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Odour receptors and neurons for detection of DEET and new insect repellents

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Summary

There are major impediments to finding improved DEET alternatives because the receptors causing olfactory repellency are unknown, and new chemistries require exorbitant costs to determine safety for human use. Here we identify DEET-sensitive neurons in a pit-like structure in the *Drosophila* antenna called the sacculus. They express a highly conserved receptor *Ir40a* and flies in which these neurons are silenced or *Ir40a* is knocked down lose avoidance to DEET. We use cheminformatics to screen >400,000 compounds and identify >100 natural compounds as candidate repellents. We test several and find that most activate *Ir40a*+ neurons and are repellents for *Drosophila*. These compounds are strong repellents in mosquitoes as well. The candidates contain chemicals that do not dissolve plastic, are affordable, smell mildly like grapes, with three being considered safe for human consumption. Our findings pave the way to discover new generations of repellents that will help fight deadly insect-borne diseases worldwide.

Blood-feeding insects transmit deadly diseases such as malaria, dengue, lymphatic filariasis, and West Nile fever, to hundreds of millions of people, causing untold suffering and more than a million deaths every year. Insect repellents can be very effective in reducing disease transmission by blocking contact between blood-seeking insects and humans.

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Author Contributions S.M.B planned and performed the chemical informatics and solubility experiment, and helped design the behaviour experiments. P.K. planned and performed the NFAT imaging, Ca^{2+} imaging and *Drosophila* behaviour experiments. S.K.T. performed Ca^{2+} imaging, electrophysiology and some behaviour experiments. T.G. performed the arm-in-cage experiments. C.P. performed behaviour analysis. S.M.B, P.K. and S.K.T helped prepare drafts of the manuscript and figures. A.D. planned experiments and helped write the manuscript. A.R. planned experiments, managed the project, and wrote the manuscript.

Author Information P.K., S.M.B., C.P. and A.R. are listed as inventors in pending patent applications filed by the University of California Riverside. Readers are welcome to comment on the online version of this article at http://www.nature.com/nature.

N,*N*-Diethyl-m-toluamide (DEET) has remained the primary insect repellent used for more than 60 years. However, DEET has little impact on disease control in endemic regions due to high costs and inconvenience of continuous application on skin at high concentrations. DEET also dissolves some plastics, synthetic fabrics, and painted surfaces[1]. Additionally, DEET inhibits mammalian acetylcholinesterase[2]. Instances of DEET resistance have also been reported in flies[3], and mosquitoes[4,5]. However, major barriers in developing improved repellents are the estimated cost for identification[6] and subsequent cost of safety analyses for new chemistries.

A significant challenge in finding improved DEET substitutes is that the target receptors through which it causes repellency in insects are unknown. Recent studies have put forth multiple models of DEET action. Pure DEET causes inhibition[7,8] or mild electrophysiological modification of neural responses to weakly-activating odours in *Drosophila* antennal olfactory neurons[9], but whether these effects contribute to repellency is unknown. Mosquitoes can also directly detect DEET[10] and mutations in the *Orco* co-receptor gene in *A. aegypti* cause reduction in repellency[11]. Some DEET-sensitive olfactory neurons have been identified in *C. quinquefasciatus*[10] and *A. aegypti*[5], but it is not yet known whether they are responsible for repellency or which odour receptors they express. A broadly tuned larval odour receptor responds to DEET[12,13], however its role in avoidance in larval or adult mosquitoes has not been demonstrated. Not only can more than one pathway contribute to olfactory repellency, analyses are further confounded by the observation that DEET also activates bitter taste neurons that mediate contact-avoidance in *Drosophila*[14,15].

DEET detected by neurons of the sacculus

In order to identify the elusive DEET-sensing neurons of the olfactory system in an unbiased manner, we used the NFAT-based system to report DEET-evoked neural activity via expression of green fluorescent protein (GFP) in *Drosophila melanogaster*[16] (Fig. 1a). Exposure to 10% DEET showed an increase in expression of GFP in neurons that innervate sensilla within the sacculus, a pit-like structure in the antenna (Fig. 1b, c, Supplementary Fig.1a, Supplementary Video 1). The dendrites of GFP+ neurons primarily innervated the most distal chamber (I) of the sacculus (Fig. 1c, Supplementary Fig. 1b). Previous studies of DEET overlooked the sacculus, since it is intractable to traditional electrophysiology methods.

Contrary to expectations from a previous report[17] we were unable to find DEET-activated reporter expression in ORNs of the maxillary palps (Fig. 1b). We therefore performed single-sensillum electrophysiology analyses and found that the previously reported *Or42a*+ pb1A neurons responded poorly to DEET, but strongly to hexane that was used as solvent in the previous study (Supplementary Fig. 2a, b).

ORNs innervating the sacculus do not express *Or* genes, but instead members of the ancient *Ir* (*Ionotropic Receptor*) gene family[18-21]. In the antennal lobes robust DEET-dependent GFP was detected in the characteristic “column” glomerulus (Fig.1d, Supplementary Fig. 3a), which is innervated by axons of *Ir40a*-expressing neurons of the sacculus[18]. Faint GFP was also observed in the *Or67d*+ DA1 glomerulus, which is likely caused by exposure to

male pheromone CVA in the assay, since the CVA-responsive at1 neuron did not respond to DEET (Supplementary Fig. 2c). The DC4 glomerulus, which is innervated by other sacculus ORNs that express *Ir64a*[19], showed a very faint signal as well (Supplementary Fig. 3a). The simplest interpretation of these results is that *Ir40a+* sacculus ORNs innervating chamber I, and projecting to the column glomerulus, may represent a major olfactory detection pathway for DEET.

Consistent with previous electrophysiological analysis[14,15], we found DEET-dependent GFP expression in gustatory neurons of the labellum (Fig. 1e). In addition we observed DEET-dependent GFP in neurons innervating the Labral Sense Organ (LSO) of the pharynx (Fig. 1e). The DEET activity mapped to neurons marked by *Gr33a* and *Gr89a*, which are bitter-sensing deterrent neurons (Supplementary Fig. 3b). Axonal projections of DEET-sensitive gustatory neurons in the sub-oesophageal ganglion (SOG) revealed arborization patterns similar to those of taste neurons originating in the labellum and the pharynx (Fig. 1e, Supplementary Fig. 3b).

In order to directly test physiological responses of the sacculus *Ir40a+* ORNs to DEET we performed *in vivo* calcium imaging in flies expressing GCaMP3 using *Ir40a-Gal4*[18,22,23]. *Ir40a* neurons show robust activation in response to a puff of DEET delivered from an atomizer but not to control DMSO (Fig. 2a,b). Moreover, DEET response is dependent on *Ir40a* (Fig. 2c).

In order to test whether the *Ir40a+* ORNs are required for DEET repellency we blocked synaptic transmission in these neurons using *Ir40a-Gal4* to express the active form of tetanus toxin (TNTG)[24]. We employed a trap lured by 10% apple cider vinegar (ACV) in which a DEET-treated filter paper was placed inside the trap. Avoidance was significantly decreased in *Ir40a-TNTG* flies as compared to various controls, including a non-functional version of the tetanus toxin (IMPTV), suggesting that *Ir40a+* neurons are required for DEET repellency (Fig. 2d). All genotypes exhibited attraction to 10% ACV in 2-choice trap assays (Supplementary Fig. 4a).

Ir40a is necessary for DEET avoidance

To test directly whether *Ir40a* is required for olfactory avoidance to DEET we examined the behaviour of flies in which *Ir40a* was knocked down pan-neuronally using an *elav-Gal4* driver to express *UAS-Ir40a-RNAi*. In 2-choice trap assays (Fig. 3a), we found a significant loss of DEET avoidance in *Ir40a-RNAi* flies as compared to control flies (Fig. 3b). Similar results were obtained when *Ir40a-RNAi* was executed selectively in *Ir40a+* ORNs using two independent *UAS-Ir40a-RNAi* transgenes (Fig. 3c). Not only was avoidance completely abolished, *Ir40a* knockdown flies in fact showed a mild attraction to the DEET trap. Attraction to ACV was unaffected (Supplementary Fig. 4b,c).

We next wanted to rule out the possibility of a developmental role for *Ir40a*. We therefore suppressed expression of *Ir40a-RNAi* during development using a temperature-sensitive $Gal80^{ts}$ transgene (Fig. 3d). Flies were raised at the permissive temperature (18°C) until just before adult eclosion, at which point they were left at 18°C (RNAi Off) or shifted to the $Gal80^{ts}$ restrictive temperature 29°C (RNAi On). Behavioural assays performed four days

after the temperature shift showed that post-developmental *Ir40a-RNAi* was sufficient to abolish DEET avoidance when RNAi was induced in *Ir40a+* ORNs (Knockdown, Fig. 3e). Moreover, DEET avoidance was completely restored when flies were returned to the $Gal80^{ts}$ permissive temperature (Recovery, Fig. 3e). Attraction to ACV was unaffected (Supplementary Fig. 4d). Taken together, these experiments demonstrate that *Ir40a* is required in adult *Ir40a+* sacculus ORNs for olfactory avoidance of DEET.

In silico prediction of new repellents

Identification of DEET receptors and neurons offer a powerful system to screen for improved repellents. However, volatile chemical space that can be exploited to find DEET substitutes is vast and therefore poses unfeasible requirements in terms of cost and time to screen. The receptor structure is unavailable for screening and the most effective repellents may require detection by both olfactory and gustatory pathways. To circumvent these limitations we developed a high-throughput chemical informatics screen. Previous studies using such structure-activity approaches have given encouraging results[25].

We identified structural features shared by DEET and other known repellents and used them to screen a vast library of compounds *in silico* for the presence of these features. We assembled a training set of known repellents that included: the two commercially approved repellents DEET and picaridin; 34 N-acyl piperidines[25] that were identified by structural relatedness to picaridin; natural repellents eucalyptol, linalool, alpha-thujone, and beta-thujone[10,26,27]; and a structurally diverse panel of other odours as negatives[28,29]. We focused on a descriptor-based computational approach and using a Sequential-Forward-Selection method[30] we incrementally identified a unique subset of 18 descriptors that were highly correlated with repellency (correlation of 0.912) (Fig. 4a, Supplementary Table 1). The repellents clustered together if the optimized descriptor subset was used to calculate Euclidean distances amongst odorants of the training set (Fig. 4b).

The optimized descriptor set was utilized to train a Support Vector Machine (SVM), which is a well-known supervised learning approach[31], to predict compounds that shared optimized structural features with known repellents (Fig. 4a). A 5-fold cross-validation on the training set of repellents was performed and a mean Receiver-Operating-Characteristic (ROC) analysis curve generated. The Area-Under-Curve was determined to be high (0.994) indicating that the *in-silico* approach was extremely effective at predicting repellents from compounds that were excluded from the training set (Fig. 4c).

We next used the 18-optimized-descriptor and SVM method to screen *in silico* a large virtual chemical library consisting of >440,000 volatile-like chemicals. Inspection of the top 1,000 predicted repellents (0.23% of hits) revealed a diverse group of chemicals that retain some structural features of the known repellents (Fig. 4d,e). We computed partition coefficient (logP) values of the 1,000 compounds to exclude those predicted to be lipophilic (logP >4.5) and therefore more likely to pass through the skin barrier in topical applications[32] (Fig. 4e). In addition, we computed predicted vapour pressures of these chemicals, since volatility may be a useful predictor of spatial volume of repellency (Fig. 4e).

Although the *in-silico* screen was feasible, a more significant challenge lies in identifying safe and effective DEET substitutes that can be rapidly approved for human use. To identify such compounds, we applied our *in-silico* screen to an assembled natural odour library consisting of >3,000 chemicals identified as originating from plants, insects, or vertebrate species, and compounds already approved for human use as fragrances, cosmetics, or flavours (Supplemental Materials). While many of the top 200 hits share structural features with known repellents from the training set, they also represent structurally diverse chemicals, allowing targeted exploration of previously untested chemical space (Fig. 4f). For example, several anthranilates and pyrazines were identified, although such compounds were largely missing from the training set.

Ir40a+ cells activated by repellents

We selected 4 compounds from the list, methyl N,N-dimethyl anthranilate (MDA), ethyl anthranilate (EA), butyl anthranilate (BA), and 2,3-dimethyl-5-isobutyl pyrazine (DIP), of which the first 3 have a mild grape-like aroma, have excellent safety profiles, have been thoroughly tested and approved for human consumption/oral inhalation by the FDA, World Health Organization and European Food Safety Authority, and have been listed in the "generally recognized as safe" (GRAS) list by the Flavour and Extract Manufacturer's Association (Fig. 4g, Supplementary Table 2). The fourth, a pyrazine, is an ant trail pheromone[33]. The anthranilate and pyrazine classes also contain a large diversity of chemicals found in nature and therefore present attractive repositories of structural substitutes.

For all 4 chemicals we found robust activation of sacculus ORNs (Fig. 5a, Supplementary Video 2) that innervate the *Ir40a+* "column" glomerulus (Fig. 5b, as shown for BA). They also activated gustatory neurons that project to similar areas of the SOG as DEET (Fig. 5b, as shown for BA). GCaMP3 imaging in *Ir40a+* neurons showed robust responses to these chemicals, while several other classes of common odorants did not (Fig. 5c, Supplementary Fig. 5). These results demonstrate that the computationally predicted chemicals activate the same chemosensory pathways as DEET and are therefore ideal candidates for new repellents.

In order to test the effect of these compounds on behaviour we used a 2-choice trap assay in which flies can sense a DEET-treated filter paper positioned at the entrance of a trap via both olfactory and gustatory systems[3,17] (Fig. 5d). All 4 compounds had strong dose-dependent repellent effects on *D. melanogaster* (Fig. 5d). Measurements were taken at 24 hours and 48 hours after the start of the assay, and were found to be consistent. Six additional predicted repellents were tested in a similar manner, at least four of which elicited strong repellency similar to DEET (Supplementary Fig. 6).

To confirm the role of *Ir40a+* neurons in mediating avoidance to these new repellents, we examined behavioural avoidance of flies in which synaptic activity of *Ir40a+* neurons was silenced using TNTG as before. We found that avoidance of chemical treated traps was substantially decreased in *Ir40a-TNTG* flies as compared to control flies (Fig. 5d), showing that *Ir40a+* neurons are required for repellency to the four chemicals.

Mosquitoes avoid predicted repellents

To test the effects of the identified chemicals on mosquito behaviour, we adapted a hand-in-glove assay that allows quantitative analysis of chemical repellency on mosquitoes attracted to a human arm (described in Supplementary Methods) (Fig. 6a, Supplementary Fig. 7). Female *A. aegypti* mosquitoes showed strong avoidance behaviour to DEET, irrespective of whether or not they could directly contact DEET (Fig. 6b). However, for sporadic landings the average time spent on the net before escape while not significant (p=0.203 for 10% DEET and p=0.06 for 1% DEET, Student's *t*-test) was reduced when direct contact with DEET was permitted, particularly at the lower concentrations (Fig. 6c). While it is difficult to asses from these experiments the direct contribution of the gustatory system alone, it demonstrates that mosquitoes can avoid DEET strongly at close range, even without making direct contact with it.

In order to test whether the 4 newly identified *Drosophila* repellents were also olfactory repellents to mosquitoes, we performed behaviour trials using the non-contact version of the assay. Notably, we found that all 4 compounds applied at 10% concentration demonstrated substantial repellency (Fig. 6d). The fraction of mosquitoes present on the net throughout the duration of the assay (Fig. 6d), as well as the cumulative number of mosquitoes present on the net were substantially decreased in the presence of the test compounds (Fig. 6e). For the mosquitoes that did land on the repellent treatment, the escape index, as measured by the frequency of take off, was substantially higher as compared to those landing on controls (Supplementary Fig. 8,9).

One of the major disadvantages of DEET is its property of solubilizing plastics and synthetic materials[1], which impacts its usefulness. We tested the ability of the 4 repellents to dissolve a 3 × 3 mm square of vinyl. While the vinyl completely disappeared in DEET within 6 hrs, there was no significant difference in the weight of the vinyl squares immersed in the 4 DEET substitutes after 6-hrs or 30-hrs (Fig. 6f).

Discussion

The unbiased strategy to use a genetic-reporter of neural activity was instrumental in identifying DEET-sensitive *Ir40a*+ neurons. These reside in the pit-like sacculus that could protect neurons from harsh chemicals. Both olfactory and gustatory systems are activated by DEET, with additional modes of detection in the antenna being mediated by *Orco*[11] and a yet to be identified tuning *Or* gene (Fig. 6h). Additionally, DEET has been reported to have a mild enhancing or suppressing effect on the activity of various *Or*-expressing neurons of antennal basiconics in *Drosophila*, although a causal relationship between this effect and repellency has not been established[9]. DEET also has a solvent effect that slows down volatile odour release, potentially also from skin[10]. Thus, multiple pathways and mechanisms are likely to participate in overall repellency.

Ir40a can account for the widespread effect of DEET olfactory repellency since it is highly conserved in species that show strong avoidance to it including *Drosophila*, mosquitoes, head lice[34], and tribolium[35], but not in the honey bee[36]. Ir40a orthologs are conserved across

multiple insects and arthropods, with several regions of amino acid similarity across the length of the protein (Supplementary Fig. 10). This degree of conservation may better explain the repellent effects of DEET across several arthropod species compared to *Or* pathways that are not as well conserved. The *Ir40a* pathway therefore has major implications in the development of safe and affordable strategies to control several types of insects and arthropods that vector diseases of animals and plants or are plant pests.

The chemical informatics enabled us to identify a number of affordable and safe potential repellents that are good candidates for regulatory approval for human use (Fig. 6g). This screen identified ∼1000 compounds and >100 additional natural compounds, many approved for use in human food and cosmetics, which may lead to other effective repellents. The repellency strategy may also have promise for use in combination with other behaviour control strategies, such as masking of CO_2-mediated attraction behaviour or population control by trapping as a part of an integrated pull-mask-push strategy[37,38]. Moreover, these DEET substitutes may be of value in controlling DEET-resistant strains as well. Since several of the new repellents are affordable, activate both the olfactory and bitter gustatory neurons, are approved for human consumption and are strong repellents for fruit flies, they may also have major implications for control of agricultural pest insects that cause enormous crop loss. Novel repellents that are safe and affordable can be used to limit insect-human contact in disease-endemic areas of the world and to provide an important line of defence against deadly vector-borne diseases.

Methods Summary

Physiological experiments

NFAT-based neural tracing[16] and GCaMP3-based Calcium imaging[22,23] were performed as previously described with some modifications (see Online Methods). Single-unit recordings from olfactory sensilla were performed as described previously[37].

Behavioural experiments

For olfactory trap assays 20 *Drosophila* were released in cylindrical arenas containing Eppendorf tube traps (Fig. 2d and Fig.3a) with 10% apple cider vinegar as a lure. Repellents were presented on filter papers placed near the trap openings in a manner that did not allow physical contact with the fly prior to its entering the trap. Trap assays to measure repellency when both olfactory and gustatory inputs were possible were performed as described previously[3]. Mosquito arm-in-cage avoidance assays were performed with 40 mated *A. aegypti* females held in a cage and presented a human arm that was inserted in a glove containing a window covered with a double-layer of netting. Test compounds were applied to the nettings. Attraction towards the arm was measured using video recordings and analysts were blind to treatments.

Chemical informatics

Optimized molecular descriptors were selected from 3,224 Dragon descriptors based on their ability to increase the correlation between descriptor values and repellency. The

repellency-optimized descriptor set was utilized to first train a Support Vector Machine to predict repellents and then applied to predict new repellents from large compound libraries.

Insects

Fly lines were obtained from the Bloomington *Drosophila* Stock Center for TNT and GCaMP3 experiments, the Vienna *Drosophila* RNAi Center for *Ir40a-RNAi*, Jing Wang (UC San Diego, CA) for NFAT tracing, and Richard Benton (University of Lausanne, Switzerland) for *Ir40a*-Gal4. Flies were grown on standard cornmeal-dextrose media, at 25°C unless otherwise noted and mosquitoes at 27°C and 70% RH.

Full Methods and associated references are available in the online version of the paper in Supplementary Information.

Methods

Fly stocks

Wild type flies were w^{1118} backcrossed to Canton-S for 5 generations. *UAS-GCaMP3* (BL#32236), *UAS-TNTG* (BL#28838), *UAS-IMPTV* (BL#28840) and *Tub-PGal80*^{ts} (BL#7017) were obtained from the Bloomington *Drosophila* Stock Center. The following stocks were generously provided: *LexAop-CD8-GFP-2A-CD8-GFP; UAS-mLexA-VP16-NFAT, LexAop-CD2-GFP* by Jing Wang (UC San Diego, CA), *Ir40a-Gal4* by Richard Benton (University of Lausanne, Switzerland), and *elav-Gal4* by Liqun Luo (Stanford, CA). *UAS-Ir40a RNAi* (1) (v101725) and *UAS-Ir40a RNAi* (2) (v3960) RNAi lines were obtained from the Vienna *Drosophila* RNAi Center. *Ir40a RNAi* is predicted to have no off-targets. Fly stocks were grown on standard cornmeal-dextrose media, at 25°C unless otherwise noted. Flies of appropriate genotypes for behaviour experiments were randomly sorted from populations before performing behavioural or electrophysiological experiments.

NFAT Based neural tracing

Late dark *Drosophila* pupae ready to emerge (~95-97 hrs) of genotype *elav-Gal4/LexAop-CD8-GFP-2A-CD8- GFP; UAS-mLexA-VP16- NFAT, LexAop-CD2-GFP/+* [16] were collected on moist filter paper strips in culture vials which contained 2 Kimwipes soaked in 5ml of water in as much of an odour free environment. A 100μl of odour at indicated concentration, dissolved in acetone, was spread on a filter strip (~1cm × 3 cm), dried for 1 minute and placed, in a vial with 10-15 pupae. The exposure was given for 24 hrs and the filter paper strip with odour was replaced at ~12-14hrs with fresh odour.

Calcium Imaging using GCaMP3

DEET, dimethyl sulfoxide (DMSO), hexane and candidate compounds were purchased from Sigma-Aldrich or the emolecules database (http://www.emolecules.com) from Enamine, Vitas M Labs or Chembridge and were of highest purity available. Approximately 10-12 days old flies raised at 29°C (to improve Gal4 expression) were anesthetized and secured by their wings on the double-sided sticky tape (ventral side up) on a petri dish (BD Falcon, 50 × 9 mm). The fly proboscis, head and body was immobilized by sticky tape as shown (Fig. S11). One antenna was stably held down using a glass electrode on thin layer of 70%

glycerol that enhanced imaging of fluorescence. The antenna was orientated with the arista and sacculus pointing upwards accessible to odours. Odorants were delivered using 5ml plastic syringes containing 2 Whatman filter paper strips (2×3 cm). A fine mist of DEET at indicated concentrations in DMSO was sprayed into the syringe using an atomizer. Fresh atomized odour syringes were prepared immediately before odour delivery. For DEET substitutes (BA, EA, MDA and DIP) a 100μl of 50% dilution in DMSO was applied to the filter paper directly and for other odorants a 100μl of (10^{-2}) solution in paraffin or water for apple cider vinegar (ACV) was applied directly on the filter paper. The odour puff (~ 2 sec) was delivered using the syringe over the antenna manually. For imaging odour-evoked activity from the antenna using GCaMP3 a Leica SP5 inverted confocal microscope was utilized. A filter block with 488 nm excitation filter and 500-535 nm emission filter was used and images were acquired at 3.3 frames per second with a resolution of 330×330 pixels using a 10× objective. The settings were optimized to capture odour-induced responses of GCaMP3 with high spatial and temporal resolution while limiting reporter bleaching.

Data analysis for calcium imaging was performed using the Leica SP5 LAS AF software (in Quantify mode) to obtain the heat map images and fluorescence intensity changes. The % ΔF/F was calculated separately for each selected cell body by taking the mean intensity value of all frames for 5 seconds prior to the odour puff (F_{pre}) and taking the mean intensity value of all frames for 5 seconds around the peak responses (F_{post}) after the end of the ~2 second of stimulus delivery period. Similarly, the mean intensity values were taken for a background area in the vicinity of the cells.

% ΔF/F was calculated according to the formula below:

$$\%\Delta \mathbf{F}/\mathbf{F} = \frac{[\mathbf{F}_{\text{post}} - \mathbf{F}_{\text{background(post)}}] - [\mathbf{F}_{\text{pre}} - \mathbf{F}_{\text{background(pre)}}]}{[\mathbf{F}_{\text{pre}} - \mathbf{F}_{\text{background(pre)}}]} \times 100$$

Immunohistochemistry

After 24hrs exposure to either odour or solvent (control), flies were anaesthetized on ice and the tissue dissected in chilled 1XPBS and fixed for 30 minutes in 4% PFA (0.3% Triton X-100) at room temperature. After washes with PBST (PBS with 0.3% Triton X-100) brains were blocked using PBST with 5% bovine serum albumin (BSA). Rabbit anti-GFP (1:1000, invitrogen) and anti-nc82 (1:10 Developmental Studies Hybridoma Bank) were used as primary antibodies and samples were incubated for 3 nights at 4 degrees. Alexa Fluor 488 anti-rabbit immunoglobulin G (IgG) (Invitrogen; 1:200) and Alexa Flour 546 anti-mouse IgG (Invitrogen; 1:200) were used as secondary antibodies, respectively followed by over night incubation at 4 degrees. Images were acquired with a Zeiss or Leica SP5 confocal microscope and images processing was done using ImageJ and Photoshop software. Data analysis was performed offline, and the investigator was blind to the treatment while counting GFP+ antennal neurons in the confocal micrographs.

Temperature sensitive Gal80^{ts} experiment

For the two-choice behaviour assay in Figure 3 and supplementary Figure S4, flies (10 males and 10 females) with genotypes *Ir40a-Gal4/+; UAS-Ir40aRNAi(2)/Gal80^{ts}* flies were grown throughout at 18°C (permissive temperature) where Gal80 is active and RNAi is off. Such flies were treated as control. In parallel, flies of the same genotype were shifted to 29°C (non-permissive temperature) from 18°C as late black pupae for 4 days to activate Gal4 and switch on RNAi. These flies were used as knockdown flies. A subset of flies that were shifted to 29°C was shifted back to 18°C for 4 additional days to turn off the RNAi and these were used as recovery flies.

Electrophysiology

Flies used were 4-7 days old and raised on cornmeal food at 25°C. Extracellular recording were made by inserting a glass electrode into the base of a palp sensillum as done previously[37,39]. Odorants were diluted in hexane or DMSO, at indicated concentrations (made fresh for every stimulus). For DEET stimulation, 10 μl of diluted odorant was applied to a filter paper strip, the hexane solvent was evaporated for 30 seconds (as in a previous study[17]) or for 5 minutes, and placed into a glass pasture pipette cartridge, and each cartridge was only used once. The evaporation of hexane from filter paper strip was much slower upon mixing with DEET and lingering dampness of the filter paper could be observed visually as well.

Behaviour testing

***Drosophila* olfactory avoidance assay for DEET—**For each trial flies that were to 3-6 day old flies (10 males and 10 females) were starved for 18 hours.

Trap Assay: Flies were transferred to a cylindrical 38.1 mm D × 84.1 mm H chamber containing a trap fashioned from an upturned 1.5 ml microcentrifuge tube with 2 mm removed from the tapered end. A pipette tip (1000 μl) was cut 2.5 cm from narrow end and 0.5 cm from top and inserted into the bottom of the inverted microcentrifuge tube. A 15mm × 16 mm #1 Whatmann filter paper was inserted in between the pipette tip and tip of microcentrifuge tube in a manner that entering flies cannot make physical contact with it. A 25 μl sample of test compound was applied to filter paper and 125 μl of 10% ACV is applied to the upturned lid of the microcentrifuge tube as attractant. Trials were run for 24 hours, and numbers of flies entering trap counted (Fig. 2d).

2-choice trap assay: For 2-choice test two 10% ACV (125 μl) lured traps as described above were placed in the cylinder, one with 50 μl solvent (DMSO) and another with 50 μl the test odorant at 50% applied to the filter paper (Fig. 3). The more volatile DIP was tested at a lower concentration of 25%. For positive control tests in Supplementary Fig. 4, 125 μl of 10% ACV in test traps and 125 μl of water in control traps is added in the upturned microcentrifuge tube lid. Both traps contained filter papers as before with 50 μl solvent (DMSO). All trials were run for 24 hours, positions randomized, and counted. Only trials with >35% participation was considered.

Preference Index = (number of flies in treated trap-number in control trap)/(number of flies in treated + control traps).

***Drosophila* olfactory and gustatory avoidance assay for DEET—**Repellency was tested in Fig. 5d and Supplementary Fig. 6 using a *Drosophila melanogaster* 2-choice trap assay as described previously[3,17] with minor modifications. Briefly, traps were made with two 1.5 ml microcentrifuge tubes (USA Scientific) and 200 µl pipette tips (USA Scientific), each cap contained standard cornmeal medium. T-shape piece of filter paper (Whatman #1) was impregnated with 5 ul of acetone (control) or 5 µl of 10%, 1%, 0.10% test odour, diluted in acetone. Traps were placed within a petri dish (100 × 15mm, Fisher) containing 10ml of 1% agarose to provide moisture. Ten wild-type Canton-S flies 4-7days old were used per trial, which lasted 48 hours by which time point nearly all flies in the assays had made a choice. For the 24 hour time point data was considered only if >35% of flies had made a choice, at 48 hours the majority of flies had made choices.

Preference Index= (number of flies in treated trap-number in control trap)/(number of flies in treated + control traps).

Mosquito arm-in-cage avoidance assay for DEET—Repellency was tested in mated and starved *A. aegypti* females using an arm-in-cage assay. *A. aegypti* mosquitoes (eggs obtained from Benzon Research Inc.) were maintained at ∼27 °C and 70% RH on 14h: 10h L: D cycle. Behavioural tests were done with 40 mated, non-blood fed, ∼24 hour starved, 4-10 day old females in 30cm × 30cm × 30cm cages with a glass top to allow for video recording (Fig. 6a, Supplementary Fig. 7). The experimental protocol was reviewed and approved by the Institutional Review Board (IRB) Compliance Analyst at UCR and determined not to require additional Human Research Review Board approval. Each test compound solution (500µl) of 10% concentration in acetone solvent was applied evenly to a white rectangular 7cm × 6cm polyester netting (mesh size 26 × 22 holes per square inch) in a glass petri-dish and suspended in the air for 30 minutes to allow solvent evaporation. The more volatile 2,3-dimethyl-5-isobutyl pyrazine was dissolved in paraffin oil. Acetone or paraffin oil (500µl) served as control. A nitrile glove (Sol-vex) was modified as described in Supplementary Fig. 7 such that a 5.8cm × 5cm window was present for skin odour exposure. A set of magnetic window frames were designed to secure the treated net ∼1.5 mm above skin, and a second untreated netting ∼4.5 mm above the treated net in a manner so that mosquitoes were attracted to skin emanations in the open window but unable to contact treated nets with tarsi, or contact and pierce skin. Additionally the test compound had minimal contact with skin. A clean set of glove and magnets were used for every trial. Care was taken that experimenter did not use cosmetics, soap etc on arms. For each trial the arm was first inserted for 5 min and the number landing or escaping test window recorded on video for 5-min period. Solvent controls were always tested prior to treatment. It was determined first that a solvent treated arm when offered to the same cage with a gap of 5 minutes slightly more mosquitoes were attracted the second time around, therefore providing a more rigorous assay for the repellents. No cage was tested more than once within 1 hour of a testing session and not more than twice on any single day. Videos were analysed blind and the numbers of mosquitoes present for a 5-sec continuous duration were

counted every minute. Mosquitoes reliably started accumulating in controls at the 2 min point, and data from this time point was considered for analysis.

Percentage present = average number of mosquitoes on window for 5 seconds at a given time-point across trials. All values were normalized to percentage of the highest value for the comparison, which was assigned a 100 percent present.

Percentage repellency = [1 - (mean cumulative number of mosquitoes on the window of treatment for 5 seconds at time points 2,3,4,5 min/mean cumulative number of mosquitoes that remained on window of solvent treatment for 5 seconds at time points 2,3,4,5 min)] × 100.

Escape Index = (Average Number of mosquitoes in treatment that landed yet left the mesh during a five second window over the following time points: 2 minutes, 3 minutes, 4 minutes, 5 minutes)/(Average Number of mosquitoes that landed yet left the mesh during a five second window over the same time points in (treatment + control)) Each time point has N=5 trials, 40 mosquitoes per trial, Except for EA, where N=4.

Chemical Informatics

Calculation of descriptors—A single energy-minimized 3-Dimensional structure was predicted for each compound using of the Omega2 software package[40]. The commercially available software package Dragon (3,224 individual descriptors) from Talete was used to calculate molecular descriptors[41]. Descriptor values were normalized across compounds to standard scores by subtracting the mean value for each descriptor type and dividing by the standard deviation. Molecular descriptors that did not show variation across compounds were removed.

Classification of repellent compounds—For our analysis, compounds from different studies were approximated into a single metric of “protection duration” as a rough indicator of repellency. The non-repellent diversifying training set of odours were assigned protection times of zero, while the approved repellents DEET and Picaridin were assigned the highest value since we made the assumption that these would have structural properties important for regulatory approval. Compounds were clustered using Euclidean distance and hierarchical clustering based on differences in repellency values, and a set of 5 compounds with the highest activity that clustered together was classified as “training repellents”.

Determination of optimized descriptor subsets—A compound-by-compound repellency distance matrix was calculated from repellency data. A separate compound-by-compound descriptor distance matrix was calculated using the 3,224 descriptor values calculated by the Dragon software package. Using a Sequential Forward Selection (SFS) approach, all descriptors are individually compared and selected for their ability to increase the correlation between descriptor values and repellency. The descriptor that correlates best is retained and each further iteration adds an additional descriptor to improve the correlation values. This process is continued until additional descriptors fail to improve the correlation value from the previous step. This process results in unique descriptor set that is optimized for repellency.

Support Vector Machine predictions from Odour compound libraries—This repellency-optimized descriptor set was utilized to train a Support Vector Machine (SVM) using regression and a radial basis function kernel available in the R package e1071, which integrates libsvm [42,43]. Optimal gamma and cost values were determined using the Tune.SVM function. The resulting trained SVM was then applied to predict activity for compounds from two libraries *in silico*, a natural compound library of ~3200 volatiles and a >440,000 compounds library.

For the natural compound library we assembled a subset of 3,197 volatile compounds from defined origins including plants, humans, insects[44], food flavours and a fragrance collection[45] including fruit and floral volatiles[46-53]. For the larger library we assembled a subset of >440,000 small molecules from the eMolecules database[54] that have properties of volatile odourants. (MW <325 and atoms: C, O, N, H, S).

We performed a 5-fold cross-validations by dividing the dataset randomly into 5 equal sized partitions. Four of the partitions were applied to train the SVM and the remaining partition, which was not used for training, was used to test predictive ability. This process is repeated five times, each trial excluding a different subset of compounds as the training set and assigning the remainder as the test set. The whole process is repeated 20 times to improve consistency. A receiver operating characteristics (ROC) analysis is then used to analyze the performance of our computational repellency prediction. The overall predictive ability was calculated as a single receiver operating characteristic (ROC) curve for all 20 independent validations.

Calculation of LogP and vapor pressure values—SMILES structures of the predicted repellent odours were used with EPI Suite (http://www.epa.gov/oppt/exposure/pubs/episuite.htm) to calculate predicted LogP and Vapour Pressure values.

Vinyl solubility test

One 3 × 3 mm square of 4 gauge vinyl was submerged in 1mL of each test compound in a glass container and stirred at a constant rate on a shaker and checked every 30 minutes until the vinyl square in DEET was completely dissolved (6 Hrs). The vinyl pieces in each of the other compounds was removed, rinsed in ethanol and weighed. The process was repeated at 30 Hrs (24 Hrs after the vinyl square completely disappeared in DEET).

Statistical analyses

For behaviour experiments with preference index, arcsine-transformed data were analyzed. Tests used are indicated in the figure legends and they are Students *t*-test,1-way ANOVA and Tukey's *post hoc* analysis. Statistical tests for each experimental category and sample trails sizes were selected on the basis of previously published studies using similar assays, which are cited throughout the manuscript. For all graphs, error bars indicate S.E.M.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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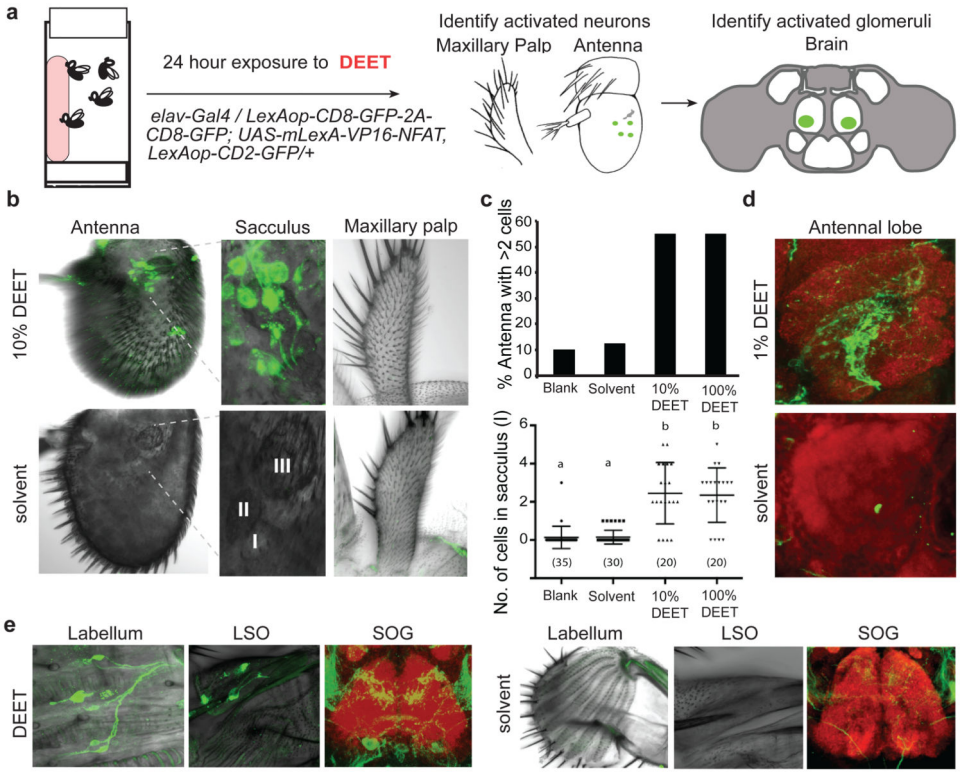


Figure 1. DEET is detected by Ir40a+ sacculus neurons

a, Schematic of the NFAT (CaLexA)-based method to label neurons activated by DEET. **b,** Confocal micrographs of olfactory organs from flies stimulated with 10% DEET or solvent (acetone). **c,** Quantification of GFP+ antennae (Top) and mean numbers of GFP+ cells in chamber I. n=35 (blank), n=30 (solvent), n=20 (10%DEET), n=20 (100%DEET). $P <$ 0.0001,1-way ANOVA with Tukey's *post hoc* test. **d,** GFP+ axonal termini in antennal lobes of flies treated as indicated. **e,f** Expression of GFP+ in the labellum, labral sense organ (LSO), the sub-esophageal ganglion (SOG). Anti-GFP (green) and anti-nc82 (red). For SOG, dorsal is top.

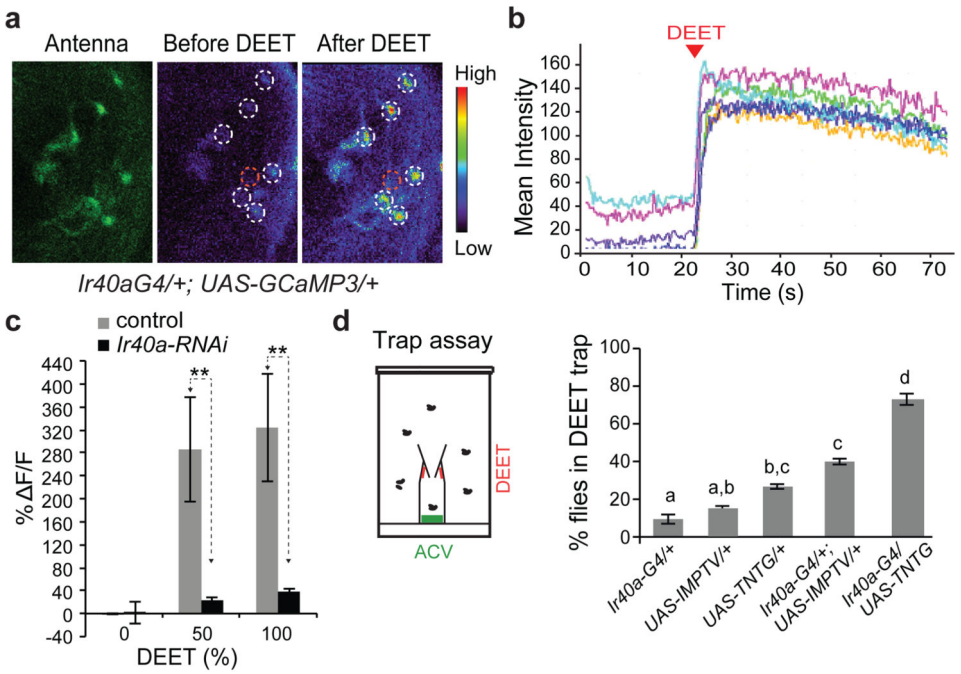


Figure 2. *Ir40a* neurons detect DEET and are required for repellency

a, Images of calcium activity in *Ir40a-Gal4/+;UAS-GCaMP3/+* neurons color-coded as indicated (right). Measurements taken from areas in dashed circles: cells (white), background (red). **b,** Mean fluorescence intensities for 6 different cells. Red arrowhead indicates onset of ∼2-sec puff of DEET. **c,** Mean percentage change in fluorescence intensity after application of ∼2-sec indicated stimulus; genotypes were *Ir40aGal4/+;UAS-GCaMP3/+* (control) and *Ir40aGal4/Ir40aGal4;UAS-GCaMP3/UAS-Ir40aRNAi(2)* (*Ir40a-RNAi*). n=10-13. ***P* < 0.01, Student's *t*-test. **d,** Schematic (left) and results (right) for DEET-treated trap assays for indicated genotypes. n=6 trials, 20 flies/trial for each genotype. Letters indicate statistical significance, $P \leq 0.008$, 1-way ANOVA with Tukey's *post hoc* analysis. Error bars=S.E.M.

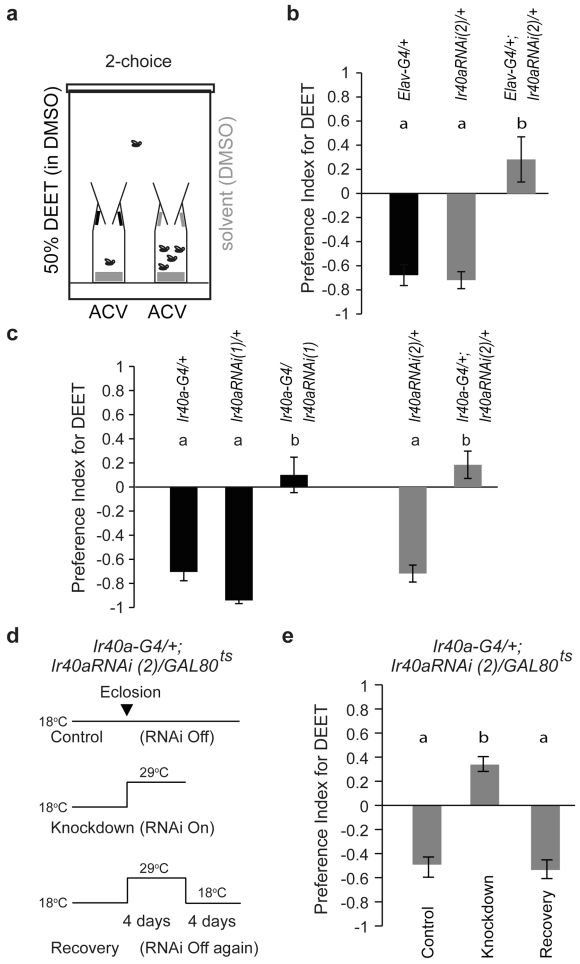


Figure 3. *Ir40a* is required for DEET avoidance

a, Set-up for behavioural 2-choice assay. **b,c** Mean preference index of indicated genotypes for DEET in 2-choice assays using **b,** *elav-Gal4*, and **c,** *Ir40a-Gal4*. n=6 trails (20 flies/trial) except *elav-Gal4/+;Ir40aRNAi(2)* n=10 trials and RNAi experiments with *Ir40a-Gal4* n=12 trials each. **d,** Genotype and schematic for post-developmental knockdown and recovery of *Ir40a*. **e,** Mean DEET preference index of flies derived from indicated treatments in 2-choice assays. n=6 trials for all conditions, with 20 flies/trial. For **b-e**, $P < 0.001$, 1-way ANOVA with Tukey's *post hoc* analysis. Error bars=S.E.M.

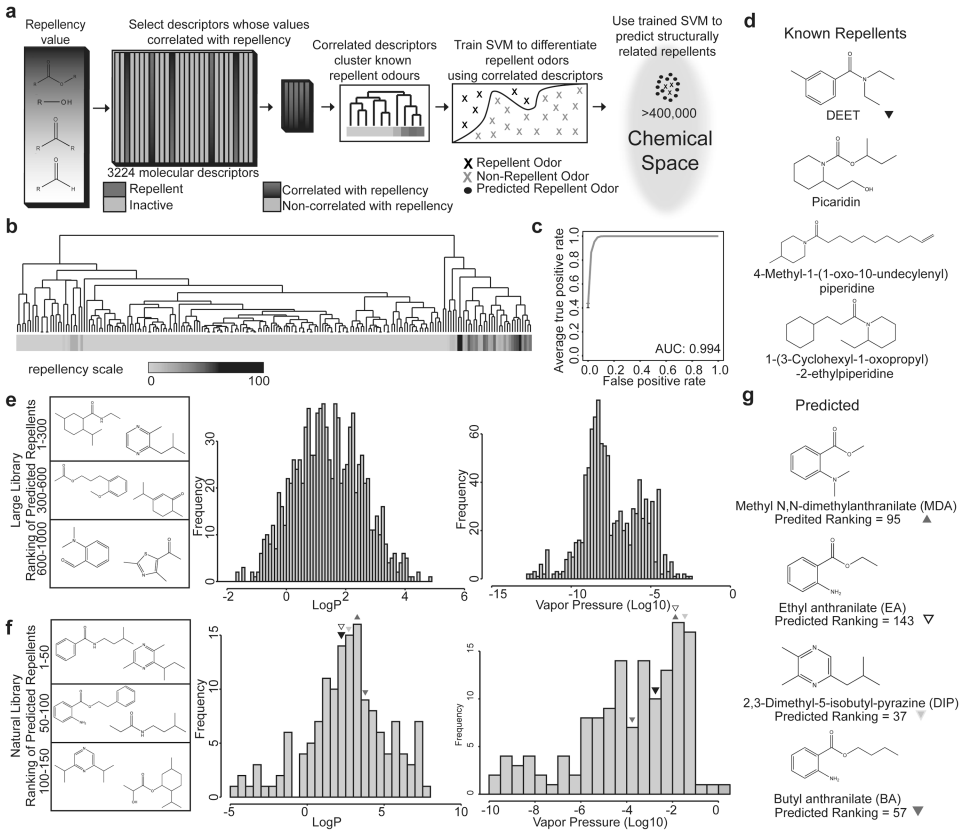


Figure 4. Chemical informatics prediction of new repellents

a, Cheminformatics discovery pipeline to identify novel DEET-like repellents. **b,** Hierarchical cluster analysis of 201 training set odorants using optimized descriptors to calculate distances in chemical space. **c,** Receiver-operating-characteristic curve (ROC) representing computational validation of repellent predictive ability from 20 independent 5-fold cross validations. AUC=Area under the curve. **d,** DEET, Picaridin, and two unapproved repellents[25]. **e,** Representative predicted repellents from >400,000 odorant library (Left) and computationally determined values for 1000 top-ranked predicted repellents (Right). **f,** Representative predicted repellents from >3,000 natural odour library (Left) and computationally determined values for 150 top-ranked predicted repellents (Right). Colour arrowheads indicate values for DEET and selected odours shown in **g**.

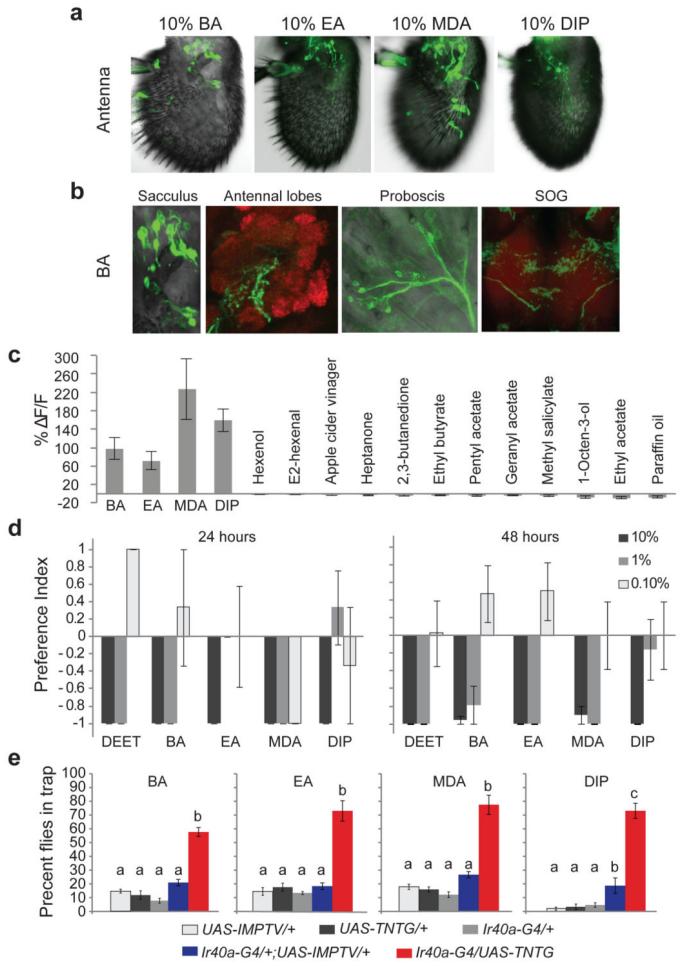
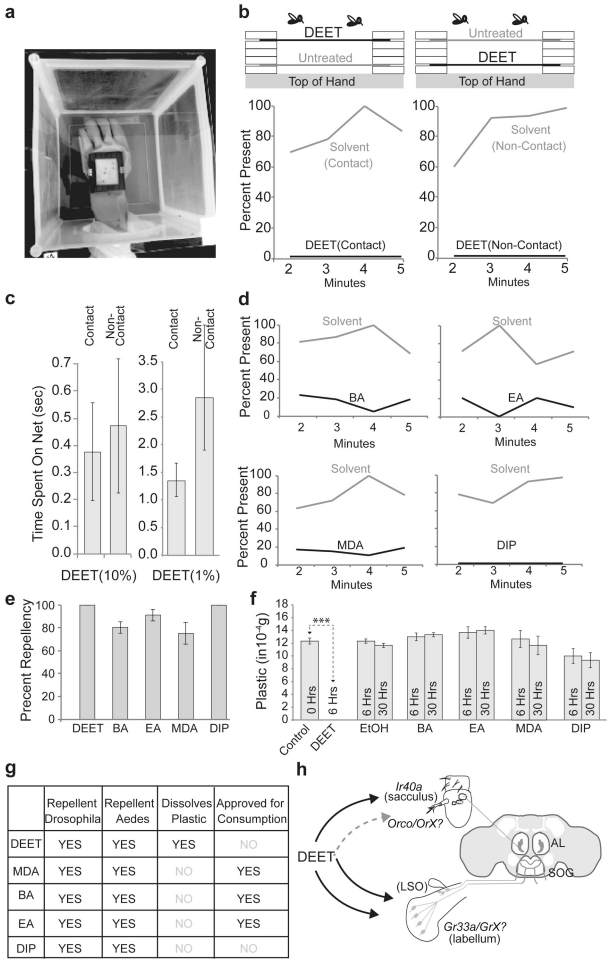


Figure 5. Predicted repellents activate Ir40a neurons and are strong repellents for *Drosophila*

a, Images of antenna of *elav-Gal4/LexAop-CD8-GFP-2A-CD8-GFP; UAS-mLexA-VP16-NFAT, LexAop-CD2-GFP/+* flies exposed to indicated stimuli for 24 hrs. **b,** BA-activated GFP+ neurons in indicated tissues. **c,** Mean changes in fluorescence intensity in *Ir40aGal4/+;UAS-GCaMP3/+* cells after ∼2-sec application of indicated odorants. n=9-17. **d,** Mean responses of flies to predicted repellents in 2-choice olfactory and gustatory trap assays measured at 24 and 48 hrs. n=3-10 trials (24 hours) and 7-10 (48 hrs); 10 flies/trial, trials with <40% participation were excluded. **e,** Quantification of flies of indicated genotypes entering repellent-treated traps. n=6 trials for each genotype, ∼20 flies for each trial. $P < 0.001$, 1-way ANOVA with Tukey's *post hoc* test. For **c-e**, error bars=S.E.M.



	Repellent Drosophila	Repellent Aedes	Dissolves Plastic	Approved for Consumption
DEET	YES	YES	YES	NO
MDA	YES	YES	NO	YES
BA	YES	YES	NO	YES
EA	YES	YES	NO	YES
DIP	YES	YES	NO	NO

Figure 6. A new class of mosquito repellents with desirable safety profiles

a, Arm-in-cage assay to measure repellency in mosquitoes. **b,** Mean percentage of female *A. aegypti* present for >5 sec on top net (Left=10% DEET, Right=solvent). Solvent controls performed separately (dark gray). **c,** Average time on net for each landing event in **b. d,** Mean percentage of female *A. aegypti* present for >5 sec on top net in non-contact assay. **e,** Cumulative repellency summed across minutes 2-5 of indicated non-contact treatment (10%) in comparison to appropriate solvent control. 40 mosquitoes/trial, n=5 trials/treatment for **b,c,d,** and **e. f,** Mean weight of vinyl pieces following submersion in indicated compounds for indicated amount of time. n=3, ***$P < 10^{-5}$, Student's *t*-test. Error bars=S.E.M. **g,** Properties of new repellents. **h,** Model for DEET detection and processing in *Drosophila*.