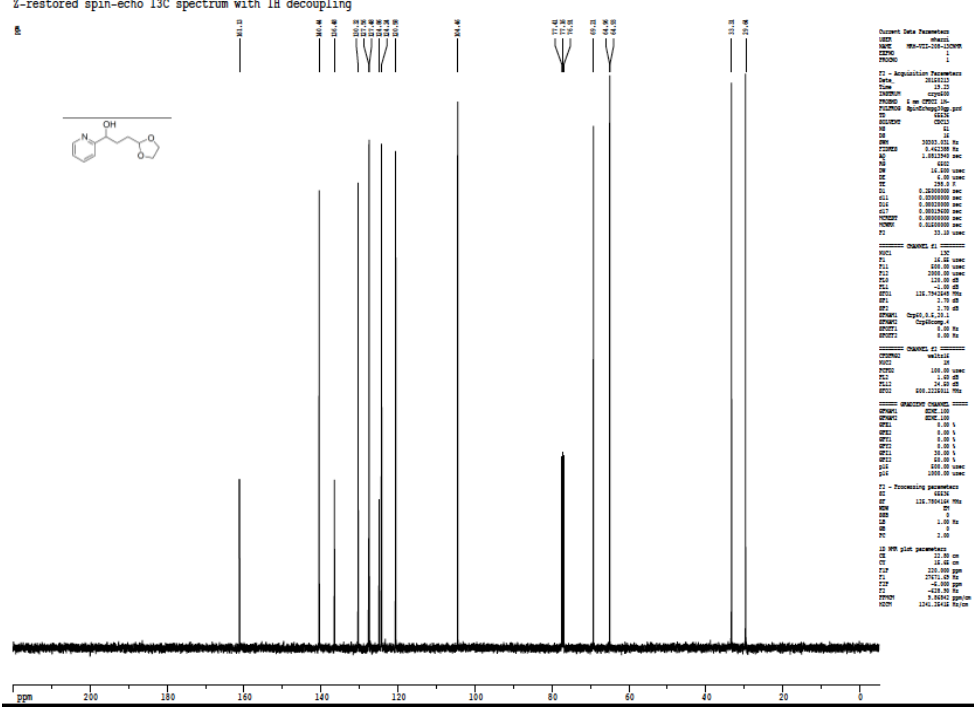
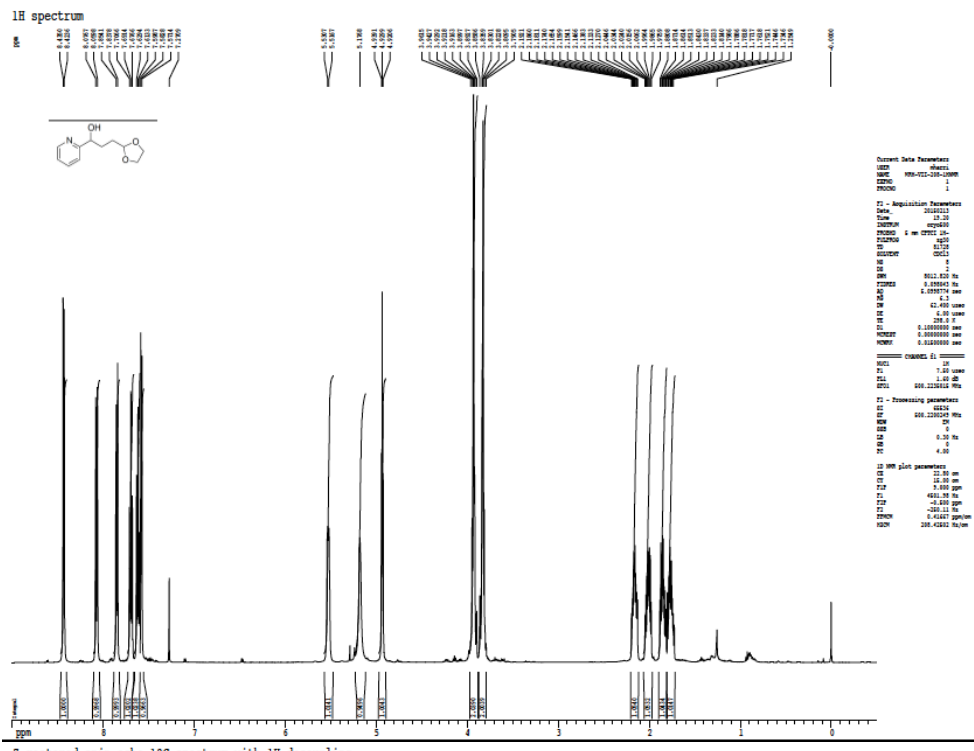
1H spectrum



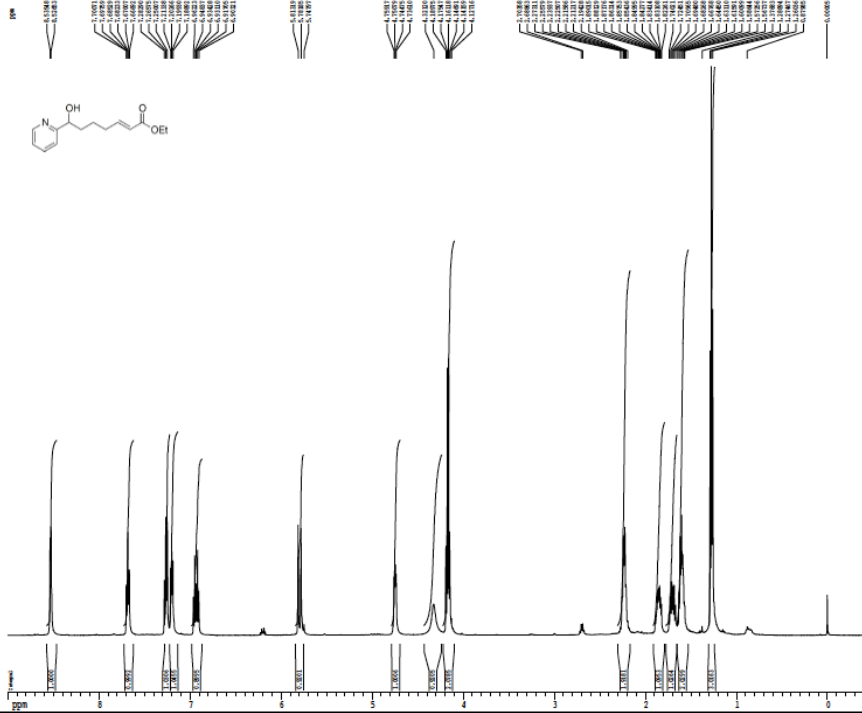
Z-restored spin-echo 13C spectrum with 1H decoupling





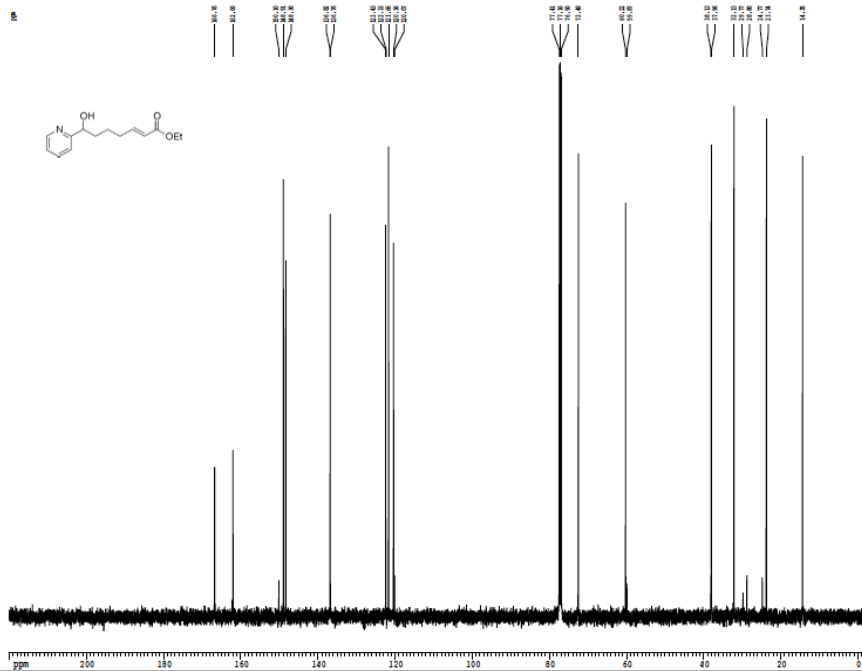


¹H spectrum



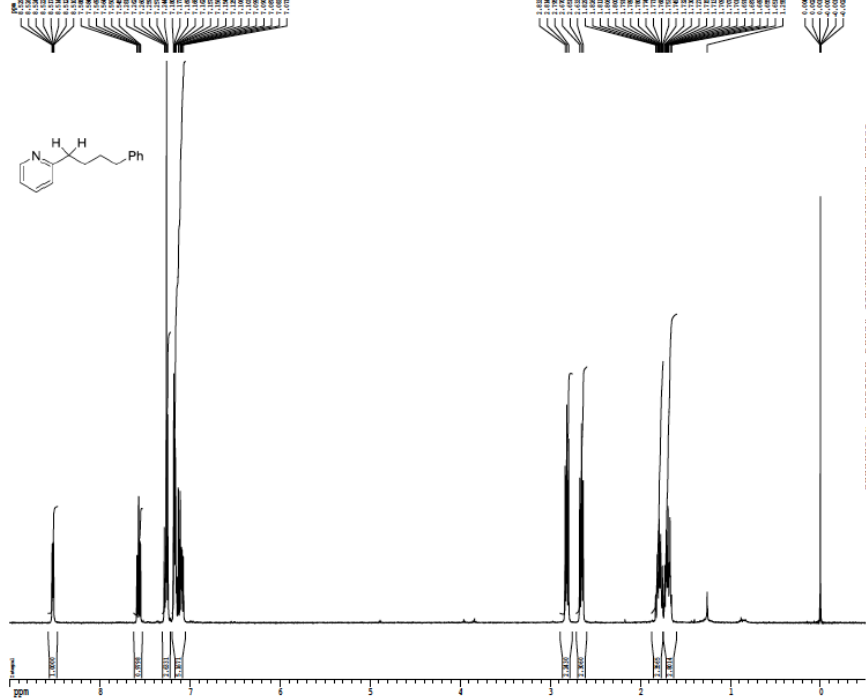
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¹³C spectrum with ¹H decoupling

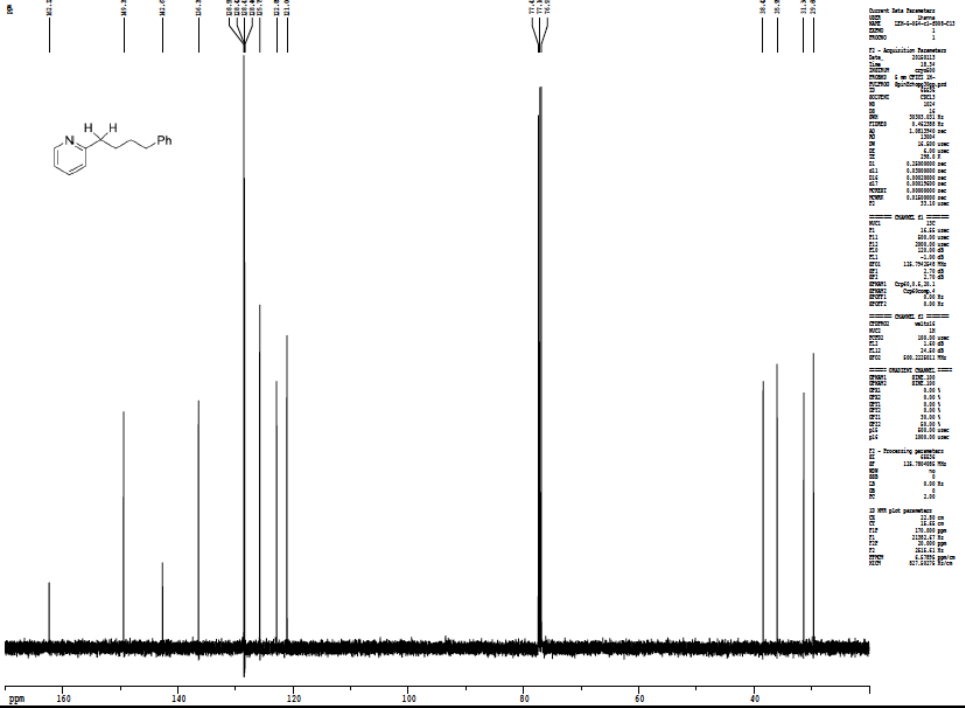


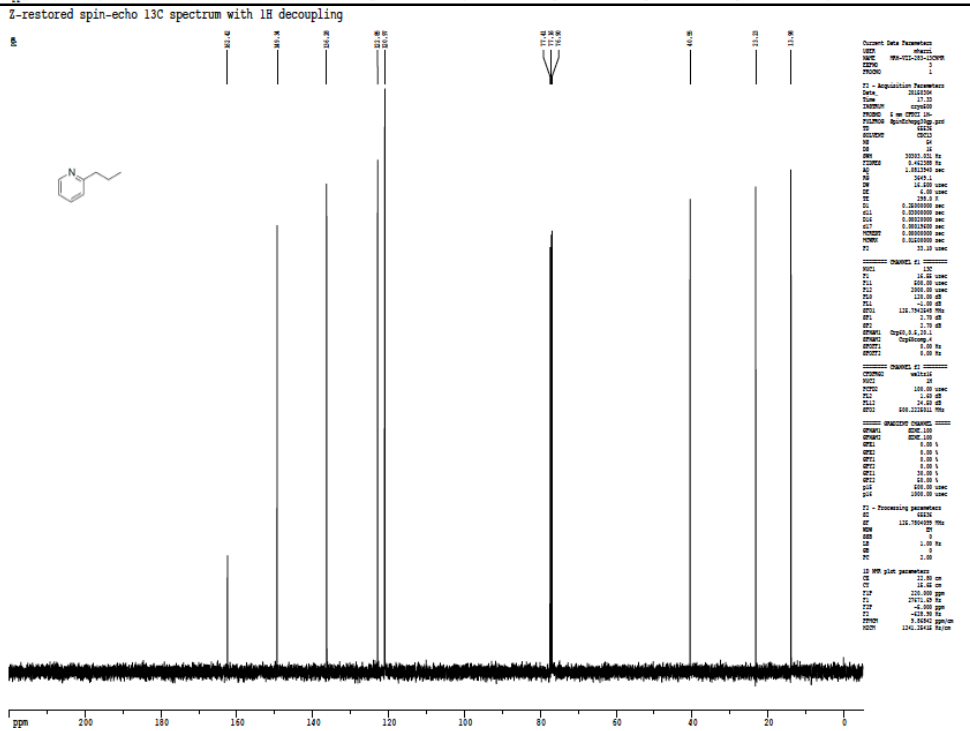
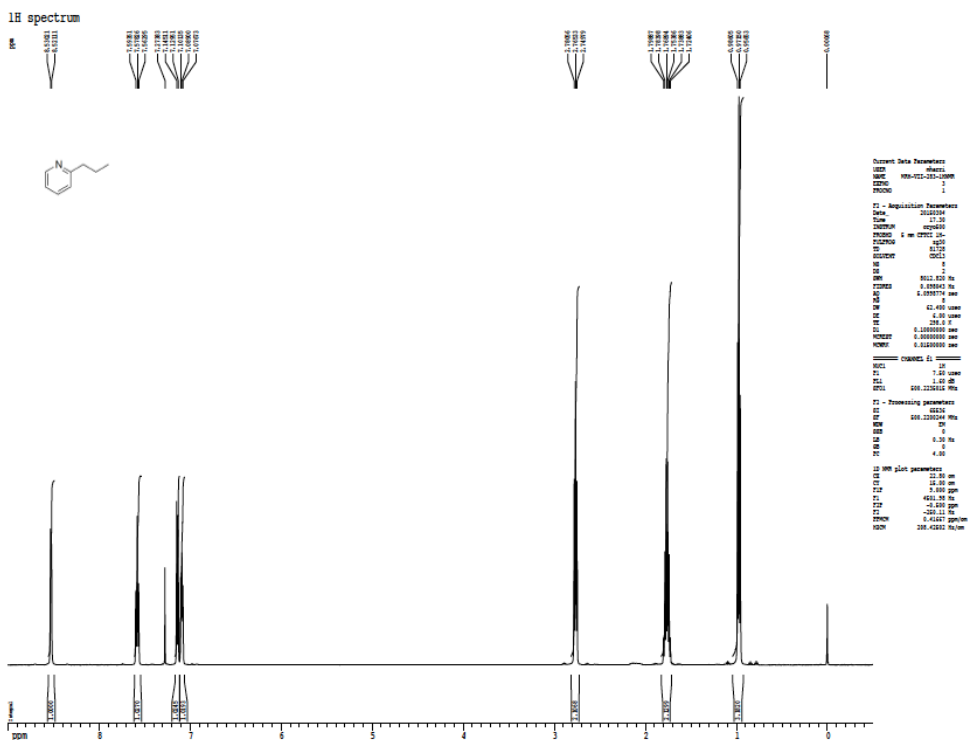
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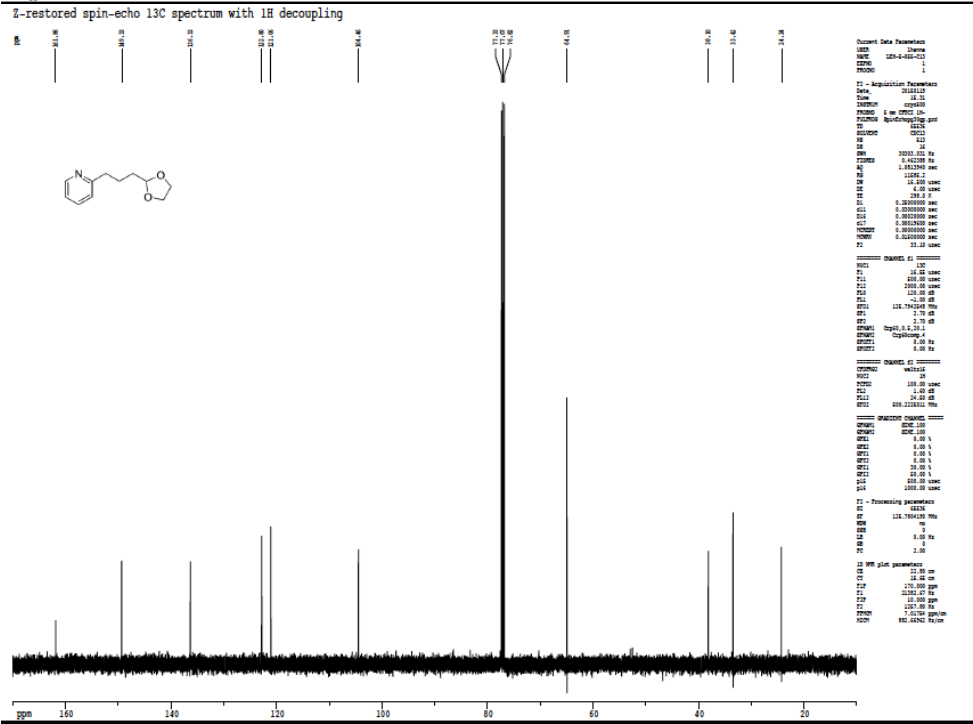
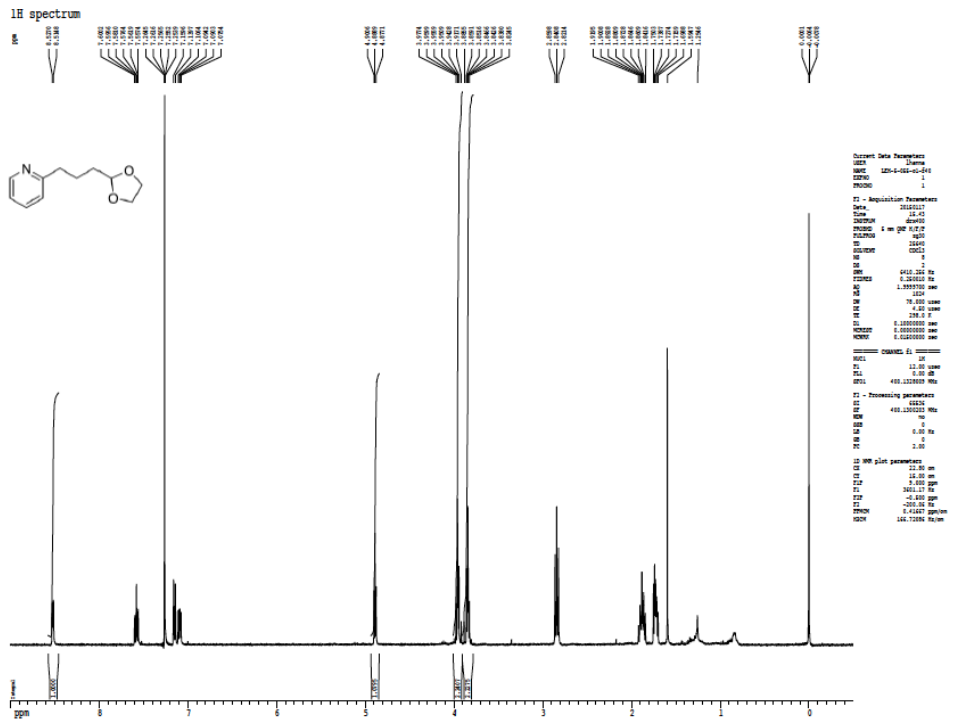
¹H spectrum

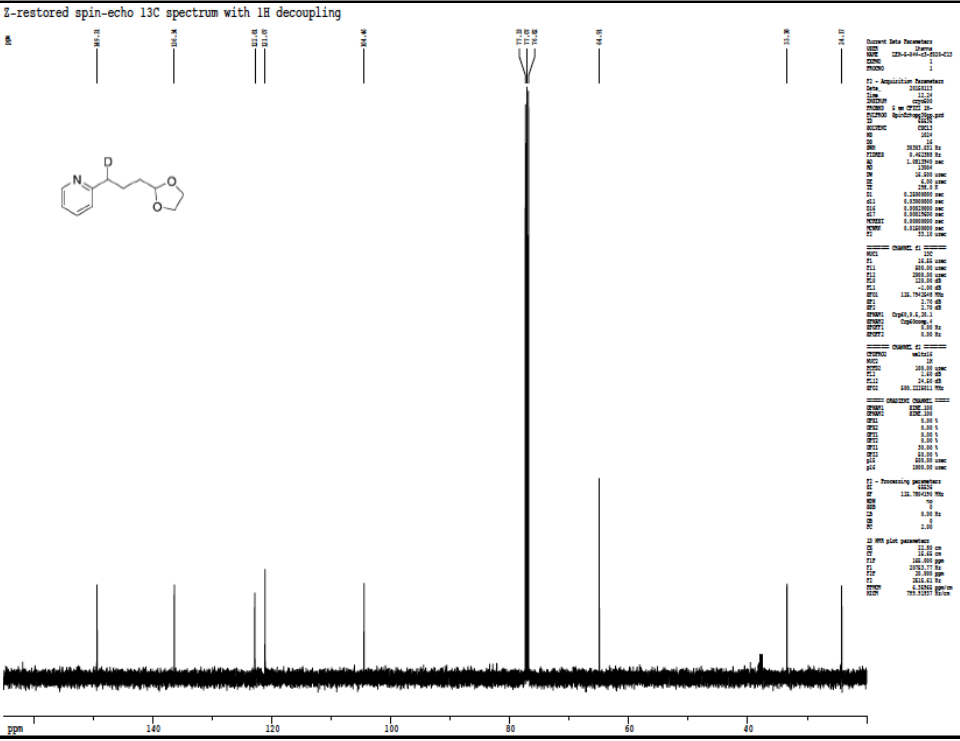
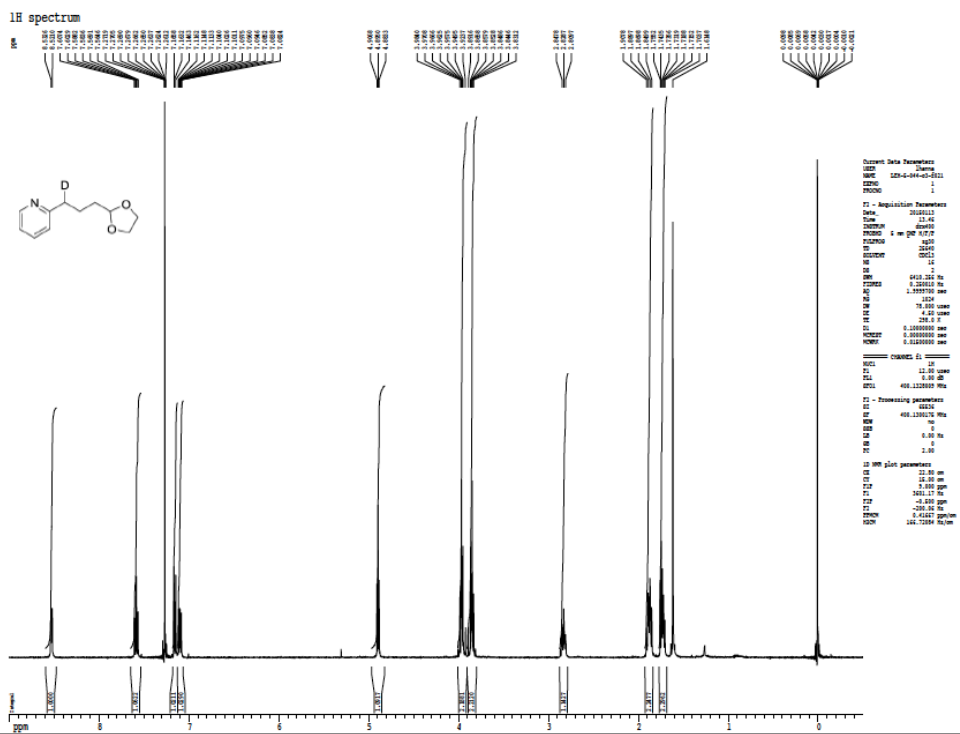


Z-restored spin-echo 13C spectrum with 1H decoupling

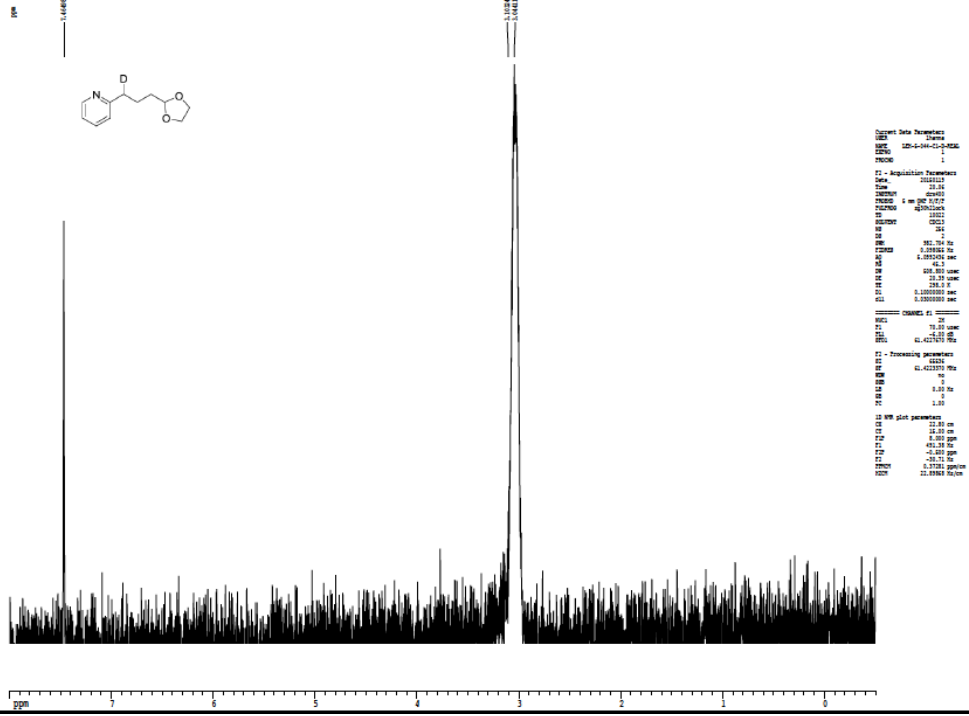


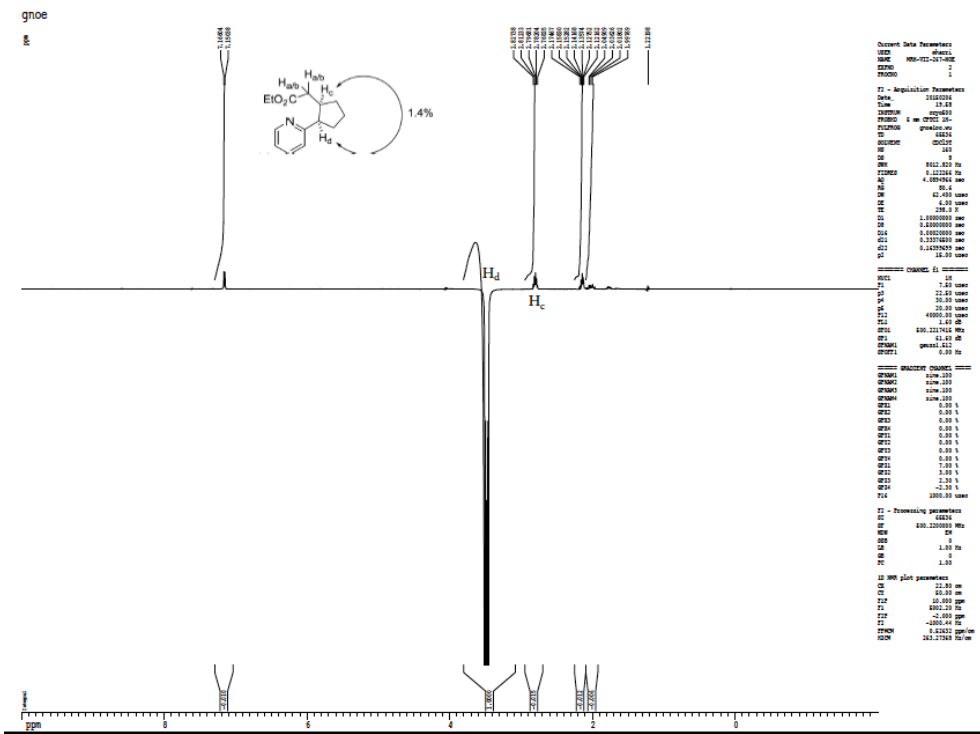




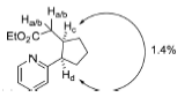


2H spectrum (measure via lock channel without changing any cables)





gnoe
g



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