UCSF

UC San Francisco Previously Published Works

Title

Reversibility of the effects of natalizumab on peripheral immune cell dynamics in MS patients

Permalink

https://escholarship.org/uc/item/62q0x2bx

Journal

Neurology, 89(15)

ISSN

0028-3878

Authors

Plavina, Tatiana
Muralidharan, Kumar Kandadi
Kuesters, Geoffrey
et al.

Publication Date

2017-10-10

DOI

10.1212/wnl.0000000000004485

Peer reviewed

Reversibility of the effects of natalizumab on peripheral immune cell dynamics in MS patients

OPEN ▲

Tatiana Plavina, PhD
Kumar Kandadi
    Muralidharan, PhD
Geoffrey Kuesters, PhD
Daniel Mikol, MD, PhD
Karleyton Evans, MD
Meena Subramanyam,
    PhD
Ivan Nestorov, PhD
Yi Chen, MS
Qunming Dong, PhD
Pei-Ran Ho, MD
Diogo Amarante, MD
Alison Adams, PhD
Jerome De Sèze, MD
Robert Fox, MD
Ralf Gold, MD
Douglas Jeffery, MD
Ludwig Kappos, MD
Xavier Montalban, MD,
    PhD
Bianca Weinstock-
    Guttman, MD
Hans-Peter Hartung, MD
Bruce A.C. Cree, MD,
    PhD, MAS

Correspondence to
Dr. Plavina:
Tatiana.plavina@biogen.com

ABSTRACT

Objective: To characterize the reversibility of natalizumab-mediated changes in pharmacokinetics/pharmacodynamics in patients with multiple sclerosis (MS) following therapy interruption.

Methods: Pharmacokinetic/pharmacodynamic data were collected in the Safety and Efficacy of Natalizumab in the Treatment of Multiple Sclerosis (AFFIRM) (every 12 weeks for 116 weeks) and Randomized Treatment Interruption of Natalizumab (RESTORE) (every 4 weeks for 28 weeks) studies. Serum natalizumab and soluble vascular cell adhesion molecule–1 (sVCAM-1) were measured using immunoassays. Lymphocyte subsets, α4-integrin expression/saturation, and vascular cell adhesion molecule–1 (VCAM-1) binding were assessed using flow cytometry.

Results: Blood lymphocyte counts (cells/L) in natalizumab-treated patients increased from 2.1×10^9 to 3.5×10^9. Starting 8 weeks post last natalizumab dose, lymphocyte counts became significantly lower in patients interrupting treatment than in those continuing treatment (3.1×10^9 vs 3.5×10^9; $p = 0.031$), plateauing at prenatalizumab levels from week 16 onward. All measured cell subpopulation, α4-integrin expression/saturation, and sVCAM changes demonstrated similar reversibility. Lymphocyte counts remained within the normal range. Ex vivo VCAM-1 binding to lymphocytes increased until ≈16 weeks after the last natalizumab dose, then plateaued, suggesting reversibility of immune cell functionality. The temporal appearance of gadolinium-enhancing lesions was consistent with pharmacodynamic marker reversal.

Conclusions: Natalizumab's effects on peripheral immune cells and pharmacodynamic markers were reversible, with changes starting 8 weeks post last natalizumab dose; levels returned to those observed/expected in untreated patients ≈16 weeks post last dose. This reversibility differentiates natalizumab from MS treatments that require longer reconstitution times. Characterization of the time course of natalizumab's biological effects may help clinicians make treatment sequencing decisions.

Classification of evidence: This study provides Class III evidence that the pharmacodynamic markers of natalizumab are reversed ≈16 weeks after stopping natalizumab. *Neurology*® 2017;89:1584–1593

GLOSSARY

AFFIRM = Safety and Efficacy of Natalizumab in the Treatment of Multiple Sclerosis; **CTS** = Clinical Trials Services; **EID** = extended interval dosing; **Gd+** = gadolinium-enhancing; **MS** = multiple sclerosis; **NK** = natural killer; **PD** = pharmacodynamics; **PK** = pharmacokinetics; **RESTORE** = Randomized Treatment Interruption of Natalizumab; **RRMS** = relapsing-remitting multiple sclerosis; **sVCAM-1** = soluble vascular cell adhesion molecule–1; **VCAM-1** = vascular cell adhesion molecule–1; **WBC** = white blood cell.

A number of therapies for relapsing-remitting multiple sclerosis (RRMS) are currently available; each has a distinct mechanism of action and benefit/risk profile and exhibits a different temporal pattern with respect to reversibility of the drug's overall biological effects. Given the expansive and complex RRMS treatment landscape, when patients experience treatment failure and

alternative therapies are being considered, treatment sequencing is an important issue. Understanding the temporal nature of the biological effect of individual therapies may facilitate treatment selection.

Natalizumab is a recombinant humanized monoclonal antibody that targets the α4 subunit of the α4β1 integrin on mononuclear leukocytes and is approved for the treatment of relapsing multiple sclerosis (MS).[1] When bound by natalizumab, α4β1 integrin is blocked from binding to vascular cell adhesion molecule–1 (VCAM-1), disrupting the homing and trafficking of mononuclear leukocytes across activated vascular endothelium of the CNS.[2–4]

The pharmacodynamics (PD), the drug effect on biological systems, of natalizumab include increased saturation/occupancy by natalizumab of its target, α4-integrin (CD49d), on the surface of circulating lymphocytes[2,5,6]; decreased surface expression of α4-integrin on lymphocytes; and decreased serum concentration of soluble VCAM-1 (sVCAM-1).[7–9] Natalizumab use is also associated with increased levels of peripheral immune cells, although these levels remain within the normal range for all investigated cell types except CD34+ progenitor cells.[5,10,11]

Natalizumab is highly efficacious at reducing both clinical and radiologic disease activity.[10] Upon natalizumab treatment interruption, disease activity returns, with the earliest MRI lesions observed 12 weeks after the last natalizumab dose.[12,13] However, the time course of reversibility of natalizumab's effects on immune cells is not well-characterized. Data from 2 large clinical trials allowed assessment of changes that occur upon natalizumab initiation (Safety and Efficacy of Natalizumab in the Treatment of Multiple Sclerosis [AFFIRM] study) and following the last natalizumab dose (Randomized Treatment Interruption of Natalizumab [RESTORE] study). We analyze the pharmacokinetics (PK), the time course of drug concentration, and the PD from these studies to further characterize the timing of the biological changes in the peripheral compartment associated with natalizumab treatment initiation and interruption.

METHODS **Study design.** AFFIRM was a randomized, multicenter, double-blind, placebo-controlled phase 3 trial conducted from November 2001 to January 2005 (NCT00027300;

clinicaltrials.gov).[10] Details of the study design and patient population were previously published.[10] Patients were randomized 2:1 to receive 300 mg natalizumab or placebo IV every 4 weeks for up to 116 weeks.

RESTORE was a randomized, multicenter, partially blinded, parallel-group exploratory study conducted between March 2010 and November 2011 (NCT01071083; clinicaltrials.gov).[12] Details of the study design and patient population were previously published.[12,13] Patients who had received 300 mg natalizumab IV for ≥12 months prior to trial entry were randomized (1:1:2) to natalizumab, placebo, or alternate therapy (IM interferon-β-1a [Avonex; Biogen, Cambridge, MA], glatiramer acetate [Copaxone; Teva Neuroscience, Kansas City, MO], or methylprednisolone). Administration of natalizumab and placebo was double-blind; alternative therapies were administered open-label. Randomization occurred at week 0 (in which all patients received their last natalizumab infusion). At week 28, patients discontinued placebo or alternate therapy and restarted open-label natalizumab.[12] Patients who experienced 1 new gadolinium-enhancing (Gd+) lesion of >0.8 cm³ in volume, ≥2 Gd+ lesions of any size, or ≥1 clinical relapses could receive high-dose corticosteroid treatment or restart open-label natalizumab treatment at the investigator's discretion. Patients who received open-label natalizumab during the treatment interruption period proceeded directly to the open-label follow-up period.

Primary research question. This analysis was conducted to characterize the timing of the PK and PD changes, including changes in peripheral immune cells, upon natalizumab treatment initiation and interruption.

Classification of evidence. This study provides Class III evidence that the PD markers of natalizumab's activity are reversed ≈16 weeks after stopping natalizumab.

Assessments. In AFFIRM, samples for PK and limited PD evaluations (total lymphocyte and white blood cell [WBC] counts) were taken from all patients at baseline (prenatalizumab) and prior to infusion at weeks 12, 24, 36, 48, 60, 72, 84, 96, 108, and 120. A cohort of 31 patients (natalizumab, n = 20; placebo, n = 11) from a subset of US sites underwent weekly measurement of α4-integrin saturation following the first dose of natalizumab; α4-integrin saturation was also measured every 24 weeks in a separate subset of 21 patients (natalizumab, n = 15; placebo, n = 6). Blood for hematology and serum chemistry were analyzed at a central laboratory (Covance Central Laboratory Services, Indianapolis, IN; Covance, Geneva, Switzerland; or Sonic Trials Laboratory, North Ryde, Australia). Natalizumab concentration was measured by Biogen using an ELISA; α4-integrin saturation on circulating lymphocytes (indicating the proportion of α4-integrin receptors on the lymphocyte cell surface membrane occupied by natalizumab) was analyzed using flow cytometry by Elan Pharmaceuticals (Dublin, Ireland). The normal range for peripheral blood cells was defined as the normal range in healthy individuals as provided by Covance (shown in table e-1 at Neurology.org).

In RESTORE, clinical, MRI, and laboratory evaluations were performed at baseline and weeks 4, 8, 12, 16, 20, 24, and 28 of the randomized treatment period. Serum natalizumab concentration was measured at Charles River Laboratories Preclinical Services (Senneville, Canada). Serum VCAM-1 concentration was measured using a LINCOplex immunoassay (Linco Research, Inc., St. Charles, MO) at Covance (Geneva, Switzerland, and Indianapolis, IN). In addition to the cell subsets analyzed in AFFIRM, the following lymphocyte populations were analyzed: CD3+CD4+ T cells, CD3+CD8+ T cells, CD19+ B cells,

natural killer (NK) cells (CD3−/CD16+CD56+), and hematopoietic stem cells (CD34+/CD45+). Lymphocyte and leukocyte subset counts, α4-integrin saturation, CD49d expression, and binding of cells to fluorescently labeled VCAM-1–immunoglobulin reagent were determined using flow cytometric analysis of whole blood by Esoterix Clinical Trials Services (CTS)–Belgium (Mechelen, Belgium) or Esoterix CTS–Cranford (Cranford, NJ), except for leukocyte–VCAM-1 binding assays, which were performed by Esoterix CTS–Cranford. Flow cytometry reagents and gating strategies are shown in appendix e-1. Because the prenatalizumab levels for lymphocyte subsets were not measured in RESTORE, values were compared with normal ranges reported in the literature.[11,14]

Standard protocol approvals, registration, and patient consent. For both the AFFIRM and RESTORE studies, the protocol was reviewed and approved by site institutional review boards. All patients provided written informed consent, and the studies were conducted in accordance with the Declaration of Helsinki and International Conference on Harmonisation Guideline on Good Clinical Practice.

Statistical analyses. RESTORE participants were categorized into 2 groups: those who continued natalizumab treatment and those whose natalizumab treatment was interrupted (i.e., switched from natalizumab to placebo or an alternative therapy). Combining the patients who switched from natalizumab into a single group provided additional statistical power to examine differences between those who remained on natalizumab and those who interrupted natalizumab treatment. Repeated-measure models were employed to calculate statistical differences in PK and PD over time between the 2 groups. Estimates were obtained for each treatment group from all available data at each collection time.

Subgroup analyses of total lymphocyte counts were conducted in RESTORE patient groups stratified by median age, level of disease activity, and sex. High disease activity in RESTORE was defined as ≥2 relapses in the year prior to natalizumab treatment initiation.

RESULTS Study population. In AFFIRM, 627 patients received natalizumab and 315 received placebo. As reported previously, no significant between-group differences in demographic or disease characteristics were observed.[10] In RESTORE, 45 patients continued natalizumab, whereas 130 patients interrupted natalizumab treatment, switching to placebo (n = 42), IM interferon-β-1a (n = 17), glatiramer acetate (n = 17), or methylprednisolone (n = 54). Baseline characteristics of patients who switched to placebo or an alternative therapy were generally similar, with the exception of Expanded Disability Status Scale score.[12] The baseline characteristics of patients who continued natalizumab and those whose natalizumab treatment was interrupted were similar (table 1).

Participants in RESTORE were older than those in AFFIRM (mean age, 41 vs 36 years) and had a longer disease duration (median duration, 11 vs 5 years), as shown in table e-2. A greater proportion of RESTORE patients (97%) than AFFIRM patients (30%) had taken a prior disease-modifying therapy. In addition, in contrast to AFFIRM patients, RESTORE patients had received natalizumab for >1 year at study entry.

PK/PD of natalizumab dosing in AFFIRM. *Serum natalizumab levels.* In AFFIRM, mean natalizumab concentration reached a plateau about 36 weeks after treatment initiation (figure e-1A).

α4-Integrin saturation. Mean α4-integrin saturation, measured in a subset of 31 AFFIRM patients, exceeded 90% ≤2 hours after the first natalizumab infusion and remained above 85% in natalizumab-treated patients over the 4 weeks following the first natalizumab dose (figure e-1B). Based on assessments of trough α4-integrin saturation taken every 24 weeks, similar levels of saturation were maintained in natalizumab-treated patients in another subset of 21 AFFIRM patients over the course of the study.

Table 1	Baseline characteristics of RESTORE patients		
Characteristic	Natalizumab treatment continued (n = 45)	Natalizumab treatment interrupted (n = 130)	Total (n = 175)
Age, y, mean (SD)	41.2 (9.70)	41.2 (9.94)	41.2 (9.85)
Female, n (%)	37 (82)	98 (75)	135 (77)
Time since MS symptom onset, y			
Mean (SD)	12.7 (7.19)	12.1 (7.94)	12.2 (7.74)
Median (range)	12 (2-40)	10 (2-41)	11 (2-41)
EDSS score at randomization			
Mean (SD)	3.0 (1.75)	3.2 (1.72)[a]	3.2 (1.73)
Median (range)	3.0 (0-6.5)	3.0 (0-8.0)[a]	3.0 (0-8.0)
Total number of natalizumab infusions prior to randomization			
Mean (SD)	29.3 (10.88)	29.1 (9.73)	29.1 (10.01)
Median (range)	29 (12, 49)	28 (12, 51)	28 (12, 51)

Abbreviations: EDSS = Expanded Disability Status Scale; MS = multiple sclerosis; RESTORE = Randomized Treatment Interruption of Natalizumab.
[a] No. = 129.

Peripheral blood cell counts. In AFFIRM, total lymphocyte counts at baseline were the same in the natalizumab and placebo groups. While minimal change was observed in placebo-treated patients during the study, the least-squares mean estimate of total blood lymphocytes in natalizumab patients increased from 2.1×10^9 cells/L at baseline to 3.5×10^9 cells/L at week 12, the first evaluated time point. These levels reached a plateau of 3.7×10^9 cells/L 24 weeks after natalizumab initiation and stayed in the 3.5×10^9–3.7×10^9 cells/L range for the remainder of the study (figure e-1C).

Levels of other peripheral blood cells were also analyzed in AFFIRM. Natalizumab treatment was associated with increases in total leukocyte (WBC), monocyte, basophil, and eosinophil counts over 120 weeks, similar to the increase in total lymphocyte count (figure e-1, D–G). Neutrophils, the only major leukocyte class with very low α4-integrin expression,[15] showed no significant difference in cell counts between natalizumab- and placebo-treated patients (figure e-1H). For all measured peripheral blood cells, mean counts remained within the normal range throughout the study period.

PK/PD after natalizumab treatment interruption in RESTORE. *Serum natalizumab.* In RESTORE, natalizumab concentrations at baseline and week 4 were similar in patients continuing natalizumab and those whose treatment was interrupted (30.6 vs 32.9 μg/mL for week 0; figure 1A). At week 8, natalizumab levels were significantly lower in patients who had interrupted treatment than in those who continued natalizumab (8.7 vs 34.7 μg/mL; $p < 0.001$). Starting at week 16, natalizumab concentrations were below the lower limit of quantification (0.25 μg/mL) for >33% of the patients who had interrupted treatment; therefore, mean natalizumab levels were not calculated for this cohort from week 16 onward.

α4-Integrin saturation and other PD markers. RESTORE patients had been treated with natalizumab for >1 year prior to study entry. Accordingly, their mean trough α4-integrin saturation levels at baseline (86%) (figure 1B) were similar to the levels observed in natalizumab-treated patients in AFFIRM (figure e-1B). RESTORE patients who continued natalizumab consistently maintained mean trough α4-integrin saturation levels near 85%–90%. At week 8, saturation levels were significantly lower in patients who had stopped natalizumab than in those who were maintained on treatment (68% vs 87%; $p < 0.001$); saturation levels in patients with natalizumab treatment interruption continued to decline at week 12 and reached a plateau of approximately 10%–15% saturation (consistent with values

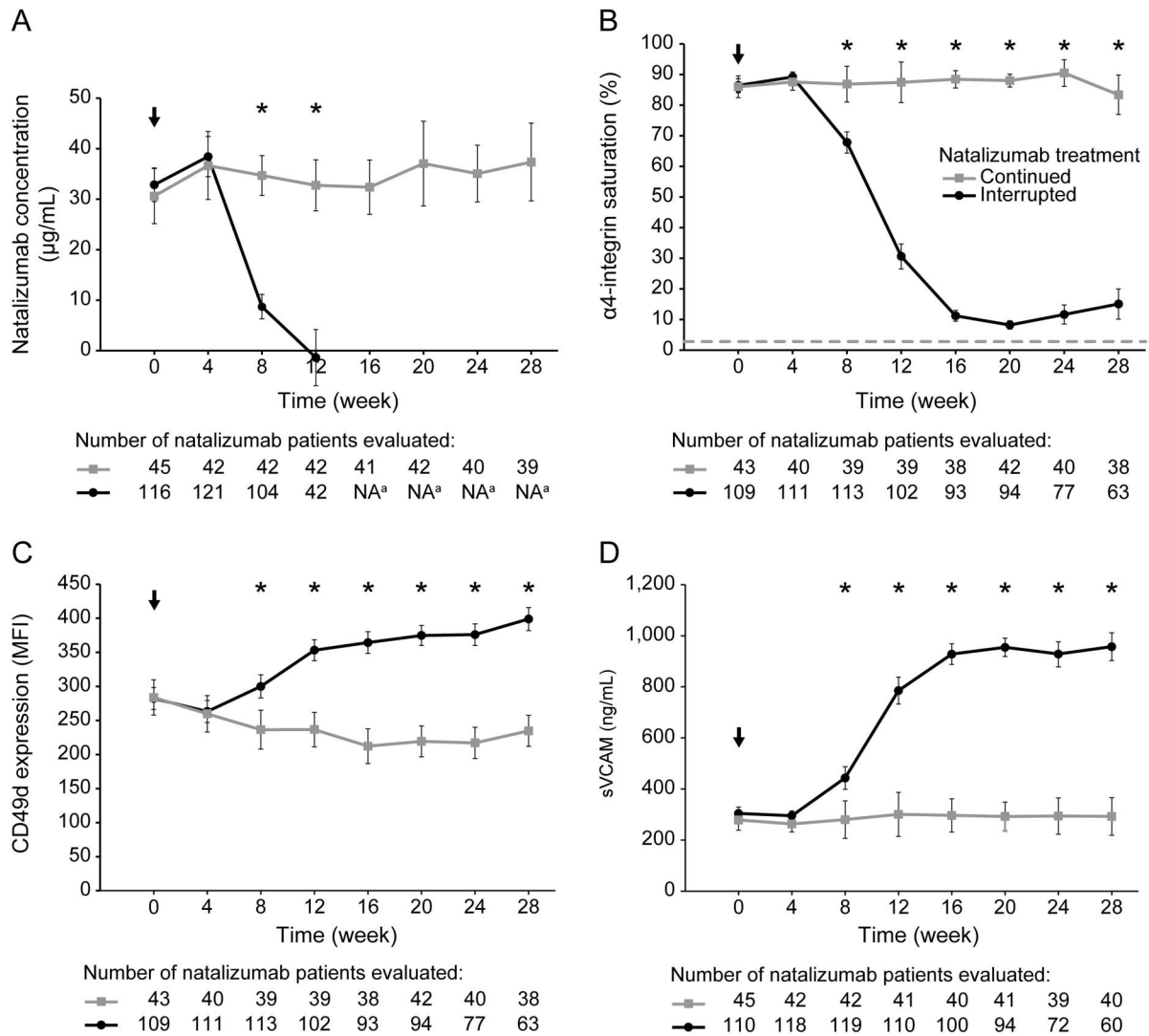observed prior to natalizumab treatment in AFFIRM) by about week 16.

In patients whose natalizumab treatment was interrupted, serum sVCAM concentration and α4-integrin (CD49d) expression each increased from baseline levels starting at week 8 and reached plateaus at approximately week 16, concurrently with decreases in natalizumab concentration and α4-integrin saturation (figure 1, C and D).

Peripheral blood cell counts. Among RESTORE patients who continued natalizumab treatment, absolute lymphocytes and lymphocyte subset cell counts did not change significantly during the randomized treatment period (figure 2, A–F). In contrast, patients who stopped natalizumab showed a significant reduction in total lymphocyte counts starting 8 weeks after the last natalizumab dose compared with patients continuing natalizumab (3.1×10^9 cells/L vs 3.5×10^9 cells/L; $p = 0.031$; figure 2A). From 16 weeks onward, total lymphocyte counts in patients who discontinued natalizumab treatment remained below those of patients who continued natalizumab (1.9×10^9 cells/L vs 3.4×10^9 cells/L; $p < 0.001$) and plateaued at levels similar to those observed prior to natalizumab initiation (2.1×10^9 cells/L, as assessed in AFFIRM prior to natalizumab treatment; figure e-1C). Similar trends were observed regardless of whether patients switched to placebo, interferon-β-1a, glatiramer acetate, or methylprednisolone (figure e-2).

All measured lymphocyte subpopulations whose levels were affected by natalizumab, including T, B, and NK cells, as well as CD34+ hematopoietic progenitor cells, demonstrated similar time courses of reversibility after the last natalizumab dose (figure 2, B–F). Compared with leukocyte levels in patients who continued natalizumab, patients whose treatment was interrupted exhibited significant decreases from baseline in total WBC, monocyte, basophil, and eosinophil counts at week 16 ($p < 0.001$; figure e-3, A–D). No notable change in cell counts or difference between the on-natalizumab and natalizumab-interruption groups was observed for neutrophils (figure e-3E).

Subgroup analyses. When RESTORE patients were grouped based on age, younger patients (≤42 years) tended to have higher total lymphocyte counts than older patients (>42 years) throughout the study (figure 2G). Similarly, total lymphocyte counts were higher in patients with high vs low disease activity (figure 2H) and in women vs men (figure 2I). In each of these subgroups, the temporal profile of reversibility was comparable to that observed in the overall population, with significantly lower levels observed starting at week 12 in all patient groups with natalizumab treatment

Least squares means estimates ± 95% confidence interval for (A) natalizumab concentration, (B) leukocyte percentage α4-integrin saturation, (C) leukocyte α4-integrin (CD49d) expression, and (D) serum vascular cell adhesion molecule (sVCAM) expression in patients continuing natalizumab or with a treatment interruption. The arrow represents the last dose before the 24-week interruption period. For percentage α4-integrin saturation (B), the gray dashed line indicates saturation prior to natalizumab treatment in the Safety and Efficacy of Natalizumab in the Treatment of Multiple Sclerosis (AFFIRM) natalizumab treatment arm (2.8%), as shown in figure e-1B. MFI = mean fluorescence intensity; RESTORE = Randomized Treatment Interruption of Natalizumab. *$p < 0.001$. [a]Mean natalizumab levels were not analyzed once ≥33% of patients had values below the lower limit of quantification.

interruption than in those who continued treatment. Total lymphocyte counts plateaued at week 16 and were maintained at levels similar to those observed prior to natalizumab (as assessed in the AFFIRM natalizumab arm; figure e-1C).

Functional assays. T cells, B cells, and monocytes from patients whose treatment was interrupted demonstrated significantly greater ex vivo binding to VCAM-1 starting at week 12 ($p < 0.001$ for B cells and monocytes; $p = 0.001$ for T cells) than in patients who remained on natalizumab (figure 3). VCAM-1 binding progressively increased through approximately 16 weeks after the last natalizumab dose and then plateaued.

Return of disease activity. When α4-integrin saturation values were evaluated in relation to the timing of the appearance of Gd+ lesions in the overall population of RESTORE, a temporal relationship was observed between decreases in α4-integrin saturation and the appearance of Gd+ lesions (figure 4).

DISCUSSION Given the growing number of available disease-modifying therapies for RRMS, treatment sequencing decisions are becoming more complex and require careful considerations of the mechanism of action, safety, efficacy, and reversibility of each drug's effects. Using data from the RESTORE study, we provide a comprehensive evaluation of the
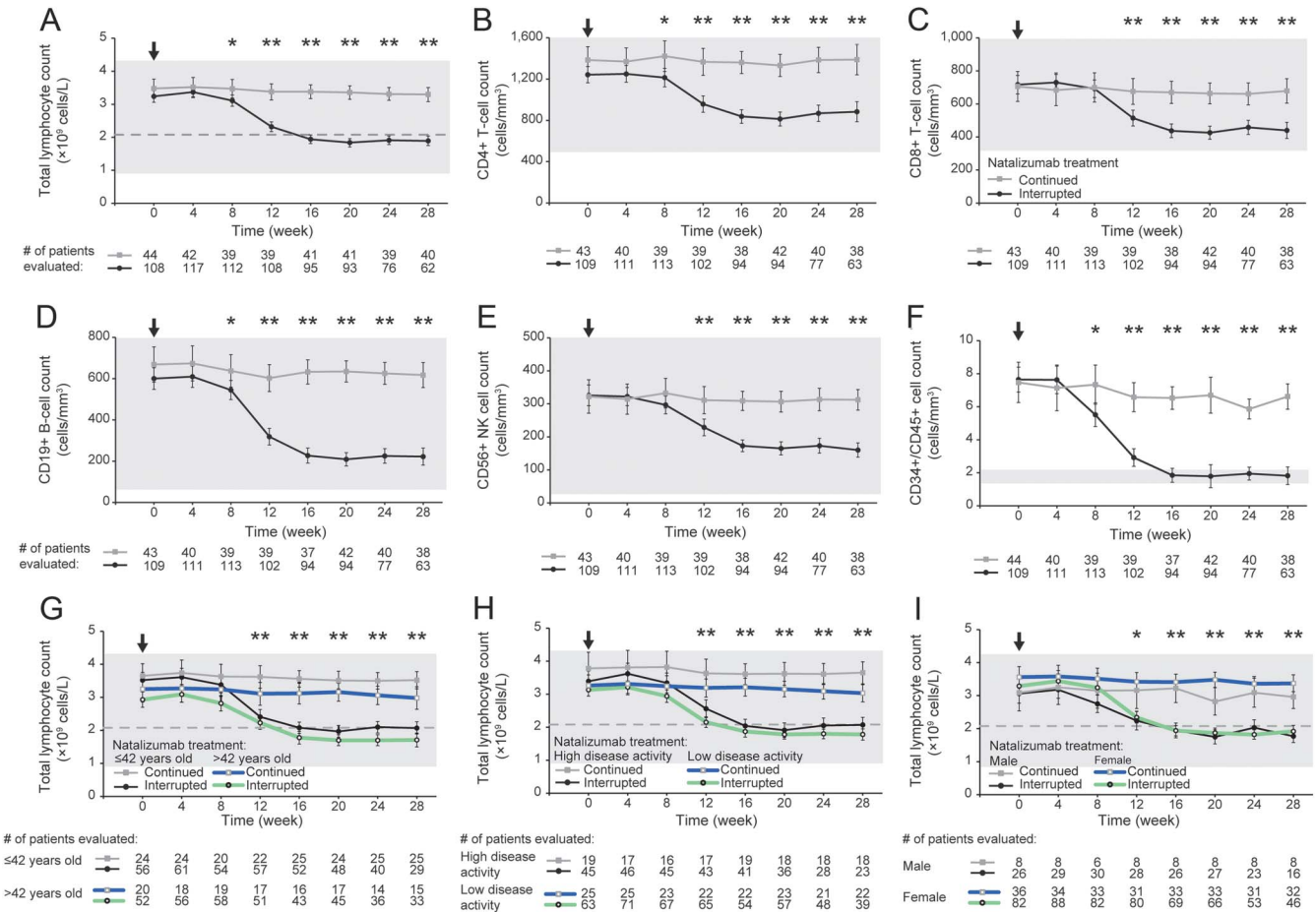
Least squares (LS) mean estimated counts ± 95% confidence interval (CI) for (A) total lymphocytes, (B) CD4+ T cells, (C) CD8+ T cells, (D) CD19+ B cells, (E) CD3−/CD16+/CD56+ natural killer (NK) cells, and (F) CD34+/CD45+ hematopoietic progenitor cells. Total lymphocyte counts in RESTORE patients stratified by demographics and disease activity. LS mean estimates ± 95% CI are shown for patients continuing and discontinuing natalizumab classified by (G) age, (H) disease activity, and (I) sex. The arrow represents the last dose before the 24-week interruption period. The shaded area shows the range of normal lymphocyte counts (total lymphocyte counts: 0.91–4.28 × 10^9 cells/L [Safety and Efficacy of Natalizumab in the Treatment of Multiple Sclerosis (AFFIRM)]; CD4+ T cells[14]: 491–2,000 cells/mm³; CD8+ T cells[14]: 314–2,087 cells/mm³; CD19+ B cells[14]: 64–800 cells/mm³; CD3−/CD16+/CD56+ NK cells[14]: 27–693 cells/mm³; CD34+/CD45+ hematopoietic progenitor cells[11]: 1.4–2.2 cells/mm³). For total lymphocytes (A, G, H, I), the gray dashed line indicates total lymphocyte counts prior to natalizumab treatment in the AFFIRM natalizumab treatment arm (2.08 × 10^9 cells/L), as shown in figure e-1C. Lymphocyte subsets were not assessed in AFFIRM. RESTORE = Randomized Treatment Interruption of Natalizumab. *p < 0.05. **p < 0.001.

reversibility of natalizumab's PK and PD effects and their relationship to clinical and MRI characteristics.

The pattern of reversibility demonstrated here adds to previous analyses[12] to offer a more complete profile of the temporal effects of natalizumab discontinuation. The earliest detectable changes in PK and PD markers were observed 8 weeks after natalizumab treatment interruption and involved serum natalizumab concentration and α4-integrin saturation on peripheral lymphocytes. By week 12, peripheral lymphocyte subset counts had decreased significantly, and by week 16 all PK and PD markers appeared similar to those observed in patients not treated with natalizumab. This reversal of natalizumab's PK and PD effects coincided with the return of clinical and radiologic disease activity. Gd+ lesions were observed in RESTORE as early as 12 weeks

after the last dose of natalizumab, with the majority of Gd+ lesions appearing at and after 16 weeks following natalizumab discontinuation,[13] suggesting that the PD and pharmacologic effects of natalizumab are tightly linked.

As some MS therapies are known to affect immune cell composition,[16–21] we assessed change in total lymphocyte counts in groups stratified by therapy being administered during the randomized treatment period in RESTORE. As results were similar regardless of whether patients were on placebo, interferon-β-1, glatiramer acetate, or methylprednisolone, data from patients on placebo or an alternative therapy were combined to assess the overall effect of natalizumab treatment interruption on PK, PD, and peripheral immune cells.

Figure 3 Functional activity of CD3+ T cells, CD19+ B cells, and monocytes from RESTORE patients

CD3+/VCAM-1 binding
— Natalizumab continued
— Natalizumab interrupted

CD19+/VCAM-1 binding
— Natalizumab continued
— Natalizumab interrupted

Monocyte/VCAM-1 binding
— Natalizumab continued
— Natalizumab interrupted

Number of patients evaluated:

Continued	42	40	39	39	38	42	40	38
Discontinued	109	111	113	102	94	94	77	63
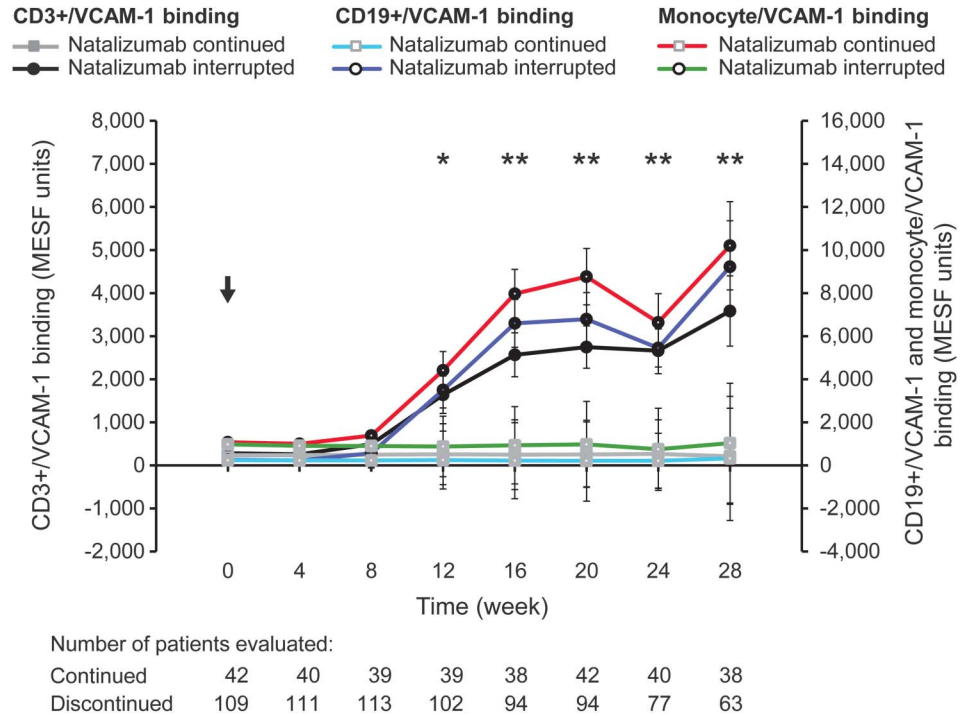
Least squares mean estimates ± 95% confidence interval are shown for ex vivo binding of cells to vascular cell adhesion molecule–1 (VCAM-1). The arrow represents the last dose before the 24-week interruption period. MESF = molecules of equivalent soluble fluorochrome; RESTORE = Randomized Treatment Interruption of Natalizumab. *p = 0.001 for T cells; $p < 0.001$ for B cells and monocytes. **$p < 0.001$ for T cells, B cells, and monocytes.

For a comprehensive understanding of natalizumab's biological effects, and to establish comparator PK/PD marker levels for patients without

Figure 4 Relationship of α4-integrin saturation and the incidence of gadolinium-enhancing (Gd+) lesions over time after the last natalizumab dose

Range of α4-integrin saturation values
Mean α4-integrin saturation
Participants with Gd+ lesions (%)

The median α4-integrin saturation level and the cumulative percentage of patients with a natalizumab treatment interruption developing Gd+ lesions are shown. The last natalizumab dose was administered on day 0. Patients with >1 Gd + lesion, %, represents the cumulative percentage of study patients with Gd + lesions out of all patients not treated with natalizumab who remained in the study at that time point.

natalizumab treatment, the present analyses employed data from the AFFIRM study. Although these findings from AFFIRM have previously been mentioned,[10] this report serves as the first presentation of those data.

As previously reported,[10] treatment with natalizumab is associated with increases in circulating immune cells from prenatalizumab levels, although these levels remain within the normal range. The magnitude and timing of the changes observed in natalizumab-treated patients were similar to those published in smaller observational studies.[11,22–29] CD34+ hematopoietic progenitor cells were previously found to increase above the normal reference levels in natalizumab-treated patients.[11] We observed similarly increased levels of CD34+ cells (mean of 7.7 cells/mm³); however, clonality (evaluated by the excess immunoglobulin light chain kappa or lambda proteins on cells) was not observed (data not shown), and counts returned to the normal range approximately 16 weeks after treatment interruption. Increases in activated T cells expressing proinflammatory cytokines were also reported for natalizumab-treated patients[30]; however, this study did not assess cytokine production.

A key limitation of the present analyses is that RESTORE patients were already receiving natalizumab at the start of the study; therefore, prenatalizumab levels

were not available for this population. Some assessments were performed at baseline in AFFIRM, yielding data that served as surrogate prenatalizumab reference levels. However, not all of the cell types assessed in RESTORE were measured in AFFIRM, and the baseline characteristics of AFFIRM and RESTORE patients were somewhat different. Specifically, AFFIRM patients were younger and had been diagnosed with MS more recently than RESTORE patients. Thus, PK and PD responses might differ between the AFFIRM and RESTORE patients. Indeed, characteristics such as age, race, and disease severity may influence circulating lymphocyte counts.[31,32] Similarly, in a subanalysis of RESTORE patients, those who were younger and those with high disease activity had higher total lymphocyte counts than older patients or those with lower disease activity. Nevertheless, the reversibility timing and pattern observed in patients with a natalizumab treatment interruption was consistent in each subgroup, suggesting that these results may be generalizable.

These data are limited in their ability to inform clinical practices other than discontinuation following monthly natalizumab administration. Of note, extending the natalizumab dosing interval up to 8 weeks has been suggested as a way to reduce costs and adverse events while maintaining efficacy.[33,34] The observations from RESTORE that PK, PD, and MRI measures only became significantly different 8 weeks after the last natalizumab dose do not permit conclusions regarding whether extended interval dosing (EID) would maintain PD and MS disease control similar to 4-week dosing. It should not be assumed that steady-state PK/PD levels will be the same after each dose of repeated EID as after discontinuation of every-4-week treatment. In AFFIRM patients receiving monthly natalizumab doses, steady-state levels were reached approximately 36 weeks after treatment initiation; the time required to reach these same levels during EID may be different. One additional limitation to the present analysis is that the reversibility of CSF lymphocyte changes was not assessed in RESTORE. Findings from a previous study by Stüve et al.[35] suggest that the effects of natalizumab on CSF lymphocyte counts may persist as long as 6 months after treatment cessation. However, the observations of that study may be limited by the small sample size, the lack of prenatalizumab CSF data, and the use of nonrandomized controls.

Overall, the results of this study demonstrate that the PD effects of natalizumab on peripheral immune cells and other markers begin to decline starting at weeks 8–12, with levels returning to those observed or expected for patients not treated with natalizumab around 16 weeks after the last natalizumab dose. The reversal of PD changes was tightly linked with the recurrence of disease activity measured by MRI. A thorough understanding of a drug's reversibility can inform the timing of initiation of an alternative therapy. Immunotherapies available or soon to be available for MS treatment such as alemtuzumab, ocrelizumab, and cladribine, whose mechanisms of action rely on depletion of specific cell populations, appear to require at least 9–18 months, and possibly up to 35 months, to reconstitute the depleted B- and T-cell populations.[17–21] Given the complexity of RRMS treatment options, the qualitative and quantitative characterization of the biological effects of natalizumab reported here may help clinicians make treatment-sequencing decisions.

REFERENCES

1. Tysabri (Natalizumab) [prescribing information]. Cambridge, MA: Biogen, Inc.; 2016.

2. Rudick RA, Sandrock A. Natalizumab: alpha 4-integrin antagonist selective adhesion molecule inhibitors for MS. Expert Rev Neurother 2004;4:571–580.

3. Yednock TA, Cannon C, Fritz LC, Sanchez-Madrid F, Steinman L, Karin N. Prevention of experimental autoimmune encephalomyelitis by antibodies against alpha 4 beta 1 integrin. Nature 1992;356:63–66.

4. Benkert TF, Dietz L, Hartmann EM, et al. Natalizumab exerts direct signaling capacity and supports a proinflammatory phenotype in some patients with multiple sclerosis. PLoS One 2012;7:e52208.

5. Miller DH, Khan OA, Sheremata WA, et al. A controlled trial of natalizumab for relapsing multiple sclerosis. N Engl J Med 2003;348:15–23.

6. Khatri BO, Man S, Giovannoni G, et al. Effect of plasma exchange in accelerating natalizumab clearance and restoring leukocyte function. Neurology 2009;72:402–409.

7. Defer G, Mariotte D, Derache N, et al. CD49d expression as a promising biomarker to monitor natalizumab efficacy. J Neurol Sci 2012;314:138–142.

8. Harrer A, Wipfler P, Einhaeupl M, et al. Natalizumab therapy decreases surface expression of both VLA-heterodimer subunits on peripheral blood mononuclear cells. J Neuroimmunol 2011;234:148–154.

9. Millonig A, Hegen H, Di Pauli F, et al. Natalizumab treatment reduces endothelial activity in MS patients. J Neuroimmunol 2010;227:190–194.

10. Polman CH, O'Connor PW, Havrdova E, et al. A randomized, placebo-controlled trial of natalizumab for relapsing multiple sclerosis. N Engl J Med 2006;354:899–910.

11. Bonig H, Wundes A, Chang KH, Lucas S, Papayannopoulou T. Increased numbers of circulating hematopoietic stem/progenitor cells are chronically maintained in patients treated with the CD49d blocking antibody natalizumab. Blood 2008;111:3439–3441.

12. Fox RJ, Cree BA, De Seze J, et al. MS disease activity in RESTORE: a randomized 24-week natalizumab treatment interruption study. Neurology 2014;82:1491–1498.

13. Kaufman M, Cree BA, De Seze J, et al. Radiologic MS disease activity during natalizumab treatment interruption: findings from RESTORE. J Neurol 2015;262:326–336.

14. Valiathan R, Deeb K, Diamante M, Ashman M, Sachdeva N, Asthana D. Reference ranges of lymphocyte subsets in healthy adults and adolescents with special mention of T cell maturation subsets in adults of South Florida. Immunobiology 2014;219:487–496.

15. Kubes P, Niu XF, Smith CW, Kehrli ME Jr, Reinhardt PH, Woodman RC. A novel beta 1-dependent adhesion pathway on neutrophils: a mechanism invoked by dihydrocytochalasin B or endothelial transmigration. FASEB J 1995;9:1103–1111.

16. Francis G, Kappos L, O'Connor P, et al. Temporal profile of lymphocyte counts and relationship with infections with fingolimod therapy. Mult Scler 2014;20:471–480.

17. Lemtrada (Alemtuzumab) [prescribing information]. Cambridge, MA: Genzyme Corporation; 2016.

18. Bar-Or A, Calabresi PA, Arnold D, et al. Rituximab in relapsing-remitting multiple sclerosis: a 72-week, open-label, phase I trial. Ann Neurol 2008;63:395–400.

19. Kappos L, Li D, Calabresi P, et al. Long-term safety and efficacy of ocrelizumab in patients with relapsing-remitting multiple sclerosis: week 144 results of a phase II, randomised, multicentre trial. Presented at the 28th Congress of the European Committee for Treatment and Research in Multiple Sclerosis; October 10–13, 2012; Lyon, France; P362.

20. Hill-Cawthorne GA, Button T, Tuohy O, et al. Long term lymphocyte reconstitution after alemtuzumab treatment of multiple sclerosis. J Neurol Neurosurg Psychiatry 2012;83:298–304.

21. Selby R, Brandwein J, O'Connor P. Safety and tolerability of subcutaneous cladribine therapy in progressive multiple sclerosis. Can J Neurol Sci 1998;25:295–299.

22. Harrer A, Pilz G, Einhaeupl M, et al. Lymphocyte subsets show different response patterns to in vivo bound natalizumab: a flow cytometric study on patients with multiple sclerosis. PLoS One 2012;7:e31784.

23. Harrer A, Pilz G, Wipfler P, et al. High interindividual variability in the CD4/CD8 T cell ratio and natalizumab concentration levels in the cerebrospinal fluid of patients with multiple sclerosis. Clin Exp Immunol 2015;180:383–392.

24. Koudriavtseva T, Sbardella E, Trento E, Bordignon V, D'Agosto G, Cordiali-Fei P. Long-term follow-up of peripheral lymphocyte subsets in a cohort of multiple
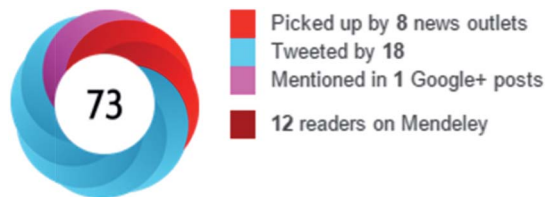
sclerosis patients treated with natalizumab. Clin Exp Immunol 2014;176:320–326.

25. Krumbholz M, Meinl I, Kumpfel T, Hohlfeld R, Meinl E. Natalizumab disproportionately increases circulating pre-B and B cells in multiple sclerosis. Neurology 2008;71:1350–1354.

26. Mattoscio M, Nicholas R, Sormani MP, et al. Hematopoietic mobilization: potential biomarker of response to natalizumab in multiple sclerosis. Neurology 2015;84:1473–1482.

27. Mellergard J, Edstrom M, Jenmalm MC, Dahle C, Vrethem M, Ernerudh J. Increased B cell and cytotoxic NK cell proportions and increased T cell responsiveness in blood of natalizumab-treated multiple sclerosis patients. PLoS One 2013;8:e81685.

28. Vennegoor A, Rispens T, Strijbis EM, et al. Clinical relevance of serum natalizumab concentration and anti-natalizumab antibodies in multiple sclerosis. Mult Scler 2013;19:593–600.

29. Zohren F, Toutzaris D, Klarner V, Hartung HP, Kieseier B, Haas R. The monoclonal anti-VLA-4 antibody natalizumab

30. mobilizes CD34+ hematopoietic progenitor cells in humans. Blood 2008;111:3893–3895.

30. Kivisakk P, Healy BC, Viglietta V, et al. Natalizumab treatment is associated with peripheral sequestration of proinflammatory T cells. Neurology 2009;72:1922–1930.

31. Jentsch-Ullrich K, Koenigsmann M, Mohren M, Franke A. Lymphocyte subsets' reference ranges in an age- and gender-balanced population of 100 healthy adults: a monocentric German study. Clin Immunol 2005;116:192–197.

32. Semra YK, Seidi OA, Sharief MK. Disease activity in multiple sclerosis correlates with T lymphocyte expression of the inhibitor of apoptosis proteins. J Neuroimmunol 2002;122:159–166.

33. Kister I, Corbory JR. Reducing costs while enhancing quality of care in MS. Neurology 2016;87:1617–1622.

34. Zhovtis Ryerson L, Frohman TC, Foley J, et al. Extended interval dosing of natalizumab in multiple sclerosis. J Neurol Neurosurg Psychiatry 2016;87:885–889.

35. Stüve O, Marra CM, Jerome KR, et al. Immune surveillance in multiple sclerosis patients treated with natalizumab. Ann Neurol 2006;59:743–747.

Discover Altmetrics

See real-time downloads and online activity for articles!

73

Picked up by **8** news outlets
Tweeted by **18**
Mentioned in **1** Google+ posts
12 readers on Mendeley

See more details

Authors and readers alike can view real-time data on articles including downloads and online activity across multiple sources. Click on the "Article Metrics" link in the right column of an article for details. To learn more about article metrics visit http://www.neurology.org/site/misc/article_usage.xhtml.