UCLA
UCLA Previously Published Works

Title

Whole-Genome Sequencing Association Analyses of Stroke and Its Subtypes in Ancestrally Diverse Populations From Trans-Omics for Precision Medicine Project

Authors

Hu, Yao
Haessler, Jeffrey W
Manansala, Regina
et al.

Whole-genome sequencing association analyses of stroke and its subtypes in ancestrally-diverse populations from TOPMed

Yao Hu, PhD[1], **Jeffrey W. Haessler, MSc**[1], **Regina Manansala, MSc**[2], **Kerri L. Wiggins, MSc**[3], **Arden Moscati, PhD**[4], **Alexa Beiser, PhD**[5,6], **Nancy L. Heard-Costa, PhD**[5], **Chloe Sarnowski, PhD**[6], **Laura M. Raffield, PhD**[7], **Jaeyoon Chung, PhD**[8,9], **Sandro Marini, PhD**[9,10], **Christopher D. Anderson, MD**[9,10,11], **Jonathan Rosand, MD**[9,10,11], **Huichun Xu, PhD**[12], **Xiao Sun, PhD**[13], **Tanika N. Kelly, PhD**[13], **Quenna Wong, MSc**[14], **Leslie A. Lange, PhD**[15], **Jerome I. Rotter, PhD**[16], **Adolfo Correa, PhD**[17], **Ramachandran S. Vasan, MD**[8], **Sudha Seshadri, MD**[5], **Stephen S. Rich, PhD**[18], **Ron Do, PhD**[4,19], **Ruth J.F. Loos, PhD**[4,20], **William T. Longstreth Jr., MD**[21], **Joshua C. Bis, PhD**[3], **Bruce M. Psaty, MD, PhD**[3,22,23], **David L. Tirschwell, MD**[24], **Themistocles L. Assimes, MD, PhD**[25], **Brian Silver, MD**[26], **Simin Liu, MD**[27], **Rebecca Jackson, MD**[28], **Sylvia Smoller, PhD**[29], **Braxton D. Mitchell, PhD**[12,30], **Myriam Fornage, PhD**[31], **Paul L. Auer, PhD**[2], **Alex P. Reiner, MD**[1,32], **Charles Kooperberg, PhD**[1] **Trans-Omics for Precision Medicine (TOPMed) Stroke Working Group, the NHLBI Trans-Omics for Precision Medicine (TOPMed) Consortium**

[1]Public Health Sciences Division, Fred Hutchinson Cancer Research Center, Seattle, WA

[2]School of Public Health, University of Wisconsin–Milwaukee, Milwaukee, WI

[3]Cardiovascular Health Research Unit, Department of Medicine, University of Washington, Seattle, WA

[4]The Charles Bronfman Institute for Personalized Medicine, Icahn School of Medicine at Mount Sinai, New York, NY

[5]Department of Neurology, Boston University School of Medicine, Boston, MA

[6]Department of Biostatistics, Boston University School of Public Health, Boston, MA

[7]Department of Genetics, University of North Carolina, Chapel Hill, NC

[8]Department of Medicine, Boston University School of Medicine, Boston, MA

[9]Center for Genomic Medicine, Massachusetts General Hospital, Boston, MA

[10]Program in Medical and Population Genetics, Broad Institute, Cambridge, MA

[11]Henry and Allison McCance Center for Brain Health, Massachusetts General Hospital, Boston, MA

[12]Department of Medicine, University of Maryland School of Medicine, Baltimore, MD

Corresponding authors: Alex P. Reiner, apreiner@uw.edu, 206.667.2710, 1100 Fairview Ave N, Seattle, WA, 98109, Charles Kooperberg, clk@fredhutch.org, 206.667.7808, 1100 Fairview Ave N, Seattle, WA, 98109.
Supplemental Materials
Expanded Materials & Methods
Online Figures I–II
Online Tables I–XVI

[13]Department of Epidemiology, Tulane University School of Public Health and Tropical Medicine, New Orleans, LA

[14]Department of Biostatistics, University of Washington, Seattle, WA

[15]Department of Medicine, University of Colorado, Denver, CO

[16]The Institute for Translational Genomics and Population Sciences, Department of Pediatrics, The Lundquist Institute for Biomedical Innovation at Harbor-UCLA Medical Center, Torrance, CA

[17]Department of Pediatrics and Medicine, University of Mississippi Medical Center, Jackson, MS

[18]Center for Public Health Genomics, University of Virginia, Charlottesville, VA

[19]Department of Genetics and Genomic Sciences, The Icahn School of Medicine at Mount Sinai, New York, NY

[20]The Mindich Child Health and Development Institute, The Icahn School of Medicine at Mount Sinai, New York, NY

[21]Departments of Neurology and Epidemiology, University of Washington, Seattle, WA

[22]Departments of Epidemiology and Health Services, University of Washington, Seattle, WA

[23]Kaiser Permanente Washington Health Research Institute, Seattle, WA

[24]Department of Neurology, University of Washington, Seattle, WA

[25]Department of Medicine, Stanford University, Stanford, CA

[26]Department of Neurology, University of Massachusetts Medical School, Worcester, MA

[27]Center for Global Cardiometabolic Health, Departments of Epidemiology, Medicine, and Surgery, Brown University, Providence, RI

[28]Division of Endocrinology Diabetes and Metabolism, The Ohio State University, Columbus, OH

[29]Department of Epidemiology and Population Health, Albert Einstein College of Medicine, New York, NY

[30]Geriatrics Research and Education Clinical Center, Baltimore Veterans Administration Medical Center, Baltimore, MD

[31]Institute of Molecular Medicine, University of Texas Health Science Center at Houston, Houston, TX

[32]Department of Epidemiology, University of Washington, Seattle, WA

Abstract

Background and Purpose—Stroke is the leading cause of death and long-term disability worldwide. Previous genome-wide association studies identified 51 loci associated with stroke (mostly ischemic) and its subtypes among predominantly European populations. Using whole-genome sequencing (WGS) in ancestrally-diverse populations from the Trans-Omics for Precision Medicine (TOPMed) Program, we aimed to identify novel variants, especially low-frequency or ancestry-specific variants, associated with all stroke (AS), ischemic stroke (IS) and its subtypes

[large artery (LAS), cardioembolic (CES), and small vessel (SVS)], and hemorrhagic stroke (HS) and its subtypes [intracerebral (ICH) and subarachnoid (SAH)].

Methods—WGS data were available for 6,833 stroke cases and 27,116 controls, including 22,315 European, 7,877 African American, 2,616 Hispanic/Latino, 850 Asian, 54 Native American and 237 other ancestry participants. In TOPMed, we performed single variant association analysis examining 40 million common variants and aggregated association analysis focusing on rare variants. We also combined TOPMed European populations with over 28,000 additional European participants from the UK BioBank (UKBB) genome-wide array data through meta-analysis.

Results—In the single variant association analysis in TOPMed, we identified one novel locus *13q33* for LAS at whole genome-wide significance (P<5.00E-9) and four novel loci at genome-wide significance (P<5.00E-8), all of which need conformation in independent studies. Lead variants in all five loci are low-frequency but are more common in non-European populations. An aggregation of synonymous rare variants within the gene *C6orf26* demonstrated suggestive evidence of association for HS (P<3.11E-6). By meta-analyzing European ancestry samples in TOPMed and UKBB, we replicated several previously reported stroke loci including *PITX2*, *HDAC9*, *ZFHX3*, and *LRCH1*.

Conclusions—We represent the first association analysis for stroke and its subtypes using WGS data from ancestrally-diverse populations. While our findings suggest the potential benefits of combining WGS data with populations of diverse genetic backgrounds to identify possible low-frequency or ancestry-specific variants, they also highlight the need to increase genome coverage and sample sizes.

Introduction

Stroke is the second leading cause of premature mortality and a leading cause of long-term disability worldwide[1, 2]. The pathogenesis of stroke is heterogeneous and multifactorial. Ischemic stroke (IS), which accounts for 87% of all stroke cases, shows an estimated heritability of approximately 38% and substantial variation across its three subtypes [cardioembolic stroke (CES), 33%; large artery stroke (LAS), 40%; and small vessel stroke (SVS), 16%][3, 4]. Hemorrhagic stroke (HS) is less common, with heritability estimated at over 40% for its two subtypes [intracerebral hemorrhage (ICH), 44%; and subarachnoid hemorrhage (SAH), 41%][5, 6].

Previous genome-wide association studies (GWAS) and Exomechip analysis have identified 51 loci associated with stroke types and subtypes, 32 of which were reported at genome-wide significance in the largest trans-ethnic meta-analysis of stroke consisting of more than half a million participants from the MEGASTROKE Consortium[7]. Most of these published studies focused on all stroke (AS), IS, and its subtypes. The majority of these studies identified loci associated with common variants [minor allele frequency (MAF) >1%] and, in aggregate, explain a limited proportion of the phenotypic variation (0.6%−1.8%)[7].

In addition, the participants included in these association studies were predominantly of European ancestry[3, 7–20]. Previous epidemiological studies have demonstrated an excess incidence of stroke cases in African American (AA) and Hispanic ancestry populations compared to European ancestry populations in the United States[21–23]. These observations reinforce the importance of exploring all stroke types and subtypes in ancestrally-diverse populations.

In the current analysis, we performed the first whole genome sequencing (WGS) analysis for multiple stroke subtypes in an ancestrally-diverse population from the Trans-Omics for Precision Medicine (TOPMed) program, aiming to uncover additional novel loci, especially those driven by low-frequency variants and variants more common in non-European populations. We also attempted to refine previously reported loci using our WGS data with more comprehensive characterizations of the genome.

Methods

Study overview and stroke adjudication

TOPMed data are available on dbGAP (ARIC, phs001211; BioMe: phs001644; CHS, phs001368; FHS, phs000974; JHS, phs000964; MESA, phs001416; WHI, phs001237). In the discovery stage, we performed two GWAS analyses, one focused on TOPMed multi-ethnic samples with denser coverage of the genome using WGS data and the other combining TOPMed and UK BioBank (UKBB) European ancestry samples to increase statistical power while focusing on relatively common variants (Fig. 1). In TOPMed, we included 6,833 incident stroke cases (5,616 IS cases and 1,080 HS cases) and 27,116 controls in our association analyses from the freeze6 data. These participants were from six cohort studies and one biobank: the Atherosclerosis Risk in Communities Study (ARIC) [24–26], the Cardiovascular Health Study (CHS) [27, 28], the Framingham Heart Study (FHS) [29–34], the Jackson Heart Study (JHS) [35–40], the Multi-Ethnic Study of Atherosclerosis (MESA) [41, 42], the Women's Health Initiative (WHI) [43], and the BioMe™ Biobank (BioMe) (Supplemental Table I). In addition, 4,474 IS cases, 959 ICH cases, 1,194 SAH cases, and up to 24,000 controls of European ancestry from the UKBB were selected for analysis. These participants in the discovery stage represented six ancestral groups based on self-reported ancestry, namely Europeans (n=22,315), AA (n=7,877), Hispanics (n=2,616), Asians (n=850), Native Americans (n=54), and others (primarily South Asian, mixed heritage, and other racial/ethnic groups, n=237, Table 1). All studies were approved by local Institutional Review Boards and written informed consent was obtained from each participant.

All stroke cases were adjudicated by physicians in each participating study in the six cohort studies. Baseline stroke cases were excluded from the analysis. In BioMe, the identification of stroke cases was based on the 10th revision of the International Statistical Classification of Diseases and Related Health Problems (ICD). IS cases in CHS, MESA, and WHI studies were further divided into CES, LAS, and SVS according to the Trial of Org 10172 in Acute Stroke Treatment (TOAST) criteria[44]. HS cases were further divided into ICH and SAH. Details are provided in the Supplemental Methods.

Whole-genome sequencing in TOPMed

A total of 106,809 samples (freeze6) underwent ~30× WGS using DNA extracted from blood samples at designated sequencing centers. Harmonization, joint calling and quality control (QC) procedures are described on the TOPMed website (https://www.nhlbiwgs.org/topmed-whole-genome-sequencing-methods-freeze-6).

Genotyping and imputation in UKBB

Genotyping of 500,000 UKBB participants was performed using either the Affymetrix UK BiLEVE Axiom array or the Affymetrix UK Biobank Axiom® array. QC procedures were performed at both the variants and the sample level, and detailed information is provided on the UKBB website (http://www.ukbiobank.ac.uk/wp-content/uploads/2014/04/UKBiobank_genotyping_QC_documentation-web-1.pdf). Imputation was performed based on reference panels from the Haplotype Reference Consortium (HRC), UK10K, and the 1000 Genome Phase 3 using MACH (http://csg.sph.umich.edu/abecasis/MACH/index.html). Genetic variants with MAF>0.1% and imputation quality score R^2>0.3 were included in the association analysis.

Single variant association analysis

Approximately 40 million genetic variants with minor allele count (MAC)>20 were included in the single variant association analysis in TOPMed at the discovery stage. We first tested the association of each variant with stroke types and subtypes treated as dichotomous outcomes using a logistic model adjusted for age, sex, ancestry, study, the first 10 principal components (PCs), and accounting for relatedness using a genetic relationship matrix (GRM) by pooling all TOPMed studies together. Due to the extremely unbalanced case/control ratios, the Scalable and Accurate Implementation of Generalized mixed model (SAIGE) software[45] was used to conduct the association analysis. In addition to the ancestry-combined analyses, ancestry-specific analyses were performed in European and AA ancestry populations. All single variant association analyses were performed on the University of Michigan ENCORE server (https://encore.sph.umich.edu). For the UKBB, association analyses of over 16 million genetic variants with IS and the two subtypes of HS (ICH and SAH) were performed after adjustment for age, sex, and the first 10 PCs using PLINK (http://zzz.bwh.harvard.edu/plink/plink2.shtml) and SAIGE (only for IS where we identified one significant locus using PLINK). We combined summary statistics from TOPMed and UKBB in European ancestry populations using fixed-effect inverse-variance-weighted meta-analysis implemented in METAL (https://genome.sph.umich.edu/wiki/METAL_Documentation, Supplemental Table II and III). Approximately 13 million variants available in both TOPMed and UKBB were included in the meta-analysis. Novel genetic variants associated with stroke outcomes were defined as those that showed *P*<5E-9 (whole genome-wide significance) [46] and *P*<5E-8 (genome-wide significance) and were located more than 1Mb of any reported loci. There have been reports about inflated odds ratios (OR) using the SAIGE algorithm[47] for very rare alleles. As an alternative analysis algorithm, we further implemented the Firth algorithm for association testing, which uses a penalized likelihood approach to estimate coefficients[48], in all unrelated TOPMed samples (removing cousins and closer relatives).

All novel loci that showed genome-wide significance (P<5E-8) were carried forward to the replication stage (Fig. 1). Replication was performed using data from the National Institute of Neurological Disorders and Stroke (NINDS) Stroke Genetics Network (NINDS-SiGN, 16,851 cases and 32,473 controls, IS, CES, LAS, and SVS were available for testing)[17]. WHI was included in both SiGN and TOPMed, and meta-analysis combining all studies in SiGN was performed after excluding samples from WHI. Since higher blood pressure is a risk factor for developing stroke, we also sought to determine whether any of our novel stroke loci were associated with blood pressure or hypertension in the TOPMed Blood Pressure Working Group. Details are provided in the Supplemental Methods.

To dissect association signals at previously established stroke loci, we performed stepwise conditional analysis within each known stroke locus that harbored at least two variants showing P<1E-5 with any stroke type or subtype in TOPMed and UKBB using individual level data.

We followed the STREGA (Strengthening the reporting of genetic association studies) reporting guideline, and a flow diagram is presented in Fig. 1.

Aggregated rare variant association analysis

In the aggregated association analysis, rare variants with MAF<1% were combined using various gene-based aggregation units that were based on high confidence loss of function (hcLoF), missense, protein-altering indels, and synonymous variants (defined based on GENCODE). Associations of aggregated units with AS, IS and HS were tested with adjustment for age, sex, ancestry, study, the first 10 PCs, and GRM using a logistic mixed model implemented in the GENESIS package[49]. We used two sampling approaches to overcome the unbalanced case/control ratios. The main approach was to include participants from WHI only, as this study contributed 70.5% of stroke cases and had a relatively balanced case/control ratio (Supplemental Table IV). The other exploratory approach was to randomly match case and control participants on a 1:3 ratio based on study, ancestry, and sex (Supplemental Table IV). Both burden test and SNP-set Sequence Kernel Association Test (SKAT) were performed for each gene region harboring more than one variant and a total MAC>20 (Supplemental Table V). Gene-wide and suggestively significant regions were defined as those with P<(0.05/(number of tested regions×four types of aggregation units×two types of association testing methods)) and P<0.05/(number of tested regions), respectively.

Functional annotation of the novel loci

Bioinformatic follow-up was performed for each novel locus using a comprehensive functional annotation database constructed with the whole genome sequence annotator (WGSA[50], including GTEx[51], DANN[52] and Eigen-PC[53] scores) and a custom UCSC analysis data hub visualizing enhancer and repressor activities, DNase I hypersensitive sites (DHS) and transcribed regions in selected tissues (Supplemental Methods).

Results

Single variant association analysis

In the discovery stage, we performed two GWAS analyses, one focused on multi-ethnic samples in TOPMed with denser coverage of the genome using WGS data and the other combining European samples in TOPMed and UKBB to increase statistical power while focusing on relatively common variants. Among 6,833 stroke cases and 27,116 controls in TOPMed, 65.7% are of European ancestry and 69.1% are females (Table 1). Genomic inflation factors ranged from 0.967 to 1.103, indicating limited evidence of population stratification in the association analyses (Supplemental Table II and III).

In the ancestry-combined association analysis in TOPMed, we identified one novel locus at whole genome-wide significance (*13q33*-rs181401679 for LAS, *P*=3.67E-9) and three additional novel loci showing genome-wide significant associations (*7q22*-rs141857337 for HS, *RAP1GAP2*-rs60380775 and *AUTS2*-rs150022429 for IS, *P*<5E-8) (Table 2, Supplemental Fig. I). The MAFs of the lead variants ranged from 0.1% (*13q33*-rs181401679) to 2.2% (*RAP2GAP2*-rs60380775) in all TOPMed samples. They are either monomorphic (*7q22*-rs141857337 and *AUST2*-rs150022429) or extremely rare (*13q33*-rs181401679 and *RAP2GAP2*-rs60380775, MAF=0.1%) in European populations. Ancestry-specific analysis in European and AA ancestry populations indicated that the identified significant associations were mainly driven by signals in AA populations at three loci (*7q22*, *AUTS2*, and *RAP1GAP2*, Supplemental Table VI). At the *13q33* locus associated with LAS in the combined analysis, no ancestry-specific results could be produced due to a MAC that was below our cut-off (MAC<20). However, we observed considerably higher MAF in Hispanic ancestry cases (MAF=10.5%, Supplemental Table VII), suggesting that the identified association might be driven by the Hispanic subgroup. In the AA-specific analysis, we identified one additional novel locus at genome-wide significance, *TEX13C*-rs145400922 on chromosome X for association with CES (*P*=2.40E-8, Table 2). Among these five novel loci, two of them (*RAP1GAP2* and *TEX13C*) harbored multiple variants with *P*<5E-8, while the top variant at each of the other three loci (*7q22*, *AUTS2*, and *13q33*) was the only one showing *P*<5E-8. All of them harbored multiple variants with *P*<1E-5, ranging from two to 21. Associations of the five novel loci with each stroke type and subtype are presented in Supplemental Table VIII. The OR values that were observed for the novel loci using the SAIGE algorithm, especially for the two extremely rare variants at *7q22* and *13q33,* were larger than those observed using the Firth algorithm in unrelated TOPMed samples (Table 2). The *P* values using both algorithms were similar, but associations at *AUTS2* and *13q33* were not genome-wide significant after Firth correction (*P*>5E-8, Table 2; recall the Firth correction analysis excluded related individuals, and included 5,564 out of 5,616 cases and 21,756 out of 27,116 controls for IS, and similar reductions for other types and subtypes).

In the European-specific meta-analysis of TOPMed and UKBB for IS, ICH, and SAH, we did not identify additional novel loci. A previously reported locus *PITX2* reached whole genome-wide significance for association with IS in the meta-analysis using the SAIGE results (rs1906611, *P*=4.68E-9, Supplemental Table IX). None of the five novel loci we

discovered in TOPMed were available in UKBB due to their extremely low MAFs or monomorphism in Europeans.

We sought replication of the five novel loci in Table 2 in the multi-ethnic SiGN Consortium. Although the coverage of variants in SiGN was improved through imputation using the TOPMed WGS data as the reference panel, it remains difficult to capture genetic variants with MAF<0.5%. As a result, only two loci (*AUTS2*-rs150022429 and *RAP1GAP2*-rs60380775) were available in SiGN and the other three loci were not available due to their low MAFs. However, neither of the two loci showed evidence of association with IS in SiGN (*P*>0.05, Supplemental Table X). The AA samples in SiGN (1,323 IS cases and 2,383 controls) provided 80% power to detect OR>1.75 for *AUTS2*-rs150022429 and OR>1.27 for *RAP1GAP2*-rs60380775. In the analysis of our five novel loci with blood pressure phenotypes in TOPMed, all loci were available for testing except for *TEX13C* on chromosome X. None of the four novel loci showed evidence of association with SBP, DBP or hypertension (*P*>0.05, Supplemental Table XI).

Assessment of previously reported stroke loci

Full association results of the 51 previously reported stroke loci (72 unique variants) in UKBB, TOPMed ancestry-combined analysis, and meta-analysis of TOPMed and UKBB are presented in Supplemental Table XI. In TOPMed alone, four of the 51 known loci were not available (polymorphic only in East Asians) and three achieved at least nominal statistical significance [*P*<1.06E-3 (0.05/47)] for the same stroke phenotype using the reported variants (*PITX2* and *ZFHX3* for CES, and *HDAC9* for LAS, Supplemental Table XII). Seven regions harbored more than two variants with *P*<1E-5 (*ALDH1A2* and *PITX2* for CES, *PMF1* for HS, *TBX3*, *CYP4F12*, and *SLC6A11* for IS, *SH3PXD2A-OBFC1* for AS), but no additional signals were identified at these loci in the stepwise conditional analysis (Supplemental Table XIII). In UKBB, seven of the 51 reported stroke loci were not available (monomorphism or extremely low MAFs in Europeans) and four loci (*PITX2*, *LRCH1*, *HDAC9*, and *ZFHX3*) with IS were nominally significant at *P*<1.14E-3 using the reported variants (0.05/44). Four regions harbored more than two variants with *P*<1E-5 (*PITX2*, *HDAC9*, and *HABP2* for IS, and *ITPK1* for ICH), but no additional variants showed *P*<1E-5 in the 1Mb regions (Data not shown).

Aggregated rare variant association analysis

In the aggregated association analysis, we focused on rare genetic variants and three stroke types (AS, IS and HS). No gene region reached gene-wide significance, but one gene region aggregated using synonymous rare variants, *C6orf26* showed suggestive significance [*P*<3.12E-6 (0.05/16,051 regions)] for HS in WHI (Supplemental Table XIV). Similar *P* values were observed in the burden and the SKAT tests (*P*=1.33E-6 and 4.59E-7, respectively, Supplemental Table XIV). In the randomly selected samples, only 4 rare variants with a total MAC of 10 were included in the analysis due to smaller sample size compared to using all WHI samples (Supplemental Table IV), which is below the MAC cutoff we used and were excluded from the analysis. Associations of this region using other aggregation units with stroke outcomes were not significant (Supplemental Table XIV). This region is located about 400kb away from a previously reported common variant

SLC22A7-rs16896398 for association with AS[7]. Among the nine rare variants included in this region, rs61747887 showed the highest MAF of 0.9%, and is more frequent in European compared to AA populations (MAF=1.5% and 0.5%, respectively). It showed nominal association with HS in the ancestry-combined and European-specific analysis (*P*=9.94E-5 and 5.23E-5, respectively) while no evidence of association was observed in AA-specific analysis (*P*=0.80). The rare variant rs61747887 we observed is not in LD with the reported common variant *SLC22A7*-rs16896398 (r^2<0.1), which showed no evidence of association with AS or HS in TOPMed ancestry-combined analysis (*P*=0.44 and 0.48, respectively). In GTEx, rs61747887 is associated with gene expression levels of *CUL7* (*P*=1.5E-5) and *RP1–20C7.6* (*P*=4.4E-5) in brain tissue. In addition, seven previously reported loci harbored nearby genes (±500kb) that showed evidence of association with at least one of the three stroke types [*P*<9.80E-4 (0.05/51 known loci), Supplemental Table XV].

Functional annotation of the novel loci

At each of the five significant or suggestive novel loci listed in Table 2, the lead variant and its LD proxies (r^2 0.4) were examined using both the functional annotation database constructed from WGSA (Supplemental Table XVI) and the customized UCSC genome browser (Supplemental Fig. II). At the *7q22* locus, the lead variant rs141857337 and its two LD proxies all showed Eigen-PC score>0 (functional, Supplemental Table XVI) and overlapped with enhancer, repressor, and DHS in brain and ventricle tissues (Supplemental Fig. II A). At the *RAP1GAP2* locus, an LD proxy rs115318048 that is in moderate LD with the lead variant (r^2=0.48) overlapped with enhancer activity in all selected tissues (Supplemental Fig. II D). At the *TEX13C* locus, the lead variant rs145400922 showed DANN scores>0.9 (deleterious, Supplemental Table XVI).

Discussion

We present the first WGS association analysis for stroke and subtypes in ancestrally-diverse populations. We identified five possible novel loci harboring low-frequency lead variants in the single variant association analysis and one suggestively gene-wide significant gene in the aggregate association analysis indicating independent signals from rare variants at an established region. We were unable to replicate two of the single variant association signals that were available using independent data from the SiGN Consortium. While our findings suggest the potential benefits of combining WGS data with populations of diverse genetic backgrounds to identify possible low-frequency or ancestry-specific variants associated with stroke, they also highlight some of the accompanying challenges including the requirement for very large numbers of stroke cases for discovery especially in the face of, phenotypic complexity compounded by the current paucity of appropriately-powered replication samples.

In our discovery analysis in TOPMed, approximately 40 million genetic variants with MAC>20 were examined in the single variant association analysis, which is five times the number of variants examined in the MEGASTROKE Consortium after imputation using the 1000 Genome reference panel[7]. The substantial improvement in the coverage of the genome using WGS coupled with ancestrally-diverse populations facilitated the identification

of ancestry-specific low-frequency variants associated with stroke especially among non-European populations. Among the five novel loci we identified in the single variant association analysis in TOPMed, two of them harbored lead variants that are monomorphic in European ancestry populations (*7q22* and *AUTS2*) and three of the lead variants show low MAFs in Europeans (MAF<0.1% for *13q33* and *RAP1GAP2,* and MAF=0.2% for *TEX13C*). Previous GWAS analyses focusing on European ancestry populations and relatively common variants (MAF 1%) would not be able to capture these low-frequency and ancestry-specific variants. Of note, *RAP1GAP2* encodes a GTPase-activating protein that activates the small guanine-nucleotide-binding protein Rap1 in platelets and interacts with synaptotagmin-like protein 1 and Rab27 and regulates secretion of dense granules from platelets at sites of endothelial damage. This gene has been reported for suggestive association with sudden cardiac arrest[54] and genome-wide significant association with white blood cell indices[55, 56], but none of the reported variants are in LD with the lead variant we identified for association with IS in TOPMed (r^2<0.02). In addition to the novel findings in the single variant association analysis, examination of rare variants (MAF<1% and total MAC>20) derived from WGS through aggregated analysis in TOPMed highlighted a rare variant, rs61747887, at an established region, *SLC22A7*, which is not in LD with the reported common variant rs16896398 (r^2<0.1)[7]. Gene expression data from GTEx helped to prioritize *CUL7* at this locus, whose encoded protein is a component of an E3 ubiquitin-protein ligase complex. Previous studies in mouse models have demonstrated its important role in vascular morphogenesis[57] and improved cardiac function after myocardial infarction[58].

Unlike published GWAS analyses focusing on limited numbers of stroke types or subtypes, a major strength of our study is our ability to perform a comprehensive analysis for AS, including the two stroke types (IS and HS), and the five subtypes (CES, LAS, SVS, ICH, and SAH). Previous studies have identified shared genetic loci across different subtypes (*SH2B3* was associated with both LAS and SVS and *ABO* was associated with both LAS and CES) as well as subtype-specific loci (*EDNRA*, *LINC01492*, *TSPAN2*, and *HDAC9* were associated with LAS only and *PITX2* and *NKX2–5* were associated with CES only)[7, 10, 11, 17, 18]. These findings indicate both shared biological pathways and risk factors across stroke subtypes and subtype-specific mechanisms. We observed similar results at *PITX2* and *HDAC9* in TOPMed. At the *PITX2* locus, all reported variants exhibiting more significant associations for CES compared to IS (smallest *P*=9.01E-7 and 0.011 for CES and IS, respectively, Supplemental Table XII). At *HDAC9* locus, reported variants showed nominal association with LAS but not IS (smallest *P*=4.24E-4 and 0.151 for LAS and IS, respectively, Supplemental Table XII). In addition, the *ZFHX3* locus has been primarily reported for association with CES, but we also observed nominal association with SVS in TOPMed (smallest *P*=5.10E-4 and 3.98E-3 for CES and SVS, respectively, Supplemental Table XII). In the evaluation of the five novel loci across stroke types and subtypes in TOPMed, the *13q33* locus significantly associated with LAS (*P*=3.67E-9) was nominally associated with SAH (*P*=0.022) and the *TEX13C* suggestively associated with CES (*P*=2.54E-8) was nominally associated with SVS (*P*=0.028, Supplemental Table VIII), suggesting these loci may impact pathways important across multiple stroke subtypes.

Our study has several limitations. First, only two out of the five novel loci are present in the SiGN replication dataset and can be attempted for replication. The fact that these novel loci are relatively rare made it difficult to find proper replication datasets for the other three loci. Second, the sample sizes in TOPMed remained limited compared to published GWAS, with the largest meta-analysis incorporating over 67,000 stroke cases and 454,000 controls[7]. This situation likely contributed to the relatively small numbers of reported loci that were confirmed in our analysis and the failure to identify independent signals at reported loci. Moreover, over 70% of our sample involves Europeans, making it challenging to definitively identify heterogeneity of the associated loci across diverse ancestral groups. Third, some of the cases were not grouped into subtypes, especially in HS where more than half of the cases did not have subtype classification. This missingness further limited statistical power to identify novel findings for these subtypes.

In conclusion, we performed the first association analysis for stroke types and subtypes using WGS data in ancestrally-diverse populations. Through single variant and aggregate association analyses, we identified five novel loci that harbored low-frequency variants and showed ancestry-specificity and confirmed one reported gene region at genome-wide significance. These findings require replication in additional well powered sample set when available. Our findings indicate that dense coverage of the genome, large sample sizes, increased representation of ancestrally-diverse participants, and detailed classification of stroke cases are essential to the identification of novel findings and better characterization of stroke-associated loci.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgements

TOPMed Accession #	TOPMed Project	Parent Study Short Name	Omics Center	Omics Support	Omics Type
phs001211	AFGen	ARIC AFGen	Broad Genomics	3R01HL092577-06S1	WGS
phs001211	VTE	ARIC	Baylor	3U54HG003273-12S2 / HHSN268201500015C	WGS
phs001644	BioMe	BioMe	Baylor	HHSN268201600033I	WGS

TOPMed Accession #	TOPMed Project	Parent Study Short Name	Omics Center	Omics Support	Omics Type
phs001644	BioMe	BioMe	MGI	HHSN268201600037I	WGS
phs001368	CHS	CHS	Baylor	HHSN268201600033I	WGS
phs001368	VTE	CHS VTE	Baylor	3U54HG003273-12S2 / HHSN268201500015C	WGS
phs000974	AFGen	FHS AFGen	Broad Genomics	3R01HL092577-06S1	WGS
phs000974	FHS	FHS	Broad Genomics	3U54HG003067-12S2	WGS
phs000964	JHS	JHS	NWGC	HHSN268201100037C	WGS
phs001416	AA_CAC	MESA AA_CAC	Broad Genomics	HHSN268201500014C	WGS
phs001416	MESA	MESA	Broad Genomics	3U54HG003067-13S1	WGS
phs001237	WHI	WHI	Broad Genomics	HHSN268201500014C	WGS

Non-standard Abbreviations and Acronyms

AA	African American
ARIC	Atherosclerosis Risk in Communities Study
AS	all stroke
BioMe	BioMe™ Biobank
CES	cardioembolic stroke
CHS	Cardiovascular Health Study
DBP	diastolic blood pressure
DHS	DNase I hypersensitive sites
FHS	Framingham Heart Study
GWAS	genome-wide association studies
GRM	genetic relationship matrix

hcLoF	high confidence loss of function
HS	hemorrhagic stroke
ICD	International Statistical Classification of Diseases and Related Health Problems
ICH	intracerebral hemorrhage
IS	ischemic stroke
JHS	Jackson Heart Study
LAS	Large artery stroke
LD	linkage disequilibrium
MAC	minor allele count
MAF	minor allele frequency
MESA	Multi-Ethnic Study of Atherosclerosis
NINDS	National Institute of Neurological Disorders and Stroke
OR	odds ratio
PC	principal component
QC	quality control
SAH	subarachnoid hemorrhage
SAIGE	Scalable and Accurate Implementation of Generalized mixed model
SBP	systolic blood pressure
SiGN	Stroke Genetics Network
SKAT	SNP-set Sequence Kernel Association Test
STREGA	Strengthening the reporting of genetic association studies
SVS	small vessel stroke
TOAST	Trial of Org 10172 in Acute Stroke Treatment
TOPMed	Trans-Omics for Precision Medicine
UKBB	UK Biobank
WGS	whole genome sequencing
WGSA	whole genome sequence annotator
WHI	Women's Health Initiative

References

1. GBD 2015 mortality and causes of death collaborators. Global, regional, and national life expectancy, all-cause mortality, and cause-specifi c mortality for 249 causes of death, 1980–2015: A systematic analysis for the global burden of disease study 2015 (vol 388, pg 1459, 2016). Lancet. 2016;388:1459–1544. [PubMed: 27733281]

2. Feigin VL, Norrving B, Mensah GA. Global burden of stroke. Circ Res. 2017;120:439–448. [PubMed: 28154096]

3. Holliday EG, Maguire JM, Evans TJ, Koblar SA, Jannes J, Sturm JW, Hankey GJ, Baker R, Golledge J, Parsons MW, et al. Common variants at 6p21.1 are associated with large artery atherosclerotic stroke. Nature Genetics. 2012;44:1147–1151. [PubMed: 22941190]

4. Bevan S, Traylor M, Adih-Samii P, Malik R, Paul NLM, Jackson C, Farrall M, Rothwell PM, Sudlow C, Dichgans M, et al. Genetic heritability of ischemic stroke and the contribution of previously reported candidate gene and genomewide associations. Stroke. 2012;43:3161–3167. [PubMed: 23042660]

5. Devan WJ, Falcone GJ, Anderson CD, Jagiella JM, Schmidt H, Hansen BM, Jimenez-Conde J, Giralt-Steinhauer E, Cuadrado-Godia E, Soriano C, et al. Heritability estimates identify a substantial genetic contribution to risk and outcome of intracerebral hemorrhage. Stroke. 2013;44:1578–1583. [PubMed: 23559261]

6. Korja M, Silventoinen K, McCarron P, Zdravkovic S, Skytthe A, Haapanen A, Faire U, Pedersen NL, Christensen K, Koskenvuo M, et al. Genetic epidemiology of spontaneous subarachnoid hemorrhage nordic twin study. Stroke. 2010;41:2458–2462. [PubMed: 20847318]

7. Malik R, Chauhan G, Traylor M, Sargurupremraj M, Okada Y, Mishra A, Rutten-Jacobs L, Giese A, Lann SW, Gretarsdottir S, et al. Multiancestry genome-wide association study of 520,000 subjects identifies 32 loci associated with stroke and stroke subtypes. Nature Genetics. 2018;50:524–537. [PubMed: 29531354]

8. Ikram MA, Seshadri S, Bis JC, Fornage M, DeStefano AL, Aulchenko YS, Debette S, Lumley T, Folsom AR, van den Herik EG, et al. Genomewide association studies of stroke. N Engl J Med. 2009;360:1718–1728. [PubMed: 19369658]

9. Gudbjartsson DF, Holm H, Gretarsdottir S, Thorleifsson G, Walters GB, Thorgeirsson G, Gulcher J, Mathiesen EB, Njølstad I, Nyrnes A, et al. A sequence variant in zfhx3 on 16q22 associates with atrial fibrillation and ischemic stroke. Nat Genet. 2009;41:876–878. [PubMed: 19597491]

10. Traylor M, Farrall M, Holliday EG, Sudlow C, Hopewell JC, Cheng YC, Fornage M, Ikram MA, Malik R, Bevan S, et al. Genetic risk factors for ischaemic stroke and its subtypes (the metastroke collaboration): A meta-analysis of genome-wide association studies. Lancet Neurol. 2012;11:951–962. [PubMed: 23041239]

11. International Stroke Genetics C, Wellcome Trust Case Control C, Bellenguez C, Bevan S, Gschwendtner A, Spencer CC, Burgess AI, Pirinen M, Jackson CA, Traylor M, et al. Genome-wide association study identifies a variant in hdac9 associated with large vessel ischemic stroke. Nat Genet. 2012;44:328–333. [PubMed: 22306652]

12. Williams FMK, Carter AM, Hysi PG, Surdulescu G, Hodgkiss D, Soranzo N, Traylor M, Bevan S, Dichgans M, Rothwell PMW, et al. Ischemic stroke is associated with the abo locus: The euroclot study. Ann Neurol. 2014;75:166–167.

13. Woo D, Falcone GJ, Devan WJ, Brown WM, Biffi A, Howard TD, Anderson CD, Brouwers HB, Valant V, Battey TWK, et al. Meta-analysis of genome-wide association studies identifies 1q22 as a susceptibility locus for intracerebral hemorrhage. American Journal of Human Genetics. 2014;94:511–521. [PubMed: 24656865]

14. Kilarski LL, Achterberg S, Devan WJ, Traylor M, Malik R, Lindgren A, Pare G, Sharma P, Slowik A, Thijs V, et al. Meta-analysis in more than 17,900 cases of ischemic stroke reveals a novel association at 12q24.12. Neurology. 2014;83:678–685. [PubMed: 25031287]

15. Traylor M, Makela KM, Kilarski LL, Holliday EG, Devan WJ, Nalls MA, Wiggins KL, Zhao W, Cheng Y, Achterberg S, et al. A novel mmp12 locus is associated with large artery atherosclerotic stroke using a genome-wide age-at-onset informed approach. Plos Genetics. 2014;10:e1004469. [PubMed: 25078452]

16. Carty CL, Keene KL, Cheng YC, Meschia JF, Chen WM, Nalls M, Bis JC, Kittner SJ, Rich SS, Tajuddin S,et al. Meta-analysis of genome-wide association studies identifies genetic risk factors for stroke in african americans. Stroke. 2015;46:2063–2068. [PubMed: 26089329]

17. NINDS Stroke Genetics Network (SiGN); International Stroke Genetics Consortium (ISGC). Loci associated with ischaemic stroke and its subtypes (sign): A genome-wide association study. Lancet Neurology. 2016;15:174–184. [PubMed: 26708676]

18. Malik R, Traylor M, Pulit SL, Bevan S, Hopewell JC, Holliday EG, Zhao W, Abrantes P, Amouyel P, Attia JR, et al. Low-frequency and common genetic variation in ischemic stroke the metastroke collaboration. Neurology. 2016;86:1217–1226. [PubMed: 26935894]

19. Traylor M, Malik R, Nalls MA, Cotlarciuc I, Radmanesh F, Thorleifsson G, Hanscombe KB, Langefeld C, Saleheen D, Rost NS, et al. Genetic variation at 16q24.2 is associated with small vessel stroke. Ann Neurol. 2017;81:383–394. [PubMed: 27997041]

20. Yamada Y, Sakuma J, Takeuchi I, Yasukochi Y, Kato K, Oguri M, Fujimaki T, Horibe H, Muramatsu M, Sawabe M, et al. Identification of six polymorphisms as novel susceptibility loci for ischemic or hemorrhagic stroke by exome-wide association studies. Int J Mol Med. 2017;39:1477–1491. [PubMed: 28487959]

21. Howard VJ, Kleindorfer DO, Judd SE, McClure LA, Safford MM, Rhodes JD, Cushman M, Moy CS, Soliman EZ, Kissela BM, et al. Disparities in stroke incidence contributing to disparities in stroke mortality. Ann Neurol. 2011;69:619–627. [PubMed: 21416498]

22. Kissela B, Schneider A, Kleindorfer D, Khoury J, Miller R, Alwell K, Woo D, Szaflarski J, Gebel J, Moomaw C, et al. Stroke in a biracial population - the excess burden of stroke among blacks. Stroke. 2004;35:426–431. [PubMed: 14757893]

23. Gardener H, Sacco RL, Rundek T, Battistella V, Cheung YK, Elkind MSV. Race and ethnic disparities in stroke incidence in the northern manhattan study. Stroke. 2020;51:1064–1069 [PubMed: 32078475]

24. The atherosclerosis risk in communities (aric) study: Design and objectives. The aric investigators. Am J Epidemiol. 1989;129:687–702. [PubMed: 2646917]

25. Rosamond WD, Folsom AR, Chambless LE, Wang CH, McGovern PG, Howard G, Copper LS, Shahar E. Stroke incidence and survival among middle-aged adults: 9-year follow-up of the atherosclerosis risk in communities (aric) cohort. Stroke. 1999;30:736–743. [PubMed: 10187871]

26. The national survey of stroke. National institute of neurological and communicative disorders and stroke. Stroke. 1981;12:I1–91. [PubMed: 7222163]

27. Fried LP, Borhani NO, Enright P, Furberg CD, Gardin JM, Kronmal RA, Kuller LH, Manolio TA, Mittelmark MB, Newman A, et al. The cardiovascular health study: Design and rationale. Ann Epidemiol. 1991;1:263–276. [PubMed: 1669507]

28. Longstreth WT, Bernick C, Fitzpatrick A, Cushman M, Knepper L, Lima J, Furberg CD. Frequency and predictors of stroke death in 5,888 participants in the cardiovascular health study. Neurology. 2001;56:368–375. [PubMed: 11171903]

29. Dawber TR, Kannel WB. The framingham study. An epidemiological approach to coronary heart disease. Circulation. 1966;34:553–555. [PubMed: 5921755]

30. Feinleib M, Kannel WB, Garrison RJ, McNamara PM, Castelli WP. The framingham offspring study. Design and preliminary data. Prev Med. 1975;4:518–525. [PubMed: 1208363]

31. Splansky GL, Corey D, Yang Q, Atwood LD, Cupples LA, Benjamin EJ, D'Agostino RB Sr, Fox CS, Larson MG, Murabito JM, et al. The third generation cohort of the national heart, lung, and blood institute's framingham heart study: Design, recruitment, and initial examination. Am J Epidemiol. 2007;165:1328–1335. [PubMed: 17372189]

32. Carandang R, Seshadri S, Beiser A, Kelly-Hayes M, Kase CS, Kannel WB, Wolf PA. Trends in incidence, lifetime risk, severity, and 30-day mortality of stroke over the past 50 years. JAMA. 2006;296:2939–2946. [PubMed: 17190894]

33. Seshadri S, Beiser A, Kelly-Hayes M, Kase CS, Au R, Kannel WB, Wolf PA. The lifetime risk of stroke: Estimates from the framingham study. Stroke. 2006;37:345–350. [PubMed: 16397184]

34. Wolf PA, Kannel WB, Dawber TR. Prospective investigations: The framingham study and the epidemiology of stroke. Adv Neurol. 1978;19:107–120. [PubMed: 742537]

35. Taylor HA Jr. The jackson heart study: An overview. Ethn Dis. 2005;15:S6–1-3.

36. Carpenter MA, Crow R, Steffes M, Rock W, Heilbraun J, Evans G, Skelton T, Jensen R, Sarpong D. Laboratory, reading center, and coordinating center data management methods in the jackson heart study. Am J Med Sci. 2004;328:131–144. [PubMed: 15367870]

37. Fuqua SR, Wyatt SB, Andrew ME, Sarpong DF, Henderson FR, Cunningham MF, Taylor HA Jr. Recruiting african-american research participation in the jackson heart study: Methods, response rates, and sample description. Ethn Dis. 2005;15:S6–18-29.

38. Payne TJ, Wyatt SB, Mosley TH, Dubbert PM, Guiterrez-Mohammed ML, Calvin RL, Taylor HA Jr, Williams DR. Sociocultural methods in the jackson heart study: Conceptual and descriptive overview. Ethn Dis. 2005;15:S6–38-48.

39. Wyatt SB, Akylbekova EL, Wofford MR, Coady SA, Walker ER, Andrew ME, Keahey WJ, Taylor HA, Jones DW. Prevalence, awareness, treatment, and control of hypertension in the jackson heart study. Hypertension. 2008;51:650–656. [PubMed: 18268140]

40. Keku E, Rosamond W, Taylor HA Jr., Garrison R, Wyatt SB, Richard M, Jenkins B, Reeves L, Sarpong D. Cardiovascular disease event classification in the jackson heart study: Methods and procedures. Ethn Dis. 2005;15:S6–62-70.

41. Bild DE, Bluemke DA, Burke GL, Detrano R, Diez Roux AV, Folsom AR, Greenland P, Jacob DR Jr, Kronmal R, Liu K, et al. Multi-ethnic study of atherosclerosis: Objectives and design. Am J Epidemiol. 2002;156:871–881. [PubMed: 12397006]

42. Kawasaki R, Xie J, Cheung N, Lamoureux E, Klein R, Klein BE, Cotch MF, Sharrett AR, Shea S, Wong TY, et al. Retinal microvascular signs and risk of stroke: The multi-ethnic study of atherosclerosis (mesa). Stroke. 2012;43:3245–3251. [PubMed: 23111439]

43. Design of the women's health initiative clinical trial and observational study. The women's health initiative study group. Control Clin Trials. 1998;19:61–109. [PubMed: 9492970]

44. Adams HP Jr., Bendixen BH, Kappelle LJ, Biller J, Love BB, Gordon DL, Marsh EE 3rd. Classification of subtype of acute ischemic stroke. Definitions for use in a multicenter clinical trial. Toast. Trial of org 10172 in acute stroke treatment. Stroke. 1993;24:35–41. [PubMed: 7678184]

45. Zhou W, Nielsen JB, Fritsche LG, Dey R, Gabrielsen ME, Wolford BN, LeFaive J, VandeHaar P, Gagliano SA, Gifford A, et al. Efficiently controlling for case-control imbalance and sample relatedness in large-scale genetic association studies. Nat Genet. 2018;50:1335–1341. [PubMed: 30104761]

46. Lin DY. A simple and accurate method to determine genomewide significance for association tests in sequencing studies. Genet Epidemiol. 2019;43:365–372. [PubMed: 30623491]

47. Mbatchou J, Barnard L, Backman J, Marketta A, Kosmicki JA, Ziyatdinov A, Benner C, O'Dushlaine C, Barber M, Boutkov B, et al. Computationally efficient whole genome regression for quantitative and binary traits. bioRxiv 2020.06.19.162354.

48. Firth D Bias Reduction of Maximum Likelihood Estimates. Biometrika. 1993; 80:27–38.

49. Gogarten SM, Sofer T, Chen H, Yu CY, Brody JA, Thornton TA, Rice KM, Conomos MP. Genetic association testing using the genesis r/bioconductor package. Bioinformatics. 2019;35:5346–5348. [PubMed: 31329242]

50. Liu X, White S, Peng B, Johnson AD, Brody JA, Li AH, Huang Z, Carroll A, Wei P, Gibbs R, et al. Wgsa: An annotation pipeline for human genome sequencing studies. J Med Genet. 2016;53:111–112. [PubMed: 26395054]

51. Gamazon ER, Segre AV, van de Bunt M, Wen XQ, Xi HS, Hormozdiari F, Ongen H, Konkashbaev A, Derks EM, Aguet F, et al. Using an atlas of gene regulation across 44 human tissues to inform complex disease- and trait-associated variation. Nature Genetics. 2018;50:956–967. [PubMed: 29955180]

52. Quang D, Chen YF, Xie XH. Dann: A deep learning approach for annotating the pathogenicity of genetic variants. Bioinformatics. 2015;31:761–763. [PubMed: 25338716]

53. Ionita-Laza I, McCallum K, Xu B, Buxbaum JD. A spectral approach integrating functional genomic annotations for coding and noncoding variants. Nature Genetics. 2016;48:214–220. [PubMed: 26727659]

54. Aouizerat BE, Vittinghoff E, Musone SL, Pawlikowska L, Kwok PY, Olgin JE, Tseng ZH. Gwas for discovery and replication of genetic loci associated with sudden cardiac arrest in patients with coronary artery disease. BMC Cardiovasc Disord. 2011;11–29. [PubMed: 21410963]

55. Astle WJ, Elding H, Jiang T, Allen D, Ruklisa D, Mann AL, Mead D, Bouman H, Riveros-Mckay F, Kostadima MA, et al. The allelic landscape of human blood cell trait variation and links to common complex disease. Cell. 2016;167:1415–1429. [PubMed: 27863252]

56. Kichaev G, Bhatia G, Loh PR, Gazal S, Burch K, Freund MK, Schoech A, Pasaniuc B, Price AL. Leveraging polygenic functional enrichment to improve gwas power. Am J Hum Genet. 2019;104:65–75. [PubMed: 30595370]

57. Arai T, Kasper JS, Skaar JR, Ali SH, Takahashi C, DeCaprio JA. Targeted disruption of p185/cul7 gene results in abnormal vascular morphogenesis. Proc Natl Acad Sci U S A. 2003;100:9855–9860. [PubMed: 12904573]

58. Hassink RJ, Nakajima H, Nakajima HO, Doevendans PA, Field LJ. Expression of a transgene encoding mutant p193/cul7 preserves cardiac function and limits infarct expansion after myocardial infarction. Heart. 2009;95:1159–1164. [PubMed: 19435717]
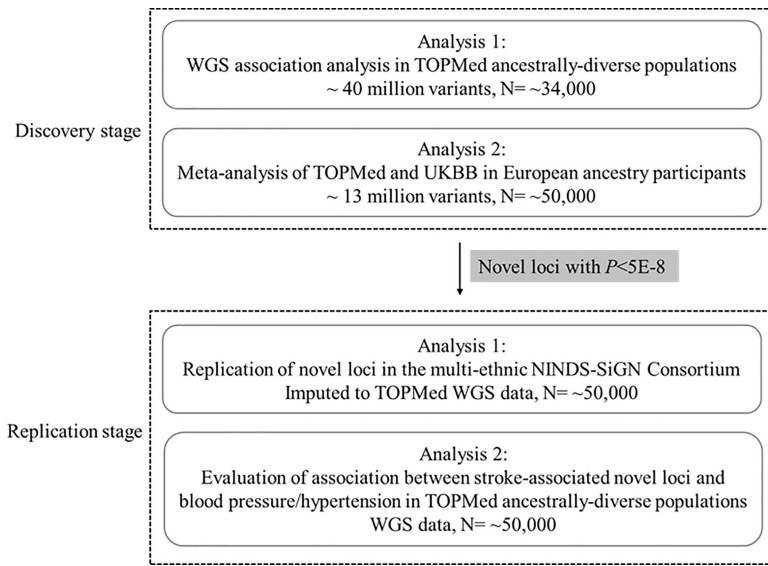
Fig. 1.
Study design.

Table 1.

Age, sex and ancestry distributions according to stroke types and subtypes among participants in TOPMed and UKBB

Stroke	Age (years)[1]	Women (%)	N	N by ancestry					
				European	AA	Hispanic/Latino	Asian	Native American	Other
TOPMed									
AS[2]	75.1±8.8	87.4	6,833	5,294	1,022	360	116	20	21
IS[2]	75.1±8.7	87.1	5,616	4,307	884	316	80	12	17
CES	78.9±7.3	92.5	1,459	1,276	122	39	17	5	-
LAS	75.5±7.2	91.8	352	296	33	19	4	-	-
SVS	74.7±7.8	93.1	898	692	132	43	29	2	-
HS[2]	74.5±8.8	90.7	1,080	862	127	43	36	8	4
ICH	75.1±8.0	94.3	716	592	68	19	31	6	-
SAH	70.5±8.3	96.6	208	167	26	9	5	1	-
Control	74.6±11.3	64.5	27,116	17,021	6,855	2,256	734	34	216
UKBB									
IS	61.4±6.6	35.3	4,474	4,474	-	-	-	-	-
ICH	60.9±6.8	43.0	959	959	-	-	-	-	-
SAH	58.1±7.2	60.7	1,194	1,194	-	-	-	-	-
Control for IS	56.6±8.1	33.3	24,000	24,000	-	-	-	-	-
Control for ICH	56.6±8.1	33.3	4,800	4,800	-	-	-	-	-
Control for SAH	56.6±8.2	33.3	5,970	5,970	-	-	-	-	-

AS, all stroke; IS, ischemic stroke; CES, cardioembolic stroke; LAS, large artery stroke; SVS, small vessel stroke; HS, hemorrhagic stroke; ICH, intracerebral hemorrhage; SAH, subarachnoid hemorrhage; AA, African American.

[1] Age of stroke cases indicated age at incident stroke and age of controls indicated age at the last follow-up.

[2] Some AS, IS and HS cases were unclassified.

Table 2.

Novel loci identified in the whole-genome single variant association analyses

Variant	Chr:pos (Build 38)	Gene/Region	CA/NCA	CAF (EA/AA/HA,%)[1]	Outcome	N (case/control)	OR	95%CI	P
TOPMed ancestry-combined analysis using SAIGE[2]									
rs141857337	7:362834	*7q22*	A/G	0/0.7/0.3	HS	1,080/27,116	455.40	50.99, 4067.00	4.26E-8
rs150022429	7:69400857	*AUTS2*	C/T	0/1.1/0.2	IS	5,616/27,116	5.28	2.91, 9.58	4.56E-8
rs181401679	13:104244508	*13q33*	G/C	0.05/0.2/0.4	LAS	352/11,274	1.52E8	2.91E5, 7.96E10	3.67E-9
rs60380775	17:2969421	*RAP1GAP2*	T/C	0.06/8.4/2.0	IS	5,616/27,116	1.74	1.44, 2.12	1.51E-8
TOPMed ancestry-combined analysis using the Firth algorithm[3]									
rs141857337	7:362834	*7q22*	A/G	0/0.7/0.3	HS	1,072/21,756	12.94	5.75, 26.98	4.90E-8
rs150022429	7:69400857	*AUTS2*	C/T	0/1.1/0.2	IS	5,564/21,756	3.39	2.20, 5.13	1.31E-7
rs181401679	13:104244508	*13q33*	G/C	0.05/0.2/0.4	LAS	349/10,721	34.01	11.33, 92.40	9.68E-8
rs60380775	17:2969421	*RAP1GAP2*	T/C	0.05/8.5/2.1	IS	5,564/21,756	1.69	1.41, 2.01	1.48E-8
TOPMed AA-specific analysis using SAIGE[2]									
rs145400922	X:125619640	*TEX13C*	G/A	0.02/3.6/0.6	CES	122/2,090	11.02	4.74, 25.63	2.40E-8
TOPMed AA-specific analysis using the Firth algorithm[3]									
rs145400922	X:125619640	*TEX13C*	G/A	0.03/3.7/0.6	CES	122/1,994	1.37	4.74, 25.63	2.90E-8

CA, coded allele; NCA, non-coded allele; CAF, coded allele frequency; EA, European ancestry; AA, African American; HA, Hispanic; HS, hemorrhagic stroke; IS, ischemic stroke; LAS, large artery stroke; CES, cardioembolic stroke.

[1] The CAF of each lead variant in each ancestral population was calculated using the associated cases and controls.

[2] Analysis using SAIGE included all TOPMed samples.

[3] Analysis using the Firth algorithm included unrelated TOPMed samples.