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Activity-based, genome-resolved metagenomics uncovers key populations and pathways involved in subsurface conversions of coal to methane

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Abstract

Microbial metabolisms and interactions that facilitate subsurface conversions of recalcitrant carbon to methane are poorly understood. We deployed an *in situ* enrichment device in a subsurface coal seam in the Powder River Basin (PRB), USA, and used BONCAT-FACS-Metagenomics to identify translationally active populations involved in methane generation from a variety of coal-derived aromatic hydrocarbons. From the active fraction, high-quality metagenome-assembled genomes (MAGs) were recovered for the acetoclastic methanogen, *Methanothrix paradoxum*, and a novel member of the *Chlorobi* with the potential to generate acetate via the Pta-Ack pathway. Members of the *Bacteroides* and *Geobacter* also encoded Pta-Ack and together, all four populations had the putative ability to degrade ethylbenzene, phenylphosphate, phenylethanol, toluene, xylene, and phenol. Metabolic reconstructions, gene analyses, and environmental parameters also indicated that redox fluctuations likely promote facultative energy metabolisms in the coal seam. The active *Chlorobi* MAG encoded enzymes for fermentation, nitrate reduction, and multiple oxygenases with varying binding affinities for oxygen. *M. paradoxum* PRB encoded an extradiol dioxygenase for aerobic phenylacetate degradation, which was also present in previously published *Methanothrix* genomes. These observations outline underlying processes for bio-methane from subbituminous coal by translationally active populations and demonstrate activity-based metagenomics as a powerful strategy in next generation physiology to understand ecologically relevant microbial populations.

Introduction

Methane is an important source of energy globally, and in recent years has undergone rapid development accounting for almost 40% of energy consumption in the United States [1]. As energy demands increase and natural gas resources are depleted, there is a heightened need for alternative and less conventional energy technologies to be developed [1]. One near-term, unconventional energy resource is biogenic coalbed methane (CBM), *i.e.*, the biological conversion of coal to methane. It has been estimated that roughly 40% of CBM in the United States is of biogenic origin, and the interest in CBM is growing due to the presence of this natural process associated with many coal reserves in the United States [2]. Biogenic CBM is a source of cleaner energy compared to coal owing to its naturally refined low molecular weight hydrocarbon content and cleaner burning properties [3, 4]. However, methane has 84 times the global warming potential of carbon dioxide over a 20-year period [5], and methane off-gassing at oil and coal wells is both a major safety concern and a serious environmental problem. Ultimately, many hydrocarbon environments can be associated with biogenic methane, and whether the goal is to stimulate methane production for harvesting cleaner fuels or mitigate methane production to restrict carbon release, current understanding indicates the likely rate-limiting step is conversion of the coal to methanogenic precursors [6, 7]. The significance of different carbon cycling pathways involved in the turnover of recalcitrant carbon to methane is still a topic of debate, and unknown carbon cycling pathways continue to be discovered. This fundamental knowledge is necessary to understand microbial processes that contribute to subsurface carbon turnover in relationship to biogenic methane and helps identify unknown pathways that link terrestrial subsurface carbon cycling with carbon dioxide and methane.

Broadly, biogenic CBM can be divided into two primary microbial steps: (i) conversion of coal or other complex organic intermediates into simpler organic precursors and (ii) conversion of simple organic intermediates to methane by methanogens. Coal is a complex

heterogeneous hydrocarbon with mixed chemical composition including high-molecular-weight polycyclic aromatic hydrocarbons and derivatives with a high mass fraction of carbon [8]. Coal is classified by ranking thermal maturity with designations increasing from lignite, subbituminous, bituminous, and anthracite. Lower rank subbituminous coals such as those found in the Powder River Basin (PRB, southeastern Montana/northeastern Wyoming, USA) are thought to be more bioavailable than higher rank coals because lower rank coals contain more oxygen, sulfur, and nitrogen and less aromaticity [9]. Laboratory studies have indicated microbial production of CBM can also increase porosity and consequently the bioavailability of coal through the utilization of oxygen-containing functional groups which reduces the degree of crystallization and increases the pore connectivity [10, 11]. Other researchers have shown that low ranking subbituminous coal can have higher concentrations of extractable acetate [12], an important intermediate in microbial conversions of coal to methane [13, 14]. Microbial communities break down coal components into simple intermediates that can be utilized by methanogens to produce methane, but the specific components of the coal that are targeted for degradation and the responsible microbial populations remain unknown.

During methanogenesis methane gas is produced as the final step of organic matter degradation in anoxic environments by taxonomically diverse archaea including seven orders of *Euryarchaeota [15, 16]*, *Verstraetearchaeota [17]*, and possibly *Korarchaeota [18, 19]*. The primary substrates for archaeal methanogenesis include carbon dioxide and hydrogen, methylated compounds, and acetate [20]. Acetoclastic methanogenesis is thought to be predominant in nature, with estimates suggesting it accounts for two-thirds of the 100 billion tons of methane produced globally by microorganisms each year [21–26]; consequently, acetate plays a crucial role in the global production of methane from organic matter. In coalbed environments, microbial community analyses and isotopic signatures have indicated the presence of acetoclastic methanogens alongside hydrogenotrophic and methylotrophic methanogens [27]. Only two identified genera of methanogens, *Methanosarcina* and *Methanothrix*, are capable of acetoclastic methanogenesis and thus are crucial to our current understanding of the global methane cycle [22]. While *Methanosarcina barkeri*, for example, is a generalist capable of producing methane from acetate as well as other substrates (*e.g.*, $CO_2 + H_2$), *Methanothrix soehngenii* is an obligate acetoclastic methanogen that outcompetes *Methanosarcina* spp. at low acetate concentrations. *Methanothrix*-like spp. are thus thought to be the predominant acetoclastic methanogens in environmental settings where acetate is limited [28, 29]. While much is known independently about complex hydrocarbon degradation and methanogenesis, our present understanding of the microbial processes that link *in situ* metabolisms remains limited.

We determined potential metabolic linkages among microbial populations engaged in coal degradation, acetate production, and methanogenesis in the PRB Flowers-Goodale coal seam using a powerful combination of four primary techniques: (i) a nine-month *in situ* enrichment with crushed coal using a subsurface environmental sampler (SES) [30], (ii) bio-orthogonal non-canonical amino acid tagging (BONCAT) [31], (iii) fluorescently active cell sorting (FACS) [32], and (iv) genome-resolved metagenomics. Previous investigations have made major strides in surveying natural microbial communities by combining BONCAT-FACS with analyses of SSU rRNA gene sequences [33, 34], but genome-resolved metagenomics has yet to be performed on active cells following BONCAT-FACS and can provide biochemical predictions for active populations. In comparison to shotgun metagenomic sequencing, which sequences total community DNA and cannot discriminate between active, dead, and dormant

organisms, sorting translationally active cells prior to metagenomic sequencing enables the identification of active microbial populations and associated genetic potential under relevant environmental conditions. We recovered high-quality, active metagenome-assembled genomes (MAGs) representing (i) a previously unidentified member of phylum *Chlorobi* with acetate-producing potential and (ii) a putative acetoclastic methanogen related to *Methanothrix paradoxum*. We hypothesize that these genomic populations (as well as members of the *Geobacter* and *Bacteroidetes*) interact in the degradation of aromatic coal byproducts and the subsequent production of methane from coal-derived acetate under fluctuating redox conditions.

Results & Discussion

Recovery of high-quality, translationally active MAGs from an in situ *coal enrichment*

The SES was filled with crushed, subbituminous coal from the PRB and deposited at 115 m depth within a coal-bearing layer of the Flowers-Goodale coal ~~bed~~ seam at the U.S. Geological Survey (USGS) Birney Test Site (**Figure 1**). Previous research demonstrated high concentrations (50 mg/L) of isotopically depleted methane (δ^{13}C-CH_4 = -67 ‰ versus VPDB) within this layer [6], indicating the presence of a microbial community engaged in CBM production. After a nine-month subsurface enrichment, the SES was retrieved maintaining *in situ* pressure and gaseous headspace conditions. We then performed BONCAT-FACS and sequenced the metagenome of translationally active and total sorted cell fractions. Metagenomic binning resulted in 24 metagenome assembled genomes (MAGs) from the translationally active fraction of the coal-enriched community (**Supplementary Table 1**) and 44 MAGs from the total cell fraction [35]. Two BONCAT-active genomic populations, Bin15 and Bin8, were recovered as high-quality MAGs with estimated completeness > 95% and estimated redundancy < 5% based on the detection of single copy genes for bacteria and archaea (**Table 1**). Robust phylogenomic analyses of concatenated archaeal ribosomal proteins indicated that Bin15 was a close neighbor to *Methanothrix paradoxum* NSM2 [36] within the *Methanosarcinales* order of methanogens (**Figure 2A**), and henceforth will be referred to as '*Methanothrix paradoxum* PRB' for Powder River Basin. Consistent with this, *M. paradoxum* PRB had the highest genome-wide average nucleotide identity with *M. paradoxum* NSM2 at 77.6% when compared with other methanogens within and outside of the *Methanosarcinales* (**Supplementary Table 2**). *M. paradoxum* PRB had a genome size of 2.90 Mb, 50.7% G+C content and 2,946 genes, similar to the type strain *M. soehngenii* GP6, which is 3.03 Mb with 51.9% G+C content and 2,925 genes (**Table 1**).

Bacterial phylogenomic analysis indicated that high-quality, translationally active MAG Bin8 belonged to phylum *Chlorobi* and classified within the poorly understood OPB56 clade (**Figure 2B**). Recently, *Chlorobi* groups were observed *in situ* via sequence analysis during long-term monitoring of an Australian coal seam post-stimulation for CBM production [37]. The *Chlorobi* phylum was first established to comprise the phototrophic Green Sulfur Bacteria, which is now considered class *Chlorobea [38]* and later revised to include the non-phototrophic class, *Ignavibacteria [39]*. OPB56 has been recognized as a third, class-level clade and was originally detected as a cluster of SSU rRNA gene sequence clones from Obsidian Pool in Yellowstone National Park [40]. Recent genomic discoveries by Hiras and colleagues [41] have confirmed the class-level OPB56 clade and, like the *Ignavibacteria*, OPB56 is non-phototrophic and contains genomes from thermophilic and non-thermophilic microbial populations. The '*Chlorobi* PRB' MAG (Bin8) was 3.34 Mb in length with 37.5% G+C content and a total of 2,777 genes, in

contrast to 2.67 Mb length, 56.0% G+C, and 2,363 genes for NICIL-2, the only other genome within the OPB56 clade that has been thoroughly evaluated [41].

The recovery of high-quality genomes for *M. paradoxum* PRB and *Chlorobi* PRB from the translationally active fraction suggests that these populations play key roles in the environment, though sequencing and/or amplification biases could also influence high genomic recoverability. We tested the hypothesis that these populations are ecologically relevant and detectable in coal-bed environments by mapping quality-filtered short reads from four additional standard shotgun (non-BONCAT) metagenomes to the *M. paradoxum* PRB and *Chlorobi* PRB MAGs (**Table 1**). The four environmental metagenomes were from (i) the same well (FG11) but a different timepoint, (ii) a different well (FGP) from the same methane-producing coal seam, (iii) a non-methane-producing well (N11) also in the PRB [35], and (iv) a CBM-producing well (CX10) from the Surat Basin, Australia [42]. After normalizing for total sequence content, mean genomic coverage values for *M. paradoxum* PRB and *Chlorobi* PRB were 22X and 4X in FG11 and 153X and 0.25X in FGP, respectively, indicating that these populations exist naturally within the methane-producing Flowers-Goodale coal seam and may fluctuate in relative abundance based on environmental conditions. Moreover, a population corresponding to *M. paradoxum* PRB was also binned in a metagenome from a separate well in the Flowers-Goodale coal seam (FG09) and is presented in a companion study [35]. Genomic sequence of *M. paradoxum* PRB and *Chlorobi* PRB were also recovered from the total community sorted fraction (in addition to the translationally active fraction) after the *in situ* enrichment (*i.e.*, Bin21 in **Figure 2A** and Bin26 in **Figure 2B**). In contrast, neither *M. paradoxum* PRB nor *Chlorobi* PRB recruited metagenomic reads from a non-methane-producing well (N11) in the PRB or a methane-producing well in the Surat Basin (CX10 [42], **Table 1**). This indicates that these populations may be endemic to high-CBM production wells in the PRB, but this hypothesis requires further testing as more metagenomic data are produced from coalbed environments. Due to the recovery of *M. paradoxum* PRB and *Chlorobi* PRB from multiple sequence sources (*i.e.*, translationally active cell sorts, total cell sorts, binned environmental metagenomes, and mapped short reads) we hypothesize that these two populations play important and interconnected roles in the accumulation of methane in the PRB subsurface coal environment.

Metabolic properties of Chlorobi PRB

Metabolic comparisons between *Chlorobi* PRB and NICIL-2 [41] demonstrated consistent properties of the OPB56 class-level clade within phylum Chlorobi (**Figure 3**). In contrast to the Green Sulfur Bacteria (class *Chlorobea*), OPB56 populations are not obligate anaerobes and do not possess genes involved in photosynthetic reactions (*i.e.*, reaction centers [*pscB, pscC, pscD*], chlorosome envelope [*csmABCDEFHIJX*], bacteriochlorophyll a [*fmoA*]). In contrast, functional genes detected in *Chlorobi* PRB and NICIL-2 genomes suggest they are obligate heterotrophs with a facultative lifestyle capable of fermentation and aerobic respiration. NICIL-2 was previously enriched under oxic conditions and, like *Chlorobi* PRB, encodes multiple subunits for cytochrome c oxidases. *Chlorobi* PRB also encodes subunits of additional oxidases with varying binding affinities for oxygen, including cbb3-type cytochromes and bd-ubiquinols [43]. These observations suggest *Chlorobi* PRB may be adapted to respiring oxygen across a range of concentrations in subsurface coal seam environments. The cbb3-type oxidase, for example, is used by pathogenic proteobacteria to colonize anoxic zones in human tissue [44].

Chlorobi PRB also has the putative ability to perform anaerobic respiration via genes for membrane-bound nitrate reduction to ammonia (*nrfAH [45]*) and nitrous oxide reductase (*nosZ [46]*). In support of these findings, previous work using BONCAT-FACS on hot spring samples from Yellowstone National Park demonstrated that the OPB56 clade increased in SSU rRNA gene relative abundance when amended with oxygen or nitrate [34]. Putative respiration pathways are supported by the detection of complete electron transport pathways in both OPB56 genomes, and all genes were detected for an F-type proton-translocating ATP synthase. Like NICIL-2, *Chlorobi* PRB lacks genes for oxidation of sulfur compounds, distinguishing the OPB56 clade from members of class *Chlorobea* such as *Chlorobaculum tepidum*. Importantly, while *Chlorobi* PRB has high estimated genome completeness > 95%, the absence of genes could represent lack of genetic potential or genes that did not assemble with the MAG for methodological reasons. Therefore, missing functional properties inferred from gene absences are considered hypotheses that require further testing.

Further distinguishing OPB56 from the photosynthetic Green Sulfur Bacteria, both *Chlorobi* PRB and NICIL-2 encoded a complete TCA cycle and lacked genes for the rTCA cycle, which is the method by which members of the *Chlorobea* fix carbon. NICIL-2 has a complete glycolysis pathway while *Chlorobi* PRB is missing the enolase gene for the conversion of 2-phosphoglycerate to phosphoenolpyruvate. Both genomes have all additional genes necessary for gluconeogenesis. While NICIL-2 and *Chlorobi* PRB are both capable of fermentation, only *Chlorobi* PRB has the putative ability to produce acetate via a combination of the phosphotransacetylase (Pta) and acetate kinase (Ack) enzymes (discussed below). In contrast, NICIL-2 lacks the *ack* gene but has an alcohol dehydrogenase for the fermentative production of ethanol, which *Chlorobi* PRB does not. Carbon sources for NICIL-2 and *Chlorobi* PRB are primarily limited to simple, short-chain carbon compounds [41]; however, consistent with its recovery from a coal seam environment, the *Chlorobi* PRB genome also possessed genes for the anaerobic degradation of aromatic hydrocarbons such as phenylphosphate dehydrogenase (*ppd*), ethylbenzene dehydrogenase (*ebd*), and phenylethanol dehydrogenase (*ped*) (**Table 2**). By contrast, NICIL-2, which was not recovered from coal, only had the *ped* gene. Evidence for anaerobic hydrocarbon degradation in *Chlorobi* PRB together with the presence of anaerobic respiration genes (*nrfAH*, *nosZ*) may indicate that under anoxic conditions *Chlorobi* PRB respires hydrocarbons using oxidized nitrogen compounds [47, 48] and/or ferments hydrocarbons to acetate.

Both genomes from the OPB56 clade lack several biosynthesis pathways for amino acids, including leucine, valine, isoleucine, serine, phenylalanine, tryptophan, tyrosine, methionine, histidine, and proline. These results suggest that NICIL-2 and *Chlorobi* PRB likely rely on exogenous sources for many amino acids. Consistent with this, Reichart and colleagues demonstrated an increase SSU rRNA gene relative abundance of the OPB56 clade in enrichments amended with isoleucine [34], and the NICIL-2 and *Chlorobi* PRB genomes both have complete degradation pathways for isoleucine. Importantly, the missing biosynthesis pathway for methionine could enhance affinity for the synthetic amino acid HPG, which is a methionine analog. However, it should be noted that HPG levels used in short-term labeling incubations were low as previously described [33], and the *Chlorobi* sequences could be mapped back to unlabeled metagenomes from the environment. Finally, *Chlorobi* PRB and NICIL-2 encode all components of a complete flagellum complex, and *Chlorobi* PRB has two additional

genes for flagellar chaperones, one that regulates flagellin polymerization (*fliS [49]*) and another that is essential to P ring formation (*flgA [50]*). These observations suggest that translationally active *Chlorobi* PRB may be motile in the subsurface coal seam, though genes for flagella are not always expressed, as is the case for cultures of *Ignavibacterium album [39, 51]*. Further investigations are needed to confirm suggested structures and functions based on gene detection/ annotation.

Acetate production by Chlorobi *PRB*

As mentioned previously, *Chlorobi* PRB has the putative ability to produce acetate as a byproduct of fermentation using the Pta-Ack pathway. During fermentation, Pta catalyzes the replacement of CoA with a phosphate group and Ack subsequently cleaves the phosphate group, thereby releasing acetate while conserving energy in the form of 1 ATP [21]. Substrates for acetate production consist of many breakdown products of complex carbon sources, including glucose, propionate, and butyrate, as well as H_2/CO_2 in the case of homoacetogens. The importance of the Pta-Ack pathway in methanogenic environments plays a crucial role in coupling the breakdown of complex carbon to a substantial fraction of global methane production [22–26, 52]. In the *Chlorobi* PRB genome, *ack* and *pta* are adjacent genes on a contig with a length of ca. 10 kbp that contains seven genes. The neighboring gene to *ack/pta* is *argE*, which encodes acetylornithine deacetylase (COG0624) within the Zinc peptidase family (cl14876), an enzyme that catalyzes another acetate-producing reaction. *Chlorobi* PRB also has an additional copy of *pta* on a separate contig. These observations further support the hypothesis that the *Chlorobi* PRB population, which was translationally active after being enriched *in situ* on coal, may release acetate as a byproduct during the degradation of coal-derived aromatics.

Acetoclastic methanogenesis by Methanothrix paradoxum *PRB*

Only two genera of methanogens, *Methanothrix* and *Methanosarcina*, have been shown to use acetate as the sole source of carbon and energy during the production of methane [53]. Unlike *Methanosarcina* spp., which are generalists that can also grow on methylated compounds or hydrogen, *Methanothrix* spp. have no known substrates for methane production apart from acetate [22, 54]. Known *Methanothrix* spp. have extremely high affinity for acetate and can outcompete *Methanosarcina* spp. by growing at lower acetate concentrations in many environments. While *Methanosarcina* spp. use the previously discussed Pta-Ack pathway in reverse to activate acetate for methane production, *Methanothrix* spp. instead use acetyl-CoA synthetase (*acs*). *M. paradoxum* PRB from the present study harbored genes consistent with the latter type of acetoclastic methanogenesis (**Table 2**; **Supplementary Table 4A**), including *acs* for acetate activation, carbon monoxide dehydrogenases (*cdh*) for cleavage of carbon groups and oxidation of CO, methyltransferases (*mtr*) for activation of the methyl group, and methyl coenzyme M reductase (*mcr*) for the final reduction step that produces methane [20, 22]. Four adjacent copies of *acs* existed on a 24,695-kbp contig and a fifth copy was detected on a separate contig. The *cdh* operon was ordered alpha, epsilon, beta, CooC1 Ni-accessory protein, delta, gamma, and was present on a single contig spanning 41,179 kbp. The *mcr* operon was on a large contig > 40 kbp in length and consisted of subunits ordered beta, D, gamma, alpha. In addition to methanogenesis from acetate, hidden Markov model (HMM) scans for anaerobic hydrocarbon degradation proteins [55] detected putative protein sequences for breaking down phenylethanol

(*ped*), toluene and xylene (*bss*), and phenol (*pps*). These observations suggest that, in addition to producing methane directly from acetate, *M. paradoxum* PRB may directly or indirectly use certain coal byproducts as additional sources of carbon and/or energy. These results coincide with the recent discovery of a different methanogen, *Methermicoccus shengliensis*, which has the ability to utilize methyl-groups from methoxylated aromatic compounds [56].

Considerations of oxygen tolerance and usage

Several important findings challenge the classical understanding that all methanogens are strict anaerobes. Possible aerotolerant methanogens have been identified in methanogenic ecosystems such as rice paddy fields, arid soils, and anaerobic digesters (*e.g.*, [57–59]). Several more observations of possible oxygen tolerance come specifically from the acetoclastic *Methanothrix* genus. First, the original cultivations of *M. soehngenii* GP6 demonstrated that growth could be attained starting from aerobic samples and on sewage exposed to pure oxygen for up to 48 hrs [54]. Second, Jetten and colleagues [22, 28] purified and characterized the Cdh enzyme from *M. soehngenii*, which was determined to be "completely insensitive to molecular oxygen," in contrast to the same enzyme from *Methanosarcina barkeri* which irreversibly decreased in activity by 90% after trace oxygen exposure [60]. Phylogenetic analysis of the Cdh enzyme from *M. paradoxum* PRB confirms placement as a neighbor to the Cdh from *M. soehngenii*, together forming a separate cluster from the *Methanosarcinales* Cdh group (**Supplementary Figure 1**). Finally, Angle *et al*. [36] observed that methane production increased by up to an order of magnitude in oxygenated wetland soils compared to anoxic soils and methanogenesis was attributed primarily to acetoclastic *M. paradoxum*. In the present study, SES oxygen measurements in the FGP well ranged from 0.25 to 1.09 mole % (n = 5) (**Supplementary Table 3**). Although we cannot rule out potential oxygen contamination from sampling or analysis, previous metagenomic analyses of coal-bed environments have indicated the importance of aerobic or microaerophilic metabolisms in such environments [61]. The observed oxygen fluctuations are consistent with the wide-ranging potential for energy conservation observed in *Chlorobi* PRB, which encodes oxygenases with varying binding affinities (high to low oxygen concentrations), nitrate reductases, and fermentation enzymes.

Remarkably, the *M. paradoxum* PRB genome harbored an extradiol dioxygenase (*elh*) gene from the LigB superfamily for the aerobic degradation of phenylacetate [62]. The presumptive protein sequence was recovered with a 40.8% amino acid similarity in the HMM scan, with an expect (e) value of 3.0×10^{-25} (e value of 5.6×10^{-86} for match to COG2078, **Supplementary Table 4A**). We ruled out sequence contamination by comparing the full 36-kb contig containing the *elh* gene to the NCBI non-redundant sequence database and observed a closest nucleotide sequence match (77.76% identity, e value = 0) to *M. soehngenii* GP6. Consistent with this, *M. soehngenii* GP6 and *M. paradoxum* NSM2 both had copies of *elh* as well. Further support for phenylacetate metabolism in *M. paradoxum* PRB was observed in a neighboring gene for phenylacetate-coenzyme A ligase (COG1451, adenylate-forming domain family), which occurred just three genes downstream of *elh* for aerobic phenylacetate degradation (**Supplementary Table 5**). Phenylacetate has been demonstrated as a key intermediate in the conversion of organic matter to methane by accumulation in peat soil enrichments when methanogenesis was inhibited; in some inhibition experiments, phenylacetate accumulated to even higher concentrations than acetate [25]. Finally, growth and methane

production were observed for *M. soehngenii* in the presence of acetate and phenylacetate, although not on phenylacetate alone [54]. Due to the (i) apparent relationship between phenylacetate and acetoclastic methanogenesis, (ii) the unique oxygen-tolerating characteristics of *Methanothrix* spp., (iii) the presence of the *elh* dioxygenase for phenylacetate degradation in *M. paradoxum* PRB, and (iv) the observed fluctuating redox conditions of the Flowers-Goodale coal seam, we hypothesize that *M. paradoxum* PRB may use trace oxygen for ring cleavage of coal-derived phenylacetate during or as an alternative (and/or supplement) to the production of methane from acetate. Further research such as methanogenic cultivations under oxic/suboxic conditions and purifications of the novel Elh enzyme would be needed to test this hypothesis.

Biological process for CBM production

The recovery of two high-quality MAGs with ostensibly related putative metabolisms (*i.e.*, acetoclastic methanogenesis in *M. paradoxum* PRB and acetate production in *Chlorobi* PRB) indicated the importance of acetate as an intermediate substrate during CBM production. We scanned the lower quality, translationally active MAGs for the putative ability to produce acetate via the Ack/Pta pathway and identified members of the *Bacteroidetes* and *Geobacter* with *ack/pta* genes (**Table 1; Supplementary Table 1**). However, the Ack/Pta pathway can be used in reverse during acetate activation to acetyl-CoA. Acetate consumption via Ack/Pta has been demonstrated for *Geobacter sulfurreducens [63]*, suggesting that *Geobacter* in the PRB may compete with *M. paradoxum* PRB for acetate dependent upon availability of potential electron acceptors. Further supporting this hypothesis, Beckmann *et al*. used DNA stable isotope probing to demonstrate carbon assimilation from acetate by *Geobacter* spp. and methanogens together in the same methane-producing coal seam in Australia [37]. In addition to reverse Ack/Pta, pyruvate:ferredoxin oxidoreductase (PFOR) has recently been suggested by *in silico* analysis to generate pyruvate from acetate in a single step in *G. sulfurreducens [64]*, representing another pathway for acetate utilization. PFOR is common among anaerobic microorganisms for the reversible oxidation of pyruvate to acetyl-CoA [65, 66]. All three bacterial MAGs (*Geobacter* PRB, *Chlorobi* PRB, and *Bacteroidetes* PRB) encode at least one PFOR as well as Ack/Pta; however, given the demonstrated reversibility of the respective reactions, directionality is difficult to predict for *in situ* conditions. Recent work has shown that the direction of the Ack/Pta pathway in *Escherichia coli* is determined by thermodynamic controls [67], suggesting redox conditions and/or metabolite availability in the Flowers-Goodale coal seam may ultimately determine whether these bacterial populations consume or produce acetate for methanogenesis by *M. paradoxum* PRB.

Deduced polypeptide sequences for *Bacteroidetes* and *Geobacter* MAGs were scanned for aerobic and anaerobic hydrocarbon degradation enzymes. Similar to *M. paradoxum* PRB, both ~~"~~*Bacteroidetes* PRB~~"~~ and ~~"~~*Geobacter* PRB~~"~~ encoded the Elh enzyme suggesting aerobic phenylacetate degradation, and all *elh* genes were on contigs taxonomically confirmed by total nucleotide matches to the same taxonomic groups. In terms of anaerobic hydrocarbon metabolism, *Bacteroidetes* PRB and *Geobacter* PRB both ~~encoded~~ had multiple copies of the gene encoding Ped for the degradation of phenylethanol. *Bacteriodetes* PRB, like *Chlorobi* PRB, encoded a phenylphosphate carboxylase (Ppc), while *Geobacter* PRB encoded the phenylphosphate synthase (Pps, like *M. paradoxum* PRB) and the Ebd for the degradation of ethylbenzene (like *Chlorobi* PRB). Members of the *Bacteroidetes* ~~is a large group of~~

~~phylogenetically diverse bacteria that can be~~have been associated with complex carbon turnover in suboxic to anoxic environments sometimes associated with methanogenesis [68, 69]. *Geobacter* sequences and/or organisms have been observed in different environments associated with the turnover of recalcitrant carbon and/or methanogenesis. For example, in recent studies, *Geobacter* were shown to be increased with biochar samples and increased methanogenesis [70], correlated to decreased polyphenolics/polycyclic aromatics in methanogenic rice paddy soils [71], and shown to catalyze the turnover of organic matter associated with Fe (hydr)oxides [72].

Our genome-resolved analyses of the translationally active community in the PRB subsurface reveal a conceptual model describing important populations and their associated biochemical capacities that contribute to microbial CBM production (**Figure 4**). By incubating coal down-well in an SES for nine months and allowing establishment of a coal-dependent microbial community under *in situ* methanogenic conditions, the coal-community was secured at depth before retrieval, and then retrieved to the surface in a sealed chamber. *M. paradoxum* PRB is likely a key methanogen in the PRB subsurface with the genomic potential to convert acetate to methane, and this population apparently becomes active in the presence of crushed coal *in situ*. Sources of acetate are likely derived from *Chlorobi* PRB via the Pta-Ack pathway, with possible additional contributions coming from *Bacteroidetes* and *Geobacter* populations, though these populations may also consume acetate depending on environmental conditions. Together, all four translationally active populations (*M. paradoxum* PRB, *Chlorobi* PRB, *Bacteroidetes* PRB, and *Geobacter* PRB) have the combined genomic potential for the anaerobic degradation of ethylbenzene, phenylphosphate, phenylethanol, toluene, xylene, and phenol. *M. paradoxum* PRB, *Bacteroidetes* PRB, and *Geobacter* PRB have the additional potential to break down phenylacetate under micro-aerobic conditions. Finally, certain hydrocarbon degradation enzymes are linked by related pathways, such as Ebd and Ped, which catalyze conversions of ethylbenzene to phenylethanol and subsequently phenylethanol to acetophenone, respectively. All four MAGs possess the *ped* gene but only *Chlorobi* PRB and *Geobacter* PRB possess strong matches to the *ebd* gene. These data provide insights into a coalbed methane community that is likely metabolically interconnected in which hydrocarbon conversions by certain community members stimulate downstream conversions by others.

Conclusions

Subsurface environments associated with different forms of hydrocarbons account for up to 10^{13} metric tons of carbon globally that can be ultimately recycled back to CO_2 and CH_4 [73]. Investigations into how microbial communities interact to complete different stages of carbon remineralization in these environments—from initial interactions and degradation of complex aromatics to ultimately the production of methane and carbon dioxide from precursor metabolites—can provide insight for potential contributions to the global carbon cycle with impacts ranging from climate change to the energy sector. Unfortunately, very little is known regarding many of the steps associated with the degradation of recalcitrant hydrocarbons in the subsurface under *in situ* conditions as these environments are extremely difficult to sample and many of the associated microorganisms are not known and/or not in cultivation. To this end, we used a unique SES device to allow an ecologically relevant community to establish under *in situ* conditions (enriched on crushed coal in the subsurface), and then used novel techniques in activity-based metagenomics to identify translationally active members of the microbial

community. Our observations indicate that in this coal-bearing subsurface ecosystem, specific microbial populations facilitate the biological conversion of coal degradation products to methane using acetate as a key intermediate. Genomic analyses of *Chlorobi* PRB (and perhaps *Bacteroidetes* PRB and *Geobacter* PRB) suggested putative abilities to degrade aromatic hydrocarbons (anaerobically or aerobically) and produce acetate for the subsequent production of methane by the putative acetoclastic methanogen, *Methanothrix paradoxum* PRB. Consequently, these microbial populations may play crucial roles cycling carbon in a shallow subsurface coal seam environment that contributes to the conversion of coal to methane gas.

Materials and Methods

Down-well sampling, BONCAT incubations

The following methods are also presented in a companion study that is a broad-scope analysis of microbial coal degradation processes (*e.g.*, fumarate addition, biosurfactant production) at multiple sites in the PRB under varying sulfate conditions [35]. In September 2017, an SES (Patent # US10704993B2) was loaded with UV-sterile crushed coal, lowered by cable to a depth of 115 m in the FG11 well in the PRB (at the U.S. Geological Survey's Birney site) [6], and opened by a control box at the surface. After nine months of down-well incubation the SES was closed, forming a gas-tight chamber, and retrieved to the surface. 10-ml SES slurries were extracted through a Swagelok device (Solon, Ohio, USA) and anoxically transferred into sterile balch tubes (95% N_2, 5% CO_2) in triplicate. L-homopropargylglycine (HPG, Click Chemistry Tools, Scottsdale, Arizona, USA) was prepared in sterile degassed water (DEPC diethyl pyrocarbonate treated filter sterilized water, pH 7) and added to each replicate at a final concentration of 250 µM. Higher HPG concentrations were used compared to previous studies (*e.g.,[33, 34]*) to overcome loss of the bioorthogonal amino acid due to sorption to porous coal. To ~~account for~~ control for sorting artifacts ~~control~~ samples were prepared the same way, except did not have HPG added (HPG negative control). All samples were incubated in the dark at 20°C for 24 hrs (compared to an *in situ* temperature range of 16 – 18°C, **Supplementary Table 3**). We note that the BONCAT methodology requires relatively short incubation times (< 48 hrs [33, 34]) to prevent over-labeling and/or cross-labeling. In this case a 24-hr incubation was selected based on experimental verification of identifiable cells by fluorescence microscopy and in attempt to minimize bottle effects for the subsamples removed from the SES. Following incubation, cells were removed from coal according to the protocol described by Couradeau *et al*. [33]. Briefly, 1 ml of slurry was removed and added to Tween® 20 at a final concentration of 0.02% (Sigma-Aldrich) in phosphate saline buffer (1X PBS). Samples were then vortexed at maximum speed for 5 min followed by centrifugation at 500 xG for 5 min [33]. The supernatant was removed, filtered through a 40-µm strainer, and spun at 14,000 xG to concentrate detached cells. The cell pellet was immediately cryopreserved at −20°C in a sterile 55% glycerol TE (11X) solution.

Fluorescent labeling, cell sorting, amplification, and metagenomic sequencing

The click reaction buffer consisted of copper sulfate ($CuSO_4$ 100 µM final concentration), tris-hydroxypropyltriazolylmethylamine (THPTA, 500 µM final concentration), and FAM picolyl azide dye (5 µM final concentration) [32]. For the click reaction, each sample (200 µl) was placed on a 25-mm 0.2-µ~~u~~m polycarbonate filter resting on a microscope slide, and 80 µl of

BONCAT click reaction was added before covering with a coverslip. The BONCAT click reaction consisted of 5 mM Sodium Ascorbate, 5mM Aminoguanidine HCl, 500 μM THPTA, 100 μM $CuSO_4$, and 5 μM FAM picolyl azide in 1X phosphate buffered saline. Incubation time was 30 min, followed by three washes in 20 ml of 1X PBS for 5 min each. Cells were recovered from the filter by vortexing in 0.02% Tween for 5 min, and then stained using 0.5 μM SYTO™59 (ThermoFisher Scientific, Invitrogen, Eugene Oregon, USA) DNA stain.

For cell sorting a BD-InfluxTM (BD Biosciences, San Jose, California, USA) specifically configured to capture total cells (SYTOTM59 [excitation = 622 nm, emission = 645 nm]) in the red region of a 640-nm laser and BONCAT active cells (FAM picolyl azide dye [excitation= 490 nm/emission = 510 nm]) in the green region of a 488-nm blue laser. The total cell population was gated for BONCAT positivity by comparing the 530/40 BP fluorescence off a 488-nm laser against an HPG negative control that had undergone the same click reaction. Two fractions (total cells and BONCAT active cells) were sorted from each replicate sample. The first faction consisted of the DNA+ cells, and the second only contained BONCAT+ cells as determined by comparison to the HPG negative control. Fractions were sorted into 394 well plates, and for each fraction 5,000 cells were collected into 4 wells and 300 cells were collected into 20 well. Following sorting plates were frozen at −80°C until further processing.

Cells were pelleted from wells containing 5,000 cells via centrifugation (6,000 xG for 1 hr at 10°C), followed by removal of the supernatant and a brief inverted spin at 6 xG. This step was necessary to avoid interference with subsequent whole genome amplification reaction chemistry. Wells containing only 300 sorted cells were not pelleted, rather were directly lysed and amplified using 5 μl WGAX reactions following optimized conditions [74]. Briefly, cells were lysed in 650 nl lysis buffer for 10 min at room temperature. The lysis buffer consisted of 300 nl TE + 350 nl of 400 mM KOH, 10 mM EDTA, and 100 mM DTT. Lysis reactions were neutralized by the addition of 350 nl of 315 mM HCl in Tris-HCl. Whole genome amplification reactions were brought to 5 μl with final concentrations of 1X EquiPhi29 reaction buffer (ThermoFisher), 0.2 U/μl EquiPhi29 polymerase (Thermo), 0.4 mM dNTPs, 50 μM random heptamers, 10 mM DTT, and 0.5 μM SYTO13. Plates were incubated at 45°C for 13 hr. Libraries for metagenomic sequencing were created using the Nextera XT v2 kit (Illumina) with 12 rounds of PCR amplification. All volumes and inputs to Nextera reactions were reduced 10-fold from the manufacturer's recommendations. Libraries were sequenced 2x150 bp mode on the Nextseq platform (Illumina).

Metagenomic assembly and binning

Raw metagenomic short reads were quality filtered using illumina-utils [75] (v1.0) with default parameters. Technical sequencing replicates were coassembled with MEGAHIT [76] (v1.2.9) for each of three biological replicates for both BONCAT-active and TOTAL sorted cells, resulting in six metagenome assemblies. These three replicate assemblies from BONCAT-active and the additional three replicates from TOTAL cell fractions were further coassembled via MEGAHIT, resulting in a single metagenome assembly for BONCAT and another for TOTAL. Assembled sequences were filtered at a minimum length cutoff of 5,000 bp and binned in anvi'o [77] (v6) based on tetranucleotide frequencies with a scaffold split size of 20,000 bp. Genome bins were scanned for single copy genes to estimate completeness and redundancy and bins were refined until estimated redundancy was < 10%. PyANIp [78] (v0.2.10) was used to compare genomic bins between BONCAT and TOTAL assemblies, and bins with alignment

lengths > 85% and average nucleotide identities (ANI) greater than 95% were considered the same microbial population recovered from both BONCAT and TOTAL. Anvi'o was used to summarize additional genomic information of all bins, including % G+C, N50, number of contigs, and cumulative sequence length.

Environmental detection of active genome bins

Three months after the BONCAT-metagenomics experiment, additional coal-enriched samples were retrieved from the FG11 well and FGP—a nearby, hydrologically connected well —for standard shotgun metagenomic sequencing. As a background control, samples were also collected from a non-methane-producing coal seam well (N11). DNA was isolated from these samples using the MP Biomedical ProDNA Spin Kit for Soil according to the manufacturer's protocol. Metagenomic sequencing was performed by the U.S Department of Energy Joint Genome Institute and raw reads were quality-filtered as above. Bowtie2 [79] (v2.2.6) was used to map FG11, FGP, and N11 short reads against BONCAT-active genome bins to retrieve sequence coverage values for these bins directly from the environment. We also downloaded metagenomic data from a methane-producing coal seam in the Surate Basin, Australia (CX10), for additional comparison (NCBI SRA: SRX1122679) [42]. Total sequence content in quality-filtered fastq files from FG11 (19.5 Gb), FGP (25.2 Gb), N11 (11.5 Gb), and CX10 (8.9 Gb) was used to normalize coverage values for comparisons across sites.

Phylogenetic analyses and taxonomic designations

For genomic bins with the highest estimated completeness *(i.e., M. paradoxum* PRB, *Chlorobi* PRB) as well as taxonomically related reference genomes, 16 ribosomal protein sequences were extracted via anvi'o and concatenated in the following order: L27A, S10, L2, L3, L4, L18p, L6, S8, L5, L24, L14, S17, S3_C, L22, S19, L16RP. Muscle [80] (v3.8.31) was used to align concatenated ribosomal protein sequences with eight maximum iterations. Phylogenetic analyses of aligned concatenated proteins were performed for archaea and bacteria with MrBayes [81] (v3.2.6) using a fixed aa model, empirical aa frequencies, eight gamma distribution categories, eight parallel chains, and a burn-in fraction of 0.25. We ran 100,000 generations for the archaeal analysis and 1,000,000 generations for the bacteria, resulting in standard deviations in split frequencies of 0.000235 and 0.000000, respectively. Taxonomies of the other bins were determined by nucleotide sequence comparisons with BLASTn [82] to the NCBI non-redundant database [83]. *Bacteroidetes*_PRB_Bin13 and *Geobacter*_PRB_Bin11 bins did not contain enough ribosomal proteins for phylogenetic analysis and were instead assigned rank level taxonomy based on nearest matches to each contig within each bin. The majority of contigs in Bin 11 (82 of 151) had strong matches to members of the *Geobacter*, while the remaining hits were closely related members of the *Deltaproteobacteria* phylum. In contrast, Bin 13 could not be resolved beyond the phylum level. Only 79 of the 126 contigs in Bin 13 had hits to the NCBI database and those hits were scattered amongst diverse members within the *Bacteroidetes* phylum (*e.g.*, *Flavobacterium*, *Draconibacterium*, *Sphingobacteriaceae*).

An additional Bayesian phylogenetic tree was calculated for the Cdh enzyme subunit alpha (approximately 800 aa in length) from methanogens. First, Cdh sequences were aligned with MAFFT [84] (maxiterate 1000, localpair) and trimmed with BMGE [85] (BLOSUM30). We used MrBayes [81] to calculate the Bayesian tree with a mixed amino acid model. The

standard deviation of split frequencies was 0.0000 after 1,000,000 iterations and all posterior probabilities were 1.00.

Metabolic analysis

Prodigal [86] was used in the anvi'o platform to identify open reading frames within genomic sequence. Initial functional annotations were conducted using the KEGG database [87] with deduced amino acid. METABOLIC [88] was also used for identification of functional genes related to major biogeochemical cycles. The *Chlorobi* PRB genome was further examined in direct comparison to the NICIL-2 genome using EC pathway annotations focused on important attributes of the *Chlorobi* phylum outlined by Hiras *et al*. [41] We used HMM scans of deduced protein sequences against the AromaDeg database [89] to uncover enzymes associated with aerobic aromatic hydrocarbon degradation. Similarly, for anaerobic conversions of aromatic hydrocarbons we scanned deduced proteins for enzymes in the AnHyDeg database [55]. For genes of interest (*e.g*., related to methane metabolism, Pta-Ack pathway, phenylacetate degradation) we examined host contigs to confirm taxonomic calls by using BLASTn [82] against the NCBI non-redundant database [83]. Contigs were further examined for neighboring genes related to the same microbial process (*e.g*., subunits of the same enzyme).

Geochemical analyses

Water samples analyzed for pH, temperature, CH_4, and $\delta^{13}C$-CH_4 were collected with a Grundfos submersible pump after three wellbore volumes were pumped and field properties stabilized and were analyzed as previously described [6, 27]. Samples for O_2 and other gases were collected with an SES. The internal substrate chamber of the SES was removed, and the SES was slowly dropped down-well to the center of the well screen depth to collect gas and water samples. The SES remained open at the center of the well screen for several minutes before closing the SES and retrieving to the surface. Gas concentrations were measured using a headspace equilibration technique developed by Isotech Laboratories, Inc. (a Stratum Reservoir brand) as described previously [90] with detailed analysis information available through Isotech Laboratories (www.isotechlabs.com). Acetate concentrations were measured by previously reported methods [91].

Conflict of Interest Statement

The authors declare no conflict of interest.

Data accessibility

Genomic sequence data associated with Total-sorted and BONCAT-sorted cells are available on the Integrated Microbial Genomes & Microbiomes (IMG) site under GOLD Study ID Gs014100. High quality MAGs for *Methanothrix paradoxum* PRB and *Chlorobi* PRB were submitted to JGI under GOLD Analysis IDs Ga0496496 and Ga0496497, respectively. Environmental metagenomes for FG11 and FGP wells are available on IMG under GOLD Project IDs Gp0406117 and Gp0406116, respectively.

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Tables:

Table 1 | Genomic characteristics of translationally active MAGs enriched on crushed coal

								Environmental Detection (Relative Coverage)			
Genomic Population	Length (Mb)	Compl (%)	Redund (%)	N50	G+C (%)	Contigs	Genes	PRB High-CH4 coal seam (FG11)	PRB High-CH4 coal seam (FGP)	PRB Low-CH4 coal seam (N11)	Surat Basin High-CH4 coal seam (CX10)
Chlorobi PRB (Bin8)	**3.34**	**97.2**	**1.4**	**34,774**	**37.5**	**164**	**2,777**	**4.1**	**0.3**	**0.1**	**0.3**
Chlorobi_NICIL-2*	2.67	95.3	N/D	168,929	56.0	152	2,363	N/A	N/A	N/A	N/A
***Methanothrix paradoxum* PRB (Bin15)**	**2.90**	**97.4**	**2.6**	**34,925**	**50.7**	**145**	**2,946**	**22.1**	**153.4**	**0.0**	**0.1**
Methanothrix paradoxum NSM2**	1.75	92.1	1.3	9,272	54.7	238	1,921	N/A	N/A	N/A	N/A
Methanothrix paradoxum ASM2**	1.17	72.4	0.0	4,915	54.9	249	1,350	N/A	N/A	N/A	N/A
Methanothrix soehngenii GP6***	3.03	97.4	0.0	3,008,626	51.9	2	2,925	N/A	N/A	N/A	N/A
Bacteroidetes PRB (Bin13)	**1.95**	**38.0**	**5.6**	**14,966**	**44.7**	**237**	**1,618**	**12.4**	**41.5**	**2.6**	**0.1**
Geobacter PRB (Bin11)	**3.04**	**40.9**	**1.4**	**12,174**	**52.5**	**393**	**2,829**	**17.7**	**32.5**	**10.5**	**0.3**

***from Hiras et al., 2016**
****from Angle et al., 2017**
*****from Patel & Sprott, 1990**

Length in megabase, estimated percent completeness and redundancy, N50, percent G + C content, number of contigs, and number of genes are provided are provided for each MAG analyzed in this study and compared to reference genomes of Chlorobi NICIL-2[41] and *M. paradoxum strains* (or MAGs) NSM2[36], ASM2[36], and GP6[54]. Environmental coverage values are displayed for quality-filtered short reads from wells FG11 and FGP mapped to each MAG. The three largest environmental metagenomes (FG11, FGP, N11) were scaled in size to the smallest metagenome (CX10) prior to relative coverage calculations.
~~Total FG11 reads were normalized to total FGP reads so environmental coverages could be compared.~~

Table 2 | Hydrocarbon degradation genes detected in translationally active coalbed populations

Population	*mcr*	*cdh*	*acs*	*ack*	*pta*	*Aerobic hydrocarbon degradation*	*Anaerobic hydrocarbon degradation*
***M. paradoxum* PRB (Bin15)**	yes	yes	yes			ELH*	PpsB*, BssD-p* Ped*, PpsB*, EbdA*, Ped*, PcmI, ApcA*
***Chlorobi* PRB (Bin8)**				yes	yes	ELH	PpcB, EbdD, Ped, Ped, Ped, Ped, Ped, Ped, CmdA, CmdB, CmdB
***Bacteroidetes* PRB (Bin13)**			yes	yes	yes	ELH*, ELH*	PpcB, Ped, PpcB, Ped, Ped, Ped, Ped, Ped
***Geobacter* PRB (Bin11)**				yes	yes	ELH*	PpsB*, EbdB*, Ped*, Ped*, Ped*, PpsB*, Ped*, ApcC*, EbdB*, PcmI*, EbdA*, CmdA*

Asterisk indicates the gene encoding this putative enzyme is on a contig with a nucleotide BLAST identity that matches the overall taxonomic identity for the MAG. Amino acid identities (AAID) to sequences in the hydrocarbon degradation databases are indicated by color (blue > 40% AAID, red > 30% AAID, and grey > 20% AAID). [**ELH** -- Extradiol Dioxygenase, LigB Superfamily, Homoprotocatechuate; **ApcA** -- Acetophenone carboxylase alpha; **ApcC** -- Acetophenone carboxylase gamma; **CmdA** -- Cymene Dehydrogenase alpha; **CmdB** -- Cymene Dehydrogenase beta; **EbdA** -- Ethylbenzene Dehydrogenase alpha; **EbdB** -- Ethylbenzene Dehydrogenase beta; **PcmI** -- p-Cresol Methylhydroxylase alpha subunit isoform; **Ped** -- Phenylethanol dehydrogenase; **PpsB** -- Phenylphosphate synthase subunit B; **BssD**-p -- Putative BssD (benzylsuccinate synthase activase); **PpcB** -- Phenylphosphate Carboxylase beta; ***mcr*** – methyl coenzyme M reductase; ***cdh*** – carbon monoxide dehydrogenase; ***acs*** – acetyl CoA synthetase; ***ack*** – acetate kinase; ***pta*** -- phosphotransacetylase]

Figures:

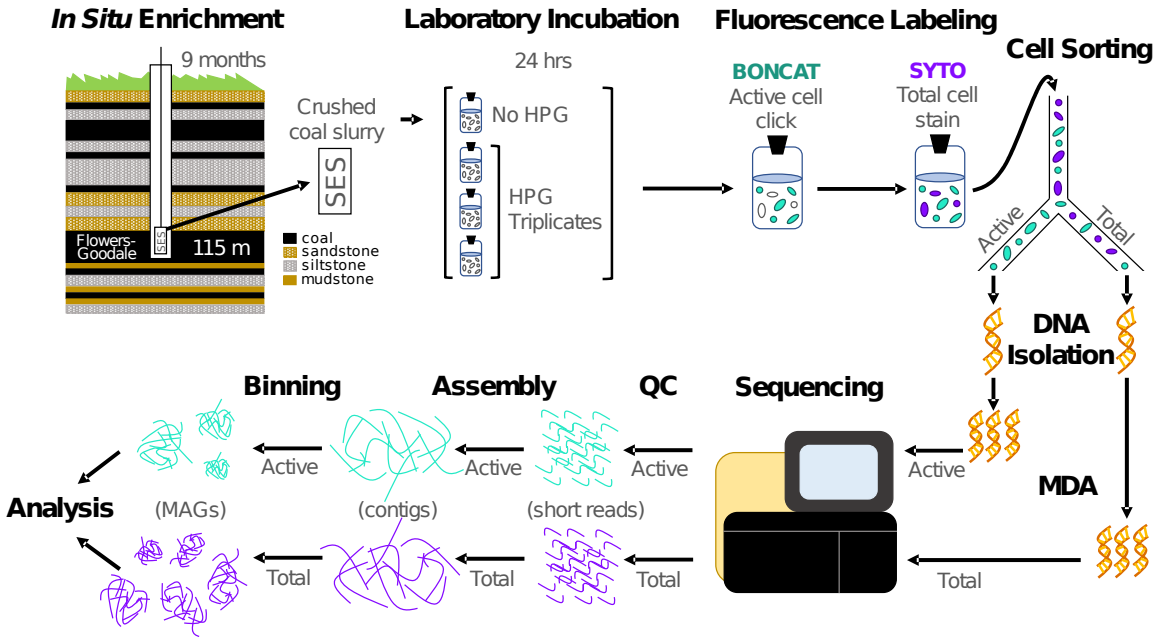


Figure 1 | Conceptual representation of BONCAT-FACS experimental setup and metagenomic sequencing workflow. We performed down-well incubation of sterile, crushed coal in the SES allowing for microbial colonization and retrieval under *in situ* pressure and anaerobic conditions. Samples were allocated into sterile gassed out serum bottles for addition of the bioorthogonal amino acid (HPG) in triplicate 24hr incubations. We then sorted click-labeled BONCAT active cells (FAM Picolyl dye; Ex: 488 nm/Em: 530 nm) and total cells (SYTO59; Ex: 640 nm/Em: 655–685 nm) from each biological replicate. This was followed by DNA extraction, MDA amplification, sequencing, and analysis. The upper left coal seam~~bed~~ stratigraphy panel was modified from Barnhart *et al*., 2016 [6].

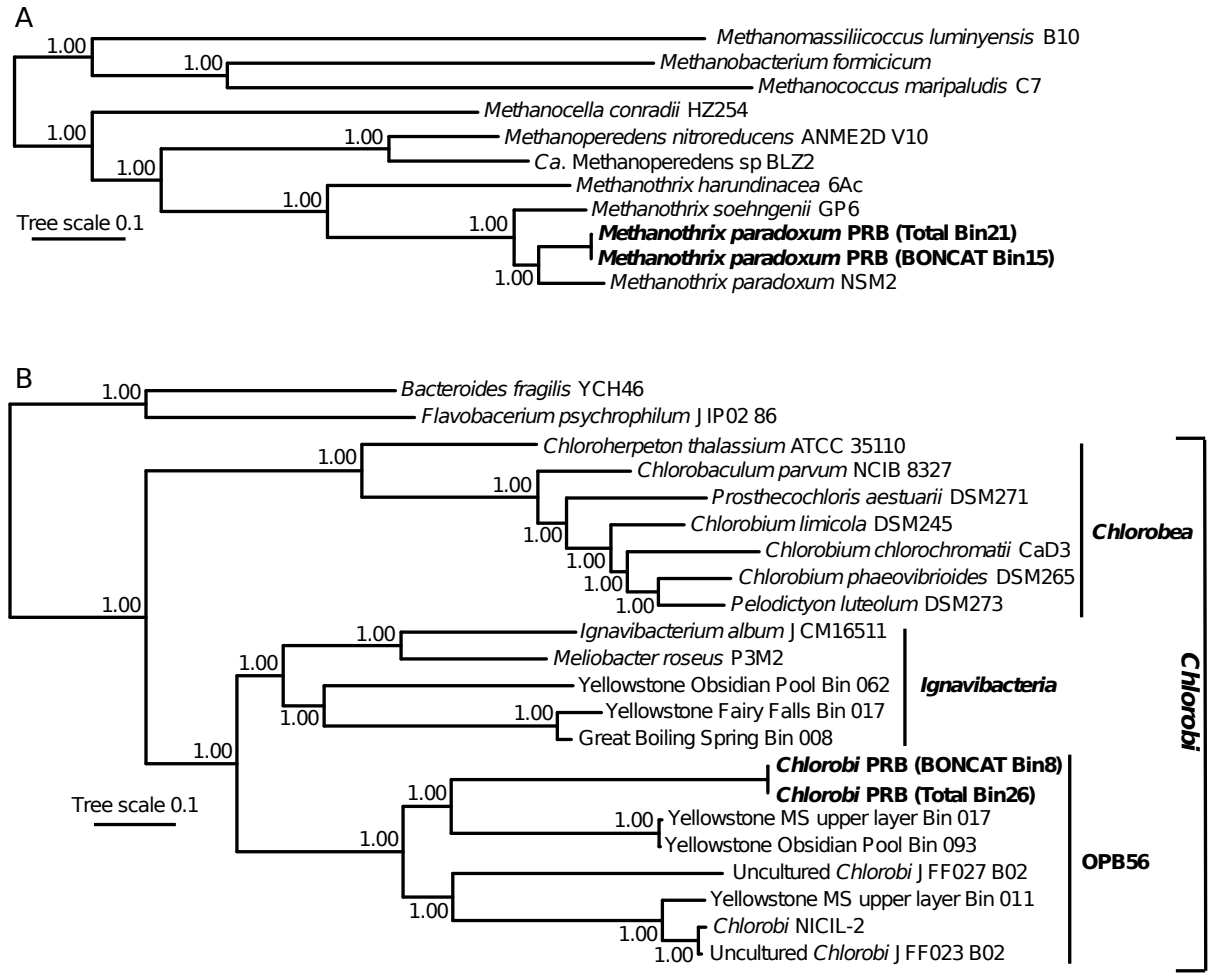
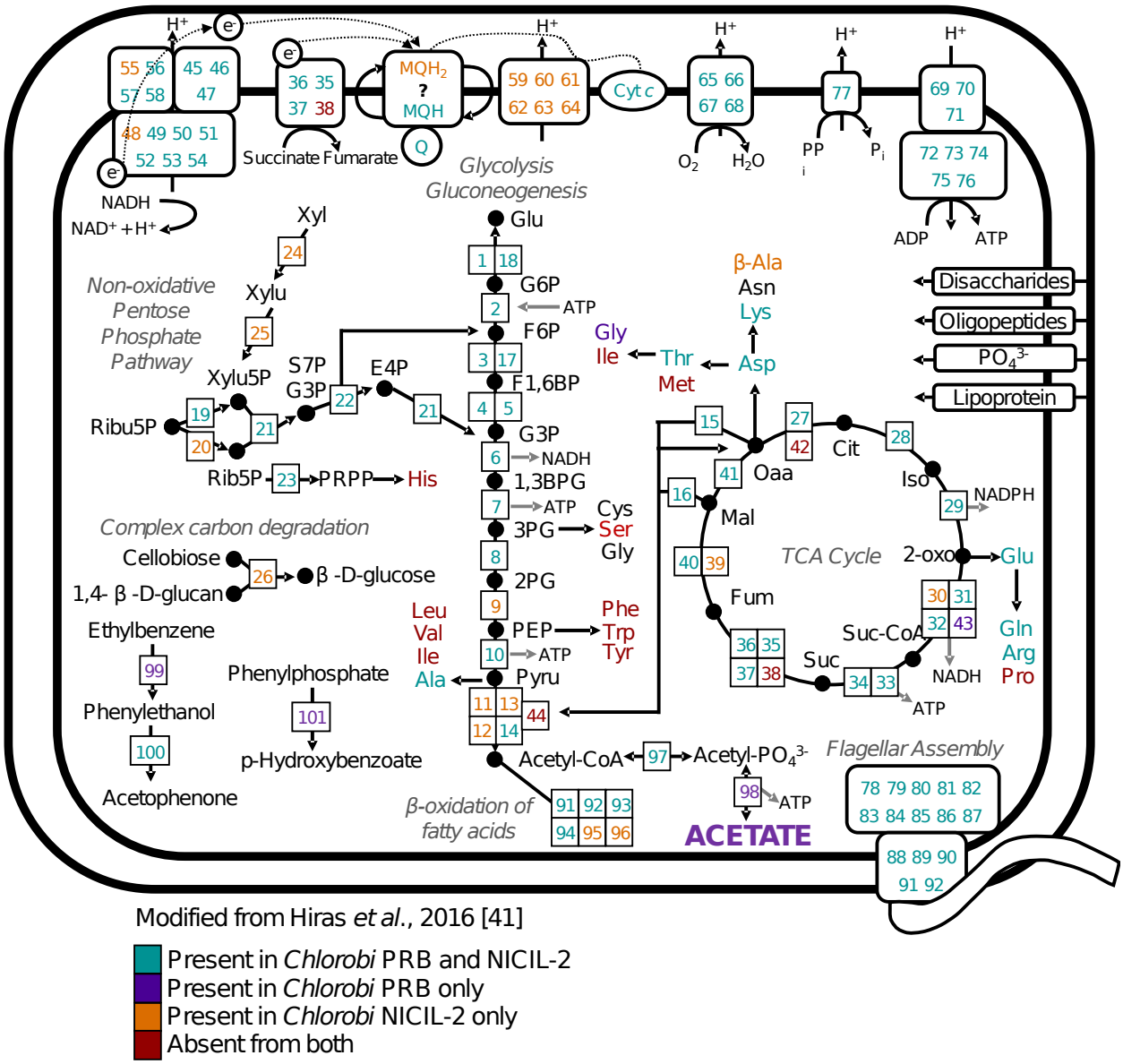


Figure 2 | Phylogenetic positions of *M. paradoxum* PRB (A) and *Chlorobi* PRB (B). Bayesian trees were constructed from concatenated alignments of 16 ribosomal proteins. Posterior probabilities (between 0.00 and 1.00) are displayed at branch nodes. The tree scale represents the average number of substitutions per site.



Modified from Hiras *et al.*, 2016 [41]

Present in *Chlorobi* PRB and NICIL-2
Present in *Chlorobi* PRB only
Present in *Chlorobi* NICIL-2 only
Absent from both

Figure 3 | Metabolic reconstruction of *Chlorobi* PRB compared to *Chlorobi* NICIL-2. Colored enzyme numbers indicate which genome contained the respective gene (teal = *Chlorobi* PRB and NICIL-2; purple = *Chlorobi* PRB only; orange = NICIL-2 only; red = neither). Enzymes are numbered and are defined in Supplementary Table 6. This figure is modified from a previous version published by Hiras and colleagues [41] for comparison with *Chlorobi* PRB. [xyl = xylose; xylu – xyluose; ribu = ribulose; rib = ribose; glu = glucose; pyru = pyruvate; standard three-letter abbreviations are used for amino acids]

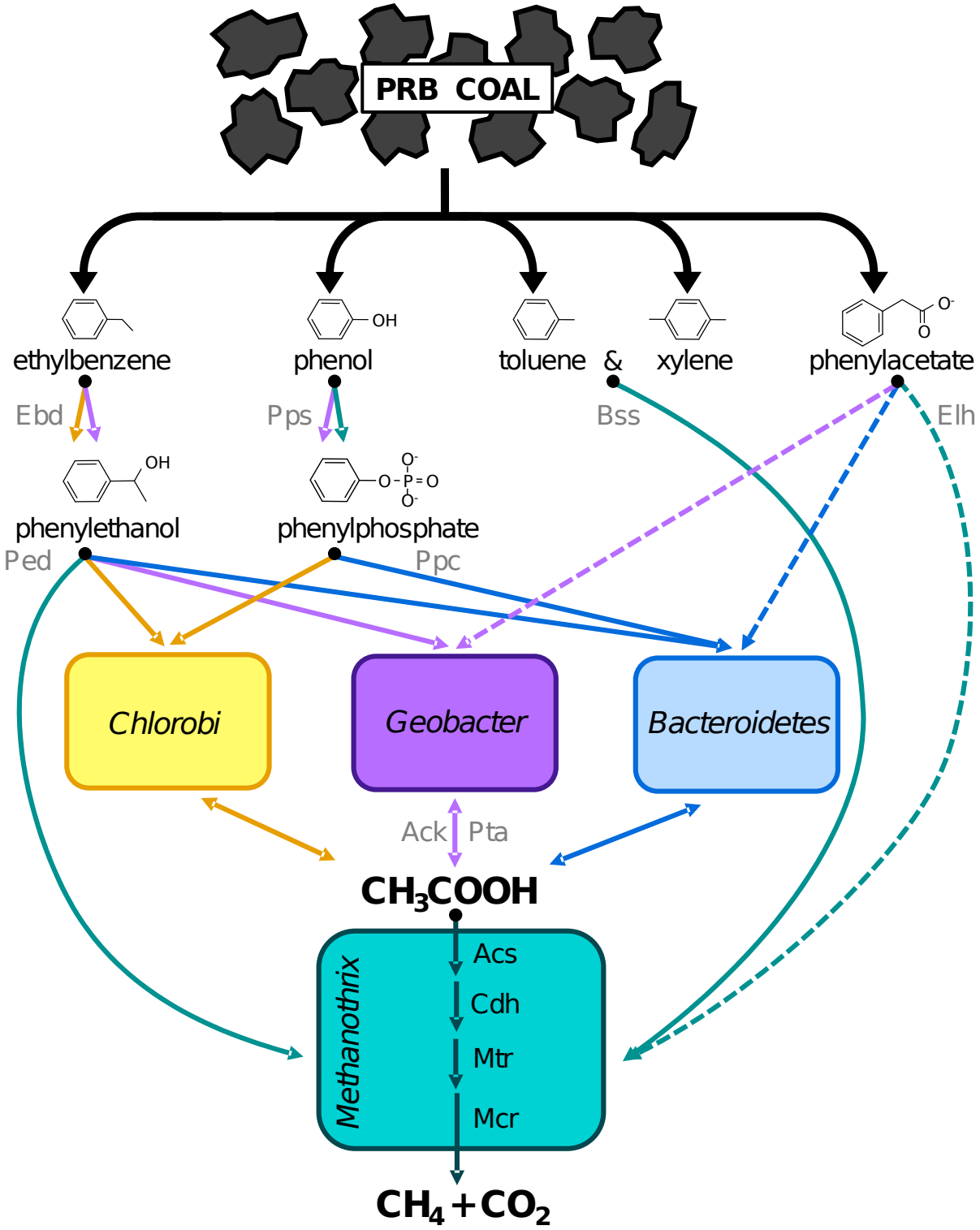


Figure 4 | Biogenic CBM production in the PRB, from aromatic hydrocarbon degradation to acetoclastic methanogenesis. Translationally active MAGs in the PRB harbor the putative ability to degrade a variety of aromatics, including ethylbenzene, phenylethanol, phenol, phenylphosphate, toluene, xylene, and phenylacetate.

Arrows representing genes or deduced enzymes are colored by the host microbial population (orange = *Chlorobi* PRB, purple = *Geobacter* PRB, blue = *Bacteroidetes* PRB, teal = *M. paradoxum* PRB) and indicate that either carbon or energy may be derived from the putative reaction. Dashed lines indicate putative oxygen-consuming reactions. Detailed metabolic potential of the *Chlorobi* PRB MAG is presented in Figure 3. [ebd = ethylbenzene dehydrogenase, ped = phenylethanol dehydrogenase, pps = phenylphosphate synthase, ppc = phenylphosphate carboxylase, bss = putative benzylsuccinate synthase, ack = acetate kinase, pta = phosphotransacetylase, acs = acetyl CoA synthase, cdh = carbon monoxide dehydrogenase, mtr = methyltransferase, mcr = methyl CoM reductase, elh = extradiol dioxygenase: LigB superfamily: homoprotocatechuate]

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