

UC Irvine

UC Irvine Previously Published Works

Title

Natural products against renin-angiotensin system for antifibrosis therapy

Permalink

<https://escholarship.org/uc/item/2cw049q4>

Authors

Yang, Tian
Chen, Yuan-Yuan
Liu, Jing-Ru
et al.

Publication Date

2019-10-01

DOI

10.1016/j.ejmech.2019.06.091

Peer reviewed



Review

Small molecules from natural products targeting the Wnt/β-catenin pathway as a therapeutic strategy

Dan Liu^a, Lin Chen^a, Hui Zhao^a, Nosratola D. Vaziri^c, Shuang-Cheng Ma^{b,*,**}, Ying-Yong Zhao^{a,*}^a School of Pharmacy, Faculty of Life Science & Medicine, Northwest University, No. 229 Taibai North Road, Xi'an, Shaanxi, 710069, China^b National Institutes for Food and Drug Control, State Food and Drug Administration, No. 2 Tiantan Xili, Beijing, 100050, China^c Division of Nephrology and Hypertension, School of Medicine, University of California Irvine, Irvine, California, 92897, USA

ARTICLE INFO

Keywords:
 Wnt/β-catenin pathway
 Natural products
 Cancer
 Renal disease
 Neurodegenerative disease

ABSTRACT

The Wnt/β-catenin signaling pathway is an evolutionarily conserved developmental signaling event that plays a critical role in regulating tissue development and maintaining homeostasis, the dysregulation of which contributes to various diseases. Natural products have been widely recognized as a treasure trove of novel drug discovery for millennia, and many clinical drugs are derived from natural small molecules. Mounting evidence has demonstrated that many natural small molecules could inhibit the Wnt/β-catenin pathway, while the efficacy of natural products remains to be determined. Therefore, this paper primarily reviews the targeting mechanism of natural small molecules for aberrant Wnt/β-catenin pathway that is intimately implicated in the pathogenesis of myriad diseases, such as cancers, renal diseases, neurodegenerative diseases and bone disorders. In addition, this review also highlights some natural products that have the potential to halt Wnt/β-catenin pathway, especially for porcupine, the receptors of Wnt ligands, β-catenin and β-catenin-dependent proteins. Additionally, a series of natural small molecules have shown good therapeutic effects against mutations of the Wnt/β-catenin pathway, which may dramatically facilitate the development of natural products in Wnt/β-catenin pathway intervention.

1. Introduction

The Wnt/β-catenin signaling pathway is an evolutionarily conserved cellular signaling cascade that plays prominent roles in various biological processes and the pathogenesis of numerous diseases, such as the early embryonic development as well as organogenesis and adult tissue homeostasis [1]. The Wnt/β-catenin pathway primarily consists of the canonical Wnt/β-catenin signaling (β-catenin/T cell factor (TCF)-dependent interaction) and non-canonical Wnt/β-catenin signaling (β-catenin/TCF-independent interaction) [2]. Canonical Wnt/β-catenin signaling includes Wnt ligand secretion, Wnt/receptors interaction, β-catenin stabilization and translocation [3,4]. Although the Wnt/β-catenin pathway shows many benefits, its dysregulation causes extensive diseases, such as cancer, cardiovascular disease and renal disease [5–9]. Many biological antibodies and chemotherapeutic reagents targeting

the aberrant Wnt/β-catenin signaling have been developed, some of which are in different clinical trials [9–17]. However, side effects and resistance of biological antibodies as well as chemotherapeutic reagents are frequently observed in clinical trials, which severely restrict their use.

Natural products have been widely used for the treatment of various cancers, cardiovascular diseases and renal diseases for many years [18–20]. It was reported that 46% of drugs approved by food and drug administration are derived from natural small molecules from 1981 to 2014 [21]. Reportedly, many natural small molecules from natural products have many effective pharmacological activities, including anti-cancer, anti-bacterial, anti-fibrotic and anti-oxidant activities, which fueling considerable enthusiasm for natural products as a source of pharmaceutical exploitation [19,22–29]. As thus, we posited that targeting the Wnt/β-catenin pathway with natural small molecules may

Abbreviations: APC, adenomatous polyposis coli; Axin, axis inhibition protein; AKT, protein kinase B; β-TrCP, β-transducin repeats containing protein; BPA, bisphenol A; c-Myc, cellular homologue of myelocytomatosis viral oncogene; CK1, casein kinase 1; Cyclin D1, cell cycle regulator D1; CKD, chronic kidney disease; Dkk, dickkopf; FZD, Frizzled; GSK3β, glycogen synthase kinase 3β; HCT, human colon cancer tissue; LRP, low-density lipoprotein receptor-related proteins; LEF, lymphoid enhancing factor; PC, *Poria cocos*; RNF43, ring finger protein 43; ROR2, receptor tyrosine kinase-like orphan receptor 2; R-Spondin, roof plate-specific spondin; Runx2, runt-related transcription factor 2; TCF, T cell factor; TGF-β1, transforming growth factor β1; TNBC, triple-negative breast cancer

* Corresponding author.

** Corresponding author at: School of Pharmacy, Faculty of Life Science & Medicine, Northwest University, Xi'an, Shaanxi, 710069, China.

E-mail addresses: masc@nifdc.org.cn (S.-C. Ma), zyy@nwu.edu.cn, zhaoyybr@163.com (Y.-Y. Zhao).

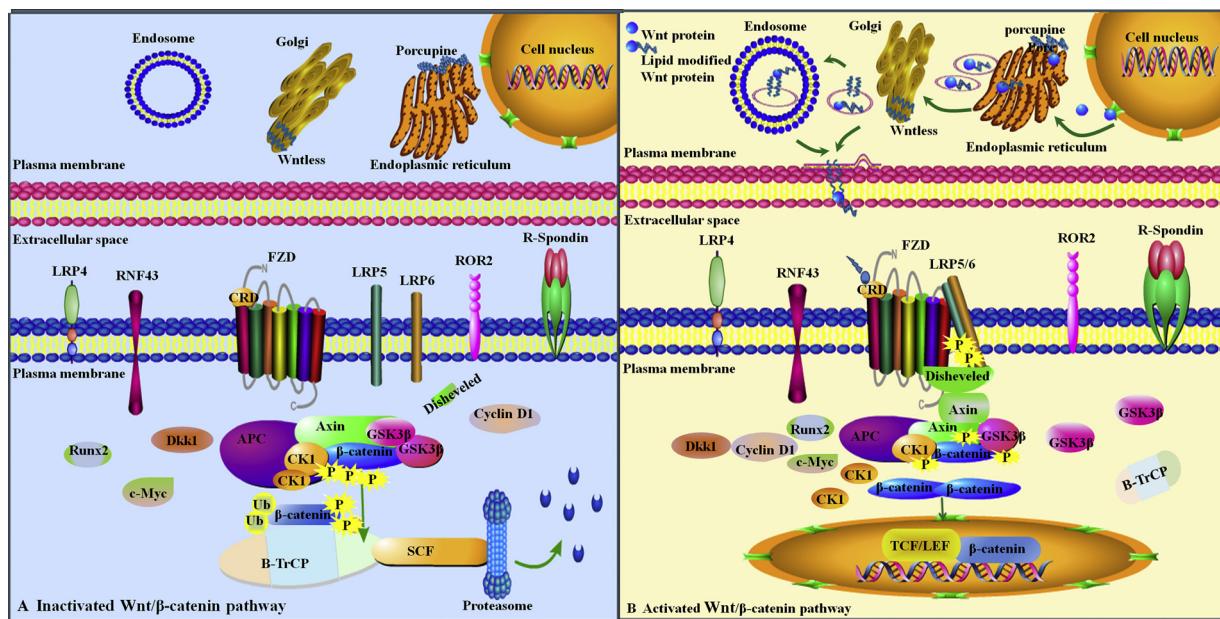


Fig. 1. Inactivated and activated Wnt/β-catenin signaling. (A) Inactivated Wnt/β-catenin signals. The destruction complex in the cytoplasm mainly consists of CK1, GSK3 β , Axin, APC, protein phosphatase 2A, and β-TrCP. When Wnt is absent, CK1 and GSK3 β phosphorylate β-catenin. The β-TrCP ubiquitinates phosphorylated β-catenin, and the latter is finally degraded by the proteasome. β-TrCP is a protein included in F-box. SCF is a complex of Skp, Cullin and F-box. (B) Activated Wnt signaling. In the presence of Wnts, Wnt proteins are lipidated by the porcupine in the endoplasmic reticulum, modified by Wntless in Golgi and ultimately transported to the plasma membrane for secretion. The binding of lipidated Wnts leads to the formation of a receptor complex, FZD and LRP5/6, which promotes the recruitment of Dishevelled to the receptor complex and the phosphorylation of LRP5/6. The Axin interacts with phosphorylated LRP5/6 and the destruction complex takes apart, which protects β-catenin from degradation. Stabilized β-catenin subsequently binds TCF in the nucleus of cells to regulate target genes.

be an effective therapy for identifying promising drug candidates. In this review, we primarily describe the Wnt/β-catenin signaling pathway in the development of multiple diseases such as cancer, renal disease, neurodegenerative disease and bone disorder. Additionally, the underlying mechanisms of small natural products that regulate the Wnt/β-catenin pathway are also highlighted.

2. The Wnt/β-catenin signaling pathway

2.1. Canonical Wnt pathway activation mechanism

In the absence of Wnts, (Fig. 1A), β-catenin in the cytoplasm is phosphorylated by glycogen synthase kinase 3 β (GSK3 β) and casein kinase 1 (CK1), both of which are parts of destruction complex that includes adenomatous polyposis coli (APC), axis inhibition protein (Axin) and β-transducin repeats containing protein (β-TrCP). In addition, phosphorylated β-catenin is ubiquitinated by β-TrCP and ultimately degraded by the proteasome [30–33]. When Wnt is present in the cytoplasm, (Fig. 1B), it will be lipidated by a special palmitoyl transferase-porcupine [34], further modified by Wntless in the Golgi and finally secreted by exocytosis [35,36]. Frizzled (FZD) receptors, the principal receptors for Wnts, consist of seven-transmembrane proteins and cysteine-rich domain (CRD) in the N-terminal [37,38]. Low-density lipoprotein receptor-related proteins (LRPs), the co-receptors of FZDs, are long single-pass transmembrane proteins. Wnt ligands bind to FZD or the CRD to induce the dimerization of FZD and LRP5/6 [4,39], as well as the phosphorylation of LRP5/6. Subsequently, phosphorylated LRP5/6 recruit Axin to membrane, and the destruction complex takes apart, which leads to the stabilization of β-catenin in cytoplasm [40]. After stabilization, β-catenin proteins translocate to the nucleus where they interact with the TCF/lymphoid enhancing factor (LEF) and sequentially activate downstream gene expression [41].

2.2. Relevant proteins and receptors in the Wnt/β-catenin pathway

Except for the above-mentioned proteins, other proteins, such as Wnt4, ring finger protein 43 (RNF43) [42], zinc and ring finger 3 [43], serine/threonine kinases [44], cellular homologue of myelocytomatosis viral oncogene (c-Myc), the cell cycle regulator D1 (cyclin D1) [45], surviving, Mitogen-activated protein kinase 1 as well as CK1e and traf2-and-nck-interacting kinase, have also been discovered to be closely associated with the Wnt signaling pathway, while different proteins appear to play different roles in biochemical signaling mechanisms. Moreover, a series of receptors are also involved in the Wnt/β-catenin pathway. Mitogen-activated protein kinase 1 [46], CK1e and traf2-and-nck-interacting kinase play pivotal roles in the activation of the Wnt/β-catenin pathway in β-catenin-dependent cancer cells. Inactivation of Wnt4 is vital for reproductive development of female mice [47]. Furthermore, Planutis *et al.* discovers that many transmembrane receptors, such as FZD1, FZD2, LRP4, roof plate-specific spondin (R-Spondin) and receptor-like tyrosine kinase/receptor tyrosine kinase-like orphan receptor 2 (ROR2), are also implicated in Wnt/β-catenin pathway [48]. However, all of corresponding signals output fully depend on the relative affinities between Wnt ligands and its receptors [49]. For instance, Wnt5a stimulates the stabilization of β-catenin target proteins by binding to FZD and LRP, while Wnt5a inhibits the β-catenin-dependent pathway via combining with ROR2. In addition, ROR2/planar cell polarity (PCP) autocrine signaling is activated when Wnt-8a binds to ROR2 [50].

3. Wnt/β-catenin pathway, diseases, natural small molecules

3.1. Cancers

3.1.1. Colon cancer

Colon cancer has high morbidity and mortality, representing the third leading cancer in men and the second leading cancer in women globally, with 1.2 million new cases and 600 000 deaths per year [51].

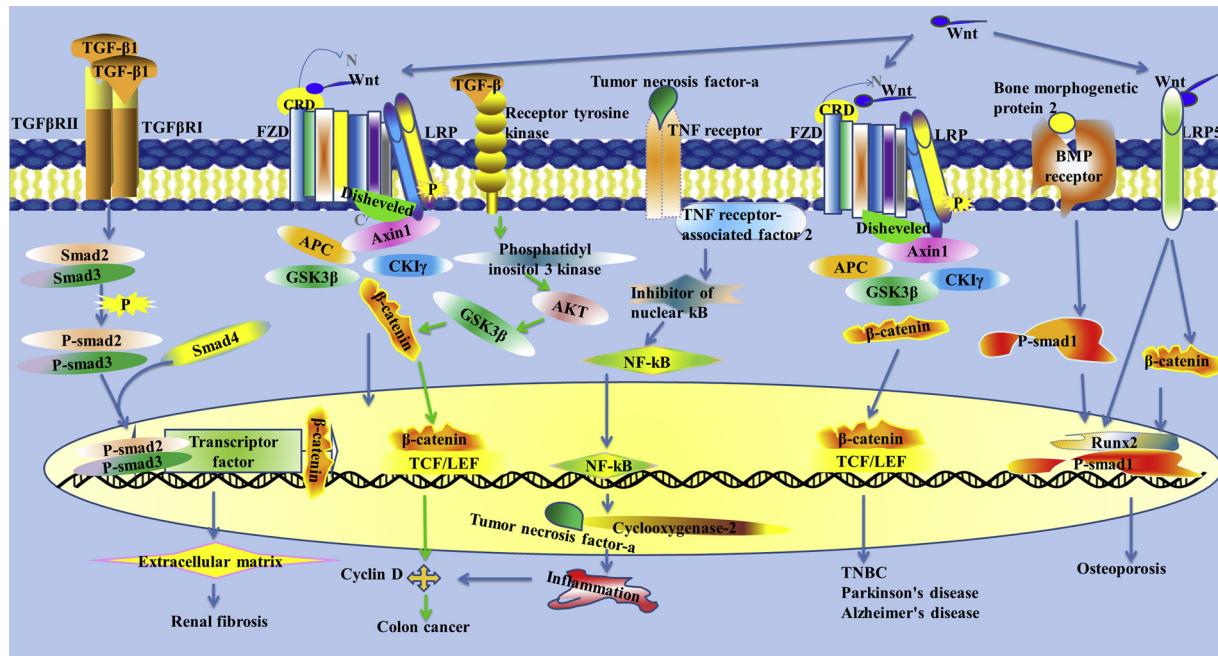


Fig. 2. Crosstalk between the Wnt/β-catenin pathway and TGF-β/Smad in renal fibrosis, phosphatidyl inositol 3 kinase/AKT and NF-κB in colon cancer, and bone morphogenetic protein 2/Smad in osteoporosis.

Table 1

The intervention effect of natural small molecules targeting Wnt/β-catenin pathway in diseases.

Diseases	Compound(s)	Therapeutic mechanisms	Reference(s)
Colon cancer	Ursolic acid	Downregulating β-catenin and TCF-dependent gene expression	[59]
	Corosolic acid		
	Toosendanin	Inhibiting AKT/GSK3β/β-catenin pathway	[60]
	Triptonide	Inhibiting Wnt/β-catenin pathway	[61]
	Silibinin	Downregulating β-catenin and cyclin D 1 expression	[63]
	Quercetin	Downregulating c-Myc and cyclin D1 expression	[62]
	Apigenin	Inhibiting β-catenin/TCF/LEF pathway	[64]
	Berberine	Downregulating β-catenin expression	[65]
	Tetrandrine	Promoting β-catenin degradation and c-Myc downregulation	[66]
	Crocin	Downregulating FZD7 mRNA expression	[83]
Breast cancer	Triptolide	Inhibiting Wnt/β-catenin pathway	[86]
	Poricoic acid (ZC, ZD, ZE, ZG and ZH)	Inhibiting Wnt/β-catenin pathway	[116,117]
Renal disease	25-O-methylalisol F	Inhibiting Wnt/β-catenin pathway	[123]
	Curcumin	Inhibiting Wnt/β-catenin pathway	[128]
	Harpagoside	Upregulating β-catenin, cyclin D1 and c-Myc expression and downregulating Dkk1 expression	[163]
Osteoporosis	Dioscin	Inhibiting AKT/GSK3/β-catenin pathway	[140]
	Wedelolactone	Upregulating phosphorylated GSK3β and Runx2 expression and enhancing β-catenin nuclear translocation	[141]
	Kirenol	Upregulating β-catenin, LRP5, disheveled2, Runx2 and p-GSK3β expression	[142]
Rheumatoid arthritis	Astragaloside I	Upregulating β-catenin and Runx2 expression	[143]
	Icarin	Upregulating Runx2 expression and enhancing β-catenin nuclear translocation	[144]
	Tricin	Upregulating Wnt3a expression and downregulating GSK3β expression	[146]
	Resveratrol	Downregulating Wnt5a expression	[154]
Hair growth	Epigallocatechin-3-gallate	Inhibiting Wnt/β-catenin pathway	[155]
	Ginsenoside F2	Upregulating β-catenin and LEF-1 expression and downregulating Dkk1 expression	[157]

A series of signaling pathways contribute to the pathogenesis of colon cancer, such as TGF-β/phosphatidyl inositol 3 kinase/protein kinase B (AKT), NF-κB and Wnt/β-catenin signaling cascade (Fig. 2), of which the Wnt/β-catenin pathway is dedicated contributor of colon cancer since the discovery of APC gene mutations [52]. Emerging studies have shown that the APC/β-catenin interaction, APC dysfunction [53] and RNF43 mutations [42,54] exacerbate aberrant Wnt signaling, leading to colon cancer, providing additionally evidence to previous studies. Moreover, R-Spondins and leucine-rich repeat containing G-protein coupled receptors 4–6 modules equally activate the Wnt/β-catenin pathway in various subtypes of colon cancer [43]. Reportedly, yes-associated protein and transcriptional co-activators with PDZ-binding

motifs leave from destruction complex and accumulate in the nucleus, decreasing the survival of patients with colon cancer [55,56]. Furthermore, forkhead box protein O 3a, a transcriptional co-activator of β-catenin, interacts with β-catenin and simultaneously increases its concentration in the nucleus, which promotes local or distant tumour metastases and significantly attenuates the survival of colon cancer patients with stages 3 and 4 [57]. Interestingly, an Axin2 mutation has been recognized as a predisposing factor to colon cancer [58].

Ursolic acid and corosolic acid are natural pentacyclic triterpenoids from various plants, both of which have been clarified as antagonists of the Wnt/β-catenin pathway in colon cancer cells. Intriguingly, a β-hydroxyl group at C-3 and carboxyl group at C-17 in ursolic acid were

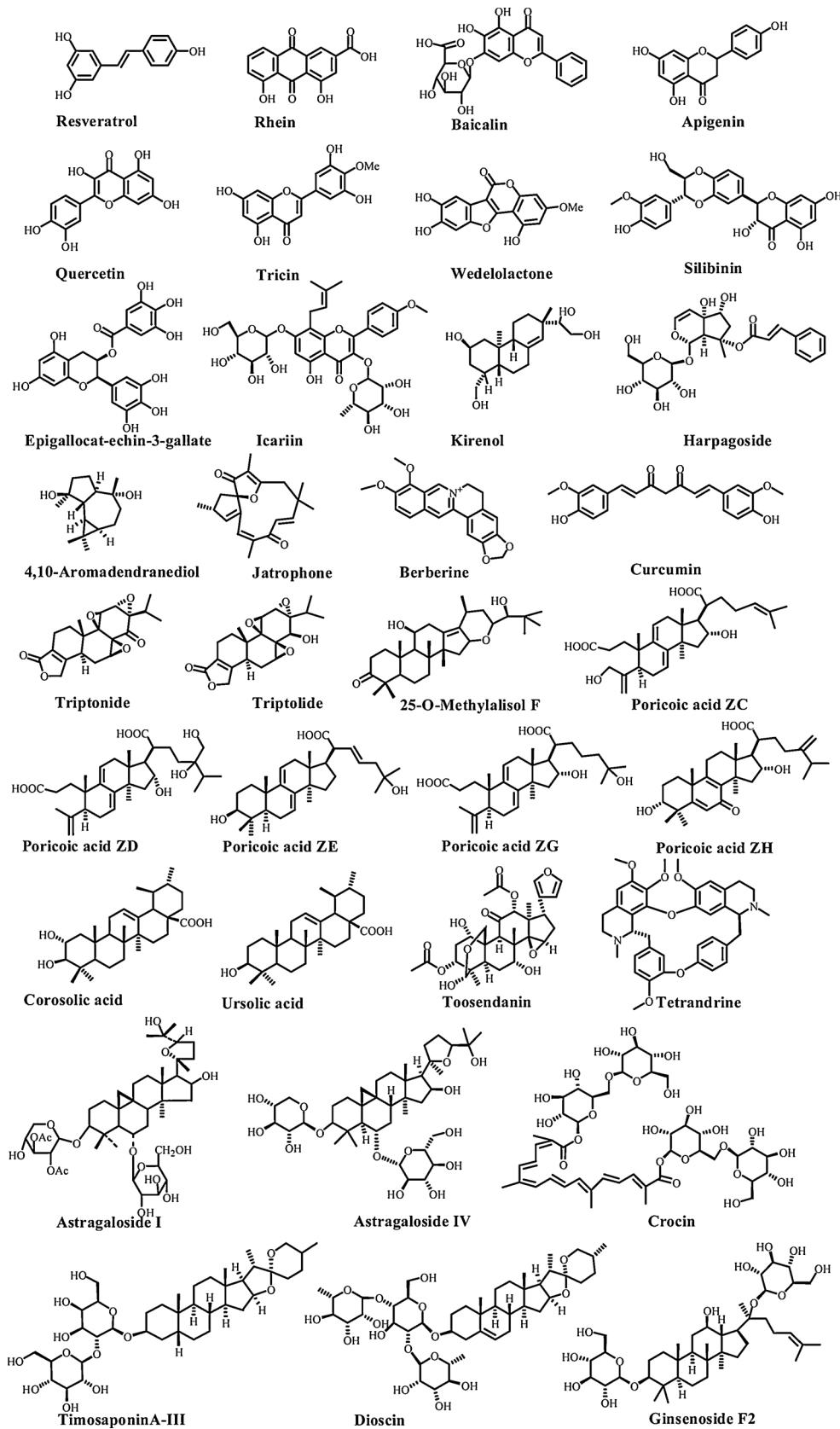


Fig. 3. Structures of natural small molecules from natural products.

previously proven to be cytotoxic to human promyelocytic leukaemia cells (HL-60), gastric cancer cells (BGC) and HeLa cells. Additionally, a carboxyl group at position C-17 and methyl group at position C-19 in

ursolic acid were positively associated with β -catenin degradation and resistance to β -catenin/TCF interaction in ursolic acid-treated human colon cancer tissue 15 cells (HCT15) with APC mutation [59].

Terpenoids, toosendanin and triptonide, possess distinct anti-cancer mechanisms. Toosendanin extracted from the fruits or bark of *Melia toosendan* Sieb et Zucc subverts the AKT/GSK3 β /β-catenin axis [60]. Triptonide in *Tripterygium wilfordii* Hook F. is identified as an inducer of apoptosis in human colon cancer cells (SW480 and RKO) and prostate cancer cell lines (PC3), which benefits from the suppression of triptonide in the Wnt pathway via targeting the C-terminal transcription domain of β-catenin or its nuclear co-factor instead of β-catenin translocation and β-catenin/TCF4 interaction [61]. Silibinin is a flavonolignan extracted from the seeds of milk thistle [62], and downregulated β-catenin as well as cyclin D 1 in polyps demonstrates the mechanism of silibinin against colon cancer in APC $^{\text{min}/+}$ mice [63]. Expression of c-Myc and cyclin D tends to be decreased in cancer stem cells and in HCT116 cells treated with quercetin, a flavonoid compound which is widely found in tea, berries, capers, onions, grapes and apples [62]. The flavonoid apigenin is widely found in fruits (orange, grapes and apples) and vegetables (parsley and onions). Recently, it is reported that apigenin could significantly inhibit proliferation, migration and invasion of colon cancer cells through suppressing activated β-catenin/TCF/LEF signaling cascades in human embryonic kidney 293T cells (HEK293 T) as well as SW480 cells. Moreover, apigenin also restrains β-catenin nuclear translocation in HCT15 cells and SW480 cells activated by LiCl in a concentration-dependent manner [64]. However, unlike ursolic acid, berberine is an isoquinoline alkaline from *Coptis chinensis* and demonstrates inhibitory effect on the expression, instead of the degradation of β-catenin in HCT116 cells [65]. Likewise, tetrrandrine, an alkaloid bis-benzylisoquinoline isolated from the dried root of *Stephanotis tetrandra* S. Moore, confers resistance to proliferation and apoptosis in colon cancer via downregulating IGF binding protein 5 expression, which promotes β-catenin degradation and downregulates c-Myc expression in dimethylhydrazine- and dextran sodium sulphate-induced colorectal cancer LoVo cells [66].

Curcumin is a phenolic compound extracted from the rhizome of turmeric (*Curcuma longa*) that is usually used in Asia as an additive, spice and pigment [67–72]. Curcumin treatment suppressed the growth of colon cancer cells through retarding cell proliferation via inhibiting the Wnt/β-catenin pathway rather than by promoting apoptosis in mice [73]. Curcumin treatment also downregulated miR-130a expression, while miR-130a overexpression abolished the anti-tumour activity of curcumin [73].

3.1.2. Endometrial cancer

Endometrial cancer occurs in the endometrium in perimenopausal and postmenopausal women. Emerging evidence indicates that curcumin possesses chemopreventive properties against various cancers [74–76]. It has been reported that curcumin treatment inhibits proliferation and apoptosis of human endometrial carcinoma cells by downregulating expression of the androgen receptor and β-catenin in a concentration- and time-dependent manner [77]. Wnt3a partially nullifies the effects of curcumin on proliferation and apoptosis in human endometrial carcinoma cells, as well as the androgen receptor expression-downregulating effect of curcumin [77]. These findings confirm that curcumin might inhibit different cancers by repressing the Wnt/β-catenin pathway via miR-130a. Hence, the Wnt/β-catenin pathway may represent a new target of curcumin in cancer treatment.

3.1.3. Triple-negative breast cancer

Triple-negative breast cancer (TNBC) is the most metastatic subtype of breast cancer and cannot be overcome by standard therapy. Oestrogen receptor, progesterone receptor and human epidermal growth factor receptor-2 are three major subtypes of breast cancer [78]. Wnt signaling is widely accepted as one of the most common sources for pro-proliferation signaling in TNBC cells, and the FZD7 protein may represent a novel target or biomarker for TNBC treatment [79]. XAV939 and clofazimine have been identified as Wnt inhibitors and promising anti-TNBC drugs, but neither of them have yet been used in

advanced clinical trials [80,81]. Fortunately, tannins, isolated from *Syzygium guineense*, directly inactivates the stabilization and transcription of Wnt3a-induced β-catenin expression, which may be pursued for novel therapies [82]. As the most potent anti-cancer carotenoids in saffron, crocin exhibits a more potent anti-metastatic ability on TNBC through downregulating FZD7 mRNA expression and upregulating E-cadherin expression in the Wnt/β-catenin pathway [83]. In contrast to crocin, crocetin fails to impart its effects on the Wnt/β-catenin pathway [84]. Proanthocyanidins, isolated from Chinese bayberry leaf, attenuate expression of β-catenin, cyclin D1 as well as c-Myc, and block the G1 cell cycle as well as self-renewal ability in ovarian cancer stem cells (OVCAR-3 SP) [85]. Additionally, triptolide, isolated from *Tripterygium wilfordii* Hook F., has an inhibitory effect on myriad cancers, such as liver cancer, non-small cell lung carcinoma, osteosarcoma, pancreatic cancer as well as breast cancer, and has the capacity to remodel the activated Wnt/β-catenin pathway [86]. Taken together, natural products play an important role in various cancer interventions by regulating the Wnt/β-catenin pathway and Wnt/β-catenin signaling may represent a specific therapeutic target of natural small molecules against cancers.

3.2. Renal disease and fibrosis

Chronic kidney disease (CKD) is an epidemic that has increased by 73% from 1990 to 2013 and represents a primary cause of death worldwide. Renal fibrosis, characterized by tubulointerstitial fibrosis and glomerulosclerosis, is an inevitable outcome and final manifestation of all kinds of progressive CKD [87–91]. Renal fibrosis is closely associated with mounting mediators and signaling pathways, such as oxidative stress, inflammation, microRNAs, transforming growth factor β1 (TGF-β1)/Smad and Wnt/β-catenin (Fig. 2), as well as the dysregulated uremic toxins, amino acids and lipid metabolism [92–100]. In recent decades, natural small molecules have gradually become an important therapeutic strategy for the prevention and treatment of renal fibrosis worldwide [22,101,102]. Emerging studies suggest that small molecular compounds isolated from natural products exert good therapeutic effects on renal disease and fibrosis [103–106]. A number of traditional Chinese medicines, such as *Poria cocos* (PC), *Alismatis rhizoma* and *Polyporus umbellatus*, promote urination and eliminate oedema, both of which are associated with the retardation of renal disease and fibrosis.

PC grows around the roots of pine trees in Asia and North America [107]. PC and its surface layer have diuretic effects [108,109], anti-hyperlipidemic activity [110,111] and CKD treatment potential [112–115]. Our study reported that poricoic acid ZG and poricoic acid ZH, isolated from the surface layer of PC, significantly ameliorated the upregulated expression of Wnt1 and β-catenin, as well as its target gene expression, including Snail1, Twist, matrix metalloproteinase 7, plasminogen activator inhibitor 1 and fibroblast specific protein 1 in TGF-β1-induced human kidney proximal epithelial cells (HK-2). Interestingly, poricoic acid ZG and poricoic acid ZH also selectively suppressed Smad3 phosphorylation through alleviating the interactions of smad anchor for receptor activation with TGF-β receptor I and Smad3 in the TGF-β/Smad pathway. Structure-function analysis indicated that the antifibrotic effect was associated with the first six-membered ring structure and the number of carboxyl groups in tetracyclic triterpenoid compounds [116]. In addition, we also obtained three new triterpenoids, including poricoic acid ZC, poricoic acid ZD and poricoic acid ZE, and discovered that they could significantly alleviate extracellular matrix production by suppressing the Wnt/β-catenin pathway and specific Smad3 phosphorylation via blocking the interaction of TGF-β receptor I with Smad3 signaling in TGF-β1 or angiotensin II-induced HK-2 cells and unilateral ureteral obstructive mice [117]. Structure-activity analysis suggested that secolanostane tetracyclic triterpenoid compounds, poricoic acid ZC and poricoic acid ZD, showed a stronger inhibitory effect than lanostane tetracyclic triterpenoid compound

poricoic acid ZE, indicating that compounds with a secolanostane skeleton exhibit stronger bioactivity than those with a lanostane skeleton.

Alismatis rhizome, the dried stem tuber of *Alisma orientale* (Sam.) Juzep., exerts powerful diuretic, anti-hyperlipidemic effects, which may be intimately associated with the anti-fibrotic effect. [118–122]. Triterpenoid compounds are the primary active components in *Alismatis rhizome* [118,123]. 25-O-methylalisol F is a new tetracyclic triterpenoid compound isolated from the *Alismatis rhizome*. Our latest study find that 25-O-methylalisol F inhibits upregulated expression of Wnt1 as well as β -catenin and its target gene expression, including Snail1, Twist, matrix metalloproteinase-7, plasminogen activator inhibitor 1 and fibroblast specific protein 1, in both TGF- β 1-induced HK-2 cells and normal rat kidney interstitial fibroblast cells (NRK-49 F). In addition, 25-O-methylalisol F inhibits Smad3 phosphorylation and maintains Smad7 expression in the TGF- β /Smad-dependent pathway, exerting a strong inhibitory effect on crosstalk between Wnt/ β -catenin and TGF- β /Smad pathways in the extracellular matrix [123]. Collectively, the Wnt/ β -catenin pathway may be a specific therapeutic target of natural small molecules against renal fibrosis.

3.3. Neurodegenerative diseases

Parkinson's disease is the second most common neurodegenerative disease, affecting approximately 1% of the population aged 65 or older worldwide [124]. Emerging management of Parkinson's disease has primarily focused on inhibitor medications, such as levodopa, dopamine agonists, monoamine oxidase type B and catechol O-methyltransferase inhibitors. In addition, surgery, rehabilitation and palliative care are used for Parkinson's disease treatment as well [125]. The causes of Parkinson's disease are attributed to age, low serum urate concentrations, smoking, α -synuclein mutations, leucine-rich repeat kinase 2, phosphatase, tension homolog-induced putative kinase 1, parkin 7, vacuolar protein sorting 35, receptor-mediated endocytosis 8 as well as coiled-coil-helix-coiled-coiled-helix domain containing 2 [124] and eukaryotic translation initiation factor 4- γ [126]. In addition, leucine-rich repeat kinase 2 mutations also exacerbate disease progression by activating the Wnt/ β -catenin pathway in adult mice and cultured fibroblasts [127]. It has been reported that curcumin isolated from the rhizome of turmeric (*Curcuma longa*) exhibits neuro-protective effects and attenuates bisphenol A (BPA)-induced neurotoxicity through inhibiting activated Wnt/ β -catenin signaling, verified by the use of Wnt specific activators, such as LiCl, GSK3 β siRNA and inhibitor dickkopf (Dkk) 1. Curcumin treatment significantly reverses BPA-mediated increased β -catenin phosphorylation, downregulated GSK3 β expression and β -catenin nuclear translocation in neural stem cells [128]. Meanwhile, neurogenesis as well as learning and memory in BPA-treated rats were improved in response to curcumin [128]. These data indicate that curcumin exhibits neuroprotection against BPA-mediated impaired neurogenesis through depressing activated Wnt/ β -catenin signaling.

Neurogenesis, the process of generating new neurons, is reduced in several neurodegenerative disorders including Alzheimer's disease. It has been reported that curcumin nanoparticles improve neuronal differentiation via decreasing the expression of GSK3 β , with enhanced β -catenin nuclear translocation and promoter activity of the TCF/LEF and cyclin D1 [129]. Treatment with curcumin nanoparticles further reverses learning and memory impairments by inducing neurogenesis in an amyloid β -induced rat model of Alzheimer's disease [129]. Moreover, molecular docking studies indicates that curcumin interacts with Wif-1, Dkk, and GSK3 β . Collectively, these findings demonstrate that curcumin treatment induces adult neurogenesis via activating the Wnt/ β -catenin pathway and improving the brain's self-repair mechanisms, which might represent a therapeutic intervention for neurodegenerative diseases, such as Parkinson's and Alzheimer's. Therefore, the Wnt/ β -catenin pathway may be exploited for a specific therapeutic target of natural small molecules in the treatment of neurodegenerative disease.

3.4. Bone disorders

The balance between osteoblasts and osteoclasts plays significant roles in bone formation and maintenance, the loss of which results in bone disorders, including osteoporosis [130]. As an age-dependent metabolic bone disorder, osteoporosis has two main characteristics, low bone mass and micro-architectural deterioration of bone tissue [131]. Unfortunately, current therapeutic strategies for osteoporosis are mainly focused on physical exercise and medications, such as bisphosphonates, teriparatide, strontium ranelate and denosumab [132]. Except for the pathogenic factors, the occurrence of osteoporosis is attributed to both non-modifiable risk factors, such as gender, race, heredity, age, and modifiable risk factors, such as medical disorders, hypogonadal states, endocrine disorders, vitamin D deficiency and long-term intake of proton pump inhibitors [133,134]. Emerging evidence suggests that bone morphogenetic protein 2/Smad and Wnt/ β -catenin pathways (Fig. 2) are associated with osteoporosis [135–137]. Enhanced β -catenin phosphorylation and GSK3 β in senile osteoporosis indicate that the Wnt/ β -catenin pathway is a possible therapeutic target in the treatment of osteoporosis [138].

Iridoid glycoside harpagoside, a major bioactive component of the radix of *Harpagophytum procumbens* var. *sublobatum* (Engl.) Stapf (Pedaliaceae), exhibits analgesic, anti-inflammatory, anti-phlogistic and anti-osteoporotic effects. Harpagoside treatment induces osteoblast differentiation via upregulating the expression of β -catenin, cyclin D1 as well as c-Myc and downregulating Dkk1 expression [139]. *In vitro* and *in vivo* experiments demonstrate that the AKT/GSK3/ β -catenin axis in osteosarcoma is probably inhibited by steroidal saponin dioscin, a major compound of Liuwei Dihuang decoction and Di'ao Xinxue kang [140].

As a derivative of coumarin, wedelolactone, isolated from *Eclipta herba* with the capability of nourishing bones, enhances osteoblastogenesis in mouse bone marrow mesenchymal stem cells through enhancing β -catenin nuclear translocation and increasing levels of phosphorylated GSK3 β protein as well as runt-related transcription factor 2 (Runx2) protein in the Wnt/GSK3 β / β -catenin pathway [141]. Additionally, it is demonstrated that the terpenoids kirenel, astragaloside I and flavone icariin have similar effects on the Wnt/ β -catenin pathway and osteoblastic differentiation. Diterpenoid kirenel isolated from *Herba Siegesbeckiae* promotes osteoblast differentiation in mouse osteoid cell lines (MC3T3-E1) and accelerates the upregulation of LRP5, disheveled2, Runx2, p-GSK3 β as well as β -catenin in the Wnt/ β -catenin pathway, contributing to osteoblast differentiation [142]. Upregulated expression of β -catenin and Runx2 has also been shown in MC3T3-E1 cells treated with Astragaloside I isolated from *Astragalus membranaceus* [143], which possesses osteogenic properties and exhibits partially similar effects as icariin on the Wnt/ β -catenin pathway with respect to osteoblastic differentiation. Icariin is extracted from *Epimedium brevicornum Maxim*, which promotes proliferation, differentiation and mineralization of osteoblasts in MC3T3-E1 cells via enhancing nuclear translocation of β -catenin and upregulating the mRNA and protein expression of Runx2 [144]. Except for increased β -catenin and Runx2, the expression of cyclin D1 and alkaline phosphatase is elevated in icariin-treated osteoblastic cells from rat mandible [145]. In addition to icariin, flavone tricin, rich in rice bran or other grass species, enhances osteoblastogenesis through upregulating Wnt3a expression while downregulating GSK3 β expression in human adult mesenchymal stem cells [146]. Taken together, the Wnt/ β -catenin pathway may be an attractive therapeutic target of natural small molecules for the treatment of bone disorders.

3.5. Other diseases

Rheumatoid arthritis is a chronic inflammatory illness that displays painful swelling or inflammation of the synovial lining in the joints and cartilage, as well as ultimate bone damage [147]. Mutations in CD28

and CD40 or genetic variants of LRP5 in the Wnt/β-catenin pathway are risk factors for bone damage in patients with rheumatoid arthritis [148]. Both upregulated expression of Wnt5a or β-catenin play prominent roles in the Wnt/β-catenin pathway in cultured fibroblast-like synoviocytes from rheumatoid arthritis patients [149]. Fortunately, newly emerging anti-sclerostin therapies (romosozumab and bloszumab) and several natural small molecules have shown a beneficial effect on the treatment of rheumatoid arthritis through targeting the Wnt/β-catenin pathway. Resveratrol, a natural polyphenolic compound isolated from plants, has various physiological effects, including anti-cancer and anti-cardiovascular, and is expected to become a promising natural product for clinical use, despite most related studies being performed in animal models [150–153]. In collagen-induced arthritis mice, levels of Wnt5a protein are decreased and the Wnt/β-catenin pathway is suppressed by resveratrol [154]. Similar to resveratrol, curcumin and epigallocatechin-3-gallate isolated from green tea suppress arthritis and have an inhibitory effect on the Wnt/β-catenin pathway [155,156]. Intriguingly, as a metabolite of ginsenoside-Rb1, ginsenoside F2 positively regulates the anagen phase and hair growth through upregulating expression of β-catenin as well as LEF-1 and downregulating expression of Dkk1 [157].

In addition, emerging studies have demonstrated that mutations in Wnt/β-catenin signals are closely linked to multiple diseases, such as synovial sarcoma, type II diabetes and myocardial fibrosis. Aberrant Wnt signaling and added β-catenin stabilization have been termed secondary changes in synovial sarcoma, but effective therapy for the treatment of synovial sarcoma remains to be determined [158], encouraging further research to block synovial sarcoma tumour formation by inhibiting the Wnt/β-catenin pathway. Mutations in Wnt5b and transcription factor 7-like 2 are observed in type II diabetes [159]. Downregulated Wnt5a, FZD-related proteins and Dkk1 are discovered in mutant pancreatic mesenchyme [160] and Wnt/β-catenin pathway is activated in intervertebral disc cells [161], which is highly consistent with previous findings. Additionally, overexpression of Wnt1 and Wnt5a or the inhibitory activities of Wnt-C59 on Wnts are all evidence of the critical role of the Wnt/β-catenin pathway in myocardial fibrosis progression and myofibroblast formation [162]. Unfortunately, thus far, no effective natural small molecules have been applied to treat these diseases that act through the Wnt/β-catenin pathway. Thereby, these studies might provide a new avenue for natural small molecules targeting the Wnt/β-catenin pathway in various diseases.

4. Concluding remarks

In summary, this systemic review examined the intervention of natural small molecules on Wnt/β-catenin pathway with respect to initiation and progression of cancers, renal diseases, neurodegenerative diseases and bone disorders. Through elaborating on the mechanisms of Wnt/β-catenin pathway using *in vitro* and *in vivo* experiments, further insights into these mechanisms enable in-depth understanding of the pathophysiology of those diseases and facilitate discovery of targeted therapeutic agents.

After a pooled analysis of clinical and non-clinical natural products, chemotherapeutic drugs and biological agents against the Wnt/β-catenin pathway, we learned that intracellular porcupine, extracellular Wnt ligands as well as their receptors, and β-catenin-dependent proteins, have become major targets of antagonists. Clinical trials have shown that porcupine is a target of a novel inhibitor, ETC-159, with poor oral bioavailability. Wnt3a is regarded as a target of microRNA-15a-5p and microRNA-195 in colon cancer. LRP6 and FZD7/FZD8-Fc are concurrently thought to be targets of sclerostin and nigericin. Moreover, β-catenin expression is regulated by natural berberine and sulphoraphane, while β-catenin-dependent proteins are targeted by natural compounds, such as epigallocatechin-3-gallate, triptonide, ursolic acid and toosendanin. In addition, interactions between β-catenin and APC, TCF or CBP are inhibited by cercosporin, baicalein and ICG-

001, respectively. Unfortunately, with further research on antagonists, a series of side effects have been discovered, which severely restrict their use. For example, R-Spondins inhibitor causes differentiation of colon tumour cells and loss of stem cell function. Long-term users of the GSK3 inhibitor lithium are diagnosed with a significantly higher incidence of renal cancers. As such, it is imperative to seek safe and effective targeted therapeutics to resolve the prominent problems that continue to puzzle the biomedical community.

With the in-depth understanding of the mechanisms of aberrant Wnt/β-catenin pathway in diseases, seeking targeted agents against mutations in Wnt/β-catenin signals have drawn increasing attentions. Of note, rich natural resources, together with advanced biotechnology, have also offered increasing possibilities for the development of novel drugs and targeted agents. Excluding the natural small molecules mentioned above (Table 1), natural rhein and baicalin, along with natural terpenoids (timosaponin AIII, astragaloside IV, jatropheone and 4,10-aromadendranediol), have also been investigated and confirmed as promising compounds against the mutant Wnt/β-catenin pathway (Fig. 3), which greatly inspires us to identify more targeted compounds from nature. Notably, the generation of targeted natural small molecules not only improves defects of existing therapeutic agents but also provides opportunities for the treatment of diseases, such as the type II diabetes, synovial sarcoma and myocardial fibrosis.

Conflict of interest

No potential conflict of interests exists.

Acknowledgment

This study was supported by Shaanxi Science and Technology Plan Project (2019ZDLSF04-04-02) and National Natural Science Foundation of China (Nos. 81673578, 81603271).

References

- [1] M. Kahn, Can we safely target the Wnt pathway? *Nat. Rev. Drug Discov.* 13 (7) (2014) 513–532.
- [2] H.W. Park, Y.C. Kim, B. Yu, T. Moroishi, J.S. Mo, S.W. Plouffe, Z.P. Meng, K.C. Lin, F.X. Yu, C.M. Alexander, C.Y. Wang, K.L. Guan, Alternative Wnt signalling activates YAP/TAZ, *Cell* 162 (4) (2015) 780–794.
- [3] L.T. Vuong, C. Iomini, S. Balmer, D. Esposito, S.A. Aaronson, M. Mlodzik, Kinesin-2 and IFT-A act as a complex promoting nuclear localization of β-catenin during Wnt signalling, *Nat. Commun.* 9 (1) (2018) 5304.
- [4] K. Zhang, Y. Zhang, L. Gu, M. Lan, C. Liu, M. Wang, Y. Su, M. Ge, T. Wang, Y. Yu, C. Liu, L. Li, Q. Li, Y. Zhao, Islr regulates canonical Wnt signalling-mediated skeletal muscle regeneration by stabilizing Dishevelled-2 and preventing autophagy, *Nat. Commun.* 9 (1) (2018) 5129.
- [5] D.Q. Chen, G. Cao, H. Chen, D. Liu, W. Su, X.Y. Yu, N.D. Vaziri, X.H. Liu, X. Bai, L. Zhang, Y.Y. Zhao, Gene and protein expressions and metabolomics exhibit activated redox signalling and Wnt/β-catenin pathway are associated with metabolite dysfunction in patients with chronic kidney disease, *Redox Biol.* 12 (2017) 505–521.
- [6] A.H. Wagner, S. Devarakonda, Z.L. Skidmore, K. Krysiak, Recurrent Wnt pathway alterations are frequent in relapsed small cell lung cancer, *Nat. Commun.* 9 (1) (2018) 3787.
- [7] X. Chen, B. Zhou, T. Yan, H. Wu, J. Feng, H. Chen, C. Gao, T. Peng, D. Yang, J. Shen, Peroxynitrite enhances self-renewal, proliferation and neuronal differentiation of neural stem/progenitor cells through activating HIF-1α and Wnt/β-catenin signalling pathway, *Free Radic. Biol. Med.* 117 (2018) 158–167.
- [8] K. Hubner, P. Cabochette, R. Dieguez-Hurtado, C. Wiesner, Y. Wakayama, K.S. Grassme, M. Hubert, S. Guenther, H.G. Belting, Wnt/β-catenin signalling regulates VE-cadherin-mediated anastomosis of brain capillaries by counteracting S1pr1 signalling, *Nat. Commun.* 9 (1) (2018) 4860.
- [9] J. Cai, L. Fang, Y. Huang, R. Li, X. Xu, Z. Hu, L. Zhang, Y. Yang, X. Zhu, H. Zhang, J. Wu, Y. Huang, J. Li, M. Zeng, E. Song, Y. He, L. Zhang, M. Li, Simultaneous overactivation of Wnt/β-catenin and TGF/β signalling by miR-128-3p confers chemoresistance-associated metastasis in NSCLC, *Nat. Commun.* 8 (2017) 15870.
- [10] J. Li, B. Yu, P. Deng, Y. Cheng, Y. Yu, K. Kevork, S. Ramadoss, X. Ding, X. Li, C.Y. Wang, KDM3 epigenetically controls tumorigenic potentials of human colorectal cancer stem cells through Wnt/β-catenin signalling, *Nat. Commun.* 8 (2017) 15146.
- [11] J. Wan, X. Hou, Z. Zhou, J. Geng, J. Tian, X. Bai, J. Nie, WT1 ameliorates podocyte injury via repression of EZH2/β-catenin pathway in diabetic nephropathy, *Free Radic. Biol. Med.* 108 (2017) 280–299.
- [12] S. Singh, A. Mishra, S.J. Mohanbai, V. Tiwari, R.K. Chaturvedi, S. Khurana, S. Shukla, Axin-2 knockdown promote mitochondrial biogenesis and

- dopaminergic neurogenesis by regulating Wnt/β-catenin signaling in rat model of Parkinson's disease, *Free Radic. Biol. Med.* 129 (2018) 73–87.
- [13] J.A. Schneider, T.W. Craven, A.C. Kasper, C. Yun, M. Haugbro, E.M. Briggs, V. Svetlov, E. Nudler, H. Knaut, R. Bonneau, M.J. Garabedian, K. Kirshenbaum, S.K. Logan, Design of peptoid-peptide macrocycles to inhibit the β-catenin TCF interaction in prostate cancer, *Nat. Commun.* 9 (1) (2018) 4396.
- [14] L. Mariotti, K. Pollock, Regulation of Wnt/β-catenin signalling by tankyrase-dependent poly(ADP-ribosylation)ation and scaffolding, *Br. J. Pharmacol.* 174 (24) (2017) 4611–4636.
- [15] S.H. Lee, S.M. Shin, P. Zhong, H.T. Kim, D.I. Kim, J.M. Kim, W. Do Heo, D.W. Kim, C.Y. Yeo, C.H. Kim, Reciprocal control of excitatory synapse numbers by Wnt and Wnt inhibitor PRR7 secreted on exosomes, *Nat. Commun.* 9 (1) (2018) 3434.
- [16] Y. Sassi, P. Avramopoulos, Cardiac myocyte miR-29 promotes pathological remodeling of the heart by activating Wnt signaling, *Nat. Commun.* 8 (1) (2017) 1614.
- [17] C. Lv, F. Li, X. Li, Y. Tian, Y. Zhang, X. Sheng, Y. Song, Q. Meng, S. Yuan, L. Luan, T. Andl, X. Feng, B. Jiao, MiR-31 promotes mammary stem cell expansion and breast tumorigenesis by suppressing Wnt signaling antagonists, *Nat. Commun.* 8 (1) (2017) 1036.
- [18] D.Q. Chen, H.H. Hu, Y.N. Wang, Y.L. Feng, G. Cao, Y.Y. Zhao, Natural products for the prevention and treatment of kidney disease, *Phytomedicine* 50 (2018) 50–60.
- [19] A.R. Guerra, M.F. Duarte, Targeting tumor metabolism with plant-derived natural products: emerging trends in cancer therapy, *J. Agric. Food Chem.* 66 (41) (2018) 10663–10685.
- [20] D.Q. Chen, G. Cao, H. Chen, C.P. Argyopoulos, H. Yu, W. Su, L. Chen, D.C. Samuels, S. Zhuang, G.P. Bayliss, S. Zhao, X.Y. Yu, N.D. Vaziri, M. Wang, D. Liu, J.R. Mao, S.X. Ma, J. Zhao, Y. Zhang, Y.Q. Shang, H. Kang, F. Ye, X.H. Cheng, X.R. Li, L. Zhang, M.X. Meng, Y. Guo, Y.Y. Zhao, Identification of serum metabolites associating with chronic kidney disease progression and anti-fibrotic effect of 5-methoxytryptophan, *Nat. Commun.* 10 (1) (2019) 1476.
- [21] D.J. Newman, G.M. Cragg, Natural products as sources of new drugs from 1981 to 2014, *J. Nat. Prod.* 79 (3) (2016) 629–661.
- [22] D.Q. Chen, Y.L. Feng, G. Cao, Y.Y. Zhao, Natural products as a source for anti-fibrosis therapy, *Trends Pharmacol. Sci.* 39 (11) (2018) 937–952.
- [23] L. Chen, T. Yang, D.W. Lu, H. Zhao, Y.L. Feng, H. Chen, D.Q. Chen, N.D. Vaziri, Y.Y. Zhao, Central role of dysregulation of TGF-β/Smad in CKD progression and potential targets of its treatment, *Biomed. Pharmacother.* 101 (2018) 670–681.
- [24] M.G. Moloney, Natural products as a source for novel antibiotics, *Trends Pharmacol. Sci.* 37 (8) (2016) 689–701.
- [25] H. Chen, G. Cao, D.Q. Chen, M. Wang, N.D. Vaziri, Z.H. Zhang, J.R. Mao, X. Bai, Y.Y. Zhao, Metabolomics insights into activated redox signaling and lipid metabolism dysfunction in chronic kidney disease progression, *Redox Biol.* 10 (2016) 168–178.
- [26] L. Chen, G. Cao, M. Wang, Y.L. Feng, D.Q. Chen, N.D. Vaziri, S. Zhuang, Y.Y. Zhao, The matrix metalloproteinase-13 inhibitor poricoic acid ZI ameliorates renal fibrosis by mitigating epithelial-mesenchymal transition, *Mol. Nutr. Food Res.* (2019) e1900132.
- [27] B. Dai, T. Yang, X. Shi, N. Ma, Y. Kang, J. Zhang, Y. Zhang, HMQ-T-F5 (1-(4-(2-aminoquinazolin-7-yl)phenyl)-3-(2bromo-5-(trifluoromethoxy)phenyl)thiourea) suppress proliferation and migration of human cervical HeLa cells via inhibiting Wnt/β-catenin signaling pathway, *Phytomedicine* 51 (2018) 48–57.
- [28] X. Shi, M. Zhu, Y. Kang, T. Yang, X. Chen, Y. Zhang, Wnt/β-catenin signaling pathway is involved in regulating the migration by an effective natural compound brucine in LoVo cells, *Phytomedicine* 46 (2018) 85–92.
- [29] W.K. Kim, D.H. Bach, H.W. Ryu, J. Oh, H.J. Park, J.Y. Hong, H.H. Song, S. Eum, T.T. Bach, S.K. Lee, Cytotoxic activities of *Telecastadium dongnaiense* and its constituents by inhibition of the Wnt/β-catenin signaling pathway, *Phytomedicine* 34 (2017) 136–142.
- [30] R. Nusse, H. Clevers, Wnt/β-catenin signaling, disease, and emerging therapeutic modalities, *Cell* 169 (6) (2017) 985–999.
- [31] A. Kazi, S. Xiang, H. Yang, D. Delitto, J. Trevino, R.H.Y. Jiang, M. Ayaz, H.R. Lawrence, P. Kennedy, S.M. Sebiti, GSK3 suppression upregulates β-catenin and c-Myc to abrogate KRas-dependent tumors, *Nat. Commun.* 9 (1) (2018) 5154.
- [32] J. Becker, J. Wilting, Wnt signaling, the development of the sympathetic-adrenomedullary system and neuroblastoma, *Cell. Mol. Life Sci.* 75 (6) (2018) 1057–1070.
- [33] C.M. Karner, F. Long, Wnt signaling and cellular metabolism in osteoblasts, *Cell. Mol. Life Sci.* 74 (9) (2017) 1649–1657.
- [34] L.S. Zhang, L. Lum, Chemical modulation of Wnt signaling in cancer, *Prog. Mol. Biol. Transl. Sci.* 153 (2018) 245–269.
- [35] K.M. Cadigan, Wnt/β-catenin signaling, *Curr. Biol.* 18 (20) (2008) R943–R947.
- [36] I.J. McGough, R.E.A. de Groot, A.P. Jellett, M.C. Betist, K.C. Varandas, C.M. Danson, K.J. Heesom, H.C. Korswagen, P.J. Cullen, SNX3-retromer requires an evolutionary conserved MON2:DOPEY2:ATP9A complex to mediate Wntless sorting and Wnt secretion, *Nat. Commun.* 9 (1) (2018) 3737.
- [37] H. Clevers, R. Nusse, Wnt/β-catenin signaling and disease, *Cell* 149 (6) (2012) 1192–1205.
- [38] V. Murillo-Garzon, I. Gorrono-Etxebarria, M. Akerfelt, M.C. Puustinen, L. Sistonen, M. Nees, J. Carton, J. Waxman, R.M. Kypta, Frizzled-8 integrates Wnt-11 and transforming growth factor-β signaling in prostate cancer, *Nat. Commun.* 9 (1) (2018) 1747.
- [39] S. Angers, R.T. Moon, Proximal events in Wnt signal transduction, *Nat. Rev. Mol. Cell Biol.* 10 (7) (2009) 468–477.
- [40] A. Voronkov, S. Krauss, Wnt/β-catenin signaling and small molecule inhibitors, *Curr. Pharm. Des.* 19 (4) (2013) 634–664.
- [41] Q. Xiao, J.B. Wu, W.J. Wang, S.Y. Chen, Y.X. Zheng, X.Q. Yu, K. Meeth, M. Sahraei, A.L.M. Bothwell, L.P. Chen, M. Bosenberg, J.F. Chen, V. Sexl, L. Sun, L. Li, W.W. Tang, D.Q. Wu, Dkk2 imparts tumor immunity evasion through β-catenin-independent suppression of cytotoxic immune-cell activation, *Nat. Med.* 24 (3) (2018) 262–270.
- [42] B.K. Koo, M. Spit, I. Jordens, T.Y. Low, D.E. Stange, M. van de Wetering, J.H. van Es, S. Mohammed, A.J. Heck, M.M. Maurice, H. Clevers, Tumour suppressor RNF43 is a stem-cell E3 ligase that induces endocytosis of Wnt receptors, *Nature* 488 (7413) (2012) 665–669.
- [43] H.X. Hao, Y. Xie, Y. Zhang, O. Charlat, E. Oster, M. Avello, H. Lei, C. Mickanin, D. Liu, H. Ruffner, X. Mao, Q. Ma, R. Zamponi, T. Bouwmeester, P.M. Finan, M.W. Kirschner, J.A. Porter, F.C. Serluca, F. Cong, ZNRF3 promotes Wnt receptor turnover in an R-Spondin-sensitive manner, *Nature* 485 (7397) (2012) 195–200.
- [44] E.C. van Kappel, M.M. Maurice, Molecular regulation and pharmacological targeting of the β-catenin destruction complex, *Br. J. Pharmacol.* 174 (24) (2017) 4575–4588.
- [45] K. Peng, L. Kou, L. Yu, C. Bai, M. Li, P. Mo, W. Li, C. Yu, Histone demethylase JMJD2D interacts with β-catenin to induce transcription and activate colorectal cancer cell proliferation and tumor growth in mice, *Gastroenterology* 156 (4) (2018) 1112–1126.
- [46] S.S. Ng, T. Mahmoudi, V.S.W. Li, P. Hatzis, P.J. Boersema, S. Mohammed, A.J. Heck, H. Clevers, MAP3K1 functionally interacts with Axin1 in the canonical Wnt signalling pathway, *Biol. Chem.* 391 (2–3) (2010) 171–180.
- [47] S. Vainio, M. Heikkila, A. Kispert, N. Chin, A.P. McMahon, Female development in mammals is regulated by Wnt-4 signalling, *Nature* 397 (6718) (1999) 405–409.
- [48] K. Planutis, M. Planutiene, A.V. Nguyen, M.P. Moyer, R.F. Holcombe, Invasive colon cancer, but not non-invasive adenomas induce a gradient effect of Wnt pathway receptor Frizzled 1 (Fz1) expression in the tumor microenvironment, *J. Transl. Med.* 11 (50) (2013).
- [49] A.K. Patel, K. Surapaneni, H. Yi, R.E. Nakamura, S.Z. Karli, S. Syeda, T. Lee, A.S. Hackam, Activation of Wnt/β-catenin signaling in Muller glia protects photoreceptors in a mouse model of inherited retinal degeneration, *Neuropharmacology* 91 (2015) 1–12.
- [50] B. Mattes, Y. Dang, G. Greicius, L.T. Kaufmann, B. Prunsche, J. Rosenbauer, J. Stegmaier, R. Mikut, S. Ozbek, G.U. Nienhaus, A. Schug, D.M. Virshup, S. Scholpp, Wnt/PCP controls spreading of Wnt/β-catenin signals by cytonemes in vertebrates, *Elife* 7 (2018) e36953.
- [51] R.L. Siegel, K.D. Miller, A. Jemal, *Cancer statistics*, 2015, *CA Cancer J. Clin.* 65 (1) (2015) 5–29.
- [52] I. Nishisho, Y. Nakamura, Y. Miyoshi, Y. Miki, H. Ando, A. Horii, K. Koyama, J. Utsunomiya, S. Baba, P. Hedge, Mutations of chromosome 5q21 genes in FAP and colorectal cancer patients, *Science* 253 (5020) (1991) 665–669.
- [53] M. Giannakis, E. Hodis, X. Jasmine Mu, M. Yamada, J. Rosenbluh, K. Cibulskis, G. Saksena, M.S. Lawrence, Z.R. Qian, R. Nishihara, E.M. Van Allen, W.C. Hahn, S.B. Gabriel, E.S. Lander, G. Getz, S. Ogino, C.S. Fuchs, L.A. Garraway, RNF43 is frequently mutated in colorectal and endometrial cancers, *Nat. Genet.* 46 (12) (2014) 1264–1266.
- [54] R. Yaeger, W.K. Chatila, M.D. Lipsky, J.F. Hechtman, A. Cerke, F. Sanchez-Vega, G. Jayakumar, S. Middha, A. Zehir, M.T.A. Donoghue, D. You, A. Viale, N. Kemeny, N.H. Segal, Z.K. Stadler, A.M. Varghese, R. Kundra, J. Gao, A. Syed, D.M. Hyman, E. Vakiani, N. Rosen, B.S. Taylor, M. Ladanyi, M.F. Berger, D.B. Solit, J. Shia, L. Saltz, N. Schultz, Clinical sequencing defines the genomic landscape of metastatic colorectal cancer, *Cancer Cell* 33 (1) (2018) 125–136.
- [55] L. Azzolini, T. Panciera, S. Soligo, E. Enzo, S. Bicciato, S. Dupont, S. Bresolin, C. Frasson, G. Basso, V. Guzzardo, A. Fassina, M. Cordenonsi, S. Piccolo, YAP/TAZ incorporation in the β-catenin destruction complex orchestrates the Wnt response, *Cell* 158 (1) (2014) 157–170.
- [56] L. Wang, S. Shi, Z. Guo, X. Zhang, S. Han, A. Yang, W. Wen, Q. Zhu, Overexpression of YAP and TAZ is an independent predictor of prognosis in colorectal cancer and related to the proliferation and metastasis of colon cancer cells, *PLoS One* 8 (6) (2013) e65539.
- [57] S.P. Tenbaum, P. Ordóñez-Moran, I. Puig, I. Chicote, O. Arques, S. Landolfi, Y. Fernandez, J.R. Herance, J.D. Gispert, L. Mendizábal, S. Aguilar, S. Ramón y Cajal, S. Schwartz Jr., A. Vivancos, E. Espín, S. Rojas, J. Baselga, J. Taberner, A. Muñoz, H.G. Palmer, β-Catenin confers resistance to PI3K and AKT inhibitors and subverts FOXO3a to promote metastasis in colon cancer, *Nat. Med.* 18 (6) (2012) 892–901.
- [58] S.M. Mazzoni, E.M. Petty, E.M. Stoffel, E.R. Fearon, An Axin2 mutant allele associated with predisposition to colorectal neoplasia has context-dependent effects on Axin2 protein function, *Neoplasia* 17 (5) (2015) 463–472.
- [59] J.H. Kim, Y.H. Kim, G.Y. Song, D.E. Kim, Y.J. Jeong, K.H. Liu, Y.H. Chung, S. Oh, Ursolic acid and its natural derivative corosolic acid suppress the proliferation of APC-mutated colon cancer cells through promotion of β-catenin degradation, *Food Chem. Toxicol.* 67 (2014) 87–95.
- [60] G. Wang, C.C. Feng, S.J. Chu, R. Zhang, Y.M. Lu, J.S. Zhu, J. Zhang, Toosendanin inhibits growth and induces apoptosis in colorectal cancer cells through suppression of AKT/GSK3β/β-catenin pathway, *Int. J. Oncol.* 47 (5) (2015) 1767–1774.
- [61] J. Chinison, J.S. Aguilar, A. Avalos, Y. Huang, Z.J. Wang, D.J. Cameron, J.J. Hao, Triptonide effectively inhibits Wnt/β-catenin signaling via c-terminal transactivation domain of β-catenin, *Sci. Rep.* 6 (2016) 32779.
- [62] D. Cianciosi, A. Varela-Lopez, T.Y. Forbes-Hernandez, M. Gasparini, S. Afrin, P. Reboreda-Rodriguez, J. Zhang, J.L. Quiles, S.F. Nabavi, M. Battino, F. Giampieri, Targeting molecular pathways in cancer stem cells by natural bioactive compounds, *Pharmacol. Res.* 135 (2018) 150–165.
- [63] S. Rajamanickam, B. Velmurugan, M. Kaur, R.P. Singh, R. Agarwal, Chemoprevention of intestinal tumorigenesis in APC^{min/+} mice by silibinin, *Cancer Res.* 70 (6) (2010) 2368–2378.
- [64] M. Xu, S. Wang, Y.U. Song, J. Yao, K. Huang, X. Zhu, Apigenin suppresses colorectal cancer cell proliferation, migration and invasion via inhibition of the Wnt/β-catenin signaling pathway, *Oncol. Lett.* 11 (5) (2016) 3075–3080.
- [65] H. Ruan, Y.Y. Zhan, J. Hou, B. Xu, B. Chen, Y. Tian, D. Wu, Y. Zhao, Y. Zhang, X. Chen, P. Mi, L. Zhang, S. Zhang, X. Wang, H. Cao, W. Zhang, H. Wang, H. Li, Y. Su, X.K. Zhang, T. Hu, Berberine binds RXRα to suppress β-catenin signaling in colon cancer cells, *Oncogene* 36 (50) (2017) 6906–6918.

- [66] K. Wu, M. Zhou, Q.X. Wu, S.X. Yuan, D.X. Wang, J.L. Jin, J. Huang, J.Q. Yang, W.J. Sun, L.H. Wan, B.C. He, The role of IGFBP-5 in mediating the anti-proliferation effect of tetrandrine in human colon cancer cells, *Int. J. Oncol.* 46 (3) (2015) 1205–1213.
- [67] R. Ganugula, M. Arora, P. Jaisamut, R. Wiwattanapatapee, H.G. Jorgensen, V.P. Venkatpurwar, B.Y. Zhou, A.R. Hoffmann, R. Basu, S.D. Guo, N.V.R.K. Majeti, Nano-curcumin safely prevents streptozotocin-induced inflammation and apoptosis in pancreatic beta cells for effective management of type 1 diabetes mellitus, *Brit. J. Pharmacol.* 174 (13) (2017) 2074–2084.
- [68] J. Trujillo, Y.I. Chirino, E. Molina-Jijon, A.C. Anderica-Romero, E. Tapia, J. Pedraza-Chaverri, Renoprotective effect of the antioxidant curcumin: recent findings, *Redox Biol.* 1 (1) (2013) 448–456.
- [69] A.B. Kunnumakkara, D. Bordoloi, G. Padmavathi, J. Monisha, N.K. Roy, S. Prasad, B.B. Aggarwal, Curcumin, the golden nutraceutical: multitargeting for multiple chronic diseases, *Br. J. Pharmacol.* 174 (11) (2017) 1325–1348.
- [70] A. Milani, M. Basirnejad, S. Shahbazi, A. Bolhassani, Carotenoids: biochemistry, pharmacology and treatment, *Br. J. Pharmacol.* 174 (11) (2017) 1290–1324.
- [71] M. Zusso, G. Mercanti, F. Bellutti, R.M.C. Di Martino, A. Pagetta, C. Marinelli, P. Brun, E. Ragazzi, R. Lo, S. Stifani, P. Giusti, S. Moro, Phenolic 1,3-diketones attenuate lipopolysaccharide-induced inflammatory response by an alternative magnesium-mediated mechanism, *Br. J. Pharmacol.* 174 (10) (2017) 1090–1103.
- [72] C.Y. Shen, J.G. Jiang, L. Yang, D.W. Wang, W. Zhu, Anti-ageing active ingredients from herbs and nutraceuticals used in traditional Chinese medicine: pharmacological mechanisms and implications for drug discovery, *Br. J. Pharmacol.* 174 (11) (2017) 1395–1425.
- [73] H.Q. Dou, R.H. Shen, J.X. Tao, L.C. Huang, H.Z. Shi, H. Chen, Y.X. Wang, T. Wang, Curcumin suppresses the colon cancer proliferation by inhibiting Wnt/β-catenin pathways via miR-130a, *Front. Pharmacol.* 8 (2017) 877.
- [74] A. Rodriguez Garcia, D. Hevia, J.C. Mayo, P. Gonzalez Menendez, L. Cocco, J. Lu, A. Holmgren, R.M. Sainz, Thioredoxin 1 modulates apoptosis induced by bioactive compounds in prostate cancer cells, *Redox Biol.* 12 (2017) 634–647.
- [75] H. Li, S. Krstic, M. Wink, Modulation of multidrug resistant in cancer cells by EGCG, tannic acid and curcumin, *Phytomedicine* 50 (2018) 213–222.
- [76] W. Zhang, H. Shi, C. Chen, K. Ren, Y. Xu, X. Liu, L. He, Curcumin enhances cisplatin sensitivity of human NSCLC cell lines through influencing Cu-Sp1-CTF1 regulatory loop, *Phytomedicine* 48 (2018) 51–61.
- [77] W. Feng, C.X. Yang, L. Zhang, Y. Fang, M. Yan, Curcumin promotes the apoptosis of human endometrial carcinoma cells by downregulating the expression of androgen receptor through Wnt signal pathway, *Eur. J. Gynaecol. Oncol.* 35 (6) (2014) 718–723.
- [78] K.T. Hwang, J. Kim, J. Jung, J.H. Chang, Y.J. Chai, S.W. Oh, S. Oh, Y.A. Kim, S.B. Park, K.R. Hwang, Impact of breast cancer subtypes on prognosis of women with operable invasive breast cancer: a population-based study using SEER database, *Clin. Cancer Res.* 25 (6) (2018) 1970–1979.
- [79] R.S. Riley, E.S. Day, Frizzled7 antibody-functionalized nanoshells enable multivalent binding for Wnt signaling inhibition in triple-negative breast cancer cells, *Small* 13 (26) (2017) UNSP 1700544.
- [80] P. De, J.H. Carlson, T. Jepperson, S. Willis, B. Leyland-Jones, N. Dey, RAC1 GTPase signals Wnt/β-catenin pathway mediated integrin-directed metastasis-associated tumor cell phenotypes in triple-negative breast cancers, *Oncotarget* 8 (2) (2017) 3072–3103.
- [81] A.V. Koval, P. Vlasov, P. Shchikova, S. Khunderyakova, Y. Markov, J. Panchenko, A. Volodina, F.A. Kondrashov, V.L. Kataeva, Anti-leprosy drug clofazimine inhibits growth of triple-negative breast cancer cells via inhibition of canonical Wnt signaling, *Biochem. Pharmacol.* 87 (4) (2014) 571–578.
- [82] A. Koval, C.A. Pieme, E.F. Queiroz, S. Ragusa, K. Ahmed, A. Blagodatski, J.L. Wolfender, T.V. Petrova, V.L. Kataeva, Tannins from Syzygium guineense suppress Wnt signaling and proliferation of Wnt-dependent tumors through a direct effect on secreted Wnts, *Cancer Lett.* 435 (2018) 110–120.
- [83] L. Arzi, A. Farahi, N. Jafarzadeh, G. Riazi, M. Sadeghzadeh, R. Hoshyar, Inhibitory effect of crocin on metastasis of triple-negative breast cancer by interfering with Wnt/β-catenin pathway in murine model, *DNA Cell Biol.* 37 (12) (2018) 1068–1075.
- [84] L. Arzi, G. Riazi, M. Sadeghzadeh, R. Hoshyar, N. Jafarzadeh, A comparative study on anti-invasion, antimigration, and antiadhesion effects of the bioactive carotenoids of saffron on 4T1 breast cancer cells through their effects on Wnt/β-catenin pathway genes, *DNA Cell Biol.* 37 (8) (2018) 697–707.
- [85] Y. Zhang, S.G. Chen, C.Y. Wei, G.O. Rankin, X.Q. Ye, Y.C. Chen, Dietary compound proanthocyanidins from Chinese bayberry (*Myrica rubra* Sieb. et Zucc.) leaves attenuate chemotherapy-resistant ovarian cancer stem cell traits via targeting the Wnt/β-catenin signaling pathway and inducing G1 cell cycle arrest, *Food Funct.* 9 (1) (2018) 525–533.
- [86] I. Nardi, T. Reno, X. Yun, T. Sztaib, J. Wang, H. Dai, L. Zheng, B. Shen, J. Kim, D. Raz, Triptolide inhibits Wnt signaling in NSCLC through upregulation of multiple Wnt inhibitory factors via epigenetic modifications to Histone H3, *Int. J. Cancer* 143 (10) (2018) 2470–2478.
- [87] A.C. Webster, E.V. Nagler, R.L. Morton, P. Masson, Chronic kidney disease, *Lancet* 389 (10075) (2017) 1238–1252.
- [88] Y.Y. Chen, D.Q. Chen, L. Chen, J.R. Liu, N.D. Vaziri, Y. Guo, Y.Y. Zhao, Microbiome-metabolome reveals the contribution of gut-kidney axis on kidney disease, *J. Transl. Med.* 17 (1) (2019) 5.
- [89] Y.Y. Zhao, X.L. Cheng, F. Wei, X. Bai, R.C. Lin, Application of faecal metabolomics on an experimental model of tubulointerstitial fibrosis by ultra performance liquid chromatography/high-sensitivity mass spectrometry with MSE data collection technique, *Biomarkers* 17 (8) (2012) 721–729.
- [90] L. Chen, W. Su, H. Chen, D.Q. Chen, M. Wang, Y. Guo, Y.Y. Zhao, Proteomics for biomarker identification and clinical application in kidney disease, *Adv. Clin. Chem.* 85 (2018) 91–113.
- [91] Z.H. Zhang, H. Chen, N.D. Vaziri, J.R. Mao, L. Zhang, X. Bai, Y.Y. Zhao, Metabolomic signatures of chronic kidney disease of diverse etiologies in the rats and humans, *J. Proteome Res.* 15 (10) (2016) 3802–3812.
- [92] Y.Y. Zhao, H.L. Wang, X.L. Cheng, F. Wei, X. Bai, R.C. Lin, N.D. Vaziri, Metabolomics analysis reveals the association between lipid abnormalities and oxidative stress, inflammation, fibrosis, and Nrf2 dysfunction in aristolochic acid-induced nephropathy, *Sci. Rep.* 5 (2015) 12936.
- [93] H. Zhao, S.X. Ma, Y.Q. Shang, H.Q. Zhang, W. Su, microRNAs in chronic kidney disease, *Clin. Chim. Acta* 491 (2019) 59–65.
- [94] Y.Y. Zhao, Metabolomics in chronic kidney disease, *Clin. Chim. Acta* 422 (2013) 59–69.
- [95] Y.Y. Zhao, R.C. Lin, Metabolomics in nephrotoxicity, *Adv. Clin. Chem.* 65 (2014) 69–89.
- [96] H. Chen, L. Chen, D. Liu, D.Q. Chen, N.D. Vaziri, X.Y. Yu, L. Zhang, W. Su, X. Bai, Y.Y. Zhao, Combined clinical phenotype and lipidomic analysis reveals the impact of chronic kidney disease on lipid metabolism, *J. Proteome Res.* 16 (4) (2017) 1566–1578.
- [97] Y.Y. Zhao, X.L. Cheng, F. Wei, X. Bai, X.J. Tan, R.C. Lin, Q. Mei, Intrarenal metabolomic investigation of chronic kidney disease and its TGF-β1 mechanism in induced-adenine rats using UPLC Q-TOF/HSMS/MSE, *J. Proteome Res.* 12 (2) (2013) 692–703.
- [98] D.Q. Chen, H. Chen, L. Chen, N.D. Vaziri, M. Wang, X.R. Li, Y.Y. Zhao, The link between phenotype and fatty acid metabolism in advanced chronic kidney disease, *Nephrol. Dial. Transplant.* 32 (7) (2017) 1154–1166.
- [99] Y.Y. Zhao, N.D. Vaziri, R.C. Lin, Lipidomics: new insight into kidney disease, *Adv. Clin. Chem.* 68 (2015) 153–175.
- [100] Z.H. Zhang, J.R. Mao, H. Chen, W. Su, Y. Zhang, L. Zhang, D.Q. Chen, Y.Y. Zhao, N.D. Vaziri, Removal of uremic retention products by hemodialysis is coupled with indiscriminate loss of vital metabolites, *Clin. Biochem.* 50 (18) (2017) 1078–1086.
- [101] Z.H. Zhang, N.D. Vaziri, F. Wei, X.L. Cheng, X. Bai, Y.Y. Zhao, An integrated lipidomics and metabolomics reveal nephroprotective effect and biochemical mechanism of *Rheum officinale* in chronic renal failure, *Sci. Rep.* 6 (2016) 22151.
- [102] Z.H. Zhang, F. Wei, N.D. Vaziri, X.L. Cheng, X. Bai, R.C. Lin, Y.Y. Zhao, Metabolomics insights into chronic kidney disease and modulatory effect of rhubarb against tubulointerstitial fibrosis, *Sci. Rep.* 5 (2015) 14472.
- [103] Y.Y. Zhao, H. Chen, T. Tian, D.Q. Chen, X. Bai, F. Wei, A pharmaco-metabolic study on chronic kidney disease and therapeutic effect of ergone by UPLC-QTOF/HDMDS, *PLoS One* 23 (9) (2014) e115467.
- [104] Y.Y. Zhao, L. Zhang, J.R. Mao, X.H. Cheng, R.C. Lin, Y. Zhang, W.J. Sun, Ergosta-4,6,8(14),22-tetraen-3-one isolated from *Polyporus umbellatus* prevents early renal injury in aristolochic acid-induced nephropathy rats, *J. Pharm. Pharmacol.* 63 (12) (2011) 1581–1586.
- [105] Y.Y. Zhao, L. Zhang, F.Y. Long, X.L. Cheng, X. Bai, F. Wei, R.C. Lin, UPLC-Q-TOF/HSMS/MS^E-based metabolomics for adenine-induced changes in metabolic profiles of rat faeces and intervention effects of ergosta-4,6,8(14),22-tetraen-3-one, *Chem. Biol. Interact.* 201 (1–3) (2013) 31–38.
- [106] L. Chen, D.Q. Chen, Unilateral ureteral obstruction causes gut microbial dysbiosis and metabolome disorders contributing to tubulointerstitial fibrosis, *Exp. Mol. Med.* 51 (3) (2019) 38.
- [107] Y.Z. Wang, J. Zhang, Y.L. Zhao, T. Li, T. Shen, J.Q. Li, W.Y. Li, H.G. Liu, Mycology, cultivation, traditional uses, phytochemistry and pharmacology of *Wolfiporia cocos* (Schwein.) Ryvarden et Gilb.: a review, *J. Ethnopharmacol.* 147 (2) (2013) 265–276.
- [108] Y.Y. Zhao, Y.L. Feng, X. Du, Z.H. Xi, X.L. Cheng, F. Wei, Diuretic activity of the ethanol and aqueous extracts of the surface layer of *Poria cocos* in rat, *J. Ethnopharmacol.* 144 (3) (2012) 775–778.
- [109] Y.L. Feng, P. Lei, T. Tian, L. Yin, D.Q. Chen, H. Chen, Q. Mei, Y.Y. Zhao, R.C. Lin, Diuretic activity of some fractions of the epidermis of *Poria cocos*, *J. Ethnopharmacol.* 150 (3) (2013) 1114–1118.
- [110] H. Miao, Y.H. Zhao, N.D. Vaziri, D.D. Tang, H. Chen, H. Chen, M. Khazaeli, M. Tarbati Boldaji, L. Hatami, Y.Y. Zhao, Lipidomics biomarkers of diet-induced hyperlipidemia and its treatment with *Poria cocos*, *J. Agric. Food Chem.* 64 (4) (2016) 969–979.
- [111] H. Miao, M.H. Li, X. Zhang, S.J. Yuan, C.C. Ho, Y.Y. Zhao, The antihyperlipidemic effect of Fu-Ling-Pi is associated with abnormal fatty acid metabolism as assessed by UPLC-HDMS-based lipidomics, *RSC Adv.* 5 (79) (2015) 64208–64219.
- [112] M. Wang, D.Q. Chen, M.C. Wang, H. Chen, L. Chen, D. Liu, H. Zhao, Y.Y. Zhao, Poricoic acid ZA, a novel RAS inhibitor, attenuates tubulo-interstitial fibrosis and podocyte injury by inhibiting TGF-β/Smad signaling pathway, *Phytomedicine* 36 (2017) 243–253.
- [113] Y.Y. Zhao, H.T. Li, Y.L. Feng, X. Bai, R.C. Lin, Urinary metabonomic study of the surface layer of *Poria cocos* as an effective treatment for chronic renal injury in rats, *J. Ethnopharmacol.* 148 (2) (2013) 403–410.
- [114] Y.Y. Zhao, P. Lei, D.Q. Chen, Y.L. Feng, X. Bai, Renal metabolic profiling of early renal injury and renoprotective effects of *Poria cocos* epidermis using UPLC Q-TOF/HSMS/MS^E, *J. Pharm. Biomed. Anal.* 81–82 (2013) 202–209.
- [115] D.Q. Chen, Y.L. Feng, L. Chen, J.R. Liu, M. Wang, N.D. Vaziri, Y.Y. Zhao, Poricoic acid A enhances melatonin inhibition of AKI-to-CKD transition by regulating Gas6/Axl/NFKB/Nrf2 axis, *Free Radic. Biol. Med.* 134 (2019) 484–497.
- [116] M. Wang, D.Q. Chen, L. Chen, D. Liu, H. Zhao, Z.H. Zhang, N.D. Vaziri, Y. Guo, Y.Y. Zhao, Novel RAS inhibitors Poricoic Acid ZG and Poricoic Acid ZH attenuate renal fibrosis via a Wnt/β-catenin pathway and targeted phosphorylation of Smad3 signaling, *J. Agric. Food Chem.* 66 (8) (2018) 1828–1842.
- [117] M. Wang, D.Q. Chen, L. Chen, G. Cao, H. Zhao, D. Liu, N.D. Vaziri, Y. Guo, Y.Y. Zhao, Novel inhibitors of the cellular renin-angiotensin system components, poricoic acids, target Smad3 phosphorylation and Wnt/β-catenin pathway against renal fibrosis, *Br. J. Pharmacol.* 175 (13) (2018) 2689–2708.
- [118] T. Tian, H. Chen, Y.Y. Zhao, Traditional uses, phytochemistry, pharmacology, toxicology and quality control of *Alisma orientale* (Sam.) Juzep: a review, *J. Ethnopharmacol.* 158 (2014) 373–387.
- [119] H. Miao, L. Zhang, D.Q. Chen, H. Chen, Y.Y. Zhao, S.C. Ma, Urinary biomarker and treatment mechanism of *Rhizoma Alismatis* on hyperlipidemia, *Biomed.*

- Chromatogr. 31 (4) (2017) e3829.
- [120] Y.L. Feng, H. Chen, T. Tian, D.Q. Chen, Y.Y. Zhao, R.C. Lin, Diuretic and anti-diuretic activities of the ethanol and aqueous extracts of *Alismatis rhizoma*, J. Ethnopharmacol. 154 (2) (2014) 386–390.
- [121] D.Q. Chen, Y.L. Feng, T. Tian, H. Chen, L. Yin, Y.Y. Zhao, R.C. Lin, Diuretic and anti-diuretic activities of fractions of *Alismatis rhizoma*, J. Ethnopharmacol. 157 (2014) 114–118.
- [122] F. Dou, H. Miao, J.W. Wang, L. Chen, M. Wang, H. Chen, A.D. Wen, Y.Y. Zhao, An integrated lipidomics and phenotype study reveals protective effect and biochemical mechanism of traditionally used *Alisma orientale* Juzepzuk in chronic kidney disease, Front. Pharmacol. 9 (2018) 53.
- [123] H. Chen, T. Yang, M.C. Wang, D.Q. Chen, Y. Yang, Y.Y. Zhao, Novel RAS inhibitor 25-O-methylalisol F attenuates epithelial-to-mesenchymal transition and tubulo-interstitial fibrosis by selectively inhibiting TGF- β -mediated Smad3 phosphorylation, Phytomedicine 42 (2018) 207–218.
- [124] D. Charvin, R. Medori, R.A. Hauser, O. Rascol, Therapeutic strategies for Parkinson disease: beyond dopaminergic drugs, Nat. Rev. Drug Discov. 17 (11) (2018) 804–822.
- [125] R. Katzenschlager, W. Poewe, O. Rascol, C. Trenkwalder, G. Deuselch, K.R. Chaudhuri, T. Henriksen, T. van Laar, K. Spivey, S. Vel, H. Staines, A. Lees, Apomorphine subcutaneous infusion in patients with Parkinson's disease with persistent motor fluctuations (TOLEDO): a multicentre, double-blind, randomised, placebo-controlled trial, Lancet Neurol. 17 (9) (2018) 749–759.
- [126] Y.C. Weng, C.M. Chen, Y.C. Chen, H.C. Fung, C.W. Chang, K.H. Chang, Y.R. Wu, Eukaryotic translation initiation factor 4- γ , 1 gene mutations are rare in Parkinson's disease among Taiwanese, J. Formos. Med. Assoc. 115 (9) (2016) 728–733.
- [127] A. Salasova, C. Yokota, D. Potesil, Z. Zdrahal, V. Bryja, E. Arenas, A proteomic analysis of LRRK2 binding partners reveals interactions with multiple signaling components of the Wnt/PCP pathway, Mol. Neurodegener. 12 (1) (2017) 54.
- [128] S.K. Tiwari, S. Agarwal, A. Tripathi, R.K. Chaturvedi, Bisphenol-A mediated inhibition of hippocampal neurogenesis attenuated by curcumin via canonical Wnt pathway, Mol. Neurobiol. 53 (5) (2016) 3010–3029.
- [129] S.K. Tiwari, S. Agarwal, B. Seth, A. Yadav, S. Nair, P. Bhatnagar, M. Karmakar, M. Kumari, L.K.S. Chauhan, D.K. Patel, V. Srivastava, D. Singh, S.K. Gupta, A. Tripathi, R.K. Chaturvedi, K.C. Gupta, Curcumin-loaded nanoparticles potently induce adult neurogenesis and reverse cognitive deficits in Alzheimer's disease model via canonical Wnt/ β -catenin pathway, ACS Nano 8 (1) (2014) 76–103.
- [130] M.N. Weitzmann, I. Ofotokun, Physiological and pathophysiological bone turnover-role of the immune system, Nat. Rev. Endocrinol. 12 (9) (2016) 518–532.
- [131] T.D. Rachner, R. Coleman, P. Hadji, L.C. Hofbauer, Bone health during endocrine therapy for cancer, Lancet Diabetes Endocrinol. 6 (11) (2018) 901–910.
- [132] M. Viswanathan, S. Reddy, N. Berkman, K. Cullen, J.C. Middleton, W.K. Nicholson, L.C. Kahwati, Screening to prevent osteoporotic fractures: updated evidence report and systematic review for the US preventive services task force, JAMA 319 (24) (2018) 2532–2551.
- [133] K.E. Hansen, J.W. Nieves, S. Nudurupati, D.C. Metz, M.C. Perez, Dexlansoprazole and esomeprazole do not affect bone homeostasis in healthy postmenopausal women, Gastroenterology 156 (4) (2019) 926–934 e6.
- [134] U.S.P.S.T. Force, S.J. Curry, A.H. Krist, D.K. Owens, M.J. Barry, A.B. Caughey, K.W. Davidson, C.A. Doubeni, J.W. Epling Jr., A.R. Kemper, M. Kubik, C.S. Landefeld, C.M. Mangione, M.G. Phipps, M. Pignone, M. Silverstein, M.A. Simon, C.W. Tseng, J.B. Wong, Screening for osteoporosis to prevent fractures: US preventive services task force recommendation statement, JAMA 319 (24) (2018) 2521–2531.
- [135] B. Kim, K.Y. Lee, B. Park, Icarin abrogates osteoclast formation through the regulation of the RANKL-mediated TRAF6/NF- κ B/ERK signaling pathway in Raw264.7 cells, Phytomedicine 51 (2018) 181–190.
- [136] I. Soundharrajan, D.H. Kim, S. Srisesharam, P. Kuppusamy, R. Sivanesan, K.C. Choi, Limonene promotes osteoblast differentiation and 2-deoxy-D-glucose uptake through p38MAPK and AKT signaling pathways in C2C12 skeletal muscle cells, Phytomedicine 45 (2018) 41–48.
- [137] J.J. Liu, Z.Z. Zhang, Q. Guo, Y.H. Dong, Q.P. Zhao, X.Q. Ma, Syringin prevents bone loss in ovariectomized mice via TRAF6 mediated inhibition of NF- κ B and stimulation of PI3K/AKT, Phytomedicine 42 (2018) 43–50.
- [138] L.D. Antika, E.J. Lee, Y.H. Kim, M.K. Kang, S.H. Park, D.Y. Kim, H. Oh, Y.J. Choi, Y.H. Kang, Dietary phlorizin enhances osteoblastogenic bone formation through enhancing β -catenin activity via GSK3 β inhibition in a model of senile osteoporosis, J. Nutr. Biochem. 49 (2017) 42–52.
- [139] H.J. Chung, W.K. Kim, J. Oh, M.R. Kim, J.S. Shin, J. Lee, I.H. Ha, S.K. Lee, Anti-osteoporotic activity of harpagoside by upregulation of the BMP2 and Wnt signaling pathways in osteoblasts and suppression of differentiation in osteoclasts, J. Nat. Prod. 80 (2) (2017) 434–442.
- [140] X. Tao, L. Yin, L. Xu, J. Peng, Dioscin: a diverse acting natural compound with therapeutic potential in metabolic diseases, cancer, inflammation and infections, Pharmacol. Res. 137 (2018) 259–269.
- [141] Y.Q. Liu, Z.L. Hong, L.B. Zhan, H.Y. Chu, X.Z. Zhang, G.H. Li, Wedelolactone enhances osteoblastogenesis by regulating Wnt/ β -catenin signaling pathway but suppresses osteoclastogenesis by NF- κ B/c-fos/NFATc1 pathway, Sci. Rep. 6 (2016) e32260.
- [142] M.B. Kim, Y. Song, J.K. Hwang, Kirenlol stimulates osteoblast differentiation through activation of the BMP and Wnt/ β -catenin signaling pathways in MC3T3-E1 cells, Fitoterapia 98 (2014) 59–65.
- [143] X. Cheng, B.F. Wei, L.J. Sun, X.F. Hu, J.C. Liang, Y. Chen, Astragaloside I stimulates osteoblast differentiation through the Wnt/ β -catenin signaling pathway, Phytother. Res. 30 (10) (2016) 1680–1688.
- [144] Y.J. Liu, L.L. Huang, B.H. Hao, H. Li, S.L. Zhu, Q.S. Wang, R.X. Li, Y.Q. Xu, X.Z. Zhang, Use of an osteoblast overload damage model to probe the effect of icariin on the proliferation, differentiation and mineralization of MC3T3-E1 cells through the Wnt/ β -catenin signalling pathway, Cell. Physiol. Biochem. 41 (4) (2017) 1605–1615.
- [145] Y. Wang, R. Wang, F. Zhang, Icarin promotes the proliferation and differentiation of osteoblasts from the rat mandible by the Wnt/ β -catenin signalling pathway, Mol. Med. Rep. 18 (3) (2018) 3445–3450.
- [146] H. Zhang, H.Q. Li, Tricin enhances osteoblastogenesis through the regulation of Wnt/ β -catenin signaling in human mesenchymal stem cells, Mech. Dev. 152 (2018) 38–43.
- [147] D. Aletaha, J.S. Smolen, Diagnosis and management of rheumatoid arthritis: a review, JAMA 320 (13) (2018) 1360–1372.
- [148] M. Bernardes, C. Duraes, A. Oliveira, M.J. Martins, R. Lucas, L. Costa, J.G. Pereira, I. Ramos, J.C. Machado, F. Simoes Ventura, LRP5 gene polymorphisms and radiographic joint damage in rheumatoid arthritis patients, Osteoporos. Int. 29 (10) (2018) 2355–2368.
- [149] B. Oz, A. Yildirim, S. Yolbas, Z.B. Celik, E.O. Etem, G. Deniz, M. Akin, Z.A. Akar, A. Karatas, S.S. Koca, Resveratrol inhibits Src tyrosine kinase, STAT3, and Wnt signaling pathway in collagen induced arthritides model, Biofactors 45 (1) (2018) 69–74.
- [150] Y.G. Zhao, W.J. Song, Z.Y. Wang, Z.Q. Wang, X. Jin, J.C. Xu, L. Bai, Y.Y. Li, J.W. Cui, L. Cai, Resveratrol attenuates testicular apoptosis in type 1 diabetic mice: role of AKT-mediated Nrf2 activation and p62-dependent Keap1 degradation, Redox Biol. 14 (2018) 609–617.
- [151] I. Orlandi, G. Stamerra, M. Strippoli, M. Vai, During yeast chronological aging resveratrol supplementation results in a short-lived phenotype Sir2-dependent, Redox Biol. 12 (2017) 745–754.
- [152] Y. Zhu, B. Feng, S. He, Z. Su, G. Zheng, Resveratrol combined with total flavones of hawthorn alleviate the endothelial cells injury after coronary bypass graft surgery, Phytomedicine 40 (2018) 20–26.
- [153] C.C. Tsai, S.L. Tey, M.C. Lee, C.W. Liu, Y.T. Su, S.C. Huang, Mechanism of resveratrol-induced relaxation of the guinea pig fundus, Phytomedicine 43 (2018) 55–59.
- [154] S.S. Koca, S. Yolbas, A. Yildirim, Z.B. Celik, E.E. Onalan, M. Akin, Resveratrol inhibits canonical Wnt signaling and ameliorates experimental arthritis, Ann. Rheum. Dis. 75 (2016) 933–933.
- [155] Z. Rasheed, N. Rasheed, O. Al Shaya, Epigallocatechin-3-O-gallate modulates global microRNA expression in interleukin-1 β -stimulated human osteoarthritis chondrocytes: potential role of EGCG on negative co-regulation of microRNA-140-3p and ADAMTS5, Eur. J. Nutr. 57 (3) (2018) 917–928.
- [156] E. Asteriou, A. Gkoutzourelas, A. Mavropoulos, C. Katsiari, L.I. Sakkas, D.P. Bogdanos, Curcumin for the management of periodontitis and early ACPA-positive rheumatoid arthritis: killing two birds with one stone, Nutrients 10 (7) (2018) e908.
- [157] H.S. Shin, S.Y. Park, E.S. Hwang, D.G. Lee, H.G. Song, G.T. Maylonov, T.H. Yi, The inductive effect of ginsenoside R1 on hair growth by altering the Wnt signal pathway in telogen mouse skin, Eur. J. Pharmacol. 730 (2014) 82–89.
- [158] J.J. Barrott, B.E. Illum, H.F. Jin, M.L. Hedberg, Y.L. Wang, A. Grossmann, M. Haldar, M.R. Capecchi, K.B. Jones, Paracrine osteoprotegerin and β -catenin stabilization support synovial sarcomagenesis in periosteal cells, J. Clin. Invest. 128 (1) (2018) 207–218.
- [159] T. Jin, Current understanding on role of the Wnt signaling pathway effector TCF7L2 in glucose homeostasis, Endocr. Rev. 37 (3) (2016) 254–277.
- [160] B.M. Larsen, S.M. Hrycaj, M. Newman, Y. Li, D.M. Wellik, Mesenchymal Hox6 function is required for mouse pancreatic endocrine cell differentiation, Development 142 (22) (2015) 3859–3868.
- [161] A. Hiyama, K. Morita, D. Sakai, M. Watanabe, CCN family member 2/connective tissue growth factor (CCN2/CTGF) is regulated by Wnt/ β -catenin signaling in nucleus pulposus cells, Arthritis Res. Ther. 20 (1) (2018) 217.
- [162] P. Blysaczuk, B. Muller Edenborn, T. Valenta, E. Ostro, M. Stellato, S. Behnke, K. Glatz, K. Basler, T.F. Luscher, O. Distler, U. Eriksson, G. Kanis, Transforming growth factor- β -dependent Wnt secretion controls myofibroblast formation and myocardial fibrosis progression in experimental autoimmune myocarditis, Eur. Heart J. 38 (18) (2017) 1413–1425.
- [163] H.J. Chung, W.K. Kim, J. Oh, M.R. Kim, J.S. Shin, J. Lee, I.H. Ha, S.K. Lee, Anti-osteoporotic activity of harpagoside by upregulation of the BMP2 and Wnt signaling pathways in osteoblasts and suppression of differentiation in osteoclasts, J. Nat. Prod. 80 (2) (2017) 434–442.