

UCLA

UCLA Previously Published Works

Title

Methionine adenosyltransferases in liver cancer

Permalink

<https://escholarship.org/uc/item/4sr0m3rw>

Journal

World Journal of Gastroenterology, 25(31)

ISSN

1007-9327

Authors

Murray, Ben

Barbier-Torres, Lucia

Fan, Wei

et al.

Publication Date

2019-08-21

DOI

10.3748/wjg.v25.i31.4300

Copyright Information

This work is made available under the terms of a Creative Commons Attribution-NonCommercial License, available at <https://creativecommons.org/licenses/by-nc/4.0/>

Peer reviewed

Methionine adenosyltransferases in liver cancer

Ben Murray, Lucia Barbier-Torres, Wei Fan, José M Mato, Shelly C Lu

ORCID number: Ben Murray (0000-0001-5375-2124); Lucia Barbier-Torres (0000-0002-7675-2888); Wei Fan (0000-0001-6462-7425); Jose M Mato (0000-0003-1264-3153); Shelly C Lu (0000-0003-2128-5407).

Author contributions: Ben Murray and Lucia Barbier-Torres contributed equally in reviewing the literature and drafting of the manuscript. Wei Fan assisted in literature review and drafting of the manuscript. José M Mato provided critical reading of the manuscript. Shelly C Lu provided critical editing and revision of the manuscript. All authors approved the final version of the manuscript.

Supported by National Institutes of Health, NIAAA, No. R01AA026759 (Lu); National Institutes of Health, NIDDK, No. R01DK107288 (Lu); National Institutes of Health, NCCIH, No. R01AT001576; National Institutes of Health, NCI, No. R01CA172086 (Lu and Mato); Agencia Estatal de Investigación MINECO, No. SAF 2017-88041-R; ISCIII PIE14/00031, No. CIBERehd-ISCIII; and Severo Ochoa Excellence Accreditation, No. SEV-2016-0644 (Mato).

Conflict-of-interest statement: None of the authors have any conflict of interest.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially,

Ben Murray, Lucia Barbier-Torres, Wei Fan, Shelly C Lu, Division of Digestive and Liver diseases, Department of Medicine, Cedars-Sinai Medical Center, Los Angeles, CA 90048, United States

José M Mato, CIC bioGUNE, Centro de Investigación Biomédica en Red de Enfermedades Hepáticas y Digestivas (Ciberehd), Technology, Park of Bizkaia, Derio 48160, Bizkaia, Spain

Corresponding author: Shelly C Lu, MD, Professor, Division of Digestive and Liver Diseases, Department of Medicine, Cedars-Sinai Medical Center, Davis Building, Room No. 2097, 8700 Beverly Blvd., Los Angeles, CA, 90048, United States. shelly.lu@cshs.org

Telephone: +1-310-4235692

Fax: +1-310-4230653

Abstract

Methionine adenosyltransferases (MATs) are essential enzymes for life as they produce S-adenosylmethionine (SAME), the biological methyl donor required for a plethora of reactions within the cell. Mammalian systems express two genes, *MAT1A* and *MAT2A*, which encode for MAT α 1 and MAT α 2, the catalytic subunits of the MAT isoenzymes, respectively. A third gene *MAT2B*, encodes a regulatory subunit known as MAT β which controls the activity of MAT α 2. *MAT1A*, which is mainly expressed in hepatocytes, maintains the differentiated state of these cells, whilst *MAT2A* and *MAT2B* are expressed in extrahepatic tissues as well as non-parenchymal cells of the liver (e.g., hepatic stellate and Kupffer cells). The biosynthesis of SAME is impaired in patients with chronic liver disease and liver cancer due to decreased expression and inactivation of MAT α 1. A switch from *MAT1A* to *MAT2A/MAT2B* occurs in multiple liver diseases and during liver growth and dedifferentiation, but this change in the expression pattern of MATs results in reduced hepatic SAME level. Decades of study have utilized the *Mat1a*-knockout (KO) mouse that spontaneously develops non-alcoholic steatohepatitis (NASH) and hepatocellular carcinoma (HCC) to elucidate a variety of mechanisms by which MAT proteins dysregulation contributes to liver carcinogenesis. An increasing volume of work indicates that MATs have SAME-independent functions, distinct interactomes and multiple subcellular localizations. Here we aim to provide an overview of MAT biology including genes, isoenzymes and their regulation to provide the context for understanding consequences of their dysregulation. We will highlight recent breakthroughs in the field and underscore the importance of MAT's in liver tumorigenesis as well as their potential as targets for cancer therapy.

Key words: Methionine adenosyltransferases; S-adenosylmethionine; Liver cancer; Hepatocellular carcinoma; Cholangiocarcinoma; Biomarkers; Therapeutic targets

and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Received: April 13, 2019

Peer-review started: April 15, 2019

First decision: May 16, 2019

Revised: May 31, 2019

Accepted: July 19, 2019

Article in press: July 19, 2019

Published online: August 21, 2019

P-Reviewer: Guo K, Trovato GMM

S-Editor: Ma YJ

L-Editor: A

E-Editor: Ma YJ



©The Author(s) 2019. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: In this review we provide the most comprehensive guide to methionine adenosyltransferases discussing their structures, functions and consequences of dysregulation in liver cancers emphasizing their potential as prognostic biomarkers for liver cancers and as targets for liver cancer therapy.

Citation: Murray B, Barbier-Torres L, Fan W, Mato JM, Lu SC. Methionine adenosyltransferases in liver cancer. *World J Gastroenterol* 2019; 25(31): 4300-4319

URL: <https://www.wjgnet.com/1007-9327/full/v25/i31/4300.htm>

DOI: <https://dx.doi.org/10.3748/wjg.v25.i31.4300>

INTRODUCTION

This review examines the roles of methionine adenosyltransferases (MATs) in liver cancers with a focus on how dysregulation of these proteins contributes to their pathogenesis. Hepatocellular carcinoma (HCC) is the most common primary liver cancer and currently the second leading cause of cancer-related death worldwide^[1]. In the majority of the cases, HCC develops in patients with underlying chronic liver disease and cirrhosis, which are mainly derived from viral hepatitis infection, alcohol abuse and non-alcoholic fatty liver disease (NAFLD)^[2]. The threat of HCC is expected to continue rising due to increasing cases of NAFLD given the obesity epidemic occurring worldwide^[3]. Even more alarming are reports of HCC from NAFLD that occurred in the absence of cirrhosis^[4]. Cholangiocarcinoma (CCA) is the second most common primary liver cancer, which typically occur in the setting of chronic biliary inflammation. MATs exert very similar roles in both types of liver cancers.

OVERVIEW OF MAT ENZYMES

MAT genes and isoenzymes

MATs (EC 2.5.1.6) belong to a family of enzymes that are essential to life as they catalyze the biosynthesis of SAME, the main methyl donor of the cell. Apart from methylation, SAME is also important as a precursor in glutathione (GSH) and polyamine synthesis^[5]. As SAME is used for the methylation of biomolecules including DNA, RNA, proteins, biological amines and phospholipids, any change in its biosynthesis and catabolism within the cell has a profound effect on cellular processes such as growth, differentiation and response to injury. Over the past decade SAME-independent roles of MAT enzymes have emerged, most notably, their ability to function as *bona fide* transcription factors as well as their ability to form parts of scaffold complexes which have been implicated in cancer development^[6-9].

In mammals, three distinct genes, *MAT1A*, *MAT2A*, and *MAT2B* give rise to the protein products MAT α 1, MAT α 2, and MAT β , respectively (Table 1, Figure 1)^[10]. *MAT1A* encodes for a 395-amino acid (396 in mouse, and 397 in rat) catalytic subunit in humans that is mainly expressed in hepatocytes (parenchymal cells of the liver) as well as bile duct epithelial cells and pancreatic acinar cells^[8,11]. MAT α 1 can oligomerize to form a homotetramer (MATIII) or homodimer (MATI). The MAT α 2 subunit, the extrahepatic catalytic protein (395 amino acids), that has an 84% sequence identity to MAT α 1, can also form dimers and tetramers but with a bias towards the dimer conformation^[10,12,13]. MAT α 2 is expressed in non-parenchymal cells of the liver as well as in all extrahepatic tissues. The regulatory subunit MAT β has four known isoforms, MAT β V1, MAT β V2, MAT β V2a, and MAT β V2b with the former two being the major splice variants^[14]. MAT β V2a and MAT β V2b are expressed at very low levels compared to MAT β V2 and have not been studied in detail^[14]. MAT β V1 is expressed in fetal liver, prostate, lung, brain, thyroid and the adrenal gland, whilst MAT β V2 is expressed in skeletal muscle and heart. Both isoforms are expressed in the kidney and thymus^[14]. MAT β V1 and MAT β V2 are 331 and 323 amino acids long, respectively, differ only by 20 amino acids at their N-terminus and share little sequence identity (7%) to the catalytic MAT proteins. MAT β has a binding pocket for the cofactor NADP⁺, which can interact with both major isoforms^[12,15].

MAT expression patterns and subcellular localization

Table 1 Mammalian MAT genes and isoenzymes

Gene	Protein	Amino acids	Alternative names	Oligomeric state	Regulatory subunit	Co-factors	K_m for methionine	K_m for ATP	K_i for SAME	X-ray structure
MAT1A	MAT α 1	395	MATIII	Dimer	No		210 μ mol/L- 7 mmol	1-3 mmol	None	20BV
			MATI MAT(α 1) ₄	Tetramer	No		23 μ mol/L-1 mmol	0.2-0.5 mmol	400 μ mol/L	
MAT2A	MAT α 2	395	MAT(α 2) ₂	Dimer			4-10 μ mol/L	70 μ mol/L	60 μ mol/L	5A19
			MAT(α 2) ₄	Tetramer						
MAT2B	MAT β V1	334		Monomeric		NADP ⁺				
	MAT β V2	323		Monomeric		NADP ⁺				2YDX

Historically, MATIII in the literature refers to complexes of MAT α 2 and MAT β .

MAT enzymes were first thought to function as SAME producing factories in the cytosol, and that SAME would be delivered to the specific compartments such as the nucleus or mitochondria for methylation reactions^[16]. However, a decade ago MAT α 1 was reported to be present within the nucleus^[17]. This was followed by publications describing MAT α 2, MAT β V1 and MAT β V2 to also be within this organelle^[6,18]. These publications showed that a nuclear location of MAT proteins was associated with enhanced histone H3K27 methylation, an epigenetic modification that leads to gene silencing^[17]. Most recently MAT α 1 was reported to be present in the mitochondrial matrix of hepatocytes, enhancing mitochondrial function and negatively regulating cytochrome P450 2E1 (CYP2E1) through methylation^[19]. These accumulating publications reinforce the concept that MATs are recruited to subcellular compartments to provide a local source of SAME.

Structural overview of MATs and MAT complexes

To date there are numerous crystal structures of MAT proteins from different species in different active site conformations, complexed with substrates, products, or their analogues^[12,20-23]. The catalytic proteins have a three-domain organization that is conserved amongst other MAT family subunits^[21,22,24]. Monomeric MAT enzymes are incapable of producing SAME as they do not contain a complete active site. Upon dimerization, both monomers contribute residues to form two active sites. The large hydrophobic surface of monomeric MAT constitutes the site of the monomer-monomer interface. A common feature of MAT enzymes is that they contain a “gating loop” (in human MAT α 1 and α 2 residues 113-131) that flanks the active site, which has been hypothesized to move dynamically to allow access to the active site^[21]. When the active site is occupied, the loop closes to form a gate over the active site, but when the active site is empty, the loop becomes disordered or open. MAT α 1 dimers can also form tetramers through the central domain of the subunits^[24]. Recent work has shown that mutation to residues of the gating loop can reduce enzyme activity and SAME formation^[25].

The regulatory β subunit is structurally very different from the α subunits, unable to produce SAME by itself. MAT β proteins contain signature motifs of the SDR (short-chain dehydrogenase/reductase) superfamily including a Rossmann fold that can bind FAD⁺ or NADP⁺ although MAT β favors the latter^[22]. MAT α 2 and MAT β interact to give rise to the MAT α 2 β complexes (also referred to as MATII)^[12,26] (Figure 1, Table 1). To date only the structure of a MAT α 2 β V₂ complex has been solved and it consists of a MAT α 2 tetramer flanked by two MAT β V2 subunits (MAT α 2₍₄₎ β V2₍₂₎). This showed that MAT α 2 can exist and function as a tetramer in the presence of MAT β ^[12]. The oligomeric state of this crystal structure, confirmed with small angle x-ray scattering^[12], is different from the suggested tetrameric form [MAT(α 2)₂(β)₂]^[27] or the proposed computational model in which MAT α β was assumed to be a trimer [MAT(α 2)₂(β)₁]^[15]. Mutational analysis showed by gel filtration that the minimum motif required for the formation of the MAT(α 2)₄(β V2)₂ complex comprises three residues at the end of the C-terminus of MAT β V2 (V₃₂₁F₃₂₂H₃₂₃)^[12]. Several publications, using recombinant purified proteins, have shown that MAT α 1 can also interact with MAT β V1, although this interaction is several orders of magnitude weaker than that of MAT α 2 and MAT β ^[12,25]. The MAT α 1 β V1 complex is not likely to occur within the cell as MAT α 1 and MAT β are generally not expressed at the same time.

MAT enzymatic mechanism

The wealth of structures available for MAT enzymes, as well as the range of

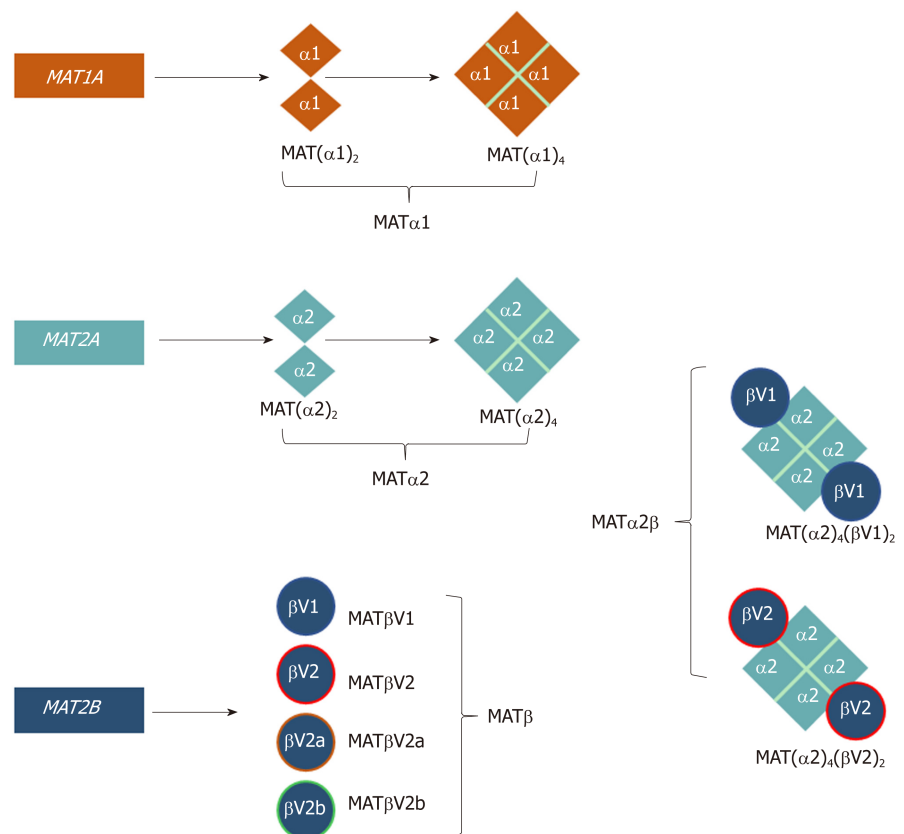


Figure 1 Schematic representation of the oligomeric states of mammalian MAT enzymes. *MAT1A* and *MAT2A* genes encode the catalytic subunits $\text{MAT}\alpha 1$ and $\text{MAT}\alpha 2$, respectively. Both $\text{MAT}\alpha 1$ and $\text{MAT}\alpha 2$ can be organized as dimers and tetramers. *MAT2B* encodes the regulatory subunit for which there are four isoforms, $\text{MAT}\beta V1$, $\text{MAT}\beta V2$, $\text{MAT}\beta V2a$, and $\text{MAT}\beta V2b$ with the former two being the major splice variants. $\text{MAT}\alpha 2$ and $\text{MAT}\beta$ interact to give rise to the $\text{MAT}\alpha 2\beta$ complexes. The $\text{MAT}(\alpha 2)_4(\beta V1)_2$ and $\text{MAT}(\alpha 2)_4(\beta V2)_2$ complexes consist of a $\text{MAT}\alpha 2$ tetramer flanked by two $\text{MAT}\beta V1$ or $\text{MAT}\beta V2$ subunits.

biochemical evidence, has provided terrific detail and insight into the enzymatic mechanism. S_{AMe} is produced by the addition of the amino acid methionine to the energy molecule ATP (Figure 2). Upon entry of the substrates ATP and methionine into the active site, the flexible gating loop becomes well-ordered closing the active site. The synthesis of S_{AMe} follows an S_N2 catalytic mechanism^[28,29], whereby the sulphur atom of methionine attacks the C5' atom of ATP displacing the triphosphosphate (PPPi) moiety to form S_{AMe}. The PPPi is then hydrolyzed giving rise to pyrophosphate (PPi) and orthophosphate (Pi), providing the energy to facilitate product release by dislodging the gating loop^[30]. For a detailed mechanism, see Komoto *et al.*^[21] 2004 and Murray *et al.*^[23] 2016.

MAT activity

Despite high sequence identity MAT enzymes exhibit different kinetic and regulatory properties for methionine, ATP, and S_{AMe}. The K_m for the methionine is lowest in $\text{MAT}\alpha 2$ followed by $\text{MAT}(\alpha 1)_2$ and is the highest for $\text{MAT}(\alpha 1)_4$ ^[5]. The K_m for ATP is also highest for $\text{MAT}(\alpha 1)_4$ (1–2 mmol), intermediate for $\text{MAT}(\alpha 1)_2$ (0.2–0.5 mmol), and lowest for $\text{MAT}\alpha 2$ (70 μM)^[31,32]. S_{AMe}, the product produced by MAT, can act as a feedback inhibitor to some of these enzymes^[33]. $\text{MAT}\alpha 2$ is the most sensitive to S_{AMe} with a 50% inhibitory concentration (IC_{50}) of 60 μM which equates to the normal physiological liver levels. $\text{MAT}(\alpha 1)_4$ is minimally inhibited by S_{AMe} (IC_{50} = 400 $\mu\text{mol/L}$) whilst $\text{MAT}(\alpha 1)_2$ can be stimulated eight-fold by high S_{AMe} levels (500 $\mu\text{mol/L}$)^[33]. These differences between $\text{MAT}\alpha 1$ and $\text{MAT}\alpha 2$ are important to allow $\text{MAT}\alpha 1$ to maintain a high S_{AMe} production in the liver (produces 6–8 g/day) compared with $\text{MAT}\alpha 2$, which does not contribute significantly to this S_{AMe} pool^[34]. Indeed, by expressing $\text{MAT}\alpha 1$ the liver is able to catabolize 50% of methionine intake via conversion to S_{AMe} and allow an up to 10-fold rise in S_{AMe} level following a methionine rich meal^[35]. Consistent with this, cells that express $\text{MAT}\alpha 1$ have much higher steady-state S_{AMe} levels than cells that express $\text{MAT}\alpha 2$ ^[36]. When either major isoform of $\text{MAT}\beta$ interacts with $\text{MAT}\alpha 2$ they increase the k_{cat} (turnover rate of an

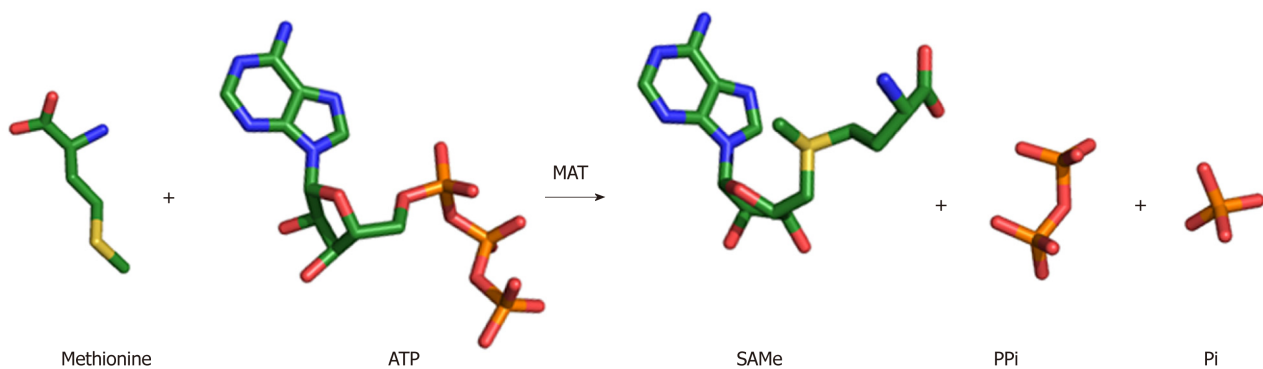


Figure 2 SAMe synthesis reaction. MAT enzyme catalyzes the biosynthesis of SAMe from the amino acid methionine and the energy molecule ATP. The sulphur atom of methionine attacks the C5' atom of ATP displacing the triphosphosphate (PPPi) moiety to form SAMe. The PPPi is then hydrolyzed giving rise to pyrophosphate (PPI) and orthophosphate (Pi).

enzyme-substrate complex) of the MAT $\alpha\beta$ complexes^[12,37,38] and increase the susceptibility to feedback inhibition by SAMe^[37].

DYSREGULATION OF MATS IN LIVER DISEASE AND CANCER

Many studies have demonstrated that MAT proteins play important roles in chronic liver disease and hepatocarcinogenesis and a switch in their expression pattern is a frequent event in liver cancers. *MAT1A*, which is mainly expressed in the liver and maintains the differentiated state of hepatocytes, is downregulated in most cirrhotic patients^[39], in patients with alcoholic hepatitis^[40], during de-differentiation and in HCC^[41,42]. Conversely, *MAT2A* and *MAT2B*, which are normally expressed only by non-parenchymal cells of the liver and extrahepatic tissues, are induced in HCC^[14,42,43]. This *MAT1A* to *MAT2A/MAT2B* switch contributes to reduced SAMe levels and is an important determinant of liver injury, fibrosis and liver cancer development in both rodents and humans^[5].

MAT gene regulation and dysregulation in HCC

While *MAT1A* is a marker for normal differentiated liver, *MAT2A* and *MAT2B* are markers for rapid liver growth and de-differentiation. *MAT2A* and *MAT2B* are transcriptionally induced in human HCC, during rapid liver growth, de-differentiation, and in response to ethanol feeding in rodents^[44-46]. Reduced hepatic MAT activity has also been observed in cirrhotic patients, which explains why many cirrhotic patients have hypermethioninemia^[39]. In human HCC, the *MAT1A*:*MAT2A* expression ratio has been inversely correlated with cell growth and genomic instability and directly correlated with HCC apoptosis and overall DNA methylation; a reduced ratio is a prognostic marker of more malignant and lower survival HCCs^[47]. MATs are regulated at transcriptional, post-transcriptional and post-translational levels by a complex network of mechanisms (Table 2). Many of these are dysregulated in HCC and participate to alter MATs expression.

MAT1A transcriptional and epigenetic control

The *MAT1A* promoter contains binding sites for multiple transcription factors, including hepatocyte nuclear factor (HNF), activator protein 1 (AP-1), CCAAT enhancer binding protein (C/EBP), c-MYC and glucocorticoids^[48]. Some of these factors are determinants of liver-specific gene expression of *MAT1A*, such as HNF and C/EBP, with the latter also control *MAT1A* expression by promoter regulation^[48,49]. Prohibitin 1 (PHB1), which is highly expressed in normal hepatocytes and downregulated in most HCCs, positively regulates *MAT1A* mRNA levels^[50]. Finally, c-MYC, MAFG and c-MAF, transcription factors that are overexpressed in human HCC and CCA, have been shown to bind to a repressive E-box element in the human *MAT1A* promoter to downregulate *MAT1A* transcription^[8,9].

MAT1A expression is also regulated by DNA epigenetic modifications. Lower *MAT1A* expression in HCC has been associated with promoter hypermethylation and histone H4 deacetylation of its promoter^[47,51]. Further investigation revealed a 750-base pair (bp) region upstream of the *MAT1A* transcriptional start site for these epigenetic

Table 2 Regulatory mechanisms of human *MAT* genes and proteins

	MAT1A	MAT2A	MAT2B
Transcriptional regulation	Glucocorticoids (+)	CpG hypomethylation (+)	AP-1 (+)
	C/EBP (+)	Histone H4 acetylation (+)	NFκB (+)
	CpG hypermethylation (-)	c-MYB (+)	SIRT1 (+)
	Histone H4 deacetylation (-)	SP-1 (+)	
	c-MYC (-)	AP-1 (+)	
	MAFG (-)	NFκB (+)	
	c-MAF (-)	HIF1α (+)	
		PPARγ (-)	
		PPARβ (+)	
		HBx (+)	
		CREB (+)	
Post-transcriptional regulation	AUF1 (-)	HuR (+)	HuR (+)
	miR-485-3p (-)	Methylated-HuR (-)	miR-21-3p (+)
	miR-495 (-)	miR-21-3p (+)	
	miR-664 (-)	miR-34a (+)	
		miR-34b (+)	
Post-translational regulation	Phosphorylation (-)	Phosphorylation (+)	Phosphorylation (+)
	Nitrosylation (-)	Sumoylation (+)	GIT1 interaction (+)
	Oxidation (-)	Acetylation (-)	MATα2 interaction (+)
		MATβ interaction (+)	

(+): Positive regulation; (-): Negative regulation.

modifications^[51]. In HepG2 cells and cirrhotic human livers, hypermethylation of sites +10 and +88, relative to the transcriptional start site have been reported to also downregulate *MAT1A* transcription^[52]. Low *MAT1A* mRNA levels and hypermethylation of both the *MAT1A* promoter and coding regions were also reported in patients with advanced NAFLD^[53].

***MAT1A* post-transcriptional control**

Binding of AU-rich RNA binding factor (AUF1) to the 3'-untranslated region (UTR) of *MAT1A* mRNA negatively regulates its stability. There is an inverse correlation between AUF1 and *Mat1a* expression; de-differentiation of rat hepatocytes in culture increases the expression of AUF1, contributing to the fall in *Mat1a* mRNA levels, whereas during liver development AUF expression falls, which coincide with increased *Mat1a* expression; AUF1 is highly expressed in human HCC and its knockdown increased *MAT1A* mRNA levels^[54].

MAT1A mRNA is also regulated by microRNAs (miRNAs) in HCC^[55,56]. Injection of 2-acetylaminofluorene in rats resulted in preneoplastic liver lesions, induction of both miR-22 and miR-29b and inhibition of *Mat1a* mRNA expression^[55]. MiR-485-3p, miR-495, and miR-664, which are increased in human HCC and responsible for the induction of LIN28B, an oncoprotein that is overexpressed in HCC and represses the tumor suppressor Let-7, have been shown to negatively regulate *MAT1A*. These specific miRNAs, through the downregulation of *MAT1A*, lowered nuclear SAME levels, leading to hypomethylation of the *LIN28B* promoter region and increased LIN28B expression^[56]. Inhibition of these miRNAs reduced tumor growth *in vitro* and *in vivo* by recovering *MAT1A* expression and inducing apoptosis^[56].

***MAT2A/MAT2B* transcriptional and epigenetic control**

MAT2A transcription is upregulated during liver growth and de-differentiation^[57-59]. Like *MAT1A*, *MAT2A* is also regulated by multiple transcription factors including c-MYB, E2F and specificity protein 1 (SP-1), all of which increase its promoter activity^[57,59]. The *MAT2A* promoter can also be induced by tumor necrosis factor-α (TNF-α) via nuclear factor κβ (NF-κβ) and AP-1 elements in the promoter region^[60]. Transforming growth factor β1 (TGF-β1) also increases the activity of the *MAT2A* promoter via NF-κβ in hepatic stellate cells (HSC)^[61]. Multiple PPAR response elements (PPRE) that bind nuclear receptors including peroxisome-proliferator activated receptors (PPAR) are present in the rat *Mat2a* promoter^[62]. In normal liver PPARγ is a marker of HSC quiescence, whilst PPARβ is induced in activated HSCs

during liver fibrogenesis^[63,64]. Both PPAR γ and PPAR β occupy the same site on the *Mat2a* promoter to cause opposite effects^[62]. PPAR γ negatively regulate *Mat2a* transcription but during HSC activation PPAR γ expression and activity fall, allowing PPAR β to bind instead and induce *Mat2a* expression^[62]. The *MAT2A* promoter is also regulated by methylation and acetylation and in human HCC, *MAT2A* promoter is hypomethylation and associate with higher histone H4 acetylation^[58]. Expression of *MAT2A* is also induced in a hypoxic tumor environment because hypoxia-inducible factor-1 α (HIF-1 α) binds to a consensus sequence within the *MAT2A* promoter activating its expression in human hepatoma cells^[65]. Finally, hepatitis B X protein (HBx) was shown to activate *MAT2A* gene transcription by facilitating NF- κ B and CREB binding to the *MAT2A* promoter, explaining *MAT2A* induction in HBV-associated HCC^[66].

Mechanism of *MAT2B* transcriptional regulation remains poorly characterized. In the human liver cancer cell line HepG2, TNF- α can upregulate *MAT2B-V1* mRNA but not *MAT2B-V2* through an AP-1 and NF- κ B dependent mechanisms^[14]. Sirtuin 1 (SIRT1), a NAD⁺-dependent deacetylase, can also induce the expression of *MAT2B*^[67]. *MAT2B-V1* mRNA expression has also been found to be regulated by leptin in HepG2 cells by mechanisms that involve extracellular signal regulated kinase (ERK) and AKT^[68].

***MAT2A/MAT2B* post-transcriptional control**

The stability of *MAT2A* mRNA can be influenced by the human RNA-binding (HuR) protein and its methylated form, methyl-HuR^[54]. The function of HuR depends on the methylated state of the protein with methyl-HuR destabilizing target mRNAs and HuR stabilizing them^[54]. During hepatocyte de-differentiation as well as HCC, HuR is induced but there is a decline in methyl-HuR, which results in a higher HuR/methyl-HuR ratio. For *MAT2A* this causes increased binding of HuR to *MAT2A* mRNA stabilizing it in HCC and de-differentiated hepatocytes^[54]. HuR has also been shown to stabilize *MAT2B* mRNA in a similar manner as *MAT2A* mRNA^[67].

Drug-induced miRNAs including miR-21-3p have been shown to control *MAT2A* and *MAT2B* stability in HepG2 cells. Either treatment with the anticancer drug berberine, which induces miR-21-3p, or overexpression of miR-21-3p itself causes apoptosis and inhibition of growth by downregulating both *MAT2A* and *MAT2B*^[69]. Most recently miR-34a and miR-34b, tumor suppressor miRNAs that are down-regulated in multiple cancers including HCC, were shown to directly target *MAT2A* mRNA and lower its expression^[70].

The N⁶-methyladenosin (m⁶A) methyltransferase METTL16, has been shown to methylate the 3'UTR of *MAT2A* hairpins in HEK293T and HeLa cells^[71,72]. In high SAME conditions methylation of hairpin 1 (hp1) of the *MAT2A* 3'UTR promotes intron retention and nuclear degradation whilst low SAME levels promote the enhanced binding of METTL16 to the hp1 of *MAT2A* 3'UTR leading to increased splicing and *MAT2A* translation^[71,72]. Regulation of *MAT2A* by METTL16 has not been reported in HCC.

***MAT* proteins post-translational modifications**

The activity and stability of *MAT* proteins can be altered by post-translational modifications including nitrosylation, phosphorylation and sumoylation. *MAT* α 1 has cysteine at position 121 (C120 in human), which lies within the flexible gating loop, can be both nitrosylated or oxidized leading to enzyme inactivation^[73]. GSH, an antioxidant, and other thiol-reducing agents can prevent and reverse this inactivation^[74]. *MAT* α 2 cannot be inhibited in this manner as glycine is at this position. Pho-sphorylation of *MAT* α 1 at threonine 342 (T341 in human) by protein kinase C was reported 25 years ago, and this modification does not alter the kinetic properties of this enzyme^[75]. However, treatment with alkaline phosphatase to dephosphorylate T342 lowered the activity of both *MAT*(α)₂ and *MAT*(α)₄ *in vitro*^[75]. Whether this is true *in vivo* has not been examined.

MAT α 2 and *MAT* β have also been reported to be phosphorylated. During liver fibrosis and HSC activation, *MAT* α 2 and *MAT* β are phosphorylated via mitogen activated protein kinase/ERK kinase (MEK) and ERK, respectively, which leads to their stabilization^[76]. Analysis revealed Y371/Y374 in *MAT* α 2 and T257/Y259 in *MAT* β to be the sites of phosphorylation and importantly, mutation of these residues inhibited HSC activation^[76]. The use of *in vitro* kinase assays, gene silencing, and chemical inhibitors have shown that MEK could be responsible for the phosphorylation of *MAT* α 2, whilst *MAT* β may be modified by ERK^[76].

Sumoylation is a post-translational modification that involves the conjugation of proteins with a small ubiquitin modifier (SUMO) that can lead to changes in the target protein's stability, localization, tertiary interaction and activity^[77]. Attachment of SUMO-1 to a protein is achieved through the sole E2-conjugating enzyme, ubiquitin-

conjugating enzyme 9 (UBC9)^[78]. The addition of SUMO-1 to a protein is generally associated with stability, and it has been shown that MAT α 2 has three SUMO-1 modifications at the lysine residues K340, K369 and K394, that increase the stability of MAT α 2 and also enhance its interaction with oncoproteins such as B-Cell CLL/lymphoma 2 (BCL-2)^[77,79]. It has been shown that treatment with SAME in liver cancer cells reduces the expression and activity of UBC9^[80].

While in normal liver MAT α 2 acetylation at K81 by the E1A binding protein (P300) causes its ubiquitination and degradation, in liver disease a lack of acetylation stabilizes the protein, and this has been associated with HCC development that is attributed to the deacetylase HDAC3^[81].

The effect of SAME on MAT expression

While the expression of MATs can influence the steady-state levels of SAME^[36], SAME level can, in return, influence MAT expression. During de-differentiation of primary hepatocytes in culture MAT1A expression falls while MAT2A expression is induced^[82]. This effect is due to a fall in SAME level since it can be blocked by the addition of SAME. Consistently, a fall in SAME level (by restricting L-methionine in medium) leads to rapid induction of MAT2A expression that is blocked upon the addition of SAME^[58,83]. Treatment with SAME results in higher methyl-HuR level leading to enhanced mRNA destabilization of MAT2A which may explain its negative effect on MAT2A expression^[54]. SAME also inhibits MAT2B expression at baseline and prevents leptin induced induction in hepatoma cells^[68].

MAT proteins interactome

MAT proteins exhibit distinct interactomes in normal and diseased liver. All three MATs interact with a variety of proteins regulating their expression and contributing to liver injury and carcinogenesis.

MAT α 1 can act as a transcription co-factor by interacting with other E-box binding regulatory proteins. MAT α 1 can heterodimerize with MAX in HCC to repress E-box-driven promoter activity, which results in the negative regulation of the transcription factors c-MYC, MAFG and c-MAF, and their oncogenic activity^[9,50]. MAT α 1 has also been found to interact with CYP2E1 to negatively regulate its protein expression by inducing its proteasomal degradation^[19]. Interestingly, MAT α 1 also interacts with p53 and DNA damage-regulated gene 1 (PDRG1) in hepatoma cells and in a mouse model of acute liver injury, which exhibited reduced total MAT α 1 expression but accumulation of nuclear MAT α 1^[84]. PDRG1 is an oncogene that is upregulated in bladder, breast and colon cancer^[85]. Interaction of PDRG1 with MAT α 1 in the nucleus resulted in a decrease in MAT activity and DNA hypomethylation^[84]. The role of PDRG1 in HCC remains unknown.

MAT α 2 not only binds to and stabilizes BCL-2 protein, it also enhances BCL-2 transcription in liver and colon cancer cell lines by binding to its promoter^[79]. MAT β is known to interact with HuR; when either of the MAT2B variants is overexpressed, cytosolic HuR content increases leading to higher mRNA levels of HuR targets such as cyclin D1 and cyclin A and proliferation^[18]. MAT β also interacts with SIRT1, and resveratrol increases this interaction by stabilizing them^[67]. MAT β also interacts with G-protein-coupled receptor kinase-interacting protein 1 (GIT1) to form a scaffold complex that interacts and activates all components of the RAS/RAF/MEK/ERK signaling pathway in liver cancer cells, promoting growth *in vitro* and *in vivo*^[7]. Interaction between MAT β and GIT1 appear to also stabilize MAT β ^[86]. Finally, both MAT α 2 and MAT β are often overexpressed in parallel in multiple cancers and part of the reason is that their interaction stabilizes both proteins^[70].

CONSEQUENCES OF MAT GENES DYSREGULATION IN THE DEVELOPMENT OF HCC

MAT genes deregulation has been widely associated with alterations that contribute to liver disease and the development of HCC. MAT1A downregulation increases oxidative stress, progenitor cells expansion, genomic instability and other mechanisms implicated in tumorigenesis, whilst MAT2A and MAT2B, which are induced in HCC, confer growth and survival advantages to cancer cells (Reviewed in Lu 2012)^[5]. In humans, the fall in MAT activity observed in cirrhotic patients is thought to contribute to the pathogenesis and progression of the disease as well as predisposition to HCC^[39].

The Mat1a-KO mouse model

The Mat1a-KO mouse model has provided important insights into the mechanisms of

how *MAT1A* might influence HCC development. This model is relevant to human liver disease since *MAT1A* expression is markedly reduced in the majority of cirrhotic patients^[87]. Mice lacking *Mat1a* have markedly increased serum methionine levels and chronically reduced hepatic SAME (70% lower) and GSH (40% lower) levels. By three months *Mat1a*-KO mice develop hepatic hyperplasia and are more susceptible to liver steatosis in response to a choline-deficient diet. By eight months, *Mat1a*-KO mice spontaneously develop NASH on a normal diet and by 18 to 20 months they develop HCC^[88]. The livers of *Mat1a*-deficient mice also exhibit oxidative stress caused by low GSH levels, impaired mitochondrial function and increased expression of CYP2E1^[34]. CYP2E1 is the principal P-450 enzyme responsible for the metabolism of hepatotoxins such as alcohol, acetaminophen and CCl₄ in the liver and has a critical role in the generation of reactive oxygen species (ROS). Because of this, *Mat1a*-KO mice are more susceptible to CCl₄-induced liver injury^[34]. Mitochondrial dysfunction is another important mechanism that sensitizes *Mat1a*-KO mice to liver injury, which was mainly attributed to low levels of both the mitochondrial chaperone PHB1 and oxidative stress^[34,89]. Our recent work adds loss of mitochondrial MAT α 1, an important regulator of mitochondrial function, as another mechanism^[49].

Genomic instability

Genomic instability, which arises from the large number of genomic mutations, chromosomal aberrations, duplications, deletions and replication errors that cancer cells carry, is a characteristic of most cancers including HCC and is considered an early step in carcinogenesis^[90,91]. Genomic instability may contribute to initiation of the malignant transformation, progression of the cancer and even resistance to therapy, influencing the overall prognosis of the cancer^[92].

MAT1A regulates DNA methylation via SAME and so *MAT1A* deficiency leads to DNA hypomethylation, which may contribute to genomic instability. Interestingly, alterations in the activity of MAT proteins and global DNA hypomethylation are prognostic markers for human HCC possibly through genomic instability^[93]. These results suggest that early dysregulation of MAT proteins could influence the progression from preneoplastic lesions to cancer. In addition, the Apurinic /Apyrimidinic Endonuclease 1 (APEX1) protein, which participates in the base excision repair of premutagenic apurinic/aprimidinic (AP) sites, the most frequent DNA lesion in cells, is markedly downregulated in *Mat1a*-KO livers. APEX1 protein level is reduced in de-differentiated hepatocytes with reduced *Mat1a* expression and low SAME levels and is recovered by SAME treatment^[94]. Taken together, these findings demonstrate that hepatic SAME depletion promotes genomic instability by different mechanisms contributing to malignant transformation.

Mitochondrial dysfunction

Along with oxidative stress, mitochondrial dysfunction represents an important trigger to hepatocarcinogenesis^[95,96]. Although mitochondrial metabolism in malignant cells is controversial, deregulated cellular energetics associated with mitochondrial dysfunction are considered a common event in cancer^[95]. As described above *Mat1a*-KO livers exhibit mitochondrial dysfunction from multiple mechanisms, including lower PHB1 expression, loss of mitochondrial MAT α 1 and increased expression of CYP2E1.

Decreased PHB1 expression

A proteomics study identified several mitochondrial proteins to be downregulated in *Mat1a*-KO mice from birth until the development of NASH^[89]. PHB1 is a well-known mitochondrial chaperone that stabilizes newly synthesized mitochondrial proteins and maintains the organization and stability of mitochondrial nucleoids. PHB1 is essential for mitochondrial function and was found significantly downregulated in the livers of *Mat1a*-KO mice as compared to wild-type animals. *In vitro* experiment suggested low SAME level promoted increased PHB1 degradation, as SAME addition prevented the fall in PHB1 protein during culture^[89]. However, a recent study showed PHB1 and *MAT1A* exert reciprocal positive regulation on each other at the mRNA level^[50], adding another mechanism to low PHB1 expression in the *Mat1a*-KO liver.

The generation of a liver specific *Phb1*-KO mouse model confirmed that PHB1 hepatic deficiency predisposes to liver injury and malignant transformation. Liver-specific *Phb1*-KO mice have liver injury at a very young age, abnormal mitochondria and increased oxidative stress. Mice lacking *Phb1* develop progressive fibrosis and multi-focal HCC by 8-10 mo of age^[97]. Although HCC could have developed as a consequence of chronic inflammation, accumulating evidence support a tumor suppressor function of PHB1 in the liver. For instance, PHB1 silencing in murine non-transformed AML12 cells increased cyclin D1, H19, and IGF2 expression and enhanced E2F binding to the cyclin D1 promoter and proliferation^[97]. PHB1 can

cooperate with CCCTC-binding transcription factor (CTCF) to negatively regulate H19 and IGF2 expression, both of which are induced in HCC^[98]. Reduced PHB1 expression has also been shown to induce *IL-8* transcription by activating NF- κ B and AP-1, resulting in enhanced *IL-8* expression and release to promote tumorigenesis^[99]. Finally, PHB1 also acts as a negative regulator of WNT signaling, and its downregulation causes the induction of multiple WNT ligands and downstream activation of canonical WNT β -catenin signaling in murine liver and human HCC cells, in part through E2F^[100].

It should be noted that the tumor suppressor role of PHB1 is highly controversial since PHB1 expression is increased in other cancers. PHB1 is also found in the nucleus, where it has been shown to interact with Rb and p53 among other proteins to bring about a change in transcriptional activities of E2F and p53^[101]. Different subcellular localizations and post-translation modifications may explain the contradictory tumor regulatory activities of PHB1 in different cell types.

Mitochondrial MAT α 1

Recently MAT α 1 was shown to be present in the mitochondrial matrix in hepatocytes^[19]. *Mat1a*-KO hepatocytes had reduced mitochondrial membrane potential and higher mitochondrial ROS, both of which were normalized when *MAT1A* was overexpressed. Oxygen consumption rate, ATP production and maximal and spare respiratory capacities were all reduced in *Mat1a*-deficient mitochondria, supporting the negative effect that *MAT1A* deficiency has on mitochondrial function. Another important finding that may contribute to mitochondrial dysfunction and liver injury in *Mat1a*-KO mice is the interaction of MAT α 1 and CYP2E1 in the mitochondria. *Mat1a* deficiency leads to higher CYP2E1 mitochondrial levels. Mitochondrial CYP2E1 also regulates the production of ROS and is known to contribute to liver injury and mitochondrial dysfunction^[102,103]. MAT α 1 negatively regulates CYP2E1 expression at mRNA and protein levels, with the latter being the dominant mechanism that involves methylation of CYP2E1 R379, promoting its proteasomal degradation^[19].

MAT α 1 was found to also interact with important mitochondrial proteins including several subunits of the electron transport chain complexes, which raises the possibility that MAT α 1 could be regulating mitochondrial function in multiple ways. These findings highlight a critical role of MAT α 1 in regulating mitochondrial function and could provide a novel target for the treatment of different liver diseases where mitochondrial dysfunction plays a key role. Taken together, reduced mitochondrial MAT α 1 could play a key role in the mitochondrial dysfunction that is often observed in HCC^[104].

Cancer stem cells

Cancer stem cells, also known as tumorinitiating cells, are known to play a central role in tumor development, metastasis and recurrence and are considered key therapeutic target for cancer treatment. Hepatic oval cells are the cancer stem cells of the liver and important contributors of hepatocarcinogenesis^[105]. They are quiescent in normal adult liver and low in number and expand during severe and prolonged injury as seen in various models of experimental carcinogenesis^[106].

Methyl-deficient diets have been used to induce oval cell proliferation and HCC formation in susceptible models such as p53^{-/-} mice^[107]. *Mat1a*-KO livers contain increased populations of liver cancer stem cells (or CD133⁺/CD49f⁺ oval cells), as they age^[108]. These cells have increased expression of several oncogenes and are tumorigenic *in vivo*. Interestingly, *Mat1a*-KO's cancer stem cells show increased MAPK signaling with enhanced ERK activity, like *Mat1a*-KO's hepatocytes, and increased oncogenic signaling (K-Ras and Survivin)^[108]. Constitutive ERK activation makes these cells resistant to the apoptotic effect of TGF- β , a well-known growth inhibitor in hepatocytes^[109,110]. How SAME deficiency allows expansion of oval cells remains unknown.

Dysregulated pathways

Increasing evidence indicates that the deregulation of various signaling pathways progressively increase with HCC progression and has a prognostic value. The *MAT1A* to *MAT2A/MAT2B* switch is also associated with activation of multiple signaling pathways including RAS/ERK^[76,86,111], I κ B kinase (IKK)/NF- κ B^[14,60], Phosphoinositide 3-kinase (PI3K)/AKT^[68,112,113] and Liver kinase B1 (LKB1)/AMPK-activated protein kinase (AMPK)^[114]. Increased sumoylation^[80] and c-MYC expression^[8], which are well-known contributors of hepatocarcinogenesis, have also been associated with reduced hepatic SAME levels.

ERK signaling

ERK signaling is tightly regulated in normal cells but uncontrollably active in cancer cells, being one of the several growth signals associated with highly malignant HCCs^[115,116]. ERK activity is regulated by the dual-specificity MAPK phosphatase (DUSP1). DUSP1 is a member of a family of dual-specificity MAPK phosphatases that dephosphorylates both serine/threonine and tyrosine residues^[117]. Interestingly, there is a reciprocal regulation between DUSP1 and ERK^[111]. Transient activation of ERK leads to catalytic activation DUSP1, which in turn inhibits ERK activity by dephosphorylation^[118]. DUSP1 feedback inhibits ERK and this activity of DUSP1 is crucial for the regulation of ERK activity in liver cells. However, prolonged ERK activation induces the phosphorylation of DUSP1 at the Ser296 residue rendering the DUSP1 protein susceptible to proteasomal degradation^[119]. In human HCC, DUSP1 expression is negatively correlated with proliferation and microvessel density and positively with survival^[118].

Hepatic DUSP1 expression is decreased in *Mat1a*-KO mice both at the mRNA and protein levels, being more pronounced at the protein level and was normalized after SAME treatment^[111]. SAME increased *DUSP1* mRNA level by enhancing p53 binding to its consensus element in the *DUSP1* promoter, which is known to activate *DUSP1* transcriptionally^[111]. Increased binding of p53 to the *DUSP1* promoter in SAME fed mice was due to the fact that SAME stabilizes APEX1, which is a known *trans*-activator of p53^[94]. DUSP1 lower protein level was attributed to its faster degradation due to increased proteasomal activity in *Mat1a*-KO mice. SAME treatment normalized proteasomal activity, increasing DUSP1 protein level and normalized ERK activity in *Mat1a*-KO mice^[111]. These findings suggest that SAME deficiency leads to uncontrolled ERK activation due at least in part to decreased DUSP1 expression during HCC development. ERK signaling was also found to be regulated by *MAT2B* since its knockdown inhibited the activation of MAPK/ERK pathway induced by leptin in liver cancer cell lines^[68]. Finally, as mentioned earlier, the MAT β -GIT1 complex efficiently binds to MEK and ERK leading to their activation. Consistent with this, overexpression of *MAT2B* or *GIT1* in the HCC cell line Huh7 enhanced tumor growth and metastasis in a mouse orthotopic HCC model^[86].

LKB1-AMPK signaling

Even though LKB1 is considered a tumor suppressor in a variety of cancers^[120], its role in liver carcinogenesis remains controversial given the fact that both reduced and increased LKB1 levels have been reported in human HCC correlating with prognosis^[121-124]. In recent years, several publications have supported the oncogenic role of LKB1 in liver cancer^[122-125].

LKB1 phosphorylates and activates AMPK, a central metabolic sensor, to control cell growth in response to environmental nutrient changes. Indeed, the LKB1/AMPK signaling pathway has tumor suppressor activity since it serves as a metabolic checkpoint arresting cell growth in conditions of nutrient deprivation or low intracellular ATP levels^[126]. In the liver the activation of LKB1 and AMPK leads to the production of nitric oxide (NO) and is required for hepatocyte proliferation, as seen in regenerating livers after partial hepatectomy and hepatocyte growth factor (HGF) treated rat hepatocytes^[114]. Interestingly, early during liver regeneration hepatic SAME level falls and exogenous SAME inhibits regeneration^[127]. The thought is that SAME level needs to fall in order to release the inhibitory tone it exerts on mitogenic pathways. Although *Mat1a*-KO mice have higher basal proliferation, they exhibit impaired liver regeneration because hepatic SAME level remained unchanged^[127]. Consistent with this, in the presence of SAME, protein phosphatase 2A (PP2A) interacts with AMPK, leading to its dephosphorylation and inactivation. One explanation for the fall in hepatic SAME level early in liver regeneration is due to increased NO formation, which can inactivate MAT α 1^[128,129].

Activation of the LKB1/AMPK pathway may contribute to hepatocarcinogenesis through other mechanisms as well. Increased LKB1 and AMPK activity results in nuclear to cytoplasmic HuR translocation and the subsequent stabilization of several cyclin mRNAs enhancing cell proliferation^[130]. LKB1/AMPK activation is also required for survival of HCC cells derived from *Mat1a*-KO livers, named SAME-D^[114]. LKB1 can regulate AKT-mediated cell survival independent of PI3K, AMPK, and mTOR2 (mammalian target of rapamycin complex). LKB1 is hyperactivated in SAME-D cells and can control survival through the phosphorylation and cytoplasmic retention of p53. In normal cells, p53 expression is maintained at a low level and is inactivated by Mdm2-mediated nuclear to cytosol transportation and proteasomal degradation. In response to oncogenic insults, p53 translocates to the nucleus to exert its tumor suppressor activity and activate pathways associated with DNA repair, cell cycle arrest and apoptosis. Notably, HuR nucleocytoplasmic shuttling also stabilizes p53 mRNA. Supporting these findings, increased cytoplasmic staining of p53 and phospho-LKB1 were found in the *Mat1a*-KO livers and in livers from human HCC

derived from both NASH and alcoholic steatohepatitis^[131]. **Figure 3** summarizes the mechanisms of HCC development in the *Mat1a*-KO mouse model.

MAT DYSREGULATION IN CCA

CCA is, after HCC, the second most common primary hepatic malignancy and like HCC, its development involves MAT genes deregulation. *MAT1A* is highly expressed also in normal bile duct epithelial cells and is repressed during chronic cholestasis and in murine and human CCA^[8]. There are common mechanisms of *MAT* genes deregulation between HCC and CCA. For example, hypermethylation of the *MAT1A* promoter has also been observed in CCA. The transcription factors c-MYC, MAFG and c-MAF, which are all induced both in HCC and CCA, also negatively regulate *MAT1A* transcription in CCA by binding to its repressor E-box promoter region^[8]. PHB1, which is also downregulated in most human CCAs, positively regulates *MAT1A* while suppressing c-MYC, MAFG, and c-MAF expression in mice^[9,50]. Consistently, reduced PHB1 expression predisposes to the development of cholestasis-induced CCA^[8,50]. **Figure 4** summarizes changes in MATs and SAME in HCC and CCA as compared to normal liver.

TARGETING MATS IN LIVER CANCER

Restoration of SAME levels

The *Mat1a*-KO mouse model demonstrates that the switch from *Mat1a* to *Mat2a* leads to hepatic SAME deficiency and hypermethioninemia^[88]. Patients with human liver cirrhosis^[39], alcoholic hepatitis^[40], advanced NAFLD^[53], as well as HCC^[42] all have lower *MAT1A* mRNA level. The restoration of hepatic SAME level appears to be an obvious therapy to overcome the fall in SAME level due to the *MAT1A*/*MAT2A* switch. There have been very few human trials that examined SAME in chronic liver disease. One study showed that SAME treatment (1,200 mg orally per day in three divided doses) for two years improved the survival or delayed liver transplantation in patients who had alcoholic liver cirrhosis with less advanced liver disease (Child's class A and B)^[132]. However, this was a post-hoc analysis, so it remains to be confirmed. Another study in patients with hepatitis B-related advanced-stage (stages B-C) HCC who received 1000 mg of SAME two hours before surgery intravenously and continued for five consecutive postoperative days showed a reduction of alanine aminotransferase and aspartate transaminase levels, delayed recurrence and a greater 24-mo survival rate as well as a lower risk of complications^[133]. However, analysis by the Cochrane Hepato-Biliary group using a total of 330 alcoholic liver disease patients treated with SAME in 8 clinical trials did not confirm the beneficial effects of SAME in human liver cirrhosis although they were small with variable quality, which resulted in the meta-analysis failing to show a significant benefit of SAME treatment^[134]. At the present time the verdict on the effectiveness of SAME in chronic liver disease is still out and its utility in chemoprevention of HCC remains to be studied.

In CCl_4 -induced liver injury SAME treatment prevented the activation of HSCs by inhibiting the collagen promoter as well as downregulating TGF- β -induced extracellular matrix protein, α -smooth muscle actin (α -SMA)^[135]. It has also been demonstrated that SAME is selectively pro-apoptotic in liver cancer cells but anti-apoptotic in normal hepatocytes^[136,137]. This property makes SAME particularly attractive as a chemopreventive agent. Indeed, SAME can chemoprevent against HCC in several preclinical models^[5]. However, SAME was ineffective at treating established tumors in an orthotopic HCC model^[138]. Investigation into this revealed that buildup of SAME in the liver was prevented by the compensatory induction of glycine-N methyl transferase (GNMT), an important enzyme abundantly expressed in the liver that is responsible for catabolizing SAME and keeping SAME level within a tight range^[5,138]. Although acute pharmacologic SAME treatment can transiently increase liver SAME level by 10-fold, prolong SAME treatment induced GNMT expression so that SAME level increased only 30%, which is insufficient to exert a pro-apoptotic effect^[138]. However, in human cirrhosis and HCC GNMT expression is often downregulated^[139], so the efficacy of SAME for HCC treatment in human warrants further investigation. At present, there are two SAME ongoing clinical trials in HCC treatment. One (NCT03178929) is designed to investigate SAME treatment (2000 mg, P.O.) in HCC patients (Barcelona Clinic Liver Cancer 0/A) after radical treatment, where tumor recurrence is the endpoint. The other (NCT02586285) aims to study the SAME treatment (500-1000 mg, iv, per day) in HCC patients after curative treatment. Although targeting SAME has been the central focus for treatment, therapies that

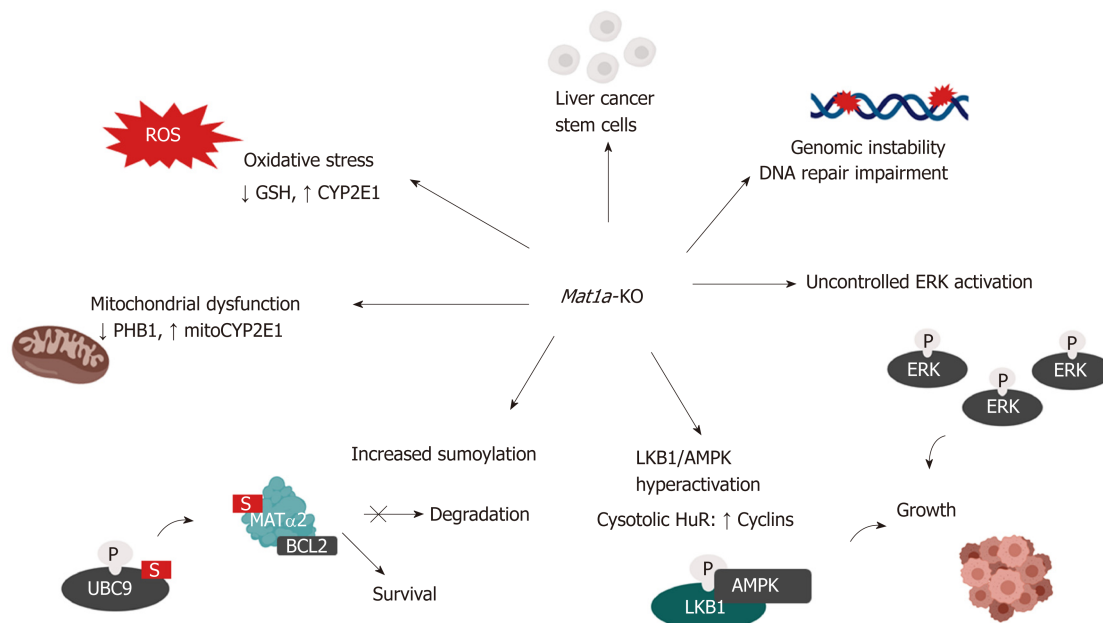


Figure 3 Mechanisms of hepatocellular carcinoma development in the *Mat1a* knockout mouse. Multiple mechanisms are known to influence hepatocellular carcinoma development in the *Mat1a*-KO mouse. These include: oxidative stress due to lower GSH levels and higher CYP2E1 expression; mitochondrial dysfunction due to reduced PHB1 and increased mitochondrial CYP2E1 levels; increased sumoylation, stabilizing MAT α 2, which acts as a transcription factor to enhance *BCL*-2 transcription as well as directly interacting with *BCL*-2 leading to its stabilization; enhanced activation of the LKB1/AMPK pathway which leads to the cytoplasmic translocation of HuR from nucleus and subsequent stabilization of cyclins; aberrant activation of ERK which promotes uncontrolled cell growth; enhanced genomic instability due to DNA hypomethylation and impaired DNA repair machinery and increased number of liver cancer stem cells with tumorigenic potential.

target the MAT proteins themselves could also offer potential benefit to which some of these are described below.

Restoring the expression of endogenous MAT1A

Overexpressing *MAT1A* in the liver cancer cell line Huh7 resulted in a stable increase in SAME levels, an induction in tumor suppressor genes, downregulation of angiogenesis genes, reduced cell growth and increased apoptosis *in vitro* and *in vivo*^[140]. This is a proof of principle that raising *MAT1A* expression in liver cancer could be an effective treatment strategy. While delivering *MAT1A* gene therapy to HCC cells is not a suitable approach, targeting miRNAs that downregulate *MAT1A* expression might be feasible. MiR-485-3p, miR-495, and miR-664 are three miRNAs that are increased in human HCC that directly target *MAT1A* at the 3'UTR^[56]. Treatment with siRNA against any of the miRNAs resulted in higher *MAT1A* expression, reduced HCC growth in an orthotopic HCC model^[56]. Higher SAME level as a result can also downregulate *MAT2A*/*MAT2B* to slow HCC growth. Taken together, raising endogenous *MAT1A* expression and hence SAME level in the HCC cells is an attractive treatment approach that deserves further study.

Targeting of MAT proteins

Therapeutic agents to target MAT α 2 have been proposed for many years. In the 1970s methionine analogs were suggested to act as substrate-competitive inhibitors and proposed as chemotherapeutic agents^[141]. Stilbene derivatives (FIDAS agents) have been proposed as inhibitors of MAT α 2, but these compounds are unlikely to specifically inhibit MAT α 2 because of their ability to bind multiple protein targets^[142]. Using molecules that directly target the active site of MAT α 2 may be problematic as this protein is expressed in most extrahepatic cells which require MAT α 2 as the SAME generator. In addition, the high level of sequence and structural identity between MAT α 1 and MAT α 2 makes it difficult to design active site molecules that would only target one of these proteins. One strategy is to target the interaction of the MAT α 2 β complexes as these complexes provide a proliferative advantage to HCC cells by lowering steady state SAME level, stabilizing each other as well as activating several mitogenic pathways^[7,43,68,86]. A novel allosteric inhibitor has been shown to bind to the interacting site of these proteins, which demonstrated promise in lung cancer cells^[13]. It remains to be examined in liver cancer models.

Targeting a posttranslational modification could also be a good strategy to only target MAT proteins that are contributing to the pathogenesis of HCC. Designing a molecule for a specific modification on MAT α 1, MAT α 2, or MAT β may be possible to

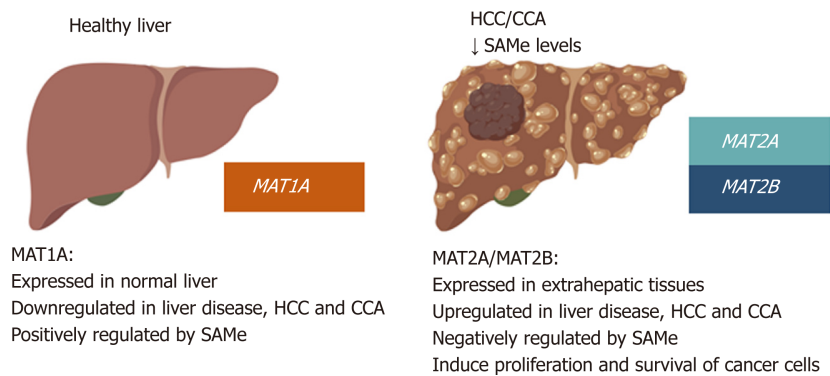


Figure 4 MAT genes expression pattern in normal liver and liver cancer. *MAT1A* is mainly expressed in normal liver by hepatocytes and bile duct epithelial cells; whereas *MAT2A* and *MAT2B* are expressed in extrahepatic tissues and by the non-parenchymal cells in the liver. During liver disease and malignant transformation there is a switch from *MAT1A* to *MAT2A/MAT2B*, being *MAT2A* and *MAT2B* the predominant MAT genes expressed in HCC and CCA. *MAT1A* maintains the differentiated state of the liver whilst *MAT2A* and *MAT2B* give proliferative and survival advantages to cancer cells. SAME positively regulates *MAT1A* and negatively regulates *MAT2A* and *MAT2B* as well as feedback inhibits MATII. Accordingly, steady state SAME levels are lower in patients with chronic liver disease and liver cancer due to the switch from *MAT1A* to *MAT2A/MAT2B*.

stabilizes MAT α 1 whilst destabilizing MAT α 2 and MAT β . For instance, sumoylation on MAT α 2 facilitated BCL-2 induction in HCC, and induced chemoresistance in HCC cells^[79]. Using small molecules to block the addition of SUMO to MAT α 2 would lower its expression as well as that of BCL-2. Similarly, MAT α 2 and MAT β are both stabilized by phosphorylation at specific residues which facilitated HSC activation^[76]. HSC activation is an important mediator of liver fibrosis, and therefore blocking the site-specific phosphorylation of these MAT proteins could be a novel treatment for liver fibrosis. As mentioned before, MAT β has been implicated to promote cell survival in HCC due to its interaction with the MEK/ERK/MAPK pathway^[68,86]. The discovery that MAT β interacted with GIT1 and caused the amplification of RAS-mediated MAPK activation provides another novel target where a molecule could block this protein-protein interaction to inhibit MAPK signaling and cell proliferation^[7].

MATs as potential molecular markers in liver cancers

Epigenetic alterations including DNA methylation are recognized as a major characteristic in HCC and may serve as diagnostic and prognostic biomarkers^[143]. MAT α 1 to MAT α 2 switch and low SAME level are associated with HCC development and strongly predict patients' survival^[47,144]. High *MAT1A* expression in human HCC is negatively correlated to tumor size (> 5 cm), whilst up-regulation of *MAT2A* is positively correlated to serum alpha-fetoprotein level and one-year recurrence after hepatectomy^[145]. Moreover, the MAT α 1 to MAT α 2 switch may promote HCC invasion and metastasis and lower patient recurrence-free survival^[146]. All these suggest MATs may serve as diagnostic and prognostic molecular markers in HCC and likely CCA as well.

CONCLUSION

Taken together, accumulating evidence demonstrates the importance of MAT genes in liver tumorigenesis. Part of the mechanism is related to a lowering in the steady state SAME level. However, all three MATs are found in the nucleus where they regulate gene expression via epigenetics as well as bona fide transcription factor and cofactors. They also have distinct interactomes and affect a myriad of signaling pathways. SAME is an attractive chemopreventive and possibly therapeutic agent in liver cancer and its efficacy in both warrant investigation. Small molecules that inhibit interactions between MAT β -GIT1 or MAT α 2- β , both of which provide proliferative advantages to liver cancer cells, and targeting posttranslational modifications that can increase *MAT1A* and/or lower *MAT2A/2B* are all exciting future directions for translating the knowledge gained from understanding mechanisms of MAT dysregulations to therapy in liver cancer.

REFERENCES

- 1 **Ryerson AB**, Ehemann CR, Altekruse SF, Ward JW, Jemal A, Sherman RL, Henley SJ, Holtzman D, Lake A, Noone AM, Anderson RN, Ma J, Ly KN, Cronin KA, Penberthy L, Kohler BA. Annual Report to the Nation on the Status of Cancer, 1975-2012, featuring the increasing incidence of liver cancer. *Cancer* 2016; **122**: 1312-1337 [PMID: 26959385 DOI: 10.1002/cncr.29936]
- 2 **Llovet JM**, Zucman-Rossi J, Pikarsky E, Sangro B, Schwartz M, Sherman M, Gores G. Hepatocellular carcinoma. *Nat Rev Dis Primers* 2016; **2**: 16018 [PMID: 27158749 DOI: 10.1038/nrdp.2016.18]
- 3 **Siegel AB**, Zhu AX. Metabolic syndrome and hepatocellular carcinoma: two growing epidemics with a potential link. *Cancer* 2009; **115**: 5651-5661 [PMID: 19834957 DOI: 10.1002/cncr.24687]
- 4 **Diehl AM**, Day C. Cause, Pathogenesis, and Treatment of Nonalcoholic Steatohepatitis. *N Engl J Med* 2017; **377**: 2063-2072 [PMID: 29166236 DOI: 10.1056/NEJMra1503519]
- 5 **Lu SC**, Mato JM. S-adenosylmethionine in liver health, injury, and cancer. *Physiol Rev* 2012; **92**: 1515-1542 [PMID: 23073625 DOI: 10.1152/physrev.00047.2011]
- 6 **Katoh Y**, Ikura T, Hoshikawa Y, Tashiro S, Ito T, Ohta M, Kera Y, Noda T, Igarashi K. Methionine adenosyltransferase II serves as a transcriptional corepressor of Maf oncoprotein. *Mol Cell* 2011; **41**: 554-566 [PMID: 21362551 DOI: 10.1016/j.molcel.2011.02.018]
- 7 **Peng H**, Li TW, Yang H, Moyer MP, Mato JM, Lu SC. Methionine adenosyltransferase 2B-GIT1 complex serves as a scaffold to regulate Ras/Raf/MEK1/2 activity in human liver and colon cancer cells. *Am J Pathol* 2015; **185**: 1135-1144 [PMID: 25794709 DOI: 10.1016/j.ajpath.2014.12.016]
- 8 **Yang H**, Liu T, Wang J, Li TW, Fan W, Peng H, Krishnan A, Gores GJ, Mato JM, Lu SC. Deregulated methionine adenosyltransferase α 1, c-Myc, and Maf proteins together promote cholangiocarcinoma growth in mice and humans (\ddagger). *Hepatology* 2016; **64**: 439-455 [PMID: 26969892 DOI: 10.1002/hep.28541]
- 9 **Liu T**, Yang H, Fan W, Tu J, Li TWH, Wang J, Shen H, Yang J, Xiong T, Steggerda J, Liu Z, Nouredin M, Maldonado SS, Annamalai A, Seki E, Mato JM, Lu SC. Mechanisms of MAFG Dysregulation in Cholestatic Liver Injury and Development of Liver Cancer. *Gastroenterology* 2018; **155**: 557-571.e14
- 10 **Gil B**, Casado M, Pajares MA, Boscá L, Mato JM, Martín-Sanz P, Alvarez L. Differential expression pattern of S-adenosylmethionine synthetase isoenzymes during rat liver development. *Hepatology* 1996; **24**: 876-881 [PMID: 8855191 DOI: 10.1002/hep.510240420]
- 11 **Lu SC**, Gukovsky I, Lugea A, Reyes CN, Huang ZZ, Chen L, Mato JM, Bottiglieri T, Pandolfi SJ. Role of S-adenosylmethionine in two experimental models of pancreatitis. *FASEB J* 2003; **17**: 56-58 [PMID: 12424217 DOI: 10.1096/fj.01-0752fje]
- 12 **Murray B**, Antonyuk SV, Marina A, Van Liempd SM, Lu SC, Mato JM, Hasnain SS, Rojas AL. Structure and function study of the complex that synthesizes S-adenosylmethionine. *IUCrJ* 2014; **1**: 240-249 [PMID: 25075345 DOI: 10.1107/S2052252514012585]
- 13 **Quinlan CL**, Kaiser SE, Bolaños B, Nowlin D, Grantner R, Karlicek-Bryant S, Feng JL, Jenkinson S, Freeman-Cook K, Dann SG, Wang X, Wells PA, Fantin VR, Stewart AE, Grant SK. Targeting S-adenosylmethionine biosynthesis with a novel allosteric inhibitor of Mat2A. *Nat Chem Biol* 2017; **13**: 785-792 [PMID: 28553945 DOI: 10.1038/nchembio.2384]
- 14 **Yang H**, Ara AI, Magilnick N, Xia M, Ramani K, Chen H, Lee TD, Mato JM, Lu SC. Expression pattern, regulation, and functions of methionine adenosyltransferase 2beta splicing variants in hepatoma cells. *Gastroenterology* 2008; **134**: 281-291 [PMID: 18045590 DOI: 10.1053/j.gastro.2007.10.027]
- 15 **González B**, Garrido F, Ortega R, Martínez-Júlvez M, Revilla-Guarinos A, Pérez-Perotejo Y, Velázquez-Campoy A, Sanz-Aparicio J, Pajares MA. NADP⁺ binding to the regulatory subunit of methionine adenosyltransferase II increases intersubunit binding affinity in the hetero-trimer. *PLoS One* 2012; **7**: e50329 [PMID: 23189196 DOI: 10.1371/journal.pone.0050329]
- 16 **Pajares MA**, Alvarez L, Pérez-Sala D. How are mammalian methionine adenosyltransferases regulated in the liver? A focus on redox stress. *FEBS Lett* 2013; **587**: 1711-1716
- 17 **Reytor E**, Pérez-Miguelsanz J, Alvarez L, Pérez-Sala D, Pajares MA. Conformational signals in the C-terminal domain of methionine adenosyltransferase I/III determine its nucleocytoplasmic distribution. *FASEB J* 2009; **23**: 3347-3360 [PMID: 19497982 DOI: 10.1096/fj.09-130187]
- 18 **Xia M**, Chen Y, Wang LC, Zandi E, Yang H, Bemanian S, Martínez-Chantar ML, Mato JM, Lu SC. Novel function and intracellular localization of methionine adenosyltransferase 2beta splicing variants. *J Biol Chem* 2010; **285**: 20015-20021 [PMID: 20421296 DOI: 10.1074/jbc.M109.094821]
- 19 **Murray B**, Peng H, Barbier-Torres L, Robinson AE, Li TWH, Fan W, Tomasi ML, Gottlieb RA, Van Eyk J, Lu Z, Martínez-Chantar ML, Liangpunsakul S, Skill NJ, Mato JM, Lu SC. Methionine Adenosyltransferase α 1 Is Targeted to the Mitochondrial Matrix and Interacts with Cytochrome P450 2E1 to Lower Its Expression. *Hepatology* 2019 [PMID: 31077594 DOI: 10.1002/hep.30762]
- 20 **González B**, Pajares MA, Hermoso JA, Guillerm D, Guillerm G, Sanz-Aparicio J. Crystal structures of methionine adenosyltransferase complexed with substrates and products reveal the methionine-ATP recognition and give insights into the catalytic mechanism. *J Mol Biol* 2003; **331**: 407-416 [PMID: 12888348 DOI: 10.1016/S0022-2836(03)00728-9]
- 21 **Komoto J**, Yamada T, Takata Y, Markham GD, Takusagawa F. Crystal structure of the S-adenosylmethionine synthetase ternary complex: a novel catalytic mechanism of S-adenosylmethionine synthesis from ATP and Met. *Biochemistry* 2004; **43**: 1821-1831 [PMID: 14967023 DOI: 10.1021/B1035611T]
- 22 **Shafqat N**, Muniz JR, Pilka ES, Papagrigoriou E, von Delft F, Oppermann U, Yue WW. Insight into S-adenosylmethionine biosynthesis from the crystal structures of the human methionine adenosyltransferase catalytic and regulatory subunits. *Biochem J* 2013; **452**: 27-36 [PMID: 23425511 DOI: 10.1042/BJ20121580]
- 23 **Murray B**, Antonyuk SV, Marina A, Lu SC, Mato JM, Hasnain SS, Rojas AL. Crystallography captures catalytic steps in human methionine adenosyltransferase enzymes. *Proc Natl Acad Sci U S A* 2016; **113**: 2104-2109 [PMID: 26858410 DOI: 10.1073/pnas.1510959113]
- 24 **Sánchez-Pérez GF**, Bautista JM, Pajares MA. Methionine adenosyltransferase as a useful molecular systematics tool revealed by phylogenetic and structural analyses. *J Mol Biol* 2004; **335**: 693-706 [PMID: 14687567 DOI: 10.1016/j.jmb.2003.11.022]
- 25 **Panmanee J**, Bradley-Clarke J, Mato JM, O'Neill PM, Antonyuk SV, Hasnain SS. Control and regulation of S-Adenosylmethionine biosynthesis by the regulatory β subunit and quinolone-based compounds. *FEBS J* 2019; **286**: 2135-2154 [PMID: 30776190 DOI: 10.1111/febs.14790]
- 26 **Koth M**, Mudd SH, Mato JM, Geller AM, Kredich NM, Chou JY, Cantoni GL. Consensus nomenclature for the mammalian methionine adenosyltransferase genes and gene products. *Trends Genet* 1997; **13**: 51-

- 52 [PMID: 9055605 DOI: 10.1016/S0168-9525(97)01013-5]
- 27 **Kotb M**, Kredich NM. S-Adenosylmethionine synthetase from human lymphocytes. Purification and characterization. *J Biol Chem* 1985; **260**: 3923-3930 [PMID: 3980460]
- 28 **Parry RJ**, Minta A. Studies of enzyme stereochemistry. Elucidation of the stereochemistry of S-adenosylmethionine formation by yeast methionine adenosyltransferase. *J Am Chem Soc* 1982; **104**: 871-872 [DOI: 10.1021/ja00367a048]
- 29 **Markham GD**, Parkin DW, Mentch F, Schramm VL. A kinetic isotope effect study and transition state analysis of the S-adenosylmethionine synthetase reaction. *J Biol Chem* 1987; **262**: 5609-5615 [PMID: 3553181]
- 30 **Markham GD**, Pajares MA. Structure-function relationships in methionine adenosyltransferases. *Cell Mol Life Sci* 2009; **66**: 636-648 [PMID: 18953685 DOI: 10.1007/s00018-008-8516-1]
- 31 **Okada G**, Teraoka H, Tsukada K. Multiple species of mammalian S-adenosylmethionine synthetase. Partial purification and characterization. *Biochemistry* 1981; **20**: 934-940 [PMID: 7213623 DOI: 10.1021/bi00507a045]
- 32 **Pajares MA**, Durán C, Corrales F, Pliego MM, Mato JM. Modulation of rat liver S-adenosylmethionine synthetase activity by glutathione. *J Biol Chem* 1992; **267**: 17598-17605 [PMID: 1517209]
- 33 **Sullivan DM**, Hoffman JL. Fractionation and kinetic properties of rat liver and kidney methionine adenosyltransferase isozymes. *Biochemistry* 1983; **22**: 1636-1641 [PMID: 6849873 DOI: 10.1021/bi00276a017]
- 34 **Martínez-Chantar ML**, Corrales FJ, Martínez-Cruz LA, García-Trevijano ER, Huang ZZ, Chen L, Kanel G, Avila MA, Mato JM, Lu SC. Spontaneous oxidative stress and liver tumors in mice lacking methionine adenosyltransferase 1A. *FASEB J* 2002; **16**: 1292-1294 [PMID: 12060674 DOI: 10.1096/fj.02-0078fje]
- 35 **Finkelstein JD**. Methionine metabolism in mammals. *J Nutr Biochem* 1990; **1**: 228-237 [PMID: 15539209 DOI: 10.1016/0955-2863(90)90070-2]
- 36 **Cai J**, Mao Z, Hwang JJ, Lu SC. Differential expression of methionine adenosyltransferase genes influences the rate of growth of human hepatocellular carcinoma cells. *Cancer Res* 1998; **58**: 1444-1450 [PMID: 9537246]
- 37 **Halim AB**, LeGros L, Geller A, Kotb M. Expression and functional interaction of the catalytic and regulatory subunits of human methionine adenosyltransferase in mammalian cells. *J Biol Chem* 1999; **274**: 29720-29725 [PMID: 10514445 DOI: 10.1074/JBC.274.42.29720]
- 38 **Nordgren KK**, Peng Y, Pelleymounter LL, Moon I, Abo R, Feng Q, Eckloff B, Yee VC, Wieben E, Weinshilboum RM. Methionine adenosyltransferase 2A/2B and methylation: gene sequence variation and functional genomics. *Drug Metab Dispos* 2011; **39**: 2135-2147 [PMID: 21813468 DOI: 10.1124/dmd.111.040857]
- 39 **Avila MA**, Berasain C, Torres L, Martín-Duce A, Corrales FJ, Yang H, Prieto J, Lu SC, Caballería J, Rodés J, Mato JM. Reduced mRNA abundance of the main enzymes involved in methionine metabolism in human liver cirrhosis and hepatocellular carcinoma. *J Hepatol* 2000; **33**: 907-914 [PMID: 11131452 DOI: 10.1016/S0168-8278(00)80122-1]
- 40 **Lee TD**, Sada MR, Mendler MH, Bottiglieri T, Kanel G, Mato JM, Lu SC. Abnormal hepatic methionine and glutathione metabolism in patients with alcoholic hepatitis. *Alcohol Clin Exp Res* 2004; **28**: 173-181 [PMID: 14745316 DOI: 10.1097/01.ALC.0000108654.77178.03]
- 41 **Lu SC**, Mato JM. S-Adenosylmethionine in cell growth, apoptosis and liver cancer. *J Gastroenterol Hepatol* 2008; **23** Suppl 1: S73-S77 [PMID: 18336669 DOI: 10.1111/j.1440-1746.2007.05289.x]
- 42 **Cai J**, Sun WM, Hwang JJ, Stain SC, Lu SC. Changes in S-adenosylmethionine synthetase in human liver cancer: molecular characterization and significance. *Hepatology* 1996; **24**: 1090-1097 [PMID: 8903381 DOI: 10.1002/hep.510240519]
- 43 **Martínez-Chantar ML**, García-Trevijano ER, Latasa MU, Martín-Duce A, Fortes P, Caballería J, Avila MA, Mato JM. Methionine adenosyltransferase II beta subunit gene expression provides a proliferative advantage in human hepatoma. *Gastroenterology* 2003; **124**: 940-948 [PMID: 12671891 DOI: 10.1053/GAST.2003.50151]
- 44 **Huang ZZ**, Mao Z, Cai J, Lu SC. Changes in methionine adenosyltransferase during liver regeneration in the rat. *Am J Physiol* 1998; **275**: G14-G21 [PMID: 9655679 DOI: 10.1152/ajpgi.1998.275.1.G14]
- 45 **Lu SC**, Huang ZZ, Yang H, Mato JM, Avila MA, Tsukamoto H. Changes in methionine adenosyltransferase and S-adenosylmethionine homeostasis in alcoholic rat liver. *Am J Physiol Gastrointest Liver Physiol* 2000; **279**: G178-G185 [PMID: 10898761 DOI: 10.1152/ajpgi.2000.279.1.G178]
- 46 **Huang ZZ**, Mato JM, Kanel G, Lu SC. Differential effect of thioacetamide on hepatic methionine adenosyltransferase expression in the rat. *Hepatology* 1999; **29**: 1471-1478 [PMID: 10216131 DOI: 10.1002/hep.510290525]
- 47 **Frau M**, Tomasi ML, Simile MM, Demartis MI, Salis F, Latte G, Calvisi DF, Seddaiu MA, Daino L, Feo CF, Brozzetti S, Solinas G, Yamashita S, Ushijima T, Feo F, Pascale RM. Role of transcriptional and posttranscriptional regulation of methionine adenosyltransferases in liver cancer progression. *Hepatology* 2012; **56**: 165-175 [PMID: 22318685 DOI: 10.1002/hep.25643]
- 48 **Zeng Z**, Huang ZZ, Chen C, Yang H, Mao Z, Lu SC. Cloning and functional characterization of the 5'-flanking region of human methionine adenosyltransferase 1A gene. *Biochem J* 2000; **346** Pt 2: 475-482 [PMID: 10677369 DOI: 10.1042/BJ3460475]
- 49 **Ikeda R**, Nishida T, Watanabe F, Shimizu-Saito K, Asahina K, Horikawa S, Teraoka H. Involvement of CCAAT/enhancer binding protein-beta (C/EBPbeta) in epigenetic regulation of mouse methionine adenosyltransferase 1A gene expression. *Int J Biochem Cell Biol* 2008; **40**: 1956-1969
- 50 **Fan W**, Yang H, Liu T, Wang J, Li TW, Mavila N, Tang Y, Yang J, Peng H, Tu J, Annamalai A, Nouredin M, Krishnan A, Gores GJ, Martínez-Chantar ML, Mato JM, Lu SC. Prohibitin 1 suppresses liver cancer tumorigenesis in mice and human hepatocellular and cholangiocarcinoma cells. *Hepatology* 2017; **65**: 1249-1266 [PMID: 27981602 DOI: 10.1002/hep.28964]
- 51 **Torres L**, Avila MA, Carretero MV, Latasa MU, Caballería J, López-Rodas G, Boukaba A, Lu SC, Franco L, Mato JM. Liver-specific methionine adenosyltransferase MAT1A gene expression is associated with a specific pattern of promoter methylation and histone acetylation: implications for MAT1A silencing during transformation. *FASEB J* 2000; **14**: 95-102 [PMID: 10627284 DOI: 10.1096/fasebj.14.1.95]
- 52 **Tomasi ML**, Li TW, Li M, Mato JM, Lu SC. Inhibition of human methionine adenosyltransferase 1A transcription by coding region methylation. *J Cell Physiol* 2012; **227**: 1583-1591 [PMID: 21678410 DOI: 10.1002/jcp.22875]
- 53 **Murphy SK**, Yang H, Moylan CA, Pang H, Dellinger A, Abdelmalek MF, Garrett ME, Ashley-Koch A,

- Suzuki A, Tillmann HL, Hauser MA, Diehl AM. Relationship between methylome and transcriptome in patients with nonalcoholic fatty liver disease. *Gastroenterology* 2013; **145**: 1076-1087
- 54 **Vázquez-Chantada M**, Fernández-Ramos D, Embade N, Martínez-Lopez N, Varela-Rey M, Woodhoo A, Luka Z, Wagner C, Anglim PP, Finnell RH, Caballería J, Laird-Offringa IA, Gorospe M, Lu SC, Mato JM, Martínez-Chantar ML. HuR/methyl-HuR and AUF1 regulate the MAT expressed during liver proliferation, differentiation, and carcinogenesis. *Gastroenterology* 2010; **138**: 1943-1953
- 55 **Koturbash I**, Melnyk S, James SJ, Beland FA, Pogribny IP. Role of epigenetic and miR-22 and miR-29b alterations in the downregulation of Mat1a and Mthfr genes in early preneoplastic livers in rats induced by 2-acetylaminofluorene. *Mol Carcinog* 2013; **52**: 318-327 [PMID: 22213190 DOI: 10.1002/mc.21861]
- 56 **Yang H**, Cho ME, Li TW, Peng H, Ko KS, Mato JM, Lu SC. MicroRNAs regulate methionine adenosyltransferase 1A expression in hepatocellular carcinoma. *J Clin Invest* 2013; **123**: 285-298 [PMID: 23241961 DOI: 10.1172/JCI63861]
- 57 **Yang H**, Huang ZZ, Wang J, Lu SC. The role of c-Myb and Sp1 in the up-regulation of methionine adenosyltransferase 2A gene expression in human hepatocellular carcinoma. *FASEB J* 2001; **15**: 1507-1516 [PMID: 11427482 DOI: 10.1096/fj.01-0040com]
- 58 **Yang H**, Huang ZZ, Zeng Z, Chen C, Selby RR, Lu SC. Role of promoter methylation in increased methionine adenosyltransferase 2A expression in human liver cancer. *Am J Physiol Gastrointest Liver Physiol* 2001; **280**: G184-G190 [PMID: 11208539 DOI: 10.1152/ajpgi.2001.280.2.G184]
- 59 **Rodríguez JL**, Boukaba A, Sandoval J, Georgieva EI, Latasa MU, García-Trevijano ER, Serviddio G, Nakamura T, Avila MA, Sastre J, Torres L, Mato JM, López-Rodas G. Transcription of the MAT2A gene, coding for methionine adenosyltransferase, is up-regulated by E2F and Sp1 at a chromatin level during proliferation of liver cells. *Int J Biochem Cell Biol* 2007; **39**: 842-850 [PMID: 17317269 DOI: 10.1016/J.BIOCEL.2007.01.009]
- 60 **Yang H**, Sadda MR, Yu V, Zeng Y, Lee TD, Ou X, Chen L, Lu SC. Induction of human methionine adenosyltransferase 2A expression by tumor necrosis factor alpha. Role of NF-kappa B and AP-1. *J Biol Chem* 2003; **278**: 50887-50896 [PMID: 14530285 DOI: 10.1074/jbc.M307600200]
- 61 **Wang K**, Fang S, Liu Q, Gao J, Wang X, Zhu H, Zhu Z, Ji F, Wu J, Ma Y, Hu L, Shen X, Gao D, Zhu J, Liu P, Zhou H. TGF- β 1/p65/MAT2A pathway regulates liver fibrogenesis via intracellular SAM. *EBioMedicine* 2019; **42**: 458-469 [PMID: 30926424 DOI: 10.1016/j.ebiom.2019.03.058]
- 62 **Ramani K**, Tomasi ML. Transcriptional regulation of methionine adenosyltransferase 2A by peroxisome proliferator-activated receptors in rat hepatic stellate cells. *Hepatology* 2012; **55**: 1942-1953 [PMID: 22271545 DOI: 10.1002/hep.25594]
- 63 **Guo YT**, Leng XS, Li T, Peng JR, Song SH, Xiong LF, Qin ZZ. Effect of ligand of peroxisome proliferator-activated receptor gamma on the biological characters of hepatic stellate cells. *World J Gastroenterol* 2005; **11**: 4735-4739 [PMID: 16094720 DOI: 10.3748/WJG.V11.I30.4735]
- 64 **Hellemans K**, Michalik L, Dittie A, Knorr A, Rombouts K, De Jong J, Heirman C, Quartier E, Schuit F, Wahli W, Geerts A. Peroxisome proliferator-activated receptor-beta signaling contributes to enhanced proliferation of hepatic stellate cells. *Gastroenterology* 2003; **124**: 184-201 [PMID: 12512042 DOI: 10.1053/gast.2003.50015]
- 65 **Liu Q**, Liu L, Zhao Y, Zhang J, Wang D, Chen J, He Y, Wu J, Zhang Z, Liu Z. Hypoxia induces genomic DNA demethylation through the activation of HIF-1 α and transcriptional upregulation of MAT2A in hepatoma cells. *Mol Cancer Ther* 2011; **10**: 1113-1123 [PMID: 21460102 DOI: 10.1158/1535-7163.MCT-10-1010]
- 66 **Liu Q**, Chen J, Liu L, Zhang J, Wang D, Ma L, He Y, Liu Y, Liu Z, Wu J. The X protein of hepatitis B virus inhibits apoptosis in hepatoma cells through enhancing the methionine adenosyltransferase 2A gene expression and reducing S-adenosylmethionine production. *J Biol Chem* 2011; **286**: 17168-17180 [PMID: 21247894 DOI: 10.1074/jbc.M110.167783]
- 67 **Yang H**, Zheng Y, Li TW, Peng H, Fernandez-Ramos D, Martínez-Chantar ML, Rojas AL, Mato JM, Lu SC. Methionine adenosyltransferase 2B, HuR, and sirtuin 1 protein cross-talk impacts on the effect of resveratrol on apoptosis and growth in liver cancer cells. *J Biol Chem* 2013; **288**: 23161-23170 [PMID: 23814050 DOI: 10.1074/jbc.M113.487157]
- 68 **Ramani K**, Yang H, Xia M, Ara AI, Mato JM, Lu SC. Leptin's mitogenic effect in human liver cancer cells requires induction of both methionine adenosyltransferase 2A and 2beta. *Hepatology* 2008; **47**: 521-531 [PMID: 18041713 DOI: 10.1002/hep.22064]
- 69 **Lo TF**, Tsai WC, Chen ST. MicroRNA-21-3p, a berberine-induced miRNA, directly down-regulates human methionine adenosyltransferases 2A and 2B and inhibits hepatoma cell growth. *PLoS One* 2013; **8**: e75628 [PMID: 24098708 DOI: 10.1371/journal.pone.0075628]
- 70 **Tomasi ML**, Cossu C, Spissu Y, Floris A, Ryo M, Iglesias-Ara A, Wang Q, Pandol SJ, Bhowmick NA, Seki E, Posadas EM, Lu SC. S-adenosylmethionine and methylthioadenosine inhibit cancer metastasis by targeting microRNA 34a/b-methionine adenosyltransferase 2A/2B axis. *Oncotarget* 2017; **8**: 78851-78869 [PMID: 29108270 DOI: 10.18632/oncotarget.20234]
- 71 **Pendleton KE**, Chen B, Liu K, Hunter OV, Xie Y, Tu BP, Conrad NK. The U6 snRNA m⁶A Methyltransferase METTL16 Regulates SAM Synthetase Intron Retention. *Cell* 2017; **169**: 824-835.e14 [PMID: 28525753 DOI: 10.1016/j.cell.2017.05.003]
- 72 **Shima H**, Matsumoto M, Ishigami Y, Ebina M, Muto A, Sato Y, Kumagai S, Ochiai K, Suzuki T, Igarashi K. S-Adenosylmethionine Synthesis Is Regulated by Selective N⁶-Adenosine Methylation and mRNA Degradation Involving METTL16 and YTHDC1. *Cell Rep* 2017; **21**: 3354-3363
- 73 **Avila MA**, Corrales FJ, Ruiz F, Sánchez-Góngora E, Mingorance J, Carretero MV, Mato JM. Specific interaction of methionine adenosyltransferase with free radicals. *Biofactors* 1998; **8**: 27-32 [PMID: 9699005 DOI: 10.1002/biof.5520080106]
- 74 **Corrales F**, Ochoa P, Rivas C, Martín-Lomas M, Mato JM, Pajares MA. Inhibition of glutathione synthesis in the liver leads to S-adenosyl-L-methionine synthetase reduction. *Hepatology* 1991; **14**: 528-533 [PMID: 1874498 DOI: 10.1002/hep.1840140320]
- 75 **Pajares MA**, Durán C, Corrales F, Mato JM. Protein kinase C phosphorylation of rat liver S-adenosylmethionine synthetase: dissociation and production of an active monomer. *Biochem J* 1994; **303**: 949-955 [PMID: 7980467 DOI: 10.1042/BJ3030949]
- 76 **Ramani K**, Donoyan S, Tomasi ML, Park S. Role of methionine adenosyltransferase α 2 and β phosphorylation and stabilization in human hepatic stellate cell trans-differentiation. *J Cell Physiol* 2015; **230**: 1075-1085 [PMID: 25294683 DOI: 10.1002/jcp.24839]
- 77 **Ghioni P**, D'Alessandra Y, Mansueto G, Jaffray E, Hay RT, La Mantia G, Guerrini L. The protein stability and transcriptional activity of p63 α are regulated by SUMO-1 conjugation. *Cell Cycle* 2005; **4**: 183-

- 190 [PMID: [15611636](#) DOI: [10.4161/cc.4.1.1359](#)]
- 78 **Gareau JR**, Lima CD. The SUMO pathway: emerging mechanisms that shape specificity, conjugation and recognition. *Nat Rev Mol Cell Biol* 2010; **11**: 861-871 [PMID: [21102611](#) DOI: [10.1038/nrm3011](#)]
- 79 **Tomasi ML**, Ryoo M, Ramani K, Tomasi I, Giordano P, Mato JM, Lu SC. Methionine adenosyltransferase $\alpha 2$ sumoylation positively regulate Bcl-2 expression in human colon and liver cancer cells. *Oncotarget* 2015; **6**: 37706-37723 [PMID: [26416353](#) DOI: [10.18632/oncotarget.5342](#)]
- 80 **Tomasi ML**, Tomasi I, Ramani K, Pascale RM, Xu J, Giordano P, Mato JM, Lu SC. S-adenosyl methionine regulates ubiquitin-conjugating enzyme 9 protein expression and sumoylation in murine liver and human cancers. *Hepatology* 2012; **56**: 982-993 [PMID: [22407595](#) DOI: [10.1002/hep.25701](#)]
- 81 **Yang HB**, Xu YY, Zhao XN, Zou SW, Zhang Y, Zhang M, Li JT, Ren F, Wang LY, Lei QY. Acetylation of MAT I α represses tumour cell growth and is decreased in human hepatocellular cancer. *Nat Commun* 2015; **6**: 6973 [PMID: [25925782](#) DOI: [10.1038/ncomms7973](#)]
- 82 **García-Trevijano ER**, Latasa MU, Carretero MV, Berasain C, Mato JM, Avila MA. S-adenosylmethionine regulates MAT1A and MAT2A gene expression in cultured rat hepatocytes: a new role for S-adenosylmethionine in the maintenance of the differentiated status of the liver. *FASEB J* 2000; **14**: 2511-2518 [PMID: [11099469](#) DOI: [10.1096/fj.00-0121.com](#)]
- 83 **Martínez-Chantar ML**, Latasa MU, Varela-Rey M, Lu SC, García-Trevijano ER, Mato JM, Avila MA. L-methionine availability regulates expression of the methionine adenosyltransferase 2A gene in human hepatocarcinoma cells: role of S-adenosylmethionine. *J Biol Chem* 2003; **278**: 19885-19890 [PMID: [12660248](#) DOI: [10.1074/jbc.M211554200](#)]
- 84 **Pérez C**, Pérez-Zúñiga FJ, Garrido F, Reytor E, Portillo F, Pajares MA. The Oncogene PDRG1 Is an Interaction Target of Methionine Adenosyltransferases. *PLoS One* 2016; **11**: e0161672 [PMID: [27548429](#) DOI: [10.1371/journal.pone.0161672](#)]
- 85 **Jiang L**, Luo X, Shi J, Sun H, Sun Q, Sheikh MS, Huang Y. PDRG1, a novel tumor marker for multiple malignancies that is selectively regulated by genotoxic stress. *Cancer Biol Ther* 2011; **11**: 567-573 [PMID: [21193842](#) DOI: [10.4161/CBT.11.6.14412](#)]
- 86 **Peng H**, Dara L, Li TW, Zheng Y, Yang H, Tomasi ML, Tomasi I, Giordano P, Mato JM, Lu SC. MAT2B-GIT1 interplay activates MEK1/ERK 1 and 2 to induce growth in human liver and colon cancer. *Hepatology* 2013; **57**: 2299-2313 [PMID: [23325601](#) DOI: [10.1002/hep.26258](#)]
- 87 **Mato JM**, Corrales FJ, Lu SC, Avila MA. S-Adenosylmethionine: a control switch that regulates liver function. *FASEB J* 2002; **16**: 15-26 [PMID: [11772932](#) DOI: [10.1096/fj.01-0401rev](#)]
- 88 **Lu SC**, Alvarez L, Huang ZZ, Chen L, An W, Corrales FJ, Avila MA, Kanel G, Mato JM. Methionine adenosyltransferase 1A knockout mice are predisposed to liver injury and exhibit increased expression of genes involved in proliferation. *Proc Natl Acad Sci U S A* 2001; **98**: 5560-5565 [PMID: [11320206](#) DOI: [10.1073/pnas.091016398](#)]
- 89 **Santamaria E**, Avila MA, Latasa MU, Rubio A, Martín-Duce A, Lu SC, Mato JM, Corrales FJ. Functional proteomics of nonalcoholic steatohepatitis: mitochondrial proteins as targets of S-adenosylmethionine. *Proc Natl Acad Sci USA* 2003; **100**: 3065-3070 [PMID: [12631701](#) DOI: [10.1073/pnas.0536625100](#)]
- 90 **Rao CV**, Asch AS, Yamada HY. Emerging links among Chromosome Instability (CIN), cancer, and aging. *Mol Carcinog* 2017; **56**: 791-803 [PMID: [27533343](#) DOI: [10.1002/mc.22539](#)]
- 91 **Coleman WB**, Tsongalis GJ. Multiple mechanisms account for genomic instability and molecular mutation in neoplastic transformation. *Clin Chem* 1995; **41**: 644-657 [PMID: [7729041](#)]
- 92 **Ferguson LR**, Chen H, Collins AR, Connell M, Damia G, Dasgupta S, Malhotra M, Meecker AK, Amedei A, Amin A, Ashraf SS, Aquilano K, Azmi AS, Bhakta D, Bilsland A, Boosani CS, Chen S, Ciriolo MR, Fujii H, Guha G, Halicka D, Helferich WG, Keith WN, Mohammed SI, Niccolai E, Yang X, Honoki K, Parslow VR, Prakash S, Rezazadeh S, Shackelford RE, Sidransky D, Tran PT, Yang ES, Maxwell CA. Genomic instability in human cancer: Molecular insights and opportunities for therapeutic attack and prevention through diet and nutrition. *Semin Cancer Biol* 2015; **35** Suppl: S5-S24
- 93 **Calvisi DF**, Ladu S, Gorden A, Farina M, Lee JS, Conner EA, Schroeder I, Factor VM, Thorgeirsson SS. Mechanistic and prognostic significance of aberrant methylation in the molecular pathogenesis of human hepatocellular carcinoma. *J Clin Invest* 2007; **117**: 2713-2722 [PMID: [17717605](#) DOI: [10.1172/JCI31457](#)]
- 94 **Tomasi ML**, Iglesias-Ara A, Yang H, Ramani K, Feo F, Pascale MR, Martínez-Chantar ML, Mato JM, Lu SC. S-adenosylmethionine regulates apurinic/aprimidinic endonuclease 1 stability: implication in hepatocarcinogenesis. *Gastroenterology* 2009; **136**: 1025-1036
- 95 **Hsu CC**, Tseng LM, Lee HC. Role of mitochondrial dysfunction in cancer progression. *Exp Biol Med (Maywood)* 2016; **241**: 1281-1295 [PMID: [27022139](#) DOI: [10.1177/1535370216641787](#)]
- 96 **Hanahan D**, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011; **144**: 646-674
- 97 **Ko KS**, Tomasi ML, Iglesias-Ara A, French BA, French SW, Ramani K, Lozano JJ, Oh P, He L, Stiles BL, Li TW, Yang H, Martínez-Chantar ML, Mato JM, Lu SC. Liver-specific deletion of prohibitin 1 results in spontaneous liver injury, fibrosis, and hepatocellular carcinoma in mice. *Hepatology* 2010; **52**: 2096-2108 [PMID: [20890892](#) DOI: [10.1002/hep.23919](#)]
- 98 **Ramani K**, Mavila N, Ko KS, Mato JM, Lu SC. Prohibitin 1 Regulates the H19-Igf2 Axis and Proliferation in Hepatocytes. *J Biol Chem* 2016; **291**: 24148-24159 [PMID: [27687727](#) DOI: [10.1074/jbc.M116.744045](#)]
- 99 **Yang JW**, Murray B, Barbier-Torres L, Liu T, Liu Z, Yang H, Fan W, Wang J, Li Y, Seki E, Mato JM, Lu SC. The mitochondrial chaperone Prohibitin 1 negatively regulates interleukin-8 in human liver cancers. *J Biol Chem* 2019; **294**: 1984-1996 [PMID: [30523154](#) DOI: [10.1074/jbc.RA118.004863](#)]
- 100 **Mavila N**, Tang Y, Berlind J, Ramani K, Wang J, Mato JM, Lu SC. Prohibitin 1 Acts As a Negative Regulator of Wingless/Integrated-Beta-Catenin Signaling in Murine Liver and Human Liver Cancer Cells. *Hepatol Commun* 2018; **2**: 1583-1600 [PMID: [30556043](#) DOI: [10.1002/hep4.1257](#)]
- 101 **Fusaro G**, Dasgupta P, Rastogi S, Joshi B, Chellappan S. Prohibitin induces the transcriptional activity of p53 and is exported from the nucleus upon apoptotic signaling. *J Biol Chem* 2003; **278**: 47853-47861 [PMID: [14500729](#) DOI: [10.1074/jbc.M305171200](#)]
- 102 **Caro AA**, Cederbaum AI. Oxidative stress, toxicology, and pharmacology of CYP2E1. *Annu Rev Pharmacol Toxicol* 2004; **44**: 27-42 [PMID: [14744237](#) DOI: [10.1146/annurev.pharmtox.44.101802.121704](#)]
- 103 **Lu Y**, Cederbaum AI. CYP2E1 and oxidative liver injury by alcohol. *Free Radic Biol Med* 2008; **44**: 723-738 [PMID: [18078827](#) DOI: [10.1016/J.FREERADBIOMED.2007.11.004](#)]
- 104 **Chagoya de Sánchez V**, Chávez E, Velasco-Loyden G, Guadalupe Lozano-Rosas M, Rusbel Aparicio-Cadena A. Interaction of Mitochondrial and Epigenetic Regulation in Hepatocellular Carcinoma. *Liver*

- Cancer*. Ahmed Lasfar: IntechOpen 2018; [DOI: [10.5772/intechopen.79923](https://doi.org/10.5772/intechopen.79923)]
- 105 **Alison MR**. Liver stem cells: implications for hepatocarcinogenesis. *Stem Cell Rev* 2005; **1**: 253-260 [PMID: [17142862](https://pubmed.ncbi.nlm.nih.gov/17142862/) DOI: [10.1385/SCR:1:3:253](https://doi.org/10.1385/SCR:1:3:253)]
- 106 **Jelnes P**, Santoni-Rugiu E, Rasmussen M, Friis SL, Nielsen JH, Tygstrup N, Bisgaard HC. Remarkable heterogeneity displayed by oval cells in rat and mouse models of stem cell-mediated liver regeneration. *Hepatology* 2007; **45**: 1462-1470 [PMID: [17538966](https://pubmed.ncbi.nlm.nih.gov/17538966/) DOI: [10.1002/hep.21569](https://doi.org/10.1002/hep.21569)]
- 107 **Dumble ML**, Croager EJ, Yeoh GC, Quail EA. Generation and characterization of p53 null transformed hepatic progenitor cells: oval cells give rise to hepatocellular carcinoma. *Carcinogenesis* 2002; **23**: 435-445 [PMID: [11895858](https://pubmed.ncbi.nlm.nih.gov/11895858/) DOI: [10.1093/carcin/23.3.435](https://doi.org/10.1093/carcin/23.3.435)]
- 108 **Rountree CB**, Senadheera S, Mato JM, Crooks GM, Lu SC. Expansion of liver cancer stem cells during aging in methionine adenosyltransferase 1A-deficient mice. *Hepatology* 2008; **47**: 1288-1297 [PMID: [18167064](https://pubmed.ncbi.nlm.nih.gov/18167064/) DOI: [10.1002/hep.22141](https://doi.org/10.1002/hep.22141)]
- 109 **Ding W**, Mouzaki M, You H, Laird JC, Mato J, Lu SC, Rountree CB. CD133+ liver cancer stem cells from methionine adenosyl transferase 1A-deficient mice demonstrate resistance to transforming growth factor (TGF)-beta-induced apoptosis. *Hepatology* 2009; **49**: 1277-1286 [PMID: [19115422](https://pubmed.ncbi.nlm.nih.gov/19115422/) DOI: [10.1002/hep.22743](https://doi.org/10.1002/hep.22743)]
- 110 **Nguyen LN**, Furuya MH, Wolfrain LA, Nguyen AP, Holdren MS, Campbell JS, Knight B, Yeoh GC, Fausto N, Parks WT. Transforming growth factor-beta differentially regulates oval cell and hepatocyte proliferation. *Hepatology* 2007; **45**: 31-41 [PMID: [17187411](https://pubmed.ncbi.nlm.nih.gov/17187411/) DOI: [10.1002/hep.21466](https://doi.org/10.1002/hep.21466)]
- 111 **Tomasi ML**, Ramani K, Lopitz-Otsoa F, Rodríguez MS, Li TW, Ko K, Yang H, Bardag-Gorce F, Iglesias-Ara A, Feo F, Pascale MR, Mato JM, Lu SC. S-adenosylmethionine regulates dual-specificity mitogen-activated protein kinase phosphatase expression in mouse and human hepatocytes. *Hepatology* 2010; **51**: 2152-2161 [PMID: [20196119](https://pubmed.ncbi.nlm.nih.gov/20196119/) DOI: [10.1002/hep.23530](https://doi.org/10.1002/hep.23530)]
- 112 **Pañeda C**, Gorospe I, Herrera B, Nakamura T, Fabregat I, Varela-Nieto I. Liver cell proliferation requires methionine adenosyltransferase 2A mRNA up-regulation. *Hepatology* 2002; **35**: 1381-1391 [PMID: [12029623](https://pubmed.ncbi.nlm.nih.gov/12029623/) DOI: [10.1053/jhep.2002.32538](https://doi.org/10.1053/jhep.2002.32538)]
- 113 **Ramani K**, Yang H, Kuhlenskamp J, Tomasi L, Tsukamoto H, Mato JM, Lu SC. Changes in the expression of methionine adenosyltransferase genes and S-adenosylmethionine homeostasis during hepatic stellate cell activation. *Hepatology* 2010; **51**: 986-995 [PMID: [20043323](https://pubmed.ncbi.nlm.nih.gov/20043323/) DOI: [10.1002/hep.23411](https://doi.org/10.1002/hep.23411)]
- 114 **Vázquez-Chantada M**, Ariz U, Varela-Rey M, Embade N, Martínez-Lopez N, Fernández-Ramos D, Gómez-Santos L, Lamas S, Lu SC, Martínez-Chantar ML, Mato JM. Evidence for LKB1/AMP-activated protein kinase/ endothelial nitric oxide synthase cascade regulated by hepatocyte growth factor, S-adenosylmethionine, and nitric oxide in hepatocyte proliferation. *Hepatology* 2009; **49**: 608-617 [PMID: [19177591](https://pubmed.ncbi.nlm.nih.gov/19177591/) DOI: [10.1002/hep.22660](https://doi.org/10.1002/hep.22660)]
- 115 **Personeni N**, Rimassa L, Pressiani T, Destro A, Ligorio C, Tronconi MC, Bozzarelli S, Carnaghi C, Di Tommaso L, Giordano L, Roncalli M, Santoro A. Molecular determinants of outcome in sorafenib-treated patients with hepatocellular carcinoma. *J Cancer Res Clin Oncol* 2013; **139**: 1179-1187 [PMID: [23568548](https://pubmed.ncbi.nlm.nih.gov/23568548/) DOI: [10.1007/s00432-013-1429-x](https://doi.org/10.1007/s00432-013-1429-x)]
- 116 **Negri FV**, Dal Bello B, Porta C, Campanini N, Rossi S, Tinelli C, Poggi G, Missale G, Fanello S, Salvagni S, Ardizzoni A, Maria SE. Expression of pERK and VEGFR-2 in advanced hepatocellular carcinoma and resistance to sorafenib treatment. *Liver Int* 2015; **35**: 2001-2008 [PMID: [25559745](https://pubmed.ncbi.nlm.nih.gov/25559745/) DOI: [10.1111/liv.12778](https://doi.org/10.1111/liv.12778)]
- 117 **Sun H**, Charles CH, Lau LF, Tonks NK. MKP-1 (3CH134), an immediate early gene product, is a dual specificity phosphatase that dephosphorylates MAP kinase in vivo. *Cell* 1993; **75**: 487-493 [PMID: [8221888](https://pubmed.ncbi.nlm.nih.gov/8221888/) DOI: [10.1016/0092-8674\(93\)90383-2](https://doi.org/10.1016/0092-8674(93)90383-2)]
- 118 **Calvisi DF**, Pinna F, Meloni F, Ladu S, Pellegrino R, Sini M, Daino L, Simile MM, De Miglio MR, Virdis P, Frau M, Tomasi ML, Seddaiu MA, Muroli MR, Feo F, Pascale RM. Dual-specificity phosphatase 1 ubiquitination in extracellular signal-regulated kinase-mediated control of growth in human hepatocellular carcinoma. *Cancer Res* 2008; **68**: 4192-4200 [PMID: [18519678](https://pubmed.ncbi.nlm.nih.gov/18519678/) DOI: [10.1158/0008-5472.CAN-07-6157](https://doi.org/10.1158/0008-5472.CAN-07-6157)]
- 119 **Lin YW**, Yang JL. Cooperation of ERK and SCFSkp2 for MKP-1 destruction provides a positive feedback regulation of proliferating signaling. *J Biol Chem* 2006; **281**: 915-926 [PMID: [16286470](https://pubmed.ncbi.nlm.nih.gov/16286470/) DOI: [10.1074/jbc.M508720200](https://doi.org/10.1074/jbc.M508720200)]
- 120 **Korsse SE**, Peppelenbosch MP, van Veelen W. Targeting LKB1 signaling in cancer. *Biochim Biophys Acta* 2013; **1835**: 194-210 [PMID: [23287572](https://pubmed.ncbi.nlm.nih.gov/23287572/) DOI: [10.1016/j.bbcan.2012.12.006](https://doi.org/10.1016/j.bbcan.2012.12.006)]
- 121 **Huang YH**, Chen ZK, Huang KT, Li P, He B, Guo X, Zhong JQ, Zhang QY, Shi HQ, Song QT, Yu ZP, Shan YF. Decreased expression of LKB1 correlates with poor prognosis in hepatocellular carcinoma patients undergoing hepatectomy. *Asian Pac J Cancer Prev* 2013; **14**: 1985-1988 [PMID: [23679304](https://pubmed.ncbi.nlm.nih.gov/23679304/) DOI: [10.7314/APJCP.2013.14.3.1985](https://doi.org/10.7314/APJCP.2013.14.3.1985)]
- 122 **Martínez-López N**, García-Rodríguez JL, Varela-Rey M, Gutiérrez V, Fernández-Ramos D, Beraza N, Aransay AM, Schlangen K, Lozano JJ, Aspichueta P, Luka Z, Wagner C, Evert M, Calvisi DF, Lu SC, Mato JM, Martínez-Chantar ML. Hepatoma cells from mice deficient in glycine N-methyltransferase have increased RAS signaling and activation of liver kinase B1. *Gastroenterology* 2012; **143**: 787-798.e13 [PMID: [22687285](https://pubmed.ncbi.nlm.nih.gov/22687285/) DOI: [10.1053/j.gastro.2012.05.050](https://doi.org/10.1053/j.gastro.2012.05.050)]
- 123 **Barbier-Torres L**, Delgado TC, García-Rodríguez JL, Zubiete-Franco I, Fernández-Ramos D, Buqué X, Cano A, Gutiérrez-de Juan V, Fernández-Domínguez I, Lopitz-Otsoa F, Fernández-Tussy P, Boix L, Bruix J, Villa E, Castro A, Lu SC, Aspichueta P, Xirodimas D, Varela-Rey M, Mato JM, Beraza N, Martínez-Chantar ML. Stabilization of LKB1 and Akt by neddylation regulates energy metabolism in liver cancer. *Oncotarget* 2015; **6**: 2509-2523 [PMID: [25650664](https://pubmed.ncbi.nlm.nih.gov/25650664/) DOI: [10.18632/oncotarget.3191](https://doi.org/10.18632/oncotarget.3191)]
- 124 **Tan X**, Liao Z, Liang H, Chen X, Zhang B, Chu L. Upregulation of liver kinase B1 predicts poor prognosis in hepatocellular carcinoma. *Int J Oncol* 2018; **53**: 1913-1926 [PMID: [30226588](https://pubmed.ncbi.nlm.nih.gov/30226588/) DOI: [10.3892/ijo.2018.4556](https://doi.org/10.3892/ijo.2018.4556)]
- 125 **Zubiete-Franco I**, García-Rodríguez JL, Lopitz-Otsoa F, Serrano-Macia M, Simon J, Fernández-Tussy P, Barbier-Torres L, Fernández-Ramos D, Gutiérrez-de Juan V, López de Davallillo S, Carlevaris O, Beguiristain Gómez A, Villa E, Calvisi D, Martín C, Berra E, Aspichueta P, Beraza N, Varela-Rey M, Ávila M, Rodríguez MS, Mato JM, Díaz-Moreno I, Díaz-Quintana A, Delgado TC, Martínez-Chantar ML. SUMOylation regulates LKB1 localization and its oncogenic activity in liver cancer. *EBioMedicine* 2019; **40**: 406-421 [PMID: [30594553](https://pubmed.ncbi.nlm.nih.gov/30594553/) DOI: [10.1016/j.ebiom.2018.12.031](https://doi.org/10.1016/j.ebiom.2018.12.031)]
- 126 **Shackelford DB**, Shaw RJ. The LKB1-AMPK pathway: metabolism and growth control in tumour suppression. *Nat Rev Cancer* 2009; **9**: 563-575 [PMID: [19629071](https://pubmed.ncbi.nlm.nih.gov/19629071/) DOI: [10.1038/nrc2676](https://doi.org/10.1038/nrc2676)]
- 127 **Chen L**, Zeng Y, Yang H, Lee TD, French SW, Corrales FJ, Garcia-Trevijano ER, Avila MA, Mato JM, Lu SC. Impaired liver regeneration in mice lacking methionine adenosyltransferase 1A. *FASEB J* 2004;

- 18:** 914-916 [PMID: [15033934](#) DOI: [10.1096/fj.03-1204fje](#)]
- 128 **Avila MA**, Mingorance J, Martínez-Chantar ML, Casado M, Martín-Sanz P, Boscá L, Mato JM. Regulation of rat liver S-adenosylmethionine synthetase during septic shock: role of nitric oxide. *Hepatology* 1997; **25**: 391-396 [PMID: [9021952](#) DOI: [10.1002/hep.510250222](#)]
- 129 **Ruiz F**, Corrales FJ, Miqueo C, Mato JM. Nitric oxide inactivates rat hepatic methionine adenosyltransferase In vivo by S-nitrosylation. *Hepatology* 1998; **28**: 1051-1057 [PMID: [9755242](#) DOI: [10.1002/hep.510280420](#)]
- 130 **Martínez-Chantar ML**, Vázquez-Chantada M, Garnacho M, Latasa MU, Varela-Rey M, Dotor J, Santamaria M, Martínez-Cruz LA, Parada LA, Lu SC, Mato JM. S-adenosylmethionine regulates cytoplasmic HuR via AMP-activated kinase. *Gastroenterology* 2006; **131**: 223-232 [PMID: [16831604](#) DOI: [10.1053/J.GASTRO.2006.04.019](#)]
- 131 **Martínez-López N**, Varela-Rey M, Fernández-Ramos D, Woodhoo A, Vázquez-Chantada M, Embade N, Espinosa-Hevia L, Bustamante FJ, Parada LA, Rodríguez MS, Lu SC, Mato JM, Martínez-Chantar ML. Activation of LKB1-Akt pathway independent of phosphoinositide 3-kinase plays a critical role in the proliferation of hepatocellular carcinoma from nonalcoholic steatohepatitis. *Hepatology* 2010; **52**: 1621-1631 [PMID: [20815019](#) DOI: [10.1002/hep.23860](#)]
- 132 **Mato JM**, Cámara J, Fernández de Paz J, Caballería L, Coll S, Caballero A, García-Buey L, Beltrán J, Benita V, Caballería J, Solà R, Moreno-Otero R, Barrao F, Martín-Duce A, Correa JA, Parés A, Barrao E, García-Magaz I, Puerta JL, Moreno J, Boissard G, Ortiz P, Rodés J. S-adenosylmethionine in alcoholic liver cirrhosis: a randomized, placebo-controlled, double-blind, multicenter clinical trial. *J Hepatol* 1999; **30**: 1081-1089 [PMID: [10406187](#) DOI: [10.1016/S0168-8278\(99\)80263-3](#)]
- 133 **Liu GY**, Wang W, Jia WD, Xu GL, Ma JL, Ge YS, Yu JH, Sun QK, Meng FL. Protective effect of S-adenosylmethionine on hepatic ischemia-reperfusion injury during hepatectomy in HCC patients with chronic HBV infection. *World J Surg Oncol* 2014; **12**: 27 [PMID: [24485003](#) DOI: [10.1186/1477-7819-12-27](#)]
- 134 **Rambaldi A**, Gluud C, Rambaldi A. S-adenosyl-L-methionine for alcoholic liver diseases. Rambaldi A. *Cochrane Database of Systematic Reviews*. Chichester, United Kingdom: John Wiley & Sons, Ltd; 2001; CD002235 [DOI: [10.1002/14651858.CD002235](#)]
- 135 **Nieto N**, Cederbaum AI. S-adenosylmethionine blocks collagen I production by preventing transforming growth factor-beta induction of the COL1A2 promoter. *J Biol Chem* 2005; **280**: 30963-30974 [PMID: [15983038](#) DOI: [10.1074/jbc.M503569200](#)]
- 136 **Ansorena E**, García-Trevijano ER, Martínez-Chantar ML, Huang ZZ, Chen L, Mato JM, Iriburu M, Lu SC, Avila MA. S-adenosylmethionine and methylthioadenosine are antiapoptotic in cultured rat hepatocytes but proapoptotic in human hepatoma cells. *Hepatology* 2002; **35**: 274-280 [PMID: [11826399](#) DOI: [10.1053/jhep.2002.30419](#)]
- 137 **Yang H**, Sadda MR, Li M, Zeng Y, Chen L, Bae W, Ou X, Runnegar MT, Mato JM, Lu SC. S-adenosylmethionine and its metabolite induce apoptosis in HepG2 cells: Role of protein phosphatase 1 and Bcl-x(S). *Hepatology* 2004; **40**: 221-231 [PMID: [15239106](#) DOI: [10.1002/hep.20274](#)]
- 138 **Lu SC**, Ramani K, Ou X, Lin M, Yu V, Ko K, Park R, Bottiglieri T, Tsukamoto H, Kanel G, French SW, Mato JM, Moats R, Grant E. S-adenosylmethionine in the chemoprevention and treatment of hepatocellular carcinoma in a rat model. *Hepatology* 2009; **50**: 462-471 [PMID: [19444874](#) DOI: [10.1002/hep.22990](#)]
- 139 **Chen YM**, Shiu JY, Tzeng SJ, Shih LS, Chen YJ, Lui WY, Chen PH. Characterization of glycine-N-methyltransferase-gene expression in human hepatocellular carcinoma. *Int J Cancer* 1998; **75**: 787-793 [PMID: [9495250](#) DOI: [10.1002/\(SICI\)1097-0215\(19980302\)75:5<787::AID-IJC20>3.0.CO;2-2](#)]
- 140 **Li J**, Ramani K, Sun Z, Zee C, Grant EG, Yang H, Xia M, Oh P, Ko K, Mato JM, Lu SC. Forced expression of methionine adenosyltransferase 1A in human hepatoma cells suppresses in vivo tumorigenicity in mice. *Am J Pathol* 2010; **176**: 2456-2466
- 141 **Lombardini JB**, Coulter AW, Talalay P. Analogues of methionine as substrates and inhibitors of the methionine adenosyltransferase reaction. Deductions concerning the conformation of methionine. *Mol Pharmacol* 1970; **6**: 481-499 [PMID: [4918253](#)]
- 142 **Sviripa VM**, Zhang W, Balia AG, Tsodikov OV, Nickell JR, Gizard F, Yu T, Lee EY, Dwoskin LP, Liu C, Watt DS. 2',6'-Dihalostyrylanilines, pyridines, and pyrimidines for the inhibition of the catalytic subunit of methionine S-adenosyltransferase-2. *J Med Chem* 2014; **57**: 6083-6091 [PMID: [24950374](#) DOI: [10.1021/jm5004864](#)]
- 143 **Anwar SL**, Lehmann U. DNA methylation, microRNAs, and their crosstalk as potential biomarkers in hepatocellular carcinoma. *World J Gastroenterol* 2014; **20**: 7894-7913 [PMID: [24976726](#) DOI: [10.3748/wjg.v20.i24.7894](#)]
- 144 **Frau M**, Feo F, Pascale RM. Pleiotropic effects of methionine adenosyltransferases deregulation as determinants of liver cancer progression and prognosis. *J Hepatol* 2013; **59**: 830-841
- 145 **An J**, Na SK, Shim JH, Park YS, Jun MJ, Lee JH, Song GW, Lee HC, Yu E. Histological expression of methionine adenosyl transferase (MAT) 2A as a post-surgical prognostic surrogate in patients with hepatocellular carcinoma. *J Surg Oncol* 2018; **117**: 892-901 [PMID: [29448301](#) DOI: [10.1002/jso.24994](#)]
- 146 **Wang R**, Jin Y, Yao XH, Fan W, Zhang J, Cao Y, Li J. A novel mechanism of the M1-M2 methionine adenosyltransferase switch-mediated hepatocellular carcinoma metastasis. *Mol Carcinog* 2018; **57**: 1201-1212 [PMID: [29749642](#) DOI: [10.1002/mc.22836](#)]



Published By Baishideng Publishing Group Inc
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA
Telephone: +1-925-2238242
Fax: +1-925-2238243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

