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### Title

Reverse chemical ecology approach for the identification of an oviposition attractant for *Culex quinquefasciatus*

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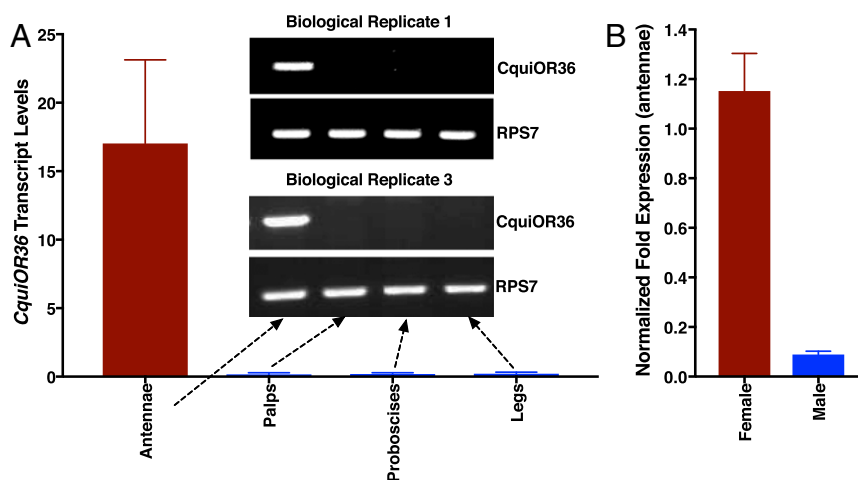
gonotrophic cycle. Although excellent progress has been made in improving trap design (see examples in refs. 4–6) and new attractants have been identified (see examples in refs. 7–9), better lures are still needed. Typically, abatement districts prepare their own fermentation recipes, which are cumbersome, have poor quality control, generate an offensive smell, and do not provide a long-term and consistent source of attractants. Since early on, several attempts have been made to isolate natural products from infusions and other natural sources (10).

To identify semiochemicals for possible applications in agriculture and medical entomology, chemical ecologists apply bioassay-guided approaches for the isolation of active ingredients, which may be augmented with electrophysiological approaches, such as gas chromatography-electroantennographic detection (GC-EAD). Although GC-EAD may provide a “shortcut” to the active ingredients, a solid and consistent bioassay is still needed to avoid false-positives (a compound may generate an electrical signal and be behaviorally inactive). The explosive advancement in our understanding of the molecular basis of insect olfaction in the last two decades has opened the door for reverse chemical ecology, a term coined for the screening of olfactory proteins as another “shortcut” toward active semiochemicals (11, 12). Two major proteins may be used for such screenings, that is, the odorant-binding proteins (OBPs) and odorant receptors (ORs). OBPs are involved in the transport of odorants through the sensillum lymph surrounding the dendrites of olfactory receptor neurons where ORs are embedded (13). ORs form ion channels with the obligatory and well-conserved coreceptor Orco, which was initially found in the fruit fly and then named OR83b (14). Semiochemicals activating OR-Orco complexes generate electrical signals that are transmitted to the brain and may ultimately lead to a behavioral response. As with GC-EAD, evidence of binding to an OBP or activation of an OR is not sufficient to infer behavioral activity. Activity must be confirmed by indoor and/or field assays. Here, we applied reverse chemical ecology in an attempt to identify semiochemicals of possible applications for monitoring or controlling populations of the southern house mosquito *Culex quinquefasciatus*. First, we identified the top 10 most-expressed OR genes in female antennae (15, 16), and then we used 7 ORs that are yet to be deorphanized, expressed them in the *Xenopus* oocyte recording system, and tested them with a panel of 230 odorants, including physiologically and behaviorally active compounds. Six of the tested ORs were silent to all tested compounds, but one of them, CquiOR36, gave the strongest responses that we have ever recorded from insect ORs to a mistakenly applied old sample of nonanal in ethanol. A careful chemical investigation showed that

CquiOR36 was responding to acetaldehyde. This was further confirmed by electroantennogram (EAG) recordings of flies carrying CquiOR36 and mosquito antennae. Behavioral assays showed that acetaldehyde is an oviposition attractant at a wide range of doses and thus has potential practical applications as a lure for gravid traps.

## Results and Discussion

**Expression Levels of the Top 10 OR Genes in *Cx. quinquefasciatus* Genome.** Previously, we identified by RNA-Seq OR genes that are significantly enriched in the antennae of the southern house mosquito compared with the legs (15) and validated our findings by analyzing expression of the top five OR genes by quantitative PCR (qPCR), which yielded transcription data in reasonable agreement with RNA-Seq analysis (15). Because the goal of this research was to complete deorphanization of the top 10 ORs and use them for reverse chemical ecology, we first ran qPCR with the top 10 ORs as selected by RNA-Seq data (15). Again, the expression pattern of most OR transcripts was in agreement with the RNA-Seq data. However, one OR in particular, CquiOR36, showed by far the highest expression levels, thus ranking number 1 by qPCR (*SI Appendix, Fig. S1*). We missed this discrepancy in our initial validation because this gene ranked as the sixth in our RNA-Seq analysis (15). Indeed, CquiOR36 expression levels by qPCR are at least threefold higher than that of CquiOR125, the OR previously ranked as number one in the *Cx. quinquefasciatus* genome (15). Next, we compared expression of CquiOR36 in olfactory and nonolfactory tissues and observed that this OR is expressed mainly in antennae, with very low levels in maxillary palps, proboscis, and legs (Fig. 1A). Moreover, CquiOR36 is expressed more in female than in male antennae (Fig. 1B). When comparing expression of olfactory genes in male and female antennae, one complication is that males have fewer olfactory sensilla than females have because olfactory sensilla are restricted to the 2 distal flagelomeres in males and distributed in all 13 flagelomeres in females (17). One way to circumvent this problem is to normalize expression using Orco as a reference. When normalized to *CquiOrco*, expression levels of *CqOR36* transcripts in female antennae were 3.74-fold higher ( $3.74 \pm 0.12$ ; mean  $\pm$  SEM) than in male antennae. Of the top 10 OR genes, CquiOR36, CquiOR125, CquiOR151, CquiOR64, CquiOR55, CquiOR132, and CquiOR93 have not been deorphanized (15, 16). By contrast, CquiOR21 (formerly CquiOR10), CquiOR95, and CquiOR136 were previously demonstrated to respond to oviposition attractants (18), ethyl 2-phenylacetate and citronellal (15), and synthetic repellents (16), respectively.

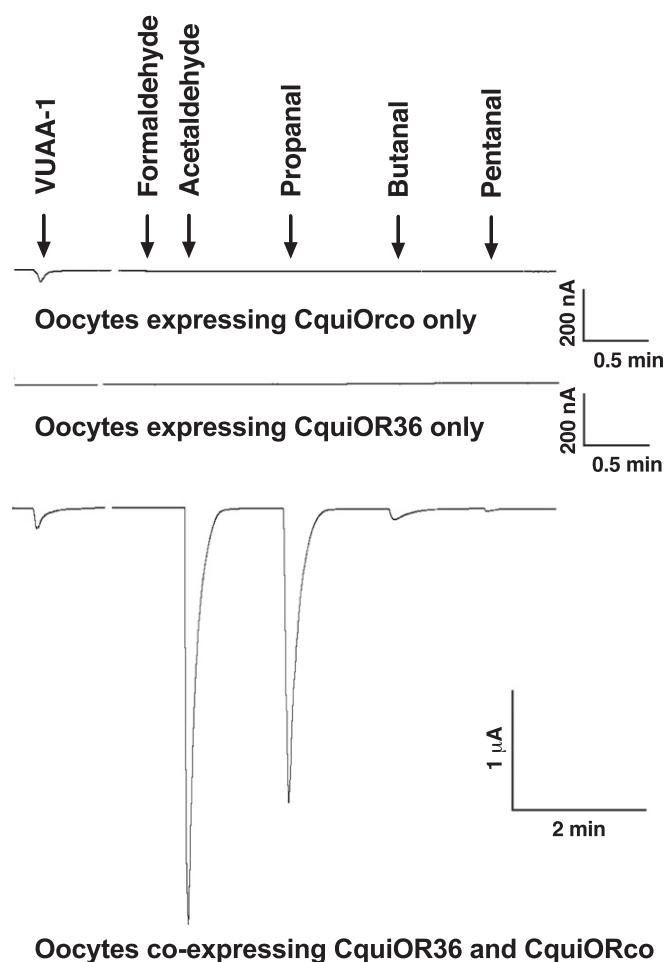


**Fig. 1.** Quantitative PCR data. (A) Transcript levels of CquiOR36 in olfactory and nonolfactory tissues from *Cx. quinquefasciatus* females. For reference, gels obtained before qPCR analysis are displayed. (B) Comparative expression of CquiOR36 in male and female antennae.

**Deorphanization Attempts.** Oocytes coexpressing CquiOR125 (GenBank accession no. KM229531), CquiOR151 (accession no. KM229534), CquiOR64 (accession no. KM229532), CuiOR55 (accession no. KM229536), CuiOR132 (accession no. KM229535), or CquiOR93 (accession no. KM229533) along with CquiOrco did not respond to any odorants in our test panel. It is not possible to conclude whether these receptors are sensitive to other compounds not included in our panel or even whether their heteromeric channels (with Orco) were properly formed. There are no current methods to unambiguously determine where heteromeric (Or-Orco) channels are formed in *Xenopus* oocytes when they failed to be activated by tested odorants. In most cases, heteromeric OR complexes are more sensitive to Orco agonists than to Orco homomers (19). Two of the best Orco agonists that may be used to infer proper channel formation are *N*-(4-ethylphenyl)-2-[(4-ethyl-5-pyridin-3-yl-1,2,4-triazol-3-yl)sulfanyl]acetamide (VUAA-1) (20) and 2-(4-ethyl-5-(4pyridinyl)-4H-1,2,4-triazol-3-yl)sulfanyl-*N*-(4-isopropylphenyl)-acetamide (OLC-12) (21). CquiOR55-CquiOrco-expressing oocytes were more sensitive to VUAA-1 than Orco homomers, thus suggesting that this receptor is functional, but the right ligands are missing in our panel. By contrast, oocytes expressing CquiOR125, CquiOR151, CquiOR64, CquiOR132, or CquiOR93 along with CquiOrco, did not elicit detectable currents when challenged with VUAA-1. We then challenged these oocytes with OLC-12 and recorded stronger responses than those elicited by VUAA-1 at the same dose (0.01 mM) in oocytes expressing CquiOR55 along with CquiOrco. We also recorded detectable currents elicited by OLC-12 on oocytes coexpressing CquiOrco with CquiOR93, CquiOR125, or CquiOR132, but CquiOR151-CquiOrco- and CquiOR64-CquiOrco-expressing oocytes were also silent to OLC-12. The receptor with highest expression levels in female antennae, CquiOR36, responded to both VUAA-1 and OLC-12 when coexpressed with CquiOrco in *Xenopus* oocytes. Previously, we considered CquiOR36 as a possible pseudogene (16), but we have now succeeded in cloning the full-length sequence of this gene by using a low annealing temperature and short primers to avoid hairpin formation. Our sequence (MG214256) differs from the sequence in VectorBase (CPIJ004162), which misses 36 internal amino acid residues. Of note, the CquiOR36 full-length sequence is contained in TCONS34486 from a previous RNA-Seq analysis (15). We therefore concluded that there could be an error in the annotation for this gene in VectorBase.

**Unusual Response of CquiOR36-CquiOrco-expressing Oocytes.** Although CquiOR36-CquiOrco-expressing oocytes were silent to our large panel of odorants, we recorded a robust current when these oocytes were mistakenly stimulated with an old solution of nonanal in ethanol. Typically, we use dimethyl sulfoxide (DMSO) as solvent, particularly in case of aldehydes that may react with the solvent and form hemiacetals. However, this was a fortuitous mistake. The current elicited by this old sample was the strongest current that we have ever recorded from oocytes, including the robust currents elicited by pheromones on pheromone receptors (22, 23). We then prepared fresh nonanal solutions using DMSO or ethanol as solvents. CquiOR36-CquiOrco-expressing oocytes were challenged with nonanal samples left at room temperature for 3 d and freshly prepared samples in these two solvents (SI Appendix, Fig. S2). These oocytes did not respond to fresh and old nonanal samples in DMSO, but again generated robust responses to old samples in ethanol and minute responses to freshly prepared nonanal samples in ethanol (SI Appendix, Fig. S2). Analysis of the nonanal samples by gas chromatography-mass spectrometry showed a small contamination with nonanoic acid. We then surmised that nonanoic acid, not nonanal, might be activating the receptor. Freshly prepared samples of nonanoic acid in DMSO did not elicit currents in the oocytes (SI Appendix, Fig. S3). Next, we tested whether hemiacetals (formed by reaction of nonanal with ethanol) might be activating the receptor. Nonanal solutions in ethanol or DMSO were kept at different conditions and tested with the same oocyte preparations. Freshly prepared samples (in ethanol) and those kept at  $-20^{\circ}\text{C}$  for 24 h elicited currents just above background levels. Of note, frozen samples had to

be brought to room temperature before use. Samples kept at  $4^{\circ}\text{C}$ , room temperature, and at  $37^{\circ}\text{C}$  generated increasing responses in this order (SI Appendix, Fig. S4). By contrast, none of the nonanal samples in DMSO kept under the conditions above generated significant currents (SI Appendix, Fig. S4). We inferred that the receptor could be responding to 1-ethoxynonan-1-ol, the hemiacetal generated by reaction of ethanol with nonanal. Because hemiacetals are unstable intermediates and, therefore, cannot be isolated, we synthesized four stable compounds having moieties resembling the structure of 1-ethoxynonan-1-ol, specifically, 3-heptyltetrahydrofuran-2-ol, 3,3-diheptyltetrahydrofuran-2-ol, 3-heptyl-5-methyltetrahydrofuran-2-ol, and 3,3-diheptyl-5-methyltetrahydrofuran-2-ol (SI Appendix, Fig. S5). None of these compounds elicited currents in the oocytes coexpressing CquiOR36 and CquiOrco (SI Appendix, Fig. S5), but these oocyte preparations responded to VUAA-1 and generated robust currents in response to 3-d-old samples of nonanal in ethanol. Next, by using solid-phase micro extraction (SPME), we analyzed old samples of nonanal in ethanol in an attempt to identify other possible contaminants. In addition to nonanal and ethanol, we detected small peaks of nonanal and acetaldehyde. We then challenged CquiOR36-CquiOrco-expressing oocytes with samples of a series of aldehydes from 1 to 10 carbons, which were dissolved in DMSO.

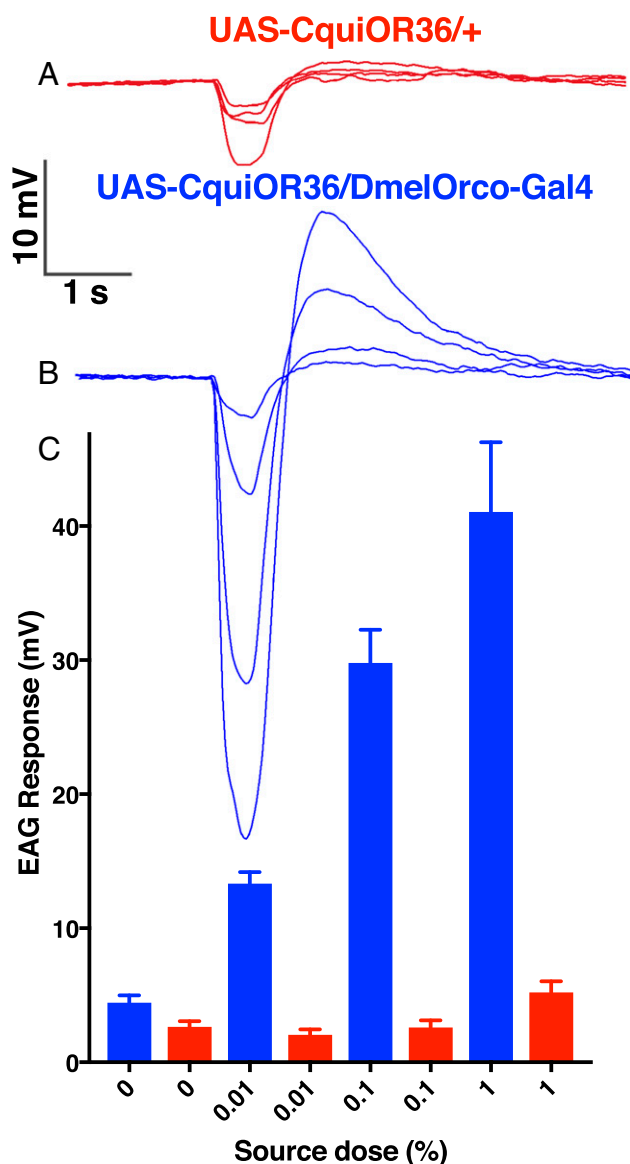


**Fig. 2.** Responses of oocytes to an Orco agonist and to a series of aldehydes. Oocytes expressing Orco only (top trace) responded to VUAA-1, but not to the aldehydes, whereas oocytes expressing CquiOR36 only (middle trace) did not respond to any tested compound. By contrast, CquiOR36-CquiOrco-expressing oocytes (lower trace) yielded robust responses to acetaldehyde and strong responses to propanal. The response to the Orco agonist was somewhat stronger in the oocytes expressing the heteromeric OR complex than the Orco homomer.

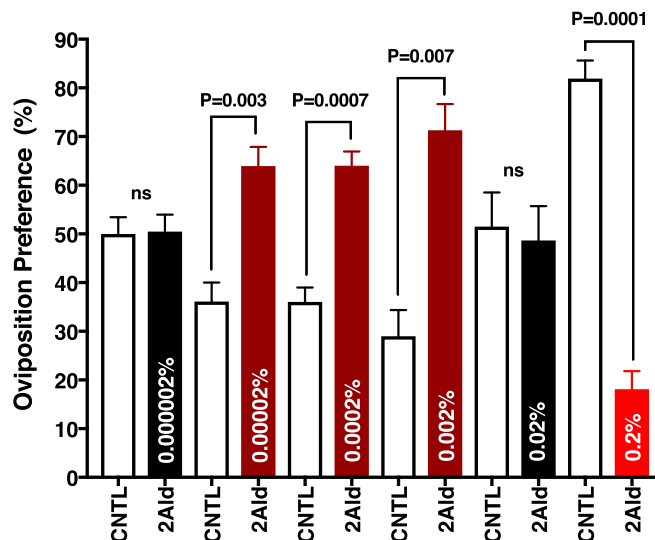


Acetaldehyde samples elicited robust responses (*SI Appendix, Fig. S6*). Propanal also generated strong responses, but none of the other aldehydes in the series, including nonanal and decanal, elicited significant responses (*SI Appendix, Fig. S6*). To ascertain that aldehyde-elicited currents were derived from CquiOR36-CquiOrco channels, we tested oocytes expressing CquiOrco only, CquiOR36 only, and those coexpressing CquiOR36 and CquiOrco (Fig. 2). Indeed, acetaldehyde and, to a lesser extent, propanal generated currents only when CquiOR36 was coexpressed with CquiOrco (Fig. 2).

**EAG Responses of Flies Expressing CquiOR36.** Using a previously reported assay (24), we expressed CquiOR36 in the fruit fly and recorded EAG responses. Flies overexpressing CquiOR36 in antennae (UAS-CquiOR36/DmelOrco-Gal4) generated robust, dose-dependent responses to acetaldehyde, whereas UAS-CquiOR36/+ flies generated minor responses even when



**Fig. 3.** EAG responses to acetaldehyde recorded from the fruit fly antennae. (A and B) Traces obtained with UAS-CquiOR36/+ and UAS-CquiOR36/DmelOrco-Gal4 flies when challenged with 0 (solvent only), 0.01, 0.1, and 1% acetaldehyde (from top to bottom). (C) Graphic representation of repetitions ( $n = 3-4$ ) of above experiments. Error bars represent SEM.



**Fig. 4.** Behavioral response from gravid females of the southern house mosquitoes to acetaldehyde in a cage oviposition assay. Each pair of bars represents one experiment with two choices: acetaldehyde (2Ald) vs. control (CNTL). Error bars represent SEM. From left to right,  $n = 15, 15, 16, 11, 16,$  and 13. Concentrations are indicated inside treatment bars. Data were analyzed with Wilcoxon matched-pair signed-rank tests.

challenged with the highest doses tested (1%) (Fig. 3). Likewise, mosquito antennae responded to acetaldehyde (*SI Appendix, Fig. S7*), thus confirming by three independent methods that both CquiOR36 is sensitive to acetaldehyde and that this semiochemical is electrophysiologically significant for mosquitoes.

**Molecular-Based Insight on Possible Biological Function.** Gene expression studies indicated that *CquiOR36* is by far the most-expressed OR in *Cx. quinquefasciatus* antennae (*SI Appendix, Fig. S1*) and that expression is restricted to female antennae (Fig. 1). Female mosquitoes rely on semiochemicals to find plants (to acquire energy for flight) and vertebrates (for a blood meal) and to locate oviposition sites. To get a better insight into how to measure mosquito behavior in response to acetaldehyde, we analyzed gene expression in an attempt to determine whether CquiOR36 is relevant for host- and blood-seeking behavior and/or oviposition behavior. We studied groups of female mosquitoes from the same cohorts, with each batch being separated into two groups. One group of 5-d-old mosquitoes received two blood meals, whereas the nonblood-fed group continued to be fed on sugar only. One, 4, and 6 d after the time of the blood meal, samples from the two groups were prepared for qPCR for comparison. Six days after the time of the blood meal, transcript levels of *CquiOR36* dropped dramatically in the groups of mosquitoes not having a blood meal (*SI Appendix, Fig. S8*), but remained unchanged in blood-fed mosquitoes that were ready for oviposition. These findings suggest that detection of acetaldehyde might be very important for oviposition behavior, but these analyses do not rule out the possibility that acetaldehyde could also be detected in a different context. We tested these two possibilities by using behavioral measurements.

**Behavioral Studies.** Using a standard cage oviposition assay (9) and testing acetaldehyde at decadic dilutions from 0.2 to  $2 \times 10^{-6}\%$ , we observed that *Cx. quinquefasciatus* females laid significantly more eggs in trays baited with acetaldehyde than in control (water only) trays (Fig. 4) at certain doses. At a very low dose ( $2 \times 10^{-6}\%$ ), there was no significant difference between treatment and control. Responses in treatments were highly significant at doses from  $2 \times 10^{-5}$  to  $2 \times 10^{-3}\%$ , but at a higher dose (0.02%) acetaldehyde was not active (Fig. 4). A clear oviposition



qPCR was performed with three biological replicates with each biological replicate replicated three times (i.e., three technical replicates per biological replicate). For comparative expression of the top 10 receptors, RNAs from female antennae and legs were used. For CquiOR36, qPCR was performed using antennae, maxillary palps, proboscis, and legs. Also, a comparison was made with RNAs from male and female antennae. The conditions for these reactions were denaturation for 3 min at 94 °C and then amplification for 30 s at 94 °C, 30 s at 55 °C, 30 s at 72 °C for 25 cycles, followed by a final 5 min of 72 °C incubation time. PCR products were loaded onto 1% agarose gels and visualized on a UV light and photographed. Data were analyzed using the 2- $\Delta\Delta$ CT method and normalized to CquiRPS7 expression levels. The mean cycle threshold (CT) of the CquiOR36 gene in female and male antennae was normalized to the mean CT of the CquiOrco gene in the respective tissues. Data were calculated using 2- $\Delta\Delta$ CT. For comparison of the CquiOR36 transcript in blood-fed and nonblood-fed mosquitoes, after normalizing with CquiRPS7, we normalized again all expression levels of each biological replicate to nonblood-fed mosquitoes 1 d after the time of the blood meal.

**Cloning of CquiOR36, -55, -64, -93, -125, -132, and -151.** Total RNA was extracted from 1,000 4- to 7-d-old female antennae with TRIzol reagent (Invitrogen). Antennal cDNA was synthesized from 1  $\mu$ g of antennal total RNA using the SMARTer RACE cDNA amplification kit according to the manufacturer's instructions (Clontech). For primers, see *SI Appendix, SI Materials and Methods*. Of note, to clone CquiOR36, short-length primers (low annealing temperature, 43 °C) were used to prevent formation of hairpin structures. PCR products were purified by a QIAquick gel extraction kit (Qiagen) and then cloned into pGEM-T vector (Promega). Of 10 colonies selected, the ones containing the target plasmids (as indicated by PCR using gene-specific primers) were extracted by a QIAprep spin mini prep kit (Qiagen) and sequenced (Davis Sequencing). For subcloning into pGEMHE, see *SI Appendix, SI Materials and Methods*.

**Electrophysiology.** The tested animals were selected from F1 progeny for *Drosophila melanogaster*, which carried both UAS-CquiOR36 and DmelOrco-Gal4 promoter (24) and nonblood-fed, 2- to 3-d-old female mosquitoes. The EAG apparatus (Syntech) was linked to a computer with an EAG2000 data acquisition interface. Recording and indifferent electrodes were made of silver wires enclosed in drawn glass capillary needles, which were filled with 1 M potassium chloride in 1% polyvinylpyrrolidone. The reference electrode was inserted in the eye of an immobilized insect, and by using a micromanipulator MP-12 (Syntech), we placed the recording electrode on the third segment of a fruit fly antenna or at the tip of the last segment of a mosquito antenna. Compounds used as stimuli were freshly dissolved in paraffin oil and loaded onto filter paper strips (1 cm<sup>2</sup>), which were placed into Pasteur pipettes. The preparation was bathed in a high-humidity air stream flowing

from a Stimulus Controller CS-55 (Syntech) at 160 mL/min to which compensatory flow or stimulus pulse (125 mL/s, 300 ms) was added. Signal from the antenna induced by stimulus or control puff was recorded for 10 s.

Two-electrode voltage-clamp technique was performed as previously described (9, 15, 16, 22, 23). Briefly, the capped cRNAs were synthesized using pGEMHE vectors and an mMESSAGE mMACHINE T7 Kit (Ambion). Purified OR cRNAs were resuspended in nuclease-free water at 200 ng/mL and microinjected with the same amount of CquiOrco cRNA into *Xenopus laevis* oocytes in stage V or VI (purchased from EcoCyte Bioscience). Then the oocytes were kept at 18 °C for 3–7 d in modified Barth's solution [NaCl 88 mM, KCl 1 mM, NaHCO<sub>3</sub> 2.4 mM, MgSO<sub>4</sub> 0.82 mM, Ca(NO<sub>3</sub>)<sub>2</sub> 0.33 mM, CaCl<sub>2</sub> 0.41 mM, Hepes 10 mM, pH 7.4] supplemented with 10 mg/mL of gentamycin and 10 mg/mL of streptomycin. Odorant-induced currents at a holding potential of -80 mV were collected and amplified with an OC-725C amplifier (Warner Instruments), low-pass-filtered at 50 Hz, and digitized at 1 kHz. Data acquisition and analysis were carried out with Digidata 1440A and pCLAMP 10 software (Molecular Devices). For the panel of odorants, see *SI Appendix, SI Materials and Methods*.

**Chemical Analyses.** For chemical synthesis, see *SI Appendix, SI Materials and Methods*. Gas chromatography-mass spectrometry was performed on a 5973 Network Mass Selective Detector linked to a 6890 GC Series Plus+ (Agilent Technologies). The GC was equipped with an HP-5MS capillary column (30 m  $\times$  0.25 mm; 0.25- $\mu$ m film; Agilent Technologies), which was operated with the following programs: 70 °C for 1 min, subsequently increased at a rate of 10 °C/min to 270 °C, and held at this final temperature for 10 min.

Injector was operated at 250 °C in pulsed splitless mode. MS transfer line was set at 280 °C, and the MS quad and MS sources were set at 150 °C and 230 °C, respectively. SPME analysis was done with 50/30 divinylbenzene/carboxen/polydimethylsiloxane (catalog # 57328-U; Supelco). Volatile compounds were collected from the headspace of the samples placed in a closed glass container with a rubber septum on the top. SPME analyses were performed with the oven starting at 35 °C for 5 min, raised to 70 °C at a rate of 2.5 °C/min, then raised at 5 °C/min to 150 °C, and subsequently raised to the final temperature of 250 °C at 10 °C/min. Under these conditions, acetaldehyde peak appeared at 1.48 min followed by ethanol peak at 1.59 min.

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- Butenandt A, Beckmann R, Stamm D, Hecker E (1959) Über Den Sexual-Lockstoff Des Seidenspinners Bombyx Mori-Reindarstellung Und Konstitution [On the sex attractant of the silkworm moth *Bombyx mori*. Isolation and structure]. *Z Naturforsch Pt B* 14: 283–284.
- World Health Organization (2012) *Handbook for Integrated Vector Management* (WHO, Geneva), p viii, 68 p.
- Johnson BJ, Ritchie SA, Fonseca DM (2017) The state of the art of lethal oviposition trap-based mass interventions for arboviral control. *Insects* 8:85.
- Cilek JE, Weston JR, Richardson AG (2017) Comparison of mosquito abundance from Biogents Sentinel 2.0 traps with and without rain covers. *J Am Mosq Control Assoc* 33: 148–150.
- Eiras AE, Buhagiar TS, Ritchie SA (2014) Development of the gravid *Aedes* trap for the capture of adult female container-exploiting mosquitoes (Diptera: Culicidae). *J Med Entomol* 51:200–209.
- Mackay AJ, Amador M, Barrera R (2013) An improved autocidal gravid ovitrap for the control and surveillance of *Aedes aegypti*. *Parasit Vectors* 6:225.
- Leal WS, et al. (2008) Reverse and conventional chemical ecology approaches for the development of oviposition attractants for *Culex* mosquitoes. *PLoS One* 3:e3045.
- Pombi M, et al. (2014) Field evaluation of a novel synthetic odour blend and of the synergistic role of carbon dioxide for sampling host-seeking *Aedes albopictus* adults in Rome, Italy. *Parasit Vectors* 7:580.
- Zhu F, Xu P, Barbosa RM, Choo YM, Leal WS (2013) RNAi-based demonstration of direct link between specific odorant receptors and mosquito oviposition behavior. *Insect Biochem Mol Biol* 43:916–923.
- Bentley MD, Day JF (1989) Chemical ecology and behavioral aspects of mosquito oviposition. *Annu Rev Entomol* 34:401–421.
- Bigiani A, Mucignat-Caretta C, Montani G, Tirindelli R (2005) Pheromone reception in mammals. *Rev Physiol Biochem Pharmacol* 154:1–35.
- Leal WS (2017) Reverse chemical ecology at the service of conservation biology. *Proc Natl Acad Sci USA* 114:12094–12096.
- Leal WS (2013) Odorant reception in insects: Roles of receptors, binding proteins, and degrading enzymes. *Annu Rev Entomol* 58:373–391.
- Larsson MC, et al. (2004) Or83b encodes a broadly expressed odorant receptor essential for *Drosophila* olfaction. *Neuron* 43:703–714.
- Leal WS, Choo YM, Xu P, da Silva CS, Ueira-Vieira C (2013) Differential expression of olfactory genes in the southern house mosquito and insights into unique odorant receptor gene isoforms. *Proc Natl Acad Sci USA* 110:18704–18709.
- Xu P, Choo YM, De La Rosa A, Leal WS (2014) Mosquito odorant receptor for DEET and methyl jasmonate. *Proc Natl Acad Sci USA* 111:16592–16597.
- Clements AN (1999) *The Biology of Mosquitoes: Sensory Reception and Behaviour* (CABI Publishing, New York).
- Hughes DT, Pelletier J, Luetje CW, Leal WS (2010) Odorant receptor from the southern house mosquito narrowly tuned to the oviposition attractant skatole. *J Chem Ecol* 36: 797–800.
- Hughes DT, et al. (2017) Functional and nonfunctional forms of CquiOR91, an odorant selectivity subunit of *Culex quinquefasciatus*. *Chem Senses* 42:333–341.
- Jones PL, Pask GM, Rinker DC, Zwiebel LJ (2011) Functional agonism of insect odorant receptor ion channels. *Proc Natl Acad Sci USA* 108:8821–8825.
- Chen S, Luetje CW (2012) Identification of new agonists and antagonists of the insect odorant receptor co-receptor subunit. *PLoS One* 7:e36784.
- Xu P, et al. (2012) Moth sex pheromone receptors and deceitful parapheromones. *PLoS One* 7:e41653.
- Xu P, Hooper AM, Pickett JA, Leal WS (2012) Specificity determinants of the silkworm moth sex pheromone. *PLoS One* 7:e44190.
- Ueira-Vieira C, Kimbrell DA, de Carvalho WJ, Leal WS (2014) Facile functional analysis of insect odorant receptors expressed in the fruit fly: Validation with receptors from taxonomically distant and closely related species. *Cell Mol Life Sci* 71:4675–4680.
- Leal HM, Hwang JK, Tan K, Leal WS (2017) Attraction of *Culex* mosquitoes to aldehydes from human emanations. *Sci Rep* 7:17965.
- Syed Z, Leal WS (2008) Mosquitoes smell and avoid the insect repellent DEET. *Proc Natl Acad Sci USA* 105:13598–13603.