

Lawrence Berkeley National Laboratory

LBL Publications

Title

Permafrost as a potential pathogen reservoir

Permalink

<https://escholarship.org/uc/item/50s30845>

Journal

One Earth, 5(4)

ISSN

2590-3330

Authors

Wu, Ruonan
Trubl, Gareth
Taş, Neslihan
[et al.](#)

Publication Date

2022-04-01

DOI

10.1016/j.oneear.2022.03.010

Peer reviewed

1 **One Earth Review: Permafrost as a Potential Pathogen Reservoir**

2

3 **Authors:** Ruonan Wu¹, Gareth Trubl², Neslihan Taş^{3,4} and Janet K. Jansson^{1*}

4

5 ¹Biological Sciences Division, Pacific Northwest National Laboratory, Richland, WA, USA

6 ²Physical and Life Sciences Directorate, Lawrence Livermore National Laboratory, Livermore, CA, USA

7 ³Earth and Environmental Sciences Area, Lawrence Berkeley National Laboratory, Berkeley, CA, USA

8 ⁴Biosciences Area, Lawrence Berkeley National Laboratory, Berkeley, CA, USA

9 *Corresponding author; e-mail: janet.jansson@pnnl.gov

10

11 **Summary**

12 The Arctic is currently warming at unprecedented rates due to global climate change, resulting in
13 thaw of large tracts of permafrost soil. A grand challenge is to understand the implications of permafrost
14 thaw on human health and the environment. Permafrost is a reservoir of mostly uncharacterized
15 microorganisms and viruses, many of which could be viable. Given our limited knowledge of resident
16 permafrost microbes, we also lack the basis to judge if they pose risks to humans, animals and plants. Here,
17 we delve into features of permafrost as a microbial habitat, and discuss what is known about the potential
18 for microbial pathogens to emerge in a warming climate as permafrost thaws. This review has broader
19 implications for human health and ecosystem sustainability in the new arctic environment that will emerge
20 in a thawed permafrost landscape.

21

22 **Keywords:** permafrost, climate change, emerging pathogen, soil microbiome, soil virus

23

24

25 **Introduction**

26 Current predictions of permafrost land coverage range between 15–24% of the Northern
27 hemisphere^{1,2}. Permafrost soils are among the largest terrestrial carbon reservoirs on the planet and in the
28 Northern hemisphere permafrost soils are estimated to contain 1014 petagrams carbon to a depth of 3
29 meters³. By definition, soils that have remained frozen for at least two consecutive years are accepted as
30 permafrost, but most formations are much older than that and originate from prior glaciation periods. Frozen
31 permafrost soils are overlain by an active layer which undergoes seasonal freeze-thaw cycles. Depending
32 on when the soil froze, permafrost can contain organic depositions from prior periods of Earth's

33 paleoclimate, including periods with a warmer and more carbon dioxide (CO₂) rich atmosphere. Active
34 layer carbon stocks are newer deposits in comparison to permafrost³.

35 Permafrost is thawing due to anthropogenic climate change, and we can already observe large scale
36 changes in permafrost regions. Soil temperatures have increased by up to 2°C over the last decade and the
37 active layer has deepened significantly⁴. Global warming has disproportionately warmed both high-latitude
38 and high-altitude permafrost (i.e., Tibetan plateau) areas, and these regions are projected to continue to
39 experience greater temperature increases compared to other regions of the world⁵. As a result, on a global
40 scale, 22–64% of permafrost is expected to thaw by 2100⁶. Thawing permafrost creates a positive feedback
41 loop by releasing nutrients such as water, allowing plants and microorganisms to grow. In low drainage
42 areas containing ice-rich permafrost or massive ground ice, gradual thaw has resulted in ground subsidence
43 and inundation where formation of bogs, fens, and thermokarst (abrupt thaw) lakes mainly support
44 microbial anaerobic fermentation and methane (CH₄) production^{7,8}. Thawed permafrost carbon stocks are
45 rapidly turned to CH₄ which further contributes to greenhouse gas accumulation^{9,10}. However, these
46 observations do not reflect the entirety of the complex changes that Arctic landscapes are currently
47 experiencing. Increasingly warmer and longer summer seasons rapidly melt ground ice where abrupt thaw
48 intensifies subsidence and deepens the active layer¹¹. Abrupt thaw in upland areas results in active layer
49 detachments or large scale landslides, and forms gullies and thaw slumps¹². Model estimations predict
50 abrupt thaw to cause carbon losses that are equivalent to 40% of those expected from the gradual thaw¹³.
51 Currently we have limited understanding of how microbes respond to such sudden changes and shifting
52 quantities in ice, carbon, and nutrient deposits⁴.

53 Here, we delve into features of permafrost that could make it a reservoir for microbes, including
54 pathogens, and discuss what is known about the potential for microbial pathogens to emerge in a warming
55 climate as permafrost thaws. In most cases we find that the threats to public health are minor. We show that
56 although there is potential for bacterial human pathogens, such as anthrax, to be problematic as permafrost
57 thaws, there is less potential for other human pathogens. Most of the microorganisms and viruses that have
58 been identified in permafrost to date target non-human hosts, such as plants and microorganisms. Plant
59 pathogens could, however, pose a risk to native and introduced crops. Although fragments of human viruses
60 have been recovered from remains preserved by permafrost, none have been shown to be infectious to
61 humans. However, many of the microorganisms and viruses residing in permafrost have not yet been
62 isolated or characterized. Therefore, their potential to pose future risks to humans, animals and plants
63 warrants consideration. These risks may increase with climate change as the landscape and vegetation
64 changes and the human population becomes increasingly exposed to thawed permafrost lands.

65

66 **MICROBES IN PERMAFROST**

67 Permafrost soils have been shown to be microbially diverse terrestrial ecosystems⁷. When frozen,
68 many of the resident microorganisms in permafrost, unlike macroorganisms — ‘think woolly mammoths’
69 — were able to adapt and survive in subzero temperatures^{7,14}. These psychrotolerant microorganisms¹⁵ also
70 have to endure other stressful conditions, including high salinity, and low water and nutrient availability¹⁶⁻
71 ¹⁸; as well as anoxia and low pH in frozen bogs and fens^{19,20}. Permafrost “microbes” include bacteria,
72 archaea, fungi, algae, protists and viruses. Protozoa have also been isolated from permafrost²¹. The types
73 of microbes and their ability to survive depend on a variety of factors including the permafrost age and
74 chemical composition. Whether any of these microbes are potentially pathogenic to humans, other animals,
75 or plants is a current concern and the topic of considerable debate²².

76 Previous reviews have summarized the potential for pathogen emergences from other cryosphere
77 environments, including glaciers and ice patches²³⁻²⁵. For example, high frequencies of influenza A viral
78 RNAs were detected in Siberian Lake ice that can potentially be transmitted via migratory birds²⁶. In another
79 study, one arthropod RNA virus and one plant DNA virus were recovered from ice patches in the Canadian
80 Arctic²⁷. Although outside the scope of this review that is focused on permafrost, these reports underscore
81 the potential for the cryosphere to serve as a reservoir of emerging pathogens.

82

83 **Bacteria and archaea in permafrost**

84 It is known that the microbial communities that inhabit active layer and permafrost soil layers are
85 quite different in composition⁷. Active layer and permafrost microbes also have different vulnerabilities to
86 increases in global temperatures. As permafrost thaws, the depth of the active layer increases. As a result,
87 there is usually an increase in microbial activity in the newly formed active layer which in return intensifies
88 greenhouse gas emissions⁷ and a transition in the composition of the microbial community²⁸. Soil moisture
89 content and overlying vegetation have been shown to be key drivers of the microbial response to permafrost
90 thaw^{20,29,30}. In addition, differences in soil conditions, e.g., temperature, redox chemistry, moisture content,
91 are clearly of importance in determining the thaw response²⁸.

92 Permafrost soils are shown to contain a wide range of soil bacteria, predominantly from the
93 Terrabacteria supergroup, including *Actinobacteria*, *Firmicutes*, *Cyanobacteria* and *Chloroflexi*, as well as
94 *Bacteroidetes* and *Proteobacteria*⁷. Some representatives of less well known phyla have also been
95 discovered in permafrost metagenomes, including metagenome-assembled genomes corresponding to
96 Candidate phylum *Eremiobacteraeota* and Candidate phylum *Dormibacteraeota* in intact and thawing
97 permafrost³¹. Similar to bacteria, many different species of archaea have been found in permafrost,
98 including representatives of *Euryarchaeota*, *Crenarchaeota* and *Thaumarchaeota*. To date, the most
99 commonly observed archaeal species in thawing permafrost are methanogenic *Euryarchaeota* that can use
100 fermentation byproducts to produce CH₄³².

101 Conditions in permafrost can preserve both nucleic acids and live cells that have mechanisms for
102 survival under long-term freezing conditions. Potential resources in permafrost, including organic carbon
103 nutrients and water are key to ensuring long-term survival. Because living organisms require water for
104 survival, permafrost poses a unique challenge due to low water availability. Even a thin layer of salt brine
105 can serve as a water reservoir because the salt increases the freezing temperature of water. Liquid water has
106 been detected in permafrost at -10°C ³³ and bacteria have been shown to be metabolically active at that
107 temperature³⁴.

108 Increasing permafrost age also increases the necessity for microbial cells to cope with extended
109 periods of stressful conditions^{28,35}. Mackelprang et al., (2017)²⁸ used 16S rRNA amplicon and metagenomic
110 sequencing to profile the permafrost microbial communities existing in a permafrost chronosequence that
111 ranged from 19000 to 33000 years ago (Pleistocene Era)²⁸. They found that the age of the permafrost
112 influenced the types of microbes present. For example, representatives of *Firmicutes* that can form resistant
113 spores increased in relative abundance with age of the permafrost. Functional analysis of metagenomes
114 revealed an enrichment in pathways for degradation of recalcitrant plant material and stress-related genes
115 in older permafrost³⁶.

116 Most research has focused on understanding how interactions among post-thaw permafrost
117 microbiomes are deterministic of observed carbon degradation rates and pathways. For example, recent
118 evidence suggests that conditions mediated by thaw could give rise to acetoclastic methanogenesis where
119 hydrogenotrophic methanogenesis was predicted to be dominant in intact permafrost³⁷. Drainage driven dry
120 soil conditions, on the other hand, can enhance aerobic processes and fuel CO_2 production especially in
121 uplands¹⁰. Under these conditions easily decomposable sugars, simple amino acids and lipids are depleted
122 due to oxidative decomposition, where hydrophobic carbon compounds are preserved in deep soils. In
123 addition, multi-omics analyses of new thermokarst bog and fen formations showed high abundances of
124 methanogenesis-related genes, transcripts and proteins, suggesting that CH_4 generation will increase in
125 these systems once permafrost thaws^{16,32}. This has important implications for greenhouse gas emissions
126 because CH_4 is a more potent greenhouse gas than CO_2 . By contrast, little is known about the risk of
127 pathogen exposure in thawed permafrost.

128

129 **Viruses in permafrost**

130 Viruses are the most numerically abundant biological entities on Earth³⁸ and we have just begun to
131 investigate the full extent of viral diversity in thawing permafrost soils. Recent surveys in the active layer
132 of permafrost soils have identified that the vast majority of viruses are double-stranded (ds) DNA
133 bacteriophages (phage; viruses that only infect bacteria, 'Bacteriophage' in **Figure 1**). These phages are
134 largely unrelated to other known phages and a large portion of the community is active³⁹. The phage

135 community composition changes with permafrost thaw from soil-virus-like assemblages in the palsas with
136 intact permafrost to a mixture of soil-like and aquatic-like in partially thawed bog to aquatic-virus-like
137 assemblages in the fully thawed inundated fen habitats^{40,41}. Phage genomes are attuned to their
138 environmental conditions by carrying auxiliary metabolic genes (AMGs) that can help their hosts overcome
139 metabolic bottlenecks such as resource limitations. For example, phages detected in thawing permafrost
140 peatlands (soils with a large quantity of organic matter due to incomplete decomposition) were found to
141 carrying an array of glycoside hydrolases that breakdown complex carbohydrates into labile carbohydrates
142 which in turn sustains a wide diversity of microorganisms^{40,41}.

143 Phages are the most intensely studied type of viruses in environmental systems, and in contrast our
144 knowledge on ssDNA and RNA viruses is limited, especially in permafrost ('Virus' in **Figure 1**). This has
145 been largely due to (1) dsDNA phages being ubiquitous and abundant, (2) common methods to capture
146 ssDNA or RNA viruses preclude quantitative ecological inferences⁴² or exclude them (e.g., metagenomes
147 don't capture RNA viruses), and (3) most ssDNA and RNA viruses appear to infect eukaryotes^{43,44}. This
148 suggests that we are missing an unknown quantity of environmental viruses that could be emerging
149 pathogens.

150 Recently, a protocol was published to increase our detection and characterization of ssDNA viruses
151 in soils underlain with permafrost⁴⁵. While ssDNA viruses were detected, they represented a small fraction
152 of the viral diversity and abundance, and the potential hosts were bacteria or micro-eukaryotes (e.g., fungi
153 and amoeba)⁴⁵. Other studies of ssDNA⁴⁶ and RNA viruses^{29,47} in non-permafrost soils showed that they
154 are abundant and diverse groups of viruses with clear differences in bulk-soil versus rhizosphere
155 communities, and also identified bacteria or micro-eukaryotes as potential hosts. However, we have yet to
156 fully understand the full extent of global viral diversity, including in permafrost soils^{29,46,47}.

157

158 **Microbial activity in permafrost**

159 Permafrost environments are subzero and often anoxic, which has led many to believe that resident
160 microbes are not active. Over the last decade there has been mounting research revealing diverse
161 assemblages of active bacteria and viruses from permafrost environments^{39,48,49}. A multi-omics approach
162 (i.e., metagenomics, metatranscriptomics and metaproteomics) was used to determine which microbial
163 populations were active along a permafrost thaw gradient¹⁶. The proteome data revealed several proteins
164 that were expressed by permafrost microorganisms to equip them to survive the harsh conditions in
165 permafrost, including cold shock proteins. The metatranscriptome data also revealed actively transcribing
166 bacterial populations in permafrost, including representatives of *Firmicutes*, *Bacteroidetes*, *Acidobacteria*
167 and *Proteobacteria* phyla¹⁶. It is noteworthy that the active microbes detected in permafrost in the study¹⁶
168 by Hultman and colleagues were representatives of 'typical' environmental bacteria, not pathogens.

169 With respect to viruses, there is a prevailing reductionist point of view that claims viruses are not
170 truly active, but rather they are like enzymes where random collisions lead to a series of chemical reactions.
171 This is because without cells, viruses cannot multiply, and they do not have the ability to generate ATP.
172 Therefore, to determine if a virus is active, evidence is needed that the virus is able to infect a host and
173 cause a change in the host's ecology. In previous work, viruses have been co-cultured with putative hosts
174 to determine activity but determining the host and culturing a virus can be difficult and time consuming⁵⁰.
175 Several advances have recently been developed to determine activity of viruses in environmental samples.
176 A few examples include: (1) bioorthogonal noncanonical amino acid tagging (BONCAT) which is a
177 technique that adds a synthetic amino acid to a sample that allows the tracking and quantification of newly
178 synthesized proteins⁵¹; (2) Stable Isotope probing (SIP) which uses an isotope with more mass to label
179 newly synthesized DNA³⁹; (3) and multi-omics (i.e., metagenomics, metatranscriptomics, and
180 metaproteomics²⁹), such as described above¹⁶, that can reveal viral presence, gene expression, and protein
181 production in a sample. Unfortunately, viruses can have several infection styles, each having variable
182 effects⁴² (warranting further investigation). While these techniques have been applied to an array of habitats,
183 few have been applied to determine viral activity in permafrost. Recently, SIP-metagenomics was used to
184 determine viral activity in bog samples³⁹ that were incubated for six to twelve months under conditions
185 simulating the freezing winter temperatures in the presence of H₂¹⁸O. Active populations incorporated ¹⁸O
186 into their DNA when replicating and could thus be distinguished from inactive populations. This study
187 revealed that only a subset of bacteria were active under the simulated winter conditions and suggested that
188 they survived by fermentation and carbon scavenging processes. However, the majority of the detected
189 phages were active and linked to the active bacterial populations, highlighting their importance in
190 structuring of the microbial community. Note that this study focused on active phages that have prokaryotic
191 hosts. Thus, the activity of potential plant, animal or human viruses was not determined.

192

193 **PATHOGENS IN PERMAFROST**

194 An estimated 5 million people currently live in Arctic regions that have underlying permafrost
195 lands⁵². This number is even higher when including tourism and trade in sub-Arctic permafrost areas^{53,54}.
196 As permafrost thaws there is thus a potential for emerging pathogens to infect humans. Building on current
197 knowledge about microbial life and activity in permafrost in general, we will focus on what has been
198 reported about potential pathogens and exposures from thawed permafrost. We will highlight examples of
199 bacterial, fungal and viral pathogens in permafrost that can pose future risks to humans, other animals and
200 plants (details of each in **Table 1**). Our survey of the literature will conclude that we still have much to
201 learn about potential pathogen risks from thawing permafrost.

202

203 **Potential bacterial pathogens in permafrost**

204 Bacterial pathogens may be preserved in a quiescent state in permafrost and become activated when
205 the permafrost thaws and conditions are favorable for growth ('Bacterial Pathogen' in **Figure 1**). One of
206 the best-known examples is that of *Bacillus anthracis*, the agent of anthrax disease that is globally
207 distributed with potential risks to humans, livestock and wildlife⁵⁵. Spores, such as those from *Bacillus*
208 *anthracis*, are notoriously robust during long-term storage in freezing conditions ('Anthrax Spores' in
209 **Figure 1**). In 2016, an outbreak of anthrax was reported in Siberia, due to exposure to infected carcasses
210 that had previously been frozen in permafrost. As the permafrost thawed the carcasses became exposed and
211 anthrax spores were released. As a result, one person and over 2000 reindeer died after ingestion of the
212 spores^{56,57}. Anthrax outbreaks have also occurred when infected cattle that were buried were re-exposed
213 after permafrost thaw⁵⁸.

214 Stella et al. (2020)⁵⁹ modeled the risk of future anthrax exposure and found that the annual risk of
215 anthrax infection is strongly correlated with active layer depth. The model distinguished between freshly
216 released spores from thawed carcasses to those that become reactivated as a result of permafrost thaw.
217 Seasonal forcing by thawing and herding was found to favor endemic disease transmission. Several
218 knowledge gaps exist that are needed to improve the model, including how spores are vertically distributed
219 in permafrost soils. Their results suggest that seasonal thawing of permafrost, together with longer periods
220 of warmer temperatures, could result in an increased prevalence of re-emergence of Anthrax spores and
221 increased risk of associated disease in the Arctic⁵⁹. A compounding reason for anthrax outbreaks is an
222 increase in reindeer with a lack of immunity⁶⁰ that could contribute to the spread and infectious capacity of
223 the pathogens. Therefore, one potential mitigation strategy to help control future outbreaks would be to
224 increase reindeer vaccinations.

225 There are several reports about prevalence of antibiotic resistance genes in permafrost bacteria⁶¹⁻
226 ⁶³ that could make known pathogens more problematic. Antibiotic resistance poses a threat to human health
227 by reducing the efficacy of antibiotic treatments. Antibiotic resistance in pathogenic bacteria is assumed to
228 be a modern phenomenon. However, targeted metagenomic analyses from 30,000-year-old Alaskan
229 permafrost showed resistance to β -lactam, tetracycline and glycopeptide predating the discovery and use of
230 medicinal antibiotics⁶⁴. This finding suggests that permafrost can serve as a reservoir of antibiotic resistance
231 genes. Prevalence of antibiotic resistance in permafrost bacteria is likely an emergent property for
232 competitive fitness rather than pathogenicity. As permafrost thaws, there is the potential for an increase in
233 bacterial populations that bear plasmids with antibiotic resistance genes that have the potential to spread.
234 However, this potential concern is likely no greater than that posed by non-permafrost soils that also harbor
235 abundant antibiotic resistance genes.

236

237 **Potential fungal pathogens in permafrost**

238 Sequencing has revealed that several fungi are present in permafrost, including species of
239 *Geomyces*, *Cladosporium*, *Aspergillus* and *Penicillium*⁶⁵. Some fungi that have been isolated from
240 permafrost are potential plant pathogens ('Fungi' in **Figure 1**). Ancient permafrost from Siberia (16,000–
241 32,000 years old) was found to contain several fungal OTUs from 3 phyla: Ascomycetes, Basidiomycetes
242 and Mucormycota⁶⁶. This study found that approximately a third of the species detected were representative
243 of plant-associated fungi, including pathogens. Within thermokarst bogs, the plant pathogens *Galerina*
244 *paludosa* and *Hyaloscypha* increased in relative abundances following permafrost thaw⁶⁷. The fungi were
245 likely trapped together with plant material as the permafrost froze. Most fungal isolates from permafrost
246 form small conidia suggesting that this is a survival mechanism for long-term incubation in the cryosphere.
247 Whether the fungi were still alive or not is not known. However, several of the fungal taxa were designated
248 to be putative psychrotrophic fungi with representatives known to produce dark pigments and spores to
249 help with survival under stress conditions.

250 Representatives of fungal insect pathogens have also been detected in ancient Siberian
251 permafrost^{66,68}. Some taxa were putative colonizers of skin or dung of mammals, potentially reflecting the
252 presence of a range of insect and other animal species that thrived at the time (Pleistocene to early Holocene
253 areas). Lydolph et al (2005)⁶⁸ studied ancient DNA from a permafrost tunnel in Siberia that ranged from
254 modern age to 400,000 years ago. They also found ancient DNA with fungal representatives of coprophilic
255 and keratophilic fungi, as well as plant pathogens. The main representatives were from Ascomycetes,
256 Basidiomycetes and Zygomycetes phyla. However, no active fungal pathogens in permafrost that directly
257 infect humans have been reported to date to our knowledge.

258

259 **Potential viral pathogens in permafrost**

260 It is currently unknown how many viruses are trapped in permafrost, but once thawed, the so-called
261 "sleeping" or "zombie" viruses may become free to infect hosts. Although viruses can be preserved in a
262 frozen environment for long periods of time⁶⁹, endurance of environment pressures during thawing without
263 bursting is critical for viruses to remain intact and infectious. One example of a class of viruses that have
264 managed to survive in ancient permafrost are giant viruses. Giant viruses or nucleocytoplasmic large DNA
265 viruses (NCDLV) are known to have durable multiple layers enclosing virion nucleoids making them
266 resistant to the harsh permafrost environment⁷⁰ ('Giant viruses' in **Figure 1**). Two members of giant viruses,
267 *Pithovirus sibericum*⁵⁰ and *Mollivirus sibericum*⁷¹ were previously isolated and revived from 30,000-year-
268 old Siberian permafrost. Although these viruses were preserved frozen in permafrost for at least ~30,000
269 years, after thaw they were able to infect a modern-day version of their host⁵⁰. The isolated giant viruses
270 were reported to have thick outer walls of 40–70 nm^{71,72} comparable to 20–80 nm for Gram positive

271 bacteria⁷³. However, the giant viruses that have been characterized to date do not pose a threat to humans
272 and other animals because their natural hosts are microeukaryotes, such as protists and algae ('Protozoa' in
273 **Figure 1**). Another giant virus, Mimivirus, that was first isolated from water samples for investigating a
274 pneumonia outbreak⁷⁴, was however, found to be capable of entering human and mouse myeloid cells
275 resulting in a significant increase in viral DNA and a decrease in macrophage viability⁷⁵. A clinical study
276 also reported a high prevalence of Mimivirus antibodies in pneumonia patients with unknown underlying
277 mechanisms⁷⁶. Although viruses often have a narrow and specific host range, these findings challenge the
278 assumption that microeukaryotes are exclusive hosts for giant viruses, highlighting a major knowledge gap
279 in virus-host relationships. Currently, however, there is no evidence that isolated and revived giant viruses
280 from permafrost can cause human infections.

281 The majority of DNA viruses in permafrost environments are phages and therefore cannot cause
282 disease in humans. In fact, phage are arguably the most well-studied type of virus and the majority of them
283 (and viruses in general) are host specific. This means that they can only infect bacteria that are the same
284 species and often only the same strain^{77,78}. Despite phages being largely bacterial host-specific, they appear
285 to be major community-structuring agents that uphold a diverse permafrost microbiome. A diverse
286 microbiome prevents any specific bacterium, e.g., a human pathogen, from growing in abundance. This
287 means that beyond phages killing pathogenic bacteria, thereby protecting us, they can create an ecosystem
288 where pathogenic bacteria are less likely to be of concern to humans⁷⁹. Given current evidence of the wide-
289 spread presence of phages in all soils, including permafrost, they are very unlikely to represent a threat to
290 humans.

291 Other types of DNA viruses, such as Variola virus which causes smallpox, that infect humans may
292 be present in permafrost environments, but they have not been detected in sufficient numbers or good
293 quality to pose a threat⁸⁰. Because DNA viruses co-evolve and replicate in the presence of their hosts, in
294 order for pathogenic human viruses to be present in sufficient quantities to cause infection, human/animal
295 remains would have to be preserved in a condition that allows for viral survival. However, this is highly
296 unlikely. For example, Variola virus DNA was recovered from several archeological sites in northeastern
297 Siberia dating back to the late 17th to 19th century. Smallpox victims mummified in the permafrost were
298 found, but the Variola virus DNA was highly degraded and thus the virus was not infectious^{81,82}. Therefore,
299 although human DNA viral pathogens can be detected in permafrost, the risk of infection is negligible and
300 only of concern if the virus is intact and in sufficient quantities to cause disease. It is noteworthy that to
301 date there have been no reported cases of viral infections from thawed permafrost in humans.

302 To our knowledge, there is no genetically informed ecological study of the RNA viral community
303 in permafrost. One reason that RNA viruses are understudied compared to DNA viruses is that RNA is less
304 stable compared to DNA⁸³ and more prone to degradation during the extraction process. One example of

305 an RNA virus is the H1N1 influenza A virus that caused the 1918 pandemic. This virus killed an estimated
306 50 million people, which overwhelmed the burial of infected humans leading to mass graves and unburied
307 corpses⁸⁴. These mass graves largely went unmarked leaving the potential for humans to interact with these
308 corpses should permafrost thaw. In the 1990s many trials were attempted to reconstruct or revive the 1918
309 pandemic influenza virus from frozen samples, but they were unsuccessful because the RNA was highly
310 degraded. In the early 2000s, advances in technology allowed for the successful reconstruction of the H1N1
311 virus from permafrost, but it was not viable⁸⁰. Complete hemagglutinin (HA) genes, encoding binding
312 proteins to initiate infection, were detected from the reverse-transcribed RNAs extracted from the frozen
313 lung tissues of 1918 influenza victims who were buried in a mass grave in permafrost⁸⁵. Although this last
314 example could be taken as a cause for concern, the viruses were not infectious. Further, ancient RNA
315 extracted from wolf tissues buried in Late Pleistocene permafrost showed no traces of RNA viruses⁸⁶. To
316 date, there is no report of intact and infectious RNA viruses directly isolated from permafrost. Therefore,
317 although RNA viruses can be preserved in permafrost, based on our current knowledge, the risk of these
318 RNA viruses being infectious to humans or other animals is unlikely.

319 Permafrost thaw is causing major changes in plant composition and productivity and will
320 undoubtedly affect plant viral pathogens⁸⁷. Previously, plant viral pathogens such as Ancient Caribou Feces
321 associated Virus (aCFV) and Tomato Mosaic Tobamovirus (ToMV) were recovered from ice^{27,88}. aCFV is
322 a DNA virus that can infect *Nicotiana benthamiana* (a close relative of tobacco) and may even infect other
323 plants or fungi (e.g., mosses, lichens), and was recovered from caribou feces dating back 700 yrs BP²⁷. To
324 test if aCFV was still infectious, a modern version of *N. benthamiana* was exposed and successfully
325 infected. Although the plant became infected, it had an asymptomatic infection, likely due to changes in
326 the plant species over time. Unlike aCFV, ToMV is one of the most common plant viruses, because it has
327 the ability to infect many different plants such as cucumber, lettuce, beet, and pepper⁸⁹. This RNA virus is
328 very stable, having previously been shown to survive UV radiation and desiccation, and was recovered
329 from ice cores dating back up to 140,000 years old⁸⁸. It is unknown if the recovered ToMV could still be
330 infectious, but it presents a potential problem because it is one of the most destructive diseases, resulting in
331 serious crop losses⁹⁰. The introduction of non-native crops to soils, such as recently thawed permafrost
332 soils, could also increase the likelihood of a viral plant epidemic^{87,90}.

333

334 CONCLUSIONS AND PERSPECTIVE

335 Although we have focused on permafrost as a potential reservoir of pathogens, there are other
336 factors to consider in a warming Arctic climate. As we have summarized in this review, although some of
337 the microorganisms and viruses that are preserved in permafrost can be active after thaw, the risks to human
338 health are generally low (**Figure 2**). In the age of COVID-19, the human population has become painfully

339 aware of the threat of emerging pathogens from 'wild' spaces. Although speculative, there is still a risk that
340 an unknown pathogen could arise from 'wild' permafrost where it has survived over a long period of time
341 in a dormant or quiescent state. A more immediate concern is expansion of modern infectious diseases via
342 vectors (e.g., ticks and mosquitoes) in areas where permafrost has thawed and the soil becomes wetter, or
343 collapses into thermokarst bogs, as the climate warms⁹¹⁻⁹³. Increased temperature and precipitation can
344 further expand the geographical distribution of animal vectors⁹⁴. Many of these vectors have been reported
345 to carry human pathogens such as *Francisella tularensis*⁹⁵, West Nile virus⁹⁶ and Zika⁹⁷. Additionally,
346 transmission vectors can facilitate the spreading of permafrost bacterial pathogens. For instance, anthrax
347 was reported to be carried by both mosquitos (*Aedes aegypti* and *Aedes taeniorhynchus*) and flies (*Stomoxys*
348 *calcitrans*)⁹⁸. Another pending concern is that increased temperature is correlated with an increase in food
349 and waterborne diseases⁹⁹. Accordingly, there have been recent reports of several enteric disease outbreaks
350 in the Arctic that were caused by bacterial pathogens, including *Campylobacter*, *E. coli* 0157:H7, *Vibrio*
351 *parahemolyticus* and *Salmonella*. Other vector-borne diseases (e.g., rodents) that could increase in
352 frequency in a warmer Arctic environment include TBE, *Nephropathia epidemia*, tularemia, borreliosis,
353 malaria, toxoplasmosis, and Sindbis Virus infections²². Finally, airborne diseases including viral respiratory
354 tract infections and influenza have been correlated to shifts in temperature and humidity. However, the
355 research underlying increased disease prevalence in the Arctic is still scarce. Therefore, we conclude with
356 a call to the broader research community to increase research efforts in this area, as recently highlighted in
357 a general call to increase research on the impact of climate change on microorganisms in the environment¹⁰⁰.

358

359 **Author Contributions**

360 J.K.J. coordinated the scope and content of the review. N.T., R.W. and G.T. drafted Figure 1. R.W.
361 drafted Figure 2. All authors contributed equally to the writing of this review.

362

363 **Acknowledgements**

364 The work of R.W. and J.K.J. was funded by Laboratory Directed Research and Development (LDRD) at
365 the Pacific Northwest National Laboratory. Pacific Northwest National Laboratory is a multi-program
366 national laboratory operated by Battelle for the Department of Energy under contract DE-AC06-
367 76RL01830. The work of G.T. was supported by the US Department of Energy (DOE) Office of Science,
368 Office of Biological and Environmental Research Genomic Science program award SCW1632, by a
369 Lawrence Livermore National Laboratory LDRD 21-LW-060, and under the auspices of the DOE under
370 contract DE-AC52-07NA27344. Funding for this work was provided to N.T. by the Office of Biological
371 and Environmental Research in the DOE Office of Science—Early Career Research Program.

372

373 **Competing interest statement**

374 The authors declare no conflict of interest.

375

376 **Additional information**

377 No additional information is available for this paper.

378

379 **Twitter handles**

380 @gtrubl

381 @Livermore_Lab

382 @berkeleylab

383 @eesalbnl

384

385 @lbnlbiosci

386

387 @cloud_zero

388

389 @Ruonan61807776

390

391 @PNNLab

392

393 @doescience

394 Relevant hashtags would be: #Climate, #Soil, #Micorbiome, #Virus, #Permafrost, #Pathogen

395 **References**

396

397 1. Dobinski, W. (2011). Permafrost. *Earth-Science Reviews* 108, 158-169.

398 2. Obu, J. (2021). How Much of the Earth's Surface is Underlain by Permafrost? *Journal of Geophysical Research: Earth Surface* 126, e2021JF006123.

399 3. Mishra, U., Hugelius, G., Shelef, E., Yang, Y., Strauss, J., Lupachev, A., Harden, J.W.,
400 Jastrow, J.D., Ping, C.-L., and Riley, W.J. (2021). Spatial heterogeneity and environmental
401 predictors of permafrost region soil organic carbon stocks. *Science advances* 7,
402 eaaz5236.

404 4. Biskaborn, B.K., Smith, S.L., Noetzli, J., Matthes, H., Vieira, G., Streletskiy, D.A.,
405 Schoeneich, P., Romanovsky, V.E., Lewkowicz, A.G., and Abramov, A. (2019). Permafrost
406 is warming at a global scale. *Nature communications* 10, 1-11.

407 5. Lu, Q., Zhao, D., and Wu, S. (2017). Simulated responses of permafrost distribution to
408 climate change on the Qinghai–Tibet Plateau. *Scientific reports* 7, 1-13.

- 409 6. Landrum, L., and Holland, M.M. (2020). Extremes become routine in an emerging new
410 Arctic. *Nature Climate Change* *10*, 1108-1115.
- 411 7. Jansson, J.K., and Taş, N. (2014). The microbial ecology of permafrost. *Nature Reviews*
412 *Microbiology* *12*, 414-425.
- 413 8. Graham, D.E., Wallenstein, M.D., Vishnivetskaya, T.A., Waldrop, M.P., Phelps, T.J.,
414 Pfiffner, S.M., Onstott, T.C., Whyte, L.G., Rivkina, E.M., and Gilichinsky, D.A. (2012).
415 Microbes in thawing permafrost: the unknown variable in the climate change equation.
416 *The ISME journal* *6*, 709-712.
- 417 9. Waldrop, M., W McFarland, J., Manies, K., Leewis, M., Blazewicz, S., Jones, M.,
418 Neumann, R., Keller, J.K., Cohen, L., and Euskirchen, E. (2021). Carbon fluxes and
419 microbial activities from boreal peatlands experiencing permafrost thaw. *Journal of*
420 *Geophysical Research: Biogeosciences* *126*, e2020JG005869.
- 421 10. Taş, N., Prestat, E., McFarland, J.W., Wickland, K.P., Knight, R., Berhe, A.A., Jorgenson,
422 T., Waldrop, M.P., and Jansson, J.K. (2014). Impact of fire on active layer and permafrost
423 microbial communities and metagenomes in an upland Alaskan boreal forest. *The ISME*
424 *journal* *8*, 1904-1919.
- 425 11. Schuur, E.A., and Abbott, B. (2011). High risk of permafrost thaw. *Nature* *480*, 32-33.
- 426 12. Turetsky, M.R., Abbott, B.W., Jones, M.C., Anthony, K.W., Olefeldt, D., Schuur, E.A.,
427 Grosse, G., Kuhry, P., Hugelius, G., and Koven, C. (2020). Carbon release through abrupt
428 permafrost thaw. *Nature Geoscience* *13*, 138-143.
- 429 13. Hugelius, G., Loisel, J., Chadburn, S., Jackson, R.B., Jones, M., MacDonald, G.,
430 Marushchak, M., Olefeldt, D., Packalen, M., and Siewert, M.B. (2020). Large stocks of
431 peatland carbon and nitrogen are vulnerable to permafrost thaw. *Proceedings of the*
432 *National Academy of Sciences* *117*, 20438-20446.
- 433 14. Gross, M. (2019). *Permafrost thaw releases problems*. Elsevier.
- 434 15. Tuorto, S.J., Darias, P., McGuinness, L.R., Panikov, N., Zhang, T., Häggblom, M.M., and
435 Kerkhof, L.J. (2014). Bacterial genome replication at subzero temperatures in
436 permafrost. *The ISME journal* *8*, 139-149.
- 437 16. Hultman, J., Waldrop, M.P., Mackelprang, R., David, M.M., McFarland, J., Blazewicz, S.J.,
438 Harden, J., Turetsky, M.R., McGuire, A.D., and Shah, M.B. (2015). Multi-omics of
439 permafrost, active layer and thermokarst bog soil microbiomes. *Nature* *521*, 208-212.
- 440 17. Mykytczuk, N.C., Foote, S.J., Omelon, C.R., Southam, G., Greer, C.W., and Whyte, L.G.
441 (2013). Bacterial growth at -15 C; molecular insights from the permafrost bacterium
442 *Planococcus halocryophilus* Or1. *The ISME journal* *7*, 1211-1226.
- 443 18. Xue, Y., Jonassen, I., Øvreås, L., and Taş, N. (2019). Bacterial and archaeal metagenome-
444 assembled genome sequences from Svalbard permafrost. *Microbiology resource*
445 *announcements* *8*, e00516-00519.
- 446 19. Yang, S., Liebner, S., Walz, J., Knoblauch, C., Bornemann, T.L., Probst, A.J., Wagner, D.,
447 Jetten, M.S., and in 't Zandt, M.H. (2021). Effects of a long-term anoxic warming
448 scenario on microbial community structure and functional potential of permafrost-
449 affected soil. *Permafrost and Periglacial Processes* *32*, 641-656.
- 450 20. Hodgkins, S.B., Tfaily, M.M., McCalley, C.K., Logan, T.A., Crill, P.M., Saleska, S.R., Rich,
451 V.I., and Chanton, J.P. (2014). Changes in peat chemistry associated with permafrost

- 452 thaw increase greenhouse gas production. *Proceedings of the National Academy of*
 453 *Sciences* *111*, 5819-5824.
- 454 21. Shatilovich, A.V., Shmakova, L.A., Mylnikov, A.P., and Gilichinsky, D.A. (2009). Ancient
 455 protozoa isolated from permafrost. In *Permafrost soils*, (Springer), pp. 97-115.
- 456 22. Hedlund, C., Blomstedt, Y., and Schumann, B. (2014). Association of climatic factors with
 457 infectious diseases in the Arctic and subarctic region—a systematic review. *Global health*
 458 *action* *7*, 24161.
- 459 23. Edwards, A. (2015). Coming in from the cold: potential microbial threats from the
 460 terrestrial cryosphere. *Frontiers in Earth Science* *3*, 12.
- 461 24. Sajjad, W., Rafiq, M., Din, G., Hasan, F., Iqbal, A., Zada, S., Ali, B., Hayat, M., Irfan, M.,
 462 and Kang, S. (2020). Resurrection of inactive microbes and resistome present in the
 463 natural frozen world: Reality or myth? *Science of The Total Environment* *735*, 139275.
- 464 25. Edwards, A., Cameron, K.A., Cook, J.M., Debonnaire, A.R., Furness, E., Hay, M.C., and
 465 Rassner, S.M. (2020). Microbial genomics amidst the Arctic crisis. *Microbial genomics* *6*.
- 466 26. Zhang, G., Shoham, D., Gilichinsky, D., Davydov, S., Castello, J.D., and Rogers, S.O.
 467 (2006). Evidence of influenza A virus RNA in Siberian lake ice. *Journal of Virology* *80*,
 468 12229-12235.
- 469 27. Ng, T.F.F., Chen, L.-F., Zhou, Y., Shapiro, B., Stiller, M., Heintzman, P.D., Varsani, A.,
 470 Kondov, N.O., Wong, W., and Deng, X. (2014). Preservation of viral genomes in 700-y-old
 471 caribou feces from a subarctic ice patch. *Proceedings of the National Academy of*
 472 *Sciences* *111*, 16842-16847.
- 473 28. Mackelprang, R., Burkert, A., Haw, M., Mahendrarajah, T., Conaway, C.H., Douglas, T.A.,
 474 and Waldrop, M.P. (2017). Microbial survival strategies in ancient permafrost: insights
 475 from metagenomics. *The ISME journal* *11*, 2305-2318.
- 476 29. Wu, R., Davison, M.R., Gao, Y., Nicora, C.D., Mcdermott, J.E., Burnum-Johnson, K.E.,
 477 Hofmockel, K.S., and Jansson, J.K. (2021). Moisture modulates soil reservoirs of active
 478 DNA and RNA viruses. *Communications biology* *4*, 1-11.
- 479 30. Hough, M., McClure, A., Bolduc, B., Dorrepaal, E., Saleska, S., Klepac-Ceraj, V., and Rich,
 480 V. (2020). Biotic and Environmental Drivers of Plant Microbiomes Across a Permafrost
 481 Thaw Gradient. *Frontiers in microbiology* *11*, 796.
- 482 31. Woodcroft, B.J., Singleton, C.M., Boyd, J.A., Evans, P.N., Emerson, J.B., Zayed, A.A.,
 483 Hoelzle, R.D., Lamberton, T.O., McCalley, C.K., and Hodgkins, S.B. (2018). Genome-
 484 centric view of carbon processing in thawing permafrost. *Nature* *560*, 49-54.
- 485 32. Mondav, R., Woodcroft, B.J., Kim, E.-H., McCalley, C.K., Hodgkins, S.B., Crill, P.M.,
 486 Chanton, J., Hurst, G.B., VerBerkmoes, N.C., and Saleska, S.R. (2014). Discovery of a
 487 novel methanogen prevalent in thawing permafrost. *Nature communications* *5*, 1-7.
- 488 33. Gilichinsky, D.A. (2002). Permafrost model of extraterrestrial habitat. In *Astrobiology*,
 489 (Springer), pp. 125-142.
- 490 34. Bakermans, C., Tsapin, A.I., Souza-Egipsy, V., Gilichinsky, D.A., and Nealson, K.H. (2003).
 491 Reproduction and metabolism at– 10 C of bacteria isolated from Siberian permafrost.
 492 *Environmental microbiology* *5*, 321-326.
- 493 35. Miner, K.R., D’Andrilli, J., Mackelprang, R., Edwards, A., Malaska, M.J., Waldrop, M.P.,
 494 and Miller, C.E. (2021). Emergent biogeochemical risks from Arctic permafrost
 495 degradation. *Nature Climate Change*, 1-11.

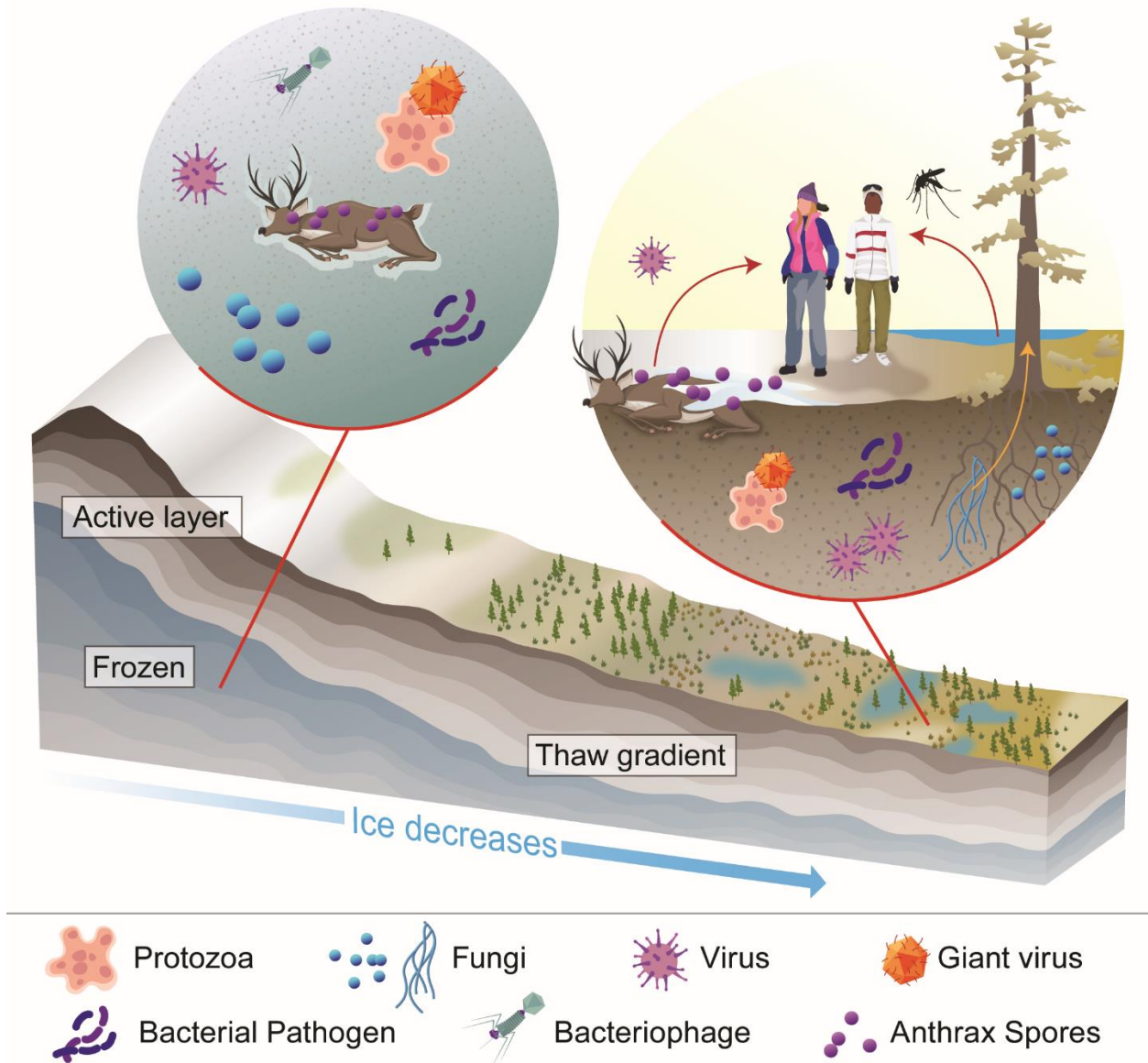
- 496 36. Zhang, D.-C., Brouchkov, A., Griva, G., Schinner, F., and Margesin, R. (2013). Isolation
497 and characterization of bacteria from ancient Siberian permafrost sediment. *Biology* 2,
498 85-106.
- 499 37. Johnston, E.R., Hatt, J.K., He, Z., Wu, L., Guo, X., Luo, Y., Schuur, E.A., Tiedje, J.M., Zhou,
500 J., and Konstantinidis, K.T. (2019). Responses of tundra soil microbial communities to
501 half a decade of experimental warming at two critical depths. *Proceedings of the*
502 *National Academy of Sciences* 116, 15096-15105.
- 503 38. Mushegian, A. (2020). Are there 10³¹ virus particles on earth, or more, or fewer?
504 *Journal of bacteriology* 202, e00052-00020.
- 505 39. Trubl, G., Kimbrel, J.A., Liquet-Gonzalez, J., Nuccio, E.E., Weber, P.K., Pett-Ridge, J.,
506 Jansson, J.K., Waldrop, M.P., and Blazewicz, S.J. (2021). Active virus-host interactions at
507 sub-freezing temperatures in Arctic peat soil. *Microbiome* 9, 1-15.
- 508 40. Emerson, J.B., Roux, S., Brum, J.R., Bolduc, B., Woodcroft, B.J., Jang, H.B., Singleton,
509 C.M., Solden, L.M., Naas, A.E., and Boyd, J.A. (2018). Host-linked soil viral ecology along
510 a permafrost thaw gradient. *Nature microbiology* 3, 870-880.
- 511 41. Trubl, G., Jang, H.B., Roux, S., Emerson, J.B., Solonenko, N., Vik, D.R., Solden, L.,
512 Ellenbogen, J., Runyon, A.T., and Bolduc, B. (2018). Soil viruses are underexplored
513 players in ecosystem carbon processing. *MSystems* 3, e00076-00018.
- 514 42. Trubl, G., Hyman, P., Roux, S., and Abedon, S.T. (2020). Coming-of-Age Characterization
515 of Soil Viruses: A User's Guide to Virus Isolation, Detection within Metagenomes, and
516 Viromics. *Soil Systems* 4, 23.
- 517 43. Koonin, E.V., Dolja, V.V., and Krupovic, M. (2015). Origins and evolution of viruses of
518 eukaryotes: the ultimate modularity. *Virology* 479, 2-25.
- 519 44. Malathi, V., and Devi, P.R. (2019). ssDNA viruses: key players in global virome.
520 *Virusdisease* 30, 3-12.
- 521 45. Trubl, G., Roux, S., Solonenko, N., Li, Y.-F., Bolduc, B., Rodríguez-Ramos, J., Eloe-Fadrosh,
522 E.A., Rich, V.I., and Sullivan, M.B. (2019). Towards optimized viral metagenomes for
523 double-stranded and single-stranded DNA viruses from challenging soils. *PeerJ* 7, e7265.
- 524 46. Bi, L., Yu, D.T., Du, S., Zhang, L.M., Zhang, L.Y., Wu, C.F., Xiong, C., Han, L.L., and He, J.Z.
525 (2021). Diversity and potential biogeochemical impacts of viruses in bulk and
526 rhizosphere soils. *Environmental microbiology* 23, 588-599.
- 527 47. Starr, E.P., Nuccio, E.E., Pett-Ridge, J., Banfield, J.F., and Firestone, M.K. (2019).
528 Metatranscriptomic reconstruction reveals RNA viruses with the potential to shape
529 carbon cycling in soil. *Proceedings of the National Academy of Sciences* 116, 25900-
530 25908.
- 531 48. Altshuler, I., Goordial, J., and Whyte, L.G. (2017). Microbial life in permafrost. In
532 *Psychrophiles: from biodiversity to biotechnology*, (Springer), pp. 153-179.
- 533 49. Malavin, S., Shmakova, L., Claverie, J.-M., and Rivkina, E. (2020). Frozen Zoo: a collection
534 of permafrost samples containing viable protists and their viruses. *Biodiversity Data*
535 *Journal* 8.
- 536 50. Legendre, M., Bartoli, J., Shmakova, L., Jeudy, S., Labadie, K., Adrait, A., Lescot, M.,
537 Poirot, O., Bertaux, L., and Bruley, C. (2014). Thirty-thousand-year-old distant relative of
538 giant icosahedral DNA viruses with a pandoravirus morphology. *Proceedings of the*
539 *National Academy of Sciences* 111, 4274-4279.

- 540 51. Pasulka, A.L., Thamatrakoln, K., Kopf, S.H., Guan, Y., Poulos, B., Moradian, A.,
541 Sweredoski, M.J., Hess, S., Sullivan, M.B., and Bidle, K.D. (2018). Interrogating marine
542 virus-host interactions and elemental transfer with BONCAT and nanoSIMS-based
543 methods. *Environmental microbiology* 20, 671-692.
- 544 52. Ramage, J., Jungsberg, L., Wang, S., Westermann, S., Lantuit, H., and Heleniak, T. (2021).
545 Population living on permafrost in the Arctic. *Population and Environment*, 1-17.
- 546 53. Karjalainen, O., Aalto, J., Luoto, M., Westermann, S., Romanovsky, V.E., Nelson, F.E.,
547 Etzelmüller, B., and Hjort, J. (2019). Circumpolar permafrost maps and geohazard indices
548 for near-future infrastructure risk assessments. *Scientific data* 6, 1-16.
- 549 54. Bjørst, L.R., and Ren, C. (2015). Steaming up or staying cool? Tourism development and
550 Greenlandic futures in the light of climate change. *Arctic Anthropology* 52, 91-101.
- 551 55. Carlson, C.J., Kracalik, I.T., Ross, N., Alexander, K.A., Hugh-Jones, M.E., Fegan, M., Elkin,
552 B.T., Epp, T., Shury, T.K., and Zhang, W. (2019). The global distribution of *Bacillus*
553 *anthracis* and associated anthrax risk to humans, livestock and wildlife. *Nature*
554 *microbiology* 4, 1337-1343.
- 555 56. Popova, A., Demina, Y.V., Ezhlova, E., Kulichenko, A., Ryazanova, A., and Maleev, V.
556 (2016). Outbreak of anthrax in the Yamalo-Nenets autonomous district in 2016,
557 epidemiological peculiarities. *Problems of Particularly Dangerous Infections* 4, 42-46.
- 558 57. Simonova, E., Kartavaya, S., Titkov, A., Loktionova, M., Raichich, S., Tolpin, V., Lupyan, E.,
559 and Platonov, A. (2017). Anthrax in the territory of Yamal: assessment of
560 epizootiological and epidemiological risks. *Problemy osobo opasnykh infektsii*, 89-93.
- 561 58. Revich, B.A., and Podolnaya, M.A. (2011). Thawing of permafrost may disturb historic
562 cattle burial grounds in East Siberia. *Global health action* 4, 8482.
- 563 59. Stella, E., Mari, L., Gabrieli, J., Barbante, C., and Bertuzzo, E. (2020). Permafrost
564 dynamics and the risk of anthrax transmission: a modelling study. *Scientific reports* 10,
565 1-12.
- 566 60. Hueffer, K., Drown, D., Romanovsky, V., and Hennessy, T. (2020). Factors contributing to
567 Anthrax outbreaks in the circumpolar north. *EcoHealth* 17, 174-180.
- 568 61. Haan, T.J., and Drown, D.M. (2021). Unearthing Antibiotic Resistance Associated with
569 Disturbance-Induced Permafrost Thaw in Interior Alaska. *Microorganisms* 9, 116.
- 570 62. Zhang, S., Yang, G., Hou, S., Zhang, T., Li, Z., and Liang, F. (2018). Distribution of ARGs
571 and MGEs among glacial soil, permafrost, and sediment using metagenomic analysis.
572 *Environmental pollution* 234, 339-346.
- 573 63. Perron, G.G., Whyte, L., Turnbaugh, P.J., Goordial, J., Hanage, W.P., Dantas, G., and
574 Desai, M.M. (2015). Functional characterization of bacteria isolated from ancient arctic
575 soil exposes diverse resistance mechanisms to modern antibiotics. *PloS one* 10,
576 e0069533.
- 577 64. D'Costa, V.M., King, C.E., Kalan, L., Morar, M., Sung, W.W., Schwarz, C., Froese, D.,
578 Zazula, G., Calmels, F., and Debruyne, R. (2011). Antibiotic resistance is ancient. *Nature*
579 477, 457-461.
- 580 65. Ozerskaya, S., Kochkina, G., Ivanushkina, N., and Gilichinsky, D.A. (2009). Fungi in
581 permafrost. In *Permafrost soils*, (Springer), pp. 85-95.
- 582 66. Bellemain, E., Davey, M.L., Kauserud, H., Epp, L.S., Boessenkool, S., Coissac, E., Geml, J.,
583 Edwards, M., Willerslev, E., and Gussarova, G. (2013). Fungal palaeodiversity revealed

- 584 using high-throughput metabarcoding of ancient DNA from arctic permafrost.
585 *Environmental microbiology* *15*, 1176-1189.
- 586 67. Schütte, U.M., Henning, J.A., Ye, Y., Bowling, A., Ford, J., Genet, H., Waldrop, M.P.,
587 Turetsky, M.R., White, J.R., and Bever, J.D. (2019). Effect of permafrost thaw on plant
588 and soil fungal community in a boreal forest: Does fungal community change mediate
589 plant productivity response? *Journal of Ecology* *107*, 1737-1752.
- 590 68. Lydolph, M.C., Jacobsen, J., Arctander, P., Gilbert, M.T.P., Gilichinsky, D.A., Hansen, A.J.,
591 Willerslev, E., and Lange, L. (2005). Beringian paleoecology inferred from permafrost-
592 preserved fungal DNA. *Applied and environmental microbiology* *71*, 1012-1017.
- 593 69. Shoham, D., Jahangir, A., Ruenphet, S., and Takehara, K. (2012). Persistence of avian
594 influenza viruses in various artificially frozen environmental water types. *Influenza*
595 *research and treatment* *2012*.
- 596 70. Fang, Q., Zhu, D., Agarkova, I., Adhikari, J., Klose, T., Liu, Y., Chen, Z., Sun, Y., Gross, M.L.,
597 and Van Etten, J.L. (2019). Near-atomic structure of a giant virus. *Nature*
598 *communications* *10*, 1-11.
- 599 71. Legendre, M., Lartigue, A., Bertaux, L., Jeudy, S., Bartoli, J., Lescot, M., Alempic, J.-M.,
600 Ramus, C., Bruley, C., and Labadie, K. (2015). In-depth study of Mollivirus sibericum, a
601 new 30,000-y-old giant virus infecting Acanthamoeba. *Proceedings of the National*
602 *Academy of Sciences* *112*, E5327-E5335.
- 603 72. Okamoto, K., Miyazaki, N., Song, C., Maia, F.R., Reddy, H.K., Abergel, C., Claverie, J.-M.,
604 Hajdu, J., Svenda, M., and Murata, K. (2017). Structural variability and complexity of the
605 giant Pithovirus sibericum particle revealed by high-voltage electron cryo-tomography
606 and energy-filtered electron cryo-microscopy. *Scientific reports* *7*, 1-12.
- 607 73. Mai-Prochnow, A., Clauson, M., Hong, J., and Murphy, A.B. (2016). Gram positive and
608 Gram negative bacteria differ in their sensitivity to cold plasma. *Scientific reports* *6*, 1-
609 11.
- 610 74. La Scola, B., Audic, S., Robert, C., Jungang, L., de Lamballerie, X., Drancourt, M., Birtles,
611 R., Claverie, J.-M., and Raoult, D. (2003). A giant virus in amoebae. *Science* *299*, 2033.
- 612 75. Ghigo, E., Kartenbeck, J., Lien, P., Pelkmans, L., Capo, C., Mege, J.-L., and Raoult, D.
613 (2008). Ameobal pathogen mimivirus infects macrophages through phagocytosis. *PLoS*
614 *pathogens* *4*, e1000087.
- 615 76. La Scola, B., Marrie, T.J., Auffray, J.-P., and Raoult, D. (2005). Mimivirus in pneumonia
616 patients. *Emerging infectious diseases* *11*, 449.
- 617 77. Hyman, P., and Abedon, S.T. (2010). Bacteriophage host range and bacterial resistance.
618 *Advances in applied microbiology* *70*, 217-248.
- 619 78. Hyman, P. (2019). Phages for phage therapy: isolation, characterization, and host range
620 breadth. *Pharmaceuticals* *12*, 35.
- 621 79. Sommers, P., Chatterjee, A., Varsani, A., and Trubl, G. (2021). Integrating Viral
622 Metagenomics into an Ecological Framework. *Annual Review of Virology* *8*.
- 623 80. El-Sayed, A., and Kamel, M. (2021). Future threat from the past. *Environmental Science*
624 *and Pollution Research* *28*, 1287-1291.
- 625 81. Stone, R. (2002). Is live smallpox lurking in the Arctic? *Science* *295*.

- 626 82. Biagini, P., Thèves, C., Balaresque, P., Geraut, A., Cagnet, C., Keyser, C., Nikolaeva, D.,
627 Gerard, P., Duchesne, S., and Orlando, L. (2012). Variola virus in a 300-year-old Siberian
628 mummy. *New England Journal of Medicine* 367, 2057-2059.
- 629 83. Tremaglio, C.Z., Barr, J.N., and Fearn, R. (2021). Genetic instability of RNA viruses. In
630 *Genome Stability*, (Elsevier), pp. 23-38.
- 631 84. Schoch-Spana, M. (2000). Implications of pandemic influenza for bioterrorism response.
632 *Clinical Infectious Diseases* 31, 1409-1413.
- 633 85. Reid, A.H., Fanning, T.G., Hultin, J.V., and Taubenberger, J.K. (1999). Origin and evolution
634 of the 1918 "Spanish" influenza virus hemagglutinin gene. *Proceedings of the National*
635 *Academy of Sciences* 96, 1651-1656.
- 636 86. Smith, O., Dunshea, G., Sinding, M.-H.S., Fedorov, S., Germonpre, M., Bocherens, H., and
637 Gilbert, M.T.P. (2019). Ancient RNA from Late Pleistocene permafrost and historical
638 canids shows tissue-specific transcriptome survival. *PLoS biology* 17, e3000166.
- 639 87. Jin, X.-Y., Jin, H.-J., Iwahana, G., Marchenko, S.S., Luo, D.-L., Li, X.-Y., and Liang, S.-H.
640 (2021). Impacts of climate-induced permafrost degradation on vegetation: A review.
641 *Advances in Climate Change Research* 12, 29-47.
- 642 88. Castello, J.D., Rogers, S.O., Starmer, W.T., Catranis, C.M., Ma, L., Bachand, G.D., Zhao, Y.,
643 and Smith, J.E. (1999). Detection of tomato mosaic tobamovirus RNA in ancient glacial
644 ice. *Polar Biology* 22, 207-212.
- 645 89. Scholthof, K.B.G., Adkins, S., Czosnek, H., Palukaitis, P., Jacquot, E., Hohn, T., Hohn, B.,
646 Saunders, K., Candresse, T., and Ahlquist, P. (2011). Top 10 plant viruses in molecular
647 plant pathology. *Molecular plant pathology* 12, 938-954.
- 648 90. Jones, R.A. (2020). Disease pandemics and major epidemics arising from new
649 encounters between indigenous viruses and introduced crops. *Viruses* 12, 1388.
- 650 91. Sokolova, O., Chashchin, V., Popova, O., Buzinov, R., Pasyukova, M., and Gudkov, A.
651 (2017). Epidemiological character of tick-borne viral encephalitis extension in the
652 Arkhangelsk region. *Ekologiya cheloveka (Human Ecology)*, 12-19.
- 653 92. Burmagina, I., Agafonov, V., and Burmagin, D. (2014). Characteristics of extreme
654 increase of vector-borne infections in the European north. *Kazan medical journal* 95,
655 731-735.
- 656 93. Waits, A., Emelyanova, A., Oksanen, A., Abass, K., and Rautio, A. (2018). Human
657 infectious diseases and the changing climate in the Arctic. *Environment international*
658 121, 703-713.
- 659 94. Reiter, P. (2001). Climate change and mosquito-borne disease. *Environmental health*
660 *perspectives* 109, 141-161.
- 661 95. Petersen, J.M., Mead, P.S., and Schriefer, M.E. (2009). *Francisella tularensis*: an
662 arthropod-borne pathogen. *Veterinary research* 40, 1.
- 663 96. Hoover, K.C., and Barker, C.M. (2016). West Nile virus, climate change, and circumpolar
664 vulnerability. *Wiley Interdisciplinary Reviews: Climate Change* 7, 283-300.
- 665 97. Pielnaa, P., Al-Saadawe, M., Saro, A., Dama, M.F., Zhou, M., Huang, Y., Huang, J., and
666 Xia, Z. (2020). Zika virus-spread, epidemiology, genome, transmission cycle, clinical
667 manifestation, associated challenges, vaccine and antiviral drug development. *Virology*
668 543, 34-42.

- 669 98. Turell, M.J., and Knudson, G.B. (1987). Mechanical transmission of *Bacillus anthracis* by
670 stable flies (*Stomoxys calcitrans*) and mosquitoes (*Aedes aegypti* and *Aedes*
671 *taeniorhynchus*). *Infection and immunity* 55, 1859-1861.
- 672 99. Semenza, J.C., Herbst, S., Rechenburg, A., Suk, J.E., Höser, C., Schreiber, C., and
673 Kistemann, T. (2012). Climate change impact assessment of food-and waterborne
674 diseases. *Critical reviews in environmental science and technology* 42, 857-890.
- 675 100. Cavicchioli, R., Ripple, W.J., Timmis, K.N., Azam, F., Bakken, L.R., Baylis, M., Behrenfeld,
676 M.J., Boetius, A., Boyd, P.W., and Classen, A.T. (2019). Scientists' warning to humanity:
677 microorganisms and climate change. *Nature Reviews Microbiology* 17, 569-586.
678



679

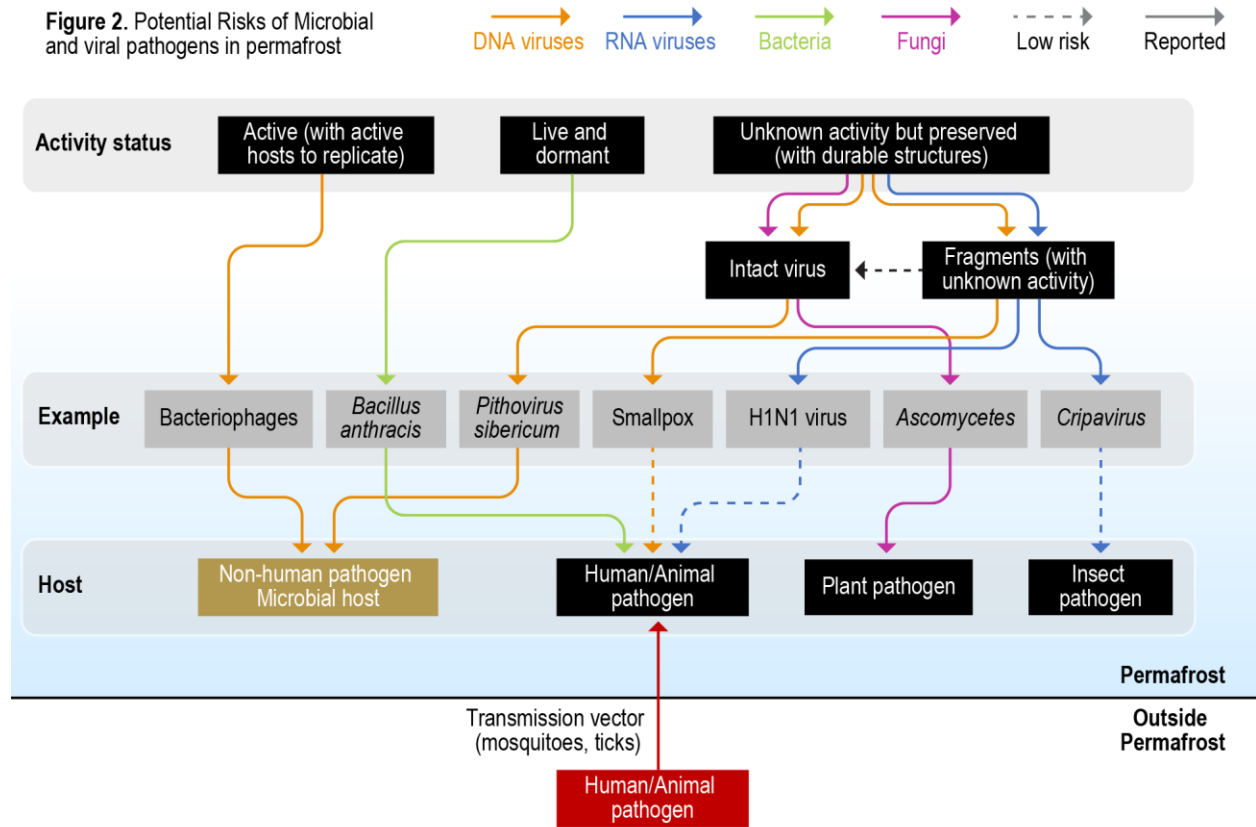
680 **Figure 1. Emergence of potential pathogens along a permafrost thaw gradient.** Left inset illustrates
 681 examples of microbes in permafrost, including non-pathogenic (e.g., ‘Bacteriophage’ and ‘Giant viruses’
 682 with bacteria and protozoa as respective hosts) and potential pathogenic microbes (e.g., spores of ‘Fungi’,
 683 ‘Anthrax spores’, other ‘Virus’). Right inset illustrates examples of pathogen emergence in thawed
 684 permafrost, including anthrax spores from animal carcasses and potential disease transmission vectors like
 685 mosquitoes.

686

687

688

689



691

692 **Figure 2. Microbes in permafrost can infect a diverse range of hosts.** Microbes in permafrost are
 693 categorized into three activity statuses: ‘Active’, ‘Live and dormant’ and ‘Unknown activity but preserved’
 694 in either intact or fragmented forms. Examples of microbes (‘DNA viruses’, orange lines; ‘RNA viruses’,
 695 blue lines; ‘Bacteria’, green lines; and ‘Fungi’, pink lines) are listed that potentially infect bacteria,
 696 protozoa, humans/animals, plants and insects. Low potential pathogenicity cases are indicated with dashed
 697 lines. Reported pathogenicity cases are indicated with solid lines.

698

699

700
701
702

Table 1. Examples of potential pathogens in permafrost and associated ice patches

*Indicates non-permafrost cryosphere samples

<i>Biological entity</i>	<i>Name</i>	<i>Disease caused</i>	<i>Type of pathogen</i>	<i>Sample detected</i>	<i>Outbreak</i>	<i>Reference</i>
<i>Bacterium</i>	<i>Bacillus anthracis</i>	Anthrax	Human or animal pathogen	Siberia permafrost	In 2016, a person and thousands of reindeer died; between 1897 and 1925, 1.5 million deer died	56-58
<i>DNA virus</i>	Variola virus	smallpox	Human or animal pathogen	Northeastern Siberia	NA	81,82
	Ancient Caribou Feces associated Virus (aCFV)	asymptomatic infection	Plant pathogen Fungal pathogen	*700-y-old caribou feces from a subarctic ice patch	NA	27
<i>Fungus</i>	Members of the genera Ascochyta, Entyloma, Epicoccum, Fusarium, Gaeumannomyces, Microdochium, Phaeosphaeria, and Phoma	Leaf spot, defoliation, and a reduction in yield and crop value	Plant pathogen	Ancient permafrost from Siberia (16,000-32,000 years old)	NA	66
	Members of Cordyceps, Paecilomyces	Paralysis or mortality	Insect pathogen	Ancient permafrost from Siberia (16,000-32,000 years old)	NA	66,68
<i>RNA virus</i>	Cripavirus	Paralysis or mortality	Insect pathogen	700-year-old caribou feces in subarctic	NA	27
	H1N1 virus	Influenza	Human or animal pathogen	frozen lung tissues of 1918 influenza victims	NA	85
	Tomato Mosaic Tobamovirus (ToMV)	Tomato mosaic disease	Plant pathogen	*140,000-year-old Greenland ice core	NA	88