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**1 Long chain ω -3 levels are associated with increased alcohol sensitivity in a
2 population-based sample of adolescents**

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26**Abstract**

27*Background*

28 Levels of the ω -3 long-chain polyunsaturated fatty acids (ω -3 LC-PUFAs), including
29 eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA), have been
30 associated with alcohol sensitivity in vertebrate and invertebrate model systems,
31 but prior studies have not examined this association in human samples despite
32 evidence of associations between ω -3 LC-PUFA levels and alcohol-related
33 phenotypes. Both alcohol sensitivity and ω -3 LC-PUFA levels are impacted by
34 genetic factors, and these influences may contribute to observed associations
35 between phenotypes. Given the potential for using EPA and DHA supplementation in
36 adjuvant care for alcohol misuse and other outcomes, it is important to clarify how
37 ω -3 LC-PUFA levels relate to alcohol sensitivity.

38*Methods*

39 Analyses were conducted using data from the Avon Longitudinal Study of Parents
40 and Children (ALSPAC). Plasma ω -3 LC-PUFA levels were measured at ages 15.5 and
41 17.5. Participants reported on their initial alcohol sensitivity using the early drinking
42 Self-Rating of Effects of Alcohol (SRE-5) scale, for which more drinks needed for
43 effects indicates lower levels of response per drink, at ages 15.5, 16.5, and 17.5.
44 Polygenic liability for alcohol consumption, alcohol problems, EPA levels, and DHA
45 levels were derived using summary statistics from large, publicly available datasets.
46 Linear regressions were used to examine the cross-sectional and longitudinal
47 associations between ω -3 LC-PUFA levels and SRE scores.

48*Results*

49 Age 15.5 ω -3 LC-PUFA levels were negatively associated with contemporaneous SRE
50 scores and with age 17.5 SRE scores. One modest association ($p=0.02$) between

51polygenic liability and SRE scores was observed, between alcohol problems-based
52PRS and age 16.5 SRE scores. Tests of moderation by genetic liability were not
53warranted.

54*Conclusions*

55Plasma ω -3 LC-PUFA levels may be related to initial sensitivity to alcohol during
56adolescence. These data indicate that diet-related factors have the potential to
57impact humans' earliest responses to alcohol exposure.

58

59**Key words:** ALSPAC, ω -3 long-chain polyunsaturated fatty acids, alcohol sensitivity,
60PUFA, HUFA, eicosapentaenoic acid, docosahexaenoic acid

61 Introduction

62 Alcohol problems are common in the US and other Western societies: A 2015
63 study of a population-based adult cohort reported a lifetime prevalence of 29.1% for
64 DSM-5 alcohol use disorder (AUD) (Grant et al., 2015) and the World Health
65 Organization estimated that 16.0% of drinkers aged 15 and older engage in heavy
66 episodic drinking (World Health Organization, 2014). Due to the impact of excessive
67 alcohol use on health and productivity, the economic consequences are substantial,
68 estimated at \$249 billion in 2010 in the US alone (Sacks et al., 2015). Clarifying the
69 biological and environmental factors that contribute to the risk of developing
70 alcohol problems is, therefore, a public health priority.

71 Long-chain ω -3 polyunsaturated fatty acid (ω -3 LC-PUFA; also known as highly
72 unsaturated fatty acids [HUFA]) levels are a primarily environmental factor that has
73 been associated with acute ethanol response behaviors in both invertebrate and
74 vertebrate models. In *C. elegans*, genetic contributors to a low level of response
75 (low LR) to alcohol have been identified (Davies et al., 2003), and the development
76 of acute functional tolerance (AFT) to ethanol requires the long chain ω -3
77 eicosapentaenoic acid (EPA) (Raabe et al., 2014). Supplementation of additional EPA
78 can enhance AFT, indicating that EPA levels can influence the acute response to
79 ethanol in *C. elegans* (Raabe et al., 2014). In mice, dietary levels of long chain ω -3s
80 interact with the genetic background to alter several acute ethanol responses
81 including low dose locomotor activation and high dose sedation. Intriguingly, in
82 C57BL/6j mice but not DBA/2j mice, dietary EPA and DHA increased voluntary
83 ethanol consumption (Wolstenholme et al., 2018)

84 Acute ethanol response behaviors, including AFT, in model organisms are a
85 model of initial alcohol sensitivity in humans. The initial acute physiological

86sensitivity to alcohol is a partially heritable phenotype (Edwards et al., 2018, Heath
87et al., 1999, Schuckit, 2018) which has been associated with later alcohol
88consumption and problems (Schuckit, 1994, Schuckit et al., 2007). Lower initial
89sensitivity to alcohol is a risk factor for higher alcohol consumption and subsequent
90problems. Improved understanding of factors that impact one's alcohol sensitivity
91may therefore be useful in understanding trajectories from early to problematic
92alcohol use and has been used as a focus for prevention programs (Schuckit et al.,
932016).

94 The effect of ω -3 LC-PUFA levels on the physiological response to ethanol may
95be of particular relevance to human alcohol use: Human EPA and DHA levels are
96primarily determined by diet, making them an easily modifiable target for alcohol
97studies. Indeed, EPA and DHA supplementation, usually from fish oil, is common,
98and can have significant impacts on the levels of ω -3 LC-PUFAs in plasma (Superko
99et al., 2013). However, in humans, little is known about how the levels of ω -3 LC-
100PUFAs may be related to alcohol sensitivity.

101 Here, we sought evidence for a relationship between measured plasma ω -3
102LC-PUFA levels and alcohol sensitivity in the Avon Longitudinal Study of Parents and
103Children (ALSPAC). We capitalized on the availability of repeated measures of
104plasma ω -3 LC-PUFA levels and self-reported initial sensitivity to alcohol (SRE-5)
105across adolescence to test whether these measures were related, and if so, if the
106relationship was contemporaneous and/or whether ω -3 LC-PUFA levels are
107associated with later alcohol sensitivity. Prior evidence indicates that both initial
108alcohol sensitivity and ω -3 LC-PUFA levels are genetically influenced (Edwards et al.,
1092018, Lemaitre et al., 2011, Steer et al., 2012). The aforementioned differences in
110the phenotypic association between ω -3 LC-PUFA levels and alcohol consumption as

111a function of genetic background in mice (Wolstenholme et al., 2018) raises the
112question of whether genetic factors may have a similar impact in humans. We
113therefore further assessed if polygenic liability for alcohol-related phenotypes
114and/or ω -3 LC-PUFA blood levels contributes to any association between ω -3 levels
115and alcohol sensitivity. Incorporation of aggregate genetic factors may clarify
116models of biological mechanism(s) contributing to the relationship between ω -3 LC-
117PUFA levels and alcohol outcomes. Furthermore, should genetic factors prove
118influential in this association, they could potentially inform the suitability of using
119EPA and DHA supplements in treatment settings.

120

121**Materials and Methods**

122*Sample*

123 There were 14,541 initial pregnancies for which the mothers enrolled in the
124Avon Longitudinal Study of Parents and Children (ALSPAC) study and had either
125returned at least 1 questionnaire or attended a “Children in Focus” clinic by July 19,
1261999. Of these initial pregnancies, there was a total of 14,062 live births and 13,988
127children who were alive at 1 year of age. Subsequent phases of enrollment
128increased the sample size over time (Fraser et al., 2013, Boyd et al., 2013). The
129phases of enrollment are described in more detail elsewhere (Fraser et al., 2013,
130Boyd et al., 2013). Only offspring genotypes were used in the current analyses.
131Participants are encouraged to contribute to assessments whenever possible even if
132not at every wave, and are permitted to skip questions within an assessment;
133accordingly, there is often variation across and within waves with respect to data
134availability for a given participant. The study website contains details of all the data
135that is available through a fully searchable data dictionary (<http://www.bristol.ac.uk/>

136 [alspac/researchers/our-data/](#)). Beginning with the age 22 assessment, online
137 questionnaires were administered using REDCap (Harris et al., 2009). Ethical
138 approval for the study was obtained from the ALSPAC Ethics and Law Committee
139 and the Local Research Ethics Committees. Informed consent for the use of data
140 collected via questionnaires and clinics was obtained from participants following the
141 recommendations of the ALSPAC Ethics and Law Committee at the time.

142 *Alcohol sensitivity*

143 Sensitivity to alcohol was assessed using the Self-Rating of the Effects of
144 Alcohol (SRE) scale (Schuckit et al., 1997). The SRE consists of 4 items; for the
145 current study, each item referred to the *first five or so times*, the SRE-5, a
146 participant used alcohol (referred to hereafter as SRE). Participants were asked to
147 report the number of standard drinks they usually needed to consume to
148 experience any effect of the alcohol, slur their speech, feel unsteady on their feet,
149 or intentionally fall asleep. Consistent with prior reports (Edwards et al., 2018)
150 responses were winsorized to limit extreme values and reduce the effect of possibly
151 spurious outliers. SRE scores were calculated by summing the drinks needed for
152 effects across items and dividing by the number of the up to four effects
153 experienced, as recommended by Schuckit and colleagues (Schuckit et al., 1997).
154 Thus, *higher SRE scores correspond to lower initial alcohol sensitivity*. The current
155 study included SRE reports from approximate ages 15.5 (n=3285), 16.5 (n=1398),
156 and 17.5 (n=942), which correspond to a time frame during which initiation of
157 alcohol use is common and thus increases the likelihood that participants are
158 reporting on their first experiences with alcohol.

159 Because we were interested in relationships between *initial* alcohol sensitivity
160 and ω -3 levels, data were coded such that only a participant's first report of

161sensitivity was used. That is, if a participant responded to the SRE questionnaire
162items at ages 15.5 and 16.5, only the age 15.5 response was included in
163regressions; this decision was made to increase the likelihood that scores more
164closely reflected the participant's first exposure to five or so drinks, as we were
165concerned that recall bias and/or more recent alcohol use experiences may impact
166responses during later assessments. Due to attrition and the fact that most
167participants had initiated alcohol use prior to age 16.5, a consequence of this
168decision was a smaller sample size in analyses for which the outcome was SRE
169score at age 16.5 or 17.5.

170 *ω -3 LC-PUFA blood levels*

171 ALSPAC participants periodically participate in clinics wherein physiological
172measures are taken in addition to typical questionnaire assessments. Fasting
173(minimum of 6 hours) plasma lipids were assessed at participant ages 15.5
174(n=3361) and 17.5 (n=3166) (an assessment at age 7.5 was excluded as it was
175temporally distant from SRE reports). Lipids were measured using a high-throughput
176nuclear magnetic resonance metabolomics platform (Soininen et al., 2015, Soininen
177et al., 2009). We restricted our analyses to total ω -3 LC-PUFAs; data are in mmol/L.

178 *Polygenic liability*

179 To assess whether genetic factors relevant to alcohol outcomes and/or ω -3
180LC-PUFA are related to any observed association between ω -3 LC-PUFA levels and
181initial alcohol sensitivity, we constructed polygenic risk scores for individuals within
182ALSPAC. PRS are derived by multiplying beta estimates (or odds ratios) for an effect
183allele at a particular locus - estimated in an independent sample - by the number of
184effect alleles an individual carries at that locus. This is repeated at the genome-wide
185level (after accounting for linkage disequilibrium). Ultimately, an individual's score

186reflects their aggregate genetic liability for a phenotype of interest (in this case,
187AUDIT scores and plasma EPA and DHA levels). Additional information on PRS is
188available in Choi et al. (2018) and Sugrue and Desikan (2019).

189 To derive PRS for the current study, we obtained publicly available summary
190statistics from the most well-powered and phenotypically appropriate GWAS
191identified through a literature search. Although a meta-analysis of two GWAS of SRE
192scores is available (Edwards et al., 2018) that study included the ALSPAC sample,
193rendering it unsuitable as a discovery dataset. We therefore used summary
194statistics from a GWAS of AUDIT (Babor et al., 2001) scores in the UK Biobank
195sample (Bycroft et al., 2018, Sanchez-Roige et al., 2019) disaggregated into the
196AUDIT-C and AUDIT-P to enable detection of potential differences in the association
197between genetic liability to each construct with ω -3 LC-PUFA levels. AUDIT-C
198consists of the first three AUDIT items and captures past-year alcohol consumption;
199AUDIT-P consists of the remaining 7 AUDIT items and captures past-year
200problematic use. Because the aim was to account for polygenic liability to alcohol
201consumption/problems rather than to dissect the impact of loci implicated at
202various levels of significance, only the inclusive $p < 0.50$ threshold PRS was derived
203for inclusion as a predictor. Note that, while the Sanchez-Roige report includes both
204UK Biobank and 23andMe participants, only summary statistics for the former were
205used in the current analyses.

206 To account for genetic factors associated with ω -3 LC-PUFA levels, we
207downloaded summary statistics from meta-analyses of GWAS on plasma EPA and
208DHA levels, made available by the CHARGE Consortium
209(<http://www.chargeconsortium.com/main/results>) and reported by Lemaitre et al.
210(2011) The CHARGE study consisted of 8,866 participants of European ancestry,

211making it suitable as a discovery sample for ALSPAC. We chose to analyze the long-
212chain polyunsaturated fatty acids EPA and DHA because these long chain ω -3 fatty
213acids had been directly tested in animal models and had been shown to affect
214ethanol sensitivity (Wolstenholme et al., 2018) In addition, EPA and DHA are the
215main constituents of fish oil, a common dietary supplement.

216 Genotypes for ALSPAC participants are available for a fee to researchers will
217an approved project (see <http://www.bristol.ac.uk/alspac/researchers/> for details).
218Genotyping and initial quality control of data were performed by ALSPAC analysts,
219unrelated to the current project. Genotyping in ALSPAC was performed on the
220Illumina HumanHap550 quad genome-wide SNP genotyping platform by 23andMe
221subcontracting the Wellcome Trust Sanger Institute, Cambridge, UK, and the
222Laboratory Corporation of America, Burlington, NC, USA. Individuals were excluded
223from analyses on the basis of excessive or minimal heterozygosity, gender
224mismatch, individual missingness (0.3%), cryptic relatedness as measured by
225identity by descent (genome-wide IBD 0.10%) and sample duplication. Individuals
226were assessed for population stratification using multi-dimensional scaling
227modelling seeded with HapMap Phase II release 22 reference populations.
228Individuals of non-European ancestry were removed from further analysis. Shapelt
229v2 was used to impute to 1000 Genomes Phase 1, Version 3, Release December
2302013. We excluded markers with $MAF < 0.01$, deviation from HWE ($p < 5 \times 10^{-6}$),
231genotyping rate < 0.95 , or $INFO < 0.80$. Polygenic risk scores were derived using the
232--score and --dosage options in Plink 1.9 (www.cog-genomics.org/plink/1.9/) (Chang
233et al., 2015) for markers with $p < 0.5$ in the discovery sample. This corresponding to
234the following numbers of SNPs contributing to the four PRS after pruning: AUDIT-C,
235222,651; AUDIT-P, 222,118; DHA, 154,759; EPA, 156,352.

236 *Statistical analyses*

237 Analyses were conducted in R version 3.4.3 using the glm function,
238 potentially using three stages of multivariable regressions. In the first stage, SRE
239 scores were regressed onto ω -3 LC-PUFA levels, including biological sex (determined
240 at birth) as a covariate. In the second stage, we added the main effects of ancestry-
241 informative principal components as well as polygenic scores for alcohol
242 consumption, alcohol problems, DHA, and EPA. In the third stage, we added
243 interaction terms between ω -3 LC-PUFA levels and any PRS variable for which a
244 main effect ($p < 0.05$) was observed for both the variables in the second stage, in
245 order to test whether polygenic liability for the trait(s) in question moderated the
246 ω -3 LC-PUFA \rightarrow SRE association.

247 Although our primary research question focused on contemporaneous ω -3 LC-
248 PUFA and SRE, we also considered the possibility that ω -3 LC-PUFA level would
249 impact later SRE scores. We therefore ran regression models in which the age 15.5
250 ω -3 LC-PUFA measure was the predictor of interest for SRE at age 16.5 or 17.5. SRE
251 scores, ω -3 LC-PUFA levels, and PRS scores were standardized prior to analysis for
252 ease of interpretation.

253

254 **Results**

255 *Descriptive statistics*

256 **Table 1** provides descriptive statistics for ω -3 LC-PUFA levels and SRE scores
257 across waves. ω -3 LC-PUFA levels at ages 15.5 and 17.5 were correlated at $r = 0.49$
258 ($p < 0.0001$). Correlations within SRE scores were not calculated since only the first
259 report was used for each individual. Both alcohol-related PRS were weakly positively
260 correlated with the first reported SRE score ($r = 0.03$, $p = 0.04-0.06$), indicating that

261 higher genetic liability to alcohol consumption/problems was correlated with
262 needing more standard units of alcohol to perceive its effects (i.e., lower alcohol
263 sensitivity). DHA/EPA PRS were weakly positively correlated with ω -3 LC-PUFA levels
264 ($r = 0.02-0.05$, $p = 0.01-0.22$). Correlations across ω -3 LC-PUFA levels and SRE
265 scores ranged from $r = -0.18$ to $r = -0.05$ ($p < 0.01-0.38$; **Figure**).

266 *Model 1 regression results*

267 **Table 2** (top panel) provides results from Model 1, in which SRE scores were
268 regressed onto ω -3 LC-PUFA levels and sex. Higher ω -3 LC-PUFA levels at age 15.5
269 were nominally associated ($p < 0.05$) with lower SRE scores (i.e., higher initial alcohol
270 sensitivity) at ages 15.5 and 17.5, but not at age 16.5. Age 17.5 ω -3 LC-PUFA levels
271 were not associated with concurrent SRE scores.

272 *Model 2 regression results*

273 We next added PRS for alcohol consumption, alcohol problems, DHA levels,
274 and EPA levels as potential predictors of initial alcohol sensitivity. Results are
275 presented in **Table 2** (bottom panel). The associations between higher age 15.5 ω -3
276 LC-PUFA levels and lower SRE scores (i.e., higher alcohol sensitivity) at ages 15.5
277 and 17.5 persisted in these adjusted models. We also observed a negative
278 association between AUDIT-P PRS and age 16.5 SRE scores. No other PRS was
279 associated with SRE score at any age (all $p \geq 0.05$). Because the only main effect of
280 PRS was observed in a model in which no significant main effect of ω -3 LC-PUFA was
281 observed, we did not test for an interaction with the PRS score.

282

283 **Discussion**

284 Prior research in model systems has demonstrated that ω -3 LC-PUFA levels
285 may influence initial physiological sensitivity to alcohol, but associations between

286 ω -3 LC-PUFA levels and sensitivity to alcohol in humans have not been reported, to
287 our knowledge. Our analyses tested for an association between plasma ω -3 LC-PUFA
288 levels and initial sensitivity to alcohol in a population-based sample of adolescents.
289 We also assessed whether genetic factors underlying to alcohol outcomes
290 (consumption or problems) or EPA/DHA levels further contributed to initial alcohol
291 sensitivity. We found that higher ω -3 LC-PUFA levels were associated with higher
292 alcohol sensitivity in some, but not all, analyses. Polygenic scores exhibited little to
293 no effect on the outcome, and moderation tests were not warranted. In conjunction
294 with results from model systems, these findings extend our understanding of the
295 relationship between ω -3 LC-PUFA levels and alcohol outcomes to include an
296 individual's earliest experiences with the drug.

297 ω -3 LC-PUFA levels assessed at age 15.5 were associated with SRE scores
298 both concurrently and two years later, but not with SRE scores in the intervening
299 year. While the effect size at age 17.5 is nearly twice that at age 15.5, the standard
300 error is much higher in the former and the corresponding significance value much
301 weaker, and we observed no association between ω -3 LC-PUFA levels and age 16.5
302 initial sensitivity. We conducted four multivariable analyses, for which a
303 corresponding conservative correction would require $p < 0.0125$ to achieve statistical
304 significance. This correction would further call into question the association between
305 age 15.5 ω -3 LC-PUFA level and age 17.5 SRE; the within-age 15.5 association
306 survives the correction. Our data, therefore, can most cautiously be interpreted as
307 supporting a relationship between contemporaneous ω -3 LC-PUFA levels and initial
308 alcohol sensitivity in early/mid-adolescence.

309 We observed a main effect of aggregate genetic liability toward alcohol
310 problems — operationalized by scores on the problems subscale of the AUDIT — on

311age 16.5 SRE scores but in no other case. This association was not in the expected
312direction: Here, higher genetic liability to alcohol problems was associated with
313lower SRE scores (i.e., higher sensitivity). There were no main effects of polygenic
314liability for alcohol consumption, EPA levels, or DHA levels. Furthermore, in the
315absence of jointly observed main effects for both polygenic liability and ω -3 LC-PUFA
316levels, moderation tests were not warranted. Thus, although both the outcome and
317predictor of interest are genetically influenced (Edwards et al., 2018, Lemaitre et
318al., 2011), it is unlikely that the degree of their phenotypic association in the
319ALSPAC sample is moderated by genetic factors. This observation is potentially
320pertinent to efforts to determine whether EPA and DHA supplementation could be
321used to modify the response to alcohol because, in the context of initial alcohol
322sensitivity in adolescence, response to ω -3 LC-PUFA supplementation is unlikely to
323depend heavily on one's underlying genetic vulnerability to alcohol problems.

324 It is important to note that the current approach does not determine whether
325the observed associations are causal in nature. In addition, because SRE scores are
326based on self-report, they may be imprecise and/or subject to recall bias. While
327ALSPAC participants are instructed to respond to the items used for the current
328study by recalling their first exposures to alcohol, these reports may be influenced
329by more recent drinking experiences. We therefore restricted our analyses to
330include only an individual's first report on the SRE items (though participants are
331administered the "first 5 or so" SRE items repeatedly across waves) in an effort to
332capture the report that was closest in time to the initial alcohol experiences.

333 We used a large, publicly available dataset of GWAS summary statistics for
334 ω -3 LC-PUFA (from the CHARGE Consortium). While a meta-analysis of initial alcohol
335sensitivity GWAS has been conducted (Edwards et al., 2018), we were unable to use

336summary statistics from that study because genetic influences were driven by the
337ALSPAC sample, and therefore were not independent. We therefore elected to use
338summary statistics from the UK Biobank, as this is a statistically well-powered,
339population-based sample of the same ancestry as ALSPAC, and prior research
340indicates that PRS are more useful when used across similarly ascertained samples
341(Savage et al., 2018) We used summary statistics based on adult phenotypes, and
342given the dynamic nature of diet and alcohol use across the life course, discovery
343samples closer in age to the ALSPAC sample will be valuable to analyze in the
344future.

345 This work adds to the growing appreciation of the effects of dietary ω -3 LC-
346PUFA levels on neurobiological outcomes, including substance use and
347psychopathology. Here we found that basal levels of ω -3 LC-PUFA were associated
348with alcohol response in adolescence, though effect sizes were small and may not
349be clinically significant. Previously, a small study of treatment-seeking substance
350abusers found that low ω -3 LC-PUFA levels were associated with an increased risk of
351relapse or study drop-out (Buydens-Branchey et al., 2009). Plasma ω -3 LC-PUFA
352levels were positively correlated with alcohol consumption in non-alcoholic people in
353IMMIDIET study (di Giuseppe et al., 2009). Together, these observations underscore
354the potential clinical implications of ω -3 LC-PUFA levels on alcohol-related outcomes.

355 Our study is particularly significant because we examined a young population
356and looked at initial responses to ethanol. In alcoholics, heavy drinking is associated
357with dysregulation of ω -3 LC-PUFA levels, probably due in part to liver damage
358(Vatsalya et al., 2016). It may therefore be difficult to distinguish between an effect
359of ω -3 LC-PUFA levels on ethanol responses, versus an effect of alcohol abuse

360 behavior on ω -3 levels. Our study of adolescents is less subject to that confound
361 than studies of adults with long-term histories of heavy drinking.

362 The mechanisms whereby ω -3 LC-PUFAs may influence low responses to
363 alcohol are beyond the scope of the current study but merit consideration. Both ω -3
364 LC-PUFAs and alcohol engage neurotransmitter systems: Deficits in ω -3 LC-PUFAs
365 adversely affect neuroinflammatory mechanisms (Laye et al., 2018) which in turn
366 impact stress hypothalamic-pituitary-adrenal (HPA) axis sensitivity and other
367 neurotransmitter systems (Levant, 2013). Effects of ω -3 LC-PUFA deficiencies on
368 dopaminergic neurons can be pronounced, especially in the ventral striatum (Healy-
369 Stoffel and Levant, 2018). The dopaminergic system is of central relevance in
370 alcohol responses (Schuckit, 2018); alcohol also has prominent simultaneous effects
371 on gamma-aminobutyric acid (GABA), glutamate, opioid, serotonin, and
372 acetylcholine systems, and on the HPA axis, each of which could contribute to
373 alcohol sensitivity (Schuckit, 2018). Future studies could potentially investigate
374 whether high versus low ω -3 LC-PUFA levels may contribute to differences in alcohol
375 sensitivity via perturbation of specific neurotransmitter systems.

376 In summary, we report an association between plasma ω -3 LC-PUFA levels
377 and initial alcohol sensitivity in a population-based cohort of adolescents, such that
378 higher ω -3 LC-PUFA levels correspond to higher alcohol sensitivity. These findings,
379 which primarily support a contemporaneous association in mid-adolescence,
380 warrant follow-up in an independent sample. While our observational study cannot
381 address causality, the results raise the possibility that dietary ω -3 LC-PUFA levels
382 could reduce low initial responses to alcohol, which has been previously associated
383 with the development of problematic alcohol outcomes (Schuckit, 1994). Our

384 findings add to the growing body of literature suggesting important associations
385 between low levels of ω -3 LC-PUFAs and increased risks for psychopathology.

386

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401

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539 **Figure caption**

540 Correlations between unstandardized ω -3 long chain polyunsaturated fatty acid
541 levels (in mmol/L; x-axis) and scores on the Self-Rating of the Effects of Alcohol
542 (SRE; y-axis). Pearson correlations and corresponding p-values are presented, along
543 with regression lines and shaded standard errors.

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546**Table 1.** Descriptive statistics for ω -3 LC-PUFA levels and Self-Rating of the Effects
547of Alcohol (SRE) scores.

	ω -3 LC-PUFA		SRE Score ¹	
	N	Mean (SD) mmol/L	N	Mean (SD)
<i>Age 15.5</i>				
Total	3361	0.28 (0.07)	3285	5.40 (2.95)
Girls	1749	0.30 (0.07)	1869	5.07 (2.83)
Boys	1612	0.26 (0.07)	1416	5.82 (3.05)
<i>Age 16.5</i>				
Total	n/a	n/a	1398	4.58 (2.43)
Girls	n/a	n/a	877	4.35 (2.32)
Boys	n/a	n/a	521	4.98 (2.56)
<i>Age 17.5</i>				
Total	3166	0.30 (0.08)	942	5.14 (2.62)
Girls	1647	0.31 (0.08)	446	4.65 (2.26)
Boys	1519	0.28 (0.07)	496	5.58 (2.84)

548¹Figures are restricted to participants' first SRE report, and represent UK standard drinks,
549one of which contains 8 grams of ethanol (a standard US drink has 14 grams of ethanol). The
550participants whose first SRE report is at age 15.5 do not overlap with those whose first SRE
551report is from a later age.

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Running Head: Omega-3s and initial alcohol sensitivity

Table 2. Results from linear models where scores on the Self-Rating of the Effects of Alcohol (SRE) scale are regressed onto concurrent or prior ω -3 LC-PUFA levels and other potential predictors/covariates. In the interest of space, results for 10 ancestry-informative principal component covariates are not shown. Continuous variables were standardized prior to analysis, and beta coefficients represent the change in SRE associated with a unit change in ω -3 LC-PUFA.

	Age 15.5 ω -3 LC-PUFA and age 15.5 SRE		Age 15.5 ω -3 LC-PUFA and age 16.5 SRE		Age 15.5 ω -3 LC-PUFA and age 17.5 SRE		Age 17.5 ω -3 LC-PUFA and age 17.5 SRE		
	Beta (SE)	<i>p</i> -value							
Model 1									
Sex ¹	-0.20 (0.05)	1.7e-5	-0.12 (0.11)	0.30	-0.51 (0.11)	1.0e-5	-0.33 (0.08)	9.6e-5	
ω -3 LC-PUFA ²	-0.07 (0.02)	9.7e-4	-0.03 (0.06)	0.58	-0.12 (0.06)	0.03	-0.01 (0.04)	0.73	
Model 2									
Sex	-0.19 (0.05)	2.35e-4	-0.10 (0.13)	0.43	-0.36 (0.12)	2.77e-3	-0.22 (0.09)	0.03	
ω -3 LC-PUFA	-0.08 (0.03)	1.67e-3	-0.01 (0.07)	0.88	-0.14 (0.06)	0.02	<0.01 (0.05)	0.94	
AUDIT-C PRS	0.03 (0.03)	0.30	0.08 (0.07)	0.25	0.02 (0.06)	0.70	0.11 (0.05)	0.05	
AUDIT-P PRS	<0.01 (0.03)	0.88	-0.17 (0.07)	0.02	0.11 (0.06)	0.09	0.04 (0.05)	0.41	
DHA PRS	-0.01 (0.03)	0.66	0.06 (0.08)	0.42	0.01 (0.06)	0.83	0.02 (0.05)	0.64	
EPA PRS	-0.02 (0.03)	0.56	-0.05 (0.07)	0.48	0.08 (0.06)	0.22	0.04 (0.05)	0.45	

¹Boys are the reference group

²mmol/L