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Article

CD40L Activates Platelet Integrin α IIb β 3 by Binding to the Allosteric Site (Site 2) in a KGD-Independent Manner and HIGM1 Mutations Are Clustered in the Integrin-Binding Sites of CD40L

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Abstract: CD40L is expressed in activated T cells, and it plays a major role in immune response and is a major therapeutic target for inflammation. High IgM syndrome type 1 (HIGM1) is a congenital functional defect in CD40L/CD40 signaling due to defective CD40L. CD40L is also stored in platelet granules and transported to the surface upon platelet activation. Platelet integrin α IIb β 3 is known to bind to fibrinogen and activation of α IIb β 3 is a key event that triggers platelet aggregation. Also, the KGD motif is critical for α IIb β 3 binding and the interaction stabilizes thrombus. Previous studies showed that CD40L binds to and activates integrins α v β 3 and α 5 β 1 and that HIGM1 mutations are clustered in the integrin-binding sites. However, the specifics of CD40L binding to α IIb β 3 were unclear. Here, we show that CD40L binds to α IIb β 3 in a KGD-independent manner using CD40L that lacks the KGD motif. Two HIGM1 mutants, S128E/E129G and L155P, reduced the binding of CD40L to the classical ligand-binding site (site 1) of α IIb β 3, indicating that α IIb β 3 binds to the outer surface of CD40L trimer. Also, CD40L bound to the allosteric site (site 2) of α IIb β 3 and allosterically activated α IIb β 3 without inside-out signaling. Two HIGM1 mutants, K143T and G144E, on the surface of trimeric CD40L suppressed CD40L-induced α IIb β 3 activation. These findings suggest that CD40L binds to α IIb β 3 in a manner different from that of α v β 3 and α 5 β 1 and induces α IIb β 3 activation. HIGM1 mutations are clustered in α IIb β 3 binding sites in CD40L and are predicted to suppress thrombus formation and immune responses through α IIb β 3.

Keywords: CD40L; integrins; α IIb β 3; High IgM syndrome type I (HIGM1); allosteric activation of integrins



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1. Introduction

Integrins are a superfamily of cell-surface heterodimers that recognize extracellular matrix (ECM), cell-surface ligands (e.g., vascular cell adhesion molecule-1 (VCAM-1) and intercellular adhesion molecule-1 (ICAM-1)), and soluble molecules (e.g., growth factors). Integrins play important roles in biological processes (e.g., wound healing and hemostasis) and in the pathogenesis of diseases [1,2]. The CD40 ligand (CD40L) is a tumor necrosis factor family member, which is expressed in activated T cells and platelets as a transmembrane form and released as a soluble form (sCD40L) by proteolytic cleavage, on both T cells and platelets. CD40L is an inflammatory mediator and a major therapeutic target [3,4]. CD40L is stored in platelet granules and rapidly transferred to the surface upon platelet activation by platelet agonists (e.g., thrombin and ADP). CD40L in activated platelets has been shown to play a role in atherosclerosis formation and stabilizing thrombus [5,6].

CD40L binds to several integrins. Previous reports suggest that integrins α IIb β 3 [5], α 5 β 1 [7], and α M β 2 [8] bind to CD40L. Each of these integrins interacts with CD40L

in a specific manner. α IIb β 3 is known to recognize the KGD motif of CD40L (residues 115–117 of CD40L) [5]. α 5 β 1 does not require the KGD motif [9]. Mutations of the CD40-binding site (Y145A, R203A, or Y145A/R203A double mutant) did not affect α 5 β 1-CD40L interaction [10], suggesting that α 5 β 1 and CD40-binding sites on CD40L are distinct. It has been reported that the N151A/Q166 mutation on the outside of trimeric CD40L suppresses α 5 β 1-mediated CD40L binding [9]. Also, the functions of the α 5 β 1-CD40L interaction are promoting tumor cell survival in the context of cancer and enhancing inflammation as well as T cell persistence in autoimmune conditions [3]. We recently showed that CD40L (residues 118–216) lacks the KGD motif bound to α v β 3, indicating that α v β 3 is a new CD40L receptor and CD40L binding to α v β 3 is not dependent on the KGD motif [11]. High IgM syndrome type 1 (HIGM1) is a congenital functional defect in CD40L/CD40 signaling due to defective CD40L [12]. HIGM1 is characterized by low or absent levels of serum IgG, IgA, and IgE, and normal or increased levels of serum IgM. These results from mutations in the pathway from B cell activation to isotype class switching due to CD40L mutations [12]. The CD40L-binding site in α v β 3 and α 5 β 1 was located in the trimeric interface inside the CD40L trimer and, interestingly, HIGM1 mutants were clustered in the trimeric interface and these HIGM1 mutants are defective in integrin binding, indicating that functional defects in these HIGM1 mutants are related to defective integrin binding of CD40L. Also, CD40L bound to the classical ligand-binding site of α v β 3 (site 1) and α v β 3 site 1 recognized the binding site in the trimeric interface, in which several HIGM1 mutants were clustered [11]. These findings indicate that CD40L interaction with integrins α v β 3 and α 5 β 1 is critical for immune responses and HIGM1 mutants affect integrin-CD40L interaction, leading to immune deficiency. Understanding the specifics of integrin-CD40L interaction, thus, is critical for new therapeutics for inflammatory diseases.

By virtual screening of a protein data bank using docking simulation, we identified the chemokine domain of CX3CL1 (fractalkine) [13] as a new ligand for integrins α v β 3, α 4 β 1, and α 5 β 1, and that CX3CL1 and integrins simultaneously bound to CX3CR1 [13]. The CX3CL1 mutant defective in integrin binding was defective in signaling and acted as an antagonist although it still bound to CX3CR1 [13], indicating that the direct integrin binding to CX3CL1 and subsequent integrin-CX3CL1-CX3CR1 ternary complex formation is required for its signaling functions. We unexpectedly discovered that CX3CL1 activated soluble integrin α v β 3 in 1 mM Ca²⁺ in cell-free conditions by binding to the allosteric ligand-binding site (site 2), which is distinct from site 1 in the integrin headpiece [14]. We assumed that the predicted allosteric site is open in the inactive form of integrin, in analogy to inactive allosteric enzymes, and we performed docking simulation of CX3CL1 and inactive (closed headpiece) integrin α v β 3 (1 JV2.pdb). CX3CL1 was shown to bind to the binding site distinct from site 1 [13]. Furthermore, we showed that peptides from site 2 of the integrin β subunit bound to CX3CL1 and blocked integrin activation by CX3CL1 [14]. Therefore, we concluded that site 2 is the allosteric binding site and involved in allosteric activation of integrins. We showed that CXCL12 (SDF-1, stromal cell-derived factor-1) activated integrins α v β 3, α 4 β 1, and α 5 β 1 by binding to site 2 [15], indicating that this mechanism of integrin activation is not limited to CX3CL1. We also found that CX3CL1, CXCL12, and CCL5 (Rantes) bound to site 1 of integrin α IIb β 3 and activated this integrin by binding to site 2, indicating that α IIb β 3 can be allosterically activated independent of the canonical inside-out signaling mechanism [16].

α IIb β 3 was identified as a receptor for CD40L [4,17] and is known to require the N-terminal KGD motif of CD40L (residues 115–117 of CD40L) for binding [4]. Using CD40L knockout (KO) mice and their wild-type (WT) littermates, CD40L was shown to be involved in thrombus stabilization, as evidenced by the increased thrombi rupture in injured arterioles of CD40L KO mice, owing to their low platelet content, compared with WT ones. Furthermore, treatment of CD40L KO platelets with recombinant sCD40L enhanced their thrombin-induced aggregation under high shear stress conditions [5,6], as well as the capacity of sCD40L to induce tyrosine phosphorylation of the integrin β chain and the abrogation of such signals in the presence of antibodies directed against β 3 tyrosine

phosphorylation or in mice with mutated $\beta 3$ tyrosine residues. By activating the integrin, CD40L promoted outside-in signaling in platelets, the formation of platelet microparticles, and enhancement of fibrinogen binding [17].

Activation of platelet integrin $\alpha \text{IIb}\beta 3$ is a key event in platelet aggregation and thrombus formation. The specifics of CD40L binding to $\alpha \text{IIb}\beta 3$ are unclear except that CD40L binding to $\alpha \text{IIb}\beta 3$ suggestively requires the KGD motif [5]. We thus studied how CD40L binds to $\alpha \text{IIb}\beta 3$ by using docking simulation and mutagenesis studies. For the present paper, we studied how CD40L binds to $\alpha \text{IIb}\beta 3$ and whether CD40L activates $\alpha \text{IIb}\beta 3$ in an allosteric manner. We showed that CD40L binds to $\alpha \text{IIb}\beta 3$ in a KGD-independent manner using CD40L that lacks the KGD motif. Two HIGM1 mutants, S128E/E129G and L155, reduced the binding of CD40L to the classical ligand-binding site (site 1) of $\alpha \text{IIb}\beta 3$, indicating that $\alpha \text{IIb}\beta 3$ binds to the outer surface of CD40L trimer. Also, CD40L bound to site 2 of $\alpha \text{IIb}\beta 3$ and allosterically activated $\alpha \text{IIb}\beta 3$ without inside-out signaling. Two HIGM1 mutants K143T and G144E on the surface of trimeric CD40L suppressed CD40L-induced $\alpha \text{IIb}\beta 3$ activation. These findings suggest that CD40L binds to $\alpha \text{IIb}\beta 3$ in a manner different from that of $\alpha \nu \beta 3$ and $\alpha 5\beta 1$ and induces $\alpha \text{IIb}\beta 3$ activation. HIGM1 mutations are clustered in $\alpha \text{IIb}\beta 3$ binding sites in CD40L and are predicted to suppress thrombus formation and immune responses through $\alpha \text{IIb}\beta 3$.

2. Materials and Method

2.1. Synthesis of Recombinant sCD40L with No KGD Motif

WT and mutant sCD40L [18] were synthesized as described [11]. sCD40L (residues 118–261) with N-terminal 6His tag (19 amino acid residues) migrated as a mostly single band with an expected molecular weight of monomer (163 amino acids, 18Kd) with a trace number of dimers and trimers. sCD40L mutants migrated as monomers with a size like that of WT CD40L (Supplemental Figure S1a).

2.2. Synthesis of Fibrinogen Fragments

Fibrinogen γ -chain C-terminal peptide. The [HHHHHHH]NRLTIGEGQQHHLGGAKQAGDV (6 His tag and Fibrinogen γ -chain C-terminal residues 390–411) was conjugated with the C-terminus of GST (glutathione S-transferase) (designated $\gamma \text{C}390\text{--}411$) in pGEXT2 vector (BamHI/EcoRI site) [19]. The protein was synthesized in *E. coli* BL21 and purified using glutathione affinity chromatography. The fibrinogen γ -chain C-terminal domain ($\gamma \text{C}151\text{--}411$) was generated as previously described [20].

2.3. Binding of Soluble $\alpha \text{IIb}\beta 3$ to CD40L

ELISA (enzyme-linked immunosorbent assay)-type binding assays were performed as described previously [13]. Briefly, wells of 96-well Immulon 2 microtiter plates (Dynatech Laboratories, Chantilly, VA, USA) were coated with 100 μL PBS containing sCD40L for 2 h at 37 $^{\circ} \text{C}$. Remaining protein-binding sites were blocked by incubating with PBS/0.1% BSA for 30 min at room temperature. After washing with PBS, soluble recombinant $\alpha \text{IIb}\beta 3$ (AgroBio, Almería, Spain, 1 $\mu \text{g}/\text{mL}$) was added to the wells and incubated in HEPES-Tyrodes buffer (10 mM HEPES, 150 mM NaCl, 12 mM NaHCO_3 , 0.4 mM NaH_2PO_4 , 2.5 mM KCl, 0.1% glucose, 0.1% BSA) with 1 mM MnCl_2 for 1 h at room temperature. After unbound $\alpha \text{IIb}\beta 3$ was removed by rinsing the wells with binding buffer, bound $\alpha \text{IIb}\beta 3$ was measured using anti-integrin $\beta 3$ mAb (AV-10) followed by HRP (horse radish peroxidase)-conjugated goat anti-mouse IgG and peroxidase substrates.

2.4. Activation of Soluble $\alpha \text{IIb}\beta 3$ by sCD40L

ELISA-type activation assays were performed as described previously [15]. Briefly, wells of 96-well Immulon 2 microtiter plates were coated with 100 μL PBS containing $\gamma \text{C}390\text{--}411$ for 2 h at 37 $^{\circ} \text{C}$. Remaining protein-binding sites were blocked by incubating with PBS/0.1% BSA for 30 min at room temperature. After washing with PBS, soluble recombinant $\alpha \text{IIb}\beta 3$ (AgroBio, 1 $\mu \text{g}/\text{mL}$) was pre-incubated with sCD40L (WT or mutant)

for 10 min at room temperature and was added to the wells and incubated in HEPES-Tyrodes buffer with 1 mM CaCl₂ for 1 h at room temperature. After unbound α IIb β 3 was removed by rinsing the wells with binding buffer, bound α IIb β 3 was measured using anti-integrin β 3 mAb (AV-10) followed by HRP-conjugated goat anti-mouse IgG and peroxidase substrates.

2.5. Activation of Cell-Surface α IIb β 3 on Chinese Hamster Ovary (CHO) Cells

α IIb β 3-CHO cells [21] were cultured in Dulbecco's modified Eagle's medium (DMEM)/10% fetal calf serum. The cells were resuspended with HEPES-Tyrodes buffer/0.02% BSA (heat treated at 80 °C for 20 min to remove contaminating cell adhesion molecules). The α IIb β 3-CHO cells were then incubated with WT or mutant sCD40L for 30 min on ice and then incubated with FITC-labeled integrin α IIb β 3 ligand (γ C390–411) for 30 min at room temperature. The cells were washed with PBS/0.02% BSA and analyzed by BD Accuri flow cytometer (Becton Dickinson, Mountain View, CA, USA). The data were analyzed using FlowJo 7.6.5.

2.6. Docking Simulation

Docking simulation of interaction between CD40L (1ALY.pdb) and integrin α IIb β 3 was performed using AutoDock3, as described [22]. In the current study, we used the headpiece (residues 1–438 of α IIb and residues 55–432 of β 3) of α IIb β 3 (5HDB.pdb). Cations were not present in α IIb β 3 during docking simulation [23,24]. The classical ligand-binding site (site 1) or the allosteric site (site 2) of α IIb β 3 were selected as the targets for CD40L.

2.7. Statistical Analysis

Treatment differences were tested using ANOVA and a Tukey multiple comparison test to control the global type I error using Prism 7 (GraphPad Software).

3. Results

3.1. sCD40L Binds to Soluble α IIb β 3 in a KGD-Independent Manner

Previous studies showed that CD40 binding to α IIb β 3 required the KGD motif (residues 215–217) of CD40L [4]. But the specifics of this interaction are unclear. The KGD motif is located at the N-terminus of soluble CD40L (sCD40L) (Figure 1a). To localize the α IIb β 3 binding site in CD40L, we performed docking simulation of interaction between the classical ligand-binding site (site 1) of α IIb β 3 (5hdb.pdb) and monomeric CD40L (1ALY.pdb) using Autodock3. The simulation predicted that α IIb β 3 binds to the outer surface of CD40L (docking energy -24.3 kcal/mol) and that the KGD motif is not in the integrin-binding interface of CD40L (Figure 1b). This is not consistent with the previous report that CD40L bound to α IIb β 3 in the KGD-motif-dependent manner [4].

We generated CD40L in which the KGD motif was deleted (residues 118–261) to study the role of the KGD motif (Supplemental Figure S1a). We studied whether the sCD40L (residues 118–261) binds to soluble α IIb β 3 in ELISA-type binding assays. Wells of a 96-well microtiter plate were coated with sCD40L, and remaining protein-binding sites were blocked with BSA. Wells were incubated with soluble α IIb β 3 for 1 h at room temperature in 1 mM Mn²⁺ (to fully activate integrin) and bound α IIb β 3 was quantified using anti- β 3 and HRP-conjugated anti-mouse IgG. α IIb β 3 bound to the soluble CD40L (residues 118–261) in a dose-dependent manner (Figure 1c). This indicates that α IIb β 3 binding to CD40L does not require the KGD motif. A known ligand to α IIb β 3 (the disintegrin domain of ADAM15) [25,26] suppressed sCD40L binding to α IIb β 3 (Figure 1d), indicating that the binding of sCD40L to α IIb β 3 is specific.

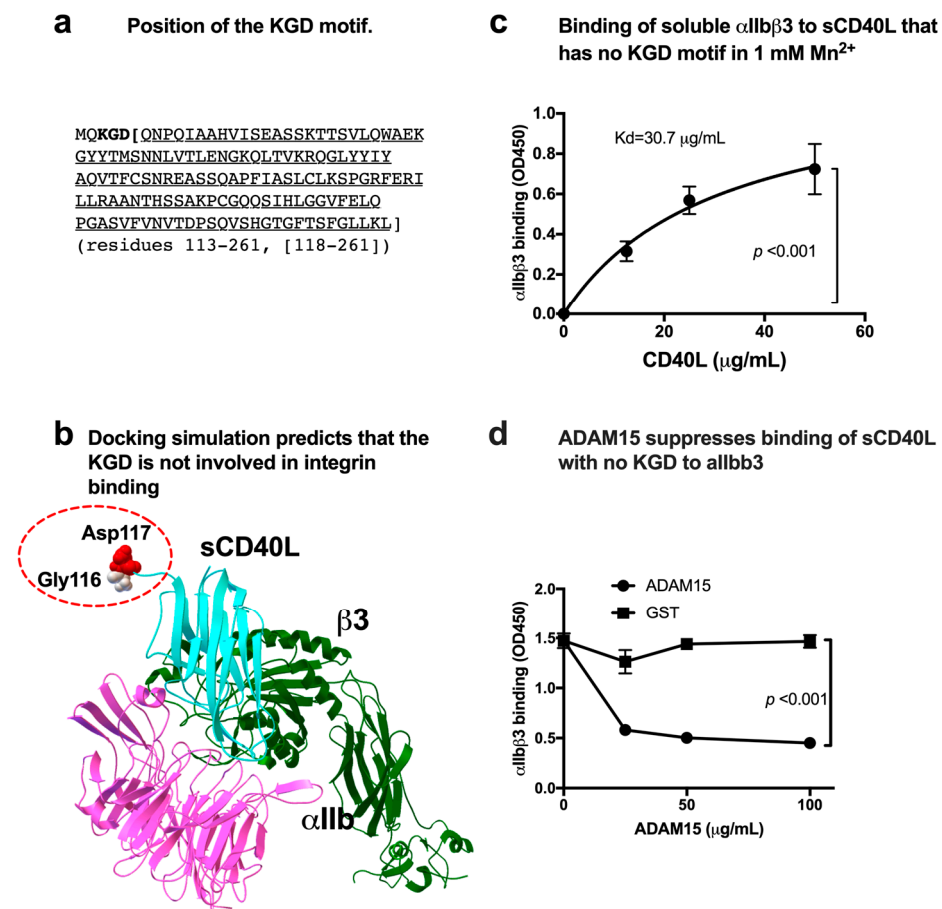


Figure 1. Binding of soluble CD40L to α IIb β 3 does not require the KGD motif. (a) Amino acid sequence of sCD40L (118–261) that lacks the KGD motif. We synthesized CD40L (118–261) to test whether the KGD motif is critical for α IIb β 3 binding. (b) Docking simulation of sCD40L and integrin α IIb β 3. Docking simulation was performed as described in the method section. Monomeric sCD40L was predicted to bind to the classical ligand-binding site of α IIb β 3 with docking energy -24.3 kcal/mol. (c) Binding of soluble integrin α IIb β 3 to sCD40L that lacks the KGD motif in 1 mM Mn^{2+} . Wells of 96-well microtiter plate were coated with sCD40L. Remaining protein-binding sites were blocked with BSA. Wells were incubated with soluble α IIb β 3 (1 μ g/mL) in Tyrode-HEPES buffer with 1 mM Mn^{2+} (to activate α IIb β 3) for 1 h at room temperature. After removing the unbound integrin by washing, bound α IIb β 3 was quantified using anti- β 3 mAb (AV10) and HRP-conjugated anti-mouse IgG. Data are shown as means \pm SD in triplicate experiments. (d) The disintegrin domain of ADAM15 (A disintegrin and metalloprotease-15) inhibits the binding of α IIb β 3 to immobilized sCD40L (118–261). The binding of soluble α IIb β 3 to CD40L was performed as described in (c), except that the disintegrin domain of ADAM15 fused to GST or parent GST was added. Data are shown as means \pm SD in triplicate experiments.

3.2. Effect of a Group of HIGM1 Mutations on the Binding of sCD40L to the Classical Ligand-Binding Site (Site 1) of α IIb β 3

Positions of amino acid residues predicted to be involved in integrin binding are shown in Figure 2a,b. Notably, several amino acid residues mutated in HIGM1 (Ser128, Glu129, Lys143, and Leu155) are clustered in the integrin-binding interface (Figure 2a,b). We incubated soluble α IIb β 3 with CD40L HIGM1 mutants (S128E/E129R, K143T, and L155P) immobilized to wells for 1 hr at room temperature in 1 mM Mn^{2+} . Bound soluble α IIb β 3 was quantified with anti- β 3 mAb and HRP-conjugated anti-mouse IgG. Notably, we found that S128E/E129R and L155P mutations (but not K143T) suppressed the binding of soluble α IIb β 3 in 1 mM Mn^{2+} (Figure 2c). These findings suggest that amino acid residues of CD40L that are critical for site 1 binding are located outside of CD40L (Figure 2e), and

that S128E/E129R and L155P HIGM1 mutants are defective in α IIb β 3 binding. A previous study reported that the binding site for α v β 3 and α 5 β 1 in CD40L was mapped in the trimeric interface of CD40L (e.g., Y170, G226, and H224) [11]. This indicates that the α v β 3 and α IIb β 3 (site 1) bind to CD40L in distinct manners (Figure 2e) and is consistent with the prediction that the KGD motif is not involved in CD40L- α IIb β 3 interaction.

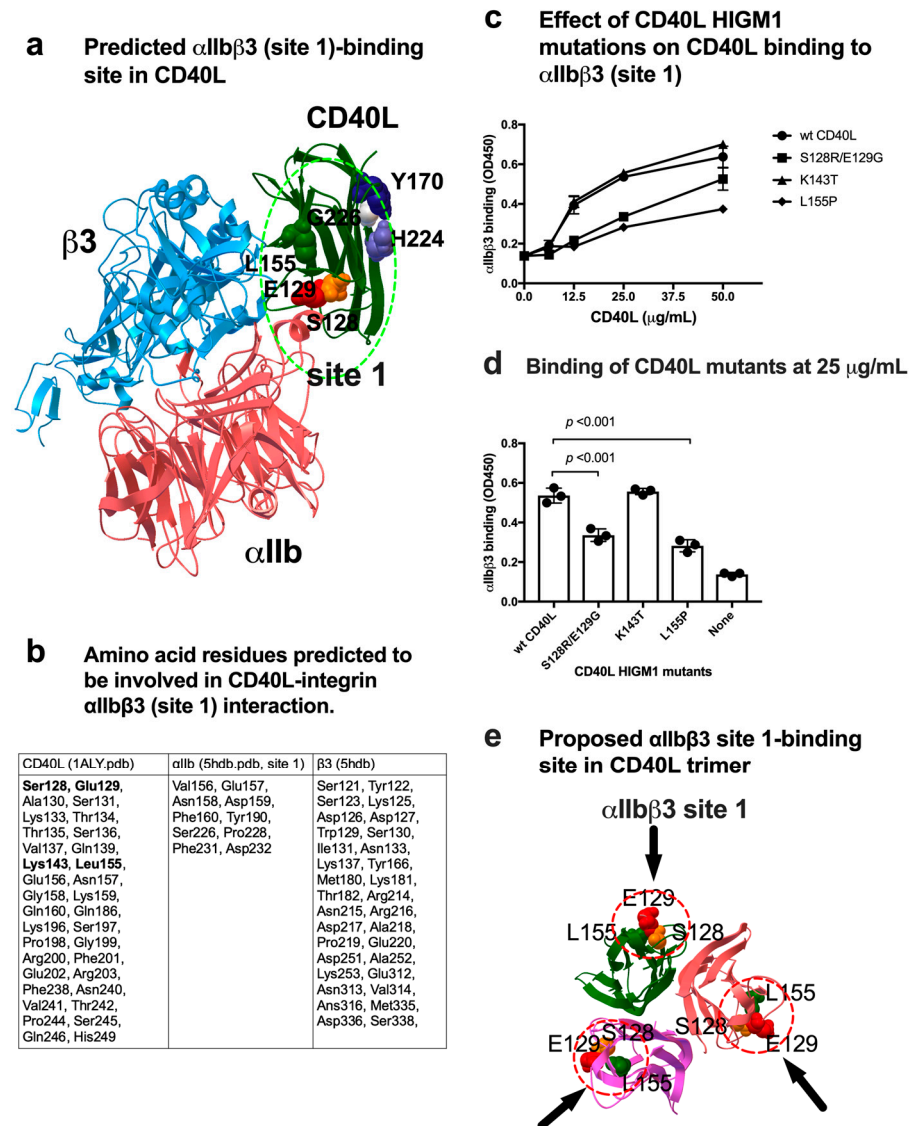


Figure 2. Effect of HIGM1 mutations on α IIb β 3 binding to CD40L. (a) Positions of HIGM1 mutations (S128, E129, and L155) in the predicted binding interface between α IIb β 3 (site 1) and CD40L monomer. Y170 and H224 are in the trimeric interface and are involved in binding to α v β 3 site 1 [11]. (b) Amino acid residues of CD40L that are predicted to be involved in binding to α IIb β 3. Amino acid residues within 0.6 nm between CD40L and α IIb β 3 in the docking model were selected using pdb viewer (version 4.1). Amino acid residues in CD40L that are mutated in HIGM1 are shown in bold. (c) Effect of the HIGM1 mutations on the binding of soluble α IIb β 3 to sCD40L. We selected several HIGM1 mutations in the site 1 binding interface of CD40L predicted by the simulation. The binding of CD40L mutants to soluble α IIb β 3 was studied as described in Figure 1. The binding of CD40L mutants at 25 μ g/mL coating condition is shown. Data are shown as means \pm SD in triplicate experiments. (d) Positions of S128, E129, and L155P that affect the binding of CD40L trimer to soluble α IIb β 3. (e) Proposed α IIb β 3 site 1-binding site in CD40L trimer.

3.3. CD40L Activates Soluble and Cell-Surface Integrin α IIb β 3 in 1 mM Ca^{2+} by Binding to Site 2

Previous studies showed that chemokines CX3CL1, CXCL12, and CCL5 activated integrin α v β 3 and α IIb β 3 in 1 mM Ca^{2+} , in which integrins are usually kept inactive, by binding to the allosteric site (site 2) [14–16]. The presence of site 2 was predicted to be on the opposite side of the classical ligand (RGD)-binding site (site 1) by docking simulation using the closed headpiece α v β 3 (1JV2.pdb) as a target. We previously showed that CD40L can activate integrins α v β 3, α 5 β 1, and α 4 β 1 by binding to site 2 [18]. CD40L is rapidly transported to the surface of platelets upon platelet activation, but it is unclear if α IIb β 3 can be activated by CD40L by binding to site 2.

We used ELISA-type activation assays, in which soluble integrins were incubated with ligands that were immobilized to plastic in the presence of chemokines or other activators in 1 mM Ca^{2+} (to keep integrins inactive) in cell-free conditions [14]. Integrin activation was defined as the increase in the binding of soluble integrins to immobilized ligand. α IIb β 3 is known to bind to the C-terminal 400HHLGGAKQAGDV411 sequence of fibrinogen γ -chain C-terminal domain (γ C) [27]. We used the C-terminal residues 390–411 of fibrinogen γ -chain fused to GST (designated γ C390–411) [16] as a ligand for α IIb β 3. To study if CD40L activates α IIb β 3, soluble α IIb β 3 was pre-incubated with sCD40L for 10 min at room temperature and then incubated with immobilized γ C390–411 for 1 hr at room temperature in the presence of sCD40L in 1 mM Ca^{2+} . Bound soluble α IIb β 3 was quantified with anti- β 3 mAb and HRP-conjugated anti-mouse IgG. Activation was defined as the increase in the binding of soluble α IIb β 3 to immobilized γ C390–411. Notably, WT sCD40L activated α IIb β 3 in a dose-dependent manner in 1 mM Ca^{2+} (Figure 3a), indicating that sCD40L activates α IIb β 3 in cell-free conditions without inside-out signaling.

Binding of integrins to their ligands requires the presence of divalent cations, and different cations can strikingly alter integrin affinities to fibronectin [28]; Mn^{2+} produces the most striking increase in integrin ligand affinity compared with other divalent cations (Mg^{2+} or Ca^{2+}) in a wide variety of integrins. Consequently, Mn^{2+} has been widely used as a positive control for integrin activation. Mn^{2+} -recreated integrin activation was thought to mimic physiologic integrin activation because it activates integrins in the absence of a bound ligand and induces similar epitope exposure [29]. We used 1 mM Mn^{2+} as a standard for integrin activation. We found that sCD40L activated α IIb β 3 at a level comparable to that of 1 mM Mn^{2+} (Figure 3b), indicating that CD40L is a potent integrin α IIb β 3 activator. Although high concentrations of sCD40L were required to detect activation of soluble α IIb β 3, CD40L is a transmembrane protein and is expected to be highly concentrated on the surface.

CHO cells that express recombinant α IIb β 3 on the cell surface (α IIb β 3-CHO cells) were incubated with FITC-labeled γ C390–411 in the presence of sCD40L and the binding of FITC-labeled γ C390–411 was assayed in 1 mM Ca^{2+} in flow cytometry (Figure 3c). We found that sCD40L activated cell-surface α IIb β 3 (Figure 3d) in a dose-dependent manner.

3.4. A Group of HIGM1 Mutations Suppress Activation of α IIb β 3 by CD40L

We performed docking simulation of interaction between monomeric CD40L and α IIb β 3 (site 2) by targeting the allosteric ligand-binding site (site 2) of α IIb β 3 (Figure 4a). Amino acid residues involved in CD40L- α IIb β 3 site 2 interaction are shown in Figure 4b. The predicted site 2 binding site in CD40L is outside of the trimer and overlaps with that of the site 1 binding site (Figure 2a). Notably, 4 HIGM1 mutations are clustered in the site 2 binding interface (S128E/E129R, K143T, G144E, and L155P) of CD40L. We found that K143T and G144E were defective in activating soluble α IIb β 3 (Figure 4c,d), which is consistent with the prediction. The positions of K143T and G144E on CD40L are shown (Figure 4e), indicating that the site 1 and site 2 binding sites overlap on the outer surface of trimeric CD40L and two HIGM1 mutants were defective in activating α IIb β 3.

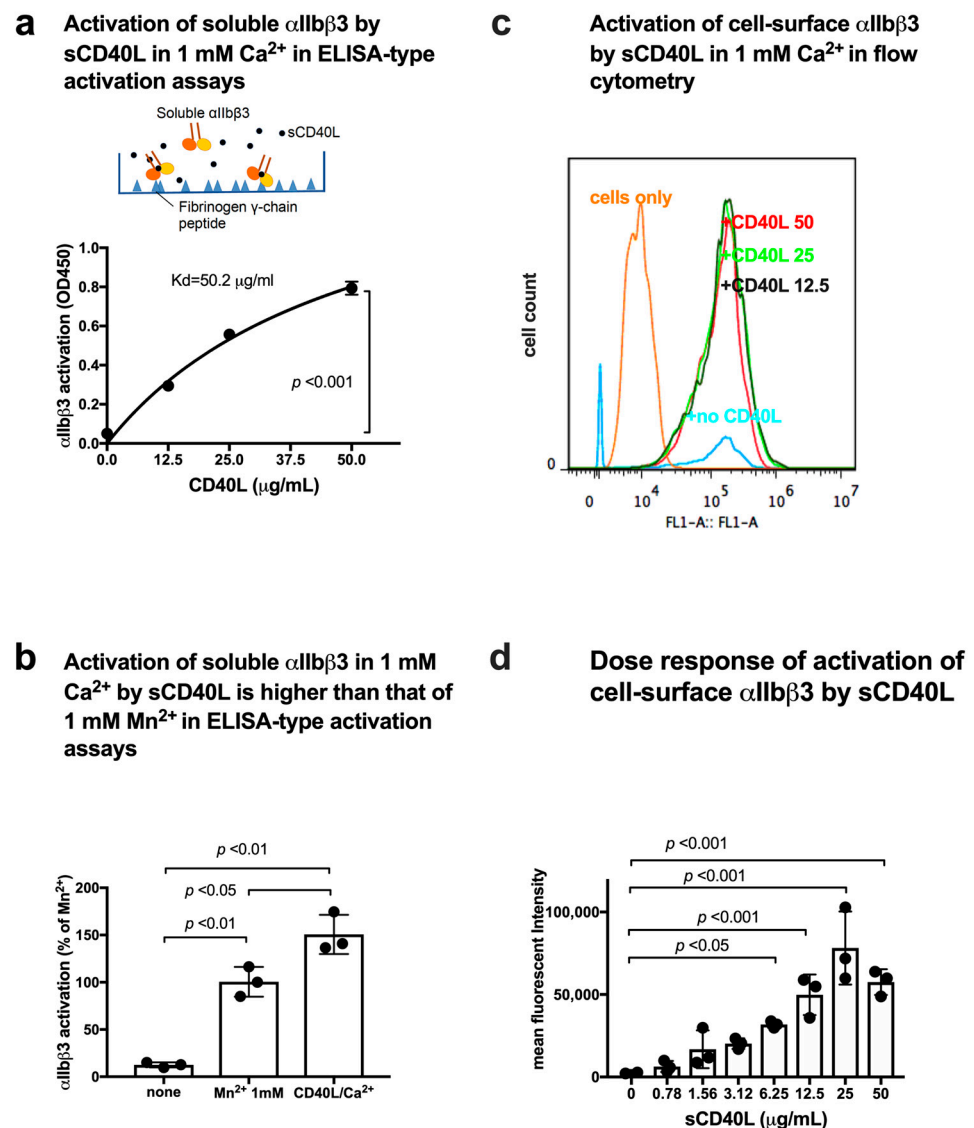
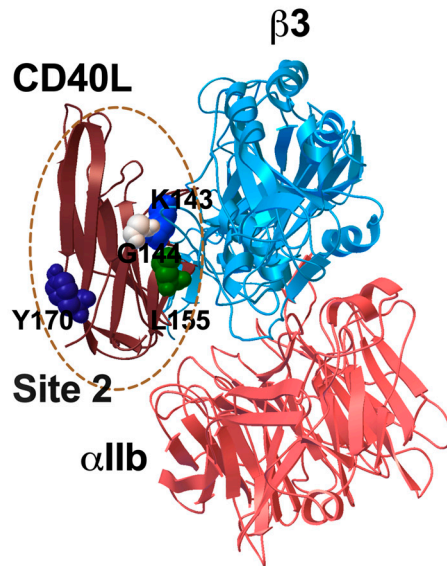


Figure 3. sCD40L activates soluble α IIb β 3 in 1 mM Ca^{2+} . (a) CD40L activated α IIb β 3 in a dose-dependent manner. Wells of 96-well microtiter plate were coated with γ C390–411 (the α IIb β 3-ligand peptide conjugated to GST) (20 $\mu\text{g}/\text{mL}$) and the remaining protein-binding sites were blocked with BSA. Wells were incubated with soluble α IIb β 3 (1 $\mu\text{g}/\text{mL}$) in Tyrode-HEPES buffer with 1 mM Ca^{2+} (to keep α IIb β 3 inactive) for 1 h at room temperature in the presence of 50 $\mu\text{g}/\text{mL}$ CD40L. After washing the unbound integrin, bound α IIb β 3 was quantified using anti- β 3 mAb (AV10) and HRP-conjugated anti-mouse IgG. Data are shown as mean \pm SD in triplicate experiments. (b) Activation of soluble α IIb β 3 in 1 mM Ca^{2+} by sCD40L was higher than that of 1 mM Mn^{2+} in ELISA-type activation assays. Activation of soluble α IIb β 3 by sCD40L was determined as described in (a). 1 mM Mn^{2+} was used instead of 1 mM Ca^{2+} as a positive control (as 100%). Data are shown as mean \pm SD in triplicate experiments. (c) Activation of cell-surface α IIb β 3 on CHO cells in flow cytometry. α IIb β 3-CHO cells were incubated with WT sCD40L (0–50 $\mu\text{g}/\text{mL}$) for 30 min on ice and then incubated with FITC-labeled integrin α IIb β 3 ligand γ C390–411 for 30 min at room temperature. The cells were washed with PBS/0.02% BSA and analyzed by BD Accuri flow cytometer (Becton Dickinson, Mountain View, CA, USA). The data were analyzed using FlowJo 7.6.5. (d) Dose response of activation of cell-surface α IIb β 3 by soluble CD40L. Activation of cell-surface α IIb β 3 was determined as described in (c). Data are shown as mean \pm SD in triplicate experiments. Values with no sCD40L were subtracted.

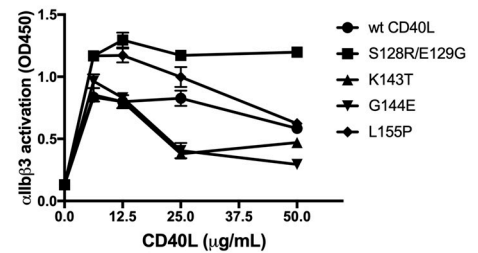
a Docking simulation of CD40L binding to site 2 of α I**l** β 3



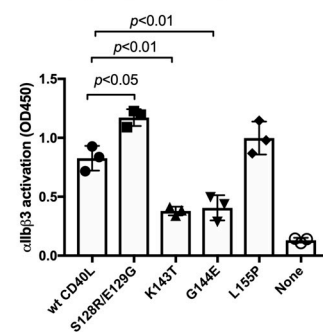
b Amino acid residues predicted to be involved in CD40L-integrin α I**l** β 3 (site 2) interaction.

CD40L (1ALY.pdb)	α I l β 3 (5hdb.pdb, site 2)	β 3 (5hdb)
Pro120, Ile127, Ser128 , Glu129 , Ala130, Ser131, Ser132, Lys133, Thr134, Gln139, Trp140, Ala141, Glu142, Lys143 , Gly144 , Tyr145, Tyr146, Asn151, Leu152, Thr154, Leu155 , Glu156, Asn157, Gly158, Lys159, Val163, Lys164, Arg165, Pro233, Gly234, His249, Gly250	Asn15, Arg386, Pro412, Thr413, Gly414, Ser415, Gly441, Ala442	Val231, Cys232, Asp233, Glu234, Lys235, Trp238, Arg239, Asn240, Asp241, His244, <u>Gln267</u> , <u>Asp270</u> , <u>Gly271</u> , <u>Gln272</u> , <u>Cys273</u> , <u>His274</u> , <u>Val275</u> , <u>Gly276</u> , <u>Ser277</u> , <u>Ser282</u> , Glu297, Lys298, Ser300, Gln301, Lys302, Asn303, Arg360, Lys417, Phe421, Lys422

c Effect of HIGM1 CD40L mutations on activation of α I**l** β 3 by CD40L in ELISA-type activation assays



d Effect of HIGM1 mutations (at 25 µg/ml) on integrin activation



e Proposed α I**l** β 3 site 2-binding site in CD40L trimer

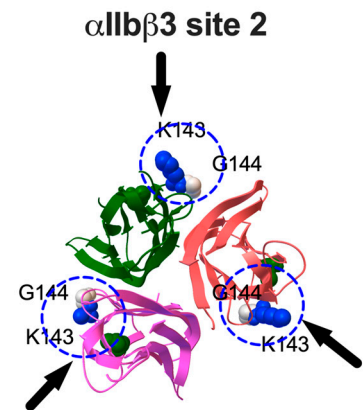


Figure 4. Effect of HIGM1 mutations on activation of α I**l** β 3 by CD40L. (a) Positions of HIGM1 mutations in the CD40L-integrin α I**l** β 3 (site 2) complex model. Y170 is in the trimeric interface of CD40L trimer. (b) Amino acid residues within 0.6 nm between CD40L and α I**l** β 3 were selected using pdb viewer (version 4.1). Amino acid residues in CD40L that are mutated in HIGM1 are shown in bold. Amino acid residues in integrin β 3 that are involved in site 2 peptide are underlined. (c) Effect of HIGM1 mutations on the CD40L-mediated activation of α I**l** β 3. We studied the effect of several HIGM1 mutations on the CD40L-mediated activation of soluble α I**l** β 3. Wells were coated with γ C390–411 and incubated with soluble α I**l** β 3 (1 μ g/mL) in Tyrode-HEPES buffer with 1 mM Ca^{2+} (to keep α I**l** β 3 inactive) for 1 h at room temperature in the presence of CD40L (wt and HIGM1 mutants). After washing the unbound integrin, bound α I**l** β 3 was quantified using anti- β 3 mAb (AV10) and HRP-conjugated anti-mouse IgG. Data are shown as mean \pm SD in triplicate experiments. (d) Effect of HIGM1 mutations (at 25 μ g/mL) on integrin activation. (e) The HIGM1 mutations that affected activation of soluble α I**l** β 3 are located outside of CD40L trimer.

4. Discussion

Previously, our studies showed that direct binding of CD40L to integrin $\alpha\nu\beta3$ plays a critical role in CD40L signaling [11]. We showed that site 1 of $\alpha\nu\beta3$ bound to the trimeric interface of CD40L. It has been reported that some of the altered residues in HIGM1 are located in the protein core or trimeric interface (Y170C, Q174R, T176I, A208D, H224Y, G226A, G227V, and L258S), pointing to the roles of these HIGM1 mutations in structural instability or disruptive trimer formation leading to defective CD40L functions [30]. We showed that, in fact, these HIGM1 mutants are defective in integrin binding, and we proposed that the defective integrin binding caused functional defects in these HIGM1 mutations in immune responses as evidenced by reduced activation in B cells when stimulated with HIGM1 mutants compared to normal CD40L [11]. Also, our studies suggest that the $\alpha\text{IIb}\beta3$ (site 1)-binding site is distinct from the site 1 binding site of $\alpha\nu\beta3$ [11].

CD40L was shown to be involved in thrombus stabilization in addition to immune responses. $\alpha\text{IIb}\beta3$ is a key component of platelet aggregation and thrombus formation and binds to CD40L. Previous studies showed that $\alpha\text{IIb}\beta3$ binding to CD40L requires the KGD motif of CD40L (residues 115–117 of CD40L) [4,17]. We showed, however, that CD40L without KGD motif did bind to this $\alpha\text{IIb}\beta3$ integrin. This was supported by the docking model, which showed that the KGD motif is not close to the $\alpha\text{IIb}\beta3$ -binding interface of CD40L. Instead, the $\alpha\text{IIb}\beta3$ (site 1)-binding site in CD40L is located outside of the CD40L trimer. Notably, a group of HIGM1 mutations (S128E/E129R and L155P), which was confirmed by mutagenesis studies with an *in vitro* binding assay, are clustered in the $\alpha\text{IIb}\beta3$ site 1 binding site, and suppressed CD40L binding to $\alpha\text{IIb}\beta3$ site 1. We previously reported that sCD40L bound to site 2 of $\alpha\nu\beta3$ and activated soluble $\alpha\nu\beta3$ [18]. Interestingly, the site 2 binding site in CD40L was in the region around the HIGM1 mutants S128R/E129G, K143T, G144E, and L155P, indicating that the $\alpha\nu\beta3$ site 2 binding site, and the $\alpha\text{IIb}\beta3$ site 1 and site 2 binding sites, overlap in CD40L. Notably, these mutants were defective in activating $\alpha\nu\beta3$ and acted as antagonists for CD40L-mediated $\alpha\nu\beta3$ activation [18]. These findings excluded the possibility that these CD40L mutants are not properly folded. Also, it would be interesting to study whether these CD40L mutants act as antagonists in CD40L-mediated $\alpha\text{IIb}\beta3$ activation and thrombus formation in future research.

Notably, in the present study, we showed that CD40L activated soluble integrin $\alpha\text{IIb}\beta3$ in cell-free conditions in 1 mM Ca^{2+} . This was clear evidence that CD40L can activate $\alpha\text{IIb}\beta3$ by binding to site 2 without inside-out signaling. CD40L also activated cell-surface $\alpha\text{IIb}\beta3$ on CHO cells that did not have machinery for inside-out signaling. The $\alpha\text{IIb}\beta3$ -binding site in CD40L is located outside of the CD40L trimer around two HIGM1 mutations, K143T and G144E, which is close to the recently mapped $\alpha\nu\beta3$ (site 2)-binding site [18]. Interestingly, this site overlaps with the $\alpha\text{IIb}\beta3$ (site 1)-binding site.

In sum, the present study suggests that the $\alpha\text{IIb}\beta3$ site 1 and site 2 binding sites, and the $\alpha\nu\beta3$ site 2 binding site in CD40L in the vicinity of the HIGM1 mutants S128R/E129G, K143T, G144E, and L155P, are critically involved in CD40L signaling functions. We propose that the defective immune responses in HIGM1 may be due to the defective integrin binding/activation, since HIGM1 mutations are clustered in the integrin-binding sites in $\alpha\text{IIb}\beta3$, $\alpha\nu\beta3$, and $\alpha5\beta1$, as shown by our present and previous studies (Figure 5).

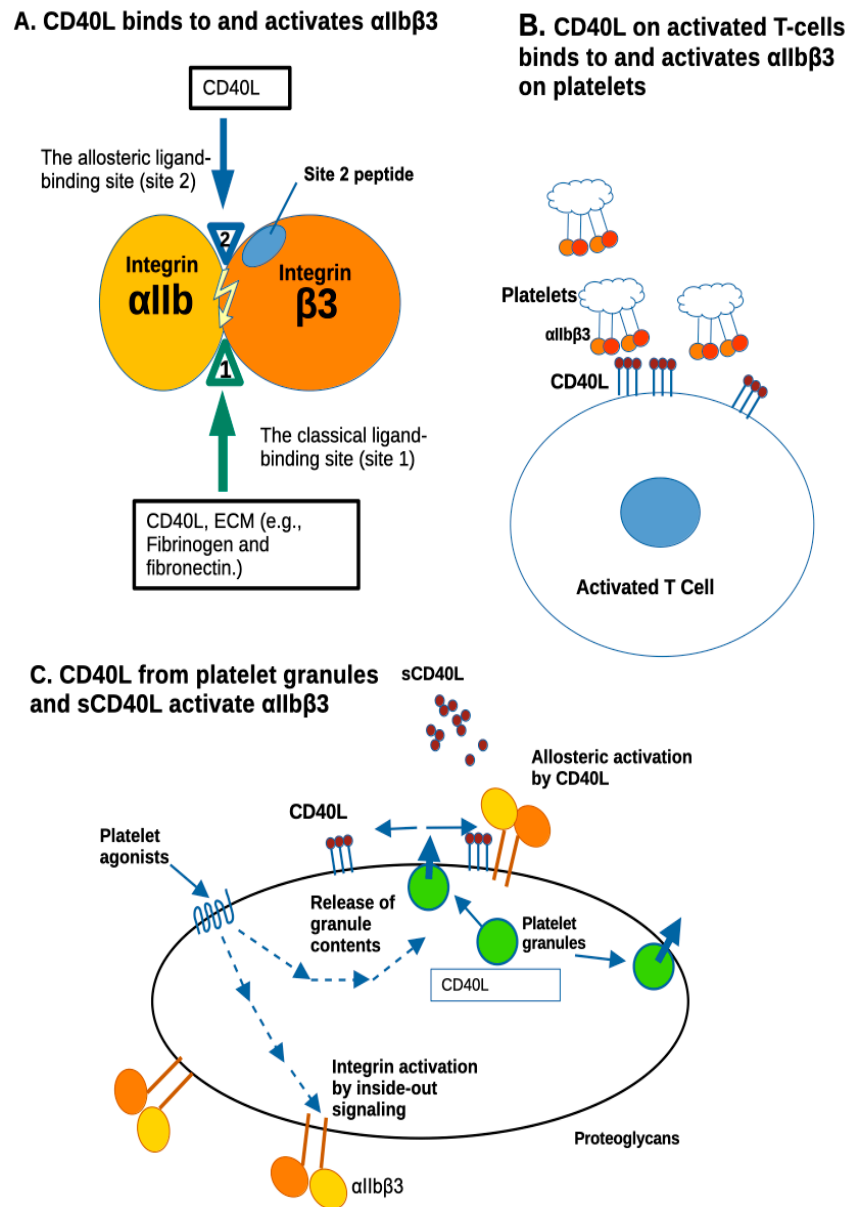


Figure 5. Allosteric activation of $\alpha\text{IIb}\beta 3$ by CD40L. (A) The present study showed that CD40L binding to $\alpha\text{IIb}\beta 3$ (site 1) does not require the KGD motif (at the N-terminus of sCD40L). CD40L also binds to site 2 and activates $\alpha\text{IIb}\beta 3$. CD40L binding interfaces to $\alpha\text{IIb}\beta 3$ (site 1 and site 2) are exposed to the surface of trimeric CD40L and overlap. Notably, HIGM1 mutations are clustered in this area. Peptide from site 2 was shown to bind to sCD40L [18], indicating that CD40L binds to site 2. CD40L binding interface to site 1 of integrins $\alpha\text{v}\beta 3$ and $\alpha 5\beta 1$ are in the trimeric interface (cryptic in CD40L trimer), indicating that CD40L monomer can bind to these integrins, but not trimer [11]. CD40L binds to site 2 and activates $\alpha\text{v}\beta 3$ and $\alpha 5\beta 1$. The binding interface to site 2 of $\alpha\text{v}\beta 3$ and $\alpha 5\beta 1$ overlap with that of $\alpha\text{IIb}\beta 3$ [18]. Notably, HIGM1 mutations are clustered in the site 1 and site 2 binding interfaces, indicating that immune deficiency of HIGM1 may be due to the defect in integrin binding and/or activation. (B) The present study predicted that CD40L on activated T cell surfaces can bind to mediate T-cell–platelet interaction by binding to and activating platelet integrin $\alpha\text{IIb}\beta 3$. Although $\alpha\text{IIb}\beta 3$ on inactive platelets is not activated, CD40L on activated T cells may activate it by binding to site 2 of $\alpha\text{IIb}\beta 3$. (C) It is known that CD40L is stored in platelet granules and rapidly transported to the platelet surface upon platelet activation. CD40L is predicted to activate $\alpha\text{IIb}\beta 3$ by binding to site 2 of $\alpha\text{IIb}\beta 3$ on the surface upon platelet activation. Soluble CD40L levels increase during inflammation and may induce allosteric activation of $\alpha\text{IIb}\beta 3$ and platelet aggregation.

It has been established that, in general, integrin $\alpha\text{IIb}\beta\text{3}$ is activated by inside-out signaling upon the activation of platelets by thrombin and other platelet agonists. However, it has been proposed that inside-out signaling does not enhance ligand binding affinity to monovalent ligand [31,32]. Inside-out signaling induces integrin binding to multivalent ligands (e.g., ligand-mimetic antibody, Pac-1 IgM specific for $\alpha\text{IIb}\beta\text{3}$, with potential 10-binding sites) [33,34]. Since the site 2-mediated integrin activation enhanced binding affinity to monovalent ligands, we propose that site 2-mediated integrin activation may be a potential mechanism for enhancing the ligand affinity of integrins. We have recently shown that several pro-inflammatory factors, CX3CL1, CCL5, and CXCL12, activate soluble integrins $\alpha\text{IIb}\beta\text{3}$ and $\alpha\text{v}\beta\text{3}$ by binding to the allosteric site (site 2) [16], and CD40L in the present study. We recently showed that CD62P (P-selectin) binds to $\alpha\text{IIb}\beta\text{3}$ and $\alpha\text{v}\beta\text{3}$ and activates them by binding to site 2 [19]. In this context, it would be interesting to study whether other cytokines and chemokines stored in platelet granules may also interact with $\alpha\text{IIb}\beta\text{3}$ and concomitantly amplify and stabilize the process of platelet activation. Such a mechanism might facilitate platelets having a multifaceted role as the amplifier of the immune responses through hemostasis and inflammatory cytokine release for recruiting immune cells such as monocytes and T cells.

CD40L is expressed on activated T cells (Figure 5). Our current findings have implications for the immune modification by platelets through direct interaction between platelets and immune cells. For example, activated platelets trigger secretion of cytokines and chemokines from monocytes and neutrophils through forming heterotypic cell aggregates [35]. Also, platelets are known to form complexes with lymphocytes [36] and fine tune their differentiation, including CD4 T helper and Treg differentiation [37–40], anti-viral cytotoxic CD8 T cell response [41], and B cell antibody production [42,43]. Importantly, platelets display major histocompatibility complex (MHC) and directly stimulate T cells [44]. Such platelet and lymphocyte interactions are thought to play an essential role in cardiovascular diseases [45] and COVID-19 [46,47]. However, the detailed mechanisms by which platelets form stable complexes with lymphocytes are not entirely clear. Notably, activated monocytes and B cells both express CD40, the receptor of CD40L. Therefore, based on our previous and current findings, and given the fact that CD40L can bind and activate other integrins such as $\alpha\text{5}\beta\text{1}$ and $\alpha\text{4}\beta\text{1}$ on lymphocytes or $\alpha\text{v}\beta\text{3}$ on vascular endothelial cells, we hypothesize that direct CD40L and integrin interaction plays a critical role in the interface of platelet and immune cell complexes for immune cell activation. Further studies will be needed to fully understand the specifics of CD40L-integrins interaction and immune cell activation in inflammatory settings, which can form the basis to develop new therapeutics for inflammatory diseases.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/cells12151977/s1>, Figure S1: SDS-PAGE of CD40L preparations.

Author Contributions: Y.K.T. performed the experiments. M.S. discussed results, interpreted the results, and edited the manuscript. Y.T. conceived the project, designed the experiments, analyzed the data, and wrote the article. All authors have read and agreed to the published version of the manuscript.

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Data Availability Statement: Research data is available upon request.

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Conflicts of Interest: The authors declare that they have no conflict of interest with the contents of this article.

References

1. Takada, Y.; Ye, X.; Simon, S. The integrins. *Genome Biol.* **2007**, *8*, 215. [[CrossRef](#)]
2. Hynes, R.O. Integrins: Bidirectional, allosteric signaling machines. *Cell* **2002**, *110*, 673–687. [[CrossRef](#)]
3. Hassan, G.S.; Salti, S.; Mourad, W. Novel Functions of Integrins as Receptors of CD154: Their Role in Inflammation and Apoptosis. *Cells* **2022**, *11*, 1747. [[CrossRef](#)]
4. Andre, P.; Prasad, K.S.; Denis, C.V.; He, M.; Papalia, J.M.; Hynes, R.O.; Phillips, D.R.; Wagner, D.D. CD40L stabilizes arterial thrombi by a beta3 integrin–dependent mechanism. *Nat. Med.* **2002**, *8*, 247–252. [[CrossRef](#)]
5. Andre, P.; Nannizzi-Alaimo, L.; Prasad, S.K.; Phillips, D.R. Platelet-derived CD40L: The switch-hitting player of cardiovascular disease. *Circulation* **2002**, *106*, 896–899. [[CrossRef](#)] [[PubMed](#)]
6. Danese, S.; Fiocchi, C. Platelet activation and the CD40/CD40 ligand pathway: Mechanisms and implications for human disease. *Crit. Rev. Immunol.* **2005**, *25*, 103–121. [[CrossRef](#)] [[PubMed](#)]
7. Leveille, C.; Bouillon, M.; Guo, W.; Bolduc, J.; Sharif-Askari, E.; El-Fakhry, Y.; Reyes-Moreno, C.; Lapointe, R.; Merhi, Y.; Wilkins, J.A.; et al. CD40 ligand binds to alpha5beta1 integrin and triggers cell signaling. *J. Biol. Chem.* **2007**, *282*, 5143–5151. [[CrossRef](#)] [[PubMed](#)]
8. Zirlik, A.; Maier, C.; Gerdes, N.; MacFarlane, L.; Soosairajah, J.; Bavendiek, U.; Ahrens, I.; Ernst, S.; Bassler, N.; Missiou, A.; et al. CD40 ligand mediates inflammation independently of CD40 by interaction with Mac-1. *Circulation* **2007**, *115*, 1571–1580. [[CrossRef](#)]
9. Alturaihi, H.; Hassan, G.S.; Al-Zoobi, L.; Salti, S.; Darif, Y.; Yacoub, D.; El Akoum, S.; Oudghiri, M.; Merhi, Y.; Mourad, W. Interaction of CD154 with different receptors and its role in bidirectional signals. *Eur. J. Immunol.* **2015**, *45*, 592–602. [[CrossRef](#)]
10. El Fakhry, Y.; Alturaihi, H.; Yacoub, D.; Liu, L.; Guo, W.; Leveille, C.; Jung, D.; Khzam, L.B.; Merhi, Y.; Wilkins, J.A.; et al. Functional interaction of CD154 protein with alpha5beta1 integrin is totally independent from its binding to alphaIIb beta3 integrin and CD40 molecules. *J. Biol. Chem.* **2012**, *287*, 18055–18066. [[CrossRef](#)]
11. Takada, Y.K.; Yu, J.; Shimoda, M.; Takada, Y. Integrin Binding to the Trimeric Interface of CD40L Plays a Critical Role in CD40/CD40L Signaling. *J. Immunol.* **2019**, *203*, 1383–1391. [[CrossRef](#)] [[PubMed](#)]
12. Piirila, H.; Valiaho, J.; Vihinen, M. Immunodeficiency mutation databases (IDbases). *Hum. Mutat.* **2006**, *27*, 1200–1208. [[CrossRef](#)]
13. Fujita, M.; Takada, Y.K.; Takada, Y. Integrins alphavbeta3 and alpha4beta1 act as coreceptors for fractalkine, and the integrin-binding defective mutant of fractalkine is an antagonist of CX3CR1. *J. Immunol.* **2012**, *189*, 5809–5819. [[CrossRef](#)] [[PubMed](#)]
14. Fujita, M.; Takada, Y.K.; Takada, Y. The chemokine fractalkine can activate integrins without CX3CR1 through direct binding to a ligand-binding site distinct from the classical RGD-binding site. *PLoS ONE* **2014**, *9*, e96372. [[CrossRef](#)] [[PubMed](#)]
15. Fujita, M.; Davari, P.; Takada, Y.K.; Takada, Y. Stromal cell-derived factor-1 (CXCL12) activates integrins by direct binding to an allosteric ligand-binding site (site 2) of integrins without CXCR4. *Biochem. J.* **2018**, *475*, 723–732. [[CrossRef](#)] [[PubMed](#)]
16. Takada, Y.K.; Fujita, M.; Takada, Y. Pro-Inflammatory Chemokines CCL5, CXCL12, and CX3CL1 Bind to and Activate Platelet Integrin alphaIIb beta3 in an Allosteric Manner. *Cells* **2022**, *11*, 3059. [[CrossRef](#)]
17. Prasad, K.S.; Andre, P.; Yan, Y.; Phillips, D.R. The platelet CD40L/GP IIb-IIIa axis in atherothrombotic disease. *Curr. Opin. Hematol.* **2003**, *10*, 356–361. [[CrossRef](#)]
18. Takada, Y.K.; Shimoda, M.; Maverakis, E.; Felding, B.H.; Cheng, R.H.; Takada, Y. Soluble CD40L activates soluble and cell-surface integrins alphavbeta3, alpha5beta1 and alpha4beta1 by binding to the allosteric ligand-binding site (site 2). *J. Biol. Chem.* **2021**, *296*, 100399. [[CrossRef](#)]
19. Takada, Y.K.; Simon, S.I.; Takada, Y. The C-type lectin domain of CD62P (P-selectin) functions as an integrin ligand. *Life Sci. Alliance* **2023**, *6*, e202201747. [[CrossRef](#)]
20. Yokoyama, K.; Zhang, X.P.; Medved, L.; Takada, Y. Specific binding of integrin alpha v beta 3 to the fibrinogen gamma and alpha E chain C-terminal domains. *Biochemistry* **1999**, *38*, 5872–5877. [[CrossRef](#)]
21. Kamata, T.; Irie, A.; Tokuhira, M.; Takada, Y. Critical residues of integrin alphaIIb subunit for binding of alphaIIb beta3 (glycoprotein IIb-IIIa) to fibrinogen and ligand-mimetic antibodies (PAC-1, OP-G2, and LJ-CP3). *J. Biol. Chem.* **1996**, *271*, 18610–18615. [[CrossRef](#)] [[PubMed](#)]
22. Ieguchi, K.; Fujita, M.; Ma, Z.; Davari, P.; Taniguchi, Y.; Sekiguchi, K.; Wang, B.; Takada, Y.K.; Takada, Y. Direct binding of the EGF-like domain of neuregulin-1 to integrins ($\alpha v \beta 3$ and $\alpha 6 \beta 4$) is involved in neuregulin-1/ErbB signaling. *J. Biol. Chem.* **2010**, *285*, 31388–31398. [[CrossRef](#)]
23. Saegusa, J.; Akakura, N.; Wu, C.Y.; Hoogland, C.; Ma, Z.; Lam, K.S.; Liu, F.T.; Takada, Y.K.; Takada, Y. Pro-inflammatory secretory phospholipase A2 type IIA binds to integrins alphavbeta3 and alpha4beta1 and induces proliferation of monocytic cells in an integrin-dependent manner. *J. Biol. Chem.* **2008**, *283*, 26107–26115. [[CrossRef](#)] [[PubMed](#)]
24. Mori, S.; Wu, C.Y.; Yamaji, S.; Saegusa, J.; Shi, B.; Ma, Z.; Kuwabara, Y.; Lam, K.S.; Isseroff, R.R.; Takada, Y.K.; et al. Direct binding of integrin alphavbeta3 to FGF1 plays a role in FGF1 signaling. *J. Biol. Chem.* **2008**, *283*, 18066–18075. [[CrossRef](#)] [[PubMed](#)]
25. Zhang, X.P.; Kamata, T.; Yokoyama, K.; Puzon-McLaughlin, W.; Takada, Y. Specific interaction of the recombinant disintegrin-like domain of MDC-15 (metargidin, ADAM-15) with integrin alphavbeta3. *J. Biol. Chem.* **1998**, *273*, 7345–7350. [[CrossRef](#)] [[PubMed](#)]
26. Langer, H.; May, A.E.; Bultmann, A.; Gawaz, M. ADAM 15 is an adhesion receptor for platelet GPIIb-IIIa and induces platelet activation. *Thromb. Haemost.* **2005**, *94*, 555–561. [[CrossRef](#)]
27. Farrell, D.H.; Thiagarajan, P. Binding of recombinant fibrinogen mutants to platelets. *J. Biol. Chem.* **1994**, *269*, 226–231. [[CrossRef](#)]

28. Gailit, J.; Ruoslahti, E. Regulation of the fibronectin receptor affinity by divalent cations. *J. Biol. Chem.* **1988**, *263*, 12927–12932. [[CrossRef](#)]
29. Ni, H.; Li, A.; Simonsen, N.; Wilkins, J.A. Integrin activation by dithiothreitol or Mn^{2+} induces a ligand-occupied conformation and exposure of a novel NH₂-terminal regulatory site on the beta1 integrin chain. *J. Biol. Chem.* **1998**, *273*, 7981–7987. [[CrossRef](#)]
30. Karpusas, M.; Hsu, Y.M.; Wang, J.H.; Thompson, J.; Lederman, S.; Chess, L.; Thomas, D. 2 A crystal structure of an extracellular fragment of human CD40 ligand. *Structure* **1995**, *3*, 1031–1039. [[CrossRef](#)]
31. Schurpf, T.; Springer, T.A. Regulation of integrin affinity on cell surfaces. *EMBO J.* **2011**, *30*, 4712–4727. [[CrossRef](#)] [[PubMed](#)]
32. Bunch, T.A. Integrin alphaIIb beta3 activation in Chinese hamster ovary cells and platelets increases clustering rather than affinity. *J. Biol. Chem.* **2010**, *285*, 1841–1849. [[CrossRef](#)]
33. Tadokoro, S.; Shattil, S.J.; Eto, K.; Tai, V.; Liddington, R.C.; de Pereda, J.M.; Ginsberg, M.H.; Calderwood, D.A. Talin binding to integrin beta tails: A final common step in integrin activation. *Science* **2003**, *302*, 103–106. [[CrossRef](#)] [[PubMed](#)]
34. Iwamoto, D.V.; Calderwood, D.A. Regulation of integrin-mediated adhesions. *Curr. Opin. Cell Biol.* **2015**, *36*, 41–47. [[CrossRef](#)]
35. von Bruhl, M.L.; Stark, K.; Steinhart, A.; Chandraratne, S.; Konrad, I.; Lorenz, M.; Khandoga, A.; Tirniceriu, A.; Coletti, R.; Kollnberger, M.; et al. Monocytes, neutrophils, and platelets cooperate to initiate and propagate venous thrombosis in mice in vivo. *J. Exp. Med.* **2012**, *209*, 819–835. [[CrossRef](#)]
36. Li, N. Platelet-lymphocyte cross-talk. *J. Leukoc. Biol.* **2008**, *83*, 1069–1078. [[CrossRef](#)]
37. Elzey, B.D.; Schmidt, N.W.; Crist, S.A.; Kresowik, T.P.; Harty, J.T.; Nieswandt, B.; Ratliff, T.L. Platelet-derived CD154 enables T-cell priming and protection against *Listeria monocytogenes* challenge. *Blood* **2008**, *111*, 3684–3691. [[CrossRef](#)]
38. Min, Y.; Hao, L.; Liu, X.; Tan, S.; Song, H.; Ni, H.; Sheng, Z.; Jooss, N.; Liu, X.; Malmstrom, R.E.; et al. Platelets fine-tune effector responses of naive CD4(+) T cells via platelet factor 4-regulated transforming growth factor beta signaling. *Cell. Mol. Life Sci.* **2022**, *79*, 247. [[CrossRef](#)]
39. Bock, M.; Bergmann, C.B.; Jung, S.; Biberthaler, P.; Heimann, L.; Hanschen, M. Platelets differentially modulate CD4(+) Treg activation via GPIIa/IIIb-, fibrinogen-, and PAR4-dependent pathways. *Immunol. Res.* **2022**, *70*, 185–196. [[CrossRef](#)]
40. Chen, X.; Xu, Y.; Chen, Q.; Zhang, H.; Zeng, Y.; Geng, Y.; Shen, L.; Li, F.; Chen, L.; Chen, G.Q.; et al. The phosphatase PTEN links platelets with immune regulatory functions of mouse T follicular helper cells. *Nat. Commun.* **2022**, *13*, 2762. [[CrossRef](#)] [[PubMed](#)]
41. Iannacone, M.; Sitia, G.; Isogawa, M.; Marchese, P.; Castro, M.G.; Lowenstein, P.R.; Chisari, F.V.; Ruggeri, Z.M.; Guidotti, L.G. Platelets mediate cytotoxic T lymphocyte-induced liver damage. *Nat. Med.* **2005**, *11*, 1167–1169. [[CrossRef](#)] [[PubMed](#)]
42. Elzey, B.D.; Tian, J.; Jensen, R.J.; Swanson, A.K.; Lees, J.R.; Lentz, S.R.; Stein, C.S.; Nieswandt, B.; Wang, Y.; Davidson, B.L.; et al. Platelet-mediated modulation of adaptive immunity. A communication link between innate and adaptive immune compartments. *Immunity* **2003**, *19*, 9–19. [[CrossRef](#)] [[PubMed](#)]
43. Cognasse, F.; Hamzeh-Cognasse, H.; Lafarge, S.; Chavarin, P.; Cogne, M.; Richard, Y.; Garraud, O. Human platelets can activate peripheral blood B cells and increase production of immunoglobulins. *Exp. Hematol.* **2007**, *35*, 1376–1387. [[CrossRef](#)]
44. Zufferey, A.; Speck, E.R.; Machlus, K.R.; Aslam, R.; Guo, L.; McVey, M.J.; Kim, M.; Kapur, R.; Boilard, E.; Italiano, J.E., Jr.; et al. Mature murine megakaryocytes present antigen-MHC class I molecules to T cells and transfer them to platelets. *Blood Adv.* **2017**, *1*, 1773–1785. [[CrossRef](#)]
45. Stark, K.; Massberg, S. Interplay between inflammation and thrombosis in cardiovascular pathology. *Nat. Rev. Cardiol.* **2021**, *18*, 666–682. [[CrossRef](#)]
46. Hottz, E.D.; Azevedo-Quintanilha, I.G.; Palhinha, L.; Teixeira, L.; Barreto, E.A.; Pao, C.R.R.; Righy, C.; Franco, S.; Souza, T.M.L.; Kurtz, P.; et al. Platelet activation and platelet-monocyte aggregate formation trigger tissue factor expression in patients with severe COVID-19. *Blood* **2020**, *136*, 1330–1341. [[CrossRef](#)] [[PubMed](#)]
47. Fard, M.B.; Fard, S.B.; Ramazi, S.; Atashi, A.; Eslamifar, Z. Thrombosis in COVID-19 infection: Role of platelet activation-mediated immunity. *Thromb. J.* **2021**, *19*, 59. [[CrossRef](#)]

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