

UC Irvine

UC Irvine Previously Published Works

Title

Precision pharmacology for Alzheimer's disease

Permalink

<https://escholarship.org/uc/item/5bg2p7pc>

Authors

Hampel, Harald
Vergallo, Andrea
Aguilar, Lisi Flores
et al.

Publication Date

2018-04-01

DOI

10.1016/j.phrs.2018.02.014

Peer reviewed



Published in final edited form as:

Pharmacol Res. 2018 April ; 130: 331–365. doi:10.1016/j.phrs.2018.02.014.

Precision pharmacology for Alzheimer's disease

Harald Hampel^{a,b,c,d,*}, Andrea Vergallo^{a,b,c,d}, Lisi Flores Aguilar^e, Norbert Benda^f, Karl Broich^g, A. Claudio Cuello^h, Jeffrey Cummingsⁱ, Bruno Dubois^{b,c,d}, Howard J. Federoff^j, Massimo Fiandaca^{k,l,m}, Remy Genthon^{b,d}, Marion Haberkampⁿ, Eric Karran^{o,p,q}, Mark Mapstone^k, George Perry^r, Lon S. Schneider^s, Lindsay A. Welikovitsh^t, Janet Woodcock^u, Filippo Baldacci^{a,b,c,d,v}, Simone Lista^{a,b,c,d,*} Alzheimer Precision Medicine Initiative (APMI)

^aAXA Research Fund & Sorbonne University Chair, Paris, France

^bSorbonne University, GRC No.21, Alzheimer Precision Medicine (APM), AP-HP, Pitié-Salpêtrière Hospital, Boulevard de l'hôpital, F-75013, Paris, France

^cBrain & Spine Institute (ICM), INSERM U 1127, CNRS UMR 7225, Boulevard de l'hôpital, F-75013, Paris, France

^dInstitute of Memory and Alzheimer's Disease (IM2A), Department of Neurology, Pitié-Salpêtrière Hospital, AP-HP, Boulevard de l'hôpital, F-75013, Paris, France

^eDepartment of Anatomy and Cell Biology, McGill University, Montreal, QC, Canada

^fBiostatistics and Special Pharmacokinetics Unit/Research Division, Federal Institute for Drugs and Medical Devices (BfArM), Bonn, Germany

^gFederal Institute for Drugs and Medical Devices (BfArM), Bonn, Germany

^hDepartment of Pharmacology and Therapeutics, McGill University, Montreal, QC, Canada

ⁱCleveland Clinic Lou Ruvo Center for Brain Health, Las Vegas, NV, USA

^jHealth Affairs CEO, UCI Health, University of California, Irvine, CA, USA

^kDepartment of Neurology, Translational Laboratory and Biorepository, University of California Irvine School of Medicine, Irvine, CA, USA

*Corresponding authors at: AXA Research Fund & Sorbonne University Chair, Sorbonne University, Department of Neurology, Institute of Memory and Alzheimer's Disease (IM2A), Brain & Spine Institute (ICM), François Lhermitte Building, Pitié-Salpêtrière Hospital, 47 Boulevard de l'hôpital, 75651 Paris CEDEX 13, France. harald.hampel@icm-institute.org (H. Hampel), simone.lista@icm-institute.org (S. Lista).

Contributors to the Alzheimer precision medicine initiative – working group (APMI-WG)

Aguilar LF (Montréal), Babiloni C (Rome), Baldacci F (Pisa), Benda N (Bonn), Black KL (Los Angeles), Bokde ALW (Dublin), Bonuccelli U (Pisa), Broich K (Bonn), Bun RS (Paris), Cacciola F (Siena), Castrillo J† (Derio), Cavedo E (Paris), Ceravolo R (Pisa), Chiesa PA (Paris), Colliot O (Paris), Coman CM (Paris), Corvol JC (Paris), Cuello AC (Montréal), Cummings JL (Las Vegas), Depypere H (Gent), Dubois B (Paris), Duggento A (Rome), Durrleman S (Paris), Escott-Price V (Cardiff), Federoff H (Irvine), Ferretti MT (Zürich), Fiandaca M (Irvine), Frank RA (Malvern), Garaci F (Rome), Genthon R (Paris), George N (Paris), Giorgi FS (Pisa), Graziani M (Roma), Haberkamp M (Bonn), Habert MO (Paris), Hampel H (Paris), Herholz K (Manchester), Karran E (Cambridge), Kim SH (Seoul), Koronyo Y (Los Angeles), Koronyo-Hamaoui M (Los Angeles), Lamari F (Paris), Langevin T (Minneapolis-Saint Paul), Lehericy S (Paris), Lista S (Paris), Lorenceau J (Paris), Mapstone M (Irvine), Neri C (Paris), Nisticò R (Rome), Nyasse-Messene F (Paris), O'Bryant SE (Fort Worth), Perry G (San Antonio), Ritchie C (Edinburgh), Rojkova K (Paris), Rossi S (Siena), Santarnecchi E (Siena), Schneider LS (Los Angeles), Sporns O (Bloomington), Toschi N (Rome), Verdooner SR (Sacramento), Vergallo A (Paris), Villain N (Paris), Welikovitsh L (Montréal), Woodcock J (Silver Spring), Younesi E (Esch-sur-Alzette).

^lDepartment of Neurological Surgery, University of California Irvine School of Medicine, Irvine, CA, USA

^mDepartment of Anatomy & Neurobiology, University of California Irvine School of Medicine, Irvine, CA, USA

ⁿNeurology/Psychiatry/Ophthalmology Unit, Federal Institute for Drugs and Medical Devices (BfArM), Bonn, Germany

^oFoundational Neuroscience Center (FNC), AbbVie Neuroscience, Cambridge, MA, USA

^pDepartment of Molecular Neuroscience, Institute of Neurology, University College London, London, UK

^qInstitute of Neurodegenerative Diseases, Catholic University of Leuven, Leuven, Belgium

^rCollege of Sciences, One UTSA Circle, The University of Texas at San Antonio, San Antonio, TX, USA

^sKeck School of Medicine of the University of Southern California, Los Angeles, CA, USA

^tDepartment of Neurology and Neurosurgery, McGill University, Montreal, QC, Canada

^uCenter for Drug Evaluation and Research, US Food and Drug Administration, Silver Spring, MD, USA

^vDepartment of Clinical and Experimental Medicine, University of Pisa, Pisa, Italy

Abstract

The complex multifactorial nature of polygenic Alzheimer's disease (AD) presents significant challenges for drug development. AD pathophysiology is progressing in a non-linear dynamic fashion across multiple systems levels – from molecules to organ systems – and through adaptation, to compensation, and decompensation to systems failure. Adaptation and compensation maintain homeostasis: a dynamic *equilibrium* resulting from the dynamic non-linear interaction between genome, epigenome, and environment. An individual vulnerability to stressors exists on the basis of individual triggers, drivers, and thresholds accounting for the initiation and failure of adaptive and compensatory responses. Consequently, the distinct pattern of AD pathophysiology in space and time must be investigated on the basis of the individual biological makeup. This requires the implementation of systems biology and neurophysiology to facilitate Precision Medicine (PM) and Precision Pharmacology (PP).

The regulation of several processes at multiple levels of complexity from gene expression to cellular cycle to tissue repair and system-wide network activation has different time delays (temporal scale) according to the affected systems (spatial scale). The initial failure might originate and occur at every level potentially affecting the whole dynamic interrelated systems within an organism. Unraveling the spatial and temporal dynamics of non-linear pathophysiological mechanisms across the *continuum* of hierarchical self-organized systems levels and from systems homeostasis to systems failure is key to understand AD. Measuring and, possibly, controlling space- and time-scaled adaptive and compensatory responses occurring during AD will represent a crucial step to achieve the capacity to substantially modify the disease course and progression at the best suitable timepoints, thus counteracting disrupting

critical pathophysiological inputs. This approach will provide the conceptual basis for effective disease-modifying pathway-based targeted therapies.

PP is based on an exploratory and integrative strategy to complex diseases such as brain proteinopathies including AD, aimed at identifying simultaneous aberrant molecular pathways and predicting their temporal impact on the systems levels. The depiction of pathway-based molecular signatures of complex diseases contributes to the accurate and mechanistic stratification of distinct subcohorts of individuals at the earliest compensatory stage when treatment intervention may reverse, stop, or delay the disease. In addition, individualized drug selection may optimize treatment safety by decreasing risk and amplitude of side effects and adverse reactions.

From a methodological point of view, comprehensive “omics”-based biomarkers will guide the exploration of spatio-temporal systems-wide morpho-functional shifts along the *continuum* of AD pathophysiology, from adaptation to irreversible failure.

The Alzheimer Precision Medicine Initiative (APMI) and the APMI cohort program (APMI-CP) have commenced to facilitate a paradigm shift towards effective drug discovery and development in AD.

Keywords

Alzheimer’s disease; Precision pharmacology; Precision medicine; Pathway-based therapy; Pathophysiology; Clinical trials

1. Introduction: precision pharmacology in the context of precision medicine

Complex chronic diseases with global unmet needs such as cancer, diabetes, immune diseases, and brain proteinopathies – including Alzheimer’s disease (AD) – primarily exhibit: I) a multifactorial nature, due to the coexistence of polygenetic/genomic/epigenomic, interactomic, and environmental susceptibility and II) altered networks, affecting relevant modules and interactomes [1,2]. The continuous failure of late stage clinical drug trials, largely developed following the traditional drug development paradigm in AD, demonstrates that a conceptual shift in Drug Discovery & Development programs is required to attain successful breakthrough developments of novel therapies [3,4]. Notably, a critical step for developing effective drugs is to explore and predict the comprehensive effect of a compound on four fundamental levels, such as I) hitting the intended target, II) altering the intended mechanism, III) altering the relevant pathophysiology, and IV) impacting clinical outcome [1].

Precision pharmacology (PP) is a novel conceptual paradigm that aims at exploring and predicting the whole effect of a molecular mechanism of action, i.e. the pharmacodynamic (PD) [5]. As a result, PP is crucial to operate from the perspective of an innovative exploratory, integrative, holistic multi-paradigm or systems level concept, at both experimental and computational level. In order to achieve the full understanding of drug action at the systems level, it is necessary to combine disease mechanism, PD and pharmacokinetic (PK) data into a single model, following the systems pharmacology

paradigm. According to the American Association of Pharmaceutical Scientists (AAPS; <https://www.aaps.org/>), systems pharmacology is defined as “the science of advancing knowledge about drug action at the molecular, cellular, tissue, organ, organism, and population levels” (available at http://www.aaps.org/Systems_Pharmacology/). Systems pharmacology is an integrative interdisciplinary model providing the potential to investigate drug action through networks of biological pathways, thus allowing the development of predictive models of PD and PK features for a certain molecule [6,7]. Therefore, traditional PK/PD procedures are integrated into the systems biology paradigm to establish predictive models of the whole effect (up-downstream regulated processes, feedback loops) of a given drug, from cell pathway signals to systems outputs [5–7]. This paradigm can lead to the characterization of pathway-based molecular signatures that will allow a mechanistic stratification of individuals and patients for a “stratified pathway-based therapy” [6–8].

The existence of high interindividual variability underlying a genetic/epigenetic different background primarily affects the mechanism of action of the drug under study (Table 1A–H). This categorization process inside drug development relies on the “omic” sciences and aims at achieving personalized predictive models of therapeutic effects, side effects, and adverse [3,9]. Applying PP is assumed to accomplish the following long-term goals: I) developing multi-target therapeutic approaches for multifactorial polygenic diseases, such as AD; and II) providing predictive models/quantitative frameworks of therapeutic efficacy and risk of adverse events for individuals, in the context of Precision Medicine (PM) [3,6]. The implementation of PP in AD is anticipated to result into an innovative and original scientific taxonomy as well as to a distinguished working lexicon and terminology (Table 2). In order to accelerate the development of the PM paradigm in AD, the international Alzheimer PM Initiative (APMI) and its related Cohort Program (APMI-CP) have been established by our consortium and conceptually associated to the U.S. Precision Medicine Initiative (PMI) (available at <https://www.whitehouse.gov/precision-medicine>) and the U.S. “All of Us Research Program” – evolved from the U.S. PMI Cohort Program (available at <https://www.nih.gov/research-training/allofus-research-program>). The research using the APMI cohorts has recently commenced to be facilitated under the structural framework of the newly established French Sorbonne University – “Clinical Research Group in Alzheimer Precision Medicine” (*Sorbonne Université – “Groupe de Recherche Clinique – Alzheimer Precision Medicine”, [GRC n° 2I]*).

Combined downstream and upstream effects on different homeostatic key molecules and pathways are commonly shared on several biological networks which, in turn, underlie apparently unrelated diseases [10]. Pathway-based therapies are anticipated to support the development of novel interventions to treat several diseases which can show misleading clinical divergence. Given the complexity and heterogeneity of many diseases, such as AD, a multi-target approach needs to be performed; in particular, the main “orchestrator” of each pathway – ultimately called target – will be identified by an integrative analysis of comprehensive multi-domain “omic” [2,3,11]. In addition, this advanced holistic systems-level approach is assumed to facilitate the drug repositioning process – also known as drug re-profiling or drug repurposing process – indicating that a drug with a recognized biological effect could be utilized to treat a disease for which it has not been registered [12].

1.1. The road to precision pharmacology: role and contribution of time and space in systems biology for research & development programs

The application of systems biology to investigate multifactorial diseases starts from the elucidation of all gene-interaction networks since complex gene-gene and gene-environment interactions upstream affect the biochemical pathways underpinning the disease with high extent of variability across a patient [3,12]. Therefore, the development of advanced computational/bioinformatic tools made the detection of statistical interactions between genetic loci possible, when examining the data *via* genome-wide association studies (GWAS) [3,9]. Exploratory computational platforms will allow quantitative and dynamic modeling of interacting biological systems active at multiple scales of organization within a *continuum*, i.e. from homeostasis to system failure. Currently available biostatistical approaches facilitate researchers in providing the profile of gene clusters related to several biological processes. There is a growing number of technologies allowing the optimization of data collection from a single biofluid or tissue sample by providing a multimodal profiling, such as genomic/epigenomic, transcriptomic, miRNAomic, proteomic, and metabolomic/lipidomic [13–16].

Charting the molecular dysregulated pathways should be accomplished using pathway-based panels that contain multiple combinations of arrays encompassing several genes, in order to track their direct expression products and the most relevant gene-gene interactions [2,9,17]. This is supposed to substantially transform the Research and Development (R&D) programs, thus paving the way for developing “molecularly” biomarker-guided targeted therapies[18] – i.e., treatments specifically adapted (“tailored”) to the individual – within a short time frame [3,12].

1.1.1. Role of time—The addition of a fourth dimension – time – to the field of structural biology will allow following-up compensatory mechanisms responsible for preserving homeostasis and its dynamic changes over time. In this regard, the identification of transcriptionally active genes and their respective products is a key signature of either active “stress responses” or dynamic loss of homeostasis. Nowadays, the role of advanced nanotechnologies able to dynamically track the time/space coordinates of molecules associated with different pathways is gaining substantial relevance. Expression profiles of genes and proteins are supposed to provide clear outcome measures, i.e., biomarkers, for target engagement as well as for predicting the response to treatment. Simultaneous gene expression and extracellular protein expression profiles can allow exploring a whole cellular species, for instance, to longitudinally investigate immune responses and cell ultrastructural alterations over time. In particular, both overactivation and changes in immune cell surface antigens occur in parallel with the progression of a wide variety of pathophysiological conditions such as in AD and [13,16,17]. As a result, biomarker-guided pathway-based therapies shaped on the comprehensive biological profile of a given subject at a given time point of the disease progression will change according to the evolving biological pattern of the individual.

1.1.2. Role of space—There is a heterogeneous cross-talk between periphery and central nervous system (CNS) pathways, based, for instance, on innate-adaptive

immune system and proteostasis networks. Interestingly, several peripheral and systemic abnormalities have been found to be associated with impaired amyloid beta (A β) peptides removal at the level of the CNS. This suggests a crucial role for brain-periphery interaction in the development and progression of brain proteinopathies, including AD [19,20]. Recently, and even more related to AD, an association between peripherally-derived neutrophils, T-regulatory lymphocytes, as well as peripheral immunity loss of function and microglial dysfunction that resulted in protein misfolding has been reported in brain or other tissues [21,22].

In summary, understanding the dynamic regulation of transcellular signals at a system level as well as the mechanisms underlying their bi-directional cross-talks is expected to restore aberrant pathways in pathophysiologically altered tissues/organs by targeting, in turn, other tissues/organs. These insights will promote the identification of remote (i.e., peripheral) key modulators of several cerebral functions, thus providing a reliable open-access to the brain. This step is essential to overcome the high degree of inaccessibility of the brain to pharmacological therapies.

2. Homeostasis and pathway-based therapy

Loss of homeostasis, ultimately leading to a dynamic pathophysiological state, consists of the breakdown of one or more homeodynamic pathways – namely the “stress responses” – originating first of all from maladaptive responses and then from failure of compensatory mechanisms (i.e. decompensation). Compensation is a self-regulatory dynamic counterbalance between regulatory defense mechanisms and disrupting stress-induced signals [23–25]. Compensation occurs through both structural and functional changes and is hierarchically organized from subcellular to cellular level, organs, and, eventually, systems. Compensatory mechanisms aim primarily at protecting the core biosynthetic processes necessary to survival. There is a *continuum* between homeostasis, metastability that precedes adaptation – compensation with an higher risk of failure of compensatory mechanisms over time – finally leading to loss of homeostasis [25–27]. In this scenario, disease is designated as a theoretical construct exhibiting successive and progressive failures (decompensation) in complex interconnected systems or brain networks, according to the notion of “systems failure” [3,9,11]. The primary descriptive concept of this model is that this construct is mostly not the linear result of a unitary etiologic factor; rather, it evolves in time in a non-linear dynamic progressive fashion across physiological and, then, pathophysiological stages – from initial adaptation to compensation and after thresholds to decompensation (leading to failure of homeostatic mechanisms) – and the convergence of failures in several networks/systems, or pathophysiological processes along a *continuum* (Fig. 1).

All living organisms, from nematodes to human beings, are continuously exposed to stress-associated signals triggered by a wide range of endogenous and external stimuli, including physical activity, temperature, UV rays, cosmic radiation, oxidants, bioenergetic restrictions, chronic cellular exposure to impaired metastable proteome and/or conformational [28,29]. Interestingly, the degree of the cellular homeodynamic “stress response” differs in terms of

amplitude and time (short-term/long-term) according to the extent of the stressful stimuli [28,29].

Cellular homeostasis represents the critical point of the individual's health span and refers to all molecular machineries needed at multiple cellular-subcellular compartments to compensate for stress-induced damage, thus finally preserving the cellular functional and metabolic stability. The existence of cellular homeostasis is ensured by "stress responses", including: I) proteostasis networks (exerting mechanisms quality control, from protein synthesis to protein degradation [30,31], II) highly conserved pro-survival and pro-apoptotic gene expression pathways (responsible for multiple level regulation, i.e., from pre-transcriptional to cell trafficking level [23,24,32]).

Several studies have shown that preserving cellular homeostasis generally affects the individual's life span while its deterioration over time underlies aging in a bidirectional way [33]. Age-related alterations affecting "stress responses" mainly occur at molecular level: glucose transport, DNA surveillance mechanisms (ensuring repair of DNA lesions), and mitochondrial electron transport play a crucial role to support pro-survival signaling as well as cell development/differentiation, apoptosis, endocytosis, microtubule stability, lipid membrane dynamics, and other key molecular processes [34–36]. As a result, DNA damage, overexpressed oxidative stress, and telomere shortening are typical patterns of aged cells displaying functional decline. This, in turn, has a significant impact on proteostasis leading to a fatal accumulation of misfolded proteins over nucleic acids, lipids, and other molecules. Notably, it is fully acknowledged – also in humans – that the uncontrolled activation of "stress response" pathways is expected to determine loss of homeostasis *via* several mechanisms, in particular through down-regulation performed by negative auto-feedback and bioenergetic depletion due to hyperactivated pathways [29,37,38].

At present, the homeostatic mechanisms have not been completely elucidated. However, it is clear that there is a complex bi-directional crosstalk among numerous anti-stress outputs (Natarajan M et al., 2006) intensifying the presence of intricate networks, where different pathways constitute central hubs coordinating various modules. As a result, the dysfunction of a single component of the network may appear as both the cause and the consequence of the dysfunction of other components, hence substantially and dynamically impacting the whole network [28,38] (Fig. 1).

The comprehensive assessment of the dynamic and mutual interplay among the various cellular "stress response" pathways modulating the individual's life span and aging, will allow disclosing novel insights on aberrant biological conditions. This, in turn, will represent a critical step for developing drugs with efficacy for unresolved medical challenges such as cancer, immune diseases, diabetes, AD (and other brain proteinopathies).

Therefore, a systems biology-based biomarker-guided multi-target therapy relies on a multi-pathway- or multi-network-based approach, which, in the case of AD, should engage selected molecular targets concerning: proteostasis network, immune response (both innate and adaptive) and endothelial dysfunction. In the perspective of a pathway-/network-based drug development strategy, similar systems failures sharing common pathophysiological

pathways appearing as “different diseases” are potentially supposed to be treated with the same molecule [38–41].

Big “omic” data need to be generated from multiple systems levels and integrated to achieve reliable information about the dynamic failures to compensate for complex disruptive signaling that can lead to the disease. Given the partly undiscovered substantial cross-talk between CNS and peripheral systems, it is acknowledged that longitudinal trajectories of blood biomarkers reflect the changes over-time in the interaction between aberrant cerebral networks and peripheral networks. For instance, blood-based inflammatory and metabolomic markers allow to *in vivo* track crucial mechanisms accounting for the pathophysiological evolution of AD along the *continuum*, from adaptation, compensation to decompensation and systems failure and from the earliest preclinical stages to late stage clinical dementia[42].

3. Current status of blood-based biomarkers – inflammatory and metabolomic – for preclinical Alzheimer’s disease

Detailed pathological analyses at autopsy, with the addition of surgical pathology and biochemical studies, have evolved to provide a basis for detecting many human diseases, especially when combined with precise clinical assessments, as in the traditional clinicopathological correlations (CPC) [43]. The traditional conception of modern AD began with such a CPC, provided in the early 20th century, by the German neuroscientist Alois Alzheimer [44]. Since his initial descriptions, the medical and research fields have primarily focused on two of his seminal neuropathological findings, the senile plaques, primarily composed of extracellular A β protein fibrils, and the intracellular neurofibrillary tangles, made up of phosphorylated tau protein species [45–47]. Not until recently, a key pathological hallmark has gained attention, i.e. the proposed “adipose inclusions” or “lipoid granules” that suggested the existence of dysregulated lipid metabolism [48,49].

Early biomarker investigations related to preclinical AD individuals featured those presenting with an autosomal dominant or familial condition (familial AD, fAD), confirmed via genetic testing and allowed the definition of associated cerebrospinal fluid (CSF), blood, and/or brain (*via* neuroimaging) abnormalities [50]. In such presymptomatic (preclinical) fAD gene mutation carriers, abnormalities in A β and tau species concentrations were confirmed compared to controls in each of the matrices, providing a time-dependent course for each protein [50,51], and suggesting different phenoconversion predictive capacity for each protein and combination analyzed. Although these preliminary biomarker investigations correlated between human fAD and certain transgenic rodent models, similar investigations of the vastly more common, late-onset polygenic form of AD (LOAD), remained incomplete. The major limitation in studying LOAD individuals was the absence of an easily attainable preclinical molecular signature that would allow accurate selection and monitoring of disease progression during the preclinical stages. Without such molecular signature for LOAD, and given the shared late neuropathological stage with fAD, a conventional partly reductionistic assumption was generated [52], hypothesizing close links between the pathophysiology of fAD and LOAD [53]. Given the lack of

a holistic understanding for the true basis and unique evolution of LOAD, therapeutic interventions based on fAD and transgenic animal model findings provided no significant evidence of clinical efficacy when tested in LOAD individuals [54]. As a result, in light of the continuous failures of late-stage clinical AD drug trials, there has been a more exploratory, integrative, and holistic reevaluation of additional factors contributing I) to AD pathophysiology, especially related to membrane damage [55], and II) in moving therapeutic interventions into the preclinical stages [56]. Both of the latter require the development of relevant biomarkers for the preclinical stages of AD, particularly targeting other pathophysiological pathways apart from the amyloidogenic one.

Neuroinflammation is such a broad pathophysiological field and has evolved by providing an etiologic explanation for brain membrane injury in AD and in a variety of neurological diseases [57–64]. As a result, the depiction of currently developed blood-based approaches needed to explore the preclinical manifestations of neuroinflammation using both direct – *via* inflammatory biomarkers – and indirect – *via* metabolomic biomarkers – measures seems to be crucial.

3.1. Inflammatory biomarkers

Even though inflammation might not classically considered an initiating factor in ND, there is emerging evidence in animal models that sustained inflammatory responses – involving microglia, the major resident immune cells in the brain, and astrocytes, glial cells with support functions – contribute to disease progression. Sustained inflammation leading to tissue pathology involves the persistence of an inflammatory stimulus or a failure in normal resolution mechanisms. A persistent stimulus may be the result of I) the presence of environmental factors and II) the formation of endogenous factors (for instance, protein aggregates) that are interpreted by the immune system as “unfamiliar” or even dangerous signals. Although some inflammatory stimuli generate positive effects for the organism, such as phagocytosis of debris and apoptotic cells, and inflammation is associated with mechanisms of tissue repair, uncontrolled/uninhibited inflammation may result in production and release of neurotoxic factors intensifying the disease states [65].

At present, the primary role of neuroinflammation in AD is unquestionable. In particular, inflammation occurs in pathologically vulnerable regions of the AD brain and it acts in this way using a plethora of local peripheral inflammatory responses. At the peripheral level, the deposition of highly insoluble abnormal materials, together with degenerating tissue, is a critical factor inducing inflammation. Similarly, at the level of the AD brain, damaged neurons and neurites as well as highly insoluble deposits of A β peptide and neurofibrillary tangles provide evident stimuli to trigger inflammation [66]. In this regard, the analysis of *post-mortem* AD brains has provided evidence for inflammatory factors and activated cell types in association with common end-stage pathophysiological features, including amyloid plaques and neurofibrillary tangles [67–69]. The primary cellular sources in the brain responsible for cytokine production are perivascular and meningeal macrophages and microglia [70] (26). Considered uniquely important to AD pathophysiology – especially in context of genetic variants of *TREM2* (encoding the triggering receptor expressed on

myeloid cells 2) gene[71,72] – microglia are known to release various soluble factors and assist in extracellular A β clearance.

Genetic factors are fully acknowledged to play a key role in AD. Notably, the search for genes involved in AD has been revolutionized by the application of GWAS, the most common approach to assess genetic variants in the genome using arrays of single nucleotide polymorphism (SNPs) to investigate the potential association with AD[73]. Interestingly, several genetic variants are involved in immune and inflammatory processes, as deeply reviewed [71,74,75]. In particular, two groups of investigators [76,77] independently identified and characterized a rare variant in the *TREM2* gene – a major microglia-specific gene in the CNS – that causes an increased susceptibility to LOAD [78].

Despite the description of the CNS components of neuroinflammation, their putative peripheral manifestations in blood have sometimes provided inconsistent results [79–84], even when comparisons between control subjects and AD patients have been performed. In particular, it has been challenging to develop informative peripheral inflammatory molecular signatures for preclinical AD. Various studies have explored biomarkers potentially associated with inflammatory processes. Cytokines – including tumor necrosis factor-alpha (TNF- α), interleukin-6 (IL-6), transforming growth factor-beta (TGF- β), and interleukin-1-beta (IL-1 β) – have been measured in CSF of AD patients, but in one *meta*-analysis the only consistent finding was the increased CSF concentrations of TGF- β in AD patients *versus* control groups [85]. TNF- α , expressed by neurons and glia, stimulates the inflammatory responses by recruiting microglia or astrocytes to lesion sites, thus leading to glial cell activation. The TNF receptor complex and its related functional proteins are supposed to be actively involved in AD pathophysiology, thus strictly associating inflammation signaling pathways with the amyloid deposition cycle in a self-propagating and destructive dynamic [86]. TNF- α binds to specific membrane glycoprotein receptors – TNF receptors (TNFRs [TNFR1 and TNFR2]) – that activate signal transduction pathways converging to a common mechanism of neuronal death. The definite function of TNFR1 as crucial mediator of inflammation, apoptosis, and amyloidogenic pathology has been scrutinized [87,88]. Remarkably, since TNFR1 and TNFR2 are both expressed and triggered differentially in AD brains *versus* non-demented brains, distinct pathophysiological mechanisms of neurodegeneration in AD brains have been proposed [89]. In addition, expression levels of TNFR1 and TNFR2 have been documented to be altered in the brains and CSF of subjects with mild cognitive impairment (MCI) and AD patients. The activity of the TNF- α converting enzyme (TACE), cleaving both pro-TNF- α and TNF receptors, is substantially increased in the CSF of AD patients and MCI subjects *versus* healthy controls. Moreover, CSF concentrations of TACE-cleaved soluble forms of TNFR1 (sTNFR1) and TNFR2 (sTNFR2) appear more elevated in AD patients *versus* healthy controls and correlated with TACE activity. Finally, greater levels of TACE activity and soluble TNFRs are present in MCI subjects *versus* AD patients, thus emphasizing an early role of TACE activity and soluble TNFRs during AD pathophysiology and a their potential usefulness as diagnostic markers in MCI and AD dementia stages [90]. In addition, the integrated CSF examination of tau protein with the constituents of the soluble IL-6 receptor complex (sIL-6RC), assumed to be a marker of neuromodulatory and brain inflammatory processes, is assumed to increase the certainty of AD detection/diagnosis [91,92]. Interestingly, peripheral blood

mononuclear cells may provide actionable longitudinal risk information [93] through increased spontaneous production of IL-1 and TNF- α associated with cognitively normal individuals with an increased risk of phenoconversion to AD. Finally, CSF YKL-40—an indicator of microglial activation – has been designated as a pathophysiological biomarker indicating immune/inflammatory mechanisms in AD and other ND, associated with tau protein pathology [64,94].

3.2. Metabolomic biomarkers

The human blood metabolome consists of thousands of small molecular species, typically less than 1500 Da (Daltons; 1.7×10^{-27} kg) in molecular weight and primarily featuring monosaccharides, acylcarnitines, biogenic amines, amino acids, fatty acids, and complex lipids. By far, lipid species make up the largest fraction (45%) of the ~50,000 metabolites currently detectable [95]. Identifiable metabolomic species, including human, pharmacologic, animal, plant, or bacterial, are currently curated in one or more of the following databases: the Human Metabolome Database (HMDB) (available at <http://www.hmdb.ca>), the METLIN database (available at <http://metlin.scripps.edu>), and the LIPID MAPS Lipidomics Gateway (available at <http://www.lipidmaps.org>). A significant number of metabolic species are yet to be annotated, with recent estimates of the total approaching 1 million [95]. Using standard reductionistic approaches, the metabolome is considered a downstream linear reflection of the genome/epigenome, transcriptome, and proteome, sequentially, and in close proximity to the clinical phenotype. Using a systems biology-based perspective, although the aforementioned might be true, complex interrelationships exist between the various “omic” layers [96], that, if properly integrated, are expected to provide an improved understanding of a complex disease state or human health.

Early metabolomic approaches to biomarker development in blood and CSF have featured either nuclear magnetic resonance-based analyses [97,98] or those utilizing mass spectrometry (MS)-based technologies[99,100]. More recent reports of metabolomic biomarkers for AD have been developed using specimens from cross-sectional investigations analyzed with MS platforms, typically comparing metabolite abundances between control subjects and individuals with either prodromal or manifest AD[101–105]. Although consensus is lacking regarding specific metabolites discovered between studies, there is a substantial preponderance for alterations in certain lipid species in blood. Analyses[106,107] from a longitudinal observational study – specifically evaluating preclinical subjects observed to phenoconvert from cognitive normality to either prodromal or manifest AD – reported significant reductions in certain plasma lipid species. Notably, reductions of some of the same species were observed in early AD subjects in an independent therapeutic trial [108].

3.3. Biomarker perspectives

3.3.1. Biomarkers as diagnostics—While there is no unanimously established consensus regarding the selection(s) of molecular biomarker panels that are most informative regarding the preclinical stages of AD, there is growing support for the use of blood-based biomarkers in helping define this crucial therapeutic window. Thanks to their classification sensitivity and specificity comparable to that provided by CSF

and neuroimaging markers, their decreased associated risk, increased patient comfort, and reduced associated cost, blood-based biomarkers are gaining *momentum* as potential screening and prediction tools and for enhanced selection, subject enrichment, and stratification of disease subsets in AD disease-modifying trials [109].

3.3.2. Biomarkers as guides to therapeutics—The ultimate objective is to develop biomarker-guided targeted therapy in AD. The potential utility of certain biomarkers as outcomes and surrogate outcomes during the preclinical stages of AD should will be a development focus. In this specific circumstance, biomarkers indicating the existence of neuroinflammatory and membrane lipid dysregulation processes may be substantially informative.

4. Cns inflammation in Alzheimer's disease stages biomarkers and therapeutic targets

The role of CNS inflammation in the development and progression of AD has been a controversial issue, more specifically whether plaque-related inflammatory and immune processes are disease-aggravating or neuroprotective [64,110–112]. The most significant advance in understanding the role of inflammation in the evolution of the AD pathophysiology is based on the observation that long-term use of non-steroidal anti-inflammatory drugs (NSAIDs) in patients with rheumatoid arthritis reduces the risk of developing AD when compared to the general population [113,114]. These findings originally supported the idea that neuroinflammation represents a key pathophysiological feature in the AD cascade, prompting the pharmaceutical industry to launch several large clinical trials on the use of classic NSAIDs, such as ibuprofen, rofecoxib, celecoxib, and R-flurbiprofen, and other anti-inflammatories, including pioglitazone, steroids, and aspirin, in symptomatic patients diagnosed with AD. Published results from these trials have been the subject of several meta-analyses, all of which have concluded that treatment with anti-inflammatories lacks efficacy in symptomatic, already clinically diagnosed AD dementia patients [115–117]. Only one trial, the Alzheimer's Disease Anti-Inflammatory Prevention Trial (ADAPT), has reported a beneficial clinical outcome when naproxen is administered before the onset of subjective cognitive impairment [118]. These findings indicate that the critical therapeutic window to target neuroinflammation is likely at preclinical AD stages [112].

Although these clinical trials have not yet produced a viable therapeutic option for the treatment or prevention of AD, they have provided insight into the dichotomous function of neuroinflammation in the progression of AD: early inflammation is likely pathogenic and disease-aggravating, whereas late inflammation appears to be dominated by tissue-resolution and phagocytic processes [112]. The idea that inflammation adopts a protective role as the disease progresses through the AD *continuum* is supported by GWAS-based analyses that have identified SNPs in the *CD33* and *TREM2* genes that are associated with an increased risk of developing AD [76,77,119–121]. Disease-relevant variants in both genes, which are primarily expressed by immune cells, result in blunted phagocytic capacity by brain macrophages, thus suggesting that clearance mechanisms likely serve to counteract

late neurodegenerative mechanisms. Inhibiting these protective inflammatory mechanisms, would theoretically exacerbate or, in the least, fail to decelerate the neurodegenerative cascade. As a result, anti-inflammatory therapy is unlikely to be disease-modifying if administered during late symptomatic stages, when the fundamental neuronal networks responsible for higher CNS functions have already been destroyed. Applying an anti-inflammatory therapy earlier in the disease process is the most promising strategy to mitigate the development of the underlying AD pathophysiology.

Recent studies in transgenic animal models of AD have revealed the presence of an early pro-inflammatory process before the development of A β plaques. For example, pre-plaque 3xTg mice exhibit increased levels of TNF- α associated with intraneuronal-A β pathophysiology in the entorhinal cortex [122]. Inhibiting TNF- α signaling prevents intraneuronal-A β accumulation and corrects pre-plaque synaptic deficits and cognitive function in the TgCRND8 and 3xTg mouse models, respectively [123,124]. The McGill-R-Thy1-APP transgenic rat model also exhibits an upregulation of pro-inflammatory molecules at the pre-plaque stage of the amyloid pathology, predominantly in neuronal cells [125,126]. Importantly, treatment with minocycline at the pre-plaque stage restores the balance of inflammatory factors and rescues cognitive deficits in a mouse model of the amyloid pathology [127,128]. Taken together, evidence in animal models suggests that early, plaque-independent inflammation contributes to the progression of the early AD-like amyloid pathology and associated cognitive deficits.

Translating these observations to the human AD pathophysiology has proven to be a major challenge. It is now understood that the underlying AD pathophysiology begins 20–30 years before the first clinical symptoms [129,130]. However, given that current technologies are not sensitive enough to detect the earliest subtle AD pathophysiological features and accompanying CNS inflammation, identifying the initial disease trajectory remains elusive. Positron emission tomography (PET)-scan technology used to measure A β plaques, tau pathology, and microglial-TSPO signaling, as well as currently-available CSF and blood biomarkers, only detect advanced AD pathophysiology with a reasonable level of certitude. The critical mass of inflammatory molecules present within the CNS during the long pre-symptomatic phase likely falls below the detection-threshold of current brain imaging techniques. In the absence of reliable early biomarkers, it is virtually unrealistic to unequivocally identify the patient population within the preclinical-AD phase that may be most amenable to anti-inflammatory therapy. It is encouraging that increased astroglial activation was observed by PET imaging 20-years before expected disease onset in patients with autosomal dominant mutations leading to fAD, suggesting that initial detection of astrogliosis may allow clinicians to flag the emergence of the asymptomatic disease phase [131]. Furthermore, a recent report indicates an association between midlife peripheral inflammation and reduction in late-life brain volume in individuals without dementia [132]. These findings suggest that early inflammatory processes could have a detrimental effect in the CNS and this might contribute to the development and progress of the pathophysiology.

It is expected that in the coming years, considerable research efforts will be focused on developing diagnostic methods able of detecting AD progression during preclinical stages, or at least early enough to substantially impact the disease with anti-inflammatory

agents, either as a single or combined therapy. Biologicals, specifically cytokine-directed monoclonal antibodies, are a particularly attractive therapeutic option given that targeting just one cytokine is often sufficient to disrupt the broader molecular cascade that culminates in chronic inflammation [133]. Several TNF- α inhibitors, including TNF- α -directed monoclonal antibodies and recombinant fusion proteins, have already been approved by the Food and Drug Administration (FDA) for the treatment of several inflammatory and auto-immune diseases, including Crohn's disease, ulcerative colitis, and rheumatoid arthritis. In one pilot study, 6-month perispinal intrathecal administration of etanercept (a decoy receptor for TNF- α) in AD patients resulted in an improvement in a variety of cognitive measures [134]; however, these results have yet to be replicated. In another trial, 6-month subcutaneous administration of etanercept in patients with mild-to-moderate AD dementia did not improve cognitive outcomes [135]. In a recent case report, infliximab, a TNF- α -directed monoclonal antibody, administered to a patient with AD led to cognitive improvement along with a decrease in AD pathophysiological biomarkers [136]. Despite these results, exploring the effects of anti-TNF- α therapy in patients with early preclinical AD are still lacking.

Besides its role as lipid sensor and involvement in metabolic pathways, peroxisome proliferator-activated receptor- γ (PPAR- γ) activation leads to the blockage of the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B)-dependent gene expression, thus inhibiting multiple inflammatory pathways [137]. PPAR- γ agonists have been shown to be beneficial when administered to AD mouse models by decreasing inflammation and AD-related pathophysiological markers [138]. Moreover, clinical trials with rosiglitazone had positive outcomes in mild-to-moderate AD patients [139,140]. The PPAR- γ agonist, pioglitazone, an approved treatment for type II diabetes, is currently under a phase 3 clinical trial being conducted in MCI individuals and AD patients (NCT02284906). Further exploration of the therapeutic effects of PPAR- γ activation is needed in early AD stages.

Inhibition of IL-1 β signaling represents another promising therapeutic option in treating preclinical AD. Currently-available IL-1 β -targeted anti-inflammatory therapies include canakinumab, an IL-1 β -directed monoclonal antibody, and anakinra, an IL-1 receptor antagonist [133,141,142]. The potential of anti-IL-1 β therapy has yet to be investigated for the treatment of early preclinical AD, either in humans or animal models. Given that IL- β processing and maturation is largely controlled by the multiprotein inflammasome complex [143–146], treatments targeting the assembly and function of the inflammasome may also lead to a reduction in IL-1 β signaling in early AD stages. Inflammasome complex activation has been reported in *post-mortem* brains of MCI individuals and AD patients as well as in AD mouse models [147–149]. Moreover, inhibiting the inflammasome in transgenic rodent models of AD leads to a reduction in AD-related pathophysiology and associated cognitive deficits [150]. Proper characterization of inflammasome activation and potential therapeutics at early stages of AD has yet to be explored.

In the absence of early biomarkers and effective therapies to diagnose and treat preclinical AD, the development of compounds targeting CD33 and TREM2 may prove effective in slowing disease progression and symptom severity in already-diagnosed patients with mild-to-moderate AD. Recent studies indicate that disease-relevant variants in *CD33* lead

to increased CD33 expression and impaired phagocytic activity of brain macrophages [151,152], whereas variants in *TREM2* leads to decreased surface cell expression or impaired functioning, also resulting in reduced macrophage phagocytosis [153,154]. The development of small compounds that either inhibit CD33 or promote TREM2 activity may represent a promising therapeutic option to promote phagocytic and clearance mechanisms within the CNS in intermediate-late AD stages. Several monoclonal antibodies targeting CD33 do in fact exist and are in development for the treatment of myeloid leukemia; however, they are currently being evaluated in clinical trials and have not been tested for the treatment of AD, either in humans or animal models [155]. Similarly, the development of a small-compound modulator enhancing TREM2 activity or prolonging its cell-surface expression may promote clearance mechanisms that are likely to be effective in decelerating late neurodegenerative mechanisms.

5. Anti-amyloid beta and anti-tau therapeutic strategies

The clearance of the A β peptide, in particular the extracellular overproduction and deposition of the 42-amino acid-long A β peptide (A β ₁₋₄₂), and the intracellular expression of tau protein, characterized by post-transcriptional phosphorylation, are recognized as critical pathophysiological mechanisms leading to AD. As a result, positivity to amyloid and tau biomarkers is mandatory to establish an effective *in vivo* diagnosis of AD [156]; for this reason, most of the currently developed disease-modifying therapies for AD are targeted on the amyloidogenic and tau pathways.

The conventional hypothesis on AD pathophysiology states that the initial neurodegenerative processes are associated with the imbalance between production and clearance of A β ₁₋₄₂ peptides resulting in cerebral accumulation of insoluble and toxic forms of aggregates of misfolded proteins [157]. In this regard, an early, fast, and efficient biomarker-guided screening of individuals during the pre-symptomatic phase might support the development of effective disease-modifying trials before the amyloid-related neurodegenerative processes become irreversible. Indeed, several longitudinal studies clearly indicated that reduced CSF A β ₁₋₄₂ concentrations combined with increased cerebral amyloid PET signal currently represent the earliest asymptomatic indicators of AD onset [129,158]. Interestingly, the early pre-symptomatic decrease of CSF A β ₁₋₄₂ concentrations or the increase of amyloid PET signal are followed by a “plateau” phase before the individuals become symptomatic (i.e., MCI and dementia stages of AD). Hence, starting an anti-amyloid trial during the dementia or even the MCI stages is predestined to fail. This is supposed to be the key reason accounting for the failure of over 100 anti-amyloid monotherapeutic trials conducted to demonstrate a benefit in slowing the progression of cognitive impairment [159]. Pharmacological anti-amyloid strategy essentially relies on modifying the dynamic balance among A β monomers, A β oligomers, and fibrils, being A β oligomers the most toxic species [160].

Modulation of A β secretase enzymes aims at I) increasing α -secretase activity by converting APP into harmless sub-metabolites as well as II) inhibiting β - and γ -secretases activity, to halt the amyloidogenic pathophysiological pathway [161,162]. However, caution should be taken when drug interventions target γ -secretase activity since this enzyme is involved

in various key physiological signaling pathways of proteins modulating cellular trafficking, apoptosis, cholesterol homeostasis, neurogenesis, and angiogenesis [163]. In particular, the amyloid precursor protein (APP) proteolytic processing generates several truncated forms of A β which have intrinsic properties providing an essential role for physiological cellular signaling mostly involved in synaptic activity-dependent modulation. Endogenous A β monomers show a potential role in the regulation of synaptic vesicles trafficking, thus finally acting as a key electrophysiological modulators of the synaptic firing [164]. Moreover, it has been reported that some A β fragments can initiate CREB-mediated cytoprotective pathways [165]. Therefore, an excessive removal of some A β monomers by a poorly calibrated pharmacological intervention may prevent hippocampal neurons from surviving aberrant pathways upstream to A β deposition.

Interestingly, an alternative option is represented by interfering with APP expression, as previously suggested in AD trials reporting the use of antidiabetic PPAR- γ agonists, including the thiazolidinediones [166]. Another potentially relevant anti-amyloid strategy is stimulating the clearance of amyloid species in the brain by increasing the activity of different proteases including angiotensin and endothelin converting enzymes, insulin degrading enzyme, metalloprotease-9, neprilysin, and plasmin [167]. The aim is to degrade amyloid metabolism byproducts by hindering their oligomerization and aggregation. Interestingly, the concentrations of amyloid degrading enzymes decrease in AD, thus possibly facilitating the deposition of toxic A β peptides. However, the modulation of amyloid proteases activity needs further assessment since it may appear as a non-specific and detrimental strategy [168]. In addition, acting on the amyloid transport modulation represents another potential approach. In particular, the multi-ligand receptor for advanced glycation end products (RAGE) efficiently binds A β in the blood and promotes its entry into the CNS through the blood brain barrier [2,161]. Finally, the apolipoprotein E, binding the A β peptides, allows their entry in the CSF circulation by the lipoprotein receptor related protein-1 and the very-low density lipoprotein receptor mediated transport [169–171].

Currently, anti-amyloid immunotherapies represent the most precisely targeted anti-amyloid treatments. The suggested therapeutic mechanism is that anti-amyloid antibodies may promote the removal of A β peptides and A β aggregates from the CNS to blood *via* a sort of “peripheral sink” [172]. Passive immunotherapy – based on the intravenous injection of anti-amyloid targeted antibodies – may induce a dosage-dependent increase of both blood and CSF A β concentrations. The use of anti-amyloid active immunotherapies has been recently proposed to design next-generation vaccines against small epitopes, instead of developing full length peptides that may generate harmful non-specific immune responses [173]. Unfortunately, the most recently published phase III trials using intravenous immunoglobulin in AD patients did not provide any clinically relevant benefit, in spite of the promising results obtained in preliminary studies [166,174]. Actually, the exact mechanisms of action of these therapies and the origin of their most common side effects, such as cerebral microbleeds, is still unexplored [175]. The ultimate response of an existing proof-of-concept in the anti-amyloid treatment strategies might come from the results of ongoing trials recruiting exclusively AD patients carrying presenilin mutations [176].

Although there is a general consensus that A β accumulation represents the initial trigger of AD pathophysiology, the continuous failures of disease-modifying anti-amyloid phase III trials encouraged the design of anti-tau therapeutic strategies. Notably, a robust correlation of tau brain pathology with the severity of the cognitive impairment in AD was reported in several longitudinal studies, thus supporting the interest on anti-tau therapies [129,177,178].

Tau is a microtubule-associated protein involved in axonal stability. It is hyperphosphorylated, separated from microtubules, and then accumulated as a misfolded protein within neurons, in tau-associated neurodegenerative diseases, including AD [179]. Tau targeted treatments can be specifically directed to the phosphorylation process, resulting in the disassembly of microtubules and, consequently, into reduced microtubule stability [180]. One strategy involves acting on the various post-transcriptional modifications monitoring tau intracellular physiological activity [179]. For instance, the up-regulation of tau O-linked glycosylation seems to decrease tau oligomerization process and leads to the deposition of toxic insoluble fibrils. Moreover, the inhibition of tau acetylation may promote tau clearance *via* the ubiquitin/proteasome system (UPS) [181]. Notably, the stimulation of the intracellular autophagy/lysosomal system may represent a way to eliminate the deposition of misfolded tau proteins in the advanced AD stages. Another possible approach includes the down-regulation of tau proteolysis mediated by distinct subtypes of cysteine proteases, namely caspases, calpains, and cathepsins [179,182]. The potential development of active or passive tau immunotherapies appears controversial, given that tau and its toxic brain inclusions have an intracellular position obstructing the tau sinking process mediated by specific anti-tau antibodies [183]. In summary, the development of tau targeted therapies is still in its infancy and, therefore, a further assessment of tau-associated pathophysiological mechanisms (also linked to other neurodegenerative diseases) is mandatory.

Notably, a novel unexplored field in the development of AD therapeutics is to investigate the relationship between the CNS – including the macromolecule circulation and removal within the glymphatic system – and the periphery [21]. The latter is involved in the clearance of potentially harmful protein byproducts, produced in the brain, that are involved in the pathogenesis of AD and other neurodegenerative diseases. In this regard, emerging data revealed that brain pathophysiological processes are reflected into the periphery; moreover, some CSF biomarkers such as neurofilament light chain [184,185], tau [185], and β -site amyloid precursor protein cleaving enzyme (BACE1) enzyme [86] are reliable blood surrogate proxies of underlying cerebral neurodegenerative mechanisms [86,186–188]. In addition, the A β species generated in the CNS can cross the brain blood barrier and be removed by peripheral organs. Actually, systemic diseases may interfere with amyloid clearance, thus contributing to AD development and progression [157]. In this regard, kidney dysfunction might be associated with the amyloidogenic pathophysiological mechanisms leading to AD and is responsible for an increased risk of cognitive/psychiatric alterations and dementia [189]. Interestingly, renal transplantation is assumed to decrease plasma A β concentrations and hemodialytic procedures reduce brain A β accumulation in subjects suffering from chronic kidney disease [190]. Another interesting observation is that A β load in liver tissue is decreased in AD *versus* healthy subjects, hence suggesting the existence of a hepatic A β -clearance dysfunction in AD [191]. Notably, epidemiological bidirectional association is evident between diabetes, pure cerebrovascular

cognitive impairment, neurodegenerative diseases such as AD, and mixed forms of dementia [192–198]. In particular, recent studies indicate that A β may have detrimental peripheral effects resulting in its atypical accumulation in pancreatic cells [191,199]. In addition, attention should be given to the impaired N-terminal processing of amylin precursor, also called islet amyloid polypeptide (IAPP), an early factor inducing the toxic accumulation and deposition of amyloid in pancreatic β -cells [200].

In conclusion, there is emerging evidence for a bidirectional interplay between brain and peripheral organs in regulating A β metabolism and other protein byproducts associated with neurodegenerative pathways. This emphasizes the need for a comprehensive and precise strategy – directed on both CNS and peripheral dysfunctions – based on the systems biology and systems neurophysiology paradigms [157,201]. The traditional “one-drug-fits-all” concept seems to be obsolete and does not reflect the heterogeneity and complexity of neurodegenerative diseases, including potential therapeutic interventions combining precise multi-target drug administrations with lifestyle changes such as diet modifications [202,203] as well as specific and “tailored” cognitive training [204]. These belong to a *spectrum* of diseases caused by the deposition of multiple misfolded proteins and cerebrovascular damage, and are unavoidably affected by systemic diseases [205]. A more flexible and adaptive multi-target strategy, taking into account the complexity of AD pathophysiology, is needed in the upcoming drug development programs.

6. Rethinking and optimizing the design of clinical trials from the precision medicine perspective

PM demands precision drug development. One cannot apply PM concepts of the right drug, in the right dose, for the right patient, without these aspects of drug treatment having been thoroughly tested in clinical trials. Although daunting, the PM approach may be precisely what is needed to resolve the current challenges facing AD drug development. No new treatments have been approved for AD since 2003 and the field has a drug development failure rate in excess of 99% [54]. All drug development programs aimed at developing disease-modifying treatments for AD or any other neurodegenerative disorder have failed [206]. PM offers a means of conceptualizing a resolution to this crisis.

The basic tenet of PM is that humans are biologically heterogeneous and that these differences express themselves in differences in the characteristics of the disease they develop, the stage and rate of progression of the disorder, and the dose needed to abrogate progression or restore function [3,4]. Conduct of clinical trials to meet the demands of PM will require different recruitment approaches, biomarker characterization of participants, dosing strategies, and data analytic approaches. A fundamental need is to better comprehend the basic biology of AD, the druggable aspects of the pathology, the heterogeneity of the disease, and the biomarkers that reveal this heterogeneity to the clinician. These are the building blocks on which precision trials and PM will be built.

6.1. The right drug

The right drug in the PM schema addresses the basic biology of AD. This requires an understanding of the heterogeneous pathology of AD and how it can be addressed pharmacologically. In a recent autopsy study of patients clinically diagnosed with AD and meeting pathologic criteria for AD, 32% of patients had AD pathology only while 68% had combinations of AD pathology, alpha-synuclein/Lewy pathology, and ischemic injury secondary to cerebrovascular disease [207]. In addition to the vascular and degenerative changes, the brains of AD patients exhibit inflammatory, oxidative, mitochondrial, transactive response DNA-binding protein 43 (TDP-43), heavy metal, and epigenetic changes that may contribute to the disease pathophysiological processes and offer opportunities of intervention [208]. The “right drug” for AD will likely be a combination regimen of agents addressing multiple types of pathology. The “right drug” may also evolve over time as the process evolves, changes become more advanced, and new elements participate in the pathophysiological cascade of AD. Clinical trials constructed around PM approach will use biomarkers (discussed below) to link the right drug/combination to the right patient.

6.2. The right dose

Dose exploration is a critical aspect of drug development and clinical trials for AD. Phase 1 should include identification of a maximum tolerated dose (MTD). Without knowledge of the MTD, later efficacy failures will inevitably raise the question of dose adequacy. In some cases, a MTD can be defined by PET occupancy studies showing target saturation thus implying that increased doses will not produce greater effects. Physical limitations, including solubility and acceptable rate of infusion, impose a maximum plausible dose for some agents. In all other cases, a MTD should be established and formulation issues should be solved prior to the Phase 1 trial if they may artificially limit the ability to define a MTD.

Dose-response characteristics will also be established in precision clinical trials. Doses that are too low to produce benefit, near the upper limit of tolerability and in the optimal range (minimal 3 doses), should be studied in early phase trials. Adaptive designs may facilitate the elimination of ineffective or toxic doses [209]. Individualization of doses, as required for PM, can be advanced through knowledge of the drug metabolism patterns of the individual, including fast and slow metabolizers and toxic response. In this regard, pharmacogenomics will play a critical role in precision trials and in the PM paradigm [210]. Pharmacogenomics can be broadly defined as the use of genomic and other “omic” information to individualize drug selection, optimize drug efficacy, and reduce adverse drug reactions. In this context, pharmacogenomic information relies on biological markers that label individuals as more or less responsive to specific medications and/or more or less susceptible to experience adverse effects. Moreover, pharmacogenomics can determine treatment response based on disease-causing variants of heterogeneous clinical conditions. Ultimately, pharmacogenomics is expected to remove the traditional “one-size-fits-all” clinical trial methodology in developing and prescribing therapeutic drugs. Hopefully, PM research and interventions will avoid this “trial and error” approach and predict who will respond to a medication and who – in turn – should avoid the same medication. Research in pharmacogenomics is also expected to provide critical information about the genomic

variations that affect response to currently recommended pharmacological treatments and future interventions. Understanding the individual variation and the implications for drug response, metabolism, and drug elimination will allow PM physicians to implement healthcare based on the individual's "omic" biomarker data.

6.3. The right patient

Different therapeutic regimens will likely be required for individuals in different phases of AD. Individuals in preclinical and prodromal AD as well as in mild, moderate, and severe forms of AD exhibit different phenotypes and different underlying "pathologies" that need to be addressed using different drugs/combinations of drugs. Cognitive enhancers are indicated in individuals with cognitive symptoms and psychotropics are indicated in those with neuropsychiatric symptoms. Different drugs/combinations of drugs will be required for those with simple *versus* complex pathology (Fig. 2) and this may evolve as the disease progresses.

The right drug will require use of biomarkers in clinical trials. Biomarkers will define the patient population for which a given therapy or combination is indicated and will link the basic pathophysiology of AD to the proper therapy. Biomarkers of alpha-synuclein, TDP-43, vascular pathology inflammation, and other CNS changes are needed to allow both the trialist and the clinician to construct treatment regimens reflective of the pathology of the individual patient. Experience in the trial will anticipate the needs of the clinician and biomarkers used in trials may evolve to companion biomarkers approved in concert with new therapies and informing their use.

Precision prevention is required for primary prevention of AD in individuals without state or trait biomarkers of the disease or for secondary prevention of those with genetic factors (presenilin-1 [*PSEN-1*], presenilin-2 [*PSEN-2*], amyloid precursor protein [*APP*] mutations or apolipoprotein E [*APOE*] homozygous state) or state biomarkers (positive amyloid PET or CSF signature of AD) indicative of impending AD. Primary prevention trials will focus on life style interventions constructed to match the genomic profile of the individual including risk for diabetes or hypercholesterolemia and other AD risk factors. Primary prevention will involve amyloid prevention agents such as BACE inhibitors. Secondary prevention trials will include lifestyle factors in combination with agents related to tau progression, inflammation, mitochondrial function, and other biological factors. Thus, precision prevention will lead eventually to "precision health".

6.4. Conduct of precision medicine trials for Alzheimer's disease

Precision trials will be structured differently from those currently conducted [211]: substantially extended biological characterization of the population using biomarkers will be required. Biological profiles will be matched to treatment/treatment combinations. Severity may represent an important parameter in selecting the right drug for the right patient. Precise matching of some AD populations to evolving therapies may allow these agents to be developed as orphan drugs for rare diseases [212]. PM trials will be more patient-centric and biomarker-guided than currently conducted trials. Large populations of well-characterized individuals will be required to allow precision trials to be performed. This will require novel

innovative strategies such as mass advertising, enlistment of large populations (e.g., those applying for retirement benefits), on-line testing, mass biomarker collection at convenient locations (such as shopping malls), development of large databases of trial-ready persons, and testing using remote assessments and virtual visits in tele-trials.

Trial analytic strategies will need to evolve to accommodate to PM requirements. PM and precision clinical trial outcomes will address individual responses in more detail. Current analytic approaches provide group data, however, little information about individual responders or non-responders is delivered. Robust analytic techniques applicable to individuals/small groups of individuals will be necessary. This might involve more dependence on well validated biomarkers than previously. Table 3 summarizes how clinical trials will be constructed in the age of PM. The time is now: PM requires precision trials and we should be pursuing these trials now. The trends are already evident with definition of different disease phases and evolution of new biomarkers. These trends need to be validated and accelerated as well as married to aggressive trial methodologies.

7. How can drug discovery programs in Alzheimer's disease accomplish a good level of translational quality to reduce the rate of failures?

7.1. Drug discovery translational for Alzheimer's disease therapeutics

In terms of drug discovery, translation is the process by which non-clinical research is performed that will give insights into the likely behavior of a therapeutic intervention in the individuals. The lack of success in demonstrating efficacy in AD patients of a very wide range of approaches may indicate that translational science is woefully inadequate in the field [54,213]. However, a more considered appraisal suggests a range of reasons for failure that can be grouped into four main – and sometimes overlapping – categories: I) inadequate drug discovery process; II) inadequate target engagement to test the therapeutic hypothesis; III) changing the therapeutic hypothesis to accommodate the compound properties; IV) acceptance of the “null hypothesis.”

7.2. Inadequate drug discovery process

In many, but not all, drug discovery programs, work is conducted as reported in Fig. 3. While this is shown as a linear process for simplicity, often steps may be omitted, feedback loops are common, and ideally human data on PK and PD properties of the therapeutic can be fed back to the non-clinical phase to inform the drug discovery program. Moving from *in vitro* to *in vivo* assays, the complexity of the assays increases, as does their relevance to human disease. While an element of this stepwise approach is to filter the large number of test compounds that may need to be synthesized, screened, and assayed in order to find those showing the appropriate potency and selectivity, there is also significant translational value in each step. Notably, biomarkers qualified for use in clinical trials to facilitate marketing authorization and regulatory decision-making should also be available as diagnostic agents. Thus, each biomarker will be useful at one stage or another stage of medical product development, i.e., from discovery to adoption in clinical practice (Fig. 4).

It is a mistake to assume that the *in vivo* model to human transition is the only important translational step: confidence in the therapeutic approach is built throughout the drug discovery process. For instance, if the potency of the potential therapeutic on the isolated target is very significantly reduced (e.g., by >30 fold) when the therapeutic is tested *versus* intact cells, the finding should be investigated and resolved. This may mean that the compound fails to enter cells if the target is intracellular, or that the cell response measured does not solely reflect target activity, and so on. Normally, the pattern of activity of multiple compounds enables trends to be delineated and understood: this is referred to as the structure-activity relationship. If the free drug level (unbound to matrix and, therefore, available to interact with the target) required for a therapeutic response in the target tissue – in this case the brain – is very different from that required for activity in cell culture assays, then this discrepancy needs to be investigated before further advance is considered.

The development and subsequent failure of tarenflurbil reveals several opportunities for significant improvement in the drug discovery process and translation into clinical studies. Tarenflurbil is the R-diastereoisomer of the racemate flurbiprofen, a non-steroidal anti-inflammatory drug approved for human use. The original preclinical data on tarenflurbil showed a dose related decrease in A β ₁₋₄₂ production from human embryonic kidney 293 (HEK293) cell line stably transfected with human Swedish mutant APP but with an incomplete dose response curve: the half maximal effective concentration (EC50) being in excess of 250 μ M [214]. Additional studies also demonstrated a reduction in A β ₁₋₄₂ production from H4 neuroglioma cells expressing Swedish mutant APP695NL but, again, the dose response was incomplete with an EC50 in excess of 250 μ M [215]. In the same study, tarenflurbil was administered to Tg2576 Swedish APP transgenic mice, for three days, at three doses: 50, 25, and 10 mg/kg od. All doses reduced A β ₁₋₄₂ in brain by a maximum of ~60% but in a non-dose related manner. The group sizes used were small (N = 4-7) and without evidence of a power calculation being employed to guide robust statistical analysis. Crucially, at the top dose of 50 mg/kg, the brain concentration of tarenflurbil was 2.5 μ M, a dose more than 100-fold lower than the EC50 in cell culture studies. This significant discrepancy should have been investigated further. In fact, subsequent studies failed to replicate the *in vivo* A β ₁₋₄₂ lowering effects of tarenflurbil.

In phase 1 human studies, the highest dose of tarenflurbil administered, 800 mg bid, produced a CSF concentration of approximately 1.2 μ M: some 200-fold lower than the EC50 concentration and without any effect on CSF A β ₁₋₄₂ concentrations [216]. After the phase 2 study [217] in which target engagement was not assessed and CSF A β metabolites were likewise not measured, a phase 3 trial enrolled 1646 mild AD patients in a randomized, double-blind, multisite, placebo controlled trial. Tarenflurbil was administered at 800 mg/kg bid in the active treatment arm for 18 months. The trial failed to meet its coprimary outcome measures of Alzheimer's Disease Assessment Scale-Cognitive Subscale (ADAS-Cog) and Alzheimer's Disease Cooperative Study Activities of Daily Living (ADCS-ADL) [218], as did a companion phase 3 trial that was discontinued early. The development of tarenflurbil clearly demonstrates an inadequate translational process during the *in vitro* to *in vivo* phase, coupled then to a likely Type 1 error in *in vivo* efficacy studies that was incorrectly used to support the clinical program.

7.3. Inadequate target engagement to test the therapeutic hypothesis

A β is released from the APP holoprotein by the sequential action of BACE and γ -secretase: inhibition of either enzyme is able to reduce the production of the A β peptide. Inhibitors of both enzymes have been tested in phase 3 clinical trials in AD patients. Semagacestat is a γ -secretase non-competitive inhibitor, binding at an allosteric site and with a complex mechanism of action [219–221].

Semagacestat inhibited A β peptide production in HEK293 cells stably expressing Swedish APPNL with an EC₅₀ of 15 nM [222]. In PDAPP transgenic mice that overexpress the hAPP717 mutant protein, dose related inhibition of brain A β production was demonstrated both acutely, and after 7 days' dosing [223]. In a 5-month chronic study, semagacestat was able to lower insoluble A β concentrations in a dose-related manner at 3, 10, and 30 mg/kg od [224]. Since this study did not incorporate a baseline group (analyzed at the commencement of dosing), it was not possible to determine whether semagacestat delayed the onset of amyloid deposition or reduced the rate of amyloid deposition, which is critically important with respect to the compound's use in AD patients [213]. In this mouse study, the 30 mg/kg dose reduced plasma A β concentrations by approximately 60%.

In a phase 1 study in humans, doses of 60, 100, and 140 mg semagacestat were administered with the peak plasma reduction in A β being ~50% at the 60 mg dose and 73% at the 140 mg dose. In this sense, there was evidence for an acceptable nonclinical to clinical translation. However, CSF samples taken 4 h after dosing in humans did not reveal a reduction of A β peptide [225]. To investigate this further, the effects of semagacestat on A β production were assessed in humans using the stable isotope labelling kinetics (SILK) protocol, which measures the production and clearance of newly synthesized proteins [226]. The oral administration of semagacestat at a single 100, 140, and 280 mg dose was able to inhibit brain A β production by 47%, 52%, and 84%, respectively, over a 12 h period, thus confirming semagacestat target engagement [227].

Subsequently, semagacestat was tested at 100 mg and 140 mg in two, Phase 3 trials – Identity 1 and Identity 2 – enrolling 2600 mild-to-moderate AD patients in 76 week, placebo-controlled, double blind, randomized, multi-site trials with ADAS-Cog and ADCS-ADL as co-primary outcome measures. Both trials, however, were discontinued following an interim analysis of Identity 1 that revealed a significant worsening of the Clinical Dementia Rating Scale Sum of Boxes (CDR-SB) and the Mini-Mental State Examination (MMSE) scores, together with an increased incidence of skin cancer as well as other adverse events [228].

There has been much discussion about the extent and time duration of A β production inhibition at the top 140 mg semagacestat dose [229]. While the plasma half-life of semagacestat is only 2.5 h, there was evidence that the PD effect of the compound exceeded this value in the brain [230]. It is likely that the adverse events, and most probably the worsening of cognition, were caused by an inhibition of γ -secretase mediated notch cleavage and, potentially, other substrates as well. It is very clear, however, that – irrespective of the unfavorable side effect profile of semagacestat – the extent of A β inhibition at the top dose of 140 mg was unlikely to have produced an inhibition higher than 25% over a 24 h period, constrained as it was by a combination of short compound half-life

and dose limitation due to preclinical toxicology findings. Thus, the potential efficacy to be derived by robust suppression of A β production was not tested because of inadequate target engagement.

7.4. Therapeutic hypothesis is changed to accommodate the compound properties

Solanezumab is a humanized IgG1 antibody derived from m266, a mouse monoclonal antibody that recognizes the mid-domain region (aa16-22) of the A β peptide with picomolar (10^{-12}) affinity [231,232]. Nonclinical *in vitro* and *in vivo* studies demonstrated that m266 was able to complex with A β so as to deplete A β from plasma. Experiments in the PDAPP transgenic mouse model demonstrated that the peripheral A β compartment was in communication with A β deposited in the brain when m266 was administered, in such a way that the amount of A β complexed by m266 in the plasma correlated with the amount of A β deposited in the hippocampus [233]. This finding gave rise to the therapeutic rationale for solanezumab for AD – a “peripheral sink hypothesis” – where capturing A β in the periphery would alter the soluble to insoluble A β equilibrium in the brain thus leading to the dissolution of amyloid plaque [232]. Nonclinical evidence for this hypothesis was, however, rather weak: in fact, it has not been demonstrated that m266 actually cleared amyloid plaque if administered after the beginning of plaque deposition [234]. In addition, reducing peripheral A β peptide to undetectable concentrations in plasma of mice using a neprilysin Fc fusion protein showed no effect on brain A β levels in wild-type mice. Performing a similar experiment in APP23 transgenic mice with existing plaque likewise was unable to reduce deposited insoluble A β levels in the brain or soluble A β concentrations in the CSF [235].

During the development of solanezumab, phase 1 clinical studies established that peripheral plasma A β increased with dose, as expected if A β was being complexed by the antibody and thereby assuming the half-life of the antibody, approximately 30 days [236]. In phase 2 studies, CSF concentrations of total A β_{1-40} peptide (i.e., antibody-bound plus unbound) increased, driven by the very small percentage of solanezumab that entered the central compartment: unbound concentrations of A β_{1-40} decreased. Total concentrations of A β_{1-42} peptide (antibody-bound plus unbound) also increased in the CSF, although unbound concentrations increased: this was taken as evidence of some mobilization of plaque A β_{1-42} [237]. In two, randomized, multisite, blinded, placebo-controlled phase 3 trials – Expedition and Expedition 2 – mild-to-moderate AD patients were administered 400 mg solanezumab by i.v. infusion every 4 weeks for 80 weeks in the active treatment arm. Expedition failed its primary outcome measures of change in ADAS-cog11 and the ADCS-ADL scale from baseline to week 80. On the basis of secondary analyses performed on Expedition, the primary outcome measure for Expedition 2 was changed to ADAS-Cog14 in mild AD patients: Expedition 2 failed this single outcome measure [238]. In secondary analyses of the two trials combined, solanezumab treatment had no effect on the concentrations of unbound CSF A β_{1-42} compared with placebo, unlike that observed in phase 2 studies [239]. More importantly, the treatment had no effect on brain amyloid as measured in a subset of patients using florbetapir PET imaging. A pooled analysis of data from Expedition and Expedition 2 including only the mild AD subset and the ADAS-cog14 as the main outcome showed some evidence for a therapeutic effect of solanezumab. This prompted a previously unplanned,

very large, third phase 3 trial, Expedition 3, in mild AD patients and with ADAS-Cog14 as the single primary outcome measure.

The dosing for Expedition 3 was the same as in Expedition and Expedition 2, i.v. infusion at 400 mg, every 4 weeks, in the active treatment arm in an 80 week trial. Thus, at some point during the clinical development phase, the concept of the “peripheral sink hypothesis” driving plaque resolution was replaced, presumably, by the hypothesis that therapeutic benefit would be mediated, in some way, by penetration of solanezumab into the central compartment and complexing free A β . Expedition 3 failed to meet its primary and secondary outcome measures: solanezumab was also shown to fail to reduce brain amyloid in a subset of patients who were assessed using florbetapir PET imaging.

The clinical development of solanezumab continues, however, as the Anti-Amyloid Treatment in Asymptomatic Alzheimer’s Disease (A4) trial ([ClinicalTrials.gov Identifier: NCT02008357](https://clinicaltrials.gov/ct2/show/study/NCT02008357)) will now test solanezumab at a dose of 1600 mg, every 4 weeks, for 240 weeks in cognitively normal individuals with evidence of brain amyloid pathology measured using florbetapir PET imaging. The primary outcome measure is the change from baseline of the Alzheimer’s Disease Cooperative Study-Preclinical Alzheimer Cognitive Composite (ADCS-PACC) to week 240. The rationale for quadrupling the dose of solanezumab from that used in the Expedition trials and lengthening the trial is, presumably, that the trend for an amelioration of cognitive impairment observed in previous phase 3 trials will reach a clinically meaningful level in a treated population that is at the earliest stages of the evolution AD pathophysiology. There are no preclinical or clinical data to support this rationale with respect to solanezumab. Thus, solanezumab’s “peripheral sink hypothesis” has clearly been disproven and the current therapeutic hypothesis for solanezumab remains unclear. One can argue that if a therapeutic benefit is ultimately shown for a drug, then the absence of a therapeutic hypothesis is somewhat irrelevant (although, in this eventuality, subsequent therapeutic approaches based on the clinical success would be difficult to enact). The clear danger of this strategy, however, is the risk of chasing a “chimera”, coupled to an uninformative clinical experiment should the trial fail.

7.5. What can we do better?

The issues to be surmounted in order to discover and develop a disease-modifying therapy for AD are clearly challenging. There are many lessons to be learned from prior studies:

1. Ensure that the nonclinical efficacy experiments mirror as far as is possible the clinical situation.
 - a. In this regard, according to the British statistician George Box, “the most that can be expected from any model is that it can supply a useful approximation to reality: all models are wrong; some models are useful.” Therefore, it is important to be aware of the differences between the *in vivo* model utilized to demonstrate therapeutic efficacy and the human disease. In particular, models can be assessed in terms of their “face”, “construct”, and “predictive” validity:

- a. **“Face validity”**: are there elements of the model that resemble the gross appearance/presentation of the human disease?
 - b. **“Construct validity”**: are there fundamental elements of model construction that are shared between the animal and human disease?
 - c. **“Predictive validity”**: do results that are derived from the model replicate in human disease?
- b. It is crucial to be rigorous in avoiding inappropriate validity assignment. Hence, an APP transgenic mouse model that over-expresses a mutant form of hAPP may well have A β amyloid plaque deposition that bears very great similarity to amyloid plaques in AD and also demonstrates learning and memory impairment. While the face and construct validity for plaque deposition is quite robust, it is absent for cognitive impairment, as such mice very often do not have neuronal loss or tau pathology that well correlate with cognitive impairment in AD, when amyloid deposition does not.
 - c. All AD therapeutics that have completed their clinical testing have been administered to patients with existing disease pathophysiology (notwithstanding patients misdiagnosed with AD). Thus, for therapeutics that might, for example, aim to slow or limit the progression of tau pathology, nonclinical experiments should be conducted in *in vivo* model systems following a therapeutic – dosing commenced after tau pathology onset – rather than preventative – dosing commenced before tau pathology onset – protocol.
 - d. Concentrations of the therapeutic required for efficacy and or evidence of pharmacological action should not significantly differ from nonclinical assay systems through to clinical testing.
 - e. c. If possible, to discover translational biomarkers, i.e., to identify changes that can be measured as a consequence of target engagement in the nonclinical efficacy or pharmacology model, that can be measured in humans that will provide confidence that the therapeutic hypothesis will be adequately tested.
- 2. Be clear on the therapeutic hypothesis and ensure that the clinical phase 3 trial will be sufficiently informative to accept the “null hypothesis”.
 - a. For instance, if the therapeutic hypothesis is that lowering A β production will provide clinical benefit, then robustly interrogate, plan for, and provide evidence for I) the extent of A β suppression will be required and II) why.
 - b. If it is impossible to replicate the conditions of the nonclinical efficacy data in man, e.g. because of adverse reactions to the therapeutic, then

endpoints are needed to determine a successful combination of drug and population and to reliably confirm truly predictive biomarkers.

Notably, the Center for Drug Evaluation and Research (CDER) at the Food and Drug Administration (FDA) (CDER/FDA) will soon release a draft guidance on clinical AD development, encouraging studies in the pre-symptomatic phase [J. Woodcock, personal communication].¹

Several biomarkers are used in AD for enrichment in clinical studies to define a restricted subpopulation that is expected to profit from treatment. For instance, amyloid PET and CSF A β ₁₋₄₂ are expected to be useful as predictive biomarkers. Although strongly correlated, both are measuring different aspects of amyloid pathology, fibrillar aggregates of A β for PET and soluble A β ₁₋₄₂ monomer concentrations for CSF A β ₁₋₄₂. Whereas both are considered acceptable for enrichment, the type of assay and a cut-off needs to be defined and justified [240]. In addition to CSF A β ₁₋₄₂, total tau (t-tau) or phospho tau (p-tau) concentrations are considered useful, since the A β ₁₋₄₂/tau ratio was found to have a higher positive predictive value than A β ₁₋₄₂ alone [158,241].

Downstream topographical markers of brain regional structural and metabolic changes – e.g. hippocampal atrophy assessed by magnetic resonance imaging (MRI) and cortical hypometabolism assessed by ¹⁸F-2-fluoro-2-deoxy-D-glucose PET (¹⁸F-FDG-PET) – while having insufficient pathological specificity were found to be better related to cognitive decline than A β itself and may be particularly valuable for detection and quantification of disease progression. Consequently, the combined use of amyloid and more downstream topographical biomarkers is expected to be more informative [242,243].

Novel biomarkers are currently investigated and may increase the utility of further stratification, e.g. tau PET imaging, biomarkers for neuroinflammation, blood or metabolic signatures [10,106,107,244,245]. Apart from that, epigenomics play an important role: for instance, *APOE* *ϵ 4* status may be used as one of the means of enrichment. Indeed, *APOE* *ϵ 4* homozygotes constitute 2–3% of the general Caucasian population and have a particularly high risk of developing symptoms of LOAD (although there seem to be substantial sex-risk differences and the presence of protected *APOE* *ϵ 4* homozygotes indicates complex individual genetic risk and protection patterns), especially in the presence of AD pathophysiology.

Pathway-related biomarkers should be identified in early development to reliably identify patients groups eligible for specific treatments. Whereas the predictivity of biomarkers expressed in terms of treatment-by-subgroup or treatment-by-biomarker interaction is usually suggested by drug action, further investigations in early phase clinical studies (possibly in surrogate endpoints) would be required to confirm the utility of the biomarker-related selection, but studies to investigate these interactions in hard clinical outcomes appear unrealistic with respect to size and duration.

¹This reflects the opinion of the author and does not necessarily reflect the position of the Food and Drug Administration.

In case of simultaneously studied diseases for the same drug, questions of how to deal with the multiplicity issue in confirmatory trials and whether and how information can be borrowed from one sub-study to the other arise. The corresponding statistical modeling usually requires additional assumptions that have to be agreed upon. However, even though these studies are considered to be explorative, they should certainly be efficient and informative enough to be justified, especially if long-term outcome is to be measured.

The precision of stratification has greatly improved in recent years, and patient treatment has significantly changed wherever the stratified medicine model has been introduced. This is due to substantial progresses in understanding the molecular basis of the disease, aided particularly by the advent of the genomic era and by the development of targeted therapies to address these new molecular targets. The introduction and refinement of key technologies has allowed these advances, through the increasingly detailed examination of the role of genes, RNAs/miRNAs, proteins/peptides, and metabolites/lipids in disease. These relevant technologies, which are set to further progress, include genomics/epigenomics, transcriptomics, proteomics/peptidomics, metabolomics/lipidomics investigations [201,246] and digital pathology analyses on clinical samples, clinical imaging studies, as well as biomedical and health informatics [247–249]. Standardized protocols for collecting and recording both types of data will be needed to allow comparing and combining samples and datasets, which is required to perform the large-sample-size research that will advance the molecular understanding of the disease. Moreover, recommendations have been recently released by the Academy of Medical Sciences (AMS) (available at <https://acmedsci.ac.uk/viewFile/51e915f9f09fb.pdf>) to safeguard the continuous development and adoption of stratified medicine products.

In essence, both the exploration and the confirmation of stratified medicine to be used in biomarker-defined subgroups requires a precise understanding of the underlying pathways, considerable amount of comparative data, efficient designs, and challenging integrative statistical modeling (integrative disease modeling, IDM), but also a well-founded appreciation of the remaining uncertainties and the likelihood of false decisions.

Acknowledgements

HH is supported by the AXA Research Fund, the “*Fondation partenariale Sorbonne Université*” and the “*Fondation pour la Recherche sur Alzheimer*”, Paris, France. Ce travail a bénéficié d’une aide de l’Etat “*Investissements d’avenir*” ANR-10-IAIHU-06. The research leading to these results has received funding from the program “*Investissements d’avenir*” ANR-10-IAIHU-06 (Agence Nationale de la Recherche-10-IA Agence Institut Hospitalo-Universitaire-6).

AV is supported by Rotary Club Livorno “Mascagni”/The Rotary Foundation (Global Grant No. GG1758249).

GP is supported by The Semmes Foundation.

Declarations of interest

HH serves as Senior Associate Editor for the Journal Alzheimer’s & Dementia; he received lecture fees from Biogen and Roche, research grants from Pfizer, Avid, and MSD Avenir (paid to the institution), travel funding from Axovant, Eli Lilly and company, Takeda and Zinfandel, GE-Healthcare and Oryzon Genomics, consultancy fees from Jung Diagnostics, Cytos Ltd., Axovant, Anavex, Takeda and Zinfandel, GE Healthcare and Oryzon Genomics, and participated in scientific advisory boards of Axovant, Eli Lilly and company, Cytos Ltd., GE Healthcare, Takeda and Zinfandel, Oryzon Genomics and Roche Diagnostics.

He is co-inventor in the following patents as a scientific expert and has received no royalties:

- *In Vitro* Multiparameter Determination Method for The Diagnosis and Early Diagnosis of Neurodegenerative Disorders Patent Number: 8916388
- *In Vitro* Procedure for Diagnosis and Early Diagnosis of Neurodegenerative Diseases Patent Number: 8298784
- Neurodegenerative Markers for Psychiatric Conditions Publication Number: 20120196300
- *In Vitro* Multiparameter Determination Method for The Diagnosis and Early Diagnosis of Neurodegenerative Disorders Publication Number: 20100062463
- *In Vitro* Method for The Diagnosis and Early Diagnosis of Neurodegenerative Disorders Publication Number: 20100035286
- *In Vitro* Procedure for Diagnosis and Early Diagnosis of Neurodegenerative Diseases Publication Number: 20090263822
- *In Vitro* Method for The Diagnosis of Neurodegenerative Diseases Patent Number: 7547553
- CSF Diagnostic in Vitro Method for Diagnosis of Dementias and Neuroinflammatory Diseases Publication Number: 20080206797
- *In Vitro* Method for The Diagnosis of Neurodegenerative Diseases Publication Number: 20080199966
- Neurodegenerative Markers for Psychiatric Conditions Publication Number: 20080131921

JC has provided consultation to AbbVie, Acadia, Actinogen, Alzheon, Anavex, Avanir, Axovant, Biogen, Boehringer-Ingelheim, Bracket, Dart, Eisai, Genentech, Kyowa, Lilly, Lundbeck, Medavante, Merck, Orion, Otsuka, Pfizer, QR, Roche, Suven, and Takeda pharmaceutical and assessment companies.

BD reports personal fees from Eli Lilly and company.

EK holds stock in AbbVie, Inc.

SL received lecture honoraria from Roche.

LSS reports grants from NIA and the State of California, and other from University of Southern California, Los Angeles, CA, USA, during the past three years; research contracts and grants from Eli Lilly, Lundbeck, Merck, Novartis, Roche/Genentech, Biogen, and TauRx; and consulting, advisory, or data monitoring committee fees from AC Immune, Accera, Allergan, Avraham, Boehringer Ingelheim, Cerespir, Cognition, Corium, Forum, Insys, Merck, Neurim, Roche, Stemmedica, Takeda, TauRx, vTv, and Toyama/FujiFilm, outside the submitted work.

AV, LEA, NB, KB, ACC, HJF, MF, RG, MH, MM, GP, LAW, JW, and FB declare no conflicts of interest.

Abbreviations:

Aβ₁₋₄₂	42-amino acid-long A β peptide
AD	Alzheimer's disease
ADAPT	Alzheimer's Disease Anti-Inflammatory Prevention Trial
ADAS-Cog	Alzheimer's Disease Assessment Scale-Cognitive Subscale
ADCS-ADL	Alzheimer's Disease Cooperative Study Activities of Daily Living
ADCS-PACC	Alzheimer's Disease Cooperative Study-Preclinical Alzheimer Cognitive Composite
AMS	Academy of Medical Sciences
APMI	Alzheimer Precision Medicine Initiative

APMI-CP	Alzheimer Precision Medicine Initiative Cohort Program
APOE	apolipoprotein E
APP	amyloid precursor protein
BACE1	β -site amyloid precursor protein cleaving enzyme
CD33	cluster of differentiation 33
CDER	Center for Drug Evaluation and Research
CDER/FDA	Center for Drug Evaluation and Research at the Food and Drug Administration
CDR-SB	Clinical Dementia Rating Scale Sum of Boxes
CNS	central nervous system
CPC	clinicopathological correlations
CSF	cerebrospinal fluid
EC50	half maximal effective concentration
fAD	familial AD
FDA	Food and Drug Administration
¹⁸F-FDG-PET	¹⁸ F-2-fluoro-2-deoxy-D-glucose
GWAS	genome-wide association studies
HEK293	human embryonic kidney 293
HMDB	Human Metabolome Database
IAPP	islet amyloid polypeptide
IL-1β	interleukin-1-beta
IL-6	interleukin-6
LOAD	late-onset AD
MMSE	Mini-Mental State Examination
MRI	magnetic resonance imaging
MS	mass spectrometry
MTD	maximum tolerated dose
NF-$\kappa$$\beta$	nuclear factor kappa-light-chain-enhancer of activated B cells
NSAIDs	non-steroidal anti-inflammatory drugs

PD	pharmacodynamics
PET	Positron emission tomography
PK	pharmacokinetic
PM	Precision Medicine
PMI	U.S. Precision Medicine Initiative
PP	Precision Pharmacology
PPAR-γ	peroxisome proliferator-activated receptor- γ
PSEN-1	presenilin-1
PSEN-2	presenilin-2
sIL-6RC	IL-6 receptor complex
SILK	stable isotope labelling kinetics
SNPs	single nucleotide polymorphism
TACE	TNF α converting enzyme
TDP-43	transactive response DNA-binding protein 43
TGF-β	transforming growth factor-beta
TNF-α	tumor necrosis factor-alpha
TNFRs	TNF receptors
TREM2	triggering receptor expressed on myeloid cells 2
UPS	ubiquitin/proteasome system

References

- [1]. Hampel H, Frank R, Broich K, Teipel SJ, Katz RG, Hardy J, Herholz K, Bokde ALW, Jessen F, Hoessler YC, Sanhai WR, Zetterberg H, Woodcock J, Blennow K, Biomarkers for Alzheimer's disease: academic, industry and regulatory perspectives, *Nat. Rev. Drug Discov* 9 (2010) 560–574, 10.1038/nrd3115. [PubMed: 20592748]
- [2]. Chen R, Mias GI, Li-Pook-Than J, Jiang L, Lam HYK, Chen R, Miriami E, Karczewski KJ, Hariharan M, Dewey FE, Cheng Y, Clark MJ, Im H, Habegger L, Balasubramanian S, O'Huallachain M, Dudley JT, Hillenmeyer S, Haraksingh R, Sharon D, Euskirchen G, Lacroute P, Bettinger K, Boyle AP, Kasowski M, Grubert F, Seki S, Garcia M, Whirl-Carrillo M, Gallardo M, Blasco MA, Greenberg PL, Snyder P, Klein TE, Altman RB, Butte AJ, Ashley EA, Gerstein M, Nadeau KC, Tang H, Snyder M, Personal omics profiling reveals dynamic molecular and medical phenotypes, *Cell* 148 (2012) 1293–1307, 10.1016/j.cell.2012.02.009. [PubMed: 22424236]
- [3]. Hampel H, O'Bryant S, Durrleman E, Younesi K, Escott-Price V, Corvol K, Broich A, Lista S, for the A.P.M. Initiative, A Precision Medicine Initiative for Alzheimer's disease: the road ahead to biomarker-guided integrative disease modeling, *Climacteric* (2017) 1–12, 10.1080/13697137.2017.1287866. [PubMed: 28105871]

- [4]. Hampel H, O'Bryant SE, Castrillo JI, Ritchie C, Rojkova K, Broich K, Benda N, Nisticò R, Frank RA, Dubois B, Escott-Price V, Lista S, PRECISION MEDICINE – The golden gate for detection, *Treat. Prev.f Alzheimer's Dis* (2016), 10.14283/jpad.2016.112.
- [5]. Iyengar R, Zhao S, Chung S-W, Mager DE, Gallo JM, Merging systems biology with pharmacodynamics, *Sci. Transl. Med* 4 (2012), 10.1126/scitranslmed.3003563, 126ps7
- [6]. Harrold JM, Ramanathan M, Mager DE, Network-based approaches in drug discovery and early development, *Clin. Pharmacol. Ther* 94 (2013) 651–658, 10.1038/clpt.2013.176. [PubMed: 24025802]
- [7]. Zhao S, Iyengar R, Systems pharmacology: network analysis to identify multiscale mechanisms of drug action, *Annu. Rev. Pharmacol. Toxicol* 52 (2012) 505–521, 10.1146/annurev-pharmtox-010611-134520. [PubMed: 22235860]
- [8]. Keiser MJ, Roth BL, Armbruster BN, Ernsberger P, Irwin JJ, Shoichet BK, Relating protein pharmacology by ligand chemistry, *Nat. Biotechnol* 25 (2007) 197–206, 10.1038/nbt1284. [PubMed: 17287757]
- [9]. Berg J, Gene-environment interplay, *Science* 354 (2016) 15, 10.1126/science.aal0219. [PubMed: 27846472]
- [10]. Cavedo E, Lista S, Khachaturian Z, Aisen P, Amouyel P, Herholz K, Jack CR, Sperling R, Cummings J, Blennow K, O'Bryant S, Frisoni GB, Khachaturian A, Kivipelto M, Klunk W, Broich K, Andrieu S, de Schotten MT, Mangin J-F, Lammertsma AA, Johnson K, Teipel S, Drzezga A, Bokde A, Colliot O, Bakardjian H, Zetterberg H, Dubois B, Vellas B, Schneider LS, Hampel H, The road ahead to cure Alzheimer's disease: development of biological markers and neuroimaging methods for prevention trials across all stages and target populations, *J. Prev. Alzheimers Dis* 1 (2014) 181–202, 10.14283/jpad.2014.32. [PubMed: 26478889]
- [11]. Bulyk ML, Walhout AJM, Chapter 4 – gene regulatory networks, in: *Handb. Syst. Biol*, Academic Press, San Diego, 2013, pp. 65–88, 10.1016/B978-0-12-385944-0.00004-6.
- [12]. Power A, Berger AC, Ginsburg GS, Genomics-enabled drug repositioning and repurposing: insights from an IOM Roundtable activity, *JAMA* 311 (2014) 2063–2064, 10.1001/jama.2014.3002. [PubMed: 24867009]
- [13]. Padmanabhan K, Shpanskaya K, Bello G, Doraiswamy PM, Samatova NF, Toward personalized network biomarkers in Alzheimer's disease: computing individualized genomic and protein crosstalk maps, *Front. Aging Neurosci* 9 (2017) 315, 10.3389/fnagi.2017.00315. [PubMed: 29085293]
- [14]. Vanneman M, Dranoff G, Combining immunotherapy and targeted therapies in cancer treatment, *Nat. Rev. Cancer* 12 (2012) 237–251, 10.1038/nrc3237. [PubMed: 22437869]
- [15]. Vogel C, Marcotte EM, Insights into the regulation of protein abundance from proteomic and transcriptomic analyses, *Nat. Rev. Genet* 13 (2012) 227–232, 10.1038/nrg3185. [PubMed: 22411467]
- [16]. Hampel H, Lista S, Teipel SJ, Garaci F, Nisticò R, Blennow K, Zetterberg H, Bertram L, Duyckaerts C, Bakardjian H, Drzezga A, Colliot O, Epelbaum S, Broich K, Lehericy S, Brice A, Khachaturian ZS, Aisen PS, Dubois B, Perspective on future role of biological markers in clinical therapy trials of Alzheimer's disease: a long-range point of view beyond 2020, *Biochem. Pharmacol* 88 (2014) 426–449, 10.1016/j.bcp.2013.11.009. [PubMed: 24275164]
- [17]. Miller JA, Woltjer RL, Goodenbour JM, Horvath S, Geschwind DH, Genes and pathways underlying regional and cell type changes in Alzheimer's disease, *Genome Med.* 5 (2013) 48, 10.1186/gm452. [PubMed: 23705665]
- [18]. Blennow K, Hampel H, Weiner M, Zetterberg H, Cerebrospinal fluid and plasma biomarkers in Alzheimer disease, *Nat. Rev. Neurol* 6 (2010) 131–144, 10.1038/nrneurol.2010.4. [PubMed: 20157306]
- [19]. Guo JL, Lee VMY, Cell-to-cell transmission of pathogenic proteins in neurodegenerative diseases, *Nat. Med* 20 (2014) 130–138, 10.1038/nm.3457. [PubMed: 24504409]
- [20]. Rawji KS, Mishra MK, Michaels NJ, Rivest S, Stys PK, Yong VW, Immunosenescence of microglia and macrophages: impact on the ageing central nervous system, *Brain J. Neurol* 139 (2016) 653–661, 10.1093/brain/awv395.

- [21]. Tarasoff-Conway JM, Carare RO, Osorio RS, Glodzik L, Butler T, Fieremans E, Axel L, Rusinek H, Nicholson C, Zlokovic BV, Frangione B, Blennow K, Ménard J, Zetterberg H, Wisniewski T, de Leon MJ, Clearance systems in the brain-implications for Alzheimer disease, *Nat. Rev. Neurol* 11 (2015) 457–470, 10.1038/nrneurol.2015.119. [PubMed: 26195256]
- [22]. Louveau A, Smirnov I, Keyes TJ, Eccles JD, Rouhani SJ, Peske JD, Derecki NC, Castle D, Mandell JW, Lee KS, Harris TH, Kipnis J, Structural and functional features of central nervous system lymphatic vessels, *Nature* 523 (2015) 337–341, 10.1038/nature14432. [PubMed: 26030524]
- [23]. Kenyon CJ, The genetics of ageing, *Nature* 464 (2010) 504–512, 10.1038/nature08980. [PubMed: 20336132]
- [24]. Fontana L, Partridge L, Longo VD, Extending healthy life span—from yeast to humans, *Science* 328 (2010) 321–326, 10.1126/science.1172539. [PubMed: 20395504]
- [25]. Lu T, Aron L, Zullo J, Pan Y, Kim H, Chen Y, Yang T-H, Kim H-M, Drake D, Liu XS, Bennett DA, Colaiácovo MP, Yankner BA, REST and stress resistance in ageing and Alzheimer’s disease, *Nature* 507 (2014) 448–454, 10.1038/nature13163. [PubMed: 24670762]
- [26]. Mair W, Morantte I, Rodrigues APC, Manning G, Montminy M, Shaw RJ, Dillin A, Lifespan extension induced by AMPK and calcineurin is mediated by CRTCL-1 and CREB, *Nature* 470 (2011) 404–408, 10.1038/nature09706. [PubMed: 21331044]
- [27]. Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, Nadon NL, Wilkinson JE, Frenkel K, Carter CS, Pahor M, Javors MA, Fernandez E, Miller RA, Rapamycin fed late in life extends lifespan in genetically heterogeneous mice, *Nature* 460 (2009) 392–395 10.1038/nature08221. [PubMed: 19587680]
- [28]. Camandola S, Mattson MP, Brain metabolism in health, aging, and neurodegeneration, *EMBO J.* 36 (2017) 1474–1492, 10.15252/embj.201695810. [PubMed: 28438892]
- [29]. Ramos FJ, Kaerberlein M, Ageing A healthy diet for stem cells, *Nature* 486 (2012) 477–478, 10.1038/486477a. [PubMed: 22739309]
- [30]. Eisenberg D, Jucker M, The amyloid state of proteins in human diseases, *Cell* 148 (2012) 1188–1203, 10.1016/j.cell.2012.02.022. [PubMed: 22424229]
- [31]. Balch WE, Morimoto RI, Dillin A, Kelly JW, Adapting proteostasis for disease intervention, *Science* 319 (2008) 916–919, 10.1126/science.1141448. [PubMed: 18276881]
- [32]. Villeda SA, Luo J, Mosher KI, Zou B, Britschgi M, Bieri G, Stan TM, Fainberg N, Ding Z, Eggel A, Lucin KM, Czirr E, Park J-S, Couillard-Després S, Aigner L, Li G, Peskind ER, Kaye JA, Quinn JF, Galasko DR, Xie XS, Rando TA, Wyss-Coray T, The ageing systemic milieu negatively regulates neurogenesis and cognitive function, *Nature* 477 (2011) 90–94, 10.1038/nature10357. [PubMed: 21886162]
- [33]. Hoeijmakers JHJ, DNA damage, aging, and cancer, *N. Engl. J. Med* 361 (2009) 1475–1485, 10.1056/NEJMra0804615. [PubMed: 19812404]
- [34]. Selman C, Tullet JMA, Wieser D, Irvine E, Lingard SJ, Choudhury AI, Claret M, Al-Qassab H, Carmignac D, Ramadani F, Woods A, Robinson ICA, Schuster E, Batterham RL, Kozma SC, Thomas G, Carling D, Okkenhaug K, Thornton JM, Partridge L, Gems D, Withers DJ, Ribosomal protein S6 kinase 1 signaling regulates mammalian life span, *Science* 326 (2009) 140–144, 10.1126/science.1177221. [PubMed: 19797661]
- [35]. Rolyan H, Scheffold A, Heinrich A, Begus-Nahrman Y, Langkopf BH, Hölter SM, Vogt-Weisenhorn DM, Liss B, Wurst W, Lie DC, Thal DR, Biber K, Rudolph KL, Telomere shortening reduces Alzheimer’s disease amyloid pathology in mice, *Brain J. Neurol* 134 (2011) 2044–2056, 10.1093/brain/awr133.
- [36]. Garinis GA, van der Horst GTJ, Vijg J, Hoeijmakers JHJ, DNA damage and ageing: new-age ideas for an age-old problem, *Nat. Cell Biol* 10 (2008) 1241–1247, 10.1038/ncb1108-1241. [PubMed: 18978832]
- [37]. Ron D, Walter P, Signal integration in the endoplasmic reticulum unfolded protein response, *Nat. Rev. Mol. Cell Biol* 8 (2007) 519–529, 10.1038/nrm2199. [PubMed: 17565364]
- [38]. Cohen P, Frame S, The renaissance of GSK3, *Nat. Rev. Mol. Cell Biol* 2 (2001) 769–776, 10.1038/35096075. [PubMed: 11584304]

- [39]. Natarajan M, Lin K-M, Hsueh RC, Sternweis PC, Ranganathan R, A global analysis of cross-talk in a mammalian cellular signalling network, *Nat. Cell Biol* 8 (2006) 571–580, 10.1038/ncb1418. [PubMed: 16699502]
- [40]. Matsunaga S, Kishi T, Annas P, Basun H, Hampel H, Iwata N, Lithium as a treatment for Alzheimer's disease: a systematic review and meta-analysis, *J. Alzheimers Dis. JAD* 48 (2015) 403–410, 10.3233/JAD-150437. [PubMed: 26402004]
- [41]. Benjamin D, Colombi M, Moroni C, Hall MN, Rapamycin passes the torch: a new generation of mTOR inhibitors, *Nat. Rev. Drug Discov* 10 (2011) 868–880, 10.1038/nrd3531. [PubMed: 22037041]
- [42]. O'Bryant SE, Mielke MM, Rissman RA, Lista S, Vanderstichele H, Zetterberg H, Lewczuk P, Posner H, Hall J, Johnson L, Fong Y-L, Luthman J, Jeromin A, Batrla-Utermann R, Villarreal A, Britton G, Snyder PJ, Henriksen K, Grammas P, Gupta V, Martins R, Hampel H, Biofluid based biomarker professional interest area, blood-based biomarkers in Alzheimer disease: current state of the science and a novel collaborative paradigm for advancing from discovery to clinic, *Alzheimers Dement. J. Alzheimers Assoc* 13 (2017) 45–58, 10.1016/j.jalz.2016.09.014.
- [43]. Gal AA, In search of the origins of modern surgical pathology, *Adv. Anat. Pathol* 8 (2001) 1–13. [PubMed: 11152089]
- [44]. Alzheimer A, Stelzmann RA, Schnitzlein HN, Murtagh FR, An english translation of Alzheimer's 1907 paper, uber eine eigenartige erkankung der hirnrinde, *Clin. Anat. N.Y* 8 (1995) 429–431, 10.1002/ca.980080612.
- [45]. Ogomori K, Kitamoto T, Tateishi J, Sato Y, Suetsugu M, Abe M, Beta-protein amyloid is widely distributed in the centra nervous system of patients with Alzheimer's disease, *Am. J. Pathol* 134 (1989) 243–251. [PubMed: 2464938]
- [46]. Braak H, Braak E, Demonstration of amyloid deposits and neurofibrillary changes in whole brain sections, *Brain Pathol. Zurich Switz* 1 (1991) 213–216.
- [47]. Crowther T, Goedert M, Wischik CM, The repeat region of microtubule-associated protein tau forms part of the core of the paired helical filament of Alzheimer's disease, *Ann. Med* 21 (1989) 127–132. [PubMed: 2504257]
- [48]. Foley P, Lipids in Alzheimer's disease: a century-old story, *Biochim. Biophys. Acta* 2010 (1801) 750–753, 10.1016/j.bbali.2010.05.004.
- [49]. Di Paolo G, Kim T-W, Linking lipids to Alzheimer's disease: cholesterol and beyond, *Nat. Rev. Neurosci* 12 (2011) 284–296, 10.1038/nrn3012. [PubMed: 21448224]
- [50]. Bateman RJ, Xiong C, Benzinger TLS, Fagan AM, Goate A, Fox NC, Marcus DS, Cairns NJ, Xie X, Blazey TM, Holtzman DM, Santacruz A, Buckles V, Oliver A, Moulder K, Aisen PS, Ghetti B, Klunk WE, McDade E, Martins RN, Masters CL, Mayeux R, Ringman JM, Rossor MN, Schofield PR, Sperling RA, Salloway S, Morris JC, Dominantly Inherited Alzheimer Network, Clinical and biomarker changes in dominantly inherited Alzheimer's disease, *N. Engl. J. Med* 367 (2012) 795–804, 10.1056/NEJMoa1202753. [PubMed: 22784036]
- [51]. Braak H, Braak E, Neuropathological stageing of Alzheimer-related changes, *Acta Neuropathol. (Berl)* 82 (1991) 239–259. [PubMed: 1759558]
- [52]. Hardy JA, Higgins GA, Alzheimer's disease: the amyloid cascade hypothesis, *Science* 256 (1992) 184–185. [PubMed: 1566067]
- [53]. Selkoe DJ, Toward a comprehensive theory for Alzheimer's disease. Hypothesis: Alzheimer's disease is caused by the cerebral accumulation and cytotoxicity of amyloid beta-protein, *Ann. N. Y. Acad. Sci* 924 (2000) 17–25. [PubMed: 11193794]
- [54]. Cummings JL, Morstorf T, Zhong K, Alzheimer's disease drug-development pipeline: few candidates, frequent failures, *Alzheimers Res. Ther* 6 (2014) 37, 10.1186/alzrt269. [PubMed: 25024750]
- [55]. Hardy J, Membrane damage is at the core of Alzheimer's disease, *Lancet Neurol.* 16 (2017) 342, 10.1016/S1474-4422(17)30091-1.
- [56]. Sperling RA, Aisen PS, Beckett LA, Bennett DA, Craft S, Fagan AM, Iwatsubo T, Jack CR, Kaye J, Montine TJ, Park DC, Reiman EM, Rowe CC, Siemers E, Stern Y, Yaffe K, Carrillo MC, Thies B, Morrison-Bogorad M, Wagster MV, Phelps CH, Toward defining the preclinical stages of Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's

- Association workgroups on diagnostic guidelines for Alzheimer's disease, *Alzheimers Dement. J. Alzheimers Assoc* 7 (2011) 280–292, 10.1016/j.jalz.2011.03.003.
- [57]. Pfeiffer RF, Neuroinflammation and Parkinson disease: the silent battleground, *Neurology* 73 (2009) 1434–1435, 10.1212/WNL.0b013e3181c2f07d. [PubMed: 19812378]
- [58]. Mattsson N, Bremell D, Anckarsäter R, Blennow K, Anckarsäter H, Zetterberg H, Hagberg L, Neuroinflammation in Lyme neuroborreliosis affects amyloid metabolism, *BMC Neurol.* 10 (2010) 51, 10.1186/1471-2377-10-51. [PubMed: 20569437]
- [59]. Heneka MT, Carson MJ, El Khoury J, Landreth GE, Brosseron F, Feinstein DL, Jacobs AH, Wyss-Coray T, Vitorica J, Ransohoff RM, Herrup K, Frautschy SA, Finsen B, Brown GC, Verkhratsky A, Yamanaka K, Koistinaho J, Latz E, Halle A, Petzold GC, Town T, Morgan D, Shinohara ML, Perry VH, Holmes C, Bazan NG, Brooks DJ, Hunot S, Joseph B, Deigendesch N, Garaschuk O, Boddeke E, Dinarello CA, Breitner JC, Cole GM, Golenbock DT, Kummer MP, Neuroinflammation in Alzheimer's disease, *Lancet Neurol.* 14 (2015) 388–405, 10.1016/S1474-4422(15)70016-5. [PubMed: 25792098]
- [60]. Filippi M, Agosta F, Does neuroinflammation sustain neurodegeneration in ALS? *Neurology* 87 (2016) 2508–2509, 10.1212/WNL.0000000000003441. [PubMed: 27837004]
- [61]. Malkki H, Multiple sclerosis: coagulation factors could mediate neuroinflammation in multiple sclerosis, *Nat. Rev. Neurol* 12 (2016) 679, 10.1038/nrneurol.2016.175.
- [62]. Simon DW, McGeachy MJ, Bayir H, Clark RSB, Loane DJ, Kochanek PM, The far-reaching scope of neuroinflammation after traumatic brain injury, *Nat. Rev. Neurol* 13 (2017) 572, 10.1038/nrneurol.2017.116. [PubMed: 28776601]
- [63]. Rossi S, Studer V, Motta C, Polidoro S, Perugini J, Macchiarulo G, Giovannetti AM, Pareja-Gutierrez L, Caldò A, Colonna I, Furlan R, Martino G, Centonze D, Neuroinflammation drives anxiety and depression in relapsing-remitting multiple sclerosis, *Neurology* 89 (2017) 1338–1347, 10.1212/WNL.0000000000004411. [PubMed: 28842450]
- [64]. Baldacci F, Lista S, Cavado E, Bonuccelli U, Hampel H, Diagnostic function of the neuroinflammatory biomarker YKL-40 in Alzheimer's disease and other neurodegenerative diseases, *Expert Rev, Proteomics* 14 (2017) 285–299, 10.1080/14789450.2017.1304217. [PubMed: 28281838]
- [65]. Glass CK, Saijo K, Winner B, Marchetto MC, Gage FH, Mechanisms underlying inflammation in neurodegeneration, *Cell* 140 (2010) 918–934, 10.1016/j.cell.2010.02.016. [PubMed: 20303880]
- [66]. Akiyama H, Barger S, Barnum S, Bradt B, Bauer J, Cole GM, Cooper NR, Eikelenboom P, Emmerling M, Fiebich BL, Finch CE, Frautschy S, Griffin WS, Hampel H, Hull M, Landreth G, Lue L, Mrak R, Mackenzie IR, McGeer PL, O'Banion MK, Pachter J, Pasinetti G, Plata-Salaman C, Rogers J, Rydel R, Shen Y, Streit W, Strommeyer R, Tooyoma I, Van Muiswinkel FL, Veerhuis R, Walker D, Webster S, Wegrzyniak B, Wenk G, Wyss-Coray T, Inflammation and Alzheimer's disease, *Neurobiol. Aging* 21 (2000) 383–421. [PubMed: 10858586]
- [67]. Duong T, Nikolaeva M, Acton PJ, C-reactive protein-like immunoreactivity in the neurofibrillary tangles of Alzheimer's disease, *Brain Res.* 749 (1997) 152–156. [PubMed: 9070642]
- [68]. Iwamoto N, Nishiyama E, Ohwada J, Arai H, Demonstration of CRP immunoreactivity in brains of Alzheimer's disease: immunohistochemical study using formic acid pretreatment of tissue sections, *Neurosci. Lett* 177 (1994) 23–26. [PubMed: 7824175]
- [69]. McGeer PL, McGeer EG, Inflammation, autotoxicity and Alzheimer disease, *Neurobiol. Aging* 22 (2001) 799–809. [PubMed: 11754986]
- [70]. van Dam AM, Brouns M, Louisse S, Berkenbosch F, Appearance of interleukin-1 in macrophages and in ramified microglia in the brain of endotoxin-treated rats: a pathway for the induction of non-specific symptoms of sickness? *Brain Res.* 588 (1992) 291–296. [PubMed: 1393581]
- [71]. Hickman SE, El Khoury J, TREM2 and the neuroimmunology of Alzheimer's disease, *Biochem. Pharmacol* 88 (2014) 495–498, 10.1016/j.bcp.2013.11.021. [PubMed: 24355566]
- [72]. Lill CM, Rengmark A, Pihlstrøm L, Fogh I, Shatunov A, Sleiman PM, Wang L-S, Liu T, Lassen CF, Meissner E, Alexopoulos P, Calvo A, Chio A, Dizdar N, Faltraco F, Forsgren L, Kirchheiner J, Kurz A, Larsen JP, Liebsch M, Linder J, Morrison KE, Nissbrandt H, Otto M, Pahnke J, Partch A, Restagno G, Rujescu D, Schnack C, Shaw CE, Shaw PJ, Tumani H, Tysnes

O-B, Valladares O, Silani V, van den Berg LH, van Rheenen W, Veldink JH, Lindenberg U, Steinhagen-Thiessen E, Consortium SLAGEN, Teipel S, Perneczky R, Hakonarson H, Hampel H, von Arnim CAF, Olsen JH, Van Deerlin VM, Al-Chalabi A, Toft M, Ritz B, Bertram L, The role of TREM2 R47H as a risk factor for Alzheimer's disease, frontotemporal lobar degeneration, amyotrophic lateral sclerosis, and Parkinson's disease, *Alzheimers Dement. J. Alzheimers Assoc* 11 (2015) 1407–1416, 10.1016/j.jalz.2014.12.009.

- [73]. Sherva R, Farrer LA, Power and pitfalls of the genome-wide association study approach to identify genes for Alzheimer's disease, *Curr. Psychiatry Rep* 13 (2011) 138–146, 10.1007/s11920-011-0184-4. [PubMed: 21312009]
- [74]. Villegas-Llerena C, Phillips A, Garcia-Reitboeck P, Hardy J, Pocock JM, Microglial genes regulating neuroinflammation in the progression of Alzheimer's disease, *Curr. Opin. Neurobiol* 36 (2016) 74–81, 10.1016/j.conb.2015.10.004. [PubMed: 26517285]
- [75]. Zhang Z-G, Li Y, Ng CT, Song Y-Q, Inflammation in Alzheimer's disease and molecular genetics: recent update, *Arch. Immunol. Ther. Exp. (Warsz.)* 63 (2015) 333–344, 10.1007/s00005-015-0351-0. [PubMed: 26232392]
- [76]. Jonsson T, Stefansson H, Steinberg S, Jonsdottir I, Jonsson PV, Snaedal J, Bjornsson S, Huttenlocher J, Levey AI, Lah JJ, Rujescu D, Hampel H, Giegling I, Andreassen OA, Engedal K, Ulstein I, Djurovic S, Ibrahim-Verbaas C, Hofman A, Ikram MA, van Duijn CM, Thorsteinsdottir U, Kong A, Stefansson K, Variant of TREM2 associated with the risk of Alzheimer's disease, *N. Engl. J. Med* 368 (2013) 107–116, 10.1056/NEJMoa1211103. [PubMed: 23150908]
- [77]. Guerreiro R, Wojtas A, Bras J, Carrasquillo M, Rogaeva E, Majounie E, Cruchaga C, Sassi C, Kauwe JSK, Younkin S, Hazrati L, Collinge J, Pocock J, Lashley T, Williams J, Lambert J-C, Amouyel P, Goate A, Rademakers R, Morgan K, Powell J, St George-Hyslop P, Singleton A, Hardy J, Alzheimer genetic analysis group, TREM2 variants in Alzheimer's disease, *N. Engl. J. Med* 368 (2013) 117–127, 10.1056/NEJMoa1211851. [PubMed: 23150934]
- [78]. Neumann H, Daly MJ, Variant TREM2 as risk factor for Alzheimer's disease, *N. Engl. J. Med* 368 (2013) 182–184, 10.1056/NEJMe1213157. [PubMed: 23151315]
- [79]. van Duijn CM, Hofman A, Nagelkerken L, Serum levels of interleukin-6 are not elevated in patients with Alzheimer's disease, *Neurosci. Lett* 108 (1990) 350–354. [PubMed: 2304653]
- [80]. Singh VK, Guthikonda P, Circulating cytokines in Alzheimer's disease, *J. Psychiatr. Res* 31 (1997) 657–660. [PubMed: 9447570]
- [81]. Lanzrein AS, Johnston CM, Perry VH, Jobst KA, King EM, Smith AD, Longitudinal study of inflammatory factors in serum, cerebrospinal fluid, and brain tissue in Alzheimer disease: interleukin-1beta, interleukin-6 interleukin-1 receptor antagonist, tumor necrosis factor-alpha, the soluble tumor necrosis factor receptors I and II, and alpha1-antichymotrypsin, *Alzheimer Dis. Assoc. Disord* 12 (1998) 215–227. [PubMed: 9772027]
- [82]. Bagli M, Papassotiropoulos A, Hampel H, Becker K, Jessen F, Bürger K, Ptok U, Rao ML, Möller H-J, Maier W, Heun R, Polymorphisms of the gene encoding the inflammatory cytokine interleukin-6 determine the magnitude of the increase in soluble interleukin-6 receptor levels in Alzheimer's disease. Results of a pilot study, *Eur. Arch. Psychiatry Clin. Neurosci* 253 (2003) 44–48, 10.1007/s00406-003-0405-x. [PubMed: 12664314]
- [83]. Richartz E, Stransky E, Batra A, Simon P, Lewczuk P, Buchkremer G, Bartels M, Schott K, Decline of immune responsiveness: a pathogenetic factor in Alzheimer's disease? *J. Psychiatr. Res* 39 (2005) 535–543, 10.1016/j.jpsychires.2004.12.005. [PubMed: 15992563]
- [84]. Dursun E, Gezen-Ak D, Hana ası H, Bilgiç B, Lohmann E, Ertan S, Atasoy L, Alaylıo lu S, Önal ÖB, Gündüz A, Apaydın H, Kızıltan G, Ulutin T, Gürvit H, Yılmaz S, The interleukin 1 alpha, interleukin 1 beta, interleukin 6 and alpha-2-macroglobulin serum levels in patients with early or late onset Alzheimer s disease, mild cognitive impairment or Parkinson s disease, *J. Neuroimmunol* 283 (2015) 50–57, 10.1016/j.jneuroim.2015.04.014. [PubMed: 26004156]
- [85]. Swardfager W, Lanctôt K, Rothenburg L, Wong A, Cappell J, Herrmann N, A meta-analysis of cytokines in Alzheimer's disease, *Biol. Psychiatry* 68 (2010) 930–941, 10.1016/j.biopsych.2010.06.012. [PubMed: 20692646]
- [86]. Shen Y, Wang H, Sun Q, Yao H, Keegan AP, Mullan M, Wilson J, Lista S, Leyhe T, Laske C, Rujescu D, Levey A, Wallin A, Blennow K, Li R, Hampel H, Increased plasma beta-secretase 1 may predict conversion to Alzheimer's disease dementia in individuals with mild cognitive

- impairment, *Biol. Psychiatry* 83 (5) (2018) 447–455, 10.1016/j.biopsych.2017.02.007. [PubMed: 28359566]
- [87]. Li R, Yang L, Lindholm K, Konishi Y, Yue X, Hampel H, Zhang D, Shen Y, Tumor necrosis factor death receptor signaling cascade is required for amyloid-beta protein-induced neuron death, *J. Neurosci* 24 (2004) 1760–1771, 10.1523/JNEUROSCI.4580-03.2004. [PubMed: 14973251]
- [88]. He P, Zhong Z, Lindholm K, Berning L, Lee W, Lemere C, Staufenbiel M, Li R, Shen Y, Deletion of tumor necrosis factor death receptor inhibits amyloid beta generation and prevents learning and memory deficits in Alzheimer's mice, *J. Cell Biol* 178 (2007) 829–841, 10.1083/jcb.200705042. [PubMed: 17724122]
- [89]. Cheng X, Yang L, He P, Li R, Shen Y, Differential activation of tumor necrosis factor receptors distinguishes between brains from Alzheimer's disease and non-demented patients, *J. Alzheimers Dis. JAD* 19 (2010) 621–630, 10.3233/JAD-2010-1253. [PubMed: 20110607]
- [90]. Jiang H, Hampel H, Prvulovic D, Wallin A, Blennow K, Li R, Shen Y, Elevated CSF levels of TACE activity and soluble TNF receptors in subjects with mild cognitive impairment and patients with Alzheimer's disease, *Mol. Neurodegener* 6 (2011) 69, 10.1186/1750-1326-6-69. [PubMed: 21978728]
- [91]. Hampel H, Teipel SJ, Padberg F, Haslinger A, Riemenschneider M, Schwarz MJ, Kötter HU, Scheloske M, Buch K, Stübner S, Dukoff R, Lasser R, Müller N, Sunderland T, Rapoport SI, Möller HJ, Discriminant power of combined cerebrospinal fluid tau protein and of the soluble interleukin-6 receptor complex in the diagnosis of Alzheimer's disease, *Brain Res.* 823 (1999) 104–112. [PubMed: 10095017]
- [92]. Hampel H, Sunderland T, Kötter HU, Schneider C, Teipel SJ, Padberg F, Dukoff R, Levy J, Möller HJ, Decreased soluble interleukin-6 receptor in cerebrospinal fluid of patients with Alzheimer's disease, *Brain Res.* 780 (1998) 356–359. [PubMed: 9507194]
- [93]. Tan ZS, Beiser AS, Vasan RS, Roubenoff R, Dinarello CA, Harris TB, Benjamin EJ, Au R, Kiel DP, Wolf PA, Seshadri S, Inflammatory markers and the risk of Alzheimer disease: the Framingham Study, *Neurology* 68 (2007) 1902–1908, 10.1212/01.wnl.0000263217.36439.da. [PubMed: 17536046]
- [94]. Baldacci F, Toschi N, Lista S, Zetterberg H, Blennow K, Kilimann I, Teipel S, Cavado E, Dos Santos AM, Epelbaum S, Lamari F, Dubois B, Floris R, Garaci F, Bonuccelli U, Hampel H, Two-level diagnostic classification using cerebrospinal fluid YKL-40 in Alzheimer's disease, *Alzheimers Dement.* 13 (9) (2017) 993–1003, 10.1016/j.jalz.2017.01.021. [PubMed: 28263742]
- [95]. Jones DP, Sequencing the exposome: a call to action, *Toxicol. Rep* 3 (2016) 29–45, 10.1016/j.toxrep.2015.11.009. [PubMed: 26722641]
- [96]. Yugi K, Kubota H, Hatano A, Kuroda S, Trans-omics: how to reconstruct biochemical networks across multiple omic layers, *Trends Biotechnol.* 34 (2016) 276–290, 10.1016/j.tibtech.2015.12.013. [PubMed: 26806111]
- [97]. Barba I, Fernandez-Montesinos R, Garcia-Dorado D, Pozo D, Alzheimer's disease beyond the genomic era: nuclear magnetic resonance (NMR) spectroscopy-based metabolomics, *J. Cell. Mol. Med* 12 (2008) 1477–1485, 10.1111/j.1582-4934.2008.00385.x. [PubMed: 18554316]
- [98]. Zhang A, Sun H, Wang P, Han Y, Wang X, Modern analytical techniques in metabolomics analysis, *Analyst* 137 (2012) 293–300, 10.1039/c1an15605e. [PubMed: 22102985]
- [99]. Sato Y, Suzuki I, Nakamura T, Bernier F, Aoshima K, Oda Y, Identification of a new plasma biomarker of Alzheimer's disease using metabolomics technology, *J. Lipid Res* 53 (2012) 567–576, 10.1194/jlr.M022376. [PubMed: 22203775]
- [100]. Trushina E, Dutta T, Persson X-MT, Mielke MM, Petersen RC, Identification of altered metabolic pathways in plasma and CSF in mild cognitive impairment and Alzheimer's disease using metabolomics, *PLoS One* 8 (2013) e63644, 10.1371/journal.pone.0063644. [PubMed: 23700429]
- [101]. Wang G, Zhou Y, Huang F-J, Tang H-D, Xu X-H, Liu J-J, Wang Y, Deng Y-L, Ren R-J, Xu W, Ma J-F, Zhang Y-N, Zhao A-H, Chen S-D, Jia W, Plasma metabolite profiles of Alzheimer's disease and mild cognitive impairment, *J. Proteome Res* 13 (2014) 2649–2658, 10.1021/pr5000895. [PubMed: 24694177]

- [102]. Graham SF, Chevallier OP, Elliott CT, Hölscher C, Johnston J, McGuinness B, Kehoe PG, Passmore AP, Green BD, Untargeted metabolomic analysis of human plasma indicates differentially affected polyamine and L-arginine metabolism in mild cognitive impairment subjects converting to Alzheimer's disease, *PLoS One* 10 (2015) e0119452, 10.1371/journal.pone.0119452. [PubMed: 25803028]
- [103]. González-Domínguez R, Rupérez FJ, García-Barrera T, Barbas C, Gómez-Ariza JL, Metabolomic-driven elucidation of serum disturbances associated with Alzheimer's disease and mild cognitive impairment, *Curr. Alzheimer Res* 13 (2016) 641–653. [PubMed: 26825096]
- [104]. Whiley L, Sen A, Heaton J, Proitsi P, García-Gómez D, Leung R, Smith N, Thambisetty M, Kloszewska I, Mecocci P, Soininen H, Tsolaki M, Vellas B, Lovestone S, Legido-Quigley C, AddNeuroMed Consortium, Evidence of altered phosphatidylcholine metabolism in Alzheimer's disease, *Neurobiol. Aging* 35 (2014) 271–278, 10.1016/j.neurobiolaging.2013.08.001. [PubMed: 24041970]
- [105]. Klavins K, Koal T, Dallmann G, Marksteiner J, Kemmler G, Humpel C, The ratio of phosphatidylcholines to lysophosphatidylcholines in plasma differentiates healthy controls from patients with Alzheimer's disease and mild cognitive impairment, *Alzheimers Dement. Amst. Neth* 1 (2015) 295–302, 10.1016/j.dadm.2015.05.003.
- [106]. Mapstone M, Cheema AK, Fiandaca MS, Zhong X, Mhyre TR, MacArthur LH, Hall WJ, Fisher SG, Peterson DR, Haley JM, Nazar MD, Rich SA, Berlau DJ, Peltz CB, Tan MT, Kawas CH, Federoff HJ, Plasma phospholipids identify antecedent memory impairment in older adults, *Nat. Med* 20 (2014) 415–418, 10.1038/nm.3466. [PubMed: 24608097]
- [107]. Fiandaca MS, Zhong X, Cheema AK, Orquiza MH, Chidambaram S, Tan MT, Gresenz CR, FitzGerald KT, Nalls MA, Singleton AB, Mapstone M, Federoff HJ, Plasma 24-metabolite panel predicts preclinical transition to clinical stages of Alzheimer's disease, *Front. Neurol* 6 (2015) 237, 10.3389/fneur.2015.00237. [PubMed: 26617567]
- [108]. Hartmann T, van Wijk N, Wurtman RJ, Olde Rikkert MGM, Sijben JWC, Soininen H, Vellas B, Scheltens P, A nutritional approach to ameliorate altered phospholipid metabolism in Alzheimer's disease, *J. Alzheimers Dis. JAD* 41 (2014) 715–717, 10.3233/JAD-141137. [PubMed: 24898653]
- [109]. Fiandaca MS, Mapstone ME, Cheema AK, Federoff HJ, The critical need for defining preclinical biomarkers in Alzheimer's disease, *Alzheimers Dement. J. Alzheimers Assoc* 10 (2014) S196–S212, 10.1016/j.jalz.2014.04.015.
- [110]. Wyss-Coray T, Mucke L, Inflammation in neurodegenerative disease—a double-edged sword, *Neuron* 35 (2002) 419–432. [PubMed: 12165466]
- [111]. Weninger SC, Yankner BA, Inflammation and Alzheimer disease: the good, the bad, and the ugly, *Nat. Med* 7 (2001) 527–528, 10.1038/87839. [PubMed: 11329045]
- [112]. Cuello AC, Early and late CNS inflammation in Alzheimer's disease: two extremes of a continuum? *Trends Pharmacol. Sci* 38 (2017) 956–966, 10.1016/j.tips.2017.07.005. [PubMed: 28867259]
- [113]. McGeer PL, McGeer E, Rogers J, Sibley J, Anti-inflammatory drugs and Alzheimer disease, *Lancet Lond. Engl* 335 (1990) 1037.
- [114]. McGeer PL, McGeer EG, The amyloid cascade-inflammatory hypothesis of Alzheimer disease: implications for therapy, *Acta Neuropathol. (Berl.)* 126 (2013) 479–497, 10.1007/s00401-013-1177-7. [PubMed: 24052108]
- [115]. Miguel-Álvarez M, Santos-Lozano A, Sanchis-Gomar F, Fiuza-Luces C, Pareja-Galeano H, Garatachea N, Lucia A, Non-steroidal anti-inflammatory drugs as a treatment for Alzheimer's disease: a systematic review and meta-analysis of treatment effect, *Drugs Aging* 32 (2015) 139–147, 10.1007/s40266-015-0239-z. [PubMed: 25644018]
- [116]. Gupta PP, Pandey RD, Jha D, Shrivastav V, Kumar S, Role of traditional nonsteroidal anti-inflammatory drugs in Alzheimer's disease: a meta-analysis of randomized clinical trials, *Am. J. Alzheimers Dis. Other Demen* 30 (2015) 178–182, 10.1177/1533317514542644. [PubMed: 25024454]
- [117]. Jaturapatporn D, Isaac MGEKN, McCleery J, Tabet N, Aspirin, steroidal and non-steroidal anti-inflammatory drugs for the treatment of Alzheimer's disease, *Cochrane Database Syst. Rev* (2012) CD006378, 10.1002/14651858.CD006378.pub2. [PubMed: 22336816]

- [118]. Breitner JC, Baker LD, Montine TJ, Meinert CL, Lyketsos CG, Ashe KH, Brandt J, Craft S, Evans DE, Green RC, Ismail MS, Martin BK, Mullan MJ, Sabbagh M, Tariot PN, ADAPT research group, extended results of the Alzheimer's disease anti-inflammatory prevention trial, *Alzheimers Dement. J. Alzheimers Assoc* 7 (2011) 402–411, 10.1016/j.jalz.2010.12.014.
- [119]. Hollingworth P, Harold D, Sims R, Gerrish A, Lambert J-C, et al. , Common variants at ABCA7, MS EPHA1, CD33 and CD2AP are associated with Alzheimer's disease, *Nat. Genet* 43 (2011) 429–435, 10.1038/ng.803. [PubMed: 21460840]
- [120]. Naj AC, Jun G, Beecham GW, Wang L-S, Vardarajan BN, et al. , Common variants at MS4A4/MS4A6E, CD2AP, CD33 and EPHA1 are associated with late-onset Alzheimer's disease, *Nat. Genet* 43 (2011) 436–441, 10.1038/ng.801. [PubMed: 21460841]
- [121]. Roussos P, Katsel P, Fam P, Tan W, Purohit DP, Haroutunian V, The triggering receptor expressed on myeloid cells 2 (TREM2) is associated with enhanced inflammation, neuropathological lesions and increased risk for Alzheimer's dementia, *Alzheimers Dement, J. Alzheimers Assoc* 11 (2015) 1163–1170, 10.1016/j.jalz.2014.10.013.
- [122]. Janelins MC, Mastrangelo MA, Oddo S, LaFerla FM, Federoff HJ, Bowers WJ, Early correlation of microglial activation with enhanced tumor necrosis factor-alpha and monocyte chemoattractant protein-1 expression specifically within the entorhinal cortex of triple transgenic Alzheimer's disease mice, *J. Neuroinflammation* 2 (2005) 23, 10.1186/1742-2094-2-23. [PubMed: 16232318]
- [123]. McAlpine FE, Lee J-K, Harms AS, Ruhn KA, Blurton-Jones M, Hong J, Das P, Golde TE, LaFerla FM, Oddo S, Blesch A, Tansey MG, Inhibition of soluble TNF signaling in a mouse model of Alzheimer's disease prevents pre-plaque amyloid-associated neuropathology, *Neurobiol Dis.* 34 (2009) 163–177. [PubMed: 19320056]
- [124]. Cavanagh C, Tse YC, Nguyen H-B, Krantic S, Breitner JCS, Quirion R, Wong TP, Inhibiting tumor necrosis factor- α before amyloidosis prevents synaptic deficits in an Alzheimer's disease model, *Neurobiol. Aging* 47 (2016) 41–49, 10.1016/j.neurobiolaging.2016.07.009. [PubMed: 27552480]
- [125]. Hanzel CE, Pichet-Binette A, Pimentel LSB, Iulita MF, Allard S, Ducatzenzeiler A, Do Carmo S, Cuello AC, Neuronal driven pre-plaque inflammation in a transgenic rat model of Alzheimer's disease, *Neurobiol. Aging* 35 (2014) 2249–2262, 10.1016/j.neurobiolaging.2014.03.026. [PubMed: 24831823]
- [126]. Pimentel LS, Allard S, Do Carmo S, Weinreb O, Danik M, Hanzel CE, Youdim MB, Cuello AC, The multi-target drug M30 shows pro-cognitive and anti-inflammatory effects in a rat model of Alzheimer's disease, *J. Alzheimers Dis. JAD* 47 (2015) 373–383, 10.3233/JAD-143126. [PubMed: 26401560]
- [127]. Ferretti MT, Allard S, Partridge V, Ducatzenzeiler A, Cuello AC, Minocycline corrects early, pre-plaque neuroinflammation and inhibits BACE-1 in a transgenic model of Alzheimer's disease-like amyloid pathology, *J. Neuroinflammation* 9 (2012) 62, 10.1186/1742-2094-9-62. [PubMed: 22472085]
- [128]. Bruno MA, Leon WC, Fragoso G, Mushynski WE, Almazan G, Cuello AC, Amyloid beta-induced nerve growth factor dysmetabolism in Alzheimer disease, *J. Neuropathol. Exp. Neurol* 68 (2009) 857–869, 10.1097/NEN.0b013e3181aed9e6. [PubMed: 19606067]
- [129]. Jack CR, Knopman DS, Jagust WJ, Petersen RC, Weiner MW, Aisen PS, Shaw LM, Vemuri P, Wiste HJ, Weigand SD, Lesnick TG, Pankratz VS, Donohue MC, Trojanowski JQ, Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers, *Lancet Neurol.* 12 (2013) 207–216, 10.1016/S1474-4422(12)70291-0. [PubMed: 23332364]
- [130]. McDade E, Bateman RJ, Stop Alzheimer's before it starts, *Nature* 547 (2017) 153–155, 10.1038/547153a. [PubMed: 28703214]
- [131]. Rodriguez-Vieitez E, Saint-Aubert L, Carter SF, Almkvist O, Farid K, Schöll M, Chiotis K, Thordardottir S, Graff C, Wall A, Långström B, Nordberg A, Diverging longitudinal changes in astrogliosis and amyloid PET in autosomal dominant Alzheimer's disease, *Brain J. Neurol* 139 (2016) 922–936, 10.1093/brain/awv404.
- [132]. Walker KA, Hoogeveen RC, Folsom AR, Ballantyne CM, Knopman DS, Windham AG, Jack CR, Gottesman RF, Midlife systemic inflammatory markers are associated with late-life brain

- volume: the ARIC study, *Neurology* 89 (2017) 2262–2270, 10.1212/WNL.0000000000004688. [PubMed: 29093073]
- [133]. Dinarello CA, Anti-inflammatory agents: present and future, *Cell* 140 (2010) 935–950, 10.1016/j.cell.2010.02.043. [PubMed: 20303881]
- [134]. Tobinick E, Gross H, Weinberger A, Cohen H, TNF-alpha modulation for treatment of Alzheimer's disease: a 6-month pilot study, *MedGenMed Medscape Gen. Med* 8 (2006) 25.
- [135]. Butchart J, Brook L, Hopkins V, Teeling J, Püntener U, Culliford D, Sharples R, Sharif S, McFarlane B, Raybould R, Thomas R, Passmore P, Perry VH, Holmes C, Etanercept in Alzheimer disease: a randomized, placebo-controlled, double-blind, phase 2 trial, *Neurology* 84 (2015) 2161–2168, 10.1212/WNL.0000000000001617. [PubMed: 25934853]
- [136]. Shi J-Q, Wang B-R, Jiang W-W, Chen J, Zhu Y-W, Zhong L-L, Zhang Y-D, Xu J, Cognitive improvement with intrathecal administration of infliximab in a woman with Alzheimer's disease, *J. Am. Geriatr. Soc* 59 (2011) 1142–1144, 10.1111/j.1532-5415.2011.03445.x. [PubMed: 21668921]
- [137]. Pascual G, Fong AL, Ogawa S, Gamliel A, Li AC, Perissi V, Rose DW, Willson TM, Rosenfeld MG, Glass CK, A SUMOylation-dependent pathway mediates transrepression of inflammatory response genes by PPAR-gamma, *Nature* 437 (2005) 759–763, 10.1038/nature03988. [PubMed: 16127449]
- [138]. Landreth G, Jiang Q, Mandrekar S, Heneka M, PPARgamma agonists as therapeutics for the treatment of Alzheimer's disease, *Neurother. J. Am. Soc. Exp. Neurother* 5 (2008) 481–489, 10.1016/j.nurt.2008.05.003.
- [139]. Watson GS, Cholerton BA, Reger MA, Baker LD, Plymate SR, Asthana S, Fishel MA, Kulstad JJ, Green PS, Cook DG, Kahn SE, Keeling ML, Craft S, Preserved cognition in patients with early Alzheimer disease and amnesic mild cognitive impairment during treatment with rosiglitazone: a preliminary study, *Am. J. Geriatr. Psychiatry* 13 (2005) 950–958, 10.1176/appi.ajgp.13.11.950. [PubMed: 16286438]
- [140]. Risner ME, Saunders AM, Altman JFB, Ormandy GC, Craft S, Foley IM, Zvartau-Hind ME, Hosford DA, Roses AD, Rosiglitazone in Alzheimer's Disease Study Group, Efficacy of rosiglitazone in a genetically defined population with mild-to-moderate Alzheimer's disease, *Pharmacogenomics J.* 6 (2006) 246–254, 10.1038/sj.tpj.6500369. [PubMed: 16446752]
- [141]. Lachmann HJ, Kone-Paut I, Kuemmerle-Deschner JB, Leslie KS, Hachulla E, Quartier P, Gitton X, Widmer A, Patel N, Hawkins PN, Canakinumab in CAPS Study Group, Use of canakinumab in the cryopyrin-associated periodic syndrome, *N. Engl. J. Med* 360 (2009) 2416–2425, 10.1056/NEJMoa0810787. [PubMed: 19494217]
- [142]. Dhimolea E, Canakinumab, *MAbs* 2 (2010) 3–13. [PubMed: 20065636]
- [143]. Martinon F, Burns K, Tschopp J, The inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proIL-beta, *Mol. Cell* 10 (2002) 417–426. [PubMed: 12191486]
- [144]. de Rivero Vaccari JP, Dietrich WD, Keane RW, Activation and regulation of cellular inflammasomes: gaps in our knowledge for central nervous system injury, *J. Cereb. Blood Flow Metab* 34 (2014) 369–375, 10.1038/jcbfm.2013.227. [PubMed: 24398940]
- [145]. Walsh JG, Muruve DA, Power C, Inflammasomes in the CNS, *Nat. Rev. Neurosci* 15 (2014) 84–97, 10.1038/nrn3638. [PubMed: 24399084]
- [146]. Latz E, Xiao TS, Stutz A, Activation and regulation of the inflammasomes, *Nat. Rev. Immunol* 13 (2013) 397–411, 10.1038/nri3452. [PubMed: 23702978]
- [147]. Heneka MT, Kummer MP, Stutz A, Delekate A, Schwartz S, Vieira-Saecker A, Griep A, Axt D, Remus A, Tzeng T-C, Gelpi E, Halle A, Korte M, Latz E, Golenbock DT, NLRP3 is activated in Alzheimer's disease and contributes to pathology in APP/PS1 mice, *Nature* 493 (2013) 674–678, 10.1038/nature11729. [PubMed: 23254930]
- [148]. Kaushal V, Dye R, Pakavathkumar P, Foveau B, Flores J, Hyman B, Ghetti B, Koller BH, LeBlanc AC, Neuronal NLRP1 inflammasome activation of Caspase-1 coordinately regulates inflammatory interleukin-1-beta production and axonal degeneration-associated Caspase-6 activation, *Cell Death Differ.* 22 (2015) 1676–1686, 10.1038/cdd.2015.16. [PubMed: 25744023]

- [149]. Yin J, Zhao F, Chojnacki JE, Fulp J, Klein WL, Zhang S, Zhu X, NLRP3 inflammasome inhibitor ameliorates amyloid pathology in a mouse model of Alzheimer's disease, *Mol. Neurobiol* (2017), 10.1007/s12035-017-0467-9.
- [150]. Daniels MJD, Rivers-Auty J, Schilling T, Spencer NG, Watremez W, Fasolino V, Booth SJ, White CS, Baldwin AG, Freeman S, Wong R, Latta C, Yu S, Jackson J, Fischer N, Koziel V, Pillot T, Bagnall J, Allan SM, Paszek P, Galea J, Harte MK, Eder C, Lawrence CB, Brough D, Fenamate NSAIDs inhibit the NLRP3 inflammasome and protect against Alzheimer's disease in rodent models, *Nat. Commun* 7 (2016) 12504, 10.1038/ncomms12504. [PubMed: 27509875]
- [151]. Bradshaw EM, Chibnik LB, Keenan BT, Ottoboni L, Raj T, Tang A, Rosenkrantz LL, Imboywa S, Lee M, Von Korff A, Alzheimer Disease Neuroimaging Initiative, Morris MC, Evans DA, Johnson K, Sperling RA, Schneider JA, Bennett DA, De Jager PL, CD33 Alzheimer's disease locus: altered monocyte function and amyloid biology, *Nat. Neurosci* 16 (2013) 848–850, 10.1038/nn.3435. [PubMed: 23708142]
- [152]. Griciuc A, Serrano-Pozo A, Parrado AR, Lesinski AN, Asselin CN, Mullin K, Hooli B, Choi SH, Hyman BT, Tanzi RE, Alzheimer's disease risk gene CD33 inhibits microglial uptake of amyloid beta, *Neuron* 78 (2013) 631–643, 10.1016/j.neuron.2013.04.014. [PubMed: 23623698]
- [153]. Kleinberger G, Yamanishi Y, Suárez-Calvet M, Czirr E, Lohmann E, Cuyvers E, Struyfs H, Pettkus N, Wenninger-Weinzierl A, Mazaheri F, Tahirovic S, Lleó A, Alcolea D, Fortea J, Willem M, Lammich S, Molinuevo JL, Sánchez-Valle R, Antonell A, Ramirez A, Heneka MT, Slegers K, van der Zee J, Martin J-J, Engelborghs S, Demirtas-Tatlidede A, Zetterberg H, Van Broeckhoven C, Gurvit H, Wyss-Coray T, Hardy J, Colonna M, Haass C, TREM2 mutations implicated in neurodegeneration impair cell surface transport and phagocytosis, *Sci. Transl. Med* 6 (2014), 10.1126/scitranslmed.3009093, 243ra86.
- [154]. Wang Y, Cella M, Mallinson K, Ulrich JD, Young KL, Robinette ML, Gilfillan S, Krishnan GM, Sudhakar S, Zinselmeier BH, Holtzman DM, Cirrito JR, Colonna M, TREM2 lipid sensing sustains the microglial response in an Alzheimer's disease model, *Cell* 160 (2015) 1061–1071, 10.1016/j.cell.2015.01.049. [PubMed: 25728668]
- [155]. Laing AA, Harrison CJ, Gibson BES, Keeshan K, Unlocking the potential of anti-CD33 therapy in adult and childhood acute myeloid leukemia, *Exp. Hematol* 54 (2017) 40–50, 10.1016/j.exphem.2017.06.007. [PubMed: 28668350]
- [156]. Dubois B, Feldman HH, Jacova C, Hampel H, Molinuevo JL, Blennow K, DeKosky ST, Gauthier S, Selkoe D, Bateman R, Cappa S, Crutch S, Engelborghs S, Frisoni GB, Fox NC, Galasko D, Habert M-O, Jicha GA, Nordberg A, Pasquier F, Rabinovici G, Robert P, Rowe C, Salloway S, Sarazin M, Epelbaum S, de Souza LC, Vellas B, Visser PJ, Schneider L, Stern Y, Scheltens P, Cummings JL, Advancing research diagnostic criteria for Alzheimer's disease: the IWG-2 criteria, *Lancet Neurol.* 13 (2014) 614–629, 10.1016/S1474-4422(14)70090-0. [PubMed: 24849862]
- [157]. Wang J, Gu BJ, Masters CL, Wang Y-J, A systemic view of Alzheimer disease – insights from amyloid- β metabolism beyond the brain, *Nat. Rev. Neurol* 13 (2017) 612–623, 10.1038/nrneurol.2017.111. [PubMed: 28960209]
- [158]. Dubois B, Hampel H, Feldman HH, Scheltens P, Aisen P, Andrieu S, Bakardjian H, Benali H, Bertram L, Blennow K, Broich K, Cavado E, Crutch S, Dartigues J-F, Duyckaerts C, Epelbaum S, Frisoni GB, Gauthier S, Genthon R, Gouw AA, Habert M-O, Holtzman DM, Kivipelto M, Lista S, Molinuevo J-L, O'Bryant SE, Rabinovici GD, Rowe C, Salloway S, Schneider LS, Sperling R, Teichmann M, Carrillo MC, Cummings J, Jack CR, Proceedings of the Meeting of the International Working Group (IWG) and the American Alzheimer's Association on The Preclinical State of AD; July 23, 2015; Washington DC, USA, Preclinical Alzheimer's disease: Definition, natural history, and diagnostic criteria, *Alzheimers Dement, J. Alzheimers Assoc* 12 (2016) 292–323, 10.1016/j.jalz.2016.02.002.
- [159]. Wang Y-J, Alzheimer disease: lessons from immunotherapy for Alzheimer disease, *Nat. Rev. Neurol* 10 (2014) 188–189, 10.1038/nrneurol.2014.44. [PubMed: 24638135]
- [160]. Jan A, Adolfsson O, Allaman I, Buccarello A-L, Magistretti PJ, Pfeifer A, Muhs A, Lashuel HA, A β 42 neurotoxicity is mediated by ongoing nucleated polymerization process rather than by discrete A β 42 species, *J. Biol. Chem* 286 (2011) 8585–8596, 10.1074/jbc.M110.172411. [PubMed: 21156804]

- [161]. Anand R, Gill KD, Mahdi AA, Therapeutics of Alzheimer's disease: past, present and future, *Neuropharmacology* 76 (Pt A) (2014) 27–50, 10.1016/j.neuropharm.2013.07.004. [PubMed: 23891641]
- [162]. Selkoe DJ, Hardy J, The amyloid hypothesis of Alzheimer's disease at 25 years, *EMBO Mol. Med* 8 (2016) 595–608, 10.15252/emmm.201606210. [PubMed: 27025652]
- [163]. Beel AJ, Sanders CR, Substrate specificity of gamma-secretase and other intramembrane proteases, *Cell. Mol. Life Sci. CMLS* 65 (2008) 1311–1334, 10.1007/s00018-008-7462-2. [PubMed: 18239854]
- [164]. Abramov E, Dolev I, Fogel H, Ciccotosto GD, Ruff E, Slutsky I, Amyloid-beta as a positive endogenous regulator of release probability at hippocampal synapses, *Nat. Neurosci* 12 (2009) 1567–1576, 10.1038/nn.2433. [PubMed: 19935655]
- [165]. Zimbone S, Monaco I, Giani F, Pandini G, Copani AG, Giuffrida ML, Rizzarelli E, Amyloid Beta monomers regulate cyclic adenosine monophosphate response element binding protein functions by activating type-I insulin-like growth factor receptors in neuronal cells, *Aging Cell* 17 (2018), 10.1111/acer.12684.
- [166]. Parsons CG, Rammes G, Preclinical to phase II amyloid beta (A β) peptide modulators under investigation for Alzheimer's disease, *Expert Opin. Investig. Drugs* 26 (2017) 579–592, 10.1080/13543784.2017.1313832.
- [167]. Nalivaeva NN, Beckett C, Belyaev ND, Turner AJ, Are amyloid-degrading enzymes viable therapeutic targets in Alzheimer's disease? *J. Neurochem* 120 (Suppl. 1) (2012) 167–185, 10.1111/j.1471-4159.2011.07510.x. [PubMed: 22122230]
- [168]. Cabrol C, Huzarska MA, Dinolfo C, Rodriguez MC, Reinstatler L, Ni J, Yeh L-A, Cuny GD, Stein RL, Selkoe DJ, Leissring MA, Small-molecule activators of insulin-degrading enzyme discovered through high-throughput compound screening, *PLoS One* 4 (2009) e5274, 10.1371/journal.pone.0005274. [PubMed: 19384407]
- [169]. Martel CL, Mackic JB, Matsubara E, Governale S, Miguel C, Miao W, McComb JG, Frangione B, Ghiso J, Zlokovic BV, Isoform-specific effects of apolipoproteins E2 E3, and E4 on cerebral capillary sequestration and blood-brain barrier transport of circulating Alzheimer's amyloid beta, *J. Neurochem* 69 (1997) 1995–2004. [PubMed: 9349544]
- [170]. Zlokovic BV, Clearing amyloid through the blood-brain barrier, *J. Neurochem* 89(2004) 807–811, 10.1111/j.1471-4159.2004.02385.x. [PubMed: 15140180]
- [171]. Erickson MA, Banks WA, Blood-brain barrier dysfunction as a cause and consequence of Alzheimer's disease, *J. Cereb. Blood Flow Metab* 33 (2013) 1500–1513, 10.1038/jcbfm.2013.135. [PubMed: 23921899]
- [172]. Golde TE, Das P, Levites Y, Quantitative and mechanistic studies of A β immunotherapy, *CNS Neurol. Disord. Drug Targets* 8 (2009) 31–49. [PubMed: 19275635]
- [173]. Winblad B, Andreasen N, Minthon L, Floesser A, Imbert G, Dumortier T, Maguire RP, Blennow K, Lundmark J, Staufenbiel M, Orgogozo J-M, Graf A, Safety, tolerability, and antibody response of active A β immunotherapy with CAD106 in patients with Alzheimer's disease: randomised, double-blind, placebo-controlled, first-in-human study, *Lancet Neurol.* 11 (2012) 597–604, 10.1016/S1474-4422(12)70140-0. [PubMed: 22677258]
- [174]. Loeffler DA, Should development of Alzheimer's disease-specific intravenous immunoglobulin be considered? *J. Neuroinflammation* 11 (2014) 198, 10.1186/s12974-014-0198-z. [PubMed: 25476011]
- [175]. Golde TE, Open questions for Alzheimer's disease immunotherapy, *Alzheimers Res. Ther* 6 (2014) 3, 10.1186/alzrt233. [PubMed: 24393284]
- [176]. Corbyn Z, New set of Alzheimer's trials focus on prevention, *Lancet Lond. Engl* 381 (2013) 614–615.
- [177]. Holtzman DM, Carrillo MC, Hendrix JA, Bain LJ, Catafau AM, Gault LM, Goedert M, Mandelkow E, Mandelkow E-M, Miller DS, Ostrowitzki S, Polydoro M, Smith S, Wittmann M, Hutton M, Tau from research to clinical development, *Alzheimers Dement. J. Alzheimers Assoc* 12 (2016) 1033–1039, 10.1016/j.jalz.2016.03.018.

- [178]. Ballatore C, Lee VM-Y, Trojanowski JQ, Tau-mediated neurodegeneration in Alzheimer's disease and related disorders, *Nat. Rev. Neurosci* 8 (2007) 663–672, 10.1038/nrn2194. [PubMed: 17684513]
- [179]. Khanna MR, Kovalevich J, Lee VM-Y, Trojanowski JQ, Brunden KR, Therapeutic strategies for the treatment of tauopathies: hopes and challenges, *Alzheimers Dement. J. Alzheimers Assoc* 12(2016) 1051–1065, 10.1016/j.jalz.2016.06.006.
- [180]. Lee VM, Daughenbaugh R, Trojanowski JQ, Microtubule stabilizing drugs for the treatment of Alzheimer's disease, *Neurobiol. Aging* 15 (Suppl. 2) (1994)S87–89. [PubMed: 7700471]
- [181]. Min S-W, Cho S-H, Zhou Y, Schroeder S, Haroutunian V, Seeley WW, Huang EJ, Shen Y, Masliah E, Mukherjee C, Meyers D, Cole PA, Ott M, Gan L, Acetylation of tau inhibits its degradation and contributes to tauopathy, *Neuron* 67 (2010) 953–966, 10.1016/j.neuron.2010.08.044. [PubMed: 20869593]
- [182]. Rissman RA, Poon WW, Blurton-Jones M, Oddo S, Torp R, Vitek MP, LaFerla FM, Rohn TT, Cotman CW, Caspase-cleavage of tau is an early event in Alzheimer disease tangle pathology, *J. Clin. Invest* 114 (2004) 121–130, 10.1172/JCI20640. [PubMed: 15232619]
- [183]. Pedersen JT, Sigurdsson EM, Tau immunotherapy for Alzheimer's disease, *Trends Mol. Med* 21 (2015)394–402, 10.1016/j.molmed.2015.03.003. [PubMed: 25846560]
- [184]. Lista S, Toschi N, Baldacci F, Zetterberg H, Blennow K, Kilimann I, Teipel SJ, Cavedo E, Dos Santos AM, Epelbaum S, Lamari F, Dubois B, Floris R, Garaci F, Hampel H, Alzheimer Precision Medicine Initiative (APMI), Diagnostic accuracy of CSF neurofilament light chain protein in the biomarker-guided classification system for Alzheimer's disease, *Neurochem. Int* 108 (2017)355–360, 10.1016/j.neuint.2017.05.010. [PubMed: 28527630]
- [185]. Olsson B, Lautner R, Andreasson U, öhrfelt A, Portelius E, Bjerke M, Hölttä M, Rosén C, Olsson C, Strobel G, Wu E, Dakin K, Petzold M, Blennow K, Zetterberg H, CSF and blood biomarkers for the diagnosis of Alzheimer's disease: a systematic review and meta-analysis, *Lancet Neurol.* (2016), 10.1016/S1474-4422(16)00070-3.
- [186]. Daniele S, Pietrobono D, Fusi J, Iofrida C, Chico L, Petrozzi L, Gerfo AL, Baldacci F, Galetta F, Siciliano G, Bonuccelli U, Santoro G, Trincavelli ML, Franzoni F, Martini C, α -Synuclein aggregates with β -amyloid or tau in human red blood cells: correlation with antioxidant capability and physical exercise in human healthy subjects, *Mol. Neurobiol* (2017), 10.1007/s12035-017-0523-5.
- [187]. Mattsson N, Andreasson U, Zetterberg H, Blennow K, Alzheimer's disease neuroimaging initiative, association of plasma neurofilament light with neurodegeneration in patients with Alzheimer disease, *JAMA Neurol.* 74 (2017) 557–566, 10.1001/jamaneurol.2016.6117. [PubMed: 28346578]
- [188]. Mattsson N, Zetterberg H, Janelidze S, Insel PS, Andreasson U, Stomrud E, Palmqvist S, Baker D, Tan Hehir CA, Jeromin A, Hanlon D, Song L, Shaw LM, Trojanowski JQ, Weiner MW, Hansson O, Blennow K, ADNI investigators, plasma tau in Alzheimer disease, *Neurology* 87 (2016) 1827–1835, 10.1212/WNL.0000000000003246. [PubMed: 27694257]
- [189]. Deckers K, Camerino I, van Boxtel MPJ, Verhey FRJ, Irving K, Brayne C, Kivipelto M, Starr JM, Yaffe K, de Leeuw PW, Köhler S, Dementia risk in renal dysfunction: a systematic review and meta-analysis of prospective studies, *Neurology* 88 (2017) 198–208, 10.1212/WNL.0000000000003482. [PubMed: 27974647]
- [190]. Gronewold J, Klafki H-W, Baldelli E, Kaltwasser B, Seidel UK, Todica O, Volsek M, Haußmann U, Wiltfang J, Kribben A, Bruck H, Hermann DM, Factors responsible for plasma β -amyloid accumulation in chronic kidney disease, *Mol. Neurobiol* 53 (2016) 3136–3145, 10.1007/s12035-015-9218-y. [PubMed: 26019016]
- [191]. Roher AE, Esh CL, Kokjohn TA, Castaño EM, Van Vickle GD, Kalback WM, Patton RL, Luehrs DC, Dausgs ID, Kuo Y-M, Emmerling MR, Soares H, Quinn JF, Kaye J, Connor DJ, Silverberg NB, Adler CH, Seward JD, Beach TG, Sabbagh MN, Amyloid beta peptides in human plasma and tissues and their significance for Alzheimer's disease, *Alzheimers Dement. J. Alzheimers Assoc* 5 (2009) 18–29, 10.1016/j.jalz.2008.10.004.
- [192]. Bharadwaj P, Wijesekara N, Liyanapathirana M, Newsholme P, Ittner L, Fraser P, Verdile G, The link between type 2 diabetes and neurodegeneration: roles for amyloid- β , amylin, and tau proteins, *J. Alzheimers Dis. JAD* 59 (2017)421–432, 10.3233/JAD-161192. [PubMed: 28269785]

- [193]. Giuntini M, Baldacci F, Del Prete E, Bonuccelli U, Ceravolo R, Diabetes is associated with postural and cognitive domains in Parkinson's disease. Results from a single-center study, *Parkinsonism Relat. Disord* 20 (2014) 671–672, 10.1016/j.parkreldis.2014.02.016. [PubMed: 24685342]
- [194]. Salvadori E, Poggesi A, Valenti R, Pracucci G, Pescini F, Pasi M, Nannucci S, Marini S, Del Bene A, Ciolli L, Ginestroni A, Diciotti S, Orlandi G, Di Donato H, De Stefano N, Cosottini M, Chiti A, Federico A, Dotti MT, Bonuccelli U, Inzitari D, Pantoni L, VMCI-Tuscany Study Group, Operationalizing mild cognitive impairment criteria in small vessel disease: the VMCI-Tuscany Study, *Alzheimers Dement, J. Alzheimers Assoc* 12 (2016) 407–418, 10.1016/j.jalz.2015.02.010.
- [195]. Salvadori E, Poggesi A, Pracucci G, Inzitari D, Pantoni L, VMCI-Tuscany Study Group, Development and psychometric properties of a neuropsychological battery for mild cognitive impairment with small vessel disease: the VMCI-Tuscany Study, *J. Alzheimers Dis. JAD* 43 (2015) 1313–1323, 10.3233/JAD-141449. [PubMed: 25147116]
- [196]. de la Monte SM, Insulin resistance and neurodegeneration progress towards the development of new therapeutics for Alzheimer's disease, *Drugs* 77 (2017) 47–65, 10.1007/s40265-016-0674-0. [PubMed: 27988872]
- [197]. Athauda D, Foltynie T, Insulin resistance and Parkinson's disease: a new target for disease modification? *Prog. Neurobiol* 145–146 (2016) 98–120, 10.1016/j.pneurobio.2016.10.001.
- [198]. Hamed SA, Brain injury with diabetes mellitus: evidence, mechanisms and treatment implications, *Expert Rev. Clin. Pharmacol* 10 (2017) 409–428, 10.1080/17512433.2017.1293521. [PubMed: 28276776]
- [199]. Wijsekara N, Gonçalves da Silva RA, De Felice FG, Fraser PE, Impaired peripheral glucose homeostasis and Alzheimer's disease, *Neuropharmacology* (2017), 10.1016/j.neuropharm.2017.11.027, pii: S0028-3908(17)30536-1.
- [200]. Moreno-Gonzalez I, Edwards Iii G, Salvadores N, Shahnawaz M, Diaz-Espinoza R, Soto C, Molecular interaction between type 2 diabetes and Alzheimer's disease through cross-seeding of protein misfolding, *Mol. Psychiatry* 22 (2017) 1327–1334, 10.1038/mp.2016.230. [PubMed: 28044060]
- [201]. Lista S, Khachaturian ZS, Rujescu D, Garaci F, Dubois B, Hampel H, Application of systems theory in longitudinal studies on the origin and progression of Alzheimer's disease, *Methods Mol. Biol. Clifton NJ* 1303 (2016) 49–67, 10.1007/978-1-4939-2627-5_2.
- [202]. Soininen H, Solomon A, Visser PJ, Hendrix SB, Blennow K, Kivipelto M, Hartmann T, LipiDiDiet clinical study group, 24-month intervention with a specific multinutrient in people with prodromal Alzheimer's disease (LipiDiDiet): a randomised, double-blind, controlled trial, *Lancet Neurol.* 16 (2017)965–975, 10.1016/S1474-4422(17)30332-0. [PubMed: 29097166]
- [203]. Anastasiou CA, Yannakoulia M, Kosmidis MH, Dardiotis E, Hadjigeorgiou GM, Sakka P, Arampatzi X, Bougea A, Labropoulos I, Scarmeas N, Mediterranean diet and cognitive health: initial results from the Hellenic Longitudinal Investigation of Ageing and Diet, *PLoS One* 12 (2017) e0182048, 10.1371/journal.pone.0182048. [PubMed: 28763509]
- [204]. Train the Brain Consortium, Randomized trial on the effects of a combined physical/cognitive training in aged MCI subjects: the Train the Brain study, *Sci. Rep* 7 (2017)39471, 10.1038/srep39471. [PubMed: 28045051]
- [205]. Baldacci F, Lista S, Garaci F, Bonuccelli U, Toschi N, Hampel H, Biomarker-guided classification scheme of neurodegenerative diseases, *J. Sport Health Sci* 5 (4) (2016) 383–387, 10.1016/j.jshs.2016.08.007. [PubMed: 30356557]
- [206]. Cummings J, Lee G, Mortsdorf T, Ritter A, Zhong K, Alzheimer's disease drug development pipeline: 2017, *Alzheimers Dement. N. Y* 3 (2017) 367–384, 10.1016/j.trci.2017.05.002. [PubMed: 29067343]
- [207]. Jellinger KA, Attems J, Neuropathological evaluation of mixed dementia, *J. Neurol. Sci* 257 (2007) 80–87, 10.1016/j.jns.2007.01.045. [PubMed: 17324442]
- [208]. Lo AW, Ho C, Cummings J, Kosik KS, Parallel discovery of Alzheimer's therapeutics, *Sci. Transl. Med* 6 (2014), 10.1126/scitranslmed.3008228, 241cm5.

- [209]. Antoniou M, Jorgensen AL, Kolamunnage-Dona R, Biomarker-guided adaptive trial designs in phase II and phase III: a methodological review, *PLoS One* 11 (2016) e0149803, 10.1371/journal.pone.0149803. [PubMed: 26910238]
- [210]. Hess GP, Fonseca E, Scott R, Fagerness J, Pharmacogenomic and pharmacogenetic-guided therapy as a tool in precision medicine: current state and factors impacting acceptance by stakeholders, *Genet. Res* 97 (2015) e13, 10.1017/S0016672315000099.
- [211]. Baldacci F, Lucchesi C, Cafalli M, Poletti M, Ulivi M, Vedovello M, Giuntini M, Mazzucchi S, Del Prete E, Vergallo A, Nuti A, Gori S, Migraine features in migraineurs with and without anxiety-depression symptoms: a hospital-based study, *Clin. Neurol. Neurosurg* 132 (2015) 74–78, 10.1016/j.clineuro.2015.02.017. [PubMed: 25804622]
- [212]. Pariser AR, Xu K, Milto J, Coté TR, Regulatory considerations for developing drugs for rare diseases: orphan designations and early phase clinical trials, *Discov. Med* 11 (2011) 367–375. [PubMed: 21524390]
- [213]. Karran E, Hardy J, A critique of the drug discovery and phase 3 clinical programs targeting the amyloid hypothesis for Alzheimer disease, *Ann. Neurol* 76 (2014) 185–205, 10.1002/ana.24188. [PubMed: 24853080]
- [214]. Morihara T, Chu T, Ubeda O, Beech W, Cole GM, Selective inhibition of abeta42 production by NSAID R-enantiomers, *J. Neurochem* 83 (2002) 1009–1012. [PubMed: 12421374]
- [215]. Eriksen JL, Sagi SA, Smith TE, Weggen S, Das P, McLendon DC, Ozols VV, Jessing KW, Zavitz KH, Koo EH, Golde TE, NSAIDs and enantiomers of flurbiprofen target gamma-secretase and lower Abeta 42 in vivo, *J. Clin. Invest* 112 (2003) 440–449, 10.1172/JCI18162. [PubMed: 12897211]
- [216]. Galasko DR, Graff-Radford N, May S, Hendrix S, Cottrell BA, Sagi SA, Mather G, Laughlin M, Zavitz KH, Swabb E, Golde TE, Murphy MP, Koo EH, Safety, tolerability, pharmacokinetics, and Abeta levels after short-term administration of R-flurbiprofen in healthy elderly individuals, *Alzheimer Dis. Assoc. Disord* 21 (2007) 292–299, 10.1097/WAD.0b013e31815d1048. [PubMed: 18090435]
- [217]. Wilcock GK, Black SE, Hendrix SB, Zavitz KH, Swabb EA, Laughlin MA, Tarenflur bil Phase II Study investigators, Efficacy and safety of tarenflur bil in mild to moderate Alzheimer's disease: a randomised phase II trial, *Lancet Neurol.* 7 (2008) 483–493, 10.1016/S1474-4422(08)70090-5. [PubMed: 18450517]
- [218]. Green RC, Schneider LS, Amato DA, Beelen AP, Wilcock G, Swabb EA, Zavitz KH, Tarenflur bil Phase 3 Study Group, Effect of tarenflur bil on cognitive decline and activities of daily living in patients with mild Alzheimer disease: a randomized controlled trial, *JAMA* 302 (2009) 2557–2564, 10.1001/jama.2009.1866. [PubMed: 20009055]
- [219]. Li T, Huang Y, Jin S, Ye L, Rong N, Yang X, Ding Y, Cheng Z, Zhang J, Wan Z, Harrison DC, Hussain I, Hall A, Lee DHS, Lau L-F, Matsuoka Y, -secretase modulators do not induce A β -rebound and accumulation of β -C-terminal fragment, *J. Neurochem* 121 (2012) 277–286, 10.1111/j.1471-4159.2011.07560.x. [PubMed: 22035227]
- [220]. Burton CR, Meredith JE, Barten DM, Goldstein ME, Krause CM, Kieras CJ, Sisk L, Iben LG, Polson C, Thompson MW, Lin X-A, Corsa J, Fiedler T, Pierdomenico M, Cao Y, Roach AH, Cantone JL, Ford MJ, Drexler DM, Olson RE, Yang MG, Bergstrom CP, McElhone KE, Bronson JJ, Macor HE, Blat Y, Grafstrom RH, Stern AM, Seiffert DA, Zaczek R, Albright CF, Toyn JH, The amyloid-beta rise and gamma-secretase inhibitor potency depend on the level of substrate expression, *J. Biol. Chem* 283 (2008) 22992–23003, 10.1074/jbc.M804175200. [PubMed: 18574238]
- [221]. Lanz TA, Karmilowicz MJ, Wood KM, Pozdnyakov N, Du P, Piotrowski MA, Brown TM, Nolan CE, Richter KEG, Finley JE, Fei Q, Ebbinghaus CF, Chen YL, Spracklin DK, Tate B, Geoghegan KF, Lau L-F, Auperin DD, Schachter JB, Concentration-dependent modulation of amyloid-beta in vivo and in vitro using the gamma-secretase inhibitor, LY-450139, *J. Pharmacol. Exp. Ther* 319 (2006) 924–933, 10.1124/jpet.106.110700. [PubMed: 16920992]
- [222]. Gitter BD, Czilli DL, Li W, Dieckman DK, Bender MH, Nissen JS, Mabry TE, Yin T, Boggs LN, McClure DB, Little SP, Johnstone EM, Audia JE, May PC, Hyslop PA, P 4-339 Stereoselective inhibition of amyloid beta peptide secretion by LY450139, a novel functional gamma secretase inhibitor, *Neurobiol. Aging* 25 (2004) S571, 10.1016/S0197-4580(04)81897-9.

- [223]. Boggs LN, Fuson KS, Gitter BD, Czilli DL, Hyslop PA, Bender MH, Li W, Audia JE, Nissen JS, Mabry TE, Ni B, Su Y, May PC, P 1-419 In vivo characterization of LY450139, a novel, stereoselective, functional gamma-secretase inhibitor, *Neurobiol. Aging* 25 (2004) S218, 10.1016/S0197-4580(04)80731-0.
- [224]. Ness DK, Boggs LN, Hepburn DL, Gitter B, Long GG, May PC, Piroozi KS, Schafer HA, Yang Z, P 2-053 Reduced β -amyloid burden, increased C-99 concentrations and evaluation of neuropathology in the brains of PDAPP mice given LY450139 dihydrate daily by gavage for 5 months, *Neurobiol. Aging. Suppl 2* (2004) S238–S239, 10.1016/S0197-4580(04)80800-5.
- [225]. Siemers ER, Dean RA, Friedrich S, Ferguson-Sells L, Gonzales C, Farlow MR, May PC, Safety, tolerability, and effects on plasma and cerebrospinal fluid amyloid-beta after inhibition of gamma-secretase, *Clin. Neuropharmacol* 30(2007)317–325, 10.1097/WNF.0b013e31805b7660. [PubMed: 18090456]
- [226]. Bateman RJ, Munsell LY, Morris JC, Swarm R, Yarasheski KE, Holtzman DM, Human amyloid-beta synthesis and clearance rates as measured in cerebrospinal fluid in vivo, *Nat. Med* 12 (2006) 856–861, 10.1038/nm1438. [PubMed: 16799555]
- [227]. Bateman RJ, Siemers ER, Mawuenyega KG, Wen G, Browning KR, Sigurdson WC, Yarasheski KE, Friedrich SW, Demattos RB, May PC, Paul SM, Holtzman DM, A gamma-secretase inhibitor decreases amyloid-beta production in the central nervous system, *Ann. Neurol* 66 (2009) 48–54, 10.1002/ana.21623. [PubMed: 19360898]
- [228]. Doody RS, Raman R, Farlow M, Iwatsubo T, Vellas B, Joffe S, Kieburtz K, He F, Sun X, Thomas RG, Aisen PS, Alzheimer's Disease Cooperative Study Steering Committee, Siemers E, Sethuraman G, Mohs R, Semagacestat Study Group, A phase 3 trial of semagacestat for treatment of Alzheimer's disease, *N. Engl. J. Med* 369 (2013)341–350, 10.1056/NEJMoa1210951. [PubMed: 23883379]
- [229]. De Strooper B, Lessons from a failed γ -secretase Alzheimer trial, *Cell* 159 (2014) 721–726, 10.1016/j.cell.2014.10.016. [PubMed: 25417150]
- [230]. Henley DB, May PC, Dean RA, Siemers ER, Development of semagacestat (LY450139), a functional gamma-secretase inhibitor, for the treatment of Alzheimer's disease, *Expert Opin. Pharmacother* 10 (2009) 1657–1664, 10.1517/14656560903044982. [PubMed: 19527190]
- [231]. Imbimbo BP, Ottonello S, Frisardi V, Solfrizzi V, Greco A, Seripa D, Pilotto A, Panza F, Solanezumab for the treatment of mild-to-moderate Alzheimer's disease, *Expert Rev. Clin. Immunol* 8 (2012) 135–149, 10.1586/eci.11.93. [PubMed: 22288451]
- [232]. DeMattos RB, Bales KR, Cummins DJ, Dodart JC, Paul SM, Holtzman DM, Peripheral anti-A beta antibody alters CNS and plasma A beta clearance and decreases brain A beta burden in a mouse model of Alzheimer's disease, *Proc. Natl. Acad. Sci. U. S. A* 98 (2001) 8850–8855, 10.1073/pnas.151261398. [PubMed: 11438712]
- [233]. DeMattos RB, Bales KR, Cummins DJ, Paul SM, Holtzman DM, Brain to plasma amyloid-beta efflux: a measure of brain amyloid burden in a mouse model of Alzheimer's disease, *Science* 295 (2002) 2264–2267, 10.1126/science.1067568. [PubMed: 11910111]
- [234]. Seubert P, Barbour R, Khan K, Motter R, Tang P, Kholodenko D, Kling K, Schenk D, Johnson-Wood K, Schroeter S, Gill D, Jacobsen JS, Pangalos M, Basi G, Games D, Antibody capture of soluble A β does not reduce cortical A β amyloidosis in the PDAPP mouse, *Neurodegener. Dis* 5 (2008) 65–71, 10.1159/000112834. [PubMed: 18182780]
- [235]. Walker JR, Pacoma R, Watson J, Ou W, Alves J, Mason DE, Peters EC, Urbina HD, Welzel G, Althage A, Liu B, Tuntland T, Jacobson LH, Harris JL, Schumacher AM, Enhanced proteolytic clearance of plasma A β by peripherally administered neprilysin does not result in reduced levels of brain A β in mice, *J. Neurosci* 33 (2013) 2457–2464, 10.1523/JNEUROSCI.3407-12.2013. [PubMed: 23392674]
- [236]. Siemers ER, Friedrich S, Dean RA, Gonzales CR, Farlow MR, Paul SM, Demattos RB, Safety and changes in plasma and cerebrospinal fluid amyloid beta after a single administration of an amyloid beta monoclonal antibody in subjects with Alzheimer disease, *Clin. Neuropharmacol* 33 (2010) 67–73, 10.1097/WNF.0b013e3181cb577a. [PubMed: 20375655]
- [237]. Farlow M, Arnold SE, van Dyck CH, Aisen PS, Snider BJ, Porsteinsson AP, Friedrich S, Dean RA, Gonzales C, Sethuraman G, DeMattos RB, Mohs R, Paul SM, Siemers ER, Safety and

- biomarker effects of solanezumab in patients with Alzheimer's disease, *Alzheimers Dement. J. Alzheimers Assoc* 8 (2012) 261–271, 10.1016/j.jalz.2011.09.224.
- [238]. Doody RS, Thomas RG, Farlow M, Iwatsubo T, Vellas B, Joffe S, Kieburtz K, Raman R, Sun X, Aisen PS, Siemers E, Liu-Seifert H, Mohs R, Alzheimer's Disease Cooperative Study Steering Committee, Solanezumab Study Group, Phase 3 trials of solanezumab for mild-to-moderate Alzheimer's disease, *N. Engl. J. Med* 370 (2014) 311–321, 10.1056/NEJMoa1312889. [PubMed: 24450890]
- [239]. Siemers ER, Sundell KL, Carlson C, Case M, Sethuraman G, Liu-Seifert H, Dowsett SA, Pontecorvo MJ, Dean RA, Demattos R, Phase 3 solanezumab trials: secondary outcomes in mild Alzheimer's disease patients, *Alzheimers Dement. J. Alzheimers Assoc* 12(2016) 110–120, 10.1016/j.jalz.2015.06.1893.
- [240]. Palmqvist S, Mattsson N, Hansson O, Alzheimer's Disease Neuroimaging Initiative, Cerebrospinal fluid analysis detects cerebral amyloid- β accumulation earlier than positron emission tomography, *Brain J. Neurol* 139 (2016) 1226–1236, 10.1093/brain/aww015.
- [241]. Medina M, Avila J, New perspectives on the role of tau in Alzheimer's disease, Implications for therapy, *Biochem. Pharmacol* 88 (2014) 540–547, 10.1016/j.bcp.2014.01.013. [PubMed: 24462919]
- [242]. Mormino EC, Kluth JT, Madison CM, Rabinovici GD, Baker SL, Miller BL, Koeppe RA, Mathis CA, Weiner MW, Jagust WJ, Alzheimer's Disease Neuroimaging Initiative, Episodic memory loss is related to hippocampal-mediated beta-amyloid deposition in elderly subjects, *Brain J. Neurol* 132 (2009) 1310–1323, 10.1093/brain/awn320.
- [243]. Jagust WJ, Landau SM, Shaw LM, Trojanowski JQ, Koeppe RA, Reiman EM, Foster NL, Petersen RC, Weiner MW, Price JC, Mathis CA, Alzheimer's Disease Neuroimaging Initiative, Relationships between biomarkers in aging and dementia, *Neurology* 73 (2009) 1193–1199, 10.1212/WNL.0b013e3181bc010c. [PubMed: 19822868]
- [244]. Villemagne VL, Burnham S, Bourgeat P, Brown B, Ellis KA, Salvado O, Szoeki C, Macaulay SL, Martins R, Maruff P, Ames D, Rowe CC, Masters CL, Australian Imaging Biomarkers and Lifestyle (AIBL) Research Group, Amyloid β deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study, *Lancet Neurol.* 12 (2013) 357–367, 10.1016/S1474-4422(13)70044-9. [PubMed: 23477989]
- [245]. O'Bryant SE, Gupta V, Henriksen K, Edwards M, Jeromin A, Lista S, Bazenet C, Soares H, Lovestone S, Hampel H, Montine T, Blennow K, Foroud T, Carrillo M, Graff-Radford N, Laske C, Breteler M, Shaw L, Trojanowski JQ, Schupf N, Rissman RA, Fagan AM, Oberoi P, Umek R, Weiner MW, Grammas P, Posner H, Martins R, STAR-B and BBBIG working groups, Guidelines for the standardization of preanalytic variables for blood-based biomarker studies in Alzheimer's disease research, *Alzheimers Dement. J. Alzheimers Assoc* 11 (2015) 549–560, 10.1016/j.jalz.2014.08.099.
- [246]. Lista S, Zetterberg H, O'Bryant SE, Blennow K, Hampel H, Evolving relevance of neuroproteomics in Alzheimer's disease, *Methods Mol. Biol. Clifton NJ* 2017(1598) 101–115, 10.1007/978-1-4939-6952-4_5.
- [247]. Neltner JH, Abner EL, Schmitt FA, Denison SK, Anderson S, Patel E, Nelson PT, Digital pathology and image analysis for robust high-throughput quantitative assessment of Alzheimer disease neuropathologic changes, *J. Neuropathol. Exp. Neurol* 71 (2012) 1075–1085, 10.1097/NEN.0b013e3182768de4. [PubMed: 23147505]
- [248]. Dunn WD, Gearing M, Park Y, Zhang L, Hanfelt J, Glass JD, Gutman DA, Applicability of digital analysis and imaging technology in neuropathology assessment, *Neuropathol. Off. J. Jpn. Soc. Neuropathol* 36 (2016) 270–282, 10.1111/neup.12273.
- [249]. Słodkowska J, García-Rojo M, Digital pathology in personalized cancer therapy, *Stud. Health Technol. Inf* 179 (2012) 143–154.
- [250]. Mullard A, BACE inhibitor bust in Alzheimer trial, *Nat. Rev. Drug Discov* 16 (2017) 155, 10.1038/nrd.2017.43.
- [251]. Kennedy ME, Stamford AW, Chen X, Cox K, Cumming JN, Dockendorf MF, Egan M, Ereshefsky L, Hodgson RA, Hyde LA, Jhee S, Kleijn HJ, Kuvelkar R, Li W, Mattson BA, Mei H, Palcza J, Scott JD, Tanen M, Troyer MD, Tseng JL, Stone JA, Parker EM, Forman MS,

The BACE1 inhibitor verubecestat (MK-8931) reduces CNS β -amyloid in animal models and in Alzheimer's disease patients, *Sci. Transl. Med* (2016) a150, 10.1126/scitranslmed.aad9704.

- [252]. Yan R, Vassar R, Targeting the β secretase BACE1 for Alzheimer's disease therapy, *Lancet Neurol.* 13 (2014)319–329, 10.1016/S1474-4422(13)70276-X. [PubMed: 24556009]
- [253]. Kandalepas PC, Sadleir KR, Eimer WA, Zhao J, Nicholson DA, Vassar R, The Alzheimer's β -secretase BACE1 localizes to normal presynaptic terminals and to dystrophic presynaptic terminals surrounding amyloid plaques, *Acta Neuropathol. (Berl)* 126 (2013) 329–352, 10.1007/s00401-013-1152-3. [PubMed: 23820808]
- [254]. Endres K, Fahrenholz F, Lotz J, Hiemke C, Teipel S, Lieb K, Tuscher O, Fellgiebel A, Increased CSFAPPs- α levels in patients with Alzheimer disease treated with acitretin, *Neurology* 83 (2014) 1930–1935, 10.1212/WNL.0000000000001017. [PubMed: 25344383]
- [255]. Marcade M, Bourdin J, Loiseau N, Peillon H, Rayer A, Drouin D, Schweighoffer F, Désiré L, Etazolate, a neuroprotective drug linking GABA(A) receptor pharmacology to amyloid precursor protein processing, *J. Neurochem* 106(2008)392–404, 10.1111/j.1471-4159.2008.05396.x. [PubMed: 18397369]
- [256]. Vellas B, Sol O, Snyder PJ, Ousset P-J, Haddad R, Maurin M, Lemarié J-C, Désiré H, Pando MP, EHT0202/002 study group, EHT0202 in Alzheimer's disease: a 3-month randomized, placebo-controlled, double-blind study, *Curr. Alzheimer Res* 8(2011) 203–212. [PubMed: 21222604]
- [257]. Rogers K, Felsenstein KM, Hrdlicka L, Tu Z, Albayya F, Lee W, Hopp S, Miller H-J, Spaulding D, Yang Z, Hodgdon H, Nolan S, Wen M, Costa D, Blain J-F, Freeman E, De Strooper B, Vulsteke V, Scrocchi L, Zetterberg H, Portelius D, Hutter-Paier B, Havas D, Ahljianian M, Flood D, Leventhal L, Shapiro G, Patzke H, Chesworth R, Koenig G, Modulation of γ -secretase by EVP-0015962 reduces amyloid deposition and behavioral deficits in Tg2576 mice, *Mol. Neurodegener* 7 (2012) 61, 10.1186/1750-1326-7-61. [PubMed: 23249765]
- [258]. Bulic B, Ness J, Hahn S, Rennhack A, Jumpertz T, Weggen S, Chemical Biology, Molecular mechanism and clinical perspective of γ -secretase modulators in Alzheimer's disease, *Curr. Neuropharmacol* 9 (2011) 598–622, 10.2174/157015911798376352. [PubMed: 22798753]
- [259]. Liang SH, Southon AG, Fraser BH, Krause-Heuer AM, Zhang B, Shoup TM, Lewis R, Volitakis I, Han Y, Greguric I, Bush AI, Vasdev N, Novel fluorinated 8-hydroxyquinoline based metal ionophores for exploring the metal hypothesis of Alzheimer's disease, *ACS Med. Chem. Lett* 6 (2015) 1025–1029, 10.1021/acsmchemlett.5b00281. [PubMed: 26396692]
- [260]. Levenson JM, Schroeter S, Carroll JC, Cullen V, Asp E, Proschitsky M, Chung AH-Y, Gilead S, Nadeem M, Dodiya HB, Shoaga S, Mufson EJ, Tsubery H, Krishnan R, Wright J, Solomon B, Fisher R, Gannon KS, NPT088 reduces both amyloid- β and tau pathologies in transgenic mice, *Alzheimers Dement. N. Y* 2 (2016) 141–155, 10.1016/j.trci.2016.06.004. [PubMed: 29067301]
- [261]. Nordberg A, Kadir A, Andreasen N, Almkvist O, Wall A, Långström B, Zetterberg H, Correlations between Alzheimer's disease cerebrospinal fluid biomarkers and cerebral glucose metabolism after 12 months of phenserine treatment, *J. Alzheimers Dis. JAD* 47 (2015) 691–704, 10.3233/JAD-132474. [PubMed: 26401704]
- [262]. Kim J, Yoon H, Horie T, Burchett JM, Restivo JL, Rotllan N, Ramírez CM, Verghese PB, Ihara M, Hoe H-S, Esau C, Fernández-Hernando C, Holtzman DM, Cirrito JR, Ono K, Kim J, MicroRNA-33 regulates ApoE lipidation and amyloid- β metabolism in the brain, *J. Neurosci* 35 (2015) 14717–14726, 10.1523/JNEUROSCI.2053-15.2015. [PubMed: 26538644]
- [263]. Morawski M, Schilling S, Kreuzberger M, Waniek A, Jäger C, Koch B, Cynis H, Kehlen A, Arendt T, Hartlage-Rübsamen M, Demuth H-U, Roßner S, Glutaminyl cyclase in human cortex: correlation with (pGlu α amyloid- β load and cognitive decline in Alzheimer's disease, *J. Alzheimers Dis. JAD* 39 (2014) 385–400, 10.3233/JAD-131535. [PubMed: 24164736]
- [264]. Hori Y, Takeda S, Cho H, Wegmann S, Shoup TM, Takahashi K, Irimia D, Elmaleh AR, Hyman BT, Hudry E, A Food and Drug Administration-approved asthma therapeutic agent impacts amyloid β in the brain in a transgenic model of Alzheimer disease, *J. Biol. Chem* 290 (2015) 1966–1978, 10.1074/jbc.M114.586602. [PubMed: 25468905]
- [265]. Butler D, Bendiske J, Michaelis ML, Karanian DA, Bahr BA, Microtubule-stabilizing agent prevents protein accumulation-induced loss of synaptic markers, *Eur. J. Pharmacol* 562 (2007) 20–27, 10.1016/j.ejphar.2007.01.053. [PubMed: 17336290]

- [266]. Davtyan H, Bacon A, Petrushina I, Zagorski K, Cribbs DH, Ghochikyan A, Agadjanyan MG, Immunogenicity of DNA- and recombinant protein-based Alzheimer disease epitope vaccines, *Hum. Vaccines Immunother* 10 (2014) 1248–1255, 10.4161/hv.27882.
- [267]. Mandler M, Santic R, Gruber P, Cinar Y, Pichler D, Funke SA, Willbold D, Schneeberger A, Schmidt W, Mattner F, Tailoring the antibody response to aggregated A β using novel Alzheimer-vaccines, *PLoS One* 10 (2015) e0115237, 10.1371/journal.pone.0115237. [PubMed: 25611858]
- [268]. Novak P, Schmidt R, Kontsekova E, Zilka N, Kovacech B, Skrabana R, Vince-Kazmerova Z, Katina S, Fialova L, Prcina M, Parrak V, Dal-Bianco P, Brunner M, Staffen W, Rainer M, Ondrus M, Ropele S, Smisek M, Sivak R, Winblad B, Novak M, Safety and immunogenicity of the tau vaccine AADvac1 in patients with Alzheimer's disease: a randomised, double-blind, placebo-controlled, phase 1 trial, *Lancet Neurol.* 16 (2017) 123–134, 10.1016/S1474-4422(16)30331-3. [PubMed: 27955995]
- [269]. Muhs A, Hickman DT, Pihlgren M, Chuard N, Giriens V, Meerschman C, van der Auwera I, van Leuven F, Sugawara M, Weingertner M-C, Bechinger B, Greferath R, Kolonko N, Nagel-Steger L, Riesner D, Brady RO, Pfeifer A, Nicolau C, Liposomal vaccines with conformation-specific amyloid peptide antigens define immune response and efficacy in APP transgenic mice, *Proc. Natl. Acad. Sci. U. S. A* 104 (2007) 9810–9815, 10.1073/pnas.0703137104. [PubMed: 17517595]
- [270]. Theunis C, Crespo-Biel N, Gafner V, Pihlgren M, Lñpez-Deber MP, Reis P, Hickman DT, Adolfsson O, Chuard N, Ndao DM, Borghgraef P, Devijver H, Van Leuven D, Pfeifer A, Muhs A, Efficacy and safety of a liposome-based vaccine against protein Tau, assessed in tau.P301L mice that model tauopathy, *PLoS One* 8 (2013) e72301, 10.1371/journal.pone.0072301. [PubMed: 23977276]
- [271]. Sevigny J, Chiao P, Bussière T, Weinreb PH, Williams L, Maier M, Dunstan R, Salloway S, Chen T, Ling Y, O'Gorman J, Qian F, Arastu M, Li M, Chollate S, Brennan MS, Quintero-Monzon O, Scannevin RH, Arnold HM, Engber T, Rhodes K, Ferrero J, Hang Y, Mikulskis A, Grimm J, Hock C, Nitsch RM, Sandrock A, The antibody aducanumab reduces A β plaques in Alzheimer's disease, *Nature* 537 (2016) 50–56, 10.1038/nature19323. [PubMed: 27582220]
- [272]. Braczynski AK, Schulz JB, Bach J-P, Vaccination strategies in tauopathies and synucleinopathies, *J. Neurochem* 143 (2017) 467–488, 10.1111/jnc.14207. [PubMed: 28869766]
- [273]. Cynis H, Frost JL, Crehan H, Lemere CA, Immunotherapy targeting pyroglutamate-3 A β : prospects and challenges, *Mol. Neurodegener* 11 (2016) 48, 10.1186/s13024-016-0115-2. [PubMed: 27363697]
- [274]. Budni J, Garcez ML, de Medeiros J, Cassaro E, Bellettini-Santos T, Mina F, Quevedo J, The anti-inflammatory role of minocycline in Alzheimer's disease, *Curr. Alzheimer Res* 13 (2016) 1319–1329. [PubMed: 27539598]
- [275]. Alam J, Blackburn K, Patrick D, Neflamapimod: clinical phase 2b-ready oral small molecule inhibitor of p38a to reverse synaptic dysfunction in early Alzheimer's disease, *J. Prev. Alzheimers Dis* 4 (2017) 273–278, 10.14283/jpad.2017.41. [PubMed: 29181493]
- [276]. Davidson JE, Lockhart A, Amos L, Stirnadel-Farrant HA, Mooser V, Sollberger M, Regeniter A, Monsch AU, Irizarry MC, Plasma lipoprotein-associated phospholipase A2 activity in Alzheimer's disease, amnesic mild cognitive impairment, and cognitively healthy elderly subjects: a cross-sectional study, *Alzheimers Res. Ther* 4 (2012) 51, 10.1186/alzrt154. [PubMed: 23217243]
- [277]. Bernier L-P, Ase AR, Boué-Grabot É, Séguéla P, Inhibition of P2 \times 4 function by P2Y6 UDP receptors in microglia, *Glia* 61 (2013) 2038–2049, 10.1002/glia.22574. [PubMed: 24123515]
- [278]. Fernandez-Martos CM, Atkinson RAK, Chuah MI, King AE, Vickers JC, Combination treatment with leptin and pioglitazone in a mouse model of Alzheimer's disease, *Alzheimers Dement. N.Y* 3 (2017) 92–106, 10.1016/j.trci.2016.11.002. [PubMed: 29067321]
- [279]. Searcy JL, Phelps JT, Pancani T, Kadish I, Popovic J, Anderson KL, Beckett TL, Murphy MP, Chen K-C, Blalock EM, Landfield PW, Porter NM, Thibault O, Long-term pioglitazone treatment improves learning and attenuates pathological markers in a mouse model of Alzheimer's disease, *J. Alzheimers Dis. JAD* 30 (2012) 943–961, 10.3233/JAD-2012-111661. [PubMed: 22495349]

- [280]. Mandrekar-Colucci S, Landreth GE, Nuclear receptors as therapeutic targets for Alzheimer's disease, *Expert Opin. Ther. Targets* 15 (2011) 1085–1097, 10.1517/14728222.2011.594043. [PubMed: 21718217]
- [281]. Lue L-F, Yan SD, Stern DM, Walker DG, Preventing activation of receptor for advanced glycation endproducts in Alzheimer's disease, *Curr. Drug Targets CNS Neurol. Disord* 4 (2005) 249–266. [PubMed: 15975028]
- [282]. Bierhaus A, Humpert PM, Morcos M, Wendt T, Chavakis T, Arnold B, Stern DM, Nawroth PP, Understanding RAGE, the receptor for advanced glycation end products, *J. Mol. Med. Berl. Ger* 83 (2005) 876–886, 10.1007/s00109-005-0688-7.
- [283]. Folch J, Petrov D, Ettcheto M, Pedrñs I, Abad S, Beas-Zarate C, Lazarowski A, Marin M, Olloquequi J, Auladell C, Camins A, Masitinib for the treatment of mild to moderate Alzheimer's disease, *Expert Rev. Neurother* 15 (2015) 587–596, 10.1586/14737175.2015.1045419. [PubMed: 25961655]
- [284]. Bennett J, Burns J, Welch P, Bothwell R, Safety and tolerability of R(+) pramipexole in mild-to-moderate Alzheimer's disease, *J. Alzheimers Dis. JAD* 49 (2016) 1179–1187, 10.3233/JAD-150788. [PubMed: 26682692]
- [285]. Sivilia S, Lorenzini L, Giuliani A, Gusciglio M, Fernandez M, Baldassarro VA, Mangano C, Ferraro L, Pietrini V, Baroc MF, Viscomi AR, Ottonello S, Villetti G, Imbimbo BP, Calzà L, Giardino L, Multi-target action of the novel anti-Alzheimer compound CHF5074: in vivo study of long term treatment in Tg2576 mice, *BMC Neurosci.* 14 (2013) 44, 10.1186/1471-2202-14-44. [PubMed: 23560952]
- [286]. Porrini V, Lanzillotta A, Branca C, Benarese M, Parrella E, Lorenzini L, Calzà L, Flaibani R, Spano PF, Imbimbo BP, Pizzi M, CHF5074 (CSP-1103) induces microglia alternative activation in plaque-free Tg2576 mice and primary glial cultures exposed to beta-amyloid, *Neuroscience* 302 (2015) 112–120, 10.1016/j.neuroscience.2014.10.029. [PubMed: 25450955]
- [287]. Xu W, Yang Y, Yuan G, Zhu W, Ma D, Hu S, Exendin-4, a glucagon-like peptide-1 receptor agonist, reduces Alzheimer disease-associated tau hyperphosphorylation in the hippocampus of rats with type 2 diabetes, *J. Investig. Med* 63 (2015) 267–272, 10.1097/JIM.000000000000129.
- [288]. Hölscher C, Central effects of GLP-1: new opportunities for treatments of neurodegenerative diseases, *J. Endocrinol* 221 (2014) T31–T41, 10.1530/JOE-13-0221. [PubMed: 23999914]
- [289]. Ma D-L, Chen F-Q, Xu W-J, Yue W-Z, Yuan G, Yang Y, Early intervention with glucagon-like peptide 1 analog liraglutide prevents tau hyperphosphorylation in diabetic db/db mice, *J. Neurochem* 135 (2015) 301–308, 10.1111/jnc.13248. [PubMed: 26183127]
- [290]. Shah RC, Matthews DC, Andrews RD, Capuano AW, Fleischman DA, VanderLugt JT, Colca JR, An evaluation of MSDC-0160, a prototype mTOT modulating insulin sensitizer, in patients with mild Alzheimer' disease, *Curr. Alzheimer Res* 11 (2014) 564–573. [PubMed: 24931567]
- [291]. Colca JR, McDonald WG, Cavey GS, Cole SL, Holewa DD, Brightwell-Conrad AS, Wolfe CL, Wheeler JS, Coulter KR, Kilkuskie PM, Gracheva E, Korshunova Y, Trusgnich M, Karr R, Wiley SE, Divakaruni AS, Murphy AN, Vigueira PA, Finck BN, Kletzien RF, Identification of a mitochondrial target of thiazolidinedione insulin sensitizers (mTOT)–relationship to newly identified mitochondrial pyruvate carrier proteins, *PLoS One* 8 (2013) e61551, 10.1371/journal.pone.0061551. [PubMed: 23690925]
- [292]. Tong M, Deochand C, Didsbury J, de la Monte SM, T3D-959, A multi-Faceted disease remedial drug candidate for the treatment of Alzheimer's disease, *J. Alzheimers Dis. JAD* 51 (2016) 123–138, 10.3233/JAD-151013. [PubMed: 26836193]
- [293]. Fisher A, Cholinergic modulation of amyloid precursor protein processing with emphasis on M1 muscarinic receptor: perspectives and challenges in treatment of Alzheimer's disease, *J. Neurochem* 120 (Suppl. 1) (2012) 22–33, 10.1111/j.1471-4159.2011.07507.x. [PubMed: 22122190]
- [294]. Ortega-Roldan JL, Ossa F, Schnell JR, Characterization of the human sigma-1 receptor chaperone domain structure and binding immunoglobulin protein (BiP) interactions, *J. Biol. Chem* 288 (2013) 21448–21457, 10.1074/jbc.M113.450379. [PubMed: 23760505]
- [295]. Kaufman AC, Salazar SV, Haas LT, Yang J, Kostylev MA, Jeng AT, Robinson SA, Gunther EC, van Dyck CH, Nygaard HB, Strittmatter SM, Fyn inhibition rescues established memory and

synapse loss in Alzheimer mice, *Ann. Neurol* 77 (2015) 953–971, 10.1002/ana.24394. [PubMed: 25707991]

- [296]. Smith LM, Zhu R, Strittmatter SM, Disease-modifying benefit of Fyn blockade persists after washout in mouse Alzheimer's model, *Neuropharmacology* 130 (2018) 54–61, 10.1016/j.neuropharm.2017.11.042. [PubMed: 29191754]
- [297]. Atherton J, Kurbatskaya K, Bondulich M, Croft CL, Garwood CJ, Chhabra R, Wray S, Jeromin A, Hanger DP, Noble W, Calpain cleavage and inactivation of the sodium calcium exchanger-3 occur downstream of A β in Alzheimer's disease, *Aging Cell* 13 (2014) 49–59, 10.1111/accel.12148. [PubMed: 23919677]
- [298]. Morales-Corraliza J, Berger JD, Mazzella MJ, Veeranna null, Neubert TA, Ghiso J, Rao MV, Staufenbiel M, Nixon RA, Mathews PM, Calpastatin modulates APP processing in the brains of β -amyloid depositing but not wild-type mice, *Neurobiol. Aging* 33 (1125) (2012) e9–18, 10.1016/j.neurobiolaging.2011.11.023.
- [299]. Izzo NJ, Staniszewski A, To L, Fa M, Teich AF, Saeed F, Wostein H, Walko T, Vaswani A, Wardius M, Syed Z, Ravenscroft J, Mozzoni K, Silky C, Rehak C, Yurko R, Finn P, Look G, Rishton G, Safferstein H, Miller M, Johanson C, Stopa E, Windisch M, Hutter-Paier B, Shamloo M, Arancio O, LeVine H, Catalano SM, Alzheimer's therapeutics targeting amyloid beta 1–42 oligomers I: Abeta 42 oligomer binding to specific neuronal receptors is displaced by drug candidates that improve cognitive deficits, *PLoS One* 9 (2014) e111898, 10.1371/journal.pone.0111898. [PubMed: 25390368]
- [300]. Yi B, Sahn JJ, Ardestani PM, Evans AK, Scott LL, Chan JZ, Iyer S, Crisp A, Zuniga D, Pierce JT, Martin SF, Shamloo M, Small molecule modulator of sigma 2 receptor is neuroprotective and reduces cognitive deficits and neuroinflammation in experimental models of Alzheimer's disease, *J. Neurochem* 140 (2017) 561–575, 10.1111/jnc.13917. [PubMed: 27926996]
- [301]. Gray SG, Epigenetic treatment of neurological disease, *Epigenomics* 3 (2011)431–450, 10.2217/epi.11.67. [PubMed: 22126204]
- [302]. Yao J, Zhao L, Mao Z, Chen S, Wong KC, To J, Brinton RD, Potentiation of brain mitochondrial function by S-equol and R/S-equol estrogen receptor β -selective phytoSERM treatments, *Brain Res.* 2013 (1514) 128–141, 10.1016/j.brainres.2013.02.021.
- [303]. Morgan TE, Finch CE, Astrocytic estrogen receptors and impaired neurotrophic responses in a rat model of perimenopause, *Front. Aging Neurosci* 7 (2015) 179, 10.3389/fnagi.2015.00179. [PubMed: 26483679]
- [304]. Simmons DA, Knowles JK, Belichenko NP, Banerjee G, Finkle C, Massa SM, Longo FM, A small molecule p75NTR ligand, LM11A-31, reverses cholinergic neurite dystrophy in Alzheimer's disease mouse models with mid- to late-stage disease progression, *PLoS One* 9 (2014) e102136, 10.1371/journal.pone.0102136. [PubMed: 25153701]
- [305]. Irwin RW, Solinsky CM, Loya CM, Salituro FG, Rodgers KE, Bauer G, Rogawski MA, Brinton RD, Allopregnanolone preclinical acute pharmacokinetic and pharmacodynamic studies to predict tolerability and efficacy for Alzheimer's disease, *PLoS One* 10 (2015) e0128313, 10.1371/journal.pone.0128313. [PubMed: 26039057]
- [306]. Boland K, Moschetti V, Dansirikul C, Pichereau S, Gheyle L, Runge F, Zimdahl-Gelling H, Sand M, A phase I, randomized, proof-of-clinical-mechanism study assessing the pharmacokinetics and pharmacodynamics of the oral PDE9A inhibitor BI 409306 in healthy male volunteers, *Hum. Psychopharmacol* 32 (2017), 10.1002/hup.2569.
- [307]. Weinreb O, Amit T, Bar-Am O, Youdim MBH, Ladostigil: a novel multimodal neuroprotective drug with cholinesterase and brain-selective monoamine oxidase inhibitory activities for Alzheimer's disease treatment, *Curr. Drug Targets* 13 (2012) 483–494. [PubMed: 22280345]
- [308]. Rinne JO, Wesnes K, Cummings JL, Hakulinen P, Hallikainen M, Hänninen J, Murphy M, Riordan H, Scheinin M, Soininen H, Rouru J, Tolerability of ORM-12741 and effects on episodic memory in patients with Alzheimer's disease, *Alzheimers Dement. N. Y. N* 3 (2017) 1–9, 10.1016/j.trci.2016.11.004.
- [309]. de Jong IEM, Mórk A, Antagonism of the 5-HT₆ receptor – Preclinical rationale for the treatment of Alzheimer's disease, *Neuropharmacology* 125 (2017)50–63, 10.1016/j.neuropharm.2017.07.010. [PubMed: 28711518]

- [310]. Calhoun A, Ko J, Grossberg GT, Emerging chemical therapies targeting 5-hydroxytryptamine in the treatment of Alzheimer's disease, *Expert Opin. Emerg. Drugs* 22 (2017) 101–105, 10.1080/14728214.2017.1293651. [PubMed: 28253832]
- [311]. Wicke K, Haupt A, Bespalov A, Investigational drugs targeting 5-HT6 receptors for the treatment of Alzheimer's disease, *Expert Opin. Investig. Drugs* 24 (2015) 1515–1528, 10.1517/13543784.2015.1102884.
- [312]. Nirogi R, Shinde A, Kambhampati RS, Mohammed AR, Saraf SK, Badange RK, Bandyala TR, Bhatta V, Bojja K, Reballi V, Subramanian R, Benade V, Palacharla RC, Bhyrapuneni G, Jayarajan P, Goyal V, Jasti V, Discovery and development of 1-[(2-bromophenyl)sulfonyl]-5-methoxy-3-[(4-methyl-1-piperazinyl)methyl]-1H-indole dimesylate monohydrate (SUVN-502): a novel, potent, selective and orally active serotonin 6 (5-HT6) receptor antagonist for potential treatment of Alzheimer's disease, *J. Med. Chem* 60(2017) 1843–1859, 10.1021/acs.J.Med.chem.6b01662. [PubMed: 28212021]
- [313]. Arnsten AFT, Wang M, Targeting prefrontal cortical systems for drug development: potential therapies for cognitive disorders, *Annu. Rev. Pharmacol. Toxicol* 56 (2016) 339–360, 10.1146/annurev-pharmtox-010715-103617. [PubMed: 26738476]

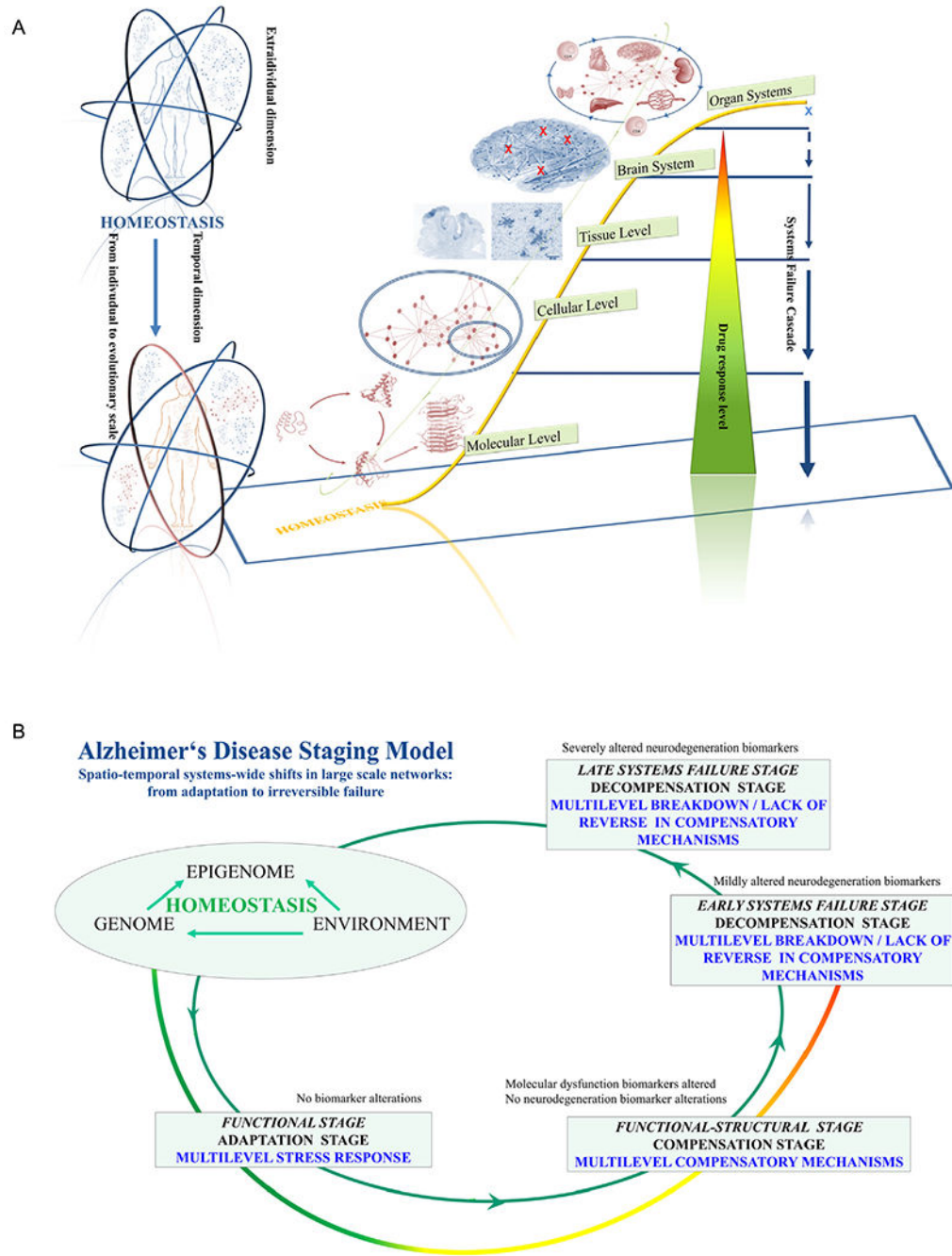


Fig. 1. (A) Trajectory of pathophysiological mechanisms across the continuum of systems multi-scale hierarchical self-organization, from systems homeostasis to systems failure: conceptual basis for molecular pathway-based therapies. The preservation of human organism homeostasis is strictly related to the interactions between human systems factors, i.e. genome/epigenome and ecosystem factors, i.e. environment (the circles). Such interactions shows a non-linear fashion with complex dynamic changes over time that are essential at the individual level for adaptation and

survival of the single organism to a certain ecosystem and at extra-individual level for adaptive (genetic) evolutionary transitions finally resulting in the trans-generational process of natural selection. For instance, the impact of a genetic mutation on a single organism may lead to wide-ranging severe maladaptive effects even though from an evolutionary trans-generational perspective this may represent a primary driver for optimized survival and reproduction. Therefore, adaptive responses are differently distributed in space and time scales, aimed at different key roles consistently with the individual, extra-individual and the trans-generational level.

Unrevealing the spatial-temporal coordinates of multilevel adaptive events across human systems (from molecular level to system level) and between these and ecosystem will uncover key notions essential for the comprehensive understanding of complex disease and at an higher level of complexity to achieve a unified theory of genetic adaptation leading to evolution. Thus, an individual vulnerability to stressors exists with an individual threshold of anti-stress response activation and failure.

The non-linear orange-shaped line represents the entire *spectrum* of pathophysiological mechanisms across all systems levels, during the course and progression of disease. Such alterations originate from initial adaptation processes leading through triggers, drivers, thresholds to a point of decompensation at both structural and functional level. The green circle surrounding the five levels represents the marked interplay among the different hierarchical self-organized systems levels. Such interactions support the hypothesis that the initial loss of homeostasis might originate and occur at every level taking into account that a single level potentially affects the whole dynamic interrelated system and, therefore, initially or ultimately the entire affected organism.

The **molecular level** shows aberrant conformational states of proteins and dysregulated molecular pathways, including: post-translational modifications, inefficient autophagic mechanisms, dysfunction of membrane dynamics. The **cellular level** originates from the sum of a number of distinct and/or interrelated aberrant molecular pathways. This has a negative impact on anti-stress responses with a subsequent overall impairment of cytoprotective and homeostatic mechanisms. The **tissue level** presents a substantial loss of structural and functional organization induced by certain categories of cells. At **brain system level**, aberrant neural oscillatory, altered metabolic, blood-flow and oxygenation activities might successively or simultaneously occur across different brain system networks, thus affecting different network integration processes and the whole functioning of the system. Therefore, brain-wide shifts in large scale network functioning allow a spatial and temporal processing resources redistribution to cope with stressors. Such hypothetical model can explain how pathophysiological alterations at the brain system level may precede, support and impact abnormal upstream to downstream molecular and cellular pathways. The **organ systems level** represents an enormous and most complex interplay among several networks of different body systems including brain. The existence of many cross-links-talks between CNS and the periphery might account for the hypothesis that brain diseases can originate or be substantially related to peripheral failure. The idea of an isolated brain disease has to be critically assessed in view of the organ systems level.

The colored pyramid represents potential outcome of effective treatment, the potential drug response at each level (from green to red and from the base to the peak there is a decreasing amplitude of effect). The arrows explain the likelihood to restore compensatory mechanisms

(i.e. disease-modifying effect) at the single level; the thicker the arrow is, the higher is the chance that the treatment is effective. The “x” positioned in correspondence of the organ systems failure indicates a hypothetical “point of no return” (pathophysiological irreversibility threshold) without any significant possibility for the drug to reverse, stop or modify the disease dynamic and progression. *Abbreviations:* CNS, central nervous system. (B) Hypothetical model of spatio-temporal systems-wide shifts in large scale networks along the continuum of AD pathophysiological processes: from adaptation to irreversible failure. Organisms are made of systems which are entities consisting in hierarchically self-organized levels with increasing structural complexity resulting in different emerging properties. Multilevel systems are strictly and dynamically interconnected through feedback and cross-talking mechanisms. As a consequence, spatial selective network activation from molecular pathways to systems large scale networks as well as time-dependent cascade of activation can allow to achieve the most effective output to copy with stressors. This, in turn is aimed to maintain homeostasis a dynamic *equilibrium* resulting from the dynamic interaction between genome, epigenome and environment. The regulation of several processes at multilevel of complexity from gene expression to cellular cycle to tissue repair and system-wide network activation has different time delays (time scale) according to the system (space scale). Thus, spatio-temporal systems-wide shifts in large scale network functioning are essential to reallocate processing resources fundamental for adaptation. The understanding of how to measure and possibly control space and time scaled adaptive and compensatory responses occurring during complex polygenic diseases with non-linear pathophysiology, as AD, will represent a crucial step for achieving the capability to effectively modify disease. Biomarkers will guide in exploring how the space and time dimensions are mechanistic involved in complex disease as AD.

Functional Stage – Adaptation Stage – Multilevel Stress Response: from metabolic reconfiguration to functional switch in cellular/tissue/systems network activity aimed to copy with different stressors/pathophysiological processes.

Functional-Structural Stage – Compensation Stage – Multilevel Compensatory Mechanism: structural and functional dynamically balancing one another in order to copy with different pathophysiological processes.

Early Systems Failure Stage – Decompensation Stage – Multilevel Breakdown/Lack of Reverse in Compensatory Mechanisms: initial and progressive loss of physiological interactions and pathophysiological compensations across multilevel systems network.

Late Systems Failure Stage – Decompensation Stage – Multilevel Breakdown/Lack of Reverse in Compensatory Mechanisms: progressed loss of physiological and pathophysiological simultaneous interactions between multilevel systems network. From the first stage to the third stage there is a decreasing chance to restore homeostatic condition (as highlighted by the colors from green to red). No option to recover homeostasis at the last stage.

Abbreviations: AD, Alzheimer’s disease.

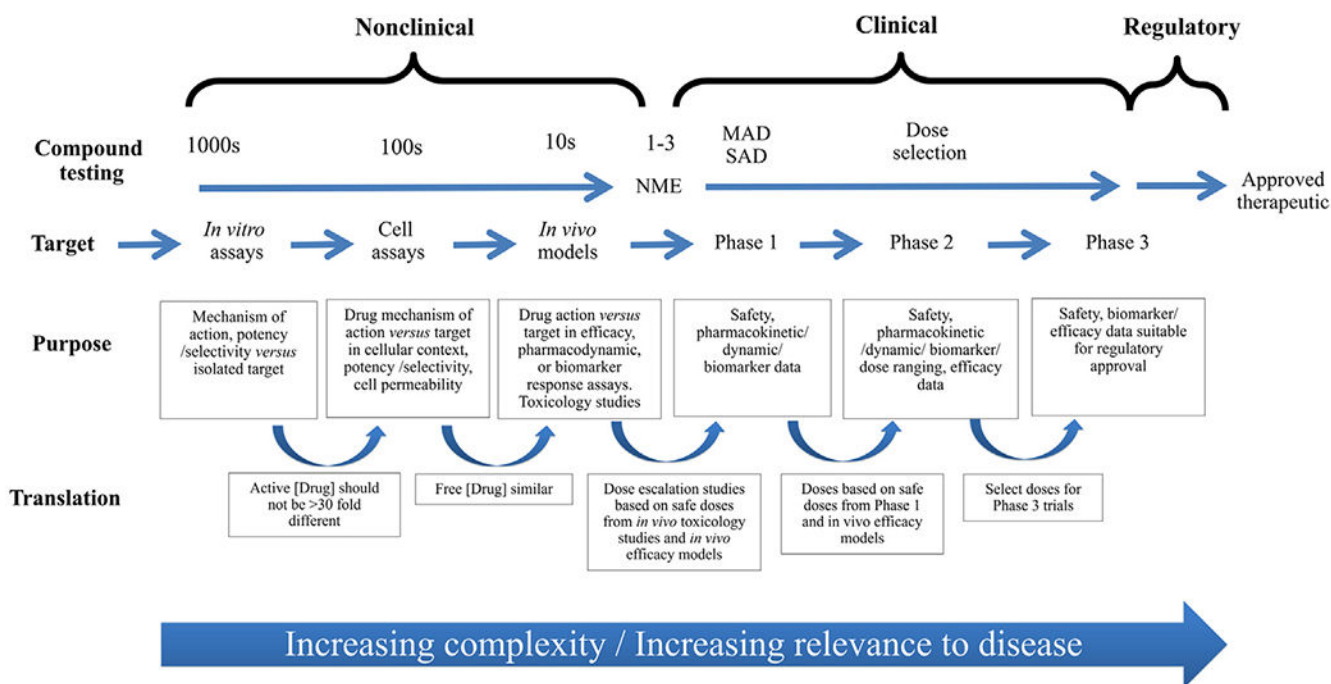
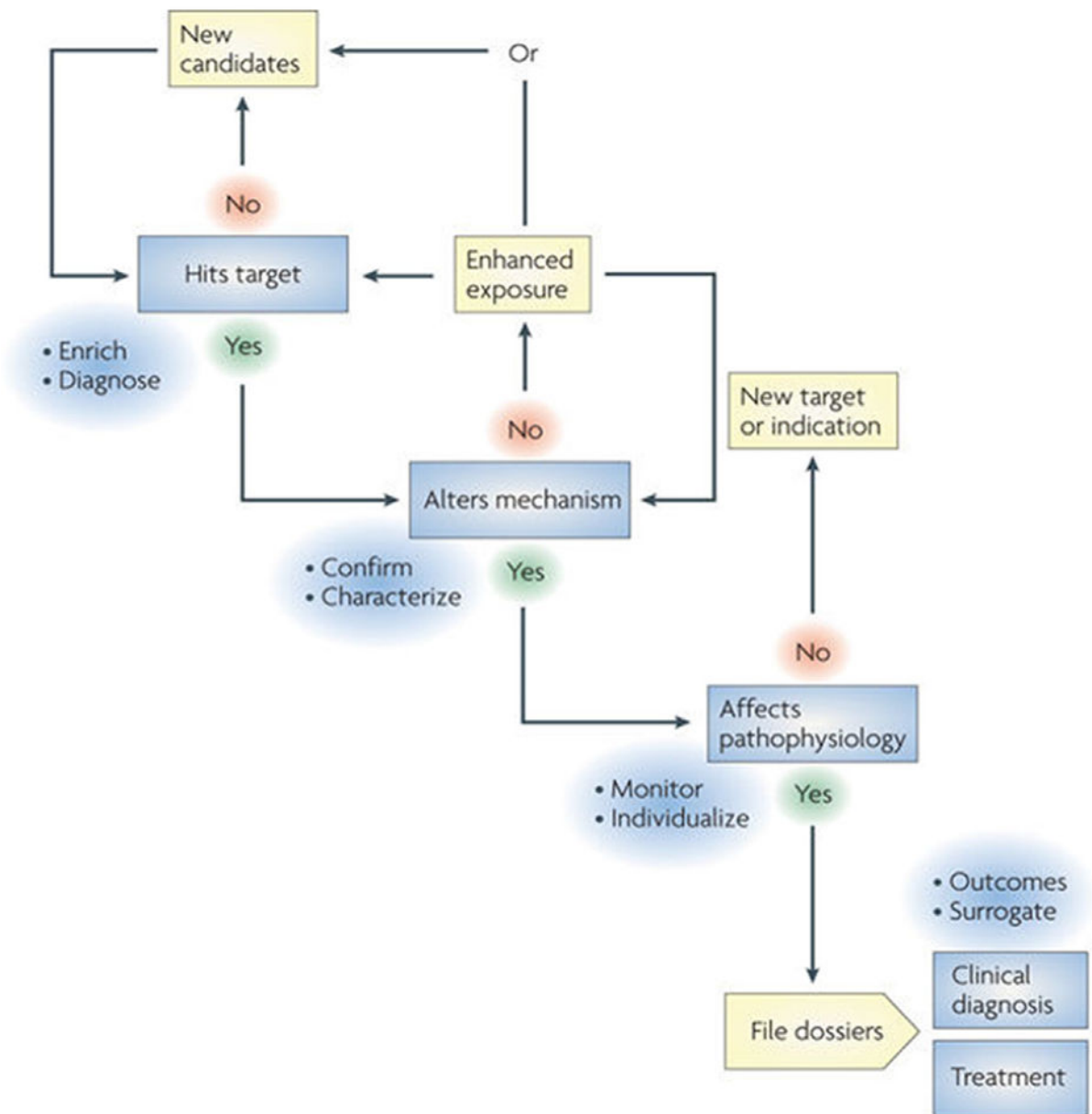


Fig. 3.

Drug discovery programs workflow. Many drug discovery programs progress through a logical sequence where the findings from one type of experiment inform the next step. Significant confidence is generated in programs where the data generated within each phase are concordant with subsequent phases. Programs that lack this translational quality are subject to increasing risk of failure.

Abbreviations: MAD, Multiple Ascending Dose; NME, New Molecular Entity; SAD, Single Ascending Dose.

Adapted from Karran E, Hardy J. "A critique of the drug discovery and phase 3 clinical programs targeting the amyloid hypothesis for Alzheimer disease." *Ann Neurol*. 2014 Aug;76(2):185-205. doi: [10.1002/ana.24188](https://doi.org/10.1002/ana.24188). Copyright © 2014 Wiley. Reprinted with permission from Wiley.



Nature Reviews | Drug Discovery

Fig. 4.

The four categories of biomarker: target, mechanism, pathophysiological, and diagnostic. Biomarkers can be categorized into four groups on the basis of their contribution to business, regulatory and clinical decision-making. Clinical decision-making can be further divided into clinical research and patient care diagnostic subcategories. The objective is to use biomarkers as early as possible in the drug development process.

– The initial step is to confirm that **a test compound hits the target** and to quantify the extent to which it does so. Next is to test three concepts in logical sequence.

- First, that hitting this target **alters the pathophysiological mechanism**.
 - Second, that altering this mechanism **affects the pathophysiology**.
 - Third, that **affecting pathophysiology** predictably improves the clinical status of the patients.
 - Biomarkers qualified to confirm the presence of the target and or extent to which the drug candidate hits the target may be validated later as diagnostic tests for early detection or diagnosis of Alzheimer’s disease (when that target is expressed differentially between healthy and diseased states).
 - Biomarkers qualified for confirming and quantifying mechanistic effects may be validated later as diagnostic tests to inform choice of therapeutic regimen, either in choice of drug or initial dosing regimen.
 - Biomarkers qualified for longitudinal quantification of patient response in terms of clinically relevant pathophysiology, may be validated later as diagnostic tests for monitoring and individualization of a therapeutic regimen.
 - Biomarkers qualified for either monitoring or individualization of therapy on clinically relevant pathophysiology may also serve as surrogate end points to support regulatory decision-making. In addition, they can be used to ensure appropriateness of use, and as quantifiers of clinical outcomes to support reimbursement decisions.
- From Hampel H et al. “Biomarkers for Alzheimer’s disease: academic, industry and regulatory perspectives.” *Nat Rev Drug Discov.* 2010 Jul;9(7):560-574. doi: [10.1038/nrd3115](https://doi.org/10.1038/nrd3115). Copyright © 2010 Springer Nature. Reprinted with permission from Springer Nature.

Table 1

Ongoing and completed clinical trials categorized by molecular targets and mechanisms of action.

A) BACE1 inhibitors					
Category	Name (Synonyms)	Company	Target type (Molecule type)	MoA – Biochemical features [Administration route]	Stage ICB Phase
BACE1 inhibitors [250–253]	AZD3293 (LY3314814, Lanabecestat)	AstraZeneca, Eli Lilly & Co.	Amyloid (SmallIM)	Selective inhibitor of BACE1 – It cleaves APP in its ectodomain, generating the amino-(N) terminus of A β and the membrane bound carboxyl-terminal fragment C99 from which γ -secretase will generate neurotoxic A β oligomers	MCI/MD aPET/CSF II-III (Recruiting)
	Verubecestat (MK-8931-009)	Merck	Amyloid (SmallIM)	(Binding of aspartyl protease BACE1 through a competitive and reversible non-covalent interactions with the active site preserving lipophilic profile useful to cross the BBB) [Oral]	MCI aPET/CSF III (Recruiting)
	CNP520	Amgen, Inc., Novartis Pharmaceuticals Corporation	Amyloid (SmallIM)		AaR α 4* II-III (Recruiting)
	JNJ-54861911	Janssen, Shionogi Pharma	Amyloid (SmallIM)		AaR aPET/CSF II-III (Recruiting)
	Elenbecestat (E2609)	Biogen, Eisai Co., Ltd.	Amyloid (SmallIM)		MCI/MD aPET II (Recruiting)
B) α -secretase modulators					
Category	Name (Synonyms)	Company	Target type (Molecule type)	MoA – Biochemical features [Administration route]	Stage ICB Phase
α -secretase modulators	Acitretin (Soriatane, Neotigason, RO 101670) [254]	Actavis, Allergan plc	Amyloid (SmallIM)	It stimulates the “non-amyloidogenic pathway” of α -secretase ADAM-10, that catalyzes the shedding of the ectodomain of APP in preventing neurotoxic A β oligomers formation and producing a neuroprotective-neurotrophic APP _{S-α} . It is a synthetic retinoic acid receptor agonist that binds RAR/RXR heterodimers which is a ligand-controlled transcription factors. Thus, it increases gene expression of α -secretase ADAM-10 [Oral]	MD n.a. II-III (Unknown)
	EHT 0202 (Etazolate)[255,256]	ExonHit Therapeutics	Amyloid/Inflammation/Transmission (SmallIM)	Selective GABA-A receptor modulator and PDE4 inhibitor by which it increases brain APP _{S-α} levels. PDE4 hydrolyzes cAMP, inhibiting the cAMP/pCREB/BDNF signaling pathway substantially involved in several pro-inflammatory responses (cytokines release, microglia activation) cAMP functionally modulates the ionotropic GABA-A receptors that potentiate the chloride efflux resulting in the depolarization of the plasma membrane followed by a rise in intracellular calcium. This, in turn, leads to the restoring of calcium homeostasis and preservation of the mitochondrial function with reduction of oxidative stress and a neuroprotective effect. In addition, GABA-A receptors have been linked to sAPP α increased production <i>via</i> ADAM10 increased expression [Oral]	MD n.a. II (Completed)
γ -secretase inhibitors or modulators	EVP-0962 (EVP 0015962)	FORUM Pharmaceuticals Inc.	Amyloid (SmallIM)	MoA not fully clear – Selective decrease of A β 1–42 production with an overall shift in the A β 1–42/A β 1–40 ratio, increasing A β 1–38 (non-toxic fragment) without affecting the total A β load as well as Notch	MCI/MD n.a. II (Completed)

A) BACE1 inhibitors						
Category	Name (Synonyms)	Company	Target type (Molecule type)	MoA – Biochemical features [Administration route]	Stage ICB Phase	
	NIC5-15	VA Office of Research and Development		receptor processing (whose suppression is associated to systemic toxicity) [257,258] γ-secretase modulators may induce subtle conformational changes of PSEN, the core catalytic site of the γ-secretase complex NIC5-15 is pinitol, a natural cyclic sugar alcohol, that may even have a potential effect as an insulin-sensitizer [Oral]	MD n.a. II (Completed)	
D) Aβ and or Tau aggregation/accumulation inhibitors						
Category	Name (Synonyms)	Company	Target type (Molecule type)	MoA – Biochemical features [Administration route]	Stage ICB Phase	
Aβ and or Tau aggregation/accumulation inhibitors	PBT2	Prana Biotechnology Limited	Amyloid (SmallM)	New generation metal chelator compound promotes cellular Zn and Cu uptake, restoring ions cellular homeostasis and promoting antioxidants cytoprotective signaling. Since cerebral Aβ plaques contain Cu and Zn ions which are involved in the Aβ oligomer aggregation, and chemical stabilization of plaque itself. Thus, the drug contributes to degrade and prevent ions-facilitated Aβ oligomers aggregation and toxicity [259] [Oral]	MD n.a. II (Completed)	
	NPT088	Proclara Biosciences, Inc.	Amyloid/Tau (?) (SmallM)	Ig-GAIM fusion protein: active fragment of g3p and human IgG1-Fc reducing the aggregation of misfolding Aβ and its precipitation in plaques. Possible MoA similar to that of Tau [260], [IV]	MD aPET I (Recruiting)	
	Posiphen (R-phenserine, (-)-Phenserine)	QR Pharma Inc.	Amyloid/α-sin/Transmission (SmallM)	It potentiates the binding of IRE to IRP1 and, in turn, the activity of the latter. IRP1 controls Fe- dependent translational processes of APP synthesis It reduces the production of Aβ, C31 and N-APP that are neurotoxic APP-derived fragments. It blocks neural SNCA mRNA translation Potential anti-neuroinflammatory and anti BACE1 mechanisms. It acts also as a selective, non-competitive AChEi enhancing cholinergic synapse [Oral]	MCI/MD CSF I-II (Recruiting) [261]	
	ISIS 814907 [262]	Ionis Pharmaceuticals, Inc.	Tau (SmallM)	Antisense oligo-nucleotides (15–25 nucleotides) that reduces brain levels of neurotoxic aggregating tau proteins by knocking-down Tau mRNA transcription. by binding it to a specific target and causing its destruction by activating the nuclear enzyme RNA hydrolases [IC]	MD CSFII (Recruiting)	
	PQ912 [263]	Probiodrug AG	Amyloid/Enzyme (SmallM)	Inhibitor of glutamyl cyclase (metalloenzyme upregulated in AD) that generates pyroglutamil-Aβ, a hydrophobic peptidases-resistant Aβ fragment, with high intrinsic toxicity and also a major component of Aβ plaques [Oral]	MCI/MD CSF II (Recruiting)	
	ALZT-OP1 (Cromolyn sodium, Intal) [264]	AZTherapies, Inc.	Amyloid (SmallM)	Cromolyn is a flavonoid derivate that inhibits Aβ aggregation into fibrils by binding Aβ monomers with hydroxyl substituents at specific sites. It may potentially act as γ-secretase modulator [Oral]	MCI/MD CSF III (Recruiting)	
	LMTM(TRx0237, LMT-X, Methylene Blue)	TauRx Therapeutics Ltd	Tau (SmallM)	It promotes the oxidation of the two cysteine residues within 4-R tau by a redox cycling mechanism, thus preventing tau aggregation or dissolving its existing aggregates [Oral]	MD – III (Completed)	

A) BACE1 inhibitors					
Category	Name (Synonyms)	Company	Target type (Molecule type)	MoA – Biochemical features [Administration route]	Stage ICB Phase
	TPI 287[265]	University of California, San Francisco	Tau (SmallM)	A tubulin-binding (at beta-tubulin site) acting as microtubule-stabilizer, thus reducing synaptic dysfunction and neuronal loss. [IV]	MD CSF. I (Not recruiting)
E) Immunotherapy (active/passive)					
Category	Name (Synonyms)	Company	Target type (Molecule type)	MoA – Biochemical features [Administration route]	Stage ICB Phase
Immunotherapy (active)	Affitope AD02 [173,266,267]	AFFIRIS AG	Amyloid (SmallM)	Six amino acids peptides that mimic the N-terminus of A β , inducing Th2-type immune response with release of anti-Tau Ab (without prominent Th1 cell activity), with subsequent Fc receptor-mediated phagocytosis and intracellular degradation [IV]	MD n.a. II (Completed)
	AADvac-1	Axon Neuroscience SE	Tau (SmallM)	An Axon peptide 108 conjugated to a KLH and aluminum hydroxide adjuvant. It is a synthetic peptide derived from amino acids 294–305 of the truncated Tau sequence inducing Th2-type immune response with release of anti-Tau Ab (without prominent Th1 cell activity), with subsequent Fc receptor-mediated phagocytosis and intracellular degradation [268] [SC]	MD n.a. I (Completed)
	CAD106 [173]	Novartis Pharmaceuticals Corporation	Amyloid (SmallM)	Multiple copies of the short A β 1–6 peptide derived from the N-terminal (B cell epitope) of A β , inducing Th2-type immune response with release of anti- A β -Ab (without prominent Th1 cell activity) binding to A β oligomers or fibrils with subsequent Fc receptor-mediated phagocytosis and intracellular degradation [IM]	AaR <i>et al</i> : homozygous I-I-III (Recruiting) MD n.a. II (Completed)
	ACI-24 (Pal1-15 acetate salt)	AC Immune SA	Amyloid (SmallM)	A β -liposomal preparation: Antigen A β 1–15 anchored on the liposomal surface, by a palmitoylated lysine tandem at each end of the peptide. It elicits A β -IgG (without prominent Th1 cell activity) by binding to A β oligomers or fibrils with following Fc receptor-mediated phagocytosis and intracellular degradation [269], [SC]	MD n.a. I-II (Recruiting)
	UB-311	United Biomedical	Amyloid (SmallM)	Equimolar mixture of two synthetic peptides, consisting of highly active UBITh [®] helper T-cell epitopes, coupled to the A β 1–14 peptide. It elicits A β -Ab (without prominent Th1 cell activity) by binding to A β oligomers or fibrils with subsequent Fc receptor mediated phagocytosis and intracellular degradation It is a proprietary vaccine delivery system (CpG oligonucleotide [159] [SC])	MD n.a. I (Completed)
	ACI-35	AC Immune SA, Janssen	Tau (SmallM)	Tau-liposomal preparation: a synthetic peptide (16 amino acids) corresponding to human protein tau sequence 393–408, with phosphorylated residues S396 and S404 derivatized with two palmitic acid chains at each terminus to enable integration into liposomes It elicits Tau-Ab without prominent Th1 cell activity [270] [SC]	MD n.a. I-II (Recruiting)
Immunotherapy (passive)[206,271–273]	Aducanumab (BIIB037)	Biogen	Amyloid (monoclonal antibody)	Human monoclonal IgG1; it binds aggregated forms of A β at the N-terminus epitope (residues 3–6) (soluble oligomers and insoluble fibrils only) and it promotes Fc-mediated microglial phagocytosis [IV]	MCI/MD aPET/CSF III (Recruiting)

A) BACE1 inhibitors					
Category	Name (Synonyms)	Company	Target type (Molecule type)	MoA – Biochemical features [Administration route]	Stage ICB Phase
	Crenezumab (MABT5102A, RG7412)	Hoffmann-La Roche	Amyloid (monoclonal antibody)	Human monoclonal IgG4; it binds oligomeric aggregated forms of A β and plaques as well with high affinity, and monomeric A β with low affinity. It promotes Fc-mediated microglial phagocytosis [IV]	MCI/MD aPET/CSF III (Recruiting)
	Gantenerumab (RO4909832, RG1450)	Hoffmann-La Roche	Amyloid (monoclonal antibody)	Human conformational monoclonal IgG1; it binds A β fibrils encompassing both N-terminal and central amino acids of A β and promotes Fc-mediated microglial phagocytosis [SC]	MCI/MD aPET/CSF III (Active, not Recruiting)
	Solanezumab (LY2062430)	Eli Lilly & Co.	Amyloid (monoclonal antibody)	Human monoclonal IgG1; it binds only soluble monomeric A β , sequestering and promoting Fc-mediated microglial phagocytosis [IV]	AaR aPET III (Recruiting)
	BAN2401	Biogen, Eisai Co., Ltd.	Amyloid (monoclonal antibody)	Human monoclonal IgG1; it binds selectively to large, soluble A β protofibrils and enhances their clearance promoting Fc-mediated microglial phagocytosis [IV]	MCI/MD aPET/CSF II (Active, not recruiting)
	Gamunex	Grifols Biologicals Inc.	Amyloid (monoclonal antibody)	Off-label used human Ig. It binds to several forms of A β and promotes Fc-mediated microglial phagocytosis [IV]	MD – II-III (Recruiting)
	AAB-003 (PF-05236812)	Pfizer, Janssen Alzheimer Immunotherapy Research & Development, LLC	Amyloid (monoclonal antibody)	Humanized monoclonal IgG1; it binds to fibrillar and soluble A β and promotes Fc-mediated microglial phagocytosis (a derivative of Bapineuzumab) [IV]	MD – I (Completed)
	LY3002813 (N3pG-A β mAb)	Eli Lilly & Co.	Amyloid (monoclonal antibody)	Humanized mE8-IgG2a Ab that binds to A β (p3-42), a pyroglutamate form of A β that is aggregated in plaques. It finally promotes Fc-mediated microglial phagocytosis [IV]	MCI aPET I (Not yet recruiting)
	SAR228810	Sanofi	Amyloid (monoclonal antibody)	Humanized monoclonal IgG1 binds to soluble protofibrillar and fibrillar A β and promotes Fc-mediated microglial phagocytosis [IV]	MD – I (Completed)
	MEDI1814	Eli Lilly & Co.	Amyloid (monoclonal antibody)	Human monoclonal IgG1; it binds only to soluble monomeric A β , sequestering and removing them [IV]	MD – I (Completed)
	LY3303560	Eli Lilly & Co.	Tau (monoclonal antibody)	Monoclonal Ab inhibiting Tau protein (MoA not disclosed by Sponsor) [SC]	MCI/MD aPET I (Recruiting)
F) Neuroinflammation modulators					
Category	Name (Synonyms)	Company	Target type (Molecule type)	MoA – Biochemical features [Administration route]	Stage ICB Phase
Neuroinflammation modulators	Sargramostim (Leukine®, GM-CSF)	University of Colorado, Denver	Microglia (SmallM)	Synthetic form of GM-CSF; it might increase phagocytosis of pathogenic protein deposits, activates microglia without increasing microglial release of pro-inflammatory cytokines [SC]	MD aPET II (Active, not recruiting)

A) BACE1 inhibitors

Category	Name (Synonyms)	Company	Target type (Molecule type)	MoA – Biochemical features [Administration route]	Stage ICB Phase
	Minocycline	Huntington Medical Research Institutes	Microglia/neurons (SmallM)	Tetracycline derivative capable of crossing the BBB; it modulates the up-regulation of iNOS and COX2, reducing pro-inflammatory response and astrocytes overactivity [274] [Oral]	MCI/MD – II (Completed)
	Neflamapimod (VX-745)	EIP Pharma, LLC	Microglia/neurons (SmallM)	It inhibits p38MAPK α , expressed in microglia and neurons. It promotes the shift of microglial activation from a pro-inflammatory to a phagocytic state, thus improving mitochondrial function, reduction of tau hyperphosphorylation, and as a consequence synaptic transmission[275]. [Oral]	MCI/MD – II (Completed)
	Rilapladib (SB-659032)	GlaxoSmithKline (GSK)	Microglia/macrophages (SmallM)	Potent and selective inhibitor of Lp-PLA2 actively secreted by monocyte-derived macrophages, T lymphocytes, and mast cells. It produces pro-inflammatory factors affecting brain microvascular endothelial cells [276] [Oral]	MCI/MD – II (Completed)
	GC 021109	GliaCure	Microglia (SmallM)	It binds the microglial P2Y6 receptor which activation leads to the synthesis and release of pro-inflammatory cytokines. In addition, purinergic signals are thought to be fundamental in shift of in microglia to an activated phenotype [277]. [Oral]	MD – I (Completed)
	Pioglitazone (AD4833)	Takeda Pharmaceutical Company, Zinfandel Pharmaceuticals Inc.	Microglia/neurons (SmallM)	PPAR γ agonists modulate, at a transcriptional level, the microglial response to A β plaques deposition increasing A β phagocytosis and decreasing cytokine release It blocks the NF- κ B-dependent gene expression, thus inhibiting multiple inflammatory pathways Effect on insulin-resistance potentially underlying AD PPAR γ regulate gene expression by forming heterodimers with RXRs and by binding to a promoter thus reducing BACE1 gene expression [278–280] [Oral]	MCI – III (Recruiting)
	Azeliragon (PF-04494700, TTP488)	Pfizer, TransTech Pharma, Inc., vTv Therapeutics LLC	Microglia/astrocytes (SmallM)	Inhibitor of RAGE (upregulated in AD astrocytes and microglial cells) that binds to AGEs, A β , S100b leading to sustained pro-inflammatory state and contributing to A β accumulation and toxicity [281,282] [Oral]	MD – III (Recruiting)
	Masitinib (AB1010)	AB Science	Mast cells/neurons (SmallM)	Inhibition of the Src family Fyn kinase involved in the pathway of tau hyperphosphorylation and tau/A β induced toxicity Inhibitor of the c-kit, the proto-oncogene receptor tyrosine kinase. This is a type 3 transmembrane receptor of the mast cell growth factor that contributes to mast cells activation. Thus, the drug preserves BBB and reduces accumulation of pro-inflammatory factors [283] [Oral]	MD – III (Completed)
	Dexpramipexole (R-pramipexole, KNS-760704)	Virginia Commonwealth University	Neurons (SmallM)	The pure R(+) optical enantiomer of the non-ergot dopamine agonist (pramipexole) with low affinity to dopaminergic receptors. It may reduce cell death induced by H2O2 and block mitochondrial permeability transition, thus reducing ROS generation, imbalance of calcium cellular homeostasis. Finally, it can downregulate mitochondrial pathway of the apoptosis cascade [284] [Oral]	MD – II (Completed)
	CHF 5074 [285,286]	CereSpir Incorporated	Microglia/Neurons (SmallM)	A non-steroidal anti-inflammatory drug that prevents the binding of astrocytic-signaling molecule soluble CD40 ligand to microglia cell surface receptor This in turn, reduce the synthesis and release of TNF- α , IL-1 β and iNOS promoting the expression of microglia M2 markers. In addition, it prevents apoptotic pathways by reducing cytochrome c release, NF- κ B pathways and caspase-3 activation (neuroprotective	MCI – II (Completed)

A) BACE1 inhibitors					
Category	Name (Synonyms)	Company	Target type (Molecule type)	MoA – Biochemical features [Administration route]	Stage ICB Phase
G) Cellular pathway signaling modulators					
Category	Name (Synonyms)	Company	Target type (Molecule type)	MoA – Biochemical features [Administration route]	Stage ICB Phase
Cellular pathway signaling modulators	Exendin-4 (Exenatide)	National Institute of Aging	Insulin signaling (SmallIM)	A protease-resistant GLP-1 analogue that exerts a long acting GLP1 receptor agonism. It induces the sequential activation of an insulin signaling pathway with increased activation of AKT/PI3K and downregulation of GSK-3 β activity, thus preventing hyperphosphorylation of tau In addition, the activation of the GLP1 receptor stimulates an adenylyl cyclase and increases cAMP levels, thus enhancing downstream kinases, as PKA, that are related to growth factor signaling [287–289] [Oral]	MCI/MD CSF II (Completed)
	Mitoglitazone (MSDC-0160)	Metabolic Solutions Development Company	Mitochondrial pathways (SmallIM)	It modulates mTOT acting as insulin sensitizer improving brain glucose metabolism without PPAR γ activation (thus reducing side-effects). In particular, the potential effect is mainly directed to Mpc2 and Mpc1, the mitochondrial pyruvate carrier complex of the mTOT, thus regulating pyruvate entry into the mitochondria [290,291] [Oral]	MD – II (Completed)
	T3D-959 (DB959)	T3D Therapeutics, Inc.	Insulin signaling (SmallIM)	Dual PPAR δ/γ nuclear receptor agonist, regulating, in neurons, dysfunctional brain glucose, lipid metabolism, gene expression of several molecules involved in A β production as BACE1 and pro-survival pathways, thus leading to neuroprotection. It also reduces pro-inflammatory response and microglia activation [292] [Oral]	MD – II (Active, not recruiting)
	Anavex 2–73	Anavex Life Science Corp.	Chaperonine pathways (SmallIM)	A mixed ligand for σ_1 /muscarinic receptors acting as agonist and enhancing most of their cytoprotective pathways: calcium homeostasis, mitochondrial function and ER stress response. In addition, it modulates the axis AKT/PI3/GSK-3 β Downregulation of GSK3 β leads to block tau hyperphosphorylation, thus enhancing ADAM17 expression and reducing BACE1 expression with reduced cellular A β toxic monomers load Potential effect of downstream transcription factors activated by GSK3 that are involved in pro-inflammatory response (STAT, NFkB) [293,294] [IV]	MD – II (Active, not recruiting)
Saracatinib (AZD0530)	AstraZeneca	Src/abl and Fyn kinases pathways (SmallIM)	Inhibitor of both the Src family Fyn kinase and Bcr-Abl tyrosine-kinase. The latter participates to several cell signaling most of them converging on transcription factors fundamental for several cytoprotective and cell-growth pathways (as STAT5 and 3). In addition, it is involved in several inflammatory pathways (inducing microglia and pro-inflammatory release) Inhibitor of the Src family Fyn kinase involved in the pathway of tau hyperphosphorylation and tau/A β induced toxicity [295,296] Inhibitor of the c-kit, the proto-oncogene receptor tyrosine kinase. This is a type 3 transmembrane receptor of the mast cell growth factor that contributes to mast cells activation [Oral]	MD aPET II (Completed)	

A) BACE1 inhibitors

Category	Name (Synonyms)	Company	Target type (Molecule type)	MoA – Biochemical features [Administration route]	Stage ICB Phase
	ABT-957	AbbVie	Calpain pathways (SmallM)	Inhibitor of calpain1, a cytosolic calcium-dependent cysteine protease (overactive in AD neurons) that activates pro-apoptotic pathways mediated by caspases. It also mediates A β toxicity by stimulating NMDR signaling cascades and has been both directly and indirectly (via GSK and cdk5) linked to hyperphosphorylation of tau [297,298] [Oral]	MD – II (Terminated)
	CT1812	Cognition Therapeutics Inc.	S2R/PGRMC1 Pathways (SmallM)	Ligand for the S2R/PGRMC1 receptor that counteracts potential perturbed calcium homeostasis causing mitochondrial dysfunction and enhanced apoptotic pathways It blocks the binding of A β oligomers to Sig2R/PGRMC1, thus interfering with A β calcium-mediated toxicity [299,300] [Oral]	MD – I (Recruiting)
	Vorinostat (suberoylamide hydroxamic acid, Zolinza)	German Center for Neurodegenerative Diseases	Apoptotic pathways (SmallM)	Epigenetic therapy through Class I HDACs inhibitor, that normalizes, in neurons and microglia, epigenetic and transcriptional activity of HD counteracting apoptotic pathways, mitotic failure and autophagic cell death The overactivation of HDACs is also associated to microglia overactivation, insulin resistance, and sustained epigenetic post-translational modifications of A β [301] [Oral]	MD – I (Recruiting)
	S-Equol (Aus-131)	Ausio Pharmaceuticals, LLC	ER- β pathway (SmallM)	S-enantiomeric metabolite of daidzein, a selective agonist of the ER- β (nuclear receptor and member of ligand-regulated transcription factor family expressed on mitochondria). Downregulation of ER- β is associated to mitochondrial dysfunction, insulin resistance, and neuroinflammation (via astrocytes overactivation) [302,303] [Oral]	MD – I (Recruiting)
	LM11A-31-BHS	Pharmatrophix Inc.	p75 neurotrophin pathway (SmallM)	p-75 neurotrophin receptor ligand promotes cell survival signal mimicking the NGF in basal forebrain cholinergic neurons. It prevents A β 1–40 to bind the receptor exerting neurotoxicity [304] [Oral]	MD CSF I-II (Recruiting)
	Allopregnanolone (3 α ,5 α -tetrahydroprogesterone)	University of Southern California	GABA-A mediated cell signaling (SmallM)	A neurosteroid acting as allosteric positive selective modulator of ionotropic GABA-A receptors potentiating the chloride efflux resulting in depolarization of the plasma membrane followed by a rise in intracellular calcium. This, in turn, leads to the restoring of calcium homeostasis and preserved mitochondrial function with reduction of oxidative stress [305] [IV]	MCI/MD – I (Recruiting)
	BI 409306 (SUB 166499)	Boehringer Ingelheim	cGMP/NO signaling (SmallM)	Phosphodiesterase9A inhibitor increasing brain levels of cGMP and NO. NO is a key molecule in anti-apoptotic/pro-survival signaling. Reduced brain cGMP levels, both basal and NMDA-coupled, are associated to AD [306] [Oral]	MD – II (Completed)
Neurotransmission modulators	Ladostigil Hemitartrate (TV-3326)	Avraham Pharmaceuticals Lid	Acetylcholine/MAO A/B (SmallM)	Multi target neuroprotective with both acetylcholinesterase and brain MAO A/B inhibitor activity. The latter prevents neuronal loss by preventing mitochondria-related oxidative stress and potentiating anti-apoptotic factors like BCl2 Potential effect on the gene expression of antioxidant scavengers [307] [Oral]	MCI/MD – II (Completed)
	ORM-12741	Orion Pharma	Noradrenaline (SmallM)	Highly selective α 2C-AR antagonist, a member of the GPCRs superfamily. It counteracts the presynaptic effects of the activated receptor: inhibiting the synthesis and release of NA and other amine). It counteracts the physiological event of inhibition of cAMP-	MD – II (Completed)

Category	Name (Synonyms)	Company	Target type (Molecule type)	MoA – Biochemical features [Administration route]	Stage ICB Phase
A) BACE1 inhibitors					
	RVT-101 (SB-742457, interpiridine)	Axovant Sciences Ltd.	Serotonin (SmallIM)	dependent closing of the voltage-gated calcium channels, and activation of MAPK signaling cascades (which leads to tau hyperphosphorylation and amyloidogenic cellular pathways) [308] [Oral]	MD – III (Completed)
	SUVN-502	Suven Life Sciences Ltd	Serotonin (SmallIM)	Selective 5-HT6 receptor antagonist. Blockade of these receptors leads to increased cholinergic firing. In addition, 5-HT6 receptors are GPCRs that positively stimulate adenylate cyclase activity, finally activating the ERK1/2 via a Fyn-dependent pathway. This, in turn, could reduce hyperphosphorylated levels of tau [309–311] [Oral]	MD – II (Recruiting)
	SAR110894D	Sanofi	Histamine (SmallIM)	Selective H3R antagonist increasing the presynaptic release of both histamine and other neurotransmitters, including acetylcholine [Oral]	MD – III (Completed)
	NorAD	Imperial College London	Noradrenaline (SmallIM)	Extended-Release Guanfacine acting as postsynaptic ARs agonist modulating neuronal excitability via regulation of ion channels, including the direct modulation of Inwardly Rectifying Potassium Channels and the indirect modulation of Hyperpolarization-Activated Channels [313] [Oral]	MD – III (Not yet recruiting)

* At least heterozygosity

Abbreviations: MoA, mechanism of action; SmallIM: small molecule; ICB: inclusion criteria biomarkers; BACE1: β -site amyloid precursor protein cleaving enzyme; APP: amyloid precursor protein; BBB: blood brain barrier; AD: Alzheimer's disease; MCI: mild cognitive impairment; MD: mild AD dementia at least; AaR: asymptomatic at risk; e4: apolipoprotein E e4 carrier; RAR/RXR: retinoid acid receptor/retinoid X receptor; ADAM-10: A disintegrin and metalloproteinase domain containing protein 10; APPs- α : soluble alpha fragment of amyloid precursor protein; PDE4: phosphodiesterase 4; cGMP: cyclic guanosine monophosphate; pCREB: phosphorylated cAMP response-element binding protein; BDNF: brain-derived neurotrophic factor; Zn: zinc; Cu: copper; Fe: ferrum; g3p: two domain fragment of the phage capsid protein; GAIM: general amyloid interaction motif; IgG: immunoglobulin G; AChEi: inhibitor of acetylcholinesterase; PPAR γ : peroxisome proliferator-activated receptor γ ; LXRs: liver X receptors; IV: intravenous; SC: subcutaneous; IM: intramuscular; IC: intrathecal; α -sin: α -sinuclein; IRP1: iron regulatory protein 1; ASOs: anti-sense oligonucleotides; KLH: keyhole limpet hemocyanin; P3K: phosphatidylinositol 3-kinase; GSK3 β : glycogen synthase kinase-3 β ; Ab: antibodies; mAb: monoclonal antibodies; 4-R tau: four microtubule binding repeats tau protein; iNOS: inducible nitric oxide synthase; COX2: cyclooxygenase-2; p38 MAPK α : the alpha isoform mitogen-activated serine/threonine protein kinase p38; Lp-PLA2: lipoprotein-associated phospholipase A2; P2Y6A: nucleotide purine metabotropic receptor; AGEs: advanced glycation end-products; RAGE: cell-surface receptor of the immunoglobulin superfamily; GLP1: glucagon-like peptide-1; mTOT: mitochondrial target of insulin sensitizers; AKT: protein kinase B; GSK-3 β : glycogen synthase kinase-3 β ; σ 1/Musc: sigma 1/muscarnic receptors; STAT: Signal Transducer and Activator of Transcription; cdk5: cyclin-dependent kinase5; S2R/PGRMC1: ligand for the sigma2 receptor, also known as the progesterone receptor membrane component 1; HDACs: histone deacetylases; ER- β : estrogen receptor- β ; TNF- α : cytokine tumor necrosis factor alpha; GDNF: glial cell line-derived neurotrophic factor; NGF: nerve growth factor; GABA-A: inhibitory γ -aminobutyric acid; 5-HT: serotonin; GPCRs: protein-coupled receptors; pERK1/2: phosphor-extracellular signal-regulated kinase1/2; cAMP: adenylyl cyclase; VGCC: voltage-gated calcium channels; NMDA: N-methyl-D-aspartate; iMAO A/B: brain monoamine oxidase A/B inhibitor; ARs: α 2c adrenergic receptor; NA: Noradrenaline; H3R: histamine 3 receptor; ROS: reactive oxygen species; NO: nitric oxide; IRKC: inwardly rectifying potassium channels; HAC: hyperpolarization-activated channels.

Table 2

Evolving lexicon and terminology of the Alzheimer Precision Medicine Initiative (APMI) paradigm.

Concept	Abbreviation	Definition
Biomarkers	BMs	A defined characteristic that is measured as an indicator of normal biological processes, pathogenic process, or response to an exposure or intervention, including therapeutic interventions. Molecular, histologic, radiographic, or physiological characteristics are types of biomarkers. A biomarker is not an assessment of how an individual feel, functions or survives. Categories of biomarkers include: susceptibility/risk biomarker, diagnostic biomarker, monitoring biomarker, prognostic biomarker, predictive biomarker, pharmacodynamics/response biomarker, and safety biomarker.
“Omics” or “Omic” disciplines		Exploratory high-throughput screening tools aimed at fully collecting, characterizing and quantifying comprehensive pools of biological molecules (DNA sequences, transcripts, miRNAs, proteins/peptides, metabolites/lipids) that relate to structure, function, metabolism and dynamics of an organism and/or whole organisms.
Stratified medicine		Medical model that uses the grouping of patients according to underlying biological mechanisms, biomarker-guidance, disease risk or likely treatment response, as established by diagnostic tests, to determine the course of care. Stratified medicine is a component of personalized medicine.
Precision Medicine	PM	Translational science paradigm related to both health and disease. PM is a biomarker-guided targeted medicine on systems-levels taking into account methodological advancements and discoveries of the comprehensive pathophysiological profiles of complex polygenic, multi-factorial neurodegenerative diseases (proteinopathies of the brain). It aims at optimizing the effectiveness of disease prevention and therapy, by considering (customized) an individual's specific “biological make-up” (e.g. genetic, biochemical, phenotypic, lifestyle, and psychosocial characteristics) for targeted interventions through PAM implementation.
Pathway-based therapy		A treatment developed following the systematic analysis of specific genes, their functions, and the interactions among them in relation to a specific complex disease. By using reliable exploratory strategies (i.e. GWAS, proteomics, and microarrays), the comprehensive understanding of the molecular mechanisms underlying complex diseases is realistic.
Systems Biology	SB	Evolving hypothesis-free, exploratory, holistic (non-reductionistic), global, integrative, and interdisciplinary paradigm using advances in multimodal high-throughput technological platforms that enable the examination of networks of biological pathways where elevated amounts of structurally and functionally different molecules are simultaneously explored over time at a system level (i.e., at the level of molecules and subcellular compartments, cells, group of cells, tissues, organs, apparatuses, or even whole organisms). According to systems biology, organisms are made of systems which are entities consisting in hierarchically self-organized levels with increasing structural complexity resulting in different emerging properties.
Systems Pharmacology	SP	Science of advancing knowledge about drug action at the molecular, cellular, tissue, organ, organism, and population levels” (available at http://www.aaps.org/Systems_Pharmacology/).
Precision Pharmacology	PP	Conceptual paradigm combining traditional data analyses as pharmacodynamic and pharmacokinetic data within the System biology approach. It encompasses the acquisition and integration of omics data operating at both experimental and computational level. Precision pharmacology aims at exploring and predicting the whole effect of a molecular mechanism of action at different systems levels.
Self-Organization	SO	Spontaneous (self-generated) interactions among components of an initially chaotic basis. Such a process may be triggered by random fluctuations that generate casual processes which are amplified by positive feedback. The result of self-organization is an entity of an increased structural-functional order that acquired the ability to survive the environment and self-repair after perturbation. Therefore, self-organization in organisms at each level is a key phenomenon for survival and evolutionary transitions.
Network		Self-organization, in biology, can be observed in protein folding, creation of lipid bilayer membranes, cell/tissue/organs genesis and development. Set of recurring motifs; each motif is a pathway; each pathway, in turn, carries out specific dynamical functions and can be modulated, i.e. up-down regulated, either upstream or downstream or both. There is a large <i>spectrum</i> of biological cross-talks between pathways and networks inside a level of a given system and among systems.
Feedback		Condition in which a component of a molecular pathway either activates or inhibits its own upstream regulators.
Cross-talk		Condition in which one or more signaling transductions of a pathway directly or indirectly affect one or more signaling transductions of other pathways. A component of one pathway can either positively and negatively modulate (cross-activation and cross-inhibition, respectively) an upstream component of another pathway, thus modulating the biological output.

Concept	Abbreviation	Definition
Signal-transduction cascades (pathway)		Circuit of interactions among molecular bioprocesses (molecular circuit) able to detect, amplify, and integrate different signals.
Emerging properties		Hierarchically self-organized levels with a certain degree of structural complexity that exhibit properties that levels with lower complexity do not show.
Homeostasis		It consists in a spontaneous tendency towards a condition of a dynamic <i>equilibrium</i> based on a continuous counterbalance between regulatory-defense mechanisms and disrupting stress-induced signals. Homeostasis is common to any biological system. Homeostatic signaling is hierarchically organized from subcellular to cellular level, across organs, and, finally, systems. Homeostasis is essential for protecting all core biosynthetic processes necessary to optimal functioning and survival.
Adaptation		Biological output arising from multi-level anti-stress response, generating advantageous morpho-functional alterations in cells and higher levels inside a system. Adaptation is essential for coping environmental stressful challenges aimed to prevent systems damage and finally promoting survival.
Compensation		Protective process in which a morpho-functional alteration is counterbalanced by another morpho-functional alteration without any change in biological output, thus preserving system homeostasis. Compensatory mechanisms are hierarchically organized through systems levels and aim at preserving the homeostasis under pathophysiological conditions. Example of compensation: myofibrillar cardiac remodeling; cell surface receptor profiles in the immune system; membrane ionic channel in neurons.
Decompensation (Failure)		Breakdown or lack of reverse of one or more compensatory mechanisms finally resulting in maladaptive morpho-functional alterations. This, in turn, reflects a homeostatic imbalance at different levels of complexity in systems and organ systems.

Table 3

Summary of clinical trial construct to conduct trials leading to PM-based interventions.

PM	Corresponding Precision Clinical Trial Features
Right Drug	<ul style="list-style-type: none"> ■ Target biologic known ■ Druggable aspect of pathology identified ■ Drug or drug/combination reflects single or multiple pathologies and stage of the process ■ Biomarker guides drug choice
Right Dose	<ul style="list-style-type: none"> ■ Maximum tolerated dose determined ■ Dose-response curve constructed ■ Multiple doses tested ■ Doses related to disease severity
Right Patient	<ul style="list-style-type: none"> ■ Phase of disorder ranging from preclinical without genetic/or amyloid risk factors, preclinical with genetic/amyloid risk factors, prodromal AD, AD dementia ■ Pathology and drug target defined by biomarkers ■ Pharmacogenetic profile of patient determined to anticipate rate of metabolism, drug-drug interactions, drug-disease interactions, and side effects ■ Socio-demographic features of individual included in planning including age, sex, caregiver status, comorbid conditions, general health, concomitant medications, etc. ■ Analysis of individual responses or response patterns of small groups of biologically well-defined individuals

Abbreviations: AD, Alzheimer's disease; PM, Precision Medicine.