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1 **Mortality and community changes **drive** sudden oak death impacts on litterfall and soil**
2 **nitrogen cycling**

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12

13 **Summary**

14

15 1) Few studies have quantified the effects of pathogen caused tree mortality to ecosystem
16 processes despite that generalist pathogen impacts differ across species and ecosystems.

17 2) We measured litterfall mass, litterfall chemistry, and soil N cycling associated with multiple
18 hosts along a gradient of mortality caused by *Phytophthora ramorum*, the cause of sudden oak
19 death.

20 3) In redwood forests, the epidemiological and ecological characteristics of the major overstory
21 species determine disease patterns and the magnitude and nature of ecosystem change. Bay laurel
22 (*Umbellularia californica*) has high litterfall N (0.992%), greater soil extractable NO₃-N, and
23 transmits infection without suffering mortality. Tanoak has moderate litterfall N (0.723%) and
24 transmits infection but also suffers extensive mortality that leads to higher extractable soil NO₃-
25 N. Redwood (*Sequoia sempervirens*) has relatively low litterfall N (0.519%), does not suffer
26 mortality or transmit the pathogen but dominates forest biomass.

27 4) The strongest impact of pathogen-caused mortality was the potential shift in species
28 composition, which will alter litterfall chemistry, patterns and dynamics of litterfall mass, and
29 increase soil NO₃-N availability. Patterns of *P. ramorum* spread and consequent mortality are
30 closely associated with bay laurel abundances suggesting this species will drive both disease
31 emergence and subsequent ecosystem function.

32

33 Key words: Emerging infectious disease, ecosystem ecology, redwood forests, community-
34 pathogen feedback, N mineralization, nitrification, *Phytophthora ramorum*

35

36 **Introduction**

37

38 Pathogens are powerful ecological and evolutionary forces that can rapidly influence the
39 structure of plant communities through landscape-to-regional tree population declines (Holt et al.
40 2003; Burdon et al. 2006; Loo 2009). Both native and exotic pathogens can be important causes
41 of tree mortality, but the respective drivers and dynamics of outbreak may be very different.
42 Widespread tree mortality can be triggered when pathogens are introduced to naïve host
43 populations where natural enemies and host defenses are absent or ineffective. In contrast,
44 widespread tree mortality caused by native pathogens or insects may follow host distribution
45 shifts, changes in management, and weather or climatic driven increases in pest or pathogen
46 populations (Raffa et al. 2008; Worrall et al. 2010; Hawkins and Hinkel 2011; McDowell et al.
47 2011). Generalized ecosystem theory predicts that pathogen outbreaks which alter host or
48 community characteristics will in turn alter ecosystem processes such as N cycling, litterfall
49 dynamics, and decomposition (Ellison et al. 2005; Lovett et al. 2006; Eviner and Likens 2008).
50 However, few field studies have quantified pathogen impacts to ecosystem processes which
51 limits understanding of the affects of pathogens on landscape-level biogeochemistry and the
52 implication of these impacts on global change (Hicke et al. 2012).

53 In contrast to the lack of empirical studies of pathogen impacts to ecosystem processes,
54 several authors have described useful conceptual frameworks that link host and pathogen
55 characteristics with mechanistic changes to functional process (Burdon et al. 2006; Lovett et al.
56 2006; Eviner and Likens 2008). The theoretical foundations of pathogen impacts on ecosystems
57 are corollaries to insect outbreak, and field study of insect outbreak provides guidance in the
58 formulation of hypotheses to study ecosystem-level pathogen impacts (Hunter 2001; Hicke et al.

59 2012). For example, foliar chemistry changes caused by foliar-feeding insects have been linked
60 to altered litterfall chemistry and decomposition rates (Lovett et al. 2002; Russell et al. 2004;
61 Chapman et al. 2006). Similarly, bark beetle outbreak has been shown to increase litterfall %N
62 under dead trees, presumably due to the arrest of nutrient resorption (Morehouse et al. 2008;
63 Griffin and Turner 2012). Mortality-related canopy damage can alter microclimate and
64 subsequent rates of soil N cycling (Classen et al. 2005; Orwig et al. 2008) while shifts in species
65 composition can cause long-term shifts in fundamental ecosystem processes that control N and C
66 dynamics (Ruess et al. 2009; Cobb 2010; Lovett et al. 2010).

67 Pathogens infect different host tissues (leaves, tree boles, roots), cause selective mortality
68 among canopy species, and may lead to species shifts within communities, suggesting that
69 epidemiological processes drive variation in ecosystem function during, and well after, the
70 emergence of disease (Burdon et al. 2006; Lovett et al. 2006). At the local scale, the timing and
71 extent of ecosystem change is [likely](#) driven by: (1) host characteristics including biomass, unique
72 function (shade tolerance, N fixation, phenology), and (2) host epidemiological characteristics
73 including susceptibility, competency to transmit infection, and consequences of infection to host
74 health (Eviner and Chapin 2003; Ellison et al. 2005; Lovett et al. 2006; Eviner and Likens 2008).
75 Although epidemiological models can be accurately applied across broad spatial scales (Gilligan
76 and van den Bosch 2008; Meentemeyer et al. 2011; Filipe et al. 2012), the lack of data on
77 ecosystem-level pathogen impacts limits our ability to test and accurately apply these models in
78 analyses of C or N cycling in landscapes shaped by disease (Lovett et al. 2006; Hicke et al.
79 2012).

80 *Phytophthora ramorum*, an oomycete pathogen which causes the forest disease sudden
81 oak death, is an example of an exotic [pathogen of unknown origin](#) which has resulted in region-

82 scale tree mortality and ecosystem change (Rizzo et al. 2005; Cobb et al. 2012a). *P. ramorum*
83 has a broad host range but susceptibility, competency to transmit infection, and impacts to host
84 health vary independently across hosts. For example, coast redwood (*Sequoia sempervirens*)
85 foliage has low-to-moderate susceptibility, supports little sporulation, and the tree does not suffer
86 mortality following infection (Davidson et al. 2005; Maloney et al. 2005). Redwood has very
87 little influence on the spread and impacts of *P. ramorum*, but is common in cool, wet
88 environments also favorable to the pathogen (Davidson et al. 2011). In contrast, susceptibility
89 and sporulation from California bay laurel (*Umbellularia californica*) foliage is high and drives
90 pathogen spread at stand-to-landscape scales, but infection has no known negative impacts on
91 bay laurel health (Davidson et al. 2008; DiLeo et al. 2009; Meentemeyer et al. 2011).
92 Susceptibility and sporulation from tanoak (*Notholithocarpus densiflorus*) twigs and foliage is
93 epidemiologically significant but, unlike redwood and bay laurel, tanoak tree boles are also
94 susceptible and infection causes bole-cankers that can lead to mortality in as little as two years
95 (Cobb et al. 2012b).

96 Predicting which exotic organisms are likely to establish and cause deleterious impacts to
97 natural resources remains an important but tremendously challenging goal of ecology.
98 Eradication of many wide-spread exotic pathogens is unrealistic and further introduction of
99 damaging microorganisms is virtually certain to continue (Balci et al. 2007; Loo 2009; Santini et
100 al. 2012). This increases the importance of understanding ecosystem-level impacts caused by
101 pathogen outbreak. In this study we focus on three mechanisms by which pathogens may alter
102 ecosystem processes that have been previously documented as drivers of ecosystem change
103 during insect outbreak: 1) direct impacts of pathogens and host mortality on litterfall chemistry,
104 2) mortality driven changes to soil N cycling and litterfall dynamics, and 3) the long-term

105 implications of pathogen-mediated community changes to litterfall and soil N cycling. Our field
106 study has three objectives which parallel these mechanisms: 1) to examine the respective effects
107 of pathogen prevalence in bay laurel and mortality in tanoak on litter N chemistry, 2) to quantify
108 the effects of disease-caused mortality to soil N cycling, litterfall amounts and litterfall
109 chemistry, and 3) to describe litter and soil N dynamics associated with each of the major
110 overstory species in redwood forests impacted by sudden oak death. *At the individual plant level*
111 *we hypothesized that *P. ramorum* infection would increase bay laurel %N and mortality would*
112 *increase tanoak litter %N given previous work demonstrating that infection increases bay laurel*
113 *leaf senescence rates (Davidson et al. 2011) and litterfall N increases in bark beetle killed trees*
114 *(Morehouse et al. 2008; Griffin and Turner 2012). We also expected stands with high levels of *P.**
115 **ramorum*-caused mortality would have increased soil N availability and mineralization rates*
116 *compared to stands without mortality given that other disease and insect caused tree mortality*
117 *has been demonstrated to alter these soil N dynamics (Hobara et al. 2001; Morehouse et al. 2008;*
118 *Orwig et al. 2008; Lovett et al. 2010; Griffin and Turner 2012). Lastly, we expected distinct*
119 *litterfall chemistry and soil N dynamics associated with the principle *P. ramorum* host species*
120 *given that species identity is a critical control over litter chemistry and soil N dynamics (Fried et*
121 *al. 1990; Finzi et al. 1998; Eviner and Chapin 2003; Cobb 2010). We accomplish these*
122 *objectives by combining litterfall and soil N cycling measured across a gradient of pathogen*
123 *prevalence and tanoak mortality with a controlled study of species influences on soil N*
124 *dynamics.*

125 **Methods**

126

127 Field sites and study design

128 We conducted measurements of litterfall from January 2007 to December 2009 (3 years)
129 and soil N cycling from December 2007 to December 2009 (2 years) at two sites where disease
130 and vegetation dynamics had been monitored during annual summer surveys from 2002-2007
131 (Cobb et al. 2012b). From a pool of potential study sites, we selected Jack London State Park
132 (Jack London) and the Marin Municipal Water District (MMWD) located in Sonoma and Marin
133 Counties (CA, USA), respectively. Both sites are notable for species composition, landuse, and
134 disease history characteristic of the broader region. Plots were selected so that soil types were
135 common at each site: Goulding clay loam at Jack London and a Tocaloma-McMullin complex at
136 MMWD. In 2002, 30 plots were established at each site; study plots are circular, 500 m², and
137 randomly located with at least 100m between each plot (see Maloney et al. 2005). At the time of
138 establishment, each stem greater than 1 cm diameter at breast height (dbh; 1.3 m height) was
139 measured for diameter, mapped, and symptomatic tissue was returned to the laboratory for
140 pathogen isolation in a *Phytophthora* selective medium (PARP; see Davidson et al. 2008). In the
141 autumn of 2006, we identified a subset of these plots (15 at each site) that span the range of
142 pathogen prevalence (number of infected hosts) and disease severity (tanoak mortality) at each
143 site. We use the strict criteria of *P. ramorum* recovery via laboratory culturing as the criteria for
144 considering an individual infected; however, mortality was assessed at the stem-level meaning
145 that stems could have been killed by *P. ramorum* yet a multi-stemmed or resprouting individual
146 could may remain living. Our study design is predicated on the expectation that changes in
147 ecosystem processes would be a function of pathogen prevalence, the local amount of host
148 biomass that could be killed by the pathogen (tanoak biomass), and the cumulative host biomass
149 which had been killed by *P. ramorum* at the initiation of measurements (dead tanoak biomass;
150 see Lovett et al. 2006). Specifically, the selected plots range in initial tanoak basal area from 0.12

151 to 35.5 m² ha⁻¹ and cumulative mortality from 0.05 to 33.7 m² ha⁻¹. We forego a two-level
152 pathogen invaded vs. non-invaded design in favor of relating the amount of variation in
153 ecosystem processes to [infection \(prevalence of infected hosts\)](#) and [mortality \(dead tanoak basal](#)
154 [area\) given the initial tanoak basal area](#) (c.f. Lovett et al. 2010). Prevalence of infection at the
155 plot level ranged from 66-100% of bay laurel stems and 6-95% of tanoak stems. Many study
156 plots are notable for almost complete tanoak mortality while other plots have suffered almost no
157 mortality even though tanoak is basal area is substantial (11-15 m² ha) and pathogen populations
158 have been present since the initial survey in 2002 (Maloney et al. 2005). [This variation forms a](#)
159 [gradient of disease impacts across plots with different host composition](#). Our study shares some
160 of the same limitations of space-for-time designs in that it does not distinguish between
161 responses of the disease to ecosystem function and ecosystem function responses to disease. To
162 address this circularity, we conducted a second measurement of N cycling under common
163 temperature and moisture conditions in the laboratory using soils collected from redwood, bay
164 laurel, tanoak, and recently killed tanoak trees located outside of our study plots. This provided
165 an independent assessment of the relative influence of dominant overstory species and tanoak
166 mortality on soil N cycling (c.f. Freid et al. 1990; Finzi et al. 1998). Further detail regarding
167 community, pathogen, and disease characteristics can be found in Table S.1 (Supplemental
168 Information).

169

170 Field litterfall and soil N cycling measurements

171 [Three 1935.48 cm² plastic litter traps were established in each plot \(~0.58 m² collection](#)
172 [area\) in January 2007 and July 2007 at the Jack London and MMWD sites respectively. Large](#)
173 [holes were cut into the trap floor, traps were lined with 1 mm mesh screen, and the trap was](#)

174 elevated 10-15 cm above the forest floor surface. This design allows free flow of precipitation
175 and air which effectively air-dried litter between collections; we found no evidence of litter
176 decomposition within our traps (e.g., discoloration, fungal hyphae). For the first two years of
177 measurement, litter was collected eight times per year (every 4 to 8 weeks) until seasonal
178 patterns of litterfall were established for each species; during the final year of measurements
179 litter was collected every 12 weeks. Litter samples were air dried in the laboratory at 45 C° for
180 48 hours when precipitation occurred between samplings and stored in paper bags 1-12 weeks
181 before processing. Foliar litter was sorted by major overstory species (redwood, tanoak, bay
182 laurel, madrone – *Arbutus menziesii*, Douglas fir – *Pseudotsuga menziesii*) and the remaining
183 material was sorted, without regard to species, into woody litter and all other material which
184 included fruit, flower parts, herbaceous plant litter, bryophytes, seeds, and occasionally insect
185 bodies. Bay laurel foliar litterfall was further assessed for the frequency of symptoms on a leaf-
186 by-leaf basis for each sample collection by qualitatively assessing the proportion of necrotic
187 tissue. After sorting, each sample was dried at 60 C° for 48 hours, weighed, and archived for
188 later chemical analysis. Litterfall chemistry was not measured for each sampling due to
189 insufficient litterfall mass at some collection dates. Rather, after the two years of measurement it
190 became clear quarterly-periods corresponding to winter (Jan-Mar), spring (Apr-Jun), summer
191 (Jul-Sept), and autumn (Oct-Dec) reflect the major seasonal changes in litterfall mass for tanoak,
192 bay laurel, and redwood in our study plots. Therefore, we composited, analyzed C and N
193 concentration, and calculated litterfall N mass on this quarterly basis.

194 We assessed soil net N mineralization and net nitrification of the surface 20 cm of
195 mineral soil with a field-incubation of intact soil cores. At two locations in each plot, we
196 removed the forest floor layer and drove a 27 cm long, 5.08 cm diameter PVC tube, 22 cm into

197 the mineral soil. The bottom 2 cm of soil was carefully removed and replaced with a nylon mesh
198 bag filled with ~10 g of IRN 150 ion exchange resin (Amberlite™) and fitted with a rubber ring
199 which held the soil in the core. This yielded an open-top, open-bottom core which allowed free
200 water movement during the 10-28 week field incubation. A second core was used to sample the
201 top 20 cm of mineral soil and establish initial NH₄-N and NO₃-N concentration. For both
202 incubated and initial cores, the PVC tube was emptied in the field, soil samples were transported
203 back to the laboratory on ice, and processed within 48 hours. Each sample (incubated and initial)
204 was sieved to pass a 2 mm screen; a subsample was dried for 48 hours at 105 C° to determine
205 moisture content and a second subsample was analyzed for inorganic N by gently shaking 10g of
206 field moist soil in 1M KCl for 0.5 hrs and filtering the extract through a 0.45µm pore-size glass-
207 fiber filter. NO₃-N and NH₄-N concentration of this extract was measured with a sulfanilamide
208 reaction after reduction in a copperized cadmium column and a salicylate method, respectively,
209 at the UC Davis Analytical Laboratory (QuikChem Methods 12-107-04-1-B and 12-107-06-2-A,
210 respectively; Lachat Instruments, Loveland, CO;).

211

212 Laboratory soil mineralization measurement

213 We conducted a laboratory incubation designed to examine the influence of individual
214 species and tanoak mortality on inorganic N availability and mineralization among species under
215 common environmental conditions. In April 2009, we selected eight redwood, healthy tanoak,
216 bay laurel, and tanoak where the main stem had been killed by *P. ramorum* (N = 32). These trees
217 were located at the Jack London site and chosen in sets of four such that each tree was between
218 10-40 m of the others in its set, and each set was separated by at least 150 m. We sampled the
219 surface 20 cm of mineral soil at eight locations within 2 m of each individual tree using a 6.60

220 cm diameter stainless steel soil-core and composited samples in the field. These samples were
221 transported, processed, and analyzed with the same methods described for N mineralization
222 measurements. Two subsamples for each tree were measured for initial soil moisture, KCL
223 extractable $\text{NO}_3\text{-N}$ and $\text{NH}_4\text{-N}$ (64 total). Soil collection occurred within two days of significant
224 rainfall, initial soil moisture content did not significantly differ among species, and soil moisture
225 was at adequate levels to support microbial processes for the five week incubation (range 0.40-
226 0.49 g g^{-1}), therefore soils were incubated at field moisture. For each tree we created 10 replicate
227 soil microcosms of ~50 g soil (sieved to pass a 2 mm screen) in 300 ml volume vented plastic
228 sample cups (320 total). Microcosms were incubated at 22 C° in a dark, climate-controlled space
229 and two microcosms from each tree were destructively sampled every week for 5 weeks to
230 estimate changes in N dynamics through time. Each microcosm was assessed for soil moisture,
231 KCL extractable $\text{NO}_3\text{-N}$ and $\text{NH}_4\text{-N}$. Tree-level data were taken as the average value from both
232 microcosms and the two initial measurements (N = 192).

233

234 Data analysis

235 We assessed the effects of disease and pathogen prevalence on litterfall and soil N
236 cycling with a series of linear models. Objective 1: To examine relationships between pathogen
237 prevalence and litter %N for bay laurel, and mortality and litter %N for tanoak we employed a
238 series of linear models for each season of collection where individual chemistry parameters (%C,
239 %N, C/N) were the dependent variables and infection (number infected bay laurel) or mortality
240 (dead basal area $\text{m}^2 \text{ ha}^{-1}$) was the independent variable. An identical model was used to assess
241 bay laurel litter chemistry and frequency of *P. ramorum* symptoms within individual samples.
242 Objective 2: We expected that disease caused changes in litterfall mass, N mass, and soil N

243 dynamics would be a joint function of the maximum potential disease impact (initial tanoak basal
244 area $\text{m}^2 \text{ha}^{-1}$) conditioned on the cumulative tanoak biomass killed by the pathogen (dead tanoak
245 basal area $\text{m}^2 \text{ha}^{-1}$). We analyzed annual litterfall mass and N amounts with a set of multivariate
246 repeated measures ANOVA models for redwood, tanoak, bay laurel, tanoak litter N, and total
247 (stand-level) litter N (N=90). We selected this ANOVA model because our litterfall parameters
248 were measured on a limited number of well defined categories (annual litterfall; c.f. Gotelli and
249 Ellison 2004) and the time-by-disease interactions are meaningful given that mortality occurs on
250 an annual basis. Here, the dependent variable (Y) for each species or litterfall component (i) at
251 time t was modeled as a function of the independent variables (X_i) conditioned on species
252 specific parameters ($b_{i,t}$), the respective annual estimated mean $\bar{Y}_{i,t}$, and a normally distributed
253 error term (ε): $Y_{i,t} = \bar{Y}_{i,t} + \sum X_i b_{i,t} + \varepsilon$. Models of soil N responses to tanoak mortality were
254 similar to those for litterfall except we used a mixed-model with time parameterized as a random
255 effect given that the timing of sampling was irregular throughout the two years of measurement
256 (c.f. Gotelli and Ellison 2004). These models also included a fixed effect of soil moisture
257 measured in the initial cores to examine potential moisture limitation to microbes among plots.
258 Objective 3: We assessed the potential impacts of species shifts by describing litter C/N and
259 local soil N dynamics associated with the major overstory species in our study plots. Differences
260 in litter %N, %C, and C/N content among species were assessed with a one-way mixed-model
261 ANOVA where species was the main effect and sampling date was a random effect; when the
262 main effect was significant, differences among species were assessed with Tukey's HSD test.
263 For our laboratory comparison of species effects on soil N dynamics, we were able to employ a
264 matched-pairs t-test for all possible pairs on the basis that each subject was grouped into
265 individual blocks. Variation in $\text{NO}_3\text{-N}$, total N pool sizes, net nitrification and N mineralization

266 rates associated with species identity and dead tanoak were assessed with identical models that
267 compared each tree-type individually with each of the others. For each linear model, normal
268 distribution and homogeneous variance of the residuals was assessed with goodness of fit tests to
269 the normal distribution and visual examination of heteroskedasticity; for the paired t-test analysis
270 normal distribution was tested for each variable. Square-root transformation was required for
271 litterfall chemistry, field-based soil N measurements, and soil N pool sizes for the laboratory
272 study. Analysis was performed with the JMP® version 8 with the critical value of $p \leq 0.05$ for
273 statistical significance.

274 **Results**

275

276 *Direct pathogen impacts on litter chemistry: Objective 1*

277 The plot-level prevalence of infected tanoak and bay laurel was not significantly
278 associated with annual litterfall mass in either species (data not shown). However, a modest,
279 positive effect of litterfall %N and prevalence of infection was found for bay laurel during the
280 spring and summer, but not during autumn and winter (Figure 1). The spring and early summer
281 seasons also correspond with peak periods of *P. ramorum* sporulation and within-tree infection at
282 the Jack London Site (see Davidson et al. 2011). We found a similar, negative, and statistically
283 significant relationship between bay laurel litterfall C/N and prevalence of infection (not shown).
284 The spring collections were also notable for relatively high %N concentrations and low C/N
285 relative to the other three seasons but this period also had the lowest mass of bay laurel litterfall
286 (Figure 2). Bay laurel %N and C/N were not significantly related to prevalence of symptomatic
287 leaves. Given that bay laurel contributed ~7-11% of overall litterfall N (Figure 2; Table S.2) and
288 that litterfall amounts were low when the pathogen may elevate foliar %N (decrease C/N), this

289 pathogen effect on the total N transfer from the canopy to the forest floor is subtle. In contrast to
290 bay laurel, no relationship between tanoak litterfall %N or C/N and tanoak mortality was found
291 for any season of comparison (Figure 1; C/N not shown).

292

293 *Mortality impacts on litterfall and soil N cycling: Objective 2*

294 Disease had significant effects on the mass of tanoak litterfall, tanoak litterfall N, and
295 total foliar litterfall N (Figure 3). Litterfall amounts were positively associated with the
296 respective pre-disease basal area for each species. But for tanoak, litterfall mass and N were also
297 negatively associated with cumulative tanoak mortality and the magnitude of these reductions
298 was also variable across years (interaction $p < 0.05$; Figure 3; Table S.3). The estimates from the
299 repeated measures model indicate *P. ramorum*-caused tanoak mortality resulted in up to 91%
300 reduction in tanoak litterfall and up to 95% reduction of tanoak litterfall N in plots with the
301 greatest amount of cumulative tanoak mortality (up to $\sim 33 \text{ m}^2 \text{ ha}^{-1}$ basal area). Even when tanoak
302 mortality was extensive, tanoak foliar litter (and tanoak foliar litter N) was still [part of the](#) overall
303 litterfall mass due to litter production from basal sprouts that frequently developed from *P.*
304 *ramorum* killed tanoak stems. Compared to other species collected in our litter traps, tanoak had
305 less seasonal variation (Figure 2). Even though redwood dominates litterfall N mass (65-78% of
306 total), total litter N (stand-level) [decreased with](#) tanoak mortality (Figure 3) probably because
307 tanoak litter has relatively high %N compared to redwood. Total foliar litterfall, woody litter,
308 and total litterfall (e.g. foliage, woody litter, and other materials) were not significantly
309 associated with disease and were relatively insensitive to forest structure across our plots (Table
310 S.4; supplemental information).

311 Total and NO₃-N pools were significantly **increased with** disease but rates of nitrification
312 and mineralization were **not affected** (Figure 4). NO₃-N and total N concentration were
313 negatively associated with pre-disease tanoak basal area and positively associated with
314 cumulative dead tanoak basal area and soil moisture (Figure 4; Table S.5). Extractable inorganic
315 N pools were dominated by NO₃-N, and were often more than 60% nitrate. The shared patterns
316 of significance between NO₃-N and total N is mostly driven by this high proportion of NO₃-N
317 (total N = NO₃-N + NH₄-N). Similarly, nitrification rates were 80 to 100% of net N
318 mineralization **for soils incubated in the field** (Figure 4). Almost identical patterns between
319 nitrification and N mineralization were driven by the dominance of nitrification in N
320 mineralization rates of our study plots. Seasonal influences on soil N concentration and
321 mineralization were weak although the sampling duration also spanned a California-wide
322 drought from 2007-2008.

323

324 *Species effects on litterfall, litterfall chemistry, and soil N cycling: Objective 3*

325 Litter chemistry was markedly different among species. %N was greatest in bay laurel,
326 lowest in redwood, and intermediate in tanoak (Figure 2; $p < 0.05$ each contrast). Litter C/N
327 followed a similar pattern with the highest C/N in redwood, the lowest in bay laurel, and
328 intermediate values for tanoak. Redwood dominated the total litterfall mass in our plots with
329 amounts followed by tanoak, madrone, bay laurel, and other species (Figure 2; Table S.2).
330 Redwood litterfall was low during the spring and summer but peaked in the late autumn/early
331 winter (Figure 2). Tanoak and bay laurel litterfall tended to peak in the mid summer and early
332 autumn, several months earlier than redwood. Despite the significant differences in litterfall

333 chemistry among species, all three followed a similar seasonal pattern of %N (and C/N) with
334 highest levels in the winter and lowest levels during peak litterfall in summer or autumn.

335 Species identity significantly affected NO₃-N availability but did not influence any other
336 soil N cycling parameter during the five week laboratory soil incubation (Figure 5). *Tanoak had*
337 *significantly lower extractable NO₃-N compared to bay laurel and dead tanoak. Soil NO₃-N*
338 *availability from redwood was significantly higher compared to tanoak and tended to be lower*
339 *than bay laurel or dead tanoak but these differences were not significant. Total N levels were*
340 *similar between species and net rates of nitrification and N mineralization also did not differ*
341 *among species (Figure 5). Soil moisture declined over the course of the incubation to an average*
342 *of 0.29 g g⁻¹ (±0.02 se) and net mineralization rates became less variable (Figure S.1). The*
343 *overall patterns of N availability from the laboratory incubation were consistent with*
344 *measurements made in the field. In both measurements tanoak mortality was positively*
345 *associated with NO₃-N availability but no changes in mineralization or nitrification rates were*
346 *found in either set of measurements.*

347

348 **Discussion**

349 This study demonstrates the potential for sudden oak death to alter litterfall and soil N
350 availability in redwood forests and provides general, *a priori* expectations of impacts to these
351 processes for many landscape-scale tree mortality events. Tanoak mortality had the greatest
352 short-term impacts on litterfall dynamics and N availability in our redwood-dominated study
353 sites, *but directional shifts in community composition mediated by *P. ramorum* will have longer-*
354 *term and perhaps greater-magnitude changes to these ecosystem features.* Our study, along with
355 several others, suggests disease-caused ecosystem changes can be driven primarily by mortality

356 and the resulting changes in plant community composition (Hobara et al. 2001; Lovett et al.
357 2006; Orwig et al. 2008; Cobb 2010). These results suggest patterns of landscape-scale tanoak
358 mortality and species shifts (Meentemeyer et al. 2008; Metz et al. 2012) are an appropriate basis
359 for predicting changes in NO₃-N availability and litterfall dynamics for sudden oak death.

360 An emerging consensus of field and modeling studies demonstrate the importance of
361 sporulation sources, especially bay laurel, on rates of *P. ramorum* spread and emergence of
362 sudden oak death (Davidson 2005; 2008; 2011; Maloney et al. 2005; Meentemeyer et al. 2008;
363 2011; Cobb et al. 2012a). Landscape-level data show increased dominance of bay laurel under
364 many conditions, especially when this species co-occurs with tanoak and redwood (Cobb et al.
365 2010; Metz et al. 2012). Shifts to greater dominance of bay laurel will increase litterfall %N as
366 well as soil NO₃-N concentration (Figure 2, 5); this increase in litter %N is likely to increase
367 overall litter decomposition rates as well (Chapman et al. 2006; Cobb 2010). Notably, tanoak
368 mortality can be extensive even when bay laurel is not present within a stand because sporulation
369 on tanoak is sufficient to cause mortality (Ramage et al. 2011; Cobb et al. 2012b; Metz et al.
370 2012). In these stands, sudden oak death is likely to favor species such as redwood or Douglas fir
371 (Cobb et al. 2010) which frequently co-occur with tanoak. These species have notably lower
372 litter quality compared to bay laurel or tanoak which is likely to result in slower litter
373 decomposition and net accumulation of forest floor mass (Figure 2; Valachovic et al. 2004). In
374 either scenario, shifts in species abundance are most likely to drive long-term changes to soil N
375 availability and litterfall dynamics in *P. ramorum* invaded forests.

376 Increased rates of soil N cycling and NO₃-N availability has been a common ecosystem
377 response following insect and pathogen outbreak (Hobara et al. 2001; Orwig et al. 2008;
378 Morehouse et al. 2008; Lovett et al. 2010; Griffin and Turner 2012). Our study departs from this

379 overall trend in that mortality increased $\text{NO}_3\text{-N}$ availability but did not change cycling rates, a
380 result that was consistent in the laboratory as well as the field (Figures 4 and 5). The majority of
381 studies examining pathogen and insect impacts to ecosystems have also focused on outbreaks
382 which result in more uniform mortality or defoliation across a stand compared to sudden oak
383 death (Hobara et al. 2001; Russell et al. 2004; Morehouse et al. 2008; Orwig et al. 2008; Lovett
384 et al. 2010; Griffin and Turner 2012). In contrast, even in our study plots with the greatest
385 amount of tanoak mortality, the majority of biomass was in redwood, bay laurel, or other species
386 which are minimally impacted by the disease. Further, survival times of *P. ramorum* infected
387 tanoak trees can vary from 2-20 years because of differences in susceptibility within populations
388 and size-specific mortality rates (Hayden et al. 2011; Cobb et al. 2012b). The resulting spatial
389 and temporal variation in mortality may dampen impacts to soil N cycling because changes in
390 canopy structure are less severe relative to homogeneous disturbances or outbreaks (Cobb et al.
391 2012a). Comparatively, Gypsy moth (*Lymantria dispar*) outbreak can cause extensive defoliation
392 with low mortality relative to other outbreaks (Lovett et al. 2002; Russell et al. 2004); this
393 defoliation can increase litterfall and litterfall N without changing N mineralization or
394 availability (Russell et al. 2004). Our study supports the general expectation that the timing and
395 uniformity of mortality is an important control over the magnitude of changes to ecosystem
396 processes following outbreak (Ellison et al. 2005; Lovett et al. 2006; Eviner and Likens 2008)
397 even though our data do not confirm our initial hypothesis that disease would increase rates of
398 soil N mineralization.

399 Direct impacts of infection on host tissues had the least significant effect on ecosystem
400 processes at the spatial scale of our study (the ecosystem; Figure 1). Unlike bark-beetle caused
401 mortality, tanoak mortality was not associated with increased litterfall %N (c.f. Morehouse et al.

2008; Griffin and Turner 2012) which may also reflect the heterogeneous timing of tanoak mortality in *P. ramorum* invaded stands (Cobb et al. 2012b). The modest positive association between bay laurel %N and prevalence of infection during the spring and summer seasons (Figure 1) could be driven by changes in plant chemistry induced by infection or by increased shedding of infected foliage (Hunter 2001; Lovett et al. 2006; Eviner and Likens 2008). In bay laurel, *P. ramorum* infection reduces photosynthetic leaf area but does not change photosynthetic rates (DiLeo et al. 2009). Additionally, the prevalence of symptoms within individual bay laurel litter samples was not significantly associated with litterfall %N or C/N suggesting direct pathogen impacts did not drive these changes in litterfall chemistry. In contrast, Davidson et al. (2011) demonstrated increased rates of leaf shedding for infected vs. uninfected bay laurel leaves and suggested *P. ramorum* can accelerate leaf senescence by 3-4 years. Increased litterfall %N is likely when leaf senescence occurs before nutrient reabsorption is maximized in evergreen species including bay laurel (Lovett et al. 2002; Chapman et al. 2006). Although this increase in litter N was small, it could be spatially extensive if other broadly distributed *Phytophthora* pathogens such as *P. nemorosa* and *P. pseudosyringae* also increase bay laurel leaf senescence rate. These other *Phytophthora* species are weak pathogens on tanoak, but have similar ecology to *P. ramorum* on bay laurel and a more extensive geographic range (Wickland et al 2008). All three *Phytophthora* species may influence bay laurel litterfall %N without eliciting disease (c.f. Eviner and Likens 2008).

P. ramorum-tanoak interactions form a relatively tractable host-pathogen system from which it is possible to build local to regional predictive models of outbreak and subsequent tree mortality (Meentemeyer et al. 2011; Cobb et al. 2012b; Filipe et al. 2012). Mortality from sudden oak death is largely driven by sporulation sources in conjunction with the distribution of

425 tanoak and susceptible oaks, the species which may be killed following *P. ramorum* infection
426 (Davidson et al. 2008; Meentemeyer et al. 2008; Lamsal et al. 2011). These patterns emerge
427 because *P. ramorum* virulence is high and resistance in tanoak is insufficient to protect many
428 tanoak populations from significant mortality (Rizzo et al. 2005; Hayden et al. 2011). Patterns of
429 mortality can be reasonably predicted for several other exotic pathogens and insects that are
430 actively spreading into naïve host populations and where community or landscape factors of
431 spread are well understood (Loo 2009; Lovett et al. 2010; Orwig et al. 2012). However,
432 predicting mortality is much more difficult for many regional tree mortality events because the
433 relationships between physiological stress and pathogen impacts are typically unknown for the
434 diverse and widespread native pathogenic flora of most temperate forests (Sinclair et al. 1987;
435 McDowell et al. 2011). Understanding how or when native pathogens and insects overcome
436 plant defenses, and what landscape, climatic, or management factors predispose hosts to greater
437 physiological stress (Raffa et al. 2008; Adams et al. 2009; McDowell et al. 2011) is likely to hold
438 greater potential to predict landscape-level tree mortality of these outbreaks.

439 For sudden oak death, many of the ecosystem changes we observed are tied to the
440 epidemiological roles of canopy tree species and their individual influences on ecosystem
441 processes. The mechanisms driving these affects included changes in host litter chemistry,
442 mortality, and shifts in community composition that are likely to be common among many
443 pathogen outbreaks in the same way that they are common drivers of ecosystem change
444 following insect outbreak. Although interactions between pathogens, hosts, and the environment
445 is a foundation of plant pathology (e.g. the disease triangle; Burdon et al. 2006), these
446 interactions are poorly understood for abundant, diverse, but broadly distributed weak pathogens
447 (Balci et al. 2007; Wickson et al. 2008; Hawkins and Henkel 2011). This lack of understanding

448 hinders prediction of tree mortality incited by regional drought but contributed to by pathogens
449 (Worrall et al. 2010; McDowell et al. 2011). However, when pathogens or insects incite or
450 substantially contribute to major tree die-offs, the longest lasting and greatest magnitude
451 ecosystem impacts can be reasonably predicted by understanding patterns of mortality and
452 subsequent changes in species composition.

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462 **Literature cited**

463 **Adams HD, Guardiola-Claramonte M, Barron-Gafford GA, Villegas JC, Breshears DD,**
464 **Zou CB, Troch PA, Huxman TE. 2009.** Temperature sensitivity of drought-induced tree
465 mortality portends increased regional die-off under global change-type drought. *Proceedings*
466 *of the National Academy of Sciences* **106**: 7063–7066

467 **Balci Y, Balci S, Eggers J, MacDonald WL, Gottschalk KW, Juzwik J, Long R. 2007.**
468 *Phytophthora* spp. associated with forest soils in eastern and north-central U.S. oak
469 ecosystems. *Plant Disease* **91**: 705–710.

- 470 **Brown LB, Allen-Diaz B. 2009.** Forest stand dynamics and sudden oak death: Mortality in
471 mixed-evergreen forests dominated by coast live oak. *Forest Ecology and Management* **257**:
472 1271–1280.
- 473 **Burdon JJ, Thrall PH, Ericson L. 2006.** The current and future dynamics of disease in plant
474 communities. *Annual Review of Phytopathology* **44**: 19–39.
- 475 **Chapman SK, Whitham TG, Powell M. 2006.** Herbivory differentially alters plant litter
476 dynamics of evergreen and deciduous trees. *Oikos* **114**: 566–574.
- 477 **Classen AT, Hart SC, Whitman TG, Cobb NS, Koch GW. 2005.** Insect Infestations Linked to
478 Shifts in Microclimate. *Soil Science Society of America Journal* **69**: 2049–2057.
- 479 **Cobb RC. 2010.** Species shift drives decomposition rates following invasion by hemlock woolly
480 adelgid. *Oikos* **119**: 1291–1298.
- 481 **Cobb RC, Chan MN, Meentemeyer RK, Rizzo DM. 2012a.** Common factors drive disease and
482 coarse woody debris dynamics in forests impacted by sudden oak death. *Ecosystems* **15**: 242-
483 255.
- 484 **Cobb RC, Filipe JAN, Meentemeyer RK, Gilligan CA, Rizzo DM. 2012b.** Ecosystem
485 transformation by emerging infectious disease: loss of large tanoak from California forests.
486 *Journal of Ecology* **100**: 712-722.
- 487 **Cobb RC, Meentemeyer RK, Rizzo DM. 2010.** Apparent competition in canopy trees
488 determined by pathogen transmission rather than susceptibility. *Ecology* **91**: 327–333.
- 489 **Davis FW, Borchert M, Meentemeyer RK, Flint A, Rizzo DM. 2010.** Pre-impact forest
490 composition and ongoing tree mortality associated with sudden oak death in the Big Sur
491 region; California. *Forest Ecology and Management* **259**: 2342–2354.

- 492 **Davidson JM, Patterson HA, Rizzo DM. 2008.** Sources of inoculum for *Phytophthora*
493 *ramorum* in a redwood forest. *Phytopathology* **98**: 860–866.
- 494 **Davidson JM, Patterson HA, Wickland AC, Fichtner EJ, Rizzo DM. 2011.** Forest type
495 influences transmission of *Phytophthora ramorum* in California oak woodlands.
496 *Phytopathology* **101**: 492–501.
- 497 **Davidson JM, Wickland AC, Patterson HA, Falk KR, Rizzo DM. 2005.** Transmission of
498 *Phytophthora ramorum* in mixed-evergreen forest in California. *Phytopathology* **95**: 587–596.
- 499 **DiLeo MV, Bostock RM, Rizzo DM. 2009.** *Phytophthora ramorum* does not cause
500 physiologically significant systemic injury to California bay laurel, its primary reservoir host.
501 *Phytopathology* **99**: 1307–1311.
- 502 **Ellison AM, Bank MS, Clinton BD, Colburn EA, Elliott K, Ford CR, Foster DR, Kloeppel**
503 **BD, Knoepp JD, Lovett GM. 2005.** Loss of foundation species: consequences for the
504 structure and dynamics of forested ecosystems. *Frontiers in Ecology and the Environment* **3**:
505 479–486.
- 506 **Eviner VT, Chapin III FS. 2003.** Functional matrix: a conceptual framework for predicting
507 multiple plant effects on ecosystem processes. *Annual Review of Ecology, Evolution, and*
508 *Systematics*: 455–485.
- 509 **Eviner VT, Likens GE. 2008.** Effects of pathogens on terrestrial ecosystem function. In: Ostfeld
510 RS, Keesing F, Eviner VT, eds. *Infectious disease ecology. Effects of ecosystems on disease*
511 *and disease on ecosystems*. Princeton, NJ: Princeton University Press, 260–283.
- 512 **Filipe JAN, Cobb RC, Meentemeyer RK, Lee CA, Valachovic YS, Cook AR, Rizzo DM,**
513 **Gilligan CA. 2012.** Landscape epidemiology and control of pathogens with cryptic and long-

- 514 distance dispersal: sudden oak death in northern Californian forests. *PLoS Computational*
515 *Biology* **8**: e1002328.
- 516 **Finzi AC, Van Breemen N, Canham CD. 1998.** Canopy tree-soil interactions within temperate
517 forests: species effects on soil carbon and nitrogen. *Ecological applications* **8**: 440–446.
- 518 **Fried JS, Boyle JR, Tappeiner II JC, Cromack Jr K. 1990.** Effects of bigleaf maple on soils
519 in Douglas-fir forests. *Canadian Journal of Forest Research* **20**: 259–266.
- 520 **Gilligan CA, Van den Bosch F. 2008.** Epidemiological models for invasion and persistence of
521 pathogens. *Annual Review of Phytopathology* **46**: 385–418.
- 522 **Griffin JM, Turner MG. 2012.** Changes to the N cycle following bark beetle outbreaks in two
523 contrasting conifer forest types. *Oecologia* **170**: 551-565.
- 524 **Hawkins AE, Henkel TW. 2011.** Native forest pathogens facilitate persistence of Douglas-fir in
525 old-growth forests of northwestern California. *Canadian Journal of Forest Research* **41**:
526 1256–1266.
- 527 **Hayden KJ, Nettel A, Dodd RS, Garbelotto M. 2011.** Will all the trees fall? Variable
528 resistance to an introduced forest disease in a highly susceptible host. *Forest Ecology and*
529 *Management* **261**: 1781–1791.
- 530 **Hicke JA, Allen CD, Desai AR, Dietze MC, Hall RJ, Hogg EH, Kashian DM, Moore D,**
531 **Raffa KF, Sturrock RN. 2012.** Effects of biotic disturbances on forest carbon cycling in the
532 United States and Canada. *Global Change Biology* **18**: 7-34.
- 533 **Hobara S, Tokuchi N, Ohte N, Koba K, Katsuyama M, Kim SJ, Nakanishi A. 2001.**
534 Mechanism of nitrate loss from a forested catchment following a small-scale, natural
535 disturbance. *Canadian Journal of Forest Research* **31**: 1326–1335.

- 536 **Holt RD, Dobson AP, Begon M, Bowers RG, Schauber EM. 2003.** Parasite establishment in
537 host communities. *Ecology Letters* **6**: 837–842.
- 538 **Hunter MD. 2001.** Insect population dynamics meets ecosystem ecology: effects of herbivory
539 on soil nutrient dynamics. *Agricultural and Forest Entomology* **3**: 77–84.
- 540 **Lamsal S, Cobb RC, Hall Cushman J, Meng Q, Rizzo DM, Meentemeyer RK. 2011.** Spatial
541 estimation of the density and carbon content of host populations for *Phytophthora ramorum* in
542 California and Oregon. *Forest Ecology and Management* **262**: 989–998.
- 543 **Loo JA. 2009.** Ecological impacts of non-indigenous invasive fungi as forest pathogens.
544 *Biological Invasions* **11**: 81–96.
- 545 **Lovett GM, Arthur MA, Weathers KC, Griffin JM. 2010.** Long-term changes in forest
546 carbon and nitrogen cycling caused by an introduced pest/pathogen complex. *Ecosystems* **13**:
547 1188–1200.
- 548 **Lovett GM, Canham CD, Arthur MA, Weathers KC, Fitzhugh RD. 2006.** Forest ecosystem
549 responses to exotic pests and pathogens in eastern North America. *BioScience* **56**: 395–405.
- 550 **Lovett GM, Christenson LM, Groffman PM, Jones CG, Hart JE, Mitchell MJ. 2002.** Insect
551 defoliation and nitrogen cycling in forests. *BioScience* **52**: 335–341.
- 552 **Maloney PE, Lynch SC, Kane SF, Jensen CE, Rizzo DM. 2005.** Establishment of an emerging
553 generalist pathogen in redwood forest communities. *Journal of Ecology* **93**: 899–905.
- 554 **Mascheretti S, Croucher PJP, Vettraino A, Prospero S, Garbelotto M. 2008.** Reconstruction
555 of the sudden oak death epidemic in California through microsatellite analysis of the pathogen
556 *Phytophthora ramorum*. *Molecular Ecology* **17**: 2755–2768.

- 557 **McDowell NG, Beerling DJ, Breshears DD, Fisher RA, Raffa KF, Stitt M. 2011.** The
558 interdependence of mechanisms underlying climate-driven vegetation mortality. *Trends in*
559 *ecology & evolution* **26**: 523–532.
- 560 **Meentemeyer RK, Cunniffe NJ, Cook AR, Filipe JAN, Hunter RD, Rizzo DM, Gilligan CA.**
561 **2011.** Epidemiological modeling of invasion in heterogeneous landscapes: spread of sudden
562 oak death in California (1990-2030). *Ecosphere* **2**: art17.
- 563 **Meentemeyer RK, Rank NE, Shoemaker DA, Oneal CB, Wickland AC, Frangioso KM,**
564 **Rizzo DM. 2008.** Impact of sudden oak death on tree mortality in the Big Sur ecoregion of
565 California. *Biological Invasions* **10**: 1243–1255.
- 566 **Metz MR, Frangioso KM, Wickland AC, Meentemeyer RK, Rizzo DM. 2012.** An emergent
567 disease causes directional changes in forest species composition in coastal California.
568 *Ecosphere* **3**: art86.
- 569 **Morehouse K, Johns T, Kaye J, Kaye M. 2008.** Carbon and nitrogen cycling immediately
570 following bark beetle outbreaks in southwestern ponderosa pine forests. *Forest Ecology and*
571 *Management* **255**: 2698–2708.
- 572 **Orwig DA, Thompson JR, Povak NA, Manner M, Niebyl D, Foster DR. 2012.** A foundation
573 tree at the precipice: *Tsuga canadensis* health after the arrival of *Adelges tsugae* in central
574 New England. *Ecosphere* **3**: art10.
- 575 **Orwig DA, Cobb RCC, D’Amato AW, Kizlinski ML, Foster DR. 2008.** Multi-year ecosystem
576 response to hemlock woolly adelgid infestation in southern New England forests. *Canadian*
577 *Journal of Forest Research* **38**: 834–843.

- 578 **Raffa KF, Aukema BH, Bentz BJ, Carroll AL, Hicke JA, Turner MG, Romme WH. 2008.**
579 Cross-scale drivers of natural disturbances prone to anthropogenic amplification: the
580 dynamics of bark beetle eruptions. *BioScience* **58**: 501–517.
- 581 **Ramage BS, O’Hara KL, Forrestel AB. 2011.** Forest transformation resulting from an exotic
582 pathogen: regeneration and tanoak mortality in coast redwood stands affected by sudden oak
583 death. *Canadian Journal of Forest Research* **41**: 763–772.
- 584 **Rizzo DM, Garbelotto M, Hansen EM. 2005.** Phytophthora ramorum: integrative research and
585 management of an emerging pathogen in California and Oregon forests. *Annual Review of*
586 *Phytopathology*. **43**: 309–335.
- 587 **Ruess RW, McFarland JM, Trummer LM, Rohrs-Richey JK. 2009.** Disease-mediated
588 declines in N-fixation inputs by *Alnus tenuifolia* to early-successional floodplains in interior
589 and south-central Alaska. *Ecosystems* **12**: 489–502.
- 590 **Russell CA, Kosola KR, Paul EA, Robertson GP. 2004.** Nitrogen Cycling in Poplar Stands
591 Defoliated by Insects. *Biogeochemistry* **68**: 365–381.
- 592 **Santini A, Ghelardini L, Pace C, Desprez-Loustau ML, Capretti P, Chandelier A, Cech T,**
593 **Chira D, Diamandis S, Gaitniekis T. 2012.** Biogeographical patterns and determinants of
594 invasion by forest pathogens in Europe. *New Phytologist* **197**: 238-250.
- 595 **Sinclair WA, Lyon HH, Johnson WT. 1987.** *Diseases of trees and shrubs*. Ithaca, NY: Cornell
596 University Press.
- 597 **Valachovic YS, Caldwell BA, Cromack Jr K, Griffiths RP. 2004.** Leaf litter chemistry
598 controls on decomposition of Pacific Northwest trees and woody shrubs. *Canadian Journal of*
599 *Forest Research* **34**: 2131–2147.

600 **Wickland AC, Jensen CE, Rizzo DM. 2008.** Geographic distribution, disease symptoms and
601 pathogenicity of *Phytophthora nemorosa* and *Phytophthora pseudosyringae* in California,
602 USA. *Forest Pathology* **38**: 288–298.

603 **Worrall JJ, Marchetti SB, Egeland L, Mask RA, Eager T, Howell B. 2010.** Effects and
604 etiology of sudden aspen decline in southwestern Colorado, USA. *Forest ecology and*
605 *management* **260**: 638–648.

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607 **Figure captions**

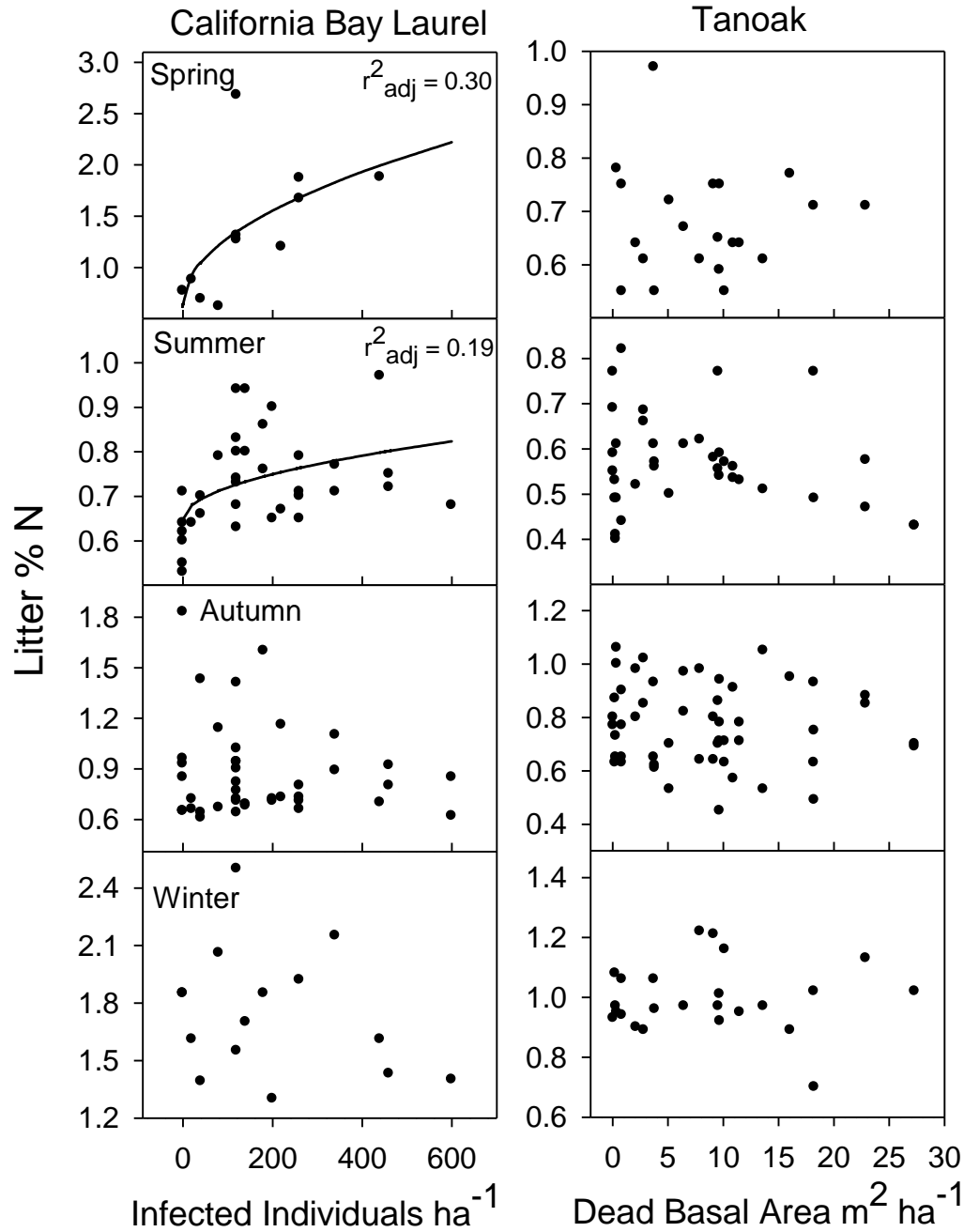
608 Figure 1. Litterfall %N vs. prevalence of *Phytophthora ramorum* in bay laurel or cumulative
609 dead tanoak basal area. When the relationship between infected hosts or mortality and litter N
610 concentration was significant ($p \leq 0.05$), the r^2 is reported along with the square root transformed
611 least squares fit. Note the differences in scale for bay laurel and tanoak (x-axis) as well as
612 differences in N concentration between seasons of measurement (y-axis).

613 Figure 2. Monthly litterfall mass (A-C) with seasonal values of carbon and nitrogen (D-F) for
614 bay laurel (top), tanoak (middle), and redwood (bottom) from two redwood forests impacted by
615 sudden oak death. Data are means from three years of litterfall monitoring presented with one
616 standard deviation for litterfall mass and standard error for seasonal litter chemistry. Note the
617 differences in scale in the panels of litterfall mass for each species. The month of collection is
618 abbreviated with the first letter.

619 Figure 3. Affects of sudden oak death-caused mortality (x-axis) on tanoak annual tanoak litterfall
620 mass (A), tanoak litterfall N (B), and total stand-level litterfall N (C). Data are observed values
621 minus those expected if the stands had not been impacted by sudden oak death (see text and table
622 S.3 for more details). Negative values on the y-axis denote the amount of litterfall reduction
623 associated with a given amount of tanoak mortality. Data are total annual amounts for each plot
624 with least squares regression lines.

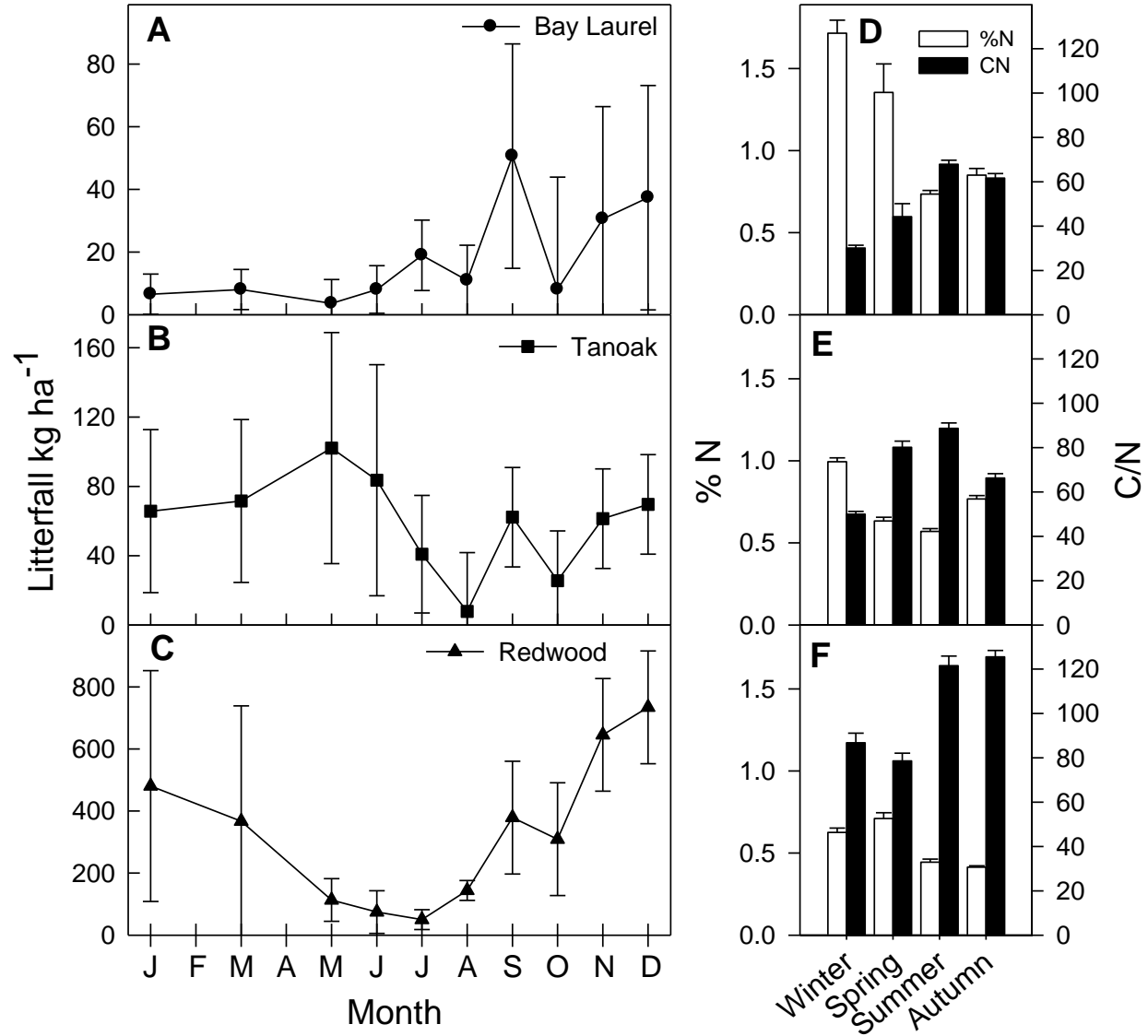
625 Figure 4. Seasonal patterns and affects of sudden oak death on soil N. Seasonal patterns of
626 inorganic N pool sizes (A) and rates of N mineralization and nitrification (B) are shown with
627 sampling date on the x-axis. Right panels; leverage plots from mixed linear models showing the
628 effect of tanoak mortality on N pool sizes (C) and rates of turnover (D). Least squares regression
629 lines are shown for statistically significant ($p \leq 0.05$) models.

630 Figure 5. Species level effects on N availability (extractable pools A and B) and cycling rates (C
631 and D). Data are results from incubation of soils collected immediately below the three focal
632 species and tanoak killed by *P. ramorum* (Dead Tanoak). Data are means with one standard
633 error. Results from a paired t-test analysis are presented above each bar with the significance
634 tests indicating differences across all possible pairs; different letters indicate statistically
635 different mean values ($p \leq 0.05$).



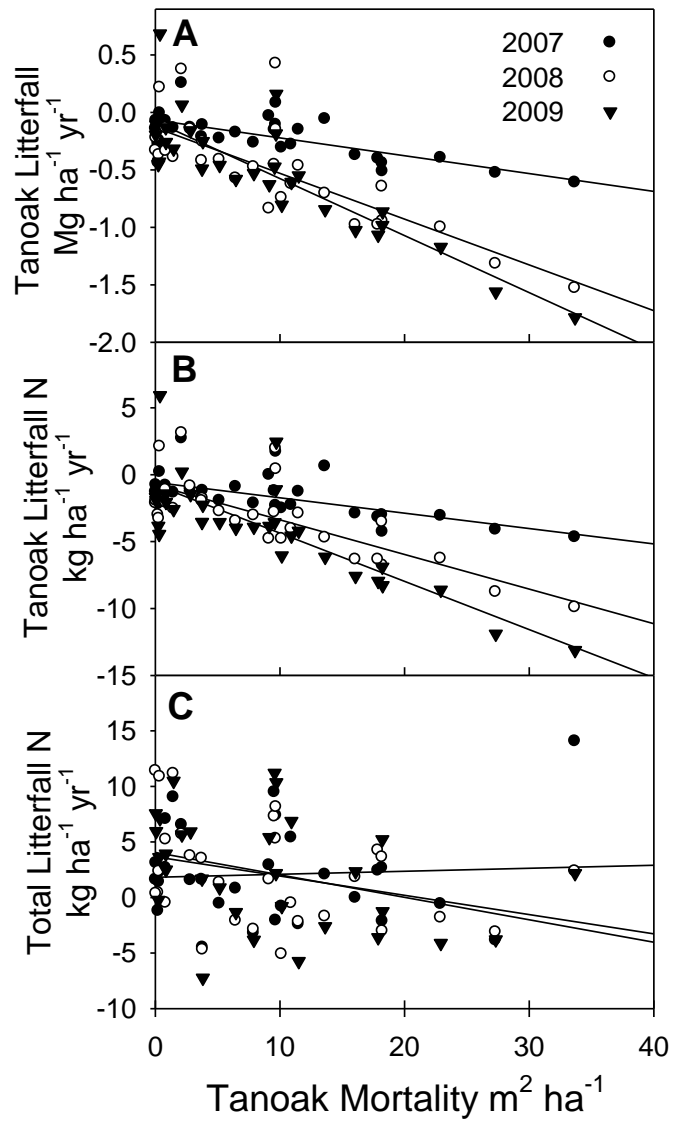
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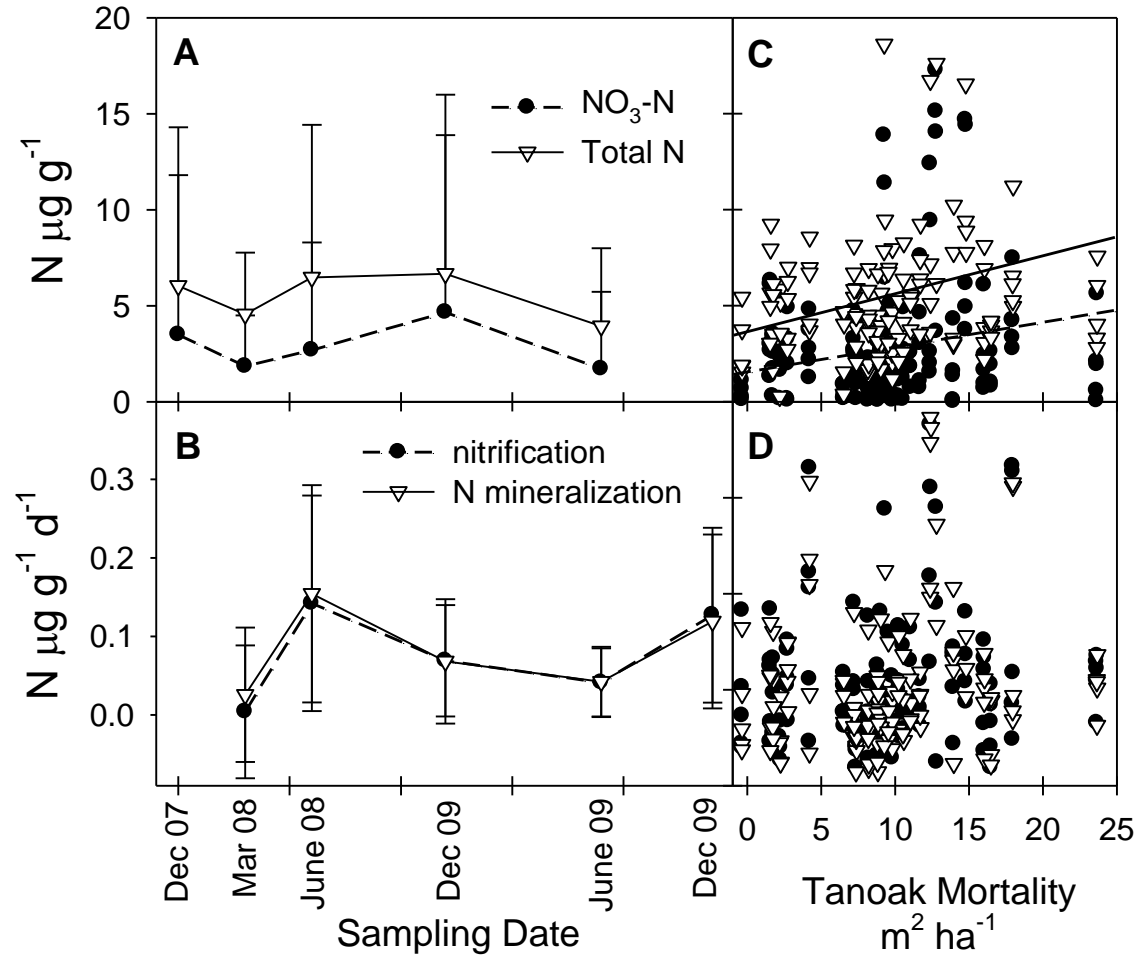
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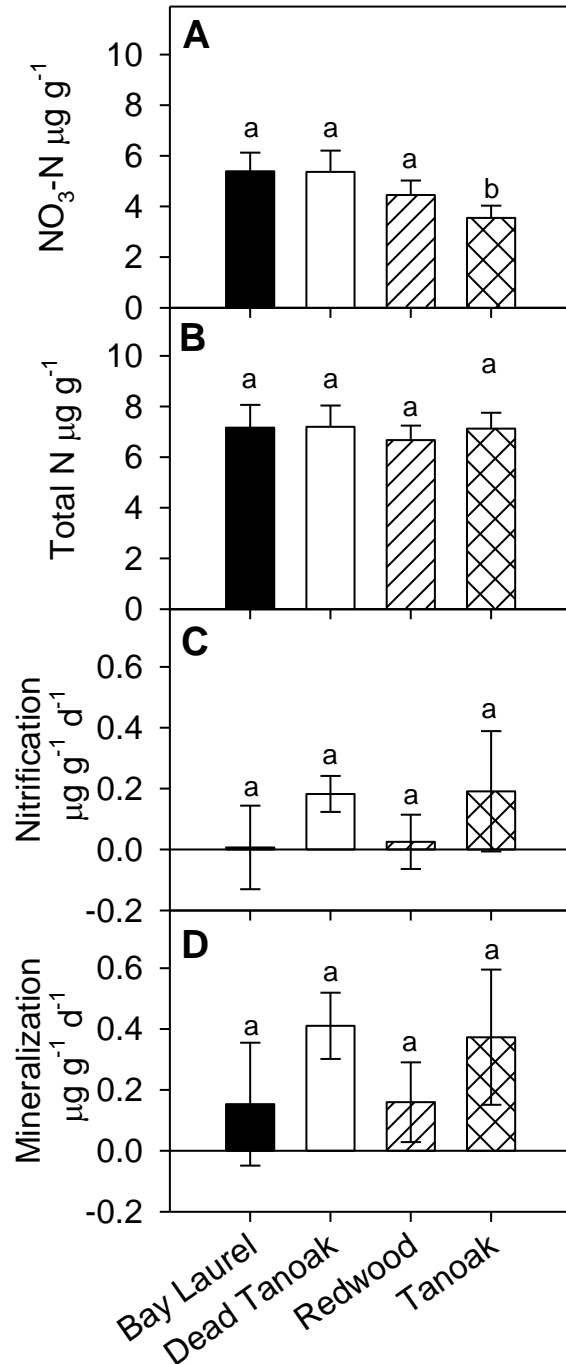
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