UC Davis UC Davis Previously Published Works

Title

Draft genome sequence of multidrug-resistant Citrobacter freundii MTR_GS_V1777 strain isolated from a spinach (Spinacia oleracea) sample in Gazipur, Bangladesh.

Permalink

https://escholarship.org/uc/item/4jr6h3qd

Journal

Microbiology Resource Announcements, 13(2)

Authors

Pramanik, Pritom Rana, Md Ramasamy, Srinivasan <u>et al.</u>

Publication Date

2024-02-15

DOI

10.1128/mra.01082-23

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <u>https://creativecommons.org/licenses/by/4.0/</u>

Peer reviewed



8 | Bacteriology | Announcement



Draft genome sequence of multidrug-resistant *Citrobacter freundii* MTR_GS_V1777 strain isolated from a spinach (*Spinacia oleracea*) sample in Gazipur, Bangladesh

Md. Saiful Islam,^{1,2} Pritom Kumar Pramanik,¹ Md. Liton Rana,¹ Srinivasan Ramasamy,³ Pepijn Schreinemachers,⁴ Ricardo Oliva,³ Md. Tanvir Rahman¹

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT We announce a genome sequence of *Citrobacter freundii* MTR_GS_V1777 strain isolated from a vegetable sample in Bangladesh. This strain had a genome size of 4,997,753 bp ($58.7 \times$ genome coverage) and contained two plasmids, typed as sequence type ST124, 38 predicted antibiotic resistance genes, and 77 predicted virulence factor genes.

KEYWORDS vegetable, whole genome, *Citrobacter freundii*, multidrug resistance, virulence, gardening system, Bangladesh

Wegetables are commonly exposed to microbial contamination due to contact with soil, dust, untreated irrigation water, and handling during harvesting and postharvest processing (1). *Citrobacter* species are among the bacteria that have been found in vegetables (2). The widespread and improper use of antimicrobial agents has led to the rise of antimicrobial resistance and multidrug resistance (MDR) in bacteria, including *Citrobacter* species (3, 4).

Between September 2022 and March 2023, fresh spinach (Spinacia oleracea) leaf samples were collected from the surface garden in the Gazipur district of Bangladesh (24.0958°N, 90.4125°E) and brought to the laboratory (24.7245°N, 90.4372°E). The collected samples were processed according to the methods outlined in a previous study (5). The samples were carefully chopped, weighed (50 g), and placed into a sterile polyethene stomacher bag with 200 mL of buffered peptone water. Subsequently, they were macerated for 5 minutes at 230 rpm using a Stomacher 400 circulator (Seward Ltd., London, UK). The processed samples were then incubated at 37°C for 24 hours, spread onto xylose-lysine deoxycholate agar plates, and incubated at 37°C overnight. The resulting colonies were subjected to Gram staining and biochemical tests to isolate Citrobacter freundii (6). Identification of C. freundii was accomplished using matrix-assisted laser desorption ionization time-of-flight mass spectrometry (7). The disk diffusion method (8) and the Clinical and Laboratory Standards Institute guidelines (9) were applied to ascertain their MDR properties. The MDR C. freundii MTR_GS_V1777 isolate, showing phenotypic resistance to gentamicin, streptomycin, ciprofloxacin, erythromycin, azithromycin, cephalexin, tetracycline, ampicillin, cotrimoxazole, and sulfonamide, was selected in this study. A single colony was then picked from the overnight culture in nutrient broth (HiMedia, India) at 37°C, and the DNA was extracted with a DNeasy Blood and Tissue Kit (QIAGEN, Hilden, Germany). Subsequently, a DNA library was prepared using the Nextera DNA Flex Library Prep Kit (Illumina, San Diego, CA, USA). Genome sequencing was performed on the Illumina NextSeq 2000 platform with 2×150 bp reads. The genome assembly involved Unicycler v.0.4.9 (10) and included initial raw paired-end read trimming (n = 1,215,420) with Trimmomatic v.0.39 (11) (leading: 20,

Editor David A. Baltrus, The University of Arizona, Tucson, Arizona, USA

Address correspondence to Md. Tanvir Rahman, tanvirahman@bau.edu.bd.

The authors declare no conflict of interest.

See the funding table on p. 2.

Received 8 November 2023 Accepted 21 December 2023 Published 11 January 2024

Copyright © 2024 Islam et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license. sliding window: 4:20:20, trailing: 20, and minlen = 36) to remove Illumina artifacts and phiX reads. Quality assessment was conducted using FastQC v.0.11.7 (12), and genome annotation was done by PGAP v.6.6 (13). Within the assembled genome, sequence type was predicted using MLST v.2.0 (14); pathogenicity index was obtained using PathogenFinder v.1.1 (15); and plasmids were identified with PlasmidFinder v.2.1 (16). Antibiotic resistance genes (ARGs) were predicted using CARD v.3.2.4 (RGI main) (17), with specific criteria for perfect (100% identity) and strict (>95% identity) matches to curated reference sequences in the CARD databases. Virulence factor genes (VFGs) were predicted through VFDB with VFanalyzer (18), and metabolic functional features were identified with RAST v.2.0 (19). Default settings were used for all software unless otherwise noted.

The genome coverage of the *C. freundii* MTR_GS_V1777 strain was 58.7×, and a total of 53 contigs were obtained. Our assembled genome had a total length of 4,997,753 bp, a guanine-cytosine content of 51.8%, four contig L50, and an N50 value of 514,523 bp. It contained a total of 4,822 genes, 4,740 coding sequences, 4,670 protein-coding sequences, 82 RNA genes, and 70 pseudogenes. This assembled genome predicted two plasmid replicons, i.e., IncFII(K) and IncR. Moreover, this genome consisted of a sequence type of ST124 and a pathogenicity index of 0.875. The strain carried 38 predicted ARGs and 77 predicted VFGs. Moreover, this genome contained 386 subsystems (having 2,259 genes) with 32% coverage.

ACKNOWLEDGMENTS

This work was conducted as part of the Consortium of International Agricultural Research Centers (CGIAR) Research Initiative on Resilient Cities Through Sustainable Urban and Peri-urban Agri-food Systems and is supported by contributors to the CGIAR Trust Fund (https://www.cgiar.org/funders).

AUTHOR AFFILIATIONS

 ¹Department of Microbiology and Hygiene, Faculty of Veterinary Science, Bangladesh Agricultural University, Mymensingh, Bangladesh
²Department of Animal Sciences, University of California—Davis, Davis, California, USA
³World Vegetable Center, Shanhua, Tainan, Taiwan
⁴World Vegetable Center, East and Southeast Asia, Bangkok, Thailand

AUTHOR ORCIDs

Md. Saiful Islam (b http://orcid.org/0000-0002-6870-4595 Md. Liton Rana (b http://orcid.org/0000-0003-1400-1250 Srinivasan Ramasamy (b http://orcid.org/0000-0002-8454-7331 Md. Tanvir Rahman (b http://orcid.org/0000-0001-5432-480X

FUNDING

Funder	Grant(s) Author(s)
Consortium of International Agricultural Research Centers (CGIAR)	Md. Tanvir Rahman

AUTHOR CONTRIBUTIONS

Md. Saiful Islam, Data curation, Formal analysis, Investigation, Methodology, Software, Writing – original draft, Writing – review and editing | Pritom Kumar Pramanik, Data curation, Formal analysis, Investigation, Software | Md. Liton Rana, Investigation, Methodology | Srinivasan Ramasamy, Conceptualization, Writing – review and editing | Pepijn Schreinemachers, Conceptualization, Writing – review and editing | Ricardo Oliva, Conceptualization, Project administration, Supervision, Writing – review and editing | Md. Tanvir Rahman, Conceptualization, Funding acquisition, Project administration, Resources, Supervision, Validation, Writing – review and editing

DATA AVAILABILITY

The whole-genome sequencing shotgun analysis of *Citrobacter freundii* MTR_GS_V1777 was deposited to GenBank under accession number JAVTVR000000000. The relevant data, including the raw reads, were also submitted with BioProject accession number PRJNA1020260, BioSample accession number SAMN37518445, and SRA accession number SRR26151867. In this paper, the specific version being referred to is identified as JAVTVR000000000.1.

REFERENCES

- Kemajou TS, Awemu GA, Digban KA, Oshoman CE, Ekundayo OI, Ajugwo AO. 2017. Microbiological studies of vegetable leaves sold in Elele market, rivers-state, Nigeria. J Transm Dis Immun 1:1–5. https://doi.org/ 10.21767/2573-0320.10004
- Adegun BR, Oluduro AO, Aregbesola OA. 2019. Isolation and molecular characterization of citrobacter species in fruits and vegetables sold for consumption in ILE-IFE, Nigeria. Scientific African 6:e00173. https://doi. org/10.1016/j.sciaf.2019.e00173
- Islam MS, Rahman A, Hassan J, Rahman MT. 2023. Extended-spectrum beta-lactamase in *Escherichia coli* isolated from humans, animals, and environments in Bangladesh: a one health perspective systematic review and meta-analysis. One Health 16:100526. https://doi.org/10. 1016/j.onehlt.2023.100526
- Ahmed T, Islam MS, Nuruzzaman M, Sadekuzzaman M, Kabir SML, Rahman MT, Khan MSR. 2023. Draft genome sequence of the multidrugresistant *Citrobacter freundii* 132-2 strain isolated from a domestic duck in Bangladesh. Microbiol Resour Announc 12:e0037823. https://doi.org/ 10.1128/mra.00378-23
- Ratshilingano MT, du Plessis EM, Duvenage S, Korsten L. 2022. Characterization of multidrug-resistant *Escherichia coli* isolated from two commercial lettuce and spinach supply chains. J Food Prot 85:122–132. https://doi.org/10.4315/JFP-21-125
- Brenner DJ, O'Hara CM, Grimont PA, Janda JM, Falsen E, Aldova E, Ageron E, Schindler J, Abbott SL, Steigerwalt AG. 1999. Biochemical identification of *Citrobacter* species defined by DNA hybridization and description of *Citrobacter gillenii* sp. nov.(formerly *Citrobacter* genomospecies 10) and *Citrobacter murliniae* sp. nov.(formerly *Citrobacter* genomospecies 11). J Clin Microbiol 37:2619–2624. https://doi.org/10.1128/JCM.37.8. 2619-2624.1999
- Kolínská R, Spanělová P, Dřevínek M, Hrabák J, Zemličková H. 2015. Species identification of strains belonging to genus *Citrobacter* using the biochemical method and MALDI-TOF mass spectrometry. Folia Microbiol (Praha) 60:53–59. https://doi.org/10.1007/s12223-014-0340-4
- Bauer AW, Kirby WMM, Sherris JC, Turck M. 1966. Antibiotic susceptibility testing by a standardized single disk method. Am J Clin Pathol 45:493– 496.
- 9. M100-S32. 2022. Performance standards for antimicrobial susceptibility testing. Clinical and Laboratory Standards Institute, Wayne, PA, USA.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595

- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible Trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/ 10.1093/bioinformatics/btu170
- 12. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. Available from: http://www.bioinformatics.babraham.ac. uk/projects/fastqc
- Li W, O'Neill KR, Haft DH, DiCuccio M, Chetvernin V, Badretdin A, Coulouris G, Chitsaz F, Derbyshire MK, Durkin AS, Gonzales NR, Gwadz M, Lanczycki CJ, Song JS, Thanki N, Wang J, Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. RefSeq: expanding the prokaryotic genome annotation pipeline reach with protein family model curation. Nucleic Acids Res 49:D1020–D1028. https://doi.org/10. 1093/nar/gkaa1105
- Larsen MV, Cosentino S, Rasmussen S, Friis C, Hasman H, Marvig RL, Jelsbak L, Sicheritz-Pontén T, Ussery DW, Aarestrup FM, Lund O. 2012. Multilocus sequence typing of total-genome-sequenced bacteria. J Clin Microbiol 50:1355–1361. https://doi.org/10.1128/JCM.06094-11
- Cosentino S, Voldby Larsen M, Møller Aarestrup F, Lund O. 2013. Pathogenfinder-distinguishing friend from foe using bacterial whole genome sequence data. PLoS One 8:e77302. https://doi.org/10.1371/ journal.pone.0077302
- Carattoli A, Zankari E, García-Fernández A, Voldby Larsen M, Lund O, Villa L, Møller Aarestrup F, Hasman H. 2014. *In silico* detection and typing of plasmids using plasmidfinder and plasmid multilocus sequence typing . Antimicrob Agents Chemother 58:3895–3903. https://doi.org/10.1128/ AAC.02412-14
- Alcock BP, Raphenya AR, Lau TTY, Tsang KK, Bouchard M, Edalatmand A, Huynh W, Nguyen A-LV, Cheng AA, Liu S, et al. 2020. CARD 2020: antibiotic resistome surveillance with the comprehensive antibiotic resistance database. Nucleic Acids Res. 48:D517–D525. https://doi.org/ 10.1093/nar/gkz935
- Liu B, Zheng D, Zhou S, Chen L, Yang J. 2022. VFDB 2022: a general classification scheme for bacterial virulence factors. Nucleic Acids Res 50:D912–D917. https://doi.org/10.1093/nar/gkab1107
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, et al. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:1–15. https: //doi.org/10.1186/1471-2164-9-75