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Emergence of colistin resistance and characterization of antimicrobial resistance and virulence factors of *Aeromonas hydrophila*, *Salmonella* spp., and *Vibrio cholerae* isolated from hybrid red tilapia cage culture

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ABSTRACT

Background. Tilapia is a primary aquaculture fish in Thailand, but little is known about the occurrence of antimicrobial resistance (AMR) in *Aeromonas hydrophila*, *Salmonella* spp., and *Vibrio cholerae* colonizing healthy tilapia intended for human consumption and the co-occurrence of these AMR bacteria in the cultivation water.

Methods. This study determined the phenotype and genotype of AMR, extended-spectrum β -lactamase (ESBL) production, and virulence factors of *A. hydrophila*, *Salmonella* spp., and *V. cholerae* isolated from hybrid red tilapia and cultivation water in Thailand. Standard culture methods such as USDA's BAM or ISO procedures were used for the original isolation, with all isolates confirmed by biochemical tests, serotyping, and species-specific gene detection based on PCR.

Results. A total of 278 isolates consisting of 15 *A. hydrophila*, 188 *Salmonella* spp., and 75 *V. cholerae* isolates were retrieved from a previous study. All isolates of *A. hydrophila* and *Salmonella* isolates were resistance to at least one antimicrobial, with 26.7% and 72.3% of the isolates being multidrug resistant (MDR), respectively. All *A. hydrophila* isolates were resistant to ampicillin (100%), followed by oxytetracycline (26.7%), tetracycline (26.7%), trimethoprim (26.7%), and oxolinic acid (20.0%). The predominant resistance genes in *A. hydrophila* were *mcr-3* (20.0%), followed by 13.3% of isolates having *floR*, *qnrS*, *sul1*, *sul2*, and *dfrA1*. *Salmonella* isolates also exhibited a high prevalence of resistance to ampicillin (79.3%), oxolinic acid (75.5%), oxytetracycline (71.8%), chloramphenicol (62.8%), and florfenicol (55.3%). The most common resistance genes in these *Salmonella* isolates were *qnrS* (65.4%), *tetA* (64.9%), *bla_{TEM}* (63.8%), and *floR* (55.9%). All *V. cholerae* isolates were susceptible to all antimicrobials tested, while the most common resistance gene was *sul1* (12.0%). One isolate of *A. hydrophila* was positive for *int1*, while all isolates of *Salmonella* and *V. cholerae* isolates were negative for integrons and *int_{SXT}*. None of the bacterial isolates

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in this study were producing ESBL. The occurrence of *mcr-3* (20.0%) in these isolates from tilapia aquaculture may signify a serious occupational and consumer health risk given that colistin is a last resort antimicrobial for treatment of Gram-negative bacteria infections.

Conclusions. Findings from this study on AMR bacteria in hybrid red tilapia suggest that aquaculture as practiced in Thailand can select for ubiquitous AMR pathogens, mobile genetic elements, and an emerging reservoir of *mcr* and colistin-resistant bacteria. Resistant and pathogenic bacteria, such as resistance to ampicillin and tetracycline, or MDR *Salmonella* circulating in aquaculture, together highlight the public health concerns and foodborne risks of zoonotic pathogens in humans from cultured freshwater fish.

Subjects Aquaculture, Fisheries and Fish Science, Microbiology, Molecular Biology

Keywords *Aeromonas hydrophila*, Antimicrobial resistance, Colistin, *Salmonella*, *Vibrio cholerae*, Hybrid red tilapia

INTRODUCTION

Thailand is one of the main producers of global freshwater fish production. Tilapia is highlighted as the main cultivated fish in Thailand with a production of at least 200,000 tons per year (FAO, 2020). The increased demand for tilapia leads to the intensification of fish farming, and this circumstance can make fish more susceptible to bacterial and viral infection. *Aeromonas* spp. and *Vibrio cholerae* are native inhabitants of the aquatic environment. *A. hydrophila* is the main causative agent of motile *Aeromonas* septicemia (MAS), while *V. cholerae* is the normal flora of freshwater fish and is postulated as an opportunistic fish pathogen (Halpern & Izhaki, 2017; Nicholson et al., 2020). These two bacteria play a particular role in foodborne pathogens. *A. hydrophila* has been implicated in foodborne diarrheal outbreaks and the presence of *V. cholerae* in freshwater fish has been associated with cholera in human cases (Liu, Whitehouse & Li, 2018; Elimian et al., 2019). *Salmonella* is one of the top five bacterial pathogens that causes foodborne and waterborne diseases in humans (Lee & Yoon, 2021). Although *Salmonella* is harmless in fish, it has been associated with many foodborne diseases related to freshwater fish (Liu, Whitehouse & Li, 2018). For example, human infection with non-typhoidal *Salmonella* (NTS) associated with contaminated fish can cause gastroenteritis, potential bacteremia and potential life-threatening infections (Akinyemi, Ajoseh & Fakorede, 2021).

Antimicrobial resistance (AMR) is of increasing concern due to its impact on therapeutic options, resulting in failure of treatment and public health impacts. AMR bacteria found in fish and aquatic environments can originate from human activities, land-based agriculture, and aquaculture (Bollache et al., 2019). Resistant bacteria from the environment can contribute to AMR in aquaculture and possibly in humans as an important One Health issue. However, AMR occurrence in fish and the surrounding aquatic environments is poorly studied. Extended-spectrum β -lactamase (ESBL)-producing bacteria are resistant to most penicillins and cephalosporins, and can also be co-resistant to other antimicrobial groups such as fluoroquinolones and colistin (Zhang et al., 2019; Gawish et al., 2021).

Multidrug resistant (MDR) *A. hydrophila*, *Salmonella* spp. and *V. cholerae* isolated from fish have been documented (Saharan, Verma & Singh, 2020), which can elevate the risk of co-infection with MDR and ESBL-producing bacteria. Colistin is a last-line antimicrobial reserved for the treatment of serious gram-negative bacterial infections in humans. In livestock, colistin is used primarily for the treatment of diarrhea and is currently banned in many countries, including Thailand due to the emergence of plasmid-borne colistin resistance genes (*mcr*) reported in meat and poultry products that indicate potential transferable resistance (Pungpian et al., 2021). The *mcr* genes are commonly detected in *Enterobacteriales*, including *E. coli* and *Salmonella*, and are occasionally found in other zoonotic gram-negative bacteria such as *A. hydrophila*, and *Klebsiella pneumonia* (Elbediwi et al., 2019; Luo, Wang & Xiao, 2020). The global dissemination of the *mcr* genes among these pathogens is highly concerning, because they can be potentially transmitted from animals to humans, resulting in unsuccessful gram-negative bacterial infection treatment. Furthermore, the presence of resistance to colistin in fish has been reported (Liu et al., 2020; Saharan, Verma & Singh, 2020), with a recent study indicating that the consumption of fish products was a risk factor for infection by bacteria harboring the *mcr-1* gene among healthy Chinese (Lv et al., 2022).

Quinolones are effective antimicrobials for the treatment of gram-negative bacterial infection. Their analogs have been applied to both humans and aquatic animals. For example, ciprofloxacin is commonly used for the treatment of gastrointestinal and urinary tract infections in humans, while oxolinic acid and enrofloxacin are approved for the treatment of columnaris and MAS in aquatic animals (Baoprasertkul, Somsiri & Boonyawiwat, 2012). Mutations in quinolone resistance determining regions (QRDRs) in the *gyrA* and *parC* regions, and plasmid-encoded quinolone resistance (PMQR) genes can confer quinolone resistance. However, the main mechanism conferred on resistance in aquatic bacteria has not been clearly investigated. Few studies of the phenotype, genotype, and its determinants of AMR related to foodborne bacteria in hybrid red tilapia have been reported in Thailand. Therefore, the aims of this study were to characterize the phenotype and genotype of AMR bacteria, virulence genes, and ESBL production of *A. hydrophila*, *Salmonella* spp., and *V. cholerae* isolated from the surface of fish, intestine, muscle, liver and kidney of hybrid red tilapia and their cultivation water, and to characterize *gyrA* and *parC* in QRDRs of ciprofloxacin-resistant isolates.

MATERIALS & METHODS

Bacterial strains

A total of 278 isolates of *A. hydrophila* ($n = 15$), *Salmonella* ($n = 188$) and *V. cholerae* ($n = 75$) were obtained from an existing bank of bacterial isolates from a previous study on bacterial pathogens from cage cultured tilapia in Thailand (Thaotumpitak et al., 2022), which were stored at the Department of Veterinary Public Health, Faculty of Veterinary Science, Chulalongkorn University (Table 1). In summary, these isolates were collected from healthy fresh tilapia (carcass rinse, intestine, muscle, liver, and kidney) and cultivation water in Kanchanaburi Province from October 2019 to November 2020, with the study

Table 1 Number of bacterial isolates tested in this study ($n = 278$).

Sample type	No. of bacterial isolate (n)		
	<i>A. hydrophila</i>	<i>Salmonella</i>	<i>V. cholerae</i>
Hybrid red tilapia			
Carcass rinse	10	24	10
Intestine	0	57	23
Meat	0	0	2
Liver and kidney	0	1	6
Cultivation water	5	106	34
Total	15	188	75

location and bacterial isolation procedures described previously ([Thaotumpitak et al., 2022](#)). All isolates had been stored in 20% glycerol at -80°C . This study was reviewed and approved by IBC No. 2031027 of the Faculty of Veterinary Science, Chulalongkorn University.

DNA preparation and polymerase chain reaction (PCR)

The DNA template was prepared using the whole cell boiling method ([Lévesque et al., 1995](#)). Briefly, the pure bacterial isolate was streaked onto nutrient agar (Difco, Franklin Lakes, NJ, USA) and incubated at 37°C for 18–24 h. Then a single colony was harvested and transferred to an Eppendorf tube containing 150 μL of rNase-free water. The well-mixed suspension was heated for 10 min and immediately placed on ice. The suspension was centrifuged at 11,000 rpm for 5 min and the supernatant was used as a DNA template.

Bacterial confirmation

Bacterial confirmation was carried out for *A. hydrophila*, *Salmonella*, and *V. cholerae* using genus-specific and species-specific primers listed in [Table 2](#). Confirmation of *A. hydrophila* was carried out by targeting genus-specific and species-specific primers (*aer* and *ah*), respectively. *Salmonella* isolates were confirmed by detection of the invasion gene (*invA*), which also served as the *Salmonella* virulence gene. The *V. cholerae* isolates were confirmed using a pair of species-specific primers (*ompW*) targeting the outer membrane protein. In addition, all *V. cholerae* isolates were serogrouped by slide-agglutination test with polyvalent *V. cholerae* O1, monoclonal *V. cholerae* O139, and monoclonal *V. cholerae* O141 from commercial antiserum (S&A reagents lab, Bangkok, Thailand).

Antimicrobial susceptibility test

The agar dilution method was used to determine antimicrobial susceptibility to twelve antimicrobial agents, including ampicillin, chloramphenicol, ciprofloxacin, enrofloxacin, florfenicol, gentamicin, oxolinic acid, oxytetracycline, streptomycin, sulfamethoxazole, tetracycline, and trimethoprim according to the CLSI standard ([CLSI, 2015](#)). These antimicrobials were chosen from common antimicrobials used in human and aquatic medicine. The clinical breakpoints and epidemiological cutoff values of *A. hydrophila*, *Salmonella*, and *V. cholerae* were determined ([CLSI, 2014](#); [CLSI, 2015](#); [CLSI, 2016](#); [CLSI,](#)

Table 2 Primers used for bacterial confirmation, AMR genotype, resistance determinants, virulence genes, and QRDRs detection.

Gene	Primer	Oligonucleotide sequences (5'–3')	Amplicon size (bp)	Reference
Genus/species confirmation				
<i>A. hydrophila</i>				
<i>aer</i>	<i>aer</i> -F	CTACTTTTGCCGGCGAGCGG	953	<i>Ahmed et al. (2018)</i>
	<i>aer</i> -R	TGATTCCCGAAGGCACTCCC		
<i>ah</i>	<i>ah</i> -F	GAAAGGTTGATGCCTAATACGTA	625	<i>Ahmed et al. (2018)</i>
	<i>ah</i> -R	CGTGCTGGCAACAAAGGACAG		
<i>Salmonella</i>				
<i>invA</i> *	<i>invA</i> -F	GTGAAATTATCGCCACGTTCCGGGCAA	284	<i>Kumar, Datta & Lalitha (2015)</i>
	<i>invA</i> -R	TCATCGCACCGTCAAAGGAACC		
<i>V. cholerae</i>				
<i>ompW</i>	<i>ompW</i> -F	CACCAAGAAGGTGACTTTATTGTG	588	<i>Sathiyamurthy, Baskaran & Subbaraj (2013)</i>
	<i>ompW</i> -R	GAACTTATAACCACCCGCG		
Resistance genes				
<i>bla</i> _{TEM}	<i>bla</i> _{TEM} -F	GCGGAACCCCTATTT	964	<i>Hasman et al. (2005)</i>
	<i>bla</i> _{TEM} -R	TCTAAAGTATATATGAGTAAACTTGGTCTGAC		
<i>bla</i> _{SHV}	<i>bla</i> _{SHV} -F	TTCGCCTGTGTATTATCTCCCTG	854	<i>Hasman et al. (2005)</i>
	<i>bla</i> _{SHV} -R	TTAGCGTTGCCAGTGYTG		
<i>bla</i> _{CTX-M}	<i>bla</i> _{CTX-M} -F	CGATGTGCAGTACCAGTAA	585	<i>Batchelor et al. (2005)</i>
	<i>bla</i> _{CTX-M} -R	AGTGACCAGAATCAGCGG		
<i>bla</i> _{NDM}	<i>bla</i> _{NDM} -F	GGTTTGGCGATCTGGTTTTTC	621	<i>Pungpian et al. (2021)</i>
	<i>bla</i> _{NDM} -R	CGGAATGGCTCATCACGATC		
<i>bla</i> _{PSE}	<i>bla</i> _{PSE} -F	GCAAGTAGGGCAGGCAATCA	422	<i>Chuanchuen et al. (2008)</i>
	<i>bla</i> _{PSE} -R	GAGCTAGATAGATGCTCACAA		
<i>bla</i> _{OXA}	<i>bla</i> _{OXA} -F	ACACAATACATATCAACTTCGC	813	<i>Costa et al. (2006)</i>
	<i>bla</i> _{OXA} -R	AGTGTGTGTTTAGAATGGTGATC		
<i>sul1</i>	<i>sul1</i> -F	CGGCGTGGGCTACCTGAACG	433	<i>Khan et al. (2019)</i>
	<i>sul1</i> -R	GCCGATCGCGTGAAGTTCCG		
<i>sul2</i>	<i>sul2</i> -F	CGGCATCGTCAACATAACCT	721	<i>Khan et al. (2019)</i>
	<i>sul2</i> -R	TGTGCGGATGAAGTCAGCTC		
<i>sul3</i>	<i>sul3</i> -F	CAACGGAAGTGGGCGTTGTGGA	244	<i>Khan et al. (2019)</i>
	<i>sul3</i> -R	GCTGCACCAATTCGCTGAACG		
<i>qnrA</i>	<i>qnrA</i> -F	AGAGGATTTCTCACGCCAGG	580	<i>Cattoir et al. (2007)</i>
	<i>qnrA</i> -R	TGCCAGGCACAGATCTTGAC		
<i>qnrB</i>	<i>qnrB</i> -F	GGMATHGAAATTCGCCACTG	264	<i>Cattoir et al. (2007)</i>
	<i>qnrB</i> -R	TTTGCGYGYCGCCAGTCGAAC		
<i>qnrS</i>	<i>qnrS</i> -F	GCAAGTTCATTGAACAGGGT	428	<i>Cattoir et al. (2007)</i>
	<i>qnrS</i> -R	TCTAAACCGTCGAGTTCGGCG		
<i>ermB</i>	<i>ermB</i> -F	AGACACCTCGTCTAACCTTCGCTC	640	<i>Raissy et al. (2012)</i>
	<i>ermB</i> -R	TCCATGTACTACCATGCCACAGG		

(continued on next page)

Table 2 (continued)

Gene	Primer	Oligonucleotide sequences (5'-3')	Amplicon size (bp)	Reference
<i>dfrA1</i>	<i>dfrA1</i> -F <i>dfrA1</i> -R	GGAGTGCCAAAGGTGAACAGC GAGGCGAAGTCTTGGGTAATAAAC	367	Shahrani, Dehkordi & Momtaz (2014)
<i>dfrA12</i>	<i>dfrA12</i> -F <i>dfrA12</i> -R	TTCGCAGACTCACTGAGGG CGGTTGAGACAAGCTCGAAT	330	Chuanchuen et al. (2008)
<i>catA</i>	<i>catA</i> -F <i>catA</i> -R	CCAGACCGTTCAGCTGGATA CATCAGCACCTTGTGCGCCT	454	Chuanchuen et al. (2008)
<i>catB</i>	<i>catB</i> -F <i>catB</i> -R	CGGATTCAGCCTGACCACC ATACGCGGTCACCTTCCTG	461	Chuanchuen et al. (2008)
<i>cmlA</i>	<i>cmlA</i> -F <i>cmlA</i> -R	TGGACCGCTATCGGACCG CGCAAGACACTTGGGCTGC	641	Chuanchuen et al. (2008)
<i>strA</i>	<i>strA</i> -F <i>strA</i> -R	TGGCAGGAGGAACAGGAGG AGGTCGATCAGACCCGTGC	405	Chuanchuen et al. (2008)
<i>strB</i>	<i>strB</i> -F <i>strB</i> -R	GGCAGCATCAGCCTTATAATTT GTGGATCCGTCATTTCATTGTT	470	Mala et al. (2017)
<i>tetA</i>	<i>tetA</i> -F <i>tetA</i> -R	GGCGGTCTTCTTCATCATGC CGGCAGGCAGAGCAAGTAGA	502	Khan et al. (2019)
<i>tetB</i>	<i>tetB</i> -F <i>tetB</i> -R	CGCCCAGTGCTGTTGTTGTC CGCGTTGAGAAGCTGAGGTG	615	Khan et al. (2019)
<i>tetD</i>	<i>tetD</i> -F <i>tetD</i> -R	AAACCATTACGGCATTCTGC GACCGGATACACCATCCATC	787	Kumai et al. (2005)
<i>addA1</i>	<i>addA1</i> -F <i>addA1</i> -R	CTCCGCAGTGGATGGCGG GATCTGCGCGCGAGGCCA	631	Chuanchuen et al. (2008)
<i>addA2</i>	<i>addA2</i> -F <i>addA2</i> -R	CATTGAGCGCCATCTGGAAT ACATTTCHCTCATCGCCGGC	500	Chuanchuen et al. (2008)
<i>aac(3)IV</i>	<i>aac(3)IV</i> -F <i>aac(3)IV</i> -R	GTGTGCTGCTGGTCCACAGC AGTTGACCCAGGGCTGTCGC	627	Stoll et al. (2012)
<i>aac(6)-Ib</i>	<i>aac(6)-Ib</i> -F <i>aac(6)-Ib</i> -R	TTGCGATGCTCTATGAGTGGCTA CTCGAATGCCTGGCGTGTTC	482	Park et al. (2006)
<i>qepA</i>	<i>qepA</i> -F <i>qepA</i> -R	GCAGGTCCAGCAGCGGGTAG CTTCTGCCCGAGTATCGTG	199	Yamane et al. (2008)
<i>floR</i>	<i>floR</i> -F <i>floR</i> -R	ATGGTGATGCTCGGCGTGGGCCA GCGCCGTTGGCGGTAACAGACACCGTGA	800	Ying et al. (2019)
<i>mcr-1</i>	<i>mcr-1</i> -F <i>mcr-1</i> -R	AGTCCGTTTGTCTTGTGGC AGATCCTTGGTCTCGGCTTG	320	Rebello et al. (2018)
<i>mcr-2</i>	<i>mcr-2</i> -F <i>mcr-2</i> -R	CAAGTGTGTTGGTCCGAGTT TCTAGCCCGACAAGCATAACC	715	Rebello et al. (2018)
<i>mcr-3</i>	<i>mcr-3</i> -F <i>mcr-3</i> -R	AAATAAAAATTGTTCCGCTTATG AATGGAGATCCCCGTTTTT	929	Rebello et al. (2018)
<i>mcr-4</i>	<i>mcr-4</i> -F <i>mcr-4</i> -R	TCACTTTCATCACTGCGTTG TTGGTCCATGACTACCAATG	1116	Rebello et al. (2018)
<i>mcr-5</i>	<i>mcr-5</i> -F <i>mcr-5</i> -R	ATGCGGTTGTCTGCATTIATC TCATTGTGGTTGTCCTTTTCTG	1644	Rebello et al. (2018)

(continued on next page)

Table 2 (continued)

Gene	Primer	Oligonucleotide sequences (5'–3')	Amplicon size (bp)	Reference
Integrations and integrative and conjugative elements				
<i>int1</i>	<i>int1</i> -F	CCTGCACGGTTCGAATG	497	<i>Kitiyodom et al. (2010)</i>
	<i>int1</i> -R	TCGTTTGTTCGCCAGC		
<i>int2</i>	<i>int2</i> -F	GGCAGACAGTTGCAAGACAA	247	<i>Kitiyodom et al. (2010)</i>
	<i>int2</i> -R	AAGCGATTTTCTGCGTGTTT		
<i>int3</i>	<i>int3</i> -F	CCGGTTCAGTCTTTCCTCAA	155	<i>Kitiyodom et al. (2010)</i>
	<i>int3</i> -R	GAGGCGTGTACTTGCCTCAT		
<i>int_{SXT}</i>	<i>int_{SXT}</i> -F	GCTGGATAGGTTAAGGGCGG	592	<i>Kitiyodom et al. (2010)</i>
	<i>int_{SXT}</i> -R	CTCTATGGGCACTGTCCACATTG		
Virulence genes				
<i>A. hydrophila</i>				
<i>aero</i>	<i>aero</i> -F	CACAGCCAATATGTCGGTGAAG	326	<i>Ahmed et al. (2018)</i>
	<i>aero</i> -R	GTCACCTTCTCGCTCAGGC		
<i>hly</i>	<i>hly</i> -F	CTATGAAAAAACTAAAAATAACTG	1500	<i>Ahmed et al. (2018)</i>
	<i>hly</i> -R	CAGTATAAGTGGGAAATGAAAAG		
<i>V. cholerae</i>				
<i>hlyA</i>	<i>hlyA</i> -F	GGCAAACAGCGAAACAAATACC	481	<i>Imani et al. (2013)</i>
	<i>hlyA</i> -R	CTCAGCGGGCTAATACGGTTTA		
<i>ctx</i>	<i>ctx</i> -F	CAGTCAGGTGGTCTTATGCCAAGAGG	167	<i>Wong, You & Chen (2012)</i>
	<i>ctx</i> -R	CCCACTAAGTGGGCACTTCTCAAAC		
<i>tcpA</i>	<i>tcpA</i> -F	CACGATAAGAAAACCGGTCAAGAG	453	<i>Singh, Isac & Colwell (2002)</i>
	<i>tcpA</i> -R	CGAAAGCACCTTCTTTCACGTTG		
QRDR				
<i>gyrA</i>	<i>gyrA</i> -F	GCTGAAGAGCTCCTATCTGG	436	<i>Chuanchuen & Padungtod (2009)</i>
	<i>gyrA</i> -R	GGTCGGCATGACGTCCGG		
<i>parC</i>	<i>parC</i> -F	GTACGTGATCATGGATCGTG	390	<i>Chuanchuen & Padungtod (2009)</i>
	<i>parC</i> -R	TTCCTGCATGGTGCCGTCG		

Notes.*Virulence gene of *Salmonella*.

2020). Reference strains, including *Staphylococcus aureus* ATCC 29213, *Escherichia coli* ATCC 25922, and *Pseudomonas aeruginosa* ATCC 27853 were used for quality control.

Detection of ESBL production

The disc diffusion method was performed according to the CLSI guideline (CLSI, 2015). In the screening test, three disks of ceftazidime (30 µg), cefotaxime (30 µg), and cefpodoxime (10 µg) (Oxoid, Basingstoke, Hampshire, UK) were used to test antimicrobial susceptibility. Isolates that exhibited resistance to at least one of these cephalosporins were further confirmed using a combination disk diffusion method. Ceftazidime (30 µg), cefotaxime (30 µg), and these two disks combined with clavulanic acid were used. The difference in inhibition zone between single cephalosporin and cephalosporins containing clavulanic acid greater than five mm indicates positive ESBL-producing isolates.

AMR gene and virulence-encoded gene detection

All primers for resistance genes, their determinants, and virulence genes are listed in [Table 2](#). Resistance genes were chosen to correspond with AMR phenotypes: *bla*_{TEM}, *bla*_{SHV}, *bla*_{CTX-M} and *bla*_{PSE} corresponding to β -lactam resistance and ESBL production; *bla*_{NDM} and *bla*_{OXA} corresponding to carbapenem resistance; *catA*, *catB*, *floR* and *cmlA* corresponding to phenicol resistance; *ermB* corresponding to erythromycin resistance; *qnrA*, *qnrB*, *qnrS*, *aac(6')-Ib*, and *qepA* corresponding to quinolone resistance; *aadA1*, *aadA2* and *aac(3)IV* corresponding to gentamicin resistance; *tetA*, *tetB* and *tetD* corresponding to tetracycline resistance; *strA* and *strB* corresponding to streptomycin resistance; *sul1*, *sul2*, and *sul3* corresponding to sulfonamide resistance; *dfrA1* and *dfrA12* corresponding to trimethoprim resistance; *mcr-1* to *mcr-5* corresponding to colistin resistance. To confirm the existence of colistin-resistant genes ($n = 3$), sequence-positive *mcr* isolates from a previous study were used as reference strains ([Pungpian et al., 2021](#)). Integrons (*int1*, *int2*, and *int3*) and integrative and conjugative elements (SXT element; *int*_{SXT}) were also detected. For the presence of virulence genes, isolates of *A. hydrophila* (aerolysin gene: *aero*; hemolysin gene: *hly*), *Salmonella* (*invA*: invasive gene) and *V. cholerae* (hemolysin gene: *hlyA*, cholera toxin: *ctx*, and co-regulated toxin: *tcpA*) were examined to determine their virulence factors.

The final volume (50 μ L) of the PCR mixture was prepared according to the manufacturer's instructions. A 5 μ L of template DNA, 25 μ L of TopTaq Master Mix (Qiagen, Hilden, Germany), 2 μ L of each forward and reverse primer, 5 μ L of coralLoad, and 11 μ L of sterile rNase free water were used. PCR amplification was carried out on a Tpersonal combi model (Biometra, Göttingen, Germany). The PCR product was separated on 1.5% (w/v) agarose gel and stained with RedsafeTM nucleic acid staining solution (Intron Biotechnology, Seongnam, Republic of Korea). The results were photographed using the Omega FluorTM gel documentation system (Aplegen, Pleasanton, CA, USA).

Determination of QRDR nucleotide sequences

Salmonella isolates ($n = 11$) were DNA sequenced for detection of the QRDRs. Two target sites at *gyrA* and *parC* were amplified by PCR using the primers listed in [Table 2](#). Eight *Salmonella* isolates were randomly selected from the intestinal tract ($n = 4$), cultivation water ($n = 2$), carcass rinses ($n = 2$) based on the resistance gradient of ciprofloxacin, and susceptible isolates to ciprofloxacin ($n = 3$) were used as a negative control. The *gyrA* and *parC* were submitted for purification and nucleotide sequencing (Bionics Co., LTD., Gyeonggi-Do, Republic of Korea). The identity of amino acid sequences were analyzed using Molecular Evolutionary Genetic Analysis (MEGA) software version 11 ([Tamura, Stecher & Kumar, 2021](#)). Reference sequences (accession number [NC_003197:2](#)) from the GenBank database are available at the National Centre for Biotechnology Information (NCBI) (<https://www.ncbi.nlm.nih.gov>).

Statistical analysis

Descriptive statistics were used to characterize the prevalence of AMR, MDR, virulence genes, integrons, SXT element, ESBL production, and QRDR mutations from *A. hydrophila*,

Salmonella, and *V. cholerae* isolates. Logistic regression was used to determine the association between resistance and its determinants, virulence factors, and ESBL production (OR >1: positive association; OR <1: negative association). Two-sided hypothesis testing, with $p < 0.05$ were considered statistically significant. All statistical analyzes were performed with Stata version 14.0 (StataCorp, College Station, TX, USA).

RESULTS

Phenotype and genotype of AMR and virulence genes of *A. hydrophila*

A. hydrophila was only detected in carcass rinses and cultivation water (Table 1). All isolates ($n = 15$) were resistant to ampicillin (Table 3), with four isolates (26.7%) resistant to oxytetracycline, tetracycline, and trimethoprim; three isolates (20.0%) were resistant to oxolinic acid. Resistance to at least one antimicrobial and MDR in *A. hydrophila* were 100% and 26.7%, respectively. Among the six AMR patterns, AMP alone (53.3%) was the most common pattern, with AMP-OTC-TET-TRI and AMP-OXO found in two isolates (13.3%) (Table 4). The most prevalent resistance gene found in *A. hydrophila* was *mcr-3* (20.0%), followed by 13.3% of isolates having *floR*, *qnrS*, *sul1*, *sul2*, and *dfrA1* (Table 3). The *mcr-3* was only detected in fish carcass rinses, and only one isolate (6.7%) was positive for *int1*. The presence of *aero* and *hly* was observed in all isolates of *A. hydrophila* (100.0%) (Table 3).

Phenotype and genotype of AMR, and virulence genes of *Salmonella* spp.

All *Salmonella* isolates ($n = 188$) were resistant to at least one antimicrobial (Table 5). There was a high prevalence of resistance to ampicillin (79.3%), oxolinic acid (75.5%), oxytetracycline (71.8%) and tetracycline (70.7%). All *Salmonella* isolates were sensitive to gentamicin (100%) and only two isolates (1.1%) were resistant to trimethoprim. More than 70% of *Salmonella* isolates exhibited MDR: of the 34 unique resistance patterns, the two most common were AMP-CHP-FFC-OTC-OXO-TET (20.7%) and AMP-CHP-ENR-FFC-OTC-OXO-TET (18.6%) (Table 6). Most *Salmonella* harbored *qnrS* (65.4%), followed by *tetA* (64.9%), *bla*_{TEM} (63.8%), and *floR* (55.9%) (Table 5). None of the colistin resistance genes was observed in the *Salmonella* isolates. In general, cultivation water exhibited higher resistance levels than any other types of fish organ or tissue sample. Based on the type of fish sample, *Salmonella* isolated from the intestine had a higher prevalence of the AMR phenotype and genotype than the fish carcass rinses. The *invA* gene was detected in all *Salmonella* isolates.

Phenotype and genotype of AMR and virulence genes of *V. cholerae*

All *V. cholerae* isolates ($n = 75$) were non-agglutinating *Vibrios* (NAGs) with O1 and O139, and non-agglutination with O141. All *V. cholerae* isolates (100%) were susceptible to the 12 antimicrobials tested. The predominant AMR gene in *V. cholerae* was *sul1* (12.0%) (Table 7). The genes *catB*, *qnrS*, *tetA*, *tetB*, *strA*, and *dfrA1* were all present at a 4.0% prevalence. Colistin resistance genes, integrons, and SXT element were not found in any *V. cholerae* isolates in this study, in contrast, all isolates had detectable *hlyA*.

Table 3 Phenotypic and genotypic resistance and virulence genes of *A. hydrophila* ($n = 15$) isolated from hybrid red tilapia carcass rinses and cultivation water.

Variable	Resistance (%)		Grand total ($n = 15$)
	Carcass rinse ($n = 10$)	Cultivation water ($n = 5$)	
Antimicrobials			
Ampicillin	10 (100.0)	5 (100.0)	15 (100.0)
Cefotaxime	0 (0)	0 (0)	0 (0)
Cefpodoxime	0 (0)	0 (0)	0 (0)
Ceftazidime	0 (0)	0 (0)	0 (0)
Chloramphenicol	0 (0)	0 (0)	0 (0)
Ciprofloxacin	0 (0)	0 (0)	0 (0)
Enrofloxacin	0 (0)	0 (0)	0 (0)
Florfenicol	0 (0)	0 (0)	0 (0)
Gentamicin	0 (0)	0 (0)	0 (0)
Oxolinic acid	3 (30.0)	0 (0)	3 (20.0)
Oxytetracycline	3 (30.0)	1 (20.0)	4 (26.7)
Streptomycin	0 (0)	0 (0)	0 (0)
Sulfamethoxazole	0 (0)	0 (0)	0 (0)
Tetracycline	3 (30.0)	1 (20.0)	4 (26.7)
Trimethoprim	3 (30.0)	1 (20.0)	4 (26.7)
MDR	3 (30.0)	1 (20.0)	4 (26.7)
AMR genes			
<i>floR</i>	2 (20.0)	0 (0)	2 (13.3)
<i>qnrS</i>	2 (20.0)	0 (0)	2 (13.3)
<i>sul1</i>	2 (20.0)	0 (0)	2 (13.3)
<i>sul2</i>	2 (20.0)	0 (0)	2 (13.3)
<i>dfrA1</i>	2 (20.0)	0 (0)	2 (13.3)
<i>mcr-3</i>	3 (30.0)	0 (0)	3 (20.0)
Integrans			
<i>int1</i>	1 (10.0)	0 (0)	1 (6.7)
Virulence genes			
<i>aero</i>	10 (100.0)	5 (100.0)	15 (100.0)
<i>hly</i>	10 (100.0)	5 (100.0)	15 (100.0)

Notes.

The table presented for positive isolates, which were only detected in carcass rinses and cultivation water. Non-detected genes in *A. hydrophila* were: *bla*_{TEM}, *bla*_{SHV}, *bla*_{CTX-M}, *bla*_{NDM}, *bla*_{PSE}, *bla*_{OXA}, *sul3*, *qnrA*, *qnrB*, *ermB*, *dfrA12*, *catA*, *catB*, *cmlA*, *strA*, *strB*, *tetA*, *tetB*, *tetD*, *addA1*, *addA2*, *aac(3)IV*, *aac(6)-Ib*, *qepA*, *mcr-1*, *mcr-2*, *mcr-4*, *mcr-5*, *int2*, *int3*, and *int*_{SXT}.

ESBL production in *A. hydrophila*, *Salmonella*, and *V. cholerae*

ESBL production was not detected in any of the bacterial strains from tilapia aquaculture or the fish tissues or organs ($n = 278$).

Association between phenotypic and genotypic AMR

Logistic regression analyses indicated there were statistically significant associations between phenotypic and genotypic resistance of tetracycline and the presence of *tetA* (O.R. = 259.0, CI [52.3–1283.6], $p < 0.0001$), sulfamethoxazole resistance and the presence of *sul2*

Table 4 Resistance patterns of *A. hydrophila* isolated from hybrid red tilapia carcass rinses and cultivation water ($n = 15$).

AMR pattern	Resistance (%)		Grand total ($n = 15$)
	Carcass rinses ($n = 10$)	Cultivation water ($n = 5$)	
AMP	4 (40.0)	4 (80.0)	8 (53.3)
AMP-OTC-TET-TRI	1 (10.0)	1 (20.0)	2 (13.3)
AMP-OTC-TRI	1 (10.0)	0 (0)	1 (6.7)
AMP-OTC-OXO-TET	1 (10.0)	0 (0)	1 (6.7)
AMP-OXO	2 (20.0)	0 (0)	2 (13.3)
AMP-TRI	1 (10.0)	0 (0)	1 (6.7)

Notes.

AMP, ampicillin; OTC, oxytetracycline; OXO, oxolinic acid; TET, tetracycline; TRI, trimethoprim.

(O.R. = 90.3, CI [5.6–1455.1], $p = 0.001$), and streptomycin resistance and the presence of *strA* (O.R. = 273.0, CI [1.8–42,372.4], $p = 0.029$).

Mutation of QRDRs

Eight isolates from the 40 *Salmonella* isolates that were resistant to ciprofloxacin were randomly selected for sequence analysis and QRDR determination. The serovars of these eight isolates were Virchow ($n = 1$), Saintpaul ($n = 5$), Neukoelln ($n = 1$) and Chartes ($n = 1$) (Table 8). Six of the eight ciprofloxacin-resistant isolates had a point mutation in *gyrA* from C to A at position 248 (Ser83Tyr); the other two resistant isolates did not have *gyrA* mutations. The *gyrA* sequences from the eight *Salmonella* isolates have been deposited at GenBank under accession numbers [OP831158–OP831165](#). All isolates carried *qnrS* without any mutations in the *parC* region.

DISCUSSION**AMR and virulence genes of *A. hydrophila***

The low prevalence of *A. hydrophila* observed in this study was consistent with previous work which found a low prevalence of 2.0% and 2.7% in the internal organs and fish muscle, respectively (Ahmed et al., 2018). This low prevalence is likely due to the samples being collected from clinically healthy hybrid red tilapia. Furthermore, fish liver is typically the main target organ for colonization by *A. hydrophila*; however, none of the liver samples in this study were positive for this bacterium (Algammal et al., 2020). In this study, all isolates (100%) were resistant to ampicillin, which was consistent with previous studies in tilapia and other aquatic animals due to their intrinsic resistance (El-ghareeb, Zahran & Abd-Elghany, 2019). More than a fourth of the *A. hydrophila* isolates (26.7%) were resistant to oxytetracycline, tetracycline, and trimethoprim, which was also consistent with previous studies (El-ghareeb, Zahran & Abd-Elghany, 2019; Yu et al., 2021). The wide range resistance of *A. hydrophila* observed in this study could result from these bacteria persisting in high concentrations of antimicrobials leading to up-regulation of the efflux pump and the production of antioxidative agents (Yu et al., 2021).

Oxytetracycline and oxolinic acid are broad-spectrum antimicrobials belonging to tetracyclines and fluoroquinolones, which have been approved for use in aquatic animals

Table 5 Phenotypic and genotypic resistance and virulence genes of *Salmonella* isolates ($n = 188$) from hybrid red tilapia and cultivation water.

Variable	Resistance (%)				
	Carcass rinse ($n = 24$)	Intestine ($n = 57$)	Liver and kidney ($n = 1$)	Cultivation water ($n = 106$)	Grand total ($n = 188$)
Antimicrobials					
Ampicillin	13 (54.2)	38 (66.7)	1 (100.0)	97 (91.5)	149 (79.3)
Cefotaxime	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)
Cefpodoxime	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)
Ceftazidime	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)
Chloramphenicol	11 (45.8)	28 (49.1)	1 (100.0)	78 (73.6)	118 (62.8)
Ciprofloxacin	3 (12.5)	14 (24.6)	0 (0)	23 (21.7)	40 (21.3)
Enrofloxacin	6 (25.0)	20 (35.1)	0 (0)	43 (40.6)	69 (36.7)
Florfenicol	12 (50.0)	22 (38.6)	1 (100.0)	69 (65.1)	104 (55.3)
Gentamicin	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)
Oxolinic acid	16 (66.7)	40 (70.2)	0 (0)	86 (81.1)	142 (75.5)
Oxytetracycline	13 (54.2)	34 (59.6)	0 (0)	88 (83.0)	135 (71.8)
Streptomycin	0 (0)	2 (3.5)	0 (0)	2 (1.9)	4 (2.1)
Sulfamethoxazole	0 (0)	1 (1.8)	0 (0)	3 (2.8)	4 (2.1)
Tetracycline	13 (54.2)	32 (56.1)	0 (0)	88 (83.0)	133 (70.7)
Trimethoprim	0 (0)	0 (0)	0 (0)	2 (1.9)	2 (1.1)
MDR	13 (54.2)	34 (59.6)	0 (0)	89 (84.0)	136 (72.3)
AMR genes					
<i>bla</i> _{TEM}	10 (41.7)	34 (59.6)	1 (100.0)	75 (70.8)	120 (63.8)
<i>floR</i>	10 (41.7)	24 (42.1)	0 (0)	71 (67.0)	105 (55.9)
<i>qnrS</i>	13 (54.2)	31 (54.4)	0 (0)	79 (74.5)	123 (65.4)
<i>tetA</i>	13 (54.2)	31 (54.4)	0 (0)	78 (73.6)	122 (64.9)
<i>tetB</i>	0 (0)	1 (1.8)	0 (0)	2 (1.9)	3 (1.6)
<i>strA</i>	0 (0)	1 (1.8)	0 (0)	2 (1.9)	3 (1.6)
<i>sul1</i>	0 (0)	0 (0)	0 (0)	2 (1.9)	2 (1.1)
<i>sul2</i>	0 (0)	1 (1.8)	0 (0)	2 (1.9)	3 (1.6)
Virulence genes					
<i>invA</i>	24 (100.0)	57 (100.0)	1 (100.0)	106 (100.0)	188 (100.0)

Notes.

This table presented only positive isolates from various tilapia samples except fish muscle, which were negative for *Salmonella*. Non-detected genes in *Salmonella* were: *bla*_{SHV}, *bla*_{CTX-M}, *bla*_{NDM}, *bla*_{PSE}, *bla*_{OXA}, *sul3*, *qnrA*, *qnrB*, *ermB*, *dfrA1*, *dfrA12*, *catA*, *catB*, *cmlA*, *strB*, *tetD*, *addA1*, *addA2*, *aac(3)IV*, *aac(6')-Ib*, *qepA*, *floR*, *mcr-1*, *mcr-2*, *mcr-3*, *mcr-4*, *mcr-5*, *int1*, *int2*, *int3*, and *int*_{SXT}.

(OIE, 2018). These antimicrobials were the priority antibiotics in use when outbreaks occurred on the hybrid red tilapia farms enrolled in this study, possibly leading to widespread selection of resistant *A. hydrophila*. A previous study found that *A. hydrophila* can harbor various *tet* genes and transfer resistance to tetracycline to *E. coli* (Harnisz, Korzeniewska & Golaś, 2015). Therefore, *A. hydrophila* might serve as an important reservoir of resistance to tetracycline in the aquatic environment. Regarding resistance in the *A. hydrophila* isolates, all isolates resistant to oxolinic acid were susceptible to both

Table 6 Resistance patterns of *Salmonella* isolates ($n = 188$) from hybrid red tilapia carcass rinses, intestine, liver and kidney, and cultivation water.

AMR pattern	No of isolates (%)				
	Fish carcass rinse ($n = 24$)	Intestine ($n = 57$)	Liver and kidney ($n = 1$)	Cultivation water ($n = 106$)	Total ($n = 188$)
Susceptible	8 (33.3)	11 (19.3)	0 (0)	4 (3.8)	23 (12.2)
AMP-CHP-CIP-ENR-FFC-OTC-OXO-TET	1 (4.2)	2 (3.5)	0 (0)	1 (0.9)	4 (2.1)
AMP-CHP-CIP-ENR-OTC-OXO-TET	0 (0)	4 (7.0)	0 (0)	6 (5.7)	10 (5.3)
AMP-CHP-CIP-FFC-OTC-OXO-TET	0 (0)	0 (0)	0 (0)	3 (2.8)	3 (1.6)
AMP-CHP-CIP-OTC-OXO-TET	0 (0)	0 (0)	0 (0)	2 (1.9)	2 (1.1)
AMP-CHP-ENR-FFC-OTC-OXO-STR-TET	0 (0)	1 (1.8)	0 (0)	0 (0)	1 (0.5)
AMP-CHP-ENR-FFC-OTC-OXO-SMZ-TET	0 (0)	0 (0)	0 (0)	1 (0.9)	1 (0.5)
AMP-CHP-ENR-FFC-OTC-OXO-TET	5 (20.8)	5 (8.8)	0 (0)	25 (23.6)	35 (18.6)
AMP-CHP-FFC	0 (0)	3 (5.3)	1 (100.0)	8 (7.5)	12 (6.4)
AMP-CHP-FFC-OTC	0 (0)	0 (0)	0 (0)	1 (0.9)	1 (0.5)
AMP-CHP-FFC-OTC-OXO	0 (0)	1 (1.8)	0 (0)	0 (0)	1 (0.5)
AMP-CHP-FFC-OTC-OXO-TET	5 (20.8)	7 (12.3)	0 (0)	27 (25.5)	39 (20.7)
AMP-CHP-FFC-OXO-STR	0 (0)	1 (1.8)	0 (0)	0 (0)	1 (0.5)
AMP-CHP-FFC-TET	0 (0)	0 (0)	0 (0)	3 (2.8)	3 (1.6)
AMP-CHP-OTC-OXO-TET	0 (0)	1 (1.8)	0 (0)	1 (0.9)	2 (1.1)
AMP-CHP-OXO	0 (0)	1 (1.8)	0 (0)	0 (0)	1 (0.5)
AMP-CHP-OXO-TET	0 (0)	1 (1.8)	0 (0)	0 (0)	1 (0.5)
AMP-CIP-ENR-OTC-OXO-SMZ-TET	0 (0)	1 (1.8)	0 (0)	0 (0)	1 (0.5)
AMP-CIP-ENR-OTC-OXO-TET	0 (0)	6 (10.5)	0 (0)	8 (7.5)	14 (7.4)
AMP-CIP-ENR-OTC-OXO-TET-TRI	0 (0)	0 (0)	0 (0)	1 (0.9)	1 (0.5)
AMP-CIP-FFC-OTC-OXO-TET	1 (4.2)	0 (0)	0 (0)	0 (0)	1 (0.5)
AMP-CIP-OTC-OXO-TET	1 (4.2)	1 (1.8)	0 (0)	1 (0.9)	3 (1.6)
AMP-CIP-TET	0 (0)	0 (0)	0 (0)	1 (0.9)	1 (0.5)
AMP-ENR-OTC-OXO-TET	0 (0)	1 (1.8)	0 (0)	0 (0)	1 (0.5)
AMP-FFC-OTC-OXO-TET	0 (0)	1 (1.8)	0 (0)	0 (0)	1 (0.5)
AMP-OTC-OXO-SMZ-STR-TET	0 (0)	0 (0)	0 (0)	1 (0.9)	1 (0.5)
AMP-OTC-OXO-TET-TRI	0 (0)	0 (0)	0 (0)	1 (0.9)	1 (0.5)
AMP-OTC-SMZ-STR-TET	0 (0)	0 (0)	0 (0)	1 (0.9)	1 (0.5)
AMP-OTC-OXO-TET	0 (0)	0 (0)	0 (0)	5 (4.7)	5 (2.7)
AMP-OTC-TET	0 (0)	1 (1.8)	0 (0)	0 (0)	1 (0.5)
CHP-FFC	0 (0)	1 (1.8)	0 (0)	0 (0)	1 (0.5)
ENR-OTC-OXO	0 (0)	0 (0)	0 (0)	1 (0.9)	1 (0.5)
OTC	0 (0)	1 (1.8)	0 (0)	2 (1.9)	3 (1.6)
OTC-OXO	0 (0)	1 (1.8)	0 (0)	0 (0)	1 (0.5)
OXO	3 (12.5)	5 (8.8)	0 (0)	2 (1.9)	10 (5.3)

Notes.

This table presented only positive isolates from various tilapia samples except fish muscle, which were negative for *Salmonella*.

AMP, ampicillin; CHP, chloramphenicol; CIP, ciprofloxacin; ENR, enrofloxacin; FFC, florfenicol; OTC, oxytetracycline; OXO, oxolinic acid; STR, streptomycin; SMZ, sulfamethoxazole; TET, tetracycline; TRI, trimethoprim.

Table 7 Genotypic resistance and virulence genes of *V. cholerae* isolates ($n = 75$) of hybrid red tilapia and cultivation water.

Genotype	Prevalence (%)					
	Fish carcass rinse ($n = 10$)	Intestine ($n = 23$)	Meat ($n = 2$)	Liver and kidney ($n = 6$)	Cultivation water ($n = 34$)	Total ($n = 75$)
AMR genes						
<i>catB</i>	1 (10.0)	0 (0)	0 (0)	0 (0)	2 (5.9)	3 (4.0)
<i>qnrS</i>	1 (10.0)	0 (0)	0 (0)	0 (0)	2 (5.9)	3 (4.0)
<i>tetA</i>	1 (10.0)	0 (0)	0 (0)	0 (0)	2 (5.9)	3 (4.0)
<i>tetB</i>	0 (0)	1 (4.3)	0 (0)	0 (0)	2 (5.9)	3 (4.0)
<i>strA</i>	0 (0)	1 (4.3)	0 (0)	0 (0)	2 (5.9)	3 (4.0)
<i>sul1</i>	1 (10.0)	0 (0)	0 (0)	0 (0)	8 (23.5)	9 (12.0)
<i>dfrA1</i>	1 (10.0)	0 (0)	0 (0)	0 (0)	2 (5.9)	3 (4.0)
Virulence genes						
<i>hlyA</i>	10 (100.0)	23 (100.0)	2 (100.0)	6 (100.0)	34 (100.0)	75 (100.0)
<i>ctx</i>	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)
<i>tcpA</i>	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)

Notes.

This table showed only positive isolates. Non-detected genes in *V. cholerae* were: *bla*_{TEM}, *bla*_{SHV}, *bla*_{CTX-M}, *bla*_{NDM}, *bla*_{PSE}, *bla*_{OXA}, *sul2*, *sul3*, *qnrA*, *qnrB*, *ermB*, *dfrA12*, *catA*, *cmlA*, *strB*, *tetD*, *addA1*, *addA2*, *aac(3)IV*, *aac(6')-Ib*, *qepA*, *floR*, *mcr-1*, *mcr-2*, *mcr-3*, *mcr-4*, *mcr-5*, *int1*, *int2*, *int3*, and *intSXT*.

Table 8 Mutations of *gyrA* in QRDRs in ciprofloxacin-resistant *Salmonella* isolates ($n = 8$).

Sample type	Serovar (n)	<i>gyrA</i> mutation	PMQR	MIC ($\mu\text{g/mL}$)		
				CIP	ENR	OXO
Carcass rinse	Virchow (1)	–	<i>qnrS</i>	4	4	8
	Saintpaul (1)	C248A	<i>qnrS</i>	4	2	16
Intestine	Saintpaul (4)	C248A	<i>qnrS</i>	4–8	2–8	4–32
Cultivation water	Neukoelln (1)	–	<i>qnrS</i>	4	2	4
	Chartes (1)	C248A	<i>qnrS</i>	>32	>64	>128

Notes.

CIP, ciprofloxacin; ENR, enrofloxacin; OXO, oxolinic acid.

ciprofloxacin and enrofloxacin, although these antimicrobials were grouped in similar quinolone derivatives. Furthermore, a study of *Aeromonas* isolated from diseased hybrid red tilapia in Thailand also showed a difference in the resistant phenotype (Mursalim et al., 2022). The explanation for this discrimination could be that different generations conferred distinct antimicrobial potency (Sinwat et al., 2018; Pham, Ziora & Blaskovich, 2019). In detail, ciprofloxacin and enrofloxacin are second-generation quinolones, therefore, they have broader bactericidal activities against Gram-negative bacteria compared to oxolinic acid, which is a first-generation quinolone. To quantify the accurate antimicrobial susceptibility testing in bacteria isolated from aquatic animals, oxolinic acid should be taken into account as a mandatory antimicrobial test regardless of whether other quinolones were selected.

In Thailand, colistin has been banned in food-producing animals as a feed additive since 2017; however, therapeutic use of colistin is still being reported in pig farms (Olaitan et al., 2021; Rueanghiran et al., 2022) and may select for colistin-resistance in these livestock

facilities. The dissemination of colistin-resistant enteric bacteria from livestock into the environment could accelerate the horizontal gene transfer of mobile genetic elements to autochthonous aquatic bacteria (Elbediwi et al., 2019). The resistance of colistin and *mcr* genes in *Aeromonas* was previously reported in healthy freshwater fish and cultivation water in China (Li et al., 2022). This study confirmed that colistin resistant *A. hydrophila* isolates already existed in hybrid red tilapia aquaculture. To our knowledge, this finding is the first report on the occurrence of *mcr-3* in freshwater fish originating in Thailand. The *mcr-3* had been previously detected in *E. coli* and *Salmonella* in pigs and pork (Pungpian et al., 2021), with the global spread of *mcr-1*, *mcr-3* and *mcr-4* is shown in Fig. 1.

In this study, all *A. hydrophila* isolates contained *aero* and *hly* genes, which were more prevalent than a previous study of freshwater fish in Egypt (Ahmed et al., 2018). These virulence genes found in *A. hydrophila* were also reported in an outbreak of diarrheal patients in Brazil (Silva et al., 2017). These findings suggest that *A. hydrophila* isolated from hybrid red tilapia and cultivation water can be both resistant and pathogenic which elevates the public health and food safety risk from these bacteria.

AMR and virulence genes of *Salmonella*

Isolates of *Salmonella* from tilapia and cultivation water exhibited a high prevalence to ampicillin (79.3%), which may be due to part to intrinsic resistance to penicillins by *Salmonella*. In addition, amoxicillin has been approved for off-label use to treat streptococcosis in tilapia aquaculture by the Thai Food and Drug Administration (Baoprasertkul, Somsiri & Boonyawiwat, 2012). The extensive use of amoxicillin in Thai aquaculture can select for β -lactam-resistant bacteria that then colonize tilapia and the immediate aquaculture environment. This study also found high *Salmonella* resistance to oxolinic acid and oxytetracycline. The improper use of oxolinic acid and oxytetracycline can select for the development of AMR and MDR *Salmonella*. In particular, high resistance to chloramphenicol (62.8%) and florfenicol (55.3%) was observed, although antimicrobials in the amphenicol groups have not been approved for use in aquaculture in Thailand. Among this collection of isolates, resistance to gentamicin and trimethoprim were not observed which was similar to a previous study in Kenya (Wanja et al., 2020). Moreover, the high prevalence (>75%) of MDR *Salmonella* isolates is consistent with previous studies in freshwater fish (Saharan, Verma & Singh, 2020; Gawish et al., 2021).

Regarding genotypic AMR, the predominant resistance genes were *qnrS* (65.4%) which was similar in tilapia and catfish in Egypt (Algammal et al., 2022), and *tetA* (64.9%) and *floR* (55.9%) which was also observed in a previous study on farmed fish in Brazil (Ferreira et al., 2021). These phenotypic and genotypic AMR findings indicate a likelihood of transmission of AMR *Salmonella* by consumption of contaminated fish products.

AMR and virulence genes of *V. cholerae*

Human infection with *V. cholerae* is a major public concern because this bacterium is a causative agent of cholera, a severe fatal diarrhea caused by the cholera toxin. Serogroups O1 and O139 have been widely reported in cholera outbreaks with high mortality in humans worldwide. On the contrary, serogroup O141 caused sporadic outbreaks of cholera-like

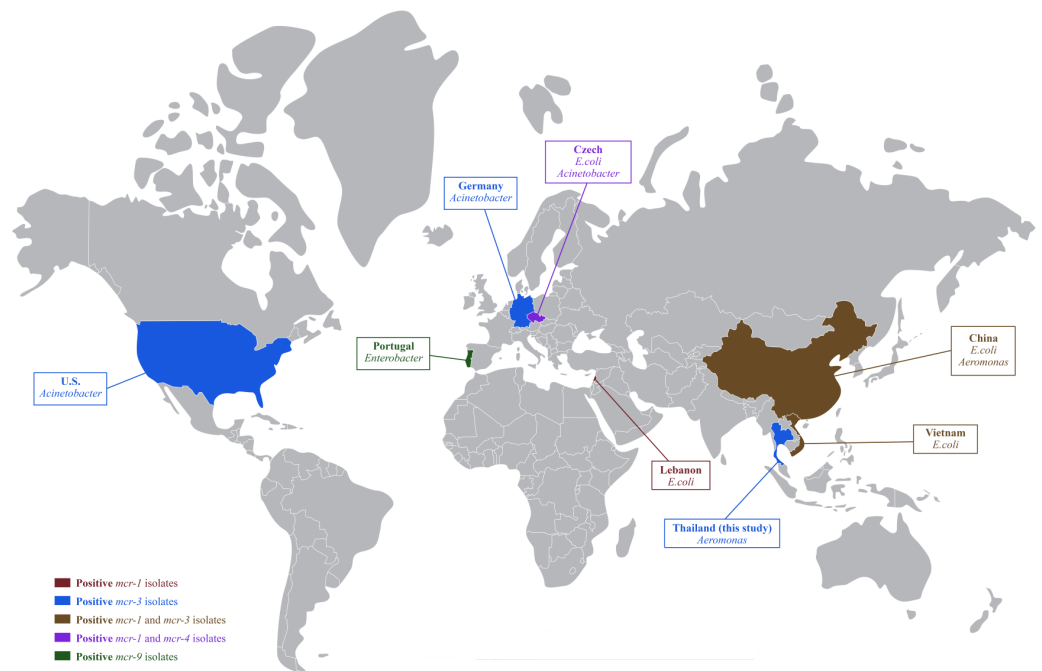


Figure 1 Global distribution of colistin resistance genes in bacteria isolated from fish.

Full-size DOI: [10.7717/peerj.14896/fig-1](https://doi.org/10.7717/peerj.14896/fig-1)

diarrhea (*Elimian et al., 2019*). All *V. cholerae* isolates found in this study belonged to non-O1/O139 and non-O141 serogroups, which is consistent with a low prevalence of these serogroups in fish and aquatic environments and in contrast to their high prevalence in human cases (*Halpern & Izhaki, 2017; Schwartz et al., 2019*). However, the infection of indigenous non-O1/O139 and non-O141 *V. cholerae* originating from aquaculture products can cause watery-to-severe diarrhea in humans due to other virulence factors (*Vezzulli et al., 2020*).

Among the *V. cholerae* isolates in this study, none of them was phenotypically resistant to antimicrobials. This finding is consistent with a previous study that found a low prevalence of AMR in non-cholera environmental strains (*Bier et al., 2015*). In contrast, a study in China found that over half of *V. cholerae* (57.6%) isolated from freshwater fish ($n = 370$) were MDR to streptomycin, ampicillin, and rifampin (*Fu et al., 2020*). Therefore, it is postulated that resistance to non-used antimicrobials in aquaculture may result from the distribution of AMR bacteria from the anthropogenic sources that then contaminate the aquaculture environment. For example, wastewater and effluent from human communities could be an important contributor to AMR in aquaculture. The most prevalent AMR genotype of *V. cholerae* in this study was *sul1* (12.0%), which encoded resistance to sulfamethoxazole. This finding could be problematic for human clinical treatment options because sulfamethoxazole is a drug of choice for the treatment of cholera (*Leibovici-Weissman et al., 2014*). In particular, a recent report showed that *V. parahaemolyticus* isolated from shrimp in China contained the *mcr-1* gene (*Lei et al.,*

2019). These findings highlight the need to monitor these bacteria in aquaculture for the emergence of colistin-resistant genes, especially in non-*Enterobacterale*.

Among the virulence genes examined in this study, *hlyA*, *ctx*, and *tcpA*, the *hlyA* genes were the most prevalent (100.0%), which was consistent with a previous study regarding *V. cholerae* isolated from fish, shellfish, and environmental samples (Shan et al., 2022). The *hlyA* encoding pore-forming toxin can cause cytotoxicity and cell vacuolation of intestinal cells leading to fluid leakage (Ramamurthy et al., 2020). The common prevalence of the *hlyA* gene was reported in clinical or epidemic strains of *V. cholerae* isolates, indicating its likely role in the pathogenesis of many cases of *V. cholerae* (Imani et al., 2013). No strains carrying *ctx* or *tcpA* were detected among *V. cholerae* isolates in this study. The *ctx* gene that encodes cholera enterotoxin leads to massive secretion of electrolytes and water into the intestinal lumen leading to severe fluid loss, while the *tcpA* gene acts as a promoter of pilus formation inducing bacterial colonization in the host's intestine (Ramamurthy et al., 2020). This indicated that the *V. cholerae* in this study were not cholera-causing strains; nonetheless, human infection with these *V. cholerae* strains can pose a risk of diarrhea due to the presence of *hlyA*.

ESBL production in *A. hydrophila*, *Salmonella*, and *V. cholerae*

In this study, no ESBL-producing pathogenic bacteria were detected in hybrid red tilapia and cultivation water. However, the increasing prevalence of ESBL-producing bacteria has been reported in environmental water, livestock animals and humans in Thailand (Runcharoen et al., 2017; Lay et al., 2020). The finding of ESBL-producing bacteria in aquaculture environments was speculated to be due to bacterial contamination of natural water resources used for aquatic animal cultivation. Several studies have reported that ESBL-producing bacteria are presented in many aquatic animals, such as tilapia, catfish, and shrimp (Hon et al., 2016; Gawish et al., 2021). Importantly, bacteria harboring ESBL genes can co-select other plasmid-mediated AMR, such as sulfamethoxazole/trimethoprim, quinolones, and colistin, which are critically important antimicrobials. Infection of these co-resistant bacteria can hamper disease treatment due to the limitation of available therapeutic options, resulting in longer hospital stays and increased antibiotic costs. The surveillance of ESBL-producing bacteria in hybrid red tilapia helps monitor and quantify the possible novel risk of ESBL transmission through consumption of cultured tilapia.

Association between phenotypic and genotypic AMR

In general, *tetA* was the predominant genotypic resistance observed in *Salmonella* and *V. cholerae* isolates. This gene can confer high phenotypic resistance to tetracycline, which was consistent with the statistical association between phenotypic and genotypic resistance to tetracycline based on the logistic regression analyses in this study. Oxytetracycline is a common antimicrobial used in tilapia farms in Thailand due to its effective treatment of aeromoniasis and francisellosis, which are endemic in Thailand (Baoprasertkul, Somsiri & Boonyawiwat, 2012). Previous study also observed a high prevalence of *tetA* in freshwater fish and water, and other *tet* genes, such as *tetL*, *tetO*, and *tetW* were also previously reported in aquaculture (Harnisz, Korzeniewska & Golaś, 2015). The statistical association

between resistance to sulfamethoxazole and the presence of *sul2*, and streptomycin and the presence of *strA* indicated that phenotypic resistance is strongly associated with the presence of the corresponding resistance genes. Other pairs of AMR phenotype-genotype did not observe statistical association. This may be due to non-examined genes or other mechanisms conferred to those resistance.

Mutation of QRDRs

Of eight ciprofloxacin resistant *Salmonella*, only a single-point mutation in *gyrA* (Ser83Tyr) was observed in six isolates. In *Salmonella*, mutations in *gyrA* of QRDRs are a substantial mechanism of resistance to quinolones, while *parC* mutations are rare (Shaheen et al., 2021). The amino acid change in *gyrA* was reported in *Salmonella* isolated from chicken, pork, and clinical isolates of humans (Sinwat et al., 2018). This study was in agreement with the frequent identification of *gyrA* mutations in environmental *Enterobacteriales* in the aquatic environment, including *Salmonella* (Johnning et al., 2015). The *qnrS*, which is the PMQR gene presented in all isolates, can mediate low resistance to quinolone and enhance resistance mediated by *gyrA* mutations. Higher MICs were mostly observed in mutant isolates. However, three isolates were resistant to ciprofloxacin and oxolinic acid, but susceptible to enrofloxacin (Table 8). The inconsistency of resistance to quinolone has previously reported and is still not conclusive (Sinwat et al., 2018). Two ciprofloxacin-resistant *Salmonella* isolates were not mutants in the *gyrA* and *parC* regions. Other genes that confer resistance to quinolones, including *aac(6′)-Ib*, the aminoglycoside acetyltransferase variant gene, and *qepA*, the plasmid-mediated nonspecific efflux pump gene, were absent from these eight ciprofloxacin-resistant *Salmonella*. These results imply that the mutations in *gyrA* and *qnrS* expression were pivotal factors driving resistance in *Salmonella*. The common use of quinolones in both animals and humans can promote horizontal gene transfer and the acquisition of quinolone-resistance genes between these two sectors through environment links. Mechanisms of quinolone resistance in *Salmonella* need to be further investigated. The rational use of common antimicrobials and rigorous antimicrobial stewardship for both animal and human therapeutic intervention can delay the spread of AMR. Increasing AMR monitoring and developing AMR control strategies in aquaculture to overcome adverse consequences of AMR should be initiated immediately.

CONCLUSIONS

Findings from this study on AMR bacteria in hybrid red tilapia suggest that aquaculture as practiced in Thailand can select for ubiquitous AMR pathogens, mobile genetic elements, and an emerging reservoir of *mcr* and colistin-resistant bacteria. Resistant and pathogenic bacteria, such as resistance to ampicillin and tetracycline, or MDR *Salmonella* circulating in aquaculture together highlight the public health concerns and foodborne risks of zoonotic pathogens to humans from cultured freshwater fish. The main sources of AMR bacteria should be evaluated to better understand of their circulation between aquaculture and their production environment. Good personal hygiene, occupational safeguards, and farm biosecurity are highly recommended to reduce AMR bacterial contamination of freshwater fish. The surveillance and monitoring of AMR in aquaculture under One Health, and

improved antimicrobial stewardship for farmers, should be promoted to better control AMR in tropical aquaculture.

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Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Varangkana Thaotumpitak performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- Jarukorn Sripradite performed the experiments, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- Edward R. Atwill conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Saharuetai Jamsripong conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.

DNA Deposition

The following information was supplied regarding the deposition of DNA sequences:

The sequences of *gyrA* are available at GenBank:

[OP831158](#) to [OP831165](#).

Data Availability

The following information was supplied regarding data availability:

The raw data and sequencing data are available in the [Supplementary Files](#).

Supplemental Information

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