

UC San Diego

UC San Diego Previously Published Works

Title

Leveraging the placenta to advance neonatal care

Permalink

<https://escholarship.org/uc/item/6pz8t8p5>

Authors

Mestan, Karen K

Leibel, Sandra L

Sajti, Eniko

et al.

Publication Date

2023

DOI

10.3389/fped.2023.1174174

Peer reviewed



OPEN ACCESS

EDITED BY

Ju Lee Oei,
University of New South Wales, Australia

REVIEWED BY

Rachana Singh,
Tufts University, United States
Dimitrios Angelis,
University of Texas Southwestern Medical
Center, United States

*CORRESPONDENCE

Karen K. Mestan
✉ kmestan@health.ucsd.edu

RECEIVED 26 February 2023

ACCEPTED 24 April 2023

PUBLISHED 15 May 2023

CITATION

Mestan KK, Leibel SL, Sajti E, Pham B, Hietalati S,
Laurent L and Parast M (2023) Leveraging the
placenta to advance neonatal care.
Front. Pediatr. 11:1174174.
doi: 10.3389/fped.2023.1174174

COPYRIGHT

© 2023 Mestan, Leibel, Sajti, Pham, Hietalati,
Laurent and Parast. This is an open-access
article distributed under the terms of the
[Creative Commons Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/).
The use, distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in this
journal is cited, in accordance with accepted
academic practice. No use, distribution or
reproduction is permitted which does not
comply with these terms.

Leveraging the placenta to advance neonatal care

Karen K. Mestan^{1,2*}, Sandra L. Leibel^{1,2}, Eniko Sajti^{1,2}, Betty Pham^{1,2},
Samantha Hietalati^{1,2}, Louise Laurent^{1,3,4} and Mana Parast^{4,5}

¹Department of Pediatrics/Division of Neonatology, University of California, San Diego School of Medicine, La Jolla, CA, USA, ²Department of Pediatrics/Division of Neonatology, Rady Children's Hospital of San Diego, San Diego, CA, USA, ³Department of Obstetrics, Gynecology and Reproductive Sciences/Division of Maternal Fetal Medicine, University of California, San Diego School of Medicine, La Jolla, CA, USA, ⁴Sanford Consortium for Regenerative Medicine, La Jolla, CA, USA, ⁵Department of Pathology, University of California, San Diego School of Medicine, La Jolla, CA, USA

The impact of placental dysfunction and placental injury on the fetus and newborn infant has become a topic of growing interest in neonatal disease research. However, the use of placental pathology in directing or influencing neonatal clinical management continues to be limited for a wide range of reasons, some of which are historical and thus easily overcome today. In this review, we summarize the most recent literature linking placental function to neonatal outcomes, focusing on clinical placental pathology findings and the most common neonatal diagnoses that have been associated with placental dysfunction. We discuss how recent technological advances in neonatal and perinatal medicine may allow us to make a paradigm shift, in which valuable information provided by the placenta could be used to guide neonatal management more effectively, and to ultimately enhance neonatal care in order to improve our patient outcomes. We propose new avenues of clinical management in which the placenta could serve as a diagnostic tool toward more personalized neonatal intensive care unit management.

KEYWORDS

neonatal intensive care unit (NICU), placental pathology, prematurity and low birth weight, preeclampsia, bronchopulmonary dysplasia (BPD), chorioamnionitis, neonatal outcome

Introduction

The placenta is the transient organ of pregnancy that plays an intricate role in the growth, development, and protection of the fetus. Historically, research on the placenta has been predominantly focused on the role of maternal health on placental development and function, as well as the downstream effects on pregnancy health, fetal development, and the transition to neonatal life. However, there has been a relative paucity of guidance aimed at applying knowledge about placental pathology to improve the management of critically ill newborn infants, in particular, premature infants, who may spend the first few weeks to months in the neonatal intensive care unit (NICU). Barriers to using the placenta to guide neonatal management include the relative lack of understanding among neonatologists about what certain placental findings may indicate about their patients and the lack of robust workflows to facilitate education, communication, and information transfer among obstetricians, pathologists, and neonatologists. In addition, the placenta is a highly complex organ system, and our fundamental understanding of the patterns of placental injury, including the timing and combined presence of multiple lesions, is still emerging (1).

The impact of placental dysfunction and placental injury on the fetus and newborn infant has become a topic of growing interest in neonatal research. However, the use of

placental pathology in directing or influencing neonatal clinical management continues to be limited for a wide range of reasons, some of which are historical and thus easily overcome today. In this review, we will summarize the most recent literature linking placental function to neonatal outcomes, focusing on clinical placental pathology findings and the most common neonatal diagnoses that have been associated with placental dysfunction. We will discuss how recent technological advances in neonatal and perinatal medicine may allow us to make a paradigm shift, in which valuable information provided by the placenta could be used to guide neonatal management more effectively, and to ultimately enhance neonatal care in order to improve our patient outcomes. We propose new avenues of clinical management in which the placenta could serve as a diagnostic tool toward more personalized NICU management.

Overview of recent literature

A recent literature search conducted by our group querying “placenta and neonatal outcome” resulted in 5,741 results since 1961 accessible through PubMed.gov (<https://pubmed.ncbi.nlm.nih.gov>). Totally, 4,703 are in the past 20 years (2003–2023), and 50% of these publications are from 2016 to 2023 alone. Thus, there has been a rapidly increasing interest in studying the placenta as it pertains to the neonate. The body of literature is quite diverse. While the bulk of studies investigate novel pathways by which the placenta may lead to or predict adverse neonatal outcomes, very few evaluate placental pathology as a diagnostic and management tool for neonatologists (2), including our previous publication in which we discussed the clinical applications of the placenta for neonatologists (3). Still to date, many of the more recent studies of placental pathology that evaluate “neonatal outcome” continue to focus on preterm birth and fetal growth restriction as the outcomes. Very few studies extend these findings to propose how the neonatologist can use placental information in caring for NICU patients—specifically in preventing or minimizing morbidities for which newer therapies and treatment strategies have since been developed.

Recent reports have linked specific lesions of the placenta to infant complications of high-risk pregnancies, including neonatal outcomes such as brain injury, heart and lung diseases (both acquired and congenital), and a wide range of diagnoses with long-term neurodevelopmental, cardiopulmonary, and metabolic sequelae. With technological advances over the years, novel structural, cellular, functional, and genetic targets have been explored, but these “pregnancy outcomes” have been largely limited to the immediate birth outcomes such as stillbirth, preterm birth, and low birth weight. Epidemiological studies led by neonatal investigators are more likely to include postnatal outcomes during the NICU course, such as the major complications of prematurity [e.g., bronchopulmonary dysplasia (BPD), necrotizing enterocolitis, and intraventricular hemorrhage, among others]. Collaboration with obstetric and pathology colleagues is critical to our understanding of how the placenta may drive these neonatal outcomes. In addition, important

mediators between maternal complications of pregnancy, placental findings, and later childhood outcomes must be taken into consideration when unraveling these complex associations.

Gross and histologic features of the placenta: relevance to neonatal management

The standard placental pathological examination conducted by experienced placental pathologists worldwide is a valuable, robust tool that has contributed significantly to our understanding of maternal and fetal health during pregnancy. The most widely used guidelines originated from the Amsterdam Placental Workshop Group, published in 2016 by Khong et al. (4), which established consensus with respect to both sampling of the placenta and criteria for major placental lesions. These definitions were recently updated in a report by Redline et al. (1) as criteria to outline four distinct domains of placental pathology, thereby facilitating classification of the so-called “patterns of placental injury” and providing a framework for understanding the developmental and pathologic processes linked to specific clinical outcomes. This reference serves as a practical resource that can be incorporated into routine neonatal management. Discussion with placental pathologists and obstetricians is essential for interpreting how the timing and presence of multiple lesions and their potential synergistic effects could have contributed to certain neonatal clinical presentations. These criteria and patterns have been leveraged over the years by various groups (5, 6) and provide significant data that the neonatologist can use to understand the intrauterine environment and exposures during fetal development, with implications for management of critically ill neonates.

Figure 1 describes the four domains of histopathology with distinct lesions reported on standard pathology reports reviewed by clinicians. In terms of relevance to newborn infants and NICU patients, the presence of acute inflammatory (AI) lesions strongly suggests recent or active intrauterine infection by bacterial, viral, or fungal pathogens and should warrant sepsis evaluation and even empiric treatment for culture-negative sepsis in the newborn period (7). Chronic inflammatory (CI) lesions are less prominently reported but may suggest more long-standing placental injury and dysfunction early in pregnancy driven by immune-mediated processes between mother and fetus (8). An intriguing CI lesion is villitis of unknown etiology (VUE), which has been seen with viral pathologies and various degrees of fetal growth restriction thought to be mediated by immune rejection (9). The presence of maternal vascular malperfusion (MVM) is understood to occur very early in pregnancy—it is linked to abnormalities in trophoblast implantation and development, and failed maternal spiral artery remodeling that leads to abnormal villous development, compromised maternal-fetal gas and nutrient exchange, and chronic fetal hypoxia. MVM is commonly seen in cases of maternal preeclampsia and severe fetal growth restriction, and is of growing interest in the management of premature infants born

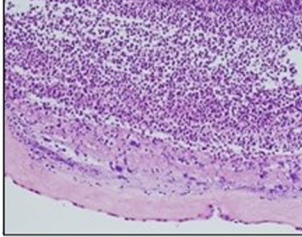
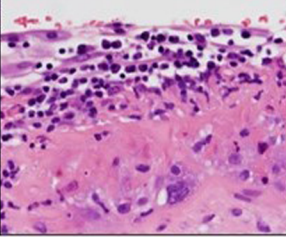
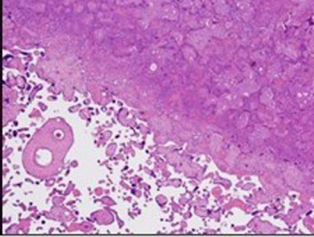
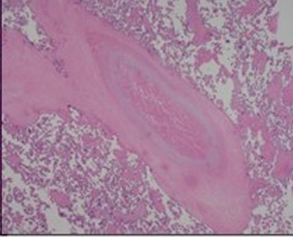
Acute Inflammation	Chronic Inflammation	Maternal Vascular Malperfusion	Fetal Vascular Pathology
<p>Maternal: Subchorionitis, Chorionitis, Chorioamnionitis, Necrotizing chorioamnionitis</p> <p>Fetal: Umbilical vasculitis, Funisitis, Chorionic plate vasculitis</p>	<p>Chronic villitis of unknown etiology (VUE); Plasma cell deciduitis; Chronic chorioamnionitis; Chronic intervillitis</p>	<p>Maternal Vessels: Fibrinoid necrosis/ Acute atherosclerosis; Muscularized basal plate arteries; Mural hypertrophy of membrane arterioles</p> <p>Chorionic Villi: Accelerated villous maturation/increased syncytial knots; Villous agglutination; Distal villous hypoplasia/small terminal villi; infarcts</p>	<p>Thrombi (mural or occlusive) in umbilical, chorionic plate, or stem villous vessels; Fetal thrombotic vasculopathy (avascular villi or villi with stromal karyorrhexis)</p>
			

FIGURE 1
Four major histologic domains of the standard placental pathology exam. AI, acute inflammation; CI, chronic inflammation; MVM, maternal vascular malperfusion; FVP, fetal vascular pathology (also referred to as fetal vascular malperfusion or FVM). Histology images provided by Northwestern University Feinberg School of Medicine, Department of Pathology.

through such exposures (10, 11). Finally, fetal vascular pathology (FVP) or malperfusion (FVM) indicates reduced or absent perfusion of the villous parenchyma by the fetus. Most common etiologies include umbilical cord obstruction leading to stasis, ischemia, and thrombosis. Other proposed causes include fetal cardiac insufficiency, hyperviscosity, and thrombophilias (12).

In addition to the above histologic domains, common gross pathologic lesions include placental weight, infarction, hemorrhage, and cord abnormalities, which can be readily identified at delivery. Alone or in conjunction with histologic lesions, these gross findings may be important in the immediate care of the critically ill neonate. For example, massive retroplacental/intraplacental hemorrhage and cord abnormalities, including hypercoiling and true knots, may prompt additional tests and monitoring for bleeding, thrombosis, or end-organ involvement not readily apparent on the newborn examination.

Neonatal outcomes of perinatal origins: opportunities to leverage the placenta

While the patient population in the NICU is highly complex, there is a growing recognition that certain neonatal diagnoses

arise from pathogenic processes that occur in the prenatal and perinatal periods of fetal development. For these diagnoses, the placenta may provide key insight into management approaches for which there are multiple pathways. In other words, more focused management plans could be tailored based on placental findings. In **Table 1**, we summarize the most recently published associations between placental gross and histologic lesions and common neonatal problems that have growing evidence of originating in the fetal stages and are considered a result of placental dysfunction (in traditional NICU presentation approach—by systems). A couple of the most prominent themes in our search of placental pathology and common NICU diagnoses are described below.

Neonatal hypoglycemia

There have been very few studies linking neonatal hypoglycemia and placental findings that could potentially enhance management. In a small case-control study, Natarajan et al. reported an association between placental syncytial knots [adjusted odds ratio (aOR) (95% CI), 2.9 (1.1–9.1); $P=0.04$] and placental calcifications [aOR (95% CI), 3 (1.1–8.7); $P=0.03$] as

TABLE 1 Common conditions treated in the NICU (by systems) with described placental associations.

System/disease	Placental association(s)	Clinical management implications
Fluids, electrolytes, and nutrition		
Hypoglycemia	Placental syncytial knots, calcifications (13) Placental weight, placental malperfusion lesions (14)	May narrow differential diagnosis: placental insufficiency/injury vs. infectious etiologies, which would shift management toward or away from prolonged antibiotic treatments. May identify cause and predict course of management for hypoglycemia in the absence of small-for-gestational age birth weight.
Postnatal growth failure	Maternal and fetal inflammatory response (15, 16)	May distinguish constitutional growth and other causes of growth delays from inflammatory processes, leading to different approaches in feeding/nutrition support and long-term follow-up.
Cardiorespiratory problems		
Apnea/bradycardia	Methylxanthines readily pass placental barrier and may effect control of breathing mechanisms in infancy (17)	Normal placental pathology may rule out pathologic causes of apnea (i.e., intrauterine infection, neurologic), thus, facilitating/confirming diagnosis of apnea of prematurity.
Respiratory distress syndrome	Histologic chorioamnionitis with pathologic causes of surfactant deficiency (18, 19)	In preterm infants, normal placenta may facilitate diagnosis of noninfectious causes of hyaline membrane disease (i.e., delayed surfactant production) vs. sepsis/pneumonia.
Meconium aspiration syndrome	Columnar epithelial changes vs. meconium-laden macrophages differentially associated with adverse neonatal outcome; funisitis (20, 21)	Placental changes could inform the extent and timing of causes, and thus guide work-up and management approaches (e.g., treat for infection, work-up for perinatal depression, supportive respiratory care).
BPD	HCA and FIR (inconclusive in meta-analyses) (22) and decreased risk of BPD and death (23) MVM increased with BPD-associated pulmonary hypertension (10, 24) Chorionic plate vascularization (25) Chronic inflammatory lesions (26)	Could help differentiate certain endotypes of BPD, such as inflammatory versus vascular, which could inform response to specific treatments of which there are currently multiple approaches with untoward side effects (e.g., steroids, pulmonary vasodilators, diuretics, chronic mechanical ventilation).
Congenital heart diseases (coarctation, septal defects, tetralogy of Fallot, transposition of great arteries, hypoplastic left heart, double outlet RV)	Abnormal villous maturation (27), increased MVM lesions, lower placental weight, abnormal umbilical cord insertion, distal villous FVM, villous edema, amnion nodosum (28, 29), DNA methylation changes (30), MVM or FVM, abnormal umbilical cord coil, placental perfusion defects (31, 32)	Could help inform the etiologies of certain heart defects, as well as intrauterine conditions and events that may further compound complex care: timing of surgical interventions, associated morbidities, anticipated clinical course, counseling for prognosis and recurrence risk in future pregnancies.
CDH	Shallow placental implantation, diffuse chronic hypoxic patterns of placental injury, lesions of fetal vascular malperfusion, likely stasis-induced (33)	As above. Severity and persistence of pulmonary hypertension may be worse with evidence of placental vascular disease, thus informing management decisions regarding need for ECMO, timing of surgical repair, prognosis.
Gastrointestinal problems		
Gastroschisis/abdominal wall defects	Placental ischemia, MVM, suggesting that vascular events in second trimester cause disruption of fetal abdominal wall formation (34)	May provide additional information on extent and chronicity of intestinal injury involvement, which could guide decisions on surgical management, feeding, and risks for intestinal dysfunction.
NEC	Multiple placental lesions, including placental inflammatory and vascular lesions, is associated with higher risk for medical or surgical NEC (35); Massive perivillous fibrin deposition with FGR (36); Histologic chorio with preeclampsia (37)	May guide NICU management decisions regarding risk for NEC, and thus timing of initiation and advancement of enteral feedings.
Neurologic problems		
HIE	Review of 26 articles: FVM, umbilical cord pathology, inflammatory changes, MVM, placental weight (38); Hypercoiled cord, abnormal insertion (39); Chronic placental pathologies associated with more severe brain injury (40); Combined acute and chronic placental changes are common, underscoring the complex causal pathways of HIE (41)	Placental findings could inform the causes of HIE, of which there are multiple. For example, acute inflammatory changes of histologic chorio should prompt infectious work-up, including for meningitis. Fetal thrombotic vasculopathy may suggest vaso-occlusive causes and prompt more neurologic work-up and supportive management. MVM and VUE may suggest chronic placental insufficiency and thus more complex involvement of multiple organ systems and slower recovery.
Neonatal seizures/stroke	Both acute and chronic placental inflammation are common in perinatal stroke (42); FIR with vascular thrombotic problem (43)	May provide key information on timing and etiology of seizures, thus prompting specific work-up leading to targeted treatments, including antibiotics, thrombolytic therapies, genetic and hematologic work-up for recurrence risks.
IVH	Massive perivillous fibrin deposition with FGR (36); MVM in premature and SGA infants (44)	May identify infants at relatively higher risk for IVH, and influence early management decisions regarding surveillance and neuroprotection strategies in “small baby” protocols.

(continued)

TABLE 1 Continued

System/disease	Placental association(s)	Clinical management implications
PVL	FIR (43); Chronic diffuse capsular deciduitis, decidual plasma cells, evidence of chronic but not acute chorioamnionitis (45)	Findings in premature infants may prompt more comprehensive early neuroimaging (MRI), early intervention services, and long-term neurodevelopment follow-up.
Infectious diseases		
Neonatal sepsis/pneumonia/meningitis	FIR (46); histologic chorioamnionitis (47); variable among many small studies, associations may be modified by antepartum prophylaxis and other clinical considerations (48)	Presence of FIR/HCA without positive blood culture may point toward partially treated infection, chronic infection, etc. prompting antibiotic treatment. Negative placental histology with positive neonatal sepsis work-up may suggest acute infection or false positive blood test in a newborn infant.
TORCH and other congenital infections	Toxoplasmosis: villous trophoblast cell apoptosis (49); Syphilis: Hydrops placentalis, funisitis, fetal vasculopathy, chronic villitis (50); Rubella: Obliteration of stem vessels, villous inflammation and sclerosis, trophoblastic necrosis (51); CMV: chronic villitis, CMV inclusions (52); HSV: Nonspecific findings (53)	See Yallapragada et al. (3)
COVID-19	Multiple lesions reported (54–56). No evidence of direct viral involvement or vertical transmission. Most recent studies report that there are no significant placental histopathologic changes that occur after dx of SARS-CoV2 infection in women during third trimester (57)	Current literature provides reassurance supporting minimal need for NICU surveillance or intervention, but long-term studies are still needed for guidance in pediatric follow-up.

CMV, cytomegalovirus; ECMO, extracorporeal membrane oxygenation; FIR, fetal inflammatory response; FVM, fetal vascular malperfusion; FGR, fetal growth restriction; HCA, histologic chorioamnionitis; HSV, herpes simplex virus; MVM, maternal vascular malperfusion; TORCH, toxoplasmosis, other, rubella, CMV, HSV; NICU, neonatal intensive care unit; BPD, bronchopulmonary dysplasia; CDH, congenital diaphragmatic hernia; NEC, necrotizing enterocolitis; HIE, hypoxic-ischemic encephalopathy; VUE, villitis of unknown etiology; IVH, intraventricular hemorrhage; PVL, periventricular leukomalacia; SGA, small for gestational age; RV, right ventricle.

predictors of small for gestational age (SGA) birth weight, in which higher rates of in-hospital morbidities included perinatal asphyxia, respiratory distress, hypoglycemia, and feeding intolerance (13). Although these associations are relatively weak, they remind us that clinical observations of hypoglycemia in SGA infants could be due to chronic placental insufficiency in some cases, allowing neonatologists to tailor the management accordingly. This might mean using fewer tests to look for other causes of hypoglycemia and feeding intolerance, such as sepsis and hyperinsulinism. Recent findings from a study by Hutcheon et al. using latent profile analysis in a much larger cohort (26,007 births from Montreal, Canada, 2001–2009) suggest that placental weight and placental malperfusion lesions may serve as novel markers to identify infants at risk for hypoglycemia due to placental insufficiency, independent of the SGA proxy (14).

Respiratory distress syndrome

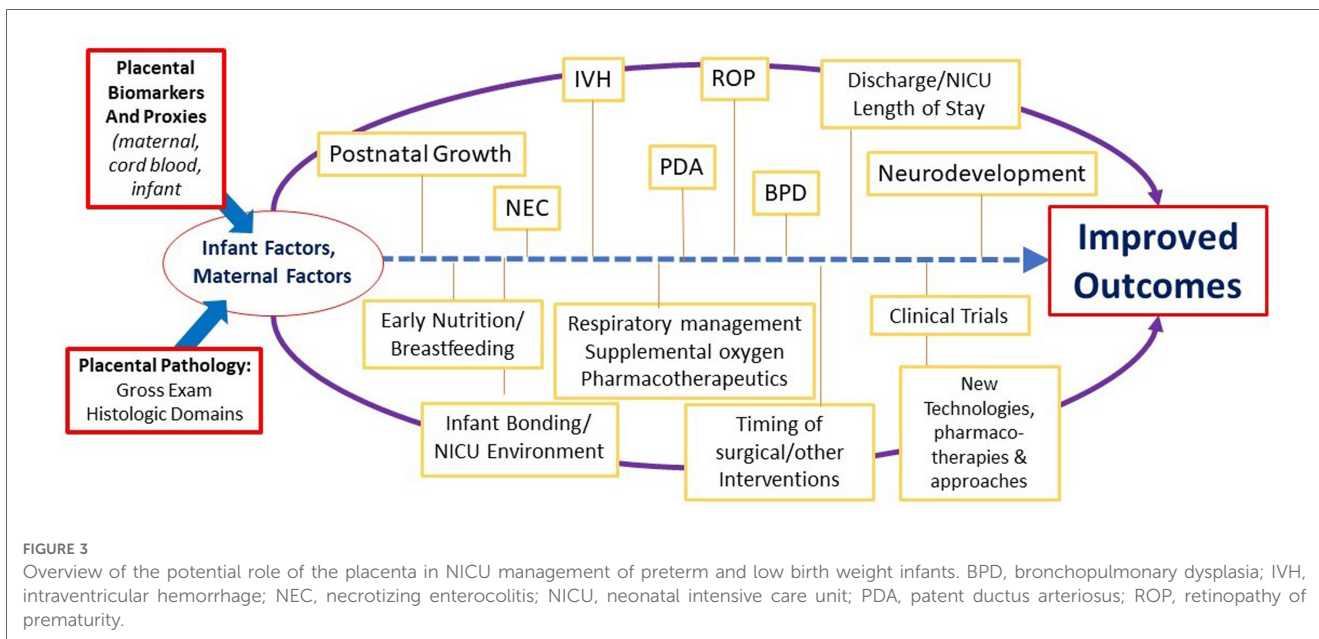
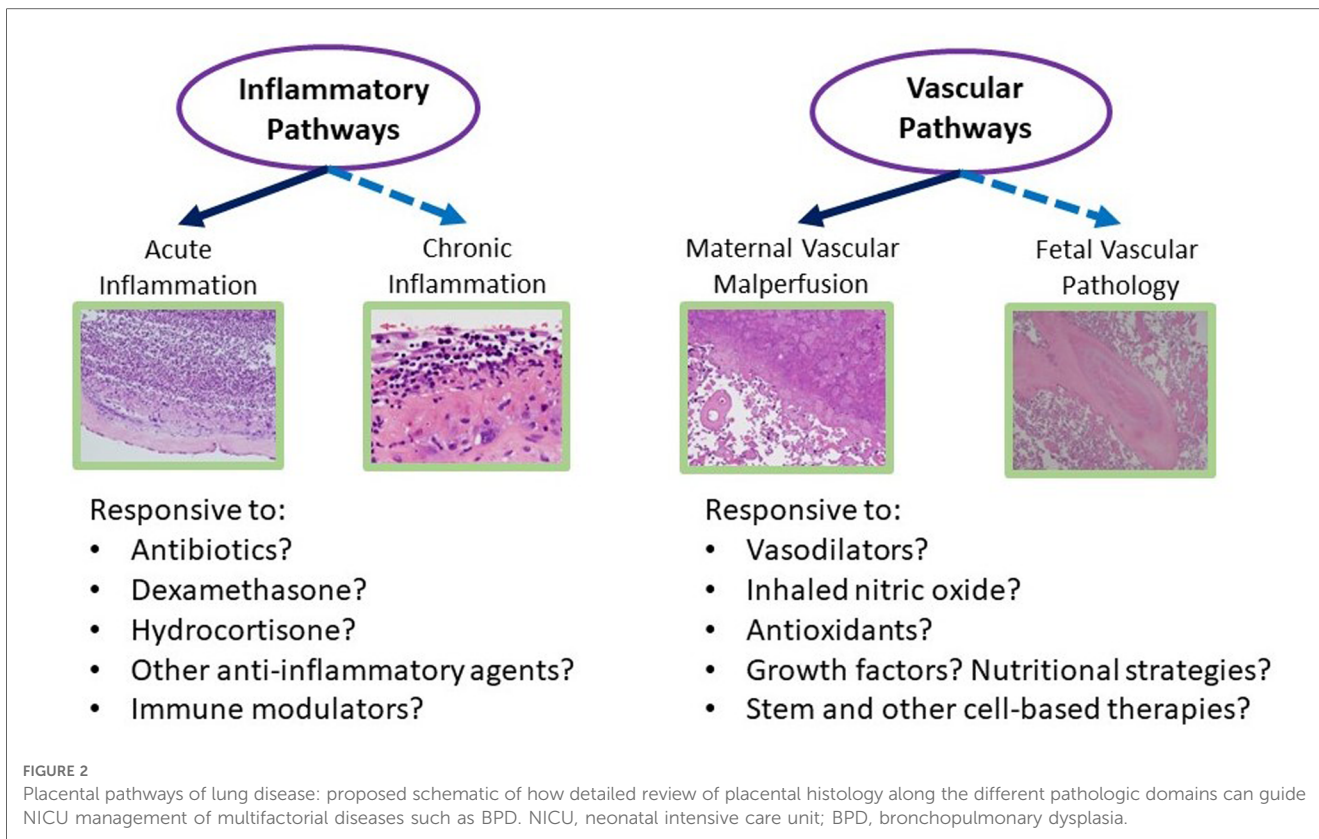
The majority of early literature on placental histopathology and neonatal outcomes was in reference to histologic chorioamnionitis (HCA), which is a relatively common finding in spontaneous preterm birth. In recent studies and large meta-analyses, the link between HCA and neonatal outcome has been reported at most as modest (58), with the exception of respiratory distress syndrome (RDS) (18)—a primary problem of surfactant deficiency in the newborn period. In a large cohort study conducted in Australia, Lahra et al. found a protective effect of maternal and fetal inflammation on RDS (59). This may suggest multiple modalities of RDS beyond surfactant deficiency that need to be investigated in ongoing studies.

Bronchopulmonary dysplasia

In our search, we found that long-standing reports linking placental inflammation to BPD are variable and appear to be refuted by several recent multicenter studies and meta-analyses. In contrast, more recent studies link placental vascular disease, specifically MVM, to sub-phenotypes of BPD-associated pulmonary hypertension (60). Inferring these “endotypes” and linking them to placental disease may have implications for neonatal management (see Figure 2) (61). Recent reports on the interaction between HCA and exposure to antenatal steroids to accelerate lung maturation and attenuate BPD suggest that infants born through intrauterine inflammation have been protected by antenatal steroids (62). Alternatively, in a recent analysis of postnatal corticosteroid response in neonates <32 weeks and their associated placental pathologies, a trend of better steroid response was observed in infants exposed to MVM (63). These and other studies provide evidence that response to certain postnatal therapies can be predicted by placental information (Figure 2).

Congenital diaphragmatic hernia

Genetic variants account for 30% of congenital diaphragmatic hernia (CDH) cases that likely occur via a complex inheritance pattern influenced by *in utero* stressors through placental function and metabolism. Genetic variants likely confer susceptibility with epigenetic modifiers (e.g., nutritional factors, toxins, hypoxia) contributing to the subsequent phenotype (64). The role of vitamin A metabolites has been well-described in normal placental maintenance and embryologic lung



development via the retinoic acid signaling pathway (65). Retinoic acid receptor responder proteins regulate antioxidant responses, angiogenesis, and anti-inflammatory processes. This signaling pathway and/or other functionally similar ones may contribute to the placental pathology that has been described in these patients. Placental phenotypes are suggestive of shallow placental implantation, lesions of stasis-associated FVM, and chronic patterns of hypoxia (33). Other work analyzing placental

pathology of CDH patients also demonstrated increased placental fetal thrombotic vasculopathy (66). While a degree of pulmonary hypertension may be explained by primary defects in lung and pulmonary vascular development, there may be a yet undefined contribution from MVM (10). Further understanding of these placental factors may be helpful to prognosticate and potentially change the continued high mortality and morbidity in these patients.

Congenital heart disease

A rapidly evolving area of neonatal research has focused on mechanisms of congenital heart disease (CHD) and placental dysfunction. The fetal heart placenta axis describes processes that regulate the development of the placenta, to both heart development and function (67, 68). Abnormal placental development may impact cardiac development, and the circulatory changes from congenital heart disease may result in placental insufficiency (69). Newborns with CHD have smaller placentas, which put them at risk for SGA. Lesions of FVM in the placenta have been associated with structural cardiac defects in the fetus (70). These studies provide compelling evidence that the placenta may play a causative role in the embryonic development of the heart, independent of, or interactively with, genetic predispositions and environmental exposures during pregnancy. However, there is still much to be investigated regarding the role of the placenta in the management of congenital heart defects. The most severe forms of CHD are rare, and the growing recognition of CHD subtypes makes our understanding of placental relationships even more challenging (71). Thus, studies that continue to use animal and *in vitro* models of placental-cardiac development are critical to our understanding of how and to what extent placental development influences fetal heart function (72, 73). Moreover, ongoing clinical research efforts to link placental findings to expanding longitudinal cohorts of CHD infants are critically needed to advance the field of neonatal-cardiac intensive care (74). As CHD is not a commonly recognized indication for placental autopsy and many subtypes/cases are not diagnosed prenatally or in the immediate postnatal period, efforts that reliably link placental features to CHD management will require well-designed, prospective studies across institutions and over many years.

Hypoxic-ischemic encephalopathy

Studies of placental pathologic findings and hypoxic-ischemic encephalopathy (HIE) continue to be published. The studies on this topic appear to have increased in the past decade due to the implementation of therapeutic hypothermia in level 3 and 4 NICUs nationwide. The question of whether the placental pathology examination may be incorporated into the decision-making process to initiate therapeutic hypothermia in a newborn infant with HIE is important, as there is ongoing controversy over the level of HIE and the effectiveness of current clinical information to predict response to therapy (e.g., cord blood gases, Apgar scores, physical examination findings). Since infants require initiation of cooling within 6 h of life, the utility of the placental examination would either need to be expedited or proxies of placental histology developed (e.g., cord blood, neonatal blood, maternal blood). Given the proximity of these biospecimens to the placenta at birth, development of these proxies seems feasible and promising. The association between placental findings and neurodevelopmental outcome in neonates with HIE is less apparent in the era of therapeutic hypothermia,

suggesting that neonatal intervention is effective in uncoupling the placental-perinatal events leading up to HIE, by attenuating the acute process of brain injury and improving long-term outcomes (75).

Neonatal sepsis

Finally, the research and clinical attempts to use placental histologic features to guide management of neonatal sepsis (and rule out sepsis) continues to be highly controversial and not universally accepted. To our knowledge, there are no recent meta-analyses evaluating the literature on the use of the placenta in neonatal sepsis management. Rather, considerable research has focused on the appropriate timing, indications, and dosage of intrapartum antibiotics given to mothers in order to prevent neonatal sepsis and its comorbidities (76). Thus, intrapartum antibiotic exposure is likely an important mediator that confounds our observations between placental findings and neonatal sepsis. As such, there are no routine uses or published protocols for incorporation of the placental examination in ruling in or ruling out neonatal infections. A recent study by Wong and Khong found umbilical cord section diagnosis of funisitis to be reasonably accurate in cases with clinical chorioamnionitis, fever, and/or premature rupture of membranes. They propose changing current laboratory practice to rapid processing of umbilical cords ahead of the rest of the other placental sections in busy pathology departments (46). These changes seem feasible and warranted, given the evidence that the presence of funisitis or vasculitis can rule in sepsis and other diagnoses requiring immediate intervention.

Current clinical limitations and research gaps

Despite the wealth of evidence accumulated over the years on associations between placental pathology and neonatal outcomes, why is the placenta not used more routinely in postnatal management of the neonate? At most large academic birth centers, standardized placental pathologic examination is routinely performed on a subset of all deliveries, based on published recommended guidelines. The criteria prompting examination, which have recently been updated (77), are historically and predominantly driven by maternal, fetal, and intrapartum indications (e.g., preeclampsia, clinical chorioamnionitis, intrauterine growth restriction, fetal distress, abruption) with neonatal indications limited to diagnoses requiring immediate care (i.e., hypoxic-ischemic encephalopathy, sepsis, meconium aspiration). Even among these acute perinatal problems, there is a lack of recommendations for how the placenta can be used to individualize care. Much needed also is consideration for how placental findings could be used to guide ongoing NICU management for infants with multifactorial conditions such as extreme prematurity (e.g., BPD, NEC, IVH), congenital heart, lung, and intestinal anomalies—for which

factors such as intrauterine infection and placental vascular disease may contribute to or complicate one's clinical course (Table 1). For some institutions, driven by state laws or hospital policies, preterm birth of a specified gestational age cut-off and/or delivery resulting in NICU admission are indications for placental pathology examination. This approach may capture the majority of diagnoses listed above. However, there is a need for more education and research to streamline, prioritize, and focus the pathologist's examination as to specific findings that may provide a more precision medicine approach for NICU patients (78).

There are several perceived and actual limitations in current clinical practice that may account for this disconnect. The turnaround time for placental reports to reach the neonatologist is relatively longer than blood and other biospecimen tests performed in clinical laboratories. In addition, the necessary expertise in placental/perinatal pathology is often limited and only readily available in tertiary care centers. Thus, requesting placental pathology results in level 2 and non-academic community NICUs is often perceived as not practical or feasible. However, well-coordinated and carefully designed workflows and infrastructures, many of which have been adopted in recent years to accommodate and facilitate patient biospecimens for research, can be implemented in a wide variety of hospital settings to preserve placental tissues for clinical care as well. Detailed protocols have been published that allow for a wide range of studies to be performed on placental tissues (79–81). Even the most common and routine methods for biopsy and archive of placental tissues, such as formalin-fixed paraffin-embedded (FFPE) tissues, can now be used to study spatial transcriptomics of the placenta (82). If properly processed, these tissues once thought useful only for routine histology could eventually be used to map the human placenta and enhance our understanding of a wide range of neonatal diseases. The above research strategies require the availability and use of specialized newer reagents, optimal workflows, and laboratory conditions, which are feasible through dedicated integration of clinical and research teams. Institutional support and funding that sustain these collaborative efforts are critical factors for success.

Leveraging the placenta at the bedside: conceptual models

Based on our ongoing research and work of others linking placental findings to neonatal outcomes, we have designed several conceptual models of how placental findings coupled with maternal data and the early perinatal course may help guide management of the critically ill infant in the later postnatal period for complex conditions such as those complicating preterm birth and low birth weight. Figure 2 illustrates such a conceptual model for BPD management, in which placental findings along pathways of acute and chronic inflammation may prompt decisions toward the use of anti-inflammatory strategies such as postnatal steroids, including type, timing, duration, and route of administration. Conversely, preterm infants with associated placental vascular pathology may be more responsive

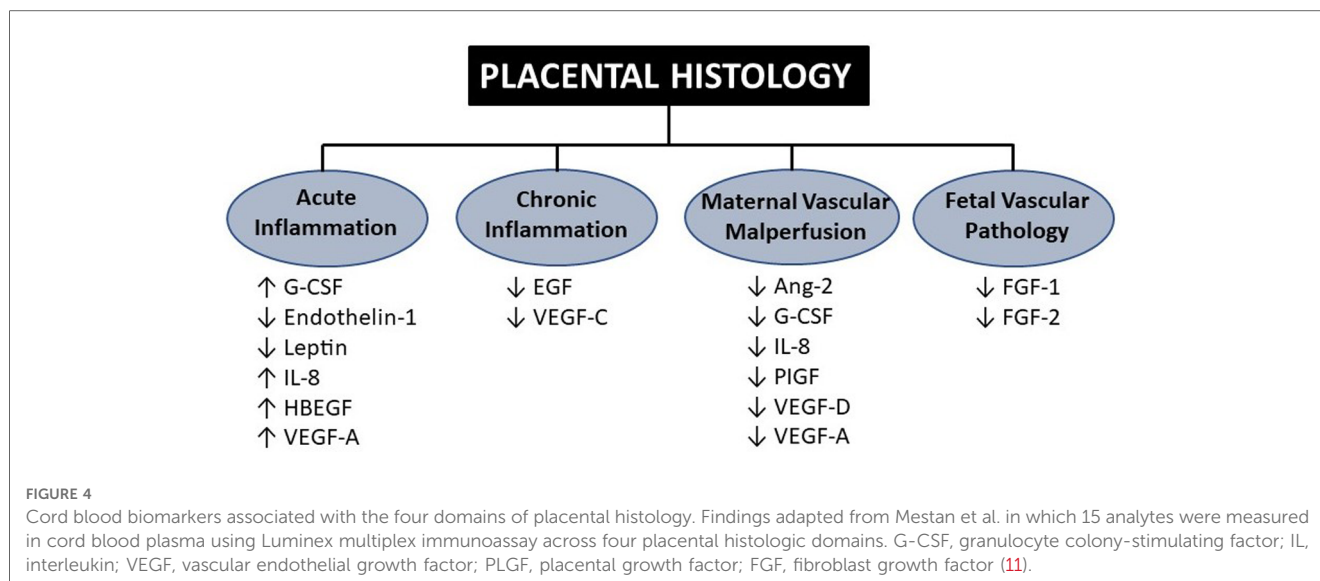
to pulmonary vasodilator therapies, and newer agents that promote pulmonary vascular growth. While these and other therapies require further investigation in larger and multicenter studies, the incorporation of placental findings into neonatal care throughout one's NICU course and in the development of newer therapies and clinical trials will help us further develop novel treatment strategies for a wide range of neonatal diseases (Figure 3).

Need for placental biomarkers and proxies to inform NICU management

Placental examination findings and histologic lesions may serve as important biomarkers of *in utero* pathology affecting the critically ill neonate. The turnaround time for placental pathology reports should not obviate routine and comprehensive assessments, as many complications of prematurity and term infant diseases often do not require immediate readings, but within the first days/weeks of life. Certain decisions, such as the initiation and durations of antibiotics and therapeutic hypothermia, may require more immediate information, which could be prioritized in the clinical setting and coordinated with effective workflows and communication between services.

Gross placental pathology in many hospitals may take up to a week and histopathology reporting may take up to 3–6 weeks depending upon the patient volumes and other priorities of a given institution. Unfortunately, there are no well-established early rapid placental diagnostic tests that can be used to evaluate or identify specific lesions of placental injury before birth or point-of-care tests at delivery. Development of such technologies is an ongoing priority of our research efforts and the goal of many investigators committed to placental research (83, 84). Point-of-care tests during pregnancy or surrounding delivery that could distinguish between maternal and placental origins of disease, such as maternal circulating extracellular vesicle (EV) microRNAs, are promising but still in development (85, 86). Rapid, more automated diagnostics using placental imaging coupled with machine learning approaches are also emerging in the new era of artificial intelligence (87). These advances could enhance neonatal management by providing earlier risk assessment, anticipation of clinical course, and earlier initiation of certain NICU interventions upon delivery.

Other strategies that could enhance the use of the placenta for neonatologists include the development of proxies and novel biomarkers that are tightly correlated with placental histology and detect placental dysfunction prior to or at birth. Maternal serum biomarkers of placental dysfunction are gradually being used in some centers to detect placental dysfunction due to maternal preeclampsia (88). In addition to the emerging use of maternal circulating sFlt-1/PlGF ratio to predict placental dysfunction-mediated preeclampsia, other potential proxies of placental function include levels of cytokines, chemokines, and growth factors circulating in cord blood at birth (11, 89–91). Studies measuring the biochemical milieu of the placenta have emerged over the years in attempts to link predictive markers of



inflammation, oxidant stress, and other pathways to neonatal outcomes. Of particular interest are outcomes related to prematurity, such as brain injury (IVH, PVL), lung injury (BPD), and neurodevelopment (cerebral palsy). As detailed in **Figure 4**, biomarkers that delineate certain placental pathologic domains could be used to specifically address problems associated with systemic inflammation (IL-6 and IL-8) and dysregulated vascular endothelial growth (VEGF-A, placental growth factor).

Future directions

The above review provides compelling evidence that the placenta is an important and valuable resource for NICU management. Recent breakthroughs and advances in technology provide even more potential approaches to leverage the placenta to enhance and improve neonatal care. Development of real-time early/rapid placental diagnostics such as placental imaging, including placental MRI during pregnancy and the use of digital image technologies linked with artificial intelligence and machine learning as mentioned above, may overcome limitations of regional availability of expertise and turnaround time for placental evaluation (87, 92). Development of more automated data entry may facilitate more rapid delivery of information to clinicians in NICU settings. The development of point-of-care testing is a recent and rapidly evolving approach to bedside diagnosis and more timely management of multiple problems in the NICU. These approaches could be applied to the evaluation of the placenta as well through closely related biomarkers in maternal blood/urine, cord blood, and postnatal biospecimens. Placental biopsies containing information on epigenomic, transcriptomic, and other multi-omics analyses with biological significance are rapidly being discovered (93, 94). Coupled with our current knowledge of placental histology and infant outcomes, the opportunities to advance neonatal care through the placenta are promising.

Author contributions

KKM contributed to the original conceptual design of the review, prepared the tables and figures, and drafted the original and final versions of the manuscript. SLL, ES, BP, SH, and LL provided their expertise in NICU and Obstetrics management, outcomes and current research, and conducted the literature reviews. MP provided her expertise in both clinical and research aspects of placental pathology and assisted with overall interpretation of findings. All co-authors contributed to drafting and revising the work for its intellectual content. All authors contributed to the article and approved the submitted version.

Funding

This work was supported by the NIH [R01 HL139798 (PI: KKM)].

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Redline RW, Ravishankar S, Bagby CM, Saab ST, Zarei S. Four major patterns of placental injury: a stepwise guide for understanding and implementing the 2016 Amsterdam consensus. *Mod Pathol*. (2021) 34(6):1074–92. doi: 10.1038/s41379-021-00747-4
- Ykema JMA, D'Haens EJ, Havenith M, van Eyck J, van Lingen RA, Hemels MAC. Pilot study demonstrates that placental histology can provide an additional tool for diagnosing early-onset neonatal sepsis. *Acta Paediatr*. (2018) 107(12):2086–91. doi: 10.1111/apa.14410
- Yallapragada S, Mestan KK, Ernst LM. The placenta as a diagnostic tool for the neonatologist. *NeoReviews*. (2016) 17(3):e131–e143. doi: 10.1542/neo.17-3-e131
- Khong TY, Mooney EE, Ariel I, Balmus NC, Boyd TK, Brundler MA, et al. Sampling and definitions of placental lesions: Amsterdam Placental Workshop Group Consensus Statement. *Arch Pathol Lab Med*. (2016) 140(7):698–713. doi: 10.5858/arpa.2015-0225-CC
- Giacometti C, Cassaro M. Clinicopathologic correlations in histopathologic report of placenta: can we do something more? *Arch Pathol Lab Med*. (2017) 141(2):184–5. doi: 10.5858/arpa.2016-0414-LE
- Freedman AA, Keenan-Devlin LS, Borders A, Miller GE, Ernst LM. Formulating a meaningful and comprehensive placental phenotypic classification. *Pediatr Dev Pathol*. (2021) 24(4):337–50. doi: 10.1177/10935266211008444
- Romero R, Pacora P, Kusanovic JP, Jung E, Panaitescu B, Maymon E, et al. Clinical chorioamnionitis at term X: microbiology, clinical signs, placental pathology, and neonatal bacteremia—implications for clinical care. *J Perinat Med*. (2021) 49(3):275–98. doi: 10.1515/jpm-2020-0297
- Enninga EAL, Leontovich AA, Fedyshyn B, Wakefield L, Gandhi M, Markovic SN, et al. Upregulation of HLA-class I and II in placentas diagnosed with villitis of unknown etiology. *Reprod Sci*. (2020) 27(5):1129–38. doi: 10.1007/s43032-019-00101-9
- Osborne B, Oltean I, Sucha E, Mitsakakis N, Barrowman N, Bainbridge S, et al. Association of distinct features of villitis of unknown etiology histopathology and fetal growth restriction diagnosis in a retrospective cohort from eastern Ontario. *Placenta*. (2022) 128:83–90. doi: 10.1016/j.placenta.2022.08.012
- Mestan KK, Check J, Minturn L, Yallapragada S, Farrow KN, Liu X, et al. Placental pathologic changes of maternal vascular underperfusion in bronchopulmonary dysplasia and pulmonary hypertension. *Placenta*. (2014) 35(8):570–4. doi: 10.1016/j.placenta.2014.05.003
- Mestan KK, Gotteiner N, Porta N, Grobman W, Su EJ, Ernst LM. Cord blood biomarkers of placental maternal vascular underperfusion predict bronchopulmonary dysplasia-associated pulmonary hypertension. *J Pediatr*. (2017) 185:33–41. doi: 10.1016/j.jpeds.2017.01.015
- Redline RW, Ravishankar S. Fetal vascular malperfusion, an update. *APMIS*. (2018) 126(7):561–9. doi: 10.1111/apm.12849
- Natarajan V, Singh P, Vigneshwar NKV, Anthony ML, Rao S, Chacham S, et al. Maternal and placental risk factors for small gestational age and fetal malnutrition. *Curr Pediatr Rev*. (2023) 19(2):187–96. doi: 10.2174/1573396318666220705154424
- Hutcheon JA, Riddell CA, Himes KP. A new approach for classifying fetal growth restriction. *Epidemiology*. (2021) 32(6):860–7. doi: 10.1097/EDE.0000000000001399
- Mestan K, Yu Y, Matoba N, Cerda S, Demmin B, Pearson C, et al. Placental inflammatory response is associated with poor neonatal growth: preterm birth cohort study. *Pediatrics*. (2010) 125(4):e891–8. doi: 10.1542/peds.2009-0313
- Trevisanuto D, Peruzzetto C, Cavallin F, Vedovato S, Cosmi E, Visentin S, et al. Fetal placental inflammation is associated with poor neonatal growth of preterm infants: a case-control study. *J Matern Fetal Neonatal Med*. (2013) 26(15):1484–90. doi: 10.3109/14767058.2013.789849
- Aden U. Methylxanthines during pregnancy and early postnatal life. *Handb Exp Pharmacol*. (2011) 200(3):73–89. doi: 10.1007/978-3-642-13443-2_14
- Sato M, Nishimaki S, Yokota S, Seki K, Horiguchi H, An H, et al. Severity of chorioamnionitis and neonatal outcome. *J Obstet Gynaecol Res*. (2011) 37(10):1313–9. doi: 10.1111/j.1447-0756.2010.01519.x
- Elimian A, Verma U, Beneck D, Cipriano R, Visintainer P, Tejani N. Histologic chorioamnionitis, antenatal steroids, and perinatal outcomes. *Obstet Gynecol*. (2000) 96(3):333–6. doi: 10.1016/s0029-7844(00)00928-5
- Tamayev L, Mor L, Herman HG, Schreiber L, Kovo M, Bar J, et al. Placental histology of acute versus continuous meconium exposure—association with obstetric and neonatal outcomes. *Placenta*. (2021) 103:214–9. doi: 10.1016/j.placenta.2020.10.002
- Yokoi K, Iwata O, Kobayashi S, Muramatsu K, Goto H. Influence of foetal inflammation on the development of meconium aspiration syndrome in term neonates with meconium-stained amniotic fluid. *PeerJ*. (2019) 7:e7049. doi: 10.7717/peerj.7049
- Lacaze-Masmonteil T. That chorioamnionitis is a risk factor for bronchopulmonary dysplasia—the case against. *Paediatr Respir Rev*. (2014) 15(1):53–5. doi: 10.1016/j.prrv.2013.09.005
- Plakkal N, Soraisham AS, Trevenen C, Freiheit EA, Sauve R. Histological chorioamnionitis and bronchopulmonary dysplasia: a retrospective cohort study. *J Perinatol*. (2013) 33(6):441–5. doi: 10.1038/jp.2012.154
- Kunjunju AM, Gopogondanahalli KR, Chan Y, Sehgal A. Bronchopulmonary dysplasia-associated pulmonary hypertension: clues from placental pathology. *J Perinatol*. (2017) 37(12):1310–4. doi: 10.1038/jp.2017.130
- Shapiro S, Trail-Burns E, Slader MG, Laptook A, De Paepe ME. Correlation between chorionic plate vascularization and risk of bronchopulmonary dysplasia in extremely preterm infants. *Placenta*. (2020) 101:154–8. doi: 10.1016/j.placenta.2020.09.016
- Sharma A, Sood BG, Qureshi F, Xin Y, Jacques SM. Chronic inflammatory placental lesions correlate with bronchopulmonary dysplasia severity in extremely preterm infants. *Pediatr Dev Pathol*. (2021) 24(5):430–7. doi: 10.1177/10935266211013625
- Snoep MC, Aliasi M, van der Meeren LE, Jongbloed MRM, DeRuiter MC, Haak MC. Placenta morphology and biomarkers in pregnancies with congenital heart disease—a systematic review. *Placenta*. (2021) 112:189–96. doi: 10.1016/j.placenta.2021.07.297
- Stanek J. Patterns of placental injury in various types of fetal congenital heart disease. *J Perinat Med*. (2022). doi: 10.1515/jpm-2022-0478. [Online ahead of print].
- Matthiesen NB, Henriksen TB, Agergaard P, Gaynor JW, Bach CC, Hjortdal VE, et al. Congenital heart defects and indices of placental and fetal growth in a nationwide study of 924 422 liveborn infants. *Circulation*. (2016) 134(20):1546–56. doi: 10.1161/CIRCULATIONAHA.116.021793
- Liu J, Wu Y, Sun H, Liu X, Gu X, Zhang Y, et al. Association between placental DNA methylation and fetal congenital heart disease. *Mol Genet Genomics*. (2023) 298(1):243–51. doi: 10.1007/s00438-022-01944-9
- Miremberg H, Gindes L, Schreiber L, Raucher Sternfeld A, Bar J, Kovo M. The association between severe fetal congenital heart defects and placental vascular malperfusion lesions. *Prenat Diagn*. (2019) 39(11):962–7. doi: 10.1002/pd.5515
- Ozcan T, Kikano S, Plummer S, Strainic J, Ravishankar S. The association of fetal congenital cardiac defects and placental vascular malperfusion. *Pediatr Dev Pathol*. (2021) 24(3):187–92. doi: 10.1177/1093526620986497
- Stanek J. Patterns of placental injury in congenital anomalies in second half of pregnancy. *Pediatr Dev Pathol*. (2019) 22(6):513–22. doi: 10.1177/1093526619852869
- Folkerth RD, Habbe DM, Boyd TK, McMillan K, Gromer J, Sens MA, et al. Gastroschisis, destructive brain lesions, and placental infarction in the second trimester suggest a vascular pathogenesis. *Pediatr Dev Pathol*. (2013) 16(5):391–6. doi: 10.2350/13-03-1316-CR.1
- Garg PM, Paschal JL, Ansari MAY, Billington L, Ware J, Adams K, et al. Association of placental pathologic findings with the severity of necrotizing enterocolitis in preterm infants—a matched case-control study. *Fetal Pediatr Pathol*. (2023) 42(2):187–97. doi: 10.1080/15513815.2022.2110340
- Spinillo A, Gardella B, Muscettola G, Cesari S, Fiandrino G, Tziialla C. The impact of placental massive perivillous fibrin deposition on neonatal outcome in pregnancies complicated by fetal growth restriction. *Placenta*. (2019) 87:46–52. doi: 10.1016/j.placenta.2019.09.007
- Duci M, Frigo AC, Visentin S, Verlato G, Gamba P, Fascetti-Leon F. Maternal and placental risk factors associated with the development of necrotizing enterocolitis (NEC) and its severity. *J Pediatr Surg*. (2019) 54(10):2099–102. doi: 10.1016/j.jpedsurg.2019.04.018
- Fox A, Doyle E, Geary M, Hayes B. Placental pathology and neonatal encephalopathy. *Int J Gynaecol Obstet*. (2023) 160(1):22–7. doi: 10.1002/ijgo.14301
- Hasegawa J, Nakao M, Ikeda T, Toyokawa S, Jojima E, Satoh S, et al. Fetal heart rate evolution patterns in cerebral palsy associated with umbilical cord complications: a nationwide study. *BMC Pregnancy Childbirth*. (2022) 22(1):177. doi: 10.1186/s12884-022-04508-2
- Hellwig L, Brada M, Held U, Hagmann C, Bode P, Frontzek K, et al. Association of perinatal sentinel events, placental pathology and cerebral MRI in neonates with hypoxic-ischemic encephalopathy receiving therapeutic hypothermia. *J Perinatol*. (2022) 42(7):885–91. doi: 10.1038/s41372-022-01356-y
- Chalak L, Redline RW, Goodman AM, Juul SE, Chang T, Yanowitz TD, et al. Acute and chronic placental abnormalities in a multicenter cohort of newborn infants with hypoxic-ischemic encephalopathy. *J Pediatr*. (2021) 237:190–6. doi: 10.1016/j.jpeds.2021.06.023
- Leon RL, Kalvacherla V, Andrews MM, Thomas JM, Mir IN, Chalak LF. Placental pathologic lesions associated with stroke in term neonates. *Front Endocrinol (Lausanne)*. (2022) 13:920680. doi: 10.3389/fendo.2022.920680

43. Ko HS, Cheon JY, Choi SK, Lee HW, Lee A, Park IY, et al. Placental histologic patterns and neonatal seizure, in preterm premature rupture of membrane. *J Matern Fetal Neonatal Med.* (2017) 30(7):793–800. doi: 10.1080/14767058.2016.1186634
44. Oh MA, Barak S, Mohamed M, Penn AA. Placental pathology and intraventricular hemorrhage in preterm and small for gestational age infants. *J Perinatol.* (2021) 41(4):843–9. doi: 10.1038/s41372-021-00954-6
45. Maleki Z, Bailis AJ, Argani CH, Askin FB, Graham EM. Periventricular leukomalacia and placental histopathologic abnormalities. *Obstet Gynecol.* (2009) 114(5):1115–20. doi: 10.1097/AOG.0b013e3181bdcfc4
46. Wong YP, Khong TY. Changing laboratory practice for early detection of a fetal inflammatory response: a contemporary approach. *Diagnostics (Basel).* (2023) 13(3):487. doi: 10.3390/diagnostics13030487
47. Rallis D, Lithoxopoulou M, Pervana S, Karagianni P, Hatzioannidis I, Soubasi V, et al. Clinical chorioamnionitis and histologic placental inflammation: association with early-neonatal sepsis. *J Matern Fetal Neonatal Med.* (2022) 35(25):8090–6. doi: 10.1080/14767058.2021.1961727
48. Berezowsky A, Romano A, Hochberg A, Krispin E, Danieli HZ, Krencel A, et al. The correlation between placental histology and microbiologic infection in the diagnosis of chorioamnionitis in preterm delivery. *Placenta.* (2022) 128:18–22. doi: 10.1016/j.placenta.2022.08.005
49. Abbasi M, Kowalewska-Grochowska K, Bahar MA, Kilani RT, Winkler-Lowen B, Guilbert LJ. Infection of placental trophoblasts by *Toxoplasma gondii*. *J Infect Dis.* (2003) 188(4):608–16. doi: 10.1086/377132
50. Sheffield JS, Sanchez PJ, Wendel GD Jr, Fong DW, Margraf LR, Zeray F, et al. Placental histopathology of congenital syphilis. *Obstet Gynecol.* (2002) 100(1):126–33. doi: 10.1016/s0029-7844(02)02010-0
51. Garcia AG, Marques RL, Lobato YY, Fonseca ME, Wigg MD. Placental pathology in congenital rubella. *Placenta.* (1985) 6(4):281–95. doi: 10.1016/s0143-4004(85)80038-2
52. Fisher S, Genbacev O, Maidji E, Pereira L. Human cytomegalovirus infection of placental cytotrophoblasts in vitro and in utero: implications for transmission and pathogenesis. *J Virol.* (2000) 74(15):6808–20. doi: 10.1128/jvi.74.15.6808-6820.2000
53. Syridou G, Spanakis N, Konstantinidou A, Piperaki ET, Kafetzis D, Patsouris E, et al. Detection of cytomegalovirus, parvovirus B19 and herpes simplex viruses in cases of intrauterine fetal death: association with pathological findings. *J Med Virol.* (2008) 80(10):1776–82. doi: 10.1002/jmv.21293
54. Baergen RN, Heller DS. Placental pathology in COVID-19 positive mothers: preliminary findings. *Pediatr Dev Pathol.* (2020) 23(3):177–80. doi: 10.1177/1093526620925569
55. Shanes ED, Mithal LB, Otero S, Azad HA, Miller ES, Goldstein JA. Placental pathology in COVID-19. *Am J Clin Pathol.* (2020) 154(1):23–32. doi: 10.1093/ajcp/aqaa089
56. Smithgall MC, Liu-Jarin X, Hamele-Bena D, Cimic A, Mourad M, Debelenko L, et al. Third-trimester placentas of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)-positive women: histomorphology, including viral immunohistochemistry and in-situ hybridization. *Histopathology.* (2020) 77(6):994–9. doi: 10.1111/his.14215
57. Gulersen M, Prasannan L, Tam Tam H, Metz CN, Rochelson B, Meirowitz N, et al. Histopathologic evaluation of placentas after diagnosis of maternal severe acute respiratory syndrome coronavirus 2 infection. *Am J Obstet Gynecol MFM.* (2020) 2(4):100211. doi: 10.1016/j.ajogmf.2020.100211
58. Been JV, Rours IG, Kornelisse RF, Lima Passos V, Kramer BW, Schneider TA, et al. Histologic chorioamnionitis, fetal involvement, and antenatal steroids: effects on neonatal outcome in preterm infants. *Am J Obstet Gynecol.* (2009) 201(6):587 e1–8. doi: 10.1016/j.ajog.2009.06.025
59. Lahra MM, Beeby PJ, Jeffery HE. Maternal versus fetal inflammation and respiratory distress syndrome: a 10-year hospital cohort study. *Arch Dis Child Fetal Neonatal Ed.* (2009) 94(1):F13–6. doi: 10.1136/adc.2007.135889
60. Abman SH. Bronchopulmonary dysplasia: “a vascular hypothesis”. *Am J Respir Crit Care Med.* (2001) 164(10 Pt 1):1755–6. doi: 10.1164/ajrccm.164.10.2109111c
61. Pierro M, Villamor-Martinez E, van Westering-Kroon E, Alvarez-Fuente M, Abman SH, Villamor E. Association of the dysfunctional placentation endotype of prematurity with bronchopulmonary dysplasia: a systematic review, meta-analysis and meta-regression. *Thorax.* (2022) 77(3):268–75. doi: 10.1136/thoraxjnl-2020-216485
62. Ahn HM, Park EA, Cho SJ, Kim YJ, Park HS. The association of histological chorioamnionitis and antenatal steroids on neonatal outcome in preterm infants born at less than thirty-four weeks' gestation. *Neonatology.* (2012) 102(4):259–64. doi: 10.1159/000339577
63. Koenders VM, Appels A, van Straaten HLM, Dutman AC, Hemels MAC. Postnatal corticosteroid response in neonates <32 weeks and relation with placental pathology. *Eur J Pediatr.* (2023) 182(1):265–74. doi: 10.1007/s00431-022-04672-9
64. Holder AM, Klaassens M, Tibboel D, de Klein A, Lee B, Scott DA. Genetic factors in congenital diaphragmatic hernia. *Am J Hum Genet.* (2007) 80(5):825–45. doi: 10.1086/513442
65. Fernandes-Silva H, Araujo-Silva H, Correia-Pinto J, Moura RS. Retinoic acid: a key regulator of lung development. *Biomolecules.* (2020) 10(1):152. doi: 10.3390/biom10010152
66. Stanek J, Sheridan RM, Le LD, Crombleholme TM. Placental fetal thrombotic vasculopathy in severe congenital anomalies prompting EXIT procedure. *Placenta.* (2011) 32(5):373–9. doi: 10.1016/j.placenta.2011.02.002
67. Mahadevan A, Tipler A, Jones H. Shared developmental pathways of the placenta and fetal heart. *Placenta.* (2022) S1043-4004(22) 00481-7. doi: 10.1016/j.placenta.2022.12.006
68. Wilson RL, Yuan V, Courtney JA, Tipler A, Cnota JF, Jones HN. Analysis of commonly expressed genes between first trimester fetal heart and placenta cell types in the context of congenital heart disease. *Sci Rep.* (2022) 12(1):10756. doi: 10.1038/s41598-022-14955-8
69. Andescavage NN, Limperopoulos C. Placental abnormalities in congenital heart disease. *Transl Pediatr.* (2021) 10(8):2148–56. doi: 10.21037/tp-20-347
70. Saleemuddin A, Tantbirojn P, Sirois K, Crum CP, Boyd TK, Tworoger S, et al. Obstetric and perinatal complications in placentas with fetal thrombotic vasculopathy. *Pediatr Dev Pathol.* (2010) 13(6):459–64. doi: 10.2350/10-01-0774-OA.1
71. Courtney J, Troja W, Owens KJ, Brockway HM, Hinton AC, Hinton RB, et al. Abnormalities of placental development and function are associated with the different fetal growth patterns of hypoplastic left heart syndrome and transposition of the great arteries. *Placenta.* (2020) 101:57–65. doi: 10.1016/j.placenta.2020.09.007
72. Ward EJ, Bert S, Fanti S, Malone KM, Maughan RT, Gkantsinikoudi C, et al. Placental inflammation leads to abnormal embryonic heart development. *Circulation.* (2023) 147(12):956–72. doi: 10.1161/CIRCULATIONAHA.122.061934
73. Radford BN, Zhao X, Glazer T, Eaton M, Blackwell D, Mohammad S, et al. Defects in placental syncytiotrophoblast cells are a common cause of developmental heart disease. *Nat Commun.* (2023) 14(1):1174. doi: 10.1038/s41467-023-36740-5
74. Danford DA, Yetman AT, Haynatzki G. Derivation and validation of a general predictive model for long term risks for mortality and invasive interventions in congenital and acquired cardiac conditions encountered in the young. *Pediatr Cardiol.* (2023). doi: 10.1007/s00246-023-03154-5. [Online ahead of print].
75. Benz LD, Bode PK, Brandt S, Grass B, Hagmann C, Liamlahi R, et al. Placental findings are not associated with neurodevelopmental outcome in neonates with hypoxic-ischemic encephalopathy—an 11-year single-center experience. *J Perinat Med.* (2022) 50(3):343–50. doi: 10.1515/jpm-2020-0583
76. Hopkins L, Smail F. Antibiotic regimens for management of intraamniotic infection. *Cochrane Database Syst Rev.* (2002) 2002(3):CD003254. doi: 10.1002/14651858.CD003254
77. Roberts DJ, Baergen RN, Boyd TK, Carreon CK, Duncan VE, Ernst LM, et al. Criteria for placental examination for obstetrical and neonatal providers. *Am J Obstet Gynecol.* (2022) 228(5):497–508.e4. doi: 10.1016/j.ajog.2022.12.017
78. Mir IN, Leon R, Chalak LF. Placental origins of neonatal diseases: toward a precision medicine approach. *Pediatr Res.* (2021) 89(2):377–83. doi: 10.1038/s41390-020-01293-6
79. Wolfe LM, Thiagarajan RD, Boscolo F, Tache V, Coleman RL, Kim J, et al. Banking placental tissue: an optimized collection procedure for genome-wide analysis of nucleic acids. *Placenta.* (2014) 35(8):645–54. doi: 10.1016/j.placenta.2014.05.005
80. Linder JE, Batey K, Johnston R, Cohen EM, Wang Y, Wang X, et al. The PathLink acquired gestational tissue bank: feasibility of project PLACENTA. *J Reprod Biotechnol Fertil.* (2018) 7:14–27.
81. Roberts VH, Gaffney JE, Lewandowski KS, Schabel MC, Morgan TK, Frias AE. A standardized method for collection of human placenta samples in the age of functional magnetic resonance imaging. *Biotechniques.* (2019) 67(2):45–9. doi: 10.2144/btn-2019-0029
82. Terry J, Bedaiwy MA. Placental interferon signaling is involved in chronic intervillitis of unknown etiology. *Placenta.* (2022) 124:5–11. doi: 10.1016/j.placenta.2022.05.006
83. Bianchi DW, Maron JL. Conversations with the editors: the past, present, and future of placental research at the Eunice Kennedy Shriver National Institute of Child Health and Human Development. *Clin Ther.* (2021) 43(2):211–7. doi: 10.1016/j.clinthera.2020.12.020
84. Weinberg DH. Real-time assessment of the development and function of the placenta across gestation to support therapeutics in pregnancy. *Clin Ther.* (2021) 43(2):279–86. doi: 10.1016/j.clinthera.2020.11.002
85. Aharon A, Rebibo-Sabbah A, Ahmad RS, Dangot A, Bar-Lev TH, Brenner B, et al. Associations of maternal and placental extracellular vesicle miRNA with preeclampsia. *Front Cell Dev Biol.* (2023) 11:1080419. doi: 10.3389/fcell.2023.1080419
86. Srinivasan S, Treacy R, Herrero T, Olsen R, Leonardo TR, Zhang X, et al. Discovery and verification of extracellular miRNA biomarkers for non-invasive prediction of Pre-eclampsia in asymptomatic women. *Cell Rep Med.* (2020) 1(2):100013. doi: 10.1016/j.xcrm.2020.100013

87. Goldstein JA, Nateghi R, Irmakci I, Cooper LAD. Machine learning classification of placental villous infarction, perivillous fibrin deposition, and intervillous thrombus. *Placenta*. (2023) 135:43–50. doi: 10.1016/j.placenta.2023.03.003
88. Kifle MM, Dahal P, Vatish M, Cerdeira AS, Ohuma EO. The prognostic utility of soluble fms-like tyrosine kinase-1 (sFlt-1) and placental growth factor (PlGF) biomarkers for predicting preeclampsia: a secondary analysis of data from the INSPIRE trial. *BMC Pregnancy Childbirth*. (2022) 22(1):520. doi: 10.1186/s12884-022-04817-6
89. Voller SB, Chock S, Ernst LM, Su E, Liu X, Farrow KN, et al. Cord blood biomarkers of vascular endothelial growth (VEGF and sFlt-1) and postnatal growth: a preterm birth cohort study. *Early Hum Dev*. (2014) 90(4):195–200. doi: 10.1016/j.earlhumdev.2014.01.003
90. Mestan K, Yu Y, Thorsen P, Skogstrand K, Matoba N, Liu X, et al. Cord blood biomarkers of the fetal inflammatory response. *J Matern Fetal Neonatal Med*. (2009) 22(5):379–87. doi: 10.1080/14767050802609759
91. Ernst LM, Mithal LB, Mestan K, Wang V, Mangold KA, Freedman A, et al. Umbilical cord miRNAs to predict neonatal early onset sepsis. *PLoS One*. (2021) 16(5):e0249548. doi: 10.1371/journal.pone.0249548
92. Pietsch M, Ho A, Bardanzellu A, Zeidan AMA, Chappell LC, Hajnal JV, et al. APPLAUSE: automatic prediction of PLacental health via U-net segmentation and statistical evaluation. *Med Image Anal*. (2021) 72:102145. doi: 10.1016/j.media.2021.102145
93. Clark J, Avula V, Ring C, Eaves LA, Howard T, Santos HP, et al. Comparing the predictivity of human placental gene, microRNA, and CpG methylation signatures in relation to perinatal outcomes. *Toxicol Sci*. (2021) 183(2):269–84. doi: 10.1093/toxsci/kfab089
94. Lapehn S, Paquette AG. The placental epigenome as a molecular link between prenatal exposures and fetal health outcomes through the DOHaD hypothesis. *Curr Environ Health Rep*. (2022) 9(3):490–501. doi: 10.1007/s40572-022-00354-8