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Columnar Specificity of Microvascular Oxygenation and Volume Responses: Implications for Functional Brain Mapping

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Cortical neurons with similar properties are grouped in columnar structures and supplied by matching vascular networks. The hemodynamic response to neuronal activation, however, is not well described on a fine spatial scale. We investigated the spatiotemporal characteristics of microvascular responses to neuronal activation in rat barrel cortex using optical intrinsic signal imaging and spectroscopy. Imaging was performed at 570 nm to provide functional maps of cerebral blood volume (CBV) changes and at 610 nm to estimate oxygenation changes. To emphasize parenchymal rather than large vessel contributions to the functional hemodynamic responses, we developed an ANOVA-based statistical analysis technique. Perfusion-based maps were compared with underlying neuroanatomy with cytochrome oxidase staining. Statistically determined CBV responses localized accurately to individually stimulated barrel columns and could resolve neighboring columns with a resolution better than 400 μm . Both CBV and early oxygenation responses extended beyond anatomical boundaries of single columns, but this vascular point spread did not preclude spatial specificity. These results indicate that microvascular flow control structures providing targeted flow increases to metabolically active neuronal columns also produce finely localized changes in CBV. This spatial specificity, along with the high contrast/noise ratio, makes the CBV response an attractive mapping signal. We also found that functional oxygenation changes can achieve submillimeter specificity not only during the transient deoxygenation (“initial dip”) but also during the early part of the hyperoxygenation. We, therefore, suggest that to optimize hemodynamic spatial specificity, appropriate response timing (using $\leq 2\text{--}3$ sec changes) is more important than etiology (oxygenation or volume).

Key words: neurovascular coupling; cerebral blood flow; cerebral blood volume; BOLD fMRI; optical imaging; initial dip; high resolution

Introduction

The pioneering studies by Mountcastle (1957) and Hubel and Wiesel (1959) demonstrated neurons in primary sensory cortices with similar properties are grouped together in columns. Columnar organization has also been described in higher association areas (Britten, 1998) and may be a general organizational principle of cerebral cortex (Rakic, 2002). Although functional architecture was originally studied with electrophysiological recordings, the role of imaging techniques such as positron emission tomography and functional magnetic resonance imaging (fMRI) is rapidly increasing. Spatial resolution of the latter has increased enough to allow columnar mapping in certain situations (Ugurbil et al., 2003). The ability to resolve individual functional units is essential for developing a deeper understanding of brain organization.

Ultimately, the spatial resolution of these imaging techniques is limited by the spatial specificity of the underlying perfusion-related signals. Activation of a cortical column increases neuronal oxidative metabolism, producing a decrease in tissue oxygenation (“initial dip”) spatially restricted to the active column (Malonek et al., 1997; Kim et al., 2000; Thompson et al., 2003). Increased activity generates a vascular response through a variety of putative mechanisms (Villringer and Dirnagl, 1995), leading to an increase in local cerebral blood flow (CBF) proportionately greater than the increase in oxygen metabolism (Fox and Raichle, 1986; Buxton and Frank, 1997). The correspondence between microvascular structures and neural columns (Woolsey et al., 1996) ensures the CBF response is also spatially specific to the activated column (Duong et al., 2001). The influx of oxygenated blood rapidly reverses the transient tissue deoxygenation, leading to a hyperoxygenation that begins in the capillary bed and drains into the venous system. Because the hyperoxygenation is most prominent in medium to large veins, this response is often considered spatially unspecific.

Increased flow also leads to increased cerebral blood volume (CBV), but their dynamic spatiotemporal relationship is not well known. We sought to determine whether the focal CBF increase also leads to a spatially specific CBV increase. Because previous

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studies suggested the volume response is highly vascular and “overspills” active cortex (Frostig et al., 1990; Malonek and Grinvald, 1996), we reasoned reduction of CBV signals from medium to large vessels might improve spatial specificity. We measured CBV changes using optical intrinsic signal (OIS) imaging in the rat barrel cortex system during stimulation of individual neighboring whiskers. To reduce the contribution of large vascular signals, we developed statistical analysis techniques that emphasize parenchymal responses.

Although many studies have found the delayed hyperoxygenation signal is too vascular to resolve submillimeter structures (Malonek and Grinvald, 1996; Kim et al., 2000), it may localize well within the first few seconds of stimulation onset (Menon and Goodyear, 1999). Similarly, Duong et al. (2000) reported the initial dip is spatially specific only within the first 2 sec. We, therefore, hypothesized that submillimeter specificity is a function of timing (observing early signals) more than etiology (oxygenation or volume). To investigate this concept further, we examined the spatiotemporal characteristics of functional oxygenation and volume changes with simultaneous OIS imaging and spectroscopy.

Materials and Methods

Animal preparation. Male Sprague Dawley rats ($n = 18$; 300–450 gm) were anesthetized with halothane (5% induction; 1.5% maintenance) and maintained at $36.5 \pm 0.5^\circ\text{C}$ with a heating blanket (Harvard Apparatus, Holliston, MA). We cannulated the tail artery for blood pressure (BP) and gas monitoring, the femoral vein for anesthesia, and the trachea for artificial ventilation. The skull was thinned, and silicone oil was applied to increase bone translucency. Anesthesia was switched to α -chloralose (60 mg/kg bolus, 30 mg/kg/hr infusion, i.v.) and pancuronium bromide (2 mg/kg bolus, 1.5 mg/kg/hr infusion), and inspired gases were adjusted to maintain physiological variables within the normal range: BP, 90–110 mmHg; $p_a\text{O}_2$, 121 ± 7 mmHg; $p_a\text{CO}_2$, 39 ± 1 mmHg (mean \pm SE). Halothane was discontinued for at least 1 hr before imaging.

Optical imaging. Animals were imaged under a SMZ1500 microscope (Nikon). Fiber optics directed white light to the preparation, and reflected light was filtered by a filter wheel (Lambda 10-2; Sutter Instruments, Novato, CA) and captured by a 16-bit CCD camera (Princeton Instruments, Trenton, NJ). We used transmission filters centered at 570 and 610 nm [full width half maximum (FWHM), 10 nm]. Imaging at 570 nm emphasizes CBV changes, and 610 nm imaging emphasizes oxygenation changes (Sheth et al., 2003). The field of view covered 5.3×7.2 mm with a 144×192 pixel array, providing a $37 \mu\text{m}$ resolution. Imaging was performed at 4 Hz with a 30 sec intertrial interval, and filters were alternated every trial.

Image analysis and histology. Ratio images were generated by subtracting and dividing a prestimulus baseline image from each image in a trial. Response time courses were calculated within a 1.0 mm diameter region of interest (ROI) in the trial-averaged data. ROI position was determined by an automated program (Matlab; The Mathworks, Natick, MA) that scanned the ROI across the peak 570 nm image and chose the position resulting in the greatest signal change. This ROI was applied to the averaged data of both wavelengths.

Functional maps were generated in two ways after Gaussian spatial filtering (three-pixel FWHM). The first method used the common approach of applying absolute thresholds to the ratio images (Chen-Bee et al., 2000). The second method used a technique that reduces the influence of large vascular fluctuations and provides statistical significance thresholds. An ANOVA test was performed at every pixel on a 64×3 matrix: the pixel value in each of 64 trials at three time points [$t = -3.50$ sec (prestimulus); $t = (\text{peak image}-1)$; $t = (\text{peak image})$]. Functional representations were displayed as the negative log of the resulting p value map. To identify the peak response area, we selected the 100 most significant pixels ($\sim 0.14 \text{ mm}^2$; approximately half the surface projection of

one barrel). After imaging, at least four fiducial lesions (10 sec, 10 μA) were made.

Animals were perfused with 0.1 M phosphate buffer and 4% paraformaldehyde. The right cortex was removed, flattened, cryoprotected, and cut tangentially in 40 μm sections. Sections were stained (Wong-Riley and Welt, 1980), photographed, and aligned to raw optical images in Photoshop (Adobe Systems, San Jose, CA).

Optical spectroscopy. We performed optical spectroscopy and imaging concurrently using a fiber bundle mounted on the microscope eyepiece that directed half the reflected light to a spectrophotometer (SpectraPro 300i; 100 μm slit; Acton Research, Acton, MA) and 16-bit CCD camera (Spec-10:400BR, Princeton Instruments). Spectra were acquired from 520 to 653 nm with 0.5 nm resolution with timing identical to that of image acquisition.

Reflectance spectra were analyzed using a modified form of the Beer–Lambert law incorporating optical attenuation attributable to absorption and scattering:

$$\log \frac{I_0^\lambda}{I(t)} = (\epsilon_{\text{Hbr}}^\lambda \times \Delta[\text{Hbr}](t) + \epsilon_{\text{HbO}_2}^\lambda \times \Delta[\text{HbO}_2](t)) \times l^\lambda + \Delta S \quad (1)$$

where I_0 is the prestimulus intensity, $I(t)$ is the intensity time course, ϵ are extinction coefficients, l is the pathlength through the tissue, S is scattering, and $\Delta[\text{Hbx}](t)$ is the time course of change in absorbers. The high degree of scattering in biological tissue distorts the Hb attenuation spectra by significantly increasing the optical pathlength in a wavelength-dependent manner. To account for this pathlength wavelength dependency (superscript λ), we used an *in vitro* phantom model to simulate the absorption and scattering properties of rat cortex (Sato et al., 2002). The phantom contained rat blood as the absorber and Intralipid (Fresenius Kabi, Clayton, NC), as the scatterer. The blood was diluted to 1% ($\sim 50 \mu\text{M}$), the approximate mean value in rat brain (Cooper et al., 1998) and within the range assumed by other studies (Jones et al., 2001; Lindauer et al., 2001). A 1% Intralipid solution provided a reduced scattering coefficient (μ_s') of 1 mm^{-1} , similar to that of brain tissue (Nakai et al., 1997). Although the contribution of scattering to optical attenuation is large, changes in scattering during functional activation are small, because the hemodynamic response and associated flow increase are more likely to affect absorption than scattering (Obrig and Villringer, 2003). We, thus, made the common simplifying assumption that scattering and pathlength wavelength dependency remained constant over time.

If we define $\alpha = \epsilon \times l$ and remove time dependency, Equation 1 becomes:

$$\log \frac{I_0^\lambda}{I^\lambda} = \alpha_{\text{Hbr}}^\lambda \times [\text{Hbr}] + \alpha_{\text{HbO}_2}^\lambda \times [\text{HbO}_2] + S \quad (2)$$

where I_0 is the *in vitro* reference spectrum with 0% hematocrit, $\alpha_{\text{Hbr}} = \epsilon_{\text{Hbr}} \times l$, and $\alpha_{\text{HbO}_2} = \epsilon_{\text{HbO}_2} \times l$. By holding hematocrit fixed and measuring the reflected intensity, we calculated α_{Hbr} and α_{HbO_2} , which represent the effective pathlength-adjusted attenuation spectra of Hbr and HbO₂. The phantom solution was fully oxygenated ([Hbr] = 0) by bubbling 100% O₂ and deoxygenated ([HbO₂] = 0) by allowing yeast (0.5% by weight) to consume O₂ and adding 30 mg of Na₂O₂S₄. Scattering was assumed to be wavelength independent, because the wavelength-dependent component (i.e., differential pathlength) was incorporated into the effective attenuation spectra. *In vitro* measurements were made with the same instrument settings and geometry as *in vivo* measurements. Functional Hbr and HbO₂ time courses were calculated from Equation 1 using a least-squares analysis and the values of α_{Hbr} and α_{HbO_2} . Because of potential differences in pathlength scale between the *in vitro* and *in vivo* conditions, results are reported as relative changes from baseline.

Experimental paradigm. We investigated spatial specificity ($n = 10$) of oxygenation and volume changes in whisker barrel cortex (Woolsey and Van der Loos, 1970). We rotated the stereotactic frame $\sim 12^\circ$ around the rostrocaudal axis to orient barrel cortex normal to the optical axis. All whiskers, except C1 and C2, were trimmed. Whisker deflection was provided by a motorized stroker, consisting of a vertical rod attached to a

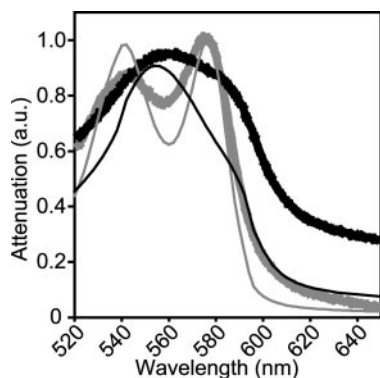


Figure 1. Phantom hemoglobin attenuation spectra. To account for the wavelength-dependent pathlength through highly scattering biological tissue, we measured reflectance spectra from an *in vitro* phantom simulating the scattering and absorption properties of brain (see Materials and Methods). Phantom attenuation spectra for HbO₂ (thick gray line) and Hbr (thick black line) were distorted relative to textbook extinction coefficients for HbO₂ (thin gray line) and Hbr (thin black line), which do not account for scattering. This difference is attributable to the inclusion of the differential pathlength factor in the highly scattering medium. Phantom and textbook spectra were normalized to facilitate comparison.

motor. Whiskers were deflected 2.8 mm in 11.5 msec in a rostral-to-caudal direction, measured ~1 cm from the face. Either C1 or C2 was deflected (2 sec, 5 Hz) during each trial, for a total of 128 trials (64 per wavelength) for each whisker.

In another eight animals, we studied the effect of response timing and etiology on spatial specificity with concurrent optical imaging and spectroscopy during electrical hindpaw stimulation (Sheth et al., 2003), which allowed us to modulate responses by varying amplitude (0.4, 0.5, 0.6, 0.8, 1.0, and 1.2 mA at 5 Hz) and frequency (2, 5, 10, 15, and 20 Hz at 0.8 mA). Stimulation (2 sec) consisted of 1 msec electrical pulses (ISO-Flex, Master-8; AMPI, Jerusalem, Israel) to the left hindpaw via two needle electrodes inserted into the plantar surface of the foot and ~10 mm away on the medial aspect of the leg. Twelve trials were collected per wavelength and stimulus condition.

Results

We assessed the spatiotemporal characteristics of functional changes in CBV and oxygenation with high resolution using OIS imaging and spectroscopy. We first confirmed that 570 nm OIS imaging reflects total hemoglobin (Hbt) and is, therefore, proportional to local changes in CBV. A novel statistical analysis procedure was used to generate functional maps with decreased vascular contribution. These maps demonstrated that volume responses could resolve columnar architecture with submillimeter precision. We provide evidence that optimal hemodynamic spatial specificity is primarily a function of timing rather than response etiology.

In vitro phantom spectra

The phantom attenuation spectra shown in Figure 1 represent the extinction coefficients of HbO₂ (α_{HbO_2}) and Hbr (α_{Hbr}) corrected for the wavelength-dependent pathlength. Inclusion of the differential pathlength distorted the phantom spectra relative to textbook absorption coefficients. Textbook spectra isosbestic points are at 522, 549, 569, and 586 nm; the HbO₂ maxima are at 542 and 577 nm; and the Hbr maximum is at 555 nm. Measured phantom spectra (0.5 nm resolution) isosbestic points were at 525, 545, 570.5, and 583 nm; the HbO₂ maxima were at 545 and 577.5 nm; and the Hbr maximum was at 557.5 nm.

Temporal characteristics

Whisker stimulation produced monophasic reflectance changes at 570 nm and biphasic changes at 610 nm (Fig. 2). The 570 nm

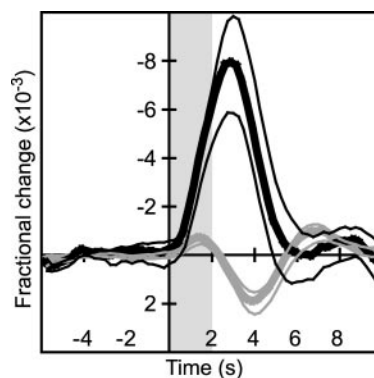


Figure 2. The 570 and 610 nm response to whisker stimulation. C1 whisker stimulation produced a monophasic reflectance decrease at 570 nm (thick black line), peaking 2.5–2.75 sec after stimulus onset. The 610 nm response (thick gray line) was biphasic, with an early negative peak occurring at 1.5 sec and a late positive peak at 4.0 sec. Negative changes are plotted upward for convention. Responses to C2 stimulation (data not shown) were similar. The shaded gray region indicates stimulus duration. The average responses are denoted by thick lines, and SEs by thin lines.

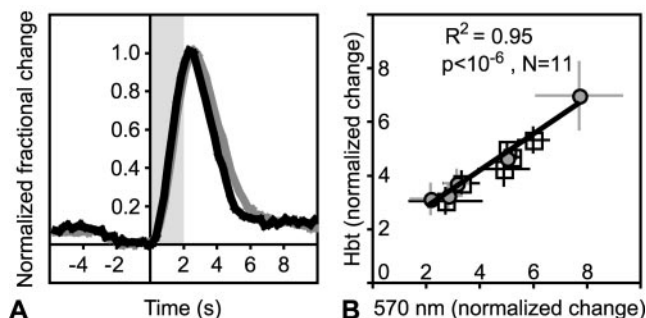


Figure 3. Etiology of 570 nm OIS responses. *A*, Normalized time courses of 570 nm (thick gray line) and Hbt (thick black line) responses from eight subjects. Both responses have similar latencies, time to peak, and overall profiles. *B*, Correlation analysis of normalized Hbt versus 570 nm peak responses to varying stimulation frequency (gray circle) and amplitude (open square) demonstrates strong correspondence between the two responses. Similar temporal profiles and peak magnitude coupling suggest that 570 nm imaging reflects changes in Hbt. Error bars represent SE.

peak occurred 2.5–2.75 sec after stimulus onset, and the 610 nm early and late phase peaks occurred at 1.5 and 4.0 sec, respectively. Baseline noise (SD of prestimulus time points) was 2.5×10^{-4} at 570 nm and 8.2×10^{-5} at 610 nm. The contrast/noise ratio (CNR) (peak magnitude divided by noise) was 30.1 for the 570 nm response, 7.2 for the 610 nm early phase, and 22.3 for the 610 nm late phase. We observed post-response hemodynamic oscillations consistent with other studies (Mayhew et al., 1996; Jones et al., 2001).

Because 569 nm is an isosbestic point of the oxyhemoglobin (HbO₂) and deoxyhemoglobin (Hbr) spectra, imaging at 570 nm is thought to emphasize changes in Hbt, which are proportional to changes in CBV under the assumption of constant hematocrit. The contribution of other wavelengths (attributable to finite width of the bandpass filter) and optical effects (such as scattering) to the imaging signal is a potential confounding factor, however. To estimate its importance, we compared the 570 nm and Hbt time courses with simultaneous OIS imaging and spectroscopy (Fig. 3). To ensure equal sampling, the imaging response was measured in the same area sampled by the spectroscopic fiber. Both time courses were normalized to their peak value ($-2.3 \pm 0.6 \times 10^{-2}$ for 570 nm and $1.0 \pm 0.1 \times 10^{-2}$ for Hbt;

mean \pm SE) to facilitate comparison. The 570 nm response latency (time at which the signal rose >2 SD above the baseline mean) was 1.0 sec after stimulation onset, and peak was at 2.75 sec. Similarly, Hbt started at 1.25 sec and peaked at 2.75 sec. We varied stimulation frequency and amplitude to modulate response size. Figure 3*B* shows a scatterplot of normalized 570 nm peak responses versus normalized Hbt peak responses, demonstrating a close correlation ($R^2 = 0.95$; $p < 10^{-6}$) between the signals. This strong linear relationship, along with similar latencies, time to peak values, and time course profiles, suggests that 570 nm imaging accurately reflects changes in Hbt. This result is consistent with previous reports that found similarities in spatial characteristics between 570 nm signals and CBV measured with fluorescent dyes (Frostig et al., 1990). Functional maps (570 nm) are, therefore, taken to represent CBV in this study.

Spatial characteristics

Figure 4 shows raw cortical images and functional activation maps using both absolute and statistical thresholding techniques for the three responses (monophasic, 570 nm; early and late phases, 610 nm) during C1 and C2 stimulation in a representative case. The 570 nm response has not only a larger magnitude than the 610 nm early phase but also a larger areal extent. Because of this spatial spread, 570 nm activation maps were thresholded at higher values. This figure highlights the difference between maps generated using absolute reflectance change thresholds and statistically determined thresholds. Absolute reflectance change maps (Fig. 4, left column) tend to accentuate vascular structures. Imaging at 610 nm emphasizes oxygenation changes, which predominate in draining veins. Venous signals were, therefore, readily apparent in 610 nm reflectance maps. Similarly, CBV-dominant 570 nm reflectance maps highlight arterioles, in which CBV fractional changes are greatest (Lee et al., 2001). Statistical maps, such as the ANOVA-based maps in this study (Fig. 4, right column), decrease vascular contributions to the functional signal. The magnitude of reflectance changes in vessels can be very large; therefore, vascular signals may linger in reflection change maps even with extensive trial averaging. Because they are not as consistent as parenchymal changes from trial to trial, however, their statistical significance is diminished relative to capillary bed changes. Statistical maps of both 570 and 610 nm early signals thus emphasize parenchymal changes (Fig. 4*D,F,H,J*). Late phase 610 nm signals are predominantly venous and, therefore, appeared similar using both analyses.

CBV spatial specificity

To determine the spatial specificity of 570 and 610 nm early phase responses, we compared statistically defined response areas with CO-stained barrels. Figure 5 shows vertical projections of C1 and C2 barrels aligned with peak 570 and 610 nm early phase responses. Peak 570 and 610 nm early response contours localized well to the barrel projections. The representations of the neighboring columns were distinct using both signals, suggesting that both could accurately resolve submillimeter structures.

To compare 570 and 610 nm early phase spatial specificity over all 10 subjects, we calculated the center of mass of the two peak response areas. The separation between the responses during C1 stimulation was $265 \pm 56 \mu\text{m}$ and during C2 stimulation was $234 \pm 49 \mu\text{m}$, less than the average barrel hollow diameter of $400\text{--}500 \mu\text{m}$ (Welker, 1971). The separation between the representations of C1 and C2 was $640 \pm 90 \mu\text{m}$ for the 570 nm response and $644 \pm 83 \mu\text{m}$ for the 610 nm early response, also consistent with the interbarrel distance of $\sim 600 \mu\text{m}$.

Vascular point spread

Hemodynamic response morphology is dictated by vascular architecture (Woolsey et al., 1996; Harrison et al., 2002) and may, therefore, extend beyond neuroanatomical boundaries (Narayan et al., 1995). To investigate the degree of vascular point spread, we compared response magnitudes over the principally stimulated barrel to those over the neighboring barrel (i.e., response to C1 stimulation over C1 vs over C2, and vice versa) in all subjects (Fig. 6). Both volume and early oxygenation responses spread beyond the principal barrel but were always significantly smaller in the neighboring barrel. The ratio of principal/neighboring barrel response size was 1.7 ± 0.3 for the 610 nm early phase (C1 and C2). The response ratio for 570 nm was 1.3 ± 0.1 for C1 and 1.4 ± 0.2 for C2. Differences between 610 and 570 nm response ratios were not significant ($p > 0.2$). Temporal characteristics of responses over the principal and neighboring column were nearly identical within our limits of temporal resolution for all phases of the hemodynamic response.

Spatial specificity and timing

Spatial specificity of the 610 nm early negative response has been confirmed with single-unit recordings (Masino et al., 1993; Shoham and Grinvald, 2001), voltage-sensitive dye imaging (Takashima et al., 2001; Slovín et al., 2002), and CO histology (Ts'o et al., 1990; Erinjeri and Woolsey, 2002). Although studies often assume that this response is analogous or even identical to the initial dip, the absorption coefficient of Hbr is only approximately three times that of HbO₂ in the 600 nm (red light) range. We performed simultaneous optical imaging and spectroscopy to determine the contribution of Hbr, HbO₂, and Hbt to the 610 nm signal over time (Fig. 7).

The first 0.75 sec of the 610 nm response followed Hbr, but whereas Hbr subsequently decreased, the 610 nm signal continued increasing for another 0.5 sec, peaking when Hbr was well below baseline. The first 0.75 sec of decreased reflectance at 610 nm thus derived from increased Hbr during the initial dip. The remainder of the response including the peak, however, occurred during the early hyperoxygenation and may be attributable to Hbt (volume) or HbO₂ rising more rapidly than the Hbr decrease. Across all animals and stimulus presentations (Fig. 7*B*), the spatially specific 610 nm early negative phase again incorporated both the initial dip and the early hyperoxygenation. Later hyperoxygenation changes comprised the 610 nm late phase and were spatially unspecific and vascular.

Discussion

Our results have two significant implications. First, they suggest the microvascular mechanisms that provide localized CBF changes also lead to localized changes in CBV. This specificity requires reducing the contribution of signals from large vessels, which are usually distant from the site of neuronal activity. Second, we found spatial specificity is predominantly a function of timing, observing early ($<2\text{--}3$ sec) changes, rather than specific response etiology, oxygenation or volume. We discuss these findings and their implications for functional brain mapping.

CBV spatial specificity

Our results indicate the CBV response is capable of resolving submillimeter functional architecture. CBV spatial specificity is comparable with that of the early oxygenation response, which may be considered a gold standard for high-resolution cortical mapping because of its well-documented specificity. The separa-

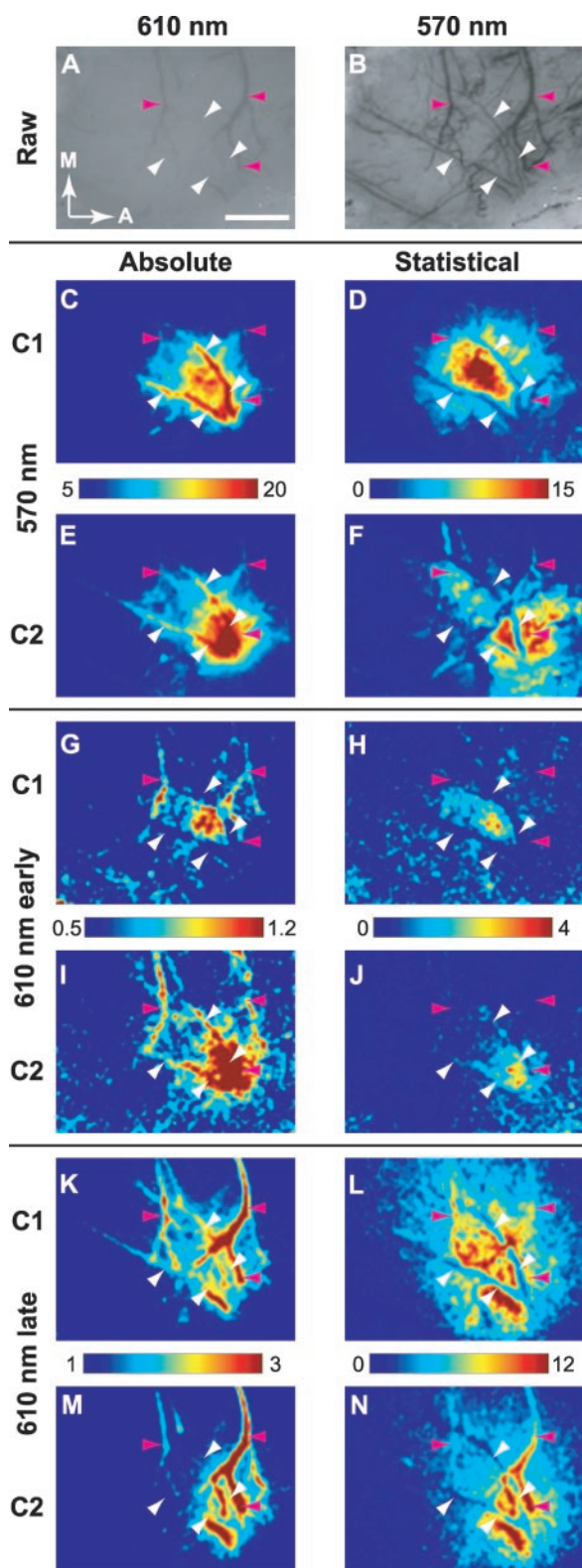


Figure 4. Absolute versus statistical thresholds. The response to C1 and C2 whisker stimulation is shown in a representative subject using absolute (left column) and statistical (right column) thresholds. Raw images of the cortex under 610 nm (A) and 570 nm (B) illumination were used to distinguish between surface veins (magenta arrowheads) and arterioles (white arrowheads), because both are equally visible at 570 nm (showing Hbt), but veins are dominant at 610 nm (emphasizing Hbr). C–F, The 570 nm response. G–J, The 610 nm early phase. K–N, The 610 nm late phase. The 570 nm absolute threshold maps (C,E) highlight activated arterioles showing large reflectance changes. This vascularity is reduced in the statistical map (D,F), which emphasizes changes in the capillary bed. Venous signals evident in absolute threshold

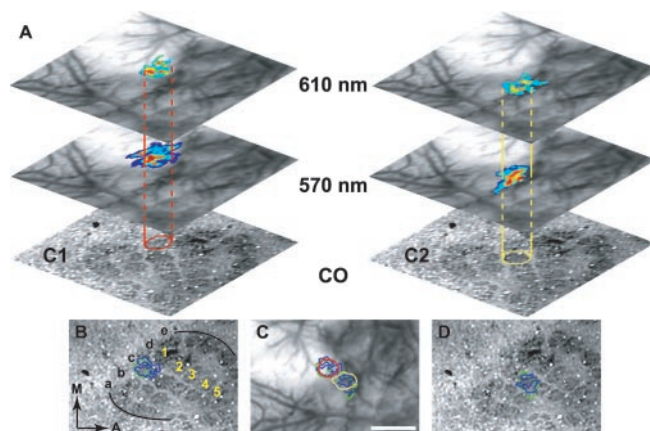


Figure 5. CBV spatial specificity and resolution. A, Vertical projection of the C1 (red) and C2 (yellow) barrel positions identified by CO staining. Barrel rows (a–e) and columns (1–5) are denoted in B. These projections pass through statistically thresholded peak 570 nm (CBV) and early 610 nm (early oxygenation) responses overlaid on raw images, demonstrating localization of both responses with underlying anatomy. Contours enclosing the peak response area for volume (green) and early oxygenation (blue) responses are overlaid on CO images of C1 (B) and C2 (D). C, Contours for both responses and whisker representations overlaid on barrel outlines demonstrate the spatially distinct representations of both hemodynamic responses. Thus, CBV responses resolve submillimeter functional architecture with accuracy comparable with early oxygenation responses. M, Medial; A, anterior. Scale bar, 1 mm.

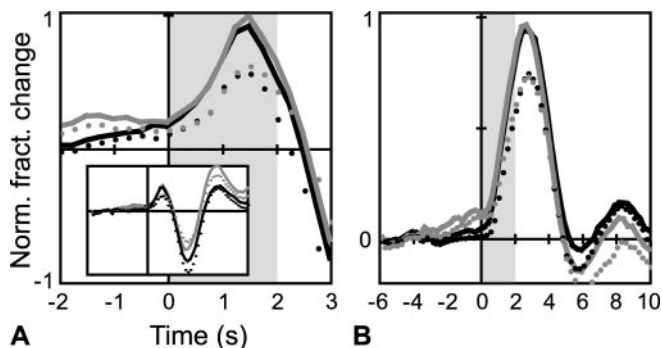


Figure 6. Vascular point spread. A, The 610 nm responses; the inset shows the full time course. B, The 570 nm responses. Black traces depict responses to C1 stimulation measured in C1 (solid) or C2 (dotted) peak contours; gray traces are analogous for C2. Time courses were normalized to peak values in the principal barrel. Both responses extended beyond the principal column but were smaller in the neighboring column. The difference in response ratio (see Results) between the two hemodynamic signals was not significant ($p > 0.2$). The temporal profile of changes was similar in the principal and neighboring barrel.

tion between the two was much less than a barrel column diameter

In contrast, previous studies reported that CBV changes exhibited excessive vascular overspill and were, therefore, inferior to early oxygenation changes for high-resolution mapping (Frostig et al., 1990; Malonek and Grinvald, 1996). We found both volume and early oxygenation responses extended beyond neuroanatomical boundaries of principal columns, probably because of horizontal intracortical connections (Petersen et al., 2003) or passive vascular processes (Narayan et al., 1995). The

maps of the 610 nm early phase (G,I) are similarly reduced in statistical maps (H,J). Because the 610 nm late phase is predominantly venous in origin, both absolute (K,M) and statistical (L,N) maps are biased toward large veins. Color bars apply to images above and below them. Absolute threshold scales represent fractional change ($\times 10^{-3}$), and statistical threshold scales represent the negative \log_{10} of the p value. M, Medial; A, anterior. Scale bar, 1 mm.

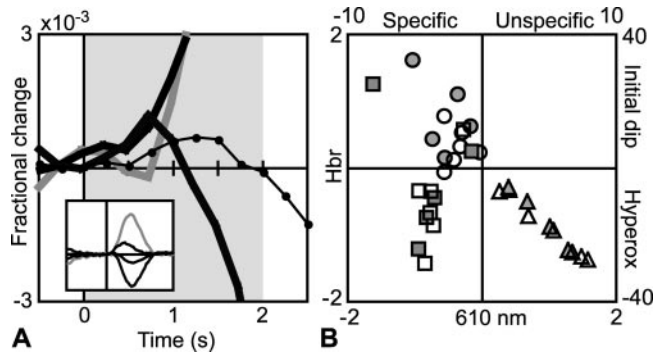


Figure 7. Etiology of spatially specific hemodynamic changes. *A*, Comparison between the 610 nm imaging time course (black circle, sign reversed for comparison) and concurrently measured Hbr (dark gray line), HbO₂ (light gray line), and Hbt (black line) for one stimulation condition (5 Hz, 0.8 mA; the inset shows the full time course). The 610 nm early phase follows Hbr for 0.75 sec but then continues to increase as Hbr rapidly decreases. The spatially specific 610 nm early negative response thus incorporates both the initial dip and early hyperoxygenation. *B*, Hbr versus the 610 nm magnitude for all stimulation conditions at 0.75 sec (white circle, amplitude variable stimulation; gray circle, frequency variable), 1.25 sec (white square, gray square), and 4.0 sec (white triangle, gray triangle). The left column of the grid (610 nm early phase) thus represents spatially specific responses, and the right column (610 nm late phase) represents spatially unspecific, vascular signals. The top row (Hbr, >0) represents the initial dip, and the bottom row (Hbr, <0) represents hyperoxygenation. Spatial specificity can, therefore, be achieved during the initial dip or early hyperoxygenation, whereas later hyperoxygenation changes are unspecific. The axis scale values are fractional change $\times 10^{-3}$. The left and bottom scale values represent data at 0.75 and 1.25 sec, and the right and top scale values represent 4.0 sec.

temporal characteristics of hemodynamic changes over principal and adjacent columns indicate the peripheral response develops concurrently with, although to a lesser degree than, the peak response and are additional evidence for vascular spread. The response ratio between principal and neighboring barrels suggests a slightly smaller point spread function for the early oxygenation change, but the difference was not significant. Duong et al. (2001) similarly found peak CBF signals were spatially precise, although the response spread into adjacent areas. The hemodynamic point spread will depend on vascular density, which may differ across species and cortex, but our results show this spread does not necessarily preclude CBV spatial specificity.

A recent study in mouse barrel cortex found 540 nm optical imaging (containing mixed contributions from oxygenation and volume etiologies) emphasized vascular structures and, therefore, was not as well confined to the active barrel as the 610 nm early response (Erinjeri and Woolsey, 2002). This vascularity is also evident in our data for both 570 nm (predominantly arteriolar) and 610 nm (predominantly venous) images analyzed using conventional subtraction-based approaches. This point underscores the importance of statistical mapping techniques, such as the ANOVA-based approach used here, that assign higher significance to pixels in the capillary bed that respond consistently across trials and lower significance to pixels in the vasculature, the variance across trials of which is high. A similar approach was used successfully by Hess et al. (2000).

CBV response specificity supports the functional relationship suggested between microvascular architecture and neural subunits (Cox et al., 1993; Woolsey et al., 1996). According to this hypothesis, individual cortical columns are supplied by one or a few penetrating arterioles, such that flow changes localize precisely to metabolic activity. Harrison et al. (2002) observed strictures on precapillary arteriole corrosion casts, suggesting the presence of pericytes capable of controlling vascular diameter.

The opportunity for fine flow control intimated by these structures is consistent with this theory, as is the columnar specificity demonstrated with CBF fMRI (Duong et al., 2001). Our results extend these findings to volume responses, which are also predominantly arteriolar (Lee et al., 2001). Therefore, spatially precise flow regulation also leads to precise volume changes.

Timing versus etiology

We found submillimeter specificity does not require focusing on the Hbr dip; it can also be achieved during the early part of the hyperoxygenation. Combined with the CBV results, these findings suggest hemodynamic spatial specificity arises not only from a spatially localized increase in neuronal oxidative metabolism but also from highly localized changes in hemoglobin oxygenation and blood volume that accompany metabolic changes. In other words, early changes in both oxygenation (regardless of direction) and volume are spatially coupled with neuronal activity. Therefore, in the context of high-resolution functional mapping, timing (imaging early signals) is more important than etiology (CBV or oxygenation signals).

Our results indicate optimal spatial precision occurs during the first few (two to three) seconds after stimulation onset, which under our conditions included the initial dip, CBV peak, and early hyperoxygenation. Studies using oxygenation-based signals report similar spatiotemporal characteristics. Duong et al. (2000) found single-condition maps of orientation columns in primary visual cortex could only be produced using the first two seconds of the blood oxygenation level-dependent (BOLD) fMRI initial dip (which was 4 sec in total duration). The latter 2 sec of the dip or later hyperoxygenation failed to identify complementary orientation structure. Using subsaturating stimuli, Menon and Goodyear (1999) generated ocular dominance maps (albeit with differential imaging) from the early part of the BOLD hyperoxygenation phase. Under their conditions, they did not detect the dip, but restricting the map to early time periods still revealed submillimeter functional architecture. These findings support the hypothesis that spatial specificity depends less on response direction than timing.

The above discussion and reports from the literature suggest the following sequence of events. Neuronal activation leads to a focal increase in oxygen consumption and relative tissue Hbr content within a few hundred milliseconds. Synaptic activity produces increased flow, confined to the active region by fine microvascular control structures. The rapidity and magnitude of the flow response, which are a function of several physiological variables, determine the degree to which Hbr increases before being washed out. A sluggish flow response would allow local Hbr to increase detectably, whereas a fast influx would quickly diminish Hbr. In either case, the oxygenation change may still be spatially specific for the first ~ 2 –3 sec. As a byproduct of the flow response, regional CBV also increases in the feeding arterioles, with a spatial distribution similar to that of CBF. Subsequent hyperoxygenation changes (>3 sec) propagate into draining veins away from the activation site. The exact timing of these hemodynamic processes will certainly depend on species, cortex, and physiological state, but the effects may be qualitatively similar.

This hypothesis generates testable predictions. By modulating the CBF response with pharmacological agents or alterations of physiological and anesthetic parameters, the relative size, timing, and spatial specificity of the Hbr increase or decrease can be assessed. Additionally, microvasculature CBF and CBV changes can be measured simultaneously to determine their activation-induced spatial and temporal coupling.

Implications for functional brain mapping

Perfusion-based functional imaging techniques offering submillimeter spatial resolution are extremely useful for studying columnar cortical organization. Several studies using conventional gradient echo T_2^* BOLD fMRI have demonstrated columnar structure, but the large venous contribution to this signal usually requires differential imaging, which presupposes common activation patterns and, therefore, limits its use (Menon et al., 1997; Menon and Goodyear, 1999; Cheng et al., 2001; Goodyear and Menon, 2001). Single-condition T_2 -weighted spin echo BOLD fMRI is less influenced by large veins and can resolve cortical columns, but the technique has low sensitivity and requires high (>4T) field strengths (Duong et al., 2002). Although BOLD fMRI using the initial dip has revealed submillimeter structure (Kim et al., 2000), the signal is elusive (Buxton, 2001), has a low CNR, and correlates weakly with the magnitude of changes in underlying neuronal activity (Sheth et al., 2003). CBF fMRI also demonstrates excellent spatial specificity and does not require high fields but suffers from a relatively low temporal resolution (Duong et al., 2001).

In comparison, CBV mapping offers several advantages. With appropriate analysis, single-condition CBV maps can resolve columnar functional organization. The increased number of arterioles supplying visual cortex columns in cats and primates compared with rat barrels suggests the capacity for even finer CBV resolution in the former. In this study, we found a fourfold and 1.5-fold increase in CNR of CBV responses relative to early and late oxygenation changes, respectively. Temporally restricting imaging to the early epoch (2–3 sec) further increases the CNR advantage of CBV over hyperoxygenation responses because CBV peaks earlier. CBV-weighted fMRI studies have similarly reported increased CNR and parenchymal specificity of CBV signals relative to BOLD signals (Mandeville and Marota, 1999). Furthermore, functional CBV changes correlate very closely with the magnitude of evoked neuronal activity (Sheth et al., 2003). This linear correspondence is desirable in any investigation using subtly graded stimuli and is essential for cognitive subtraction-based studies. Although CBV fMRI is currently limited to animal studies because of the requirement for exogenous contrast, human trials are underway (Sharma et al., 1999).

In conclusion, we found functional CBV changes were capable of resolving columnar organization. We hypothesize that spatial localization with submillimeter accuracy is a function of temporal characteristics (using early signals) more than specific response etiology (oxygenation or volume responses). Spatial specificity requires reducing the contribution of vascular signals, which can be achieved by observing early responses and statistical thresholding. The CBV response additionally offers high CNR and linear coupling with neuronal activity, making it an attractive candidate for high-resolution mapping.

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