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Inter-Rater Agreement of Anal Cytology

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Abstract

Most anal cancers are caused by persistent infections with carcinogenic human papillomaviruses (HPV). Similar to cervical carcinogenesis, the progression from HPV infection to anal cancer goes through precancerous lesions that can be treated to prevent invasion. In analogy to cervical cytology, anal cytology has been proposed as a screening tool for anal cancer precursors in high-risk populations. We analyzed the inter-observer reproducibility of anal cytology in a population of 363 HIV-infected men who have sex with men (MSM). Liquid-based cytology (LBC) specimens were collected in the anal dysplasia clinic before performing high-resolution anoscopy (HRA) on all subjects. Papanicolaou-stained, LBC slides were evaluated by two cytopathologists, blinded to clinical outcome and the other pathologist's results, using the revised Bethesda terminology. Overall agreement between two observers was 66% (kappa 0.54, linear weighted kappa 0.69). Using dichotomizing cytology results (ASC-US or worse vs. less than ASC-US), the agreement increased to 86% (kappa 0.69). We observed an increasing likelihood of testing positive for markers associated with HPV-related transformation, p16/Ki-67 and HPV oncogene mRNA, with increasing severity of cytology results both for individual cytologists and for consensus cytology interpretation ($p_{\text{trend}} < 0.0001$ for all). In summary, we observed moderate to good agreement between two cytopathologists evaluating anal cytology samples collected in HIV-positive MSM. A higher severity of anal cytology was associated with biomarkers of anal precancers. Anal cytology may be used for anal cancer screening in high-risk populations, and biomarkers of HPV-related transformation can serve for quality control of anal cytology.

Keywords

anal cancer screening; cytology; human papillomavirus (HPV); human immunodeficiency virus (HIV); men who have sex with men (MSM); biomarkers

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Introduction

Anal cancer is uncommon in the general population, with incidence rates of about 2/100,000 in the United States (1). In certain high-risk populations, such as men who have sex with men (MSM), and human immunodeficiency virus (HIV)-positive men and women, the risks for anal cancer can be much higher and may approach the cervical cancer risk in unscreened populations of women. In MSM, anal cancer rates are estimated to be 40/100,000 (2-4), and in HIV-positive MSM the risk of anal cancer may be 2-4-fold higher (2;3;5;6) or more than in HIV-negative MSM. A recent analysis of 13 cohorts found that HIV-positive MSM were at the highest risk of anal cancer, followed by HIV-positive men or women, and all were at much higher risk than HIV-uninfected populations (7).

Analogous to cervical cytology, anal cytology has been recommended as a method for screening for the prevention of anal cancer through detection of precancerous lesions, anal intraepithelial neoplasia grade 3 (AIN3) and grade 2 (AIN2), and treatment. Surprisingly, there are limited data on the inter-observer or inter-rater agreement of anal cytology, unlike for cervical cytology. A previous study of 120 cytology slides from HIV-infected men reported a weighted Kappa for agreement between 4 pathologists evaluating the slides independently was 0.54 (8).

Since anal cancer is caused by the same causal factor as cervical cancer, persistent infection by high-risk human papillomavirus (HR-HPV), HPV measurements and related biomarkers might be used as objective measures potentially for quality control of anal cytology, just as HR-HPV is used for cervical cytology, specifically for ASC-US (9-11). Examples of other potentially useful biomarkers include detection of HPV E6/E7 oncogene mRNA, p16^{INK4a}, and HPV16 (12), the most carcinogenic HPV genotype. Comparisons of anal cytology and histology results from laboratory data might also provide benchmarks for anal cytology (8;13;14).

To examine the issue of inter-rater agreement of anal cytology and the relationship of biomarkers with anal cytologic interpretations, we conducted an analysis in a population of HIV-positive MSM enrolled at an anal cancer-screening clinic in the Kaiser Permanente Northern California HMO.

Methods

Study population

The study was based at the San Francisco Kaiser Permanente Northern California (KPNC) Anal Cancer Screening Clinic. We enrolled men who were identified as HIV-positive through the Kaiser HIV registry, who were 18 years or older, who were not diagnosed with anal cancer prior to enrollment and who provided informed consent. In total, 363 men were enrolled between August 2009 and June 2010. The study was reviewed and approved by the institutional review boards at KPNC and at NCI. All participants were asked to complete a self-administered questionnaire to collect risk factor information. Additional information on HIV status and medication, sexually transmitted diseases, and histopathology results were abstracted from the KPNC clinical database.

For 87 subjects of 271 without biopsy-proven AIN2 or AIN3 at the enrollment visit, follow-up information on outcomes from additional clinic visits up to 12/2011 was available and included in the analysis to correct for possible imperfect sensitivity of high-resolution anoscopy (13;15).

Clinical exam, evaluation, and results

During the clinical examination, two specimens were collected by inserting a wetted flocked nylon swab (16) into the anal canal up to the distal rectal vault and withdrawing with rotation and lateral pressure. Both specimens were transferred to PreservCyt medium (Hologic, Bedford, MA). A third specimen was collected for routine Chlamydia Trachomatis and Neisseria Gonorrhoea testing. After specimen collection, participants received a digital anorectal exam followed by high-resolution anoscopy (HRA). All suspicious-appearing lesions in HRA were biopsied and sent for routine histopathological review by KPNC pathologists, which was graded as condyloma, and AIN 1-3. No cancers were observed in this study population.

From the first specimen, a ThinPrep slide (Hologic) was prepared for routine Pap staining and evaluation. Two pathologists (T.D. and D.T.) reviewed the slides independently. Cytology results were reported analogous to the Bethesda classification (17) for cervical cytology except otherwise noted. The following categories were used: NILM (negative), atypical squamous cells of undetermined significance (ASC-US), atypical squamous cells cannot rule out HSIL (ASC-H), low-grade squamous intraepithelial lesion (LSIL), high-grade squamous intraepithelial lesion (HSIL), favor AIN2 (HSIL-AIN2), and HSIL-AIN3. ASC-H, HSIL-AIN2, and HSIL-AIN3 were combined into a single, high-grade cytology category for this analysis.

Biomarker testing—Using the residual specimen from the first collection, mtm Laboratories (Heidelberg, Germany) performed the p16^{INK4a}/ki-67 dual immunostaining (“p16/ki-67 staining”) using their CINtec Plus cytology kit according to their specifications. A Thinprep 2000 processor (Hologic) was used to prepare a slide, which then stained for according to the manufacturer's instructions. The CINtec Plus cytology kit was then applied to the unstained cytology slide for p16/ki-67 staining.

On the second collected specimen, Roche Molecular Systems (Pleasanton, CA) tested for HRHPV, including separate detection of HPV16, and HPV18 DNA, using their cobas 4800 HPV assay. To prepare DNA for cobas, automated sample extraction was performed as follows: 500 μ L of the PreservCyt specimen was pipetted into a secondary tube (Falcon 5-ml polypropylene round-bottom tube, 12- by-75-mm style, non-pyrogenic, sterile). The tube was capped, mixed by vortexing, uncapped, placed on the x-480 specimen rack and loaded onto the x-480 sample extraction module of the cobas 4800 system. The x-480 extraction module then inputs 400 μ L of this material into the specimen preparation process. The extracted DNA was then tested as previously described (16).

Norchip also tested the second specimen for HPV16, 18, 31, 33, and 45 HPV E6/E7 mRNA using their Prelect HPV-Proofer assay according to their specifications. All testing was done masked to the results of the other assays, clinical outcomes, and patient characteristics.

Statistical analysis—For the agreement between the two cytology readers, we calculated the total agreement with a binomial 95% confidence interval (95%CI). We calculated Cohen's kappa with 95%CI as a chance-corrected measure of agreement as described in (18). Since kappa does not account for the degree of disagreement between categories and treats any disagreement equally, we calculated linear weighted kappa with 95%CI for the ordered cytology categories. Thus, disagreement between adjacent categories results in lower reduction of kappa values than disagreement between non-adjacent categories. Kappa values of <0.20 were interpreted as poor, between 0.21-0.40 as fair, between 0.41-0.60 as moderate, between 0.61-0.80 as good and above 0.80 as very good. Exact versions of symmetry (4 category) and McNemar (2 category) chi-square tests were used to test for statistically significant differences in the distribution of the cytologic interpretations

between readers. A non-parametric test of trend was used to assess the trend in the proportion of positives for each biomarker for risk of AIN2 or more severe (AIN2+) with increasing severity of the cytologic interpretation (19). Finally, a Fisher's exact test was used to test for differences in the proportion of positives for each biomarker between sub-groups defined by the paired cytologic interpretations.

Results

The 363 men enrolled in the study had median, mean, and range of age of 53 years, 53 years, and 26-79 years, respectively. Most men were users of highly active anti-retroviral therapy (93%), 89% of men had an HIV viral load <75 copies, and 97% had a CD4 count higher than 200 (82% higher than 350) at the time of enrollment. Of the 363 men who enrolled in the study, 339 (93%) men had cytologic interpretations available from both study cytopathologists and were the basis of this analysis. The 24 men who were not included in the analysis due to missing cytology interpretations had a non-significantly lower percentage of HR-HPV DNA (65% vs. 80%; $p=0.09$).

Table 1 shows the comparison of the cytologic interpretations by the two cytopathologists (readers). The first reader called 33% as negative, 22% as ASC-US, 20% as LSIL, and 26% as high-grade cytology. The second reader called 43% as negative, 10% as ASC-US, 24% as LSIL, and 23% as high-grade cytology. The crude agreement was 66% (95%CI = 61%-71%), kappa was 0.54 (95%CI = 0.47-0.60), and linear weighted was 0.69 (95%CI = 0.63-0.74). Reader 1 was more likely to interpret the cytology more severe ($p < 0.0001$). Re-categorized the cytology as negative or ASC-US or more severe, the crude agreement was 86% (95%CI = 82%-9-%) and the kappa was 0.69 (95%CI = 0.61-0.76). Reader 1 was more likely to interpret the cytology as ASC-US or more severe ($p < 0.0001$).

In **Table 2**, we show the relationships of various biomarkers and risk of having a histologic diagnosis of AIN2+ with the individual and paired cytologic interpretations. There was a significant trend ($p_{\text{trend}} < 0.0001$) of an increasing likelihood of testing positive for any of the biomarkers and or having an AIN2+ diagnosis with increasing severity of the cytologic interpretation for each rater individually. Similarly, there was a significant trend ($p_{\text{trend}} < 0.0001$) of increasingly likelihood of testing positive for any of the biomarkers and or having an AIN2+ diagnosis with increasing severity of consensus cytologic interpretation. Although the numbers for specific pairs of discordant cytologic interpretations were small, making generalization difficult, there was a tendency for these paired results to reflect a mixture of both over-called and under-called cytologic interpretations, as indicated by the intermediate positivity of the biomarker results compared to the consensus paired results i.e., ASC-US/ASC-US<ASCUS/LSIL or LSIL/ASC-US<LSIL/LSIL.

However, we observed a large number of the discordant pair results of ASC-US/Negative (rater 1/rater 2). Comparing the profiles of biomarker positivity and risk of AIN2+ (**Figure 1**), we noted that the profile of the ASC-US/Negative subgroup was more akin to Negative/Negative than to ASC-US/ASC-US. Specifically, the % positive for HR-HPV DNA and p16/ki-67 staining for ASC-US/Negative was significantly lower than for ASC-US/ASC-US ($p = 0.02$ and 0.03 , respectively) but not significantly higher than for Negative/Negative ($p = 0.6$ and $p = 1$, respectively).

Discussion

In our analysis, we found moderate to good agreement between two cytopathologists evaluating anal cytology using samples from HIV-infected MSM. When compared to Lytwyn *et al.* (8), we found a better linear-weighted kappa, 0.69 vs. 0.54 (overall for 4

pathologists), but a worse un-weighted kappa, 0.54 vs. 0.69 (median). Thus, in Lytwyn *et al.* (8) there was better exact agreement but when there was disagreement in the severity of the cytology the discrepancies were more pronounced compared to this analysis. Any differences in inter-rater agreement between studies may be due to differences in the screening and treatment between populations, resulting in differences in the size of the lesions and the number of diagnostically informative cells on a slide. We also used a different collection device, flocked nylon swab (16), than the typical Dacron swab, which may have altered the number of diagnostic cells on a slide. Finally, reader 2, an experienced cytopathologist who had only read cervical cytology prior to the study, received training for anal cytology from reader 1 before the study started, which might have also influenced the agreement between the cytopathologists. Of note, in our study, the histologic confirmation even of consensus HSIL cytology results was limited, due to the limited performance of high resolution anoscopy that is widely recognized (13).

With increasing annual rates of anal cancer in the U.S. (**Figure 2**), it will be important to establish screening programs targeting high-risk populations such as HIV-positive MSM and HIV-infected men and women (7). Although there is no established method of anal cancer screening, cytology has been recommended (2) and its use may be cost effective in high-risk populations (20). We also showed that the detection of several biomarkers and the diagnosis of AIN2+ increased with increasing severity of anal cytology, as been shown for cervical cytology. These biomarkers therefore might be useful as objective standards to help monitor and maintain performance of anal cytology. For example, retrospectively reviewing anal cytology interpreted as HSIL in conjunction with biomarker results may improve individual pathologists diagnostic accuracy and identify false negative and false positive diagnoses.

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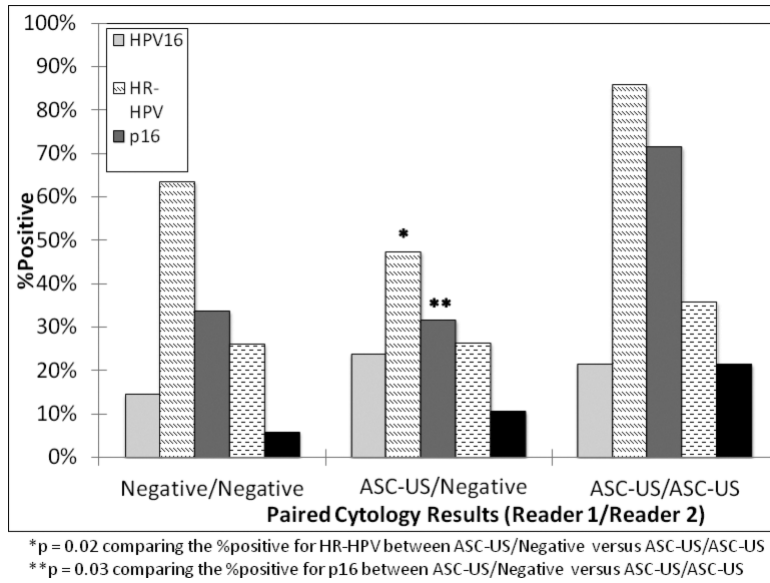


Figure 1.

The relationship of biomarker results for paired cytology results of Negative/Negative, ASC-US/Negative, and ASC-US/ASC-US (Reader 1/Reader 2). For each paired cytology results, the percent positive for HPV16 DNA, high-risk HPV (HR-HPV) DNA, p16^{INK4a} immunocytochemistry, HPV16, 18, 31, 45, and 58 E6/E7 mRNA, or had anal intraepithelial neoplasia grade 2 or more severe diagnosis (AIN2+) is shown. Abbreviations: ASC-US, atypical squamous cells of undetermined significance

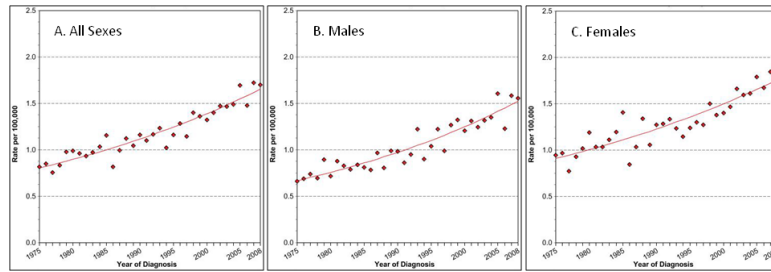


Figure 2. Annual age-adjusted anal cancer incidence rates in the U.S. for both sexes (A), males (B), and females (C). Data are from <http://seer.cancer.gov/>. Incidence source: SEER 9 areas (San Francisco, Connecticut, Detroit, Hawaii, Iowa, New Mexico, Seattle, Utah, and Atlanta). Rates are per 100,000 and are age-adjusted to the 2000 US Std Population (19 age groups - Census P25-1130). The modeled rates are the point estimates for the regression lines calculated by the Joinpoint Regression Program (Version 3.5, April 2011, National Cancer Institute).

Table 1

Inter-rater agreement for cytologic interpretation by two readers. Bold type highlights exact agreement, italic type indicates the cells the contribute the greatest to disagreement.

	Reader 2				Total
	Negative	ASC-US	LSIL	High-Grade*	
Reader 1					
Negative	104	4	3	1	112
ASC-US	38	14	14	7	73
LSIL	2	8	47	10	67
High-Grade*	3	8	17	59	87
Total	147	34	81	77	339

Abbreviations: ASC-US, atypical squamous cells of undetermined significance; LSIL, low-grade squamous intraepithelial lesion

* high-grade cytology includes high-grade squamous intraepithelial lesion (HSIL) and atypical squamous cells cannot rule out HSIL (ASC-H)

Table 2

The relationship of biomarker results and paired cytology results from the two raters. For each paired cytology results, the number and percent positive for HPV16 DNA, high-risk HPV (HR-HPV) DNA, p16^{INK4a}/ki-67 immunocytochemistry (p16), HPV16, 18, 31, 45, and 58E6/E7 mRNA (mRNA), or had anal intraepithelial neoplasia grade 2 or more severe diagnosis (AIN2+) is presented. Bold type with gray background indicates exact agreement for cytologic interpretation.

	Reader 2											
	Negative		ASC-US		LSIL		High-Grade*		Total			
	n	%	n	%	n	%	n	%	n	%		
Negative	%HPV16+	15	14%	0	0%	0	0%	0	0%	15	13%	
	%HR-HPV+	66	63%	4	100%	3	100%	1	100%	74	66%	
	%p16+	35	34%	4	100%	2	67%	0	0%	41	37%	
	%mRNA+	27	26%	3	75%	1	33%	0	0%	31	28%	
	% AIN2	6	6%	1	25%	2	67%	0	0%	9	8%	
ASC-US	%HPV16+	9	24%	3	21%	4	29%	1	14%	17	23%	
	%HR-HPV+	18	47%	12	86%	13	93%	7	100%	50	68%	
	%p16+	12	32%	10	71%	10	71%	7	100%	39	53%	
	%mRNA+	10	26%	5	36%	7	50%	3	43%	25	34%	
	% AIN2	4	11%	3	21%	2	14%	2	29%	11	15%	
Reader 1	%HPV16+	1	50%	3	38%	16	34%	7	70%	27	40%	
	%HR-HPV+	2	100%	7	88%	40	85%	10	100%	59	88%	
	%p16+	1	50%	6	75%	39	83%	9	90%	55	82%	
	%mRNA+	1	50%	5	63%	25	53%	9	90%	40	60%	
	% AIN2	0	0%	0	0%	14	30%	3	30%	17	25%	
High-Grade*	%HPV16+	1	33%	3	38%	6	35%	35	59%	45	52%	
	%HR-HPV+	3	100%	8	100%	16	94%	59	100%	86	99%	
	%p16+	3	100%	6	75%	15	88%	58	98%	82	94%	
	%mRNA+	1	33%	5	63%	10	59%	50	85%	66	76%	
	% AIN2	2	67%	5	63%	3	18%	31	53%	41	47%	
Total	%HPV16+	26	18%	9	26%	26	32%	43	56%	104	31%	
	%HR-HPV+	89	61%	31	91%	72	89%	77	100%	269	79%	

	Reader 2											
	Negative		ASC-US		LSIL		High-Grade*		Total			
	n	%	n	%	n	%	n	%	n	%		
%p16+	51	35%	26	76%	66	81%	74	96%	217	64%		
%mRNA+	39	27%	18	53%	43	53%	62	81%	162	48%		
% AIN2	12	8%	9	26%	21	26%	36	47%	78	23%		

Abbreviations: ASC-US, atypical squamous cells of undetermined significance; LSIL, low-grade squamous intraepithelial lesion

* high-grade cytology includes high-grade squamous intraepithelial lesion (HSIL) and atypical squamous cells cannot rule out HSIL (ASC-H)