Title
The crystal structure of a cardiac calsequestrin filament reveals an atomic mechanism of calsequestrin-associated catecholaminergic polymorphic ventricular tachycardia

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The crystal structure of a cardiac calsequestrin filament reveals an atomic mechanism of calsequestrin-associated catecholaminergic polymorphic ventricular tachycardia

by

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To my parents, John Joseph Titus and Catherine Anne Wilcox-Titus, and to my partner, Tene Aneka Cage. Thank you for encouraging me and supporting me at every turn.

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The subject of my Ph.D. work is the structure and function of the calcium-binding protein, calsequestrin, and its relationship to cardiac physiology and disease. Much of my work on calsequestrin originated in the clinic with academic electrophysiologists who recognized a case of rare disease with basic science implications. I thank Jason Roberts and Melvin Scheinman for initiating projects that have led in exciting directions. I am grateful to Jason for our collaboration and for his advice on our shared clinical and scientific work.

My wife and young son have generously tolerated my many absences while I completed my Ph.D. work, and my parents have been a source of continuous encouragement as I ventured out in new professional and academic directions. I dedicate my work to them.

The crystal structure of a cardiac calsequestrin filament reveals an atomic mechanism of calsequestrin-associated catecholaminergic polymorphic ventricular tachycardia

Erron Wilcox Titus

Abstract

Calcium homeostasis is essential to cardiac and skeletal muscle physiology, where contractile function requires tight but dynamic control of calcium levels across different cellular compartments. The major calcium storage protein in muscle tissues is calsequestrin, a highly acidic protein responsible for buffering up to 50 % of total sarcoplasmic reticulum (SR) calcium. Mutations in cardiac calsequestrin cause a highly lethal familial arrhythmia, catecholaminergic polymorphic ventricular tachycardia (CPVT), while mutations in skeletal muscle calsequestrin have been linked to myopathies. Calsequestrin's high density calcium storage is facilitated by homomultimerization of the protein into filaments, but a compelling atomic-resolution structure of a calsequestrin filament is lacking. This gap in knowledge has limited our understanding of calsequestrin biochemistry, SR calcium storage, and molecular mechanisms of calseqestrin-associated diseases. We report here a crystal structure of a cardiac calsequestrin filament with supporting mutation analysis by an *in vitro* filamentation assay. We also report and characterize a novel disease-associated mutation, S173I, which localizes to the structure's filament-forming interface. In addition, we show that a previously reported dominant calsequestrin-associated CPVT mutation, K180R, maps to the same multimerization surface. Both mutations disrupt filamentation in vitro, suggesting a model where dominant disease arises from mutations that disrupt multimer formation. Finally, a ytterbium-derivatized structure pinpoints multiple credible calcium sites at filament-forming interfaces, explaining the atomic basis of calsequestrin filamentation in the presence of calcium. This work advances our basic understanding of calsequestrin biochemistry and also provides a unifying structure-function molecular mechanism by which dominant-acting calsequestrin mutations provoke lethal arrhythmias.

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Introduction

The Ca^{2+} ion is a ubiquitous chemical messenger in eukaryotic cells. Passive flows of calcium from regions of high concentration to low produce transient changes in intracellular calcium, which are then coupled to a diverse set of signal transduction pathways. In facilitation of its diverse roles as an intracellular messenger, calcium exhibits by far the largest concentration difference across compartments, from approximately 2 mM extracellularly to 100 nM intracellularly - a range of up to 5 orders of magnitude.[1] This calcium gradient exists across the plasma membrane separating the cytosol from the extracellular space but also between the cytosol and the endoplasmic reticulum (ER) lumen, with the latter maintaining an ionic milieu resembling extracellular conditions. Direct physical coupling between the ER lumen and the extracellular space allows some degree of passive equilibration between these two compartments[2], so that the biochemical work of establishing and maintaining the potential is not done twice.

In muscle, the transduction of an electrical signal at the membrane into a calcium flow that activates the contractile apparatus is known as excitation-contraction coupling. Under conditions of high muscle loading, the calcium flows required by the contractile apparatus become much more substantial than other biological calcium fluxes. In order to minimize the energy expended per contractile cycle, muscle cells possess enhanced calcium buffering at the sites of storage and release, allowing a high total calcium content with much lower free calcium. Calsequestrin (CSQ) is a 44 kD highly acidic protein that serves as the principal calcium buffer of the sarcoplasmic reticulum (SR, a muscle-specific extension of the ER) in cardiac and skeletal muscle, storing up to 50 % of SR calcium in a bound state, with each calsequestrin monomer storing up to 40-50 calcium ions.[3][4][5][6][7][8][9] Calsequestrin is complexed to the SR calcium channel, the ryanodine receptor (RyR), thereby ensuring that calcium is stored at the site of its release.[10]

Since their initial identification, in 1971, a substantial field of research has formed around calsequestrins. Calsequestrins (CASQ1 in skeletal muscle and CASQ2 in cardiac muscle) are highly homologous in structure and function, and 64 % identical in sequence, with skeletal

muscle calsequestrin appearing to have higher calcium capacity.[11] Calcium-binding propensity is explained in large part by calsequestrin's remarkable fraction of negatively-charged acidic residues (26 % glutamate or aspartate in CASQ2 and 28 % in CASQ1, with corresponding average isoelectric points of 4.2 and 4.0, respectively). In both cardiac and skeletal muscle, calsequestrin localizes to the junctional SR (jSR) of muscle and forms multimers that are anchored to the luminal SR membrane. From low resolution electron micrographs of isolated skeletal muscle fibers, these multimers appear as a dense collection of filaments.[7][12] Anchoring of calsequestrin filaments occurs within a complex consisting of RyR and the single-pass transmembrane proteins triadin and junctin.[10] Extended homo-multimerization provides high density calcium storage, but multimerization appears to be essential for localization, too, in that multimerization-defective mutants are trafficked along the secretory pathway and lost from the ER/SR.[13][14][15] Additional insights into calsequestrin biochemistry and cell biology encompass post-translational modifications[16][17], calcium-storage capacity[11], and interactions with the calcium release unit.[18][19][20]. Finally, as our understanding of calsequestrin biology has advanced, so has our understanding of its role in human disease. Mutations in skeletal muscle calsequestrin have been putatively linked to malignant hyperthermia and to vacuolar myopathies[21], while mutations in cardiac calsequestrin are well known to cause catecholaminergic polymorphic ventricular tachycardia (CPVT), a highly lethal familial arrhythmia. CPVT is caused by a state of cardiac calsequestrin deficiency, whether arising from null/hypomorphic alleles or from point mutants that disrupt calsequestrin multimerization.[22][23] Consistent with a mechanism where by deficiency leads to disease, most calsequestrin-associated CPVT mutations have recessive inheritance. For a protein whose function depends on multimerization, there is, at least to date, a surprising paucity of mutations with putative dominant negative mechanism.

Despite the rich body of work concerning the calsequestrin biology, no compelling high-resolution candidate structure for a calsequestrin filament has emerged. Although sixteen crystal structures of calsequestrin have been published[8][11][24][16][25][21][26], none of these contain a filament-like assembly. All 16 prior structures reveal calsequestrin dimers that are nearly-identical to one another, but a search for dimer-to-dimer interfaces within and across

these crystal unit cells reveals only weak crystallographic packing contacts that appear incompatible with robust biological multimerization. In addition, the observed dimer-to-dimer interfaces vary substantially from one lattice to another. Lack of mutagenesis studies supporting proposed oligomerization interfaces in the prior published structures further calls into question whether the relevant biological multimer has ever been observed. These prior studies have established that calsequestrins dimerize in a wide variety of conditions, with *intra*-dimer interactions that are largely the same across published structures, but the mechanism by which dimers assemble into higher order multimers (*inter*-dimer assembly) remains elusive.

A structure of a calsequestrin filament would advance our understanding of calsequestrin biology and permit a comprehensive mapping of disease-causing mutations to biologically-relevant multimerization interfaces. We report here an investigation of dominant-acting cardiac calsequestrin disease mutants that culminated in a new X-ray structure of cardiac calsequestrin, one that we believe reveals the biologically relevant filament-forming interface. We show that known dominant-acting mutants - one previously reported and one described for the first time in this study - map to the newly reported multimerization interface. Furthermore, we provide supporting biochemical analysis of likely calcium-binding sites where the protein multimerizes. These findings fundamentally advance our understanding of the mechanism by which calsequestrin contributes to calcium homeostasis and provide insight into mechanisms of calsequestrin-associated familial diseases.

Results

Autosomal Dominant CASQ2 Disease Mutations Disrupt Calsequestrin Multimerization

We encountered a proband from a family with CPVT-like tachy-arrhythmias and multiple cases of sudden, unexplained death at a young age (Figure 1A). The proband presented at age 33 with a biventricular tachycardia on ECG. She underwent EP study that showed an inducible ventricular tachycardia with focal origination next to the anterior fascicle, and an inducible

atypical AVNRT that was successfully ablated. Targeted sequencing of channelopathy genes (KCNQ1, KCNH2, SCN5A, ANK2, KCNE1, KCNE2, KNCJ2, CAV3, RYR2, and CASQ2) revealed only a heterozygously-carried isoleucine-for-serine substitution at position 173 in cardiac calsequestrin. The proband's son and multiple other family members report a tachycardic phenotype (ranging from self-reported palpitations to diagnosed episodes of tachycardia). The family history is also notable for multiple cases of sudden cardiac death at a young age. Overall, the distribution of affected individuals in the pedigree is potentially consistent with dominant inheritance. As the family did not consent to follow-up genetic testing or clinical phenotyping, we were unable to rigorously assess disease/mutation co-segregation.

The genetic evidence for pathogenicity of the S173I novel variant is not conclusive, but since known pathogenic point mutations in CASQ2 exhibit defective filamentation in a simple turbidity assay, we elected to investigate the S173I mutation biochemically. The turbidity assay for S173I reveals a profound decrease in multimerization rate (Figure 1B). Minimal improvement in the multimerization rate is obsered in a non-physiologic 0 mM potassium condition (Figure S1), demonstrating that the protein is intact but defective in multimerization.

The sole known CASQ2 disease mutation with strong evidence for dominant inheritance, K180R, was not reported to cause defects in multimerization and was proposed to act via alternative mechanisms.[27]. The striking effect of S173I on CASQ2 mutimerization, combined with the fact that other known CASQ2 point mutants exhibit the same biochemical defect, prompted us to reexamine multimerization capacity of the K180R mutant. The turbidity assay for K180R under the same conditions used for S173I shows little difference from the wild type CASQ2 variant (Figure 1C). However, prior reports suggested that calsequestrin maintains distinct magnesium and calcium binding sites.[28] Therefore, we investigated filamentation kinetics of the K180R mutant in the presence of magnesium. Extended incubation of the K180R mutant with magnesium (2 mM MgCl2) prior to addition of calcium yields a profound filamentation defect (Figure 1D). In vivo, calsequestrin would likely encounter similar levels of free magnesium throughout the SR.

As the dominant inheritance pattern is classically associated with disease mutations that disrupt protein-protein interactions, we might expect to find dominant-acting disease mutations

at cardiac calsequestrin's dimer or multimer interfaces. However, neither S173 nor K180 fall at credible, previously identified candidate multimerization interfaces: they are not near the intra-dimer interface, nor are they near candidate inter-dimer interfaces in the prior crystal structures. We therefore elected to pursue another structure in the belief that the biologically relevant multimer has not yet been observed.

Overview of The New Candidate Structure of a Calsequestrin Filament

We have determined a new crystal structure of human cardiac calsequestrin obtained from a full-length construct in a very low-pH (3-3.5) crystallization condition. The previously characterized calsequestrin dimer is again observed, but now in an arrangement that produces a closely-packed filament (Figure 2). The new structure provides a compelling candidate for a biologically-plausible higher-order multimer. Crystallographic data collection and processing statistics are summarized in (Table S1) for the native structure as well as a ytterbium-soaked condition used to identify probable calcium-binding sites. The repeating unit of the native crystal resides in a higher-symmetry point group compared to prior calsequestrin structures (Table S2). The oligomer-forming contacts that exist between dimers are novel, differing significantly from all previously reported calsequestrin crystal structures (Table S4). The inter-dimer interface in our structure encompasses significantly greater buried surface area than previously observed (Table S4).

The cardiac calsequestrin monomer, like its skeletal calsequestrin equivalent, consists of an N terminal loop, 3 thioredoxin domains, and a disordered acidic tail. Within the dimer, there is two-fold symmetry with mutual exchange of N-terminal loops, as previously observed (Figure 3A). In our structure, the dimers are stacked along a screw axis to form the filament, with each dimer rotated 90° with respect to its neighbors. Although the dimers are positioned at discrete 90° rotations with respect to one another, the underlying architecture of the filament is in fact helical at the level of thioredoxin domains (Figure 3A). Thioredoxin domains II and III form a double helix at the core of the filament (Figure 3B). An outer helix or "collar" of larger diameter, consisting only of thioredoxin domain I, then winds around the inner double

helix. All helices are left-handed, corresponding to the left-handed screw axis at the level of dimer-stacking.

The 3-helix configuration appears to promote the close packing of globular thioredoxin domains. Helical packing permits each domain to contact multiple other domains in multiple other protomers, which is in stark contrast to other reported candidate structures (PDB ID 1A8Y, rabbit skeletal muscle calsequestrin; PDB ID 1SJI, canine cardiac calsequestrin), which lack the helical pitch and have much more limited interacting surface area (Figure 4). The close-packing of the new filament candidate is starkly visible when thioredoxin domains are represented as equally-sized spheres centered at the domain center of mass (Figure 4, right-hand side).

Notably, our crystallization condition produced crystals only at a low pH (3-3.5, measured by applying a crystallization drop to litmus paper) resulting from the use of aged PEG reagents (presumably having undergone degradation to glycolic acid) or from direct addition of concentrated HCl. This restriction obtained irrespective of calcium in concentrations ranging from trace to 14 mM (higher calcium concentrations led to precipitation, likely due to the presence of sulfate, and were therefore not assessed). Given calsequestrin's remarkable overall acidity and low isoelectric point, an explanation for the role of low pH in promoting calsequestrin filamentation could be the neutralization of substantial negative charge on acidic side chains that extend across the filament interfaces.

Lanthanide Substitution Identifies Cation-Binding Sites that Participate in Dimer Formation

Prior work has identified putative calcium binding sites at the dimer interface of calsequestrin[25]. Our new filament structure permits us to examine the role of multivalent ions at all surfaces relevant for filamentation - both the intra-dimer surfaces as well as the inter-dimer surfaces responsible for higher-order multimers. To localize candidate calcium ligand sites within the context of the new filament structure, we collected data from a Ytterbium (Yb)-soaked crystal.(Table S1) From the anomalous map, we identified approximately 8 sites with strong Yb

signal per CASQ2 chain (¿ 3.8 σ in the anomalous map and difference density in the Fo-Fc map). As the prior work to identify calsequestrin's calcium-binding sites was based largely on the indirect method of inference from metal coordination geometry with nearby side chains and waters, the use of Yb provides a direct approach to confirming the prior findings, as well as extending our understanding to the entire filament.

We first assessed the presence of Yb at the intra-dimer interface. We identified several high-occupancy Yb sites, most of which are clustered in a narrow region between protomers, where they are coordinated by multiple, highly-conserved acidic residues (Figure 5 and Figure S2). Consistent with the location of a putative calcium ion in the prior study, we find a Yb atom coordinated primarily by E147 of chain A and D278 of chain B. In addition, we find a Yb atom coordinated primarily by E143 and E275, as well as another Yb atom coordinated by D310 and a putative sulfate ion within a solvent cavity enclosed by the dimer. These Yb-bridging sites stabilize the interaction between thioredoxin domain II from protomer A and thioredoxin domain III of protomer B. The interactions between domains result in close juxtaposition of acidic side chains (Figure 5), requiring neutralization of the charge by the multivalent counterions or, in the case of the native structure, protonation. In the native structure, the E147/D278 interaction provides an example of a carboxyl-carboxylate bond stabilized by low pH.[29][28] Anomalous signal is found at this site in the derivative structure, but consistently on just one side of the otherwise symmetric dimer. This subtle asymmetry, with a cation bound on one side and a carboxyl-carboxylate bond on the other, likely reflects a degree difference in occupancy rather than a sharp distinction, but it is a consistent feature of the density map across all 4 dimers in the crystal asymmetric unit and may be conformationally conducive to filament formation.

In both the native structure and Yb-soaked structure, the dimer subunits have undergone substantial inward rotation in comparison to the other high-resolution cardiac calsequestrin structure (PDB ID 1SJI) (Figure 5B). The inward domain movement is produced largely by rotation of the monomer as a rigid body and results in a conformation where domains II and III are more closely packed. This conformational shift recapitulates a similar finding from the prior study where a 15° rotation within the skeletal calsequestrin dimer was observed for structures

crystallized in a high-calcium buffer.[25] Upon inspection of all prior published calsequestrin structures, it is apparent that 6 prior calsequestrin structures belong to this ”tightly-packed dimer” group, while the remaining 10 contain dimers that are more loosely-packed, without inward rotation of chains (Figure S3). The more tightly-packed structures were all crystallized in the presence of multivalent cations (usually calcium), or in one case (PDB ID 2VAF) a monovalent cation at extremely high concentration (2 M NaCl. The other group were crystallized with no added multivalent cations, with the exception of PDB ID 3TRP, which contained calcium in the crystallization drop at approximately 5-fold lower than lowest concentration from the tightly-packed group. Within the tightly-packed group, there is a greater degree of conformational disorder in Domain I (Figure S4). This is accompanied by a modest loss of contact in Domain I, while multiple hydrophobic side chains from Domains II and III that were not buried now obtain buried surface area (Figure S5). Thus, our data provide additional evidence for conformational change within the dimer upon calcium-binding (likely induced by closer approximation of acidic side chains), independent confirmation of intra-dimer calcium binding sites, and an explanation for why an altered conformation of the dimer becomes energetically tolerable in the presence of calcium.

Cation-Binding Promotes Filament Formation by Dimer-Dimer Stacking

Like the dimer interface, the inter-dimer filament-forming interface is bridged by Yb sites surrounded by closely-apposed acidic side chains (Figure 6 and Figure S6). In contrast to the dimer interface, where carboxyl-carboxylate bonds are still seen in the Yb-derivative structure, at the inter-dimer interface the sites of carboyxl-carboylate interaction have been throughly substituted with a cation. Identification of these coordination sites provides a testable model for the biological correctness of the new filament structure. To test this model, we mutated the aspartate or glutamate side chains involved in Yb binding to alanine and examined the effect of the mutations on filamentation kinetics. The most pronounced effect resulted from mutations targeting the symmetrical interaction involving E184 and E187 (Figure 6A). The Yb site formed by these residues lies on the edge of a solvent cavity enclosed by the inter-dimer

interface, partially shielded from exposure to the bulk solvent. Glutamates 184 and 187 belong to an alpha helix that provides a linkage between domains of the outer thioredoxin collar. This helix, belonging to thioredoxin domain 2, sits between thioredoxin I domains of different dimers and interacts with inter-dimer salt bridges on either side. Mutating E184 and E187 to alanine effectively prevents filamentation within the time period of observation.(Figure 6B). Alanine mutagenesis of the D50 residue that participates in a salt bridge with K180 at the N-terminal end of this helix produces a similar defect (Figure 6C).

Two more sites of strong Yb signal are notable. Residues D348 and D350 are oriented along with their symmetry mates to form a cluster of 4 acidic side chains that coordinate a single cation. As rendered in (Figure 6A), this Yb site forms the base of the solvent cavity enclosed by the inter-dimer interface. Outside this cavity, on the fully solvent-exposed exterior of the filament, residues D351 and E357 and their symmetry mates form bidentate interactions with two Yb atoms, adopting conformations in which opposing acidic rotamers are bent away from one another, thereby alleviating electrostatic repulsion that would otherwise disrupt multimer formation (Figure 6A). In summary, the bound cation at these sites appears to facilitate filamentation by neutralizing the abundant negative surface charge of calsequestrin. Mutating these sites to alanine would be expected to relieve mutal repulsion of closely-packed acidic side chains. Consistent with this, alanine mutagenesis of these residues has a largely net neutral effect on filamentation kinetics (Figure S7).

Intriguingly, the vast majority of Yb sites that we identify are located within a continuous solvent cavity that winds through the interior of the filament, forming a hollow core with solvent exposure at the dimer and inter-dimer interfaces. Each dimer contains a solvent pocket within its interior, with a second solvent cavity located at inter-dimer interface (Figure 7A-B). The stacking of dimers forms a continuous solvent cavity along the entire length of the filament (Figure 7). The calcium ions that would appear to be bound in the dimer's interior would constitute a separate store of calcium that is more slowly-mobilized than the highly-accessible pool of ions bound to the surface and the solvated acidic tail.

Dominant Disease Mutations Disrupt Cardiac Calsequestrin's Inter-Dimer Interface

The newly observed filament structure permits us to assess the possible pathogenic mechanism of the S173I mutation that began this investigation. Remarkably, S173 occupies a critical position at the filament-forming interface - a charged pocket formed by the interaction of K87, S173, and D325 (Figure 8A and Figure S8. This pocket, formed of residues from 3 different protomers and also 3 different thioredoxin domains, enforces an interaction between the 3 thioredoxin domains at a single site. This interaction explains the disruptive effect of the S173I mutation discussed above (turbidity assay from Figure 1). On the basis of the apparent importance of this site, as well as the absence of D325 in any other previously described candidate filament interface, we also mutated residue D325 to D325I. As expected, D325I exhibits similar profound loss of function in the turbidity assay (Figure 8B).

The afore-mentioned K180 residue has recently been implicated in CPVT[27]. Specifically, a lysine-to-arginine mutation at this position provided the first strong genetic evidence for a dominant CPVT-causing mutation in cardiac calsequestrin. Although this substitution is relatively conservative, the genetic evidence for the pathogenicity of K180R as a dominant mutation is compelling on the basis of a comprehensive segregation analysis in a large family. As shown in Figure 1, the K180R mutation also results in a filamentation defect. Our new structure provides a mechanistic explanation for this effect. In the new structure, K180 participates in helix-stapling salt bridges at the E184/E187 putative calcium site/interface region (Figure 6A) shown by us to be essential for robust filamentation.(Figure 6B) The high conservation of K180 in evolution (Figure S9) and key role of K180 and its interacting partners in supporting calsequestrin filamentation provide additional support for the physiological relevance of our new structure.

Discussion

We report here a crystal structure of a calsequestrin filament. We also elucidate the biochemical basis of divalent cation-induced filamentation. Using Yb substitution, we confirm previous findings of intra-dimer cation binding sites, and we reveal additional inter-dimer cation sites that have a substantial effect on the rate of multimerization. Furthermore, we provide three forms of evidence in support of the biological relevance of the new structure: biophysical (buried surface area, all-by-all interactions of protomers and domains), biochemical (disruption of multimerization by targeted mutagenesis), and biomedical (disease mutations with dominant inheritance are located at the newly-identified filament-forming interfaces).

What is the proximal source of free energy for calsequestrin filamentation? The prevailing view has been that calsequestrin filamentation is driven by an increase in solvent entropy: ions are bound at filament interfaces, and solvent entropy increases as ion hydration shells are lost.[28] Under this view, the net contribution from solvent entropy is much greater than the net enthalpic contribution from calcium binding after considering the energetic cost of dehydration. This hypothesis is complicated, however, by the fact that we observe stable filamentation in the near-absence of ligand (only trace concentrations of divalent cations were present in the low-pH crystallization condition). The low-pH filament is stabilized instead by carboxyl-carboxylate interactions between closely juxtaposed acidic residues. While increased solvent entropy likely contributes to filamentation, we postulate instead that protein conformational entropy is likely to be the major driving force. The structure we report is remarkably disordered by conventional B-factor metrics, and especially so in Domain I. In fact, all tightly-packed calsequestrin dimers (7 crystallographic results total, including this study) exhibit increased disorder in the solvent-exposed loops of Domain I (Figure S4). The consistency of this observation within the group makes it less likely that conformational disorder within Domain I is a feature of our specific crystallization condition and more likely that it is a result of the dimer having adopted the tightly-packed conformation favorable for filamentation.

Although low pH may not be necessary for filamentation, calsequestrin's pH-sensitivity prompts questions about the possible role of intra-luminal pH changes in the regulation of cal-

cium uptake and release. Carboxyl-carboxylate side chain interactions modulated by prevailing pH are a common feature of calcium-binding proteins, wherein a shared proton provides a stabilizing interaction when the cation is unavailable.[30][28] These carboxyl-carboxylate interactions have a higher pKa than a solitary carboxylate, permitting them to play a significant role at pH ranges closer to physiologic.[31][28] Even at neutral pH, studies of calsequestrin and calmodulin have revealed that protons are released upon calcium binding, consistent with loss of carboxyl-carboxylate bonds.[30][28] Notably, the SERCA pump, in addition to being an ATPase, is a Ca^{2+}/H^{+} antiporter, so it is reasonable to hypothesize that proton flows over the course of a cycle of CICR have downstream regulatory effects on calsequestrin's filamentation state, or its affinity for calcium, or both. A small decrease in SR luminal pH during calcium release has been observed[32], and multiple groups have proposed that proton influx into the SR constitutes a small but important fraction of the counterion flow required to maintain charge neutrality when large calcium fluxes occur. Proton efflux from the SR during calcium reuptake, by way of the SERCA pump's antiporter function, would increase the availability of calcium binding sites. Conversely, protein influx into the SR during calcium release - possibly via a second, independent proton transport pathway within the SERCA pump[33] - could conceivably stabilize calsequestrin filaments until high levels of calcium are restored. Prior work showed a change in calsequestrin's intrinsic fluorescence at a pH of 6.0 [34], suggesting that dynamic effects on calsequestrin are not limited to the low pH regime used in our crystallization experiments. Since calsequestrin is present at high concentration and likely acts non-trivially as a proton buffer in its own right, effects due to total free proton concentration may manifest as protons move from an SR-based buffer system to one in the cytosol and back with little change in detectable pH. It is important to note that presumed regulatory of effects of dynamic SR pH are speculative and require further elucidation.

Strikingly, most known CASQ2 mutations that disrupt filamentation at the intra-dimer interface (e.g. R33Q, D307H, and P308L) are associated with recessive inheritance, yet the mutations investigated here (S173I, K180R) that disrupt filamentation at the inter-dimer interface are associated with dominant inheritance. The apparent discrepancy in inheritance patterns, mapping cleanly to distinct interfaces albeit with limited sampling, is notable. To date, the field

has explained the recessive mechanism of intra-dimer interface mutations by way observing that calsequestrins have no classical ER retention signal: filamentation is necessary for calsequestrin to remain in the jSR.[13][14][15] In the setting of a heterozygously-carried mutation that inhibits dimer formation, mutant monomers are eliminated by trafficking, such that the pool of wild type protein would be largely unaffected. Under this model, filamentation defects overlap mechanistically with a class of calsequestrin-deficient conditions arising from null or hypomorphic alleles, all leading to a final common endpoint of decreased calcium-buffering capacity, and a resulting susceptibility to diastolic calcium leak.

In seeming contrast to this model, poisoning of multimerization in classical dominant fashion would require mutant-incorporating dimers to remain jSR-resident, where they could interfere with assembly. There is no reason, however, based on current understanding of secretory pathways, that the dimer would be any less susceptible than the monomer to export. How, then, can the ER export mechanism be reconciled with dominant-acting filamentation-defect mutants? If filamentation defects are equivalent at the molecular level to calsequestrin deficiency, how do we explain the finding of filamentation-disrupting mutations with likely dominant inheritance? We would propose that all filamentation-defective mutants within unincoporated dimers are trafficked out of the jSR in much the same way that unincoporated mutant monomers are. The resulting disease mechanism for dominant mutations is therefore more complex than a classical dominant negative effect: for as long as mutant-containing dimers may be present in the jSR, they interfere with multimerization, but crucially, when they leave, they steal half of the total WT protein with them. The result is paradoxical and perhaps somewhat novel: insufficiency by way of underlying dominant negative biochemistry.

In sum, taking into account calsequestrin's demonstrated susceptibility to ER export, we can combine the present work with the existing rich body of calsequestrin research to neatly explain the inheritance puzzle associated with CPVT-causing calsequestrin mutations. Dimer-defective mutants produce monomers which are trafficked away, but the heterozygous state is rescued because WT protein is unaffected by the pool of defective monomeric protein. In contrast, mutations that interfere with the inter-dimer interaction result in depletion of a substantial fraction of WT protein, as unincorporated dimers containing a mix of WT and mutant

protein are continually lost. Future work should seek to confirm this hypothetical mechanism using cell biological investigative methods.

Materials and Methods

Human Subjects

The patient included in the study provided informed consent as part of a research protocol approved by the University of California, San Francisco Committee on Human Research.

Cloning and Generation of Plasmids

Full-length cardiac calsequestrin was cloned from human cardiac mRNA by reverse transcription, PCR, A-tailing of a PCR product, and TA ligation. A clone lacking the signal peptide sequence was subcloned by PCR and Gibson Assembly into a T7-based bacterial overexpression vector (pET28a) in front of a 6His site and TEV protease cleavage sequence. Site-directed mutagenesis was performed to generate alanine mutants and CPVT mutants. All constructs were transformed in NEB Stable or XL-1 Blue *E. coli*, purified by miniprep, and retransformed into Rosetta (DE3)pLysS *E. coli* for overexpression. Selection for the pET28a vector was performed using 50 µg/L kanamycin. Selection for pLysS was performed by adding 25 µg/L chlorampenicol.

Molecular Cloning

The human cardiac calsequestrin coding sequence was obtained by RT-PCR. Subsequent cloning was performed by Gibson Assembly. The construct for protein expression was cloned into a pET28a-based expression vector. This construct excluded the N-terminal signal peptide but retained the remaining coding sequence, including the C-terminal acidic tail. Point mutants were generated using the protocol from the Q5 Site-Directed Mutagenesis Kit (New England BioLabs), using either the Q5 or Phusion polymerases. All constructs were verified by Sanger sequencing. Primers used for cloning and mutagenesis are provided in Table S1.

Expresssion and Purification of Cardiac Calsequestrin

pET28a-based expression constructs were miniprepped from a cloning strain and transformed into Rosetta (DE3)pLysS *E. coli*. Overnight starter cultures were used to inoculate large cultures (typically 750 mL of broth per 2.8 L flask), which were grown to OD 0.4 and then induced with 0.25 mM IPTG. Upon induction, temperature was reduced from 37 °C to 24 °C. Cultures were grown for 6-9 hours post-induction or overnight and then spun down (optimal yields were obsered from shorter durations of growth). All cultures were grown in standard LB in 50 µg/L kanamycin and 25 µg/L chlorampenicol. Pellets were resuspended in lysis buffer (20 mM Tris pH 7.4, 500 mM NaCl, 10 mM imidazole, 1 EDTA-free protease inhibitor tablet per 50 mL) and frozen at –80 °C. Frozen suspensions were thawed, sonicated on ice (5 min at 1 s on/1 s off), and clarified (15,000 *g*, 45 min, 4 °C). The clarified supernatant was filtered (0.2 µm), and calsequestrin-containing fractions were isolated by IMAC using a 5 mL HisTrap FF column attached to a GE Akta FPLC (IMAC Buffer A: 20 mM Tris pH 7.4, 500 mM NaCl, 10 mM imidazole; IMAC Buffer B: 20 mM Tris pH 7.4, 500 mM NaCl, 300 mM imidazole). Protein was eluted in 10% steps of Buffer B. The first eluted fraction (10% Buffer B) was always discarded (consistently observed to be impure as determined by SDS-PAGE). Remaining protein-containing fractions were pooled. TEV protease was added to the pooled fractions at a concentration of 1:40 by mass, and the protein was dialyzed overnight at 4 °C in TEV protease dialysis buffer (50 mM Tris pH 8.0, 0.5 mM EDTA, 1 mM DTT). The cleaved protein was further dialyzed for several hours in EDTA dialysis buffer (20 mM HEPES pH 7.3, 100 mM NaCl, 5 mM EDTA) and then overnight into Anion Exchange Buffer A (20 mM HEPES pH 7.3, 100 mM NaCl). Anion exchange polishing was performed using a HisTrap FF column in series with 3x1 mL Mono Q columns. (Buffer A: 20 mM HEPES pH 7.3, 100 mM NaCl; Buffer B: 20 mM HEPES pH 7.3, 1 M NaCl). Protein was eluted in a continuous gradient up to 100% Buffer B), with calsequestrin-rich fractions consistently eluting at 40-50% Buffer B. Fractions were collected and analyzed for purity by SDS-PAGE and A260/A280 ratio. Fractions that were optimally pure and free of A260 contamination were pooled, concentrated to 20 mg/mL, and frozen at –80 °C.

Alanine mutants and the D325I mutant were purified as described above, except that phos-

phate IMAC buffers were employed (Buffer A: phosphate buffer at pH 7.4, 500 mM NaCl, 10 mM imidazole; Buffer B: phosphate buffer at pH 7.4, 500 mM NaCl, 300 mM imidazole), and an on-column high-salt wash was performed (phosphate buffer at pH 7.4, 2 M NaCl). In addition, the TEV protease dialysis buffer used for these purifications contained 100 mM NaCl.

Crystallization of Cardiac Calsequestrin (Native Conditions)

Crystallization screens were carried out in 96-well hanging-drop format and monitored using a Formulatrix Rock Imager automated imaging system. Conditions conducive to crystal growth were optimized and then reproduced in a 24-well format. The best diffraction was obtained by mixing thawed protein (10–20 mg mL^{-1} in 20 mM HEPES pH 7.3, 400–500 mM NaCl) 1:1 with 15% PEG 4000 and 400 mM Li_2SO_4. The pH of the PEG 4000 solution used to produce the best-diffracting crystals was tested by litmus paper and found to be approximately 3-3.5. Despite the presence of 20 mM HEPES in the protein reagent, the pH of the drops in which cystals grew was controlled by the PEG and remained 3-3.5. Freshly-made PEGs were incompatible with calsequestrin crystal growth except when concentrated HCl was added to the mother liquor, producing crystals similar to those observed with benchtop-aged PEGs. Interestingly, only unbuffered conditions yielded crystals. Multiple attempts to grow crystals at a buffered low pH (using acetate or glycine-based buffers) failed.

Ytterbium Soak

We initially attempted to identify calcium sites using anomalous signal from calcium ($CaCl_2$) added to the crystallization condition described above. We were unsuccessful, likely due to a combination of several factors. The calcium absorption edge is unreachable with conventional tunable x-ray sources and in normal atmosphere; thus it can only be approached, with resulting weakened anomalous signal. In addition, calsequestrin has an average K_d for calcium of 1 mM. Thus occupancy at a typical site would be expected to be lower as compared to other calcium-binding proteins. Presence of sulfate in the crystallization condition limited calcium concentrations to approximately 14 mM and below, above which a precipitate was observed.

Although this limit is above the K_d, it was insufficient for robust anomalous signal. Crystals of calsequestrin that formed in trace calcium were therefore soaked in $YbCl_3$. Hanging drops containing calsequestrin crystals were uncovered and an Eppendorf Microloader was used to inject 2 µL drops (1 µL protein and 1 µL mother liquor) with 200 µL of 2 M $YbCl_3$. Data were collected within 5 minutes with no back-soaking.

Crystal Data Collection and Structure Determination

Hanging drops were uncovered and submerged in a drop of Parabar 10312 (previously known as Paratone). For Yb-soaks, Yb was quickly injected prior to application of the oil. Crystals were looped and pulled through the oil. Excess oil was blotted away, and the loop was mounted directly into the cryostream of the Tom-Alber-Tron endstation at ALS beamline 8.3.1. Frames were collected at 1.116 keV (1.386 keV for Yb-soaked crystals) using the endstation's Pilatus3 S 6M detector using a strategy designed to balance redunancy against radiation damage.

Structure Determination

Diffraction images were processed with xia2 using the DIALS integration pipeline using a resolution cutoff of $CC_{1/2}$ ¿ 0.3.[35][36][37][38][39] For the native structure, the merged diffraction intensities were used to find a molecular replacement solution in Phaser[40] with the previously published canine cardiac calsequestrin structure, 1SJI[11], serving as the initial model. This resulted in solution in space group $P4_322$ containing one calsequestrin chain per AU. This solution was refined in PHENIX[41] with PHENIX AutoBuild[41][42][43][44][45], with extensive manual model-building in Coot[46]. For the Yb-complexed dataset, data were processed as above but with preservation of anomalous signal (no merging of Friedel pairs). The refined native structure was used a molecular replacement model, and a solution was found in space group $P4_32_12$ containing a dimer in the AU. This solution refined poorly. The Yb-complexed dataset was reprocessed in P1, and a molecular replacement solution was found in P1 with 16 chains in the AU. Refinement of this model was tested using Zanuda[47], and the best R-free

was found to be in space group $P12_11$. The Yb-complexed dataset was reprocessed in space group $P12_11$, and a molecular replacement solution was found with 8 chains in the AU. This solution refined well. The anomalous map was used in refinement to place Yb atoms in the structure.

Turbidity Assays

Recombinant protein samples were thawed, diluted in 2 mL–3 mL of Turbidity Assay Buffer (15 mM Tris pH 7.4, 20 mM NaCl, 85 mM KCl) and dialyzed in Turbidity Assay Buffer plus 10 mM EDTA. Samples were then redialyzed overnight in the same buffer without EDTA. Approximately half of the sample was then dialyzed in Zero-Potassium Turbidity Assay Buffer (15 mM Tris pH 7.4, 20 mM NaCl). Protein A280 was measured in triplicate (Nanodrop) using the appropriate matching buffer as background, and protein was diluted to 2.25 µM in a 140 µL volume in half-area wells of a µClear 96-well plate. The plate was covered, and protein in the wells was allowed to equilibrate on the benchtop for 20 minutes. The turbidity assay was performed using a BioTek Synergy 2 plate reader equipped with reagent injectors. Seven µL of 20 mM $CaCl_2$ solution was injected into each well, and the plate underwent shaking for 20 s. Absorbance at 350 nm was monitored for 45 min. The protocol was performed in plate synchronized mode for consistent well-to-well timing. A 100 mM ion-selective electrode calcium standard (Sigma, cat no.) stock solution was used for all $CaCl_2$ dilutions.

Quantification and Statistical Methods

Data points in figures represent mean values, with error bars representing standard deviation. All turbidity assay data points are mean of 3 technical replicates.

Data and Software Availability

The structures determined as part of this work are deposited in the Protein Data Bank (PDB) under identifiers 6OWV (native) and 6OWW (ytterbium-soaked). The raw diffraction dataset for the native structure is deposited in Zenodo under doi:10.5281/zenodo.2941360. The raw

diffraction dataset for the ytterbium-soaked structure is likewise deposited in Zenodo under doi:10.5281/zenodo.2943248. Protein structure figures were generated using PyMOL.[48] The interior cavity of the calsequestrin filament was traced using HOLLOW.[49] Sequence alignments were generated using TEXshade.[50] Plots were generated using python matplotlib.[51] The manuscript and figure layouts were constructed entirely in LATEX using PGF/Tik*Z*. Data and code to generate all figures are freely available at `https://github.com/errontitus/casq2-structure-function`.

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Figures

Figure 1

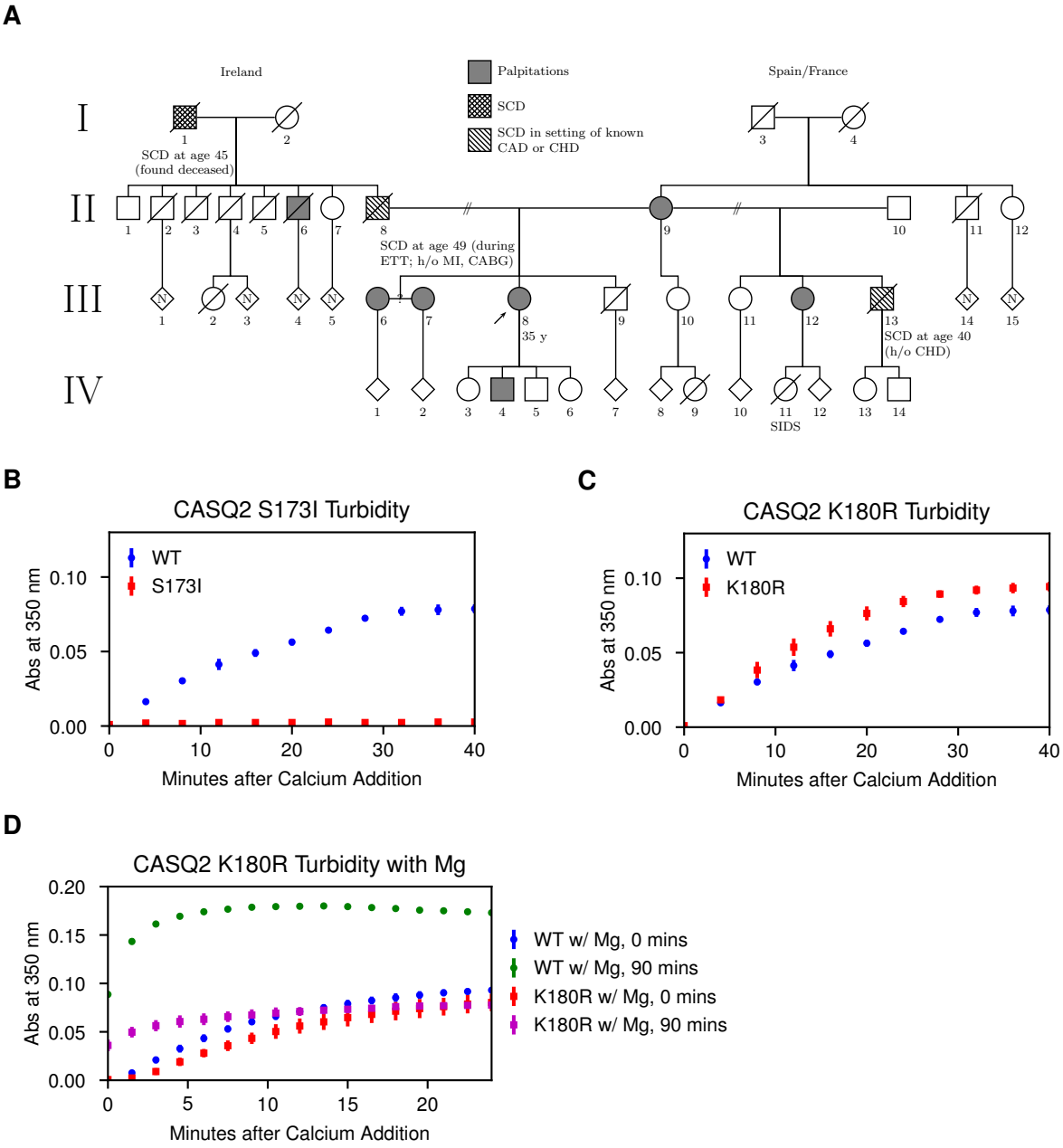


Figure 1: Dominant-acting CASQ2 disease mutations disrupt calsequestrin multimerization.
(A) Pedigree of a large extended family with the S173I mutation and a CPVT-like phenotype. CABG = coronary artery bypass graft; CAD = coronary artery disease; CHD = congenital heart disease; ETT = exercise treadmill test; MI = myocardial infarction; SCD = suddent cardiac death; SIDS = sudden infant death syndrome.
(B) Multimerization kinetics of the S173I mutant observed using a turbidity assay after addition of 1 mM $CaCl_2$ to purified protein (pH 7.4, 20 mM NaCl, 85 mM KCl).
(C) Multimerization kinetics of the K180R mutant observed using a turbidity assay (same conditions as in B).
(D) Multimerization kinetics of the K180R mutant observed using a turbidity assay (same conditions as in B, but with 2 mM $MgCl_2$ added prior to calcium).

Figure 2

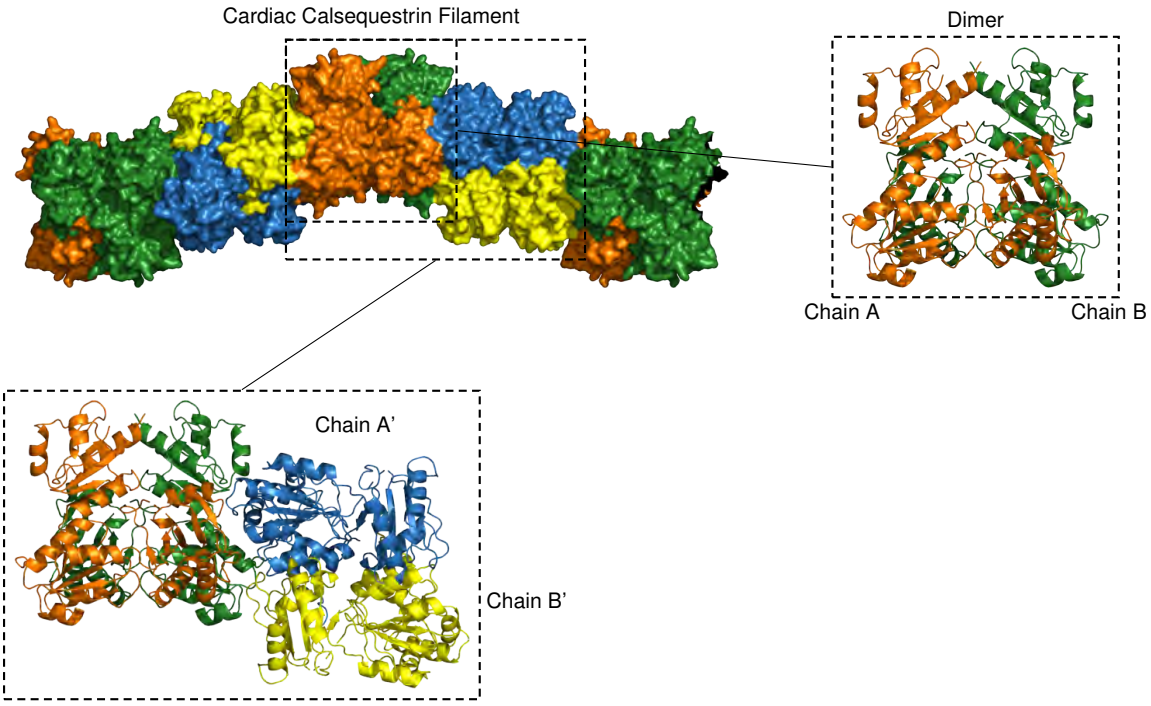


Figure 2: The cardiac calsequestrin filament candidate structure is assembled from calsequestrin dimers assembled on a screw axis with 90 degrees of rotation per dimer. The cardiac calsequestrin candidate filament (PDB ID 6OWV) is shown, along with a representative dimeric and tetrameric assembly.

Figure 3

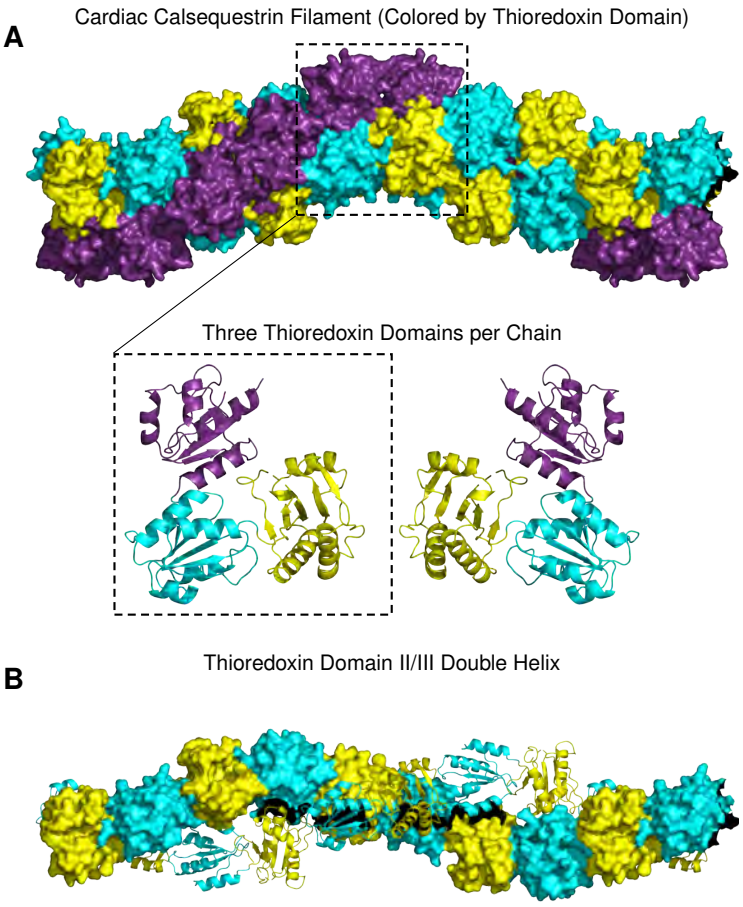


Figure 3: Viewed at the level of its thioredoxin domains (3 per protomer), the cardiac calsequestrin filament consists of an inner thioredoxin double helix with an outer thioredoxin single helix wrapping the double helical core.
(A) The cardiac calsequestrin filament rendered with thioredoxin domain colors, revealing the helical character of the filament. The outer single helix, consisting of domain I, winds around an inner double helix consisting of domains II and III. Cardiac calsequestrin monomers are colored by thioredoxin domain (domain I, purple; domain I, cyan; domain III, yellow). The monomers are translated but remain in their dimer-forming orientation with two-fold symmetry.
(B) The inner double helix of the cardiac calsequestrin filament consisting of thioredoxin domains II and III.

Figure 4

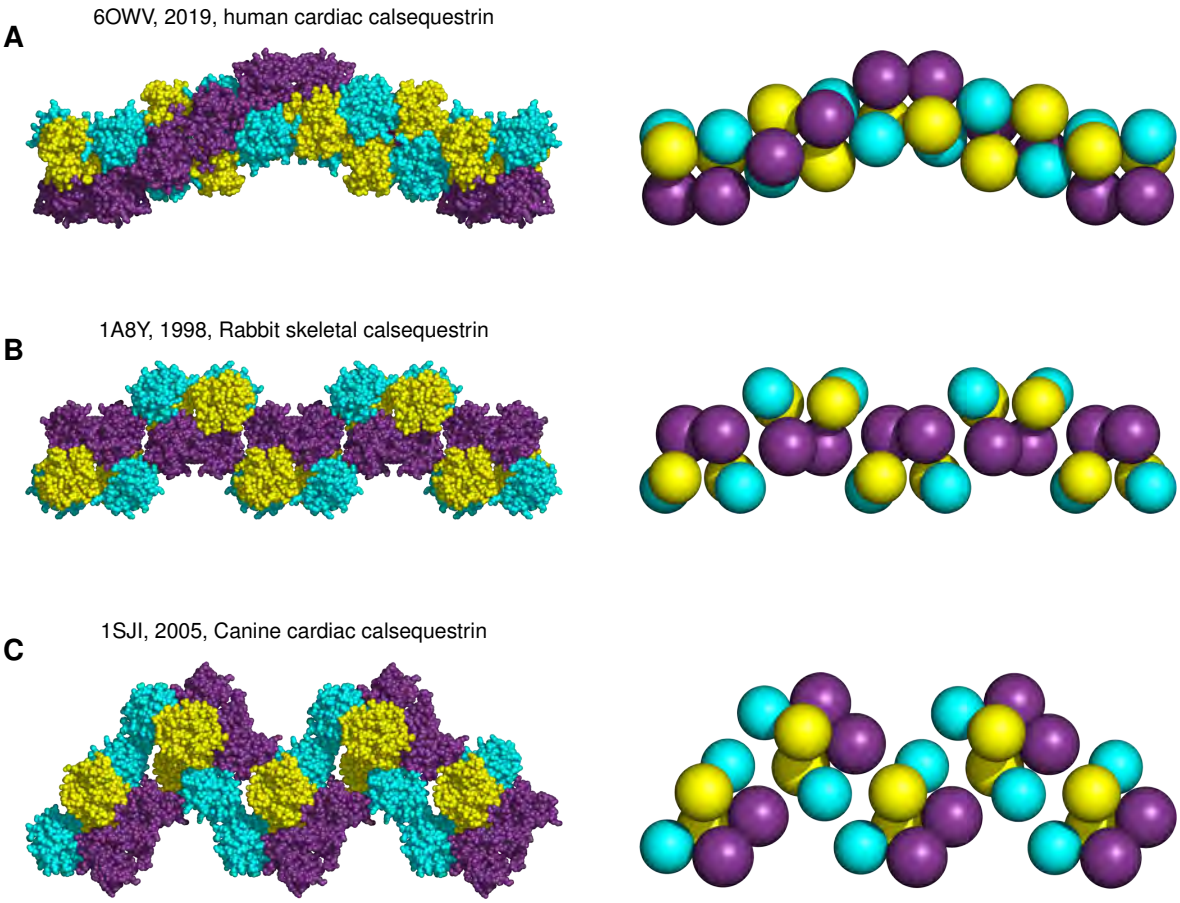


Figure 4: Prior putative calsequestrin filament candidates have fewer inter-domain contacts, fewer inter-protomer contacts, and substantially less buried surface area compared to the new candidate. The new candidate CASQ2 filament exhibits tight packing of protomers and thioredoxin domains (shown on the right using equal-size spheres placed at the center of mass of each thioredoxin domain).
(A) The new candidate cardiac calsequestrin filament assembled from crystallographic symmetry operations on 6OWV (human CASQ2, 2019).
(B) A putative skeletal calsequestrin filament assembled from crystallographic symmetry operations on 1A8Y (rabbit CASQ1, 1998).
(C) A putative skeletal calsequestrin filament assembled from crystallographic symmetry operations on 1SJI (canine CASQ2, 2005).

Figure 5

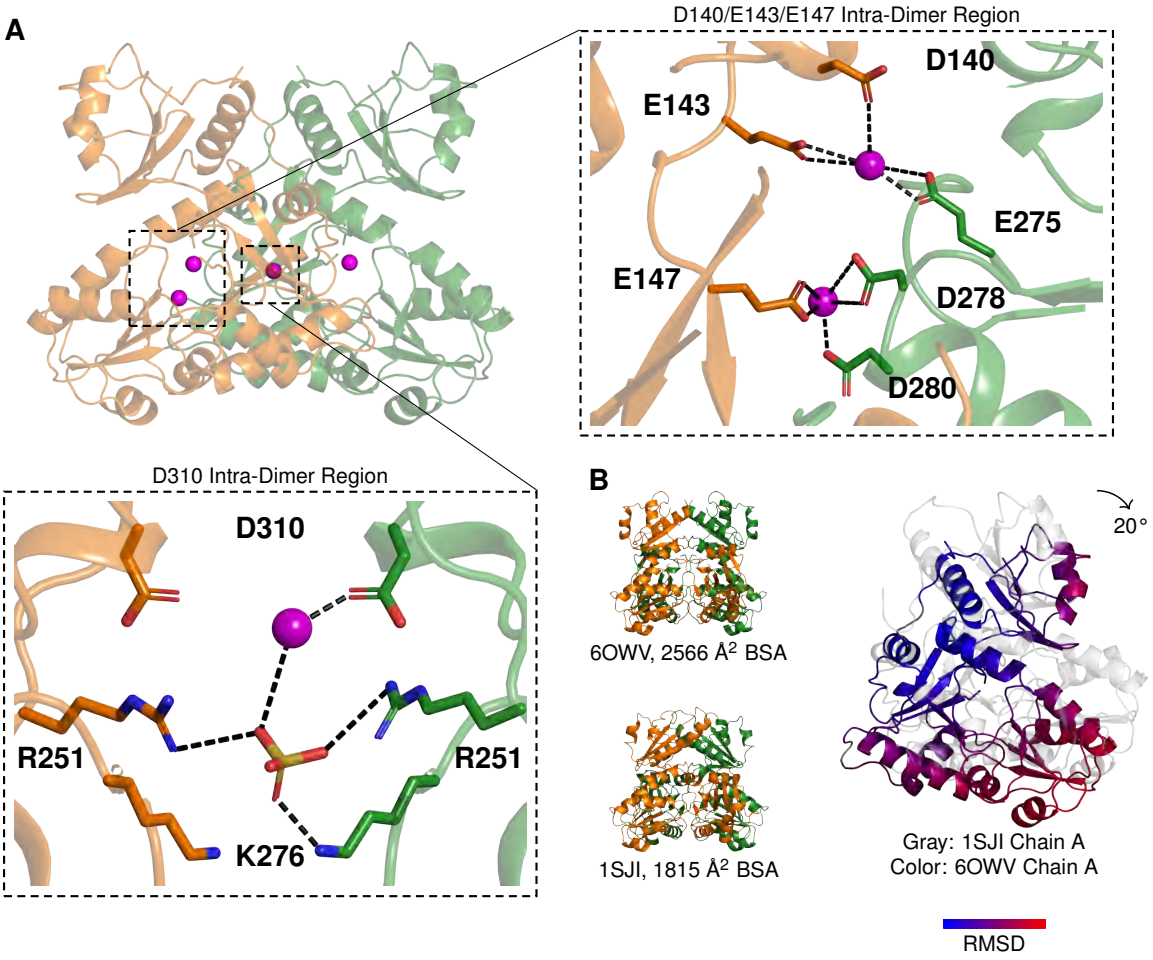


Figure 5: The intra-dimer interface of cardiac calsequestrin contains likely sites of calcium binding (identified by ytterbium anomalous signal).
(A) Dimer with Yb sites (magenta spheres) within its interior cavity. Closeups focus on Yb positions that bridge dimer chains A and B.
(B) Comparison of a previously published cardiac calsequestrin dimer (1SJI) to the more tightly-packed dimer that we report. The tightly-packed dimer results primarily from rigid body rotation of the dimer chains inward (for a single chain, we observe 20° counter-clockwise rotation in the plane of the page when the other chain is fixed to the reference dimer). The inward rotation produces an increase in buried surface area (BSA) in thioredoxin domains II and III.

Figure 6

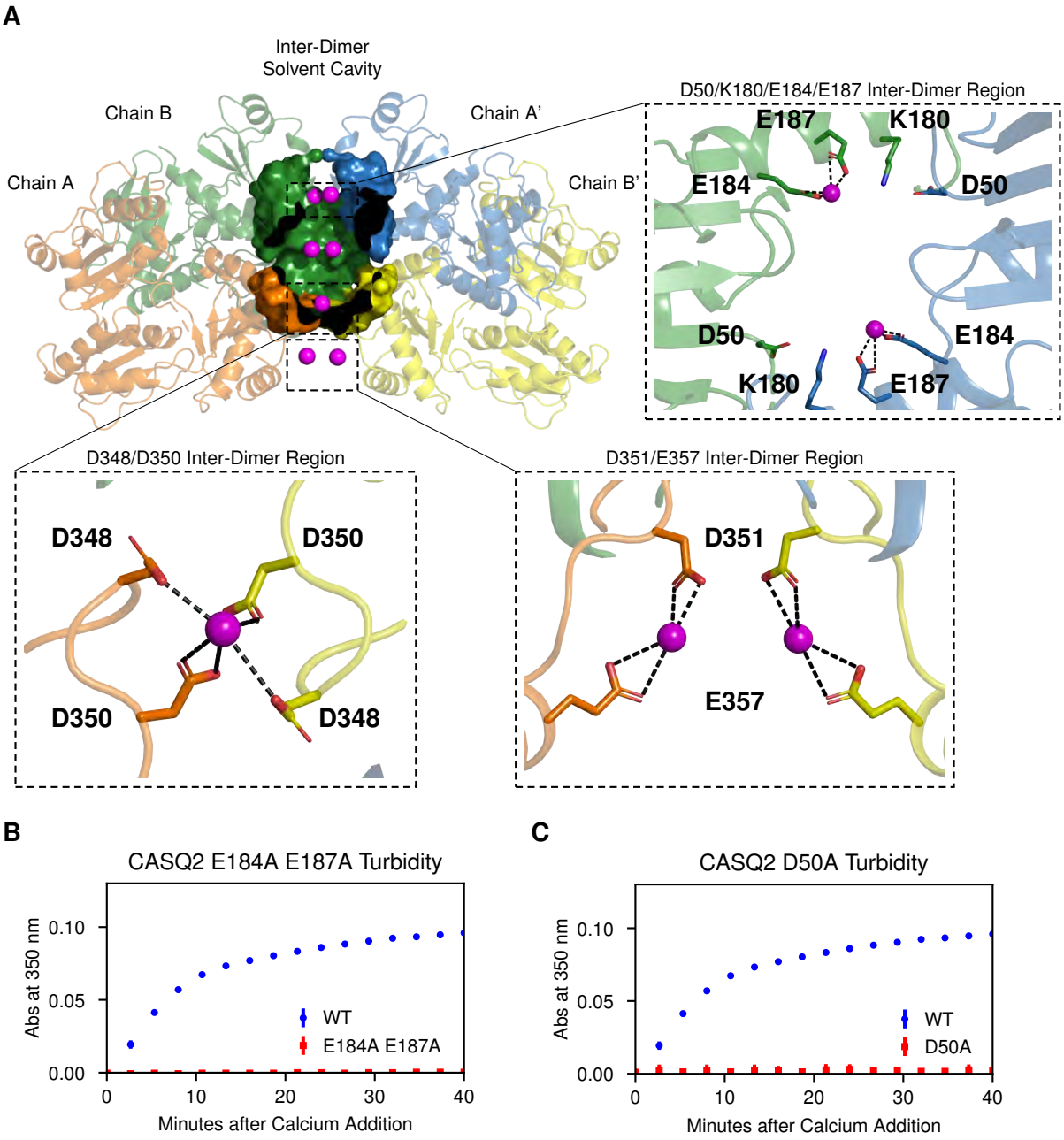


Figure 6: The inter-dimer interface of cardiac calsequestrin contains likely sites of calcium binding (identified by ytterbium anomalous signal). Disruption of these sites leads to filamentation defect.
(A) Yb (magenta spheres) bound within the walled pocket formed by the inter-dimer interface, with closeups of ytterbium site at E184 and E187, D348 and D350, and D351 and D357. Thioredoxin domain II of chain A' (blue) is omitted to allow visualization of the interior of the solvent pocket formed by the inter-dimer interface.
(B) Turbidity assay after alanine mutagenesis of the D50 residue that participates in a salt bridge with K180, adjacent to the E184/E187 ligand site.
(C) Turbidity assay after alanine mutagenesis of putative calcium-binding residues E184 and E187.

Figure 7

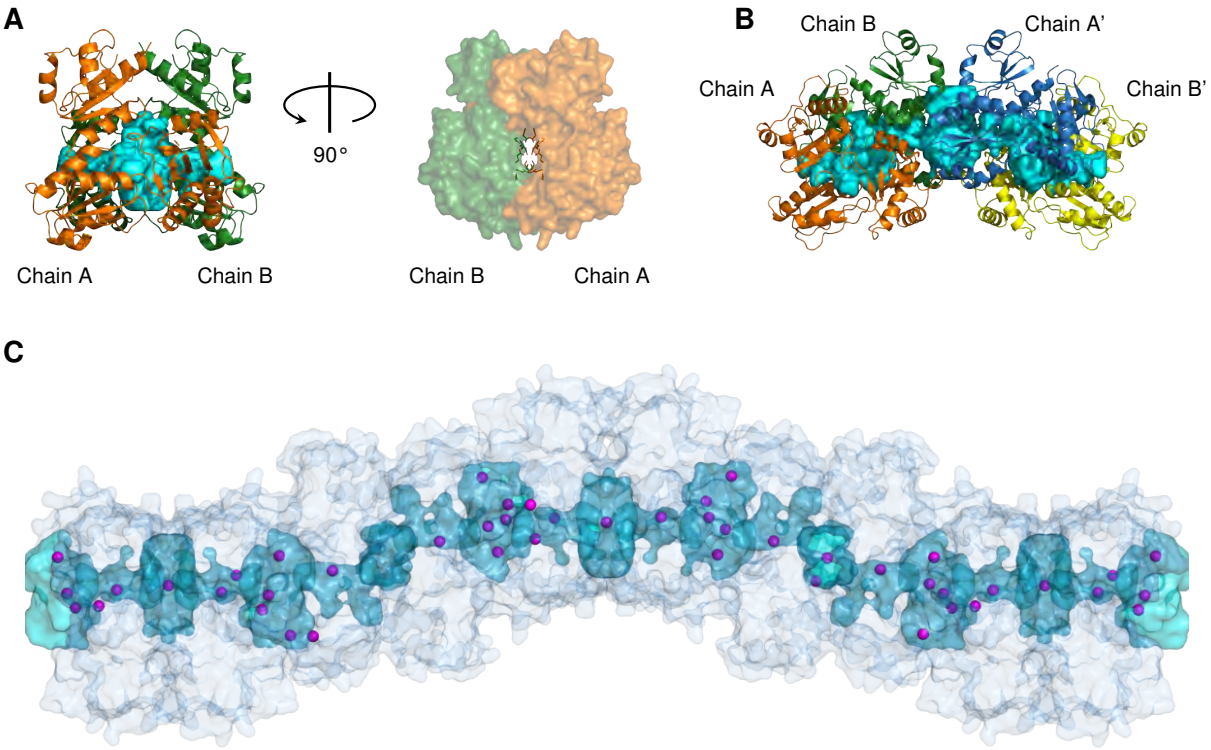


Figure 7: The cardiac calsequestrin filament possesses a continuous solvent-accessible interior cavity that winds along the filament long axis.
(A) The interior cavity of the dimer (colored cyan) as traced by HOLLOW using a 1.4 Åprobe. Also shown is the interior cavity viewed down its long axis. Residues that participate in the coordination of Yb atoms within the intra-dimer cleft (Figure 5) are shown as sticks. All other residues are rendered as surface.
(B) The lumen of the dimer is continuous with a large solvent pocket formed by the tetramer interface.
(C) View of the filament and its continuous interior cavity, with Yb sites shown as purple spheres.

Figure 8

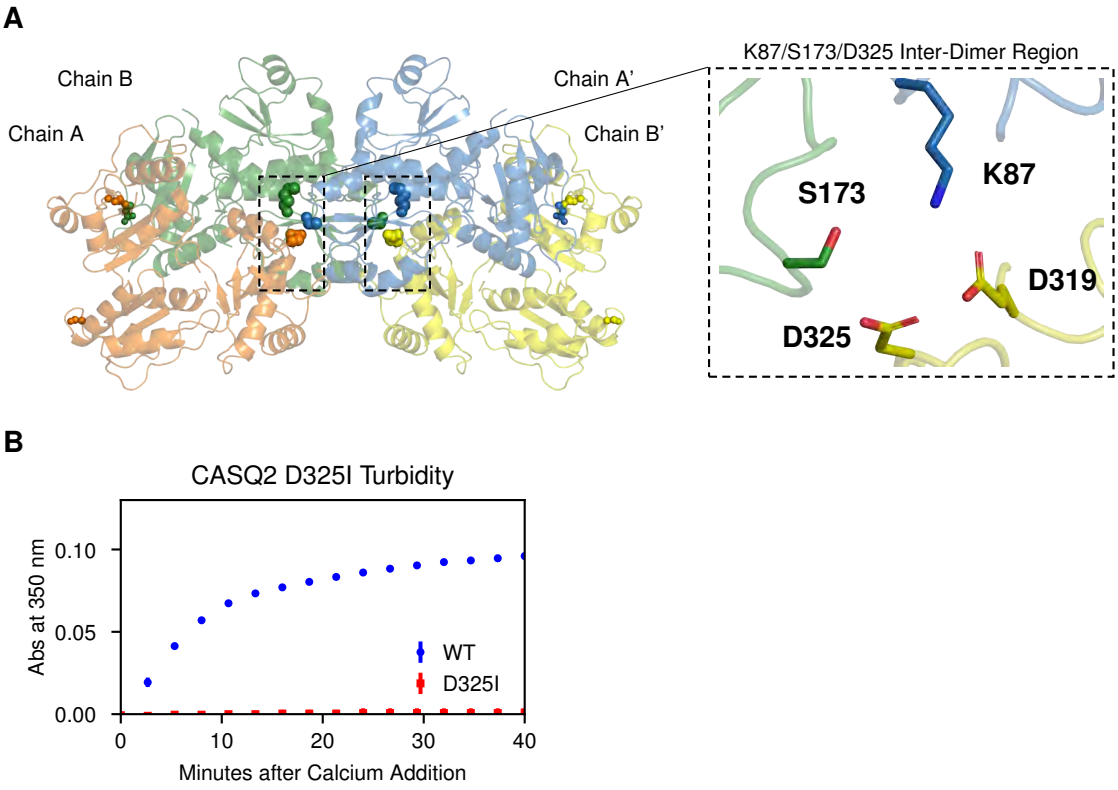


Figure 8: The cardiac calsequestrin S173 residue is located at a highly hydrophilic 3-protomer interface within the inter-dimer contact region. Disruption of this hydrophilic pocket by hydrophobic substitution leads to filamentation defect.
(A) Inter-dimer interface with with interface residues in the S173 region rendered as spheres. the closeup panel shows a charged pocket at the inter-dimer interface where the CPVT-associated S173I mutation is found. In this pocket, 3 different thioredoxin domains from 3 distinct chains interact (K87, S173, D325).
(B) Turbidity assay with the D325I mutation.

Supplemental Figures

Figure S1

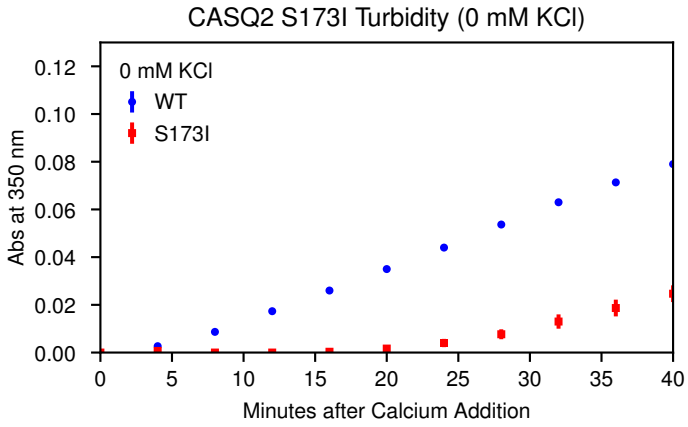


Figure S1: Multimerization kinetics of the S173I mutant observed in 0 mM KCl.

Figure S2

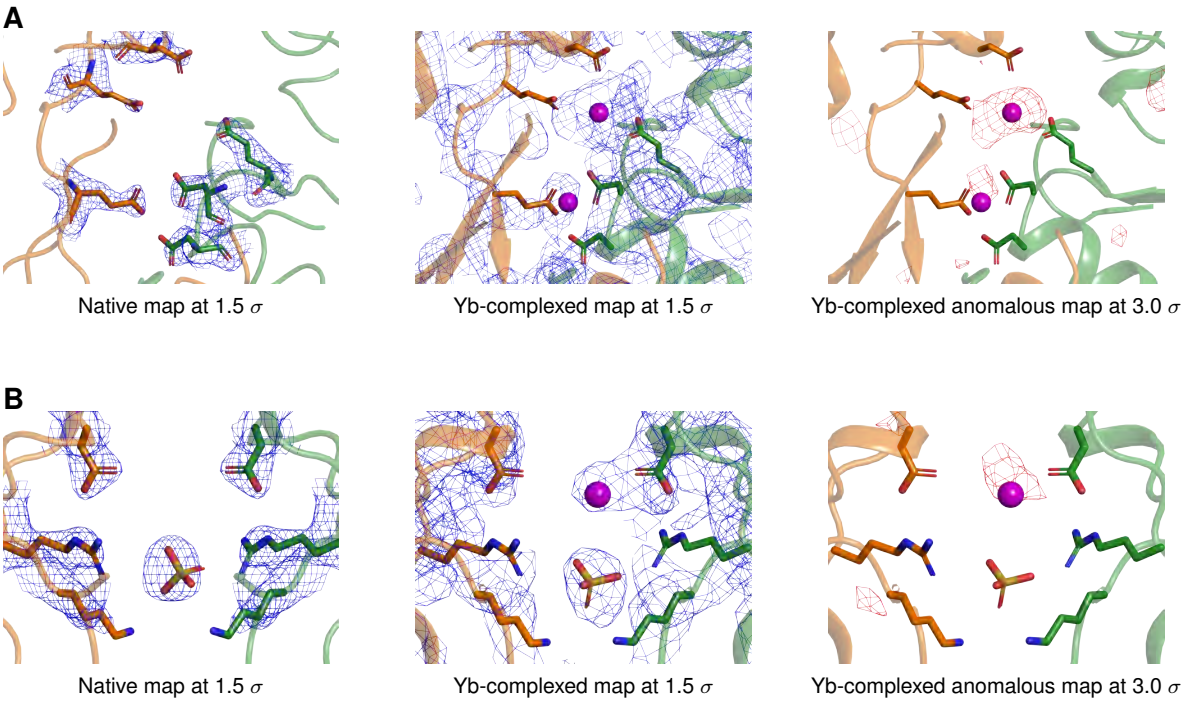


Figure S2: Electron density maps for Yb-binding sites at the cardiac calsequestrin intra-dimer interface.
(A) Electron density maps for the D140/E143/E147 region of interest.
(B) Electron density maps for the D310 region of interest.

Figure S3

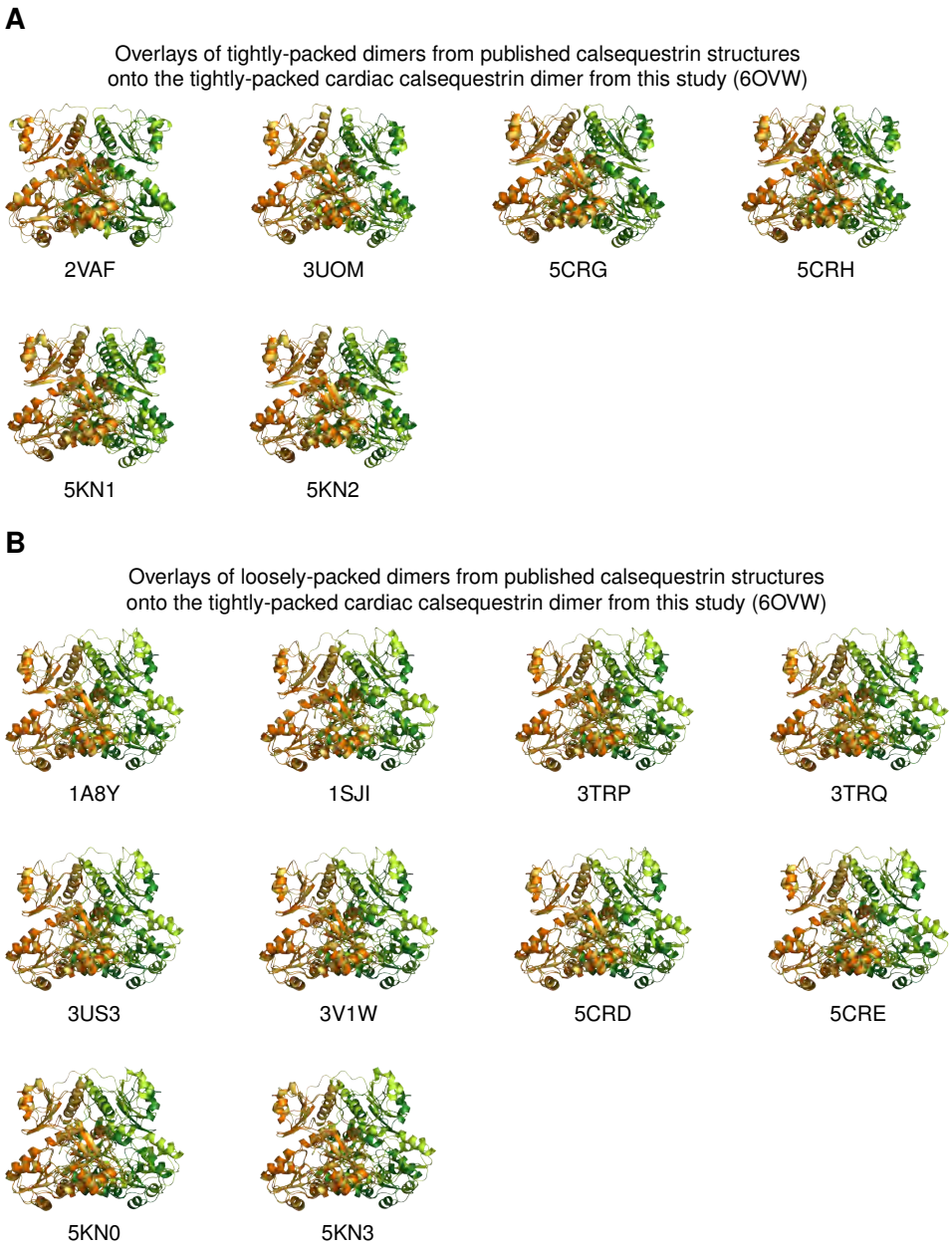


Figure S3: Overlays of dimers from published calsequestrin structures (ligher orange and green) onto the tightly-packed dimer from this study (darker orange and green) reveals two distinct conformational groupings. The more tightly-packed conformation with inwardly-rotated chains is correlated with presence of neutralizing cations. Dimers are aligned on one chain (chain A arbitrarily chosen) to show degree of inward rotation in the other chain.
(A) Overlays of tightly-packed dimer structures.
(B) Overlays of loosely-packed dimer structures.

Figure S4

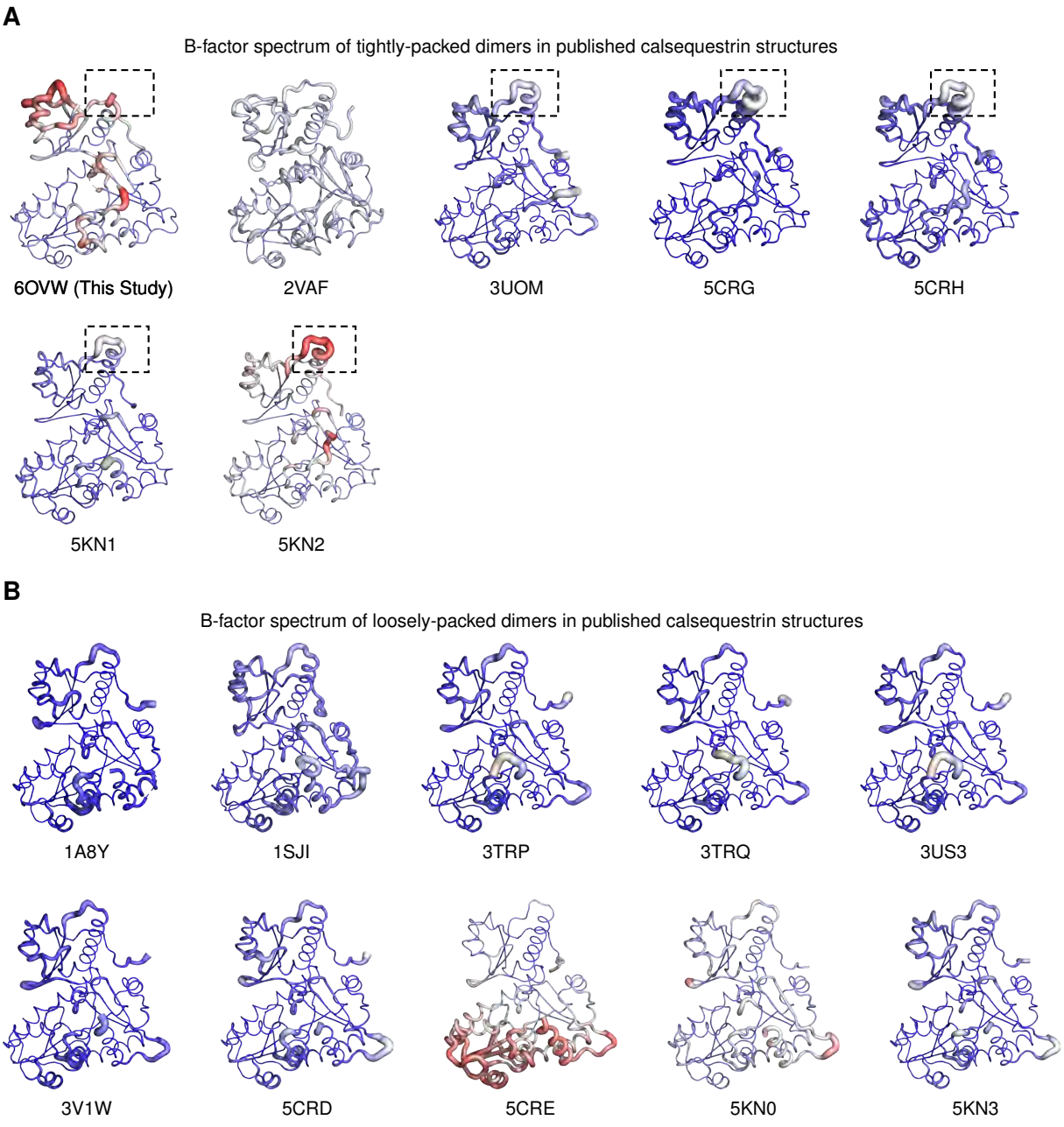
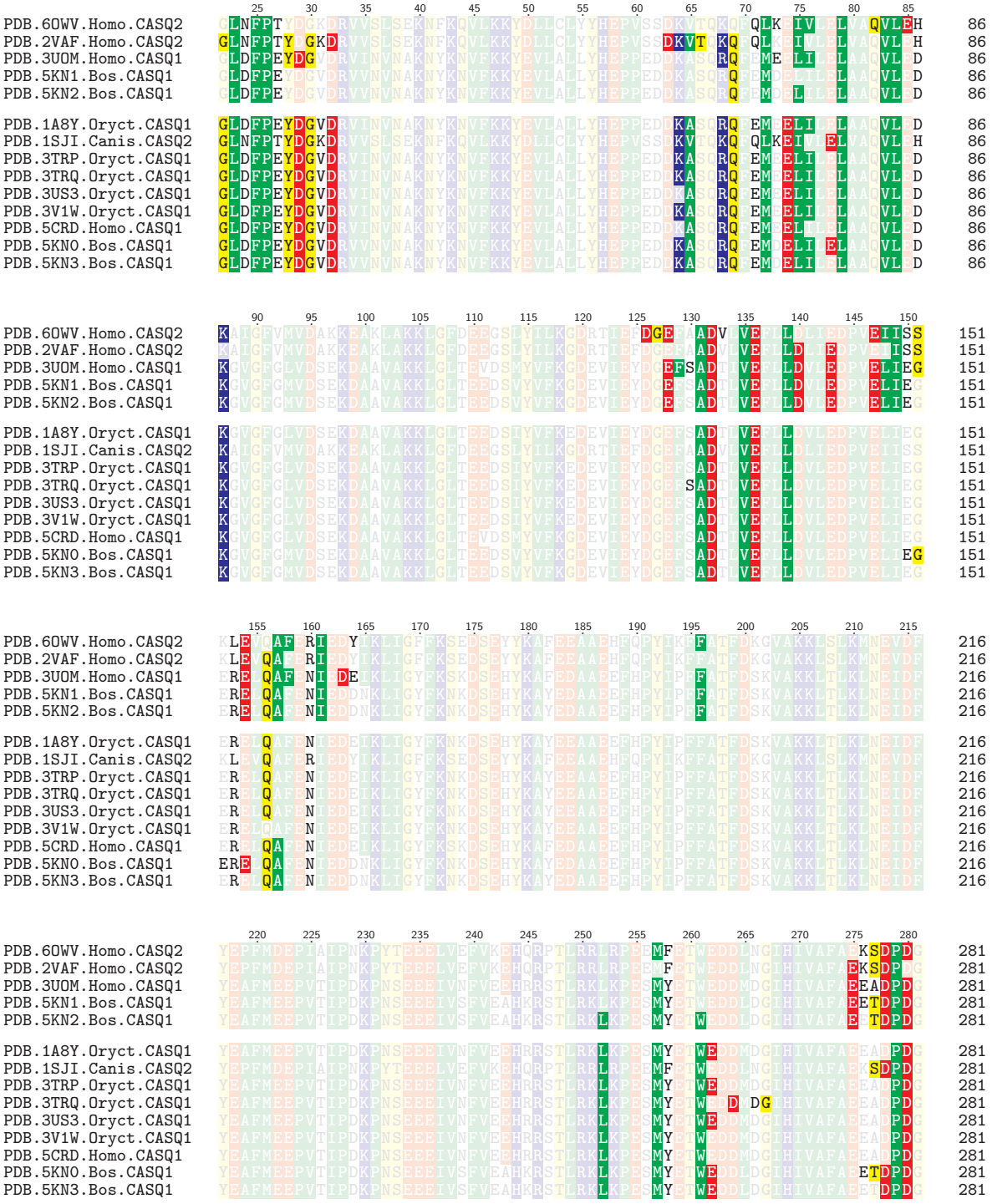


Figure S4: Comparing B-factors between tightly-packed and loosely-packed calsequestrin dimers reveals that the tightly-packed dimer exhibits increased conformational disorder in domain I.
(A) B-factor spectrum of tightly-packed dimers in published calsequestrin structures, with region of increased disorder boxed. In PDB 6OVW, corresponding to this study, the disordered loop region is omitted entirely due to the high level of disorder.
(B) B-factor spectrum of loosely-packed dimers in published calsequestrin structures.

Figure S5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PDB.2VAF.Homo.CASQ2
PDB.3UOM.Homo.CASQ1
PDB.5KN1.Bos.CASQ1
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PDB.1A8Y.Oryct.CASQ1
PDB.1SJI.Canis.CASQ2
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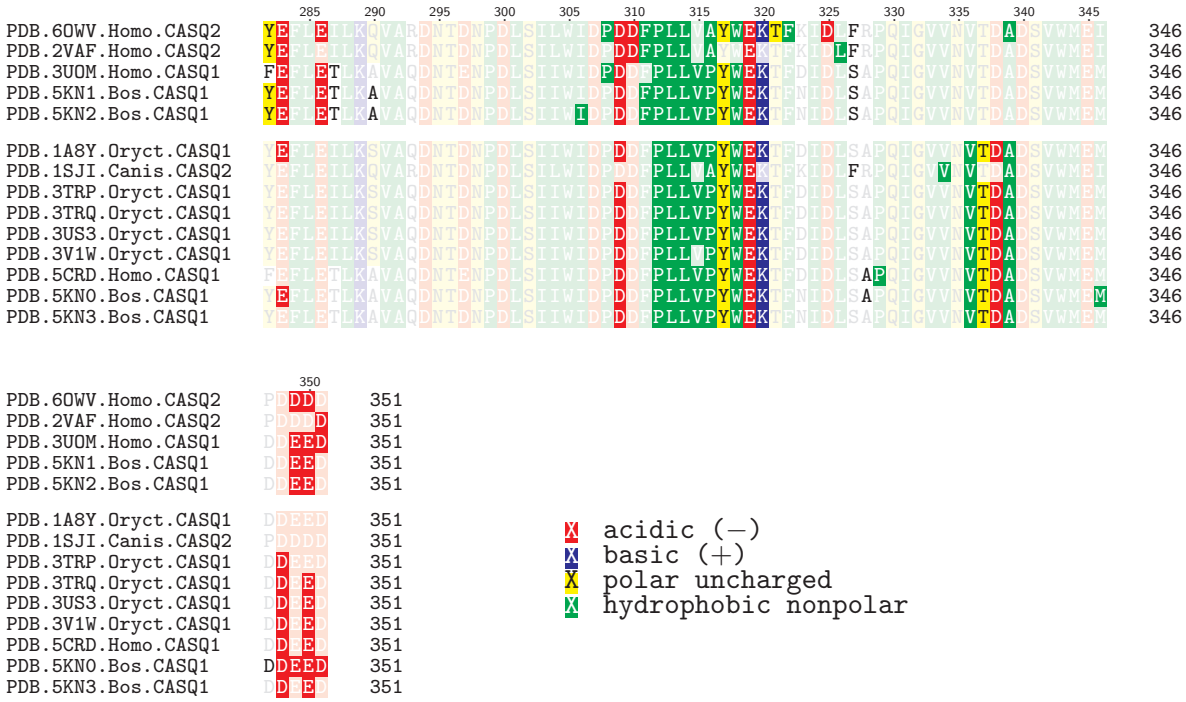


Figure S5: Multiple sequence alignment from published calsequestrin structures, grouped by dimer conformational class (top group: compact dimer; bottom group: relaxed dimer). Intra-dimer interface residues are highlighted; color represents hydropathy. Rotation of chains in compacted dimers leads to loss of contacts near the N terminus but gain of contacts elsewhere. Calsquestrin structures 5CRE, 5CRG, and 5CRH are omitted (point mutants belonging to the same investigation as 5CRD).

Figure S6

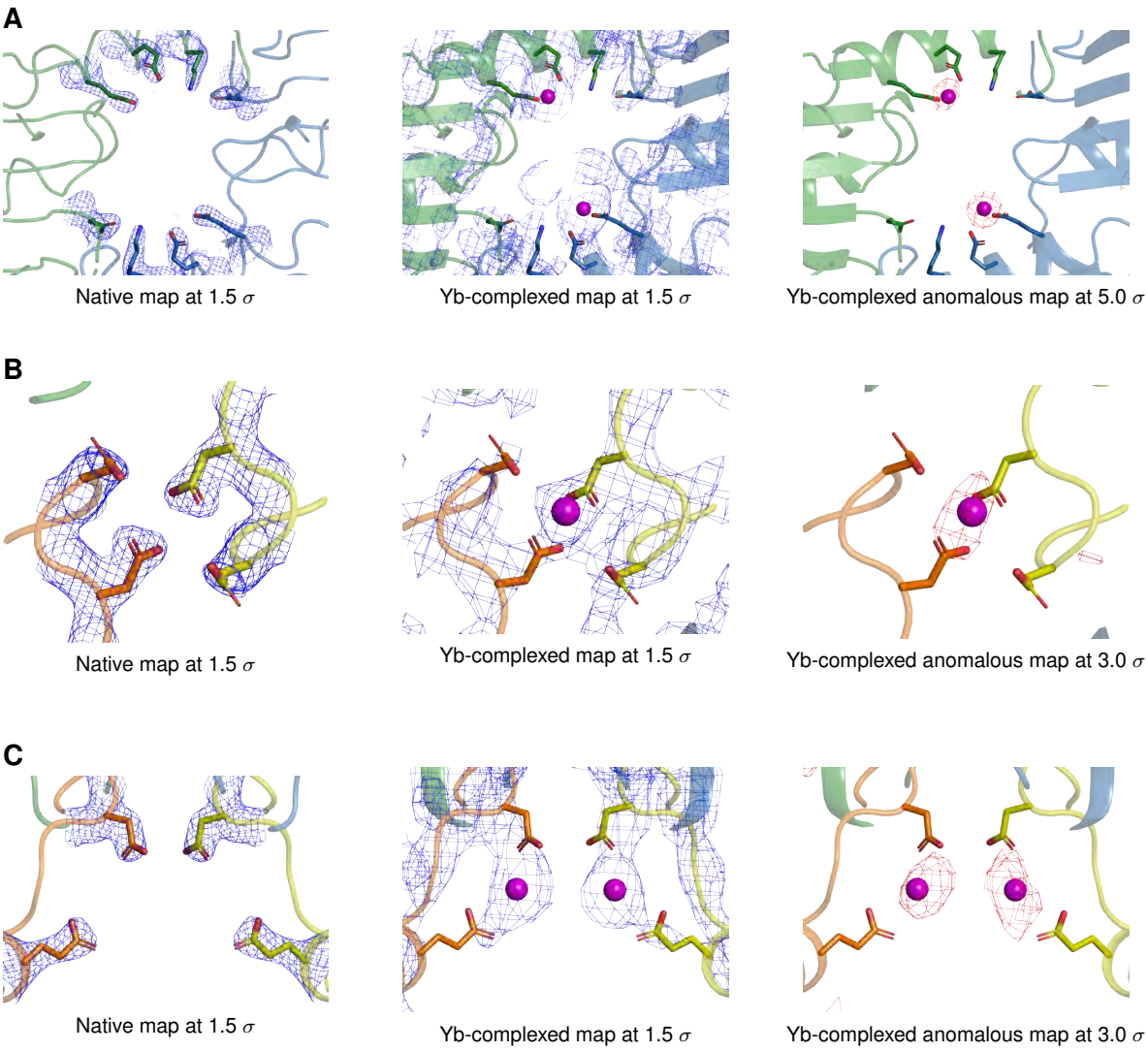


Figure S6: Electron density maps for Yb-binding sites at the cardiac calsequestrin inter-dimer interface.
(A) Electron density maps for the D50/K180/E184/E187 region of interest.
(B) Electron density maps for the D348/D350 region of interest.
(C) Electron density maps for the D351/E357 region of interest.

Figure S7

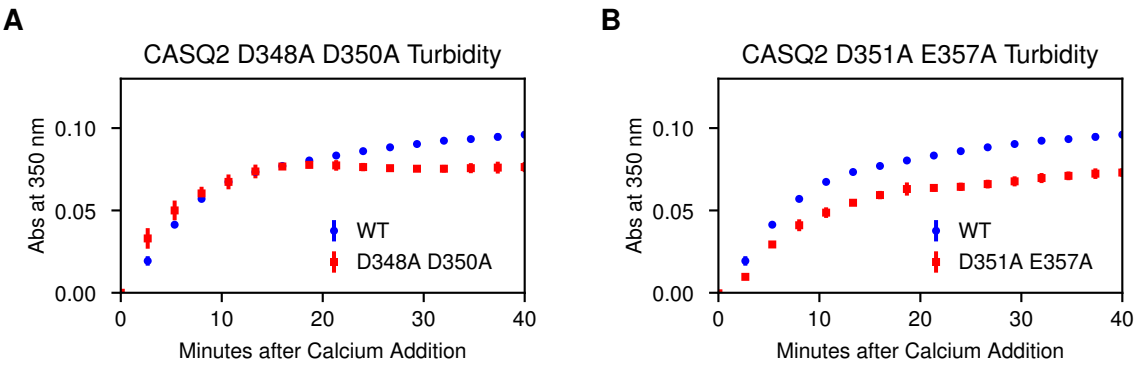
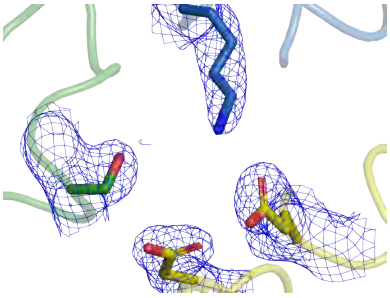


Figure S7: Turbidity assays showing effect of alanine mutagenesis of additional Yb-binding sites at the cardiac calsequestrin inter-dimer interface.
(A) Turbidity assay after alanine mutagenesis of the putative calcium-coordinating residues D348 and D350.
(B) Turbidity assay after alanine mutagenesis of the putative calcium-coordinating residues D351 and E357.

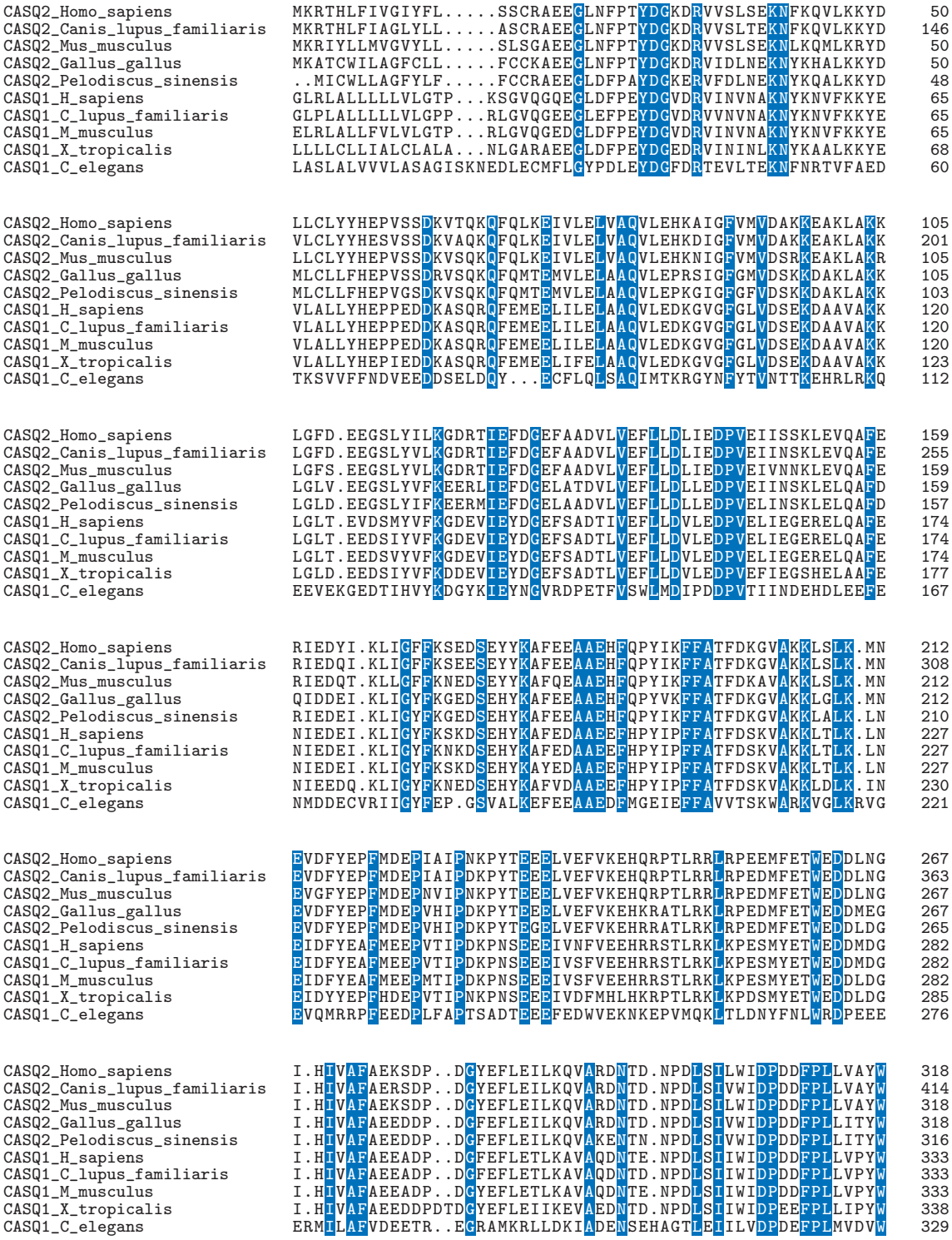
Figure S8



Native map at 1.5 σ

Figure S8: Electron density map for the S173 region at the cardiac calsequestrin inter-dimer interface.

Figure S9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CASQ2_Mus_musculus MKRIYLLMVGVYLL.....SLSGAEEGLNFPTYDGKDRVVSLSEKNLKQMLKRYD 50
CASQ2_Gallus_gallus MKATCWILAGFCLL.....FCCKAEEGLNFPTYDGKDRVIDLNEKNYKHALKKYD 50
CASQ2_Pelodiscus_sinensis ..MICWLLAGFYLF.....FCCRAEEGLDFPAYDGKERVFDLNEKNYKQALKKYD 48
CASQ1_H_sapiens GLRLALLLLLVLGTP...KSGVQGQEGLDFPEYDGVDRVINVNAKNYKNVFKKYE 65
CASQ1_C_lupus_familiaris GLPLALLLLLVLGPP...RLGVQGEEGLEFPEYDGVDRVVNVNAKNYKNVFKKYE 65
CASQ1_M_musculus ELRLALLFVLVLGTP...RLGVQGEDGLDFPEYDGVDRVINVNAKNYKNVFKKYE 65
CASQ1_X_tropicalis LLLLCLLIALCLALA...NLGARAEEGLDFPEYDGEDRVININLKNYKAALKKYE 68
CASQ1_C_elegans LASLALVVVLASAGISKNEDLECMFLGYPDLEYDGFDRTEVLTEKNFNRTVFAED 60
CASQ2_Homo_sapiens LLCLYYHEPVSSDKVTQKQFQLKEIVLELVAQVLEHKAIGFVMVDAKKEAKLAKK 105
CASQ2_Canis_lupus_familiaris VLCLYYHESVSSDKVAQKQFQLKEIVLELVAQVLEHKDIGFVMVDAKKEAKLAKK 201
CASQ2_Mus_musculus LLCLYYHEPVSSDKVSQKQFQLKEIVLELVAQVLEHKNIGFVMVDSRKEAKLAKR 105
CASQ2_Gallus_gallus MLCLLFHEPVSSDRVSQKQFQMTEMVLELAAQVLEPRSIGFGMVDSKKDAKLAKK 105
CASQ2_Pelodiscus_sinensis MLCLLFHEPVGSDKVSQKQFQMTEMVLELAAQVLEPKGIGFGFVDSKKDAKLAKK 103
CASQ1_H_sapiens VLALLYHEPPEDDKASQRQFEMEELILELAAQVLEDKGVGFGLVDSEKDAAVAKK 120
CASQ1_C_lupus_familiaris VLALLYHEPPEDDKASQRQFEMEELILELAAQVLEDKGVGFGLVDSEKDAAVAKK 120
CASQ1_M_musculus VLALLYHEPPEDDKASQRQFEMEELILELAAQVLEDKGVGFGLVDSEKDAAVAKK 120
CASQ1_X_tropicalis VLALLYHEPIEDDKASQRQFEMEELIFELAAQVLEDKGVGFGLVDSEKDAAVAKK 123
CASQ1_C_elegans TKSVVFFNDVEEDDSELDQY...ECFLQLSAQIMTKRGYNFYTVNTTKEHRLRKQ 112
CASQ2_Homo_sapiens LGFD.EEGSLYILKGDRTIEFDGEFAADVLVEFLLDLIEDPVEIISSKLEVQAFE 159
CASQ2_Canis_lupus_familiaris LGFD.EEGSLYVLKGDRTIEFDGEFAADVLVEFLLDLIEDPVEIINSKLEVQAFE 255
CASQ2_Mus_musculus LGFS.EEGSLYVLKGDRTIEFDGEFAADVLVEFLLDLIEDPVEIVNNKLEVQAFE 159
CASQ2_Gallus_gallus LGLV.EEGSLYVFKEERLIEFDGELATDVLVEFLLDLLEDPVEIINSKLELQAFD 159
CASQ2_Pelodiscus_sinensis LGLD.EEGSLYIFKEERMIEFDGELAADVLVEFLLDLLEDPVELINSKLELQAFD 157
CASQ1_H_sapiens LGLT.EVDSMYVFKGDEVIEYDGEFSADTIVEFLLDVLEDPVELIEGERELQAFE 174
CASQ1_C_lupus_familiaris LGLT.EEDSIYVFKGDEVIEYDGEFSADTLVEFLLDVLEDPVELIEGERELQAFE 174
CASQ1_M_musculus LGLT.EEDSVYVFKGDEVIEYDGEFSADTLVEFLLDVLEDPVELIEGERELQAFE 174
CASQ1_X_tropicalis LGLD.EEDSIYVFKDDEVIEYDGEFSADTLVEFLLDVLEDPVEFIEGSHELAAFE 177
CASQ1_C_elegans EEVEKGEDTIHVYKDGYKIEYNGVRDPETFVSWLMDIPDDPVTIINDEHDLEEFE 167
CASQ2_Homo_sapiens RIEDYI.KLIGFFKSEDSEYYKAFEEAAEHFQPYIKFFATFDKGVAKKLSLK.MN 212
CASQ2_Canis_lupus_familiaris RIEDQI.KLIGFFKSEESEYYKAFEEAAEHFQPYIKFFATFDKGVAKKLSLK.MN 308
CASQ2_Mus_musculus RIEDQT.KLLGFFKNEDSEYYKAFQEAAEHFQPYIKFFATFDKAVAKKLSLK.MN 212
CASQ2_Gallus_gallus QIDDEI.KLIGYFKGEDSEHYKAFEEAAEHFQPYVKFFATFDKGVAKKLGLK.MN 212
CASQ2_Pelodiscus_sinensis RIEDEI.KLIGYFKGEDSEHYKAFEEAAEHFQPYIKFFATFDKGVAKKLALK.LN 210
CASQ1_H_sapiens NIEDEI.KLIGYFKSKDSEHYKAFEDAAEEFHPYIPFFATFDSKVAKKLTLK.LN 227
CASQ1_C_lupus_familiaris NIEDEI.KLIGYFKNKDSEHYKAFEDAAEEFHPYIPFFATFDSKVAKKLTLK.LN 227
CASQ1_M_musculus NIEDEI.KLIGYFKSKDSEHYKAYEDAAEEFHPYIPFFATFDSKVAKKLTLK.LN 227
CASQ1_X_tropicalis NIEEDQ.KLIGYFKNEDSEHYKAFVDAAEEFHPYIPFFATFDSKVAKKLDLK.IN 230
CASQ1_C_elegans NMDDECVRIIGYFEP.GSVALKEFEEAAEDFMGEIEFFAVVTSKWARKVGLKRVG 221
CASQ2_Homo_sapiens EVDFYEPFMDEPIAIPNKPYTEEELVEFVKEHQRPTLRRLRPEEMFETWEDDLNG 267
CASQ2_Canis_lupus_familiaris EVDFYEPFMDEPIAIPDKPYTEEELVEFVKEHQRPTLRRLRPEDMFETWEDDLNG 363
CASQ2_Mus_musculus EVGFYEPFMDEPNVIPNKPYTEEELVEFVKEHQRPTLRRLRPEDMFETWEDDLNG 267
CASQ2_Gallus_gallus EVDFYEPFMDEPVHIPDKPYTEEELVEFVKEHKRATLRKLRPEDMFETWEDDMEG 267
CASQ2_Pelodiscus_sinensis EVDFYEPFMDEPVHIPDKPYTEGELVEFVKEHRRATLRKLRPEDMFETWEDDLDG 265
CASQ1_H_sapiens EIDFYEAFMEEPVTIPDKPNSEEEIVNFVEEHRRSTLRKLKPESMYETWEDDMDG 282
CASQ1_C_lupus_familiaris EIDFYEAFMEEPVTIPDKPNSEEEIVSFVEEHRRSTLRKLKPESMYETWEDDMDG 282
CASQ1_M_musculus EIDFYEAFMEEPMTIPDKPNSEEEIVSFVEEHRRSTLRKLKPESMYETWEDDLDG 282
CASQ1_X_tropicalis EIDYYEPFHDEPVTIPNKPNSEEEIVDFMHLHKRPTLRKLKPDSMYETWEDDLDG 285
CASQ1_C_elegans EVQMRRPFEEDPLFAPTSADTEEEFEDWVEKNKEPVMQKLTLDNYFNLWRDPEEE 276
CASQ2_Homo_sapiens I.HIVAFAEKSDP..DGYEFLEILKQVARDNTD.NPDLSILWIDPDDFPLLVAYW 318
CASQ2_Canis_lupus_familiaris I.HIVAFAERSDP..DGYEFLEILKQVARDNTD.NPDLSIVWIDPDDFPLLVAYW 414
CASQ2_Mus_musculus I.HIVAFAEKSDP..DGYEFLEILKQVARDNTD.NPDLSILWIDPDDFPLLVAYW 318
CASQ2_Gallus_gallus I.HIVAFAEEDDP..DGFEFLEILKQVARDNTD.NPDLSIVWIDPDDFPLLITYW 318
CASQ2_Pelodiscus_sinensis I.HIVAFAEEDDP..DGFEFLEILKQVAKENTN.NPDLSIVWIDPDDFPLLITYW 316
CASQ1_H_sapiens I.HIVAFAEEADP..DGFEFLETLKAVAQDNTE.NPDLSIIWIDPDDFPLLVPYW 333
CASQ1_C_lupus_familiaris I.HIVAFAEEADP..DGFEFLETLKAVAQDNTD.NPDLSIIWIDPDDFPLLVPYW 333
CASQ1_M_musculus I.HIVAFAEEADP..DGYEFLETLKAVAQDNTE.NPDLSIIWIDPDDFPLLVPYW 333
CASQ1_X_tropicalis I.HIVAFAEEDDPDTDGYEFLEIIKEVAEDNTD.NPDLSIIWIDPEEFPLLIPYW 338
CASQ1_C_elegans ERMILAFVDEETR..EGRAMKRLLDKIADENSEHAGTLEIILVDPDEFPLMVDVW 329

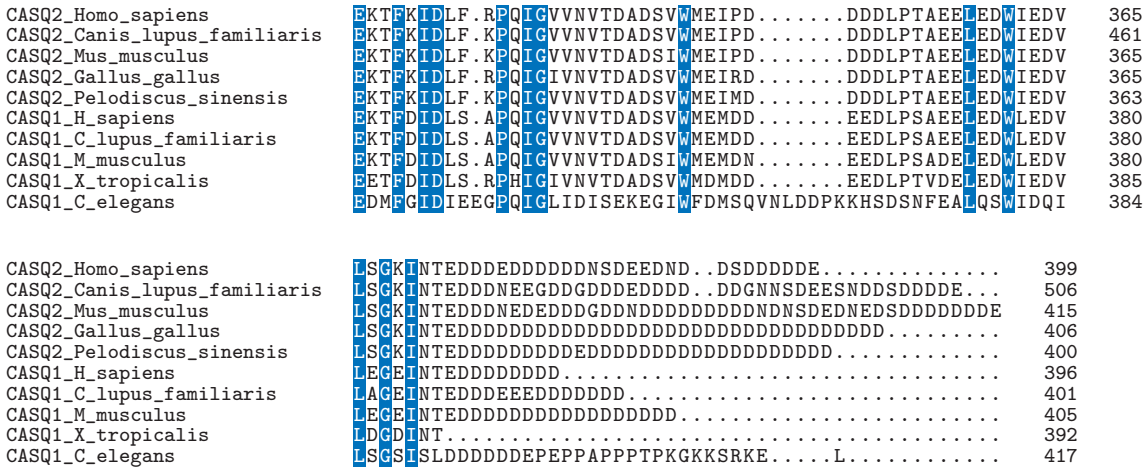


Figure S9: Multiple sequence alignment showing conservation across calsequestrins.

Supplemental Tables

Table S1: Crystallographic Data Collection and Refinement

	Native (6OWV)	**Ytterbium-Complexed (6OWW)**
Data Collection		
Wavelength (Å)	1.116	1.386
Space Group	P 43 2 2	P 1 21 1
Cell Dimensions		
a, b, c (Å)	62.5329, 62.5329, 213.189	85.8318, 86.0152, 214.34
α,β,γ (°)	90, 90, 90	90, 89.9072, 90
Resolution (Å)	53.94 - 1.88 (1.947 - 1.88)	107.2 - 3.84 (3.977 - 3.84)
R_{pim} (%)	1.8 (194.8)	16.2 (85.0)
$CC_{1/2}$	0.999 (0.422)	0.992 (0.616)
Mn(I)/σ(I)	14.82 (0.52)	4.52 (0.96)
Completeness (%)	97.76 (79.77)	98.45 (98.05)
Multiplicity	22.5 (12.6)	11.2 (11.5)
No. Unique Reflections	35501 (3425)	29897 (2924)
Refinement		
R_{work} / R_{free} (%)	21.4 / 25.4	29.1 / 34.2
Resolution (Å)	53.94 - 1.88	107.2 - 3.84
No. of chains in AU	1	8
Number of non-hydrogen atoms	2779	21752
Protein	2718	21665
Ligand	27	87
Solvent	34	N/A
B-Factors		
Protein	81.93	97.36
Ligand	112.54	135.46
Solvent	55.84	N/A
R.m.s. deviations		
Bond lengths (Å)	0.009	0.008
Bond angles (°)	1.25	1.14

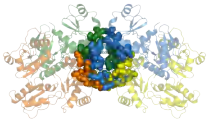
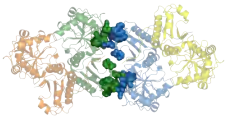
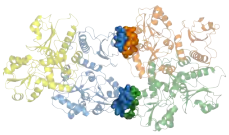
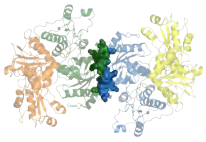
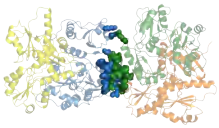
Table S2: Comparison of Crystallographic Space Group and Unit Cell Across All Published Calsequestrin Structures

PDB Code	Year	Protein	Space Group	Unit Cell Edges	Unit Cell Angles
6OWV	2019	CASQ2 (Homo)	P 43 2 2	62.52, 62.52, 213.15	90, 90, 90
1A8Y	1998	CASQ1 (Oryctolagus)	C 2 2 21	59.740, 145.560, 111.790	90.00, 90.00, 90.00
1SJI	2004	CASQ2 (Canis)	I 4	145.188, 145.188, 99.82	90, 90, 90
2VAF	2007	CASQ2 (Homo)	I 41 2 2	150.650, 150.650, 227.470	90.00, 90.00, 90.00
3TRP	2012	CASQ1 (Oryctolagus)	C 2 2 21	59.122, 144.863, 110.376	90.00, 90.00, 90.00
3TRQ	2012	CASQ1 (Oryctolagus)	C 2 2 21	59.271, 144.565, 111.170	90.00, 90.00, 90.00
3UOM	2012	CASQ1 (Homo)	P 1	89.799, 89.793, 119.161	90.13, 89.90, 60.05
3US3	2012	CASQ1 (Oryctolagus)	C 2 2 21	59.214, 144.811, 110.471	90.00, 90.00, 90.00
3V1W	2012	CASQ1 (Oryctolagus)	C 2 2 21	59.082, 144.592, 110.965	90.00, 90.00, 90.00
5CRD	2015	CASQ1 (Homo)	C 2 2 21	59.170, 145.132, 110.242	90.00, 90.00, 90.00
5CRE	2015	CASQ1 (Homo)	P 21 21 2	66.106, 82.815, 89.269	90.00, 90.00, 90.00
5CRG	2015	CASQ1 (Homo)	P 1 21 1	91.179, 67.462, 158.062	90.00, 96.48, 90.00
5CRH	2015	CASQ1 (Homo)	P 1 21 1	65.681, 68.553, 99.262	90.00, 92.84, 90.00
5KN0	2016	CASQ1 (Bos)	P 1	60.342, 92.994, 101.849	71.12, 84.57, 73.48
5KN1	2015	CASQ1 (Bos)	C 2 2 21	135.669, 165.604, 156.626	90.00, 90.00, 90.00
5KN2	2016	CASQ1 (Bos)	C 2 2 21	130.363, 169.194, 155.477	90.00, 90.00, 90.00
5KN3	2016	CASQ1 (Bos)	C 2 2 21	59.393, 146.060, 110.340	90.00, 90.00, 90.00

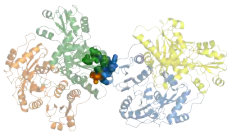
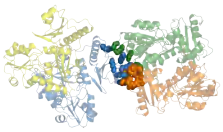
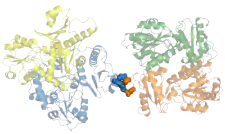
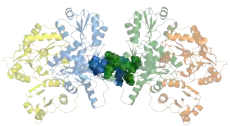
Table S3: Comparison of buried surface area (BSA) at dimer interfaces across all published calsequestrin structures. All calsequestrin dimers observed to date have the same symmetry but can be divided into two classes based on rigid body rotations of the chains (see Figure 5 for additional details).

PDB Code	CASQ2/CASQ1	Packing	BSA (Å^2)	DeltaG
6OWV	CASQ2	Tightly-Packed	2566.0	-29.2
1A8Y	CASQ1	Loosely-Packed	2051.0	-25.0
1SJI	CASQ2	Loosely-Packed	1815.0	-27.6
2VAF	CASQ2	Tightly-Packed	2178.0	-25.4
3TRP	CASQ1	Loosely-Packed	2201.0	-25.5
3TRQ	CASQ1	Loosely-Packed	2326.0	-23.7
3UOM	CASQ1	Tightly-Packed	2463.0	-24.5
3US3	CASQ1	Loosely-Packed	2174.0	-25.7
3V1W	CASQ1	Loosely-Packed	2163.0	-25.6
5CRD	CASQ1	Loosely-Packed	2190.0	-25.1
5CRE	CASQ1	Loosely-Packed	2058.0	-25.3
5CRG	CASQ1	Tightly-Packed	2447.0	-29.7
5CRH	CASQ1	Tightly-Packed	2458.0	-30.9
5KN0	CASQ1	Loosely-Packed	2482.0	-23.3
5KN1	CASQ1	Tightly-Packed	2038.0	-22.7
5KN2	CASQ1	Tightly-Packed	2265.0	-28.1
5KN3	CASQ1	Loosely-Packed	2219.0	-23.4

Table S4: Comparison of buried surface area (BSA) at putative dimer-dimer multimerization interfaces observed in all published calsequestrin structures. Residues with buried surfaces area at a dimer-dimer interface are rendered as spheres.

PDB Code	CASQ2/CASQ1	Interface	BSA (Å^2)	Oriented View	Equivalent PDBs[1]
6OWV	CASQ2	Tetramer (BC)	734.0		6OWW
		Tetramer (AC)	382.0		
		Tetramer (BD)	381.0		
		Tetramer (AD)	101.0		
1A8Y	CASQ1	Tetramer (BC)	715.0		3TRP, 3TRQ, 3US3, 3V1W, 5CRD, 5KN0, 5KN3
1SJI	CASQ2	Tetramer (BC)	305.0		
		Tetramer (AC)	188.0		
2VAF	CASQ2	Tetramer (BC)	509.0		
3UOM	CASQ1	Tetramer (BC)	724.0		

. . . continued

PDB Code	CASQ2/CASQ1	Interface	BSA (Å^2)	Oriented View	Equivalent PDBs
5CRE	CASQ1	Tetramer (BC)	255.0		
		Tetramer (AC)	162.0		
5CRG	CASQ1	Tetramer (AC)	304.0		
		Tetramer (BC)	74.0		
5CRH	CASQ1	Tetramer (AC)	102.0		
5KN1	CASQ1	Tetramer (BC)	436.0		5KN2

[1]Where equivalent PDB codes are listed, space group and unit cell, and consequently interfaces, are isomorphous (with the exception of 5KN0, which contains a tetramer equivalent to 1A8Y, but the structure was determined in P1).

Table S5: Oligonucleotides used in this study.

Oligonucleotide	**Sequence**
Primer to amplify H. sapiens CASQ2, forward	ATGAAGAGAACTCACTTGTTTATTGT
Primer to amplify H. sapiens CASQ2, reverse	CTATTCATCATCATCGTCATCACTGT
Primer to amplify H. sapiens CASQ2 with homology tails for the pET28 vector, forward (skips signal peptide sequence)	TTCCAGGGCCATATG GCTAGC AGGGCAGAAGAGGGGCTTA
Primer to amplify H. sapiens CASQ2 with homology tails for the pET28 vector, reverse	CGGAGCTCGAATTC GGATCC CTATTCATCATCATCGTCATCACTGT
Primer for site-directed mutagenesis of H. sapiens CASQ2, D50A, forward	AAAGAAATATgccTTGCTTTGCC
Primer for site-directed mutagenesis of H. sapiens CASQ2, D50A, reverse	AAAACCTGCTTGAAGTTCTTC
Primer for site-directed mutagenesis of H. sapiens CASQ2, D144A, forward	CTAATTGAAGcCCCAGTGGAG
Primer for site-directed mutagenesis of H. sapiens CASQ2, D144A, reverse	ATCCAAGAGGAACTCCAC
Primer for site-directed mutagenesis of H. sapiens CASQ2, E174A, forward	TTCAAGAGTGccGACTCAGAATACTAC
Primer for site-directed mutagenesis of H. sapiens CASQ2, E174A, reverse	AAAGCCAATGAGTTTGATG
Primer for site-directed mutagenesis of H. sapiens CASQ2, S173I, forward	CTTTTTCAAGattGAGGACTCAGAATAC
Primer for site-directed mutagenesis of H. sapiens CASQ2, S173I, reverse	CCAATGAGTTTGATGTAGTC
Primer for site-directed mutagenesis of H. sapiens CASQ2, K180R, forward	AGAATACTACcggGCTTTTGAAGAAGC

. . . continued

Oligonucleotide	**Sequence**
Primer for site-directed mutagenesis of H. sapiens CASQ2, K180R, reverse	GAGTCCTCACTCTTGAAAAAG
Primer for site-directed mutagenesis of H. sapiens CASQ2, E184A, forward	GCTTTTGAAGcAGCAGCTGAAC
Primer for site-directed mutagenesis of H. sapiens CASQ2, E184A, reverse	CTTGTAGTATTCTGAGTCC
Primer for site-directed mutagenesis of H. sapiens CASQ2, E187A on E184A background, forward	GCAGCAGCTGcACACTTCCAG
Primer for site-directed mutagenesis of H. sapiens CASQ2, E187A on E184A background, reverse	TTCAAAAGCCTTGTAGTATTCTG
Primer for site-directed mutagenesis of H. sapiens CASQ2, D325I, forward	TTTCAAGATTatCCTATTCAGGCCACAG
Primer for site-directed mutagenesis of H. sapiens CASQ2, D325I, reverse	GTCTTCTCCCAGTAGGCA
Primer for site-directed mutagenesis of H. sapiens CASQ2, D348A, forward	GAGATTCCAGccGATGACGATCTTCC
Primer for site-directed mutagenesis of H. sapiens CASQ2, D348A, reverse	CATCCAGACACTGTCAGC
Primer for site-directed mutagenesis of H. sapiens CASQ2, D350A on D348A background, forward	CCAGCCGATGcCGATCTTCCA
Primer for site-directed mutagenesis of H. sapiens CASQ2, D350A on D348A background, reverse	AATCTCCATCCAGACACTG
Primer for site-directed mutagenesis of H. sapiens CASQ2, D351A, forward	GATGATGACGccCTTCCAACTGC

. . . continued

Oligonucleotide	**Sequence**
Primer for site-directed mutagenesis of H. sapiens CASQ2, D351A, reverse	TGGAATCTCCATCCAGAC
Primer for site-directed mutagenesis of H. sapiens CASQ2, E357A on D351A background, forward	ACTGCTGAGGccCTGGAGGACTG
Primer for site-directed mutagenesis of H. sapiens CASQ2, E357A on D351A background, reverse	TGGAAGGGCGTCATCATC

References

[1] E. Carafoli and J. Krebs, "Why calcium? how calcium became the best communicator," *J. Biol. Chem.*, vol. 291, pp. 20849–20857, Sept. 2016.

[2] M. Prakriya and R. S. Lewis, "Store-Operated calcium channels," *Physiol. Rev.*, vol. 95, pp. 1383–1436, Oct. 2015.

[3] D. H. MacLennan and P. T. Wong, "Isolation of a calcium-sequestering protein from sarcoplasmic reticulum," *Proc. Natl. Acad. Sci. U. S. A.*, vol. 68, pp. 1231–1235, June 1971.

[4] D. H. MacLennan, "Isolation of a second form of calsequestrin," *J. Biol. Chem.*, vol. 249, pp. 980–984, Feb. 1974.

[5] T. J. Ostwald and D. H. MacLennan, "Isolation of a high affinity calcium-binding protein from sarcoplasmic reticulum," *J. Biol. Chem.*, vol. 249, pp. 974–979, Feb. 1974.

[6] B. Costello, C. Chadwick, A. Saito, A. Chu, A. Maurer, and S. Fleischer, "Characterization of the junctional face membrane from terminal cisternae of sarcoplasmic reticulum," *J. Cell Biol.*, vol. 103, pp. 741–753, Sept. 1986.

[7] C. Franzini-Armstrong, L. J. Kenney, and E. Varriano-Marston, "The structure of calsequestrin in triads of vertebrate skeletal muscle: a deep-etch study," *J. Cell Biol.*, vol. 105, pp. 49–56, July 1987.

[8] S. Wang, W. R. Trumble, H. Liao, C. R. Wesson, A. K. Dunker, and C. H. Kang, "Crystal structure of calsequestrin from rabbit skeletal muscle sarcoplasmic reticulum," *Nat. Struct. Biol.*, vol. 5, pp. 476–483, June 1998.

[9] B. C. Knollmann, N. Chopra, T. Hlaing, B. Akin, T. Yang, K. Ettensohn, B. E. C. Knollmann, K. D. Horton, N. J. Weissman, I. Holinstat, W. Zhang, D. M. Roden, L. R. Jones, C. Franzini-Armstrong, and K. Pfeifer, "Casq2 deletion causes sarcoplasmic reticulum volume increase, premature ca2+ release, and catecholaminergic polymorphic ventricular tachycardia," *J. Clin. Invest.*, vol. 116, pp. 2510–2520, Sept. 2006.

[10] D. M. Bers, “Macromolecular complexes regulating cardiac ryanodine receptor function,” *J. Mol. Cell. Cardiol.*, vol. 37, pp. 417–429, Aug. 2004.

[11] H. Park, I. Y. Park, E. Kim, B. Youn, K. Fields, A. K. Dunker, and C. Kang, “Comparing skeletal and cardiac calsequestrin structures and their calcium binding: a proposed mechanism for coupled calcium binding and protein polymerization,” *J. Biol. Chem.*, vol. 279, pp. 18026–18033, Apr. 2004.

[12] S. Perni, M. Close, and C. Franzini-Armstrong, “Novel details of calsequestrin gel conformation in situ,” *J. Biol. Chem.*, vol. 288, pp. 31358–31362, Oct. 2013.

[13] M. L. Milstein, T. D. Houle, and S. E. Cala, “Calsequestrin isoforms localize to different ER subcompartments: evidence for polymer and heteropolymer-dependent localization,” *Exp. Cell Res.*, vol. 315, pp. 523–534, Feb. 2009.

[14] T. P. McFarland, M. L. Milstein, and S. E. Cala, “Rough endoplasmic reticulum to junctional sarcoplasmic reticulum trafficking of calsequestrin in adult cardiomyocytes,” *J. Mol. Cell. Cardiol.*, vol. 49, pp. 556–564, Oct. 2010.

[15] B. C. Knollmann, “A “rough” journey to the sarcoplasmic reticulum–implications of altered calsequestrin trafficking for cardiac arrhythmia,” *J. Mol. Cell. Cardiol.*, vol. 49, pp. 554–555, Oct. 2010.

[16] E. J. Sanchez, K. M. Lewis, G. R. Munske, M. S. Nissen, and C. Kang, “Glycosylation of skeletal calsequestrin: implications for its function,” *J. Biol. Chem.*, vol. 287, pp. 3042–3050, Jan. 2012.

[17] U. Kirchhefer, D. Wehrmeister, A. V. Postma, G. Pohlentz, M. Mormann, D. Kucerova, F. U. Müller, W. Schmitz, E. Schulze-Bahr, A. A. Wilde, and J. Neumann, “The human CASQ2 mutation K206N is associated with hyperglycosylation and altered cellular calcium handling,” *J. Mol. Cell. Cardiol.*, vol. 49, pp. 95–105, July 2010.

[18] L. Zhang, J. Kelley, G. Schmeisser, Y. M. Kobayashi, and L. R. Jones, “Complex formation between junctin, triadin, calsequestrin, and the ryanodine receptor. proteins of the cardiac

junctional sarcoplasmic reticulum membrane," *J. Biol. Chem.*, vol. 272, pp. 23389–23397, Sept. 1997.

[19] S. Rani, C. S. Park, P. K. Sreenivasaiah, and D. H. Kim, "Characterization of Ca(2+)-Dependent Protein-Protein interactions within the ca(2+) release units of cardiac sarcoplasmic reticulum," *Mol. Cells*, vol. 39, pp. 149–155, Feb. 2016.

[20] A. Handhle, C. E. Ormonde, N. L. Thomas, C. Bralesford, A. J. Williams, F. A. Lai, and S. Zissimopoulos, "Calsequestrin interacts directly with the cardiac ryanodine receptor luminal domain," *J. Cell Sci.*, Sept. 2016.

[21] K. M. Lewis, L. A. Ronish, E. Ríos, and C. Kang, "Characterization of two human skeletal calsequestrin mutants implicated in malignant hyperthermia and vacuolar aggregate myopathy," *J. Biol. Chem.*, vol. 290, pp. 28665–28674, Nov. 2015.

[22] N. C. Bal, A. Sharon, S. C. Gupta, N. Jena, S. Shaikh, S. Gyorke, and M. Periasamy, "The catecholaminergic polymorphic ventricular tachycardia mutation R33Q disrupts the n-terminal structural motif that regulates reversible calsequestrin polymerization," *J. Biol. Chem.*, vol. 285, pp. 17188–17196, May 2010.

[23] N. C. Bal, N. Jena, D. Sopariwala, T. Balaraju, S. Shaikh, C. Bal, A. Sharon, S. Gyorke, and M. Periasamy, "Probing cationic selectivity of cardiac calsequestrin and its CPVT mutants," *Biochem. J*, vol. 435, pp. 391–399, Apr. 2011.

[24] E. Kim, B. Youn, L. Kemper, C. Campbell, H. Milting, M. Varsanyi, and C. Kang, "Characterization of human cardiac calsequestrin and its deleterious mutants," *J. Mol. Biol.*, vol. 373, pp. 1047–1057, Nov. 2007.

[25] E. J. Sanchez, K. M. Lewis, B. R. Danna, and C. Kang, "High-capacity ca2+ binding of human skeletal calsequestrin," *J. Biol. Chem.*, vol. 287, pp. 11592–11601, Mar. 2012.

[26] K. M. Lewis, G. R. Munske, S. S. Byrd, J. Kang, H.-J. Cho, E. Ríos, and C. Kang, "Characterization of Post-Translational modifications to calsequestrins of cardiac and skeletal muscle," *Int. J. Mol. Sci.*, vol. 17, Sept. 2016.

[27] B. Gray, R. D. Bagnall, L. Lam, J. Ingles, C. Turner, E. Haan, A. Davis, P.-C. Yang, C. E. Clancy, R. W. Sy, and C. Semsarian, "A novel heterozygous mutation in cardiac calsequestrin causes autosomal dominant catecholaminergic polymorphic ventricular tachycardia," *Heart Rhythm*, May 2016.

[28] K. H. Krause, M. Milos, Y. Luan-Rilliet, D. P. Lew, and J. A. Cox, "Thermodynamics of cation binding to rabbit skeletal muscle calsequestrin. evidence for distinct ca(2+)- and mg(2+)-binding sites," *J. Biol. Chem.*, vol. 266, pp. 9453–9459, May 1991.

[29] L. Sawyer and M. N. James, "Carboxyl-carboxylate interactions in proteins," *Nature*, vol. 295, pp. 79–80, Jan. 1982.

[30] M. Milos, J. J. Schaer, M. Comte, and J. A. Cox, "Calcium-proton and calcium-magnesium antagonisms in calmodulin: microcalorimetric and potentiometric analyses," *Biochemistry*, vol. 25, pp. 6279–6287, Oct. 1986.

[31] L. Sawyer and M. N. G. James, "Carboxyl–carboxylate interactions in proteins," *Nature*, vol. 295, pp. 79–80, Jan. 1982.

[32] F. Kamp, P. Donoso, and C. Hidalgo, "Changes in luminal ph caused by calcium release in sarcoplasmic reticulum vesicles," *Biophys. J.*, vol. 74, pp. 290–296, Jan. 1998.

[33] L. M. Espinoza-Fonseca, "The Ca2+-ATPase pump facilitates bidirectional proton transport across the sarco/endoplasmic reticulum," *Mol. Biosyst.*, vol. 13, pp. 633–637, Mar. 2017.

[34] C. Hidalgo, P. Donoso, and P. H. Rodriguez, "Protons induce calsequestrin conformational changes," *Biophys. J.*, vol. 71, pp. 2130–2137, Oct. 1996.

[35] G. Winter, "*xia2*: an expert system for macromolecular crystallography data reduction," *Journal of Applied Crystallography*, vol. 43, pp. 186–190, Feb 2010.

[36] W. D. G. P. J. M. B. A. S. G. R. J. G. M. F.-M. L. V. M. M.-C. T. Y. I. D. S. N. K. Winter, G. and G. Evans, "DIALS: implementation and evaluation of a new integration package," *Acta Crystallographica Section D*, vol. 74, pp. 85–97, Feb 2018.

[37] P. Evans, “Scaling and assessment of data quality,” *Acta Crystallographica Section D*, vol. 62, pp. 72–82, Jan 2006.

[38] P. R. Evans and G. N. Murshudov, “How good are my data and what is the resolution?,” *Acta Crystallographica Section D*, vol. 69, pp. 1204–1214, Jul 2013.

[39] M. D. Winn, C. C. Ballard, K. D. Cowtan, E. J. Dodson, P. Emsley, P. R. Evans, R. M. Keegan, E. B. Krissinel, A. G. W. Leslie, A. McCoy, S. J. McNicholas, G. N. Murshudov, N. S. Pannu, E. A. Potterton, H. R. Powell, R. J. Read, A. Vagin, and K. S. Wilson, “Overview of the *CCP*4 suite and current developments,” *Acta Crystallographica Section D*, vol. 67, pp. 235–242, Apr 2011.

[40] A. J. McCoy, R. W. Grosse-Kunstleve, P. D. Adams, M. D. Winn, L. C. Storoni, and R. J. Read, “Phaser crystallographic software,” *J. Appl. Crystallogr.*, vol. 40, pp. 658–674, Aug. 2007.

[41] P. D. Adams, P. V. Afonine, G. Bunkóczi, V. B. Chen, I. W. Davis, N. Echols, J. J. Headd, L.-W. Hung, G. J. Kapral, R. W. Grosse-Kunstleve, A. J. McCoy, N. W. Moriarty, R. Oeffner, R. J. Read, D. C. Richardson, J. S. Richardson, T. C. Terwilliger, and P. H. Zwart, “PHENIX: a comprehensive python-based system for macromolecular structure solution,” *Acta Crystallogr. D Biol. Crystallogr.*, vol. 66, pp. 213–221, Feb. 2010.

[42] P. V. Afonine, R. W. Grosse-Kunstleve, N. Echols, J. J. Headd, N. W. Moriarty, M. Mustyakimov, T. C. Terwilliger, A. Urzhumtsev, P. H. Zwart, and P. D. Adams, “Towards automated crystallographic structure refinement with phenix.refine,” *Acta Crystallogr. D Biol. Crystallogr.*, vol. 68, pp. 352–367, Apr. 2012.

[43] T. Terwilliger, “SOLVE and RESOLVE: automated structure solution, density modification and model building,” 2004.

[44] T. C. Terwilliger, R. W. Grosse-Kunstleve, P. V. Afonine, N. W. Moriarty, P. H. Zwart, L.-W. Hung, R. J. Read, and P. D. Adams, “Iterative model building, structure refinement

and density modification with the PHENIX AutoBuild wizard," *Acta Crystallogr. D Biol. Crystallogr.*, vol. 64, no. 1, pp. 61–69, 2008.

[45] P. H. Zwart, R. W. Grosse-Kunstleve, and P. D. Adams, "Xtriage and fest: automatic assessment of x-ray data and substructure structure factor estimation," *CCP4 Newsl*, vol. 43, pp. 27–35, 2005.

[46] P. Emsley, B. Lohkamp, W. G. Scott, and K. Cowtan, "Features and development of coot," *Acta Crystallogr. D Biol. Crystallogr.*, vol. 66, pp. 486–501, Apr. 2010.

[47] A. A. Lebedev and M. N. Isupov, "Space-group and origin ambiguity in macromolecular structures with pseudo-symmetry and its treatment with the program zanuda," *Acta Crystallogr. D Biol. Crystallogr.*, vol. 70, pp. 2430–2443, Sept. 2014.

[48] Schrödinger, LLC, "The PyMOL molecular graphics system, version 1.8." November 2015.

[49] B. K. Ho and F. Gruswitz, "HOLLOW: generating accurate representations of channel and interior surfaces in molecular structures," *BMC Struct. Biol.*, vol. 8, p. 49, Nov. 2008.

[50] E. Beitz, "TeXshade: shading and labeling of multiple sequence alignments using LaTeX2e," *Bioinformatics*, vol. 16, pp. 135–139, Feb. 2000.

[51] J. D. Hunter, "Matplotlib: A 2d graphics environment," *Computing In Science & Engineering*, vol. 9, no. 3, pp. 90–95, 2007.

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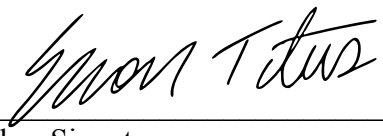

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