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# Toxicity Inhibitors Protect Lipid Membranes from Disruption by A $\beta$ 42

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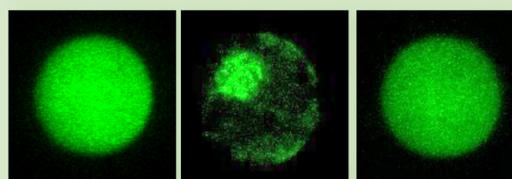
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## Supporting Information

**ABSTRACT:** Although the precise molecular factors linking amyloid  $\beta$ -protein (A $\beta$ ) to Alzheimer's disease (AD) have not been deciphered, interaction of A $\beta$  with cellular membranes has an important role in the disease. However, most therapeutic strategies targeting A $\beta$  have focused on interfering with A $\beta$  self-assembly rather than with its membrane interactions. Here, we studied the impact of three toxicity inhibitors on membrane interactions of A $\beta$ 42, the longer form of A $\beta$ , which is associated most strongly with AD. The inhibitors included the four-residue C-terminal fragment A $\beta$ (39–42), the polyphenol (–)-epigallocatechin-3-gallate (EGCG), and the lysine-specific molecular tweezer, CLR01, all of which previously were shown to disrupt different steps in A $\beta$ 42 self-assembly. Biophysical experiments revealed that incubation of A $\beta$ 42 with each of the three modulators affected membrane interactions in a distinct manner. Interestingly, EGCG and CLR01 were found to have significant interaction with membranes themselves. However, membrane bilayer disruption was reduced when the compounds were preincubated with A $\beta$ 42, suggesting that binding of the assembly modulators to the peptide attenuated their membrane interactions. Importantly, our study reveals that even though the three tested compounds affect A $\beta$ 42 assembly differently, membrane interactions were significantly inhibited upon incubation of each compound with A $\beta$ 42, suggesting that preventing the interaction of A $\beta$ 42 with the membrane contributes substantially to inhibition of its toxicity by each compound. The data suggest that interference with membrane interactions is an important factor for A $\beta$ 42 toxicity inhibitors and should be taken into account in potential therapeutic strategies, in addition to disruption or remodeling of amyloid assembly.

**KEYWORDS:** Molecular tweezer, membrane interactions, amyloid  $\beta$ -protein (A $\beta$ ), polyphenols, fibril inhibitors



Effects of A $\beta$ 42 inhibitors on membrane interactions

The transformation of soluble proteins into toxic oligomers and amyloid fibrils is a key pathologic process in devastating medical disorders, including Alzheimer's disease (AD), Parkinson's disease, and type-II diabetes.<sup>1,2</sup> Though the presence of fibrillar aggregates appears to be a universal phenomenon in amyloid diseases, the relationship between amyloid formation, disease progression, and pathogenicity remains unclear. Amyloid plaques, in which the main component is the amyloid  $\beta$ -protein (A $\beta$ ), particularly its longer form, A $\beta$ 42, are a pathologic hallmark of AD. A $\beta$  oligomers, which form as intermediates in the plaque-formation process and are considered the proximal neurotoxins in AD, have been reported to cause membrane leakage, either through nonspecific pore formation or via other mechanisms.<sup>3–5</sup> A two-step mechanism for A $\beta$ 42–membrane interactions has been recently reported.<sup>6</sup> Accordingly, understanding the mechanistic aspects of A $\beta$ 42 interactions with cellular membranes has been the focus of intensive research.<sup>7–10</sup>

Diseases associated with protein misfolding and aggregation, including AD, are currently incurable. Therefore, extensive research effort has been directed at developing inhibitors and modulators of protein aggregation and exploring their therapeutic potential. Notably, however, despite strong evidence for involvement of lipid and membrane interactions of misfolded proteins in the cytotoxicity of amyloidogenic proteins,<sup>11,12</sup> development of therapeutic drugs targeting protein misfolding and aggregation largely has neglected this aspect of the pathologic mechanism and focused almost exclusively on substances that interfere with the self-assembly processes of amyloid proteins, such as A $\beta$ 42.

Numerous molecules have been evaluated for their effect on A $\beta$  self-assembly and toxicity.<sup>13–16</sup> Peptide fragments compris-

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ing discrete short sequences within  $A\beta_{42}$  have been shown to reduce cytotoxicity in cell culture and in vivo, presumably through binding to  $A\beta_{42}$  at early aggregation stages, thereby inhibiting formation of the toxic oligomers.<sup>17,18</sup> Polyphenols, such as resveratrol (found in red grape skin and seeds)<sup>19,20</sup> and (–)-epigallocatechin-3-gallate (EGCG, a component of green tea),<sup>21,22</sup> have been among the most widely studied inhibitors of cytotoxicity and fibril formation of amyloid proteins. These molecules have attracted much attention because they are nutraceuticals; they exist in different food sources and therefore are considered bioavailable and safe. Another attractive feature is that many of them also have antioxidant and anti-inflammatory activities.<sup>23,24</sup> EGCG, in particular, is a broad-spectrum inhibitor, previously shown to interfere in the  $A\beta_{42}$  fibrillation process at different stages, blocking toxic oligomer assembly,<sup>22,25,26</sup> and inducing fibril disassembly.<sup>27,28</sup> Amyloid inhibitory effects of small compounds derived from natural products have also been recently reported.<sup>29</sup>

Using a different approach, recent studies have identified a synthetic lysine-binding “molecular tweezer” called CLR01 as a powerful modulator of the self-assembly and a general inhibitor of the toxic effect of amyloid proteins, including  $A\beta_{42}$ .<sup>30,31</sup> The mechanism of action of CLR01 is reversible binding to exposed lysine residues, which disrupts electrostatic and hydrophobic interactions involving lysines and leads to remodeling of the assembly process into formation of nontoxic and non-amyloidogenic structures that can be effectively degraded by the natural clearance mechanisms. Because the binding is highly labile and occurs with micromolar affinity and because in misfolded proteins lysines tend to be more exposed to the solvent than in typical globular proteins,<sup>32</sup> at the concentrations needed for inhibition, CLR01 does not interfere with normal protein structure or function.<sup>31,33</sup>

Here, we specifically address the participation of membranes in amyloid toxicity inhibition through investigating the effect of three modulators,  $A\beta(39-42)$ , EGCG, and CLR01, on membrane interactions of  $A\beta_{42}$ . These three inhibitors were chosen because they represent a variety of features some of which overlap and some that are unique to each compound (Table 1). Different than most assembly modulation studies

**Table 1. Details of the Assembly Modulators Studied**

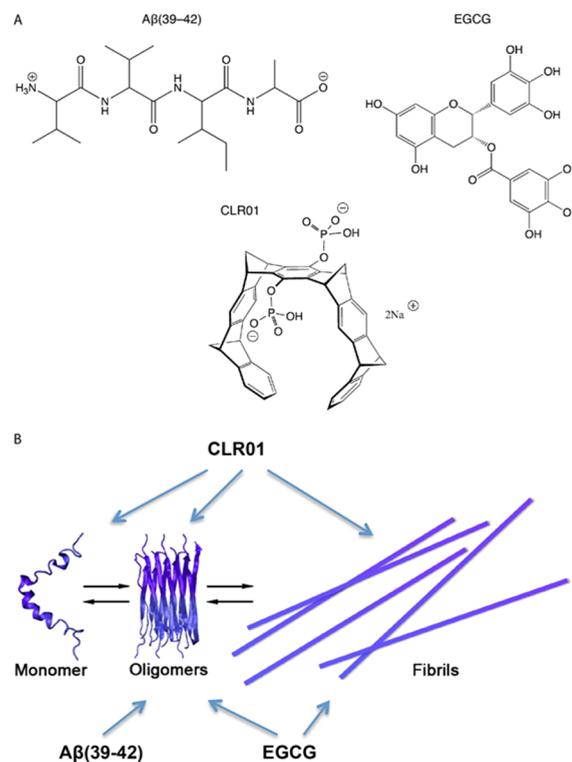
compound	EGCG	$A\beta(39-42)$	CLR01
type	small molecule	peptide	small molecule
specificity <sup>a</sup>	nonspecific	$A\beta_{42}$	nonspecific
binds monomers	no	no	yes
binds oligomers	yes	yes	yes
specific binding site	no	mainly N-terminal region	Lys16 > Lys28 > Arg5

<sup>a</sup>Specificity refers to whether the compound binds one particular target or multiple targets, as well as targeting particular assembly states, monomers, oligomers, or fibrils.<sup>22,30,34</sup>

reported to date, the focus of our study was on the ternary interactions among  $A\beta_{42}$ , the assembly modulators, and membrane lipid bilayers. As such, our work illuminates a rarely studied angle of assembly modulators' activity, their effect on membrane interactions of  $A\beta$ . The data reveal that the interactions between  $A\beta_{42}$  and each assembly modulator effectively “shield” the membrane, not only blocking membrane interactions of  $A\beta_{42}$  but also, surprisingly, inhibiting bilayer disruption by the assembly modulators themselves.

## RESULTS AND DISCUSSION

Figure 1 shows the assembly modulators studied here and their putative effects on the  $A\beta_{42}$  assembly process. We selected



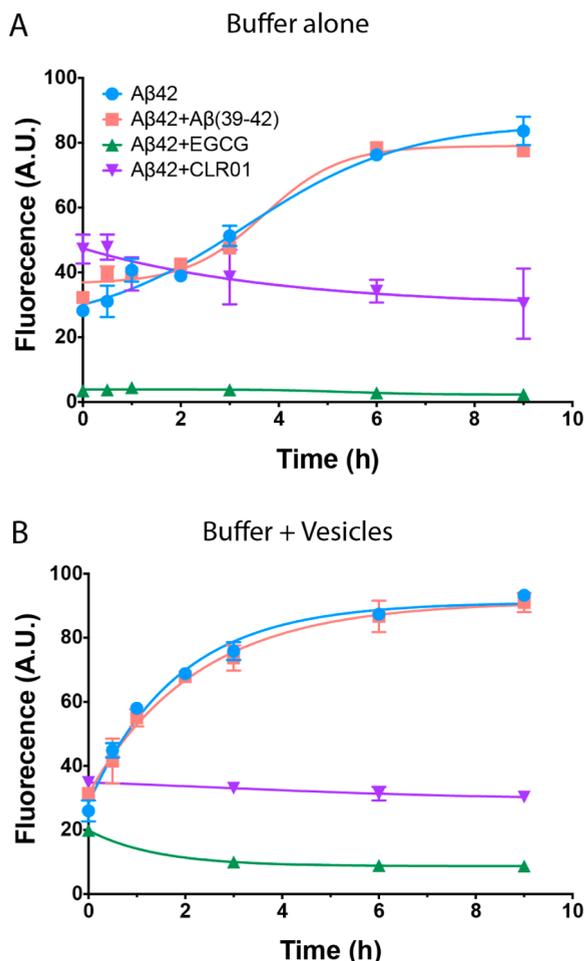
**Figure 1.** Assembly modulators studied and their putative activity. (A) The molecules investigated in this study:  $A\beta(39-42)$ , EGCG, and CLR01. (B) Stages of  $A\beta_{42}$  fibrillation putatively modulated by compounds 1–3.

these modulators because they belong to distinct molecular classes and they remodel  $A\beta_{42}$  using different mechanisms. Specifically,  $A\beta(39-42)$  (Figure 1A) comprises the four-residue C-terminal sequence of  $A\beta_{42}$  (Val-Val-Ile-Ala), which was shown to reduce  $A\beta_{42}$  neurotoxicity by remodeling its oligomerization but did not affect  $A\beta_{42}$  fibrillation.<sup>18,34–36</sup> EGCG is a polyphenol extracted from green tea known to bind  $A\beta$  oligomers, modify them covalently,<sup>37</sup> and inhibit their toxicity.<sup>22,38</sup> CLR01 is a molecular tweezer that binds  $A\beta$  monomers and oligomers noncovalently, thereby remodeling  $A\beta_{42}$  assembly and inhibiting its toxicity.<sup>30,39,40</sup> CLR01 also can disaggregate preformed  $A\beta$  fibrils, albeit with much slower kinetics than EGCG. Both EGCG and CLR01 are broad-spectrum assembly modulators that have been shown to be effective inhibitors of the toxicity of multiple amyloidogenic proteins,<sup>38,41</sup> whereas  $A\beta(39-42)$  inhibits  $A\beta_{42}$  selectively.<sup>34,35</sup>

Figure 1B summarizes the putative stages of the  $A\beta_{42}$  assembly process affected by each modulator.<sup>34,35,39,42</sup> It should be emphasized, however, that although the inhibition mechanisms depicted in Figure 1B are based on experimental analyses, the effects of the modulators in the presence of membranes and the interplay between assembly modulation and membrane interactions have not been addressed previously.

We compared first the fibrillation kinetics of  $A\beta_{42}$  modulator mixtures in buffer alone and in the presence of DMPC/DMPG vesicles. This vesicle composition has been employed widely as

a mimetic of cellular membranes.<sup>43,44</sup> The kinetics of  $\beta$ -sheet formation was monitored using the thioflavin-T (ThT) fluorescence assay (Figure 2). ThT is a commonly used dye



**Figure 2.** Fibrillation kinetics of  $A\beta_{42}$ . ThT fluorescence was recorded in the absence (A) or presence (B) of DMPC/DMPG vesicles. The data are presented as mean  $\pm$  SEM of two to four reactions with 10 readings of each sample at every time point.

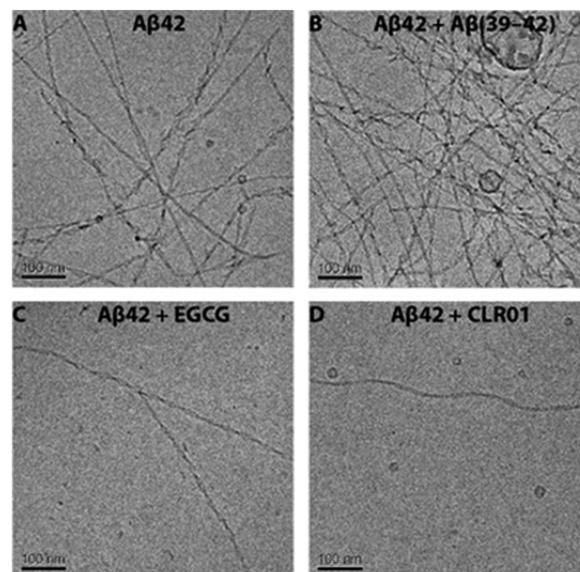
exhibiting enhanced fluorescence upon binding to  $\beta$ -sheets in amyloid fibrils.<sup>45,46</sup> Importantly, ThT is not suitable for measurement of  $\beta$ -sheet formation and aggregation in the presence of some compounds, for example, curcumin, quercetin, and resveratrol, which have significant absorbance in the 440–482 nm range used for ThT excitation and emission.<sup>47</sup> However, none of the compounds that we used absorb in this range,<sup>48,49</sup> and therefore the ThT assay is suitable for measuring the formation of  $\beta$ -sheet-rich fibrils in the presence of these assembly modulators.

Figure 2 shows different fibrillation kinetics of  $A\beta_{42}$  in the absence or presence of DMPC/DMPG vesicles. In the absence of vesicles, a lag phase of  $\sim 2$  h was observed followed by an increase in fluorescence up to 6 h, at which point the fluorescence signal plateaued (Figure 2A). In contrast, in the vesicle-containing solution, the ThT fluorescence increased rapidly already within the first hour of incubation with no apparent lag phase (Figure 2B), suggesting that the presence of a membrane-like environment accelerated  $A\beta_{42}$  fibril nucleation, as reported previously.<sup>50</sup>

The assembly modulators had distinct effects on the fibrillation kinetics (Figure 2). The ThT fluorescence curves corresponding to the  $A\beta_{42}/A\beta(39-42)$  mixture, both in buffer and in vesicle solutions, were almost identical to those of  $A\beta_{42}$  alone, in agreement with a previous report that  $A\beta(39-42)$  did not significantly affect the extent or kinetics of  $A\beta_{42}$  fibril formation.<sup>34</sup> In contrast, no increase in ThT fluorescence was observed in the presence of either EGCG or CLR01. In fact, in the presence of EGCG in buffer alone the initial fluorescence was substantially lower than in any of the other cases (Figure 2A). These data are in agreement with a previous report indicating that in addition to preventing  $A\beta$  fibril formation, EGCG competes with ThT binding to  $A\beta$ ,<sup>37</sup> presumably leading to the apparent low fluorescence. In the presence of the lipid vesicles, the initial ThT fluorescence of the  $A\beta_{42}/EGCG$  mixture was similar to that of the other reactions and then decreased gradually over time (Figure 2B), suggesting that initially the vesicles might shield, at least partially, the binding sites on  $A\beta$  from EGCG but not from ThT.

A lack of increase in ThT fluorescence intensity was observed also in the presence of CLR01 (Figure 2), which is not known to affect ThT binding to  $A\beta$ . Therefore, these data likely point simply to inhibition of  $\beta$ -sheet formation of  $A\beta_{42}$  by CLR01 regardless of the presence of the vesicles. Overall, the ThT fluorescence data suggest that the three modulators behave similarly in the presence of lipid vesicles to the way they behave in simple buffer solutions; namely,  $A\beta(39-42)$  does not affect fibril formation,<sup>34</sup> CLR01 prevents  $A\beta_{42}$  fibrillogenesis, whereas conclusions regarding the effect of EGCG on fibrillogenesis cannot be made due to its competition with ThT.<sup>39</sup>

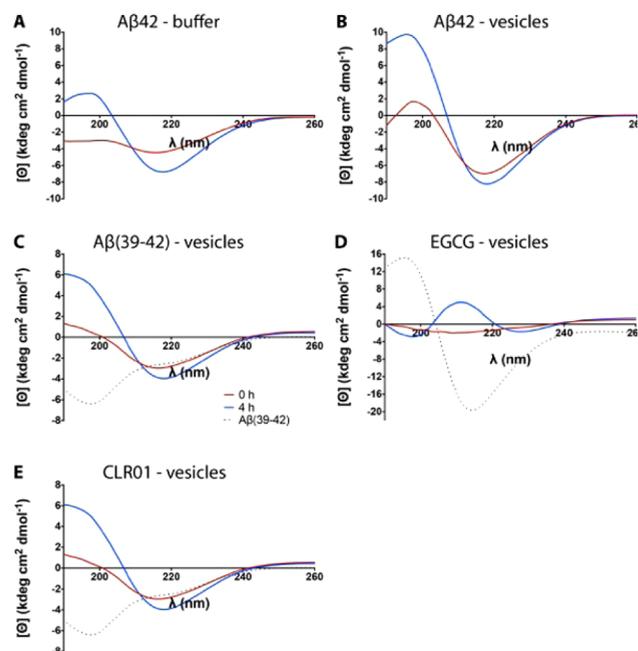
Cryo-TEM analysis (Figure 3) corroborated the ThT data and provided a visual depiction of the effect of the modulators on  $A\beta_{42}$  in the presence of lipid vesicles. TEM examination of  $A\beta_{42}$  in the presence of each modulator in simple buffers has been reported previously<sup>30,34,51</sup> and therefore is not shown



**Figure 3.**  $A\beta_{42}$  fibril morphology in the presence of membrane mimetics. Cryo-TEM images of  $A\beta_{42}$  samples after 12 h incubation with DMPC/DMPG vesicles. (A)  $A\beta_{42}$ ; (B)  $A\beta_{42}$  co-incubated with  $A\beta(39-42)$ ; (C)  $A\beta_{42}$  co-incubated with EGCG; (D)  $A\beta_{42}$  co-incubated with CLR01.

here.  $A\beta$ 42 alone or in the presence of  $A\beta$ (39–42) (as well as in the presence of DMPC/DMPG vesicles) formed abundant fibrils (Figure 3A,B), whereas when co-incubated with EGCG or CLR01 only scarce fibrils were observed (Figure 3C,D). Together, the ThT (Figure 2) and microscopy (Figure 3) experiments indicate that the effects of the three assembly modulators on  $A\beta$ 42 fibrillation in the presence of lipid vesicles are similar to their effects in the absence of membranes.

To gain further insight into the impact of the ternary interactions among  $A\beta$ 42, lipid vesicles, and assembly modulators on the changes in the secondary structure of  $A\beta$ 42 during its self-assembly, we used CD spectroscopy (Figure 4). The CD spectra of  $A\beta$ 42 in buffer alone showed a



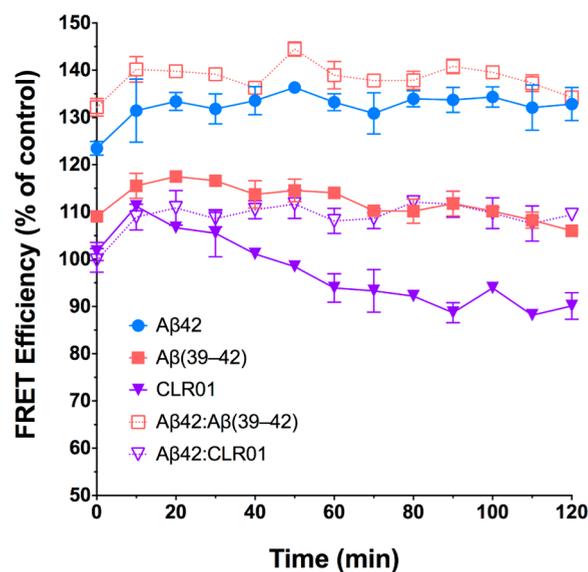
**Figure 4.**  $A\beta$ 42 secondary structure. Time-dependent secondary structure change of  $A\beta$ 42 monitored by CD spectroscopy in the absence (A) or presence (B–E) of vesicles and toxicity inhibitors. (A, B)  $A\beta$ 42; (C)  $A\beta$ 42 +  $A\beta$ (39–42); (D)  $A\beta$ 42 + EGCG; (E)  $A\beta$ 42 + CLR01. The red spectra were recorded at time = 0, while the blue spectra were taken after 4 h incubation. The spectra of the buffer and modulators were subtracted from each presented spectrum. Modulator spectra are presented as dotted lines for reference. Note that the y-axis scale is different for each modulator.

change from an initial mixture of statistical coil and  $\beta$ -sheet to a predominantly  $\beta$ -sheet conformation, reflected in spectral maxima at 195–198 nm and minima at 217–218 nm (Figure 4A).<sup>52</sup> Consistent with previous studies,<sup>53</sup> the presence of the lipid vesicles promoted  $\beta$ -sheet formation of  $A\beta$ 42, giving rise particularly to increased intensity of the maxima at 195–198 nm (Figure 4B).

The three inhibitors had distinct effects on the folding of  $A\beta$ 42 in the presence of vesicles (Figure 4C–E).  $A\beta$ (39–42) appeared not to disrupt  $\beta$ -sheet formation, although signal intensity was reduced by the peptide fragment (Figure 4C). Both EGCG and CLR01, however, significantly disrupted the  $\beta$ -sheet conformation of  $A\beta$ 42. In the case of EGCG addition, the initial spectrum was nearly flat with a shallow minimum at 209 nm. Upon incubation, a maximum developed at 211 nm and a minimum at 228 nm pointing to irregular coil conformations. The CD spectra observed in the presence of

CLR01 suggested traces of  $\beta$ -sheet initially (red spectrum, Figure 4E). By 4 h, a conformational transition took place leading to a shift of the minimum to 210 nm and a decrease in its amplitude, suggesting development of disordered structures. Overall, the CD spectra indicate that EGCG and CLR01 but not  $A\beta$ (39–42) perturbed the structure of  $A\beta$ 42 during the first hours of the assembly reaction and attenuated formation of the typical cross- $\beta$  structure of amyloid fibrils. Similar conformational rearrangements were recorded without DMPC/DMPG vesicles present (Supplementary Figure 1).

In the next part of the study, we focused on the effect of  $A\beta$ 42, the assembly modulators, and their mixtures on the membrane-bilayer structure and properties. First, we examined how addition of  $A\beta$ 42 in the absence or presence of each modulator might affect membrane structure and dynamics. To address this question, we measured Förster resonance energy transfer (FRET) between a lipid-attached fluorescent donor, N-NBD-PE, and acceptor, N-Rh-PE, embedded in DMPC/DMPG vesicles. FRET has been widely used for investigating bilayer dynamics and effects of membrane-active compounds.<sup>54,55</sup> Addition of  $A\beta$ 42 to the vesicles caused an increase of  $\sim$ 30% in FRET efficiency for the duration of the experiment (Figure 5). This increase was not due to interaction



**Figure 5.** Effects  $A\beta$ 42 in the presence or absence of  $A\beta$ (39–42) or CLR01 on energy transfer between fluorescence donor and acceptor embedded within membrane bilayers. Time-dependent changes in the FRET efficiency relative to untreated vesicles, for which the FRET efficiency was considered 100%. The concentration of  $A\beta$ 42 was 30 mM, whereas concentrations of the modulators (together with the peptide or alone) were 150  $\mu$ M. The data are representative of four independent experiments performed in triplicate and are presented as mean  $\pm$  SEM.

of  $A\beta$ 42 with the donor or acceptor themselves, because on their own, both N-NBD-PE and N-Rh-PE displayed a  $\sim$ 10% decrease in fluorescence in the presence of  $A\beta$ 42 (data not shown). The increase above the baseline FRET efficiency (e.g., 100%) suggested that some clustering of the fluorescence donor and acceptor molecules might have occurred, leading to more efficient energy transfer.

Figure 5 shows that addition of  $A\beta$ (39–42) had a smaller effect, increasing FRET efficiency by  $\sim$ 10% relative to baseline, suggesting weaker interaction of the tetrapeptide with the

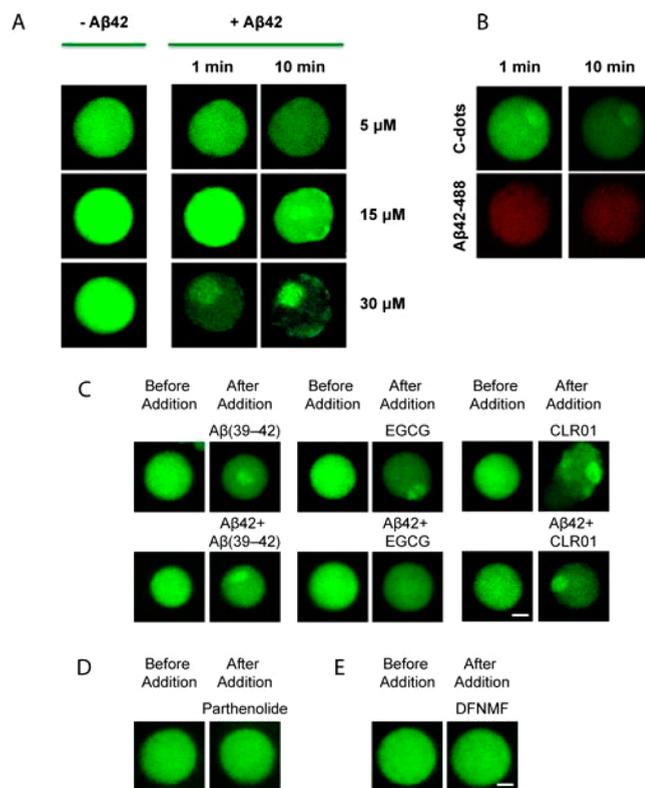
vesicles. The FRET efficiency in the presence of the  $A\beta_{42}$ / $A\beta_{(39-42)}$  mixture appeared to reflect the cumulative effect of both peptides, reaching  $\sim 40\%$  increase in efficiency. CLR01 caused a gradual decrease in FRET efficiency to  $\sim 90\%$  over the duration of the experiment (Figure 5). This effect might be ascribed to weak quenching of the fluorescence by CLR01 or to perturbation of the membrane by CLR01. Interestingly, when a mixture of CLR01 and  $A\beta_{42}$  was incubated with the vesicles, the FRET efficiency was  $\sim 10\%$  above baseline, suggesting that the interaction between  $A\beta_{42}$  and CLR01 negated the individual effect of the peptide ( $\sim 30\%$  increased FRET) and the molecular tweezer ( $\sim 10\%$  decreased FRET) on their own. The data also suggested that the decrease in FRET efficiency in the presence of CLR01 alone was due to membrane perturbation rather than quenching. FRET experiments examining the effect of EGCG were inconclusive, since EGCG had a significant quenching effect of both membrane-embedded fluorophores (Supplementary Figure 2).

To gain further insight into the impact of  $A\beta_{42}$  and the modulators on the membrane structure and dynamics, we used giant vesicles decorated with amphiphilic fluorescent carbon dots and examined them by confocal fluorescence microscopy (Figure 6). Carbon dots are bright, multicolored imaging agents that can be coupled readily to biological assemblies such as cells and vesicles, enabling diverse spectroscopic and microscopic visualization capabilities.<sup>56</sup> A recent study demonstrated that amphiphilic carbon dots could be adsorbed onto the lipid bilayers of giant unilamellar vesicles (GUVs) thereby providing a tool for visualization of membrane-associated processes and vesicle dynamics.<sup>56</sup>

The lipid vesicles used for the confocal fluorescence microscopy experiments in Figure 6 were PC/PG GUVs, which are substantially larger than the small unilamellar DMPC/DMPG vesicles employed in all the experiments described above, to enable microscopic visualization. However, the basic lipid types and polarity of the GUVs were maintained similar to those of the DMPC/DMPG vesicles to allow meaningful comparison with the previous experiments.

Using confocal fluorescence microscopy, we visualized for the first time the effect of  $A\beta_{42}$  on the membrane bilayer within minutes after addition of  $A\beta_{42}$  to lipid vesicles. Prior to addition of  $A\beta_{42}$ , the vesicles exhibited a spherical morphology and uniform surface distribution of the carbon dots (Figure 6A). Addition of  $A\beta_{42}$  caused a dramatic time- and concentration-dependent modification of the vesicle surface organization. Addition of  $5 \mu\text{M}$   $A\beta_{42}$  resulted in slight distortion of the spherical vesicle shape after 10 min (the decrease of the fluorescence signal shown in the figure was recorded also in control vesicles without addition of the peptide and likely is related to bleaching of the fluorescent carbon dots over time). Addition of  $15 \mu\text{M}$   $A\beta_{42}$  modified the vesicle lipid arrangement more strongly giving rise to shape distortion and formation of distinct brighter and darker domains on the vesicle surface. A remarkable fluorescent “spot” was observed upon incubation of the carbon-dot-labeled vesicles with  $30\text{-}\mu\text{M}$   $A\beta_{42}$ .

The clustering of the carbon dots into bright domains is consistent with the FRET results reported above. If similar clustering of the FRET donor and acceptor occurred, FRET efficiency would be higher in the equivalent of the bright domains and lower in the darker domains visualized by the carbon dots. The sum of the change would depend on the specific size and organization of the membrane and, as



**Figure 6.** Interaction of  $A\beta_{42}$  or assembly modulators with GUVs labeled with amphiphilic carbon dots. The images show confocal microscopy images of giant unilamellar PC/PG vesicles labeled with amphiphilic fluorescent carbon dots. (A) GUVs incubated in the absence or presence of unlabeled  $A\beta_{42}$ ; (B) GUVs incubated with carbon dots; (C) GUVs incubated with carbon dots in the absence or presence of  $A\beta_{42}$  labeled with AlexaFluor 488 ( $A\beta_{42}$ –488). The red color was chosen arbitrarily to distinguish the  $A\beta_{42}$ –488 from the green carbon dots. (C) GUVs incubated in the absence or presence of  $A\beta_{42}$  and each of the modulators. (D) GUVs were incubated in the absence or presence of the sesquiterpene lactone parthenolide as a negative control. (E) GUVs were incubated in the absence or presence of the pentapeptide Asp-Phe-Asn-Met-Phe (DFNMF) as a negative control. In all cases,  $A\beta_{42}$  was added at  $30 \mu\text{M}$  and the modulator at  $150 \mu\text{M}$ . The images in panels C–E were taken at  $t = 10 \text{ min}$ . The scale bars correspond to  $5 \mu\text{m}$ .

discussed above, gave rise to an overall increase of  $\sim 30\%$  in FRET efficiency in the presence of  $A\beta_{42}$ .

The observed reorganization of the carbon dots upon addition of  $A\beta_{42}$  suggested two possible scenarios: (1) the carbon-dot reorganization reflected the changes in lipid organization within the membrane vesicles rather than direct interactions between  $A\beta_{42}$  and the vesicle-attached carbon dots or (2) the clustering of the carbon dots paralleled and colocalized with self-association and aggregation of  $A\beta_{42}$ . To distinguish between these two scenarios, we incubated GUVs with carbon dots under the same conditions as in the previous experiment, but this time added  $A\beta_{42}$  fluorescently labeled at the N-terminal amino group by AlexaFluor 488 (Figure 6B). Examination of the images revealed that unlike the carbon dots,  $A\beta_{42}$  did not aggregate during the short period of interaction with the membrane and remained uniformly dispersed in the vesicle. Thus, the data support the first scenario:  $A\beta_{42}$  induces perturbation of the membrane and causes clustering of the carbon dots but does not coassociate with the carbon dots itself.

Addition of each of the modulators to the PC/PG GUVs caused rearrangement of the carbon dots and in the case of EGCG and CLR01 also distortion of the vesicles.  $A\beta(39-42)$  alone induced moderate aggregation of the fluorescent carbon dots on the vesicle surface relative to  $A\beta42$  and did not change the overall spherical morphology of the vesicle. When  $A\beta42$  and  $A\beta(39-42)$  were coadded to the GUVs, similar vesicle surface reorganization was apparent (Figure 6C), suggesting that regardless of its own membrane activity,  $A\beta(39-42)$  offered some protection of the vesicles from the perturbation caused by  $A\beta42$ . A fluorescent “spot” was apparent, similar to the one induced by  $A\beta42$  alone, yet the remaining surface of the vesicle did not become dark to the same extent as with  $A\beta42$  alone (Figure 6A).

In our study design,  $A\beta42$  and the modulators were mixed together prior to addition to the vesicles. This strategy was used to ascertain that neither  $A\beta42$  nor the modulators exhibit (their intrinsic) membrane interactions, and accordingly the analysis would reveal whether new modes of membrane interactions occurred when the peptide and modulators interacted in solution. To validate that the effect of the compounds indeed was on each other, we also tested sequential addition, first  $A\beta42$  and then each of the modulators or vice versa. In all the cases, the initial effect on the distribution of the carbon dots and vesicle shape persisted and was not affected by addition of the second compound ( $A\beta42$  or any of the modulators, data not shown).

Similar to  $A\beta42$  and  $A\beta(39-42)$ , EGCG on its own also modified the distribution of the fluorescent carbon dot on the vesicle surface (Figure 6C). In addition, EGCG caused some distortion of the spherical morphology of the vesicle. The area in which the carbon dots concentrated appeared to protrude above the vesicle surface. Interestingly, when EGCG and  $A\beta42$  were coadded to the GUVs, the vesicle morphology and surface appearance were almost identical to the control vesicles (Figure 6C), suggesting that interaction between  $A\beta42$  and EGCG inhibited membrane disruption and reorganization by both molecules.

CLR01 had the strongest impact on the membrane among the three modulators, inducing pronounced distortion of the spherical vesicle morphology. As observed with EGCG, when CLR01 was co-incubated with  $A\beta42$ , the effect of the molecular tweezer on the vesicles was substantially less pronounced. In the presence of the  $A\beta42$ /CLR01 mixture the vesicles retained their spherical morphology and the clumping of the carbon dots into a bright patch resembled the effect of  $A\beta42$  by itself (Figure 6C).

Because all three modulators appeared to perturb the lipid vesicles, we asked whether this was due to general nonspecific interactions with the vesicles or the carbon dots. To answer this question, we used two unrelated compounds, the anti-inflammatory sesquiterpene lactone parthenolide and the pentapeptide Asp-Phe-Asn-Met-Phe (a short fragment within the protein hormone calcitonin). Neither compound affected the distribution of the carbon dots under identical conditions (Figures 6D,E, respectively), suggesting that the observed membrane perturbation was not due to nonspecific interactions.

This study examined the relationship between the action of  $A\beta42$  toxicity inhibitors and membrane interactions. This aspect of the antitoxic activity of compounds with therapeutic potential in amyloid diseases often has been overlooked although membrane interactions likely have important roles

in mediating the toxic effects of amyloid proteins. Our data clearly show that for the three molecules examined, each representing a distinct class of antitoxic agents, incubation and association with  $A\beta42$  substantially modulated membrane interactions.

The data confirmed that lipid vesicles promote  $\beta$ -sheet formation and fibril assembly by  $A\beta42$ . The distinct role of the vesicle environment was particularly apparent in the ThT fluorescence assay, which demonstrated elimination of the lag phase in the presence of vesicles, in contrast to the buffer solution (Figure 2). Similarly, the CD data showed accelerated and enhanced  $\beta$ -sheet formation when  $A\beta42$  was incubated with DMPC/DMPG vesicles (Figure 4A,B). In the reverse direction, that is, the impact of  $A\beta42$  on the membrane, FRET analysis showed an increase in FRET efficiency between lipid-embedded donor and acceptor upon addition of  $A\beta42$  to DMPC/DMPG vesicles (Figure 5A), suggesting that  $A\beta42$  induced clustering of the donor and acceptor. This interpretation was supported by fluorescence microscopy imaging of PC/PG GUVs labeled with amphiphilic carbon dots. Taken together, the data suggest that  $A\beta42$  causes a substantial and rapid (within a few minutes) lipid reorganization.

Each of the three assembly modulators examined displayed its known effect in solutions containing buffer alone, confirming previous results. The presence of the lipid vesicles had a relatively minor effect on the activity of the modulators. Thus, EGCG and CLR01 prevented the ThT fluorescence increase by  $A\beta42$  regardless of the presence of the lipid vesicles, whereas  $A\beta(39-42)$  had no effect, similar to the findings in buffer alone (Figure 2). Cryo-TEM examination of  $A\beta42$  in the presence of vesicles revealed formation of scarce, occasional fibrils in reaction mixtures with EGCG or CLR01 (Figure 3). Similarly, the presence of lipid vesicles induced only minor changes in the CD spectra of  $A\beta42$  in the presence of each modulator, and the kinetics of secondary-structure transition remained largely the same in each case (Figure 4). Overall, the data supported the notion that  $A\beta42$  aggregation is facilitated by lipid vesicles but suggested that disruption of the aggregation by effective assembly modulators, such as EGCG or CLR01, is not affected by the membranes. In other words, in the push-pull competition between the membranes and the modulators, the modulators have the upper hand.

In the case of EGCG, this conclusion contradicts the observations by Engel et al., who reported that EGCG was a substantially less efficient inhibitor of amyloid formation by islet amyloid polypeptide (IAPP) in the presence of phospholipids than in simple buffer solutions.<sup>57</sup> The difference between our results and those of Engel et al. may simply reflect the fact that  $A\beta42$  and IAPP are different peptides. In addition, conceivably, the differences could also be related to the distinct charge states of IAPP and  $A\beta42$ . In both studies, the vesicles had a negatively charged surface. We used a mixture of neutral DMPC and negatively charged DMPG, and Engel et al. used negatively charged dipalmitoylphosphatidylglycerol (DPPG) vesicles. In contrast to  $A\beta42$ , which has a net charge of  $-3$  at physiologic pH, IAPP has a net charge of  $+3$ , which makes its attraction to the vesicles substantially higher than that of  $A\beta$ , providing a plausible explanation for the lower impact of EGCG on IAPP than on  $A\beta42$  in the presence of negatively charged vesicles. If this explanation is correct, the opposite would be expected in the presence of positively charged lipid membranes, that is, EGCG would be expected to have a lower effect on  $A\beta42$

aggregation than on IAPP aggregation. Moreover, different lipid binding mechanisms may also play role. IAPP contains an amphipathic  $\alpha$ -helix that can participate in binding membranes; whereas, the  $A\beta$  peptide seems to anchor into membranes more directly via the hydrophobic C-terminus.

In contrast to the unremarkable impact of the membrane vesicles on the capability of the modulators to impact  $A\beta$ 42 assembly, both  $A\beta$ 42 and each modulator appeared to have a substantial impact on the organization and dynamics of the lipid molecules within the vesicles. This was apparent by FRET measurements, which suggested that  $A\beta$ 42 and, to a lower extent,  $A\beta$ (39–42) induced rapid clustering of the FRET donor and acceptor. In contrast, CLR01 caused slow reduction of the FRET efficiency reaching a  $\sim$ 10% decrease by 2 h, suggesting perturbation of the vesicles in a manner that increased the average distance between the donor and acceptor molecules. Interestingly, this effect was reversed when CLR01 was added to the vesicles in the presence of  $A\beta$ 42, even though the concentration of CLR01 was 5-times that of  $A\beta$ 42.

These observations could be explained by the fact that  $A\beta$ 42 may bind up to three CLR01 molecules<sup>30</sup> and suggest that the affinity of CLR01 for  $A\beta$ 42 is substantially higher than that for the vesicles, which would be expected, because the molecular tweezer is known to bind with low micromolar affinity to lysine residues. In addition, CLR01 binds arginine residues with 5–10 times lower affinity. Upon binding of CLR01 to the lysine and arginine residues in  $A\beta$ , the charge of these residues is reversed for each lysine or arginine residue from positive (+1) to negative (ca. – 2) depending on the protonation state of the CLR01 phosphate groups,<sup>58</sup> increasing the net charge of  $A\beta$ 42 from –3 up to –6. Thus, the complex of  $A\beta$ 42 and CLR01 is expected to have a lower affinity for the membrane vesicles than  $A\beta$ 42 alone. Similarly, the tendency of CLR01 to interact with the phospholipid molecules was reduced, likely due to engagement of its hydrophobic “arms” with the peptide, making them are less available for interaction with the membrane.

The fluorescence microscopy experiments utilizing carbon-dot-labeled GUVs revealed that when each of the compounds we used (including  $A\beta$ 42 itself) was added individually to the vesicles it induced bilayer reorganization and in some cases caused different degrees of deformation of the globular structure of the vesicle. These membrane interactions were weaker in each case when instead of the individual compound, the mixture of each modulator with  $A\beta$ 42 was added to the lipid vesicles, supporting the notion that in the interplay of the interaction of  $A\beta$ 42, the lipid vesicles, and the toxicity inhibitors, the binding of the inhibitors to  $A\beta$ 42 was the predominant interaction, overshadowing the interaction between  $A\beta$ 42 and the membrane or the modulators and the membrane. These data suggest that in addition to their impact on  $A\beta$ 42 assembly, an important mechanism by which the molecules examined here inhibit  $A\beta$ 42-induced toxicity may be by reducing the interaction of  $A\beta$ 42 with cell membranes. In particular, this mechanism may contribute to the inhibitory effect of  $A\beta$ (39–42), which was shown to inhibit the toxicity of  $A\beta$ 42 in several cell culture tests, including cell viability and electrophysiologic assays,<sup>18,59</sup> without affecting  $A\beta$ 42 assembly into amyloid fibrils.<sup>34</sup>

In summary, our study illuminates a rarely studied angle of toxicity inhibitors' activity, their effect on membrane interactions of  $A\beta$ 42. The involvement of *membranes* and *membrane interactions* in  $A\beta$  toxicity in general,  $A\beta$ 42 toxicity in

particular, have become a widely accepted paradigm, and there have been numerous studies focusing on this aspect of amyloid biology. However, there is no consensus yet as to what exactly are the underlining links between membrane disruption and amyloid toxicity. The clear and significant modulation of membrane reorganization upon the interactions of  $A\beta$ 42 and the modulators constitute an important contribution to understanding the activity of the three inhibitors and their possible therapeutic uses.

The data suggest that the interaction of  $A\beta$ 42 with effective assembly modulators, EGCG and CLR01, is stronger than its interaction with membranes and that interaction with all three toxicity inhibitors partially shields  $A\beta$ 42 from interacting with the membranes, contributing to the protective effect of these compounds. An open and intriguing question, which will require additional future investigation, is why the interaction of each of the compounds with the lipid vesicles, which in certain cases appeared to induce stronger perturbation than that of  $A\beta$ 42, does not lead to apparent toxicity. Further high-resolution structural insights into the inhibitor–peptide–membrane system will contribute to better understanding of these issues.

## METHODS

**Materials.**  $A\beta$ 42 was purchased from AnaSpec (USA) in a lyophilized form at >95% purity.  $A\beta$ (39–42) was purchased from Pepton (South Korea) in a lyophilized form at >90% purity. *L*- $\alpha$ -Phosphatidylcholine (egg, chicken), *L*- $\alpha$ -phosphatidylglycerol (egg, chicken, sodium salt), 1,2-dimyristoyl-*sn*-glycero-3-phosphocholine (DMPC), 1,2-dimyristoyl-*sn*-glycero-3-[phospho-*rac*-(1-glycerol)] (DMPG), 1,2-dimyristoyl-*sn*-glycero-3-phosphoethanolamine-*N*-(7-nitro-2-1,3-benzoxadiazol-4-yl) (*N*-NBD-PE), and 1,2-dimyristoyl-*sn*-glycero-3-phosphoethanolamine-*N*-(lissamine rhodamine B sulfonyl) (*N*-Rh-PE) were purchased from Avanti Polar Lipids. Thioflavin T (ThT), 1,1,1,3,3,3-hexafluoro-2-propanol (HFIP), sodium hydro-sulfite, sodium phosphate monobasic, and EGCG, >95% purity, were purchased from Sigma-Aldrich (Rehovot, Israel). CLR01 was prepared and purified as a  $\text{Li}^+$  salt as described previously.<sup>30,49</sup>

**Peptide and Sample Preparations.**  $A\beta$ 42 was dissolved in HFIP at a concentration of 1 mg/mL and stored at –20 °C until use to prevent aggregation. For each experiment, the solution was thawed, and the required amount was dried by evaporation for 6–7 h to remove the HFIP. The dried peptide sample was dissolved in 10 mM sodium phosphate, pH 7.4, at room temperature. Stock solutions of  $A\beta$ (39–42), EGCG, and CLR01 were prepared at 5 mM in deionized water and diluted into the  $A\beta$ 42 solutions at the required concentration.

**Thioflavin T (ThT) Fluorescence Assay.** ThT fluorescence measurements were conducted at 37 °C using 96-well path cell culture plates on a Varioskan plate reader (Thermo, Finland). Measurements were made on samples containing 30  $\mu\text{M}$   $A\beta$ 42 in the absence or presence of  $A\beta$ (39–42), EGCG, or CLR01 at 1:5  $A\beta$ /inhibitor concentration ratio and in the absence or presence of lipid vesicles (final concentration 1 mM). A 192- $\mu\text{L}$  aliquot of the aggregation reaction was mixed with 48  $\mu\text{L}$  of 100  $\mu\text{M}$  ThT in sodium phosphate, pH 7.4, at different time points. The fluorescence intensity was measured following a 10 min incubation at  $\lambda_{\text{ex}} = 440$  and  $\lambda_{\text{em}} = 485$  nm.

**Cryogenic Transmission Electron Microscopy (Cryo-TEM).** Cryo-TEM imaging of aliquots taken from the same reaction mixtures used in the ThT experiments after 12-h incubation was carried out as follows: A 3- $\mu\text{L}$  droplet of the reaction mixture was deposited on a glow-discharged TEM grid (300 mesh Cu Lacey substrate grid; Ted Pella). The excess liquid was blotted with a filter paper, and the specimen was rapidly plunged into liquid ethane precooled with liquid nitrogen in a controlled environment (Leica EM GP). The vitrified samples were transferred to a cryo-specimen holder (Gatan model

626) and examined at  $-181\text{ }^{\circ}\text{C}$  using a FEI Tecnai 12 G2 TWIN TEM operated at 120 kV in low-dose mode. Grids were imaged a few micrometers under focus to increase phase contrast. The images were recorded with a Gatan charge-coupled device camera (model 794) and analyzed by Digital Micrograph software, Version 3.1.

**Circular Dichroism (CD) Spectroscopy.** CD spectra were recorded in the range of 190–260 nm at room temperature on a Jasco J-715 spectropolarimeter, using 1 mm quartz cuvettes. Solutions composed of 400  $\mu\text{L}$  contained 50  $\mu\text{M}$  A $\beta$ 42 in the absence or presence of 5-fold excess A $\beta$ (39–42), EGCG, or CLR01 and in the absence or presence of 1 mM lipid vesicles. Spectra were recorded every 1 h for 4 h. CD signals resulting from vesicles and buffer were subtracted from the corresponding spectra.

**Förster Resonance Energy Transfer (FRET).** Small unilamellar vesicles (SUVs, DMPC/DMPG at 1:1 molar ratio) were prepared by dissolving the lipid components in chloroform/ethanol and drying together under vacuum, followed by dissolution in sodium phosphate, pH 7.4, and sonication of the aqueous lipid mixture at room temperature for 10 min using a Sonics vibracell VCX130 ultrasonic cell disrupter. Prior to drying, the lipid vesicles were supplemented with N-NBD-PE and N-Rh-PE at a 500:1:1 molar ratio, respectively. A $\beta$ 42 (30  $\mu\text{M}$ ) in the absence or presence of A $\beta$ (39–42), EGCG, or CLR01 at 1:5 A $\beta$ 42/modulator concentration ratio was added to the vesicles (final vesicle concentration 1 mM) at  $t = 0$ . Fluorescence emission spectra were acquired at different time points up to 2 h ( $\lambda_{\text{ex}} = 469\text{ nm}$ ) in the range of 490–650 nm using a Varioskan 96-well plate reader (Thermo, Finland).

To calculate the extent of FRET efficiency, the following equation was used:

$$\text{efficiency} = \frac{R_1 - R_{100\%}}{R_0 - R_{100\%}} \times 100\%$$

in which  $R$  is a ratio of fluorescence emission of NBD-PE (531 nm)/rhodamine B-PE (591 nm).  $R_1$  is the ratio in the peptide/vesicle mixtures,  $R_{100\%}$  was measured following the addition of 20% Triton X-100 to the vesicles, which causes complete dissolution of the vesicles, and  $R_0$  corresponds to the ratio recorded for vesicles without any additives.

**Giant Unilamellar Vesicles Labeled with Amphiphilic Carbon Dots.** Amphiphilic carbon dots were prepared according to a published protocol.<sup>56</sup> Briefly, preparation of the carbon dots was carried out in an aqueous solution, and started with *O,O'*-dilauroyl tartaric acid anhydride produced through reacting *L*-tartaric acid with lauryl chloride. Subsequent reaction with *D*-glucose and hydrothermal carbonization yielded carbon dots exhibiting inner graphitic cores coated with an amphiphilic layer comprising alkyl chains and carboxylic acid moieties.<sup>56</sup>

Giant unilamellar vesicles (GUVs) were prepared through the rapid evaporation method.<sup>60</sup> Briefly, GUVs comprising egg-PC and egg-PG (1:1 mol ratio) were prepared through dissolving the lipid constituents with 1 mg of carbon quantum dots dissolved in 500  $\mu\text{L}$  of chloroform through vortexing and sonication. The mixture was then transferred to a 250 mL round-bottom flask, and the aqueous phase (2.5 mL of 0.1 M sucrose, 0.1 mM KCl, 50 mM Tris solution, pH 7.4) was added carefully with a pipet and stirred gently for  $\sim 5$  min. The organic solvent was removed in a rotary evaporator under reduced pressure (final pressure 20 mbar) at room temperature. After evaporation for 4–5 min, an opalescent fluid was obtained with a volume of approximately 2.5 mL.

**Confocal Fluorescence Microscopy.** GUVs were imaged in the absence or presence of A $\beta$ 42, inhibitors, or their mixtures using a PerkinElmer UltraVIEW system equipped with an Axiovert-200 M (Zeiss, Germany) microscope and a Plan-Neofluar 63 $\times$ /1.4 oil objective. The excitation wavelengths of 440 and 488 nm were generated by an Ar/Kr laser.

## ■ ASSOCIATED CONTENT

### 📄 Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acschemneuro.5b00200.

CD data of A $\beta$ 42 and inhibitors in buffer and FRET data for EGCG (PDF)

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### Notes

The authors declare no competing financial interest.

## ■ ABBREVIATIONS

AD, Alzheimer's disease; A $\beta$ , amyloid  $\beta$ ; EGCG, (–)-epigallocatechin-3-gallate

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