**Title**

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**Authors**

Kremeyer, B
García, J
Müller, H
et al.

# Original Paper

**Human Heredity**

# Genome-Wide Linkage Scan of Bipolar Disorder in a Colombian Population Isolate Replicates Loci on Chromosomes 7p21–22, 1p31, 16p12 and 21q21–22 and Identifies a Novel Locus on Chromosome 12q

B. Kremeyer[a]   J. García[b]   H. Müller[a]   M.W. Burley[a]   I. Herzberg[a]   M.V. Parra[c]

C. Duque[c]   J. Vega[c]   P. Montoya[b]   M.C. López[b]   G. Bedoya[c]   V. Reus[d]

C. Palacio[b]   C. López[b]   J. Ospina-Duque[b]   N.B. Freimer[e–g]   A. Ruiz-Linares[a, c]

[a]Department of Genetics, Evolution and Environment, University College London, London, UK;
[b]Grupo de Investigación en Psiquiatría, Departamento de Psiquiatría, and [c]Laboratorio de Genética Molecular,
Universidad de Antioquia, Medellín, Colombia; [d]Department of Psychiatry, University of California San Francisco,
San Francisco, Calif., [e]Center for Neurobehavioral Genetics, [f]Department of Psychiatry and Behavioral Sciences,
School of Medicine, and [g]The Jane and Terry Semel Institute for Neuroscience and Human Behavior, University of
California, Los Angeles, Calif., USA

## Abstract

***Background/Aims:*** Bipolar disorder (BP) is a severe psychiatric illness, characterised by alternating episodes of depression and mania, which ranks among the top ten causes of morbidity and life-long disability world-wide. We have previously performed a whole-genome linkage scan on 6 pedigrees segregating severe BP from the well-characterised population isolate of Antioquia, Colombia. We recently collected genotypes for the same set of 382 autosomal microsatellite markers in 9 additional Antioquian BP pedigrees. Here, we report the analysis of the combined pedigree set. ***Methods:*** Linkage analysis using both parametric and nonparametric approaches was conducted for 3 different diagnostic models: severe BP only (BPI); mood disorders (BPI, BPII and major depression); and psychosis (operationally defined by the occurrence of at least 1 episode of hallucinations and/or delusions). ***Results and Conclusion:*** For BPI only, the most interesting result was obtained for chromosome 7p21.1–p22.2 under a recessive model of inheritance (heterogeneity LOD score = 2.80), a region that had previously been linked to BP in a study on Portuguese Island families. For both BPI and mood disorders, nonparametric analyses identified a locus on chromosome 12ct–q14 (nonparametric linkage = 2.55 and 2.35, respectively). This locus has not previously been reported as a candidate region for BP. Additional candidate regions were found on chromosomes 1p22–31 (mood disorders) and 21q21–22 (BPI), 2 loci that have repeatedly been implicated in BP susceptibility. Linkage analysis of psychosis as a phenotype identified candidate regions on chromosomes 2q24–31 and 16p12–q12. The finding on chromosome 16p is noteworthy because the same locus has been implicated by genome-wide association analyses of BP.

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Barbara Kremeyer
Research Department of Genetics, Evolution and Environment
University College London, Wolfson House
4 Stephenson Way, London NW1 2HE (UK)
Tel. +44 20 7679 5093, Fax +44 20 7679 5052, E-Mail b.kremeyer@ucl.ac.uk

## Introduction

Bipolar disorder (BP) is a severe and complex psychiatric condition characterised by alternating episodes of depression and mania [1]. Based on clinical severity, 2 variants of this disorder are commonly distinguished: BP type I (BPI), including at least 1 episode of full-blown mania, typically requiring hospitalization, and BP type II (BPII), characterised by hypomanic episodes, representing a milder form of mania [1]. With a life-time prevalence of 1–1.5% across populations and genders [2], BP is among the top ten causes of morbidity and life-long disability in both the developing and the developed world [3]. Family, twin and adoption studies provide strong evidence for a genetic contribution to the aetiology of this disorder [4, 5]; however, progress in the identification of disease-causing variants has been slow. Family-based linkage studies have produced inconsistent results, and the first genome-wide association scans (GWAS) in BP have failed to live up to the expectations generated by the GWAS findings in other complex diseases [6, 7]. These disappointing results are most likely related to the genetic heterogeneity and phenotypic variability resulting from the descriptive nature of psychiatric diagnoses. The study of a rigorously phenotyped pedigree collection from a population isolate should help reduce this phenotypic and genetic heterogeneity. We have previously implemented this approach by carrying out a whole-genome linkage scan in 6 extended pedigrees segregating severe BP from a well-characterised population isolate (Antioquia in Colombia) [8, 9], and found genome-wide significant evidence of linkage on chromosome 5q31–34 [10]. This signal has subsequently been confirmed through fine-mapping of pedigrees and trios from both Antioquia and the related isolate of the Central Valley of Costa Rica [10, 11].

Since reporting the results of this initial linkage scan, we have collected genome-wide linkage data for 9 additional Antioquian BP families. Here, we report the results of the joint analysis of our original linkage data [10] and the data for these additional pedigrees. In these extended analyses we also explore additional diagnostic and genetic models.

## Subjects and Methods

### Subjects

Fifteen extended pedigrees segregating severe BP were collected as part of an ongoing psychiatric genetics programme in the population isolate of Antioquia, Colombia (fig. 1). Six of these had been included in a previous linkage scan (CO3, CO4, CO7, CO14, CO18, CO27; fig. 1b) [10]. Index cases were recruited in the municipalities of Medellín and Envigado (Antioquia, Colombia) at the Hospital Mental de Antioquia, the Hospital San Vicente de Paúl, the Clínica Sameín, the Clínica Insam, and the Mental Health Centre of Envigado. All index cases had at least 6 great-grandparents born in Antioquia. Families with at least 3 individuals with a clinical diagnosis of BP were chosen for pedigree extension by a social worker or psychiatric nurse using the Family Interview for Genetic Studies [12]. Family CO24 includes only 2 cases of BP; it was studied because it also includes 1 case of major depression and 1 case of schizophrenia, which are phenotypes also considered in the analyses (see below).

Family members were assessed by a psychiatrist using the Spanish version of the Diagnostic Interview for Genetic Studies (DIGS version 3) that we validated in Colombia [12, 13]. Final DSM-IV-TR diagnoses [14] were reached through consensus between 2 expert psychiatrists in a best estimate procedure, as described by Freimer and colleagues [15]. The occurrence of psychotic episodes was determined based on both the DIGS and medical records. Family members with a clinical diagnosis of BPI who were unavailable for interview (including deceased individuals) were considered affected if they had undergone at least 2 hospitalizations and clinical records allowed confirmation of symptoms through the best estimate process. No psychiatric diagnoses other than BPI were considered in family members that were unavailable for interview. In order to maintain a strict phenotypic definition, and to rule out cases where psychiatric symptoms might be secondary to other medical conditions, any psychiatric diagnoses in patients with mental retardation and/or neurological lesions, as well as cases related to substance abuse, were disregarded for the analyses. Altogether, the sample comprised 90 cases of BPI (37.8% males, 62.2% females; age at onset 22.2 ± 7.6 years), 22 cases of major depression (18.2% males, 82.8% females; age at onset 26.6 ± 13.8 years), and 1 case each of schizophrenia (female; age at onset 27 years) and BPII (male; age at onset could not be determined reliably). A total of 46 individuals (30.4% males, 69.6% females) had suffered from psychosis, operationally defined by the occurrence of at least 1 episode of hallucinations and/or delusions. These included 43 of the 90 BPI cases (47.8%), the schizophrenic individual and 2 individuals with a psychotic mood disorder that did not meet the full diagnostic criteria for BP (these 2 individuals are included in families CO14 and CO15; see fig. 1). Written informed consent was obtained from all subjects prior to enrolment in the study. This research was approved by the Ethics Committees of all participating institutions.

### Genotyping

DNA was extracted from whole blood using standard laboratory procedures. Genotype data for 9 newly ascertained pedigrees from Antioquia were obtained for 382 evenly spaced autosomal microsatellite markers from the ABI Linkage Mapping Set v2.5 (Applied Biosystems, Warrington, UK; average distance ~10 cM). Fragment length analysis was performed on a 96-capillary 3730xl DNA Analyzer (Applied Biosystems), and raw genotyping data were analysed using the GeneMapper® software v3.7 (Applied Biosystems). Genotypes were scored independently by 2 researchers blinded to affection status and relatedness between subjects ('double scoring'). Any discordant genotypes were disregarded and, if necessary, repeated. Genotype data were checked for in-
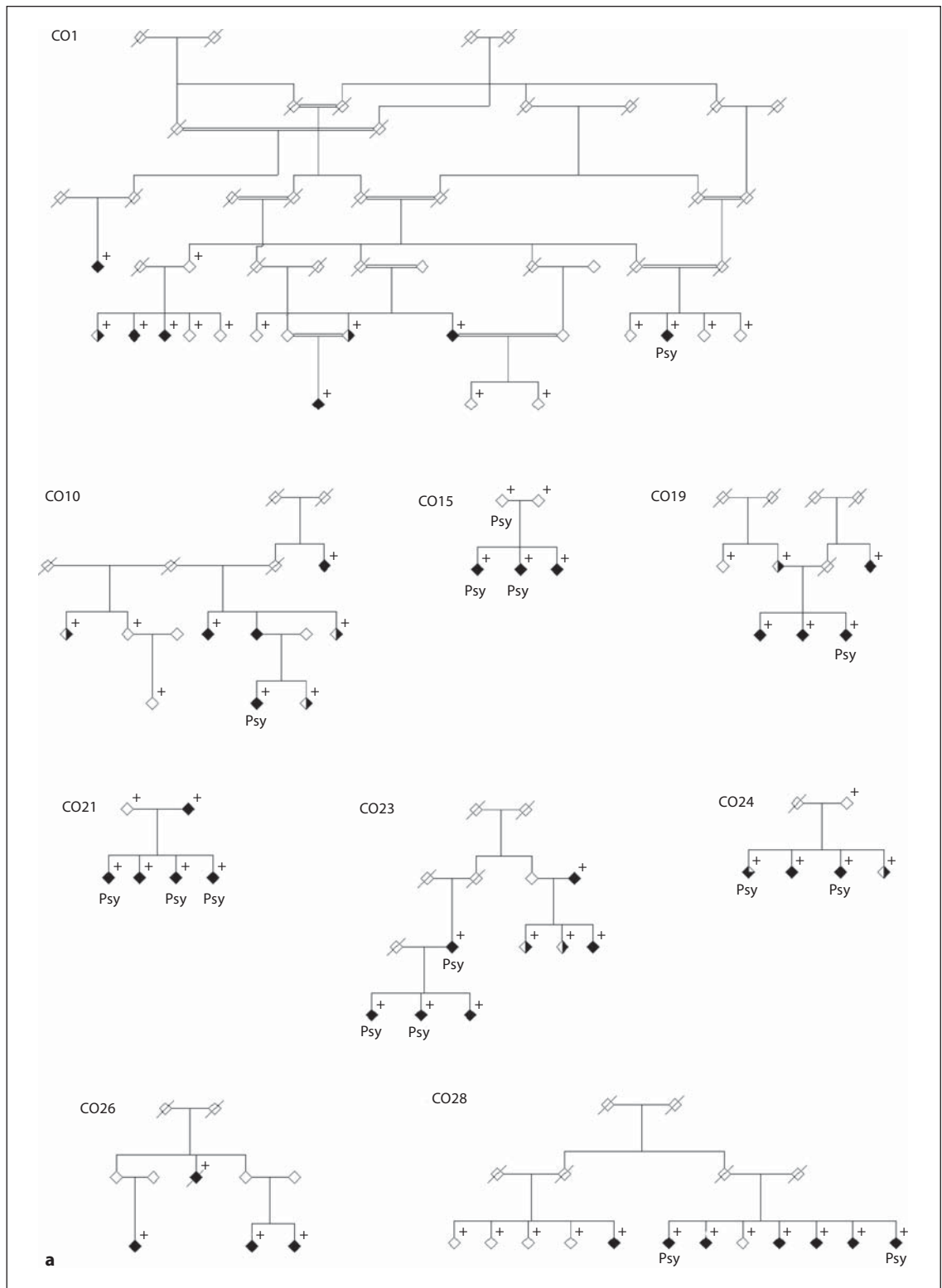
**Fig. 1. a** Set of 9 Antioquian pedigrees segregating severe BP genotyped for this study. Individuals marked with a cross were available for genotyping. Both sets of pedigrees (shown here and in fig. 1b) were analysed together for this study. Filled symbols indicate BPI, half-filled symbols indicate major depression (right half filled) and BPII (left half filled). Filled symbols with a blank upper right quarter indicate schizophrenia. Individuals labelled 'Psy' have suffered episodes of psychosis.
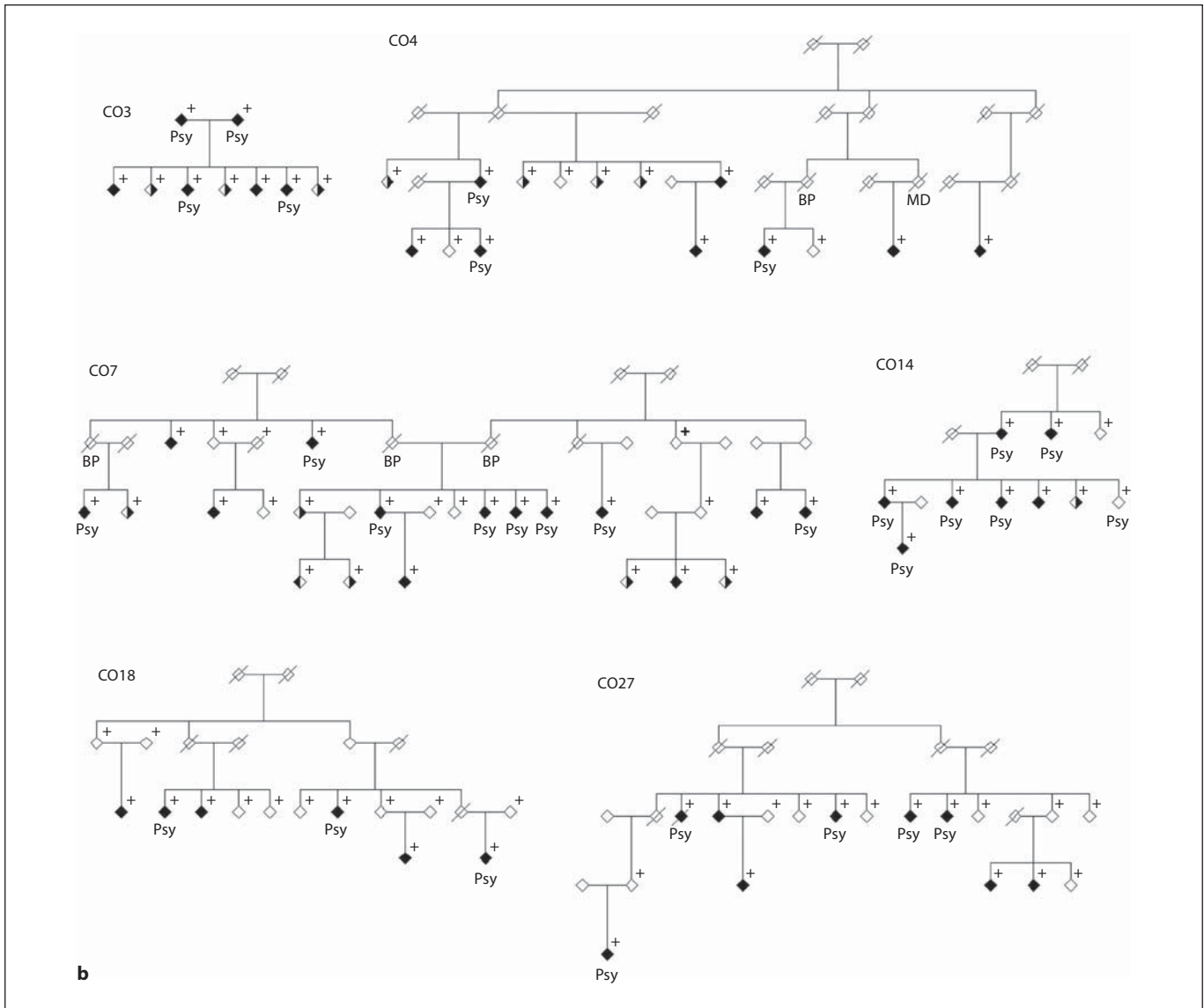
**Fig. 1. b** Set of 6 Antioquian pedigrees genotyped in an initial linkage scan in the Antioquian population [10]. Both sets of pedigrees (shown here and in fig. 1a) were analysed together for this study. Individuals marked with a cross were available for genotyping. Filled symbols indicate BPI, half-filled symbols indicate major depression (right half filled) and BPII (left half filled). Filled symbols with a blank upper right quarter indicate schizophrenia. Individuals labelled 'Psy' have suffered episodes of psychosis.

consistencies with mendelian inheritance using PedCheck [16], and the occurrence of non-mendelian errors was tested with Sim-Walk2 v2.9.1 [17, 18]. The quality-checked genotype data for the 9 new families were combined with those from our previous study of 6 extended BPI families [10] by calibrating all genotypes using a CEPH sample of known genotype (CEPH 1347-02). All analyses were carried out on the combined data set. Because the patterns of disease transmission did not support an X-chromosomal mode of inheritance in the Antioquian pedigrees, the X-chromosome was not analysed. The present study therefore represents an autosome-wide linkage scan.

*Data Analysis and Statistical Methods*

We focused our ascertainment strategy on families segregating BPI. However, several other psychiatric diagnoses segregate in these pedigrees, most notably major depression. We therefore analysed our data under 3 different diagnostic models. In a conservative model, only cases of BPI were considered affected ('narrow model'). In the second model, cases of BPI, BPII and major depression were considered affected ('broad model'). In a third model, the affection status was defined by the occurrence of psychosis ('psychosis model'). Because not all families included 2 or more individuals affected by psychosis, the data set for this last

model comprised only 11 pedigrees. For analyses under all 3 models, all remaining family members were considered to be of unknown phenotype.

Multipoint parametric heterogeneity LOD (HLOD) score and nonparametric linkage (NPL) analyses were performed using SimWalk2 v2.9.1 [17]. Due to the uncertainties involved in specifying an inheritance model underlying the aetiologically complex phenotypes analysed here, NPL analysis was conducted for all 3 phenotypic models. For the narrow (BPI) model only, we also carried out parametric analyses in order to enable a direct comparison of the results from the present study to those from our previous study, which involved part of the families included here [10]. Parametric linkage analysis was performed under both a dominant and a recessive model. Analysis under the dominant model was carried out employing the same parameters used in our previous linkage scan; these parameters were estimated in previous studies of families from the Central Valley of Costa Rica [10, 19]. The frequency of the disease allele was set at 0.003, the phenocopy rate at 0.01, and the penetrances for heterozygous and homozygous carriers of the disease allele were set to 0.81 and 0.9, respectively. Under the recessive model, the frequency of the disease allele was set to 0.1, the phenocopy rate to 0.01, and the penetrance for homozygous carriers of the disease allele was set to 0.80. These parameter values have been used in previous studies (e.g. [20]). In order to reduce the number of tests carried out and to facilitate comparison with our previous analysis of a subset of these families, the model-based linkage analysis was performed only for the narrow phenotypic model. Marker allele frequencies for all analyses were estimated from the complete pedigree data set using Mendel v8.0.1, correcting for relatedness between individuals [21]. Three-digit genotypes were recoded to a 2-digit format, and data files converted to different formats, as required, using Mega2 v4.0 [22]. All analyses used the Marshfield genetic map (http://research.marshfieldclinic.org/genetics/home/index.asp).

## Results

Genotype data for 382 autosomal microsatellite markers were obtained for 75 samples from 9 pedigrees. Genotype data for a further 91 samples from 6 pedigrees from our previously published linkage study [10] were available for the same set of markers. For 10 markers, the genotypes from both data sets could not be calibrated unambiguously; these were excluded from further analyses (D1S498, D1S2836, D5S418, D6S262, D11S905, D13S159, D15S994, D17S798, D19S221, and D21S263). The overall data completeness was 94% across chromosomes and samples.

The genome-wide results of the parametric and nonparametric analyses are shown in figures 2 and 3, respectively. The most noticeable results have been extracted into table 1 (results for individual families are summarised in table 2).

The most interesting study-wide signals were found on chromosome 7p21.1–p22.2, with a HLOD score of 2.80 for the recessive analysis under the narrow diagnostic model, and at marker D12S85 on chromosome 12q13.11, where NPL scores of 2.55 and 2.35 were observed in nonparametric analysis under narrow and broad diagnostic models, respectively. A HLOD score of 2.14 was observed on chromosome 21q at marker D21S1914 in the dominant parametric analysis under the narrow diagnostic model. For this marker, a parametric LOD score of 3.21 was observed in family CO7, and a NPL score of 2.32 in family CO14 (see table 2). For the dominant analysis, HLOD scores >1.3 were also observed on chromosomes 13q and 1p.

Nonparametric analysis under the narrow diagnostic model identified additional candidate regions on chromosomes 1p31.1–p21.2, 1q31.1, 5q12.3, and 9p21.2. Chromosome 1p was also highlighted by the parametric analysis (see above), and by the nonparametric analysis under the broad diagnostic model; results from all 3 analyses defined a common candidate region on chromosome 1p31.1–p21.2. Candidate regions on chromosomes 1q31.1 and 9p21.2 were also supported by the nonparametric analysis under the broad diagnostic model.

For the psychosis phenotype, the highest NPL scores were obtained for chromosomes 2q24.3 and 16p12.1 (2.09 and 2.05, respectively); NPL scores ≥1.3 were also identified on chromosomes 6q, 10q, and 12q. The signal on chromosome 12q was 34cM telomeric of the most significant signal obtained under narrow and broad models.

## Discussion

We performed a whole-genome linkage scan in 15 extended pedigrees segregating severe BP from the population isolate of Antioquia, Colombia. While we identified no signals at a genome-wide level of significance, we found suggestive evidence for a range of candidate regions for BP, mood disorders in general, and psychosis. A lack of significant results at a genome-wide level has characterised linkage studies of BP for the past decades, reflecting the challenges implicated in the mapping of susceptibility variants for complex psychiatric disease, and

**Fig. 2.** Results of the parametric linkage analysis under the narrow diagnostic model and near-dominant (**a**) and recessive (**b**) inheritance models. Multipoint HLOD scores (solid black line) and the estimated proportion of linked families (α; dotted red line) are plotted against the cumulative genetic distance in Haldane cM. Chromosomes are separated by dotted black lines. (For figures see next pages.)
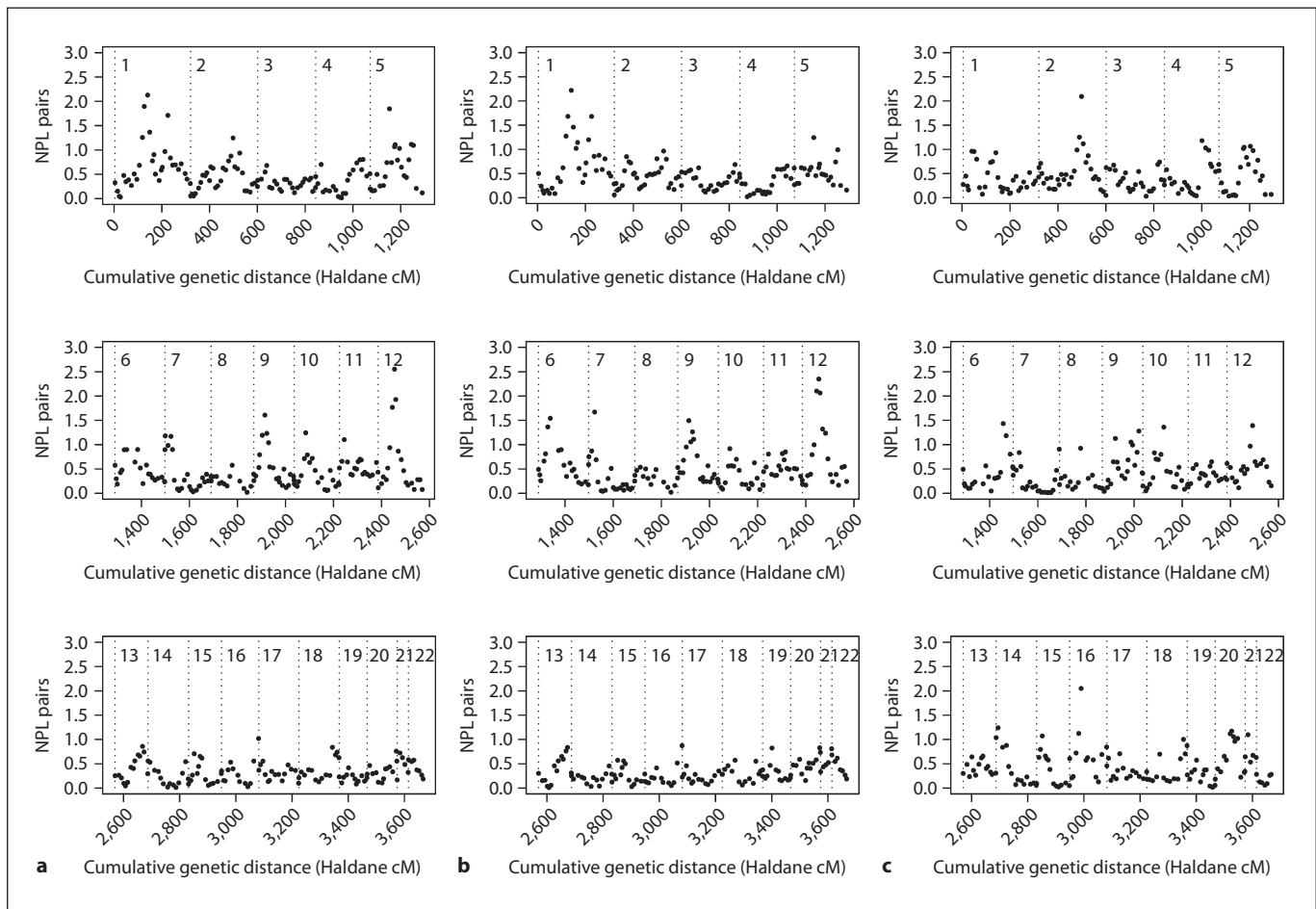
Hum Hered 2010;70:255–268

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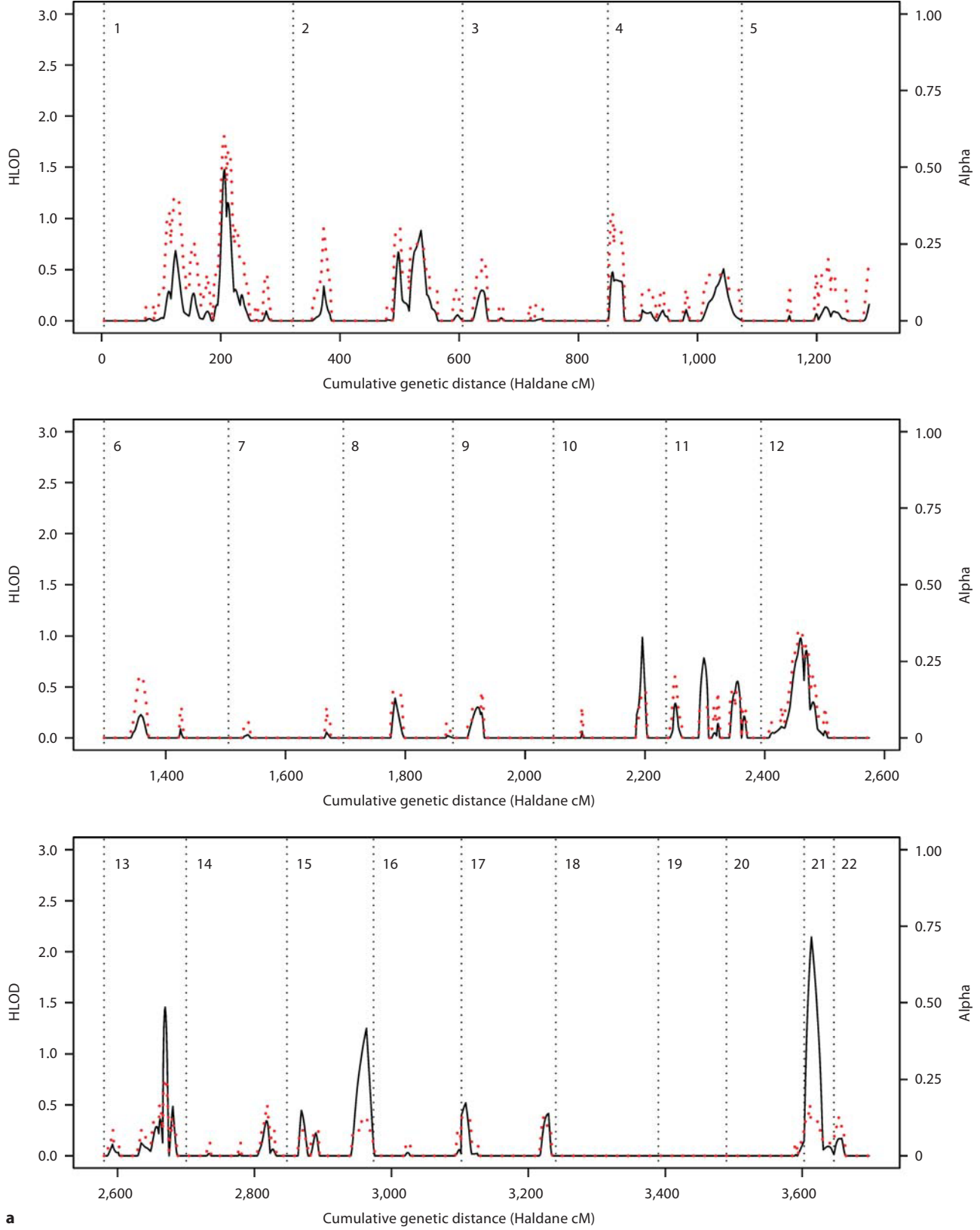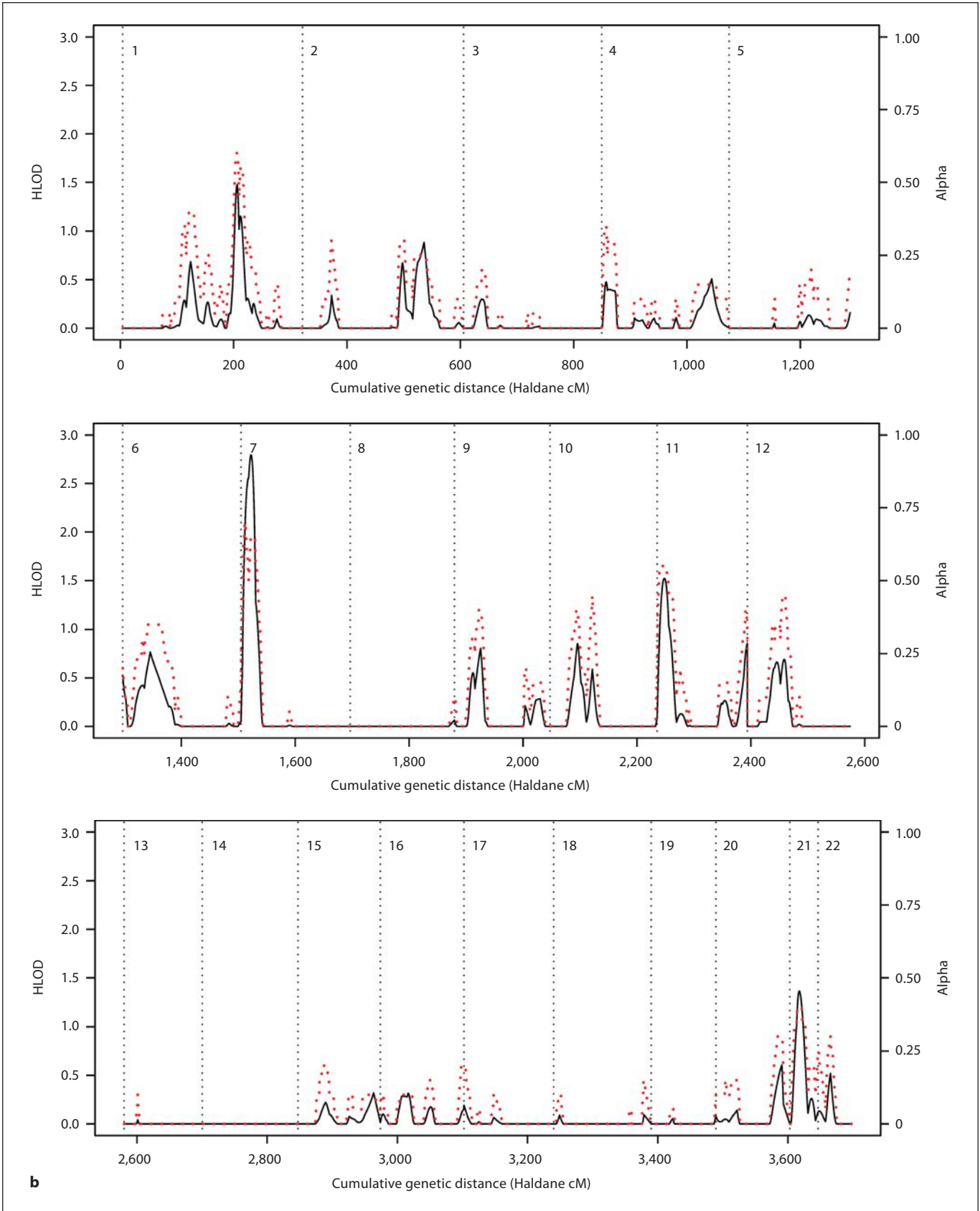
**Fig. 3.** Results of the NPL analysis under narrow (**a**), broad (**b**) and psychosis models (**c**). NPL scores are plotted against the cumulative genetic distance in Haldane cM. Chromosomes are separated by dotted lines.

making the replication of results all the more important. Interestingly, and encouragingly, many of our results replicate findings from previous studies, as discussed in detail below.

BP is a complex and heterogeneous condition, whose mode of inheritance does not easily conform to standard linkage analysis. The use of nonparametric methods provides a way of overcoming these difficulties, while the use of different diagnostic models can help explore the possible genetic overlap between BP, mood disorders and schizophrenia. Consequently, we performed NPL analysis under 3 different diagnostic models: BPI only; BPI, BPII and major unipolar depression; and psychosis (i.e. 'narrow', 'broad', and 'psychosis models', respectively). Additionally, parametric linkage analysis was used for the narrow diagnosis, using both a near-dominant and a

recessive inheritance model. A similar combination of diagnostic definitions and genetic models has been previously used in a number of linkage studies of BP (e.g. [20]).

There was a substantial overlap in the results obtained with the narrow and broad diagnostic models, as both analyses highlighted loci on chromosomes 1p13–31, 1q25–31 and 12p11–q14. An overlap of linkage signals under these 2 diagnostic models has been previously reported [23–28]. These observations suggest the existence of loci predisposing to mood disorders in general rather than to BP in particular, consistent with the observation of an increased rate of major depression in relatives of BP patients [29].

The narrow diagnostic model (BPI) also produced some specific linkage signals on chromosomes 7p21–22, 5q12, 13q33 and 21q21–22, although results differed

**Table 1.** Chromosomal regions suggestive of linkage (LOD or NPL ≥1.3) in the collection of 15 Antioquian BP pedigrees

| Chromosome | Diagnostic model | Inheritance model | Maximum HLOD | Proportion of linked families, α | Marker interval | Chromosomal region | Position Mb |
|---|---|---|---|---|---|---|---|
| *Parametric analysis* | | | | | | | |
| 1 | narrow | dominant | 1.32 | 0.35 | D1S2841–D1S2868 | 1p31.1–p22.1 | 79.49–93.35 |
| 1 | narrow | recessive | 1.47 | 0.6 | D1S196–D1S218 | 1q24.2–q25.1 | 167.60–174.50 |
| 7 | narrow | recessive | 2.80 | 0.65 | D7S517–D7S507 | 7p22.2–p21.1 | 4.50–17.60 |
| 13 | narrow | dominant | 1.46 | 0.3 | D13S158–D13S173 | 13q33.1–q33.3 | 103.98–107.81 |
| 21 | narrow | dominant | 2.14 | 0.15 | D21S1256–D21S1252 | 21q21.1–q22.13 | 19.32–37.83 |

| Chromosome | Diagnostic model | Inheritance model | Maximum NPL | p value | Marker interval | Chromosomal region | Position Mb |
|---|---|---|---|---|---|---|---|
| *Nonparametric analysis* | | | | | | | |
| 1 | narrow | model-free | 2.13 | 0.0074 | D1S207–D1S206 | 1p31.1–p21.2 | 82.54–101.69 |
| 1 | broad | model-free | 2.22 | 0.0060 | D1S207–D1S206 | 1p31.1–p21.2 | 82.54–101.69 |
| 1 | narrow | model-free | 1.71 | 0.0194 | D1S238 | 1q31.1 | 188.15 |
| 1 | broad | model-free | 1.68 | 0.0207 | D1S238 | 1q31.1 | 188.15 |
| 2 | psychosis | model-free | 2.09 | 0.0081 | D2S2330 | 2q24.3 | 166.70 |
| 5 | narrow | model-free | 1.85 | 0.0142 | D5S647 | 5q12.3 | 66.25 |
| 6 | broad | model-free | 1.54 | 0.0286 | D6S276–D6S1610 | 6p22.2–p21.2 | 24.19–39.26 |
| 6 | psychosis | model-free | 1.44 | 0.0367 | D6S441 | 6q25.2 | 153.81 |
| 7 | broad | model-free | 1.67 | 0.0212 | D7S507 | 7p21.1 | 17.60 |
| 9 | narrow | model-free | 1.61 | 0.0246 | D9S161 | 9p21.2 | 27.63 |
| 9 | broad | model-free | 1.50 | 0.0319 | D9S161 | 9p21.2 | 27.63 |
| 10 | psychosis | model-free | 1.36 | 0.0434 | D10S1652 | 10q21.2 | 64.41 |
| 12 | narrow | model-free | 2.55 | 0.0028 | D12S345–D12S368 | 12p11.21–q13.13 | 32.32–52.63 |
| 12 | broad | model-free | 2.35 | 0.0044 | D12S345–D12S83 | 12p11.21–q14.1 | 32.32–60.89 |
| 12 | psychosis | model-free | 1.39 | 0.0403 | D12S351 | 12q21.33 | 91.90 |
| 16 | psychosis | model-free | 2.05 | 0.0089 | D16S3068 | 16p12.1 | 25.56 |

Physical marker positions are from the February 2009 assembly of the UCSC genome browser (http://genome.ucsc.edu).

somewhat between parametric and nonparametric analyses (see table 1). The loci on chromosomes 7p21–22, 13q33 and 21q21–22 were highlighted by the parametric analysis, possibly implicating that segregation patterns at these loci were broadly consistent with the model used for analysis. Segregation on chromosome 7p21–22 may hence best fit the recessive model, while segregation of loci on chromosomes 13q33 and 21q21–22 may best fit the near-dominant mode of inheritance. It is possible that these regions were not consistently picked up by the nonparametric analysis because of its reduced power (although NPL analysis did produce a p value of 0.0212 for the chromosome 7 region under the broad phenotypic model).

Results under the psychosis phenotype implicated different regions from those suggested by the 2 other diagnostic models (chromosomes 2q24–31, 10q21 and 16p12). There are very few published analyses of the psychosis phenotype in bipolar pedigrees; available results are con-

sistent with our findings in that they implicate regions different from those linked to BP as such [30, 31], suggesting that there are genetic loci conferring susceptibility to psychosis independent of affective disorders *per se*. The concept of a genetic predisposition to psychosis is in line with evidence for a shared genetic susceptibility between BP and schizophrenia [32, 33].

Several of the loci identified here have been reported in previous studies of BP. Both the 13q33 and 21q21–22 regions have repeatedly been implicated in susceptibility to BP [23, 28, 34–37]. The region on 21q21 was one of the best-supported loci in our previous linkage scan [10], and the present study provides further support for this region. Interestingly, most of the studies showing evidence for a locus on chromosome 21q also used a narrow diagnostic model similar to the one used here [23, 28, 35]. Lin and colleagues [38] also reported linkage of chromosome 21q22 for early-onset BP, a subtype of the disorder associ-

**Table 2.** Chromosomal regions suggestive of linkage (LOD ≥2 or NPL ≥2) in individual families

| Chromosome | Diagnostic model | Inheritance model | Maximum LOD | Marker interval | Chromosomal region | Position Mb | Pedigree ID |
|---|---|---|---|---|---|---|---|
| *Parametric analysis* | | | | | | | |
| 1 | narrow | dominant | 2.06 | D1S2890–D1S230 | 1p32.2–p31.3 | 57.87–62.60 | CO27 |
| 3 | narrow | dominant | 2.57 | D3S1565–D3S1601 | 3q26.31–q28 | 173.48–191.68 | CO7 |
| 11 | narrow | dominant | 2.19 | D11S905–D11S987 | 11p12–q13.2 | 40.97–67.89 | CO1 |
| 15 | narrow | dominant | 2.46 | D15S127–D15S120 | 15q26.1–15tel | 91.40–99.59 | CO18 |
| 21 | narrow | dominant | 3.21 | D21S1256–D21S1252 | 21q21.1–q22.13 | 19.32–37.83 | CO7 |

| Chromosome | Diagnostic Model | Inheritance model | Maximum NPL (p value) | Marker interval | Chromosomal region | Position Mb | Pedigree ID |
|---|---|---|---|---|---|---|---|
| *Nonparametric analysis* | | | | | | | |
| 1 | broad | model-free | 2.00 (0.0101) | D1S207 | 1p31.1 | 82.54 | CO27 |
| 2 | broad | model-free | 2.23 (0.0059) | D2S117–D2S325 | 2q32.3–q33.3 | 195.62–208.27 | CO23 |
| 3 | narrow | model-free | 2.32 (0.0048) | D3S1580 | 3q28 | 188.54 | CO7 |
| 4 | psychosis | model-free | 2.30 (0.0051) | D4S424 | 4q31.21 | 142.20 | CO14 |
| 16 | narrow | model-free | 2.49 (0.0032) | D16S3046–D16S3068 | 16p12.2–12.1 | 20.89–25.56 | CO7 |
| 16 | psychosis | model-free | 2.40 (0.0040) | D16S3046 | 16p12.2 | 20.89 | CO7 |
| 16 | broad | model-free | 2.12 (0.0075) | D16S3068 | 16p12.1 | 25.56 | CO7 |
| 21 | narrow | model-free | 2.32 (0.0048) | D21S1914 | 21q21.2 | 25.62 | CO14 |

Physical marker positions are from the February 2009 assembly of the UCSC genome browser (http://genome.ucsc.edu).

ated with increased severity of clinical symptoms [39]. Interestingly, the 13q33 region harbours the *DAOA* gene which has been found to be associated with both BP and schizophrenia [40–44].

The loci on 1p13–31 and 1q25–31 implicated here have also both been reported to be involved in BP susceptibility [23, 45–50]. The chromosome 1p13–31 region had been highlighted by our initial genome scan [10], and the present study has further strengthened this signal. The locus on the long arm of chromosome 1 has frequently been linked to schizophrenia [33, 51–54]. This region includes the genes encoding the regulator of G signalling protein, *RGS4*, and the nitric oxide synthase 1 (neuronal) adaptor protein, *NOS1AP*, which have both been associated with BP or schizophrenia [55–59]. Interestingly, NOS1AP expression has been found to be increased in the prefrontal cortex of bipolar subjects [60].

The strongest support for linkage in our study was observed for regions on chromosome 7p21.1–p22.2 (under the narrow affection model and assuming recessive inheritance), and 12q13 (under both narrow and broad affection models).

Evidence of linkage of BP to the chromosome 7p21 region has been reported in a set of 16 extended Portuguese Island pedigrees [61]. The replication of this finding in the Colombian families is of interest because both populations share ancestral contributions from the Iberian Peninsula.

The chromosome 12q13 region has been implicated in a study of French Canadian BP pedigrees. Like the present study, the signal from the Canadian pedigrees was obtained using a broad diagnostic model including BPI, BPII, schizoaffective disorder and recurrent unipolar depression [34]. However, the strongest support for linkage in the Canadian families appears to be in a region telomeric to the locus described here. An interesting candidate gene in the region identified here is *Timeless*. This gene plays a crucial role in the regulation of circadian rhythms [62], the dysfunction of which is a widespread phenomenon in BP [63].

Amongst the regions implicated under the psychosis model, both chromosomes 2q24 and 16p12 have previously been linked to psychiatric disease. The 2q24 region has been implicated in both BP and schizophrenia [53, 64, 65], and the chromosome 16p12 region has been implicated in severe psychotic BP [31]. Interestingly, the most significant signal in the WTCCC BP genome-wide association study was for a SNP in the 16p12 region [6]. This

region harbours several interesting candidate genes, including *DCTN5*, whose product appears to interact with that of *DISC1* (Disrupted in Schizophrenia 1), a gene repeatedly linked to schizophrenia [66, 67].

In this expanded pedigree sample we failed to find further linkage support for the 5q31–34 region implicated by a subset of the families analysed previously [10]. A possible explanation is interfamily heterogeneity. In particular the two largest families examined here (CO1 and CO28) stem from an isolated village, which could represent a subisolate within Antioquia. Neither of these two families provides support for a locus on chromosome 5q31–34, and due to their size, these two pedigrees have a strong impact on the total LOD-scores calculated. For a complex disorder like BP, genetic heterogeneity is not unlikely even within a population isolate.

Such difficulties highlight the need to readjust how results from linkage studies should be interpreted. Most individual studies might not have sufficient power to detect susceptibility loci for psychiatric disease on their own, and replication studies and meta-analyses are essential to evaluate the true significance of linkage findings. In this respect, it is interesting that regions highlighted by meta-analyses of genome-wide linkage studies of BP often do not coincide with the most significant sig-

nals of the individual studies included [47, 68, 69]. Results from an individual study might therefore represent relatively larger genetic effects of importance only for a specific family or population. On the other hand, individually weaker signals could show greater consistency across studies, hence only reaching statistical significance through meta-analyses. Family studies have contributed, and continue to contribute, important information to the emerging picture of genetic predisposition to BP. The present study, which has provided further supporting evidence for a range of previously reported loci, but in which we have also identified a possible novel locus on chromosome 12q13, adds to this picture.

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