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Cuticle Integrity and Biogenic Amine Synthesis in *Caenorhabditis elegans* Require the Cofactor Tetrahydrobiopterin (BH4)

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ABSTRACT Tetrahydrobiopterin (BH4) is the natural cofactor of several enzymes widely distributed among eukaryotes, including aromatic amino acid hydroxylases (AAAHs), nitric oxide synthases (NOSs), and alkylglycerol monooxygenase (AGMO). We show here that the nematode *Caenorhabditis elegans*, which has three AAAH genes and one AGMO gene, contains BH4 and has genes that function in BH4 synthesis and regeneration. Knockout mutants for putative BH4 synthetic enzyme genes lack the predicted enzymatic activities, synthesize no BH4, and have indistinguishable behavioral and neurotransmitter phenotypes, including serotonin and dopamine deficiency. The BH4 regeneration enzymes are not required for steady-state levels of biogenic amines, but become rate limiting in conditions of reduced BH4 synthesis. BH4-deficient mutants also have a fragile cuticle and are generally hypersensitive to exogenous agents, a phenotype that is not due to AAAH deficiency, but rather to dysfunction in the lipid metabolic enzyme AGMO, which is expressed in the epidermis. Loss of AGMO or BH4 synthesis also specifically alters the sensitivity of *C. elegans* to bacterial pathogens, revealing a cuticular function for AGMO-dependent lipid metabolism in host–pathogen interactions.

KEYWORDS bipterin; epidermis; serotonin; dopamine; GTPCH; alkylglycerol monooxygenase; AGMO

TETRAHYDROBIOPTERIN (BH4; 6R-5,6,7,8-tetrahydrobiopterin) is the natural cofactor of three distinct classes of enzymes including the aromatic amino acid hydroxylases (AAAHs), nitric oxide synthases (NOSs), and alkylglycerol monooxygenase (AGMO) (Werner *et al.* 2011). BH4 is therefore critical for a variety of cellular processes, being essential

for the conversion of L-Phe to L-Tyr, for alkyl ether lipid metabolism, for synthesis of nitric oxide (NO), and synthesis of the neurotransmitters serotonin (5-hydroxytryptamine, 5HT) and dopamine (DA) and their derivatives. BH4 is present in many eukaryotes (Werner-Felmayer *et al.* 2002), including the nematode *Caenorhabditis elegans* (Calvo *et al.* 2008), in which the functions of the AAAHs phenylalanine hydroxylase (PAH, gene *pah-1*), tyrosine hydroxylase (TH, gene *cat-2*), and tryptophan hydroxylase (TPH, gene *tph-1*) are well established (Lints and Emmons 1999; Loer *et al.* 1999; Sze *et al.* 2000; Calvo *et al.* 2008). *C. elegans* lacks an endogenous NOS (Gusarov *et al.* 2013); as shown below, *C. elegans* encodes a single ortholog of the recently characterized AGMO (Watschinger *et al.* 2010).

In mammals, BH4 is synthesized *de novo* in four steps from GTP by at least three enzymes: GTP cyclohydrolase I (GTPCH1, human gene *GCH1*), 6-pyruvoyl tetrahydrobiopterin synthetase (PTPS, human gene *PTS*), and either sepiapterin reductase (SR), carbonyl reductase, and/or aldose reductase (Figure 1,

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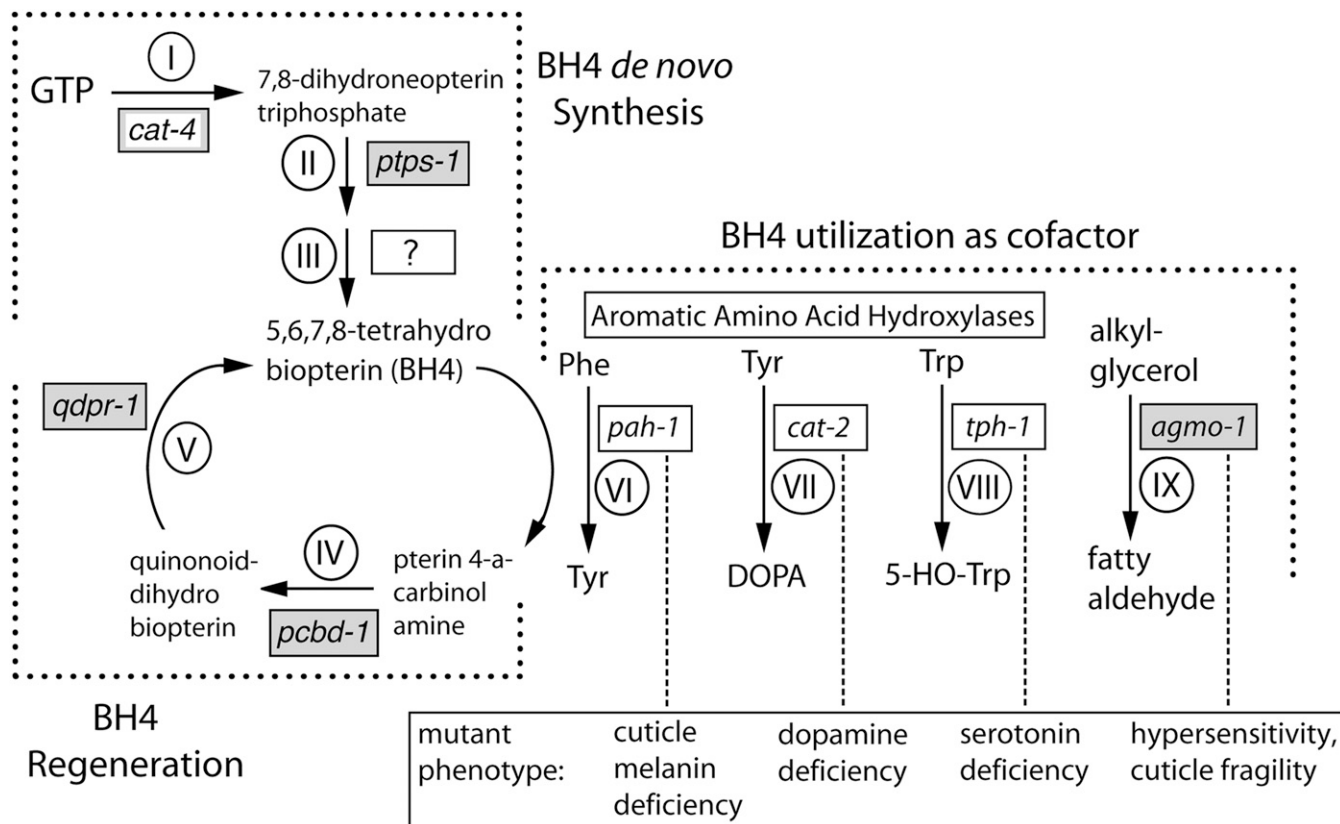


Figure 1 Biosynthesis, regeneration and utilization of tetrahydrobiopterin (BH4), and functions of BH4-dependent enzymes in *C. elegans*. Enzymes catalyzing pathway reactions are indicated by Roman numerals (Werner *et al.* 2011). (I) GTP cyclohydrolase I (E.C. 3.5.4.16); (II) 6-pyruvoyl tetrahydropterin synthase (E.C. 4.2.3.12); (III) sepiapterin reductase (E.C. 1.1.1.153), (IV), pterin-4a-carbinolamine dehydratase (E.C. 4.2.1.96); (V) [quinoid] dihydropteridine reductase (E.C. 1.6.99.7); (VI) phenylalanine hydroxylase (E.C. 1.14.16.1); (VII) tyrosine hydroxylase (E.C. 1.14.16.2); (VIII) tryptophan hydroxylase (E.C. 1.14.16.4); and (IX) alkylglycerol monooxygenase (E.C. 1.14.16.5). Names of *C. elegans* genes encoding these enzymes are shown in boxes adjacent to Roman numerals; gray boxes indicate genes for which knockout mutants are described for the first time in this work. Although *cat-4* mutant phenotypes have been described previously, we demonstrate here that the gene encodes GTPCH1. Top left: Pathway for *de novo* BH4 synthesis. The gene encoding the enzyme catalyzing the final step(s) in BH4 synthesis is unknown. Bottom left: Pathway for BH4 regeneration. Right: Four enzymes that use BH4. Mutants in *cat-4* and *ptps-1* genes (BH4-deficient) have all phenotypes listed in the box at bottom. Mutants in individual BH4-dependent enzyme genes have the indicated subset of BH4-deficiency phenotype (dashed line).

Table 1; Werner *et al.* 2011). BH4 synthesis is regulated through the action of the GTPCH1 feedback regulatory protein (GFRP), known to mediate the activation or inhibition of mammalian GTPCH1 by L-Phe or BH4, respectively. In humans, mutations in the *GCH1* gene can be recessive or cause a dominant Dopa-responsive dystonia, with or without hyperphenylalaninemia (HPA) (Ichinose *et al.* 1999). Mutations in the *PTS* gene lead to BH4-deficient HPA (also called atypical HPA or malignant phenylketonuria) (Thöny and Blau 1997). In BH4-deficient HPA patients, the neurological symptoms vary in severity depending on the degree of reduction in biogenic amine and nitric oxide levels. These conditions are manageable by carefully monitored biopterin supplementation and other treatments (Blau *et al.* 2001; Longo 2009). *Pts* knockout mice die within 48 hr if untreated with BH4 and neurotransmitter precursors (Sumi-Ichinose *et al.* 2001; Elzaouk *et al.* 2003).

As well as being required for neurotransmitter synthesis, BH4 and its derivatives are important for the synthesis of pigments and quinones involved in cross-linking external cuticle layers in invertebrates (Iino *et al.* 2000; Kato *et al.*

2006). The molecular genetics of biopterin synthesis and biogenic amine metabolism have been extensively characterized in *Drosophila melanogaster* (Wright 1987; O'Donnell *et al.* 1989). In *Drosophila*, where dopa and dopamine are required for sclerotization and tanning of the cuticle (Neckameyer and White 1993), knockout *Punch* (GTPCH1) and *purple* (PTPS) mutants die as embryos due to severe cuticle abnormalities and/or a requirement for serotonin in germband extension (Mackay and O'Donnell 1983; Colas *et al.* 1999). Interestingly, GFRP is not found in *Drosophila* (Funderburk *et al.* 2006).

When BH4 is used by the AAHs and AGMO in their respective hydroxylation reactions, it is oxidized to pterin 4-a-carbinolamine (Figure 1). This oxidized biopterin can be regenerated in mammals to BH4 by two reduction steps catalyzed by pterin carbinolamine dehydratase (PCBD) and quinoid dihydropteridine reductase (QDPR) (Werner *et al.* 2011). In humans, mutations in *QDPR* have severe effects like those in *GCH1* and *PTS* genes, whereas mutation in the human *PCBD1* gene yields a milder clinical picture (Opladen *et al.* 2012). BH4 can also be oxidized nonenzymatically and subsequently

Table 1 Pterin synthesis, regeneration, and related genes in *C. elegans*

Protein	<i>C. elegans</i> gene name	Genetic location / Genomic position (coding)	BLAST2 Expected-value vs. human and fly proteins*	Mutant allele(s)
GTP cyclohydrolase I (GTPCH1)	<i>cat-4</i> / F32G8.6	V: 2.59 V: 10,564,851-10,567,502 bp	<i>Hs</i> : 7×10^{-84} <i>Dm</i> : 6×10^{-81}	<i>e1141</i> , <i>e3015</i> , <i>e3030</i> , <i>ok342</i> , <i>tm773</i>
Pyruvoyl tetrahydropteridine synthase (PTPS)	<i>ptps-1</i> / B0041.6	I: -1.03 I: 4,652,907-4,652,187 bp	<i>Hs</i> : 9×10^{-39} <i>Dm</i> : 8×10^{-44}	<i>e3042</i> , <i>tm1984</i>
Sepiapterin reductase (SR)	No ortholog	NA	**Best matches in Ce by BLASTP <i>Hs</i> SR: 6×10^{-9} <i>Dm</i> SR: 9×10^{-12}	NA
Carbonyl reductase (CR)	No ortholog	NA	**Best match in Ce by BLASTP <i>Hs</i> CBR1: 2×10^{-12}	NA
Aldose reductase (AR)	Y39G8B.1	II: 21.67 II: 13,970,583-13,972,946 bp	<i>Hs</i> AKR1B1: 2×10^{-97} <i>Hs</i> AKR1C3: 1×10^{-87}	<i>ok1682</i>
Pterin carbinolamine dehydratase (PCBD)	<i>pcbd-1</i> / T10B11.1	I: 1.57 I: 6,951,134-6,951,946 bp	<i>Hs</i> : 9×10^{-39} <i>Dm</i> : 2×10^{-37}	<i>tm5924</i>
Quinoid dihydropteridine reductase (QDPR)	<i>qdpr-1</i> / T03F6.1	III: 21.21 III: 13,393,783-13,394,837 bp	<i>Hs</i> : 3×10^{-70} <i>Dm</i> : 3×10^{-77}	<i>tm2337</i> , <i>tm2373</i>
Dihydrofolate reductase (DHFR)	<i>dhfr-1</i> / C36B1.7	I: 3.04 I: 8,736,060-8,737,028 bp	<i>Hs</i> : 3×10^{-34} <i>Dm</i> : 3×10^{-36}	None
GTP cyclohydrolase I feedback regulatory protein (GFRP)	<i>gfrp-1</i> / Y38C1AA.13	IV: -26.81 IV: 203,882-207,984 bp	<i>Hs</i> : 5×10^{-28} <i>Dm</i> : no GFRP	None

*BLAST2: For multiple isoforms, the score shown is the best match between a Ce predicted protein and *Hs* (human) or *Dm* (fruit fly) protein. **Best match of sepiapterin reductase (SR) or carbonyl reductase (CR) via BLASTP to Ce proteins (nr database). Both CR and aldose reductase are possible partial substitutes for SR. NA, not applicable.

regenerated to BH4 by dihydrofolate reductase (DHFR) (Werner *et al.* 2011; Xu *et al.* 2014).

Analysis of the complete genomic sequence of *C. elegans* reveals orthologs of genes encoding bipterin synthesis, regulation, and regeneration enzymes known from other animals (Figure 1 and Table 1; see also Hobert 2013). *C. elegans* has clear orthologs encoding the first two BH4 synthetic enzymes, GTP cyclohydrolase I (GTPCH1, gene *cat-4*) and 6-pyruvoyl-tetrahydropterin synthase (PTPS, gene *ptps-1*). *C. elegans* appears to lack an ortholog of sepiapterin reductase (Kallberg *et al.* 2002), but does encode an aldose reductase (Table 1), which can partially substitute for SR in mammals (Park *et al.* 1991; Iino *et al.* 2003). The *C. elegans* genome also contains genes encoding bipterin regeneration enzymes PCBD, QDPR, and DHFR (genes *pcbd-1*, *qdpr-1*, and *dhfr-1*) and a clear ortholog of GFRP (*gfrp-1*).

Although *cat-4* mutants were isolated in 1975 (Sulston *et al.* 1975), the biochemical genetics of BH4 in *C. elegans* has not been previously examined. *cat-4* mutants were found based on their lack of the catecholamine DA (Sulston *et al.* 1975) and were subsequently found to be 5HT deficient (Desai *et al.* 1988). These neurotransmitter deficiencies in *cat-4* mutants cause a variety of subtle behavioral abnormalities, including defective locomotory rate regulation and male mating (Loer and Kenyon 1993; Sawin *et al.* 2000). We and others have also found that *cat-4* mutants are hypersensitive to a variety of

agents, suggesting they might have a generally “leaky” cuticle (Loer 1995; Weinshenker *et al.* 1995; Cronin *et al.* 2005; Baker *et al.* 2012). A mechanistic explanation for the cuticle defects in *cat-4* mutants, however, has been lacking.

Here we characterize the *C. elegans* pathway for BH4 biosynthesis, regulation, and regeneration. We find that *cat-4* and *ptps-1* mutants are bipterin-, 5HT- and DA-deficient and lack GTPCH1 and PTPS activities, as predicted. BH4-deficient animals have a fragile cuticle that is more permeable to small molecules, resulting in hypersensitivity to multiple chemicals. As deletion mutants in the AAAH genes do not display cuticle fragility or chemical hypersensitivity, we inferred that these phenotypes might reflect impaired function in another BH4-dependent enzyme. We show here that loss of function in the bipterin-dependent lipid metabolic enzyme AGMO (Watschinger *et al.* 2010) results in chemical hypersensitivity and cuticle fragility like that observed in BH4-deficient mutants. We find that *agmo-1* is expressed in the epidermis, consistent with its requirement for a BH4 cofactor and its role in cuticle integrity. Furthermore, *agmo-1* and the BH4-deficient mutants share a common phenotype of sensitivity to bacterial infection by *Leucobacter* Verde1. Our studies provide the first *in vivo* evidence for a role for AGMO in epidermal lipid metabolism and in pathogen defense, with implications for the function of this enzyme in other animals and in humans.

Materials and Methods

C. elegans culture, strains, and transgenes

Routine culturing of *C. elegans* was performed as described (Brenner 1974); strains were grown at 20° for all experiments, although acute analyses were at room temperature (RT) (21°–23°). Nomenclature used for *C. elegans* genetics conforms to conventions (Horvitz *et al.* 1979). Worm strains used are listed in Supporting Information. Deletion mutant strains of *cat-4*, *ptps-1*, *q DPR-1*, *p CBD-1*, and *agmo-1* were outcrossed three to five times before most analyses. Homozygosity of deletion strains was confirmed by PCR.

We generated transgenes for *cat-4*, *ptps-1*, *p CBD-1*, *q DPR-1*, *gfrp-1*, *pah-1*, and *agmo-1* by the duplex PCR method (Hobert 2002); see Table S1 on transgenes and transgenics. Amplified genomic DNAs were fused to amplified GFP; duplex products were co-injected with *Ptx-3::RFP* marker plasmid into wild type to generate transgenics. We also examined reporter transgenics previously described by others (Table S2).

The *cat-4* mutant allele sequencing and bioinformatics

The *cat-4(e1141)* mutation was identified by PCR-amplifying exons from genomic CB1141 DNA, then Sanger sequencing the purified PCR product as described (Hare and Loer 2004). The mutation was confirmed by sequencing both strands for two independent PCR reactions. The *cat-4(gk245686)* mutation (strain VC20144) was confirmed in the same manner. PCR and sequencing primers were designed using Primer3 (<http://www-genome.wi.mit.edu/cgi-bin/primer/primer3.cgi>).

C. elegans genomic and predicted cDNA sequences were retrieved from WormBase and/or GenBank. Blast searches and Blast2 comparisons were performed using the NCBI Blast server. Multiple sequence alignments used CLUSTALW (www.ebi.ac.uk/clustalw/).

Detection of serotonin and dopamine in situ

Serotonin was assessed in whole-mount worms by immunofluorescence as previously described (Desai *et al.* 1988; Loer and Kenyon 1993) using a polyclonal antiserum against 5HT paraformaldehyde conjugated to BSA (Sigma, St. Louis; catalog no. E5545, lot 091K4831) previously tested for specificity (Loer and Rivard 2007). DA was assessed in whole-mount worms via formaldehyde-induced fluorescence (FIF) as previously described (Lints and Emmons 1999). Treatment of worms with 5-hydroxytryptophan (5-HTP) or L-dopa prior to staining was done as described (Rivard *et al.* 2010).

Chemical sensitivity and cuticle integrity tests and male mating assays

Most chemicals were obtained from Sigma-Aldrich, including levamisole (T512), 5-hydroxytryptophan, and L-dopa. BH4 and sepiapterin were obtained from B. Schircks Laboratories (Jona, Switzerland). We tested adult hermaphrodites grown for 3 days at 20° from synchronized L1 worms hatched in M9. Eggs isolated by bleaching gravid adult hermaphrodites were washed extensively with M9, then hatched in M9 at RT (22°–23°).

Starved L1 worms were used immediately or stored at 12°–13°. Synchronized L1s were used no later than 7 days after egg isolation. Adult worms were washed from plates with M9 and 500- μ l aliquots placed in 24-well plates. Because of “swimming-induced paralysis,” which can vary based on biogenic amine levels (McDonald *et al.* 2007), wells were scored after approximately the same length of time in M9 buffer. Chemicals (SDS, levamisole) were made as 2 \times solutions in M9; 500 μ l of 2 \times solution was added to a well with 500 μ l worms (50–100) to yield a working concentration. Individual worms in a well were observed through a stereomicroscope and scored as immobile if no movement was seen during the brief moment of observation (≤ 1 sec).

Rapid bleach hypersensitivity tests were done with standard *C. elegans* “alkaline bleach” [4.5% sodium hypochlorite (household bleach)/1M NaOH]. Gravid adult hermaphrodites were tested on NGM plates by applying a ~ 5 - μ l drop of bleach on a worm and assaying time to rigidity using a stereomicroscope. Cuticle disintegration assays were carried out as described (Calvo *et al.* 2008). Briefly, gravid adults placed in a milder alkaline bleach solution (1% sodium hypochlorite, 0.25 M NaOH) were observed under a dissecting stereomicroscope, and the time of first major break in the cuticle was recorded. Plates were agitated manually every 30 sec. A total of 15–30 worms were scored in every experiment.

Preparation of worm homogenates and the effect of BH4 supplementation

Nematode cultures were grown on NGM plates supplemented with 5 mM ascorbate, 200 μ M BH4 for 3–5 days (until food was almost depleted). Mixed-stage populations (nonsynchronized, with predominance of adult worms) were recovered from plates with sterile M9 buffer and cleaned of bacteria by successive centrifugations at 1000 rpm. Finally, worms were incubated 30 min in M9 with agitation to reduce bacteria in the gut and centrifuged again; the resulting pellet was immediately frozen in liquid nitrogen for later use. The frozen worm pellet was resuspended in 400 μ l distilled water containing 5 mM dithioerythritol, homogenized with an Ultraturrax (Iba, Stauffen, Germany), frozen in liquid nitrogen, thawed, homogenized again, and centrifuged 5 min at 16,000 $\times g$ at 4°.

Determination of GTPCH1 and PTPS activity and BH4 content

GTPCH1 and PTPS enzymatic assays were carried out as described (Werner *et al.* 1997; Heller *et al.* 2001), with modifications to reduce the amount of required material. Results were related to protein concentrations measured in homogenates and eluates by Bradford assay (Biorad, Vienna). For BH4 determinations, 50 μ l of homogenate was used for both iodine oxidation in acid and alkaline medium assays (Heller *et al.* 2001), and BH4 concentration was calculated from the difference of resulting biopterin from the two oxidation schemes (Fukushima and Nixon 1980). For acidic oxidation, 50 μ l homogenate was mixed with 5 μ l 1 M HCl and 5 μ l 0.1 M iodine solution (prepared in 0.25 M potassium iodide). For basic

oxidation, HCl was replaced by 1 M NaOH. After 60 min at RT in the dark, 10 μ l 1 M HCl was added to the alkaline oxidation only, both incubations were centrifuged for 5 min at 16,000 \times g at 4°, and supernatants added to 10 μ l freshly prepared 0.1 M ascorbic acid. Biopterin was measured by HPLC after injection of 10 μ l on a Nucleosil 10 SA column (250 mm long, 4 mm inner diameter, 10 μ m particle size; Macherey Nagl, Düren, Germany), elution with 50 mM potassium phosphate, pH 3.0 at 35° and fluorescence detection (Excitation 350 nm, Emission 440 nm) with an Agilent 1200 HPLC (Agilent, Vienna).

For determining GTPCH1 activity, 80 μ l of homogenate was separated from low MW compounds using Micro Bio-Spin 6 columns (Biorad) equilibrated to GTPCH1 assay buffer (100 mM Tris-HCl, pH 7.8, 2.5 mM EDTA, 300 mM KCl, 10% (v/v) glycerol), incubated with 1.5 mM GTP for 90 min at 37° in 85.7 μ l total volume. Reaction was stopped by addition of 2.85 μ l 1 M HCl and 2.85 μ l 0.1 M iodine. Oxidation of resulting 7,8-dihydroneopterin triphosphate to neopterin triphosphate was achieved by incubation for 60 min at RT in the dark. After 2 min centrifugation at 16,000 \times g at 4°, 2.85 μ l 0.1 M ascorbic acid was added to the supernatant. After neutralization by addition of 2.85 μ l 1 M NaOH, neopterin phosphates were cleaved to neopterin by incubation with 6.4 units alkaline phosphatase for 30 min at 37°. Neopterin was quantified by HPLC using 10 μ l final reaction mixture injected on a reversed phase C-18 column (250 mm long, 4 mm inner diameter, 5 μ m particle size; Lichrosphere, Merck, Darmstadt, Germany), eluted with 15 mM potassium phosphate, pH 6.0, at 25° and fluorescence detection (Ex 350 nm, Em 440 nm).

For measuring PTPS activity, 80 μ l of homogenate was separated from low MW compounds using Micro Bio-Spin 6 columns (Biorad) equilibrated to PTPS assay buffer (0.1 M Tris-HCl, pH 7.4, 20 mM MgCl₂), and incubated with 40 μ M freshly prepared 7,8-dihydroneopterin triphosphate (using recombinant *Escherichia coli* GTPCH1) and 2 mM NADPH in the presence of *E. coli*-expressed recombinant mouse SR (2 nmol/min) in 100 μ l total volume for 1 h at 37° (Werner *et al.* 1997). Reaction was stopped by addition of 5 μ l 1 M HCl, and 5 μ l 0.1 M iodine. Following further incubation for 1 h at RT in the dark, resulting biopterin was quantified by HPLC as for neopterin in the GTPCH1 assay described above.

Isolation of *Leucobacter Verde1* resistance mutants and whole genome sequencing

Selections for increased resistance to *Leucobacter Verde1* exploited the hypersensitivity of certain *bus* and *srf* mutants to this bacterial pathogen, which is completely lethal to such mutants but not to wild-type *C. elegans* (Hodgkin *et al.* 2013). Mutants in three such genes (*bus-10*, *srf-2*, and *srf-5*) were used initially. Populations of these mutants were mutagenized with 0.05 M EMS (Brenner 1974). After mutagenesis, 50–80 individual L4 worms were picked to separate plates and grown on *E. coli* OP50 for two generations. About 200 F₂ progeny from each plate were then transferred to mixed *E. coli*/Verde1 (10:1) lawns and incubated at RT for a further 7–10 days, after which most (273/300) plates contained only dead or dying

worms. Plates with surviving fertile worms were retained, and single worms were picked from these to establish independent resistant lines. Outcrossing established that most resistant lines carried recessive extragenic mutations conferring resistance to Verde1 and increased bleach sensitivity. Genetic mapping and complementation tests, utilizing the bleach sensitivity phenotype, assigned 27 independent mutations to nine complementation groups. The largest complementation group (eight alleles) was mapped genetically to the right arm of LGIII. DNA from a mutant strain carrying one of these mutations (*e3016*) was prepared for whole genome sequencing using a standard library preparation without amplification, followed by Illumina 50-bp paired end sequencing on the HiSeq2000. Candidate mutations were identified using MAQGene (Bigelow *et al.* 2009), which revealed a nonsense mutation (W130opal) in *agmo-1*. Specific sequencing of *agmo-1* in strains for other alleles in this complementation group showed that all carried predicted severe mutations in this gene. Similarly, two other complementation groups were found to correspond to *cat-4* (alleles *e3015* and *e3030*) and *ptps-1*(*e3042*). Detailed analyses of six other complementation groups will be presented elsewhere.

Results

The C. elegans cat-4 (catecholamine defective) gene encodes GTP cyclohydrolase I (GTPCH1)

The *cat-4* gene maps genetically near a predicted GTPCH1-encoding gene (F32G8.6) on chromosome V. Sequencing F32G8.6 exons from the original allele *cat-4*(*e1141*), revealed a missense mutation that changes an amino acid (T66I) that is 100% conserved in GTPCH1 proteins (Figure 2, Figure S1). Two putative knockout mutants with deletions (*ok342* and *tm773*) in F32G8.6 failed to complement *e1141*, were also 5HT- and DA-deficient, and were hypersensitive with a fragile cuticle (Table 2). We examined additional *cat-4* mutants with missense mutations altering different highly conserved amino acids (Figure 2); these mutants showed a range of phenotypes. *cat-4*(*e3030*) mutants had little or no apparent 5HT or DA, whereas *e3015* mutants had reduced 5HT and DA.

Bleach hypersensitivity and reduced melanin phenotypes of *cat-4*(*tm773*) mutant worms are rescued in transgenics carrying a plasmid containing F32G8.6 coding sequence, 3'-UTR, and 1500 bp upstream (Baker *et al.* 2012). We tested these transgenic worms for 5HT and DA expression; in two independent transgenic lines carrying extrachromosomal arrays, a high percentage of worms were rescued for DA and 5HT in all normal dopaminergic and serotonergic neurons (Figure S2).

Mutants defective in BH4 synthesis display biogenic amine deficiency, chemical hypersensitivity, and cuticle fragility

We confirmed the role of a predicted PTPS gene (B0041.6; *ptps-1*; Figure S3) by examining the phenotypes of worms with a deletion (*tm1984*) or nonsense mutation (*e3042*) in coding sequence. We found that both *ptps-1* mutants were

Table 2 Phenotypes of *C. elegans* pterin-related gene mutants

<i>C. elegans</i> strain	Alleles tested	Serotonin ^a	Dopamine ^b	Bleach hypersensitivity
N2 (wild type)	NA	+ ^c	+	Non Hyp
<i>cat-4(-)</i> [GTPCH1]	<i>e1141, ok342, tm773, e3030</i>	- ^d	-	Hyp
<i>ptps-1(-)</i>	<i>tm1984, e3042</i>	-	-	Hyp
<i>pcbd-1(-)</i>	<i>tm5924</i>	+/-	+/-	Non Hyp
<i>qdpr-1(-)</i>	<i>tm2337, tm2373</i>	+/-	+/-	Non Hyp
<i>tph-1(-)</i>	^e	-	+	Non Hyp
<i>cat-2(-)</i> [TH]	^e	+	-	Non Hyp
<i>bas-1(-)</i> [AADC]	^e	-	-	Non Hyp
<i>pah-1(-)</i>	^e	+	+	Non Hyp
<i>agmo-1(-)</i>	<i>e3016, e3019, e3029, e3047</i>	+	+	Hyp
Y39G8B.1(-) [AR] ^f	<i>ok1682</i>	+	+	Non Hyp

NA, not applicable.

^a Tested by immunoreactivity (IR).

^b Tested by formaldehyde induced fluorescence (FIF).

^c Wild type.

^d Deficient.

^e Reported previously.

^f Y39G8B.1 encodes an aldose reductase (AR) ortholog, a possible partial substitute for sepiapterin reductase. The putative mutant, however, shows no phenotypes consistent with BH4 deficiency.

5HT-deficient, DA-deficient, and hypersensitive with a fragile cuticle, just like *cat-4* mutants (Figure 3, Figure 4, Table 2). Also like *cat-4* mutants, *ptps-1* mutants can be rescued for 5HT immunoreactivity (5HT-IR) by treating worms with 5-HTP, the immediate precursor to 5HT and product of TPH activity, and rescued for DA (as seen by FIF) by treatment with L-dopa, the immediate precursor to DA and product of TH activity (Figure 3, C, F, I, and L). Both treatments bypass the need for TH and TPH function in synthesis of the neurotransmitters, and also show that serotonergic and dopaminergic neurons are present and appear morphologically normal in *cat-4* and *ptps-1* mutants. We also determined that *ptps-1* mutant male worms, like *cat-4* mutant males (Loer and Kenyon 1993), were defective in the “turning” step of male mating behavior (Figure S4).

The hypersensitivity and cuticle fragility phenotypes of *cat-4* and *ptps-1* mutants are seen most dramatically when worms are placed in a standard “alkaline bleach” solution typically used to kill and dissolve gravid adult worms to isolate the more bleach-resistant eggs. Whereas wild-type worms die and become rigid after many seconds, and take minutes to dissolve, we observed that *cat-4* and *ptps-1* mutant adult worms died in a few seconds or less, ruptured quickly, and their cuticles dissolved completely in <2 min (Figure 4A). To quantify the cuticle fragility phenotype of *cat-4* and *ptps-1* mutants, we tested worms using a mild alkaline bleach treatment. Similar to the effect of standard bleach treatment, *cat-4* and *ptps-1* worms ruptured more quickly: a major break in the cuticle

occurred after about 4 min in the solution, compared with 8 min for wild-type worms (Figure 4B). *cat-4* and *ptps-1* mutants showed comparably increased sensitivity to various chemicals and drugs, including the detergent SDS and the anthelmintic levamisole, which in nematodes acts as a cholinergic agonist (Figure S5, A and C). In tests of acute exposure causing worm immobility, the biopterin synthesis mutants were approximately twice as sensitive to SDS compared to wild type, and 5–20 times more sensitive to levamisole.

To demonstrate that *CAT-4* and *PTPS-1* are *bona fide* BH4 synthetic enzymes, we measured GTPCH1 and PTPS activity in soluble protein extracts from homogenized worms, as well as total biopterin and BH4 content. *cat-4* null mutants lacked GTPCH1 activity, but displayed PTPS activity, whereas *ptps-1* mutants lacked PTPS activity, but had normal GTPCH1, confirming loss of the predicted functions of these genes (Figure 5, A and B). In addition, very low levels of total biopterin and BH4 were detected in the mutants, compared with levels in wild-type worms (Figure 5, C and D). As expected, BH4 supplementation did not alter enzymatic activities of GTPCH1 or PTPS (Figure 5, A and B), which were determined in homogenates freed from low molecular weight compounds by gel filtration. Rearing worms with exogenous BH4 also failed to increase levels of BH4 in mutants, although total biopterins did increase, indicating some uptake of biopterin (Figure 5, C and D). We observed no rescue of cuticle fragility (Figure S8) or neurotransmitter synthesis in

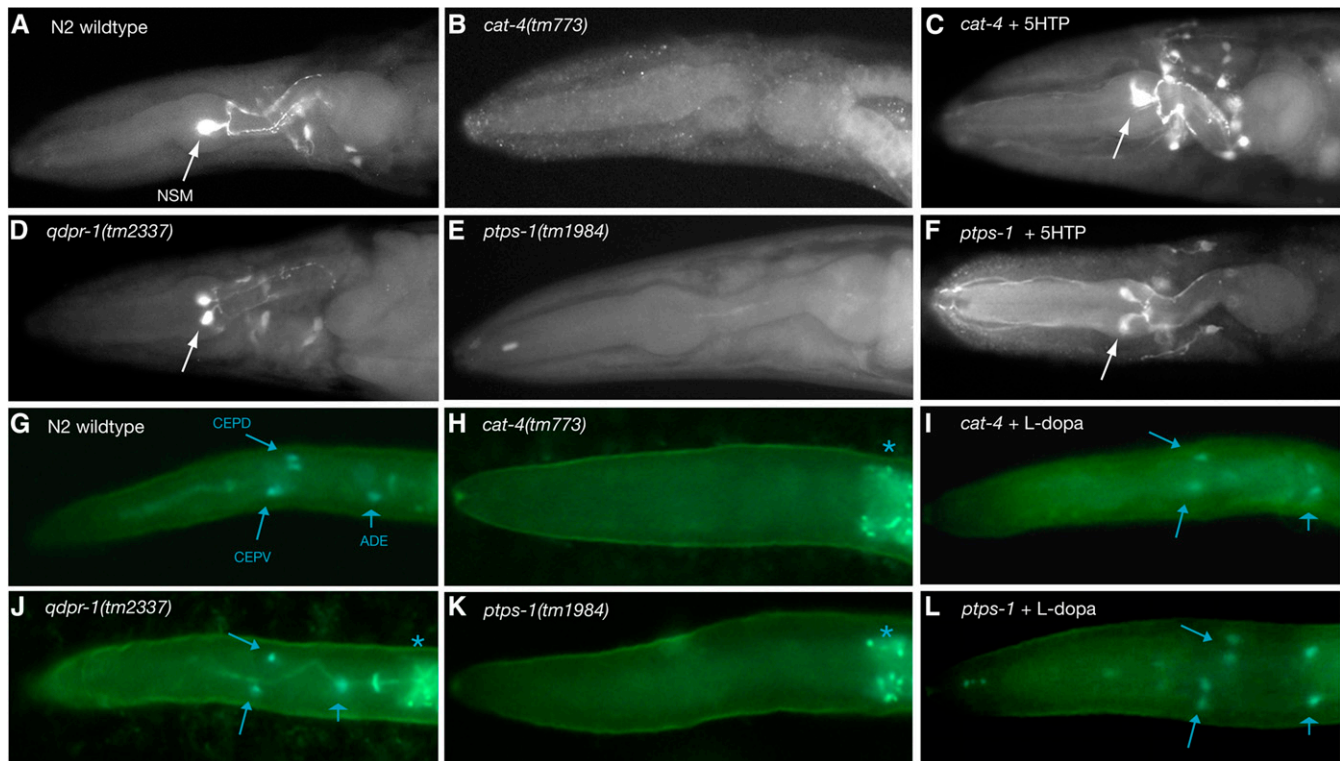


Figure 3 Neurotransmitter phenotypes of *cat-4*, *ptps-1*, and *qdpr-1* mutants. (A–F) Anti-5HT immunofluorescence of adult worm heads. One prominent 5HT neuron cell body (neurosecretory motoneuron, NSM) is marked with an arrow. Anterior is to the left. (A) Wild type (N2) and (D) *qdpr-1* mutants have normal 5HT; (B) *cat-4* and (E) *ptps-1* mutants lack 5HT. (C and F) 5HT-IR is restored in *cat-4* and *ptps-1* mutants by treatment with 5-hydroxytryptophan (5-HTP). 5-HTP also causes dopaminergic neurons to make 5HT. [Rescue of *cat-4* mutants by 5-HTP has been shown previously (Loer and Kenyon 1993).] (G–L) FIF showing dopamine (DA) in larval worm heads. DA-containing cells have characteristic blue-green fluorescence; background is more yellow-green. DA cells in the head are indicated with arrows (CEPD and CEPV, smaller arrowhead; and ADE, broad arrowhead). Asterisks indicate nonspecific intestinal fluorescence. In some heads, one can see both right and left side DA neurons. Anterior is to the left. (G) Wild type (N2) and (J) *qdpr-1* mutants have normal DA; (H) *cat-4* and (K) *ptps-1* mutants lack DA. (I and L) DA fluorescence is restored in *cat-4* and *ptps-1* mutants by treatment with L-dopa.

cat-4 and *ptps-1* mutants supplemented with exogenous biopterins.

The chemical hypersensitivity and cuticle fragility defects of biopterin mutants result from loss of function in alkyglycerol monooxygenase AGMO

As shown previously, knockout mutants in individual AAAH genes (*pah-1*, *cat-2*, and *tph-1*) are normal with respect to chemical sensitivity and cuticle strength (Table 2); *pah-1* mutants display cuticle defects only in sensitized backgrounds in which the cuticle is already compromised (Calvo *et al.* 2008). These observations suggested that deficiency in function of the newly identified BH4-dependent enzyme AGMO might explain the hypersensitivity and cuticle fragility phenotypes of *cat-4* and *ptps-1* mutants. We therefore examined other mutants with similar phenotypes of cuticle fragility. Genetic screens for *C. elegans* mutants with altered sensitivity to bacterial infections of the cuticle have recovered many mutants with associated chemical hypersensitivity and cuticle fragility phenotypes, presumably due to changes in the cuticle or surface properties of the worm (Gravato-Nobre *et al.* 2005). Among these hypersensitive mutants are those that fail to show the characteristic re-

sponse to *Microbacterium nematophilum* infection, a swollen rectal epidermis; such mutants have the “Bus” phenotype (bacterially unswollen). Interestingly, mutations in certain *bus* genes confer greater sensitivity to cuticle infection by bacterial species to which wild-type worms are resistant. The bacterial sensitivity of *bus* mutants can be suppressed by mutation in **suppressor of bus**, or *subs* genes (Hodgkin *et al.* 2013, see *Materials and Methods*). Among *subs* mutants, we found eight independent mutant alleles of *agmo-1*, the *C. elegans* ortholog of the human alkyglycerol monooxygenase (AGMO) gene. Similar to *cat-4* and *ptps-1* mutants, *agmo-1* mutants displayed rapid cuticle disintegration in standard alkaline bleach (Figure 4A). Four *agmo-1* alleles contain premature stop codons, so are likely complete loss of function (Figure 6). Four missense alleles alter amino acids highly conserved among putative AGMO orthologs from other metazoans (Figure 6B, Figure S6), so may abolish or greatly reduce protein function; two of these affect conserved histidines that are among the eight conserved histidines characteristic of fatty acid hydroxylases.

Two new mutant alleles of *cat-4* and one *ptps-1* allele were also isolated via this screen, including a hypomorphic/reduction-of-function allele *cat-4(e3015)*, indicating

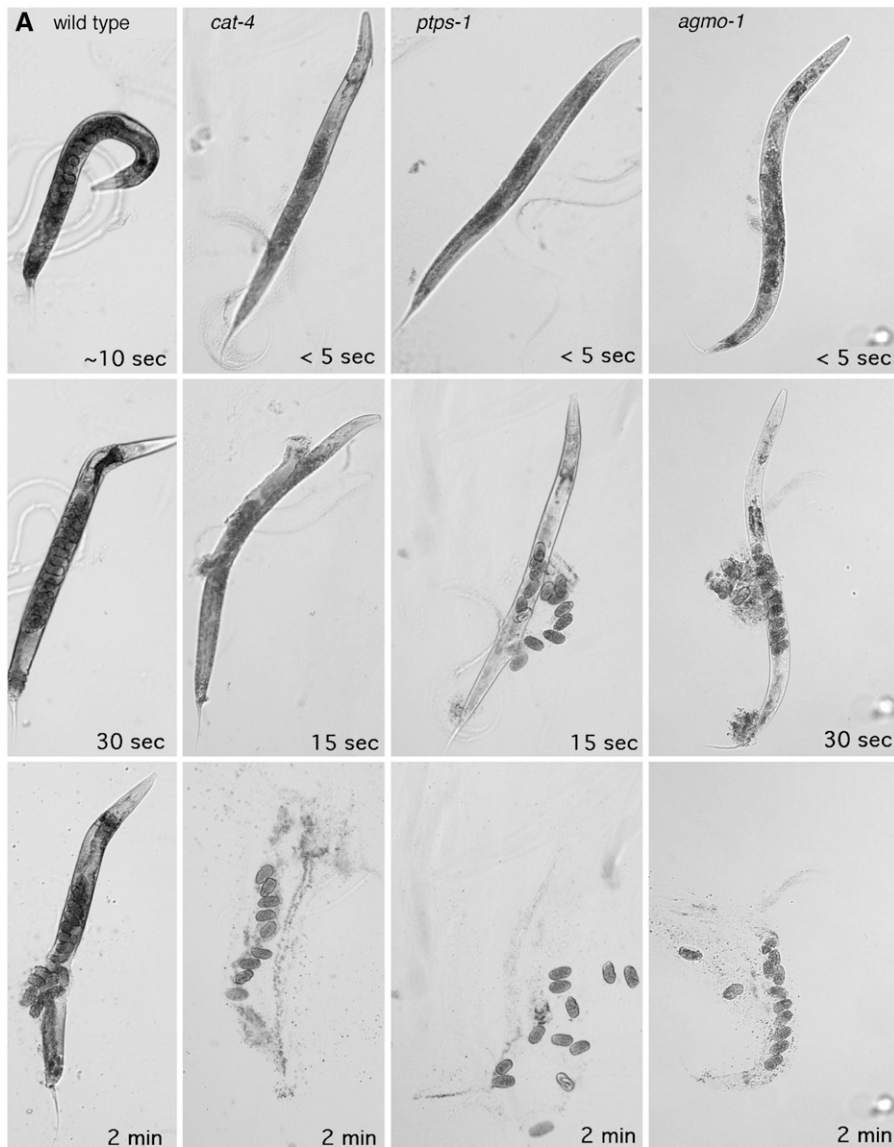
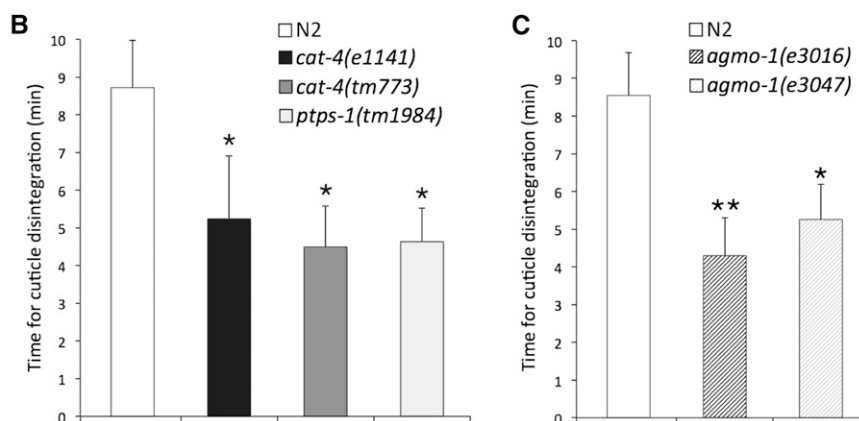


Figure 4 *cat-4*, *ptps-1*, and *agmo-1* mutants share a common hypersensitivity to exogenous chemicals. (A) Hypersensitivity and cuticle fragility shown by rapid death and disintegration of *cat-4*, *ptps-1*, and *agmo-1* mutants in standard alkaline bleach. Adult gravid hermaphrodites were photographed after addition of bleach drop. Top panels: N2 (wild type) worms were still wriggling after 10 sec in bleach, whereas *cat-4(tm773)*, *ptps-1(tm1984)*, and *agmo-1(e3016)* mutants were immobile/dead in a few seconds (<5 sec). Middle panels: N2 worm is dead but intact 30 s after bleaching. Mutants have exploded at multiple sites by 15–30 sec, releasing internal contents. Bottom panels: N2 worm has ruptured, but remains largely intact at 2 min; mutant worms have completely disintegrated and cuticles vanished, leaving eggs and internal debris. (B and C) Cuticle fragility of *cat-4*, *ptps-1* (B), and *agmo-1* (C) mutants. Time (mean \pm SD) for cuticle disintegration in mild alkaline bleach, scored for the first major break in the worm cuticle. Groups for each experiment compared with one-factor ANOVA followed by planned pairwise comparisons made with Scheffé's *F*-test; each experiment showed significant differences among the groups (overall ANOVA, $P < 0.0001$). (B) Representative experiment with worms from mixed stage cultures ($n = 15$); *cat-4* and *ptps-1* compared to wild type (N2). Asterisks (*) indicate significant differences (*F*-test, $P < 0.0001$) between each mutant and wild type. Mutants were not significantly different from one another ($P > 0.05$). (C) Two *agmo-1* mutants compared to wild type (N2), synchronized gravid adult hermaphrodites ($n = 17$ – 27). Both mutants were significantly different from wild type ($P < 0.0001$); double asterisks (**) indicate *e3016* was also significantly different from *e3047* ($P = 0.002$).



that complete loss of bipterin synthesis is not required for the Subs phenotype. All *agmo-1* mutants tested were hypersensitive to a variety of chemicals, including alkaline bleach, SDS, and levamisole, comparable to *cat-4* and *ptps-1* mutants

(Figure 4C, Figure S5, B and D). As expected, *agmo-1* mutants displayed normal levels of DA and 5HT in neurons (Figure S7), indicating the independence of the neuronal and epidermal roles of BH4.

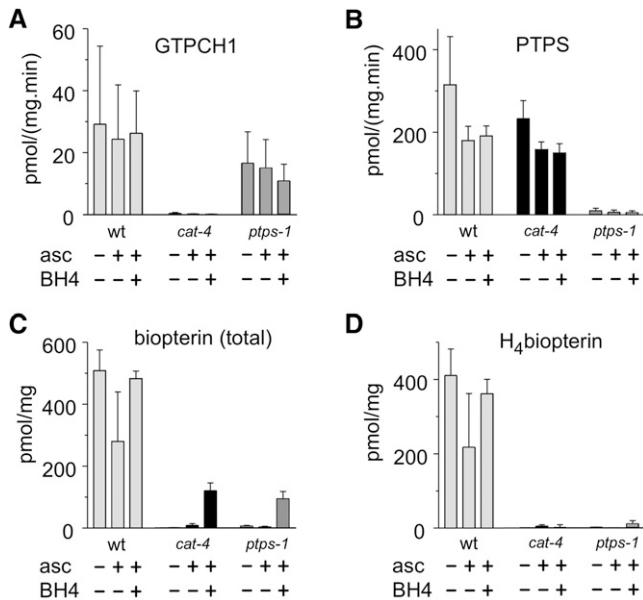


Figure 5 Bioprotein synthetic enzyme activity and bioprotein levels in wild-type and mutant *C. elegans*. Enzyme activity or bioprotein content from worm homogenates from mixed stage cultures. Untreated (-/-), with ascorbate (asc, 5 mM) alone, or asc plus BH4 (200 μ M), as indicated below each column. Enzyme activities and BH4 levels from worm homogenates were determined by HPLC with fluorescence detection (see *Materials and Methods*). All levels are expressed relative to protein mass. Mean \pm SEM are shown for three to four independent measurements (two to three for *cat-4* mutant). (A) GTPCH1 activity. (B) PTPS activity. (C) Total bioprotein derivatives concentration. (D) BH4 concentration. GTPCH1 and PTPS activities were measured in soluble fraction of worm extracts. Total bioprotein and BH4 measurements were taken from the same samples. Worm strains: N2, wild type; *cat-4(tm773)*; and *pts-1(tm1984)*.

The BH4 regeneration cycle maintains biogenic amine levels under conditions of limiting BH4

We further tested putative knockout alleles of the worm bioprotein regeneration enzyme genes *qdpr-1* and *pcbd-1*. These mutants showed no obvious morphological or behavioral phenotypes, and initially appeared wild type for 5HT, DA, and bleach sensitivity (Figure 3, Table 2). Because loss of BH4 regeneration in the context of normal *de novo* BH4 synthesis may not reduce BH4 levels sufficiently to cause an obvious phenotype, we also tested *qdpr-1* and *pcbd-1* mutants in combination with the *cat-4* reduction-of-function allele, *cat-4(e3015)*. Reduction of neurotransmitters in *cat-4(e3015)* worms was most apparent in young larvae; *e3015* adult worms were nearly wild type, suggesting that functional BH4 accumulates over the life of the worm (Figure S9). We found that both 5HT and DA were very strongly reduced in double mutants with *qdpr-1* and *pcbd-1* in comparison to the single mutant *cat-4(e3015)*. The double mutants also showed the strongest reduction in young larvae (Figure 7), although differences were also apparent among adults (Figure S10). For example, although 5HT-IR was absent in only 7–20% of L1–L2 *cat-4* worms, 55% of *pcbd-1(tm5924); cat-4(e3015)* worms lacked 5HT staining (Figure 7, A and C). Similarly, 82% of *qdpr-1(tm2373); cat-4(e3015)* and 88% of *qdpr-1(tm2337); cat-4(e3015)* lacked

5HT staining (Figure 7, B and D). Similar results were obtained using FIF staining to detect DA (Figure S11). These results demonstrate an important role for bioprotein regeneration under conditions in which bioprotein levels may be limiting for neurotransmitter synthesis.

BH4 synthesis genes are expressed in biogenic amine neurons and in the epidermis

Consistent with the known role of GTPCH1 in synthesizing BH4, required for the function of AAHs, GFP reporter constructs show *cat-4* expression in identified serotonergic and dopaminergic neurons (where TPH and TH are expressed, respectively), and in the epidermis (where both PAH and AGMO are expressed, see below). We examined *cat-4* reporter constructs from three different sources, including some previously described by others (see *Materials and Methods*; Table S1 and Table S2). Overall, in larval and adult worms, we observed strong expression in serotonergic and dopaminergic neurons, in most of the epidermis—especially the large epidermal syncytium (*hyp7*)—and more weakly in some intestinal cells (Figure 8A). Given the phenotypes of *pts-1* mutants, and the predicted function of the gene, we expected that the gene's expression would likely match that of *cat-4*/GTPCH1. We made and examined three independent *pts-1::GFP* reporter transgenics (translational fusion in the final coding exon plus 2.0 kb of sequence upstream of the predicted ATG), as well as a *pts-1* reporter transgene (~2.6 kb upstream sequence) described elsewhere (Zhang *et al.* 2014). Transgenics with the shorter upstream sequence resembled the *cat-4* expression pattern, with expression in some epidermal cells, and a few serotonergic neurons (Figure 8B). The *pts-1* reporters, however, were also observed in additional cells, and were not expressed in other epidermal cells or in all 5HT and DA neurons, suggesting these reporters may lack important positive or negative regulatory elements.

We made and examined reporter transgenics for the *pcbd-1*, *qdpr-1*, and *gfrp-1* genes, and also examined some described by others (Zhang *et al.* 2014). All were expressed in the epidermis, similar to that observed in *cat-4* transgenics (Figure 8, C–E), but were not highly expressed in 5HT and DA neurons. The epidermal expression, when observed, was similar in localization, intensity, and developmental timing in larvae. All of these transgenics also showed some expression of varying intensity in other cells (nonepidermal cells, and non-5HT and non-DA neurons in the head and body). When we examined one *qdpr-1* transgenic with 4.9-kb genomic sequence (full-length translational fusion with ~4.0 kb upstream), we observed expression also in several known 5HT and DA neurons (Figure 8D, bottom panel). This transgenic showed some reduction in epidermal expression. With the exception of the larger *qdpr-1* construct, the other previously described transgenics (Zhang *et al.* 2014) were expressed similarly to those reported here.

agmo-1 is expressed throughout the epidermis beginning in embryogenesis

We examined *agmo-1::GFP* reporter fusion transgenics and found that an *agmo-1* transgene with ~2300 bp of upstream

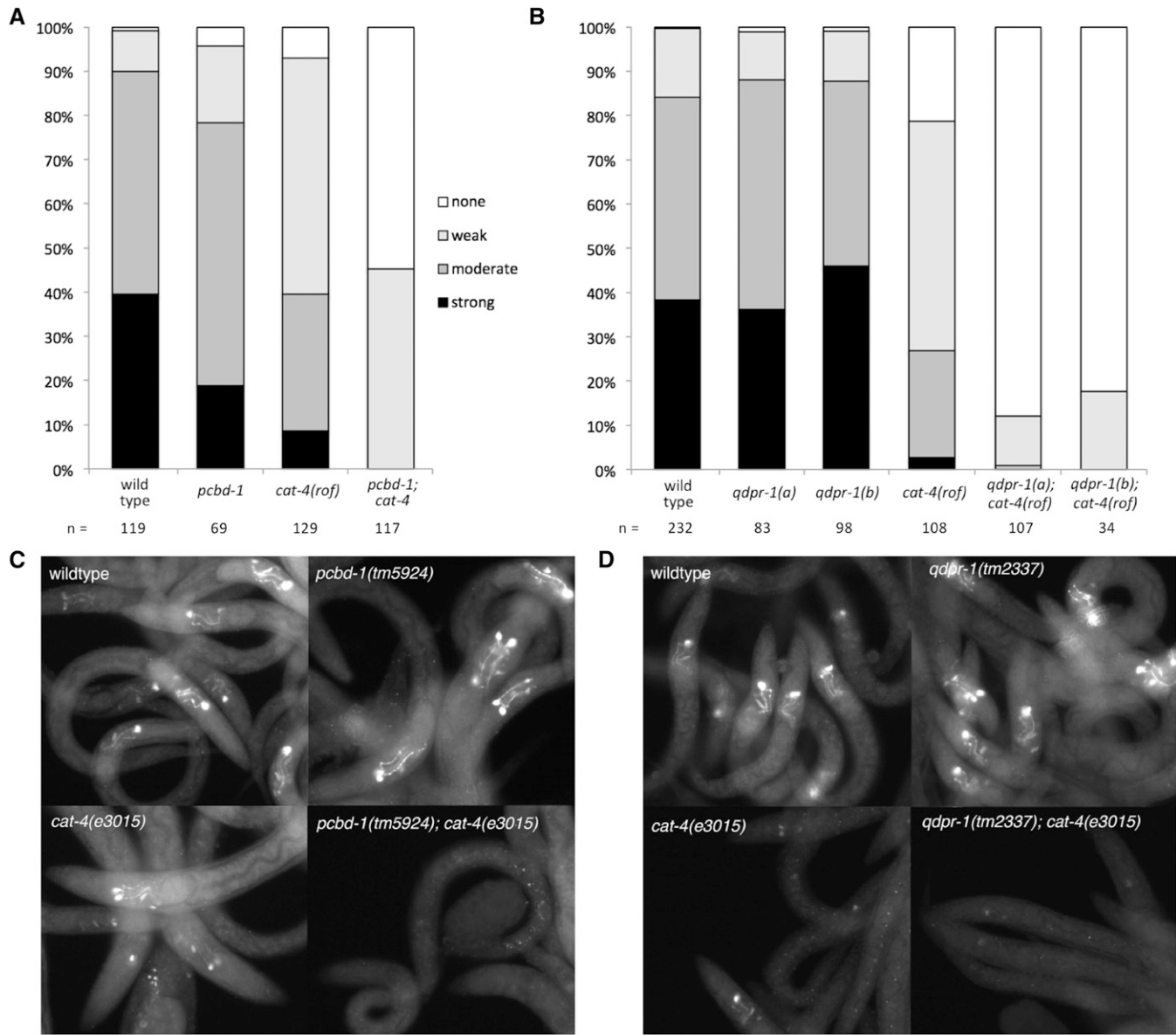


Figure 7 5HT synthesis is dependent on the bioptryin regeneration pathway. Serotonin immunoreactivity (5HT-IR) in NSM somas and neurites of young (L1–L2) worms. Mixed populations of wild type (N2), single, and double mutant worms were scored by serotonin antibody staining. Staining definitions are as follows: strong, somas bright (saturated staining, no internal features apparent) and neurites bright; medium, somas not saturated, may show some internal structure (i.e., a nucleus may be seen), neurites present; weak/faint, somas visible (may be just above background), neurites absent or very faint; and none, no stained structures apparent. (A) *pcb-1(tm5924)* mutants display at most a mild reduction in 5HT-IR. Double mutant *pcb-1; cat-4(rof)* worms display almost no 5HT-IR; *cat-4(rof)* = reduction of function allele = *e3015*. (B) Loss of 5HT-IR in *qdpr-1* mutant worms. Wild type (N2), *qdpr-1*, *cat-4*, and double mutant *qdpr-1; cat-4* worms scored as described in A. *qdpr-1(a)* = *tm2337*; *qdpr-1(b)* = *tm2373*. (C) Examples of staining in mutants including *pcb-1(tm5924)*. (D) Examples of staining in mutants including *qdpr-1(tm2337)*. All images in C and D were taken using the same exposure.

cuticle is being constructed prior to hatching. *cat-4* and *pah-1* reporters are also expressed in embryos, although expression appears earlier in embryogenesis (Figure S13).

Discussion

We have characterized the genetic basis of bioptryin/BH4 biosynthesis and function in *C. elegans*. We have demonstrated that the mutationally identified *cat-4* gene encodes GTPCH1, and that mutations in a PTPS-encoding gene cause phenotypes

identical to those of *cat-4* mutants. *cat-4* and *ptps-1* mutants lack their respective enzymatic activities, have greatly decreased BH4 content, and have phenotypes consistent with their predicted roles in BH4 synthesis, including loss of the neurotransmitters 5HT and DA. Bioptryin synthesis mutants were not rescued by supplemental BH4 or sepiapterin. Although some exogenous bioptryin was taken up, we could only detect the inactive form 7,8-dihydrobiopterin (BH2). In mammalian cell culture and in mice, BH4 is taken up as BH2 and then reduced to BH4 by DHFR (Hasegawa *et al.* 2005). In *C. elegans*, we

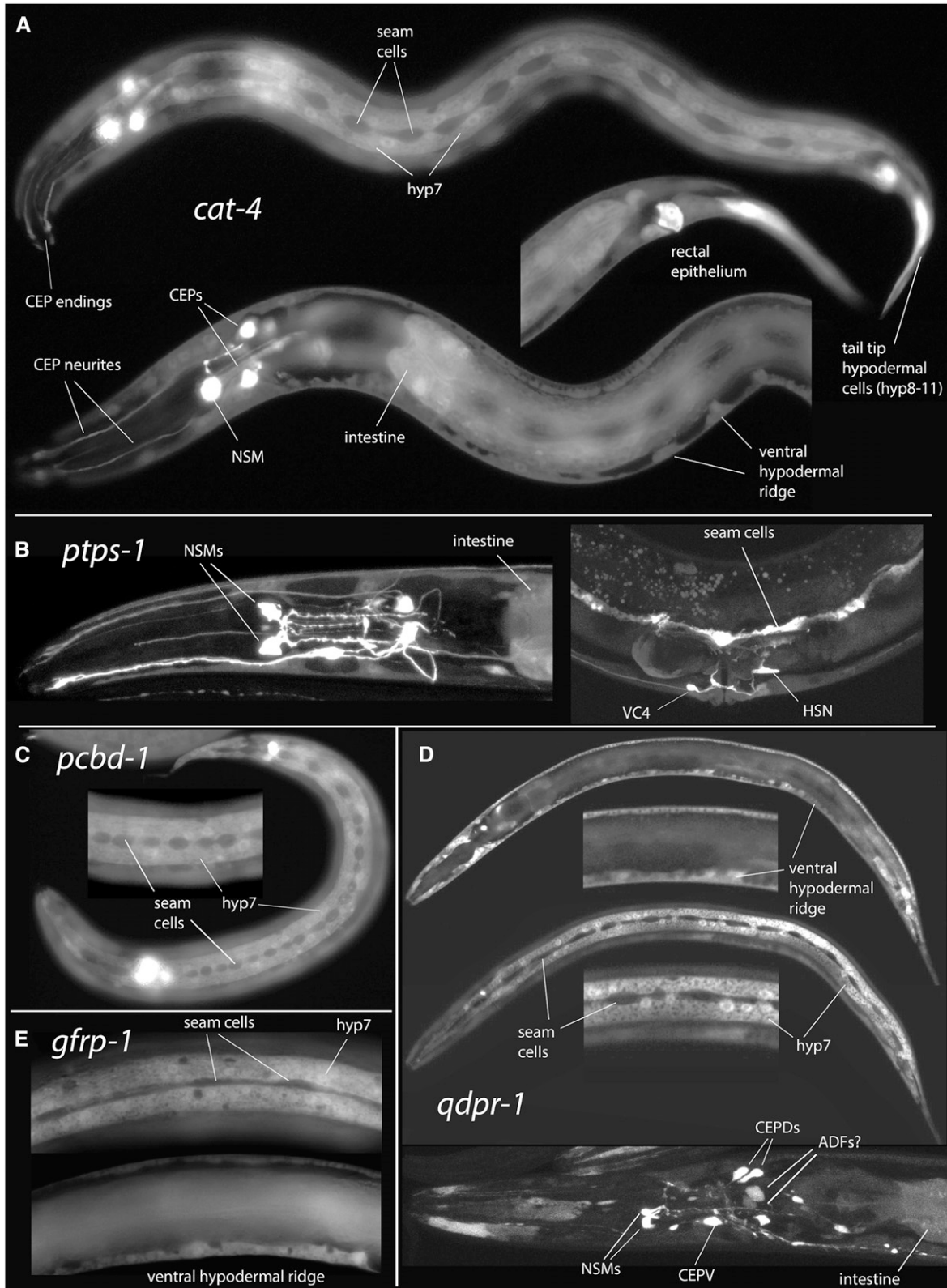


Figure 8 Bipterin synthetic genes are expressed in biogenic amine neurons and in the epidermis. (A) Expression of *cat-4* GFP reporters in larval stage 2 (L2) worms, construct with ~2.7 kb upstream sequence (strain C29719). Top: Superficial focal plane showing epidermal expression, especially in the *hyp7* syncytium. Seam cells have undergone doubling division and can be seen along the lateral side as darker regions among the brightly staining *hyp7* cell. Dendritic endings of CEP neurons can be seen at the tip of the “nose.” Middle: Medial focal plane showing anal cells, strongly expressing tail epidermal cells and posterior intestinal cells expressing GFP. Bottom: Medial focal plane showing epidermal expression in the body and head. In these

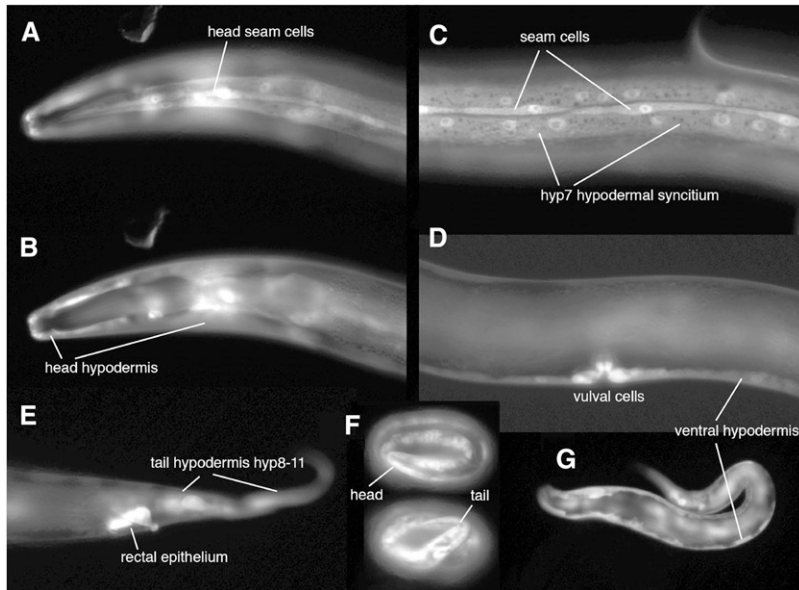


Figure 9 *agmo-1* is expressed in the *C. elegans* epidermis. (A–G) Expression of *agmo-1::GFP* reporter fusions in the epidermis; all worms at same magnification, standard epifluorescence. (A–E) Young adult hermaphrodites; anterior to the left. (A) Head, surface focal plane showing expression in epidermal syncytial cells and seam. (B) Head, mid-depth focal plane showing expression in all head epidermal cells (same head as A). (C) Lateral body surface showing expression in epidermal syncytium hyp7 and lateral seam cells. (D) Mid-depth focal plane showing expression in vulval cells and ventral epidermal ridge. (E) Tail showing strong expression in rectal epithelial cells and other tail epidermis. (F) Two 3-fold-stage embryos with very strong expression in epidermis. (G) Young larva, mid-depth focal plane, showing expression like that seen in adult (and all other larval stages).

hypothesize that exogenous BH4 is similarly taken up as BH2, but for unknown reasons is not subsequently reduced to BH4 by the *C. elegans* DHFR. In any case, the lack of functional rescue of the BH4-deficient mutants by exogenous biopterins is consistent with the general requirement for biopterin synthesis in cells and tissues that use BH4-dependent enzymes (Thöny *et al.* 2000).

Our observations, using 5HT and DA synthesis as a proxy for biopterin levels, demonstrate that *pcbd-1* and *qdpr-1* genes, predicted to encode BH4 regeneration enzymes, function to maintain biopterin needed for neurotransmitter synthesis. The biopterin regeneration mutants showed clear effects on neurotransmitter synthesis when combined with a reduction-of-function *cat-4* mutation. Because of the inherent variability of both techniques—immunofluorescence (for 5HT) and FIF (for DA)—possible slight reductions in staining observed in the *pcbd-1* or *qdpr-1* single mutants could not be definitively assessed. Thus in *C. elegans*, function of these BH4 regeneration genes may be specifically revealed when BH4 biosynthesis is impaired.

Besides lacking 5HT and DA, *cat-4* and *ptps-1* mutants display increased sensitivity to many chemicals, and a less sturdy cuticle. The cuticle defects of *cat-4* have been a long-standing conundrum, as they are not seen in mutants in

known BH4-dependent AAAH enzymes (*pah-1*, *cat-2*, and *tph-1*). Solution of this puzzle awaited the cloning of the last remaining BH4-dependent enzyme, alkylglycerol mono-oxygenase/AGMO (Watschinger *et al.* 2010), allowing identification of the *C. elegans* AGMO ortholog. Mutations in *agmo-1* were predicted to cause cuticle fragility and hypersensitivity, and such mutations were indeed subsequently found in screens for altered resistance to a worm bacterial pathogen that infects animals via the cuticle (Hodgkin *et al.* 2013). Consistent with a role in establishing outer surface properties of the worm, *agmo-1* reporters are expressed throughout the epidermis, which secretes the cuticle, a complex extracellular matrix (Chisholm and Xu 2012). The epidermis is also most likely the cellular origin of the epicuticle, a poorly understood extracellular lipid-rich layer that has been hypothesized to function as a permeability barrier and in pathogen defense (Chisholm and Xu 2012). Identification of AGMO as required for cuticle integrity further supports the idea that lipid metabolism is critical for surface properties of nematodes.

Our analysis of biopterin-related gene expression in *C. elegans* by reporter fusion transgenics is generally consistent with BH4 synthesis being required cell autonomously for AAAH function in 5HT and DA neurons and for AGMO function in the epidermis.

transgenics, expression in the head is seen in hyp6, but not the more anterior hyp5. NSM and CEP neuron somas are seen in the head, plus some neuronal processes (especially CEP processes). A few other neuronal somas stain less brightly. The anterior intestine also shows GFP expression, as do some rectal epithelial cells (likely B and Y cells). (B) *ptps-1*-GFP reporters. Left: head showing NSM expression plus other neurons and nonneuronal cells, including anterior intestine (dorsal–ventral view, maximum intensity projection) (MIP); strain OH11619, described by Zhang *et al.* (2014). Right: vulval region showing expression in neurons VC4, 5 (weak) and HSN, plus lateral seam cells (lateral view, MIP); strain CZ18321. (C) *pcbd-1* reporter in larva showing expression in epidermal syncytium hyp7, tail and head epidermis, but not in seam cells; strain CZ19212. Out of focus: pharyngeal muscle and rectal epithelium expression. Inset: enlargement of midbody region. (D) *qdpr-1* reporters. Top images: single optical sections from larva showing broad epidermal expression, but little expression in neurons; strain CZ19213. Insets: enlargements of midbody regions. Bottom image: Head showing expression in identified 5HT and DA neurons: NSMs, ADFs, CEPs, and other cells, with reduced epidermal expression; strain CZ19215 (lateral view, MIP). (E) *gfrp-1* reporter showing expression in epidermal syncytium hyp7 (top: superficial focal plane) and other epidermal cells (bottom: central focal plane). (A–E) Anterior is to the left in all worms. (A, C, and E) Standard epifluorescence. (B and D) Laser scanning confocal imaging; (B and D, bottom) images are maximal intensity projection of Z-stack; and (D, top and middle) single confocal image planes.

Not all our reporter transgenics, however, were expressed in patterns expected based on mutant phenotypes of the genes examined. Transgenes reported here are mostly transcriptional fusions and may lack positive or negative regulatory elements. Further work, using rescuing transgenes or under endogenous or tissue-specific control, will be required to fully define the cellular requirements for BH4 synthesis.

Our studies reveal the first *in vivo* biological role for the BH4-dependent lipid metabolic enzyme AGMO, which is the only enzyme known to degrade the ether lipid bond in alkylglycerols and alkylglycerol-lyso-phospholipids (Watschinger and Werner 2013). The precise biochemical role of AGMO in the cuticle and permeability barrier in the worm remains to be established. We do not know whether alkyl ether lipid metabolism in *C. elegans* serves primarily an anabolic or catabolic function: *agmo-1* mutant phenotypes could result from failure to synthesize a needed product or by accumulation of toxic intermediates. Failure of the AGMO reaction might lead to accumulation of ether lipids, somehow destabilizing the lipid-rich epicuticle. Alternatively, AGMO-1 substrates might alter cuticle development via signaling pathways, analogous to the effects of antitumor ether lipids on mammalian tumor cells (Arthur and Bittman 1998).

Other mutants with altered pathogen sensitivity in *C. elegans* display chemical hypersensitivity and cuticle fragility. The *bus* mutants are partially resistant to bacterial rectal infection by certain worm pathogens, apparently due to alterations in cuticle surface features (Gravato-Nobre *et al.* 2005). Some of these mutants are bleach hypersensitive, dying and rupturing more quickly than N2 wild type. Several *bus* genes have now been identified, and some encode lipid metabolic enzymes expressed in cells that overlap with *cat-4* expression, suggesting a pathway association with *agmo-1*. For example, *bus-18* (aka *acl-10*) encodes a lysocardiolipin acyltransferase expressed in the epidermis (Gravato-Nobre and Hodgkin 2008; Imae *et al.* 2010). However, in general, the cuticle defects of such *bus* mutants are more severe than those of bipterin synthesis or *agmo-1* mutants and extend to outer cuticle sloughing during handling, rupture at the vulva, and “skiddy” locomotion. These more severe cuticle defects suggest additional, widespread roles for lipid metabolism in the cuticle.

In conclusion, our systematic analysis of BH4 function in *C. elegans* confirms its essential role as a cofactor for TH and TPH in neurotransmitter synthesis in neurons. In addition to this well-established function, we elucidate here a new role for BH4 as a cofactor for AGMO in epidermal cells. Our studies reveal an unexpected *in vivo* role for AGMO in supporting cuticle stability and sensitivity to bacterial infection, presumably via ether lipid metabolism. These observations raise the question of whether AGMO might play comparable roles in epidermal function in other animals or in humans. The presence of a complete enzymatic pathway for *de novo* synthesis of BH4 has been established in the human epidermis (Chavan *et al.* 2006), but at present no mutations in mammalian or human AGMO have been reported, nor have severe skin defects been described in patients deficient in

BH4 synthesis. Nevertheless, we note that the products of AGMO catalysis are converted to fatty acids by fatty aldehyde dehydrogenase (FALDH), and that FALDH deficiency in humans results in a combination of skin permeability barrier defects, ichthyosis, and neurological disease known as Sjögren-Larsson syndrome (Rizzo 2011). It would be interesting to explore whether AGMO contributes to lipid-based permeability barrier function in other organisms or in humans.

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GENETICS

Supporting Information

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Cuticle Integrity and Biogenic Amine Synthesis in *Caenorhabditis elegans* Require the Cofactor Tetrahydrobiopterin (BH₄)

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SL1 spliced leader

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M S R I E N E S G F L S S D A A S V G S

gaagacgataaagttgagatgaagaaaagaaacggaacgattccaaaggaagatcatttg
E D D K V E M K K R N G T I P K E D H L

aaatcaatgtgcaatgcctatcagagcatcattcaacatgtcggagaagacatcaatcgt
K S M C N A Y Q S I I Q H V G E D I N R

cagggacttctgaaaactccagaacgtgctgccaagcaatgatggcattcaciaagggga
Q G L L K T P E R A A K A M M A F T K G

tacgatgatcaacttgatgagctcctcaacgaggcagtattcgacgaggatcacgatgag
Y D D Q L D E L L N E A V F D E D H D E

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M V I V K D I E M F S L C E H H L V P F

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M G K V H I G Y I P N K K V L G L S K L

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A R I V E M F S R R L Q V Q E R L T K Q

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I A T A M V Q A V Q P S G V A V V I E A

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S H M C M V M R G V Q K I N A S T T T S

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C M L G V F R D D P K T R E E F L N L I

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N K R

tactaacttttcataactttccatgtgaatctactaacatcttatgcttcagctttttaa
attcatacaacatgttcattgcttg

Figure S1 *cat-4* cDNA sequence encodes a 223 amino acid protein. We isolated, cloned and sequenced *cat-4* cDNAs by RT-PCR from mixed stage *C. elegans him-5* worms as previously described (Hare and Loer 2004); cDNA clones were also obtained from the ORFeome project (Reboul et al. 2001). For *cat-4*, twelve independent clones from pooled ORFeome DNA were partially sequenced; 11/12 were identical F32G8.6 cDNAs, 1/12 was a different non-F32G8.6 sequence. We found that *cat-4* message is trans-spliced to SL1 (underlined at the 5' end). All cDNA clones we examined and those found in databases from various transcriptomics projects showed the same pattern of splicing, indicating there is likely a single type of mRNA and therefore a single isoform of the worm GTPCH1 protein with 223 amino acids. Introns are located in genomic sequence following the base indicated by a red arrow. Genbank accession numbers associated with these *cat-4* cDNA clones are KP290890-KP290893.

We also used RT-PCR to isolate *cat-4* cDNAs from *C. briggsae* which were very similar to those from *C. elegans*. RT-PCR from the diplogastrid nematode *Pristionchus pacificus* yielded 2 different cDNAs: a 9 exon transcript encoding a 270 aa protein, and an 8 exon transcript encoding a 245 aa protein. The longer cDNA matches some gene predictions, whereas the shorter cDNA encodes a more typical GTPCH1 protein.

It is notable that GTPCH1 transcripts are among the most abundant transcripts in some nematodes. For example, among SL1-spliced ESTs from a mixed stage library from the plant parasitic nematode *Pratylenchus penetrans*, 6% were from GTPCH1 (Mitreva et al. 2004). In several mammalian gastrointestinal parasites (e.g., *Ostertagia ostertagia*, *Haemonchus contortus*, *Teladorsagia circumcincta*), a significant fraction of transcripts – up to 30% – from infective L3 stage larvae encoded GTPCH1 (Hoekstra et al. 2000; Moore et al. 2000; Nisbet et al. 2008).

We also examined worms with the *cat-4* missense allele *gk245686* from the Million Mutation Project (Thompson et al. 2013). In this mutant, there is a relatively conservative change (M185I), although the alteration is in an amino acid that is 100% conserved in metazoans, and likely within the active site (see Fig 1). Perhaps surprisingly, worms with this mutation had no obvious neurotransmitter deficiency or bleach hypersensitivity, although the change is conservative.

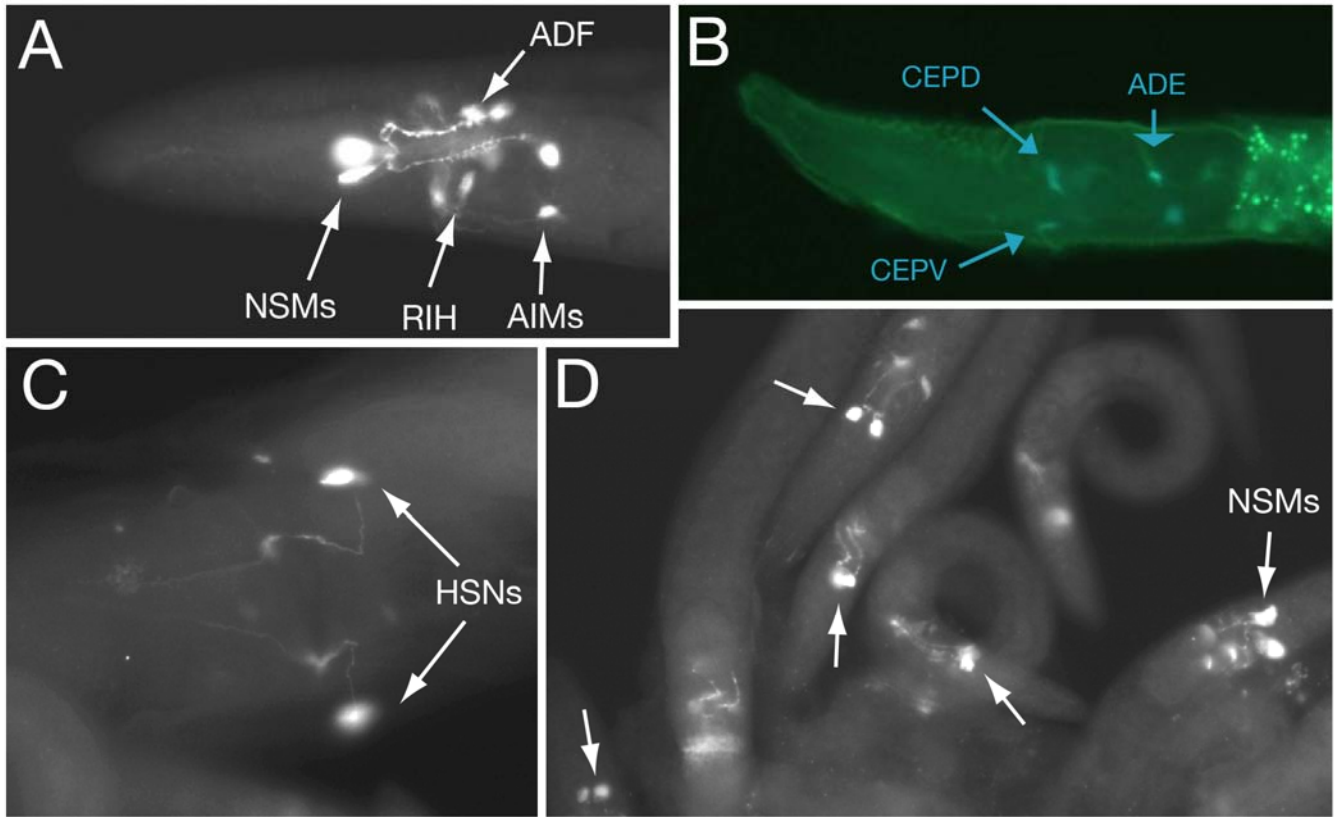


Figure S2 Rescue of neurotransmitter synthesis in *cat-4* mutants by genomic F32G8.6-containing sequence. Anti-5HT immunofluorescence (A, C, D) or FIF (B) of adult and larval *cat-4(tm773)* worms rescued with F32G8.6-containing plasmid (from Baker et al. 2012). (A) Head of adult worm showing presence of all normal serotonergic neurons; somas indicated with arrows. NSM neurites are apparent. (B) FIF of larval worm head showing presence of normal complement of DA neurons. (C) Adult hermaphrodite serotonergic HSN egg-laying neurons innervating vulval region and extending neurites anteriorly to the head. (D) Many rescued (5HT positive) larvae. NSM neuron somas indicated with arrow.

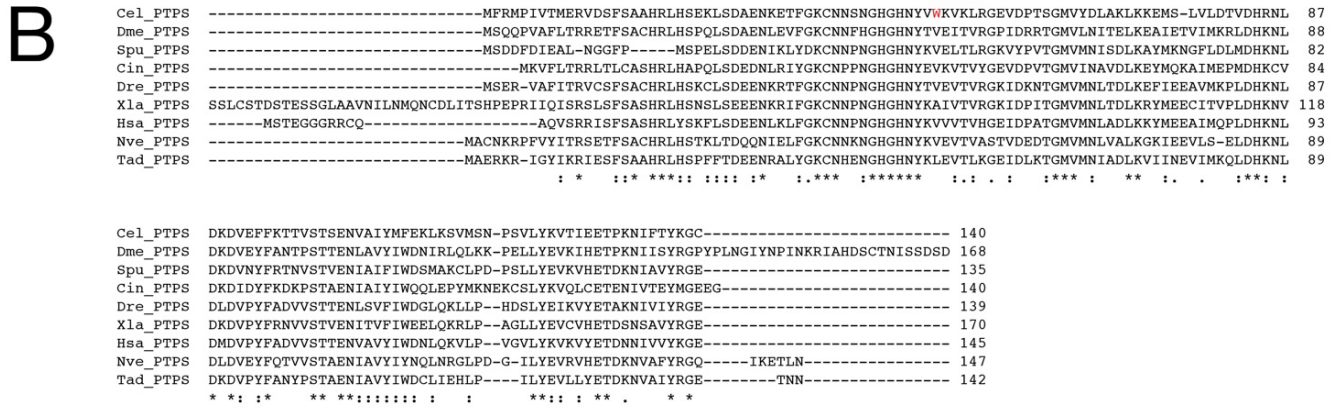
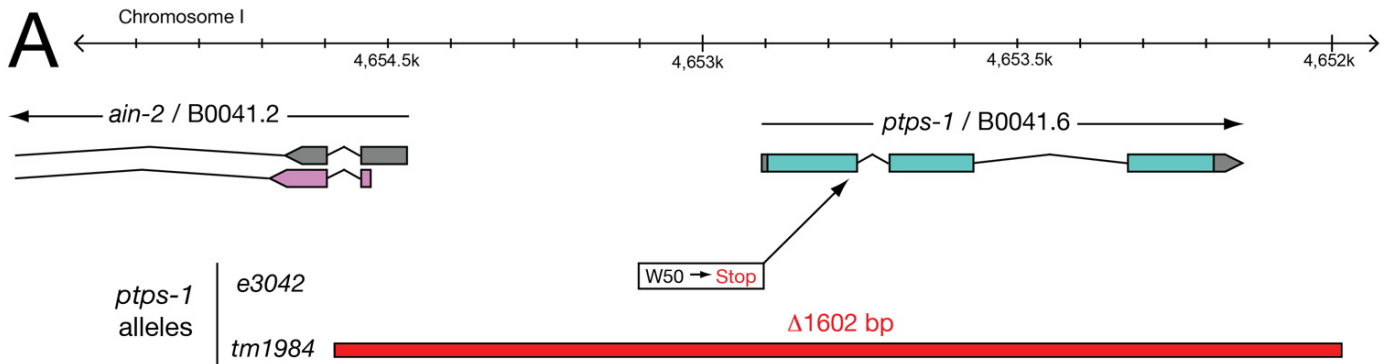


Figure S3 The *pts-1*/B0041.6 gene encodes 6-Pyruvoyl Tetrahydropterin Synthase (PTPS). (A) Physical map of *pts-1* region with mutant alleles and gene model. Nature of *pts-1* alleles is shown below and approximate locations indicated with arrow (point mutation) or red bar (deletion). Various EST, OST, and transcriptome sequencing support the gene model shown, and the predicted protein sequence. Image partly derived from WormBase genome browser editable SVG. (B) Alignments of *C. elegans* PTPS-1 predicted protein with PTPS proteins from other metazoans. Asterisks below alignment show 100% conserved amino acids, colon indicates conserved highly similar aa's, period indicates conserved weakly similar aa's. Location of *pts-1* mutant allele (*e3042*) marked with red letter. Species abbreviations: Cel – *C. elegans*, Dme – *Drosophila melanogaster*, Spu – *Strongylocentrotus purpuratus*, Cin – *Ciona intestinalis*, Dre – *Danio rerio*, Xla – *Xenopus laevis*, Hsa – *Homo sapiens*, Nve – *Nematostella vectensis*, Tad – *Trichoplax adherens*.

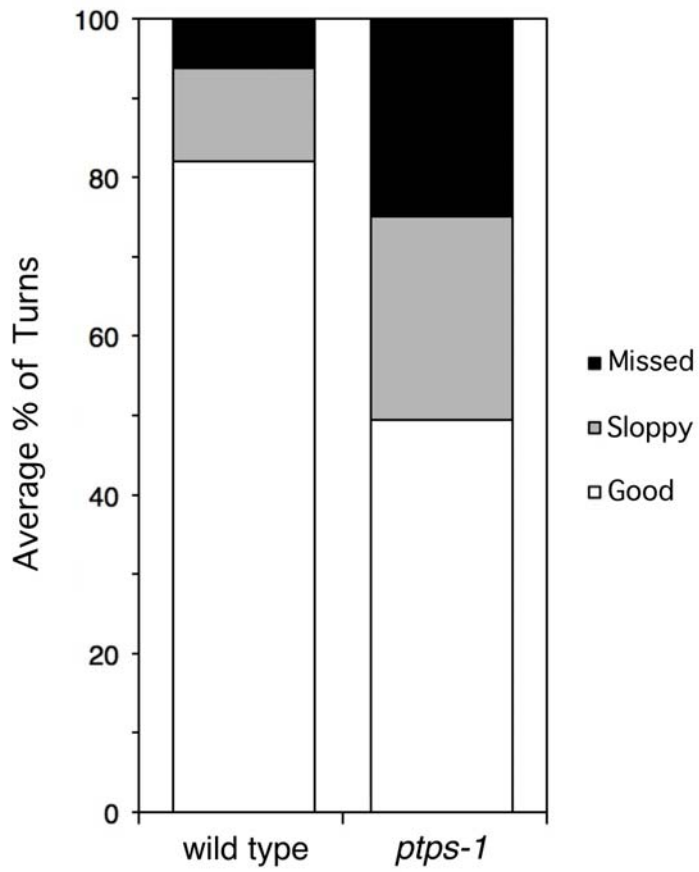


Figure S4 Male turning behavior is defective in *ptps-1* mutants. Columns show average percentage of each type of turn (good, sloppy, missed; definition of turn types from (Loer and Kenyon 1993) for each individual male, each observed for 5 min (wild type, n = 10 *him-5(e1490)* males; n = 12 *ptps-1(tm1984)*; *him-5(e1490)* males).

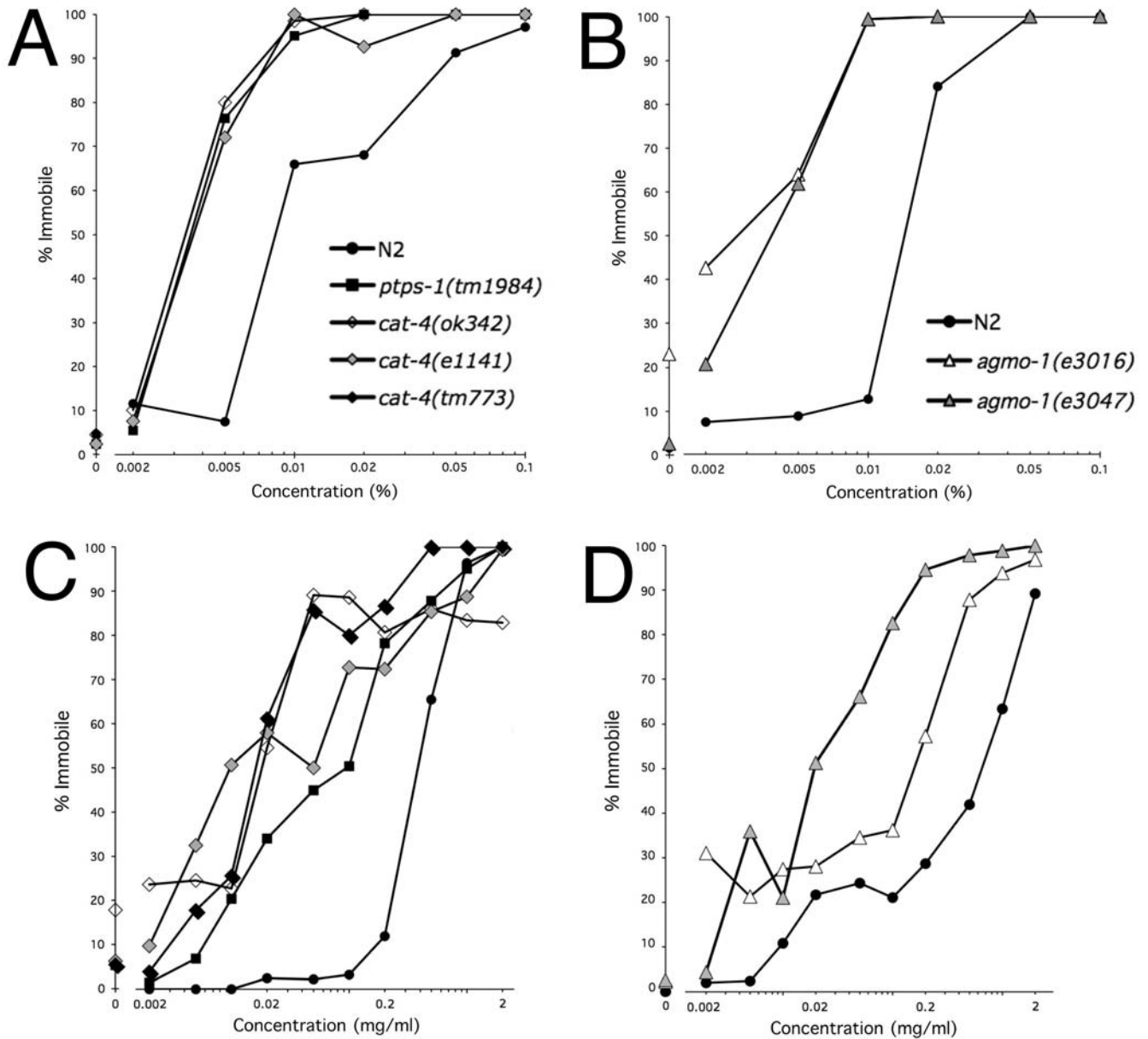


Figure S5 Similar hypersensitivity phenotypes of *cat-4*, *ptps-1* and *agmo-1* mutants. (A, B) SDS hypersensitivity of *cat-4*, *ptps-1* (A) and *agmo-1* (B) mutants. Synchronized adult gravid hermaphrodites were exposed to SDS in M9 buffer for 30 min, then examined for movement. Legend indicates alleles tested. (C, D) Levamisole hypersensitivity of *cat-4*, *ptps-1* (C) and *agmo-1* (D) mutants. Worms were exposed to levamisole in M9 for 5 min, then examined for movement. Legends as in A, B.

SL1 spliced leader

```

1  ggtttaattacccaagtttgagaactcttctccacaaaattttaaccaaagaaaagaggatgaacgagacggca 75
                                     M N E T A
76  tggttggatagggatttctcaaatacgtcactgggacatcgattgctggatagactaactctcacgaatcttcgt 150
    W L D R V F S N T S L G H R L L D R L T L T N L R
151  cacgcattctaccttatctcaccttacgagaccaccgctcgagtcgattgatgatgttccgaactataacgctgaa 225
    H A F Y L I S P Y E T T V E S I D D V P N Y N A E
226  gtttctgctgggtggctgggtgtttcttactgccgagttttcattttattcatttccggacatgaagacagattt 300
    V S A W W L V F L T A E F F I L F I S G H E D R F
301  gcactcaatgattcaataacgtcaatttgtgctggaatgctcagccaatgcttcaaatgggtggctcgagctgtc 375
    A L N D S I T S I C A G M L S Q C F K F G G R A V
376  gcgatattcttgtagctgattgtgtgggataactggcgaatattagaacctccatgggattccccgtggacatgg 450
    A I F L Y V I V W D N W R I L E P P W D S P W T W
451  atttttgcctgttctttcaagattttatgtattatctggggcatcgggctgtgcatggaagccggcttcttctgg 525
    I F C L F F Q D F M Y Y L G H R A V H E A G F F W
526  ggtcttcacacaattcaccatagctccgaataactacaatttctcaactgctctacgacaagctgccatacaagat 600
    G L H T I H H S S E Y Y N F S T A L R Q A A I Q D
601  gctggattagcgatctatgactgtattcagccattcttcatccctccatcaatatttttagttcatcgatatttt 675
    A G L A I Y D C I Q A F F I P P S I F L V H R Y F
676  tcggagattttccaatttatcatgcatacctcgttgggtggacaccatgggacccttggcttagtattcaacact 750
    S E I F Q F I M H T S L V D T M G P L G L V F N T
751  ccgtctcatcatcgagtacatcatggaagaaatccgtattgtattgacaagaattatggaggagttttcattatt 825
    P S H H R V H H G R N P Y C I D K N Y G G V F I I
826  tgggataagatgttcaacacatttgaagccgaacgtcagcatgaccaccaatctatggattagttaccaacgag 900
    W D K M F N T F E A E R H D D P P I Y G L V T N E
901  aacactttcaatcaaactctacctccaattccatgctctttgggacatttttaattttcaaaggattcacaaaagat 975
    N T F N Q I Y L Q F H A L W D I L I F K G F T K D
976  gtgaaaggagagcccattgtttcctggagttgtgaacaaattgaaagcaaccgtattcccgccgggctggttcca 1050
    V K G E P M F P G V V N K L K A T V F P P G W F P
1051  ggggttctgtcaccctgttctttcattggatgagcatgggtaatccagctcacggagtacctgagccagagaaa 1125
    G V P V T P F F H W M S M V N P A H G V P E P E K
1126  cctgttctcagatacagcccacctgcgaggatcctagtgaagtttacgtggcatcgtcattcttgttgttgttg 1200
    P V L R Y S P P A R I L V K V Y V A S S F L L L L
1201  gctatattcttccattttgaatacagaccggaatcatttgagctacttggattgtacagtcaagattgcatacttt 1275
    A I F F H F E Y D R N H L S Y L D C T V K I A Y F
1276  gtggttacgatgcaatgttttggagcattttttgatatgaaatgggtatgcccggtacattgaaattgctcgttgt 1350
    V V T M Q C F G A F F D M K W Y A R Y I E I A R C
1351  tgtggagttctcatctattacggagtactcatgttcgatcatattgggtgcaggaactcatcgtctttttgtcatt 1425
    C G V L I Y Y G V L M F D H I G A G T H R L F V I
1426  tcaactgcataatcatggctattgcattgtggacgactgatgttttgggtggagaaactctcccaatgctgctcaaag 1500
    S L H I M A I A L W T T D V L V E K L S Q C C S K
1501  aatcaatcaataaatccagaaaaaggtgacctggaacgggctccagaaattgcatcgatctcgaaaaatgttcaa 1575
    N Q S I N P E K G D L E R A P E I A S I S K N V Q
1576  taatatgatttttatagagttgttgtttccattgtctttccaggctgtcatattctagtcaactcttttccctttt 1650
    *
1651  ttgtttgatggtttttcttaaatgaagattttttctctcaaaaaa 1725

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Figure S6 An *agmo-1* cDNA encodes a 505 amino acid protein. For *agmo-1*, we completed sequencing of cDNA yk1597b01 (previously partially sequenced, accession BJ763208) kindly provided by Yuji Kohara. The clone had a single nucleotide difference from genomic sequence that would result in a single amino acid difference from that predicted from genomic sequence (nt: T425C; aa: L122P). Both differences are marked in red. Introns located following the base indicated by a red arrow. The trans-spliced leader SL1 is underlined at the 5' end; the likely polyadenylation signal is underlined in the 3' UTR. We also sequenced twelve independent ORFeome clones (Reboul et al. 2001). We found clones of two different lengths, but both types appeared to be abnormal with an unspliced intron #4 which would result in a severely truncated protein. Genbank accession number for this *agmo-1* cDNA clone is KP290894.

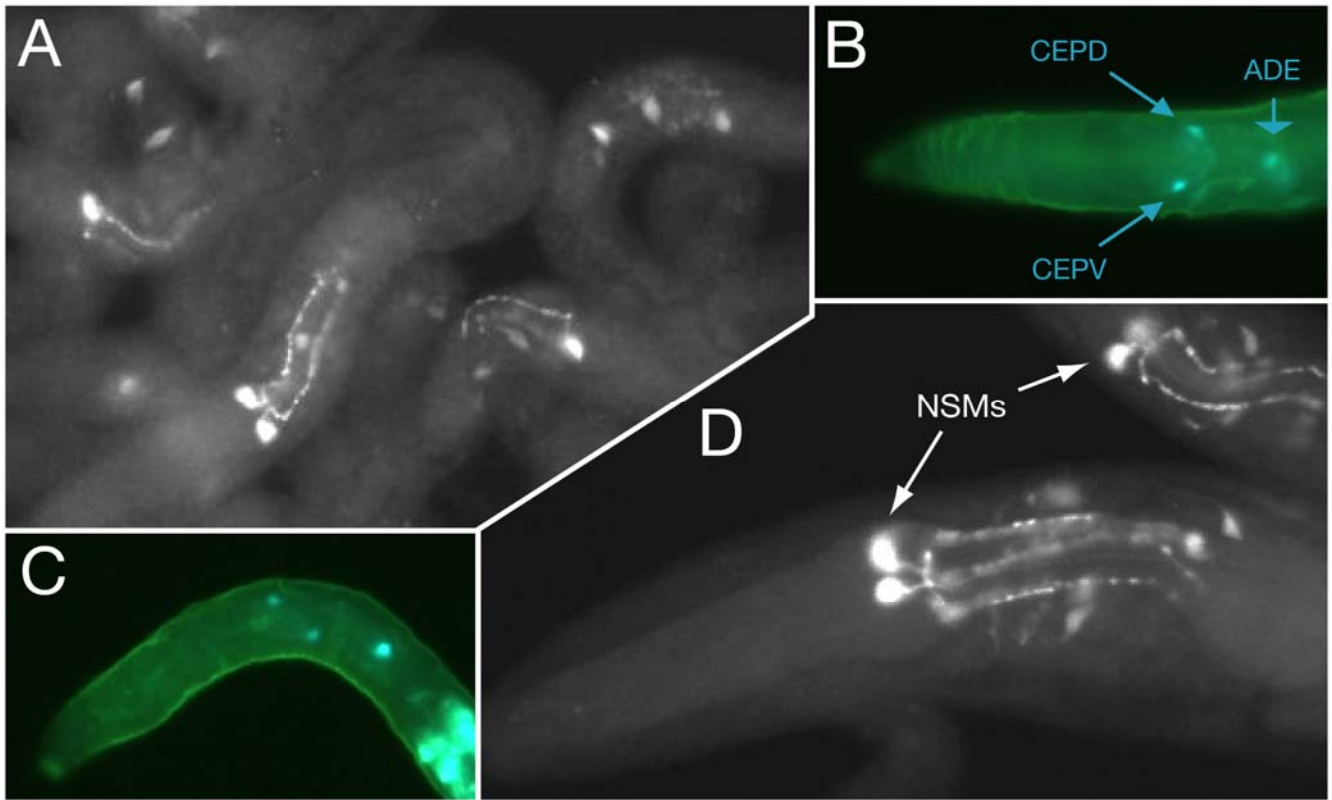


Figure S7 *agmo-1* mutants have normal 5HT and DA. (A, B) *agmo-1(e3047)*; (C, D) *agmo-1(e3016)*. Anti-5HT immunofluorescence (A, D) of worm heads of larvae (A, D) and adult (D) showing normal 5HT neurons. FIF (B, C) of larval heads showing normal DA neuron staining.

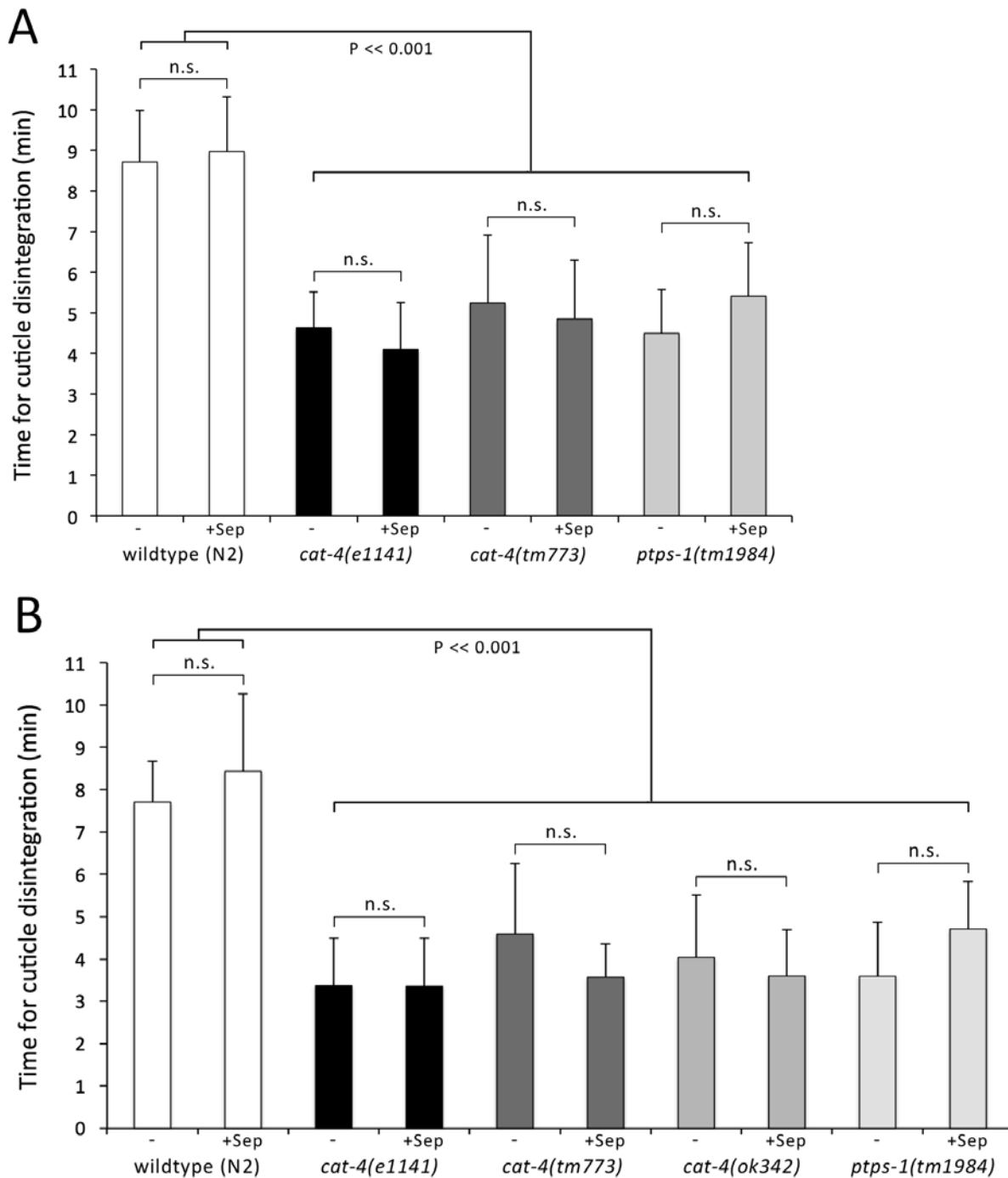


Figure S8 Exogenous biopterins do not rescue cuticle fragility in BH4-deficient mutants. (A-E) Cuticle disintegration time (mean \pm S.D.) during mild alkaline bleach treatment, scored as first major cuticle rupture; worms from mixed stage cultures ($n = 15$), as in Fig 4 (B, C). For all experiments, groups were compared with 1-factor ANOVA followed by planned pairwise comparisons made with Scheffé's F-test (Sokal and Rohlf 1981). (A) Wild-type worms compared to biopterin deficient worms, with or without supplemental sepiapterin (100 μ M). There were significant differences among the groups (overall ANOVA, $P < 0.0001$). All pairwise comparisons between a genotype with sepiapterin (+Sep) or without (-) were not significant (n.s.). All pairwise comparisons of wild type with mutants were significantly different ($P < 0.001$). No biopterin deficient genotype was significantly different from another. (B) Additional experiment comparing wild-type to biopterin deficient worms, with or without supplemental sepiapterin, including *cat-4(ok342)*. Results like in (A). A customary approach in mammalian cells to increase BH4 intracellularly is to use sepiapterin (Werner-Felmayer et al. 2002; Aguado et al. 2006), since SR can effectively convert sepiapterin to 7,8 dihydrobiopterin (BH2) intracellularly, which is then reduced to BH4 by dihydrofolate reductase (DHFR). In *C. elegans*, we believe that other reductases substitute for SR and carry out the last steps in the synthesis of BH4 (since there is no ortholog); these enzymes should also be able to convert sepiapterin to BH2, the latter being further reduced to BH4 by DHFR in the so-called salvage pathway (Thöny et al. 2000).

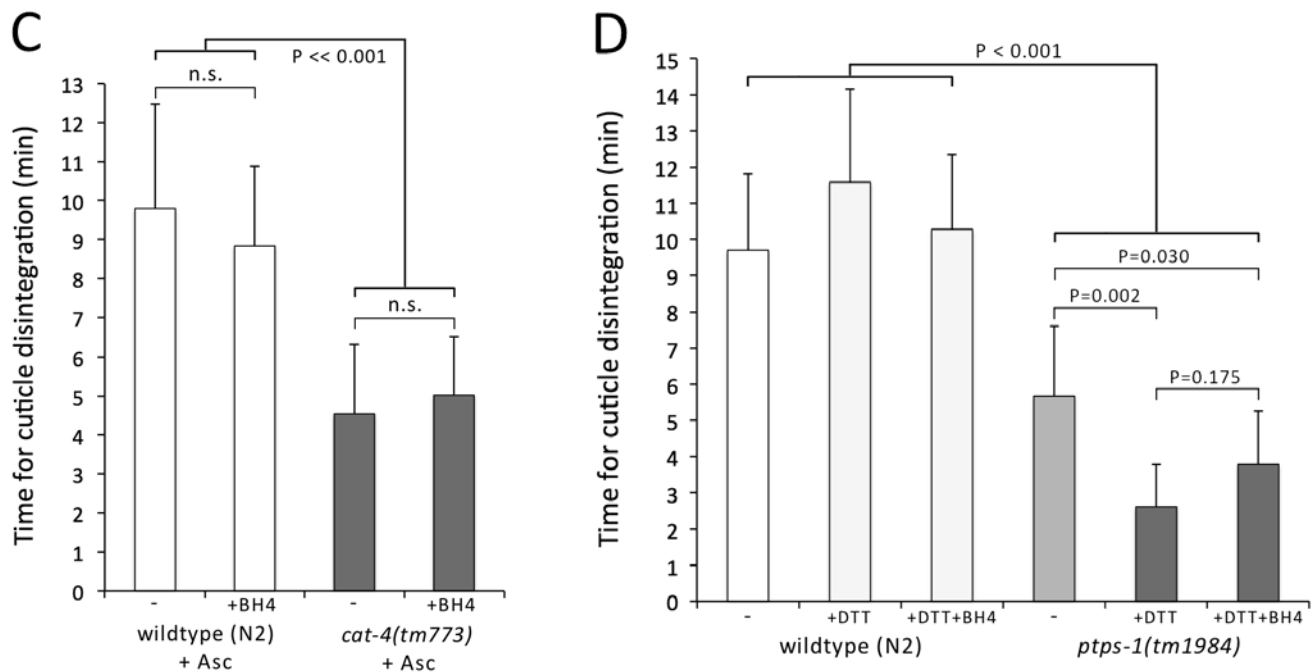


Figure S8 Exogenous biopterins do not rescue cuticle fragility in BH4-deficient mutants. (C) Supplementation with BH4 (200 μ M), including ascorbate (Asc, 5 mM added to reduce oxidation) does not alter cuticle fragility in wild type or *cat-4(tm773)* mutant. Comparisons as in (A). (D) Supplementation with BH4 (200 μ M) with or without dithiothreitol (DTT, 5 mM added to reduce oxidation) does not alter cuticle fragility in wild type (left columns, all N2 group comparisons not significant). Addition of DTT makes *pts-1(tm1984)* mutant cuticles significantly more fragile ($P < 0.05$ in pairwise comparisons with or without DTT). BH4 may reduce damage caused by DTT although the difference is not statistically significant ($P = 0.175$). DTT can reduce disulfide bonds in the highly cross-linked *C. elegans* cuticle (Stenvall et al. 2011).

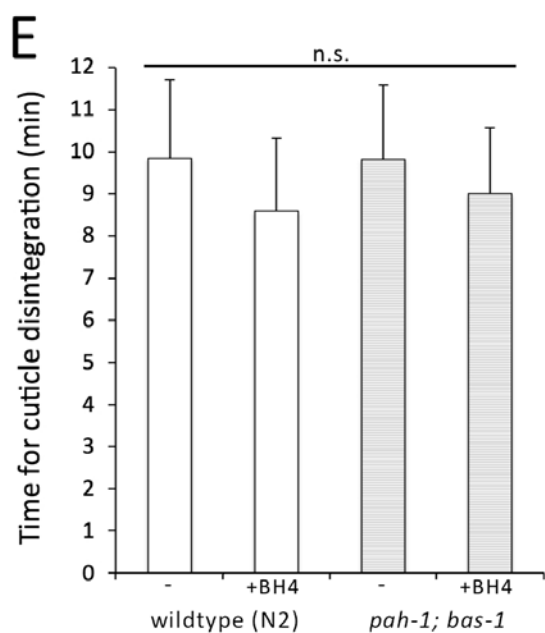


Figure S8 Exogenous biopterins do not rescue cuticle fragility in BH4-deficient mutants. (E) Loss of PAH activity, 5HT and DA do not affect cuticle fragility; supplementation with BH4 does not affect cuticle fragility of wild type or *pah-1; bas-1* mutant. There were no significant differences among the groups in the overall 1-factor ANOVA. Double mutant *pah-1(tm520); bas-1(ad446)* lacks PAH and aromatic amino acid decarboxylase activities; *bas-1* mutants are 5HT- and DA-deficient (Hare and Loer 2004; Calvo et al. 2008).

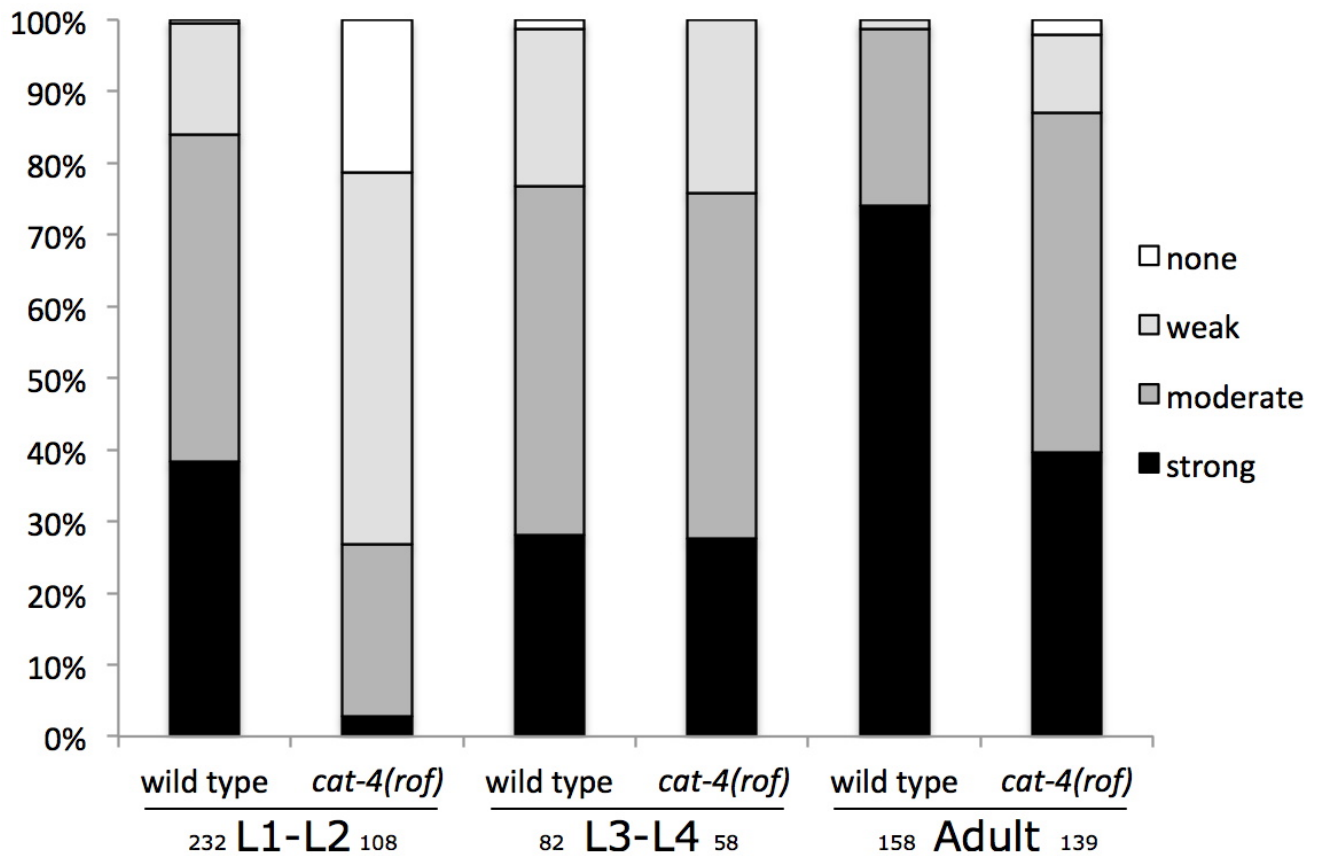


Figure S9 *cat-4* reduction-of-function mutants accumulate 5HT during larval development. Anti-5HT immunoreactivity differences in wild type and *cat-4(e3015)* reduction of function (*rof*) are most apparent in young larvae. 85% of wild-type worms are strongly to moderately stained whereas only 27% of *cat-4(e3015)* L1-L2 worms are strongly to moderately stained. Mutants in mixed populations of young (L1-L2), older larvae (L3-L4), and adults, scored for intensity of NSM cell body staining (as in Fig. 7A, B). Numbers of worms scored shown below the column.

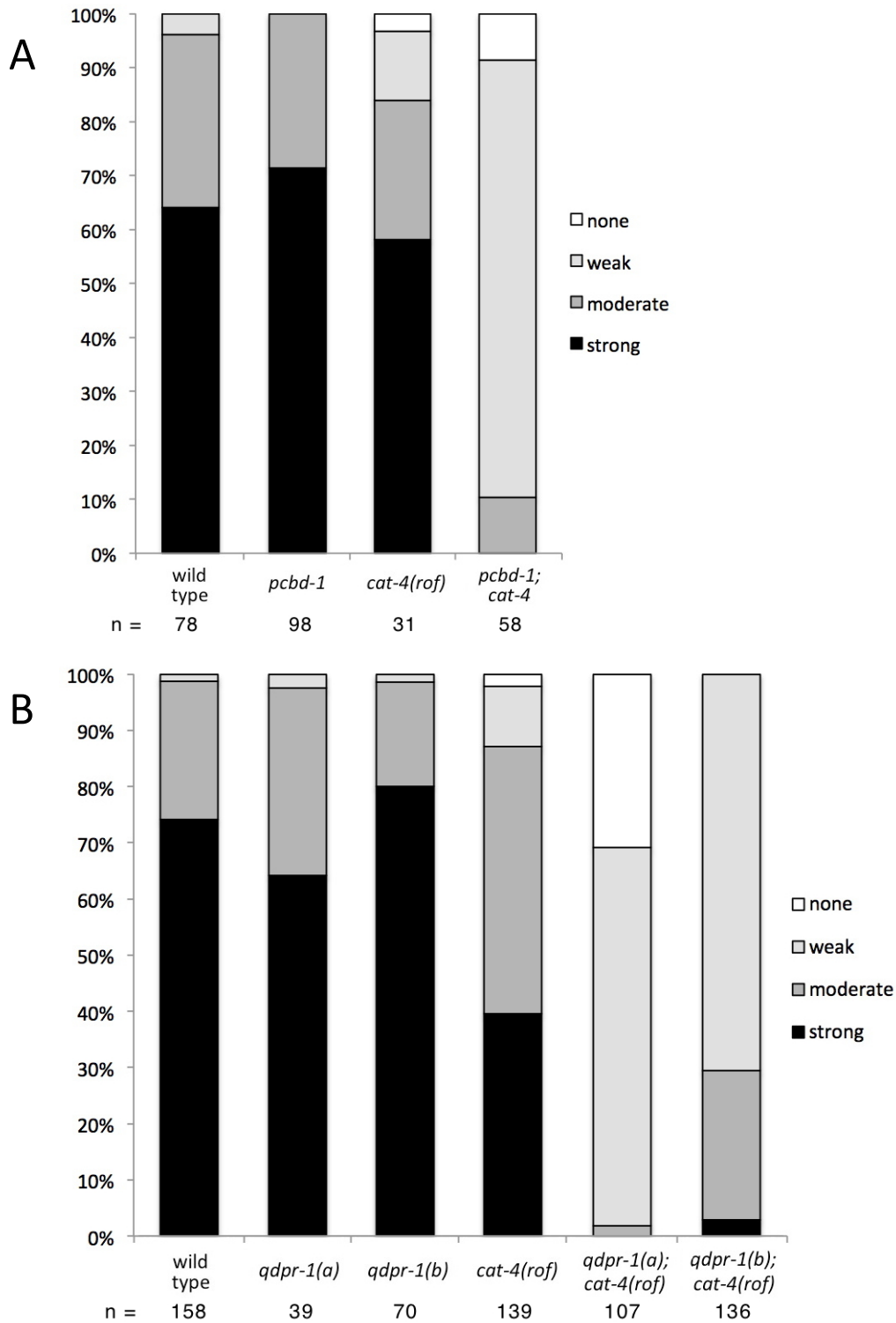


Figure S10 Bioppterin regeneration genes *pcbd-1* and *qdpr-1* function in 5HT synthesis in adult worms. Anti-5HT immunoreactivity differences in wild type, single mutant, and double mutants in adult worms (see legend for Fig 7A, B). Numbers of worm scored shown below the columns. (A) *pcbd-1* single and double mutants with *cat-4(e3015)*. (B) *qdpr-1* single and double mutants with *cat-4(e3015)*.

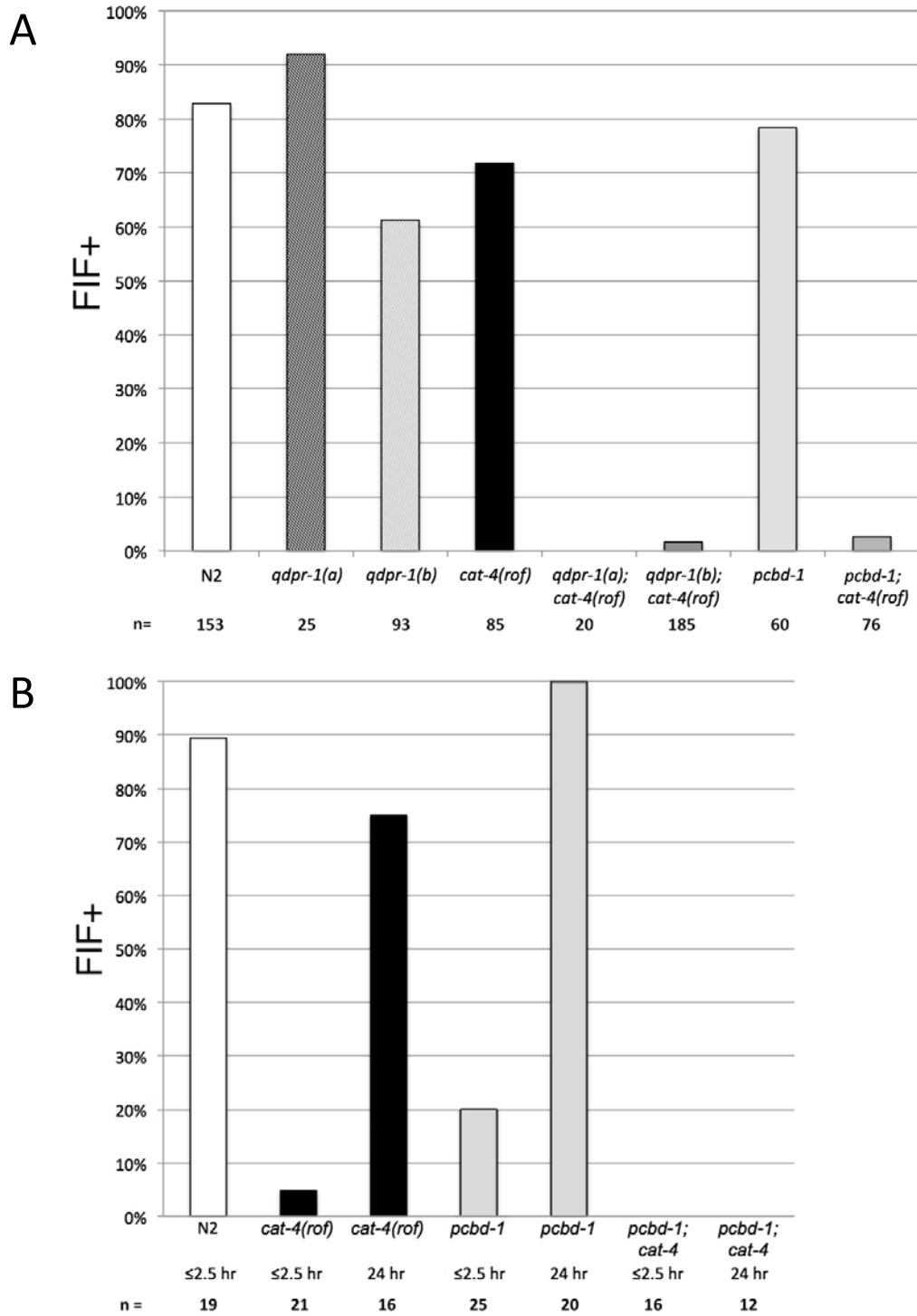


Figure S11 Bioprotein regeneration genes *pcb-1* and *qdpr-1* function in DA synthesis. (A) Mixed populations, young worms (L1-L2) scored for DA by formaldehyde induced fluorescence (FIF). Y axis – percentage of worms with one or more FIF+ head neurons (without regard to brightness). Alleles used: *cat-4(rof)* = 'reduction of function' = *e3015*; *qdpr-1(a)* = *tm2337*; *qdpr-1(b)* = *tm23373*; *pcb-1(tm5924)*. (B) Loss of DA in BH4 synthesis and regeneration mutants is most apparent in very young worms. Staged worms scored for DA by FIF – early L1s (≤ 2.5 hr post-hatching), mid-L2s (~ 24 post-hatching). Y axis and alleles used as in (A). Eggs were picked to a seeded plate and allowed to hatch for 2.5 hr, at which time hatchling worms were immediately subjected to FIF staining, or transferred to a new seeded plate, incubated at 20°, and tested by FIF 24 hr later.

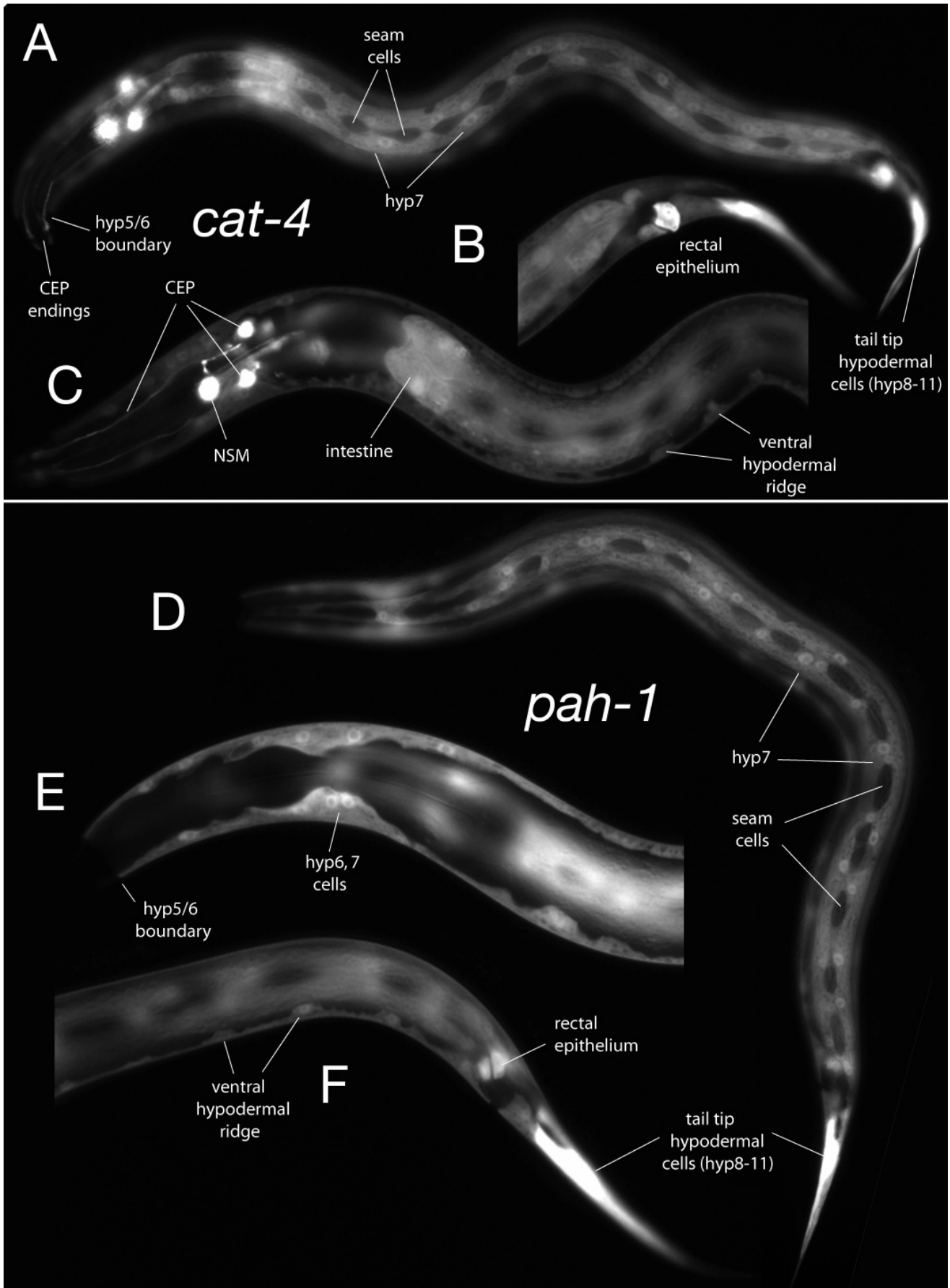


Figure S12 Expression of *cat-4* and *pah-1* GFP reporters. Anterior is to the left in all worms. (A-C) *cat-4*::GFP transgenic L2 stage worms, repeated from Fig 8A to facilitate comparison with *pah-1*::GFP expression (D-F). (A) Superficial focal plane showing epidermal expression, especially in hyp7 syncytium. Seam cells have undergone doubling division and can be seen along the lateral side as darker regions among the brightly staining hyp7 cell. Dendritic endings of CEP neurons can be seen at the tip of the 'nose.' (B) Medial focal plane showing anal cells, strongly expressing tail epidermal cells and posterior intestinal cells expressing GFP. (C) Medial focal plane showing epidermal expression in the body and head, and the boundary between hyp6 and unstained hyp5. NSM and CEP neuron somas are seen in the head, plus some neuronal processes (especially CEP processes). A few other neuronal somas stain less brightly. The anteriormost intestine also shows GFP expression, as do some rectal epithelial cells (here in what appear to be B & Y cells). (D-F) *pah-1*::GFP transgenic L2 stage worms. (D) Superficial focal plane showing especially hyp7 expression, with dark seam cells (post-doubling division) lying over hyp7. (E) Medial focal plane showing anterior epidermal expression. The hyp6 (expressing) / hyp5 (dark, not expressing) boundary is very clear here. (E) Medial focal plane in the posterior showing intestine, some rectal epithelial cells, and strongly expressing epidermal tail cells. (F) Medial focal plane showing ventral hypodermal ridge, rectal epithelium cells, and strongly expressing tail epidermal cells expressing GFP.

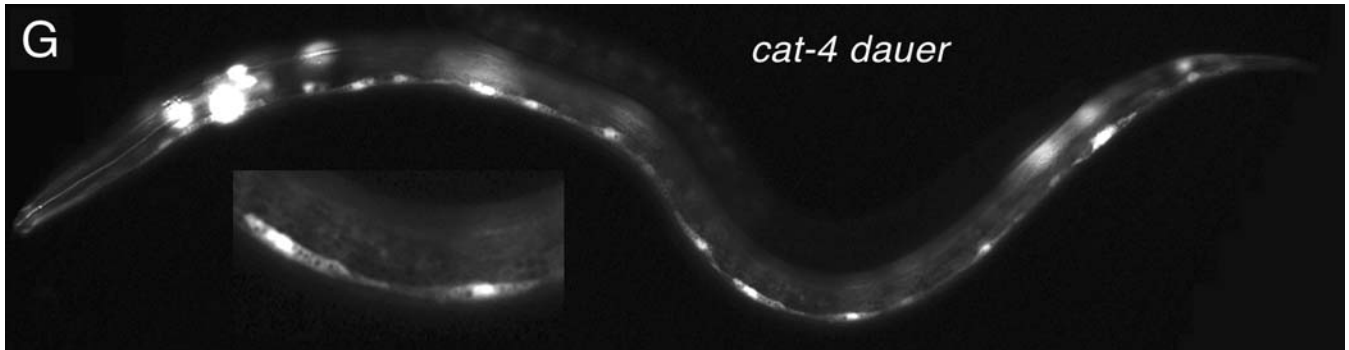


Figure S12 (G) *cat-4*::GFP worm showing expression in lateral epidermal seam cells during dauer stage. Anterior to the left. Inset: closeup of two seam cells in midbody region.

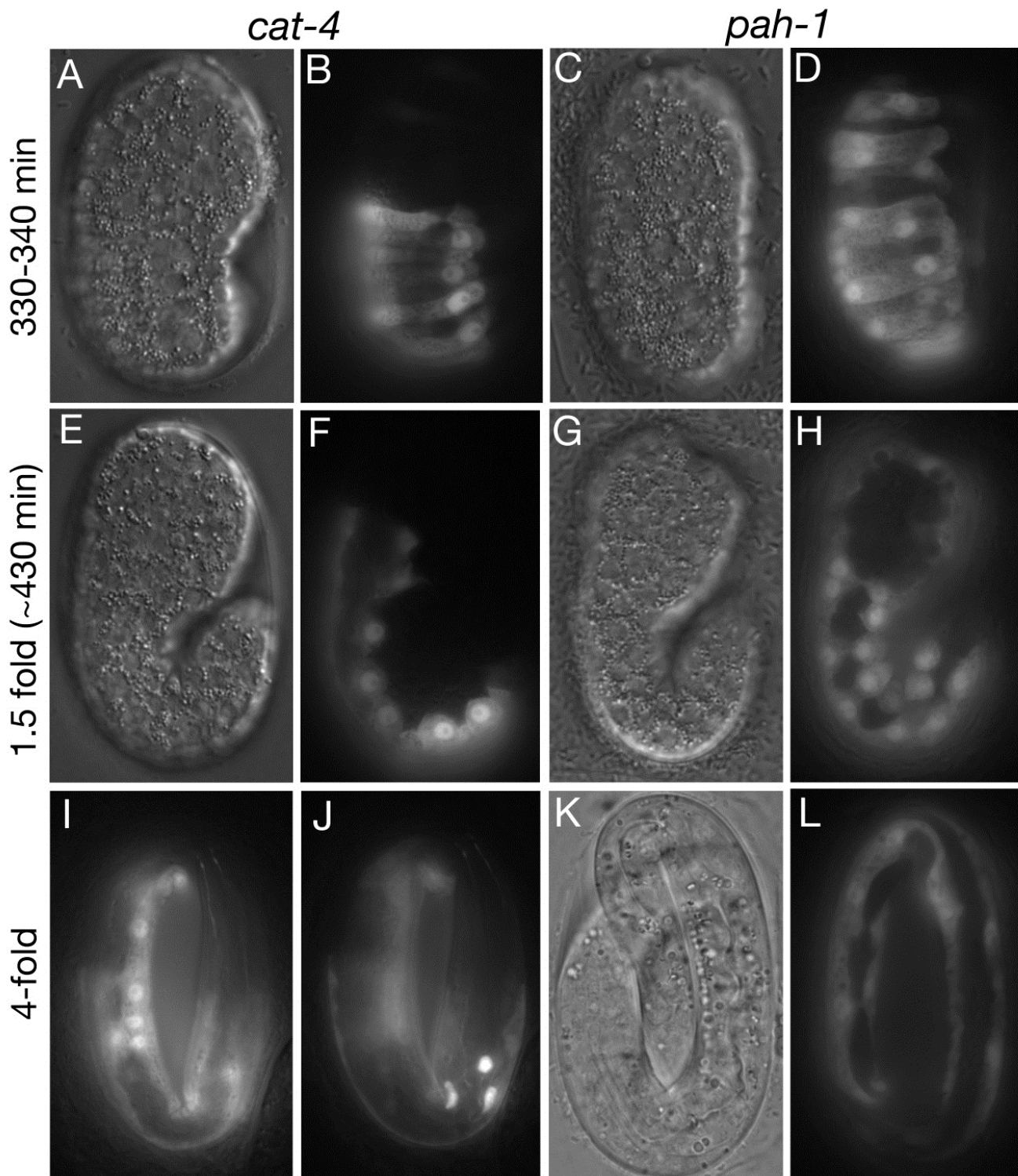


Figure S13 Expression of *cat-4*, *pah-1* and *pts-1* reporter constructs in embryos. Left hand columns (A, B, E, F, I, J) - *cat-4*::GFP embryos; Right hand columns (C, D, G, H, K, L) *pah-1*::GFP embryos. Pairs of photos (e.g., A+B) have DIC view of embryos (A) matched with GFP fluorescence (B) in same superficial focal plane. Top row (A-D): 330-340 min, post nuclear counter-migration. Embryos have just begun elongation, dorsal view, anterior up. (B) *cat-4*::GFP embryo shows expression in posterior dorsal epidermal cells. (D) *pah-1*::GFP is expressed in both anterior and posterior dorsal epidermal cells. Middle row (E-H). 1.5-fold stage, lateral view. (F) *cat-4*::GFP expression is seen in posterior dorsal epidermal cells. (H) *pah-1*::GFP is expressed in most dorsal and ventral epidermal cells, but not in lateral seam cells. Bottom row (I-L). Late (4-fold) embryos. (I) Superficial focal plane of anterior of worm showing *cat-4*::GFP epidermal staining (except in seam cells) similar to that seen in larvae. CEP neurons dendritic endings are seen in tip of nose. (J) Deeper focal plane of same worm showing NSM and CEP neurons expressing GFP. (K) DIC view of *pah-1*::GFP late embryo (4-fold). (L) Different, superficial focal plane of same worm.

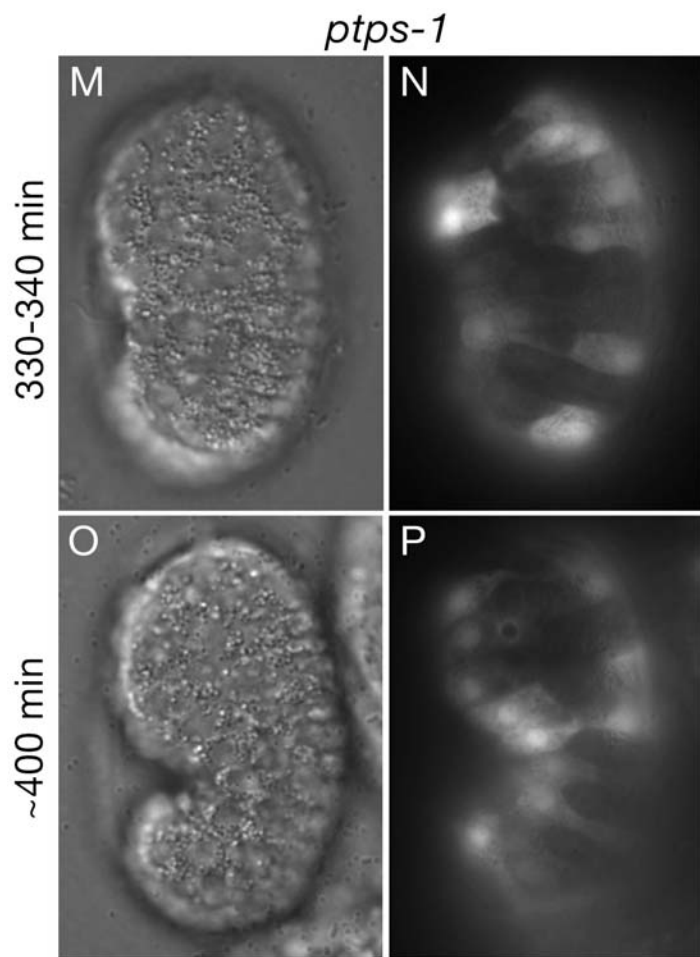


Figure S13 Expression of *cat-4*, *pah-1* and *ptps-1* reporter constructs in embryos. (M-P) *ptps-1*::GFP embryos (strain OH11619). Pairs of matching photos showing DIC view of embryo with GFP fluorescence at the indicated times. Expression seen in both anterior and posterior dorsal epidermal cells

File S1

Supporting Materials and Methods

Worms strains used in this study:

N2 wild type	CB1490 <i>him-5(e1490)V</i>
CB369 <i>unc-51(e369)</i>	CB1141 <i>cat-4(e1141)V</i>
HA1335 <i>cat-4(ok342)V</i>	LC81 <i>cat-4(tm773)V</i>
VC20144 <i>cat-4(gk245686)V</i>	CLB49 [line 1] <i>glsEx1 [rol-6(dom) cat-4(+)]; cat-4(tm773)V</i>
CLB49 [line 4] <i>glsEx3 [rol-6(dom) cat-4(+)]; cat-4(tm773)V</i>	CB7107 <i>him-8(e1489)IV; cat-4(e3015)V</i>
CB7130 <i>him-8(e1489)IV; cat-4(e3030)V</i>	LC80 <i>ptps-1(tm1984)I</i>
LC84 <i>ptps-1(tm1984)I; him-5(e1490)V</i>	CB7094 <i>ptps-1(e3042)I</i>
LC87 <i>qdpr-1(tm2337)III</i>	LC90 <i>qdpr-1(tm2373)III</i>
LC133 <i>pcbd-1(tm5924)I</i>	LC129 <i>qdpr-1(tm2337)III; cat-4(e3015)V</i>
LC130 <i>qdpr-1(tm2373)III; cat-4(e3015)V</i>	LC131 <i>pcbd-1(tm5924)I; cat-4(e3015)V</i>
LC83 <i>pah-1(tm520); bas-1(ad446)</i>	CB7014 <i>agmo-1(e3016)III</i>
CB7127 <i>agmo-1(e3019)III</i>	CB7128 <i>agmo-1(e3029)III</i>
CB7129 <i>agmo-1(e3047)III</i>	VC1198 <i>Y39G8B.1(ok1682)II</i>

See also transgenic reporter fusion strains listed in Tables S1, S2.

Table S1 Biopterin-related GFP reporter gene transgenics (this work) – primers and characteristics

Gene	Primers [†]	Sequence	Fusion type	Size (bp)**	Transgenics‡
<i>cat-4</i>	AC1763 (A)	AAAGGTTGCATGTTGCAGATG	Transcriptional, through ATG	2654	CZ9718, -19 (<i>juEx2054</i> , -5)
	AC1764 (A*)	TGCATGTTGCAGATGGAAATTG			
	AC1765 (B)	CATTTTGATATTATGATGTTGATAGA			
<i>ptps-1</i>	AC3051 (A)	ATAGTCCGGTCTTGTACCAC	Translational, in final coding exon	2751 (~2000)	CZ18320, -21, -22 (<i>juEx5474</i> , -5, -6)
	AC3050 (B)	ACATCCTTTATAAGTGAAAATATT			
	AC3049 (A) [§]	TCGAATTTTCGCGGACAAGG	Translational, in final coding exon	1160 (~450)	CZ18092, -93 (<i>juEx5477</i> , -8)
<i>pcbd-1</i>	AC3217 (A)	ATCAGAGTAGGAGTCAGGGAG	Transcriptional, upstream of ATG	1505	CZ19212 (<i>juEx5785</i>)
	AC3218 (B)	TGAATGAGAAGATGCGTTGAGAAG			
<i>qdpr-1</i>	AC3219 (A)	CGGTATCCTTGTGCCCCAAC	Transcriptional, upstream of ATG	1048	CZ19213, -14 (<i>juEx5786</i> , -7)
	AC3220 (B)	TGTTCGAAACTGCAAGGAAAAAGT			
<i>qdpr-1</i>	qdpr1AL1 (A)	TGGCGGAATCGATTTATTTG	Translational, in final coding exon	4887 (~4000)	CZ19215 (<i>juEx5788</i>)
	qdpr1AL2 (A*)	GCCATATTGCGTTCAATGAG			
	qdpr1BR (B)	TCGATGTTCCATTTTCAGTGG			
<i>grfp-1</i>	AC3221 (A)	AATACGGTGCCAGGTGTCAG	Transcriptional, upstream of ATG	1088	CZ19216 (<i>juEx5789</i>)
	AC3222 (B)	TTCTTGATTTTTTTTGTTCGGAAT			
<i>agmo-1</i>	AC3030 (A)	ACTTGCAAACAGTTGGAAGC	Transcriptional, upstream of ATG	2281	CZ17563, -64, -65 (<i>juEx5238</i> , -39, -40)
	AC3031 (B)	CCTCTTTTCATTTGGTTAAAATTT			
<i>pah-1</i>	AC1769 (A)	ATGGAAGGTCAGATTGGATATC	Transcriptional, through ATG of an isoform	2005	CZ9720, -21 <i>juEx2056</i> , -7
	AC1770 (A*)	TCAGATTGGATATCTTCCACG			
	AC1771 (B)	CATTTTCAGAGAACAAGATTTGGT			

† - Primer name ('style' of primer, ala Hobert, 2002); not shown above, all 'B' primers begin with:

AGTCGACCTGCAGGCATGCAAGCT (sequence overlapping GFP coding to create fusion)

§ - Primer paired with the same B (AC3050)

** - Length of sequence upstream to GFP (length upstream of predicted translation start if translational fusion)

‡ - Strain number (beginning with CZ) and allele designation of extrachromosomal transgenes (*juEx*)

Table S2 Biopterin-related GFP reporter gene transgenics from other sources

Gene	Fusion type	Size (bp)**	Transgenic	Reference
<i>cat-4</i>	translational, exon 2	~2800 (~2000)	JY739 [†]	Sze et al. 2002 [†]
<i>cat-4</i>	transcriptional	~630	OH8482	Flames and Hobert 2009
<i>ptps-1</i>	transcriptional	~2600	OH11619	Zhang et al. 2014
<i>pcbd-1</i>	transcriptional	~800	OH12385	Zhang et al. 2014
<i>qdpr-1</i>	transcriptional	~700	OH12386	Zhang et al. 2014
<i>gfrp-1</i>	translational, exon 2	~2500 (~1400)	OH12387	Zhang et al. 2014

** - length of sequence upstream to GFP (length upstream of predicted translation start if translational fusion).

† - this strain shows expression in biogenic amine neurons and the *epidermis*, *not in muscle cells* as reported.

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