UC Davis

UC Davis Previously Published Works

Title

Xuezhikang, Extract of Red Yeast Rice, Improved Abnormal Hemorheology, Suppressed Caveolin-1 and Increased eNOS Expression in Atherosclerotic Rats

Permalink

https://escholarship.org/uc/item/3nr30482

Journal

PLOS ONE, 8(5)

ISSN

1932-6203

Authors

Zhu, Xin-Yuan Li, Pei Yang, Ya-Bing et al.

Publication Date

2013

DOI

10.1371/journal.pone.0062731

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at https://creativecommons.org/licenses/by/4.0/

Peer reviewed



Iron: The Forgotten Driver of Nitrous Oxide Production in Agricultural Soil

Xia Zhu^{1,2,3}, Lucas C. R. Silva³, Timothy A. Doane³*, William R. Horwath³

1 Chengdu Institute of Biology, Chinese Academy of Sciences, Chengdu, China, 2 University of Chinese Academy of Sciences, Beijing, China, 3 Department of Land, Air, and Water Resources, University of California Davis, Davis, California, United States of America

Abstract

In response to rising interest over the years, many experiments and several models have been devised to understand emission of nitrous oxide (N_2O) from agricultural soils. Notably absent from almost all of this discussion is iron, even though its role in both chemical and biochemical reactions that generate N_2O was recognized well before research on N_2O emission began to accelerate. We revisited iron by exploring its importance alongside other soil properties commonly believed to control N_2O production in agricultural systems. A set of soils from California's main agricultural regions was used to observe N_2O emission under conditions representative of typical field scenarios. Results of multivariate analysis showed that in five of the twelve different conditions studied, iron ranked higher than any other intrinsic soil property in explaining observed emissions across soils. Upcoming studies stand to gain valuable information by considering iron among the drivers of N_2O emission, expanding the current framework to include coupling between biotic and abiotic reactions.

Citation: Zhu X, Silva LCR, Doane TA, Horwath WR (2013) Iron: The Forgotten Driver of Nitrous Oxide Production in Agricultural Soil. PLoS ONE 8(3): e60146. doi:10.1371/journal.pone.0060146

Editor: Ben Bond-Lamberty, DOE Pacific Northwest National Laboratory, United States of America

Received January 18, 2013; Accepted February 23, 2013; Published March 29, 2013

Copyright: © 2013 Zhu et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: Support was provided by the California Department of Resources Recycling and Recovery (www.calrecycle.ca.gov), Agreement IWM09027; the J. G. Boswell Endowed Chair in Soil Science; and the University of California Agricultural Experiment Station (http://caes.ucdavis.edu/research/agexpstn). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

1

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: tadoane@ucdavis.edu

Introduction

Emission of N2O from soils is an extensively studied environmental process, given that N₂O is "at the heart of debates" [1] on several prevalent current issues. Approximately two-thirds of total global emission comes from soils; most of the emission from soils is in turn attributed to agriculture [2]. The intrinsic soil properties (as opposed to temporary changes) most commonly mentioned in research studies and models as controlling emission of N2O are texture, pH, organic matter, and ability to supply inorganic nitrogen [3-12]. Production of N₂O in soil is generally attributed to microbiological processes [1,2,13-17], and therefore the factors that regulate the activity of N2O-producing microorganisms should be the same factors that regulate N₂O production. These controlling factors are generally thought to be well recognized, but as research and related commentary on N2O emission from agricultural soils continue to accumulate, the possible role of iron is rarely considered. This is in spite of its known involvement in enzymatic reactions [2,18,19] and non-enzymatic reactions [20-23] that generate N₂O. The connection between iron and N₂O may have been neglected because iron has never figured prominently in routine evaluations of soil for agronomic research or practical management decisions. Unlike the other soil properties cited above, iron does not have a direct and immediate bearing on the growth of most crops or on the agricultural suitability of a soil from either a physical or a chemical point of view. When it is considered, this is in instances of suspected plant deficiency or toxicity, not in the context of its potential connection with the nitrogen cycle. In addition, compared to other intrinsic properties, soil iron does not dramatically affect the short-term changes in microbiological activity generally associated with N_2O production. For these reasons, once interest in N_2O began to intensify, the previously reported connection with iron was already out of sight. The intent of our work was to reconsider the potential significance of iron in emission of N_2O from agricultural soils.

Materials and Methods

Ethics statement

The soils used in this study were collected under consent of the land owners, and the compost used was collected under consent of the compost facility management.

Soil characterization

Soils were collected from the top 15 cm in 10 agricultural fields throughout California, and were sieved to 2 mm following collection. Soil pH was measured in 1 M KCl (1:1 w:v). Percent clay, silt, and sand were determined by a modified pipet method [24]. Total carbon and nitrogen were determined on ball-milled samples by combustion-GC (Costech ECS 4010). Just prior to setting up the experiment, inorganic nitrogen (ammonium plus nitrate) was extracted by 0.5 M $\rm K_2SO_4$ and determined colorimetrically [25,26]. Dissolved organic carbon (DOC) was determined in the same extract by UV-persulfate digestion (Teledyne-Tekmar Phoenix 8000).

We chose two commonly used, contrasting indices to characterize soil iron: that extractable by acid hydroxylamine (FeA), an index of reactive iron(III) minerals [27]; and that extractable by pyrophosphate (FeP), representing iron complexed with soil

organic matter [28–30]. FeA was extracted by shaking 0.8 g soil for one hour with 40 ml 0.25 M hydroxylamine hydrochloride in 0.25 M HCl, followed by centrifugation for 30 minutes at $15600 \times$ G. FeP was extracted by shaking 1 g soil with 100 ml 0.1 M tetrasodium pyrophosphate for 16 hours, followed by centrifugation for 30 minutes at $15600 \times$ G; further centrifugation did not result in any difference in measured iron concentration, indicating that all fine iron colloids had been removed, an important consideration when using this extractant [29,30]. The concentration of iron in all extracts was determined colorimetrically [31]; pyrophosphate extracts were neutralized by a small addition of HCl prior to this determination. There was no interference from pyrophosphate in the colorimetric analysis. All analyses of soil properties were performed in duplicate. These properties are reported in Table 1.

Experimental treatments

As stated above, the properties most commonly believed to control emission of N_2O from agricultural soil include texture, pH, organic matter, and the inherent ability of the soil to release inorganic nitrogen. These are intrinsic properties which are not abruptly altered by environmental conditions; in contrast, our treatments were designed to manipulate the most common temporary extrinsic changes that influence N_2O production: water content, fertilization, and organic amendments. Since these can vary across a range of values, we necessarily limited our choice of treatments. Fertilizer and compost (as a model organic amendment) were either withheld or added at a rate typical of agriculture in California, and two water contents were chosen according to the range expected in agricultural soils. Field capacity, the amount

of water a soil can retain against gravity, was chosen as the upper reference point. This is not uncommon, as soil moisture can temporarily exceed field capacity following irrigation or rainfall events [32,33]. In practice we used water holding capacity (WHC) to represent field capacity. As a contrasting treatment, we chose 50% WHC. This is near the permanent wilting point of most soils [34], and it is not likely that soil moisture will fall below this in the field except during unmanaged dry seasons. Although many intermediate values could have been selected as treatments, we chose to use both ends of a typical spectrum of values in order to present a broad yet concise study.

Experimental set-up

Prior to set-up, WHC was determined as follows: a soil sample was placed into a funnel lined with filter paper, which was then placed into a beaker of water such that just the tip of the funnel was always in contact with water; after the sample ceased to take up water, the sample was allowed to drain, and the moisture content measured. To begin the experiment, the equivalent of 50 g dry soil was placed into cups, which were themselves placed into larger jars containing a small amount of water to avoid desiccation. The larger jars were sealed with lids containing a small foam plug to allow gas exchange with the atmosphere. To imitate the timing typical of agricultural operations, 2 g finely ground finished green waste compost (corresponding approximately to a field application of 60 t ha⁻¹ in the top 15 cm) were mixed with the soils and incubated at 40% WHC for seven days. Treatments not receiving compost were similarly incubated. Following this preincubation, each soil received a fertilizer addition according to treatment: none, ammonium sulfate, or

Table 1. Characterization of the soils used in this study.

Location	Classification ^a	FeA ^b	FeP ^c	DOC^d	Inorganic N	Total N	Total C	Sand	Silt	Clay	рН
		mg kg ⁻¹	mg kg ⁻¹	mg kg ⁻¹	mg kg ⁻¹	%	%	%	%	%	
Sacramento Valley											
Davis	Fine, montmorillonitic, thermic Mollic Haploxeralf	1800	170	17	2	0.09	0.85	30	42	24	5.4
Dixon 1	Fine-silty, mixed, nonacid, thermic Typic Xerorthent	2150	290	30	11	0.14	1.60	23	49	28	5.6
Dixon 2	Fine-silty, mixed, nonacid, thermic Typic Xerorthent	1900	210	19	5	0.11	1.18	15	41	44	5.5
Salinas Valley											
Castroville	Fine, montmorillonitic, thermic Ultic Palexerol	710	550	88	32	0.08	0.75	72	15	13	6.4
Salinas 1	Fine, montmorillonitic, thermic Pachic Argixeroll	390	150	44	5	0.07	0.66	64	23	13	7.2
Salinas 2	Fine, montmorillonitic, thermic Typic Pelloxerert	1890	240	88	28	0.16	1.78	22	36	42	7.4
Spence	Fine-loamy, mixed, thermic, Typic Argixeroll	670	270	63	18	0.11	1.28	50	29	21	6.6
San Joaquin Valley											
Five Points	Fine-loamy, mixed, superactive, thermic Typic Haplocambid	850	60	57	4	0.08	0.67	36	32	32	6.8
Modesto	Fine-loamy, mixed, superactive, thermic Typic Argixeroll	410	240	164	130	0.11	0.97	72	18	10	6.9
Sanger	Coarse-loamy, mixed, nonacid, thermic Typic Xerorthent	390	260	28	4	0.03	0.30	61	32	7	4.2

^aUnited States Department of Agriculture official soil series description, ^b acid hydroxylamine-extractable iron, ^c pyrophosphate-extractable iron, ^d dissolved organic carbon. doi:10.1371/journal.pone.0060146.t001

potassium nitrate. The amount of nitrogen added was 100 mg ${\rm kg}^{-1}$ soil, corresponding approximately to a field rate of 150 kg ${\rm kg}^{-1}$. Fertilizer solution was sprayed onto the soils to reach a water content of 50% or 100% WHC, depending on the treatment. For each soil there were three replicates per treatment. Samples were incubated for 14 days at 22 degrees C.

Samples for N_2O analysis were taken on days 0, 1, 2, 3, 5, 9, and 14 following addition of fertilizer. The jars containing the soil cups were closed with lids containing septa and allowed to stand for one hour. Gas samples were taken at 0, 30, and 60 minutes after closure and transferred to evacuated gas sampling vials. N_2O concentration was determined by gas chromatography-ECD detection (Shimadzu GC-2014). At each sampling date, the rate of N_2O emission (flux) was determined by linear interpolation of the 0, 30, and 60 minute measurements. Cumulative N_2O emission over the course of the incubation was calculated using these data, taking the flux measured at a given date to be the average flux for the interval represented by that date.

Statistical analysis

To identify the soil properties that most strongly explained N₂O emission in each experimental treatment, we studied the data using partial least squares (PLS) multivariate analysis, a form of structural equation modeling. This tool is particularly suitable when the number of predicting variables is greater than the number of observed variables, when multicolinearity is expected among predicting variables, and when multivariate normality can not be assumed [35–37]. PLS ranks the predicting variables by importance based on linear regression models that project the predicting variables and the observed variables to a new, multivariate space. Prior to subjecting the data to PLS analysis, predicting variables (soil properties) and the observed response (cumulative N2O emission) were standardized by centering and scaling the data to have a mean of zero and a standard deviation of one. This ensures that the predicting variables are ranked based on how much of the variation is explained when all variables have the same weight.

Although correlations among variables are possible, especially in studies that involve soil properties, this does not change the interpretation given by PLS, which depicts the relative importance of each variable separately, independently of intrinsic links between variables. Nevertheless, a correlation matrix is presented (Table 2) as an aid in understanding the relationships between the soil properties used in our study.

Following the exploratory PLS analysis, linear regressions between iron and N_2O emission were calculated using unweighted, untransformed data, and were considered significant enough to report at P<0.1. All statistical analyses were performed using JMP 10 software.

Results and Discussion

The results of the PLS analysis are shown in Figure 1, where each soil property is ranked according to its ability to explain cumulative N_2O emission across all soils. This ranking was performed for each of the 12 different treatments studied. In five of these treatments, iron (as either FeA or FeP) ranked higher than any other measured soil characteristic in explaining observed emissions. In four additional treatments, iron was among the top four predictors.

As a complementary approach to further investigate the relationship between iron and N₂O emission, simple linear regressions were calculated in which N₂O data were compared against FeP and FeA. Whereas PLS was used to arrange a suite of soil properties according to their ability to explain N₂O emission, regressions indicate, by the value of r², how much of the variability in N₂O emission can be explained by a single property; regressions also indicate the direction of the effect (positive or negative slope) and degree of importance of the effect (absolute value of the slope). In most cases, a significant relationship between N₂O emission and a given variable can be expected when that variable is ranked highly by PLS. In certain cases, however, a variable ranked highly by PLS may not necessarily yield a significant linear relationship when that variable is considered apart from the other variables; conversely, certain treatments in which a variable is not ranked highly by PLS may nonetheless yield a significant regression. The primary reason for this occasional discrepancy is the nature of the PLS procedure: by considering all predicting variables together, new predictors are generated which are composites of the original variables. Table 3 reports the results of the regressions for treatments that showed a significant relationship between N2O emission and either iron index. Despite a dataset of values for N₂O emission which spanned more than three orders of magnitude across soils, several notable connections between iron and N₂O emission emerged.

FeP was significantly related to N_2O emission in four treatments, in which it explained between 16 and 62 percent of the variability, with a positive slope in all cases (i.e. greater

Table 2. Correlation matrix of the soil properties evaluated in this study.

	FeA ^a	FeP ^b	DOC	Inorganic N	Total N	Total C	Sand	Silt	Clay	рН
FeA	-	-0.07	-0.41	-0.37	0.68	0.70	-0.91	0.84	0.79	-0.15
FeP	-0.07	-	0.25	0.05	0.04	0.08	0.38	-0.37	-0.31	-0.10
DOC	-0.41	0.25	-	0.93	0.25	0.12	0.53	-0.68	-0.29	0.59
Inorganic N	-0.37	0.05	0.93	_	0.18	0.02	0.45	-0.54	-0.28	0.43
Total N	0.68	0.04	0.25	0.18	-	0.98	-0.57	0.37	0.66	0.47
Total C	0.70	0.08	0.12	0.02	0.98	-	-0.61	0.46	0.66	0.36
Sand	-0.91	0.38	0.53	0.45	-0.57	-0.61	-	-0.89	-0.91	0.13
Silt	0.84	-0.37	-0.68	-0.54	0.37	0.46	-0.89	-	0.63	-0.44
Clay	0.79	-0.31	-0.29	-0.28	0.66	0.66	-0.91	0.63	-	0.17
pH	-0.15	-0.10	0.59	0.43	0.47	0.36	0.13	-0.44	0.17	-

^aacid hydroxylamine-extractable iron, ^b pyrophosphate-extractable iron, ^c dissolved organic carbon. doi:10.1371/journal.pone.0060146.t002



Figure 1. Relative importance of soil properties in explaining cumulative emission of N_2O under different conditions. Result of partial least squares multivariate analysis performed across ten soils for each of 12 different treatments. Two indices of soil iron (FeA: acid hydroxylamine-extractable iron and FeP: pyrophosphate-extractable iron) were ranked alongside other soil properties commonly considered to control soil N_2O emission. The size of each bar is given by the variable importance in the projection (VIP) value, and indicates the relative strength of each variable in explaining emission in that treatment. WHC= water holding capacity; DOC= dissolved organic carbon. doi:10.1371/journal.pone.0060146.g001

emission was associated with more FeP). This influence was greatest under 100% WHC when ammonium was present and compost was absent. Such a condition may be reasonably expected on occasion, since most fertilizers supply ammonium, and since this may occur close in time to irrigation or rainfall. In this treatment, an increase in FeP of 1 mg kg⁻¹ corresponded to an increase in cumulative emission of 11.9 ng N₂O-N g⁻¹ soil (averaged across all soils) during the course of the incubation (Table 3).

Like FeP, the connection between FeA and N_2O emission was also significant under several different conditions. Unlike FeP, however, which was positively related to N_2O emission, FeA was always negatively related to N_2O emission. There was no treatment in which both iron indices were significantly related to N_2O emission (Table 3). Considering that FeP and FeA bear almost no relationship to each other (Table 2), this difference in behavior suggests that these two indices indeed reflect two forms of iron that differ in reactivity. Also notable in Table 3 is the effect of

Table 3. Results of simple linear regression of cumulative N_2O emission (as ng N_2O -N g^{-1} soil) against iron, across ten soils and under 12 different conditions.

	50% WHC	50% WHC + compost	100% WHC	100% WHC + compost
No fertilizer	NS	FeP: 0.37, 0.38	FeA: 0.12, -0.09	NS
Ammonium	NS	FeA: 0.28, -0.20	FeP: 0.62, 11.9	FeA: 0.23, -0.62
Nitrate	FeP: 0.19, 0.46	NS	FeP: 0.16, 2.1	NS

The first value given is that of r^2 , and the second value is the slope of the regression. NS = regression was not significant for either iron index. WHC = water holding capacity; FeA = acid hydroxylamine-extractable iron; FeP = pyrophosphate-extractable iron. doi:10.1371/journal.pone.0060146.t003

compost in fertilized treatments: the observed negative association between N_2O emission and FeA occurred only in the presence of compost, while the stimulating effect of FeP was observed only without compost.

The contrasting relationships of FeA and FeP with N2O emission could be due to differences in the reaction of either form of iron with nitrogen compounds in the soil matrix. For example, hydroxylamine is produced from biological oxidation of ammonia, and is known to generate N₂O upon chemical reaction with iron(III) [20,38]. Reaction with FeP versus FeA, or locally high concentrations of either hydroxylamine or iron, could lead to more or less N₂O compared to other reaction products [38]. The ability of aerobic microorganisms to acquire iron can likewise depend on its chemical nature, consequently influencing the amount of reactive compounds produced or consumed through reactions that use iron-dependent enzymes. As soil water content increases, reducing conditions may develop, especially when the depletion of oxygen is accelerated by easily metabolized organic matter. The chemical nature of existing iron(III) may determine the ease with which it is reduced to iron(II) in anaerobic microsites. This will in turn control its participation in other reactions that produce N₂O, such as chemodenitrification, which includes the abiotic reduction of nitrite to N₂O by iron(II) [39,40]. Chemodenitrification can also produce other gases, and the relative amount of N2O released may be affected by the form of iron present. A related anaerobic process is nitrate-dependent iron(II) oxidation [41]; a recent review [42] has highlighted, in the context of this process, how the simultaneous presence of nitrate-reducing and iron(III)-reducing areas can potentially be important to nitrogen cycling. Under anaerobic conditions, iron(III) can also be linked to ammonium oxidation [43,44]. If reactions that generate N₂O are active in any of the above processes, they may be stimulated or suppressed by different forms of iron, such as the two indices examined in this study. The degree of this influence under different conditions will then determine the importance of iron relative to other soil properties.

Our treatments consisted of two contrasting values for soil moisture and addition of amendments. This was done in order to explore the importance of iron across a wide range of conditions while at the same time avoiding a cumbersome dataset. It is clear from Figure 1 that the importance of iron can change between the two limits of each treatment variable. For example, between 50 and 100% WHC under ammonium fertilization, iron moves from a position of modest relevance to become the highest-ranked driver. Since our results show the importance of iron only at two distinct values, we do not know how its importance under intermediate conditions changes between the two end values. Even without such intermediate data, the differences between contrasting treatments can aid in understanding the mechanisms at work in generating N₂O. In the above example, the importance of iron rises markedly under ammonium fertilization as soil moisture increases from 50 to 100% WHC; FeP surpasses FeA in strength as well. As mentioned earlier, ammonia is oxidized to hydroxylamine, and this can react with iron(III) to produce N₂O. In a wetter soil, solutes are more mobile, which can lead to greater production of hydroxylamine as well as greater contact of

References

- Reay DS, Davidson EA, Smith KA, Smith P, Melillo JM, et al. (2012) Global agricultural and nitrous oxide emissions. Nat Clim Change 2: 410-416.
- Thomson AJ, Giannopoulos G, Pretty J, Baggs EM, Richardson DJ (2012) Biological sources and sinks of nitrous oxide and strategies to mitigate emissions. Phil Trans R Soc B 367: 1157-1168.
- Sahrawat KL, Keeney DR (1986) Nitrous oxide emissions from soils. Advances in Soil Sci 4: 103-148.

hydroxylamine with iron. FeP is also likely to be more soluble than FeA. Any combination of these effects might elevate the importance of iron and change which form is more relevant in explaining the associated $N_2{\rm O}$ data.

The overall position of iron among other drivers of N₂O emission is determined by both its reactivity and the presence of processes subject to its influence. Ample opportunity for inquiry exists for defining the extent of the relationship between iron and N₂O in managed as well as unmanaged ecosystems, and this can provide useful practical and theoretical information. For example, including iron in current models of N₂O emission may strengthen their predictive ability. In addition, inasmuch as certain indices of iron can be related to its physical or chemical characteristics, observing the relationship between a given index and N₂O production, and how this changes under different conditions, may provide insight into the specific reactions at work. As stated earlier, production of N₂O is generally accepted to be a microbial affair, and it is logical to assume that the factors that regulate the activity of N2O-producing microorganisms should be the same factors that regulate N2O production. This is not incorrect, but is perhaps a somewhat restrictive rendering; a more accurate framework might include "biotic-abiotic reaction sequences" [39] that generate N₂O, such as those outlined above. Indeed, "the complex interactions that occur between microorganisms and other biotic and abiotic factors" have been suggested to be a key part of further understanding greenhouse gas production and improving predictions [17].

Conclusion

It has been recently emphasized [45] that solutions to environmental problems require explicit consideration of the couplings between element cycles. The environmental chemistry of iron has been well researched, as have many of the interrelated details of the nitrogen cycle. The specific connection between iron and N_2O in soil has also been recognized in both older and recent studies. However, iron and nitrogen have yet to be brought together in agricultural systems, the foremost source of soil N_2O emission. Our most important conclusion is simple: iron does indeed figure prominently among the soil properties controlling N_2O emission in contrasting conditions across diverse soils. Studies concerned with the potential of agricultural soil to emit N_2O will gain new momentum by remembering this "key biogeochemical engine" [46], building on a connection identified a long time ago but largely overlooked since then.

Acknowledgments

We gratefully acknowledge the assistance of those who collected the soil samples, as well as the constructive comments received during review, which were very helpful in improving our paper.

Author Contributions

Conceived and designed the experiments: XZ WRH. Performed the experiments: XZ. Analyzed the data: XZ LCRS. Wrote the paper: TAD.

- Eichner M (1990) Nitrous oxide emissions from fertilized soils: summary of available data. J Environ Qual 19: 272-280.
- Bouwman AF, Fung I, Matthews E, John J (1993) Global analysis of the potential for N₂O production in natural soils. Global Biogeochem Cycles 7: 557-597.
- Robertson K (1994) Nitrous oxide emission in relation to soil factors at low to intermediate moisture levels. J Environ Qual 23: 805-809.

- Dobbie KE, McTaggart IP, Smith KA (1999) Nitrous oxide emissions from intensive agricultural systems: variations between crops and seasons, key driving variables, and mean emission factors. J Geophys Res 104: 26891-26899.
- Li C, Aber J, Stange F, Butterbach-Bahl H (2000) A process-oriented model of N₂O and NO emissions from forest soils: 1. Model development. J Geophys Res 105: 4369-4384.
- Skiba U, Smith KA (2000) The control of nitrous oxide emissions from agricultural and natural soils. Chemosphere Global Change Sci 2: 379-386.
- 10. Bouwman AF, Boumans LJM, Batjes NH (2002) Modeling global annual $\rm N_2O$ and NO emissions from fertilized fields. Global Biogeochem Cycles 16: 1080.
- Freibauer A, Kaltschmitt M (2003) Controls and models for estimating direct nitrous oxide emissions from temperate and sub-boreal agricultural mineral soils in Europe. Biogeochemistry 63: 93-115.
- Stehfest E, Bouwman L (2006) N₂O and NO emission from agricultural fields and soils under natural vegetation: summarizing available measurement data and modeling of global annual emissions. Nutr Cycl Agroecosys 74: 207-228.
- 13. Williams EJ, Hutchinson GL, Fehsenfeld FC (1992) NO_x and N_2O emissions from soil. Global Biogeochem Cycles 6: 351-388.
- Bremner JM (1997) Sources of nitrous oxide in soils. Nutr Cycl Agroecosys 49: 7-16.
- Freney JR (1997) Emission of nitrous oxide from soils used for agriculture. Nutr Cycl Agroecosys 49: 1-6.
- Davidson EA, Keller M, Erickson HE, Verchot LV, Veldkamp E (2000) Testing a conceptual model of soil emissions of nitrous and nitric oxides. BioScience 50: 667-680.
- Singh BK, Bardgett RD, Smith P, Reay DS (2010) Microorganisms and climate change: terrestrial feedbacks and mitigation options. Nat Rev Microbiol 8: 779-790.
- 18. Meiklejohn J (1953) Iron and the nitrifying bacteria. J Gen Microbiol 8: 58-65.
- Glass JB, Orphan VJ (2012) Trace metal requirements for microbial enzymes involved in the production and consumption of methane and nitrous oxide. Front Microbiol doi: 10.3389/fmicb.2012.00061.
- Chao TT, Kroontje W (1966) Inorganic nitrogen transformations through the oxidation and reduction of iron. Soil Sci Soc Am Proc 30: 193-196.
- Buresh RJ, Moraghan JT (1976) Chemical reduction of nitrate by ferrous iron. J Environ Qual 5: 320-325.
- Chalamet A, Bardin R (1976) Action of ferrous ions on reduction of nitrous acid in hydromorphous soils. Soil Biol Biochem 9: 281-285.
- Bremner JM, Blackmer AM, Waring SA (1980) Formation of nitrous oxide and dinitrogen by chemical decomposition of hydroxylamine in soils. Soil Biol Biochem 12: 263-269.
- Burt R, editor (1992) Soil Survey Laboratory Methods Manual. Soil Survey Investigations Report no. 42.WashingtonDC:USDA.
- Verdouw H, van Echteld CJA, Dekkers EMJ (1978) Ammonia determination based on indophenol formation with sodium salicylate. Water Res 12: 399-402.
- Doane TA, Horwath WR (2003) Spectrophotometric determination of nitrate with a single reagent. Anal Lett 36: 2713-2722.
- Lovley DR, Phillips EJP (1987) Rapid assay for microbially reducible ferric iron in aquatic sediments. Appl Environ Microbiol 53: 1536-1540.

- Bremner JM, Heintze SG, Mann PJG, Lees H (1946) Metallo-organic complexes in soil. Nature 158: 790-791.
- Schuppli PA, Ross GJ, McKeague JA (1983) The effective removal of suspended materials from pyrophosphate extracts of soil from tropical and temperate regions. Soil Sci Soc Am J 47: 1026-1032.
- Loveland PJ, Digby P (1984) The extraction of Fe and Al by 0.1 M pyrophosphate solutions: a comparison of some techniques. J Soil Sci 35: 243-250.
- Dominik P, Kaupenjohann M (2000) Simple spectrophotometric determination of Fe in oxalate and HCl soil extracts. Talanta 51: 701-707.
- Cassel DK, Nielsen DR (1986) Field capacity and available water capacity. In: Klute A, editor. Methods of Soil Analysis.Part 1.Physical and Mineralogical Methods.Madison,WI:Soil Science Society of America. pp. 901-926.
- Veihmeyer FJ, Hendrickson AH (1931) The moisture equivalent as a measure of the field capacity of soils. Soil Sci 32: 181-193.
- Hendrickson AH, Veihmeyer FJ (1945). Permanent wilting percentages of soils obtained from field and laboratory trials. Plant Physiol 20: 517-539.
- Tenenhaus M, Vinzi VE, Chatelin YM, Lauro C (2005) PLS path modeling. Comput Statd Data Anal 48: 159-205.
- Marcoulides G, Chin WW, Saunders C (2009) A critical look at partial least squares modeling. MIS Quart 33: 171-175.
- Vinzi V, Chim WW, Henseler J, Wang H, editors (2010) Handbook of Partial Least Squares: Concepts, Methods, and Applications. Heidelberg: Springer.
- Bengtsson G, Fronaeus S, Bengtsson-Kloo L (2002) The kinetics and mechanism of oxidation of hydroxylamine by iron(III). J Chem Soc Dalton Trans 12: 2548-2552.
- Burger M, Venterea RT (2011) Effects of nitrogen fertilizer types on nitrous oxide emissions. In: Guo L, Gunasekara AS, McConnel LL, editors. Understanding Greenhouse Gas Emissions from Agricultural Management.Wa-Washington DC:American Chemical Society. pp. 179-202.
- Hanse HCB, Borggaard OK, Sorensen J (1994) Evaluation of the free energy of formation of Fe(II)-Fe(III) hydroxide-sulphate (green rust) and its reduction of nitrite. Geochim Cosmochim Acta 58: 2599-2608.
- Straub KL, Benz M, Schink B, Widdel F (1996) Anaerobic, nitrate-dependent microbial oxidation of ferrous iron. Appl Environ Microbiol 62: 1458-1470.
- Weber KA, Achenbach LA, Coates JD (2006) Microorganisms pumping iron: anaerobic microbial iron oxidation and reduction. Nat Rev Microbiol 4: 752-764.
- Clement JC, Shrestha J, Ehrenfeld JG, Jaffe PR (2005) Ammonium oxidation coupled to dissimilatory reduction of iron under anaerobic conditions in wetland soils. Soil Biol Biochem 37: 2323-2328.
- Shrestha J, Rich JJ, Ehrenfeld JG, Jaffe PR (2009) Oxidation of ammonium to nitrite under iron-reducing conditions in wetland soils. Soil Sci 174: 156-164.
- Finzi AC, Cole JJ, Doney SC, Holland EA, Jackson RB (2011) Research frontiers in the analysis of coupled biogeochemical cycles. Front Ecol Environ 9: 74-80.
- Silver WL, Hall SJ, Liptzin D, Yang WH (2011) The iron redox engine drives carbon, nitrogen, and phosphorus cycling in terrestrial ecosystems. American Geophysical Union Abstracts, http://adsabs.harvard.edu/abs/2011AGUFM. B11F.01S, accessed Oct.28, 2012.