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Authors

Walentek, Peter
Hagenlocher, Cathrin
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Publication Date

2015-09-01

DOI

10.1016/j.dib.2015.04.003

Peer reviewed



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Contents lists available at ScienceDirect

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Data Article

ATP4 and ciliation in the neuroectoderm and endoderm of *Xenopus* embryos and tadpoles

Peter Walentek^{a,b,*}, Cathrin Hagenlocher^a, Tina Beyer^{a,1}, Christina Müller^a, Kerstin Feistel^a, Axel Schweickert^a, Richard M. Harland^b, Martin Blum^a

^a Institute of Zoology, University of Hohenheim, Garbenstrasse 30, 70593 Stuttgart, Germany

^b Department of Molecular and Cell Biology, Center for Integrative Genomics, University of California at Berkeley, Berkeley, California 94720, USA

ARTICLE INFO

Article history:

Received 27 March 2015

Received in revised form

4 April 2015

Accepted 7 April 2015

Available online 20 April 2015

Keywords:

Cilia

ATP4a

Gastric H+/K+ATPase

Wnt signaling

Xenopus

ABSTRACT

During gastrulation and neurulation, *foxj1* expression requires ATP4a-dependent Wnt/ β -catenin signaling for ciliation of the gastrocoel roof plate (Walentek et al. *Cell Rep.* 1 (2012) 516–527.) and the mucociliary epidermis (Walentek et al. *Dev. Biol.* (2015)) of *Xenopus laevis* embryos. These data suggested that ATP4a and Wnt/ β -catenin signaling regulate *foxj1* throughout *Xenopus* development. Here we analyzed whether *foxj1* expression was also ATP4a-dependent in other ciliated tissues of the developing *Xenopus* embryo and tadpole. We found that in the floor plate of the neural tube ATP4a-dependent canonical Wnt signaling was required for *foxj1* expression, downstream of or in parallel to Hedgehog signaling. In the developing tadpole brain, ATP4-function was a prerequisite for the establishment of cerebrospinal fluid flow. Furthermore, we describe *foxj1* expression and the presence of multiciliated cells in the developing tadpole gastrointestinal tract. Our work argues for a general requirement of ATP4-dependent Wnt/ β -catenin signaling for *foxj1* expression and motile ciliogenesis throughout *Xenopus* development.

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DOI of original article: <http://dx.doi.org/10.1016/j.ydbio.2015.03.013>

* Corresponding author at: Department of Molecular and Cell Biology, Center for Integrative Genomics, University of California at Berkeley, Berkeley, California 94720, USA.

E-mail address: walentek@berkeley.edu (P. Walentek).

¹ Current address: Medical Proteome Center, Institute for Ophthalmic Research, University of Tübingen, Nägelestrasse 5, 72074 Tübingen, Germany.

<http://dx.doi.org/10.1016/j.dib.2015.04.003>

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Specification Table

Subject area	Biology
More specific subject area	Cell and developmental biology
Type of data	Text file, figures, movies
How data was acquired	Microscopy (fluorescent, confocal, bright-field)
Data format	Analyzed and annotated figures and movies
Experimental factors	NA
Experimental features	Xenopus embryos were manipulated by morpholino oligonucleotide-mediated knockdown and application of pharmacological inhibitors. Gene expression, morphology and cilia function were analyzed by in situ hybridization, immunofluorescence, and quantification of extracellular fluid flow
Data source location	NA
Data accessibility	The data described here is presented in this article in form of figures and supplemental movies

Value of the data

- Our results indicate that the ATP4/Wnt/ β -catenin module is required for neural *foxj1* expression downstream of, or in parallel to, Hedgehog signaling.
- ATP4 function is required for the generation of cerebrospinal fluid flow.
- *atp4a* and *foxj1* are co-expressed in the gastrointestinal tract.
- The tadpole stomach is lined by multiciliated cells, which generate an extracellular fluid flow.

1. Data, experimental design and methods*1.1. Analysis of ATP4a/Wnt-dependent foxj1 expression in floor plate of the neural tube*

The floor plate and the brain represent additional sites of vertebrate *foxj1* expression [1,5,6,13]. We tested whether floor plate expression of *foxj1* required ATP4 and Wnt/ β -catenin signaling in *Xenopus* by injection of 1 pmol/injection of *atp4a* morpholino oligonucleotide (*atp4aMO*) targeted to dorso-medial regions of developing embryos. Embryos were injected at the two- to four-cell stage using a Harvard Apparatus or Picospritzer setup in $1 \times$ modified Barth's solution (MBSH) with 4% Ficoll (BioChemica) and transferred to $0.1 \times$ MBSH 15 min after injection. Gene expression was analyzed by whole mount in situ hybridization (WMISH). *atp4a* morphants showed a reduction of *foxj1* expression in the floor plate ($p < 0.001$; Fig. 1A,C and G), which was rescued by co-injection of $1 \text{ ng}/\mu\text{l}$ β -catenin DNA ($p < 0.01$; Fig. 1E and G).

1.2. Monitoring floor plate formation in atp4a morphants

Formation of the floor plate in *atp4a* morphants was analyzed histologically and by analysis of gene expression. Embryos were embedded in gelatin–albumin and sectioned on a vibratome ($30 \mu\text{m}$). The floor plate was present, as judged by concentration of pigment due to apical constriction of medial neural plate cells, both in *atp4a* morphants and in specimens co-injected with β -catenin DNA (Fig. 2A–C, A'–C'). Floor plate-specific *sonic hedgehog* expression (*shh*; [12]) was also present in *atp4a* morphants (Fig. 2D and E).

1.3. Analysis of Hedgehog-dependent foxj1 expression in the floor plate of the neural tube

To analyze if *Xenopus foxj1* expression depended on Hedgehog (HH) signaling, as reported for zebrafish *foxj1* [3,20], embryos were incubated with the HH signaling inhibitor cyclopamine

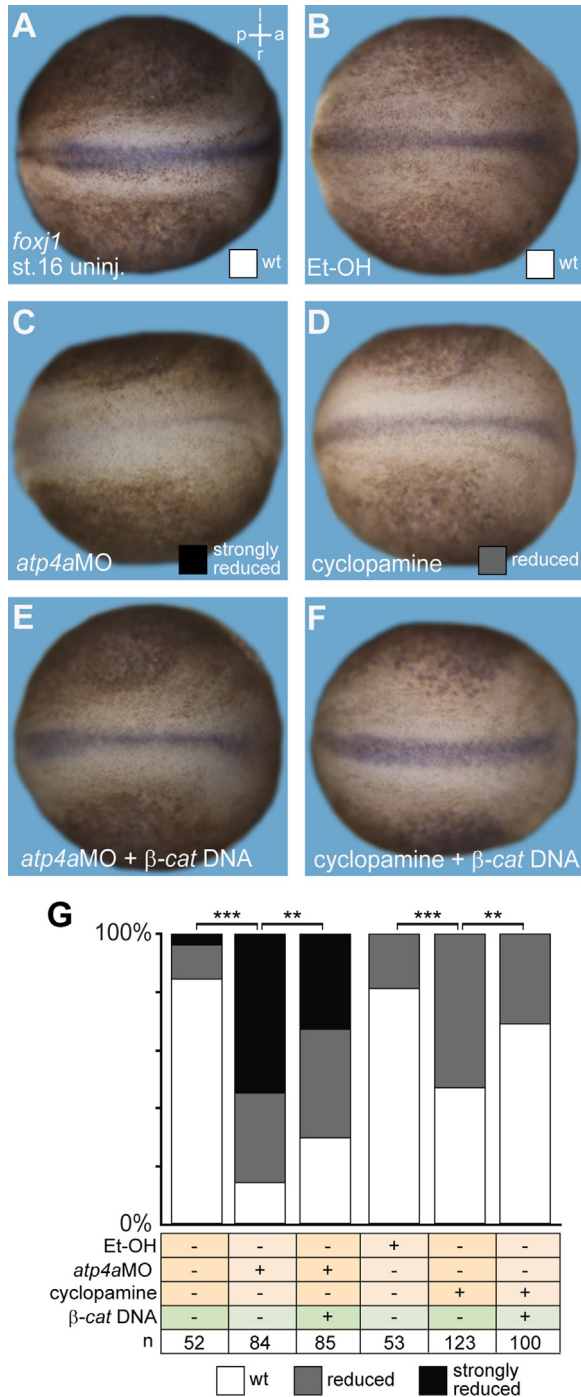


Fig. 1. Floor plate *foxj1* expression requires ATP4a and β -catenin downstream or in parallel of Hedgehog signaling. (A–F) WMISH for *foxj1* expression in control and manipulated embryos at stage 16. (A, B) Normal *foxj1* expression in the floor plate of control uninjected (uninj.; A) and ethanol (1%; EtOH; B) treated specimens. (C) Strong reduction of *foxj1* signals in *atp4a* morphants was partially rescued by co-injection of β -catenin DNA (β -cat.; E). (D) Inhibition of Hedgehog signaling by cyclopamine treatment decreased *foxj1* expression in the floor plate and was partially rescued by injection of β -catenin DNA (β -cat.; F). (G) Quantification of results. a, anterior; l, left; n, number of embryos; p, posterior; r, right; st., stage.

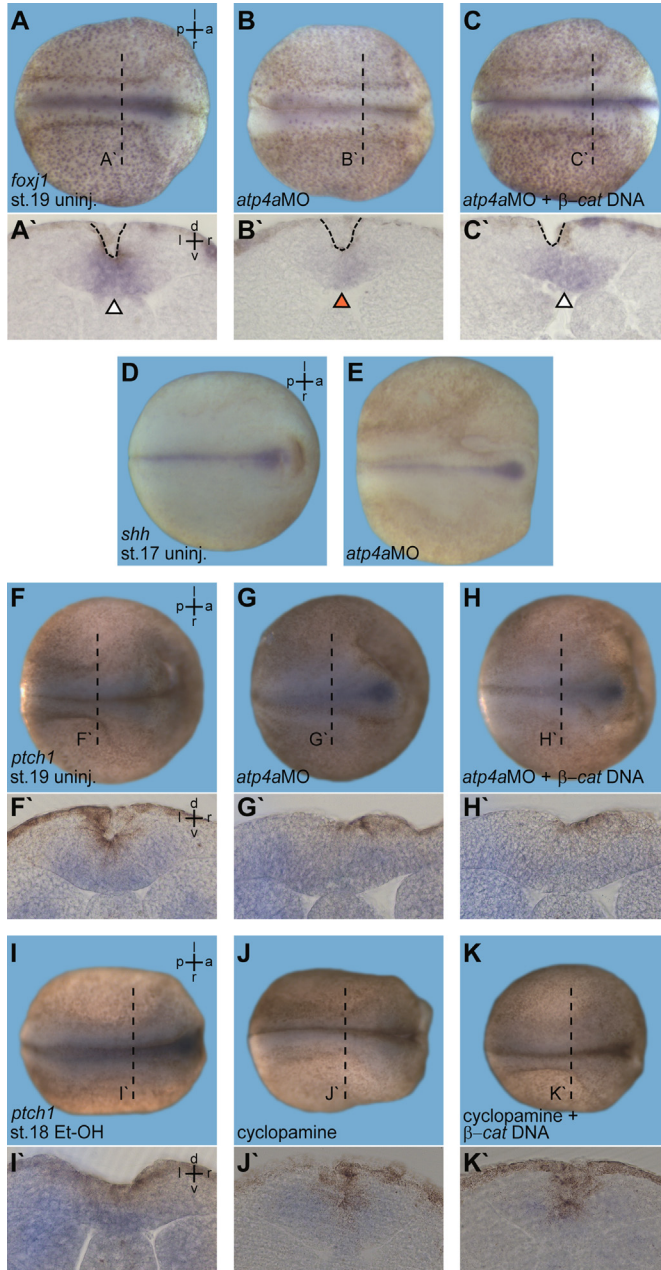


Fig. 2. Normal floor plate induction and Hedgehog signaling in ATP4a-deficient embryos. (A–E) Normal floor plate formation in *atp4a* morphants. WMISH for *foxj1* (A–C) and *shh* (D, E) revealed attenuated *foxj1* expression but unaffected floor plate formation, as judged by apically constricted cells (histological vibratome sections in A–C; planes indicated in A–C) and *shh* expression in *atp4a* morphants. (F–K) Unaffected Hedgehog signaling in *atp4a* morphants with (H) or without (G) co-injection of β -catenin (β -cat.) DNA, as judged by WMISH for *ptch1*, a direct Hedgehog signaling target, to monitor activity of Hedgehog signaling, as compared to uninjected controls (uninj.). *ptch1* expression was decreased in cyclopamine-treated embryos (J, K), as compared to ethanol (1%) controls (I), but independent of β -catenin DNA injection (K, K'). a, anterior; l, left; p, posterior, right; st., stage.

(Selleckchem; solvent: ethanol; concentration: 100 μ M). Incubations were performed according to standard procedures [7] from stage 8 until fixation, on embryos treated with Proteinase K (as described; [15]) in order to permeabilize the fertilization membrane. Proteinase K-treated embryos incubated with corresponding concentrations of ethanol were used as controls. Cyclopamine treatment reduced *foxj1* expression in the floor plate ($p < 0.001$; Fig. 1B, D and G). Injection of β -catenin DNA partially rescued floor plate *foxj1* expression in cyclopamine-treated specimens ($p < 0.01$; Fig. 1F and G).

1.4. Monitoring hedgehog signaling state in *atp4a* morphants and cyclopamine treated embryos

Analysis of *patched 1* (*ptch1*) expression by WMISH was used to monitor HH signaling activity [9]. Embryos were embedded in gelatin–albumin and sectioned on a vibratome (30 μ m). *ptch1* expression was downregulated in cyclopamine treated specimens, but unaffected by *atp4a*MO or β -catenin DNA injection (Fig. 2F–K).

2. Analysis of cerebrospinal fluid flow in *atp4a* morphant tadpoles

To investigate whether ATP4 was required for brain cilia, we analyzed cerebrospinal fluid (CSF) flow as a proxy. For imaging and calculation of ependymal flow cf. Hagenlocher et al. [6] and Walentek et al. [18]. To facilitate late analysis of *atp4a* morphants, we used 1 pmol/injection of a splice-site MO (*atp4a*SplMO) which targeted the second exon/intron boundary of zygotically expressed pre-mRNA. Injection of fluorescent beads into the brain ventricles at stage 45 revealed a significant reduction of CSF flow velocity in *atp4a* morphants ($p < 0.001$; Fig. 3; Movie 1). In contrast, velocity of CSF flow in *atp4a* morphants was increased by co-injection of either *atp4a* or *foxj1* DNA constructs ($p < 0.05/0.001$; Fig. 3; Movie 1).

Supplementary material related to this article can be found online at <http://dx.doi.org/10.1016/j.dib.2015.04.003>.

2.1. Analysis of *atp4a*SplMO-induced intron retention

4-cell stage embryos were injected four times with 1 pmol/injection of *atp4a*SplMO into the vegetal halve (Fig. 5O), thereby targeting the developing gastrointestinal tract including the stomach, where zygotic *atp4a* expression was previously confirmed [17]. At stage 45 total RNAs were isolated from injected and uninjected tadpoles and *atp4a* intron 2 retention was confirmed (Fig. 4) by standard RT-PCR using the following primers on cDNA and genomic DNA extracts:

*atp4a*Ex2-for 5'-GCATGAAAAAATGGAC-3';
*atp4a*Int2-rev 5'-TCCTGTCTGCCAATAAACCC-3';

RT-PCR for *elongation factor 1 α* (*ef1a*) was used as loading control employing the following primers:

forward 5'-CAGATTGGTCTGGATATGC-3';
 reverse 5'-ACTGCCTTGATGACTCCTAG-3'.

3. Analysis of *atp4a* and *foxj1* expression in the gastrointestinal tract

High levels of *atp4a* transcripts were found in the stomach of the tadpole (Fig. 5A and C), where ATP4 localization and function have been previously described [8]. We also observed weaker *atp4a* expression in the embryonic esophagus and the proximal small intestine (Fig. 5D). Analysis of *foxj1* mRNA transcription in stage 45 tadpoles revealed expression in the very same regions of the gastrointestinal (GI) tract (Fig. 5E and F).

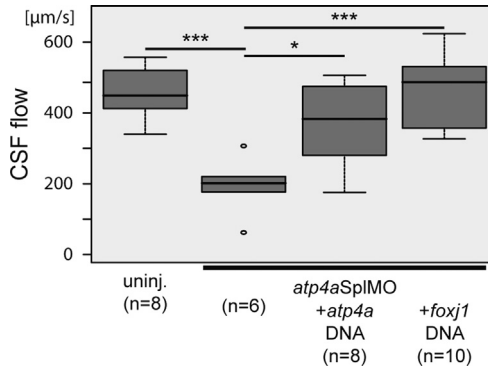


Fig. 3. ATP4a is required for *foxj1*-dependent cerebrospinal fluid in the tadpole brain. To investigate whether ATP4 was required for brain cilia, we analyzed cerebrospinal fluid (CSF) flow as a proxy. Injection of fluorescent beads into the brain ventricles at stage 45 revealed a significant reduction of CSF velocity in *atp4a* morphants. In contrast, velocity of CSF flow in *atp4a* morphants was partially rescued by co-injection of either *atp4a* or *foxj1* DNA constructs. cf. Movie 1.

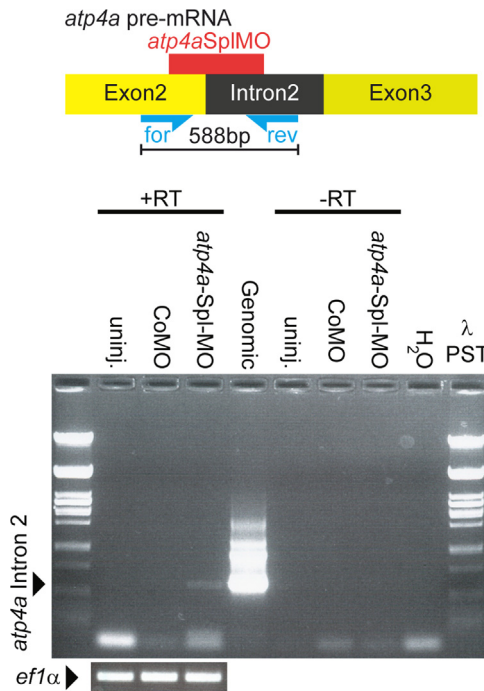


Fig. 4. The *atp4a* splice-site MO causes *atp4a* intron 2 retention. To facilitate late analysis of *atp4a* morphants, a splice-site MO (*atp4aSplMO*) was used, which targeted the second exon/intron boundary of zygotically expressed mRNA and caused *atp4a* intron 2 retention. *atp4aSplMO* targeted to gastrointestinal tract caused intron2 retention, as shown by RT-PCR using primers (blue arrows) which bind to exon 2 (yellow) and intron 2 (black). Genomic DNA served as positive control. Total RNA extracts without reverse transcription (-RT) and water (H₂O) served as negative controls. RT-PCR of *ef1α* served as loading control. λ-phage DNA digested with Pst1 (λ PST) served as size marker.

3.1. Analysis of multiciliated cells in the gastrointestinal tract

Gastrointestinal (GI) tract cilia have previously only been reported in the esophagus of *Xenopus* tadpoles [4]. In order to test whether other parts of the GI tract were ciliated, we stained for

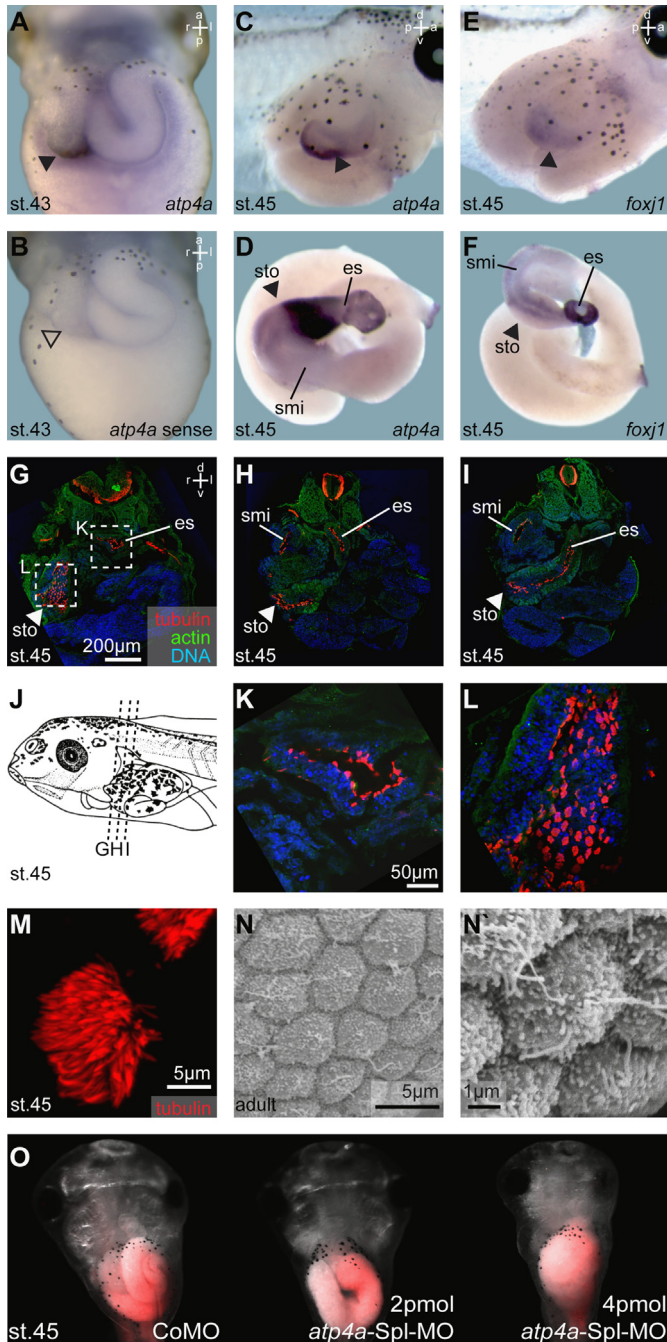


Fig. 5. *foxj1* and *atp4a* are co-expressed in the gastrointestinal tract which transiently harbors motile cilia in the stomach. (A–F) WMISH for *atp4a* (A,C,D) and *foxj1* (E,F) in the GI tract of stage 43–45 tadpoles (A–C, E; stomach highlighted by arrowheads; ventral views) and isolated GI tracts (D, F). (B) *atp4a* sense control revealed no staining. Note the co-expression of *atp4a* and *foxj1* at stage 45 (C–F). (G–M) GI tract ciliation as shown by immunofluorescent staining of cilia/tubulin (acetylated- α -tubulin staining, red) and staining for actin (phalloidin, green) as well as nuclei (DAPI, blue) on cryosections from stage 45 tadpoles (planes indicated in J). MCCs (M) were found in the esophagus (es; G–I,K), stomach (sto; G–I,L) and the proximal part of the small intestine (smi; H, I). (N, N') Scanning electron microscopy analysis of gastric epithelia from adult frogs revealed the presence of short monocolia, but lack of MCCs. (O) Injection of *atp4a*-SplMO targeted to the endoderm prevented normal development of the GI tract. Targeting was monitored by co-injection of fluorescent rhodamine dextrane (red). Embryos are shown in ventral view. a, anterior; d, dorsal; es, esophagus; l, left; p, posterior; r, right; st., stage; smi, small intestine; sto, stomach; v, ventral.

acetylated- α -tubulin by immuno-histochemistry (Fig. 5G–L). GI tract ciliation was analyzed on cryosections (40 μ m) of embryos embedded in O.C.T. (Tissue-Tek) using standard procedures. This analysis confirmed the presence of multiciliated cells (MCCs) in the esophagus and identified the stomach and the proximal small intestine as additional sites of ciliated cells in tadpoles (Fig. 5G–M). In contrast to tadpoles, only short monocilia of about 2 μ m length were detected in the adult gastric epithelium by scanning electron microscopy (Fig. 3N and N').

3.2. Assessment of extracellular fluid flow in the gastrointestinal tract

In order to assess motility of endodermal MCCs, the anterior portion of the GI tract of anesthetized stage 45 tadpoles was dissected and incubated in 0.1 \times MBSH containing benzocaine (Sigma) to prevent peristaltic movements. FITC-conjugated latex beads (FluoSpheres[®] carboxylate-modified microspheres, 0.5 μ m, yellow–green fluorescence (505/515), 2% solids, Life Technologies; diluted to 0.04% in 0.1 \times MBSH) were diluted to 0.04% in 0.1 \times MBSH and applied to the anterior esophagus using a Harvard Apparatus injector. The chamber was sealed and imaged for 2 min using epifluorescent illumination at 10 \times magnification on a Zeiss Axioskop 2 microscope (Movie 2). Beads were transported through the esophagus and stomach until they reached the small intestine, i.e. the region where ciliation starts to decline.

Supplementary material related to this article can be found online at <http://dx.doi.org/10.1016/j.dib.2015.04.003>.

3.3. ATP4a loss of function in the gastrointestinal tract

To test the functional relevance of ATP4a in the context of GI tract ciliation, *atp4a*SpIMO was injected vegetally to target the endoderm (Fig. 5O). Although inhibition of endodermal ATP4a function reduced intestinal *foxj1* expression (not shown), it also interfered with normal development of the GI tract (Fig. 5O), a phenotype reminiscent of failure in Wnt-dependent specification of the proximal GI tract [21].

4. Ethics statement

All animals were treated according to the German regulations and laws for care and handling of research animals, and experimental manipulations according to §6, article 1, sentence 2, no. 4 of the animal protection act were approved by the Regional Government Stuttgart, Germany (Vorhaben A 365/10 ZO “Molekulare Embryologie”).

This work was also done with approval of University of California, Berkeley's Animal Care and Use Committee. University of California, Berkeley's assurance number is A3084-01, and is on file at the National Institutes of Health Office of Laboratory Animal Welfare.

4.1. Statistical evaluation of results

Statistical evaluation of experiments represented by bar graphs was performed using chi-square tests (<http://www.physics.csbsju.edu/stats/contingency.html>). Statistics of experiments represented by box plots were calculated by Wilcoxon sum of ranks (Mann–Whitney) tests (http://www.fon.hum.uva.nl/Service/Statistics/Wilcoxon_Test.html).

4.2. Constructs used for Manipulation of embryos

Morpholino oligonucleotides (MOs) were obtained from Gene Tools:

- *atp4a*MO (5'-GTCATATTGTTCCCTTTTCCCCATC-3') 1 pmol,
- *atp4a*SpIMO (5'-CCCCCCCCCATTTCTTACAATGT-3') 1 pmol.

The following DNA constructs were used for injections after purification using a PureYield Plasmid Midiprep kit (Promega):

- *atp4a*-CS2⁺ MT [17] 1 ng/μl,
- *foxj1*-CS2⁺ [16] 0.5 ng/μl,
- *β-catenin-gfp*-CS2⁺ [10] 1 ng/μl.

Drop size was calibrated to about 7–8 nl per injection. Rhodamine-B or Cascade-blue dextran (0.5–1.0 mg/ml; Molecular Probes) were co-injected and used as lineage tracer.

4.3. Whole-mount *in situ* hybridization

Embryos were fixed in MEMFA for 1–2 h and processed following standard protocols. Digoxigenin-labeled (Roche) RNA probes (*atp4a* and *foxj1*, [17]; *shh* probe was generated according to NM_001088313; *ptch1*, [11]) were prepared from linearized plasmids using SP6, T3, or T7 RNA polymerase (Promega). *In situ* hybridization was conducted following standard procedures.

4.4. Immuno-histochemistry and scanning electron microscopy

Immuno-histochemistry followed standard protocols, using antibodies specific for acetylated- α -tubulin (mouse, 1:700; Sigma), anti-mouse Cy3 (sheep, 1:250; Sigma); anti-rabbit Alexa-555 (1:250; Invitrogen), anti-mouse Alexa-555 (1:250; Invitrogen). Cell boundaries were visualized by Alexa 488-conjugated phalloidin (Invitrogen), which stained the actin cytoskeleton. DAPI (Invitrogen) was used to visualize nuclei. Imaging was performed on a Zeiss LSM710. Maximum intensity projections of confocal z-scans were computed using ImageJ [14]. Scanning electron microscopy was as previously described [2].

Funding

Work in the Blum lab was supported by a grant from the Deutsche Forschungsgemeinschaft (BL-285/9) to MB. *Xenopus* work in the Harland lab was supported by the National Institutes of Health (NIH) (GM42341).

PW and TB were recipients of Ph.D. fellowships from the Landesgraduiertenförderung Baden-Württemberg. PW and open-access publication of this article were supported by a postdoctoral fellowship from the Deutsche Forschungsgemeinschaft (Wa 3365/1-1). CH and KF are indebted to the Baden-Württemberg Stiftung for the financial support of their research by the Eliteprogramme for Postdocs. KF was supported by a Margarete-von-Wrangell fellowship, funded by the European Social Fund and by the Ministry Of Science, Research and the Arts in Baden-Württemberg.

Authors contribution

PW initiated, conducted and analyzed experiments. TB performed electron microscopy. CH and KF helped significantly with ependymal flow. CM contributed to GI analysis. PW, AS and MB planned experiments and interpreted the results. RMH helped with interpretation of data and writing of the manuscript. PW and MB wrote the manuscript with input from all authors.

Acknowledgments

Helpful discussions and sharing of reagents are gratefully acknowledged: C. Kintner, B. Mitchell, D. Gradl, E. Pera, N. Papalopulu, K. Geehring, B. Ulmer, I. Schneider, T. Machen, H. Fischer, B. Illek and J. Wallingford. We thank S. Bogusch and A. Schäfer for expert technical support.

References

- [1] E. Amar, I.B. Dawid, Isolation and expression analysis of *foxj1* and *foxj1.2* in zebrafish embryos, *Int. J. Dev. Biol.* 52 (2008) 985–991, <http://dx.doi.org/10.1387/jidb.072477ea>.
- [2] T. Beyer, M. Danilchik, T. Thumberger, P. Vick, M. Tisler, I. Schneider, S. Bogusch, P. Andre, B. Ulmer, P. Walentek, B. Niesler, M. Blum, A. Schweickert, Serotonin signaling is required for Wnt-dependent GRP specification and leftward flow in *Xenopus*, *Curr. Biol.* 22 (2012) 1–7, <http://dx.doi.org/10.1016/j.cub.2011.11.027>.
- [3] A. Caron, X. Xu, X. Lin, Wnt/ β -catenin signaling directly regulates *Foxj1* expression and ciliogenesis in zebrafish Kupffer's vesicle, *Development* 139 (2012) 514–524, <http://dx.doi.org/10.1242/dev.071746>.
- [4] A.D. Chalmers, J.M. Slack, Development of the gut in *Xenopus laevis*, *Dev. Dyn.* 212 (1998) 509–521, [http://dx.doi.org/10.1002/\(SICI\)1097-0177\(199808\)212:4<509::AID-AJA4>3.0.CO;2-L](http://dx.doi.org/10.1002/(SICI)1097-0177(199808)212:4<509::AID-AJA4>3.0.CO;2-L).
- [5] C. Cruz, V. Ribes, E. Kutejova, J. Cayuso, V. Lawson, D. Norris, J. Stevens, M. Davey, K. Blight, F. Bangs, A. Mynett, E. Hirst, R. Chung, N. Balaskas, S.L. Brody, E. Marti, J. Briscoe, *Foxj1* regulates floor plate cilia architecture and modifies the response of cells to sonic hedgehog signalling, *Development* 137 (2010) 4271–4282, <http://dx.doi.org/10.1242/dev.051714>.
- [6] C. Hagenlocher, P. Walentek, C. Müller, T. Thumberger, K. Feistel, Ciliogenesis and cerebrospinal fluid flow in the developing *Xenopus* brain are regulated by *foxj1*, *Cilia* 2 (2013) 12, <http://dx.doi.org/10.1186/2046-2530-2-12>.
- [7] T. Hollemann, E. Tadjuidje, K. Koebernick, T. Pieler, Manipulation of hedgehog signaling in *Xenopus* by means of embryo microinjection and application of chemical inhibitors, *Methods Mol. Biol.* 397 (2007) 35–45.
- [8] M. Ikuzawa, S. Yasumasu, K.-I. Kobayashi, T. Inokuchi, I. Iuchi, Stomach remodeling-associated changes of H⁺/K⁺-ATPase beta subunit expression in *Xenopus laevis* and H⁺/K⁺-ATPase-dependent acid secretion in tadpole stomach, *J. Exp. Zool. A. Comp. Exp. Biol.* 301 (2004) 992–1002, <http://dx.doi.org/10.1002/jez.a.127>.
- [9] J. Jiang, C.C Hui, Hedgehog signaling in development and cancer, *Dev. Cell* 15 (6) (2008) 801–812, <http://dx.doi.org/10.1016/j.devcel.2008.11.010>.
- [10] J.R. Miller, R.T. Moon, Analysis of the signaling activities of localization mutants of beta-catenin during axis specification in *Xenopus*, *J. Cell Biol.* 139 (1997) 229–243.
- [11] T.H. Min, M. Kriebel, S. Hou, E.M. Pera, The dual regulator Sufu integrates Hedgehog and Wnt signals in the early *Xenopus* embryo, *Dev. Biol.* 358 (2011) 262–276, <http://dx.doi.org/10.1016/j.ydbio.2011.07.035>.
- [12] M. Placzek, J. Briscoe, The floor plate: multiple cells, multiple signals, *Nat. Rev. Neurosci.* 6 (2005) 230–240, <http://dx.doi.org/10.1038/nrn1628>.
- [13] B.S. Pohl, W. Knöchel, Isolation and developmental expression of *Xenopus Foxj1* and *FoxK1*, *Dev. Genes Evol.* 214 (2004) 200–205, <http://dx.doi.org/10.1007/s00427-004-0391-7>.
- [14] J. Schindelin, I. Arganda-Carreras, E. Frise, V. Kaynig, M. Longair, T. Pietzsch, S. Preibisch, C. Rueden, S. Saalfeld, B. Schmid, J.-Y. Tinevez, D.J. White, V. Hartenstein, K. Eliceiri, P. Tomancak, A. Cardona, Fiji: an open-source platform for biological-image analysis, *Nat. Methods* 9 (2012) 676–682, <http://dx.doi.org/10.1038/nmeth.2019>.
- [15] H.L. Sive, R.M. Grainger, R.M. Harland, Early Development of *Xenopus laevis*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, USA <http://dx.doi.org/10.1101/pdb.prot5536>.
- [16] J.L. Stubbs, I. Oishi, J.C. Izpisua Belmonte, C. Kintner, The forkhead protein *Foxj1* specifies node-like cilia in *Xenopus* and Zebrafish embryos, *Nat. Genet.* 40 (2008) 1454–1460, <http://dx.doi.org/10.1038/ng.267>.
- [17] P. Walentek, T. Beyer, T. Thumberger, A. Schweickert, M. Blum, ATP4a is required for Wnt-dependent *Foxj1* expression and leftward flow in *Xenopus* left-right development, *Cell Rep* 1 (2012) 516–527, <http://dx.doi.org/10.1016/j.celrep.2012.03.005>.
- [18] P. Walentek, S. Bogusch, T. Thumberger, P. Vick, E. Dubaissi, T. Beyer, M. Blum, A. Schweickert, A novel serotonin-secreting cell type regulates ciliary motility in the mucociliary epidermis of *Xenopus* tadpoles, *Development* 141 (2014) 1–8, <http://dx.doi.org/10.1242/dev.102343>.
- [19] P. Walentek, T. Beyer, C. Hagenlocher, C. Müller, K. Feistel, A. Schweickert, R.M. Harland, M. Blum, ATP4a is required for development and function of the *Xenopus* mucociliary epidermis – a potential model to study proton pump inhibitor-associated pneumonia, *Dev. Biol.* (2015). <http://dx.doi.org/10.1016/j.ydbio.2015.03.013>.
- [20] X. Yu, C.P. Ng, H. Habacher, S. Roy, *Foxj1* transcription factors are master regulators of the motile ciliogenic program, *Nat. Genet.* 40 (2008) 1445–1453, <http://dx.doi.org/10.1038/ng.263>.
- [21] Z. Zhang, S.A. Rankin, A.M. Zorn, Different thresholds of Wnt-Frizzled 7 signaling coordinate proliferation, morphogenesis and fate of endoderm progenitor cells, *Dev. Biol.* 378 (2013) 1–12, <http://dx.doi.org/10.1016/j.ydbio.2013.02.024>.