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OPEN Quantification of tumor microenvironment acidity in glioblastoma using principal component analysis of dynamic susceptibility contrast enhanced MR imaging

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Glioblastoma (GBM) has high metabolic demands, which can lead to acidification of the tumor microenvironment. We hypothesize that a machine learning model built on temporal principal component analysis (PCA) of dynamic susceptibility contrast-enhanced (DSC) perfusion MRI can be used to estimate tumor acidity in GBM, as estimated by pH-sensitive amine chemical exchange saturation transfer echo-planar imaging (CEST-EPI). We analyzed 78 MRI scans in 32 treatment naive and post-treatment GBM patients. All patients were imaged with DSC-MRI, and pH-weighting that was quantified from CEST-EPI estimation of the magnetization transfer ratio asymmetry (MTR_{asym}) at 3 ppm. Enhancing tumor (ET), non-enhancing core (NC), and peritumoral T2 hyperintensity (namely, edema, ED) were used to extract principal components (PCs) and to build support vector machines regression (SVR) models to predict MTR_{asym} values using PCs. Our predicted map correlated with MTR_{asym} values with Spearman's r equal to 0.66, 0.47, 0.67, 0.71, in NC, ET, ED, and overall, respectively ($p < 0.006$). The results of this study demonstrates that PCA analysis of DSC imaging data can provide information about tumor pH in GBM patients, with the strongest association within the peritumoral regions.

Glioblastoma (GBM) is the most common malignant primary brain tumor in adults, characterized with vascular proliferation, diffuse infiltration in the adjacent brain parenchyma, and resistance to the standard therapies¹. The tumor microenvironment plays an important role in abundant infiltration of GBM tumor cells, its resistance to standard therapies, recurrence and therefore, poor patient prognosis². Due to rapid growth of GBM tumors and actively migrating cell population, hypercellular regions are formed typically surrounding the necrotic foci tissues and have a high metabolic demand³. When the tumor grows, the lack of sufficient circulation compared to the cell population of the tumor results in ischemia and secretion of angiogenic factors, which in turn leads to proliferation of new vessels⁴.

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Total no. of patients	32
Pre-surgery timepoints	12
Post-surgery timepoints	89
Age (years)	
Mean	64.6 ± 10.11
Median	66.5
Range	40
Sex	
Male	19
Female	13
IDH status	
Wild type	29
Mutant	3
MGMT promoter methylation status	
Methylated	20
Unmethylated	12

Table 1. Patient demographics.

Neo-angiogenesis forms a tortuous and branched vascular structure with increased blood volume and permeability, and impaired cerebral perfusion with subsequent necrosis¹⁷. These alterations promote tumor growth, decrease oxygen, increase glycolysis and lactic acid, decrease extracellular pH, facilitate cell invasion¹⁸. This augments the probability of mutations, such as vascular endothelial growth factor (VEGF) gene expression triggered by the hypoxia-inducible factor (HIF) family of transcription factors¹⁹. Even in presence of abundant oxygen, glycolysis is often enhanced in cancers due to elevated concentration of lactic acid, resulting in a substantial decrease in extracellular pH which leads to escalated invasion and aggressiveness of the tumor, and decreased immune response²⁰.

Various MRI techniques have been used to measure tumor pH. Magnetic resonance spectroscopy (MRS) reveals the amount and spatial distribution of particular biochemical substances involved in metabolic processes in tumoral tissues²¹. ¹H MRS, ¹³C MRS, and phosphorus MRS (³¹P MRS) can estimate levels of cellular metabolites and tumoral pH^{21,22}. Chemical exchange saturation transfer (CEST) is a relatively new MR technique that images certain compounds at very low concentrations that are too low to be directly detected by MRS technique²³. Tumor blood perfusion may also reflect the underlying tumor pH, as well-perfused tumors have more efficient removal of excess lactic acid, protons, CO₂, and other metabolic byproducts. Dynamic susceptibility contrast (DSC) perfusion is a widespread clinical MRI approach for quantification of cerebral perfusion and is often collected for assessment of brain tumors²⁴. As DSC-MRI can measure tissue perfusion and compromised microvasculature in GBMs, it might be able to quantify tumor acidity.

Amine CEST echo-planar imaging (CEST-EPI) is a fast molecular imaging MRI technique to measure tumor pH²⁵. In a recent study, a positive linear correlation between cerebral blood volume (CBV) obtained from DSC perfusion MRI and acidity was demonstrated in areas of T2-hyperintense, non-enhancing tumor in glioma patients²⁵. We hypothesize that principal component analysis (PCA) of DSC-MRI perfusion images in conjunction with machine learning (ML) techniques in patients with GBM may quantify microvascular structure at the voxel level and infer capillary-level hemodynamics that correlates with tumor acidity. PC analysis of DSC-MRI has shown potential in predicting the location of future recurrence^{26,27}, patient's survival²⁸, arteriovenous shunting²⁹, and *EGFR*/*MGMT* status³⁰. The aim of this analysis is to use ML methods based on perfusion MRI scans to uncover unique tissue characteristics that correlate with tissue acidity and might provide insights about the tumoral and peritumoral tissue metabolism to guide treatment planning.

Results

In this prospective study, we included 32 patients (19 males, 13 females; age, 64.6 ± 10.11 years old), who were confirmed to have GBM tumors (Table 1). A total of 101 CEST-MRI scans were acquired from the study subjects (12 patients had pre-surgical and 89 had imaging during active treatment with radiation and/or chemotherapy), of which, 78 scans with their corresponding DSC-MRI scans passed our data preparation and pre-processing steps. The SVR machine learning method based on PCA was applied in a cohort of 78 cases.

Principal component analysis of perfusion time-series revealed that the tumor subregions, i.e., ET, NC, ED, form characteristic clusters (Fig. 1B), which facilitate specification of the tissues and allow for mapping the heterogeneity within a specific tissue. Figure 2 illustrates structural MRI scans, including T1, T1-Gd, T2, T2-FLAIR images for a male patient (58 years old) with GBM. Furthermore, relative cerebral blood volume (rCBV) map generated from DSC-MRI scans using CAPS software, PC1–PC3 images derived from PC analysis of the hemodynamic curves, the MTR_{param} image constructed using our proposed approach, along with the actual MTR_{param} image quantified from CEST imaging are shown in the whole pathogenic region. As it can be inferred, our constructed MTR_{param} provides more accurate voxel-wise mapping of the actual MTR_{param} image, compared to the rCBV map. Among the seven PCs used in building our model, the visual similarity is most striking for the first three PC images since the components progressively capture less variance.

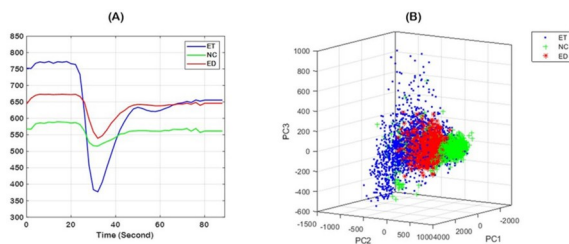


Figure 1. An illustration of the perfusion time-series in tumorous subregions, i.e., ET, NC, and ED (A); and the clustering of each tissue type using PCA analysis (B), signifying the potential of the PCs in capturing tissue characteristics. PC1, PC2, and PC3 represent the first, second, and third principal components, respectively. *ET* Enhancing tumor, *NC* Necrotic core, *ED* peritumoral edema.

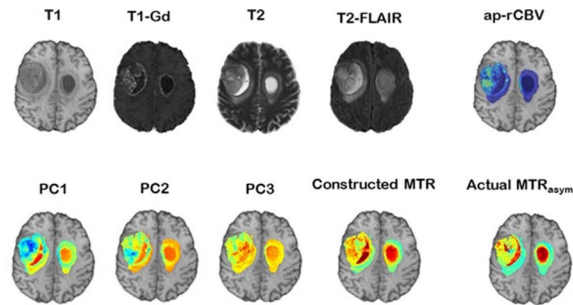


Figure 2. Conventional MRI, including T1, T1-Gd, T2, and T2-FLAIR, scans of a 58-year-old male patient included in our study. Map of a proxy to relative cerebral blood volume (ap-rCBV) derived from DSC-MRI scans with CaPTk software. Three principal components (PCs), PC1 to PC3, calculated using PCA of the hemodynamic perfusion curves, along with the MTR_{sym} image constructed using the seven PCs in association with the actual MTR_{sym} image. CaPTk version 1.8.1 (www.med.upenn.edu/cbic/captk/).

The MTR_{sym} image constructed from perfusion PCs using our proposed regression method showed moderate to strong agreement with the MTR_{sym} image, with R of 0.47 ($p = 0.006$), 0.66 ($p = 0.0009$), 0.67 ($p = 0.00001$) in the ET, NC, and ED regions, respectively, and 0.71 ($p < 10^{-6}$) in the whole pathologic region, as a union of ET, NC, and ED areas, averaged over all the patients. Figure 3 demonstrates a strong association of the MTR_{sym} image built from the perfusion PCs and the actual MTR_{sym} image, implying the potential of ML in distinguishing the tumorous regions with specific metabolism characteristics.

This method outperforms the conventional DSC-MRI analysis, as displayed in Fig. 4, where panel (A) presents the signal intensity–time curves of the voxels located in the high (red) and low (blue) MTR_{sym} image. While the voxels have been selected from the regions with different levels of acidic pH, the perfusion curves are not discriminant. Applying PCA analysis differentiates the clusters of the same high and low MTR_{sym} , as it can be observed from panel (B) of Fig. 4. The bottom panel (part C) shows three PCs quantified for the higher and lower MTR_{sym} regions. Figure 4C reveals that the first principal component (PC1) is primarily related to the level of the perfusion signal, as evidenced by the large variance throughout the signal time course. The second principal

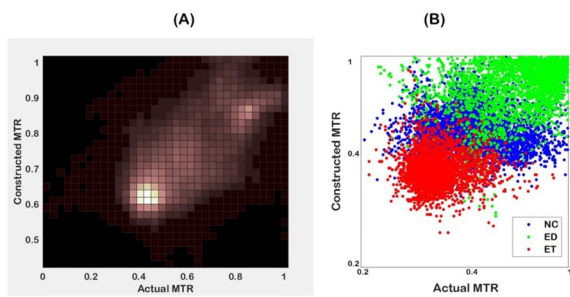


Figure 3. Demonstration of (A) bivariate histogram of the constructed in comparison with actual $MTR_{0.0m}$ images; and (B) association of the clusters of tumor tissues in the constructed versus actual $MTR_{0.0m}$ image.

component (PC2) is related to the depth of the perfusion signal drop, in relation to the baseline level, and the third principal component (PC3) relates to the shape of the drop of the perfusion signal, e.g., steepness of the signal drop and recovery. As a comparison, Fig. 5 displays the signal intensity–time curves in the highest (red) and lowest (blue) voxels in PC1, PC2, and PC3 images, suggesting the differentiability of the tissues based on PCA analysis. Specifically, PC1 provides a noticeable discrimination between the areas with different hemodynamic properties. The discrimination diminishes in larger PCs as evidenced by this illustration.

Discussion

Our study showed that high-resolution pH-sensitive imaging in brain tumors can be achieved on clinical 3T MR systems using DSC-MRI with strong correlation to CEST-EPI pH imaging. DSC-MRI can characterize microvascular circulation in GBM patients, and the respective acidity can be extracted using PCA and ML techniques. The proposed techniques are ideally suited for evaluation of malignant gliomas since GBM remains the most angiogenic primary brain tumor and therefore exhibits extensive neovascularization, compromised brain blood barrier, and heterogeneous acidity. DSC-MRI without proper processing cannot discern the pH heterogeneity of the tumoral regions, but as proposed in this study, PCA of the perfusion time-series can be used to differentiate tissue acidity.

GBMs have unique pathophysiological characteristics such as significant invasiveness, fast growth, and rapid seeding. Also, similar to the other cancers, GBMs prefer glycolysis over oxidative phosphorylation even in the presence of ample oxygen (Warburg effect) that results in increased intracellular lactate and elevated acidification. These tumors also cause direct destruction of surrounding tissue, including both neuronal death via glutamate excitotoxicity³⁸ and degradation of the extracellular matrix via metalloproteinases and other proteases³⁹ that are pH-dependent^{40,41}. Since, ion movement is directly coupled to movement of other ions, pH not only serves as a regulator of cell activity, but also an indirect surrogate marker of other cellular functions. pH heterogeneity in the tumor microenvironment is critical for surgical and chemo-radiation planning. Weak base and weak acidic drugs get trapped in either the intracellular or extracellular spaces due to “ion trapping” phenomena⁴².

Wang et al. investigated association between pH-sensitive CEST-EPI and relative cerebral blood volume (CBV) measurements obtained from DSC-MRI in patients with gliomas¹³. They reported a strong correlation between acidity and CBV in the T2-hyperintense regions, but not in the areas of enhancing tumor. In agreement with this study, our results demonstrated that the correlation between our constructed $MTR_{0.0m}$ map and the actual $MTR_{0.0m}$ as quantified by CEST-EPI, is highest in the peritumoral T2-hyperintense areas, followed by necrotic tumor core, and contrast-enhancing tumor regions. However, in our study, in different subregions of each tumor, a heterogeneous distribution of high-acidity tissues was observed.

The proposed PCA-derived features integrated with SVR method uses temporal dynamics of DSC-MR imaging. Our results indicate that a more comprehensive analysis of DSC along with machine learning, can unravel useful information related to tumor acidity, which is typically not obtained with traditional CBV or CBF maps. Specifically, higher acidotic tissues demonstrate higher level of DSC signal based on PC1, consistent with higher necrosis and free water mobility. PC2 mainly represents the depth of drop in DSC signal, and deeper drop in the signal shows higher acidity. Integrative information of signal drop and recovery is stored in PC3, where lower signal recovery is associated with higher acidotic tissues.

CEST-EPI has been shown to be effective as a non-invasive biomarker to distinguish between IDH1 mutant and wild type gliomas, and also 1p/19q codeleted from intact IDH1 mutant gliomas^{12,43}. In addition, it has been shown to be of value as an early imaging biomarker for bevacizumab treatment response and failure in recurrent

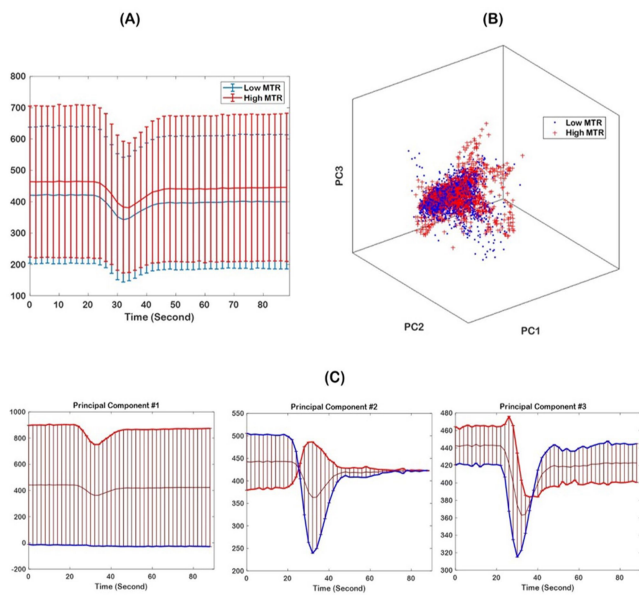


Figure 4. (A) Perfusion curves calculated within regions of low and high MTR_{app} (shown in blue and red colors, respectively), suggesting poor discrimination of the regions solely based on hemodynamic curves. (B) Discrimination of low and high MTR_{app} regions based on PC analysis; PC1 = principal component 1; PC2 = principal component 2; PC3 = principal component 3. (C) The three principal components for high MTR_{app} regions, yielding a marked differentiation of these regions based on the PCs.

glioblastoma²⁷. Our proposed approach could support measuring tumor acidity with DSC-MRI, as a more widely-accessible imaging method compared with CEST-EPI.

There are limitations to our study, including limited sample size and single institutional data collection; these results need to be validated in a large multi-institutional study.

In conclusion, the results of this study indicate that PCA analysis of DSC-MRI in conjunction with machine learning techniques, can potentially enable better localization of highly acidic regions. In turn, this information may be used for tumor prognostication and treatment response evaluation.

Methods

Patients. Institutional review board (IRB) approval of the University of Pennsylvania was obtained for this prospective study and informed consent was collected from the participants. All methods were carried out in accordance with relevant guidelines and regulations. 32 subjects with intra-axial brain mass suggestive of high-grade glioma, who were referred to the radiology department of hospital of University of Pennsylvania from March 2018 to February 2020 and were subsequently proven by histopathology to be GBM were included in this study.

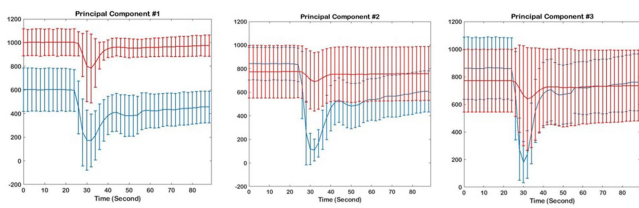


Figure 5. The perfusion curves calculated from the regions with highest (red) and lowest (blue) values on individual Principal Component images: (left) Principal Component 1; (middle) Principal Component 2; and (right) Principal Component 3.

Image acquisition. All MRI scans were performed on a Magnetom Tim Trio 3 Tesla scanner (Siemens, Erlangen, Germany) using a 12-channel phased array head coil. Conventional MRI sequences included axial T1-weighted (T1) before and after administration of gadolinium contrast agent (T1-Gd) with matrix size = $192 \times 256 \times 192$, resolution = $0.98 \times 0.98 \times 1.00$ mm³, repetition time (TR in ms)/echo time (TE in ms) = 1760/3.1, T2-weighted (T2) with matrix size = $208 \times 256 \times 64$, resolution = $0.94 \times 0.94 \times 3.00$, TR/TE = 4680/85, T2 fluid-attenuated inversion recovery (T2-FLAIR) with matrix size = $192 \times 256 \times 60$, resolution = $0.94 \times 0.94 \times 3.00$, TR/TE = 9420/141.

DSC-MR imaging was performed by a gradient-echo planar (GE-EPI) imaging sequence during a second 0.1 mmol/kg bolus of Dotarem (Gadoterate Meglumine) with the following parameters: TR/TE = 2000/45 ms, FOV = 22×22 cm², resolution = $1.72 \times 1.72 \times 3$ mm³, 20 sections. A bolus of contrast agent with a dose of 0.1 mmol/kg which was done for DCE (dynamic-contrast-enhanced) imaging served as a preload dose for DSC imaging to reduce the effect of contrast agent leakage on relative cerebral blood volume (rCBV) measurements.

Acquisition of pH-sensitive information was performed through an amine contrast specific for single-echo CEST-EPI sequence^{3,39}. MR imaging acquisition parameters included the following: FOV = $240 \times 256 \times 217 \times 256$ mm, matrix size = $128 \times 116 \times 128$, slice thickness = 4 mm with no inter-slice gap, 25 consecutive slices, excitation pulse flip angle = 90°, TE = 27 ms, bandwidth = 1628 Hz, and generalized auto-calibrating partially parallel acquisition factor = 2–3. Off-resonance saturation was applied using a pulse train of 3×100 ms Gaussian pulses with a peak amplitude of 6 microtesla. A total of 29 off-resonance or z-spectral points were sampled at frequency offsets of -3.5 to -2.5 ppm, -0.3 to $+0.3$ ppm, and -2.5 to $+3.5$ ppm, all in increments of 0.1 ppm. A reference scan (S0) was obtained with the same acquisition parameters, without the saturation pulses. Total scan time for CEST-EPI was approximately 6 min.

MRI pre-processing. For each patient, all MRI volumes (T1, T2, T2-FLAIR, DSC-MRI and MTR_{min}) were rigidly co-registered with their corresponding T1-Gd using the Greedy registration method⁴⁰ (<https://github.com/pushkevich/greedy>). Subsequently, all conventional MRI scans (T1, T1-Gd, T2, T2-FLAIR) were smoothed to remove any high frequency intensity variations (i.e., noise)⁴¹, corrected for magnetic field inhomogeneities using N4ITK method⁴² and skull-stripped using FSL BET⁴³ followed by manual revision when needed. For brain tumor segmentation in the images, DeepMedic⁴⁴, a Deep Learning (DL)-based segmentation algorithm in Cancer Imaging Phenomics Toolkit (CaPTK) v1.7.8⁴⁵ which had been trained on BraTS 2017 training data, was applied to the co-registered conventional MRI scans. Brain tumor segmentation delineated three regions of interest (ROIs), i.e., enhancing tumor (ET), necrosis (NC), and peritumoral edema (ED), in the GBM tumors.

Amine CEST-EPI post-processing. Clinical post-processing of CEST-EPI consisted of affine motion correction (MCFLIRT; FSL, <https://fsl.fmrib.ox.ac.uk/fsl/fslwiki/MCFLIRT>) and B0 correction via a z-spectra-based K-means clustering and Lorentzian fitting algorithm⁴⁶. An integral of the width of 0.4 ppm was then obtained around both the -3.0 and $+3.0$ ppm (-3.2 to -2.8 and $+2.8$ to $+3.2$ ppm, respectively) spectral points of the inhomogeneity-corrected data. These data points were combined with the S0 image to calculate the asymmetry in the magnetization transfer ratio (MTR_{asym}) at 3.0 ppm as defined by equation $MTR_{\text{asym}}(\omega) = S(-\omega) - S(\omega)/S$, where ω is the offset frequency of interest (3.0 ppm). All resulting maps were registered to high-resolution post-contrast T1-weighted images for subsequent analyses.

Temporal principal component analysis. Principal component analysis (PCA) is a dimensionality reduction method⁴⁷ which was used in this study to distill the DSC-MRI time series down to a few components that capture the temporal dynamics of blood perfusion. All hemodynamic perfusion curves were aligned and normalized for the baseline and maximum drop across the patients³⁷. We randomly selected voxels in each tumor subregion, i.e., ET, NC, and ED, and generated their signal intensity–time curves (Fig. 1A). PCA was subsequently applied to capture the variance of the time series in all the ROIs and all subjects. Because of the relative

consistency in the perfusion patterns of the various ROIs, seven principal components were sufficient to capture more than 99% of the variance in the perfusion signal for all tumor subregions and all patients.

Generation of MTR_{syn} images based on PCs using machine learning. We built several regression models for tumor subregions using support vector machine regression (SVR) aiming to predict the MTR_{syn} values from the seven PCs on a voxel-by-voxel basis to create a PC-derived MTR_{syn} image, referred to as constructed MTR_{syn} image. Leave-one-subject-out cross-validation of these predictive models was performed to ensure that the model and the associate estimates of accuracy would likely generalize to new patients. We trained the SVR models separately in ET, NC, and ED regions using Gaussian kernel functions with an automatic kernel scale and sequential minimal optimization (SMO) configuration. Performance of the SVR method was evaluated using Spearman's correlation. All machine learning and statistical analyses was performed in MATLAB 9.4.0.949201 (R2018a) Update 6.

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References

- Ostrom, Q. T. et al. CIBTRUS statistical report: Primary brain and other central nervous system tumors diagnosed in the United States in 2011–2015. *Neuro Oncol.* **20**, 181–186. <https://doi.org/10.1093/neuonc/nqz131> (2018).
- Cooper, L. A. et al. The tumor microenvironment strongly impacts master transcriptional regulators and gene expression class of glioblastoma. *Am. J. Pathol.* **180**, 2108–2119. <https://doi.org/10.1016/j.ajpath.2012.01.040> (2012).
- Brat, D. J. et al. Pseudopalisades in glioblastoma are hypoxic, express extracellular matrix processes, and are formed by an actively migrating cell population. *Cancer Res.* **64**, 920–927. <https://doi.org/10.1158/0008-5472.can-03-2073> (2004).
- Nabaviadeh, S. A. et al. Arterial spin labeling and dynamic susceptibility contrast-enhanced MR imaging for evaluation of arteriovenous shunting and tumor hypoxia in glioblastoma. *Sci. Rep.* **9**, 6747. <https://doi.org/10.1038/s41598-019-45312-x> (2019).
- Brat, D. J. & Van Meir, E. G. Vaso-occlusive and prothrombotic mechanisms associated with tumor hypoxia, necrosis, and accelerated growth in glioblastoma. *Lab. Invest. J. Tech. Methods Pathol.* **84**, 397–405. <https://doi.org/10.1038/labinvest.3700070> (2004).
- Harris, R. J. et al. Simultaneous pH-sensitive and oxygen-sensitive MRI of human gliomas at 3 T using multi-echo amine proton chemical exchange saturation transfer gradient echo echo-planar imaging (CEST-SAGE-EPI). *Magn. Reson. Med.* **80**, 1962–1978. <https://doi.org/10.1002/mrm.27204> (2018).
- Fathi Kazerooni, A., Bakas, S., Salghheh Rad, H. & Davatzikos, C. Imaging signatures of glioblastoma molecular characteristics: A radiogenomics review. *J. Magn. Reson. Imaging (MRI)* **52**, 54–69. <https://doi.org/10.1002/jmri.26907> (2020).
- Maxwell, P. H., Pugh, C. W. & Ratcliffe, P. J. Activation of the HIF pathway in cancer. *Curr. Opin. Genet. Dev.* **11**, 293–299. [https://doi.org/10.1016/s0959-4374\(00\)00193-3](https://doi.org/10.1016/s0959-4374(00)00193-3) (2001).
- Gillies, R. J. & Gatenby, R. A. Metabolism and its sequelae in cancer evolution and therapy. *Cancer J. (Sudbury Mass.)* **21**, 88–96. <https://doi.org/10.1097/tpc.0000000000000102> (2015).
- Estrella, V. et al. Acidity generated by the tumor microenvironment drives local invasion. *Cancer Res.* **73**, 1524–1535. <https://doi.org/10.1158/0008-5472.can-12-2796> (2013).
- Ciechocka, M., Korub, J. & Urbank, A. pH measurements of the brain using phosphorus magnetic resonance spectroscopy (31P MRS) in healthy men—Comparison of two analysis methods. *Pol. J. Radiol.* **80**, 509–514. <https://doi.org/10.12659/pjr.895178> (2015).
- Najac, C. & Rosen, S. M. MR molecular imaging of brain cancer metabolism using hyperpolarized ^{13}C magnetic resonance spectroscopy. *Top. Magn. Reson. Imaging (TMRI)* **25**, 187–196. <https://doi.org/10.1097/rmr.0000000000000104> (2016).
- Wu, B. et al. An overview of CEST MRI for non-MR physicists. *ENMMP Phys.* **3**, 19. <https://doi.org/10.1186/s40658-016-0155-2> (2016).
- Bouerman, J. L. et al. Consensus recommendations for a dynamic susceptibility contrast MRI protocol for use in high-grade gliomas. *Neuro Oncol.* **22**, 1262–1275. <https://doi.org/10.1093/neuonc/nosa141> (2020).
- Wang, Y. L. et al. Association between tumor acidity and hypervascularity in human gliomas using pH-weighted amine chemical exchange saturation transfer echo-planar imaging and dynamic susceptibility contrast perfusion MRI at 3T. *ANR Am. J. Neuro-radiol.* **40**, 979–986. <https://doi.org/10.3174/ajnr.A6063> (2019).
- Akbhari, H. et al. Imaging surrogates of infiltration obtained via multiparametric imaging pattern analysis predict subsequent location of recurrence of glioblastoma. *Neurosurgery* **78**, 572–580. <https://doi.org/10.1227/nec.0000000000001202> (2016).
- Akbhari, H. et al. Pattern analysis of dynamic susceptibility contrast-enhanced MR imaging demonstrates peritumoral tissue heterogeneity. *Radiology* **273**, 502–510. <https://doi.org/10.1148/radiol.14132458> (2014).
- Macyszyn, L. et al. Imaging patterns predict patient survival and molecular subtype in glioblastoma via machine learning techniques. *Neuro Oncol.* **18**, 417–425. <https://doi.org/10.1093/neuonc/nou127> (2016).
- Bakas, S. et al. In vivo detection of EGFRvIII in glioblastoma via perfusion magnetic resonance imaging signature consistent with deep peritumoral infiltration: The q-index. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **23**, 4724–4734. <https://doi.org/10.1158/1078-0432.ccr-16-1871> (2017).
- Ye, Z. C. & Sontheimer, H. Glioma cells release excitotoxic concentrations of glutamate. *Cancer Res.* **59**, 4383–4391 (1999). <https://doi.org/10.2741/1046> (2003).
- Fasciglione, G. F., Martin, S., D'Alessio, S., Politi, V. & Colletta, M. pH- and temperature-dependence of functional modulation in metalloproteinases. A comparison between neutrophil collagenase and gelatinases A and B. *Biophys. J.* **79**, 2138–2149. [https://doi.org/10.1016/s0006-3495\(00\)76461-7](https://doi.org/10.1016/s0006-3495(00)76461-7) (2000).
- Gioia, M. et al. pH dependence of the enzymatic processing of collagen I by MMP-1 (fibroblast collagenase), MMP-2 (gelatinase A), and MMP-14 ectodomain. *J. Biol. Inorg. Chem. IBC Publ. Soc. Biol. Chem.* **15**, 1219–1232. <https://doi.org/10.1007/s00775-010-0680-8> (2010).
- Raghunand, N. & Gillies, R. J. pH and drug resistance in tumors. *Drug Resist. Updates Rev. Comment. Antimicrob. Anticancer Chemother.* **3**, 39–47. <https://doi.org/10.1054/drug.2000.0119> (2000).
- Yao, J. et al. Metabolic characterization of human IDH mutant and wild type gliomas using simultaneous pH- and oxygen-sensitive molecular MRI. *Neuro Oncol.* **21**, 1184–1196. <https://doi.org/10.1093/neuonc/nou078> (2019).
- Yao, J. et al. Human IDH mutant 1p19q co-deleted gliomas have low tumor acidity as evidenced by molecular MRI and PET: A retrospective study. *Sci. Rep.* **10**, 11922. <https://doi.org/10.1038/s41598-020-68733-5> (2020).
- Yao, J. et al. pH-weighted amine chemical exchange saturation transfer echoplanar imaging (CEST-EPI) as a potential early biomarker for bevacizumab failure in recurrent glioblastoma. *J. Neurooncol.* **142**, 587–595. <https://doi.org/10.1007/s11060-019-03132-z> (2019).

28. Harris, R. J. *et al.* Simulation, phantom validation, and clinical evaluation of fast pH-weighted molecular imaging using amine chemical exchange saturation transfer echo planar imaging (CEST-EPI) in glioma at 3 T. *NMR Biomed.* **29**, 1563–1576. <https://doi.org/10.1002/nbm.3611> (2016).
29. Yushkevich, P. A. *et al.* Fast automatic segmentation of hippocampal subfields and medial temporal lobe subregions in 3 Tesla and 7 Tesla T2-weighted MRI. *Alzheimer's Dement.* **7**, 126–127 (2016).
30. Smith, S. M. & Brady, J. M. SUSAN—A new approach to low level image processing. *Int. J. Comput. Vis.* **23**, 45–78 (1997).
31. Sled, J. G., Zijdenbos, A. P. & Evans, A. C. A nonparametric method for automatic correction of intensity nonuniformity in MRI data. *IEEE Trans. Med. Imaging* **17**, 87–97. <https://doi.org/10.1109/42.668698> (1998).
32. Smith, S. M. Fast robust automated brain extraction. *Hum. Brain Mapp.* **17**, 143–155. <https://doi.org/10.1002/hbm.1002> (2002).
33. Kamnitsas, K. *et al.* Efficient multi-scale 3D CNN with fully connected CRF for accurate brain lesion segmentation. *Med. Image Anal.* **36**, 61–78. <https://doi.org/10.1016/j.media.2016.10.004> (2017).
34. Davatzikos, C. *et al.* Cancer imaging phenomics toolkit: quantitative imaging analytics for precision diagnostics and predictive modeling of clinical outcome. *J. Med. Imaging (Bellingham, Wash.)* **5**, 011018. <https://doi.org/10.1117/1.jmi.5.1.011018> (2018).
35. Fathi Kazerooni, A. *et al.* Cancer imaging phenomics via CaPTK: Multi-institutional prediction of progression-free survival and pattern of recurrence in glioblastoma. *JCO Clin. Cancer Inform.* **4**, 234–244. <https://doi.org/10.1200/cci.19.00121> (2020).
36. Tso, J. *et al.* Improving B(0) correction for pH-weighted amine proton chemical exchange saturation transfer (CEST) imaging by use of k-means clustering and Lorentzian estimation. *Tomography (Ann Arbor, Mich.)* **4**, 123–137. <https://doi.org/10.18383/jtom.2018.00017> (2018).
37. Akbari, H. *et al.* Histopathology-validated machine learning radiographic biomarker for noninvasive discrimination between true progression and pseudo-progression in glioblastoma. *Cancer* **126**, 2625–2636. <https://doi.org/10.1002/cncr.32790> (2020).

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Competing interests


The authors declare no competing interests.

Additional information

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