

UCSF

UC San Francisco Previously Published Works

Title

Neuroprotective Properties of Xenon

Permalink

<https://escholarship.org/uc/item/6h44s12d>

Journal

Molecular Neurobiology, 57(1)

ISSN

0893-7648

Authors

Maze, Mervyn
Laitio, Timo

Publication Date

2020

DOI

10.1007/s12035-019-01761-z

Peer reviewed

Dear Author

Here are the proofs of your article.

- You can submit your corrections **online** or by **fax**.
- For **online** submission please insert your corrections in the online correction form. Always indicate the line number to which the correction refers.
- Please return your proof together with the permission to publish confirmation.
- For **fax** submission, please ensure that your corrections are clearly legible. Use a fine black pen and write the correction in the margin, not too close to the edge of the page.
- Remember to note the journal title, article number, and your name when sending your response via e-mail, fax or regular mail.
- **Check** the metadata sheet to make sure that the header information, especially author names and the corresponding affiliations are correctly shown.
- **Check** the questions that may have arisen during copy editing and insert your answers/corrections.
- **Check** that the text is complete and that all figures, tables and their legends are included. Also check the accuracy of special characters, equations, and electronic supplementary material if applicable. If necessary refer to the *Edited manuscript*.
- The publication of inaccurate data such as dosages and units can have serious consequences. Please take particular care that all such details are correct.
- Please **do not** make changes that involve only matters of style. We have generally introduced forms that follow the journal's style. Substantial changes in content, e.g., new results, corrected values, title and authorship are not allowed without the approval of the responsible editor. In such a case, please contact the Editorial Office and return his/her consent together with the proof.
- If we do not receive your corrections **within 48 hours**, we will send you a reminder.

Please note

Your article will be published **Online First** approximately one week after receipt of your corrected proofs. This is the **official first publication** citable with the DOI.

Further changes are, therefore, not possible.

After online publication, subscribers (personal/institutional) to this journal will have access to the complete article via the DOI using the URL:

<http://dx.doi.org/10.1007/s12035-019-01761-z>

If you would like to know when your article has been published online, take advantage of our free alert service. For registration and further information, go to:

<http://www.springerlink.com>.

Due to the electronic nature of the procedure, the manuscript and the original figures will only be returned to you on special request. When you return your corrections, please inform us, if you would like to have these documents returned.

The **printed version** will follow in a forthcoming issue.

Metadata of the article that will be visualized in OnlineFirst

1	Article Title	Neuroprotective Properties of Xenon	
2	Article Sub- Title		
3	Article Copyright - Year	Springer Science+Business Media, LLC, part of Springer Nature 2019 (This will be the copyright line in the final PDF)	
4	Journal Name	Molecular Neurobiology	
5	Corresponding Author	Family Name	Maze
6		Particle	
7		Given Name	Mervyn
8		Suffix	
9		Organization	UCSF
10		Division	Center for Cerebrovascular Research, Department of Anesthesia and Perioperative Care
11		Address	San Francisco, CA, USA
12		e-mail	Mervyn.Maze@ucsf.edu
13	Author	Family Name	Laitio
14		Particle	
15		Given Name	Timo
16		Suffix	
17		Organization	University ofTurku
18		Division	Division of Perioperative Services, Intensive Care Medicine and Pain Management, Turku University Hospital
19		Address	Turku, Finland
20		e-mail	
21	Schedule	Received	29 August 2019
22		Revised	
23		Accepted	29 August 2019
24	Abstract	Note: This data is mandatory. Please provide.	
25	Keywords separated by ' - '	Xenon - Neuroprotection - Medical application	
26	Foot note information	Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.	

1
3
2
4
5
6
7
8
9

Neuroprotective Properties of Xenon

Mervyn Maze¹ · Timo Laitio²

Received: 29 August 2019 / Accepted: 29 August 2019
 © Springer Science+Business Media, LLC, part of Springer Nature 2019

Q3 10
11
Q4 12

Abstract
 Note: This data is mandatory. Please provide.
Keywords Xenon · Neuroprotection · Medical application

13
14

Introduction

Q5 15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32

Xenon is a colorless, odorless, tasteless, mono-atomic, and inert gas with a relative molecular weight of 131.3, and the empirical formula is Xe. Xenon is an extremely rare gas that represents no more than 0.0875 ppm in the atmosphere; this feature led the discoverers William Ramsay and Morris Travers to name it xenon from the Greek word “ξένος” (xenos) for stranger or foreign. Because of xenon’s rarity, it is extremely expensive to produce from the residue left from air separation units that are used to produce oxygen; therefore, its commercial applications have been limited to high-priced applications such as the ultimate “clean gas” in the electronics/semi-conductor industry, an ion propellant for space travel, and a bright lighting source, and for medical applications, notably anesthesia, imaging, and neuroprotection following acute ongoing injury. In this review, the authors trace the development of xenon for medical applications from the physico-chemical properties to the initial preclinical studies, and conclude in randomized clinical trials (RCTs).

33
34
35

Inert but Biologically Active

Because xenon is enshrouded by five filled electron shells, it is incapable of covalent bonding and forming adducts under

Q1
Q2

✉ Mervyn Maze
 Mervyn.Maze@ucsf.edu

¹ Center for Cerebrovascular Research, Department of Anesthesia and Perioperative Care, UCSF, San Francisco, CA, USA
² Division of Perioperative Services, Intensive Care Medicine and Pain Management, Turku University Hospital, University of Turku, Turku, Finland

biological conditions as electrons cannot be donated or accepted. However, because of xenon’s relatively high polarizability [1], with a value of 4 compared with 0.2 for helium, it can form dipoles and has an affinity for amino acid residues surrounding preformed hydrophobic cavities thereby changing the functional properties of neighboring proteins by London dispersion forces. In this manner, xenon has been shown to have activity on many proteins, governed mostly by the size and shape of xenon atoms. Because of its chemical non-reactivity, xenon is not biotransformed, which results in two important features governing its future clinical use. Most xenobiotic drugs are converted into metabolites that may be toxic; as xenon is not metabolized, the dangers posed by toxic metabolites are obviated. Furthermore, xenon in the inspired and expired gases are identical, permitting recirculation of exhaled xenon and thereby limiting the need for a fresh supply of this scarce resource.

Xenon for Anesthesia 53

Xenon was reported to have anesthetic properties in 1951 [2] and comes closest to exhibiting all of the ideal properties of an inhaled general anesthetic (Table 1). Compared with nitrous oxide, the other non-potent gaseous anesthetic, xenon is 1.5 times more potent; xenon is more suitable than nitrous oxide for anesthesia because of its lower blood/gas solubility and the consequent extremely rapid inflow and washout from the body. Despite this, xenon is infrequently used as an anesthetic even though European market authorization has been in effect for more than a decade; low utilization is attributed to the high cost involved in manufacturing this rare gas from the atmosphere. Therefore, the expense of using xenon as an anesthetic for routine adult surgery appears not to be justified given the available alternatives [3].

t1.1	Table 1 Properties of an	
t1.2	“ideal” inhalation agent	Obtainable in pure form at a reasonable cost
t1.3		Inherently stable
t1.4		Not biotransformable
t1.5		Lacks organ-specific toxicity
t1.6		Minimal cardiorespiratory effects
t1.7		Non-flammable
t1.8		Low blood-gas partition coefficient for rapid uptake, elimination, and titratability
t1.9		Sufficiently potent allowing an enriched inspired oxygen concentration
t1.10		Lacks long-term adverse effects with chronic exposure
t1.11		Lacks unpleasant smell and irritation to airway
t1.12		Possesses analgesic and hypnotic properties
t1.13		Readily reversible central nervous system effects with no stimulation

68 Drugs produce the anesthetic state via interaction with re-
 69 ceptor targets which potentiate inhibitory neurotransmission
 70 and/or inhibit excitatory neurotransmission [4]. Xenon is
 71 thought to exert anesthetic action by potent non-competitive
 72 inhibition of the excitatory NMDA receptors [5] through an
 73 action at the binding site of the co-agonist, glycine [6]. Xenon
 74 also exerts potent effects on the neuronal background potassi-
 75 um channels including two-pore domain potassium channels
 76 such as TREK and TASK, which modulate neuronal excitabil-
 77 ity [7], and on ATP-sensitive potassium channels [8].

78 The first reported clinical experience with xenon for anes-
 79 thesia was published in 1951 by Cullen and Gross, who re-
 80 ported on two patients who underwent surgical procedures
 81 (orchietomy in an 81-year-old man and fallopian tube liga-
 82 tion in a 38-year-old woman who was 24-h postpartum) while
 83 receiving a xenon–oxygen (80:20) mixture to achieve the first
 84 plane of the third stage of anesthesia [2]. Induction was com-
 85 pleted within 5 min and both patients maintained normal
 86 blood pressure, pulse rate, and pulse character and had good
 87 color throughout the procedures. Within 2 min of
 88 discontinuing xenon, both patients were oriented to time,
 89 place, and person; several hours later, they were able to recol-
 90 lect information given to them at this time. Since then, numer-
 91 ous clinical studies have investigated the effects of xenon in
 92 humans.

93 Safety and tolerability information regarding xenon for in-
 94 halation stem mostly from published literature describing clin-
 95 ical studies investigating the anesthetic properties of xenon
 96 gas, and the publications contain only summary information.
 97 In aggregate, the literature suggests that administration of xe-
 98 non, as a general anesthetic, to patients both with and without
 99 cardiovascular disease is associated with hemodynamic

stability that is unparalleled in critical care settings. Side ef- 100
 101 fects identified in the literature that are frequently associated
 102 with the use of xenon gas for inhalation as a general anesthetic
 103 include raised intracranial pressure [9], bradycardia [3], and
 104 nausea and vomiting [10]. Although bradycardia is a safety
 105 concern identified for xenon anesthesia, if heart rate slows to
 106 the point that systemic blood pressure decreases, then standard
 107 positive chronotropic agents such as anti-muscarinic agents
 108 (e.g., glycopyrrolate or atropine) and β_1 adrenergic agonists
 109 (e.g., isoproterenol) can be administered to reverse it. Xenon is
 110 not known to interfere with oxygenation, but in an oxygen and
 111 xenon mixture, the greater the percentage of inhaled xenon
 112 administered to a subject, the lower the fraction of inspired
 113 oxygen that can be administered. Under circumstances in
 114 which lung oxygenation is compromised (e.g., from pulmo-
 115 nary edema), a higher fraction of inspired oxygen may be
 116 required to prevent arterial hypoxemia.

Xenon has a favorable pharmacokinetic (PK) profile for 117
 118 anesthesia with fast induction and emergence, which is inde-
 119 pendent of the duration of exposure. This PK effect is attrib-
 120 utable to its low blood-gas partition coefficient of 0.115 [11],
 121 which is significantly lower than those of other inhalational
 122 anesthetics (nitrous oxide, 0.47; sevoflurane, 0.65; desflurane,
 123 0.42). As xenon is excreted by the lungs with no biotransfor-
 124 mation by the renal or hepatic systems, it may prove to be the
 125 anesthetic of choice in certain circumstances when liver or
 126 kidney function decrements.

Xenon has an oil/water solubility coefficient of 20, which is 127
 128 the highest coefficient of all noble gases, and it is the only
 129 noble gas with anesthetic properties at atmospheric pressures.
 130 The physico-chemical properties of xenon are detailed in
 131 Table 2.

Cardiovascular Effects of Xenon in Patients 132
Without Cardiac Diseases 133

In 1990, Lachmann and associates published a randomized 134
 135 double-blind trial comparing the efficacy and potency of xe-
 136 non with those of nitrous oxide, with special focus on the
 137 cardiovascular and respiratory systems [12]. The authors

Table 2 Physico-chemical properties: Ostwald solubility coefficients of xenon at 37 °C	Ostwald solubility coefficients (mL gas/ mL liquid) at 37 °C		t2.1
			t2.2
	Water/gas	0.075	t2.3
	Oil/gas	1.8	t2.4
	Blood/gas	0.115	t2.5
	Oil/water	20	t2.6
	Muscle/liver/kidney	0.10	t2.7
	Adipose tissue	1.3	t2.8
	Brain, gray substance	0.13	t2.9
	Brain, white substance	0.23	t2.10

138 concluded that xenon is a more potent anesthetic than nitrous
 139 oxide in suppressing response to surgical stimuli and main-
 140 taining hemodynamic stability. Lachmann's group also com-
 141 pared the effect of xenon and nitrous oxide on the neurohu-
 142 moral response and hemodynamics of 32 ASA class I–II pa-
 143 tients, with the same protocol as described above [13]. The
 144 investigators concluded that xenon has more favorable hemo-
 145 dynamic, neurohumoral, and antinociceptive properties than
 146 nitrous oxide. Luttrupp and associates investigated the effects
 147 of xenon on in vivo cardiac function using transesophageal
 148 echocardiography and hemodynamic measurements [14]. The
 149 fractional area in a short-axis view of the left ventricle at the
 150 level of the papillary muscles remained unchanged, suggest-
 151 ing that xenon anesthesia had no adverse effect on myocardial
 152 function as well as hemodynamics. The first multicenter ran-
 153 domized control trial, involving 224 patients in six centers,
 154 compared xenon/oxygen with isoflurane/nitrous oxide anes-
 155 thesia and concluded that xenon anesthesia is as safe and
 156 effective as the isoflurane/nitrous oxide regimen, with the ad-
 157 vantage that xenon exhibited more rapid recovery [15]. Also,
 158 significantly fewer xenon-anesthetized patients required ino-
 159 tropic support than the isoflurane group. In a single center
 160 study involving 160 patients, the hemodynamic effects were
 161 compared between those randomized to receive either xenon
 162 or propofol [16]. While systolic blood pressure was well
 163 maintained after induction with xenon at near baseline levels,
 164 propofol caused a significant post-induction decline in pres-
 165 sure that persisted throughout maintenance of general anes-
 166 thesia. Heart rates were significantly lower in the patients who
 167 received xenon. In a multicenter study involving 252 patients
 168 scheduled for elective non-cardiovascular surgery, hemody-
 169 namic stability, including transesophageal echocardiography,
 170 was compared between patients randomized to receive either
 171 xenon or isoflurane [17]. While isoflurane decreased the myo-
 172 cardial contractile index, no such change was noted in the
 173 xenon-anesthetized patients, leading the authors to opine that
 174 xenon enables cardiovascular stability.

175 **Cardiovascular Effects of Xenon in Patients With**
 176 **Cardiovascular Diseases**

177 Ten patients after coronary artery bypass graft surgery (per-
 178 formed on cardiopulmonary bypass) were randomized to re-
 179 ceive either propofol or xenon for sedation while being venti-
 180 lated in the ICU [18]. The patients were crossed over to the
 181 alternative sedative after some hours. Compared with propofol
 182 sedation, xenon sedation did not change the heart rate or blood
 183 pressure; left ventricular stroke work index was similar.

184 Effects of xenon on hemodynamics in patients scheduled
 185 for coronary artery bypass graft surgery have also been
 186 assessed [19]. Statistically significant differences were found
 187 between the two groups' mean arterial pressure (MAP), frac-
 188 tional area change of the left ventricle, and end-diastolic area

of the left ventricle. Xenon decreased the MAP and fractional
 area change significantly less and increased end-diastolic area
 significantly more than nitrous oxide. In a safety and feasibil-
 ity study involving 20 patients undergoing coronary artery
 bypass surgery, xenon, at varying concentrations (0%, 20%,
 35%, and 50% v/v), was administered while on cardiopulmo-
 nary bypass [20]. Despite theoretical concerns about expan-
 sion of gas bubbles, the cerebral embolic load, measured by
 middle cerebral artery Doppler, was no higher in patients who
 received xenon. Troponin levels tended to be lower 24 h after
 surgery in patients who received xenon. Twenty-six patients
 scheduled for implantation of an internal cardioverter-
 defibrillator were randomized to receive either xenon or
 propofol (both with remifentanyl) for maintenance of general
 anesthesia [21]. Most of these patients had heart failure from
 ischemic heart disease or dilated cardiomyopathy. In contrast
 to propofol, surgical patients maintained on xenon had no
 changes in either the MAP or the left ventricular ejection
 fraction.

Central Nervous System Effects of Xenon

Volunteers ($n = 12$) were randomized to receive general anes-
 thesia with xenon or propofol and the cerebral metabolic rate
 was assessed with the positron emission tomography (PET)
 ligand ^{18}F -fluorodeoxyglucose [22]. The xenon-exposed vol-
 unteers had cerebral metabolic rates globally reduced by 26%
 compared with those exposed to propofol alone. In another
 study, using ^{15}O -labeled water, the regional cerebral blood
 flow was monitored by PET scanning during xenon anesthesia
 in nine volunteers [23]. Xenon statistically significantly de-
 creased the regional cerebral blood flow in several of the gray
 matter areas studied while regional cerebral blood flow in-
 creased by 22.1% ($\pm 13.6\%$) in the white matter. A follow-
 up PET study, involving five healthy subjects, assessed re-
 gional cerebral blood flow and regional cerebral glucose me-
 tabolism using ^{15}O -labeled water and ^{18}F -labeled
 fluorodeoxyglucose, respectively [24]. In general, the regional
 reduction in cerebral metabolism was greater than the regional
 decrement in cerebral blood flow. Luttrupp et al. [14] investi-
 gated the effects of inhalation of 65% xenon on cerebral blood
 flow velocities, using Doppler sonography in 17 ASA class I
 patients undergoing abdominal surgery; they found that cere-
 bral blood flow velocity was unchanged during the first 5 min
 of xenon anesthesia, but was significantly increased in the left
 and right, middle, and the right anterior cerebral arteries after
 15 and 30 min. In addition, Giller et al. [25] noted that admin-
 istration of 25%, 30%, or 35% of xenon for 5 min to normal
 volunteers resulted in an increase in cerebral blood flow, mea-
 sured by Doppler velocity, in 85% of subjects and a decrease
 in cerebral blood flow in 15% of subjects. These findings are
 in contrast to the findings of the PET studies described in the
 preceding paragraph. Reasons for these discrepancies could

240 be differences in the patient populations (i.e., healthy volun- 288
241 teers versus patients undergoing surgery), differences in the 289
242 duration of xenon administration, and differences in the meth- 290
243 odology used to assess blood flow. In a trial involving supple- 291
244 mentation of therapeutic hypothermia with administration of 292
245 xenon to neonates suffering from hypoxic ischemic encephal- 293
246 opathy, Azzopardi and colleagues reported a significant re- 294
247 duction in seizure activity in patients randomized to receive 295
248 30% xenon [26]. 296

249 Neuroprotection 297

250 Xenon is thought to exert neuroprotective action by acting as an 301
251 antagonist at NMDA receptors. Excessive entry of calcium, 302
252 mediated by NMDA receptors, triggers biochemical cascades 303
253 that ultimately lead to neuronal cell death. NMDA-induced neu- 304
254 rotoxicity is through “excitotoxicity” from overactivation of 305
255 NMDA receptors that underlies the acute neuronal injury ob- 306
256 served following insults such as stroke, cardiac arrest, and trau- 307
257 matic brain injury. NMDA receptor antagonists are neuropro- 308
258 tective in in vitro and in vivo brain injury models [27]. 309
Q6 259 Following the discovery that xenon inhibits NMDA receptors 310
260 [5], it was shown that xenon could protect neuronal cell cultures 311
261 against injury induced by NMDA, glutamate, or oxygen- 312
262 glucose deprivation [28]. The same study showed xenon to be 313
263 neuroprotective in vivo against neuronal injury caused by sub- 314
264 cutaneous injection of *N*-methyl (D, L)-aspartate in rats. 315
265 Subsequently, this finding was corroborated by Petzelt et al., 316
266 in an in vitro model of hypoxia [29] and in an in vivo model 317
267 of stroke [30]. Other NMDA receptor antagonists such as ni- 318
268 trous oxide, ketamine, and dizocilpine (MK-801) have intrinsic 319
269 neurotoxicity, but xenon not only appears to be devoid of these 320
270 neurotoxic effects but also ameliorates the injury produced by 321
271 other NMDA antagonists [31]. Furthermore, xenon upregulates 322
272 the transcription factor hypoxia inducible factor 1 alpha (HIF 323
273 1α) and its downstream cytoprotective effectors including eryth- 324
274 ropoietin [32, 33]. Xenon has now been shown to afford neuro- 325
275 protection in a variety of mammalian in vitro and in vivo models 326
276 and meets the Stroke Treatment Academic Industry Roundtable 327
277 recommendation for proceeding to clinical trials [34]. 328

278 Phase II Clinical Trial in Out-of-Hospital Cardiac Arrest 329 279 Patients 330

280 Based upon successful animal studies investigating the effects 331
281 of xenon in the setting of cardiac arrest [35, 36] and because of 332
282 the synergistic interaction between xenon and therapeutic hy- 333
283 pothemia [37, 38], the Xe-Hypotheca trial (NCT 00879892; 334
284 May 2009–September 2014) was initiated at a single academ- 335
285 ic site (University of Turku Hospital, Finland) to determine the 336
286 feasibility and cardiac safety of inhaled xenon when added to 337
287 therapeutic hypothermia for successfully resuscitated out-of- 338

hospital cardiac arrest (OHCA) patients [39]. Feasibility was 288
established after the first 36 patients were randomized to re- 289
ceive either therapeutic hypothermia alone ($n = 18$) or thera- 290
peutic hypothermia in combination with xenon by inhalation 291
($n = 18$), with a target concentration of at least 40% xenon for 292
24 h. In the xenon group, the median end-tidal xenon concen- 293
tration was 47% and duration of xenon inhalation was 25.5 h. 294
Xenon did not induce significant conduction, repolarization, 295
or rhythm abnormalities. Median dose of norepinephrine dur- 296
ing hypothermia was 2.95 mg in xenon-treated patients and 297
5.30 mg in patients treated with therapeutic hypothermia alone 298
($p = 0.06$). Heart rate was statistically significantly lower in 299
xenon-treated patients than that in patients treated with thera- 300
peutic hypothermia alone ($p = 0.04$). From the initial results of 301
this trial, the investigators concluded that xenon treatment in 302
combination with hypothermia is feasible and has favorable 303
cardiac features in OHCA patients. The Xe-Hypotheca trial 304
was extended to a second site in 2013 (University of Helsinki 305
Hospital, Finland) with an expanded cohort; the effect of xe- 306
non on ischemic white matter damage was assessed by frac- 307
tional anisotropy from diffusion tensor magnetic resonance 308
imaging (MRI) [40]. Neurological outcome and mortality at 309
6 months were also assessed. A total of 224 patients were 310
screened for eligibility. One hundred and ten OHCA patients, 311
aged 24–76 years, were randomized to receive either hypo- 312
themia treatment alone for 24 h (control group, $n = 55$) or 313
inhaled xenon, administered to achieve an end-tidal xenon 314
concentration of at least 40%, combined with hypothermia 315
(33 °C) for 24 h (xenon group, $n = 55$). The primary endpoint 316
was severity of ischemic white matter brain injury as evaluat- 317
ed by fractional anisotropy from diffusion tensor MRI; MRIs 318
were scheduled within 16 h after rewarming of a patient (rang- 319
ing between 36 and 52 h after OHCA). Secondary endpoints 320
were neurological outcome, assessed with cerebral perfor- 321
mance category score (from 1 = conscious, alert, able to work, 322
might have mild cognitive deficit, to 5 = death) and modified 323
Rankin Scale (score from 0 = no symptoms at all to 6 = death), 324
mortality at 6 months, and complication rate within 7 days of 325
post-CA. However, the trial was not powered to detect statisti- 326
cally significant differences in clinical efficacy (i.e., mortal- 327
ity at 6 months and neurological outcome) between groups. 328
The primary endpoint was assessed in the complete case pop- 329
ulation. Survival at 6 months and complication rate were an- 330
alyzed in the intention-to-treat population. Kaplan–Meier sur- 331
vival curves and a Cox proportional hazards model were used 332
to compare mortality at 6 months between groups. 333

Of the randomized patients, six patients in the control 334
group and seven patients in the xenon group were missing 335
MRI data and were excluded from the complete case popula- 336
tion. The mean (\pm SD) global fractional anisotropy value of all 337
voxels in the xenon group (0.433 [\pm 0.028]) was significantly 338
different than that in the control group (0.419 [\pm 0.033]) ($p =$ 339
0.03). The age-, gender-, and site-adjusted mean global 340

341 fractional anisotropy values were 3.8% higher in the xenon
 342 group than those in the control group (adjusted mean difference
 343 0.016 [95% CI, 0.005 to 0.027]; $p = 0.006$). The severity
 344 of observed widespread injury was demonstrated; on average,
 345 41.7% of the white matter tracts, including major commissural,
 346 associative, and projection fibers, were significantly more
 347 severely injured in the control group than in the xenon group.
 348 These fibers are involved in multiple important cognitive
 349 functions such as attention, memory, language, emotions, auditory,
 350 visual and executive processing, and motor functions
 351 of the body.

352 At 6 months, 75 patients (68.2%) were alive and able to
 353 provide follow-up data. In ordinal analysis of modified
 354 Rankin Scale, median (interquartile range) value was 1 (0 to
 355 6) in the xenon group and 1 (0 to 6) in the control group
 356 (median difference = 0 [95% CI, 0 to 0]; $p = 0.68$). The
 357 Kaplan–Meier survival estimate (panel A) after 6 months
 358 was 27.7% (15/55 patients) in the xenon group and 34.5%
 359 (19/55 patients) in the control group (adjusted hazard ratio =
 360 0.49 [95% CI, 0.23 to 1.01]; $p = 0.053$) (Fig. 1).

361 It was concluded that among comatose survivors of
 362 OHCA, treatment with inhaled xenon combined with hypothermia
 363 resulted in less white matter damage, as measured by
 364 fractional anisotropy of diffusion tensor MRI, than treatment
 365 with hypothermia alone. In contrast, there was no statistically

366 significant difference between groups in neurological out-
 367 comes or mortality at 6 months. However, the study was un-
 368 derpowered to detect a statistically significant difference in
 369 clinical outcome due to the rarity of severe neurological im-
 370 pairment in long-term survivors after CA; about 90% of CA
 371 patients who are alive at the 6-month follow-up have experi-
 372 enced a good neurological outcome (cerebral performance
 373 category 1–2). While there was no statistically significant dif-
 374 ference in neurological outcomes or mortality at 6 months,
 375 unpublished data demonstrates that there was a trend to a
 376 survival benefit.

377 A predefined secondary objective was to assess the effect of
 378 inhaled xenon on myocardial ischemic damage [41]. Troponin-
 379 T levels were measured at hospital admission, and at 24 h, 48 h,
 380 and 72 h post-CA. Among comatose OHCA patients, inhaled
 381 xenon combined with hypothermia resulted in less severe myo-
 382 cardial injury than with hypothermia alone, as demonstrated by
 383 the significantly reduced release of troponin-T.

384 Rates of serious adverse events (SAEs) in the xenon group
 385 were not significantly different from the rates of SAEs in the
 386 standard of care group [40]. SAEs seen in both the xenon and
 387 standard of care groups include status epilepticus, acute kid-
 388 ney injury (in the “risk,” “injury,” or “failure,” RIFLE cate-
 389 gories), pulmonary edema, ventricular fibrillation, ventricular
 390 tachycardia, atrial fibrillation, coronary stent thrombosis,

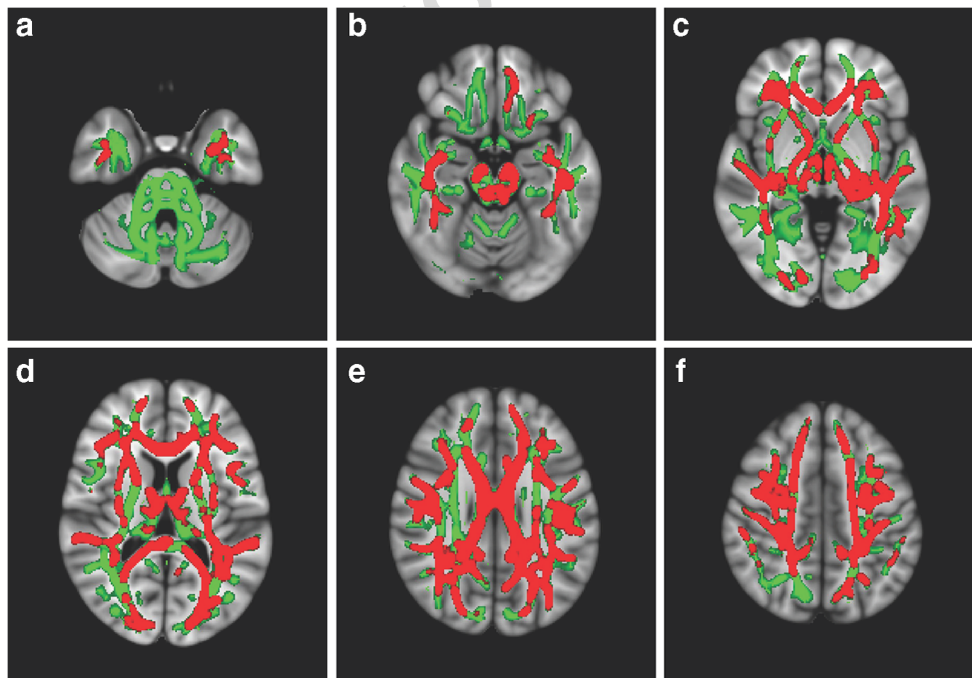


Fig. 1 Whole-brain fiber tractography of fractional anisotropy. Fractional anisotropy (FA) is a scalar value representing directionality of water diffusion. White matter damage leads to a loss of microstructural organization that can be quantified by the loss of directionality in the diffusion of water molecules in the white matter tracts. Using data from a diffusion tensor imaging sequence of an MRI scan performed within 72 h of rewarming, panels a–f represent sequential ascending horizontal planes of the major tracts in the brain. The visualization presents the

results of the voxel-wise tract-based spatial statistics analysis of FA values between the xenon group and the control group. Voxels with significantly ($p < 0.05$, family-wise error corrected for multiple comparisons) higher fractional anisotropy values in the xenon group were identified and are shown in red in the statistical visualization (i.e., 41.7% of all 119,013 analyzed voxels), whereas areas in which there were no significant difference in fractional anisotropy values between the groups are shown in green (modified from reference [40])

391 sepsis, pneumonia, multi-organ failure, adult respiratory dis-
 392 tress syndrome, and subarachnoid hemorrhage SAEs only ob-
 393 served in the xenon group included bradycardia treated with
 394 pacemaker ($n = 1$ event) and serious bleeding (gastrointesti-
 395 nal, $n = 1$ event). SAEs only observed in the standard of care
 396 group included third-degree atrioventricular block ($n = 1$
 397 event), carotid dissection ($n = 1$ event), carotid thrombosis
 398 ($n = 1$ event), and serious bleeding (intracranial, $n = 1$ event).

399 Conclusion and Potential Future Applications

400 Xenon exhibits many features of a putative neuroprotective
 401 agent with an ideal pharmacokinetic profile for use following
 402 acute neurological injury. Studies involving several different
 403 acute neurological injury models in a variety of animal species
 404 from four laboratories have consistently demonstrated the
 405 neuroprotective efficacy of xenon even when administered
 406 as long as 6 h following neurological injury. The mechanisms
 407 for neuroprotection appear to involve (i) antagonism of the
 408 NMDA receptor whose activation is pivotal for the excitotoxic
 409 damage that follows neurologic injury, and (ii) upregulation of
 410 HIF 1 α and the resulting cytoprotection from erythropoietin, a
 411 downstream effector of the transcription factor.

412 A phase 2 RCT (Xe-Hypotheca) demonstrated significant-
 413 ly less white matter brain damage, as reflected by higher glob-
 414 al fractional anisotropy values, in subjects randomized to re-
 415 ceive xenon by inhalation during the 24-h period of targeted
 416 temperature management.

417 The stage is now set for a pivotal phase 3 RCT,
 418 XePOCHAS (NCT03176186), to determine the efficacy
 419 (using endpoints of good functional outcome and survival at
 420 30 and 90 days), safety, and cost-effectiveness of xenon, at a
 421 dose of 50% of 1 atmosphere, in the management of post-
 422 cardiac arrest syndrome patients. The trial is likely to report
 423 its outcome in 2021 or before depending on the interim anal-
 424 ysis at the halfway point of the 1436-patient trial to be con-
 425 ducted in 7 countries in Europe and North America.

426 In the event that xenon exhibits neuroprotective efficacy in
 427 the XePOCHAS trial, then subsequent trials are likely to be
 428 conducted in other acute neurological injury settings including
 429 stroke and traumatic brain injury. As with the XePOCHAS trial,
 430 the maximal dose of xenon is likely to be restricted to $\leq 50\%$
 431 of 1 atmosphere as these patients may also have lung injury that
 432 will require an F_iO_2 of no less than 0.5 to avoid hypoxemia.

433 A further clinical application may be in pediatric surgical
 434 settings to obviate the occurrence of anesthetic-induced devel-
 435 opmental neurotoxicity (AIDN). For AIDN, xenon may be
 436 particularly effective both by reducing exposure to high con-
 437 centrations of neurotoxic volatile anesthetics by virtue of xe-
 438 non's anesthetic properties and because the neurotoxicity may
 439 be obviated by xenon's neuroprotective properties.

References

1.	Dmochowski I (2009) Xenon out of its shell. <i>Nat Chem</i> 1(3):250	441
2.	Cullen SC, Gross EG (1951) The anesthetic properties of xenon in animals and human beings, with additional observations on krypton. <i>Science</i> 113(2942):580–582	442 443 444
3.	Law LS, Lo EA, Gan TJ (2016) Xenon anesthesia: a systematic review and meta-analysis of randomized controlled trials. <i>Anesth Analg</i> 122(3):678–697	445 446 447
4.	Franks NP (2008) General anaesthesia: from molecular targets to neuronal pathways of sleep and arousal. <i>Nat Rev Neurosci</i> 9(5):370–386	448 449 450
5.	Franks NP, Dickinson R, de Sousa SL, Hall AC, Lieb WR (1998) How does xenon produce anaesthesia? <i>Nature</i> 396(6709):324	451 452
6.	Dickinson R, Peterson BK, Banks P, Similliss C, Martin JC, Valenzuela CA, Maze M, Franks NP (2007) Competitive inhibition at the glycine site of the N-methyl-D-aspartate receptor by the anesthetics xenon and isoflurane: evidence from molecular modeling and electrophysiology. <i>Anesthesiology</i> 107(5):756–767	453 454 455 456 457
7.	Gruss M, Mathie A, Lieb WR, Franks NP (2004) The two-pore-domain K(+) channels TREK-1 and TASK-3 are differentially modulated by copper and zinc. <i>Mol Pharmacol</i> 66(3):530–537	458 459 460
8.	Bantel C, Maze M, Trapp S (2010) Noble gas xenon is a novel adenosine triphosphate-sensitive potassium channel opener. <i>Anesthesiology</i> 112(3):623–630	461 462 463
9.	Plougmann J, Astrup J, Pedersen J, Gyldensted C (1994) Effect of stable xenon inhalation on intracranial pressure during measurement of cerebral blood flow in head injury. <i>J Neurosurg</i> 81(6):822–828	464 465 466 467
10.	Lo EA, Law LS, Gan TJ (2016) Paradox of the incidence of post-operative nausea and vomiting after xenon-based anaesthesia. <i>Br J Anaesth</i> 116(6):881–883	468 469 470
11.	Goto T, Suwa K, Uezono S, Ichinose F, Uchiyama M, Morita S (1998) The blood-gas partition coefficient of xenon may be lower than generally accepted. <i>Br J Anaesth</i> 80(2):255–256	471 472 473
12.	PMID: 27199321 (Lo, 2016, <i>Br J Anaesth</i> , Paradox of the incidence of postoperative nausea and vomiting after xenon-based anaesthesia) Need PMID for Lachmann 1990	474 475 476
13.	Boomsma F, Rupprecht J, Man in 't Veld AJ, de Jong FH, Dzoljic M, Lachmann B (1990) Haemodynamic and neurohumoral effects of xenon anaesthesia. A comparison with nitrous oxide. <i>Anaesthesia</i> 45(4):273–278	477 478 479 480
14.	Luttrupp HH, Romner B, Perhag L, Eskilsson J, Fredriksen S, Werner O (1993) Left ventricular performance and cerebral haemodynamics during xenon anaesthesia. A transoesophageal echocardiography and transcranial Doppler sonography study. <i>Anaesthesia</i> 48(12):1045–1049	481 482 483 484 485
15.	Rossaint R, Reyle-Hahn M, Schulte Am Esch J, Scholz J, Scherpereel P, Vallet B, Giunta F, Del Turco M et al (2003) Multicenter randomized comparison of the efficacy and safety of xenon and isoflurane in patients undergoing elective surgery. <i>Anesthesiology</i> 98(1):6–13	486 487 488 489 490
16.	Coburn M, Kunitz O, Baumert JH, Hecker K, Haaf S, Zühlsdorff A, Beeker T, Rossaint R (2005) Randomized controlled trial of the haemodynamic and recovery effects of xenon or propofol anaesthesia. <i>Br J Anaesth</i> 94(2):198–202	491 492 493 494
17.	Wappler F, Rossaint R, Baumert J, Scholz J, Tonner PH, van Aken H, Berendes E, Klein J et al (2007) Multicenter randomized comparison of xenon and isoflurane on left ventricular function in patients undergoing elective surgery. <i>Anesthesiology</i> 106(3):463–471	495 496 497 498
18.	Dingley J, King R, Hughes L, Terblanche C, Mahon S, Hepp M, Youhana A, Watkins A (2001) Exploration of xenon as a potential cardiostable sedative: a comparison with propofol after cardiac surgery. <i>Anaesthesia</i> 56(9):829–835	499 500 501 502

503 19. Goto T, Hanne P, Ishiguro Y, Ichinose F, Niimi Y, Morita S (2004) Cardiovascular effects of xenon and nitrous oxide in patients during
504 fentanyl-midazolam anaesthesia. *Anaesthesia* 59(12):1178–1183 552
505
506 20. Lockwood GG, Franks NP, Downie NA, Taylor KM, Maze M (2006) Feasibility and safety of delivering xenon to patients under-
507 going coronary artery bypass graft surgery while on cardiopulmonary
508 bypass: phase I study. *Anesthesiology* 104(3):458–465 553
509
510 21. Baumert JH, Hecker KE, Hein M, Reyle-Hahn M, Horn NA, Rossaint R (2005) Effects of xenon anaesthesia on the circulatory
511 response to hypoventilation. *Br J Anaesth* 95(2):166–171 554
512
513 22. Rex S, Schaefer W, Meyer PH, Rossaint R, Boy C, Setani K, Büll U, Baumert JH (2006) Positron emission tomography study of re-
514 gional cerebral metabolism during general anesthesia with xenon in
515 humans. *Anesthesiology* 105(5):936–943 555
516
517 23. Laitio RM, Kaisti KK, Långsjö JW, Aalto S, Salmi E, Maksimow
518 A, Aantaa R, Oikonen V et al (2007) Effects of xenon anesthesia on
519 cerebral blood flow in humans: a positron emission tomography
520 study. *Anesthesiology* 106(6):1128–1133 556
521
522 24. Laitio RM, Långsjö JW, Aalto S, Kaisti KK, Salmi E, Maksimow
523 A, Aantaa R, Oikonen V et al (2009) The effects of xenon anesthe-
524 sia on the relationship between cerebral glucose metabolism and
525 blood flow in healthy subjects: a positron emission tomography
526 study. *Anesth Analg* 108(2):593–600 557
527
528 25. Giller CA, Purdy P, Lindstrom WW (1990) Effects of inhaled stable
529 xenon on cerebral blood flow velocity. *AJNR Am J Neuroradiol*
530 11(1):177–182 558
531
532 26. Azzopardi D, Robertson NJ, Kapetanakis A, Griffiths J, Rennie JM,
533 Mathieson SR, Edwards AD (2013) Anticonvulsant effect of xenon
534 on neonatal asphyxial seizures. *Arch Dis Child Fetal Neonatal Ed*
535 98(5):F437–F439 559
536
537 27. Choi DW, Koh JY, Peters S (1988) Pharmacology of glutamate
538 neurotoxicity in cortical cell culture: attenuation by NMDA antagon-
539 ists. *J Neurosci* 8(1):185–196 560
540
541 28. Wilhelm S, Ma D, Maze M, Franks NP (2002) Effects of xenon on
542 in vitro and in vivo models of neuronal injury. *Anesthesiology*
543 96(6):1485–1491 561
544
545 29. Petzelt C, Blom P, Schmehl W, Müller J, Kox WJ (2003) Prevention
546 of neurotoxicity in hypoxic cortical neurons by the noble gas xenon.
547 *Life Sci* 72(17):1909–1918 562
548
549 30. David HN, Leveille F, Chazalviel L, MacKenzie ET, Buisson A,
550 Lemaire M, Abraini JH (2003) Reduction of ischemic brain damage
551 by nitrous oxide and xenon. *J Cereb Blood Flow Metab* 23(10):
552 1168–1173 563
553
554 31. Nagata A, Nakao Si S, Nishizawa N, Masuzawa M, Inada T, Murao
555 K, Miyamoto E, Shingu K (2001) Xenon inhibits but N(2)O en-
556 hances ketamine-induced c-Fos expression in the rat posterior cin-
557 gulate and retrosplenial cortices. *Anesth Analg* 92(2):362–368 564
558
559 32. Ma D, Lim T, Xu J, Tang H, Wan Y, Zhao H, Hossain M, Maxwell
560 PH et al (2009) Xenon preconditioning protects against renal
561 ischemic-reperfusion injury via HIF-1alpha activation. *J Am Soc*
562 *Nephrol* 20(4):713–720 565
563
564 33. Stoppe C, Ney J, Brenke M, Goetzenich A, Emontzpohl C, Schälte
565 G, Grottko O, Moeller M et al (2016) Sub-anesthetic xenon in-
566 creases erythropoietin levels in humans: a randomized controlled
567 trial. *Sports Med* 46(11):1753–1766 568
568
569 34. Albers GW, Goldstein LB, Hess DC, Wechsler LR, Furie KL,
570 Gorelick PB, Hurn P, Liebeskind DS et al (2011) Stroke
571 Treatment Academic Industry Roundtable (STAIR) recommenda-
572 tions for maximizing the use of intravenous thrombolytics and
573 expanding treatment options with intra-arterial and neuroprotective
574 therapies. *Stroke* 42(9):2645–2650 575
575
576 35. Fries M, Nolte KW, Coburn M, Rex S, Timper A, Kottmann K,
577 Siepmann K, Häusler M et al (2008) Xenon reduces
578 neurohistopathological damage and improves the early neurologi-
579 cal deficit after cardiac arrest in pigs. *Crit Care Med* 36(8):2420–
580 2426 576
581
582 36. Fries M, Brücken A, Çizen A, Westerkamp M, Löwer C, Deike-
583 Glindemann J, Schnorrenberger NK, Rex S et al (2012) Combining
584 xenon and mild therapeutic hypothermia preserves neurological
585 function after prolonged cardiac arrest in pigs. *Crit Care Med*
586 40(4):1297–1303 577
587
588 37. Ma D, Hossain M, Chow A, Arshad M, Battson RM, Sanders RD,
589 Mehmet H, Edwards AD et al (2005) Xenon and hypothermia com-
590 bine to provide neuroprotection from neonatal asphyxia. *Ann*
591 *Neurol* 58(2):182–193 578
592
593 38. Martin JL, Ma D, Hossain M, Xu J, Sanders RD, Franks NP, Maze
594 M (2007) Asynchronous administration of xenon and hypothermia
595 significantly reduces brain infarction in the neonatal rat. *Br J*
596 *Anaesth* 98(2):236–240 579
597
598 39. Arola OJ, Laitio RM, Roine RO, Grönlund J, Saraste A, Pietilä M,
599 Airaksinen J, Perttilä J et al (2013) Feasibility and cardiac safety of
600 inhaled xenon in combination with therapeutic hypothermia follow-
601 ing out-of-hospital cardiac arrest. *Crit Care Med* 41(9):2116–2124
602 582
603
604 40. Laitio R, Hynninen M, Arola O, Virtanen S, Parkkola R,
605 Saunavaara J, Roine RO, Grönlund J et al (2016) Effect of inhaled
606 xenon on cerebral white matter damage in comatose survivors of
607 out-of-hospital cardiac arrest: a randomized clinical trial. *JAMA*
608 315(11):1120–1128 583
609
610 41. Arola O, Saraste A, Laitio R, Airaksinen J, Hynninen M, Bäcklund
611 M, Ylikoski E, Wennervirta J et al (2017) Inhaled xenon attenuates
612 myocardial damage in comatose survivors of out-of-hospital cardiac
613 arrest: the Xe-HypotheCa trial. *J Am Coll Cardiol* 70(21):2652–
614 2660 584
615
616
617
618
619
620
621
622
623
624
625
626
627
628
629
630
631
632
633
634
635
636
637
638
639
640
641
642
643
644
645
646
647
648
649
650
651
652
653
654
655
656
657
658
659
660
661
662
663
664
665
666
667
668
669
670
671
672
673
674
675
676
677
678
679
680
681
682
683
684
685
686
687
688
689
690
691
692
693
694
695
696
697
698
699
700
701
702
703
704
705
706
707
708
709
710
711
712
713
714
715
716
717
718
719
720
721
722
723
724
725
726
727
728
729
730
731
732
733
734
735
736
737
738
739
740
741
742
743
744
745
746
747
748
749
750
751
752
753
754
755
756
757
758
759
760
761
762
763
764
765
766
767
768
769
770
771
772
773
774
775
776
777
778
779
780
781
782
783
784
785
786
787
788
789
790
791
792
793
794
795
796
797
798
799
800
801
802
803
804
805
806
807
808
809
810
811
812
813
814
815
816
817
818
819
820
821
822
823
824
825
826
827
828
829
830
831
832
833
834
835
836
837
838
839
840
841
842
843
844
845
846
847
848
849
850
851
852
853
854
855
856
857
858
859
860
861
862
863
864
865
866
867
868
869
870
871
872
873
874
875
876
877
878
879
880
881
882
883
884
885
886
887
888
889
890
891
892
893
894
895
896
897
898
899
900
901
902
903
904
905
906
907
908
909
910
911
912
913
914
915
916
917
918
919
920
921
922
923
924
925
926
927
928
929
930
931
932
933
934
935
936
937
938
939
940
941
942
943
944
945
946
947
948
949
950
951
952
953
954
955
956
957
958
959
960
961
962
963
964
965
966
967
968
969
970
971
972
973
974
975
976
977
978
979
980
981
982
983
984
985
986
987
988
989
990
991
992
993
994
995
996
997
998
999
1000

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

AUTHOR QUERIES

AUTHOR PLEASE ANSWER ALL QUERIES.

- Q1. Please check if the corresponding author's telecommunications data are correctly captured/ indicated.
- Q2. Please check if the affiliations are presented correctly.
- Q3. An abstract section is desired for this manuscript, but none is provided in the source files. Please then consider providing an appropriate abstract that substantively meets the requirements declared in the "Instructions for Authors" page of this journal.
- Q4. Keywords are required. The following are suggested: xenon, neuroprotection, medical application. Please check if appropriate; otherwise, please provide.
- Q5. Please check if the section headings are assigned to appropriate levels.
- Q6. The sentence "Following the discovery that xenon inhibits..." has been modified. Please check if the intended meaning was retained.
- Q7. A citation for Fig. 1 was inserted in the sentence "The Kaplan–Meier survival estimate (panel A)..." Please check if appropriate. Otherwise, please advise.

UNCORRECTED PROOF