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Crop Response Functions Integrating Water, Nitrogen, and Salinity

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Abstract

Process-based simulation models are used to generate seasonal crop yield and nitrate leaching datasets for several important crops. The simulated data is then used to estimate novel three-input crop response functions that account for the effects of interactions and feedback mechanisms in the whole plant-water-nitrogen-salinity system. Comparisons with available field data show that this appears to be a reliable approach for estimating analytical crop response functions with water, nitrogen, and salinity as input factors. Results also demonstrate the shortcomings of using simpler two-input functions. The estimated functions are continuously differentiable and can be easily incorporated into comprehensive agricultural-economic-environmental optimization models, thus facilitating greater utilization of process-based models by a wider range of disciplines.

Keywords: crop response functions, crop yield, nitrate leaching, irrigation, nitrogen, salinity

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1 **1. Introduction**

2 Our ability to efficiently manage agricultural water has benefitted in re-
3 cent years from the development of process-based simulation models that are
4 capable of predicting the effects of varying conditions and management prac-
5 tices on crop yield and the environment. Examples of such models include
6 GLEAMS, EPIC, APSIM, SMCR_N, CropSyst, SWAP, ENVIRO-GRO and
7 HYDRUS (Knisel and Turtola, 2000; Williams et al., 1995; Keating et al.,
8 2003; Zhang et al., 2010; Stöckle et al., 2003; Kroes et al., 2008; Pang and
9 Letey, 1998; Šimůnek et al., 2008). Models such as these typically are based
10 on the specific agronomic and biophysical processes that occur at the plant or
11 plot level in short time steps throughout a growing season, and thus represent
12 our best scientific understanding of those processes.

13 These models are potentially very useful for researchers in other disci-
14 plines who are investigating questions that require accurate representation
15 of agronomic and biophysical processes, possibly at larger spatial and time
16 scales. A prime example is economics which is often concerned with pre-
17 dicting the effects of changes in environmental, economic, or regulatory con-
18 ditions on grower behavior and welfare, usually at the farm level and over
19 multiple growing seasons. Such predictions invariably require solving a math-
20 ematical optimization problem that represents the grower’s decision-making
21 process. Although it is possible to link an economic optimization model
22 directly with an external process-based simulation model such that the eco-
23 nomic model calls the simulation model each time the optimization routine
24 needs to calculate a level or derivative of one of the simulated variables,
25 this is uncommon in practice due to the requisite programming skills and

26 the substantial computational burden. A recent example of this approach
27 is Lehmann et al. (2013) in which a genetic algorithm is used to bridge
28 the models. Although the authors acknowledge that “the full potential of
29 [process-based] models is only tapped when as many different management
30 variables as possible are considered simultaneously” (p.56), they must limit
31 their choice set to twelve discrete decision variables in order to achieve rea-
32 sonable computation times. While a decision set of this dimension may be
33 adequate for some single period problems, notwithstanding the lack of con-
34 tinuous choice variables, multi-period problems can easily involve hundreds
35 of decision variables (e.g. Baerenklau et al., 2008).

36 A far more common approach that is more widely accessible, more com-
37 putationally feasible, and allows for a richer set of decision variables is to
38 embed in the economic model analytical functions that have been fitted to
39 data generated either from field experiments or by the external simulation
40 model. This amounts to an indirect linkage of the models via the analytical
41 functions, as shown in Figure 1. A recent example of this approach is Finger
42 (2012) who uses simulated yield data from CropSyst to estimate production
43 functions that are then used to predict changes in water and fertilizer appli-
44 cation rates by corn producers in response to changing economic conditions.
45 In general terms, such crop response functions relate output variables (e.g.,
46 crop yield, pollutant emissions) to the quantity and/or quality of at least one
47 input factor. Crop yield functions have a long history, likely dating back to
48 von Liebig’s “law of the minimum” in the mid-1800’s, and continue to play
49 an important role in economic analysis of agricultural production (Hexem
50 et al., 1978; Lanzer and Paris, 1981; Letey and Dinar, 1986; Griffin et al.,

51 1987; Berck and Helfand, 1990; Tembo et al., 2008). Tembo et al. (2008)
52 provides an overview. Common applications include yield response to water,
53 salinity, fertilizer, pesticide, or some combination of these.

54 As concerns about the effects of agricultural pollution have increased,
55 emission functions have been developed to augment crop yield functions
56 (Tanji et al., 1979; Peralta et al., 1994; Pang and Letey, 1998; Knapp and
57 Schwabe, 2008). With both yield and emission functions in hand, economic
58 analysis can be extended to include not only market inputs and outputs but
59 also the nonmarket effects of agricultural production on natural resources and
60 environmental quality. In the case of nitrogen fertilizer, nitrate leaching typ-
61 ically is estimated as a function of applied water and applied nitrogen. When
62 the response functions are embedded in an economic optimization model, the
63 effects of a fertilizer tax, for example, can be estimated on irrigation water
64 use, fertilizer use, crop yield, farm income, nitrate leaching, and ultimately
65 groundwater quality.

66 Standard practice for empirical specification of such agri-environmental
67 crop response functions has converged on two-input models, typically either
68 water and salinity, or water and nutrients (as in Finger, 2012), or water
69 and pesticides depending on the desired application.¹ Incorporating mul-
70 tiple inputs allows modeling of potentially important interaction effects on
71 crop yield and pollutant emissions. For example, applied irrigation water

¹Here we refer to the variable inputs for which decisions must be made throughout a growing season. Many other choices by a producer affect yield and emissions, such as planting, harvest, and irrigation technologies. However standard practice is to treat these as fixed factors of production and to estimate crop response functions conditionally on these choices.

72 is at least as important as applied nitrogen for determining nitrate leaching
73 because water is the main transport medium for dissolved salts (Pang and
74 Letey, 1998). Therefore, in areas where nitrate pollution is a potential threat
75 to public health and the environment, proper evaluation of pollution control
76 policies requires information on the response of both crop yield and nitrate
77 leaching to both water and nitrogen. Another example is the effect of saline
78 irrigation water on nitrate leaching. Total leached nitrogen has been shown
79 to increase due to the effects of salinity stress on water and nutrient uptake
80 (e.g., Pang and Letey, 1998; Ramos et al., 2011).

81 We are not aware of any previously published crop response functions
82 with three input factors, but such functions would be particularly useful
83 for addressing persistent and emerging problems from irrigated agriculture.
84 Therefore the purpose of this study is to develop, demonstrate, and test a
85 methodology for estimating integrated crop response functions with three in-
86 put factors; and to disseminate the estimated functions for several important
87 crops that use water, nitrogen and salinity as inputs. In order to address
88 the lack of field experimental data that would support estimation of such
89 functions, we utilize simulations. Novel and generally applicable response
90 functions are derived from the simulated data that account for the effects
91 of interactions and feedback mechanisms in the whole plant-water-nitrogen-
92 salinity system.

93 2. Methodology

94 2.1. Function Inputs Specification

95 Most studies estimate models of crop yield and nitrate leaching using
96 *applied* water and nitrogen fertilizer as inputs (e.g., Helfand and House, 1995;
97 Llewelyn and Featherstone, 1997). From an agronomic perspective, it is
98 the combination of management practices like these and pre-existing soil
99 conditions that determine yield and leaching; yet only a few studies include
100 variables such as soil nitrogen stock as an additional input (Vickner et al.,
101 1998; Martínez and Albiac, 2006). Neglecting to account for soil conditions
102 does not necessarily lead to biased estimation results but it does limit the
103 transferability of the response functions to other regions or even to the same
104 field under different conditions. Our crop response functions use available
105 water, available nitrogen, and exposed salinity as inputs and are thus more
106 general and transferrable. Below we show how to navigate between our input
107 variables and those that are more commonly used.

108 Water that is available for crop uptake includes irrigation (e.g., surface
109 water, groundwater, recycled drainage water), precipitation, and initial water
110 content in soil. Initial water content is relatively small compared to the
111 amount of applied water, and thus can be assumed away from crop available
112 water (Letey and Knapp, 1995). Denoting the remaining water sources as
113 $w_i, i = 1, \dots, I$ (cm), crop-available water, w (cm), can be specified as the
114 summation shown in equation (1).

$$w = \sum_{i=1}^I w_i \quad (1)$$

115 Crop-available nitrogen includes only inorganic nitrogen, mainly in the
 116 forms of ammonium and nitrate. Direct sources include soil, atmospheric
 117 deposition, irrigation, and fertilizer. Additionally, the process of mineral-
 118 ization can slowly convert organic nitrogen to ammonium and thus increase
 119 crop-available nitrogen. Following Pang and Letey (1998), it is assumed that
 120 nitrification is rapid so that all mineral nitrogen is NO_3 . Loss of nitrogen
 121 includes volatilization when inorganic fertilizers containing urea are applied
 122 to the field and denitrification when nitrate-nitrogen in soil is converted to
 123 nitrogen gas through microbial processes. Denoting the average seasonal
 124 denitrification rate as λ , the volatilization rate of applied inorganic fertil-
 125 izer as β , and the average seasonal mineralization rate of organic nitrogen
 126 as δ , equation (2) specifies crop available nitrogen n (kg/ha) as a function
 127 of soil inorganic nitrogen in^{soil} (kg/ha), soil organic nitrogen on^{soil} (kg/ha),
 128 applied inorganic fertilizer in^{fl} (kg/ha) and organic fertilizer on^{fl} (kg/ha),
 129 water source w_i (cm) and its nitrogen concentration n_i^w (kg/ha-cm), and the
 130 seasonal rate of atmospheric nitrogen deposition d (kg/ha). This equation
 131 includes the main processes in and above the root zone that can affect the
 132 nitrate leaching rate. Therefore our nitrate leaching functions focus on the
 133 amount of nitrate leached out of the root zone. Once this information is avail-
 134 able, further steps (beyond our analysis) can be taken to incorporate other
 135 processes in the unsaturated and saturated zones to simulate downstream
 136 nitrate emissions.

$$n = (1 - \lambda) \left(in^{soil} + (1 - \beta) in^{fl} + \delta (on^{soil} + on^{fl}) + \sum_{i=1}^I w_i n_i^w + d \right) \quad (2)$$

137 A few models have been developed for predicting salt concentration of soil
 138 solution under intra-seasonal irrigation (Bresler, 1967; Knapp, 1984) and have
 139 been applied to studies on saline water irrigation (e.g., Knapp, 1992; Kan
 140 et al., 2002; Kan, 2008). Following Knapp (1992) and Kan et al. (2002), we
 141 adapt the salt balance model in Knapp (1984) to obtain the seasonal average
 142 for crop exposed salinity s (dS/m), as shown in equation (3). Here, ν (cm)
 143 is the field capacity for soil moisture, s^{soil} (dS/m) is the salt concentration
 144 of soil solution at the beginning of a growing season, w and w_i are defined
 145 in equation (1), s_i^w (dS/m) is the salt concentration of water source w_i , and
 146 w_{up} (cm) is the amount of water absorbed by crops. Crop exposed salinity
 147 equals the total amount of salt in the soil and from irrigation divided by the
 148 total amount of water that is not taken up by the crop.

$$s = \frac{\nu s^{soil} + \sum_{i=1}^I w_i s_i^w}{\nu + w - w_{up}} \quad (3)$$

149 The crop response functions can be summarized as

$$w_{up} = \Psi_{w_{up}}(w, n, s) \quad (4)$$

$$n_{up} = \Psi_{n_{up}}(w, n, s) \quad (5)$$

$$ry = \Psi_{ry}(w, n, s) \quad (6)$$

$$nl = \Psi_{nl}(w, n, s) \quad (7)$$

150 where w , n , and s are defined above; and w_{up} (cm), n_{up} (kg/ha), ry , and
 151 nl (kg/ha) are respectively water uptake, nitrogen uptake, relative yield (the
 152 ratio of actual yield to maximum attainable yield), and nitrate leaching.

153 *2.2. Alternative Function Specification*

154 The crop response functions defined above depend on absolute values of
155 water, nitrogen, and salinity. As Letey and Dinar (1986) point out, for pur-
156 poses of transferring such relationships among different geographical areas, it
157 is helpful if these inputs can be expressed in relative terms. We can achieve
158 this by specifying local scaling factors and conducting function transforma-
159 tions.

160 Relative input values are equal to absolute input values divided by scaling
161 factors, which are listed in Table 1. The three critical scaling factors for a crop
162 in a certain region are maximum water uptake (i.e., potential transpiration)
163 w_{up}^* (cm), maximum nitrogen uptake n_{up}^* (kg/ha), and the salinity critical
164 value \overline{EC} (dS/m), each of which depends on climate, soil conditions, and
165 farming practices. Table 2 summarizes the scaling factors for corn, cotton,
166 and small grains in our study region. These scaling factors can be derived for
167 other regions of interest, and once this information is available, the functions
168 we develop here are transferable to those regions.

169 Take the relative yield function as an example. We can transform the
170 relative yield function in equation (6) by using an appropriate set of scaling
171 factors to get equation (8), a new function Ω_{ry} that only relies on relative
172 input values. Since local climate conditions, soil conditions, and farming
173 practices are incorporated into local values of w_{up}^* , n_{up}^* , and \overline{EC} , this func-
174 tion is independent of these characteristics. The same is true for our water
175 uptake, nitrogen uptake, and nitrate leaching functions when they are ex-
176 pressed similarly in relative terms.

$$\begin{aligned}
ry &= \Psi_{ry}(w, n, s) \\
&= \Psi_{ry}(rw \cdot w_{up}^*, rn \cdot n_{up}^*, rs \cdot \overline{EC}) \\
&= \Omega_{ry}(rw, rn, rs \mid w_{up}^*, n_{up}^*, \overline{EC})
\end{aligned} \tag{8}$$

177 3. Data

178 Data required for estimating our crop response functions include: applied
179 water, applied nitrogen, soil nitrogen, soil salinity, crop water uptake, crop
180 nutrient uptake, crop yield, and nitrate leaching. We are not aware of any
181 field experiments that have generated a full set of data that could be used to
182 simultaneously estimate the effects water, nitrogen and salinity on yield and
183 leaching. This generally limited availability of field data is likely due to the
184 high cost of experimentally quantifying the combined effects of multiple input
185 factors on yield and solute leaching. This motivates our use of simulation
186 models to generate the required data and our subsequent use of available
187 field data to validate the approach.

188 3.1. Model selection

189 Many models have been developed to simultaneously deal with plant
190 growth, water flow, and solute movement at the field level. Among those
191 models, CropSyst, SWAP, ENVIRO-GRO, and HYDRUS contain salinity
192 modules. The four models are similar in terms of their simulation capa-
193 bilities and each has had its individual modules successfully tested under
194 various empirical conditions (e.g., Pang and Letey, 1998; Stöckle et al., 2003;

195 Zhang et al., 2010; Bonfante et al., 2010). However, there are few evalu-
196 ations of these models that simultaneously test both nutrient and salinity
197 modules. One exception is a study by Pang and Letey (1998) that evaluates
198 ENVIRO-GRO against field experiment data and demonstrates good agree-
199 ment. ENVIRO-GRO might have been an ideal model for the purpose of this
200 study, but the nitrogen module was subsequently modified and is no longer
201 valid (J. Letey, personal communication). Another exception is a recent
202 study by Ramos et al. (2011) that evaluates HYDRUS-1D using data from
203 a field experiment in which corn is irrigated with water of varying nitrogen
204 and salt concentrations. HYDRUS-1D is a software modeling environment
205 for analysis of water flow and solute transport in variably saturated porous
206 media (Šimůnek et al., 2008). It is used worldwide and has been shown to be
207 reliable for modeling water flow and solute transport, especially for processes
208 in soil and groundwater. The results in Ramos et al. (2011) show HYDRUS-
209 1D to be an effective tool for simulating concentrations of both salinity and
210 nitrogen species in soil, and thus we elect to use it here for datast generation.

211 Ramos et al. (2011) do not utilize the active mechanism of root nutrient
212 uptake in HYDRUS-1D, which is reasonable given their objective to simulate
213 field conditions in a relatively simple way and to provide indicative values.
214 Given our objective of quantifying crop yield and solute leaching, we need
215 to include both active and passive root nitrogen uptake. We simulate the
216 transport of two solutes (salt and nitrogen) in HYDRUS-1D. For salts we
217 assume that there is no uptake by plant roots, while for nitrogen we utilize
218 the compensated root water and nutrient uptake modules through both pas-
219 sive and active mechanisms (as described in Šimůnek and Hopmans (2009)).

220 Simulation of active solute uptake with multiple solutes is a standard feature
221 of the latest version of HYDRUS-1D.

222 The outputs from HYDRUS-1D include estimated water uptake, solute
223 uptake, and solute leaching but not crop yield. External functions thus
224 are required to convert water and nutrients uptake from HYDRUS-1D into
225 crop yield. Following Pang and Letey (1998), relative yield is specified as a
226 function of relative water uptake and relative nitrogen uptake:

$$ry = \min [ry_w, ry_n] = \min \left[\frac{w_{up}}{w_{up}^*}, \Phi \left(\frac{n_{up}}{n_{up}^*} \right) \right] \quad (9)$$

227 Here Φ represents a quadratic relationship (Pang and Letey, 1998; Feng
228 et al., 2005). Combining HYDRUS-1D outputs with the agronomic model
229 in equation (9) allows us to generate a full set of estimates for crop water
230 uptake, nitrogen uptake, nitrate leaching, and crop yield.

231 3.2. Validation of Simulated Data

232 To our knowledge, the best available field experiment data for validating
233 this study are from a corn trial in Davis, California from 1973 to 1976 (Tanji
234 et al., 1979). The field was treated with a wide range of water and nitrogen
235 applications: four different rates of nitrogen fertilizer (0, 90, 180, and 360
236 kgN/ha) and three different irrigation regimes (20, 60, and 100 cm), with
237 each replicated four times. In addition, nitrogen in harvested grain and stover
238 (i.e., nitrogen uptake) was accurately measured. See Tanji et al. (1979) and
239 Broadbent and Carlton (1980) for detailed descriptions. This high quality
240 dataset has been used previously for model validation (e.g., Tanji et al., 1979;
241 Pang and Letey, 1998, 2000) and continues to be used in more recent work

242 (e.g. Knapp and Schwabe, 2008).

243 Broadbent and Carlton (1980) report that the Davis corn field trial was
244 established on Yolo fine sandy loam (alluvial, Typic Xerorthent). Therefore
245 in the water flow module of HYDRUS-1D we select sandy loam under the
246 soil catalog, which gives soil hydraulic parameters for typical soil types. The
247 soil bulk density is assumed to be 1.40 Mg/m³ (Pang and Letey, 1998). The
248 longitudinal dispersivity is assumed to be 20 cm and the molecular diffusion
249 coefficient of nitrates and salts in free water is set to zero because the fluxes
250 by diffusion are negligible compared to dispersive transport (Appendix H,
251 Chang et al., 2005). Tanji et al. (1979) also report the maximum root depth
252 for corn in the Yolo soil to be around 2.4 m so we use this value in the root
253 growth module, where the logistic growth function is used to simulate root
254 growth. The root growth factor is specified as 50% after half of the growing
255 season. Because the field trial used nonsaline irrigation water and all plots
256 received pre-irrigation prior to planting, initial soil salinity is assumed to be
257 0.01 dS/m for all simulations.

258 Results are presented in Figures 2 and 3. Linear regression equations
259 are reported along with the coefficients of determination. Figure 2 displays
260 field measured nitrogen uptake versus the simulated nitrogen uptake from
261 the model of Tanji et al. (1979) and from HYDRUS-1D. The HYDRUS-1D
262 model shows overall better performance than the widely used Tanji model.
263 The slope coefficient is closer to one and the intercept term is closer to zero
264 and quite small relative to the range of nitrogen uptake magnitudes. The
265 null hypotheses that these coefficients are respectively equal to one and zero
266 cannot be rejected at 95% confidence levels. Figure 3 compares field mea-

267 sured relative yield to the simulated relative yield calculated from equation
268 (9). Although the fit is not as good as that for nitrogen uptake, it is still
269 quite good given the complexities and uncertainties associated with the whole
270 plant-water-nitrogen-salinity system. The simulation results also compare fa-
271 vorably against the results reported by Pang and Letey (1998) in a test of the
272 ENVIRO-GRO model against the same data, where the slope, intercept, and
273 R^2 for nitrogen uptake and relative yield are respectively [0.87, 22.55, 0.75]
274 and [0.85, 0.05, 0.84]. Experimental data on nitrate leaching are not avail-
275 able from the Davis trial so we cannot conduct a similar comparison for our
276 simulated leaching results. However Ramos et al. (2011) demonstrate that
277 HYDRUS-1D can accurately simulate soil water nitrate concentrations in a
278 similar empirical setting. Overall these results suggest that our approach can
279 be used to model nitrogen uptake and relative yield with acceptable levels of
280 accuracy.

281 *3.3. Linearization of Nitrogen Uptake Curves*

282 Inputs for our HYDRUS-1D simulation include raw data on crops (e.g.,
283 transpiration rate, salt tolerance, maximum root depth, maximum nitrogen
284 uptake, and maximum water uptake) and soil (e.g., evaporation rate, hy-
285 draulic properties, and solute transport parameters).² We also must provide
286 crop-specific uptake curves (daily potential uptake) for both water and nitro-
287 gen. Most of this information can be obtained locally through either direct

²Climate data (e.g., precipitation and average daily temperature) can be input directly into simulation models, or it can be incorporated indirectly through other variable inputs. We have shown in the previous section why the latter is a better approach for improving the transferability of crop response functions.

288 field measurement or indirect estimation methods. However there is little
289 information on nitrogen uptake curves for crops other than the small grain
290 forages that are discussed in Crohn et al. (2009). To bridge this gap, we
291 use the available data on small grain forages to test the possibility of ap-
292 proximating nitrogen uptake curves with a linear relationship that can be
293 extended to a wider range of crops under the assumption that those uptake
294 curves are approximately linear, as well.

295 Figure 4 depicts the nitrogen uptake curves for eight small grain for-
296 ages commonly grown in California in the winter months. These curves are
297 based on the logistic function developed by Crohn et al. (2009), where the
298 nitrogen content of a crop is a function of cumulative growing degree-days
299 (GDD, or thermal time). For ease of comparison, and consistent with the
300 literature (Crohn et al. (2009)), all curves in Figure 4 terminate with the
301 maximum nitrogen uptake $n_{up}^* = 250$ kgN/ha and the harvest thermal time
302 $d_{harvest} = 2500$ GDD, which are common values for small grain crops in the
303 San Joaquin Valley of California. In practice, the nitrogen uptake of a crop
304 might be higher than the crop's nitrogen content, since some nitrogen can
305 be lost to the atmosphere. Here we assume that the nitrogen uptake and
306 the nitrogen content of a crop are identical. The curves fall into two cate-
307 gories: exponential for two ryegrass crops and sigmoid for the other crops.
308 As Crohn et al. (2009) point out, the exponential curves are most likely due
309 to forage quality and harvest schedule constraints. Sigmoid curves are widely
310 recognized and applied.

311 In order to investigate whether the shape of a nitrogen uptake curve
312 significantly affects estimated total nitrogen uptake and relative yield, sim-

313 ulations of Swan oat (top curve in Figure 4), Longhorn oat (middle curve
 314 in Figure 4), and Bartali Italian ryegrass (bottom curve in Figure 4) with
 315 nonlinear uptake curves are compared to simulations for the same crops with
 316 linear uptake curves. For each crop, we specify combinations of five levels
 317 of available water ($[0.25, 0.5, 1, 1.5, 2] \times w_{up}^*$), five levels of available nitrogen
 318 ($[0.25, 0.5, 1, 1.5, 2] \times n_{up}^*$), and five levels of exposed salinity ($[0, 0.25, 0.5, 0.75, 1] \times$
 319 \overline{EC}), which produces 125 input scenarios for each crop. \overline{EC} is the critical
 320 value of soil salinity at which crop yield decreases to zero. It is calculated
 321 from the salt tolerance parameters of the crop following the approach in
 322 Maas and Hoffman (1977). The linear nitrogen uptake curve of a crop is
 323 constructed from the crop's maximum nitrogen uptake, the harvest thermal
 324 time, and the cumulative thermal time over the season d (GDD), as shown
 325 in equation (10). All other HYDRUS-1D specifications are held constant for
 326 each pair of simulations.

$$n_{up}^d = \frac{n_{up}^*}{d_{harvest}} d \quad (10)$$

327 Figure 5 displays a comparison of the simulation results. Most results
 328 closely track the 45-degree line implying little discernible effect of linearizing
 329 the nitrogen uptake curve. The points that visibly lie off the 45-degree line
 330 are scenarios in which crops receive sufficient water with low salinity but little
 331 nitrogen, which rarely happens in practice. The t-statistics for the three re-
 332 lationships (not shown) suggest that the relative yields of Longhorn oat and
 333 Bartali Italian ryegrass are not statistically different from their linearized
 334 versions at the 95% level; the relative yield of Swan Oat is statistically differ-
 335 ent from its linearized version at the 95% level due to a very small standard

336 error on the slope coefficient, but the difference is not practically significant
337 (about 1.3% of the estimated yield). We conclude that any nonlinear terms
338 in a nitrogen uptake curve can be safely disregarded for our purposes, and
339 thus adopt the linear relationship in equation (10) that can be applied to any
340 crop for which an estimate of the maximum nitrogen uptake is available.

341 4. Results

342 We estimate the crop response functions in equations (4)-(7) for corn, cot-
343 ton, and winter small grains using simulated datasets. For each crop, we spec-
344 ify combinations of at least five levels of available water ($[0.25, 0.5, 1, 1.5, 2] \times$
345 w_{up}^*), five levels of available nitrogen ($[0.25, 0.5, 1, 1.5, 2] \times n_{up}^*$), and six lev-
346 els of exposed salinity ($[0, 0.2, 0.4, 0.6, 0.8, 1] \times \overline{EC}$), which produces at least
347 150 input scenarios for each crop. Some of these input values may seem ex-
348 treme, but they are selected in order to cover most, if not all, of a producer's
349 possible operating scenarios, including potentially stringent environmental
350 regulations. The highest level of available water is double the maximum wa-
351 ter uptake, taking into account that the operator might apply excess water
352 to flush salts out of the root zone. The highest level of available nitrogen
353 is also double the maximum nitrogen uptake, since both animal waste and
354 commercial fertilizers tend to be over-applied to crop fields. And exposed
355 salinity covers the entire range in which it is possible for a crop to grow.

356 For each combination of input values, we use HYDRUS-1D and the agro-
357 nomic model in equation (9) to simulate water uptake, nitrogen uptake, rel-
358 ative yield, and nitrate leaching. We then use this data to estimate crop re-
359 sponse and nitrate leaching as smooth functions of the input values. Drainage

360 water and salt leaching are beyond the scope of this paper but can be calcu-
361 lated using mass balance relationships (see (Knapp and Baerenklau, 2006)).

362 The forms of the water and nitrogen uptake and relative yield functions
363 are developed from the traditional Mitscherlich-Baule functional form, which
364 is discussed below. Because the estimation methods for these three func-
365 tions are similar, we only present the procedure for estimating the crop rel-
366 ative yield function in the following section. The nitrate leaching function
367 is adapted from Knapp and Schwabe (2008). We use corn as an example to
368 illustrate the whole estimation procedure.

369 *4.1. Relative Yield Function*

370 Various forms have been proposed for crop yield functions. Griffin et al.
371 (1987) provide a thorough review of twenty traditional and popular functional
372 forms. They also discuss guidelines for form selection, one of which pertains
373 to application-specific characteristics. Since we expect the resulting functions
374 in this paper to be incorporated into economic optimization models, continu-
375 ous differentiability is a desirable property. Llewelyn and Featherstone (1997)
376 compare five functional forms using corn yield data from the CERES-Maize
377 simulator for western Kansas. Corn yield is estimated as a function of ni-
378 trogen and irrigation water. Their results favor the Mitscherlich-Baule (MB)
379 form over all other specifications. Shenker et al. (2003) measure the yield
380 response of sweet corn to the combined effects of nitrogen fertilization and
381 water salinity over a wide range of nitrogen and salinity levels. Two func-
382 tional forms (Liebig-Sprengel and MB) are evaluated based on the measured
383 data. The results suggest that either functional form can successfully predict
384 water needs, nitrogen needs, and yield. Liebig-Sprengel is a minimization

385 function derived from von Liebig’s “law of the minimum”. It results in a step-
 386 wise response curve that is not differentiable. Therefore, MB is chosen as the
 387 base functional form for crop relative yield in this study. We also compare
 388 our MB function with quadratic and square root functions of corn using pair-
 389 wise P-tests, as used in Frank et al. (1990) and Llewelyn and Featherstone
 390 (1997). The MB function is found to be favored over the other functional
 391 forms.

392 The traditional MB function is usually expressed as equation (11), where
 393 a calibrates the upper limit on yield and the b_i are parameters for the input
 394 factors X_i . This function exhibits continuously positive marginal productiv-
 395 ities of input factors and allows for factor substitution.

$$Y = a \prod_i \left[1 - e^{-b_i^1(X_i + b_i^0)} \right] \quad (11)$$

396 Either absolute yield or relative yield can be the dependent variable, with
 397 a equal to the maximum attainable yield when estimating the absolute yield
 398 and equal to one when estimating the relative yield. Using this form, relative
 399 yield as a function of three inputs can be written as equation (12), where
 400 $b_w, b_n,$ and b_s are parameters for crop available water, available nitrogen, and
 401 exposed salinity.

$$\begin{aligned} ry &= \Psi_{ry}(w, n, s) \\ &= \left[1 - e^{-b_w^1(w - b_w^0)} \right] \left[1 - e^{-b_n^1(n - b_n^0)} \right] \left[1 - e^{-b_s^1(s - b_s^0)} \right] \end{aligned} \quad (12)$$

402 Our efforts to estimate equation (12) directly from the simulated data

403 did not produce good results. Inspection of the simulated data for each
 404 salinity level revealed that relative yield is roughly “bell shaped” in the water
 405 dimension. We thus modify the water parameter in equation (12) so that it,
 406 too, is bell shaped. To do this, we use a parameterized variant of the logistic
 407 probability density function (preferred over the normal distribution because
 408 of its heavier tails) and define it as the water coefficient φ shown in equations
 409 (13) and (14).³ For each salinity level, the crop relative yield function is then
 410 defined as

$$\begin{aligned}
 ry &= \psi_{ry}(w, n | s) \\
 &= \left[1 - e^{-b_w^1(\varphi(w)w - b_w^0)} \right] \left[1 - e^{-b_n^1(n - b_n^0)} \right]
 \end{aligned} \tag{13}$$

411 where

$$\varphi(w) = \frac{4e^{d_1w + d_0}}{(1 + e^{d_1w + d_0})^2} + d_2 \tag{14}$$

412 As shown in Figure 6, this approach produces very good results ($R^2 >$
 413 0.99 for each salinity level). However it produces a set of coefficient estimates
 414 $\Upsilon \equiv \{b_w^1, b_w^0, b_n^1, b_n^0, d_0, d_1, d_2\}$ and thus a yield function that is specific to
 415 each simulated salinity level. To get a single continuously differentiable yield
 416 function that operates over all water, nitrogen, and salinity levels, we note
 417 that each coefficient estimate is effectively a function of salinity so we proceed
 418 to estimate a parametric function of salinity for each coefficient. Salinity thus

³The coefficient 4 is applied to approximately standardize the range of the function to the $[0, 1]$ interval.

419 enters the yield function indirectly through its influence on the water and
420 nitrogen parameters rather than directly as a separate multiplicative term.

421 This approach also reduces the computational requirement by breaking
422 down the problem into two subproblems. First, estimate equation (13) once
423 for each value of $s = [0, 0.2, 0.4, 0.6, 0.8, 1] \times \overline{EC}$, each time using the subset of
424 simulated data points that corresponds to the selected salinity value. These
425 estimations produce the surfaces shown in Figure 6 for the case of corn. As
426 noted by Griffin et al. (1987), convergence problems may arise when estimat-
427 ing the (modified) MB functions during the first step of this procedure. We
428 found that standard methods were sufficient for overcoming such difficulties.
429 These include: (1) increasing iteration limits for the regression models, (2)
430 specifying alternative starting values for the parameters, and (3) using data
431 visualization as a complementary method for goodness-of-fit assessment (e.g.,
432 Figures 6 and 9). Second, estimate each parameter $\Upsilon_i \in \Upsilon$ as a polynomial
433 function of salinity using the coefficient estimates from the first subproblem
434 as data. Figure 7 depicts the regression curves for the case of corn. Again,
435 agreement between the data and functions is generally very good. Finally,
436 substitute the fitted polynomial equations into equations (13) and (14) to
437 get the crop relative yield function with three input variables, as shown in
438 equation (15). For the case of corn, this approach is verified by the excellent
439 agreement between the simulated data and the fitted relative yield shown in
440 Figure 8.

$$\begin{aligned}
ry &= \Psi_{ry}(w, n, s) \\
&= \left[1 - e^{-b_w^1(s)(\varphi(w|d_0(s), d_1(s), d_2(s)) w - b_w^0(s))} \right] \left[1 - e^{-b_n^1(s)(n - b_n^0(s))} \right] \quad (15)
\end{aligned}$$

441 Table 3 summarizes the parameter estimates for the corn relative yield
442 function. Each row shows the coefficients that specify each parameter $\Upsilon_i \in \Upsilon$
443 as a polynomial function of salinity, as well as the R^2 values for each polyno-
444 mial regression. The last column shows the R^2 value for a linear regression
445 of the estimated relative yield from equation (15) versus the simulated data.
446 As can be seen in the table, the R^2 values are generally very good. Similarly,
447 Tables 4 and 5 summarize the parameter estimates for the corn water uptake
448 function and nitrogen uptake function.

449 4.2. Nitrate Leaching Function

450 We use the same comparison tests as for the crop relative yield function
451 to compare four forms of nitrate leaching functions (test results available
452 upon request). A function adapted from Knapp and Schwabe (2008) outper-
453 forms the quadratic, cubic, and square root functions, mainly because of its
454 convex-concave properties and guarantee of a plateau maximum. In Knapp
455 and Schwabe (2008), nitrate leaching is specified as a function of soil nitro-
456 gen, applied nitrogen, infiltrated water, and a set of estimable parameters.
457 Equation (16) shows their function with our notation.

$$nl = \psi_{nl}(w, n | s) = \frac{\vartheta_n \cdot n}{1 + e^{-\vartheta_w^1(w - \vartheta_w^0)}} \quad (16)$$

458 Salinity is notably absent from this leaching equation. Thus the param-
 459 eter vector $\boldsymbol{\vartheta} \equiv \{\vartheta_w^1, \vartheta_w^0, \vartheta_n\}$ implicitly applies to a fixed salinity level. To
 460 incorporate varying salinity levels, we again specify parameters as functions
 461 of salinity levels as in the crop relative yield functions. Our nitrate leaching
 462 function with three input factors is specified in equation (17).

$$nl = \Psi_{nl}(w, n, s) = \frac{\vartheta_n(s) \cdot n}{1 + e^{-\vartheta_w^1(s)(w - \vartheta_w^0(s))}} \quad (17)$$

463 Here we adopt the same estimation procedure as for the crop relative yield
 464 functions. For the case of corn, first estimate equation (16) for each value of
 465 $s = 0, 6, 12, 18, 24, 30$ dS/m to get the surfaces shown in Figure 9. Second,
 466 estimate each parameter $\vartheta_i \in \boldsymbol{\vartheta}$ as a polynomial function of salinity. Figure
 467 10 depicts the regression curves. Finally, substitute the fitted polynomial
 468 equations into equation (16) to get the nitrate leaching function with three
 469 input variables, as shown in equation (17). Figure 11 demonstrates that the
 470 estimated nitrate leaching function fits the simulated data very well for the
 471 case of corn.

472 Table 6 summarizes the parameter estimates for the corn nitrate leaching
 473 function. Each row shows the coefficients that specify each parameter $\vartheta_i \in$
 474 $\boldsymbol{\vartheta}$ as a polynomial function of salinity, as well as the R^2 values for each
 475 polynomial regression. The last column shows the R^2 value for a linear
 476 regression of the estimated nitrate leaching from equation (17) versus the
 477 simulated data. As in Table 3, the R^2 values are again very good. We
 478 follow the same procedure to estimate the response functions (water uptake,
 479 nitrogen uptake, relative yield, and nitrate leaching) for cotton and small
 480 grains. The estimation results are reported in Tables 7 and 8.

481 5. Discussions

482 The datasets we use for estimating our crop response functions are sim-
483 ulated from a hydrologic model consisting of water flow, solute transport,
484 and root growth modules, in conjunction with a simple but effective ana-
485 lytical agronomic model. Therefore, our crop response functions are able
486 to account for the effects of interactions and feedback mechanisms in the
487 whole plant-water-nitrogen-salinity system. Figures 12 and 13 present two
488 examples illustrating the importance of using these integrated crop response
489 functions as opposed to a traditional function with only two inputs.

490 Figure 12 demonstrates the relationship between corn relative yield and
491 available water under three different salinity levels (0.2, 2, and 10 dS/m),
492 given a fixed level of available nitrogen (200 kg/ha). For each salinity level,
493 the relative yield gradually increases to a plateau and then declines. This
494 is consistent with the findings in Pang and Letey (1998) and Knapp and
495 Schwabe (2008). With fixed nitrogen and excessive water, more nitrate is
496 leached out of the root zone and thus less nitrogen is available for crop
497 uptake. Moreover the figure demonstrates the significant effect of salinity on
498 yield that would otherwise be omitted from a standard two-input (water and
499 nitrogen) crop response model.

500 Figure 13 demonstrates the relationship between nitrate leaching and
501 available water under three different salinity levels (0.2, 2, and 10 dS/m),
502 given a fixed level of available nitrogen (200 kg/ha). For each salinity level,
503 nitrate leaching significantly increases when available water exceeds the plant
504 uptake capacity and eventually reaches a plateau. Comparing the three
505 curves shows that high salinity levels tend to generate high nitrate leaching.

506 The underlying mechanisms are well explained in Pang and Letey (1998):
507 “Salinity leads to reduced plant growth, which leads to reduced evapotran-
508 spiration, which leads to more leaching, which leads to salt removal from the
509 root zone. However, the leaching also removes other chemicals such as N and
510 pesticides. Reduced N leads to reduced plant growth, which leads to less
511 evapotranspiration, which leads to more leaching, which leads to even less N
512 in the root zone” (p. 1426). In short, the interactions and feedbacks between
513 plant growth, evapotranspiration, and leaching are complex. Therefore im-
514 portant information can be lost and incorrect nitrate leaching estimates can
515 be generated if common two-input crop response functions are applied in
516 cases where water, nitrogen, and salinity actually interact.

517 One such case where it is important to account for interactions among all
518 three factors is land-application of animal manure. The consolidation trend
519 in animal agriculture has resulted in waste generation rates that far exceed
520 the ability of crops to utilize waste nutrients as fertilizer (Gollehon et al.,
521 2001). In the absence of regulation, the most cost-effective disposal option
522 for farmers is to over-apply manure to crops, resulting in both groundwater
523 and surface water pollution (Harter et al., 2002). Because animal manure
524 usually contains high concentrations of nutrients and salts, both of which
525 affect crop growth and the ability of crops to uptake nitrogen, evaluations
526 of animal waste management practices and policies should be based on in-
527 tegrated crop response functions that relate crop yield and pollutant emis-
528 sion to water, nitrogen, and salinity. Such an application is demonstrated
529 in Wang and Baerenklau (2014), where the crop response functions devel-
530 oped here are incorporated into a dynamic environmental-economic model

531 for policy analysis. These crop response functions also would be useful more
532 broadly for economic analyses of irrigated agriculture in arid and semi-arid
533 regions where problems of water scarcity, excess nutrients, and high salinity
534 commonly coexist.

535 **6. Conclusions**

536 Integrated models of agri-environmental systems are potentially very use-
537 ful for evaluating proposed or anticipated changes in operating conditions
538 (e.g., policies, technologies, prices, and climate). The ability to predict both
539 economic and environmental outcomes within these models is crucial for mak-
540 ing accurate evaluations. Although process-based simulations models poten-
541 tially can be linked to economic optimization models in order to address
542 these questions, this approach can be problematic and remains uncommon.
543 Instead, analytical crop response functions have provided the foundation for
544 such evaluations for many years, but have been limited to only one or two
545 input factors largely due to the limited availability of data for estimation.

546 This article uses a process-based model to generate simulated crop yield
547 and nitrate leaching datasets that are then used to estimate novel three-
548 input crop response functions for several important crops. The functions
549 account for the effects of interactions and feedback mechanisms in the whole
550 plant-water-nitrogen-salinity system, and thus facilitate greater utilization of
551 the knowledge contained in process-based models by other disciplines. Com-
552 parisons with available field data show that this appears to be a reliable
553 approach for estimating integrated crop response functions with water, ni-
554 trogen, and salinity as input factors. Comparisons with simpler two-input

555 functions demonstrate the shortcomings of those functions, which continue
556 to be widely used in disciplines such as economics.

557 Because the crop response functions developed here use available water,
558 available nitrogen, and exposed salinity as inputs, they are more general than
559 functions based only on the characteristics of applied inputs. It is straight-
560 forward to navigate between our input variables and those that are more
561 commonly used, provided sufficient information about soil characteristics is
562 available. Furthermore we demonstrate how to express our inputs in relative
563 terms to facilitate transfer of our crop response functions across different
564 geographic areas.

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Table 1: Scaling factors for calculating relative value

Absolute Value	Scale	Relative value
Available water w [cm]	Maximum water uptake w_{up}^* [cm]	$rw = \frac{w}{w_{up}^*}$
Available nitrogen n [kgN/ha]	Maximum nitrogen uptake n_{up}^* [kgN/ha]	$rn = \frac{n}{n_{up}^*}$
Soil salinity s [dS/m]	Salinity critical value \overline{EC} [dS/m]	$rs = \frac{s}{\overline{EC}}$

Table 2: Scaling factors for corn, cotton, and small grains in the study region

Scale	Corn	Cotton	Small Grains
Maximum water uptake w_{up}^* [cm]	63	68.48	40
Maximum nitrogen uptake n_{up}^* [kgN/ha]	300	187.50	250
Salinity critical value \overline{EC} [dS/m]	30	53.85	85

*Data source: Maas and Hoffman (1977), Pang and Letey (1998), and Crohn et al. (2009).

Table 3: Corn relative yield function

Function	Estimated Parameters							Ψ
	1	s	s ²	s ³	R ²	R ²		
b_w^1	1.8143E-02	-7.0787E-04	1.4785E-05	-6.5115E-08	0.9973			
b_w^0	-1.0316E+01	-2.4081E-01	-2.6960E-02	8.0481E-04	0.9815			
b_n^1	9.3959E-02	2.1530E-01	-1.5375E-02	2.9392E-04	0.7995			
b_n^0	2.3456E-04	-4.9809E-05	2.9616E-06	-5.2352E-08	0.9524	0.9973		
d_0	-2.1708E+00	3.9813E-02	3.7506E-03	-1.8434E-04	0.7736			
d_1	6.2726E-02	1.3402E-04	-2.6225E-05	5.0871E-06	0.9846			
d_2	2.9870E-01	4.9733E-03	-1.3716E-03	2.3810E-05	0.9993			

† Relative yield function: $\Psi_{ry}(w, n, s) = (1 - \exp(-b_w^1(s) (\varphi(w) w - b_w^0(s)))) (1 - \exp(-b_n^1(s) (n - b_n^0(s))))$
 where $\varphi(w) = \frac{4 \exp(d_1(s)w + d_0(s))}{(1 + \exp(d_1(s)w + d_0(s)))^2} + d_2(s)$

Table 4: Corn water uptake function

Function	Estimated Parameters							Ψ
	1	s	s^2	s^3	R^2	R^2	R^2	
b_w^1	1.8525E-02	-7.8960E-04	1.9666E-05	-1.5166E-07	0.9977			
b_w^0	-1.0092E+01	-2.9076E-01	-2.3904E-02	7.4959E-04	0.9816			
b_n^1	1.2619E+01	-1.0002E+00	5.6978E-02	-1.1916E-03	0.7032			
b_n^0	-8.3833E+00	-8.5954E-01	4.4055E-02	-3.6489E-04	0.5158	0.9973		
d_0	-2.3167E+00	7.0209E-02	1.9723E-03	-1.5336E-04	0.8084			
d_1	6.4865E-02	-3.0723E-04	-7.3700E-07	4.6484E-06	0.9854			
d_2	2.9041E-01	6.7238E-03	-1.4749E-03	2.5620E-05	0.9990			

† Water uptake function: $\Psi_{w_{up}}(w, n, s) = (1 - \exp(-b_w^1(s)(\varphi(w)w - b_w^0(s))))(1 - \exp(-b_n^1(s)(n - b_n^0(s))))$
where $\varphi(w) = \frac{4 \exp(d_1(s)w + d_0(s))}{(1 + \exp(d_1(s)w + d_0(s)))^2} + d_2(s)$

Table 5: Corn nitrogen uptake function

Function	Estimated Parameters							Ψ
	1	s	s^2	s^3	R^2	R^2		
b_w^1	4.1660E-02	-6.2435E-04	1.9684E-04	-2.6517E-06	0.9989			
b_w^0	-3.8416E+01	3.8479E-01	6.0445E-02	-1.4881E-03	0.9992			
b_n^1	7.9577E-03	1.6436E-05	-2.6500E-06	4.6767E-08	0.9961			
b_n^0	-1.1849E+00	-7.2459E-03	1.4038E-03	-3.1992E-05	0.8800	0.9956		
d_0	-1.0656E+00	-2.8598E-02	5.4957E-03	-1.1587E-04	0.9941			
d_1	5.2347E-02	7.3267E-04	-4.7909E-05	1.3255E-06	0.9778			
d_2	2.8552E-03	6.0586E-04	-1.1421E-04	2.9139E-06	0.7705			

† Nitrogen uptake function: $\Psi_{n_{up}}(w, n, s) = (1 - \exp(-b_w^1(s)(\varphi(w)w - b_w^0(s))))(1 - \exp(-b_n^1(s)(n - b_n^0(s))))$
 where $\varphi(w) = \frac{4 \exp(d_1(s)w + d_0(s))}{(1 + \exp(d_1(s)w + d_0(s)))^2} + d_2(s)$

Table 6: Corn nitrate leaching function

Function	Estimated Parameters					Ψ
	1	s	s^2	R^2	R^2	
Nitrate Leaching (ml) [§] [kg/ha]	ϑ_w^1 4.1465E-01	ϑ_w^0 8.8139E+01	ϑ_n 8.8789E-02	s -2.3226E-02	s^2 4.0842E-04	R^2 0.9067
						R^2 0.9734
						R^2 0.9878

[§] Nitrate leaching function: $\Psi_{nl}(w, n, s) = (\vartheta_n(s) \cdot n) (1 + \exp(-\vartheta_w^1(s) (w - \vartheta_w^0(s))))^{-1}$

Table 7: Cotton response functions

Functions	Estimated Parameters							Ψ	
	1	s	s^2	s^3	\sqrt{s}	R^2	R^2		
Water Uptake (w_{up}) [†] [cm]	b_w^1	2.4758E-02	-5.5610E-02	4.6684E-04	-	2.3827E-01	0.9983		
	b_w^0	2.9370E+00	3.7825E-01	-3.8065E-03	-	-1.7042E+00	0.9672		
	b_n^1	3.4761E-02	-9.5432E-04	9.7281E-06	-	3.3671E-03	0.9865		
	b_n^0	-1.0048E+03	-6.9806E+01	4.3059E-01	-	4.4738E+02	0.9913	0.9850	
	d_0	-1.9310E-01	-4.8508E-01	3.6268E-03	-	2.4483E+00	0.9997		
	d_1	3.0048E-02	1.3073E-03	1.6158E-05	-	-7.4888E-03	0.9932		
	d_2	2.2604E-01	2.3395E-02	-1.7739E-04	-	-1.3251E-01	0.9749		
	b_w^1	4.1347E-02	-9.5788E-04	1.3248E-04	-2.0094E-06	-	0.7922		
	b_w^0	-8.1342E+01	3.5931E+01	-1.1290E+00	8.4346E-03	-	0.9218		
Nitrogen Uptake (n_{up}) [‡] [kg/ha]	b_n^1	1.0014E-02	8.4480E-05	-1.5166E-05	2.5512E-07	-	0.9233		
	b_n^0	-5.7776E-02	-4.0057E-02	-4.5261E-02	9.4218E-04	-	0.8831	0.9301	
	d_0	-2.7723E+00	7.0878E-01	-2.5691E-02	2.3649E-04	-	0.9786		
	d_1	2.3927E-02	1.4263E-02	-6.7201E-04	7.7165E-06	-	0.7584		
	d_2	3.1032E+00	-2.2247E+00	1.4816E-01	-1.9775E-03	-	0.9799		
	b_w^1	4.2910E-02	4.1989E-02	-7.8219E-04	-	-	0.9969		
	b_w^0	4.5197E+00	9.8234E-02	-	-	-1.3980E+00	0.9924		
	b_n^1	2.8070E-02	2.1351E-03	-2.6165E-06	-	-	0.9586		
	b_n^0	4.1684E-02	8.3436E-04	-	-	-1.2605E-02	0.9749	0.9135	
Relative Yield (ry) [‡] [-]	d_0	4.3794E-01	2.8471E-01	-5.2277E-03	-	-	0.9618		
	d_1	2.7633E-02	-7.8865E-04	3.8162E-05	-	-	0.9992		
	d_2	4.0487E-01	6.4077E-03	-	-	-1.0180E-01	0.9909		
	ϑ_w^1	1.2103E-01	-1.6859E-03	8.7523E-06	-	-	0.9285		
	ϑ_w^0	1.0767E+02	-1.3533E-01	-5.5516E-03	-	-	0.9966	0.9932	
	ϑ_n	2.4911E-01	3.7287E-03	-1.1199E-05	-	-	0.9510		
	[†] Water uptake function: $\Psi(w, n, s) = (1 - \exp(-b_w^1(s)(\varphi(w)w - b_w^0(s))))(1 - \exp(-b_n^1(s)(n - b_n^0(s))))$ where $\varphi(w) = \frac{4 \exp(d_1(s)w + d_0(s))}{(1 + \exp(d_1(s)w + d_0(s)))^2} + d_2(s)$								
	[‡] Nitrogen uptake function or relative yield function: $\Psi(w, n, s) = (1 - \exp(-b_w^1(s)(\varphi(w)w - b_w^0(s))))(1 - \exp(-b_n^1(s)(n - b_n^0(s))))$ where $\varphi(w) = \frac{\Psi(w, n, s)}{(1 + \exp(d_1(s)w + d_0(s)))^2} +$ $(d_2(s))^2$								
	[§] Nitrate leaching function: $\Psi_{nl}(w, n, s) = (\vartheta_n(s) \cdot n)(1 + \exp(-\vartheta_w^1(s)(w - \vartheta_w^0(s))))^{-1}$								

Table 8: Small Grains response functions

Functions	Estimated Parameters							Ψ
	1	s	s^2	s^3	\sqrt{s}	R^2	R^2	
Water Uptake (w_{up}) [†] [cm]	b_w^1	5.4887E-02	8.8365E-04	-	-	-1.2660E-02	0.9992	
	b_w^0	-1.4706E+00	3.8352E-01	-	-	-4.6480E+00	0.9995	
	b_n^1	3.3459E-02	3.6858E-04	-	-	-5.2111E-03	0.9887	
	b_n^0	-1.0577E+03	2.2656E+01	-	-	-3.2040E+02	0.9999	0.9895
	d_0	-1.1065E+01	-1.7822E-01	-	-	2.6840E+00	0.9995	
	d_1	2.3098E-01	4.9613E-03	-	-	-5.5987E-02	0.9969	
	d_2	7.3158E-01	-2.3317E-02	-	-	1.2677E-01	0.9699	
	b_w^1	2.3093E-02	1.0020E-03	-5.4338E-05	8.2254E-07	-	0.9937	
Nitrogen Uptake (n_{up}) [†] [kg/ha]	b_w^0	-2.7373E+01	3.0280E-01	2.4283E-02	-2.8201E-04	-	0.9995	
	b_n^1	7.5630E-03	-1.4065E-04	2.6628E-06	-1.1513E-08	-	0.9914	
	b_n^0	3.5255E+00	-9.6064E-01	2.2256E-02	-1.0843E-04	-	0.9979	0.9760
	d_0	3.4820E+00	-5.9384E+00	2.4480E-01	-2.0653E-03	-	0.9677	
	d_1	-7.8489E-02	8.8699E-03	-4.2253E-03	4.8533E-05	-	0.9799	
	d_2	6.9359E-01	8.8347E-02	8.0244E-04	-2.2655E-05	-	0.9995	
	b_w^1	2.1015E-02	-3.7979E-04	3.1736E-06	-	-	0.9999	
	b_w^0	-9.7590E+00	-2.4221E-01	2.5489E-03	-	-	0.9990	
Relative Yield (ry) [†] [-]	b_n^1	1.7967E-02	-8.0471E-04	6.1540E-05	-	-	0.9997	
	b_n^0	6.9662E-02	-2.6187E-03	2.1165E-05	-	-	0.9899	0.9920
	d_0	-3.3623E+00	8.7670E-02	-7.6724E-04	-	-	0.9931	
	d_1	8.8470E-02	-1.4634E-03	2.3777E-05	-	-	0.9945	
	d_2	9.4826E-01	-1.4023E-02	2.2275E-05	-	-	0.9918	
	ϑ_w^1	7.2178E-01	8.0706E-03	-	-	-1.2705E-01	0.9987	
	ϑ_w^0	8.1202E+01	-2.8048E-01	-	-	6.7213E-01	0.8654	0.9732
	ϑ_n	2.8377E-02	1.8152E-03	-	-	-5.6472E-03	0.9998	

[†] Water uptake function, nitrogen uptake function, or relative yield function: $\Psi(w, n, s) = (1 - \exp(-b_w^1(s)(\varphi(w)w - b_w^0(s))))(1 - \exp(-b_n^1(s)(n - b_n^0(s))))$ where $\varphi(w) = \frac{4 \exp(d_1(s)w + d_0(s))}{(1 + \exp(d_1(s)w + d_0(s)))^2} + d_2(s)$

[§] Nitrate leaching function: $\Psi_{nl}(w, n, s) = (\vartheta_n(s) \cdot n)(1 + \exp(-\vartheta_w^1(s)(w - \vartheta_w^0(s))))^{-1}$

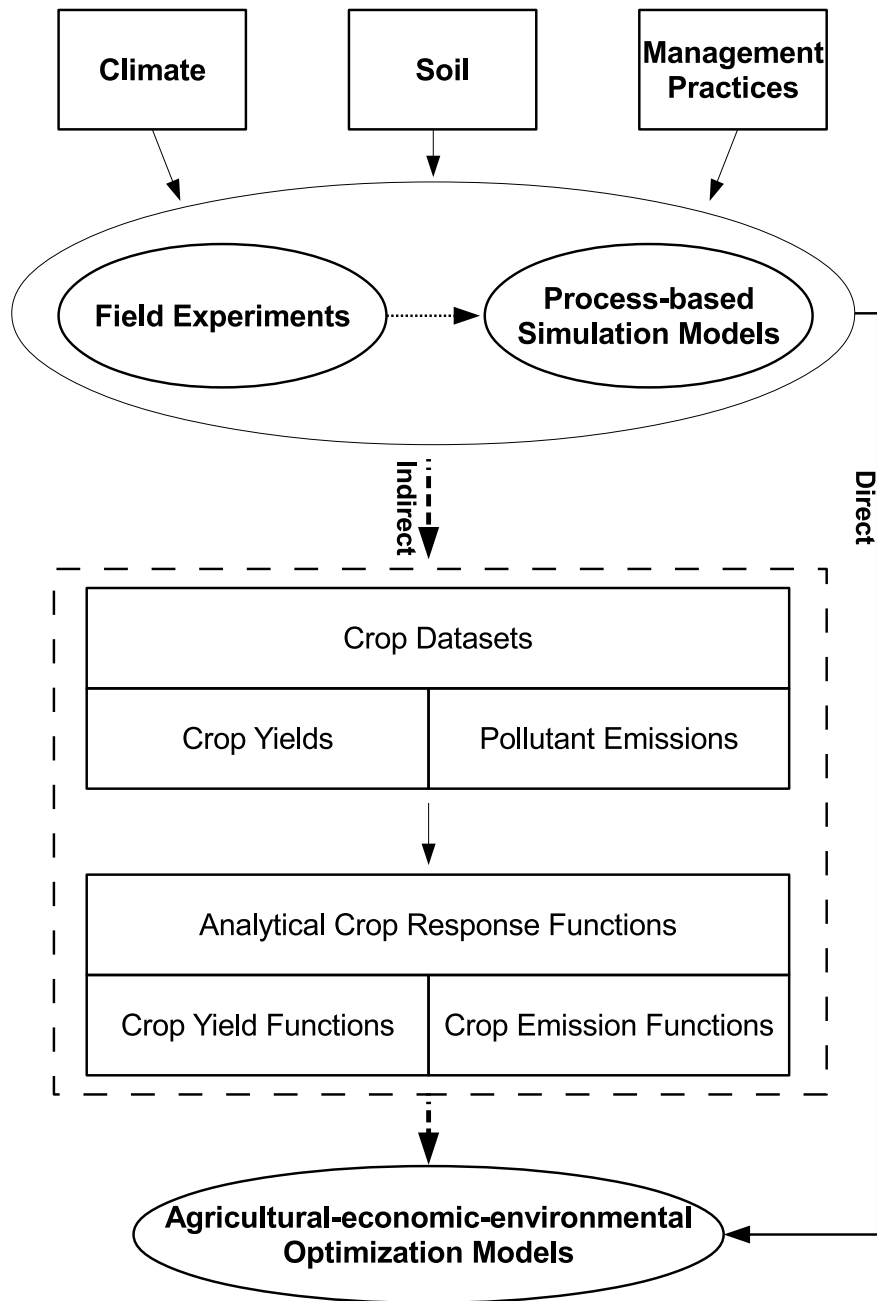


Figure 1: Alternative approaches for linking process-based simulation models with optimization models

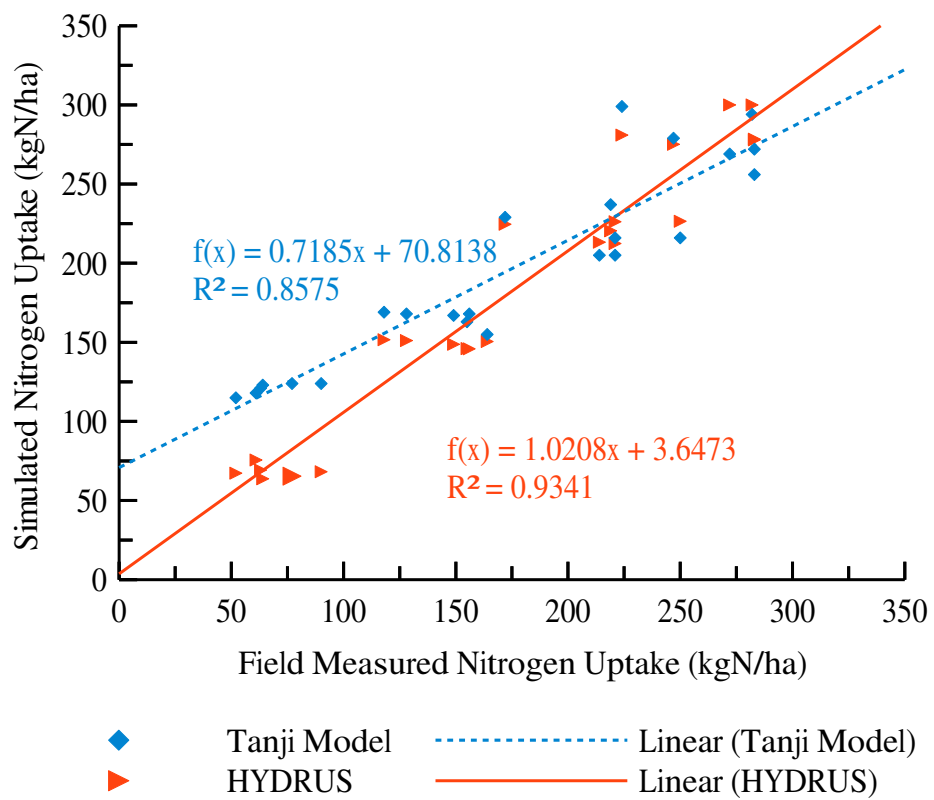


Figure 2: Field measured nitrogen uptake vs. simulated nitrogen uptake from Tanji et al. (1979) model & from HYDRUS-1D

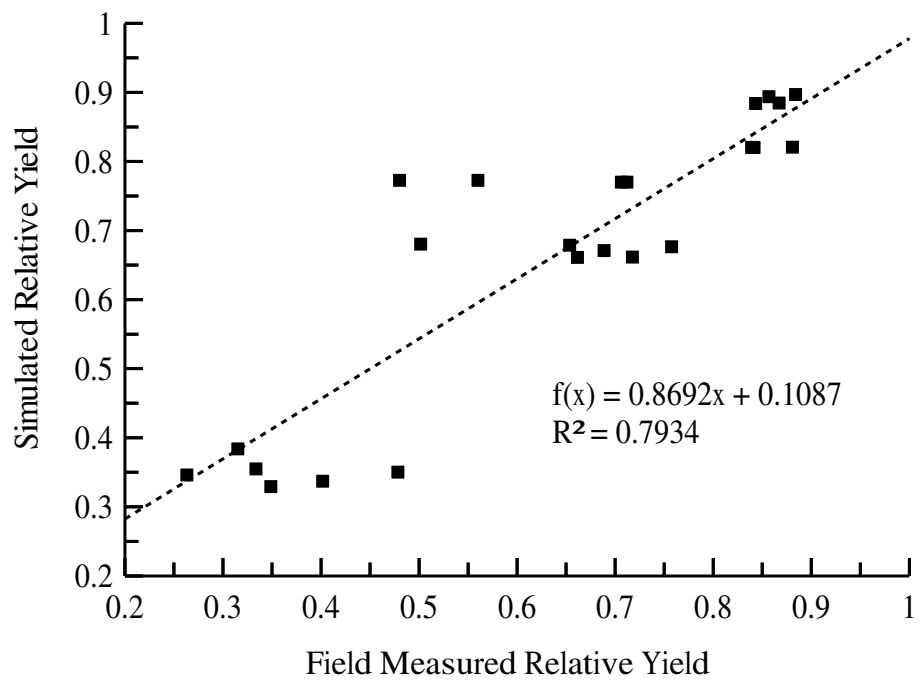


Figure 3: Field measured relative yield vs. simulated relative yield from our model

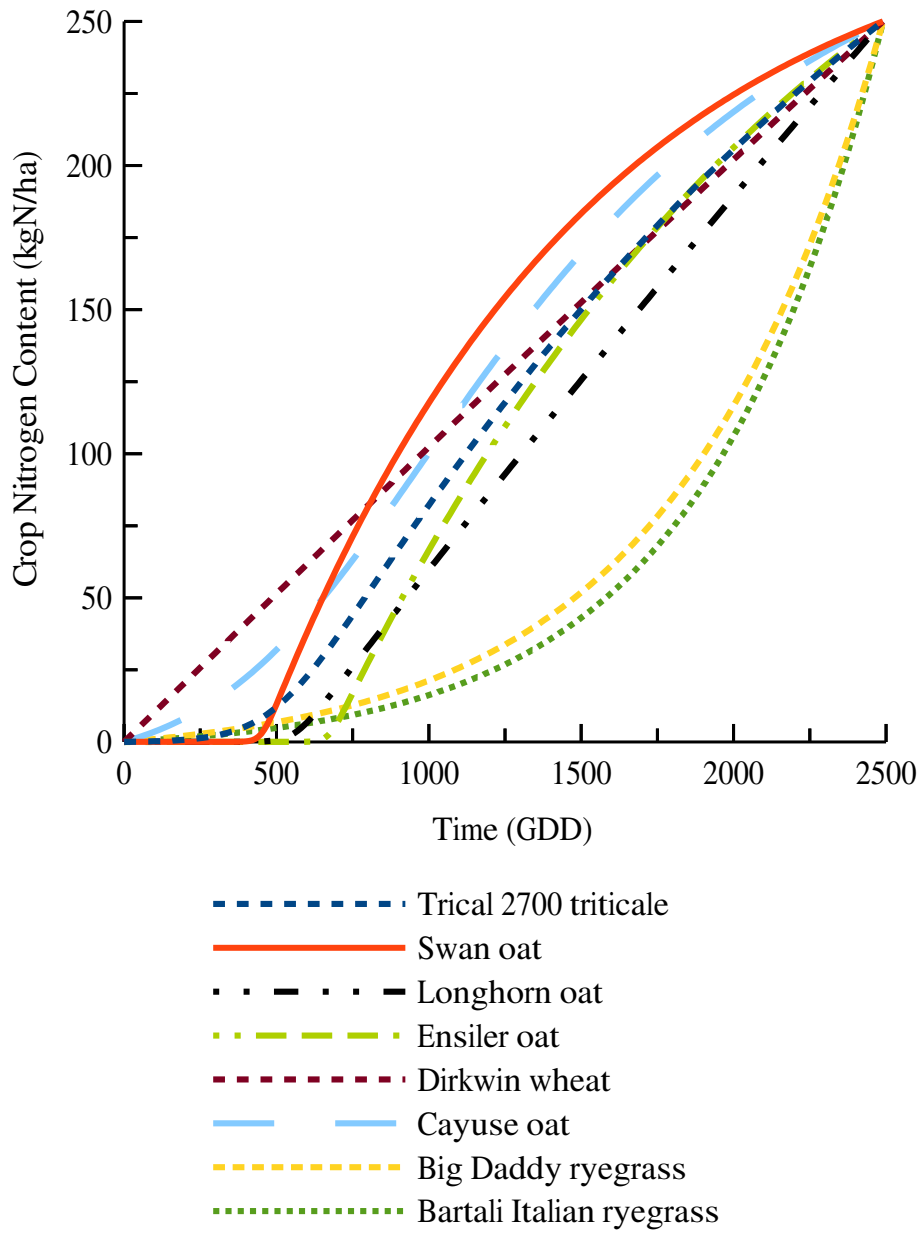


Figure 4: Nitrogen uptake curves for eight small grain forages commonly grown in California. Source: Crohn et al. (2009).

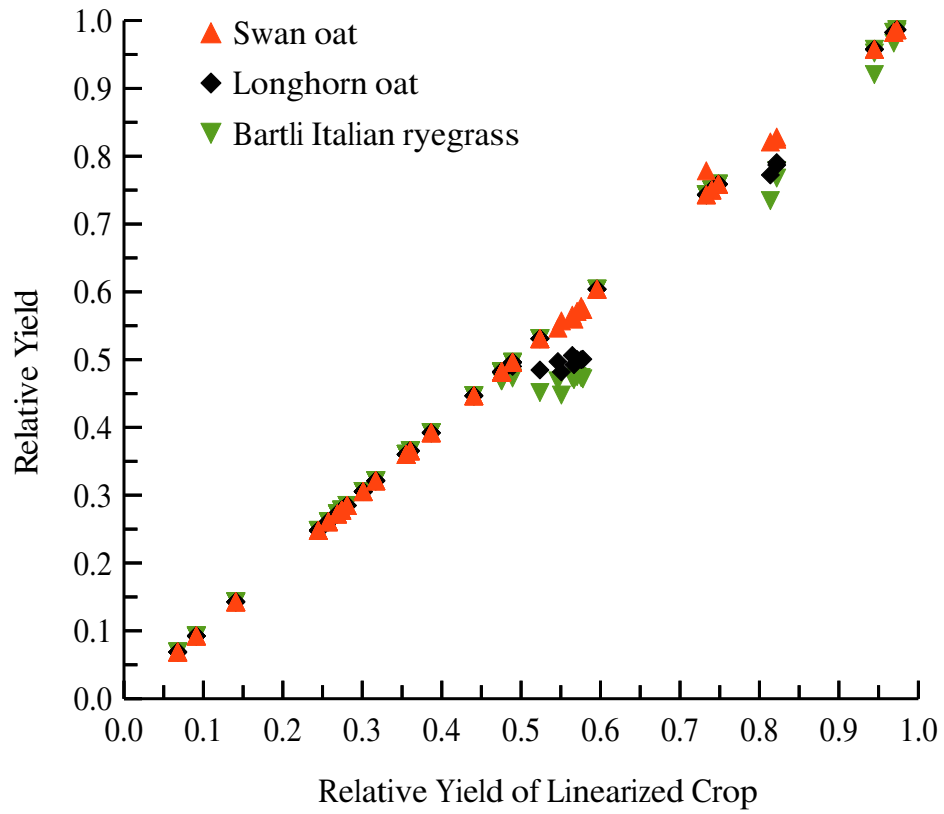


Figure 5: Relative yield of the linearized crop vs. relative yield of Swan oat, Longhorn oat, and Bartali Italian ryegrass

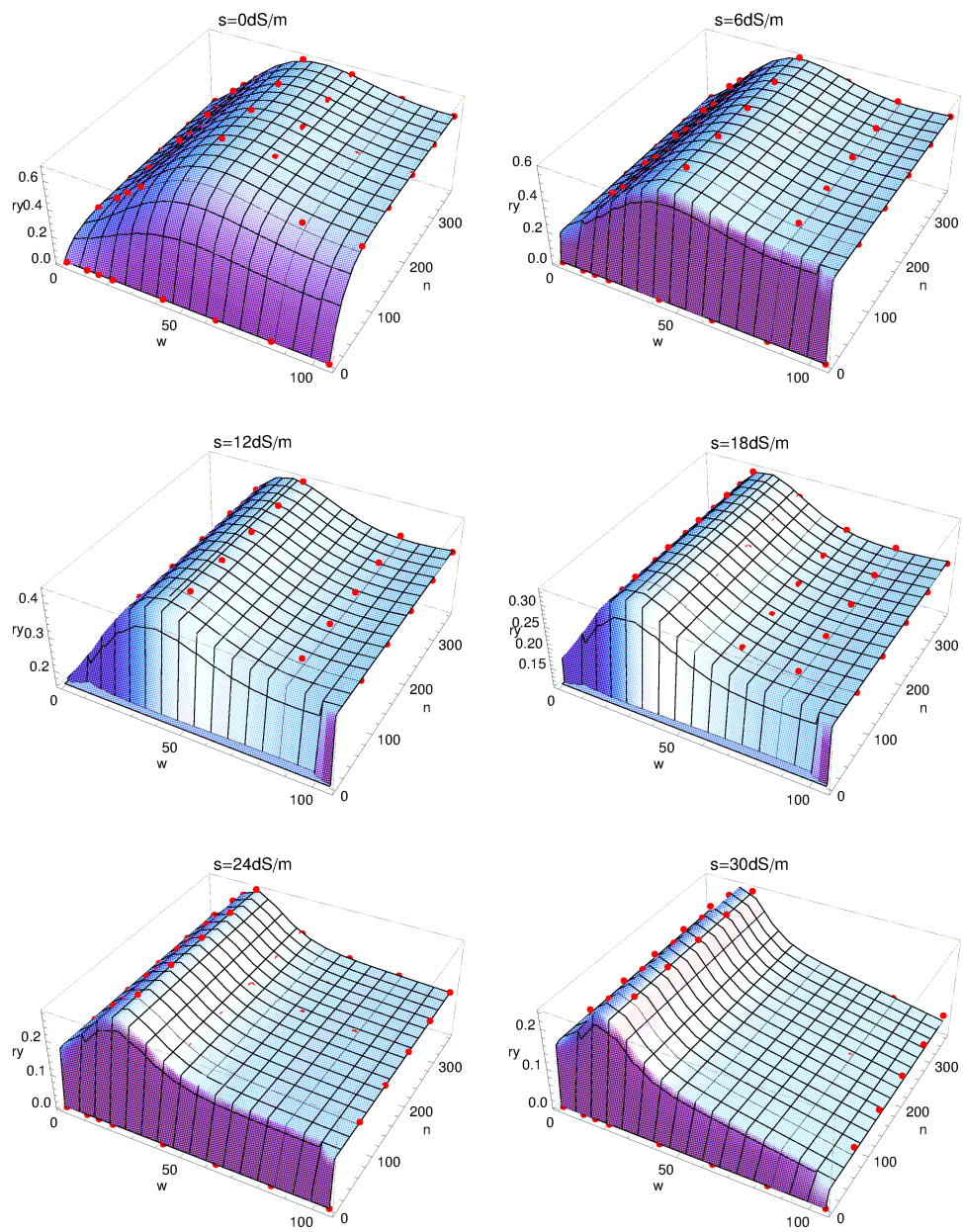


Figure 6: Relative yield vs. available water and available nitrogen for corn when soil salinity is 0, 6, 12, 18, 24, and 30 dS/m. Points: simulated data. Surfaces: fitted functions.

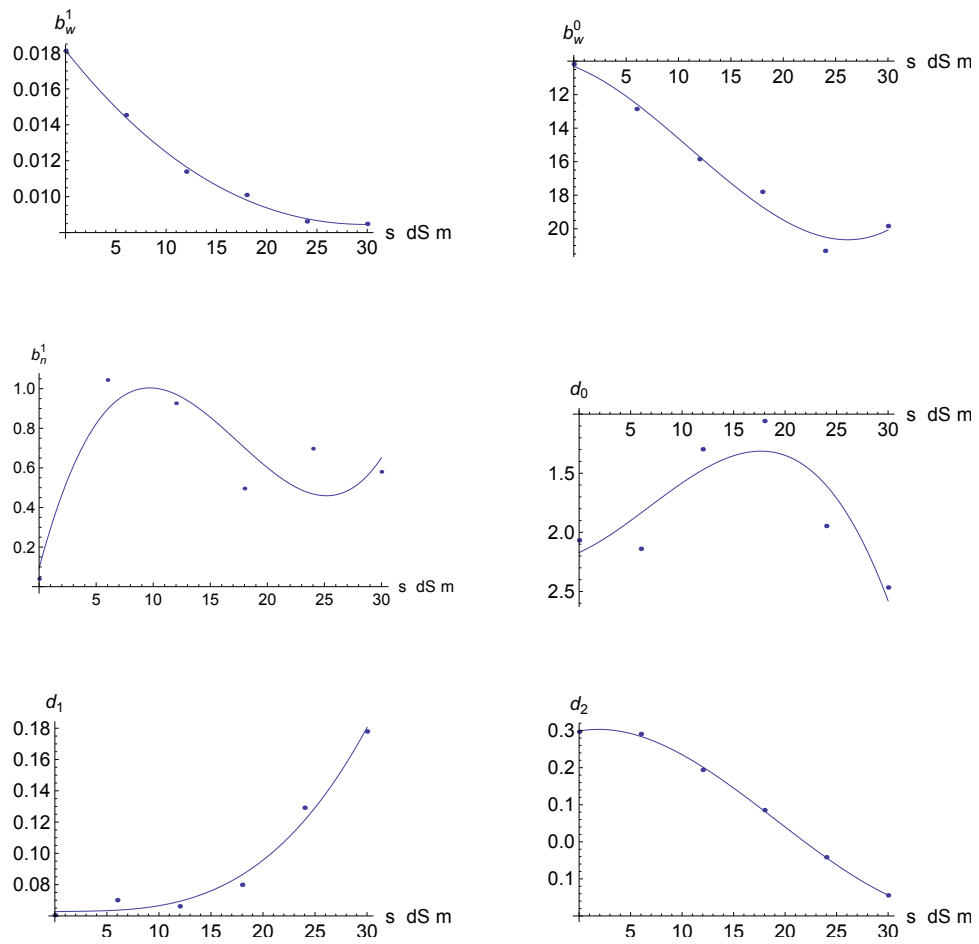


Figure 7: Polynomial regression of water and nitrogen parameters in the relative yield function for corn

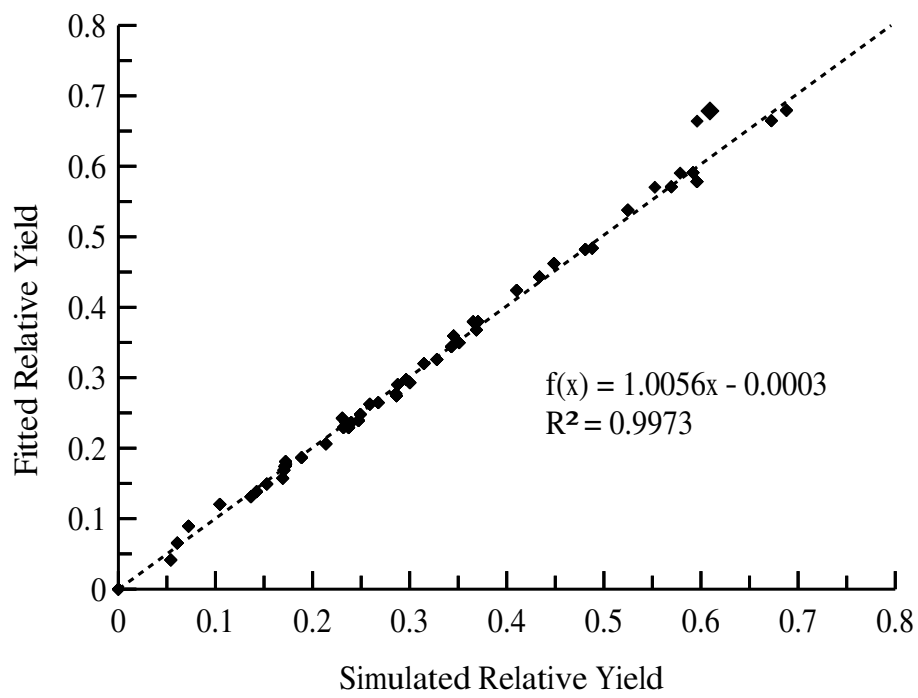


Figure 8: Relative yield function for corn: simulated data vs. fitted data

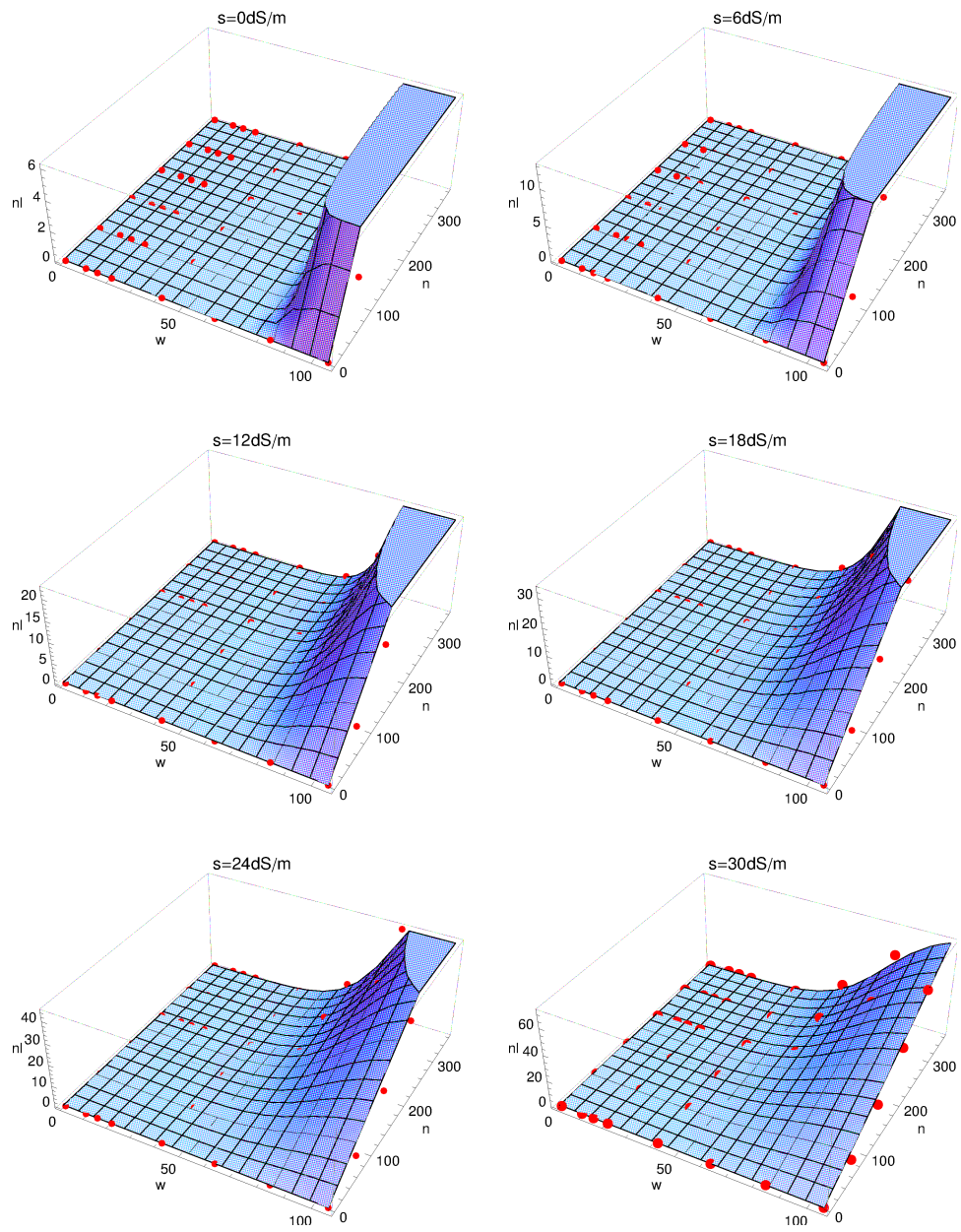


Figure 9: Nitrate leaching vs. available water and available nitrogen for corn when soil salinity is 0, 6, 12, 18, 24, and 30 dS/m. Points: simulated data. Surfaces: fitted functions.

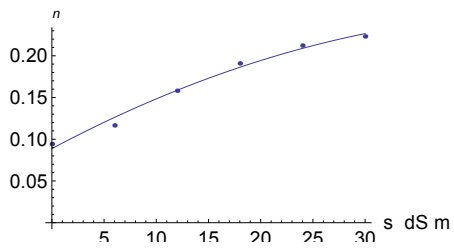
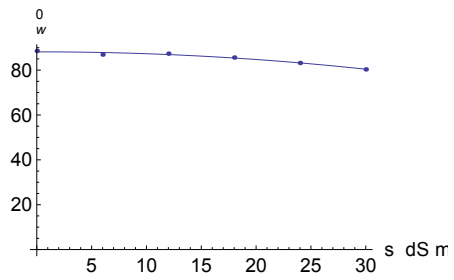
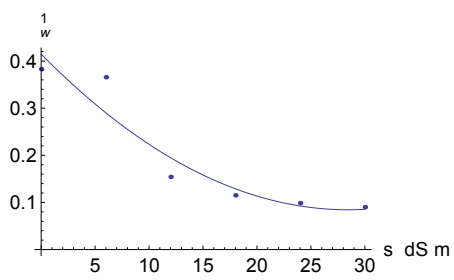


Figure 10: Polynomial regression of water and nitrogen parameters in the nitrate leaching function for corn

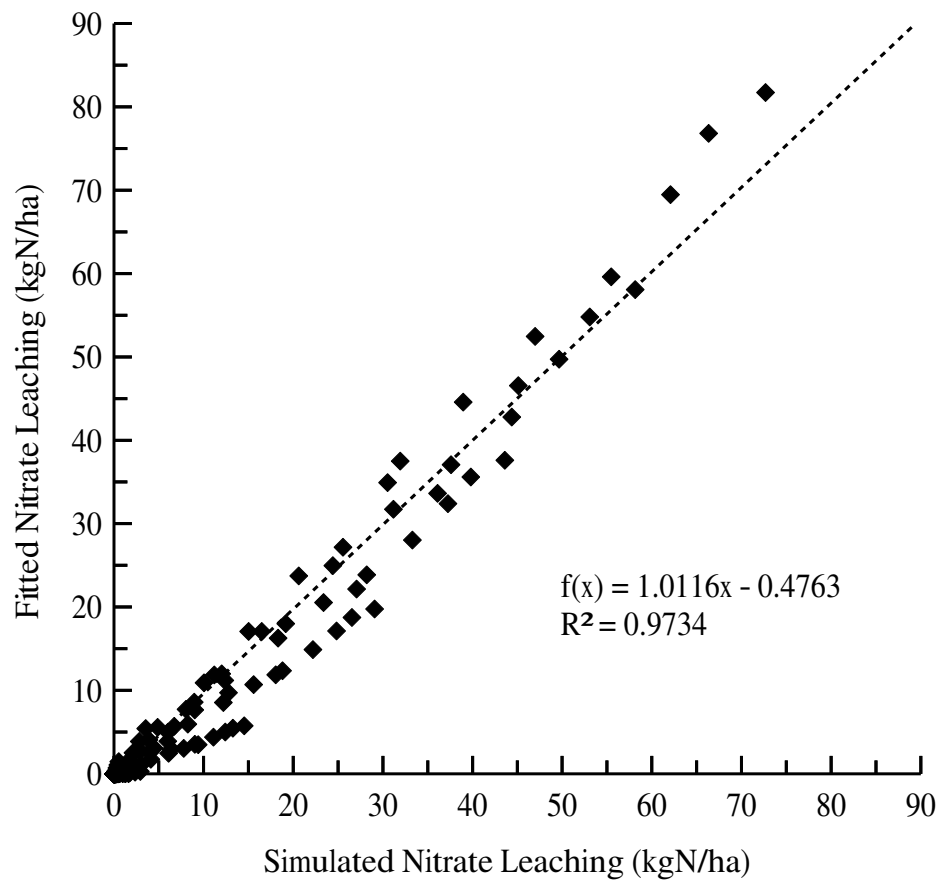


Figure 11: Nitrate leaching function for corn: fitted data vs. simulated data

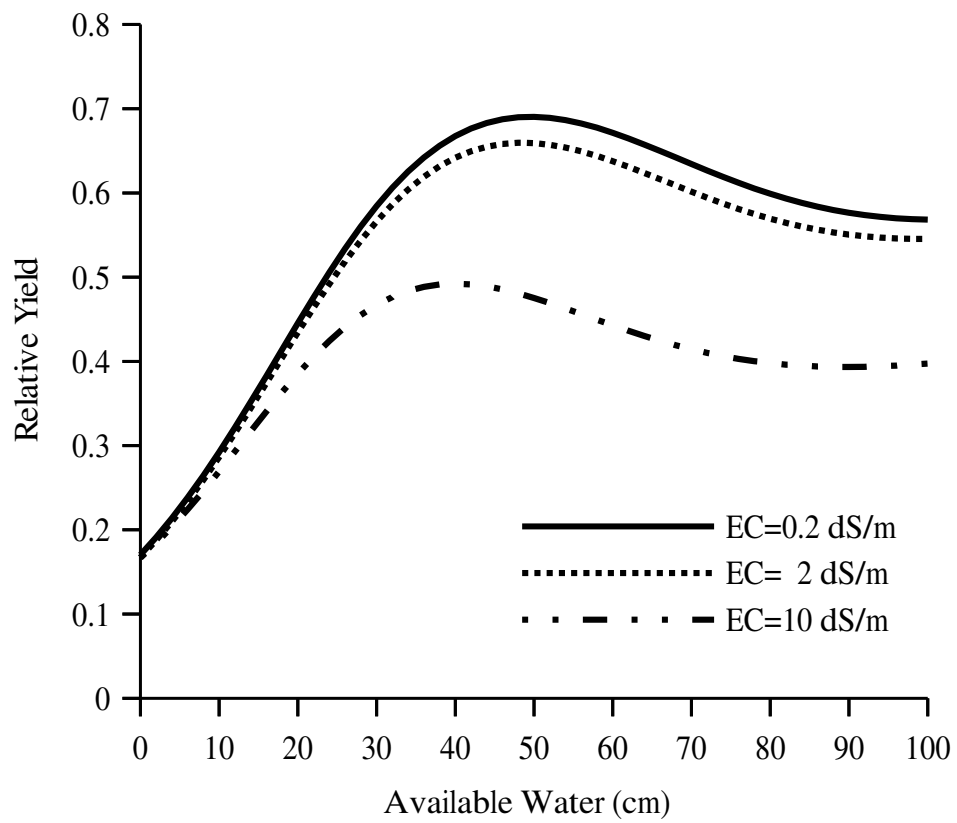


Figure 12: Interactions of water, nitrogen, and salinity for corn relative yield

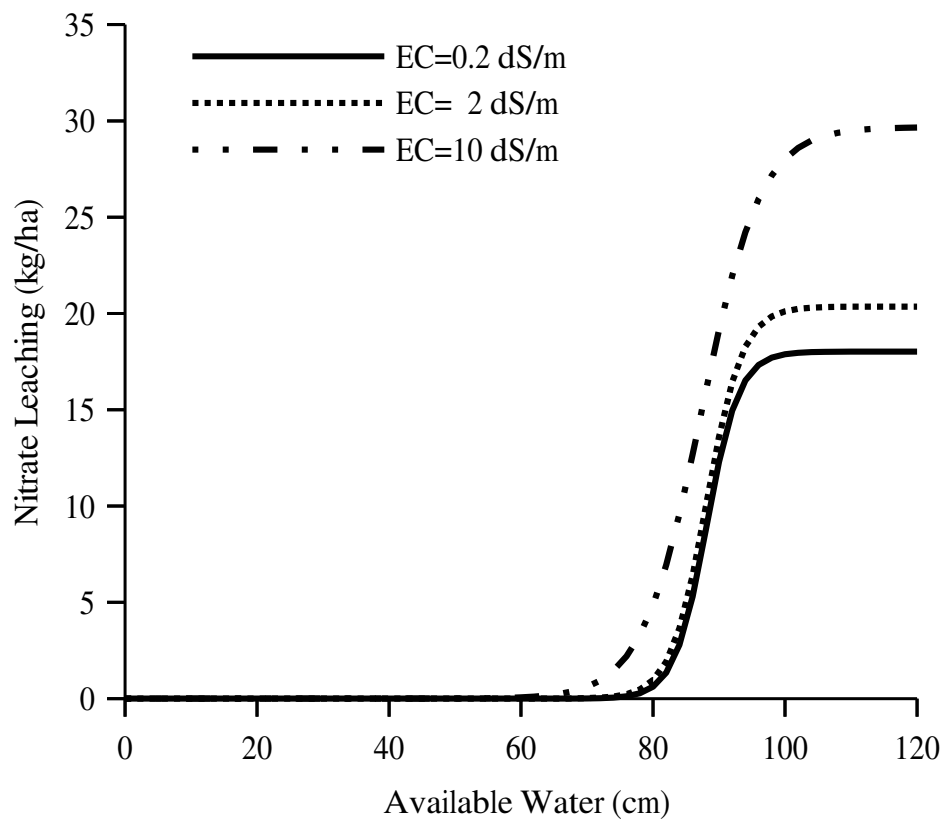


Figure 13: Interactions of water, nitrogen, and salinity for corn nitrate leaching