

UCLA

UCLA Previously Published Works

Title

Evaluation of the Performance of the Cobas CT/NG Test for Use on the Cobas 6800/8800 Systems for Detection of Chlamydia trachomatis and Neisseria gonorrhoeae in Male and Female Urogenital Samples

Permalink

<https://escholarship.org/uc/item/7m7718qd>

Journal

Journal of Clinical Microbiology, 57(4)

ISSN

0095-1137

Authors

Van Der Pol, Barbara
Fife, Kenneth
Taylor, Stephanie N
[et al.](#)

Publication Date

2019-04-01

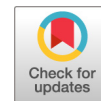
DOI

10.1128/jcm.01996-18


Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <https://creativecommons.org/licenses/by/4.0/>

Peer reviewed



Evaluation of the Performance of the Cobas CT/NG Test for Use on the Cobas 6800/8800 Systems for Detection of *Chlamydia trachomatis* and *Neisseria gonorrhoeae* in Male and Female Urogenital Samples

 Barbara Van Der Pol,^a Kenneth Fife,^b Stephanie N. Taylor,^c Melinda B. Nye,^d Steven E. Chavoustie,^e David L. Eisenberg,^f LaShonda Crane,^g Gregory Hirsch,^h Rodney Arcenas,ⁱ Elizabeth M. Marloweⁱ

^aUniversity of Alabama at Birmingham, Birmingham, Alabama, USA

^bIndiana University School of Medicine, Indianapolis, Indiana, USA

^cLouisiana State University, New Orleans, Louisiana, USA

^dLaboratory Corporation of America Holdings, Burlington, North Carolina, USA

^eHealthcare Clinical Data, Inc., North Miami, Florida, USA

^fPlanned Parenthood of St. Louis Region and Southwest Missouri, St. Louis, Missouri, USA

^gPlanned Parenthood Gulf Coast, Inc., Houston, Texas, USA

^hPlanned Parenthood of Northern, Central and Southern New Jersey, Inc., Morristown, New Jersey, USA

ⁱRoche Molecular Systems, Inc., Pleasanton, California, USA

ABSTRACT The clinical performance of the Cobas CT/NG assay on the Cobas 6800/8800 systems (Cobas) for the detection of *Chlamydia trachomatis* and *Neisseria gonorrhoeae* was established in a multisite, prospective collection study using male and female urogenital specimens; supportive data from archived specimens were also included. The results obtained with the Cobas assay were compared with the patient infected status derived from a combination of U.S. Food and Drug Administration-approved nucleic acid amplification tests to determine the sensitivity and specificity of detection from each sample type. The sensitivity of Cobas for the detection of *C. trachomatis* in female specimens was 95.6% (95% confidence interval [CI], 92.4% to 97.4%) for urine; 98.6% (95% CI, 95.2% to 99.6%) and 99.2% (95% CI, 95.4% to 99.9%) for clinician- and self-collected vaginal swab specimens, respectively; 93.3% (95% CI, 89.6% to 95.7%) for endocervical swabs; and 92.5% (95% CI, 88.7% to 95.1%) for cervical swab samples in PreservCyt. The specificity for the detection of *C. trachomatis* was $\geq 98.8\%$ for all female sample types. Sensitivity and specificity estimates of Cobas for the detection of *C. trachomatis* in male urine samples were 100% (96.8% to 100.0%) and 99.7% (95% CI, 99.2% to 99.9%), respectively. The sensitivity of Cobas for the detection of *N. gonorrhoeae* in female specimens was 94.8% (95% CI, 89.6% to 97.4%) for urine; 100.0% (95% CI, 87.9% to 100.0%) and 100.0% (95% CI, 87.9% to 100.0%) for clinician- and self-collected vaginal swab specimens, respectively; 97.0% (95% CI, 91.5% to 99.0%) for endocervical swabs; and 96.6% (95% CI, 90.6% to 98.8%) for cervical samples in PreservCyt; the specificity for all female sample types was $>99.0\%$. The sensitivity and specificity of Cobas for detecting *N. gonorrhoeae* in male urine were 100.0% (95% CI, 95.8% to 100.0%) and 99.5% (95% CI, 98.8% to 99.8%), respectively. Fully automated assays help fill the clinical need for a sensitive, high-throughput screening tool to aid public health efforts to control *C. trachomatis* and *N. gonorrhoeae* infections.

KEYWORDS *Chlamydia trachomatis*, *Neisseria gonorrhoeae*, PCR, Cobas CT/NG assay, genital infection, molecular diagnostics, nucleic acid amplification test, sexually transmitted infection

Citation Van Der Pol B, Fife K, Taylor SN, Nye MB, Chavoustie SE, Eisenberg DL, Crane L, Hirsch G, Arcenas R, Marlowe EM. 2019. Evaluation of the performance of the Cobas CT/NG test for use on the Cobas 6800/8800 systems for detection of *Chlamydia trachomatis* and *Neisseria gonorrhoeae* in male and female urogenital samples. J Clin Microbiol 57:e01996-18. <https://doi.org/10.1128/JCM.01996-18>.

Editor Erik Munson, Marquette University

Copyright © 2019 Van Der Pol et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Barbara Van Der Pol, bvanderpol@uabmc.edu.

Received 19 December 2018

Returned for modification 6 January 2019

Accepted 11 January 2019

Accepted manuscript posted online 16 January 2019

Published 28 March 2019

Chlamydia trachomatis and *Neisseria gonorrhoeae* are the leading bacterial causes of sexually transmitted infections (STIs) worldwide, with an estimated 131 million and 78 million new cases, respectively, each year (1). In the United States, approximately 1.7 million cases of *C. trachomatis* infection and 555,608 cases of *N. gonorrhoeae* infection were reported in 2017, representing increases of 4.7% and 18.5%, respectively, since 2015 (2). Both *C. trachomatis* and *N. gonorrhoeae* are major causes of pelvic inflammatory disease, ectopic pregnancy, and tubal factor infertility, and they facilitate the acquisition and transmission of human immunodeficiency virus (3–6). The importance of routine screening for *C. trachomatis* and *N. gonorrhoeae* to reduce the risk of these complications is well established (4, 7–10), and countries worldwide have implemented routine screening guidelines for populations with the highest risk of infection (11–15). Screening programs have been improved by expansion of acceptable sample types, such as self-collected vaginal swab samples, which have been shown to be equivalent in performance to clinician-collected samples and to be well accepted by patients (16, 17). The increasing use of preexposure prophylaxis (PrEP) for human immunodeficiency virus in high-risk individuals adds to the demand for routine STI screening programs (18–21); the Centers for Disease Control and Prevention (CDC) recommendation of quarterly STI screening for individuals on PrEP increases the demands of high-volume screening programs. These guidelines specifically include testing for *C. trachomatis* and *N. gonorrhoeae* for those sexually active adults that are prescribed PrEP. Furthermore, with shifting health care provision options and reduced funding for STI-specific health care clinics, offering STI screening in primary health care settings, such as general medicine, family planning, and obstetrics/gynecology clinics, is becoming more common. Diagnostic solutions that support the increasing need for STI screening across multiple populations and health care settings are clearly relevant to today's STI control efforts.

The CDC STI laboratory diagnostics recommendations (22) state that infection with *C. trachomatis* or *N. gonorrhoeae* should be detected using highly sensitive and specific nucleic acid amplification tests (NAATs). High-throughput NAATs can support national and community-based screening programs, leading to the earlier detection and treatment of infections, reduced transmission and prevalence, and reduced complications and associated health care costs (10, 22–25).

While automated STI screening solutions exist, a substantial need remains for fully automated, high-throughput molecular assays to address the increasing demands for STI screening services (26, 27). In this study, we evaluated the Cobas CT/NG test for use on the Cobas 6800/8800 systems (Cobas), which may address the needs of high-throughput laboratories (28, 29). The objective of this study was to evaluate the clinical performance of Cobas in urogenital samples from symptomatic and asymptomatic men and women.

MATERIALS AND METHODS

Patient population. Samples were prospectively collected from participants attending 8 geographically diverse family planning, obstetrics/gynecology, and STI clinics in the United States (Houston, TX; Miami, FL; Indianapolis, IN; New Orleans, LA; Seattle, WA; Morristown, NJ; St. Louis, MO; and Birmingham, AL). Eligible participants were aged ≥ 14 years, had been sexually active within the past 6 months, and were willing and able to provide written informed consent. Individuals were excluded if any of the following conditions were met: (i) they had previously been enrolled in the study; (ii) they had received antimicrobial therapy during the previous 21 days; (iii) they had used Replens (Lil'Drug Store Products, Inc., Cedar Rapids, IA), RepHresh odor-eliminating vaginal gel or RepHresh clean balance (Church & Dwight, Co, Inc., Princeton, NJ), or products containing metronidazole within the previous 3 days; (iv) they had a history of a full hysterectomy; or (v) they had a contraindication to Papanicolaou testing or cervical sampling. Participants were classified as symptomatic if they self-reported at least one of the following symptoms: dysuria; coital pain, postcoital bleeding; abnormal vaginal discharge or bleeding; pelvic, uterine, or ovarian pain; urethral discharge; testicular pain; and scrotal pain or swelling. All other participants were classified as asymptomatic. This study was conducted in compliance with the International Conference on Harmonisation good clinical practice guidelines and the regulations of the U.S. Food and Drug Administration (FDA). Institutional review board approval was obtained from each participating study site prior to the start of the study.

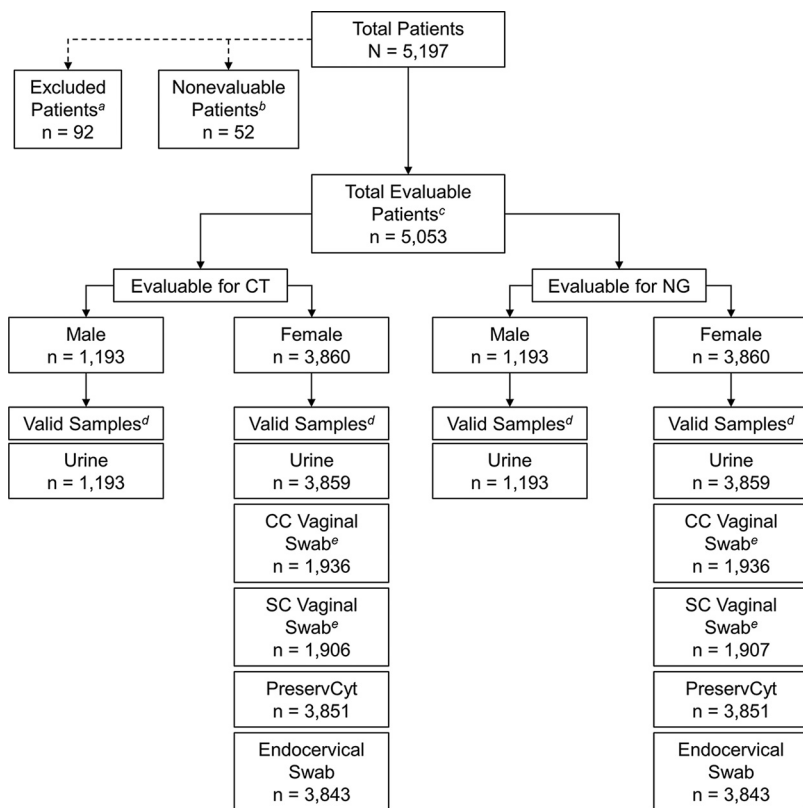


FIG 1 Patient disposition and specimen evaluability in the prospective population. ^a, reasons for exclusion included clinician decision ($n = 3$), the patient did not meet the eligibility criteria ($n = 21$), the patient was unwilling to provide informed consent ($n = 37$), pregnancy ($n = 1$), protocol deviation ($n = 14$), screen failure ($n = 1$), and withdrawal by the patient ($n = 15$); ^b, nonevaluable subjects were those without an assigned *C. trachomatis*/*N. gonorrhoeae* patient infected status (PIS; $n = 51$) or invalid/failed Cobas test results ($n = 1$). ^c, patients were considered evaluable if they had both a designated *C. trachomatis*/*N. gonorrhoeae* PIS, based on the prespecified PIS algorithm, and valid Cobas results for at least one sample type; ^d, valid samples were those with valid Cobas results and were included in the analysis; samples with missing, invalid, or failed Cobas results were excluded from the analysis; ^e, prospectively enrolled women were randomly assigned to the clinician-collected (CC) or self-collected (SC) vaginal swab specimen group. CT, *C. trachomatis*; NG, *N. gonorrhoeae*.

Due to the low prevalence of *N. gonorrhoeae* among women, a subset of archived samples collected in support of the U.S. clinical trial for the Cobas CT/NG assay on the Cobas 4800 system (Cobas 4800) (30–32) was retested on Cobas (6800/8800 systems), and the results were included in the analysis.

Specimen collection. The following specimens were collected (in order) from women: 1 first-catch urine (FCU) specimen; 1 self-collected (SC) vaginal swab specimen placed in Cobas PCR medium (from approximately 50% of participating women); up to 3 clinician-collected (CC) vaginal swab specimens; 1 endocervical swab specimen; and 1 cervical swab sample in PreservCyt solution, with collection of these samples alternating between the use of a spatula/cytobrush and a broom. The comparator assay vaginal swab collection order was randomized and was performed by clinicians. For women not randomized to self-obtain the Cobas vaginal swab specimen, clinicians collected this last among all of the vaginal swab samples collected. Archived sample types for the *N. gonorrhoeae* analyses included FCU, samples in PreservCyt, and endocervical swabs. One FCU sample was collected from each participating man.

Specimen testing. The Cobas CT/NG test is a qualitative nucleic acid amplification test performed on the Cobas 6800/8800 system; the system’s features have been previously described in detail (28, 29). Prospectively collected samples from each participant were tested with Cobas at 1 of 3 test sites. The archived samples were tested internally at Roche Molecular Systems, Pleasanton, CA. The FDA-approved NAATs used to determine patient infected status (PIS) included the BD ProbeTec CT Q^x and GC Q^x amplified DNA assays (CTQ/GCQ; BD, Sparks, MD), the Hologic Aptima Combo 2 CT/NG assay (AC2; Hologic Inc., San Diego, CA), and the Abbott m2000 RealTime CT/NG assay (m2000; Abbott Molecular, Des Plaines, IL). m2000 was used only for male urine testing to provide a third result. For all systems, samples were tested and data were interpreted according to the respective manufacturers’ instructions for use.

Patient infected status for *C. trachomatis* and *N. gonorrhoeae*. Four comparator test results were available for women: urine and vaginal swab specimen results from both CTQ/GCQ and AC2. Women were categorized as being infected with *C. trachomatis* or *N. gonorrhoeae* (PIS) if both NAATs (CTQ/GCQ

TABLE 1 Patient demographics and baseline characteristics

| Characteristic | Values for: | |
|--|--|--|
| | Prospective population ^a (n = 5,053) | Archived samples ^b (n = 295) |
| Age (yr) | | |
| Mean (SD) | 30.4 (9.8) | 27.5 (9.2) |
| Median (range) | 28 (14–79) | 25 (18–70) |
| No. (%) of subjects by sex | | |
| Female | 3,860 (76.4) | 295 (100) |
| Male | 1,193 (23.6) | 0 |
| No. (%) of subjects by race | | |
| Black/African American | 2,843 (49.1) | 159 (53.9) |
| White | 1,946 (38.5) | 116 (39.3) |
| Asian | 91 (1.8) | 6 (2.0) |
| American Indian/Alaska Native | 26 (0.5) | 0 |
| Native Hawaiian/Pacific Islander | 8 (0.2) | 1 (0.3) |
| Multiple/other | 323 (6.4) | 13 (4.4) |
| Not reported | 176 (3.5) | 0 |
| No. (%) of subjects by ethnicity | | |
| Not Hispanic/Latino | 3,541 (70.1) | 226 (76.6) |
| Hispanic/Latino | 1,379 (27.3) | 58 (19.7) |
| Unknown | 52 (1.0) | 2 (0.7) |
| Not reported | 81 (1.6) | 9 (3.1) |
| No. (%) of subjects by self-reported symptom status | | |
| Asymptomatic | 3,305 (65.4) | 145 (49.2) |
| Symptomatic | 1,747 (34.6) | 150 (50.8) |
| No. (%) of subjects by pregnancy status (women only) | | |
| Not pregnant | 3,811 (98.7) | 281 (95.3) |
| Pregnant | 43 (1.1) | 9 (3.1) |
| Unknown | 6 (0.2) | 5 (1.7) |
| No. (%) of subjects by clinic type | | |
| Family planning | 3,130 (61.9) | 134 (45.4) |
| STI ^c | 973 (19.3) | 120 (40.7) |
| Obstetrics/gynecology | 797 (15.8) | 41 (13.9) |
| Family planning/STI | 153 (3.0) | 0 |

^aParticipants enrolled in the prospective study population with designated infection status (infected or not infected) and final valid Cobas CT/NG test results.

^bArchived samples from study COB-CTNG-282 from female participants with a designated *N. gonorrhoeae* PIS (infected or not infected), final valid Cobas CT/NG test results, and an adequate sample volume for testing.

^cSTI, sexually transmitted infection.

and AC2) generated positive results in at least one sample type (urine and/or vaginal swab specimens). Endocervical swab and PreservCyt specimens were tested only with Cobas and were therefore not included in the calculation of the PIS. The endocervical and PreservCyt test results were compared with the PIS of women. Men were classified as infected if 2 of 3 NAATs (CTQ/GCQ, AC2, and m2000) gave positive results using FCU. Each NAAT was also evaluated using a rotating PIS analysis, constructed from the results obtained from the other NAATs used in the study, including Cobas results, based on the above-mentioned criteria for defining PIS. The rotating PIS analyses incorporated the results from the prospectively collected specimens only.

Data analysis and interpretation of results. All data analyses were performed using SAS/STAT software. The sensitivity, specificity, positive predictive value (PPV), and negative predictive value (NPV) of Cobas were calculated overall and by symptom status for detection of *C. trachomatis* and *N. gonorrhoeae* compared with the PIS for each sex (see Table S1 in the supplemental material). Specimens with invalid test results and/or for which further retests could not be performed due to a lack of sample volume were excluded from the data analyses.

The sensitivity, specificity, PPV, and NPV of Cobas were estimated separately for each sample type and symptom status. Two-sided 95% confidence intervals (CIs) were calculated using the Wilson method for proportions. Positive percent agreement (PPA), negative percent agreement (NPA), and overall percent agreement (OPA) were assessed by pairwise comparison between Cobas and the comparator

TABLE 2 Clinical performance of Cobas versus PIS by symptomatic status for *C. trachomatis*^a

| Sex, sample type | Symptom status | Total | % sensitivity (no. of samples positive/ total no. tested) | 95% CI for sensitivity | % specificity (no. of samples positive/ total no. tested) | 95% CI | Prevalence (%) | PPV (%) | NPV (%) |
|-------------------|----------------|--------|---|------------------------|---|------------|----------------|---------|---------|
| Women | | | | | | | | | |
| Urine | Symptomatic | 1,441 | 96.0 (119/124) | 90.9–98.3 | 99.8 (1,315/1,317) | 99.4–100.0 | 8.6 | 98.3 | 99.6 |
| | Asymptomatic | 2,418 | 95.2 (140/147) | 90.5–97.7 | 99.6 (2,262/2,271) | 99.2–99.8 | 6.1 | 94.0 | 99.7 |
| | Overall | 3,859 | 95.6 (259/271) | 92.4–97.4 | 99.7 (3,577/3,588) | 99.5–99.8 | 7.0 | 95.9 | 99.7 |
| CC vaginal swab | Symptomatic | 711 | 100.0 (63/63) | 94.3–100.0 | 99.2 (643/648) | 98.2–99.7 | 8.9 | 92.6 | 100.0 |
| | Asymptomatic | 1,225 | 97.6 (83/85) | 91.8–99.4 | 99.0 (1,129/1,140) | 98.3–99.5 | 6.9 | 88.3 | 99.8 |
| | Overall | 1,936 | 98.6 (146/148) | 95.2–99.6 | 99.1 (1,772/1,788) | 98.6–99.4 | 7.6 | 90.1 | 99.9 |
| SC vaginal swab | Symptomatic | 720 | 100.0 (59/59) | 93.9–100.0 | 98.8 (653/661) | 97.6–99.4 | 8.2 | 88.1 | 100.0 |
| | Asymptomatic | 1,186 | 98.4 (60/61) | 91.3–99.7 | 99.2 (1,116/1,125) | 98.5–99.6 | 5.1 | 87.0 | 99.9 |
| | Overall | 1,906 | 99.2 (119/120) | 95.4–99.9 | 99.0 (1,769/1,786) | 98.5–99.4 | 6.3 | 87.5 | 99.9 |
| PreservCyt | Symptomatic | 1,438 | 95.1 (116/122) | 89.7–97.7 | 99.5 (1,309/1,316) | 98.9–99.7 | 8.5 | 94.3 | 99.5 |
| | Asymptomatic | 2,413 | 90.3 (131/145) | 84.4–94.2 | 99.7 (2,261/2,268) | 99.4–99.9 | 6.0 | 94.9 | 99.4 |
| | Overall | 3,851 | 92.5 (247/267) | 88.7–95.1 | 99.6 (3,570/3,584) | 99.3–99.8 | 6.9 | 94.6 | 99.4 |
| Endocervical swab | Symptomatic | 1,433 | 95.9 (116/121) | 90.7–98.2 | 99.1 (1,300/1,312) | 98.4–99.5 | 8.4 | 90.6 | 99.6 |
| | Asymptomatic | 2,410 | 91.1 (133/146) | 85.4–94.7 | 99.5 (2,253/2,264) | 99.1–99.7 | 6.1 | 92.4 | 99.4 |
| | Overall | 3,843 | 93.3 (249/267) | 89.6–95.7 | 99.4 (3,553/3,576) | 99.0–99.6 | 6.9 | 91.5 | 99.5 |
| Men, urine | Symptomatic | 305 | 100.0 (63/63) | 94.3–100.0 | 99.6 (241/242) | 97.7–99.9 | 20.7 | 98.4 | 100.0 |
| | Asymptomatic | 887 | 100.0 (55/55) | 93.5–100.0 | 99.8 (830/832) | 99.1–99.9 | 6.2 | 96.5 | 100.0 |
| | Overall | 1,192 | 100.0 (118/118) | 96.8–100.0 | 99.7 (1,071/1,074) | 99.2–99.9 | 9.9 | 97.5 | 100.0 |
| Overall | | 16,587 | 95.5 (1,138/1,191) | 94.2–96.6 | 99.5 (15,312/15,396) | 99.3–99.6 | 7.2 | 93.1 | 99.7 |

^aCC, clinician collected; Cobas, Cobas CT/NG test for use on the Cobas 6800/8800 systems; NPV, negative predictive value; PIS, patient infected status; PPV, positive predictive value; PreservCyt, cervical sample collected in PreservCyt cytology medium; SC, self-collected.

tests with matched clinical specimen types (i.e., the same sample type from women). For the comparison of performance between assays, the Fisher exact test was performed, and *P* values are presented.

RESULTS

Patient population. A total of 5,197 participants were enrolled in the prospective population; of these, 92 were excluded because they did not meet the inclusion criteria and 52 were excluded due to invalid test results and/or the inability to retest specimens (Fig. 1). A total of 5,053 participants were evaluable and included in the analyses. Archived samples from an additional 295 women were included in the *N. gonorrhoeae* analyses. Patient demographics and baseline characteristics are shown in Table 1. Overall, there were 16,587 *C. trachomatis* test results (prospective population) and 16,959 *N. gonorrhoeae* (prospective and archived population) test results that were included in the analyses.

Chlamydia trachomatis. Of the 3,860 evaluable, prospectively enrolled women, 3,834 had valid urine sample results and 3,820 had valid vaginal swab sample results with Cobas, AC2, and CTQ/GCQ; 1,180 of 1,193 evaluable, prospectively enrolled men had valid urine sample results with Cobas, AC2, CTQ/GCQ, and m2000. In total, 271 (7.0%) of 3,860 women in the prospective population were defined as *C. trachomatis* infected based on the PIS. Sensitivity and specificity by sample type are shown in Table 2. Overall the sensitivity and specificity of *C. trachomatis* detection for all female sample types were ≥92.5 and ≥98.8%, respectively. A total of 118 (9.9%) of 1,193 men were defined as being infected with *C. trachomatis*, with sensitivity and specificity estimates for Cobas of 100% (95% CI, 96.8% to 100.0%) and 99.7% (95% CI, 99.2% to 99.9%), respectively. Venn diagrams comparing the *C. trachomatis*-positive results across all NAATs, regardless of PIS, in female urine, female vaginal swab, and male urine samples are shown in Fig. 2. The OPA between Cobas and each predicate NAAT for the detection of *C. trachomatis* in all sample types was >98%, with a PPA of >92% and an NPA of >99%.

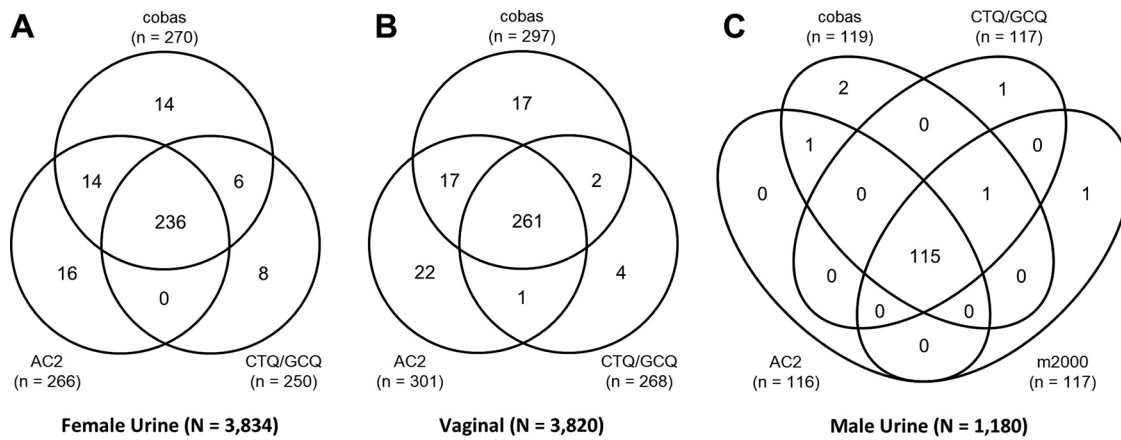


FIG 2 Venn diagrams comparing *C. trachomatis*-positive results across NAATs with urine samples from women (A), vaginal swab samples (B), and urine samples from men (C).

Overall, there were 1,936 CC and 1,906 SC prospective vaginal swab specimens. The Cobas sensitivity performances for the detection of *C. trachomatis* from CC and SC vaginal swabs were very similar at 98.6% (95% CI, 95.2% to 99.6%) and 99.2% (95% CI, 95.4% to 99.9%), respectively. The specificity performances were also very similar at 99.1% (95% CI, 98.6% to 99.4%) and 99.0% (95% CI, 98.5% to 99.4%), respectively (Table 2).

The sensitivity and specificity for *C. trachomatis* detection with Cobas, AC2, CTQ/GCQ, and m2000 (male urine only) were estimated using a rotating PIS analyses that incorporated the results obtained with prospectively collected female urine, female vaginal swab, and male urine specimens. The sensitivity of the Cobas assay was significantly higher than that of CTQ/GCQ for female urine ($P < 0.001$) and vaginal swabs ($P = 0.006$) (Table 3). The specificities of all assays for the detection of *C. trachomatis* in both female urine and vaginal swab samples were $\geq 99.0\%$ across the rotating PIS assay combinations (Table 3). The sensitivities and specificities for the detection of *C. trachomatis* in male urine samples were comparable with Cobas, AC2, CTQ/GCQ, and m2000 ($\geq 98.3\%$ and $\geq 99.7\%$, respectively; Table 4).

***Neisseria gonorrhoeae*.** Of the 3,860 evaluable, prospectively enrolled women, 3,835 had valid urine sample results and 3,826 had valid vaginal swab sample results on Cobas, AC2, and CTQ/GCQ; 1,180 of 1,193 evaluable, prospectively enrolled men had valid urine sample results on Cobas, AC2, CTQ/GCQ, and m2000. Of the 3,860 prospectively enrolled women, 57 (1.5%) were considered *N. gonorrhoeae* infected based on the PIS; of the 295 women who contributed archived specimens, 82 were *N. gonorrhoeae* infected. Sensitivity and specificity by sample type are shown in Table 5. Overall, among the prospective and archived female specimens, the sensitivity and specificity of Cobas

TABLE 3 Rotating PIS analysis of sensitivity and specificity of NAATs for detecting *Chlamydia trachomatis* in urine and clinician-collected vaginal swab samples from women^a

| Comparison, sample type | No. of samples tested | No. of <i>C. trachomatis</i> PIS-positive samples tested/total no. | % sensitivity (95% CI) | P value for comparison with Cobas sensitivity | No. of <i>C. trachomatis</i> PIS-negative samples tested/total no. | % specificity (95% CI) | P value for comparison with Cobas specificity |
|---------------------------------------|-----------------------|--|------------------------|---|--|------------------------|---|
| AC2 vs PIS (Cobas and CTQ/GCQ) | | | | | | | |
| Urine from women | 3,839 | 254/272 | 93.4 (89.8–95.8) | 0.348 | 3,555/3,567 | 99.7 (99.4–99.8) | 0.838 |
| Vaginal swab (clinician) | 3,836 | 266/270 | 98.5 (96.3–99.4) | 1.000 | 3,531/3,566 | 99.0 (98.6–99.3) | 0.882 |
| CTQ/GCQ vs PIS (Cobas and AC2) | | | | | | | |
| Urine from women | 3,852 | 243/285 | 85.3 (80.7–88.9) | <0.001 | 3,560/3,567 | 99.8 (99.6–99.9) | 0.480 |
| Vaginal swab (CC) | 3,825 | 262/283 | 92.6 (88.9–95.1) | 0.006 | 3,536/3,542 | 99.8 (99.6–99.9) | <0.001 |

^aAC2, Hologic Aptima Combo 2 CT/NG assay; CC, clinician collected; Cobas, Cobas CT/NG test for use on the Cobas 6800/8800 systems; CTQ/GCQ, BD ProbeTec CT Q^x and GC Q^x amplified DNA assays; NAAT, nucleic acid amplification test; PIS, patient infected status.

TABLE 4 Rotating PIS analysis of sensitivity and specificity of NAATs for detecting *Chlamydia trachomatis* in urine samples from men^a

| Comparison | No. of samples tested | No. of <i>C. trachomatis</i> PIS-positive samples tested/total no. | % sensitivity (95% CI) | No. of <i>C. trachomatis</i> PIS-negative samples tested/total no. | % specificity (95% CI) |
|--|-----------------------|--|------------------------|--|------------------------|
| AC2 vs PIS (Cobas, CTQ/GCQ, and m2000) | 1,187 | 117/119 | 98.3 (94.1–99.5) | 1,067/1,068 | 99.9 (99.5–100.0) |
| CTQ/GCQ vs PIS (Cobas, AC2, and m2000) | 1,187 | 116/117 | 99.1 (95.3–99.8) | 1,069/1,070 | 99.9 (99.5–100.0) |
| m2000 vs PIS (Cobas, AC2, and CTQ/GCQ) | 1,189 | 118/119 | 99.2 (95.4–99.9) | 1,069/1,070 | 99.9 (99.5–100.0) |

^aAC2, Hologic Aptima Combo 2 CT/NG assay; Cobas, Cobas CT/NG test for use on the Cobas 6800/8800 systems; CTQ/GCQ, BD ProbeTec CT Q^x and GC Q^x amplified DNA assays; m2000, Abbot m2000 RealTime CT/NG assay; NAAT, nucleic acid amplification test; PIS, patient infected status.

for detecting *N. gonorrhoeae* were ≥94.8% and ≥99.7%, respectively. A total of 87 (7.3%) of 1,193 men were considered *N. gonorrhoeae* infected based on the PIS, with sensitivity and specificity estimates of Cobas for the detection of *N. gonorrhoeae* in male urine of 100.0% (95% CI, 95.8% to 100.0%) and 99.5% (95% CI, 98.8% to 99.8%), respectively. Venn diagrams comparing the *N. gonorrhoeae*-positive results across all NAATs, regardless of PIS, in female urine, female vaginal swab, and male urine samples are shown in Fig. 3. The OPA between Cobas and each predicate NAAT for *N. gonorrhoeae* in all sample types was >98%, with a PPA of >73% and an NPA of >99%.

Overall, there were 1,936 CC and 1,906 SC prospective vaginal swab samples. The Cobas sensitivity and specificity performances for the detection of *N. gonorrhoeae* from CC and SC vaginal swab specimens were the same (100% and 99.7%, respectively) (Table 5).

The rotating PIS analysis results for prospectively collected female urine samples and CC vaginal swabs are shown in Table 6. The sensitivity of Cobas for *N. gonorrhoeae* detection in female urine samples was higher than the sensitivities of AC2 (*P* = 0.012) and CTQ/GCQ (*P* = 0.001); all 3 assays showed specificities of ≥99.6% for female urine specimens. The sensitivity of detecting *N. gonorrhoeae* in prospectively collected CC vaginal swabs was also higher with Cobas than with AC2 and CTQ/GCQ, but the

TABLE 5 Clinical performance of Cobas versus PIS by symptomatic status for *N. gonorrhoeae*^a

| Sex, sample type | Symptom status | Total | % sensitivity (no. of samples positive/total no. tested) | 95% CI for sensitivity | % specificity (no. of samples positive/total no. tested) | 95% CI for specificity | Prevalence (%) | PPV (%) | NPV (%) |
|-------------------|----------------|--------|--|------------------------|--|------------------------|----------------|---------|---------|
| Women | | | | | | | | | |
| Urine | Symptomatic | 1,535 | 96.7 (59/61) | 88.8–99.1 | 99.8 (1,471/1,474) | 99.4–99.9 | 4.0 | 95.2 | 99.9 |
| | Asymptomatic | 2,519 | 93.2 (68/73) | 84.9–97.0 | 100.0 (2,446/2,446) | 99.8–100.0 | 2.9 | 100.0 | 99.8 |
| | Overall | 4,054 | 94.8 (127/134) | 89.6–97.4 | 99.9 (3,917/3,920) | 99.8–100.0 | 3.3 | 97.7 | 99.8 |
| CC vaginal swab | Symptomatic | 711 | 100.0 (11/11) | 74.1–100.0 | 99.7 (698/700) | 99.0–99.9 | 1.5 | 84.6 | 100.0 |
| | Asymptomatic | 1,225 | 100.0 (17/17) | 81.6–100.0 | 99.8 (1,205/1,208) | 99.3–99.9 | 1.4 | 85.0 | 100.0 |
| | Overall | 1,936 | 100 (28/28) | 87.9–100.0 | 99.7 (1,903/1,908) | 99.4–99.9 | 1.4 | 84.8 | 100.0 |
| SC vaginal swab | Symptomatic | 720 | 100.0 (14/14) | 78.5–100.0 | 99.7 (704/706) | 99.0–99.9 | 1.9 | 87.5 | 100.0 |
| | Asymptomatic | 1,187 | 100.0 (14/14) | 78.5–100.0 | 99.7 (1,169/1,173) | 99.1–99.9 | 1.2 | 77.8 | 100.0 |
| | Overall | 1,907 | 100.0 (28/28) | 87.9–100.0 | 99.7 (1,873/1,879) | 99.3–99.9 | 1.5 | 82.4 | 100.0 |
| PreservCyt | Symptomatic | 1,486 | 97.9 (44/48) | 89.1–99.6 | 99.9 (1,437/1,438) | 99.6–100.0 | 3.2 | 97.9 | 99.9 |
| | Asymptomatic | 2,436 | 95.1 (39/41) | 83.9–98.7 | 100.0 (2,394/2,395) | 99.8–100.0 | 1.7 | 97.5 | 99.9 |
| | Overall | 3,922 | 96.6 (86/89) | 90.6–98.8 | 99.9 (3,831/3,833) | 99.8–100.0 | 2.3 | 97.7 | 99.9 |
| Endocervical swab | Symptomatic | 1,484 | 100.0 (45/45) | 92.1–100.0 | 99.9 (1,438/1,439) | 99.6–100.0 | 3.0 | 97.8 | 100.0 |
| | Asymptomatic | 2,464 | 94.5 (52/55) | 85.1–98.1 | 100.0 (2,408/2,409) | 99.8–100.0 | 2.2 | 98.1 | 99.9 |
| | Overall | 3,948 | 97.0 (97/100) | 91.5–99.0 | 99.9 (3,846/3,848) | 99.8–100.0 | 2.5 | 98.0 | 99.9 |
| Men, urine | Symptomatic | 305 | 100.0 (82/82) | 95.5–100.0 | 98.7 (220/223) | 96.1–99.5 | 26.9 | 96.5 | 100.0 |
| | Asymptomatic | 887 | 100.0 (5/5) | 56.6–100.0 | 99.7 (879/882) | 99.0–99.9 | 0.6 | 62.5 | 100.0 |
| | Overall | 1,193 | 100.0 (87/87) | 95.8–100.0 | 99.5 (1,099/1,105) | 98.8–99.8 | 7.3 | 93.5 | 100.0 |
| Overall | | 16,959 | 97.2 (453/466) | 95.3–98.4 | 99.9 (16,469/16,493) | 99.8–99.9 | 2.7 | 95.0 | 99.9 |

^aCC, clinician collected; Cobas, Cobas CT/NG test for use on the Cobas 6800/8800 systems; NPV, negative predictive value; PIS, patient infected status; PPV, positive predictive value; PreservCyt, cervical sample collected in PreservCyt cytology medium; SC, self-collected.

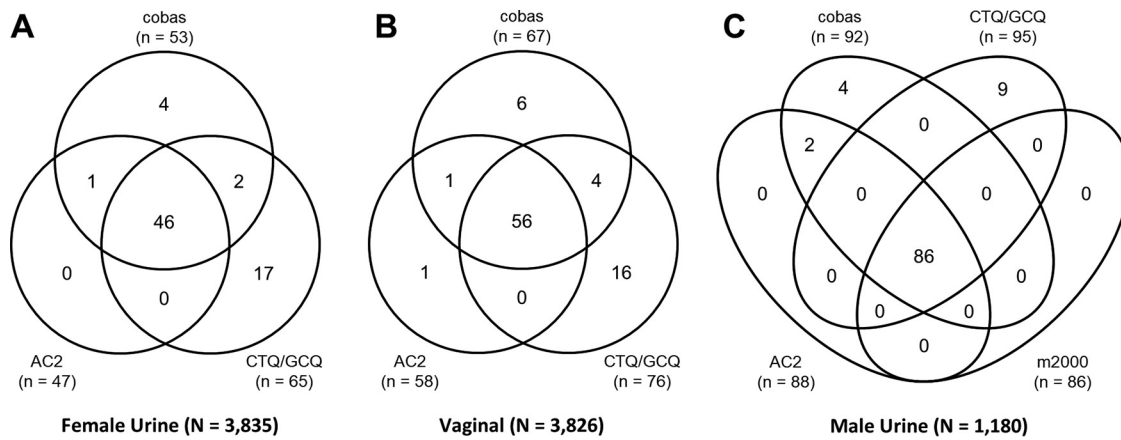


FIG 3 Venn diagrams comparing *N. gonorrhoeae*-positive results across NAATs with urine samples from women (A), vaginal swab samples (B), and urine samples from men (C) (prospective population).

difference was not statistically significant ($P = 1.000$). All 3 assays showed specificities of $\geq 99.5\%$ for vaginal swabs. The sensitivities and specificities for *N. gonorrhoeae* detection in male urine samples were comparable to those of Cobas, AC2, CTQ/GTQ, and m2000 ($\geq 97.7\%$ and $\geq 99.2\%$, respectively; Table 7).

DISCUSSION

This multicenter study evaluated the clinical performance of Cobas at detecting *C. trachomatis* and *N. gonorrhoeae* in urogenital samples from men and women. Based on a PIS determined using multiple FDA-approved NAATs, the overall sensitivity and specificity of Cobas for the detection of *C. trachomatis* were $>95\%$ and $>99\%$, respectively, in female urogenital samples and 100% and $>99\%$, respectively, in male urine samples. The overall sensitivity and specificity for detecting *N. gonorrhoeae* across all female urogenital sample types were $>96\%$ and $>99\%$, respectively, and those for male urine samples were 100% and $>95\%$, respectively. Sensitivity and specificity were comparable between symptomatic and asymptomatic individuals for all sample types and were generally consistent across the 9 collection sites. The Cobas assay had a high OPA ($>98\%$) with all predicate FDA-approved NAATs tested for their ability to detect *C. trachomatis* and *N. gonorrhoeae* in vaginal swab samples and in urine samples from both women and men.

Previous studies with multiple NAATs have shown that the presence or absence of symptoms does not significantly affect test performance (31, 33, 34). Consistent with these studies, the sensitivities and specificities of Cobas for detecting *C. trachomatis* and *N. gonorrhoeae* in our study were comparable between symptomatic and asymptomatic patients. Additionally, Cobas showed a high sensitivity and specificity, regard-

TABLE 6 Rotating PIS analysis of sensitivity and specificity of NAATs for detecting *Neisseria gonorrhoeae* in urine and clinician-collected vaginal swab samples from women^a

| Comparison, sample type | No. of samples tested | No. of <i>N. gonorrhoeae</i> PIS-positive samples tested/total no. | Sensitivity (95% CI) | P value for comparison with Cobas sensitivity | No. of <i>N. gonorrhoeae</i> PIS-negative samples tested/total no. | Specificity (95% CI) | P value for comparison with Cobas specificity |
|---------------------------------------|-----------------------|--|----------------------|---|--|----------------------|---|
| AC2 vs PIS (Cobas and CTQ/GCQ) | | | | | | | |
| Urine from women | 3,853 | 49/59 | 83.1 (71.5–90.5) | 0.012 | 3,778/3,794 | 99.6 (99.3–99.7) | 0.002 |
| Vaginal swab (CC) | 3,830 | 56/58 | 96.6 (88.3–99.0) | 1.000 | 3,752/3,772 | 99.5 (99.2–99.7) | 0.203 |
| CTQ/GCQ vs PIS (Cobas and AC2) | | | | | | | |
| Urine from women | 3,839 | 46/60 | 76.7 (64.6–85.6) | 0.001 | 3,778/3,779 | 100 (99.9–100) | 0.625 |
| Vaginal swab (CC) | 3,839 | 56/60 | 93.3 (84.1–97.4) | 0.302 | 3,777/3,779 | 99.9 (99.8–100) | 0.046 |

^aAC2, Hologic APTIMA Combo 2 CT/NG Assay; CC, clinician collected; Cobas, Cobas CT/NG test for use on the Cobas 6800/8800 systems; CTQ/GCQ, BD ProbeTec CT Q^x and GC Q^x amplified DNA assays; NAAT, nucleic acid amplification test; PIS, patient infected status.

TABLE 7 Rotating PIS analysis of sensitivity and specificity of NAATs for detecting *Neisseria gonorrhoeae* in urine samples from men^a

| Comparison | No. of samples tested | No. of <i>N. gonorrhoeae</i> PIS-positive samples tested/total no. | % sensitivity (95% CI) | No. of <i>N. gonorrhoeae</i> PIS-negative samples tested/ total no. | % specificity (95% CI) |
|--|-----------------------|--|------------------------|---|------------------------|
| AC2 vs PIS (Cobas, CTQ/GCQ, and m2000) | 1,187 | 86/88 | 97.7 (92.1–99.4) | 1,090/1,099 | 99.2 (98.5–99.6) |
| CTQ/GCQ vs PIS (Cobas, AC2, and m2000) | 1,187 | 87/87 | 100 (95.8–100.0) | 1,098/1,100 | 99.8 (99.3–100.0) |
| m2000 vs PIS (Cobas, AC2, and CTQ/GCQ) | 1,189 | 87/89 | 97.8 (92.2–99.4) | 1,100/1,100 | 100 (99.7–100.0) |

^aAC2, Gen-Probe APTIMA Combo 2 CT/NG assay; Cobas, Cobas CT/NG test for use on the Cobas 6800/8800 systems; CTQ/GCQ, BD ProbeTec CT Q^x and GC Q^x amplified DNA assays; m2000, Abbot m2000 RealTime CT/NG assay; NAAT, nucleic acid amplification test; PIS, patient infected status.

less of symptomatic status, with a variety of sample types routinely used in clinical practice, including the preferred and minimally invasive vaginal swabs and male urine samples, which are in line with the specimen types recommended by the CDC and European guidelines for *C. trachomatis* and *N. gonorrhoeae* detection (11, 12, 22). Of note was the comparable performance of self-collected vaginal swab specimens and clinician-collected vaginal swab specimens. With a performance equivalent to that for clinician-collected specimens, self-collection options decrease barriers to routine STI screening and increase patient satisfaction, thus increasing the likelihood of getting tested and treated in a timely fashion and reducing overall rates of STIs. The ability of Cobas to detect *C. trachomatis* and *N. gonorrhoeae* in multiple sample types from both symptomatic and asymptomatic patients indicates that Cobas is suitable for both screening and diagnostic efforts.

We used a rotating PIS, in which 1 of the reference tests served as the investigational device, while the remaining 2 tests (for women) or 3 tests (for men) served as the reference tests in order to construct the PIS. This type of analysis can be helpful in understanding how each of the reference tests used within this study could affect the estimates of assay performance. The rotating PIS also provides a means of showing how each test compares with the others. In this analysis, the sensitivities of Cobas and AC2 for *C. trachomatis* detection were comparable with both female urine and clinician-collected vaginal swab samples, whereas CTQ/GTQ had a lower sensitivity with both sample types; specificities were comparable across assays. The sensitivity for detecting *N. gonorrhoeae* in female urine and vaginal swab samples was the highest for Cobas, followed by CTQ/GTQ and AC2; specificities were comparable across the assays. The sensitivities and specificities for detecting both *C. trachomatis* and *N. gonorrhoeae* in male urine samples were >97% and >99%, respectively, for all assays.

Conclusion. In this multicenter clinical study, Cobas showed a high sensitivity, specificity, PPV, and NPV for the direct, qualitative detection of *C. trachomatis* and *N. gonorrhoeae* in a variety of urogenital samples from symptomatic and asymptomatic men and women. The Cobas CT/NG assay for use on the Cobas 6800/8800 systems helps fill the clinical need for a sensitive, high-throughput screening solution to aid in the diagnosis of *C. trachomatis* and *N. gonorrhoeae* infections and supports public health efforts to control these infections.

SUPPLEMENTAL MATERIAL

Supplemental material for this article may be found at <https://doi.org/10.1128/JCM.01996-18>.

SUPPLEMENTAL FILE 1, PDF file, 0.1 MB.

ACKNOWLEDGMENTS

We are grateful to Laurel Kuehl for participating in data collection.

This study was funded by Roche Molecular Systems, Inc., Pleasanton, CA.

B. Van Der Pol discloses grants from BD Diagnostics, Binx Health, Hologic, Rheonix, Roche Molecular Systems, and SpeeDx and personal fees from Abbott Molecular, BioFire Diagnostics, Binx Health, and SpeeDx. K. Fife has received research grants from BD Diagnostics, Hologic, Rheonix, and Roche Molecular Systems. S. N. Taylor has received research grants from Beckman Coulter, Becton, Dickinson, GlaxoSmithKline,

Hologic, Melinta, and Roche Molecular Diagnostics and personal fees from GlaxoSmith-Kline. M. B. Nye received compensation for clinical trial services rendered from the Laboratory Corporation of America. S. E. Chavoustie received compensation for clinical trial services rendered from Health Care Clinical Data, Inc. D. L. Eisenberg receives research funding and serves as scientific advisor to Medicines360, is a trainer for Merck's contraceptive implant, and is a consultant for Femasys. L. Crane and G. Hirsch received institutional support for this study from Roche Molecular Systems. R. Arcenas and E. M. Marlow are employees of Roche Molecular Systems, Inc. Planned Parenthood received compensation for this study from Roche Molecular Systems, Inc. Support for third-party writing assistance for the manuscript, furnished by Elizabeth Ohneck of Health Interactions, Inc., was provided by Roche Molecular Systems.

REFERENCES

- Newman L, Rowley J, Vander Hoorn S, Wijesooriya NS, Unemo M, Low N, Stevens G, Gottlieb S, Kiarie J, Temmerman M. 2015. Global estimates of the prevalence and incidence of four curable sexually transmitted infections in 2012 based on systematic review and global reporting. *PLoS One* 10:e0143304. <https://doi.org/10.1371/journal.pone.0143304>.
- Centers for Disease Control and Prevention. 2018. Sexually transmitted disease surveillance 2017. U.S. Department of Health and Human Services, Atlanta, GA.
- Bebear C, de Barbeyrac B. 2009. Genital Chlamydia trachomatis infections. *Clin Microbiol Infect* 15:4–10. <https://doi.org/10.1111/j.1469-0691.2008.02647.x>.
- Fleming DT, Wasserheit JN. 1999. From epidemiological synergy to public health policy and practice: the contribution of other sexually transmitted diseases to sexual transmission of HIV infection. *Sex Transm Infect* 75:3–17. <https://doi.org/10.1136/sti.75.1.3>.
- Holder NA. 2008. Gonococcal infections. *Pediatr Rev* 29:228–234. <https://doi.org/10.1542/pir.29-7-228>.
- Kelly H, Coltart CEM, Pant Pai N, Klausner JD, Unemo M, Toskin I, Peeling RW. 2017. Systematic reviews of point-of-care tests for the diagnosis of urogenital Chlamydia trachomatis infections. *Sex Transm Infect* 93: S22–S30. <https://doi.org/10.1136/sextrans-2016-053067>.
- Centers for Disease Control and Prevention. 1993. Recommendations for the prevention and management of Chlamydia trachomatis infections, 1993. *MMWR Recommend Rep* 42(RR-12):1–39.
- Centers for Disease Control and Prevention. 1998. HIV prevention through early detection and treatment of other sexually transmitted diseases—United States. Recommendations of the Advisory Committee for HIV and STD Prevention. *MMWR Recommend Rep* 47(RR-12):1–24.
- Howell MR, Gaydos JC, McKee KT, Jr, Quinn TC, Gaydos CA. 1999. Control of Chlamydia trachomatis infections in female army recruits: cost-effective screening and treatment in training cohorts to prevent pelvic inflammatory disease. *Sex Transm Dis* 26:519–526. <https://doi.org/10.1097/00007435-199910000-00007>.
- Jones CA, Knaup RC, Hayes M, Stoner BP. 2000. Urine screening for gonococcal and chlamydial infections at community-based organizations in a high-morbidity area. *Sex Transm Dis* 27:146–151. <https://doi.org/10.1097/00007435-200003000-00005>.
- Bignell C, Unemo M, European STI Guidelines Editorial Board. 2013. 2012 European guideline on the diagnosis and treatment of gonorrhoea in adults. *Int J STD AIDS* 24:85–92. <https://doi.org/10.1177/0956462412472837>.
- Lanjouw E, Ouburg S, de Vries HJ, Stary A, Radcliffe K, Unemo M. 2016. 2015 European guideline on the management of Chlamydia trachomatis infections. *Int J STD AIDS* 27:333–348. <https://doi.org/10.1177/0956462415618837>.
- Lee KC, Ngo-Metzger Q, Wolff T, Chowdhury J, LeFevre ML, Meyers DS. 2016. Sexually transmitted infections: recommendations from the U.S. Preventive Services Task Force. *Am Fam Physician* 94:907–915.
- New Zealand Sexual Health Society. 2017. NZSHS STI management guidelines for use in primary care 2017. <https://www.nzshs.org/guidelines>. Accessed 22 October 2018.
- Expert Working Group for the Canadian Guidelines on Sexually Transmitted Infections. 2018. Canadian guidelines on sexually transmitted infections. <https://www.canada.ca/en/public-health/services/infectious-diseases/sexual-health-sexually-transmitted-infections/canadian-guidelines/sexually-transmitted-infections.html>. Accessed 22 October 2018.
- Arias M, Jang D, Gilchrist J, Luinstra K, Li J, Smieja M, Chernesky MA. 2016. Ease, comfort, and performance of the HerSwab vaginal self-sampling device for the detection of Chlamydia trachomatis and Neisseria gonorrhoeae. *Sex Transm Dis* 43:125–129. <https://doi.org/10.1097/OLQ.0000000000000406>.
- Paudyal P, Llewellyn C, Lau J, Mahmud M, Smith H. 2015. Obtaining self-samples to diagnose curable sexually transmitted infections: a systematic review of patients' experiences. *PLoS One* 10:e0124310. <https://doi.org/10.1371/journal.pone.0124310>.
- Grant RM, Lama JR, Anderson PL, McMahan V, Liu AY, Vargas L, Goicochea P, Casapia M, Guanira-Carranza JV, Ramirez-Cardich ME, Montoya-Herrera O, Fernandez T, Veloso VG, Buchbinder SP, Chariyalertsak S, Schechter M, Bekker LG, Mayer KH, Kallas EG, Amico KR, Mulligan K, Bushman LR, Hance RJ, Ganoza C, Defechereux P, Postle B, Wang F, McConnell JJ, Zheng JH, Lee J, Rooney JF, Jaffe HS, Martinez AI, Burns DN, Glidden DV, iPrEx Study Team. 2010. Preexposure chemoprophylaxis for HIV prevention in men who have sex with men. *N Engl J Med* 363:2587–2599. <https://doi.org/10.1056/NEJMoa1011205>.
- Grant RM, Anderson PL, McMahan V, Liu A, Amico KR, Mehrotra M, Hosek S, Mosquera C, Casapia M, Montoya O, Buchbinder S, Veloso VG, Mayer K, Chariyalertsak S, Bekker LG, Kallas EG, Schechter M, Guanira J, Bushman L, Burns DN, Rooney JF, Glidden DV, iPrEx Study Team. 2014. Uptake of pre-exposure prophylaxis, sexual practices, and HIV incidence in men and transgender women who have sex with men: a cohort study. *Lancet Infect Dis* 14:820–829. [https://doi.org/10.1016/S1473-3099\(14\)70847-3](https://doi.org/10.1016/S1473-3099(14)70847-3).
- Volk JE, Marcus JL, Phengrasamy T, Blechinger D, Nguyen DP, Follansbee S, Hare CB. 2015. New HIV infections with increasing use of HIV pre-exposure prophylaxis in a clinical practice setting. *Clin Infect Dis* 61: 1601–1603. <https://doi.org/10.1093/cid/civ778>.
- Scott HM, Klausner JD. 2016. Sexually transmitted infections and pre-exposure prophylaxis: challenges and opportunities among men who have sex with men in the US. *AIDS Res Ther* 13:5. <https://doi.org/10.1186/s12981-016-0089-8>.
- Papp JR, Schachter J, Gaydos CA, Van Der Pol B, Centers for Disease Control and Prevention. 2014. Recommendations for the laboratory-based detection of Chlamydia trachomatis and Neisseria gonorrhoeae—2014. *MMWR Recommend Rep* 63(RR-02):1–19.
- Howell MR, Quinn TC, Gaydos CA. 1998. Screening for Chlamydia trachomatis in asymptomatic women attending family planning clinics. A cost-effectiveness analysis of three strategies. *Ann Intern Med* 128: 277–284. <https://doi.org/10.7326/0003-4819-128-4-199802150-00005>.
- Howell MR, McKee KTJ, Gaydos JC, Quinn TC, Gaydos CA. 2000. Point-of-entry screening for C. trachomatis in female army recruits. Who derives the cost savings? *Am J Prev Med* 19:160–166. [https://doi.org/10.1016/S0749-3797\(00\)00202-6](https://doi.org/10.1016/S0749-3797(00)00202-6).
- Kamwendo F, Forslin L, Bodin L, Danielsson D. 1996. Decreasing incidences of gonorrhoea- and chlamydia-associated acute pelvic inflammatory disease. A 25-year study from an urban area of central Sweden. *Sex Transm Dis* 23:384–391. <https://doi.org/10.1097/00007435-199609000-00007>.
- Ledeboer NA, Dallas SD. 2014. The automated clinical microbiology laboratory: fact or fantasy? *J Clin Microbiol* 52:3140–3146. <https://doi.org/10.1128/JCM.00686-14>.

27. Zaninotto M, Plebani M. 2010. The “hospital central laboratory”: automation, integration and clinical usefulness. *Clin Chem Lab Med* 48: 911–917. <https://doi.org/10.1515/CCLM.2010.192>.
28. Cobb B, Simon CO, Stramer SL, Body B, Mitchell PS, Reisch N, Stevens W, Carmona S, Katz L, Will S, Liesenfeld O. 2017. The Cobas(R) 6800/8800 system: a new era of automation in molecular diagnostics. *Expert Rev Mol Diagn* 17:167–180. <https://doi.org/10.1080/14737159.2017.1275962>.
29. Marlowe EM, Hardy D, Krevolin M, Gohl P, Bertram A, Arcenas R, Seiverth B, Schneider T, Liesenfeld O. 2017. High-throughput testing of urogenital and extragenital specimens for detection of *Chlamydia trachomatis* and *Neisseria gonorrhoeae* with Cobas CT/NG. *Eur J Microbiol Immunol (Bp)* 7:176–186. <https://doi.org/10.1556/1886.2017.00018>.
30. Taylor SN, Liesenfeld O, Lillis RA, Body BA, Nye M, Williams J, Eisenhut C, Hook EW, Van Der Pol B. 2012. Evaluation of the Roche Cobas CT/NG test for detection of *Chlamydia trachomatis* and *Neisseria gonorrhoeae* in male urine. *Sex Transm Dis* 39:543–549. <https://doi.org/10.1097/OLQ.0b013e31824e26ff>.
31. Van Der Pol B, Liesenfeld O, Williams JA, Taylor SN, Lillis RA, Body BA, Nye M, Eisenhut C, Hook EW. 2012. Performance of the Cobas CT/NG test compared to the Aptima AC2 and Viper CTQ/GCQ assays for detection of *Chlamydia trachomatis* and *Neisseria gonorrhoeae*. *J Clin Microbiol* 50: 2244–2249. <https://doi.org/10.1128/JCM.06481-11>.
32. Van Der Pol B, Taylor SN, Liesenfeld O, Williams JA, Hook EW. 2013. Vaginal swabs are the optimal specimen for detection of genital *Chlamydia trachomatis* or *Neisseria gonorrhoeae* using the Cobas 4800 CT/NG test. *Sex Transm Dis* 40:247–250. <https://doi.org/10.1097/OLQ.0b013e3182717833>.
33. Gaydos CA, Van Der Pol B, Jett-Goheen M, Barnes M, Quinn N, Clark C, Daniel GE, Dixon PB, Hook EW, III, CT/NG Study Group. 2013. Performance of the Cepheid CT/NG Xpert rapid PCR test for detection of *Chlamydia trachomatis* and *Neisseria gonorrhoeae*. *J Clin Microbiol* 51: 1666–1672. <https://doi.org/10.1128/JCM.03461-12>.
34. Van Der Pol B, Ferrero DV, Buck-Barrington L, Hook E, Lenderman C, Quinn T, Gaydos CA, Lovchik J, Schachter J, Moncada J, Hall G, Tuohy MJ, Jones RB. 2001. Multicenter evaluation of the BDProbeTec ET system for detection of *Chlamydia trachomatis* and *Neisseria gonorrhoeae* in urine specimens, female endocervical swabs, and male urethral swabs. *J Clin Microbiol* 39:1008–1016. <https://doi.org/10.1128/JCM.39.3.1008-1016.2001>.