

UC Irvine

UC Irvine Previously Published Works

Title

Draft Genome of Janthinobacterium sp. RA13 Isolated from Lake Washington Sediment

Permalink

<https://escholarship.org/uc/item/70h985pj>

Journal

Microbiology Resource Announcements, 3(1)

ISSN

2169-8287

Authors

McTaggart, Tami L

Shapiro, Nicole

Woyke, Tanja

et al.

Publication Date

2015-02-26

DOI

10.1128/genomea.01588-14

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <https://creativecommons.org/licenses/by/4.0/>

Peer reviewed

Draft Genome of *Janthinobacterium* sp. RA13 Isolated from Lake Washington Sediment

Tami L. McTaggart,^{a*} Nicole Shapiro,^b Tanja Woyke,^b Ludmila Chistoserdova^a

Department of Chemical Engineering, University of Washington, Seattle, Washington, USA^a; DOE Joint Genome Institute, Walnut Creek, California, USA^b

* Present address: Tami L. McTaggart, University of California, Irvine, Irvine, California, USA.

Sequencing the genome of *Janthinobacterium* sp. RA13 from Lake Washington sediment is announced. From the genome content, a versatile life-style is predicted, but not *bona fide* methylotrophy. With the availability of its genomic sequence, *Janthinobacterium* sp. RA13 presents a prospective model for studying microbial communities in lake sediments.

Received 29 December 2014 Accepted 5 January 2015 Published 12 February 2015

Citation McTaggart TL, Shapiro N, Woyke T, Chistoserdova L. 2015. Draft genome of *Janthinobacterium* sp. RA13 isolated from Lake Washington sediment. *Genome Announcements* 3(1):e01588-14. doi:10.1128/genomeA.01588-14.

Copyright © 2015 McTaggart et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](http://creativecommons.org/licenses/by/3.0/).

Address correspondence to Ludmila Chistoserdova, milachis@u.washington.edu.

When natural microbial communities from Lake Washington sediment are incubated under the atmosphere of methane, simple, semi-stable communities are formed, consisting of *bona fide* methanotroph species and of nonmethanotrophic satellite species. One of the types found to persist in such methane-fed microcosms is the *Janthinobacterium* species (1). Strain *Janthinobacterium* sp. RA13 was isolated from such an enrichment culture that was incubated at 10°C in a minimal salts medium, with multiple transfers and dilutions, for approximately 18 months (1), by plating onto Nutrient Broth (NB) agar medium (Difco). Axenic culture was obtained by selecting a single colony, followed by multiple re-streaking onto fresh NB plates. The culture features a typical violet color caused by violacein (2).

The draft genome of *Janthinobacterium* sp. RA13 was generated at the DOE Joint genome Institute (JGI), Walnut Creek, CA, USA using Pacific Biosciences (PacBio) sequencing technology (3). All general aspects of library construction and sequencing performed at the JGI can be found at <http://www.jgi.doe.gov>. The raw reads were assembled using HGAP (version 2.2.0.p1) (4). The final draft assembly contains one contig, totaling 6,421,258 bp in size. Genes were identified using Prodigal (5), followed by a round of manual curation using GenePRIMP (6). The predicted coding sequences (CDSs) were translated and used to search the National Center for Biotechnology Information (NCBI) nonredundant database, UniProt, TIGRFam, Pfam, KEGG, COG, and InterPro databases. The tRNAscanSE tool (7) was used to find tRNA genes, whereas rRNA genes were found by searches against models of the rRNA genes built from SILVA (8). Other noncoding RNAs such as the RNA components of the protein secretion complex and the RNase P were identified by searching the genome for the corresponding Rfam profiles using Infernal (<http://infernal.janelia.org>). Additional gene prediction analysis and manual functional annotation was performed within the Integrated Microbial Genomes (IMG) platform (<http://img.jgi.doe.gov>) developed by the JGI (9).

From the genome content, a versatile lifestyle can be predicted for *Janthinobacterium* sp. RA13, including the potential for anaerobic metabolism linked to denitrification. However, with the ex-

ception of the C1 transfer pathway linked to tetrahydrofolate, no traditional methylotrophy pathways (10) are identifiable. With the availability of its genomic sequence, *Janthinobacterium* sp. RA13 presents a prospective model for studying microbial communities in lake sediments.

Nucleotide sequence accession number. The genome sequence has been deposited in GenBank under the accession no. JQNP01000001.

ACKNOWLEDGMENTS

This material is based upon work supported by the National Science Foundation (grant MCB-0950183) and by the U.S. Department of Energy, Office of Science, Office of Biological and Environmental Research under award DE-SC-0010556. The work conducted by the U.S. Department of Energy Joint Genome Institute was supported by the Office of Science of the U.S. Department of Energy under contract DE-AC02-05CH11231.

REFERENCES

- Oshkin IY, Beck DA, Lamb AE, Tchesnokova V, Benuska G, McTaggart TL, Kalyuzhnaya MG, Dedysh SN, Lidstrom ME, Chistoserdova L. 21 October 2014. Methane-fed microbial microcosms show differential community dynamics and pinpoint taxa involved in communal response. *ISME J*. <http://dx.doi.org/10.1038/ismej.2014.203>.
- Pantanello F, Berlutti F, Passariello C, Sarli S, Morea C, Schippa S. 2007. Violacein and biofilm production in *Janthinobacterium lividum*. *J Appl Microbiol* 102:992–999. <http://dx.doi.org/10.1111/j.1365-2672.2006.03155.x>.
- Eid J, Fehr A, Gray J, Luong K, Lyle J, Otto G, Peluso P, Rank D, Baybayan P, Bettman B, Bibillo A, Bjornson K, Chaudhuri B, Christians F, Cicero R, Clark S, Dalal R, Dewinter A, Dixon J, Foquet M, Gaertner A, Hardenbol P, Heiner C, Hester K, Holden D, Kearns G, Kong X, Kuse R, Lacroix Y, Lin S, Lundquist P, Ma C, Marks P, Maxham M, Murphy D, Park I, Pham T, Phillips M, Roy J, Sebra R, Shen G, Sorenson J, Tomaney A, Travers K, Trulson M, Vieceli J, Wegener J, Wu D, Yang A, Zaccarin D, Zhao P, Zhong F, Korlach J, Turner S. 2009. Real-time DNA sequencing from single polymerase molecules. *Science* 323:133–138. <http://dx.doi.org/10.1126/science.1162986>.
- Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <http://dx.doi.org/10.1038/nmeth.2474>.

5. Hyatt D, Chen GL, LaCascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics* 11:119. <http://dx.doi.org/10.1186/1471-2105-11-119>.
6. Pati A, Ivanova NN, Mikhailova N, Ovchinnikova G, Hooper SD, Lykidis A, Kyrpides NC. 2010. GenePRIMP: a gene prediction improvement pipeline for prokaryotic genomes. *Nat Methods* 7:455–457. <http://dx.doi.org/10.1038/nmeth.1457>.
7. Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res* 25:955–964. <http://dx.doi.org/10.1093/nar/25.5.0955>.
8. Pruesse E, Quast C, Knittel, Fuchs BM, Ludwig W, Peplies J, Glöckner FO. 2007. Silva: a comprehensive online resource for quality checked and aligned ribosomal RNA sequence data compatible with ARB. *Nucleic Acids Res* 35:2188–7196. <http://dx.doi.org/10.1093/nar/gkm864>.
9. Markowitz VM, Mavromatis K, Ivanova NN, Chen IMA, Chu K, Kyrpides NC. 2009. IMG ER: a system for microbial genome annotation expert review and curation. *Bioinformatics* 25:2271–2278. <http://dx.doi.org/10.1093/bioinformatics/btp393>.
10. Chistoserdova L. 2011. Modularity of methylotrophy, revisited. *Environ Microbiol* 13:2603–2622. <http://dx.doi.org/10.1111/j.1462-2920.2011.02464.x>.