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# Journal

Neuron, 110(16)

# Authors

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# **Publication Date**

2022-08-17

## DOI

10.1016/j.neuron.2022.04.030

Peer reviewed



# **HHS Public Access**

Author manuscript *Neuron.* Author manuscript; available in PMC 2023 August 17.

Published in final edited form as:

Neuron. 2022 August 17; 110(16): 2545-2570. doi:10.1016/j.neuron.2022.04.030.

# The Emergence and Influence of Internal States

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#### Abstract

Animal behavior is shaped by a variety of "internal states" – partially hidden variables that profoundly shape perception, cognition, and action. The neural basis of internal states, such as fear, arousal, hunger, motivation, aggression, and many others, is a prominent focus of research efforts across animal phyla. Internal states can be inferred from changes in behavior, physiology, and neural dynamics and are characterized by properties such as pleiotropy, persistence, scalability, generalizability, and valence. To date, it remains unclear how internal states and their properties are generated by nervous systems. Here we review recent progress, which has been driven by advances in behavioral quantification, cellular manipulations, and neural population recordings. We synthesize research implicating defined subsets of state-inducing cell types, widespread changes in neural activity, and neuromodulation in the formation and updating of internal states. In addition to highlighting the significance of these findings, our review advocates for new approaches to clarify the underpinnings of internal brain states across the animal kingdom.

#### In Brief

In this review, Flavell, Gogolla, Lovett-Barron, and Zelikowsky synthesize research across animal models to discuss the classification of internal states, the roles of state-modulating neurons, and the impact of diverse states on neural dynamics and behavior

#### Keywords

Internal States; Neuromodulation; Neural Circuits; Brain-Wide Activity

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#### Introduction

Nervous systems are in a constant state of flux, with rich internal dynamics that determine how brains respond to inputs and produce outputs. The hidden processes that underlie these dynamics can be described as "internal states", and include arousal, motivation, emotion, and varying homeostatic needs. Internal states allow us to integrate information about our external environment and internal physiological conditions into centralized brain states, which shape how sensory information is processed and orchestrate appropriate behavioral and physiological responses (Anderson, 2016; Bolles, 1967; Tinbergen, 1951).

While internal states are difficult to observe directly, they can be inferred from observations of an animal's overt behavior and systemic physiology, or from within the brain, such as by investigating neuronal dynamics or perturbing neural function. For instance, an animal's state of hunger can be determined based on caloric deficit and circulating hormones, or its state of aggression inferred from observing attacks elicited by conspecifics. Likewise, several recent studies have discovered consistent changes in neuronal dynamics encompassing multiple cell types and brain systems concomitant to behavioral and/or physiological state changes (Grundemann et al., 2019; Lovett-Barron et al., 2020; Xu et al., 2020). A wide variety of animals – from jellyfish to humans – appear to organize their behavior in a state-like fashion, suggesting that the neural mechanisms that underlie the generation of internal brain states are evolutionarily ancient (Nath et al., 2017; Weissbourd et al., 2021). In humans, changes in state representation, switching, and timing are thought to occur in many psychiatric and neurological diseases. Here our focus is on the study of experimentally tractable animal models; but, the ubiquity of internal states across animal species suggests that general principles found in animals will hold relevance for understanding the human condition in health and disease.

Several recent technical advances have spurred remarkable progress in our ability to describe and investigate internal states in animal models. These include new and improved methods for tracking animal behavior, manipulating neurons, and analyzing population-level neural activity. Studies across a range of animal models now provide evidence that internal brain states can be controlled by the actions of small subsets of neurons, but can influence activity across broad swaths of the brain, often in parallel. Across organisms, neuromodulators have been repeatedly identified as central elements in the generation of internal states, with a wide range of circuit organizations that deploy neuromodulators in distinct manners (Bargmann, 2012; Getting, 1989; Harris-Warrick and Marder, 1991; Marder, 2012; McGinley et al., 2015b).

Here, we begin by defining internal states, focusing on the features which characterize them. Next, we review the experimental approaches used to study internal states, the neural basis of internal states, and the central role that neuromodulation plays in the formation and function of internal states. Finally, we close by highlighting key emerging themes of internal state control across species, including the ability of states to influence multiple circuits and cell types in parallel, the action of neuromodulators to mediate states in concert, the neural properties governing state transitions, and the persistence of states via

recurrent dynamics. The principles discussed here derive from a large and diverse literature, growing out of psychology, neuroscience, cognitive science, biology and ethology over many decades. As we cannot provide an exhaustive accounting of this work, we instead focus on specific principles that are common across organisms and highlight recent findings that have relevance for scientists currently studying internal states.

#### **Defining internal states**

Internal brain states can be defined from changes in physiology, behavior, and/or brain activity. We use the term "internal state" to refer to a state that can be independently controlled and which can occur simultaneously with other states within the same animal. For example, hunger and fear represent distinct internal states. The states that we discuss here all consist of changes in nervous system function that can be inferred from an animal's behavior (though such inference can be challenging, since states are not entirely overt; see below). In addition, some internal states involve changes in other parts of the body. For example, hunger involves changes in gut metabolism, hormone levels, and more. These interactions between the brain and the periphery can be bi-directional. We consider these peripheral changes to be important aspects of the state. We expect that the definition of "internal state" will become more precise as the field evolves and we return to the complexities of this definition at the end of the review. In this review, we will start by discussing characteristic features of internal states, how they can be inferred from behavioral and physiological changes, and then discuss their neuronal correlates.

#### Features of internal states

Internal states enable us to produce flexible and adaptive behavioral and physiological responses in a wide range of different settings. These internal states are stable enough to organize behaviors over long timescales, and flexible enough to facilitate adaptive (or maladaptive) responses to different circumstances or changing environments. To be both flexible and stable, internal states often possess the following features: *pleiotropy*, persistence, scalability, generalizability, and valence (Figure 1) (Adolphs and Anderson, 2013; Anderson, 2016; Darwin, 1872; Tye, 2018). Pleiotropy refers to the feature that each state influences multiple aspects of behavior and physiology in parallel, such as body temperature, respiration, locomotion, sensory responsiveness, and more (Figure 1). *Persistence* describes the ability of internal states to produce behavioral and physiological responses that outlast the termination of the stimulus that initiated the response. We do not consider individual motor actions to be states, but persistent sequences of motor actions may be classified as states. Scalability indicates the ability of these responses to scale with the magnitude of the stimulus. Generalizability refers to the degree to which an internal state can produce responses to stimuli that are distinct from the original stimulus that elicited the response. Valence describes the positive or negative affect associated with that state. Taken together, the multifaceted and flexible nature of internal states provides evolutionary advantages for organisms across the animal kingdom.

#### A prototypical internal state: fear

The above mentioned properties of internal states can be conceptualized in the context of emotion, and can be well illustrated using one of the most well studied states in neuroscience and psychology – fear (Adolphs, 2008; Dukes et al., 2021; Fanselow, 2018; Fanselow and Pennington, 2018; Janak and Tye, 2015; LeDoux and Daw, 2018; LeDoux, 2017; 2020; LeDoux and Brown, 2017; Mobbs et al., 2019; Tovote et al., 2015; Tye and Deisseroth, 2012). For example, if you are afraid of flying on a plane, you might display a set of *pleiotropic* changes including an increase in heart rate, galvanic skin response, and feelings of anxiety, which *persist* well beyond the time in which you are exposed to the plane (stimulus). These neural and peripheral responses might *scale* with the strength of the stimulus, such that they increase during turbulence, and they may *generalize* to other similar stimuli, such as helicopters or cars. The *valence* of this state is negative, causing you to avoid flying in a plane as much as possible.

In laboratory settings, the internal state of fear is often investigated using classical conditioning (Pavlov, 1927) in which an animal, often a rodent, is conditioned to fear a previously neutral cue (e.g. auditory tone) which, through training, comes to predict the occurrence of an aversive stimulus (e.g. foot shock). These classical conditioning paradigms allow for precise control over experimental parameters and their effects on fear. In both controlled, as well as more naturalistic settings, an animal may display a wide variety of fear-related behaviors - fleeing, freezing, fighting - depending on the imminence of the threat and the shape of the environment (Fanselow, 2018; Fanselow et al., 2019; Fanselow and Lester, 1988; Perusini and Fanselow, 2015). These fear behaviors demonstrate hallmark characteristics of an internal state. For example, in rats and mice, freezing behavior scales with the magnitude of the foot shock (Fanselow and Bolles, 1979), generalizes to similar auditory cues, and can *persist* well beyond termination of the auditory stimulus (Quinn et al., 2002). These behavioral readouts correspond to physiological findings, which identify neurons that are active during fear conditioning and/or expression, *persist* in their activity beyond termination of a fear-eliciting stimulus, *generalize* their activity to similar stimuli, and scale the intensity of their activity depending on stimulus magnitude (e.g. Ciocchi et al., 2010; Haubensak et al., 2010).

Nevertheless, it is important to note that despite being heavily studied, fear represents one of the most hotly contested internal states, with many questions currently unanswered (see Mobbs et al., 2019 for a review of some of these issues). For example, what are the behavioral readouts that best capture the internal state of fear? How exactly is fear distinct from other similar states, such as anxiety? Do these states lie on the same continuum, and thus, collectively represent a larger internal state of defense? How does this internal state interact with prior experience? And finally, some have even argued that it may not be possible to truly study fear in non-human animals (LeDoux, 2020; 2021). Thus, while fear is a powerful, well-studied example of an internal state, fear also represents some of the challenges facing the field of internal states.

While fear in rodents exemplifies many of the characteristics of an internal state – at both the behavioral and neurobiological level – examples of numerous behaviors influenced by internal states can be found in almost every species studied. In the sections below, we

discuss a variety of internal states across different model organisms. Like many areas of biology searching for general principles, we believe that our understanding of internal states will benefit enormously from integrating results across multiple organisms and behavioral conditions (Jourjine and Hoekstra, 2021; Katz, 2016; Laurent, 2020; Yartsev, 2017).

#### Experimental approaches to studying internal states

Investigating the neural basis of internal states requires the accurate inference of such states, extracted from measurements and manipulations of behavior, physiological parameters, and environmental context (Figure 2A). Here, we discuss different approaches for inducing and measuring internal states in a laboratory setting.

#### Experimentally inducing need states

Many studies rely on manipulating environmental or physiological variables in order to induce internal states. For instance, exposing animals to specific stimuli, environments, or physiological conditions has proven useful to induce binary global state changes; this includes induction of anxious states with threatening environments (Calhoon et al., 2018; Tovote *et al.*, 2015), induction of hunger with food or nutrient deprivation (Livneh et al., 2020; Sayin et al., 2019; Vogt et al., 2021), and induction of thirst with water deprivation (Allen et al., 2019; Livneh *et al.*, 2020; Zimmerman et al., 2017) (Figure 2B). These studies often rely on single characteristic behaviors as a readout (approach versus avoidance, exploiting versus roaming, attack versus mounting), and the robustness of these need state-induced behaviors allow for averaging results across individuals. Such approaches have been useful in identifying key characteristics of deprivation-induced need- states, enabling the exploration of their neurobiological underpinnings (Sternson, 2013).

#### Inferring internal states from overt locomotor behavior

Locomotion represents a key observable variable from which internal states can be inferred. When observing locomotion over time, experimenters can classify epochs of fast-timescale actions into slower-timescale states distinguished by the probability and content of the animal's motion (Flavell et al., 2020; Ji et al., 2021; Marques et al., 2020; Poulet and Petersen, 2008) (Figure 2C). Many organisms, including mammals, zebrafish, flies, and worms display stable, global changes in behavioral patterns such as switches between active and inactive locomotor states. Active states, characterized by longer movement trajectories, include exploration and roaming. Inactive states, characterized by little or short locomotor bouts, include idling, dwelling or exploiting (Flavell et al., 2013; Ji *et al.*, 2021; Marques *et al.*, 2020). These global patterns have been shown to also exist in more complex organisms, such as rodents (Grundemann *et al.*, 2019). Similar state-dependent switches in active versus passive behaviors have been described in the contexts of active sensing versus quiescence (Poulet and Petersen, 2008), running versus resting (Keller et al., 2012), or high versus low arousal (Rodriguez-Romaguera et al., 2020).

Measuring such bi-modal changes in 'state' can be achieved by tracking entire animals in space (Flavell *et al.*, 2020; Ji *et al.*, 2021; Marques *et al.*, 2020) and measuring course locomotion parameters or spatial coverage. Movement can also be characterized in a more

detailed manner, by tracking the position of the body and limbs over time to classify states; these studies are enabled by a recent proliferation of methods for tracking body posture

(Box 1). States can also be inferred from their effects on the performance of repeatable motor behaviors with trial-like structures. For instance, the response rate and reaction time to sensory stimuli can be used to infer arousal or alertness across species (Harris and Thiele, 2011; Lovett-Barron et al., 2017; Maimon, 2011; McGinley *et al.*, 2015b; Moore and Zirnsak, 2017; Musall et al., 2019).

#### Inferring internal states from higher-order behavior

Beyond classifying states from coarse locomotor behavior, recent studies have also focused on extracting more complex behavioral patterns to describe internal states. While methods to track animal behavior are increasingly powerful (see Box 1), it remains challenging to analyze and understand the high-dimensional behavioral data arising from these tools (Berman, 2018; Datta *et al.*, 2019). Towards this goal, machine learning (ML) has become key. For example, from the kinematic features extracted over long time scales, ML algorithms are able to extract and classify behavioral patterns and sequences, their variation across time and individuals, and their perturbation by drugs and disease models.

One such ML approach is Motion Mapper (Berman et al., 2014) which identifies behavioral modules by low-dimensional embedding and clustering. Recent evidence testing different unsupervised approaches for behavioral mapping and clustering argues that keeping the data in as many dimensions as possible for clustering is preferable (Todd et al., 2017). Other techniques use intuitive behavior annotation by the experimenter, which allows supervised ML algorithms to quantify these behaviors (e.g. JAABA (Kabra *et al.*, 2013)). Another approach that has also been successful is to measure multiple behavioral parameters and infer underlying state(s) using probabilistic approaches. For instance, Hidden Markov models (HMM) have been employed to infer behavioral states in many organisms (Calhoun et al., 2019; Cermak et al., 2020; Marques *et al.*, 2020). However, these techniques rely on variables that are quantified and identified by the experimenter as being state-relevant.

Making use of the temporal sequence of behavioral actions over time has been a particularly powerful approach to infer internal states (Figure 2D) (Berman et al., 2016; Luxem et al., 2020; Wiltschko et al., 2015; York et al., 2021). For example, two recent studies using this approach were able to classify the behavioral sequences that comprise the larval zebrafish's hunting behavior from specific eye and tail movements in the context of available prey (Johnson et al., 2020; Mearns et al., 2020). Another such technique, Motion Sequencing (MoSeq (Wiltschko *et al.*, 2015)), is an ethologically-inspired behavioral analysis method. In a recent landmark study, Wiltschko et al. (Wiltschko et al., 2020) automatically and effectively deconstructed behavioral differences and similarities elicited by a panel of neuroactive and psychoactive drugs in mice. MoSeq was able to distinguish the behavioral changes elicited by the drugs, which each elicit movement reductions through different mechanisms, such as distinguishing catalepsy and sedation, and are often confused in traditional behavioral assays. MoSeq was even able to predict drug dosage. These studies reveal that temporal sequence-based approaches can capture spontaneous transitions between diverse internal states across highly variable and diverse datasets.

#### Approaches for considering the co-existence and interactions of internal states

Despite the advances discussed above, one complication is that animals can be under the influence of multiple states at once. For instance, individuals may exist in one coherent state that integrates or selects from multiple internal needs and outside stimuli. For example, individuals may be influenced by diverse physiological and affective need states in parallel, such as thirst, hunger, fear, social isolation, and environmental conditions (availability of food, social or predator encounters). These needs and contextual changes elicit drives that compete or may be mutually reinforcing depending on the context (Duistermars et al., 2018; Eiselt et al., 2021; Thornquist and Crickmore, 2020) (Figure 2E). Together these parameters may result in integrated and complex internal states, which manifest as behavioral switches when one drive overcomes another, or may serve to generate entirely unique behavior patterns. Indeed, recent work has highlighted the overlap between distinct states such as hunger and thirst (Eiselt *et al.*, 2021; Gong et al., 2020). Interestingly, the lateral hypothalamus of the mouse has been found to be a key hub in organizing behavioral switches in response to multiple diverse internal states (Nieh et al., 2016), emphasizing the complex interactions between different need and motivational states.

To further understand the dynamics and organization of multiple internal states, such as whether they are organized hierarchically or in parallel, it may become necessary to study animal behavior over longer time scales in naturalistic settings, where animals are exposed to multiple needs and stimuli (Burnett et al., 2019; Burnett et al., 2016; Thornquist and Crickmore, 2020). For instance, can multiple states stably co-exist, or do brains exist in a unitary state that is a combination of multiple lower-level states? Are some states more likely to "win" control over behavior compared to other states? Such questions highlight the field's long-standing interest in understanding distinct need-states and how they sit in a hierarchy, with each basic need emerging once a central need is met (Maslow, 1943). In turn, these questions generate new ones - what are the rules governing the hierarchy of state control over behavior? Do different states adhere to different rules? Further experiments are required to address these interesting questions.

#### Studying individuals to address the subjectivity of internal states

A particular challenge in studying internal states arises from individuality. Past experiences, social hierarchies, contextual factors, genetic background, and hormonal influences may determine the 'personality' of individual animals and strongly shape how each individual reacts in common circumstances. Results from worms (Stern et al., 2017), flies (Honegger and de Bivort, 2018), zebrafish (Pantoja et al., 2016; Pantoja et al., 2020), and mice (Forkosh et al., 2019) argue that the neuronal underpinnings of internal states may best be addressed by studying individuals in detail (Figure 2F).

As an example of how detailed and individualized behavioral readouts may help the study of internal states, a recent study found evidence that facial expressions might represent innate and sensitive reflections of the subjective emotion state of individual mice (Dolensek et al., 2020). Employing machine-vision and ML algorithms, Dolensek et al. were able to categorize mouse facial expressions objectively and quantitatively at millisecond time scales. Notably, the authors demonstrate that the facial expressions revealed individual variability in intensity, value, and persistence of subjective emotion states (Anderson and Adolphs, 2014). Furthermore, other recent studies have found that a large fraction of the brain's activity can be explained by movement variables, read out from the face or the body (Musall *et al.*, 2019; Steinmetz et al., 2019; Stringer et al., 2019). These results highlight how powerful each individual's idiosyncratic behavior is in driving brain-wide activity changes, independent of task or stimulus involvement. This emphasizes the challenges of summarizing data across multiple animals without the ability to control for these variables.

In a powerful example of how prior experience can shape individual differences and contribute to variability in internal states, Remedios and Kennedy et al. (Remedios et al., 2017) found that exposure to social experience results in a shift in both a mouse's subsequent behavior and neuronal ensemble activity in the ventromedial hypothalamus (VMH). More specifically, naïve male mice with no prior sexual experience demonstrate a lack of aggression towards male conspecifics, which correlates with an overlap in the neural ensembles which represent male versus female conspecifics. As males are exposed to repeated social experience, aggressive behavior emerges, coupled with a separation in the neuronal ensembles which represent male versus female conspecifics. Interestingly, this shift to aggressive behavior and separable male/female ensembles in the VMH varies across mice, highlighting that the neural populations driving aggression are subject to plasticity and sensitive to additional factors controlling individual differences.

Taken together, these findings collectively argue that experiences, as well as changes in bodily condition or physiological need, exert powerful influences on the neuronal machinery from which internal states emerge. Consequently, the internal states evoked by the same set of influences may differ depending on an individual's history and current contextual standing. An important question for future research will be to ask how endocrine, genetic, plasticity and potentially further mechanisms may drive individual differences in internal state. It will be crucial to have individualized readouts of internal states at hand to tackle this important question.

#### Approaches towards improved state definitions

As mentioned above, internal states induce pleiotropic effects, impacting multiple behaviors and physiological paramaters in parallel. Thus, to improve and refine the description and detection of changes in internal states, integrated multidimensional analyses including behavioral but also physiological measurements may be key. The available measures, and ease of using them, vary depending on the species being studied. For instance, the transparent larval zebrafish may be useful for videography of the body (heartbeat, muscle tone, blood flow, respiratory movements), but less useful for testing circulating hormones (limited volume of blood to test). Larger animals, in contrast, can allow for chronically inserted devices that monitor metabolism and systemic physiology.

Future improvements in the methods to classify behaviors and internal states will likely involve making more measurements – simultaneous posture recording, physiological measures, and descriptions of the sensory environment and individual animal history. Importantly, ensuring tools for collecting and integrating such multi-modal information are "user-friendly" will be critical in their widespread use, an essential component for the

field's understanding of a given internal state. These approaches can provide more rigorous definitions of states that have already been extensively studied (arousal, fear, hunger) and may also reveal currently unknown 'states' that explain trends in behavior, but do not yet have a clear label. For instance, recent studies have identified previously unrecognized connections between neural dynamics and metabolic state (Tingley et al., 2021). Ultimately, states may be best described directly from the brain itself. We next discuss common signatures of internal states across the brains of different species.

#### The neural basis of internal states

Internal states have the capacity to influence multiple aspects of sensation, cognition, action, and systemic physiology. Here we discuss recent work highlighting how distinct populations of neurons can generate different internal states, and the influence of such states on the rest of the nervous system.

#### A neuronal population code of behavioral states

Several recent studies across different species and brain regions have highlighted that the behavioral state of an animal can be predicted and thus read-out from the activity dynamics of neuronal populations that either span brain wide networks or dominate single brain regions. For example, a study in the rodent basolateral amygdala found that two distinct neuronal populations of principle neurons predicted the switches between exploratory versus nonexploratory defensive states (Grundemann *et al.*, 2019). Similarly, networks of neurons encoding exploitation versus exploration states have been identified in fish (Marques *et al.*, 2020) and worms (Ji *et al.*, 2021). Interestingly, behavioral states can be decoded with high accuracy from the combinatorial activity of diverse molecularly defined cell types, but not from the activity of single cell types (Lovett-Barron *et al.*, 2020; Xu *et al.*, 2020). These and similar findings highlight that internal states are represented in neuronal population dynamics that recruit neurons across multiple different cell types, brain regions and neuromodulatory systems.

#### Small subsets of neurons can drive state transitions

As described above, internal states are represented in combinatorial and complex activity dynamics of entire neuronal populations. Nevertheless, the use of methods to precisely activate neurons (Luo et al., 2018) has revealed that even small subsets of neurons can drive persistent brain states with influence over a variety of behavioral features in multiple different species. Dramatic examples abound in the study of rodent behavior, where optogenetic or chemogenetic activation of genetically- and anatomically-defined subsets of neurons can evoke specific behaviors and associated brain states (Anderson, 2016; Sternson, 2013; Yizhar et al., 2011). This includes the induction of behaviors associated with hunger upon stimulation of Agouti-related peptide (AGRP) neurons in the arcuate nucleus of the hypothalamus (Aponte et al., 2011; Chen et al., 2016; Krashes et al., 2011), thirst-related behavior with stimulating neurons in the lamina terminalis (Allen et al., 2017; Augustine et al., 2018; Leib et al., 2017; Oka et al., 2015), or aggressive behaviors with stimulation of neurons in the ventromedial hypothalamus (Falkner et al., 2016; Lee et al., 2014; Lin et al., 2011), among many other examples.

These experiments have revealed some important shared features of diverse state-inducing neural populations: brief activation of these cells drives persistent states, and these cells project to multiple brain regions to induce different aspects of the core brain state (Figure 3A). For instance, activation of hunger-associated AGRP neurons induces an aversive motivational state (Berrios et al., 2021; Betley et al., 2015), promoting mice to eat food when available (Aponte et al., 2011; Krashes et al., 2011). Feeding is driven by AGRP neuron projections to the paraventricular hypothalamus (PVH), lateral hypothalamus (LH), paraventricular thalamus (PVT), and bed nucleus of the stria terminalis (BNST) (Atasoy et al., 2012; Betley et al., 2013; Horio and Liberles, 2021), but also primes mice to eat more later through its projection to the PVH (Chen et al., 2019; Chen et al., 2016; Jikomes et al., 2016), increases attention to visual and olfactory food cues through projections to the PVT (Horio and Liberles, 2021; Livneh et al., 2017; Livneh et al., 2020), suppresses fear and aggressive behavior through projections to the medial amygdala (Padilla-Coreano et al., 2016), and inhibits inflammatory nociception and the effects of appetite suppressants through projections to the parabrachial nucleus (PBN) (Alhadeff et al., 2018; Essner et al., 2017). Similarly, activation of thirst-associated neurons in the medial preoptic nucleus (MPON) that project to the PVT, PVH, or LH induce drinking behavior when water is present and induce a negative motivational drive (Allen et al., 2017a; Leib et al., 2017), in addition to increasing blood pressure through the hypothalamic projections (Leib et al., 2017). Furthermore, stimulation of aggression-associated neurons in the ventrolateral division of the ventromedial hypothalamus (VMHvl) can produce defensive behaviors through projections to the anterior hypothalamus and midbrain (Wang et al., 2015), inhibit mounting behaviors and ultrasonic vocalizations through projections to the medial preoptic area (MPOA) (Karigo et al., 2021), drive biting through outputs to the periaqueductal grey (PAG) (Falkner et al., 2020), and possesses a number of other output projections (Lo et al., 2019). These features allow a small set of neurons to influence a diversity of behavioral outcomes through specialized projections, a collateralization that is also present in the control of arousal (Poe et al., 2020), anxiety (Kim et al., 2013), and parenting (Kohl et al., 2018) in rodent brains.

The projections of putative state-control neurons are particularly well studied in rodents, but these principles have been found across multiple model systems, where stimulation of small sets of neurons with broad projections can influence internal states (Figure 3B, C). In the compact *C. elegans* nervous system, the activation of one or few neurons can induce state transitions, including the initiation of roaming and dwelling by PDF- and serotonin-releasing neurons, respectively (Churgin et al., 2017; Flavell *et al.*, 2013; Ji *et al.*, 2021), and the induction of low arousal/sleep states by peptidergic neurons (Nath et al., 2016; Turek et al., 2016; Turek et al., 2013). In *Drosophila*, aggression can be induced by activation of tachykinin-expressing neurons (Asahina et al., 2014), and threat displays are evoked by a small subset of anterior inferior protocerebrum neurons (Duistermars *et al.*, 2018). A set of male-specific P1 neurons evokes a persistent internal state of social arousal, which enhances either aggression or courtship behaviors depending on context (Anderson, 2016; Bath et al., 2014; Clowney et al., 2015; Hindmarsh Sten et al., 2021; Inagaki et al., 2014a; Jung et al., 2020; von Philipsborn et al., 2011; Zhang et al., 2016); analogous neurons in female *Drosophila* have also been found to promote persistent behavior (Deutsch et al., 2020).

While these activation studies are informative, it is important to consider the natural dynamics of state-triggering neurons as well, which may contribute to internal states in a dynamic regime not explored by artificial stimulation (Jazayeri and Afraz, 2017; Wolff and Olveczky, 2018) (Box 2).

#### Internal states influence neurons across the brain

While internal states can be initiated by small subsets of neurons, their broad effects on behavior and systemic physiology suggest that states can have wide-ranging influence over the nervous system. Across model systems, internal states have been found to influence broad swaths of the brain—findings made possible through the application of optical and electrical techniques for large-scale cellular-level recording of neurons across multiple brain regions in behaving animals (Ahrens and Engert, 2015; Engel and Steinmetz, 2019; Lin et al., 2022; Urai et al., 2022).

One class of internal state that has been studied extensively is a state of arousal associated with movement, where awake animals transition between periods of overt movement and/or enhanced alertness and periods of relative quiescence. In C. elegans, motor activity drives a large number of neurons across the head ganglia (Hallinen et al., 2021; Ji et al., 2021; Nguyen et al., 2016), while extended quiescence broadly suppresses activity (Nichols et al., 2017). In Drosophila, locomotion or tethered flight increases the activity of neurons across multiple brain regions (Aimon et al., 2019; Mann et al., 2021) including identified neurons with roles in visual processing (Chiappe et al., 2010; Hindmarsh Sten et al., 2021; Kim et al., 2017a; Kim et al., 2015; Maimon et al., 2010; Strother et al., 2018; Suver et al., 2012), and motor control (Ache et al., 2019). During zebrafish swimming, whole-brain imaging has revealed broad engagement of neurons across the forebrain, midbrain, and hindbrain (Ahrens et al., 2012; Chen et al., 2018; Dunn et al., 2016; Lovett-Barron et al., 2020; Naumann et al., 2016), with widespread suppression of neurons during quiescence (Andalman et al., 2019; Mu et al., 2019). In behaving mice, locomotion and/or movement of the face or limbs influences the activity of neurons across multiple regions of dorsal neocortex (Allen et al., 2017b; Kauvar et al., 2020; Makino et al., 2017; Niell and Stryker, 2010) and subcortical areas (Musall et al., 2019; Steinmetz et al., 2019; Stringer et al., 2019), even including the axons of retinal ganglion cells (Liang et al., 2020; Schroder et al., 2020). Overall, an animal's brain displays dramatic and widespread neural activity changes during movement versus quiescence.

Despite the convenience of measuring locomotion alone, states of high arousal can occur without overt movements of the limbs or face (Lovett-Barron *et al.*, 2017; McGinley et al., 2015a; Reimer et al., 2014; Vinck et al., 2015). Therefore, it remains to be seen whether the neural dynamics in a rapidly moving animal reflect the internal state of the animal (McGinley *et al.*, 2015b), efference copy-like feedback of motor actions (Ji *et al.*, 2021; Kim *et al.*, 2017a; Kim *et al.*, 2015; Schneider et al., 2014), or a combination thereof (Liu and Dan, 2019; McGinley *et al.*, 2015b; Reimer *et al.*, 2014; Vinck *et al.*, 2015). In cases where large populations of neurons could be recorded simultaneously, these locomotion/ arousal-associated behavioral states are characterized by the evolution of a low-dimensional population state (Ahrens *et al.*, 2012; Ji *et al.*, 2021; Kato et al., 2015; Mu *et al.*, 2019;

Stringer *et al.*, 2019). Whether such states appear at the cellular level in larger primate brains remains presently unknown, but there is evidence for broadly synchronized brain regions in humans (Fox et al., 2005; Raichle, 2015).

In addition to locomotion-related arousal, need states such as hunger and thirst are also shown to modulate large-scale neural activity. Hunger influences multiple aspects of *Drosophila* behavior (Kim et al., 2017c), through modulation of olfactory neurons (Ko et al., 2015; Root et al., 2011), gustatory neurons (Inagaki et al., 2014b), motor-control neurons (Jourjine et al., 2016; Yu et al., 2016), and other central brain populations (Inagaki et al., 2012; Krashes et al., 2009; Park et al., 2016; Tsao et al., 2018; Yapici et al., 2016). In zebrafish larvae, food restriction biases fish towards hunting behavior (Johnson *et al.*, 2020), with hunger increasing the activity of serotonergic neurons in the raphe (Filosa et al., 2016) and caudal hypothalamus (Wee et al., 2019b), potentially by sensitizing visually responsive neurons in the optic tectum (Filosa *et al.*, 2016; Yokogawa et al., 2012). In mice, hunger can influence cue-evoked activity in association cortices, amygdala, and brainstem (Burgess et al., 2016; Calhoon *et al.*, 2018; Gong *et al.*, 2020; Livneh *et al.*, 2017; Livneh *et al.*, 2020; Lutas et al., 2019).

One particularly informative study (Allen *et al.*, 2019) examined the impact of thirst state on a mouse's performance in a water-motivated behavioral task. Using large-scale electrophysiological recordings from populations of neurons across dozens of brain regions, the authors found that the state of thirst was widely encoded as a low-dimensional population state. This state influences both spontaneous and cue-evoked neural activity – largely increasing the rates and durations of task-responsive neurons (Figure 3D). Notably, thirst-related dynamics across multiple brain regions – but not all – were reinstated by optogenetic activation of dehydration-sensitive neurons in the subfornical organ. This suggests that both natural and optogenetic induction of an internal state can influence the activity of neurons throughout the brain, but subtle differences in the set of influenced brain regions distinguish between the two conditions. Whether natural or optogenetically-evoked thirst states produce comparable subjective experiences for the animal, or are capable of modulating the same set of behaviors, is presently unclear.

As techniques for large-scale recording in freely-moving animals advance (Cong et al., 2017; Grover et al., 2020; Ji *et al.*, 2021; Juavinett et al., 2019; Kim et al., 2017b; Nguyen *et al.*, 2016; Steinmetz et al., 2021), we expect that investigators will find that other internal states also exert a brain-wide influence, including those that evolve over longer timescales (Hrvatin et al., 2020; Stern *et al.*, 2017) or whose classification is more complex, including parental behavior (Carcea et al., 2021; Kohl *et al.*, 2018; Marlin et al., 2015; Wu et al., 2014), emotional regulation (Anderson and Adolphs, 2014; Dolensek *et al.*, 2020), and the multiple effects of social deprivation (Anneser et al., 2020; Matthews et al., 2016; Tunbak et al., 2020; Zelikowsky et al., 2018).

It remains to be seen whether such brain-wide concerted activity patterns are important for the execution of state-dependent behavior, or are a mere consequence of shared activity across recurrently connected circuits that span multiple brain regions. This could be tested in future studies by independently manipulating state-dependent population activity in different

brain regions and measuring the effects on state-dependent behaviors and activity in other regions. To understand these mechanisms, better knowledge of how the cellular actions of neuromodulators collectively produce global brain state-dynamics is needed.

#### A central role for neuromodulation

Perhaps the largest unifying factor identified in the control of distinct internal states and their impact on behavior is the role of neuromodulators (Bargmann, 2012; Bargmann and Marder, 2013; Flavell *et al.*, 2013; Harris-Warrick and Marder, 1991; Kennedy et al., 2014; Marder, 2012; Nusbaum and Blitz, 2012; Taghert and Nitabach, 2012; Zelikowsky *et al.*, 2018).

Neuromodulators occupy an ideal position with respect to the control of internal states – they modulate synaptic and cellular function over long time scales due to their impact on biochemical signaling and ion channel function, they can titrate their effects via magnitude of modulator release, and they can act locally as well as send far-reaching diffuse signals across multiple brain regions (van den Pol, 2012). This makes them prime candidates for the flexible, scalable, and persistent control of behavior – key requirements for an internal state.

#### Foundational principles discovered in reduced invertebrate circuits

While much of this review focuses on the nervous systems of animals amenable to behavioral study of internal states, it is important to recognize that much of our understanding of neuromodulation derives from the study of invertebrate circuits in reduced preparations - including the stomatogastric ganglion of crustaceans, the swimming central pattern generator of the mollusc, the motor system of the leech, the abdominal and buccal ganglia of the sea slug *Aplysia*, and others (Bargmann, 2012; Bargmann and Marder, 2013; Getting, 1989; Harris-Warrick and Marder, 1991; Kristan and Calabrese, 1976; Marder, 2002; 2012; Marder and Calabrese, 1996; Marder and Thirumalai, 2002; Nusbaum and Blitz, 2012; Taghert and Nitabach, 2012). The experimental access of these circuits, often exhibiting complex and flexible rhythmic dynamics *in vitro*, enable detailed electrophysiological and biochemical analysis of functioning neural networks across states of experimentally-induced modulation.

Pioneering studies using these preparations have established that neuromodulators are capable of switching functional networks between different modes of population activity ((Dickinson et al., 1990; Eisen and Marder, 1984; Getting, 1989; Getting and Dekin, 1985; Nusbaum and Beenhakker, 2002; Nusbaum et al., 2001; Powell et al., 2021), through extrinsic and local sources of neuromodulation (Katz, 1998; Katz and Frost, 1995; 1996; Katz et al., 1994) that act upon membrane excitability and synaptic transmission (Katz *et al.*, 1994; Martin et al., 1997; Nadim and Bucher, 2014). These neuromodulators exert their effects on multiple neurons and networks in parallel (Brezina, 2010; Harris-Warrick and Johnson, 2010; Harris-Warrick and Marder, 1991; Marder, 2012; Schwarz et al., 1980; Taghert and Nitabach, 2012), and each neuron or synapse is subject to modulation by multiple sources (Flamm et al., 1987; Hempel et al., 1996; Kintos et al., 2016; Swensen and Marder, 2000; 2001).

While we cannot fully discuss the breadth and influence of this literature here, we would like to emphasize how its influence has greatly shaped subsequent work on state-dependent behavior and neuromodulation in larger animals. As we will discuss in the remainder of this section, these pioneering studies identified themes that are present across small and large circuits alike, and raise still-unanswered questions about how to interpret the complexity and behavioral significance of heavily modulated networks (Getting, 1989; Marder, 2012).

#### Neuromodulatory systems possess a Fan-In/Fan-Out organization

Most ascending neuromodulatory systems display a characteristic organization in which a relatively small group of neuromodulator-producing neurons receives diverse synaptic inputs and sends diffuse projections to many brain regions (Figure 4) (Ren et al., 2018; Saper et al., 2010; Weissbourd et al., 2014). This gives rise to a "fan-in" organization where signals converge onto the neuromodulator-producing neurons and a "fan-out" organization in which the modulators impact many downstream brain regions. This fan-out organization of neuromodulatory systems is observed at the anatomical level in diverse organisms (Figure 5A). For example, in *C. elegans* the serotonergic neuron NSM releases serotonin at non-synaptic neurosecretory terminals that are apposed to the nerve ring - the main neuropil of the worm's brain (Nelson and Colon-Ramos, 2013). In zebrafish, oxytocin neurons project from the hypothalamus to influence multiple regions across the forebrain, midbrain, brainstem, and spinal cord (Herget et al., 2017; Lovett-Barron et al., 2020; Wee et al., 2019a). In mice, multiple monoaminergic neuron types project across the brain (Ren et al., 2019; Schwarz et al., 2015). These are just a few of many examples. This overall organization likely allows neuromodulatory systems to encode the brain state's by integrating multiple inputs, and exert coordinated control by broadly influencing multiple brain regions simultaneously.

A notable alternative to this organization is local processing distributed across multiple sites, controlled by single (Zelikowsky et al., 2018a, see "Theme 1" below), or multiple neuropeptide systems. Such distributed effects could be far more prominent than is currently appreciated, driven by widespread expression of neuropeptides and receptors, which has been observed in *C. elegans* (Taylor et al., 2021) and in mammalian striatum (Castro and Bruchas, 2019) and neocortex (Smith et al., 2019). See Theme #1 below for more on this topic.

# Volume transmission allows neuromodulatory systems to signal diffusely and over long timescales

Another feature of neuromodulatory systems that may endow them with a specialized ability to control internal states is their action through volume transmission. Decades ago, electron microscopy studies of neurons that release biogenic amines, such as dopamine, serotonin, and norepinephrine, revealed that these cells often display putative active zones at non-synaptic varicosities along their axons (Calas et al., 1974; Descarries and Mechawar, 2000; Descarries et al., 1996). These observations, which have also been made for dense core vesicle release sites in neuropeptide-releasing neurons, suggest that these transmitters can be released extrasynaptically (Oti et al., 2021; Persoon et al., 2018; van de Bospoort et al., 2012). In the case of neuropeptides, release from dendrites has even been observed

(Ludwig and Leng, 2006). Many of these transmitters also function at classical synapses and the degree to which they act via synaptic versus extrasynaptic volume transmission varies by brain region (Moukhles et al., 1997). In invertebrate systems, extrasynaptic release sites for amines and neuropeptides are also widely observed (White et al., 1986). In addition, these transmitters can be released into circulating fluid, which allows them to act as neurohormones (Kravitz, 2000; Reiter et al., 2014; White *et al.*, 1986).

Extrasynaptic release of neuromodulators could allow these transmitters to diffuse and persist in brain tissue, which might allow for long timescale modulation of target cells. Indeed, the receptors and transporters for these transmitters are commonly localized microns or tens of microns away from active zones (Callado and Stamford, 2000; Liu et al., 2021). Measurements of extracellular amines and neuropeptides, via voltammetry and newer fluorescent sensors (Sabatini and Tian, 2020), support the view that neuromodulators persist in extracellular space for 100s of milliseconds to many seconds (Bunin and Wightman, 1998; Callado and Stamford, 2000; Park et al., 2011). Work in this area has been most extensive for dopamine and, while recent results support the idea that dopamine can act through volume transmission, the presence of dopamine at levels sufficient to activate its receptors likely only occurs over a micron away from an active zone during synchronous release from multiple nearby active zones (Beyene et al., 2019; Jan et al., 1979; Liu et al., 2021). Estimates of neuropeptide diffusion based on photo-uncaging suggest potentially longer-range diffusion (Banghart and Sabatini, 2012). Further studies using recently developed neuromodulator sensors will more precisely clarify these dynamics, which may be critical to internal state control.

#### Neuromodulators stably alter neuronal excitability to control persistent internal states

In addition to slow diffusion of the ligand, the long timescale action of neuromodulators is also thought to be due the fact that amines and neuropeptides primarily act through metabotropic receptors, which activate biochemical signaling pathways that remain active after receptor activation (Figure 5B, C). The activation of these pathways can modulate cellular excitability and a variety of other cellular processes. As described above, the effects of metabotropic signaling on neuronal activity have perhaps been best characterized in the stomatogastric ganglia of crustaceans, where metabotropic pathways converge onto a number of different currents to modulate neuronal excitability. However, classical neurotransmitters can also act through metabotropic receptors (Ringstad et al., 2009; Thompson and Lummis, 2006), so this feature does not fully distinguish the neuromodulatory systems from other neurotransmitters. Nevertheless, neuromodulator-dependent activation of metabotropic signaling has been directly linked to the generation of internal states.

Related to persistent internal states, neuromodulator-induced activation of metabotropic signaling is known to regulate persistent neural activity in many systems. For example, in the presence of a muscarinic agonist, current injection into mammalian layer V entorhinal neurons elicits a remarkably stable increase in firing rate that can occur in a graded manner (Egorov et al., 2002). In the presence of serotonin, spinal motoneurons display

bi-stable activity (Hounsgaard and Kiehn, 1989). In *Drosophila*, dopamine acting through the Dop1R2 receptor and downstream potassium channels can stably alter the excitability of the dorsal fan-shaped body neurons to control sleep (Pimentel et al., 2016). In the striatum, dopamine persistently elevates the excitability of D1 receptor-expressing striatal projection neurons (Lahiri and Bevan, 2020). Indeed, metabotropic regulation of firing modes appears to be a common property of neurons (Derjean et al., 2003). *In vivo* electrophysiological studies of thalamic and cortical contributions to arousal states also support a role for neuromodulatory systems in eliciting stable activity (McCormick, 1992; McCormick and Prince, 1986; Pape and McCormick, 1989; Steriade et al., 1993). Behavioral state-correlated activation of cholinergic and noradrenergic axons in cortex is associated with sustained depolarizations in pyramidal cells (Goard and Dan, 2009; Meir et al., 2018; Pinto et al., 2013; Polack et al., 2013). Overall, these studies provide evidence that neuromodulatory control of persistent neural activity contributes to the generation of internal states.

#### Neuromodulators stably alter biochemical signaling to control persistent internal states

Studies linking neuromodulator-induced biochemical signaling to internal states have been most extensive for the cAMP-PKA pathway. Fluorescent sensors of cAMP levels and PKA activation have revealed persistent increases in cAMP levels and downstream signaling with kinetics on the order of tens of seconds to minutes in freely-moving flies (Thornquist et al., 2021), and mice (Lee et al., 2019; Zhang et al., 2021). These kinetics have been tied to internal state generation in several organisms.

One example is the set of Corazonin neurons in Drosophila, a small group of neurons controlling the animal's drive to copulate. Graded accumulation of cAMP in these neurons over minutes during successive activity bouts can trigger a synchronous burst of network activity, or eruption, that changes the motivational state of the fly such that its copulation drive is reduced (Thornquist et al., 2021). Optogenetic elevation of cAMP levels in Corazonin neurons can elicit this state transition. Another example is from the zebrafish brainstem, where stable accumulation of evidence also occurs downstream of alpha-1B adrenergic receptors in radial glia, where noradrenaline release during successive futile actions stably increases glial calcium levels to elicit a transition to a passive behavioral state (Mu et al., 2019). Long-lasting activation of astrocytic signaling in mammalian circuits has also been linked to stable states of neural activity (Deemyad et al., 2018), suggesting that this may be a recurring mechanism for stable accumulation of persistent activity. Finally, a recent study of mating drive in male mice showed that stable increases in cAMP occur in MPOA neurons after transient hypothalamic dopamine release activated by a social encounter with a female (Zhang et al., 2021). This then triggers a stable state of motivation to mate, whose kinetics match cAMP kinetics in MPOA neurons. Together, these studies highlight how the timescale of biochemical signaling is closely linked to the persistence of internal states.

Other stable neuronal signaling pathways also contribute to behavioral state generation. Activation of the calcium-dependent protein kinase CaMKII in *Drosophila* Corazonin neurons delays a motivational state change that terminates copulation until 5–7 min after copulation begins (Thornquist et al., 2020). Interestingly, previous work has shown that

CaMKII activation initially requires elevated calcium levels, but the activation of the 12subunit CaMKII holoenzyme can be sustained in a calcium-independent manner through autophosphorylation of adjacent subunits, allowing for stable, minutes-long activation of the enzyme (Lisman et al., 2012; Miller and Kennedy, 1986). Sustained activation of CaMKII in Corazonin neurons detected through fluorescent reporter imaging was shown to have a causal effect on the timing of the motivational state transition of the fly. This work demonstrates how stable biochemical pathways within neurons can influence network activity and internal states.

#### Gene expression changes across internal states

While stable, activity-induced changes in gene expression are essential for lasting behavioral changes during long-term memory and circadian timing (Dubowy and Sehgal, 2017; Yap and Greenberg, 2018), the role of dynamic gene expression in persistent internal states is less well studied. However, changes in gene expression have been notably detected across feeding states. For example, feeding state-dependent changes in neuromodulator (Entchev et al., 2015) and chemoreceptor (Sengupta, 2013) expression in *C. elegans* have been linked to satiety-related behavioral changes. Similarly, food deprivation alters the expression of hundreds of genes in AGRP neurons of the hypothalamus (Henry et al., 2015). Gene expression changes in lateral hypothalamus are even associated with the onset of obesity over days (Rossi et al., 2019).

Gene expression changes have also been linked to other motivational drives, for example the drive to copulate in Drosophila. Abstinence from copulation elicits an increase in activation of the neural activity-dependent transcription factor CREB in a group of neurons that form a recurrent loop (Zhang et al., 2019). The stable expression of a CREB-induced potassium channel then influences mating behavior for hours to days after animals have mated and CREB activation has subsided. Given that activity-dependent transcription is a ubiquitous feature of neuronal gene expression and that it can reflect historical patterns of neural activity in a surprisingly precise manner (Brigidi et al., 2019), it may play a similar role in the control of other drive states. Given that these activity-dependent pathways are also known to regulate structural plasticity, future work may be aimed at examining whether internal states are accompanied by structural changes in neural circuits. Overall, the links between neuromodulator-induced biochemical signaling and internal state generation are now becoming apparent, but our understanding of this relationship is still in its infancy.

#### Emerging themes of internal state control across species

Despite substantial variability amongst internal states within an organism and across different organisms, there exists a striking commonality in how some of these states are organized in the brain. Indeed, recent studies have identified several examples of common neural mechanisms that contribute to internal state control.

#### Theme 1: Internal states influence multiple circuits and cell types in parallel

While the predominant view of internal states favors a "hub and spoke" type of "fan-out" mechanism (highlighted above), there is evidence for the control of internal states in a more distributed, parallel action manner. Here, we highlight a few key examples.

Above, we highlighted how neuromodulators can act locally within a given brain region to exert control over behavior. However, there is growing evidence that neuromodulators can exert their state-like control over behavior in a distributed manner across numerous brain regions simultaneously. For example, Zelikowsky and colleagues identified a role for the neuropeptide Tachykinin 2 (Tac2) in the control of an internal brain state produced by prolonged social isolation stress (Zelikowsky *et al.*, 2018). Using a multiplex approach employing a variety of loss-of-function techniques and testing multiple behaviors, the authors discovered that Tac2 signaling is necessary and sufficient for the effects of social isolation to produce enhanced aggression, persistent fear, and acute fear responses. Importantly, the authors found that each isolation-altered behavior was independently controlled by Tac2 signaling in distinct brain regions This "web-like" distributed, local circuit organization has also been shown to control additional states and systems.

One prominent example is the role of the neuropeptide pigment-dispersing factor (PDF) in the control of circadian rhythms. Indeed, PDF has been shown to coordinate the phase and amplitude of circadian rhythms through its action on separate populations of cells across the fly brain (Lin et al., 2004). Importantly, PDF operates in a distributed manner across the fly brain, providing unified and organized control over circadian rhythms in flies despite the unique effects that PDF exerts in a region-specific manner (Taghert and Nitabach, 2012). Local, distributed neuromodulation has also been recently studied in the context of rodent fear behavior, where disinhibitory interneurons in several neocortical regions have been found to be excited by local and afferent sources of the neuropeptide Gastrin-Releasing Peptide (GRP) (Melzer et al., 2021). In the auditory cortex, GRP receptor signaling facilitates auditory fear conditioning, and the role of GRP signaling in other regions remains to be investigated.

Collectively, these studies highlight the potential biological benefit of a dispersed internal state, wherein separate behaviors can be controlled via distinct brain regions, yet remain in concert with each other through overarching control by a single neuropeptide system. While it is highly likely that in such examples additional signaling molecules are co-released along with these neuropeptides (see Theme 2 below), the ability of a single neuropeptide to exert large-scale effects across the brain and behavior is nevertheless striking.

Recent work has also shown that single neuromodulators are capable of controlling distinct internal states in different contexts. For example, while Tac2 has been implicated in the control of the state produced by prolonged social isolation (see above), work by Andero and colleagues has also identified a role for Tac2 signaling in the CeA in the fear state produced by exposure to footshock (Andero et al., 2016; Andero et al., 2014). Similarly, while PDF has been implicated in the regulation of circadian rhythms (see above), additional work by Flavell and colleagues using genetic screens, quantitative behavioral analyses, and optogenetics also identified a role for PDF in the control of roaming behavior in worms

(Flavell *et al.*, 2013). This pattern of neuropeptidergic "multi-purposing" can be found in the identification of oxytocin in pair-wise bonding (Donaldson and Young, 2008; Froemke and Young, 2021; Insel and Young, 2001), but also maternal behavior (Marlin *et al.*, 2015), fear (Pisansky et al., 2017), and other states. Finally, in a series of seminal studies, Galanin<sup>+</sup> neurons in the medial preoptic area were identified in the control of parental behavior in both males and females (Kohl *et al.*, 2018; Kohl and Dulac, 2018; Wu *et al.*, 2014), while Galanin<sup>+</sup> neurons in the ventrolateral preotic area have been found to promote sleep and heat loss (Kroeger et al., 2018).

Overall, these examples highlight diversity in function and internal state control for single neuropeptides operating across the brain to control a single state, as well as the ability of a single neuropeptide to be "repurposed" to serve in the formation of multiple internal states. This diversity can range across brain regions and even species. Importantly, while it is tempting to assign one-to-one pairings between individual neuromodulators and internal states, this appears to be an oversimplification. In particular, neuromodulatory repurposing further reinforces the notion that neuromodulators – with their physiological properties, brain-wide networks, region-specificity, and slow-release, persistent signaling properties – are ideal candidates for the control of internal states and their effects on behavior.

#### Theme 2: Neuromodulators act in concert

Many of the studies discussed in this review highlight the functional role of individual cell types and neuromodulatory transmitters, suggesting that each of these neuromodulatory systems plays a unique role in whatever state or behavior was examined. This is unlikely to be the case. One of the most salient lessons from the study of small invertebrate circuits is that neurons and synapses are modulated by multiple substances (Getting 1989; Harris-Warrick and Marder, 1991; Marder, 2012), and their interactions produce emergent effects that are not easily predicted from the actions of one modulator alone (Flamm *et al.*, 1987; Hempel *et al.*, 1996; Kintos *et al.*, 2016; Swensen and Marder, 2000; 2001).

Why this discrepancy between the small-circuit literature and more recent studies of neuromodulatory systems? A possible reason may be the bias of common laboratory techniques. Modern studies of neuromodulation often use genetic model systems, such as those discussed extensively here (worms, flies, fish, mice), whose power comes from the specificity they afford: the ability to study a single genetically or anatomically-defined cell type, or analyze the actions of specific transmitters and receptors (Luo *et al.*, 2018; Sabatini and Tian, 2020). In contrast, classical studies in small invertebrate circuits primarily used bath-applied neuromodulatory transmitters and hormones, allowing for the study of multiple transmitter actions.

We have reason to believe, however, that an accounting for ubiquitous co-modulation will become more prominent in genetic model systems as well. For instance, in rodents, singlecell RNA sequencing has emphasized the fact that each cell expresses a large number of neuromodulatory receptors (Campbell et al., 2017; Henry *et al.*, 2015; Kim et al., 2019; Moffitt et al., 2018; Saunders et al., 2018; Smith *et al.*, 2019) and viral strategies allow investigators to control multiple independent cell types in the same animal (Luo *et al.*, 2018). Furthermore, recent studies combining live functional imaging with *post hoc* registration to

multiple gene expression markers (Bugeon et al., 2021; Lovett-Barron *et al.*, 2017; Lovett-Barron *et al.*, 2020; von Buchholtz et al., 2021; Xu *et al.*, 2020) provides the opportunity to image multiple genetically-defined cell types at once. In larval zebrafish, this approach has demonstrated that multiple neuromodulatory cell types are co-active during states of heightened alertness (Lovett-Barron et al., 2017), and many hypothalamic neuropeptide-producing cell types are co-active across various homeostatic threats (Lovett-Barron et al., 2020).

We believe that an appreciation of co-modulation will move the field away from the perspective of studying neural circuits as "labeled lines" – an approach so useful in the understanding of sensory systems and reflexes – and towards an understanding of modulated circuits as an emergent state produced by multiple interacting neuromodulatory effects (Getting 1989; Harris-Warrick and Marder, 1991; Marder, 2012).

#### Theme 3: State transitions engage mutually-exclusive neural populations

One common mechanism in the neural encoding of global brain states is the switching between largely mutually-exclusive populations of neurons that encode opposing states. This is observed across species and brain states, including well-studied examples of sleep-state switching in mammals (Saper *et al.*, 2010; Weber and Dan, 2016), zebrafish (Oikonomou and Prober, 2017), and invertebrates (Shafer and Keene, 2021) as well as mutually-exclusive populations of neurons encoding hunger states in the zebrafish hypothalamus (Wee *et al.*, 2019b), and distinct populations that encode separable internal states of social engagement in the mouse (Karigo *et al.*, 2021).

The distinction between roaming and dwelling has been studied across species, where distinct neural populations produce these opposing states: exploration of large spaces in search of resources ("roaming") versus exploiting local resources by staying in place ("dwelling"). In freely-moving C. elegans, the roaming-inducing neuropeptide PDF and dwelling-inducing monoamine serotonin (Flavell et al., 2013) recruit distinct populations of neurons that are active in a mutually-exclusive manner to promote each behavior (Ji et al., 2021) (Figure 6A). Of note, the neurons that generate these opposing neuromodulators mutually inhibit one another to generate this two-state system. Similarly, brain-wide imaging in freely-swimming zebrafish larvae (Kim et al., 2017b) also revealed a pattern of mutuallyexclusive populations across the midbrain, diencephalon, and brainstem that encode longlasting roaming and dwelling states during hunting behavior, as well as neurons that signal the transition from roaming/exploration to dwelling/feeding (Marques et al., 2020) (Figure 6B). As in *C. elegans*, serotonergic neurons were implicated in initiating dwelling states. Finally, population imaging in the mouse amygdala revealed that, across behavioral contexts, mutually-exclusive populations of neurons encode general states of roaming-like exploratory movement and dwelling-like defensive behaviors (Grundemann et al., 2019) (Figure 6C).

Together, these studies indicate that mutually-exclusive internal states can be encoded in the opposing activity of neuronal populations. However, these "flip-flop" dynamics may not generalize to internal states that exhibit continuous variation or interactions with other states that are not mutually-exclusive. The population dynamics and switching mechanisms underlying these states are not yet well explored.

#### Theme 4: State persistence through recurrent dynamics

It has long been recognized that neural circuits with recurrent excitation might be able to generate stable neural responses to transient inputs (Joshua and Lisberger, 2015). For example, transient motor signals that move the position of the eye are received by a recurrently-connected neural integrator circuit whose activity is persistently altered to maintain the position of the eye (Aksay et al., 2007; Miri et al., 2011). Recent work has now highlighted the importance of recurrent excitation for the generation of persistent internal states.

Studies of a neural circuit that controls behavioral states in female *Drosophila* provide new evidence that recurrent excitation is important for the generation of internal states. Activation of pC1 neurons in female flies elicits increased female receptivity to males and increased shoving and chasing, even several minutes after the optogenetic stimulus has terminated (Deutsch *et al.*, 2020). Distinct subsets of pC1 neurons control female receptivity versus shoving and chasing behaviors. Interestingly, a brain-wide imaging approach revealed that activation of the pC1d/e neurons that control shoving and chasing induced persistent activity in many downstream brain regions, in addition to pC1 neurons themselves. A connectomic analysis showed that pC1 neurons are part of a recurrently connected neural circuit, with prominent reciprocal connections to aIPg-b and aIPg-c cells, which are also interconnected with one another. As all of these cell types are excitatory (Schretter et al., 2020), this suggests that pC1 is a functionally important node in a recurrently connected circuit that elicits a persistent behavioral state.

In male Drosophila, activation of a stable, recurrently active circuit also underlies behavioral state generation. Activation of the P1 interneurons elicits a minutes-long internal state that consists of elevated courtship and aggression (Clowney *et al.*, 2015; Hoopfer et al., 2015). While P1 neurons are not persistently active during this state, a group of downstream neurons, named pCD neurons, exhibit long-lasting activation during this internal state (Figure 7A) (Jung *et al.*, 2020). Activity in these neurons is required for stable behavioral changes during the P1-induced state and transient inactivation of pCD neurons attenuates their persistent neural response to P1 activation, providing evidence that continued pCD activity supports its own persistence. Transient inactivation of pCD neurons also suppresses persistent aggressive behavior elicited by recent exposure to a female fly. This study highlights how neural circuits with recurrent excitation can maintain a persistent internal state.

Studies in mammals have also implicated recurrent connectivity in the control of internal states. Activation of VMHdmSF1 neurons in the ventromedial hypothalamus can elicit a state of fear or anxiety (Kunwar et al., 2015). As a group, the VMHdmSF1 neurons show persistent activation in response to social sensory cues that can evoke an anxiety state (Kennedy et al., 2020). However, the dynamics of the neurons within this population vary, with some neurons displaying immediate onset activation and others ramping slowly. Moreover, neurons in the population respond differently to different social cues. Several computational models were constructed to determine whether they could recapitulate features of the population activity. Interestingly, only the models that included recurrent connectivity and neuromodulation were able to do so, suggesting that recurrent connectivity

and neuromodulation may co-occur in this circuit to support stable population dynamics (Figure 7B). It is worth noting that there is an additional similarity between P1 interneurons and VMH neurons, which is that they can both induce different behavioral states in different sensory contexts. This specific topic has been reviewed previously in Anderson, 2016.

While we note examples here of state persistence driven by recurrent circuits, persistence can also be achieved by neuromodulatory control of cellular excitability (as discussed above). It is not well understood whether these mechanisms are interdependent or used in different cases to achieve similar outcomes depending on the contexts, circuits, or timescales involved.

#### Conclusions

In this review, we have discussed our current understanding of internal states: how they are defined, measured, generated by neurons, as well as how they affect the brain and behavior. Building upon the insights from many other authoritative reviews about internal states (Anderson, 2016; Bargmann, 2012; Bargmann and Marder, 2013; Getting, 1989, etc.; Lee and Dan, 2012; Marder, 2012; McCormick *et al.*, 2020; McGinley *et al.*, 2015a; Taghert and Nitabach, 2012; Tye, 2018, etc.), here we have emphasized advances in the classification of internal states, the insights from studying brain-wide populations, and some of the many biological mechanisms through which neuromodulators can influence states. Importantly, we have emphasized common principles found across model species.

While the field has made enormous progress, many fundamental questions about internal states and their neural basis remain unanswered or completely unexplored. How do sensorimotor circuits integrate state-relevant information to drive adaptive behavioral responses? To what extent do neuromodulators have unique versus redundant effects? Are brain-wide dynamics required for the expression of states or just a consequence of a massively interconnected brain? Why are some states controlled by a handful of neurons while others are controlled by neurons distributed across multiple brain regions?

As the field resolves these mechanistic questions, it may be important to reflect on the challenges of defining internal states. How do different co-occurring states interact with each other, and would it be more useful in certain instances to simply refer to the animal's overall state? Can states always be inferred from behavior and/or physiology? When do measurements of the brain, behavior, and physiology reflect the same underlying state and when do they reveal unexpected distinctions? Is there a true distinction between motor actions, sequences of motor actions, and states, or does behavior simply unfold along a continuum of timescales? Can behavior in natural environments be adaptive in the absence of long timescale state organization?

One key issue regarding the definition of internal states is their degree of independence. How do we know that fear represents a unique internal state, distinct from others such as anxiety? Is the ability to distinguish such states dependent on the tools we use for measuring their observable output? Would we be able to further splinter internal states into smaller sub-states if we had better tools? How does selection of model organism affect our ability to

Page 23

isolate and define an internal state? Given the wide variability in model organisms as well as experimental approaches, would we benefit from a definition of internal states as they pertain to biological relevance and their importance to survival?

These questions and more can be addressed using the emerging methodological approaches discussed herein, including more rigorous quantification of states using integrated datasets and ML approaches, precise observation and control of electrical and biochemical activity across entire nervous systems, and better theoretical frameworks understanding the utility of internal states.

As with any search for common principles in biology, this field of neuroscience will benefit greatly from studying an expanded set of animal species, challenging animals with more natural and varied behavioral conditions, and welcoming scientists to approach these questions with diverse views, expertise, and experiences.

#### Acknowledgements

We thank Julia Kuhl for artwork, and our labs for fruitful discussion.

SWF is supported by the JPB Foundation, a Sloan Research Fellowship, a McKnight Scholars Award, a Brain Research Foundation Seed Grant, the NSF (award #1845663), and the NIH (R01GM135413, R01NS104892).

NG is supported by the Max-Planck Society and the European Research Council (ERC) under the European Union's Horizon 2020 research and innovation programme (ERC-2017-STG, grant agreement n° 758448 to N.G.).

MLB is supported by a Searle Scholars Award, a Sloan Research Fellowship, a Klingenstein-Simons Fellowship, a Packard Foundation Fellowship, and the NIMH (R00MH112840).

MZ is supported by a Sloan Research Fellowship, a LOREAL FWIS award, a Klingenstein-Simons Fellowship, a Whitehall Fellowship, and the NIMH (R00MH108734).

#### References

- Ache JM, Namiki S, Lee A, Branson K, and Card GM (2019). State-dependent decoupling of sensory and motor circuits underlies behavioral flexibility in Drosophila. Nat Neurosci 22, 1132–1139. 10.1038/s41593-019-0413-4. [PubMed: 31182867]
- Adolphs R (2008). Fear, faces, and the human amygdala. Current opinion in neurobiology 18, 166– 172. 10.1016/j.conb.2008.06.006. [PubMed: 18655833]
- Adolphs R, and Anderson D (2013). Social and emotional neuroscience. Current opinion in neurobiology 23, 291–293. 10.1016/j.conb.2013.04.011. [PubMed: 23669616]
- Ahrens MB, and Engert F (2015). Large-scale imaging in small brains. Curr Opin Neurobiol 32, 78–86. 10.1016/j.conb.2015.01.007. [PubMed: 25636154]
- Ahrens MB, Li JM, Orger MB, Robson DN, Schier AF, Engert F, and Portugues R (2012). Brainwide neuronal dynamics during motor adaptation in zebrafish. Nature 485, 471–477. 10.1038/ nature11057. [PubMed: 22622571]
- Aimon S, Katsuki T, Jia T, Grosenick L, Broxton M, Deisseroth K, Sejnowski TJ, and Greenspan RJ (2019). Fast near-whole-brain imaging in adult Drosophila during responses to stimuli and behavior. PLoS Biol 17, e2006732. 10.1371/journal.pbio.2006732. [PubMed: 30768592]
- Aksay E, Olasagasti I, Mensh BD, Baker R, Goldman MS, and Tank DW (2007). Functional dissection of circuitry in a neural integrator. Nature neuroscience 10, 494–504. 10.1038/nn1877. [PubMed: 17369822]
- Alhadeff AL, Su Z, Hernandez E, Klima ML, Phillips SZ, Holland RA, Guo C, Hantman AW, De Jonghe BC, and Betley JN (2018). A Neural Circuit for the Suppression of Pain by a Competing Need State. Cell 173, 140–152 e115. 10.1016/j.cell.2018.02.057. [PubMed: 29570993]

- Allen WE, Chen MZ, Pichamoorthy N, Tien RH, Pachitariu M, Luo L, and Deisseroth K (2019). Thirst regulates motivated behavior through modulation of brainwide neural population dynamics. Science 364, 253. 10.1126/science.aav3932. [PubMed: 30948440]
- Allen WE, DeNardo LA, Chen MZ, Liu CD, Loh KM, Fenno LE, Ramakrishnan C, Deisseroth K, and Luo L (2017a). Thirst-associated preoptic neurons encode an aversive motivational drive. Science 357, 1149–1155. 10.1126/science.aan6747. [PubMed: 28912243]
- Allen WE, Kauvar IV, Chen MZ, Richman EB, Yang SJ, Chan K, Gradinaru V, Deverman BE, Luo L, and Deisseroth K (2017b). Global Representations of Goal-Directed Behavior in Distinct Cell Types of Mouse Neocortex. Neuron 94, 891–907 e896. 10.1016/j.neuron.2017.04.017. [PubMed: 28521139]
- Andalman AS, Burns VM, Lovett-Barron M, Broxton M, Poole B, Yang SJ, Grosenick L, Lerner TN, Chen R, Benster T, et al. (2019). Neuronal Dynamics Regulating Brain and Behavioral State Transitions. Cell 177, 970–985 e920. 10.1016/j.cell.2019.02.037. [PubMed: 31031000]
- Andero R, Daniel S, Guo JD, Bruner RC, Seth S, Marvar PJ, Rainnie D, and Ressler KJ (2016). Amygdala-Dependent Molecular Mechanisms of the Tac2 Pathway in Fear Learning. Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology 41, 2714–2722. 10.1038/npp.2016.77. [PubMed: 27238620]
- Andero R, Dias BG, and Ressler KJ (2014). A role for Tac2, NkB, and Nk3 receptor in normal and dysregulated fear memory consolidation. Neuron 83, 444–454. 10.1016/j.neuron.2014.05.028. [PubMed: 24976214]
- Anderson DJ (2016). Circuit modules linking internal states and social behaviour in flies and mice. Nature reviews. Neuroscience 17, 692–704. 10.1038/nrn.2016.125. [PubMed: 27752072]
- Anderson DJ, and Adolphs R (2014). A Framework for Studying Emotions across Species. Cell 157, 187–200. 10.1016/j.cell.2014.03.003. [PubMed: 24679535]
- Anneser L, Alcantara IC, Gemmer A, Mirkes K, Ryu S, and Schuman EM (2020). The neuropeptide Pth2 dynamically senses others via mechanosensation. Nature 588, 653–657. 10.1038/s41586-020-2988-z. [PubMed: 33268890]
- Aponte Y, Atasoy D, and Sternson SM (2011). AGRP neurons are sufficient to orchestrate feeding behavior rapidly and without training. Nat Neurosci 14, 351–355. 10.1038/nn.2739. [PubMed: 21209617]
- Asahina K, Watanabe K, Duistermars BJ, Hoopfer E, Gonzalez CR, Eyjolfsdottir EA, Perona P, and Anderson DJ (2014). Tachykinin-expressing neurons control male-specific aggressive arousal in Drosophila. Cell 156, 221–235. 10.1016/j.cell.2013.11.045. [PubMed: 24439378]
- Atasoy D, Betley JN, Su HH, and Sternson SM (2012). Deconstruction of a neural circuit for hunger. Nature 488, 172–177. 10.1038/nature11270. [PubMed: 22801496]
- Augustine V, Gokce SK, Lee S, Wang B, Davidson TJ, Reimann F, Gribble F, Deisseroth K, Lois C, and Oka Y (2018). Hierarchical neural architecture underlying thirst regulation. Nature 555, 204–209. 10.1038/nature25488. [PubMed: 29489747]
- Banghart MR, and Sabatini BL (2012). Photoactivatable neuropeptides for spatiotemporally precise delivery of opioids in neural tissue. Neuron 73, 249–259. 10.1016/j.neuron.2011.11.016. [PubMed: 22284180]
- Bargmann CI (2012). Beyond the connectome: how neuromodulators shape neural circuits. BioEssays : news and reviews in molecular, cellular and developmental biology 34, 458–465. 10.1002/bies.201100185. [PubMed: 22396302]
- Bargmann CI, and Marder E (2013). From the connectome to brain function. Nature methods 10, 483–490. [PubMed: 23866325]
- Bath DE, Stowers JR, Hormann D, Poehlmann A, Dickson BJ, and Straw AD (2014). FlyMAD: rapid thermogenetic control of neuronal activity in freely walking Drosophila. Nat Methods 11, 756–762. 10.1038/nmeth.2973. [PubMed: 24859752]
- Ben-Shaul Y (2017). OptiMouse: a comprehensive open source program for reliable detection and analysis of mouse body and nose positions. BMC Biol 15, 41. 10.1186/s12915-017-0377-3. [PubMed: 28506280]
- Berman GJ (2018). Measuring behavior across scales. BMC Biol 16, 23. 10.1186/s12915-018-0494-7. [PubMed: 29475451]

- Berman GJ, Bialek W, and Shaevitz JW (2016). Predictability and hierarchy in Drosophila behavior. Proc Natl Acad Sci U S A 113, 11943–11948. 10.1073/pnas.1607601113. [PubMed: 27702892]
- Berman GJ, Choi DM, Bialek W, and Shaevitz JW (2014). Mapping the stereotyped behaviour of freely moving fruit flies. J R Soc Interface 11. 10.1098/rsif.2014.0672.
- Berrios J, Li C, Madara JC, Garfield AS, Steger JS, Krashes MJ, and Lowell BB (2021). Food cue regulation of AGRP hunger neurons guides learning. Nature 595, 695–700. 10.1038/ s41586-021-03729-3. [PubMed: 34262177]
- Betley JN, Cao ZF, Ritola KD, and Sternson SM (2013). Parallel, redundant circuit organization for homeostatic control of feeding behavior. Cell 155, 1337–1350. 10.1016/j.cell.2013.11.002. [PubMed: 24315102]
- Betley JN, Xu S, Cao ZFH, Gong R, Magnus CJ, Yu Y, and Sternson SM (2015). Neurons for hunger and thirst transmit a negative-valence teaching signal. Nature 521, 180–185. 10.1038/nature14416. [PubMed: 25915020]
- Beyene AG, Delevich K, Del Bonis-O'Donnell JT, Piekarski DJ, Lin WC, Thomas AW, Yang SJ, Kosillo P, Yang D, Prounis GS, et al. (2019). Imaging striatal dopamine release using a nongenetically encoded near infrared fluorescent catecholamine nanosensor. Sci Adv 5, eaaw3108. 10.1126/sciadv.aaw3108. [PubMed: 31309147]
- Bohnslav JP, Wimalasena NK, Clausing KJ, Dai YY, Yarmolinsky DA, Cruz T, Kashlan AD, Chiappe ME, Orefice LL, Woolf CJ, and Harvey CD (2021). DeepEthogram, a machine learning pipeline for supervised behavior classification from raw pixels. Elife 10. 10.7554/eLife.63377.
- Bolanos LA, Xiao D, Ford NL, LeDue JM, Gupta PK, Doebeli C, Hu H, Rhodin H, and Murphy TH (2021). A three-dimensional virtual mouse generates synthetic training data for behavioral analysis. Nat Methods 18, 378–381. 10.1038/s41592-021-01103-9. [PubMed: 33820989]
- Bolles RC (1967). Theory of Motivation (Harper and Row).
- Branson K, Robie AA, Bender J, Perona P, and Dickinson MH (2009). High-throughput ethomics in large groups of Drosophila. Nat Methods 6, 451–457. 10.1038/nmeth.1328. [PubMed: 19412169]
- Brezina V (2010). Beyond the wiring diagram: signalling through complex neuromodulator networks. Philos Trans R Soc Lond B Biol Sci 365, 2363–2374. 10.1098/rstb.2010.0105. [PubMed: 20603357]
- Brigidi GS, Hayes MGB, Delos Santos NP, Hartzell AL, Texari L, Lin PA, Bartlett A, Ecker JR, Benner C, Heinz S, and Bloodgood BL (2019). Genomic Decoding of Neuronal Depolarization by Stimulus-Specific NPAS4 Heterodimers. Cell 179, 373–391 e327. 10.1016/j.cell.2019.09.004. [PubMed: 31585079]
- Bugeon S, Duffield J, Dipoppa M, Ritoux A, Prankerd I, Nicolout-sopoulos D, Orme D, Shinn M, Peng H, Forrest H, et al. (2021). A transcriptomic axis predicts state modulation of cortical interneurons. bioRxiv, 2021.2010.2024.465600. 10.1101/2021.10.24.465600.
- Bunin MA, and Wightman RM (1998). Quantitative evaluation of 5-hydroxytryptamine (serotonin) neuronal release and uptake: an investigation of extrasynaptic transmission. The Journal of neuroscience : the official journal of the Society for Neuroscience 18, 4854–4860. [PubMed: 9634551]
- Burgess CR, Ramesh RN, Sugden AU, Levandowski KM, Minnig MA, Fenselau H, Lowell BB, and Andermann ML (2016). Hunger-Dependent Enhancement of Food Cue Responses in Mouse Postrhinal Cortex and Lateral Amygdala. Neuron 91, 1154–1169. 10.1016/j.neuron.2016.07.032. [PubMed: 27523426]
- Burnett CJ, Funderburk SC, Navarrete J, Sabol A, Liang-Guallpa J, Desrochers TM, and Krashes MJ (2019). Need-based prioritization of behavior. Elife 8. 10.7554/eLife.44527.
- Burnett CJ, Li C, Webber E, Tsaousidou E, Xue SY, Bruning JC, and Krashes MJ (2016). Hunger-Driven Motivational State Competition. Neuron 92, 187–201. 10.1016/j.neuron.2016.08.032. [PubMed: 27693254]
- Calas A, Alonso G, Arnauld E, and Vincent JD (1974). Demonstration of indolaminergic fibres in the media eminence of the duck, rat and monkey. Nature 250, 241–243. 10.1038/250241a0. [PubMed: 4211906]

- Calhoon GG, Sutton AK, Chang C-J, Libster AM, Glober GF, Lévêque CL, Murphy GD, Namburi P, Leppla CA, Siciliano CA, et al. (2018). Acute Food Deprivation Rapidly Modifies Valence-Coding Microcircuits in the Amygdala. bioRxiv, 285189. 10.1101/285189.
- Calhoun AJ, Pillow JW, and Murthy M (2019). Unsupervised identification of the internal states that shape natural behavior. Nat Neurosci 22, 2040–2049. 10.1038/s41593-019-0533-x. [PubMed: 31768056]
- Callado LF, and Stamford JA (2000). Spatiotemporal interaction of alpha(2) autoreceptors and noradrenaline transporters in the rat locus coeruleus: implications for volume transmission. J Neurochem 74, 2350–2358. 10.1046/j.1471-4159.2000.0742350.x. [PubMed: 10820195]
- Campbell JN, Macosko EZ, Fenselau H, Pers TH, Lyubetskaya A, Tenen D, Goldman M, Verstegen AM, Resch JM, McCarroll SA, et al. (2017). A molecular census of arcuate hypothalamus and median eminence cell types. Nat Neurosci 20, 484–496. 10.1038/nn.4495. [PubMed: 28166221]
- Carcea I, Caraballo NL, Marlin BJ, Ooyama R, Riceberg JS, Mendoza Navarro JM, Opendak M, Diaz VE, Schuster L, Alvarado Torres MI, et al. (2021). Oxytocin neurons enable social transmission of maternal behaviour. Nature 596, 553–557. 10.1038/s41586-021-03814-7. [PubMed: 34381215]
- Castro DC, and Bruchas MR (2019). A Motivational and Neuropeptidergic Hub: Anatomical and Functional Diversity within the Nucleus Accumbens Shell. Neuron 102, 529–552. 10.1016/ j.neuron.2019.03.003. [PubMed: 31071288]
- Cermak N, Yu SK, Clark R, Huang YC, Baskoylu SN, and Flavell SW (2020). Whole-organism behavioral profiling reveals a role for dopamine in state-dependent motor program coupling in C. elegans. Elife 9. 10.7554/eLife.57093.
- Chen X, Mu Y, Hu Y, Kuan AT, Nikitchenko M, Randlett O, Chen AB, Gavornik JP, Sompolinsky H, Engert F, and Ahrens MB (2018). Brain-wide Organization of Neuronal Activity and Convergent Sensorimotor Transformations in Larval Zebrafish. Neuron 100, 876–890 e875. 10.1016/j.neuron.2018.09.042. [PubMed: 30473013]
- Chen Y, Essner RA, Kosar S, Miller OH, Lin YC, Mesgarzadeh S, and Knight ZA (2019). Sustained NPY signaling enables AgRP neurons to drive feeding. Elife 8. 10.7554/eLife.46348.
- Chen Y, Lin YC, Kuo TW, and Knight ZA (2015). Sensory detection of food rapidly modulates arcuate feeding circuits. Cell 160, 829–841. 10.1016/j.cell.2015.01.033. [PubMed: 25703096]
- Chen Y, Lin YC, Zimmerman CA, Essner RA, and Knight ZA (2016). Hunger neurons drive feeding through a sustained, positive reinforcement signal. Elife 5. 10.7554/eLife.18640.
- Chiappe ME, Seelig JD, Reiser MB, and Jayaraman V (2010). Walking modulates speed sensitivity in Drosophila motion vision. Curr Biol 20, 1470–1475. 10.1016/j.cub.2010.06.072. [PubMed: 20655222]
- Churgin MA, McCloskey RJ, Peters E, and Fang-Yen C (2017). Antagonistic Serotonergic and Octopaminergic Neural Circuits Mediate Food-Dependent Locomotory Behavior in Caenorhabditis elegans. J Neurosci 37, 7811–7823. 10.1523/JNEUROSCI.2636-16.2017. [PubMed: 28698386]
- Ciocchi S, Herry C, Grenier F, Wolff SB, Letzkus JJ, Vlachos I, Ehrlich I, Sprengel R, Deisseroth K, Stadler MB, et al. (2010). Encoding of conditioned fear in central amygdala inhibitory circuits. Nature 468, 277–282. 10.1038/nature09559. [PubMed: 21068837]
- Clowney EJ, Iguchi S, Bussell JJ, Scheer E, and Ruta V (2015). Multimodal Chemosensory Circuits Controlling Male Courtship in Drosophila. Neuron 87, 1036–1049. 10.1016/j.neuron.2015.07.025. [PubMed: 26279475]
- Coddington LT, and Dudman JT (2018). The timing of action determines reward prediction signals in identified midbrain dopamine neurons. Nature neuroscience 21, 1563–1573. 10.1038/ s41593-018-0245-7. [PubMed: 30323275]
- Cong L, Wang Z, Chai Y, Hang W, Shang C, Yang W, Bai L, Du J, Wang K, and Wen Q (2017). Rapid whole brain imaging of neural activity in freely behaving larval zebrafish (Danio rerio). Elife 6. 10.7554/eLife.28158.
- Darwin C (1872). The Expressions of the Emotions in Man and Animals (University of Chicago Press).
- Datta SR, Anderson DJ, Branson K, Perona P, and Leifer A (2019). Computational Neuroethology: A Call to Action. Neuron 104, 11–24. 10.1016/j.neuron.2019.09.038. [PubMed: 31600508]

- Deemyad T, Luthi J, and Spruston N (2018). Astrocytes integrate and drive action potential firing in inhibitory subnetworks. Nat Commun 9, 4336. 10.1038/s41467-018-06338-3. [PubMed: 30337521]
- Derjean D, Bertrand S, Le Masson G, Landry M, Morisset V, and Nagy F (2003). Dynamic balance of metabotropic inputs causes dorsal horn neurons to switch functional states. Nature neuroscience 6, 274–281. 10.1038/nn1016. [PubMed: 12592405]
- Descarries L, and Mechawar N (2000). Ultrastructural evidence for diffuse transmission by monoamine and acetylcholine neurons of the central nervous system. Prog Brain Res 125, 27–47. 10.1016/S0079-6123(00)25005-X. [PubMed: 11098652]
- Descarries L, Watkins KC, Garcia S, Bosler O, and Doucet G (1996). Dual character, asynaptic and synaptic, of the dopamine innervation in adult rat neostriatum: a quantitative autoradiographic and immunocytochemical analysis. J Comp Neurol 375, 167–186. 10.1002/ (SICI)1096-9861(19961111)375:2<167::AID-CNE1>3.0.CO;2-0. [PubMed: 8915824]
- Deutsch D, Pacheco D, Encarnacion-Rivera L, Pereira T, Fathy R, Clemens J, Girardin C, Calhoun A, Ireland E, Burke A, et al. (2020). The neural basis for a persistent internal state in Drosophila females. Elife 9. 10.7554/eLife.59502.
- Dickinson PS, Mecsas C, and Marder E (1990). Neuropeptide fusion of two motor-pattern generator circuits. Nature 344, 155–158. 10.1038/344155a0. [PubMed: 2308633]
- Dolensek N, Gehrlach DA, Klein AS, and Gogolla N (2020). Facial expressions of emotion states and their neuronal correlates in mice. Science 368, 89–94. 10.1126/science.aaz9468. [PubMed: 32241948]
- Donaldson ZR, and Young LJ (2008). Oxytocin, vasopressin, and the neurogenetics of sociality. Science 322, 900–904. 10.1126/science.1158668. [PubMed: 18988842]
- Dubowy C, and Sehgal A (2017). Circadian Rhythms and Sleep in Drosophila melanogaster. Genetics 205, 1373–1397. 10.1534/genetics.115.185157. [PubMed: 28360128]
- Duistermars BJ, Pfeiffer BD, Hoopfer ED, and Anderson DJ (2018). A Brain Module for Scalable Control of Complex, Multi-motor Threat Displays. Neuron 100, 1474–1490 e1474. 10.1016/ j.neuron.2018.10.027. [PubMed: 30415997]
- Dukes D, Abrams K, Adolphs R, Ahmed ME, Beatty A, Berridge KC, Broomhall S, Brosch T, Campos JJ, Clay Z, et al. (2021). The rise of affectivism. Nat Hum Behav. 10.1038/ s41562-021-01130-8.
- Dunn TW, Marshall JD, Severson KS, Aldarondo DE, Hildebrand DGC, Chettih SN, Wang WL, Gellis AJ, Carlson DE, Aronov D, et al. (2021). Geometric deep learning enables 3D kinematic profiling across species and environments. Nat Methods 18, 564–573. 10.1038/s41592-021-01106-6. [PubMed: 33875887]
- Dunn TW, Mu Y, Narayan S, Randlett O, Naumann EA, Yang CT, Schier AF, Freeman J, Engert F, and Ahrens MB (2016). Brain-wide mapping of neural activity controlling zebrafish exploratory locomotion. Elife 5, e12741. 10.7554/eLife.12741. [PubMed: 27003593]
- Egorov AV, Hamam BN, Fransen E, Hasselmo ME, and Alonso AA (2002). Graded persistent activity in entorhinal cortex neurons. Nature 420, 173–178. 10.1038/nature01171. [PubMed: 12432392]
- Eiselt AK, Chen S, Chen J, Arnold J, Kim T, Pachitariu M, and Sternson SM (2021). Hunger or thirst state uncertainty is resolved by outcome evaluation in medial prefrontal cortex to guide decisionmaking. Nature neuroscience 24, 907–912. 10.1038/s41593-021-00850-4. [PubMed: 33972802]
- Eisen JS, and Marder E (1984). A mechanism for production of phase shifts in a pattern generator. J Neurophysiol 51, 1375–1393. 10.1152/jn.1984.51.6.1375. [PubMed: 6145759]
- Engel TA, and Steinmetz NA (2019). New perspectives on dimensionality and variability from large-scale cortical dynamics. Curr Opin Neurobiol 58, 181–190. 10.1016/j.conb.2019.09.003. [PubMed: 31585331]
- Entchev EV, Patel DS, Zhan M, Steele AJ, Lu H, and Ch'ng Q (2015). A gene-expression-based neural code for food abundance that modulates lifespan. eLife 4, e06259. 10.7554/eLife.06259. [PubMed: 25962853]
- Essner RA, Smith AG, Jamnik AA, Ryba AR, Trutner ZD, and Carter ME (2017). AgRP Neurons Can Increase Food Intake during Conditions of Appetite Suppression and Inhibit Anorexigenic

Parabrachial Neurons. J Neurosci 37, 8678–8687. 10.1523/JNEUROSCI.0798-17.2017. [PubMed: 28821663]

- Falkner AL, Dollar P, Perona P, Anderson DJ, and Lin D (2014). Decoding ventromedial hypothalamic neural activity during male mouse aggression. J Neurosci 34, 5971–5984. 10.1523/ JNEUROSCI.5109-13.2014. [PubMed: 24760856]
- Falkner AL, Grosenick L, Davidson TJ, Deisseroth K, and Lin D (2016). Hypothalamic control of male aggression-seeking behavior. Nat Neurosci 19, 596–604. 10.1038/nn.4264. [PubMed: 26950005]
- Falkner AL, Wei D, Song A, Watsek LW, Chen I, Chen P, Feng JE, and Lin D (2020). Hierarchical Representations of Aggression in a Hypothalamic-Midbrain Circuit. Neuron 106, 637–648 e636. 10.1016/j.neuron.2020.02.014. [PubMed: 32164875]
- Fanselow MS (2018). The Role of Learning in Threat Imminence and Defensive Behaviors. Curr Opin Behav Sci 24, 44–49. 10.1016/j.cobeha.2018.03.003. [PubMed: 30276224]
- Fanselow MS, and Bolles RC (1979). Naloxone and shock-elicited freezing in the rat. J Comp Physiol Psychol 93, 736–744. 10.1037/h0077609. [PubMed: 479405]
- Fanselow MS, Hoffman AN, and Zhuravka I (2019). Timing and the transition between modes in the defensive behavior system. Behav Processes 166, 103890. 10.1016/j.beproc.2019.103890. [PubMed: 31254627]
- Fanselow MS, and Lester LS (1988). A functional behavioristic approach to aversively motivated behavior: predatory imminence as a determinant of the topography of defensive behavior. In Evolution and Learning, Bolles RC, and Beecher MD, eds. pp. 185–211.
- Fanselow MS, and Pennington ZT (2018). A return to the psychiatric dark ages with a two-system framework for fear. Behav Res Ther 100, 24–29. 10.1016/j.brat.2017.10.012. [PubMed: 29128585]
- Filosa A, Barker AJ, Dal Maschio M, and Baier H (2016). Feeding State Modulates Behavioral Choice and Processing of Prey Stimuli in the Zebrafish Tectum. Neuron 90, 596–608. 10.1016/ j.neuron.2016.03.014. [PubMed: 27146269]
- Flamm RE, Fickbohm D, and Harris-Warrick RM (1987). cAMP elevation modulates physiological activity of pyloric neurons in the lobster stomatogastric ganglion. J Neurophysiol 58, 1370–1386. 10.1152/jn.1987.58.6.1370. [PubMed: 2449516]
- Flavell SW, Pokala N, Macosko EZ, Albrecht DR, Larsch J, and Bargmann CI (2013). Serotonin and the neuropeptide PDF initiate and extend opposing behavioral states in C. elegans. Cell 154, 1023–1035. 10.1016/j.cell.2013.08.001. [PubMed: 23972393]
- Flavell SW, Raizen DM, and You YJ (2020). Behavioral States. Genetics 216, 315–332. 10.1534/ genetics.120.303539. [PubMed: 33023930]
- Forkosh O, Karamihalev S, Roeh S, Alon U, Anpilov S, Touma C, Nussbaumer M, Flachskamm C, Kaplick PM, Shemesh Y, and Chen A (2019). Identity domains capture individual differences from across the behavioral repertoire. Nat Neurosci 22, 2023–2028. 10.1038/s41593-019-0516-y. [PubMed: 31686022]
- Fox MD, Snyder AZ, Vincent JL, Corbetta M, Van Essen DC, and Raichle ME (2005). The human brain is intrinsically organized into dynamic, anticorrelated functional networks. Proc Natl Acad Sci U S A 102, 9673–9678. 10.1073/pnas.0504136102. [PubMed: 15976020]
- Froemke RC, and Young LJ (2021). Oxytocin, Neural Plasticity, and Social Behavior. Annual review of neuroscience 44, 359–381. 10.1146/annurev-neuro-102320-102847.
- Getting PA (1989). Emerging principles governing the operation of neural networks. Annual review of neuroscience 12, 185–204. 10.1146/annurev.ne.12.030189.001153.
- Getting PA, and Dekin MS (1985). Mechanisms of pattern generation underlying swimming in Tritonia. IV. Gating of central pattern generator. J Neurophysiol 53, 466–480. 10.1152/ jn.1985.53.2.466. [PubMed: 2984350]
- Goard M, and Dan Y (2009). Basal forebrain activation enhances cortical coding of natural scenes. Nature neuroscience 12, 1444–1449. 10.1038/nn.2402. [PubMed: 19801988]
- Gong R, Xu S, Hermundstad A, Yu Y, and Sternson SM (2020). Hindbrain Double-Negative Feedback Mediates Palatability-Guided Food and Water Consumption. Cell 182, 1589–1605 e1522. 10.1016/j.cell.2020.07.031. [PubMed: 32841600]

- Graving JM, Chae D, Naik H, Li L, Koger B, Costelloe BR, and Couzin ID (2019). DeepPoseKit, a software toolkit for fast and robust animal pose estimation using deep learning. Elife 8. 10.7554/ eLife.47994.
- Grover D, Katsuki T, Li J, Dawkins TJ, and Greenspan RJ (2020). Imaging brain activity during complex social behaviors in Drosophila with Flyception2. Nat Commun 11, 623. 10.1038/ s41467-020-14487-7. [PubMed: 32001689]
- Grundemann J, Bitterman Y, Lu T, Krabbe S, Grewe BF, Schnitzer MJ, and Luthi A (2019). Amygdala ensembles encode behavioral states. Science 364. 10.1126/science.aav8736.
- Hallinen KM, Dempsey R, Scholz M, Yu X, Linder A, Randi F, Sharma AK, Shaevitz JW, and Leifer AM (2021). Decoding locomotion from population neural activity in moving C. elegans. Elife 10. 10.7554/eLife.66135.
- Harris KD, and Thiele A (2011). Cortical state and attention. Nature reviews. Neuroscience 12, 509– 523. 10.1038/nrn3084. [PubMed: 21829219]
- Harris-Warrick RM, and Johnson BR (2010). Checks and balances in neuromodulation. Front Behav Neurosci 4. 10.3389/fnbeh.2010.00047.
- Harris-Warrick RM, and Marder E (1991). Modulation of neural networks for behavior. Annual review of neuroscience 14, 39–57. 10.1146/annurev.ne.14.030191.000351.
- Haubensak W, Kunwar PS, Cai H, Ciocchi S, Wall NR, Ponnusamy R, Biag J, Dong HW, Deisseroth K, Callaway EM, et al. (2010). Genetic dissection of an amygdala microcircuit that gates conditioned fear. Nature 468, 270–276. 10.1038/nature09553. [PubMed: 21068836]
- Hempel CM, Vincent P, Adams SR, Tsien RY, and Selverston AI (1996). Spatiotemporal dynamics of cyclic AMP signals in an intact neural circuitm. Nature 384, 166–169. 10.1038/384166a0. [PubMed: 8906791]
- Henry FE, Sugino K, Tozer A, Branco T, and Sternson SM (2015). Cell type-specific transcriptomics of hypothalamic energy-sensing neuron responses to weight-loss. eLife 4. 10.7554/eLife.09800.
- Herget U, Gutierrez-Triana JA, Salazar Thula O, Knerr B, and Ryu S (2017). Single-Cell Reconstruction of Oxytocinergic Neurons Reveals Separate Hypophysiotropic and Encephalotropic Subtypes in Larval Zebrafish. eNeuro 4. 10.1523/ENEURO.0278-16.2016.
- Hindmarsh Sten T, Li R, Otopalik A, and Ruta V (2021). Sexual arousal gates visual processing during Drosophila courtship. Nature 595, 549–553. 10.1038/s41586-021-03714-w. [PubMed: 34234348]
- Honegger K, and de Bivort B (2018). Stochasticity, individuality and behavior. Curr Biol 28, R8–R12. 10.1016/j.cub.2017.11.058. [PubMed: 29316423]
- Hong YK, Lacefield CO, Rodgers CC, and Bruno RM (2018). Sensation, movement and learning in the absence of barrel cortex. Nature 561, 542–546. 10.1038/s41586-018-0527-y. [PubMed: 30224746]
- Hoopfer ED, Jung Y, Inagaki HK, Rubin GM, and Anderson DJ (2015). P1 interneurons promote a persistent internal state that enhances inter-male aggression in Drosophila. eLife 4. 10.7554/ eLife.11346.
- Horio N, and Liberles SD (2021). Hunger enhances food-odour attraction through a neuropeptide Y spotlight. Nature 592, 262–266. 10.1038/s41586-021-03299-4. [PubMed: 33658716]
- Hounsgaard J, and Kiehn O (1989). Serotonin-induced bistability of turtle motoneurones caused by a nifedipine-sensitive calcium plateau potential. The Journal of physiology 414, 265–282. 10.1113/ jphysiol.1989.sp017687. [PubMed: 2607432]
- Hrvatin S, Sun S, Wilcox OF, Yao H, Lavin-Peter AJ, Cicconet M, Assad EG, Palmer ME, Aronson S, Banks AS, et al. (2020). Neurons that regulate mouse torpor. Nature 583, 115–121. 10.1038/ s41586-020-2387-5. [PubMed: 32528180]
- Inagaki HK, Ben-Tabou de-Leon S, Wong AM, Jagadish S, Ishimoto H, Barnea G, Kitamoto T, Axel R, and Anderson DJ (2012). Visualizing neuromodulation in vivo: TANGO-mapping of dopamine signaling reveals appetite control of sugar sensing. Cell 148, 583–595. 10.1016/ j.cell.2011.12.022. [PubMed: 22304923]
- Inagaki HK, Jung Y, Hoopfer ED, Wong AM, Mishra N, Lin JY, Tsien RY, and Anderson DJ (2014a). Optogenetic control of Drosophila using a red-shifted channelrhodopsin reveals experiencedependent influences on courtship. Nature methods 11, 325–332. 10.1038/nmeth.2765. [PubMed: 24363022]

- Inagaki HK, Panse KM, and Anderson DJ (2014b). Independent, reciprocal neuromodulatory control of sweet and bitter taste sensitivity during starvation in Drosophila. Neuron 84, 806–820. 10.1016/j.neuron.2014.09.032. [PubMed: 25451195]
- Insel TR, and Young LJ (2001). The neurobiology of attachment. Nat Rev Neurosci 2, 129–136. 10.1038/35053579. [PubMed: 11252992]
- Jan YN, Jan LY, and Kuffler SW (1979). A peptide as a possible transmitter in sympathetic ganglia of the frog. Proc Natl Acad Sci U S A 76, 1501–1505. 10.1073/pnas.76.3.1501. [PubMed: 35789]
- Janak PH, and Tye KM (2015). From circuits to behaviour in the amygdala. Nature 517, 284–292. 10.1038/nature14188. [PubMed: 25592533]
- Jazayeri M, and Afraz A (2017). Navigating the Neural Space in Search of the Neural Code. Neuron 93, 1003–1014. 10.1016/j.neuron.2017.02.019. [PubMed: 28279349]
- Ji N, Madan GK, Fabre GI, Dayan A, Baker CM, Kramer TS, Nwabudike I, and Flavell SW (2021). A neural circuit for flexible control of persistent behavioral states. eLife 10. 10.7554/eLife.62889.
- Jikomes N, Ramesh RN, Mandelblat-Cerf Y, and Andermann ML (2016). Preemptive Stimulation of AgRP Neurons in Fed Mice Enables Conditioned Food Seeking under Threat. Curr Biol 26, 2500–2507. 10.1016/j.cub.2016.07.019. [PubMed: 27568593]
- Johnson RE, Linderman S, Panier T, Wee CL, Song E, Herrera KJ, Miller A, and Engert F (2020). Probabilistic Models of Larval Zebrafish Behavior Reveal Structure on Many Scales. Curr Biol 30, 70–82 e74. 10.1016/j.cub.2019.11.026. [PubMed: 31866367]
- Joshua M, and Lisberger SG (2015). A tale of two species: Neural integration in zebrafish and monkeys. Neuroscience 296, 80–91. 10.1016/j.neuroscience.2014.04.048. [PubMed: 24797331]
- Jourjine N, and Hoekstra HE (2021). Expanding evolutionary neuroscience: insights from comparing variation in behavior. Neuron 109, 1084–1099. 10.1016/j.neuron.2021.02.002. [PubMed: 33609484]
- Jourjine N, Mullaney BC, Mann K, and Scott K (2016). Coupled Sensing of Hunger and Thirst Signals Balances Sugar and Water Consumption. Cell 166, 855–866. 10.1016/j.cell.2016.06.046. [PubMed: 27477513]
- Juavinett AL, Bekheet G, and Churchland AK (2019). Chronically implanted Neuropixels probes enable high-yield recordings in freely moving mice. Elife 8. 10.7554/eLife.47188.
- Jung Y, Kennedy A, Chiu H, Mohammad F, Claridge-Chang A, and Anderson DJ (2020). Neurons that Function within an Integrator to Promote a Persistent Behavioral State in Drosophila. Neuron 105, 322–333 e325. 10.1016/j.neuron.2019.10.028. [PubMed: 31810837]
- Kabra M, Robie AA, Rivera-Alba M, Branson S, and Branson K (2013). JAABA: interactive machine learning for automatic annotation of animal behavior. Nat Methods 10, 64–67. 10.1038/ nmeth.2281. [PubMed: 23202433]
- Karigo T, Kennedy A, Yang B, Liu M, Tai D, Wahle IA, and Anderson DJ (2021). Distinct hypothalamic control of same- and opposite-sex mounting behaviour in mice. Nature 589, 258– 263. 10.1038/s41586-020-2995-0. [PubMed: 33268894]
- Kato S, Kaplan HS, Schrodel T, Skora S, Lindsay TH, Yemini E, Lockery S, and Zimmer M (2015). Global brain dynamics embed the motor command sequence of Caenorhabditis elegans. Cell 163, 656–669. 10.1016/j.cell.2015.09.034. [PubMed: 26478179]
- Katz PS (1998). Neuromodulation intrinsic to the central pattern generator for escape swimming in Tritonia. Annals of the New York Academy of Sciences 860, 181–188. 10.1111/ j.1749-6632.1998.tb09048.x. [PubMed: 9928311]
- Katz PS (2016). 'Model organisms' in the light of evolution. Curr Biol 26, R649–650. 10.1016/ j.cub.2016.05.071. [PubMed: 27458905]
- Katz PS, and Frost WN (1995). Intrinsic neuromodulation in the Tritonia swim CPG: the serotonergic dorsal swim interneurons act presynaptically to enhance transmitter release from interneuron C2. The Journal of neuroscience : the official journal of the Society for Neuroscience 15, 6035–6045. [PubMed: 7666187]
- Katz PS, and Frost WN (1996). Intrinsic neuromodulation: altering neuronal circuits from within. Trends Neurosci 19, 54–61. 10.1016/0166-2236(96)89621-4. [PubMed: 8820868]
- Katz PS, Getting PA, and Frost WN (1994). Dynamic neuromodulation of synaptic strength intrinsic to a central pattern generator circuit. Nature 367, 729–731. 10.1038/367729a0. [PubMed: 8107867]

- Kauvar IV, Machado TA, Yuen E, Kochalka J, Choi M, Allen WE, Wetzstein G, and Deisseroth K (2020). Cortical Observation by Synchronous Multifocal Optical Sampling Reveals Widespread Population Encoding of Actions. Neuron 107, 351–367 e319. 10.1016/j.neuron.2020.04.023. [PubMed: 32433908]
- Keller GB, Bonhoeffer T, and Hubener M (2012). Sensorimotor mismatch signals in primary visual cortex of the behaving mouse. Neuron 74, 809–815. 10.1016/j.neuron.2012.03.040. [PubMed: 22681686]
- Kennedy A, Asahina K, Hoopfer E, Inagaki H, Jung Y, Lee H, Remedios R, and Anderson DJ (2014). Internal States and Behavioral Decision-Making: Toward an Integration of Emotion and Cognition. Cold Spring Harbor symposia on quantitative biology 79, 199–210. 10.1101/ sqb.2014.79.024984. [PubMed: 25948637]
- Kennedy A, Kunwar PS, Li LY, Stagkourakis S, Wagenaar DA, and Anderson DJ (2020). Stimulusspecific hypothalamic encoding of a persistent defensive state. Nature 586, 730–734. 10.1038/ s41586-020-2728-4. [PubMed: 32939094]
- Kim AJ, Fenk LM, Lyu C, and Maimon G (2017a). Quantitative Predictions Orchestrate Visual Signaling in Drosophila. Cell 168, 280–294 e212. 10.1016/j.cell.2016.12.005. [PubMed: 28065412]
- Kim AJ, Fitzgerald JK, and Maimon G (2015). Cellular evidence for efference copy in Drosophila visuomotor processing. Nat Neurosci 18, 1247–1255. 10.1038/nn.4083. [PubMed: 26237362]
- Kim DH, Kim J, Marques JC, Grama A, Hildebrand DGC, Gu W, Li JM, and Robson DN (2017b). Pan-neuronal calcium imaging with cellular resolution in freely swimming zebrafish. Nat Methods 14, 1107–1114. 10.1038/nmeth.4429. [PubMed: 28892088]
- Kim DW, Yao Z, Graybuck LT, Kim TK, Nguyen TN, Smith KA, Fong O, Yi L, Koulena N, Pierson N, et al. (2019). Multimodal Analysis of Cell Types in a Hypothalamic Node Controlling Social Behavior. Cell 179, 713–728 e717. 10.1016/j.cell.2019.09.020. [PubMed: 31626771]
- Kim SM, Su CY, and Wang JW (2017c). Neuromodulation of Innate Behaviors in Drosophila. Annual review of neuroscience 40, 327–348. 10.1146/annurev-neuro-072116-031558.
- Kim SY, Adhikari A, Lee SY, Marshel JH, Kim CK, Mallory CS, Lo M, Pak S, Mattis J, Lim BK, et al. (2013). Diverging neural pathways assemble a behavioural state from separable features in anxiety. Nature 496, 219–223. 10.1038/nature12018. [PubMed: 23515158]
- Kintos N, Nusbaum MP, and Nadim F (2016). Convergent neuromodulation onto a network neuron can have divergent effects at the network level. J Comput Neurosci 40, 113–135. 10.1007/ s10827-015-0587-z. [PubMed: 26798029]
- Ko KI, Root CM, Lindsay SA, Zaninovich OA, Shepherd AK, Wasserman SA, Kim SM, and Wang JW (2015). Starvation promotes concerted modulation of appetitive olfactory behavior via parallel neuromodulatory circuits. Elife 4. 10.7554/eLife.08298.
- Kohl J, Babayan BM, Rubinstein ND, Autry AE, Marin-Rodriguez B, Kapoor V, Miyamishi K, Zweifel LS, Luo L, Uchida N, and Dulac C (2018). Functional circuit architecture underlying parental behaviour. Nature 556, 326–331. 10.1038/s41586-018-0027-0. [PubMed: 29643503]
- Kohl J, and Dulac C (2018). Neural control of parental behaviors. Curr Opin Neurobiol 49, 116–122. 10.1016/j.conb.2018.02.002. [PubMed: 29482085]
- Krashes MJ, DasGupta S, Vreede A, White B, Armstrong JD, and Waddell S (2009). A neural circuit mechanism integrating motivational state with memory expression in Drosophila. Cell 139, 416– 427. 10.1016/j.cell.2009.08.035. [PubMed: 19837040]
- Krashes MJ, Koda S, Ye C, Rogan SC, Adams AC, Cusher DS, Maratos-Flier E, Roth BL, and Lowell BB (2011). Rapid, reversible activation of AgRP neurons drives feeding behavior in mice. J Clin Invest 121, 1424–1428. 10.1172/JCI46229. [PubMed: 21364278]
- Kravitz EA (2000). Serotonin and aggression: insights gained from a lobster model system and speculations on the role of amine neurons in a complex behavior. J Comp Physiol A 186, 221– 238. 10.1007/s003590050423. [PubMed: 10757238]
- Kristan WB Jr., and Calabrese RL (1976). Rhythmic swimming activity in neurones of the isolated nerve cord of the leech. J Exp Biol 65, 643–668. 10.1242/jeb.65.3.643. [PubMed: 1018167]
- Kroeger D, Absi G, Gagliardi C, Bandaru SS, Madara JC, Ferrari LL, Arrigoni E, Munzberg H, Scammell TE, Saper CB, and Vetrivelan R (2018). Galanin neurons in the ventrolateral preoptic

area promote sleep and heat loss in mice. Nat Commun 9, 4129. 10.1038/s41467-018-06590-7. [PubMed: 30297727]

- Kunwar PS, Zelikowsky M, Remedios R, Cai H, Yilmaz M, Meister M, and Anderson DJ (2015). Ventromedial hypothalamic neurons control a defensive emotion state. Elife 4. 10.7554/ eLife.06633.
- Lahiri AK, and Bevan MD (2020). Dopaminergic Transmission Rapidly and Persistently Enhances Excitability of D1 Receptor-Expressing Striatal Projection Neurons. Neuron 106, 277–290 e276. 10.1016/j.neuron.2020.01.028. [PubMed: 32075716]
- Laurent G (2020). On the value of model diversity in neuroscience. Nature reviews. Neuroscience 21, 395–396. 10.1038/s41583-020-0323-1. [PubMed: 32514109]
- LeDoux J, and Daw ND (2018). Surviving threats: neural circuit and computational implications of a new taxonomy of defensive behaviour. Nature reviews. Neuroscience 19, 269–282. 10.1038/ nrn.2018.22. [PubMed: 29593300]
- LeDoux JE (2017). Semantics, Surplus Meaning, and the Science of Fear. Trends Cogn Sci 21, 303– 306. 10.1016/j.tics.2017.02.004. [PubMed: 28318937]
- LeDoux JE (2020). Thoughtful feelings. Current biology : CB 30, R619–R623. 10.1016/ j.cub.2020.04.012. [PubMed: 32516605]
- LeDoux JE (2021). What emotions might be like in other animals. Curr Biol 31, R824–R829. 10.1016/ j.cub.2021.05.005. [PubMed: 34256909]
- LeDoux JE, and Brown R (2017). A higher-order theory of emotional consciousness. Proceedings of the National Academy of Sciences of the United States of America 114, E2016–E2025. 10.1073/ pnas.1619316114. [PubMed: 28202735]
- Lee H, Kim DW, Remedios R, Anthony TE, Chang A, Madisen L, Zeng H, and Anderson DJ (2014). Scalable control of mounting and attack by Esr1+ neurons in the ventromedial hypothalamus. Nature 509, 627–632. 10.1038/nature13169. [PubMed: 24739975]
- Lee SH, and Dan Y (2012). Neuromodulation of brain states. Neuron 76, 209–222. 10.1016/ j.neuron.2012.09.012. [PubMed: 23040816]
- Lee SJ, Chen Y, Lodder B, and Sabatini BL (2019). Monitoring Behaviorally Induced Biochemical Changes Using Fluorescence Lifetime Photometry. Front Neurosci 13, 766. 10.3389/fnins.2019.00766. [PubMed: 31417343]
- Leib DE, Zimmerman CA, Poormoghaddam A, Huey EL, Ahn JS, Lin YC, Tan CL, Chen Y, and Knight ZA (2017). The Forebrain Thirst Circuit Drives Drinking through Negative Reinforcement. Neuron 96, 1272–1281 e1274. 10.1016/j.neuron.2017.11.041. [PubMed: 29268095]
- Liang L, Fratzl A, Reggiani JDS, El Mansour O, Chen C, and Andermann ML (2020). Retinal Inputs to the Thalamus Are Selectively Gated by Arousal. Curr Biol 30, 3923–3934 e3929. 10.1016/ j.cub.2020.07.065. [PubMed: 32795442]
- Lin A, Witvliet D, Hernandez-Nunez L, Linderman SW, Samuel ADT, and Venkatachalam V (2022). Imaging whole-brain activity to understand behaviour. Nature Reviews Physics. 10.1038/ s42254-022-00430-w.
- Lin D, Boyle MP, Dollar P, Lee H, Lein ES, Perona P, and Anderson DJ (2011). Functional identification of an aggression locus in the mouse hypothalamus. Nature 470, 221–226. 10.1038/ nature09736. [PubMed: 21307935]
- Lin Y, Stormo GD, and Taghert PH (2004). The neuropeptide pigment-dispersing factor coordinates pacemaker interactions in the Drosophila circadian system. The Journal of neuroscience : the official journal of the Society for Neuroscience 24, 7951–7957. 10.1523/ JNEUROSCI.2370-04.2004. [PubMed: 15356209]
- Lisman J, Yasuda R, and Raghavachari S (2012). Mechanisms of CaMKII action in long-term potentiation. Nature reviews. Neuroscience 13, 169–182. 10.1038/nrn3192. [PubMed: 22334212]
- Liu C, Goel P, and Kaeser PS (2021). Spatial and temporal scales of dopamine transmission. Nat Rev Neurosci 22, 345–358. 10.1038/s41583-021-00455-7. [PubMed: 33837376]
- Liu D, and Dan Y (2019). A Motor Theory of Sleep-Wake Control: Arousal-Action Circuit. Annual review of neuroscience 42, 27–46. 10.1146/annurev-neuro-080317-061813.

- Livneh Y, Ramesh RN, Burgess CR, Levandowski KM, Madara JC, Fenselau H, Goldey GJ, Diaz VE, Jikomes N, Resch JM, et al. (2017). Homeostatic circuits selectively gate food cue responses in insular cortex. Nature 546, 611–616. 10.1038/nature22375. [PubMed: 28614299]
- Livneh Y, Sugden AU, Madara JC, Essner RA, Flores VI, Sugden LA, Resch JM, Lowell BB, and Andermann ML (2020). Estimation of Current and Future Physiological States in Insular Cortex. Neuron 105, 1094–1111 e1010. 10.1016/j.neuron.2019.12.027. [PubMed: 31955944]
- Lo L, Yao S, Kim DW, Cetin A, Harris J, Zeng H, Anderson DJ, and Weissbourd B (2019). Connectional architecture of a mouse hypothalamic circuit node controlling social behavior. Proceedings of the National Academy of Sciences of the United States of America 116, 7503– 7512. 10.1073/pnas.1817503116. [PubMed: 30898882]
- Lovett-Barron M, Andalman AS, Allen WE, Vesuna S, Kauvar I, Burns VM, and Deisseroth K (2017). Ancestral Circuits for the Coordinated Modulation of Brain State. Cell 171, 1411–1423 e1417. 10.1016/j.cell.2017.10.021. [PubMed: 29103613]
- Lovett-Barron M, Chen R, Bradbury S, Andalman AS, Wagle M, Guo S, and Deisseroth K (2020). Multiple convergent hypothalamus-brainstem circuits drive defensive behavior. Nature neuroscience 23, 959–967. 10.1038/s41593-020-0655-1. [PubMed: 32572237]
- Ludwig M, and Leng G (2006). Dendritic peptide release and peptide-dependent behaviours. Nature reviews. Neuroscience 7, 126–136. 10.1038/nrn1845. [PubMed: 16429122]
- Luo L, Callaway EM, and Svoboda K (2018). Genetic Dissection of Neural Circuits: A Decade of Progress. Neuron 98, 865. 10.1016/j.neuron.2018.05.004. [PubMed: 29772206]
- Lutas A, Kucukdereli H, Alturkistani O, Carty C, Sugden AU, Fernando K, Diaz V, Flores-Maldonado V, and Andermann ML (2019). State-specific gating of salient cues by midbrain dopaminergic input to basal amygdala. Nat Neurosci 22, 1820–1833. 10.1038/s41593-019-0506-0. [PubMed: 31611706]
- Luxem K, Fuhrmann F, Kürsch J, Remy S, and Bauer P (2020). Identifying Behavioral Structure from Deep Variational Embeddings of Animal Motion. bioRxiv, 2020.2005.2014.095430. 10.1101/2020.05.14.095430.
- Maimon G (2011). Modulation of visual physiology by behavioral state in monkeys, mice, and flies. Curr Opin Neurobiol 21, 559–564. 10.1016/j.conb.2011.05.001. [PubMed: 21628097]
- Maimon G, Straw AD, and Dickinson MH (2010). Active flight increases the gain of visual motion processing in Drosophila. Nat Neurosci 13, 393–399. 10.1038/nn.2492. [PubMed: 20154683]
- Makino H, Ren C, Liu H, Kim AN, Kondapaneni N, Liu X, Kuzum D, and Komiyama T (2017). Transformation of Cortex-wide Emergent Properties during Motor Learning. Neuron 94, 880– 890 e888. 10.1016/j.neuron.2017.04.015. [PubMed: 28521138]
- Mandelblat-Cerf Y, Ramesh RN, Burgess CR, Patella P, Yang Z, Lowell BB, and Andermann ML (2015). Arcuate hypothalamic AgRP and putative POMC neurons show opposite changes in spiking across multiple timescales. Elife 4. 10.7554/eLife.07122.
- Mann K, Deny S, Ganguli S, and Clandinin TR (2021). Coupling of activity, metabolism and behaviour across the Drosophila brain. Nature 593, 244–248. 10.1038/s41586-021-03497-0. [PubMed: 33911283]
- Marder E (2002). Non-mammalian models for studying neural development and function. Nature 417, 318–321. 10.1038/417318a. [PubMed: 12015611]
- Marder E (2012). Neuromodulation of neuronal circuits: back to the future. Neuron 76, 1–11. 10.1016/ j.neuron.2012.09.010. [PubMed: 23040802]
- Marder E, and Calabrese RL (1996). Principles of rhythmic motor pattern generation. Physiological reviews 76, 687–717. 10.1152/physrev.1996.76.3.687. [PubMed: 8757786]
- Marder E, and Thirumalai V (2002). Cellular, synaptic and network effects of neuromodulation. Neural Netw 15, 479–493. 10.1016/s0893-6080(02)00043-6. [PubMed: 12371506]
- Marlin BJ, Mitre M, D'Amour J A, Chao MV, and Froemke RC (2015). Oxytocin enables maternal behaviour by balancing cortical inhibition. Nature 520, 499–504. 10.1038/nature14402. [PubMed: 25874674]
- Marques JC, Li M, Schaak D, Robson DN, and Li JM (2020). Internal state dynamics shape brainwide activity and foraging behaviour. Nature 577, 239–243. 10.1038/s41586-019-1858-z. [PubMed: 31853063]

Martin KC, Casadio A, Zhu H, Yaping E, Rose JC, Chen M, Bailey CH, and Kandel ER (1997).
Synapse-specific, long-term facilitation of aplysia sensory to motor synapses: a function for local protein synthesis in memory storage. Cell 91, 927–938. 10.1016/s0092-8674(00)80484-5.
[PubMed: 9428516]

Maslow AH (1943). A Theory of Human Emotion. Psychological Review 50, 430-437.

- Mathis A, Mamidanna P, Cury KM, Abe T, Murthy VN, Mathis MW, and Bethge M (2018). DeepLabCut: markerless pose estimation of user-defined body parts with deep learning. Nat Neurosci 21, 1281–1289. 10.1038/s41593-018-0209-y. [PubMed: 30127430]
- Mathis MW, and Mathis A (2020). Deep learning tools for the measurement of animal behavior in neuroscience. Curr Opin Neurobiol 60, 1–11. 10.1016/j.conb.2019.10.008. [PubMed: 31791006]
- Matthews GA, Nieh EH, Vander Weele CM, Halbert SA, Pradhan RV, Yosafat AS, Glober GF, Izadmehr EM, Thomas RE, Lacy GD, et al. (2016). Dorsal Raphe Dopamine Neurons Represent the Experience of Social Isolation. Cell 164, 617–631. 10.1016/j.cell.2015.12.040. [PubMed: 26871628]
- McCormick DA (1992). Neurotransmitter actions in the thalamus and cerebral cortex and their role in neuromodulation of thalamocortical activity. Progress in neurobiology 39, 337–388. 10.1016/0301-0082(92)90012-4. [PubMed: 1354387]
- McCormick DA, Nestvogel DB, and He BJ (2020). Neuromodulation of Brain State and Behavior. Annual review of neuroscience 43, 391–415. 10.1146/annurev-neuro-100219-105424.
- McCormick DA, and Prince DA (1986). Acetylcholine induces burst firing in thalamic reticular neurones by activating a potassium conductance. Nature 319, 402–405. 10.1038/319402a0. [PubMed: 2418361]
- McGinley MJ, David SV, and McCormick DA (2015a). Cortical Membrane Potential Signature of Optimal States for Sensory Signal Detection. Neuron 87, 179–192. 10.1016/ j.neuron.2015.05.038. [PubMed: 26074005]
- McGinley MJ, Vinck M, Reimer J, Batista-Brito R, Zagha E, Cadwell CR, Tolias AS, Cardin JA, and McCormick DA (2015b). Waking State: Rapid Variations Modulate Neural and Behavioral Responses. Neuron 87, 1143–1161. 10.1016/j.neuron.2015.09.012. [PubMed: 26402600]
- Mearns DS, Donovan JC, Fernandes AM, Semmelhack JL, and Baier H (2020). Deconstructing Hunting Behavior Reveals a Tightly Coupled Stimulus-Response Loop. Curr Biol 30, 54–69 e59. 10.1016/j.cub.2019.11.022. [PubMed: 31866365]
- Meir I, Katz Y, and Lampl I (2018). Membrane Potential Correlates of Network Decorrelation and Improved SNR by Cholinergic Activation in the Somatosensory Cortex. The Journal of neuroscience : the official journal of the Society for Neuroscience 38, 10692–10708. 10.1523/ JNEUROSCI.1159-18.2018. [PubMed: 30373769]
- Melzer S, Newmark ER, Mizuno GO, Hyun M, Philson AC, Quiroli E, Righetti B, Gregory MR, Huang KW, Levasseur J, et al. (2021). Bombesin-like peptide recruits disinhibitory cortical circuits and enhances fear memories. Cell 184, 5622–5634 e5625. 10.1016/j.cell.2021.09.013. [PubMed: 34610277]
- Miller SG, and Kennedy MB (1986). Regulation of brain type II Ca2+/calmodulin-dependent protein kinase by autophosphorylation: a Ca2+-triggered molecular switch. Cell 44, 861–870. 10.1016/0092-8674(86)90008-5. [PubMed: 3006921]
- Miri A, Daie K, Arrenberg AB, Baier H, Aksay E, and Tank DW (2011). Spatial gradients and multidimensional dynamics in a neural integrator circuit. Nature neuroscience 14, 1150–1159. 10.1038/nn.2888. [PubMed: 21857656]
- Mobbs D, Adolphs R, Fanselow MS, Barrett LF, LeDoux JE, Ressler K, and Tye KM (2019). Viewpoints: Approaches to defining and investigating fear. Nat Neurosci 22, 1205–1216. 10.1038/s41593-019-0456-6. [PubMed: 31332374]
- Moffitt JR, Bambah-Mukku D, Eichhorn SW, Vaughn E, Shekhar K, Perez JD, Rubinstein ND, Hao J, Regev A, Dulac C, and Zhuang X (2018). Molecular, spatial, and functional single-cell profiling of the hypothalamic preoptic region. Science 362. 10.1126/science.aau5324.
- Moore T, and Zirnsak M (2017). Neural Mechanisms of Selective Visual Attention. Annu Rev Psychol 68, 47–72. 10.1146/annurev-psych-122414-033400. [PubMed: 28051934]

- Moukhles H, Bosler O, Bolam JP, Vallee A, Umbriaco D, Geffard M, and Doucet G (1997). Quantitative and morphometric data indicate precise cellular interactions between serotonin terminals and postsynaptic targets in rat substantia nigra. Neuroscience 76, 1159–1171. 10.1016/ s0306-4522(96)00452-6. [PubMed: 9027876]
- Mu Y, Bennett DV, Rubinov M, Narayan S, Yang CT, Tanimoto M, Mensh BD, Looger LL, and Ahrens MB (2019). Glia Accumulate Evidence that Actions Are Futile and Suppress Unsuccessful Behavior. Cell 178, 27–43 e19. 10.1016/j.cell.2019.05.050. [PubMed: 31230713]
- Musall S, Kaufman MT, Juavinett AL, Gluf S, and Churchland AK (2019). Single-trial neural dynamics are dominated by richly varied movements. Nat Neurosci 22, 1677–1686. 10.1038/ s41593-019-0502-4. [PubMed: 31551604]
- Nadim F, and Bucher D (2014). Neuromodulation of neurons and synapses. Current opinion in neurobiology 29, 48–56. 10.1016/j.conb.2014.05.003. [PubMed: 24907657]
- Nath RD, Bedbrook CN, Abrams MJ, Basinger T, Bois JS, Prober DA, Sternberg PW, Gradinaru V, and Goentoro L (2017). The Jellyfish Cassiopea Exhibits a Sleep-like State. Curr Biol 27, 2984–2990 e2983. 10.1016/j.cub.2017.08.014. [PubMed: 28943083]
- Nath RD, Chow ES, Wang H, Schwarz EM, and Sternberg PW (2016). C. elegans Stress-Induced Sleep Emerges from the Collective Action of Multiple Neuropeptides. Curr Biol 26, 2446–2455. 10.1016/j.cub.2016.07.048. [PubMed: 27546573]
- Naumann EA, Fitzgerald JE, Dunn TW, Rihel J, Sompolinsky H, and Engert F (2016). From Whole-Brain Data to Functional Circuit Models: The Zebrafish Optomotor Response. Cell 167, 947–960 e920. 10.1016/j.cell.2016.10.019. [PubMed: 27814522]
- Nelson JC, and Colon-Ramos DA (2013). Serotonergic neurosecretory synapse targeting is controlled by netrin-releasing guidepost neurons in Caenorhabditis elegans. The Journal of neuroscience : the official journal of the Society for Neuroscience 33, 1366–1376. 10.1523/ JNEUROSCI.3471-12.2012. [PubMed: 23345213]
- Nguyen JP, Shipley FB, Linder AN, Plummer GS, Liu M, Setru SU, Shaevitz JW, and Leifer AM (2016). Whole-brain calcium imaging with cellular resolution in freely behaving Caenorhabditis elegans. Proc Natl Acad Sci U S A 113, E1074–1081. 10.1073/pnas.1507110112. [PubMed: 26712014]
- Nichols ALA, Eichler T, Latham R, and Zimmer M (2017). A global brain state underlies C. elegans sleep behavior. Science 356. 10.1126/science.aam6851.
- Nieh EH, Vander Weele CM, Matthews GA, Presbrey KN, Wichmann R, Leppla CA, Izadmehr EM, and Tye KM (2016). Inhibitory Input from the Lateral Hypothalamus to the Ventral Tegmental Area Disinhibits Dopamine Neurons and Promotes Behavioral Activation. Neuron 90, 1286– 1298. 10.1016/j.neuron.2016.04.035. [PubMed: 27238864]
- Niell CM, and Stryker MP (2010). Modulation of visual responses by behavioral state in mouse visual cortex. Neuron 65, 472–479. 10.1016/j.neuron.2010.01.033. [PubMed: 20188652]
- Nusbaum MP, and Beenhakker MP (2002). A small-systems approach to motor pattern generation. Nature 417, 343–350. 10.1038/417343a. [PubMed: 12015615]
- Nusbaum MP, and Blitz DM (2012). Neuropeptide modulation of microcircuits. Current opinion in neurobiology 22, 592–601. 10.1016/j.conb.2012.01.003. [PubMed: 22305485]
- Nusbaum MP, Blitz DM, Swensen AM, Wood D, and Marder E (2001). The roles of co-transmission in neural network modulation. Trends Neurosci 24, 146–154. 10.1016/s0166-2236(00)01723-9. [PubMed: 11182454]
- Oikonomou G, and Prober DA (2017). Attacking sleep from a new angle: contributions from zebrafish. Current opinion in neurobiology 44, 80–88. 10.1016/j.conb.2017.03.009. [PubMed: 28391131]
- Oka Y, Ye M, and Zuker CS (2015). Thirst driving and suppressing signals encoded by distinct neural populations in the brain. Nature 520, 349–352. 10.1038/nature14108. [PubMed: 25624099]
- Otchy TM, Wolff SB, Rhee JY, Pehlevan C, Kawai R, Kempf A, Gobes SM, and Olveczky BP (2015). Acute off-target effects of neural circuit manipulations. Nature 528, 358–363. 10.1038/ nature16442. [PubMed: 26649821]
- Oti T, Satoh K, Uta D, Nagafuchi J, Tateishi S, Ueda R, Takanami K, Young LJ, Galione A, Morris JF, et al. (2021). Oxytocin Influences Male Sexual Activity via Non-synaptic Axonal Release in

the Spinal Cord. Current biology : CB 31, 103–114 e105. 10.1016/j.cub.2020.09.089. [PubMed: 33125871]

- Padilla-Coreano N, Bolkan SS, Pierce GM, Blackman DR, Hardin WD, Garcia-Garcia AL, Spellman TJ, and Gordon JA (2016). Direct Ventral Hippocampal-Prefrontal Input Is Required for Anxiety-Related Neural Activity and Behavior. Neuron 89, 857–866. 10.1016/j.neuron.2016.01.011. [PubMed: 26853301]
- Pantoja C, Hoagland A, Carroll EC, Karalis V, Conner A, and Isacoff EY (2016). Neuromodulatory Regulation of Behavioral Individuality in Zebrafish. Neuron 91, 587–601. 10.1016/j.neuron.2016.06.016. [PubMed: 27397519]
- Pantoja C, Larsch J, Laurell E, Marquart G, Kunst M, and Baier H (2020). Rapid Effects of Selection on Brain-wide Activity and Behavior. Curr Biol 30, 3647–3656 e3643. 10.1016/ j.cub.2020.06.086. [PubMed: 32763165]
- Pape HC, and McCormick DA (1989). Noradrenaline and serotonin selectively modulate thalamic burst firing by enhancing a hyperpolarization-activated cation current. Nature 340, 715–718. 10.1038/340715a0. [PubMed: 2475782]
- Park J, Takmakov P, and Wightman RM (2011). In vivo comparison of norepinephrine and dopamine release in rat brain by simultaneous measurements with fast-scan cyclic voltammetry. Journal of neurochemistry 119, 932–944. 10.1111/j.1471-4159.2011.07494.x. [PubMed: 21933188]
- Park JY, Dus M, Kim S, Abu F, Kanai MI, Rudy B, and Suh GSB (2016). Drosophila SLC5A11 Mediates Hunger by Regulating K(+) Channel Activity. Curr Biol 26, 2550. 10.1016/ j.cub.2016.08.027.
- Pavlov IP (1927). Conditioned reflexes: an investigation of the physiological activity of the cerebral cortex. (Oxford University Press).
- Pereira TD, Aldarondo DE, Willmore L, Kislin M, Wang SS, Murthy M, and Shaevitz JW (2019). Fast animal pose estimation using deep neural networks. Nat Methods 16, 117–125. 10.1038/ s41592-018-0234-5. [PubMed: 30573820]
- Pereira TD, Shaevitz JW, and Murthy M (2020). Quantifying behavior to understand the brain. Nat Neurosci 23, 1537–1549. 10.1038/s41593-020-00734-z. [PubMed: 33169033]
- Persoon CM, Moro A, Nassal JP, Farina M, Broeke JH, Arora S, Dominguez N, van Weering JR, Toonen RF, and Verhage M (2018). Pool size estimations for dense-core vesicles in mammalian CNS neurons. EMBO J 37. 10.15252/embj.201899672.
- Perusini JN, and Fanselow MS (2015). Neurobehavioral perspectives on the distinction between fear and anxiety. Learning & memory 22, 417–425. 10.1101/lm.039180.115. [PubMed: 26286652]
- Pimentel D, Donlea JM, Talbot CB, Song SM, Thurston AJF, and Miesenbock G (2016). Operation of a homeostatic sleep switch. Nature 536, 333–337. 10.1038/nature19055. [PubMed: 27487216]
- Pinto L, Goard MJ, Estandian D, Xu M, Kwan AC, Lee SH, Harrison TC, Feng G, and Dan Y (2013). Fast modulation of visual perception by basal forebrain cholinergic neurons. Nature neuroscience 16, 1857–1863. 10.1038/nn.3552. [PubMed: 24162654]
- Pisansky MT, Hanson LR, Gottesman II, and Gewirtz JC (2017). Oxytocin enhances observational fear in mice. Nat Commun 8, 2102. 10.1038/s41467-017-02279-5. [PubMed: 29235461]
- Poe GR, Foote S, Eschenko O, Johansen JP, Bouret S, Aston-Jones G, Harley CW, Manahan-Vaughan D, Weinshenker D, Valentino R, et al. (2020). Locus coeruleus: a new look at the blue spot. Nature reviews. Neuroscience 21, 644–659. 10.1038/s41583-020-0360-9. [PubMed: 32943779]
- Polack PO, Friedman J, and Golshani P (2013). Cellular mechanisms of brain state-dependent gain modulation in visual cortex. Nature neuroscience 16, 1331–1339. 10.1038/nn.3464. [PubMed: 23872595]
- Poulet JF, and Petersen CC (2008). Internal brain state regulates membrane potential synchrony in barrel cortex of behaving mice. Nature 454, 881–885. 10.1038/nature07150. [PubMed: 18633351]
- Powell DJ, Marder E, and Nusbaum MP (2021). Perturbation-specific responses by two neural circuits generating similar activity patterns. Current biology : CB 31, 4831–4838 e4834. 10.1016/ j.cub.2021.08.042. [PubMed: 34506730]
- Quinn JJ, Oommen SS, Morrison GE, and Fanselow MS (2002). Post-training excitotoxic lesions of the dorsal hippocampus attenuate forward trace, backward trace, and delay fear conditioning

in a temporally specific manner. Hippocampus 12, 495–504. 10.1002/hipo.10029. [PubMed: 12201634]

- Raichle ME (2015). The brain's default mode network. Annual review of neuroscience 38, 433–447. 10.1146/annurev-neuro-071013-014030.
- Reimer J, Froudarakis E, Cadwell CR, Yatsenko D, Denfield GH, and Tolias AS (2014). Pupil fluctuations track fast switching of cortical states during quiet wakefulness. Neuron 84, 355–362. 10.1016/j.neuron.2014.09.033. [PubMed: 25374359]
- Reiter RJ, Tan DX, Kim SJ, and Cruz MH (2014). Delivery of pineal melatonin to the brain and SCN: role of canaliculi, cerebrospinal fluid, tanycytes and Virchow-Robin perivascular spaces. Brain Struct Funct 219, 1873–1887. 10.1007/s00429-014-0719-7. [PubMed: 24553808]
- Remedios R, Kennedy A, Zelikowsky M, Grewe BF, Schnitzer MJ, and Anderson DJ (2017). Social behaviour shapes hypothalamic neural ensemble representations of conspecific sex. Nature 550, 388–392. 10.1038/nature23885. [PubMed: 29052632]
- Ren J, Friedmann D, Xiong J, Liu CD, Ferguson BR, Weerakkody T, DeLoach KE, Ran C, Pun A, Sun Y, et al. (2018). Anatomically Defined and Functionally Distinct Dorsal Raphe Serotonin Sub-systems. Cell 175, 472–487 e420. 10.1016/j.cell.2018.07.043. [PubMed: 30146164]
- Ren J, Isakova A, Friedmann D, Zeng J, Grutzner SM, Pun A, Zhao GQ, Kolluru SS, Wang R, Lin R, et al. (2019). Single-cell transcriptomes and whole-brain projections of serotonin neurons in the mouse dorsal and median raphe nuclei. Elife 8. 10.7554/eLife.49424.
- Ringstad N, Abe N, and Horvitz HR (2009). Ligand-gated chloride channels are receptors for biogenic amines in C. elegans. Science 325, 96–100. 10.1126/science.1169243. [PubMed: 19574391]
- Rodriguez-Romaguera J, Ung RL, Nomura H, Otis JM, Basiri ML, Namboodiri VMK, Zhu X, Robinson JE, van den Munkhof HE, McHenry JA, et al. (2020). Prepronociceptin-Expressing Neurons in the Extended Amygdala Encode and Promote Rapid Arousal Responses to Motivationally Salient Stimuli. Cell reports 33, 108362. 10.1016/j.celrep.2020.108362. [PubMed: 33176134]
- Root CM, Ko KI, Jafari A, and Wang JW (2011). Presynaptic facilitation by neuropeptide signaling mediates odor-driven food search. Cell 145, 133–144. 10.1016/j.cell.2011.02.008. [PubMed: 21458672]
- Rossi MA, Basiri ML, McHenry JA, Kosyk O, Otis JM, van den Munkhof HE, Bryois J, Hubel C, Breen G, Guo W, et al. (2019). Obesity remodels activity and transcriptional state of a lateral hypothalamic brake on feeding. Science 364, 1271–1274. 10.1126/science.aax1184. [PubMed: 31249056]
- Sabatini BL, and Tian L (2020). Imaging Neurotransmitter and Neuromodulator Dynamics In Vivo with Genetically Encoded Indicators. Neuron 108, 17–32. 10.1016/j.neuron.2020.09.036. [PubMed: 33058762]
- Saper CB, Fuller PM, Pedersen NP, Lu J, and Scammell TE (2010). Sleep state switching. Neuron 68, 1023–1042. 10.1016/j.neuron.2010.11.032. [PubMed: 21172606]
- Saunders A, Macosko EZ, Wysoker A, Goldman M, Krienen FM, de Rivera H, Bien E, Baum M, Bortolin L, Wang S, et al. (2018). Molecular Diversity and Specializations among the Cells of the Adult Mouse Brain. Cell 174, 1015–1030 e1016. 10.1016/j.cell.2018.07.028. [PubMed: 30096299]
- Sayin S, De Backer JF, Siju KP, Wosniack ME, Lewis LP, Frisch LM, Gansen B, Schlegel P, Edmondson-Stait A, Sharifi N, et al. (2019). A Neural Circuit Arbitrates between Persistence and Withdrawal in Hungry Drosophila. Neuron 104, 544–558 e546. 10.1016/j.neuron.2019.07.028. [PubMed: 31471123]
- Schneider DM, Nelson A, and Mooney R (2014). A synaptic and circuit basis for corollary discharge in the auditory cortex. Nature 513, 189–194. 10.1038/nature13724. [PubMed: 25162524]
- Schretter CE, Aso Y, Robie AA, Dreher M, Dolan MJ, Chen N, Ito M, Yang T, Parekh R, Branson KM, and Rubin GM (2020). Cell types and neuronal circuitry underlying female aggression in Drosophila. Elife 9. 10.7554/eLife.58942.
- Schroder S, Steinmetz NA, Krumin M, Pachitariu M, Rizzi M, Lagnado L, Harris KD, and Carandini M (2020). Arousal Modulates Retinal Output. Neuron 107, 487–495 e489. 10.1016/ j.neuron.2020.04.026. [PubMed: 32445624]

- Schwarz LA, Miyamichi K, Gao XJ, Beier KT, Weissbourd B, DeLoach KE, Ren J, Ibanes S, Malenka RC, Kremer EJ, and Luo L (2015). Viral-genetic tracing of the input-output organization of a central noradrenaline circuit. Nature 524, 88–92. 10.1038/nature14600. [PubMed: 26131933]
- Schwarz TL, Harris-Warrick RM, Glusman S, and Kravitz EA (1980). A peptide action in a lobster neuromuscular preparation. J Neurobiol 11, 623–628. 10.1002/neu.480110611. [PubMed: 6108351]
- Segalin C, Williams J, Karigo T, Hui M, Zelikowsky M, Sun JJ, Perona P, Anderson DJ, and Kennedy A (2021). The Mouse Action Recognition System (MARS) software pipeline for automated analysis of social behaviors in mice. Elife 10. 10.7554/eLife.63720.
- Sengupta P (2013). The belly rules the nose: feeding state-dependent modulation of peripheral chemosensory responses. Current opinion in neurobiology 23, 68–75. 10.1016/ j.conb.2012.08.001. [PubMed: 22939570]
- Shafer OT, and Keene AC (2021). The Regulation of Drosophila Sleep. Current biology : CB 31, R38–R49. 10.1016/j.cub.2020.10.082. [PubMed: 33434488]
- Smith SJ, Sumbul U, Graybuck LT, Collman F, Seshamani S, Gala R, Gliko O, Elabbady L, Miller JA, Bakken TE, et al. (2019). Single-cell transcriptomic evidence for dense intracortical neuropeptide networks. eLife 8. 10.7554/eLife.47889.
- Steinmetz NA, Aydin C, Lebedeva A, Okun M, Pachitariu M, Bauza M, Beau M, Bhagat J, Bohm C, Broux M, et al. (2021). Neuropixels 2.0: A miniaturized high-density probe for stable, long-term brain recordings. Science 372. 10.1126/science.abf4588.
- Steinmetz NA, Zatka-Haas P, Carandini M, and Harris KD (2019). Distributed coding of choice, action and engagement across the mouse brain. Nature 576, 266–273. 10.1038/s41586-019-1787-x. [PubMed: 31776518]
- Steriade M, McCormick DA, and Sejnowski TJ (1993). Thalamocortical oscillations in the sleeping and aroused brain. Science 262, 679–685. 10.1126/science.8235588. [PubMed: 8235588]
- Stern S, Kirst C, and Bargmann CI (2017). Neuromodulatory Control of Long-Term Behavioral Patterns and Individuality across Development. Cell 171, 1649–1662 e1610. 10.1016/ j.cell.2017.10.041. [PubMed: 29198526]
- Sternson SM (2013). Hypothalamic survival circuits: blueprints for purposive behaviors. Neuron 77, 810–824. 10.1016/j.neuron.2013.02.018. [PubMed: 23473313]
- Stih V, Petrucco L, Kist AM, and Portugues R (2019). Stytra: An open-source, integrated system for stimulation, tracking and closed-loop behavioral experiments. PLoS Comput Biol 15, e1006699. 10.1371/journal.pcbi.1006699. [PubMed: 30958870]
- Stringer C, Pachitariu M, Steinmetz N, Carandini M, and Harris KD (2019). High-dimensional geometry of population responses in visual cortex. Nature 571, 361–365. 10.1038/ s41586-019-1346-5. [PubMed: 31243367]
- Strother JA, Wu ST, Rogers EM, Eliason JLM, Wong AM, Nern A, and Reiser MB (2018). Behavioral state modulates the ON visual motion pathway of Drosophila. Proc Natl Acad Sci U S A 115, E102–E111. 10.1073/pnas.1703090115. [PubMed: 29255026]
- Suver MP, Mamiya A, and Dickinson MH (2012). Octopamine neurons mediate flight-induced modulation of visual processing in Drosophila. Curr Biol 22, 2294–2302. 10.1016/ j.cub.2012.10.034. [PubMed: 23142045]
- Swensen AM, and Marder E (2000). Multiple peptides converge to activate the same voltagedependent current in a central pattern-generating circuit. J Neurosci 20, 6752–6759. [PubMed: 10995818]
- Swensen AM, and Marder E (2001). Modulators with convergent cellular actions elicit distinct circuit outputs. J Neurosci 21, 4050–4058. [PubMed: 11356892]
- Taghert PH, and Nitabach MN (2012). Peptide neuromodulation in invertebrate model systems. Neuron 76, 82–97. S0896–6273(12)00801-X [pii] 10.1016/j.neuron.2012.08.035. [PubMed: 23040808]
- Taylor SR, Santpere G, Weinreb A, Barrett A, Reilly MB, Xu C, Varol E, Oikonomou P, Glenwinkel L, McWhirter R, et al. (2021). Molecular topography of an entire nervous system. Cell 184, 4329–4347 e4323. 10.1016/j.cell.2021.06.023. [PubMed: 34237253]

- Thompson AJ, and Lummis SC (2006). 5-HT3 receptors. Curr Pharm Des 12, 3615–3630. 10.2174/138161206778522029. [PubMed: 17073663]
- Thornquist SC, and Crickmore MA (2020). Behavioural choice emerges from nonlinear all-to-all interactions between drives. bioRxiv, 2020.2003.2012.989574. 10.1101/2020.03.12.989574.
- Thornquist SC, Langer K, Zhang SX, Rogulja D, and Crickmore MA (2020). CaMKII Measures the Passage of Time to Coordinate Behavior and Motivational State. Neuron 105, 334–345 e339. 10.1016/j.neuron.2019.10.018. [PubMed: 31786014]
- Thornquist SC, Pitsch MJ, Auth CS, and Crickmore MA (2021). Biochemical evidence accumulates across neurons to drive a network-level eruption. Mol Cell 81, 675–690 e678. 10.1016/ j.molcel.2020.12.029. [PubMed: 33453167]
- Tinbergen N (1951). The Study of Instinct (Clarendon Press).
- Tingley D, McClain K, Kaya E, Carpenter J, and Buzsaki G (2021). A metabolic function of the hippocampal sharp wave-ripple. Nature 597, 82–86. 10.1038/s41586-021-03811-w. [PubMed: 34381214]
- Todd JG, Kain JS, and de Bivort BL (2017). Systematic exploration of unsupervised methods for mapping behavior. Phys Biol 14, 015002. 10.1088/1478-3975/14/1/015002. [PubMed: 28166059]
- Tovote P, Fadok JP, and Luthi A (2015). Neuronal circuits for fear and anxiety. Nature reviews. Neuroscience 16, 317–331. 10.1038/nrn3945. [PubMed: 25991441]
- Tsao CH, Chen CC, Lin CH, Yang HY, and Lin S (2018). Drosophila mushroom bodies integrate hunger and satiety signals to control innate food-seeking behavior. Elife 7. 10.7554/eLife.35264.
- Tunbak H, Vazquez-Prada M, Ryan TM, Kampff AR, and Dreosti E (2020). Whole-brain mapping of socially isolated zebrafish reveals that lonely fish are not loners. Elife 9. 10.7554/eLife.55863.
- Turek M, Besseling J, Spies JP, Konig S, and Bringmann H (2016). Sleep-active neuron specification and sleep induction require FLP-11 neuropeptides to systemically induce sleep. Elife 5. 10.7554/ eLife.12499.
- Turek M, Lewandrowski I, and Bringmann H (2013). An AP2 transcription factor is required for a sleep-active neuron to induce sleep-like quiescence in C. elegans. Curr Biol 23, 2215–2223. 10.1016/j.cub.2013.09.028. [PubMed: 24184105]
- Tye KM (2018). Neural Circuit Motifs in Valence Processing. Neuron 100, 436–452. 10.1016/ j.neuron.2018.10.001. [PubMed: 30359607]
- Tye KM, and Deisseroth K (2012). Optogenetic investigation of neural circuits underlying brain disease in animal models. Nature reviews. Neuroscience 13, 251–266. 10.1038/nrn3171. [PubMed: 22430017]
- Urai AE, Doiron B, Leifer AM, and Churchland AK (2022). Large-scale neural recordings call for new insights to link brain and behavior. Nature neuroscience 25, 11–19. 10.1038/ s41593-021-00980-9. [PubMed: 34980926]
- van de Bospoort R, Farina M, Schmitz SK, de Jong A, de Wit H, Verhage M, and Toonen RF (2012). Munc13 controls the location and efficiency of dense-core vesicle release in neurons. J Cell Biol 199, 883–891. 10.1083/jcb.201208024. [PubMed: 23229896]
- van den Pol AN (2012). Neuropeptide transmission in brain circuits. Neuron 76, 98–115. 10.1016/ j.neuron.2012.09.014. [PubMed: 23040809]
- Vinck M, Bos JJ, Van Mourik-Donga LA, Oplaat KT, Klein GA, Jackson JC, Gentet LJ, and Pennartz CM (2015). Cell-Type and State-Dependent Synchronization among Rodent Somatosensory, Visual, Perirhinal Cortex, and Hippocampus CA1. Front Syst Neurosci 9, 187. 10.3389/ fnsys.2015.00187. [PubMed: 26834582]
- Vogt K, Zimmerman DM, Schlichting M, Hernandez-Nunez L, Qin S, Malacon K, Rosbash M, Pehlevan C, Cardona A, and Samuel ADT (2021). Internal state configures olfactory behavior and early sensory processing in Drosophila larvae. Sci Adv 7. 10.1126/sciadv.abd6900.
- von Buchholtz LJ, Ghitani N, Lam RM, Licholai JA, Chesler AT, and Ryba NJP (2021). Decoding Cellular Mechanisms for Mechanosensory Discrimination. Neuron 109, 285–298 e285. 10.1016/ j.neuron.2020.10.028. [PubMed: 33186546]
- von Philipsborn AC, Liu T, Yu JY, Masser C, Bidaye SS, and Dickson BJ (2011). Neuronal control of Drosophila courtship song. Neuron 69, 509–522. 10.1016/j.neuron.2011.01.011. [PubMed: 21315261]

- Walter T, and Couzin ID (2021). TRex, a fast multi-animal tracking system with markerless identification, and 2D estimation of posture and visual fields. Elife 10. 10.7554/eLife.64000.
- Wang L, Chen IZ, and Lin D (2015). Collateral pathways from the ventromedial hypothalamus mediate defensive behaviors. Neuron 85, 1344–1358. 10.1016/j.neuron.2014.12.025. [PubMed: 25754823]
- Weber F, and Dan Y (2016). Circuit-based interrogation of sleep control. Nature 538, 51–59. 10.1038/ nature19773. [PubMed: 27708309]
- Wee CL, Nikitchenko M, Wang WC, Luks-Morgan SJ, Song E, Gagnon JA, Randlett O, Bianco IH, Lacoste AMB, Glushenkova E, et al. (2019a). Zebrafish oxytocin neurons drive nocifensive behavior via brainstem premotor targets. Nature neuroscience 22, 1477–1492. 10.1038/s41593-019-0452-x. [PubMed: 31358991]
- Wee CL, Song EY, Johnson RE, Ailani D, Randlett O, Kim JY, Nikitchenko M, Bahl A, Yang CT, Ahrens MB, et al. (2019b). A bidirectional network for appetite control in larval zebrafish. Elife 8. 10.7554/eLife.43775.
- Weissbourd B, Momose T, Nair A, Kennedy A, Hunt B, and Anderson DJ (2021). A genetically tractable jellyfish model for systems and evolutionary neuroscience. Cell 184, 5854–5868 e5820. 10.1016/j.cell.2021.10.021. [PubMed: 34822783]
- Weissbourd B, Ren J, DeLoach KE, Guenthner CJ, Miyamichi K, and Luo L (2014). Presynaptic partners of dorsal raphe serotonergic and GABAergic neurons. Neuron 83, 645–662. 10.1016/ j.neuron.2014.06.024. [PubMed: 25102560]
- White JG, Southgate E, Thomson JN, and Brenner S (1986). The structure of the nervous system of the nematode Caenorhabditis elegans. Philos Trans R Soc Lond B Biol Sci 314, 1–340. 10.1098/ rstb.1986.0056. [PubMed: 22462104]
- Wiltschko AB, Johnson MJ, Iurilli G, Peterson RE, Katon JM, Pashkovski SL, Abraira VE, Adams RP, and Datta SR (2015). Mapping Sub-Second Structure in Mouse Behavior. Neuron 88, 1121– 1135. 10.1016/j.neuron.2015.11.031. [PubMed: 26687221]
- Wiltschko AB, Tsukahara T, Zeine A, Anyoha R, Gillis WF, Markowitz JE, Peterson RE, Katon J, Johnson MJ, and Datta SR (2020). Revealing the structure of pharmacobehavioral space through motion sequencing. Nat Neurosci 23, 1433–1443. 10.1038/s41593-020-00706-3. [PubMed: 32958923]
- Wolff SB, and Olveczky BP (2018). The promise and perils of causal circuit manipulations. Current opinion in neurobiology 49, 84–94. 10.1016/j.conb.2018.01.004. [PubMed: 29414070]
- Wu Z, Autry AE, Bergan JF, Watabe-Uchida M, and Dulac CG (2014). Galanin neurons in the medial preoptic area govern parental behaviour. Nature 509, 325–330. 10.1038/nature13307. [PubMed: 24828191]
- Xu S, Yang H, Menon V, Lemire AL, Wang L, Henry FE, Turaga SC, and Sternson SM (2020). Behavioral state coding by molecularly defined paraventricular hypothalamic cell type ensembles. Science 370. 10.1126/science.abb2494.
- Yap EL, and Greenberg ME (2018). Activity-Regulated Transcription: Bridging the Gap between Neural Activity and Behavior. Neuron 100, 330–348. 10.1016/j.neuron.2018.10.013. [PubMed: 30359600]
- Yapici N, Cohn R, Schusterreiter C, Ruta V, and Vosshall LB (2016). A Taste Circuit that Regulates Ingestion by Integrating Food and Hunger Signals. Cell 165, 715–729. 10.1016/ j.cell.2016.02.061. [PubMed: 27040496]
- Yartsev MM (2017). The emperor's new wardrobe: Rebalancing diversity of animal models in neuroscience research. Science 358, 466–469. 10.1126/science.aan8865. [PubMed: 29074765]
- Yizhar O, Fenno LE, Prigge M, Schneider F, Davidson TJ, O'Shea DJ, Sohal VS, Goshen I, Finkelstein J, Paz JT, et al. (2011). Neocortical excitation/inhibition balance in information processing and social dysfunction. Nature 477, 171–178. 10.1038/nature10360. [PubMed: 21796121]
- Yokogawa T, Hannan MC, and Burgess HA (2012). The dorsal raphe modulates sensory responsiveness during arousal in zebrafish. J Neurosci 32, 15205–15215. 10.1523/ JNEUROSCI.1019-12.2012. [PubMed: 23100441]

- York RA, Carreira-Rosario A, Giocomo LM, and Clandinin TR (2021). Flexible analysis of animal behavior via time-resolved manifold embedding. bioRxiv, 2020.2009.2030.321406. 10.1101/2020.09.30.321406.
- Yu Y, Huang R, Ye J, Zhang V, Wu C, Cheng G, Jia J, and Wang L (2016). Regulation of starvationinduced hyperactivity by insulin and glucagon signaling in adult Drosophila. Elife 5. 10.7554/ eLife.15693.
- Zelikowsky M, Ding K, and Anderson DJ (2018). Neuropeptidergic Control of an Internal Brain State Produced by Prolonged Social Isolation Stress. Cold Spring Harbor symposia on quantitative biology 83, 97–103. 10.1101/sqb.2018.83.038109. [PubMed: 30948452]
- Zhang SX, Lutas A, Yang S, Diaz A, Fluhr H, Nagel G, Gao S, and Andermann ML (2021). Hypothalamic dopamine neurons motivate mating through persistent cAMP signalling. Nature 597, 245–249. 10.1038/s41586-021-03845-0. [PubMed: 34433964]
- Zhang SX, Rogulja D, and Crickmore MA (2016). Dopaminergic Circuitry Underlying Mating Drive. Neuron 91, 168–181. 10.1016/j.neuron.2016.05.020. [PubMed: 27292538]
- Zhang SX, Rogulja D, and Crickmore MA (2019). Recurrent Circuitry Sustains Drosophila Courtship Drive While Priming Itself for Satiety. Current biology : CB 29, 3216–3228 e3219. 10.1016/ j.cub.2019.08.015. [PubMed: 31474539]
- Zimmerman CA, Leib DE, and Knight ZA (2017). Neural circuits underlying thirst and fluid homeostasis. Nature reviews. Neuroscience 18, 459–469. 10.1038/nrn.2017.71. [PubMed: 28638120]

#### Box 1:

#### Methods for computational analysis of animal behavior.

There has been a recent proliferation of techniques aimed at providing high throughput, automated behavioral tracking and classification. These advances in behavioral analyses have been especially aided by the expansion of computational tools. Particularly, recent technological advances in machine-vision and machine-learning have revolutionized the capacities to automatically track, classify, and decode animal behavior. Artificial deep neuronal networks are a rich addition to the field of behavioral assessment and may be the foundation of a totally new field of computational neuroethology (Datta et al., 2019). Recently developed methods to measure animal behavior in different species include Stytra (Stih et al., 2019), TRex (Walter and Couzin, 2021), Ctrax (Branson et al., 2009), JAABA (Kabra et al., 2013), Optimouse (Ben-Shaul, 2017), LEAP (Pereira et al., 2019), DeepLabCut (Mathis et al., 2018), DeepEthogram (Bohnslav et al., 2021), DeepPoseKit (Graving et al., 2019), DANNCE (Dunn et al., 2021), MARS (Segalin et al., 2021) or a 3D virtual mouse (Bolanos et al., 2021). These methods allow for tracking everything from body parts to multi-action behavioral motifs. Details of these novel approaches can be found in a number of authoritative reviews published recently (Datta et al., 2019; Mathis and Mathis, 2020; Pereira et al., 2020).

#### Box 2:

#### Challenges and caveats for the manipulation of state-triggering neurons.

Optogenetic, chemogenetic, and thermogenetic techniques can allow for targeted manipulation of state-promoting neurons, but these approaches may not reproduce the natural dynamics of these cells recorded in vivo. While some molecularly-defined subpopulations of neurons show concerted neural activity that can be reasonably approximated with optogenetic perturbations (i.e.: mouse AGRP neurons; (Betley et al., 2015; Chen et al., 2015; Mandelblat-Cerf et al., 2015)), other populations show complex dynamics within a molecularly-defined subpopulation (i.e. mouse VMHvl neurons (Falkner et al., 2014; Karigo et al., 2021; Remedios et al., 2017)). In addition, state-triggering neurons may fluctuate on various timescales, from slow tracking of homeostatic features (Sternson, 2013; Zimmerman et al., 2017) to faster activity of arousal-associated neurons, which can track bias in behavioral (i.e. reaction time) and physiological (i.e. pupil diameter) measures (Maimon, 2011; McCormick et al., 2020; McGinley et al., 2015b). Manipulating the activity of neurons across fast and slow timescales, while accounting for their potentially different effects (Hong et al., 2018; Otchy et al., 2015; Wolff and Olveczky, 2018), remains a challenge. In addition, many neurons with state-related activity may not necessarily be able to evoke the same state upon stimulation (Lovett-Barron et al., 2017).

With these caveats in mind, we should be critical about whether or not artificial activation appears to trigger seemingly "normal" behavioral manifestations of internal states. Are many manipulations sufficiently natural enough, or constrained by the properties of downstream circuits to remain within the relevant neural population space (Jazayeri and Afraz, 2017; Wolff and Olveczky, 2018)? Are conventional manipulations of neuromodulatory cell types routinely achieving saturating effects on downstream populations (Coddington and Dudman, 2018)? Are our measurements too coarse to discern the difference between natural and unnatural triggered states (eg., measuring effects through neuron spike rates, overt behavior, or cortical EEG, for example), and would more nuanced measurements resolve these distinction (eg., measuring effects through ionic conductance, context-dependent ethograms, or manifold of population dynamics)?

In general, a better capacity to precisely match and perturb aspects of natural activity should reveal which components of neural dynamics are important or dispensable for the initiation, persistence, and multiplexing of internal states.



#### Figure 1. Features of an example internal state

Using fear in rodents as an example, we show how a central internal state can exhibit multiple features and influence a number of behavioral and physiological processes. Hallmark characteristics of an internal state, including persistence, scalability, and generalizability, are illustrated at left and pleiotropic effects associated with the state of fear are displayed on the right.



# **Figure 2.** Approaches to infer the presence of internal states from observable behavior (A) Measuring overt behavior by tracking animal movement (examples: keypoint-based pose tracking in lemurs and nematodes).

(B) Inducing need states through environmental control (examples: social or caloric deprivation in rodents).

(C) Inferring internal state from transitions in observable movements (example: fly wing extension during courtship).

(D) Inferring states from the co-occurrence of multiple behavioral features (example: hunting states of larval zebrafish).

(E) Multiple states can interact with one another (example: a hungry rodent may show less fear when foraging under predation).

(F) State expression can vary across individuals (example: a rodent's position in a social hierarchy influences their aggressivity and response to stress).



Figure 3. Collateralized projections and brain-wide influence of state-inducing neurons (A) Schematic of projections from AGRP+ hunger-promoting neurons (red) in the arcuate nucleus of the mouse hypothalamus.

(B) Schematic of projections from P1 social arousal-promoting neurons (red) in the fly.

(C) Schematic of projections from the serotonergic NSM neuron (red) that promotes dwelling states in the nematode.

(D) Stimulating thirst-promoting neurons in the lamina terminals recapitulates the effects of natural thirst on behavior (bottom left) and neural populations recorded in multiple brain regions (right; from Allen et al., 2019).



**Figure 4. Fan-in and fan-out organization of internal states and neuromodulatory neurons** Top: internal states are influenced by the integration of multiple sensory, motor, and internal factors and themselves influence multiple behaviors and physiological processes. Bottom: similarly, many state-inducing neuromodulatory cell types integrate inputs from multiple brain regions and send outputs to multiple downstream regions.



#### Figure 5. The broad reach and diverse cellular effects of neuromodulators

(A) Examples of broadly projecting neuromodulatory neurons in larval zebrafish (Herget et al., 2017). adult fly (Deng et al., 2019), and mouse (Li et at., 2018).

(B) Neuromodulation can target neurons across the spatial extent of the brain but. within target regions, acts at the scale of intracellular signaling.

(C) Schematics of various neuromodulatory signaling mechanisms in neurons, from rapid (top) to persistent (bottom).

Flavell et al.

Page 49



#### Figure 6. Opposing brain states engage mutually exclusive neural populations

(A) Roaming and dwelling states in *C. elegans* are supported by opposing sets of neurons that mutually inhibit each other (Ji et al., 2021).

(B) Separate brain-wide populations regulate roaming versus dwelling states in hunting larval zebrafish (Marques et al., 2020).

(C) Exploration versus anxiety engage different populations of neurons in the mouse amygdala (Gründemann et al., 2019).





(A) Schematics of persistent neural and behavioral responses to transient sensory stimuli.(B) One potential mechanism for generating neuronal persistence is stowty evolving biochemical signaling within neurons, which has been demonstrated to control the persistence of internal states in flies and mammals (Zhang et al. 2019, 2021; Thornquist et al. 2021).

(C) Another potential mechanism is recurrent excitation among interconnected neurons, as has been recently demonstrated to maintain persistent defensive behaviors in flies and rodents (Jung et al., 2020; Kennedy et al., 2020).