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MELANOMA (RJ SULLIVAN, SECTION EDITOR)

Prognostic Biomarkers for Melanoma Immunotherapy

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10 Abstract

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Purpose of Review Recent developments in immunotherapy have transformed the landscape of melanoma therapy. Here, we review markers for response to immunotherapy.

13 **Recent Findings** Current immunotherapies disable immune checkpoints on T cells and other immune cells and allow immune

14 rejection of tumor. This process depends crucially on a preexisting response to the development of the melanoma. Here we

describe the complexity of the anti-tumor immune response and the links to the development of markers that are currently used or under investigation in the clinic.

17 Summary We describe immune response biomarkers along with new developments that could translate into advances.

18 **Keywords** Immune checkpoint inhibitors · Programmed death-1 (PD-1) · Programmed death ligand-1 (PD-L1) · Exhausted T cells

19 (Tex) · Tumor microenvironment (TME) · Memory T cells (Tmem) · Tumor mutation burden · Circulating tumor DNA

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21 Introduction

Melanoma treatment has been transformed recently by the 22development of rapidly accelerated fibrosarcoma (RAF) and 23mitogen-activated protein (MAP) kinase inhibitors [1-4] and 24by immune checkpoint Inhibitors (CPI) such as anti-PD-1 and 25anti-CTLA-4 [5], with many patients deriving long-term clin-26ical benefit [1, 3, 6]. However, these durable responses still 27occur only in a fraction of patients and can be associated with 2829significant toxicity, particularly when used in combination. In this review, we focus on our current understanding of the 30 mechanism of action of immunotherapy and on biomarkers 31

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to select patients for treatment on clinical trials and for particular therapies in the clinic. 33

The stages of neoplastic transformation and associated mo-34 lecular alterations have been well described for melanoma 35[7–9], yet complex tumoral/stromal/immune interactions re-36 sult in tumor heterogeneity that is evident in patients with the 37 same histological signatures as well as between tumors within 38 the same patient and even within different areas of a single 39 tumor [10-12]. The mechanisms fundamental to CPI and spe-40 cifically to anti-PD-1/PD-L1 activity in a varied and often 41 complex tumor microenvironment (TME) have led to the 42identification of a multifactorial process dependent on the in-43teractions of specific cell types with diverse functions. 44

Current Understanding of the Mechanism(s) of Action45of PD-1 Check Point Inhibitors46

Tumor immunogenicity or the ability of the tumor to trigger a 47 productive immune response is arguably fundamental to all 48 effective anti-cancer therapies, including some chemo/radio 49therapies and targeted therapies but especially immunother-50apies. While melanoma is widely recognized as an immuno-51genic tumor overall, great variability in immunogenicity is 52evident during disease progression and between patients or 53even lesions, which may shed light on the effectiveness of 54anti-PD-1 therapies [13]. Ultraviolet radiation coupled with 55key molecular changes which are the primary drivers in the 56

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malignant transformation of melanocytes often produces an 57exceptionally high rate of somatic mutations [7, 14]. These 58mutations which promote tumorigenesis by coordinated dys-5960 regulation of cellular processes are also central to its immuno-61 genicity with the emergence of neoantigens, as well as increased cancer/testis or differentiation antigens [15]. 62 63 Importantly, immunogenicity is not static; indeed, coevolution of the immune system with the tumor initiates be-64 fore neoplastic transformation. Immune pressure either elimi-65 66 nates developing tumors or steers them towards an equilibri-67 um and ultimately to tumor escape by immune evasion as well 68 as by direct and indirect subversion of the immune response 69 itself [16, 17]. The process of immunoediting highlights not only the dynamic nature of immunogenicity but also provides 70insight into the complexity and evolving spatial/temporal in-71terplay between the tumor and immune response, which lies at 7273the heart of the effectiveness of CPI immunotherapy.

74Activated lymphocytes including NK and T cells transient-75ly express PD-1 on their cell surface, which in the melanoma setting may represent recently engaged tumor-specific T cells. 76Significantly, translational analysis of adoptive cellular thera-77py patients has identified that PD-1+ rather than PD-1- CD8+ 7879tumor infiltrating lymphocytes (TIL) conferred superior oligoclonal expansion of tumor-reactive TCRB clonotypes, 80 suggesting that PD-1 expression may mark a population of 81 82 anti-tumor CTL [18]. However, chronic TCR signaling can lead to sustained PD-1 expression and the triggering of im-83 mune adaptation, a physiological reaction to curb an inappro-84 priate or autoimmune response that can be usurped by tumor 85 cells to promote peripheral tolerance. Sustained PD-1 expres-86 sion along with increasing co-expression of additional 87 88 markers such as CTLA-4, TIM-3, LAG-3, TIGIT, and VISTA denotes the transition from an activated effector 89 90 (Teff) to an exhausted T cell (Tex). In a typical melanoma setting replete with chronic antigen stimulation, T cells tend 9192to exist on a continuum that ranges from a state of stemness towards dysfunction with effector or memory-like states being 93 94key intermediates. Identification and interrogation of these progressively differentiating T cell subsets are paramount to 95appreciating an effective anti-PD-1 therapy [19, 20]. 96

97 The use of a single or limited set of markers is likely to be 98 inaccurate in discriminating between transitioning immune populations as some markers such as PD-1 are shared between 99 100immune subsets. Instead, linked functional characteristics can more readily delineate these discrete intratumoral states with a 101102 transition from high levels of cytotoxic molecules 103(granzymes/perforin) and effector cytokines (Interferons, IL-2, IL-12, and TNF), a high proliferative capacity and anabolic 104metabolism associated with an effector subset to limited or 105absent cytotoxic/effector molecules, low proliferative capaci-106107ty, and catabolic metabolism associated with dysfunctional or 108highly exhausted T cells [21]. Additionally, specific transcription factors, gene expression profiles, and epigenetic 109

signatures vield an even finer picture of these subsets while 110providing insight into their respective functions [19]. T-bet, a 111 transcription factor associated with Th1-biased response, clas-112sically associated with Teff cells also plays a role in Tex cells 113[22]. The transcription factors NR4A, EOMES, and TOX all 114 have been associated with a Tex lineage while TCF1 addition-115ally drives a stem-like progenitor lineage capable of self-re-116newal, while seeding Teff cells and memory T cells (Tmem) 117cells [19, 23, 24]. Recent advances in transcriptomic (scRNA-118 seq) and epigenetic (ATAC-seq) analyses effects of anti-PD-1 119blockade have identified gene and epigenetic signatures asso-120ciated with these subsets along with their key regulators [25•, 121 26]. 122

Understanding the functional role that these discrete T cell 123populations play during PD-1 blockade continues to be a high 124priority with the initial focus being on exhausted T cells. 125Interestingly, a high frequency of tumor infiltrating CD8+ T 126cells expressing PD-1+/CTLA-4+, a subset of exhausted T 127cells is highly predictive of anti-PD-1 CPI response in meta-128static melanoma patients [27•, 28]. Conversely, a low 129intratumoral frequency of these Tex cells exhibited a negative 130predictive value with single-agent anti-PD1 therapy yet this 131low frequency was not associated with poor outcomes in pa-132tients treated with combination anti-CTLA-4 and anti-PD-1, 133suggesting a non-redundant mechanism with the combination 134CPI [28]. Similarly, circulating peripheral Tex cells (PD-1+/ 135CTLA-4+ CD8+ T cells) were found to be "reinvigorated" 136during anti-PD-1 therapy of melanoma, which was associated 137with a positive clinical outcome particularly in patients with a 138larger ratio of proliferating Tex cells to tumor burden, defined 139as the sum of the long axis of all measurable lesions reported 140on the pre-therapy imaging reports [29•]. While these studies 141suggest that tumor-reactive Tex are a major target of PD-1 CPI 142with functional roles both in the TME and systemically, a 143chronic LCMV model demonstrated that the anti-PD-1 rein-144vigoration is transient, and exhaustion persists shortly after 145PD-1 CPI treatment due to a stable epigenetic signature 146[25•,30]. This functional maintenance of exhaustion may ac-147count for the narrow proliferative burst of Tex when on treat-148ment and clarify its relationship with tumor burden; beyond 149the clear prognostic consideration, a larger tumor burden can 150more easily withstand the effector response associated with a 151transient proliferative burst while readily providing chronic 152antigenic stimulation in addition to PD-L1 or other inhibitory 153immune signals in the TME required to maintain T cell ex-154haustion. The dysfunctional state associated with Tex epige-155netic program, particularly in the EomeshiPD-1hi subset, is 156unable to sustain a memory-like response with PD-1 block-157ade, which is critical for effective tumor immunity and re-158mains distinct from naïve T cells (Tnaive), Teff, and Tmem 159cells [25•, 26]. The fate of Tex cells underscores that while 160 exhausted T cells can be useful to predict response and can 161certainly contribute to the efficacy of PD-1 blockade when 162

163 tumor burden is low or if PD-1 blockade occurs before
164 reaching a late dysfunctional state, immune subsets capable
165 of self-renewal and persistence in the presence of chronic
166 antigen are likely required for sustained responses.

167 The necessary role of Teff cells in a productive anti-tumor immune response has been well documented but the cell types 168 169 that maintain this pool of effectors while enabling memorylike subsets continue to be defined. Expression of the tran-170scription factor TCF-1 in T cells has identified a self-171renewing precursor population critical for response to immu-172notherapy both in preclinical models and in patients with mel-173174anoma [31, 32]. In a chronic LCMV model, IL-12 or other inflammatory mediators could blunt TCF-1 expression via 175STAT-4, allowing for the differentiation of KLRG1+ Teff 176[33]. However, TCF-1 in a PD-1-dependent fashion could 177suppress TCF-1- T-bet+/KLRG1+ Teff differentiation while 178179establishing a CD8+ Eomes+ Tex precursor population [24]. 180 The factors underlying the generation, maintenance, or fate 181 choices of these stem-like TCF-1+ T cells are actively being investigated. A recent study has revealed that the metabolic 182state of the TME, specifically elevated extracellular potassium 183 which induces a starvation response/autophagy and catabolic 184185metabolism, promotes stem cell-like TCF-1 expressing T cells via an epigenetic-dependent stemness-associated program 186[21]. 187

188 Other studies have demonstrated innate immune mechanisms that can also be instrumental in effective CPI therapies. 189PD-1 is expressed on natural killer (NK) cells and when en-190 191 gaged with PD-L1 can limit NK cytotoxicity. In tumors with 192 loss of MHC class I, rejection was dependent on these innate effectors which was significantly enhanced with PD-1 block-193194ade [34]. Even in models where CD8+ T cells routinely mediate tumor regression, PD-1+ NK cells demonstrated a mean-195ingful contribution to anti-PD-1 therapy with notable selection 196 197 of PD-L1+ tumor cells [34]. Further, NK cells were shown to 198 play a significant role in the efficacy of PD-1 blockade in melanoma by producing the cytokine FLT3L and forming 199200stable conjugates with CD141+ cDC1, resulting in increased frequencies of the DCs [35]. Similarly, cDC1 subsets were 201 shown to play an additional key role in the efficacy of PD-1 202203 blockade by producing IL-12, triggering IFN- γ secretion from PD-1+ T cells, which further engaged IL-12 secretion from 204DCs. The resulting IL-12/IFN- γ feed-forward loop which ini-205206 tiated with anti-PD-1 treatment helped further license theses PD-1+ T cells and enhance PD-1 blockade [36]. A separate 207study demonstrated the role of benefit in PD-1 blockade of 208209PD-1+ on CD103+ DCs that engage and activate intratumoral T cells via production of CXCL9/CXCL10 [37]. Conversely, 210increased oncogenic, *β*-catenin signaling in TME leads to 211downregulation of CCL4, which blunts the frequency of a 212213similar subset of DCs, ultimately limiting T cell recruitment [38, 39]. Beyond NK and DC subsets, PD-1+ tumor-214associated macrophages (TAM) with an M2-like phenotype, 215

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which generally are associated with poor patient outcomes, 216 can contribute to anti-tumor immunity via tumor phagocytosis 217 when in the presence of anti-PD-1 in combination with other 218 therapeutics like anti-CSFR1 or anti-SIRP α blocking antibodies [40, 41]. 220

The mechanisms fundamental to CPI and specifically anti-221PD-1/PD-L1 activity in a varied and often complex tumor 222microenvironment (TME) have led to the identification of a 223multifactorial process dependent on the interactions of specif-224ic cell types with diverse functions. While this complexity can 225pose a challenge to identify the relevant parameter(s) specific 226for a given patient or even a specific lesion, many of these 227seemingly distinct mechanisms converge on the TME with 228specific immune subsets driving a collective immunogenicity 229in turn creating a foundation for effective anti-PD-1 therapy. 230

Biomarkers of Response to CPI

PD-L1 Expression

Following the discovery of PD-1 expression on lymphocytes 233[42], the B7 family member, B7-H1, was identified as the 234ligand for PD-1 [43]. This protein, called B7-H1, was identi-235fied by Dong et al. based on its similarity to the co-stimulatory 236ligands B7-1 and B7-2 on immune cells [44]. Ligation of this 237protein in the context of antigen binding on T cells caused IL-23810 secretion. The pattern of expression of B7-H1 in malignant 239neoplasms such as melanoma was very interesting. Notably, 240B7-H1 was strongly co-localized with tumor infiltrating lym-241phocytes [45•]. In addition, interferon- γ was found at the in-242terface of B7-H1 expressing tumor cells and TILs. In this 243same study, B7-H1 (now more commonly referred to as PD-244L1) was also found to be a prognostic marker, predicting ex-245tended survival. When tumor tissue was analyzed in patients 246treated with the PD-1 blocking antibody, nivolumab, PD-L1 247expression was found to correlate with response [46]. A sim-248ilar observation was made with the PD-L1 antibody, 249atezolizumab, in a phase I multiple solid tumor trial [47]. 250These observations were confirmed with pembrolizumab 251[48]. 252

Given the clinical and translational data supporting the use 253of PD-L1 as a biomarker for response to PD-1 blockade, many 254recent trials have explored this prospectively and retrospec-255tively. In a retrospective analysis of patients treated on the 256Keynote 001 trial, tumors from 451 patients (out of 655 pa-257tients treated) were stained with the 22C3 monoclonal anti-258body [48]. Samples were assessed by a quantitative membra-259nous staining called the MEL score which incorporated inten-260sity and frequency of staining on tumor and tumor adjacent 261stromal and immune cells. A positive score was anyone with 262MEL ≥ 2 (staining in $\geq 1\%$ of cells). Of the 451 patients eval-263uated, 344 (76%) had PD-L1-positive tumors. A higher MEL 264score was associated with a higher response rate and longer 265

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266PFS (hazard ratio, 0.76: 95% CI, 0.71 to 0.82) and OS (hazard ratio, 0.76; 95% CI, 0.69 to 0.83) (P < .001 for each). The 267objective response rate was 8% for MEL 0 and ranged up to 268 26957% for MEL 4 showing the dynamic range of this marker. In 270a prospective nivolumab vs dacarbazine clinical trial [49], using the rabbit monoclonal 28-8 antibody, 5% or greater tu-271272mor cell staining was considered "positive" [50] with 2 pathologists independently scoring using an automated Dako 273stainer. In the PD-L1-positive group, 52.7% had an objective 274response to nivolumab versus 33.1% in the PD-L1-negative 275276group. Other trials in melanoma have consistently shown a 277higher response rate and higher PFS (and in some cases a higher OS) in PD-L1 high patients [6, 51-53]. Some of the 278questions that remain with PD-L1 IHC have to do with the 279difference between tumor and/or stroma and different mono-280clonal antibodies although recent data has shown that most 281282 widely used PD-L1 monoclonal antibodies are quite consis-283tent, reproducible, and inter-comparable regardless of the spe-284cific methodology used in the hands of trained experienced pathologists using recent rapidly processed specimens [54]. 285Also, PD-L1 expression has differing predictive value in ma-286lignancies arising from differing sites. In non-small cell lung 287288 cancer, in the Keynote 001 clinical trial, the response rate of pembrolizumab varied from 8.1% in the < 1% PD-L1 group to 28929.6% in the PD-L1 50-74% group [55]. In Keynote 010, 290291where chemo-naïve patients were randomized to pembrolizumab at 2 mg/kg or 10 mg/kg (or docetaxel), PD-292L1 staining of 1-49% was associated with a response rate of 293 29410% while \geq 50% PD-L1 had a response rate of 30% (all for 295the pembrolizumab cohort). Contrast these findings to nivolumab in renal cell cancer in the Checkmate 025 study 296297where PD-L1 expression was not found to significantly predict benefit from PD-1 blockade (<1%, OS was 27.4 months 298while PD-L1 \geq 1%, OS was 21.8 months). Similar results 299 300 were seen in Keynote 427, which examined first line 301 pembrolizumab in renal cell cancer; no difference in response 302 rate with PD-L1 expression [56].

Immune Cell Infiltration and "Exhausted" T (Tex) Cellsin the Tumor Microenvironment

Tumor infiltrating lymphocytes (TIL) have been shown to 305correlate with prognosis in melanoma (as well as in many 306 307 other tumor types) [57–59]. An important study, by Tumeh et al., showed that CD8+ TIL density in tumor samples was 308 higher in responding patients than in patients with disease 309 310 progression [60•]. Subsequently, it was demonstrated that CD8+ cells with dual PD-1/CTLA4 expression, or 311"exhausted" CD8+ cells, were predictive of PD-1 monother-312apy response in melanoma [27•]. Furthermore, lower levels of 313 314"exhausted" CD8+ cells were associated with response to dual 315PD-1/CTLA4 inhibitor therapy but were insufficient for monotherapy PD-1 blockade [28]. Another important study 316

by Broz et al. showed that the presence of cDC1 dendritic 317 cells in melanoma was predictive of response [61•]. These 318 findings have been extended by Spranger et al. who showed 319 that BATF + dendritic cells are characteristic of immune infil-320 trated melanoma [62]. More recently, the Krummel group 321 demonstrated that there are 2 axes in "immunogenic" tumors, 322 one a NK-cDC1 axis [35] that operates in checkpoint respon-323 sive tumors and a CD4-cDC2 axis [63]. Other axes may op-324erate in other tumor types as well. 325

Recent translational studies have shed light on the biology, 326 location, and surroundings of Tex cells. Tex have a distinctive 327 transcriptional profile that is maintained stably via large-scale 328 epigenetic programming and transcription factors [25•, 30]. 329 The presence of Tex cells in tumors can predict responses once 330 tumor burden is factored in [29•]. Thommen et al. reported 331 that these cells produce chemokines that attract B cells and 332 TFH cells that produce tertiary lymphoid structures [64]. 333 More recently, the transcription factor Tox, highly expressed 334 in exhausted T cells, appears to be critical to maintain their 335 tissue presence but not the dysfunction associated with them 336 [65–67]. There remain unanswered questions about precursor 337 and terminal exhausted T cells and the transitions possible 338 between these states [68]. 339

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Tumor Mutation Burden and MSI

Tumors with high mutation burdens appear to have an in-341 creased response rate and better survival in response to PD-1 342 immunotherapy [69]. In non-small cell lung cancer, a higher 343 non-synonymous mutation burden was associated with better 344 PFS, OS, and objective response when treated with anti-PD-1 345[70]. While some neoantigens are clonal (shared by multiple 346 sites), others are present in a more localized fashion (branch). 347 It has been hypothesized that because of the selection pressure 348 that neoantigen-directed T cells put on tumor cells, that non-349 clonal neoantigen bearing tumor cells could be edited out 350while clonal neoantigens persist and can predict for greater 351response to PD-1 inhibitors [71]. The use of tumor mutation 352burden to select patients for PD-1 therapy has not yielded 353consistent results and at present is experimental [72]. These 354studies need additional replication to be widely accepted. At 355 present, it is unclear how tumor mutation burden impacts mel-356anoma although it has been noted that uveal melanoma, which 357has a low tumor mutation burden has a low response rate to 358 PD-1 blockade [73, 74]. 359

Extremely high rates of tumor mutation burden are seen in 360 patients with mismatch repair deficiency [75]. These tumors 361also have a very high response rate to PD-1 immunotherapy 362 [76, 77] and anti-PD-1 therapy is approved for use in these 363 patients regardless of primary site. Widespread and continual 364 mutation resulting from a deficiency in DNA repair is thought 365 to generate neoantigens which in turn prime T cells [78]. 366 While melanoma is not part of Lynch syndrome and mismatch 367

368 repair deficiency is uncommon in melanoma, desmoplastic melanoma, which can have a high UV mutagenesis signature, 369 is also associated with a high response rate to anti-PD-1 ther-370 371apy [79, 80]. Basal cell cancer of the skin is the most mutated 372 non-mismatch repair deficient cancer [81]. Squamous cell cancer of the skin also has a very high mutation burden [82]. 373 374 Squamous cell cancers respond well to PD-1 blockade, while 375response rates for basal call cancers are lower [83, 84]. Merkel Cell Cancer, an uncommon skin neoplasm, also responds to 376 377 PD-1 blockade regardless of polyoma viral status [85] although only the merkel cell polyoma virus negative tumors 378 379 have a high mutation burden, presumably due to UV damage [86]. These data illustrate the complexity of this field and the 380 continuing research into the impact mutation burden has on 381PD-1 response. 382

383 Peripheral Blood Biomarkers

There is great interest in identifying peripheral blood biomarkers associated with favorable response to immunotherapy in melanoma, as these could be serially collected and offer significant safety, cost, and convenience advantages. Peripheral biomarkers could also allow for profiling of the systemic immune response in a way that tumor biopsies cannot.

Since basic peripheral blood laboratory variables are col-391lected routinely in standard clinical care, it is possible to study 392 393 these variables in large retrospective clinical studies. For ex-394 ample, Martens et al. analyzed peripheral blood biomarkers of 209 patients with advanced melanoma on ipilimumab. They 395396 found that a baseline signature of low lactate dehydrogenase (LDH), absolute monocyte count (AMC), and myeloid-397 derived suppressor cells (MDSC), as well as high absolute 398 399 eosinophil count (AEC), regulatory T cells, and relative lymphocyte count (RLC) were associated with improved out-400comes with ipilimumab therapy [87]. Similarly, Weide et al. 401 402 analyzed peripheral blood biomarkers of patients with advanced melanoma treated with pembrolizumab and found that 403 high relative eosinophil count (REC), high relative lympho-404 405 cyte count (RLC), low LDH, and absence of metastasis other than soft-tissue/lung metastases are independent baseline 406characteristics associated with favorable overall survival 407 408 [88]. Most recently, Rosner et al. evaluated peripheral blood clinical laboratory variables associated with outcomes follow-409 ing combination nivolumab and ipilimumab immunotherapy 410in melanoma. They found that significant independent vari-411 ables for favorable OS included the following: high relative 412eosinophils, high relative basophils, low absolute monocytes, 413low LDH, and a low neutrophil-to-lymphocyte ratio [89]. 414 415Further work is needed to validate these peripheral blood biomarkers in randomized controlled clinical trials. Ultimately, 416 such biomarkers could be a simple and cost-effective way to 417

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define which patients with metastatic melanoma may derive418the most benefit from immunotherapy.419

Immunotherapy and the Microbiome

Complex microbial communities, known as the microbiota, 421 colonize the mammalian host and contribute to the health of 422 the host [90]. Over the last few decades, there has been in-423 creasing evidence to suggest that the bacterial microbiome 424 plays an important role in carcinogenesis as well as the body's 425response to cancer treatment [91] [92]. While CPI therapy has 426 revolutionized the treatment of metastatic melanoma, re-427 sponse to CPI therapy is variable, with some patients achieve 428 a robust response while other patients have minimal or no 429response. One hypothesis that has emerged recently is that 430 the gut microbiome may affect response to CPI therapy, and 431thus the study of the gut microbiome can yield important clues 432 about which patients will derive the most benefit from 433 immunotherapy. 434

There is evidence in mouse models that modulation of the 435 gut microbiome may enhance responses to immune check-436point blockade, so several groups have studied whether the 437 human microbiome affects response to CPI therapy. In a study 438by Gopalakrishnan et al., the authors examined the oral and 439gut microbiome of 112 melanoma patients undergoing anti-440 PD-1 immunotherapy [93]. The authors observed significant 441 differences in the diversity and composition of the gut, but not 442 oral, microbiome of patients who responded to PD-1 therapy 443 versus those patients who did not respond, namely responders 444 had higher alpha diversity (P < 0.01), relative abundance of 445bacteria of the Ruminococcaceae family (P < 0.01), and 446 showed significantly higher alpha diversity (P < 0.01) and rel-447 ative abundance of bacteria of the Ruminococcaceae family 448 (P < 0.01) in responding patients. In a similar study analyzing 449the stool microbiota before and after anti-PD-1 therapy, 450Matson et al. showed that patients who responded to anti-451PD-1 therapy had an abundance of certain bacteria, including 452Bifidobacterium longum, Collinsella aerofaciens, and 453Enterococcus faecium compared with non-responders [94]. 454One hypothesis is that the increased bacterial diversity in some 455 patients leads to increased immune cell infiltration. Wargo 456et al. performed immune profiling of stool samples from mel-457anoma patients and demonstrated increased tumor immune 458 infiltrates in responding patients, with a higher density of 459CD8+ T cells which correlated with abundance of specific 460bacteria enriched in the gut microbiome [95]. Together, these 461studies suggest that the commensal microbiome of patients 462 may have a mechanistic impact on anti-tumor immunity. 463 Further studies are needed to better understand the precise 464 mechanisms mediating this effect, and specifically to deter-465 mine whether there are ways to modulate the microbiome to 466 affect response to treatment. 467

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468 Aside from the ways in which the gut microbiome affects response to therapy, another interesting observation is that the 469gut microbiome may influence which patients are most at risk 470 for checkpoint blockade-induced colitis. In a prospective 471 472 study of patients with metastatic melanoma undergoing ipilimumab treatment, authors correlated the pre-473 474 inflammation fecal microbiota and microbiome composition with subsequent development of colitis [96]. They observed 475that patients with a paucity of bacteria involved in polyamine 476 477 transport and B vitamin biosynthesis was associated with an 478 increased risk of colitis, whereas patients with increased rep-479 resentation of bacteria in the Bacteroidetes phylum were more 480 resistant to the development of colitis.

481 **Conclusions**

482 Recently, we have seen a rapid increase in our understanding of the mechanism of action of CPI. Melanoma has served as a 483484 model system for many functional and analytical studies. 485 While some of these laboratory advances have translated into clinical and translational studies, in many instances, the com-486 487 plexity of the immune response to tumor has stymied attempts 488 to develop markers that accurately and comprehensively profile the immune response to tumor. While it is unlikely that a 489 single biomarker or a simple combination of biomarkers can 490provide the profile we need as clinicians and translational 491492 researchers, rapid advances are underway and we expect some 493 of these advances to translate into trial and clinical use.

494 **Compliance with Ethical Standards**

495 Conflict of Interest Christopher G. Twitty declares that he has no con496 flict of interest. Laura A. Huppert declares that she has no conflict of
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498 OncoSec, Bristol-Myers Squibb, Incyte, Checkmate, and Pfizer.

Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects performed by any of the authors.

502 **References**

503 Papers of particular interest, published recently, have been504 highlighted as:

- 505 Of importance
- 5061.Holderfield M, Deuker MM, McCormick F, McMahon M.507Targeting RAF kinases for cancer therapy: BRAF-mutated mela-508noma and beyond. Nat Rev Cancer. 2014 Jul;14(7):455–67.
- Bollag G, Hirth P, Tsai J, Zhang J, Ibrahim PN, Cho H, et al. Clinical efficacy of a RAF inhibitor needs broad target blockade in BRAF-mutant melanoma. Nature. 2010 Sep 30;467(7315):596– 512

3.

4.

5.

6

7.

8.

9

10

11.

12.

13.

14.

15.

16

17

18

19

20.

21.

22.

23.

Robert C, Grob JJ, Stroyakovskiy D, Karaszewska B, Hauschild A, 513Levchenko E, et al. Five-year outcomes with dabrafenib plus 514trametinib in metastatic melanoma. N Engl J Med. 2019;381(7): 515516626-36. Flaherty KT, Robert C, Hersey P, Nathan P, Garbe C, Milhem M, 517et al. Improved survival with MEK inhibition in BRAF-mutated 518melanoma. N Engl J Med. 2012;367(2):107-14. 519Ribas A, Wolchok JD. Cancer immunotherapy using checkpoint 520blockade. Science. 2018;359(6382):1350-5. 521Larkin J, Chiarion-Sileni V, Gonzalez R, Grob JJ, Cowey CL, Lao 522CD, et al. Combined nivolumab and ipilimumab or monotherapy in 523untreated melanoma. N Engl J Med. 2015;373(1):23-34. 524Miller AJ, Mihm MC. Melanoma. N Engl J Med. 2006;355(1):51-525526 65. Shain AH, Joseph NM, Yu R, Benhamida J, Liu S, Prow T, et al. 527 Genomic and transcriptomic analysis reveals incremental disrup-528tion of key signaling pathways during melanoma evolution. 529Cancer Cell. 2018;34(1):45-55 e4. 530Shain AH, Bastian BC. From melanocytes to melanomas. Nat Rev 531Cancer. 2016;16(6):345-58. 532Angelova M, Mlecnik B, Vasaturo A, Bindea G, Fredriksen T, 533Lafontaine L, et al. Evolution of metastases in space and time under 534immune selection. Cell. 2018;175(3):751-65 e16. 535Boddupalli CS, Bar N, Kadaveru K, Krauthammer M, 536Pornputtapong N, Mai Z, et al. Interlesional diversity of T cell 537receptors in melanoma with immune checkpoints enriched in 538tissue-resident memory T cells. JCI Insight. 2016;1(21):e88955. 539Lehmann B, Biburger M, Brückner C, Ipsen-Escobedo A, Gordan 540S, Lehmann C, et al. Tumor location determines tissue-specific 541recruitment of tumor-associated macrophages and antibody-542dependent immunotherapy response. Sci Immunol. 2017;6:2(7). 543Roh W, Chen P-L, Reuben A, Spencer CN, Prieto PA, Miller JP, 544et al. Integrated molecular analysis of tumor biopsies on sequential 545CTLA-4 and PD-1 blockade reveals markers of response and resis-546tance. Sci Transl Med. 2017:01:9(379). 547Schumacher TN, Schreiber RD. Neoantigens in cancer immuno-548therapy. Science. 2015;348(6230):69-74. 549Gubin MM, Zhang X, Schuster H, Caron E, Ward JP, Noguchi T, 550et al. Checkpoint blockade cancer immunotherapy targets tumour-551specific mutant antigens. Nature. 2014;515(7528):577-81. 552Dunn GP, Old LJ, Schreiber RD. The immunobiology of cancer 553immunosurveillance and immunoediting. Immunity. 2004;21(2): 554137-48. 555O'Donnell JS, Teng MWL, Smyth MJ. Cancer immunoediting and 556resistance to T cell-based immunotherapy. Nat Rev Clin Oncol. 5572019;16(3):151-67. 558Gros A, Robbins PF, Yao X, Li YF, Turcotte S, Tran E, et al. PD-1 559identifies the patient-specific CD8⁺ tumor-reactive repertoire infil-560trating human tumors. J Clin Invest. 2014;124(5):2246-59. 561Li H, van der Leun AM, Yofe I, Lubling Y, Gelbard-Solodkin D, 562van Akkooi ACJ, et al. Dysfunctional CD8 T cells form a prolifer-563ative, dynamically regulated compartment within human melano-564ma. Cell. 2019;176(4):775-89 e18. 565Sade-Feldman M, Yizhak K, Bjorgaard SL, Ray JP, de Boer CG, 566 Jenkins RW, et al. Defining T cell states associated with response to 567checkpoint immunotherapy in melanoma. Cell. 2019;176(1-2): 568404. 569Vodnala SK, Eil R, Kishton RJ, Sukumar M, Yamamoto TN, Ha N-570H, et al. T cell stemness and dysfunction in tumors are triggered by a 571common mechanism. Science. 2019;363(6434). 572Anderson AC, Lord GM, Dardalhon V, Lee DH, Sabatos-Peyton 573574CA, Glimcher LH, et al. T-bet, a Th1 transcription factor regulates the expression of Tim-3. Eur J Immunol. 2010;40(3):859-66. 575Liu X, Wang Y, Lu H, Li J, Yan X, Xiao M, et al. Genome-wide 576analysis identifies NR4A1 as a key mediator of T cell dysfunction. 577 Nature. 2019;567(7749):525-9. 578

- 579 24. Chen J, López-Moyado IF, Seo H, Lio C-WJ, Hempleman LJ,
 580 Sekiya T, et al. NR4A transcription factors limit CAR T cell func581 tion in solid tumours. Nature. 2019;567(7749):530–4.
- 582 25.• Pauken KE, Sammons MA, Odorizzi PM, Manne S, Godec J, Khan
 583 O, et al. Epigenetic stability of exhausted T cells limits durability of
 584 reinvigoration by PD-1 blockade. Science. 2016;354(6316):1160–5
 585 Establishes the concepts and findings relating to "exhausted" T
 586 cells.
- 587 26. Bengsch B, Johnson AL, Kurachi M, Odorizzi PM, Pauken KE,
 588 Attanasio J, et al. Bioenergetic insufficiencies due to metabolic
 589 alterations regulated by the inhibitory receptor PD-1 are an early
 590 driver of CD8(+) T cell exhaustion. Immunity. 2016;45(2):358–73.
- 591 27.• Daud AI, Loo K, Pauli ML, Sanchez-Rodriguez R, Sandoval PM, 592 Taravati K, et al. Tumor immune profiling predicts response to anti-593 PD-1 therapy in human melanoma. J Clin Invest. 2016;126(9): 3447–52 Describes the connection between "exhausted" T cells 595 and PD-1 response in melanoma.
- Loo K, Tsai KK, Mahuron K, Liu J, Pauli ML, Sandoval PM, et al.
 Partially exhausted tumor-infiltrating lymphocytes predict response to combination immunotherapy. JCI Insight [Internet]. 2017;2(14)
 Available from: https://insight.jci.org/articles/view/93433.
- 400 29.• Huang AC, Postow MA, Orlowski RJ, Mick R, Bengsch B, Manne
 401 S, et al. T-cell invigoration to tumour burden ratio associated with
 402 anti-PD-1 response. Nature. 2017;545(7652):60–5 Points to rein403 vigoration of exhausted T cells in melanoma.
- 60430.Im SJ, Hashimoto M, Gerner MY, Lee J, Kissick HT, Burger MC,605et al. Defining CD8(+) T cells that provide the proliferative burst606after PD-1 therapy. Nature. 2016;537(7620):417–21.
- Kurtulus S, Madi A, Escobar G, Klapholz M, Nyman J, Christian E, et al. Checkpoint blockade immunotherapy induces dynamic changes in PD-1-CD8+ tumor-infiltrating T cells. Immunity. 2019;50(1):181–94 e6.
- 611 32. Siddiqui I, Schaeuble K, Chennupati V, Fuertes Marraco SA,
 612 Calderon-Copete S, Pais Ferreira D, et al. Intratumoral Tcf1+PD613 1+CD8+ T cells with stem-like properties promote tumor control in
 614 response to vaccination and checkpoint blockade immunotherapy.
 615 Immunity. 2019;50(1):195–211 e10.
- 516 33. Danilo M, Chennupati V, Silva JG, Siegert S, Held W. Suppression
 of Tcf1 by inflammatory cytokines facilitates effector CD8 T cell
 differentiation. Cell Rep. 2018;22(8):2107–17.
- 619 34. Hsu J, Hodgins JJ, Marathe M, Nicolai CJ, Bourgeois-Daigneault
 620 M-C, Trevino TN, et al. Contribution of NK cells to immunother621 apy mediated by PD-1/PD-L1 blockade. J Clin Invest.
 622 2018;128(10):4654-68.
- Barry KC, Hsu J, Broz ML, Cueto FJ, Binnewies M, Combes AJ, et al. A natural killer-dendritic cell axis defines checkpoint therapyresponsive tumor microenvironments. Nat Med. 2018;24(8):1178– 91.
- 627 36. Garris CS, Arlauckas SP, Kohler RH, Trefny MP, Garren S, Piot C,
 628 et al. Successful anti-PD-1 cancer immunotherapy requires T cell629 dendritic cell crosstalk involving the cytokines IFN-γ and IL-12.
 630 Immunity. 2018;49(6):1148–61 e7.
- 631 37. Chow MT, Ozga AJ, Servis RL, Frederick DT, Lo JA, Fisher DE,
 632 et al. Intratumoral activity of the CXCR3 chemokine system is
 633 required for the efficacy of anti-PD-1 therapy. Immunity.
 634 2019;50(6):1498–512 e5.
- 635 38. Spranger S, Bao R, Gajewski TF. Melanoma-intrinsic β-catenin
 636 signalling prevents anti-tumour immunity. Nature.
 637 2015;523(7559):231-5.
- 638
 39. Spranger S, Gajewski TF. Tumor-intrinsic oncogene pathways mediating immune avoidance. Oncoimmunology. 2016 Mar;5(3):
 e1086862.
- 641 40. Gordon SR, Maute RL, Dulken BW, Hutter G, George BM, McCracken MN, et al. PD-1 expression by tumour-associated macrophages inhibits phagocytosis and tumour immunity. Nature. 644 2017;545(7655):495–9.

- 41. Neubert NJ, Schmittnaegel M, Bordry N, Nassiri S, Wald N, Martignier C, et al. T cell-induced CSF1 promotes melanoma resistance to PD1 blockade. Sci Transl Med. 2018;10(436).
 42. Likik N A and Strand St
- 42. Ishida Y, Agata Y, Shibahara K, Honjo T. Induced expression of 648 PD-1, a novel member of the immunoglobulin gene superfamily, 649 upon programmed cell death. EMBO J. 1992;11(11):3887–95. 650
- 43. Freeman GJ, Long AJ, Iwai Y, Bourque K, Chernova T, Nishimura H, et al. Engagement of the PD-1 immunoinhibitory receptor by a novel B7 family member leads to negative regulation of lymphocyte activation. J Exp Med. 2000;192(7):1027–34.
 654
- 44. Dong H, Zhu G, Tamada K, Chen L. B7-H1, a third member of the B7 family, co-stimulates T-cell proliferation and interleukin-10 secretion. Nat Med. 1999 Dec;5(12):1365–9.
 45. Content of the secret s
- 45.• Taube JM, Anders RA, Young GD, Xu H, Sharma R, TL MM, et al. Colocalization of inflammatory response with B7-H1 expression in human melanocytic lesions supports an adaptive resistance mechanism of immune escape. Sci Transl Med. 2012;4(127):127ra37
 bescribes PD-L1 staining in melanoma as a marker of PD-1 response.
- 46. Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. N Engl J Med. 2012;366(26):2443– 54.
- 47. Herbst RS, Soria J-C, Kowanetz M, Fine GD, Hamid O, Gordon MS, et al. Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A in cancer patients. Nature. 2014;515(7528): 670 563–7. 671
- Daud AI, Wolchok JD, Robert C, Hwu W-J, Weber JS, Ribas A, et al. Programmed death-ligand 1 expression and response to the anti-programmed death 1 antibody pembrolizumab in melanoma. J Clin Oncol Off J Am Soc Clin Oncol. 2016;34(34):4102–9.
 Robert C, Long GV, Brady B, Dutriaux C, Maio M, Mortier L, et al.
- 49. Robert C, Long GV, Brady B, Dutriaux C, Maio M, Mortier L, et al. Nivolumab in previously untreated melanoma without BRAF mutation. N Engl J Med. 2015 Jan 22;372(4):320–30.
 678
- 50. Wolchok JD, Kluger H, Callahan MK, Postow MA, Rizvi NA, 679 Lesokhin AM, et al. Nivolumab plus ipilimumab in advanced melanoma. N Engl J Med. 2013;369(2):122–33.
 51. O. J. Med. 2013;369(2):122–33.
- 51. Carlino MS, Long GV, Schadendorf D, Robert C, Ribas A, Richtig E, et al. Outcomes by line of therapy and programmed death ligand 1 expression in patients with advanced melanoma treated with pembrolizumab or ipilimumab in KEYNOTE-006: a randomised 685 clinical trial. Eur J Cancer Oxf Engl. 2018;101:236–43.
- Tawbi HA, Forsyth PA, Algazi A, Hamid O, Hodi FS, Moschos SJ, et al. Combined nivolumab and ipilimumab in melanoma metastatic to the brain. N Engl J Med. 2018;379(8):722–30.
- 53. Hodi FS, Chiarion-Sileni V, Gonzalez R, Grob J-J, Rutkowski P, Cowey CL, et al. Nivolumab plus ipilimumab or nivolumab alone versus ipilimumab alone in advanced melanoma (CheckMate 067): 4-year outcomes of a multicentre, randomised, phase 3 trial. Lancet Oncol. 2018;19(11):1480–92.
- 54. Adam J, Le Stang N, Rouquette I, Cazes A, Badoual C, Pinot-Roussel H, et al. Multicenter harmonization study for PD-L1 IHC testing in non-small-cell lung cancer. Ann Oncol Off J Eur Soc Med Oncol. 2018;29(4):953–8.
- 55. Garon EB, Rizvi NA, Hui R, Leighl N, Balmanoukian AS, Eder JP, et al. Pembrolizumab for the treatment of non-small-cell lung cancer. N Engl J Med. 2015;372(21):2018–28.
 56. Value Content of the treatment of non-small-cell lung cancer. N Engl J Med. 2015;372(21):2018–28.
- 56. Lee J-L, Ziobro M, Gafanov R, Matveev VB, Suarez C, Donskov F, et al. KEYNOTE-427 cohort B: first-line pembrolizumab (pembro) 703 monotherapy for advanced non-clear cell renal cell carcinoma (NCC-RCC). J Clin Oncol. 2019;37(15_suppl):4569. 705
- 57. Thomas NE, Busam KJ, From L, Kricker A, Armstrong BK, Anton-Culver H, et al. Tumor-infiltrating lymphocyte grade in primary melanomas is independently associated with melanoma specific survival in the population-based genes, environment and 709

AUTHIP102 RIDS6 POR O 601 P020

Page 8 of 9

- melanoma study. J Clin Oncol Off J Am Soc Clin Oncol.
 2013;31(33):4252–9.
 58. Huh JW, Lee JH, Kim HR. Prognostic significance of tumorinfiltrating lymphocytes for patients with colorectal cancer. Arch Surg. 2012;147(4):366–72.
 59. Zeng D-Q, Yu Y-F, Ou Q-Y, Li X-Y, Zhong R-Z, Xie C-M, et al. Promostic and predictive value of tumor-infiltrating lymphocytes
- 716Prognostic and predictive value of tumor-infiltrating lymphocytes717for clinical therapeutic research in patients with non-small cell lung718cancer. Oncotarget. 2016;7(12):13765–81.
- Tumeh PC, Harview CL, Yearley JH, Shintaku IP, EJM T, Robert L,
 et al. PD-1 blockade induces responses by inhibiting adaptive immune resistance. Nature. 2014;515(7528):568–71 Describes T cell
 proliferation in response to PD-1 blockade.
- 723 61.• Broz ML, Binnewies M, Boldajipour B, Nelson AE, Pollack JL,
 724 Erle DJ, et al. Dissecting the tumor myeloid compartment reveals
 725 rare activating antigen-presenting cells critical for T cell immunity.
 726 Cancer Cell. 2014;26(5):638–52 Shows the importance of DC in
 727 determining an inflamed tumor.
- Spranger S, Dai D, Horton B, Gajewski TF. Tumor-residing Batf3
 dendritic cells are required for effector T cell trafficking and adoptive T cell therapy. Cancer Cell. 2017;31(5):711–23 e4.
- 63. Binnewies M, Mujal AM, Pollack JL, Combes AJ, Hardison EA,
 Barry KC, et al. Unleashing type-2 dendritic cells to drive protective antitumor CD4+ T cell immunity. Cell. 2019;177(3):556–71
 e16.
- 735 64. Thommen DS, Koelzer VH, Herzig P, Roller A, Trefny M, Dimeloe
 736 S, et al. A transcriptionally and functionally distinct PD-1+ CD8+ T
 737 cell pool with predictive potential in non-small-cell lung cancer
 738 treated with PD-1 blockade. Nat Med. 2018;24(7):994–1004.
- 5. Scott AC, Dündar F, Zumbo P, Chandran SS, Klebanoff CA,
 Shakiba M, et al. TOX is a critical regulator of tumour-specific T
 cell differentiation. Nature. 2019;571(7764):270–4.
- 66. Khan O, Giles JR, McDonald S, Manne S, Ngiow SF, Patel KP,
 et al. TOX transcriptionally and epigenetically programs CD8+ T
 cell exhaustion. Nature. 2019;571(7764):211–8.
- 745 67. Yao C, Sun H-W, Lacey NE, Ji Y, Moseman EA, Shih H-Y, et al.
 746 Single-cell RNA-seq reveals TOX as a key regulator of CD8+ T cell
 747 persistence in chronic infection. Nat Immunol. 2019;20(7):890–
 748 901.
- 68. Blank CU, Haining WN, Held W, Hogan PG, Kallies A, Lugli E, et al. Defining "T cell exhaustion.". Nat Rev Immunol. 2019;19(11):665–74.
- Samstein RM, Lee C-H, Shoushtari AN, Hellmann MD, Shen R,
 Janjigian YY, et al. Tumor mutational load predicts survival after
 immunotherapy across multiple cancer types. Nat Genet.
 2019;51(2):202–6.
- 756 70. Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V,
 757 Havel JJ, et al. Cancer immunology. Mutational landscape deter758 mines sensitivity to PD-1 blockade in non-small cell lung cancer.
 759 Science. 2015;348(6230):124–8.
- 760 71. McGranahan N, Furness AJS, Rosenthal R, Ramskov S, Lyngaa R,
 761 Saini SK, et al. Clonal neoantigens elicit T cell immunoreactivity
 762 and sensitivity to immune checkpoint blockade. Science.
 763 2016;351(6280):1463–9.
- 764 72. Hellmann MD, Paz-Ares L, Bernabe Caro R, Zurawski B, Kim S765 W, Carcereny Costa E, et al. Nivolumab plus ipilimumab in ad766 vanced non-small-cell lung cancer. N Engl J Med. 2019;381(21):
 767 2020–31.
- 768 73. Algazi AP, Tsai KK, Shoushtari AN, Munhoz RR, Eroglu Z, Piulats
 769 JM, et al. Clinical outcomes in metastatic uveal melanoma treated
 770 with PD-1 and PD-L1 antibodies. Cancer. 2016;122(21):3344–53.
- 771 74. Johnson DB, Bao R, Ancell KK, Daniels AB, Wallace D, Sosman JA, et al. Response to anti-PD-1 in uveal melanoma without high-volume liver metastasis. J Natl Compr Cancer Netw JNCCN. 2019;17(2):114–7.

- 75. Mandal R, Samstein RM, Lee K-W, Havel JJ, Wang H, Krishna C, et al. Genetic diversity of tumors with mismatch repair deficiency influences anti-PD-1 immunotherapy response. Science. 2019;364(6439):485–91.
 778
- Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, 779 et al. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. Science. 2017;357(6349):409–13. 781
- 77. Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H, Eyring AD, et al. PD-1 blockade in tumors with mismatch-repair deficiency. N Engl J Med. 2015;372(26):2509–20.
 784
- Germano G, Lamba S, Rospo G, Barault L, Magrì A, Maione F, et al. Inactivation of DNA repair triggers neoantigen generation and impairs tumour growth. Nature. 2017;552(7683):116–20.
- Froglu Z, Zaretsky JM, Hu-Lieskovan S, Kim DW, Algazi A, Johnson DB, et al. High response rate to PD-1 blockade in desmoplastic melanomas. Nature. 2018;553(7688):347–50.
- Kaunitz GJ, Cottrell TR, Lilo M, Muthappan V, Esandrio J, Berry S, et al. Melanoma subtypes demonstrate distinct PD-L1 expression profiles. Lab Investig J Tech Methods Pathol. 2017;97(9):1063–71. 793
- Jayaraman SS, Rayhan DJ, Hazany S, Kolodney MS. Mutational landscape of basal cell carcinomas by whole-exome sequencing. J Invest Dermatol. 2014;134(1):213–20.
- Martincorena I, Roshan A, Gerstung M, Ellis P, Van Loo P, McLaren S, et al. Tumor evolution. High burden and pervasive positive selection of somatic mutations in normal human skin. Science. 2015;348(6237):880–6.
- 83. Falchook GS, Leidner R, Stankevich E, Piening B, Bifulco C, 801
 Lowy I, et al. Responses of metastatic basal cell and cutaneous squamous cell carcinomas to anti-PD1 monoclonal antibody 803
 REGN2810. J Immunother Cancer. 2016;4:70. 804
- 84. Migden MR, Rischin D, Schmults CD, Guminski A, Hauschild A, 805
 Lewis KD, et al. PD-1 blockade with cemiplimab in advanced cutaneous squamous-cell carcinoma. N Engl J Med. 2018;379(4): 807
 341–51. 808
- Nghiem PT, Bhatia S, Lipson EJ, Kudchadkar RR, Miller NJ, Annamalai L, et al. PD-1 blockade with pembrolizumab in advanced Merkel-cell carcinoma. N Engl J Med. 2016;374(26): 2542–52.
 812
- 86. Goh G, Walradt T, Markarov V, Blom A, Riaz N, Doumani R, et al.
 813 Mutational landscape of MCPyV-positive and MCPyV-negative Merkel cell carcinomas with implications for immunotherapy.
 815 Oncotarget. 2016;7(3):3403–15.
 816
- 87. Martens A, Wistuba-Hamprecht K, Geukes Foppen M, Yuan J, Postow MA, Wong P, et al. Baseline peripheral blood biomarkers associated with clinical outcome of advanced melanoma patients treated with ipilimumab. Clin Cancer Res Off J Am Assoc Cancer Res. 2016;22(12):2908–18.
 821
- Weide B, Martens A, Hassel JC, Berking C, Postow MA, Bisschop K, et al. Baseline biomarkers for outcome of melanoma patients treated with pembrolizumab. Clin Cancer Res Off J Am Assoc Cancer Res. 2016;22(22):5487–96.
- 89. Rosner S, Kwong E, Shoushtari AN, Friedman CF, Betof AS, 826
 Brady MS, et al. Peripheral blood clinical laboratory variables associated with outcomes following combination nivolumab and ipilimumab immunotherapy in melanoma. Cancer Med. 829 2018;7(3):690–7. 830
- 90. Caballero S, Pamer EG. Microbiota-mediated inflammation and antimicrobial defense in the intestine. Annu Rev Immunol.
 831

 2015;33(1):227–56.
 833
- 91. Schwabe RF, Jobin C. The microbiome and cancer. Nat Rev 834 Cancer. 2013;13(11):800–12. 835
- 92. Gopalakrishnan V, Helmink BA, Spencer CN, Reuben A, Wargo
 836

 JA. The influence of the gut microbiome on cancer, immunity, and
 837

 cancer immunotherapy. Cancer Cell. 2018;33(4):570–80.
 838
- Gopalakrishnan V, Spencer CN, Nezi L, Reuben A, Andrews MC, 839 Karpinets TV, et al. Gut microbiome modulates response to anti– 840

- PD-1 immunotherapy in melanoma patients. Science.
 2018;359(6371):97–103.
- 843 94. Matson V, Fessler J, Bao R, Chongsuwat T, Zha Y, Alegre M-L,
 844 et al. The commensal microbiome is associated with anti–PD-1
 845 efficacy in metastatic melanoma patients. Science.
 846 2018;359(6371):104–8.
- 847 95. Wargo JA, Gopalakrishnan V, Spencer C, Karpinets T, Reuben A,
 848 Andrews MC, et al. Association of the diversity and composition of
- the gut microbiome with responses and survival (PFS) in metastatic
- 858

melanoma (MM) patients (pts) on anti-PD-1 therapy. J Clin Oncol. 850 2017;35(15 suppl):3008. 851

96. Dubin K, Callahan MK, Ren B, Khanin R, Viale A, Ling L, et al. 852
Intestinal microbiome analyses identify melanoma patients at risk for checkpoint-blockade-induced colitis. Nat Commun. 2016;7(1): 854
1–8. 855

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