# **Lawrence Berkeley National Laboratory**

**LBL Publications**

## **Title**

Photosynthetic responses to temperature across the tropics: a meta-analytic approach

## **Permalink**

<https://escholarship.org/uc/item/2cm2c75g>

## **Authors**

Carter, Kelsey R Cavaleri, Molly A Atkin, Owen K [et al.](https://escholarship.org/uc/item/2cm2c75g#author)

## **Publication Date**

2024-12-12

## **DOI**

10.1093/aob/mcae206

## **Copyright Information**

This work is made available under the terms of a Creative Commons Attribution-NonCommercial License, available at <https://creativecommons.org/licenses/by-nc/4.0/>

Peer reviewed

#### Photosynthetic responses to temperature across the tropics: a meta-analytic approach

Thank you for agreeing to review this paper for Annals of Botany. The Annals of Botany aims to be among the very top of plant science journals and as we receive over 1000 submissions every year we need to be very selective in deciding which papers we can publish. In making your assessment of the manuscript's suitability for publication in the journal please consider the following points.

#### **Scientific Scope**

Annals of Botany welcomes papers in all areas of plant science. Papers may address questions at any level of biological organization ranging from molecular through cells and organs, to whole organisms, species, communities and ecosystems. Its scope extends to all flowering and non-flowering taxa, and to evolutionary and pathology research. Many questions are addressed using comparative studies, genetics, genomics, molecular tools, and modeling.

To merit publication in Annals of Botany, contributions should be substantial, concise, written in clear English and combine originality of content with potential general interest.

- We want to publish papers where our reviewers are enthusiastic about the science: is this a paper that you would keep for reference, or pass on to your colleagues? If the answer is "no" then please enter a low priority score when you submit your report.
- We want to publish papers with novel and original content that move the subject forward, not papers that report incremental advances or findings that are already well known in other species. Please consider this when you enter a score for originality when you submit your report.

#### **Notes on categories of papers:**

All review-type articles should be novel, rigorous, substantial and "make a difference" to plant science. The purpose is to summarise, clearly and succinctly, the "cutting edge" of the subject and how future research would best be directed. Reviews should be relevant to a broad audience and allshould havea**strong conclusion and illustrations** including diagrams.

- *Primary Research* articles should report on original research relevant to the scope of the journal, demonstrating an important advance in the subject area, and the results should be clearly presented, novel and supported by appropriate experimental approaches. The Introduction should clearly set the context for the work and the Discussion should demonstrate the importance of the results within that context. Concise speculation, models and hypotheses are encouraged, but must be informed by the results and by the authors' expert knowledge of the subject.
- *Reviews* should place the subject in context, add significantly to previous reviews in the subject area and moving forward research in the subject area. Reviews should be selective, including the most important and best, up-to-date, references, not a blow-by-blow and exhaustive listing.
- Research in Context should combine a review/overview of a subject area with original research, often leading to new ideas or models; they present a hybrid of review and research. Typically a Research in Context article contains an extended Introduction that provides a general overview of the topic before incorporating new research results with a Discussion proposing general models and the impact of the research.
- *Viewpoints* are shorter reviews, presenting clear, concise and logical arguments supporting the authors' opinions, and in doing so help to stimulate discussions within the topic.
- *Botanical Briefings* are concise, perhaps more specialised reviews and usually cover topical issues, maybe involving some controversy.



<sup>16</sup> U.S. Geological Survey, Southwest Biological Science Center, Moab, UT, United States; <sup>17</sup> Institute for Global Change Biology, and School for the Environment and Sustainability, University of Michigan, Ann Arbor, MI 48109, United States; <sup>18</sup> Department of Forest Resources, University of Minnesota, St. Paul, MN, United States; <sup>19</sup> Biospheric Sciences Laboratory, NASA Goddard Space Flight Center, Greenbelt, Maryland, United States; <sup>20</sup> Smithsonian Tropical Research Institute, Apartado Postal 0843-03092, Panama, Republic of Panama; <sup>21</sup> Instituto de Biodiversidade *e* Florestas, Universidade Federal do Oeste do Pará (UFOPA, CEP 68035-110, Santarém, PA, Brazil <sup>22</sup> Department of Biological and Environmental Sciences, University of Gothenburg, Gothenburg, Sweden; 53 <sup>23</sup> School of Biological Sciences, University of Hong Kong, Pokfulam, Hong Kong. *This manuscript has been co-authored by UT-Battelle, LLC under Contract No. DE-AC05- 00OR22725 with the U.S. Department of Energy. The United States Government retains and the publisher, by accepting the article for publication, acknowledges that the United States Government retains a non-exclusive, paid-up, irrevocable, worldwide license to publish or reproduce the published form of this manuscript, or allow others to do so, for United States Government purposes. The Department of Energy will provide public access to these results of federally sponsored research in accordance with the DOE Public Access Plan (*<http://energy.gov/downloads/doe-public-access-plan>*).*

- **Abstract**
- 

### **Background and Aims**

69 Tropical forests exchange more carbon dioxide  $(CO<sub>2</sub>)$  with the atmosphere than any other

terrestrial biome. Yet, uncertainty in the projected carbon balance over the next century is

roughly three-times greater for the tropics than other ecosystems. Our limited knowledge of

tropical plant physiological responses, including photosynthetic, to climate change is a

substantial source of uncertainty in our ability to forecast the global terrestrial carbon sink.

#### **Methods**

 We used a meta-analytic approach, focusing on tropical photosynthetic temperature responses, to address this knowledge gap. Our dataset, gleaned from 18 independent studies, included leaf- level light saturated photosynthetic (*Asat*) temperature responses from 108 woody species, with additional temperature parameters (35 species) and rates (250 species) of both maximum rates of electron transport (*Jmax*) and Rubisco carboxylation (*Vcmax*). We investigated how these 81 parameters responded to mean annual temperature (MAT), temperature variability, aridity, and elevation, as well as also how responses differed among successional strategy, leaf habit, and light environment.

#### **Key Results**

86 Optimum temperatures for  $A_{sat}$  ( $T_{optA}$ ) and  $J_{max}$  ( $T_{optJ}$ ) increased with MAT but not for  $V_{cmax}$  (*ToptV*). Although photosynthetic rates were higher for "light" than "shaded" leaves, light conditions did not generate differences in temperature response parameters. *ToptA* did not differ 89 with successional strategy, but early successional species had  $\sim$  4 °C wider thermal niches than



 2015). In addition, these forests are projected to surpass their historical climate margin, entering into novel climate conditions within the next quarter century (Williams *et al.* 2007; but see Jaramillo *et al.* 2010), a trend anticipated to occur sooner for the tropics than other global regions (Diffenbaugh and Scherer 2011; Mora *et al.* 2013; Doughty *et al.* 2023). Some tropical forests are already believed to be operating near or beyond their photosynthetic thermal optima (Doughty and Goulden 2008; Vårhammar *et al.* 2015; Mau *et al.* 2018; Dusenge *et al.* 2021; Doughty *et al.* 2023), making them particularly vulnerable to the effects of climate warming on carbon uptake.

 Due to the significant uncertainties around how the tropical forest biome will respond to continued global change, better representation of vegetation processes is needed to more accurately inform Earth system and dynamic vegetation models (Friedlingstein *et al.* 2006; Matthews *et al.* 2007; Booth *et al.* 2012; Rogers *et al.* 2017; Fisher *et al.* 2018). In particular, quantifying photosynthetic temperature responses of tropical species will help to reduce model uncertainty (Matthews *et al.* 2007; Booth *et al.* 2012). Photosynthesis has a peaked response to temperature, where the rate of photosynthesis increases and then declines after the optimum temperature (*ToptA*; Table 1) is reached. The components of photosynthetic decline beyond the thermal optimum can be examined by exploring stomatal conductance and the underlying biochemical processes that control photosynthesis. These biochemical processes include the 131 maximum rate of carbon dioxide  $(CO_2)$  fixation by Rubisco  $(V_{cmax})$  and the maximum rate of 132 photosynthetic electron transport  $(J_{max})$ , both of which are derived by a well-established biochemical model (Farquhar *et al.* 1980; von Caemmerer and Farquhar 1981). Global vegetation models use the temperature response parameters of these biochemical processes

- controlling photosynthesis to predict carbon uptake at wider scales (Kattge *et al.* 2009; Lin *et al.* 2012; Smith and Dukes 2013; Mercado *et al.* 2018; Oliver *et al.* 2022).
- Considerable efforts have been made to quantify these photosynthetic response parameters at the global scale (Medlyn *et al.* 2002; Kattge and Knorr 2007; Yamori *et al.* 2014; Kumarathunge *et al.* 2019; Crous *et al.* 2022). These studies show that species can (but may not) acclimate to their growth environment, and algorithms developed in Kattge and Knorr (2007) have been implemented in some Earth system and vegetation models for more accurate representation of photosynthetic acclimation (e.g. Arneth *et al.* 2012; Lombardozzi *et al.* 2015; Smith *et al.* 2016; Mercado *et al.* 2018). However, Kattge and Knorr (2007) did not have enough data to represent tropical species in their meta-analysis. As a result, carbon models are likely biased in projecting tropical biome temperature responses. More recently, Kumarathunge *et al.* (2019) published updated algorithms including six datasets from tropical forests which will undoubtedly improve global carbon models (Zarakas *et al.* 2024). Even so, because tropical forests cycle a disproportionate amount of carbon, specific investigations of tropical photosynthetic responses to temperature based on plant function and growth strategy will further minimize uncertainty for this crucial biome (Booth *et al.* 2012).

151 There is strong evidence suggesting that, across the globe,  $T_{opt}$  is determined by the plant's current growth temperature (Berry and Björkman 1980; Kattge and Knorr 2007; Kumarathunge *et al.* 2019). Genetic variation also plays an important role in determining species' ability to acclimate and adjust to their growth temperatures (Berry and Björkman 1980; Yamori *et al.* 2014; Crous *et al.* 2022; but see Kumarathunge *et al.* 2019). However, it is still unclear whether this holds true within tropical ecosystems. Studies of photosynthetic temperature responses of tropical forest species provide evidence that *Topt* is either closely associated with



rarely incorporated into vegetation models (Lombardozzi *et al.* 2015; Smith *et al.* 2016; Mercado

 *et al.* 2018). Growth strategies are often characterized by their successional strategy, with some forms, such as early successional species and lianas, incorporating fast growth as juveniles, while late successional and evergreen species employ slower growth as juveniles (Bloom *et al.* 1985, Box *et al.* 1996; Wright *et al.* 2004; Michaletz *et al.* 2016). Due to higher radiation reaching deeper into the canopy, early successional forests have more variable land surface temperature fluxes than late successional forests (Cao and Sanchez-Azofeifa 2017), suggesting that seedlings 187 adapted to this environment may have a greater plasticity to adjust  $T_{opt}$  to their fluctuating growth environment. Studies of canopy species in Panama found that early successional seedlings had a higher *Topt* than late successional seedlings (Slot *et al.* 2016, Slot and Winter, 2018). However, those results were not replicated for mature canopy trees (Slot and Winter 2017a), suggesting that successional type *Topt* differences are primarily driven by trees at the immature seedling and sapling stages.

 Tropical trees with differing leaf habits (i.e., evergreen vs. deciduous) may also employ different temperature responses. For example, species with shorter-lived leaves have a greater variability in leaf phenotypes, making them more responsive to seasonal changes (Kitajima *et al.* 1997). Compared to longer-lived evergreen leaves, shorter-lived deciduous leaves are hypothesized to have broader photosynthetic temperature response curves (i.e., thermal niches; Michaletz *et al.* 2016). Broad- and needle-leaf evergreen species have been found less able to increase their growth rates in warmer temperatures than deciduous species (Way and Oren 2010; Way and Yamori 2014; Yamori *et al.* 2014; Reich *et al.* 2022). Recently, Crous *et al.* (2022) found that needleleaf evergreen species' photosynthetic and respiration rates declined more with warming compared to broadleaf evergreen species. This, in addition to longer-lived leaves having lower photosynthetic capacity (Niinemets 2007), and lower rates of photosynthesis

 (Wright *et al.* 2004), suggests that evergreen and deciduous species may have different capabilities to respond to their growth environment.

 Light availability may also play a role in modulating plant photosynthetic responses to temperature (Niinemets 2007). Models of canopy photosynthesis and global primary productivity often separate leaves into 'sun' and 'shade' leaves, as they have different photosynthetic responses to irradiance (Sinclair *et al.* 1976; De Pury and Farquhar 1997; Wang and Leuning 1998; Ryu *et al.* 2011). Because leaf temperature is strongly influenced by irradiance (Rey- Sánchez *et al.* 2016; Fauset *et al.* 2018; Miller *et al.* 2021; Crous *et al.* 2023), it should follow that sun leaves that have developed under higher irradiance are acclimated to operate at higher temperatures. However, comparisons of leaves growing in different light environments in tropical forests have found large differences in photosynthetic capacity but little to no differences in photosynthetic temperature response (Pearcy 1987; Hernández *et al.* 2020), or thermotolerance (Slot *et al.* 2019), between sun and shade leaves. The limited evidence that we have comparing tropical temperature responses of sun and shade leaves suggests that light may 218 play a large role in determining overall carbon gain but only a minor role for leaves' photosynthetic temperature responses.

 Rainfall and moisture regimes also play a role in controlling plant photosynthesis, which can lead to restrictions on temperature response parameters. In general, drier conditions can induce stomatal closure, slowing the rate of photosynthesis and decreasing tropical forest productivity (Cavaleri *et al.* 2017; Santos *et al.* 2018; Van Schaik *et al.* 2018; Kumarathunge *et al.* 2020; Mujawamariya *et al.* 2023). However, drier conditions are also associated with less rainfall and cloud cover, and a higher light environment can directly increase ecosystem productivity (Carswell *et al.* 2002). Ecosystem scale studies show gross primary productivity

 (GPP) can either increase in the dry season (Goulden *et al.* 2004; Yan *et al.* 2013; Wu *et al.* 2016; Green *et al.* 2020) or remain constant between seasons (Carswell *et al.* 2002; Yan *et al.* 2013; Guan *et al.* 2015), suggesting that tropical forests can sustain higher GPP during the higher dry-season atmospheric water stress if they are not stomatal conductance limited. Across two Panamanian tropical systems, a leaf level study showed that, when compared to a wet forest, seasonally dry forests can have higher rates of photosynthesis and higher optimum temperatures that correspond to their higher growth temperatures (Slot and Winter 2017a). Within a Puerto Rican tropical forest, drier soil was associated with higher optimum temperatures but lower rates of photosynthesis (Carter *et al.* 2020). These studies suggest that optimum temperatures could be positively correlated with drier tropical systems.

 In order to better understand tropical net photosynthetic and biochemical responses to temperature, we used a meta-analytic approach to quantify how photosynthetic temperature response parameters respond to different climate and growth environment factors using already established temperature response functions (Medlyn *et al.* 2002; June *et al.* 2004). We hypothesize that (1) light saturated photosynthetic optimum temperatures (*ToptA*) will be positively correlated with mean annual temperatures (MAT) due to positive shifts in *Vcmax* temperature response parameters. We similarly hypothesize that, due to indirect environmental effects of higher light availability, (2) temperature optima will decrease with rising aridity index (*AI*) (decrease in wetter ecosystems). We also compare temperature response variables of leaves grown in different light environments (sun vs. shade), growth environments (*in situ* vs. *ex situ* or field vs chamber/glasshouse), leaf habits (evergreen vs. drought semi-deciduous), and successional strategy (early vs. mid-late). We predicted that (3) sun leaves would have higher photosynthetic rates than shade leaves; but that *Topt* would not differ between different light

 environments. Additionally, we predicted that (4) early successional species *Topt* will not differ from late successional species and (5) broadleaf evergreen leaves would have a narrower thermal niche and lower *Topt* than semi-deciduous species. Lastly, we aimed to estimate the most important individual environmental drivers to best predict the temperature parameters of both net photosynthesis and the biochemical reactions driving photosynthesis.

*Methods*

*Meta-analysis data collection and selection*

 For this meta-analysis, we gathered datasets where photosynthetic measurements were collected at different leaf temperatures on woody (trees, shrubs, and lianas) tropical species. These data come in the form of net photosynthesis measured at saturating light conditions (*Asat*) vs. leaf temperature (*Tleaf*) response curves, *Asat* vs. *Tleaf* estimated from photosynthetic light response curves at different temperatures, biochemical parameters (*Vcmax* and *Jmax*) vs. *Tleaf* 264 response curves (estimated from net assimilation response to different leaf internal  $CO<sub>2</sub>$  concentrations, *A-C<sup>i</sup>* curves, measured at different temperatures), and measurements of *Asat* and *A-C<sup>i</sup>* curves at multiple ambient temperatures through time. Data were gathered from woody 267 species in forested systems within the tropical latitudes  $(23^{\circ}26'10.6" N, 23^{\circ}26'10.6" S)$ , including tropical montane systems. We obtained our data by approaching research groups for unpublished data and searching "photosynthesis" "tropical" "temperature" on Web of Science (Fig S1). This resulted in 18 datasets with representation in Africa (2), Oceana (6), North America (8), and South America (3). No studies were identified from the Asian continent. Site-specific climate data from the years 1970-2000 were collected from the WorldClim database

 (Fick and Hijmans, 2017) using provided latitude and longitudinal data. Latitude and longitude were designated as the location where plants grew, except for data from Read (1990), which were obtained with plants that were grown in a chamber. In this specific case, seeding source location was used for latitude and longitude and mean annual temperature (MAT) was designated as the growth chamber temperature. Data were extracted from the WorldClim database using the 'getData' function in the 'raster' package in R version 3.5.0 (R Core Team 2020). Aridity index (*AI*) was calculated as mean annual precipitation divided by mean annual potential evapotranspiration (Greve and Seneviratne 2015), where both variables were collected from WorldClim. Higher *AI* indicates a less arid system. *AI* was only used from *in situ* datasets, i.e*.,* we excluded greenhouse, growth chamber, and arboretum grown individuals from this analysis. Successional stage and leaf habit (raingreen semi-deciduous or evergreen; Poulter *et al.* (2015)) were either provided by the contributing data author or extracted from the literature. Species that were classified as "pioneer" and "shade-intolerant" were designated as "early successional". If the species was classified as "shade-tolerant" the species was considered "mid/late successional". When light environment information was available, we used author designations or classified ourselves; where growth chamber, greenhouse, "open" or "upper" canopy was considered "sun," and "understory" was considered shade. All samples grown in growth chambers, greenhouses, or transplant studies in arboretums were considered "*ex situ*". All other growth environments (i.e., "field collected") were designated as "*in situ*". We gathered photosynthetic data in two ways: 1) raw data in the form of photosynthetic response curves or 2) extraction from published articles. Data were digitized from published articles using Digitize It 2016 version 4.2.0 software (Alcasa). Raw data were provided from both published and

295 unpublished sources. Some of the datasets that were shared with us also included a 'warming'

296 treatment. For these data, we only used leaves grown in the 'control' environment.

297

### 298 *Net photosynthesis parameter extraction*

299 Within individual datasets, means of different species and canopy class (shaded or sun) 300 from the same study were treated as separate, independent samples (Curtis and Wang 1998). 301 The net photosynthetic temperature optimum of each sample was extracted from a peaked

302 curve (June *et al.* 2004):

$$
A_{sat} = A_{opt} \times e^{-\left(\frac{T_{leaf} - T_{optA}}{\Omega}\right)^2}
$$
 (1)

304

305 where  $A_{sat}$  (µmol m<sup>-2</sup> s<sup>-1</sup>) is the rate of net assimilation at the leaf temperature ( $T_{leaf}$ ) in  ${}^{\circ}C$ ,  $T_{optA}$ 306 (°C) is the optimum temperature for photosynthesis, and  $A_{opt}$  ( $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>) is the rate of 307 photosynthesis at  $T_{optA}$ .  $\Omega$ , or net photosynthetic thermal niche, is the temperature difference 308 from  $T_{optA}$  where photosynthesis declines to 37% of  $A_{opt}$ .  $\Omega$  (°C) describes the width of the 309 response curve peak, where wide curves have a higher  $\Omega$  and narrower curves have a lower  $\Omega$ . 310 Prior to fitting Equation 1, *Asat* from each dataset was individually inspected for outliers. Outliers 311 were removed only when they were clearly erroneous, such as  $A_{sat}$  < 0  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup> that were not 312 clearly caused by high temperatures. In addition, data points with  $C_i < 0$  were removed as they 313 were considered bad measurements. In total, we removed 402 data points, 2.79% of our *Asat* data. 314 To compare the rates of net photosynthesis across studies, we extracted the rate at 25 °C 315  $(A_{25})$  by allowing  $T_{leaf}$  to equal 25 in Equation 1 for each set of extracted temperature parameters. 316 This standard temperature was selected because it is similar to the average MAT (25.5  $\degree$ C) in our 317 dataset and is often used as a standard so photosynthetic rates are widely comparable across



*Biochemical parameter extraction*

 Biochemical rates, *Jmax* and *Vcmax*, were estimated from *A-C<sup>i</sup>* curves. Most datasets 330 collected  $A - C_i$  curves starting at an ambient  $CO_2$  concentration, 360-410 ppm.  $A - C_i$  curves were 331 obtained by gradually decreasing the  $CO<sub>2</sub>$  below ambient concentrations (to as low as zero ppm). CO<sub>2</sub> concentrations were then brought back up to ambient levels and then gradually increased to saturating concentrations (up to 2100 ppm). Prior to fitting the *A-C<sup>i</sup>* curves, data points outside 0  $\langle C_i \rangle$  < 2200 ppm were removed from the dataset as they were beyond the range of  $CO_2$  concentration given to the leaf. We further removed datapoints where *Asat* was smaller than -10 336 and greater than 70 µmol m<sup>-2</sup> s<sup>-1</sup> as they were not considered reasonable  $A_{sat}$  rates. In total we 337 removed less than 0.5% of total  $A - C_i$  datapoints.  $J_{max}$  and  $V_{cmax}$  were obtained using the default fit method with "Tcorrect = FALSE" in the 'fitaci' function from the 'plantecophys' package (Duursma 2015) in R version 3.5.0 (R Core Team, 2020), which extracts parameters using the Farquhar, von Caemmerer, and Berry model (FvCB model; (Farquhar *et al.* 1980, von

 Caemmerer and Farquhar 1981). We further looked at the fitted A-*C<sup>i</sup>* curves and individually removed curves with poor fits. We further removed curves where fitted *Jmax* and *Vcmax* values 343 were less than 0  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>, as this is not possible for correctly fit curves. After the initial data exclusion, we removed outliers where *Jmax* or *Vcmax* were clearly erroneous by looking at qqplots and histograms of each dataset. In total, 7.8% or 102 A-*C<sup>i</sup>* curves were removed from the initial 346 dataset.

347 Biochemical temperature response parameters for *Jmax* and *Vcmax* were extracted using the 348 peaked Arrhenius function (Medlyn *et al.* 2002):

349

$$
T_k = (k_{opt}) \frac{H_d \exp\left(\frac{E_d(T_k - T_{opt})}{(T_k \exp(t))}\right)}{H_d - E_d \left[1 - \exp\left(\frac{H_d(T_k - T_{opt})}{(T_k \exp(t))}\right)\right]}
$$
(2)

351

352 where  $T_k$  is the measured leaf temperature in Kelvin,  $(k_{opt})$  is the value of  $J_{max}$  or  $V_{cmax}$  at the 353 optimum temperature ( $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>),  $E_a$  is the activation energy in the Arrhenius function (kJ 354 mol<sup>-1</sup>), or exponential increase in  $J_{max}$  or  $V_{cmax}$  before  $T_{opt}$ ,  $H_d$  is the deactivation energy of  $J_{max}$  or 355  $V_{cmax}$  after  $T_{opt}$  (kJ mol<sup>-1</sup>), and *R* is the universal gas constant (8.314 JK<sup>-1</sup>mol<sup>-1</sup>). To avoid over-356 parameterization of the temperature response function, we set  $H_d = 200 \text{ kJ mol}^{-1}$  and estimated 357 *Topt*, *kopt*, and *E<sup>a</sup>* from Equation 2. Each individual curve was examined and curves were removed 358 if  $T_{opt}$ ,  $k_{opt}$ , or  $E_a$  values were over or underestimated, e.g. visually estimated  $T_{opt}$  was clearly 359 higher or lower than model estimations which was often due to too few temperatures used to 360 produce the curve, resulting in 35 *Vcmax* and 35 *Jmax* temperature response curves. 361 We extracted the rate of  $V_{cmax}$  ( $V_{25}$ ) and  $J_{max}$  ( $J_{25}$ ) at 25 °C from *A-C<sub>i</sub>* curves measured from

362 temperatures ranging from 20-30  $\degree$ C by setting Tcorrect = "TRUE" in the "fitaci" function. The

363 ratio between  $J_{25}$  and  $V_{25}$  (*J:V*) was calculated by dividing  $J_{25}$  by  $V_{25}$  for each individual sample. 364 This resulted in 295 samples in our *V<sup>25</sup>* and *J<sup>25</sup>* datasets. Version 1.4 of the "plantecophys" 365 package defaults to using temperature fitting parameters estimated from a global analysis of 366 photosynthetic temperature responses that estimated values using (Medlyn *et al.* 2002): 367

368 
$$
T_k = k_{25} exp \left[ \frac{E_a (T_k - 298)}{(298RT_k)} \right] \frac{1 + exp \left( \frac{298\Delta S - H_d}{298R} \right)}{1 + exp \left( \frac{T_k \Delta S - H_d}{T_k R} \right)}
$$
(3)

369

370 Where  $\Delta S$  is an entropy term. We estimated  $J_{25}$  and  $V_{25}$  using the default "global" parameters 371 and this study's tropical estimations of  $E_a$  and  $\Delta S$  (Table S3) and made comparisons of the two 372 fitting estimations.

373

### 374 *Meta-analytic statistical analyses*

375 Biases for sample size were accounted for by weighting each extracted parameter with 376 the number of observations that were used in each temperature response curve. The weighting 377 factor was calculated as (Hedges and Olkin 1985; Gurevitch *et al.* 1992):

378

$$
J = 1 - \left(\frac{3}{4(n-1)}\right) \tag{4}
$$

380

 where *J* is the weighting factor and n is the number of data points used to fit each temperature response curve (Fig. S2). The weighted mean was incorporated into the linear model by adding *J* into the 'weights' weighting factor component of the 'lmer' function the 'lme4' package in R (Bates *et al.* 2015). All data analyses were performed in R version 3.5.0 (R Core Team, 2021).

 Mixed effects models were used to compare global and tropical *Vcmax* and *Jmax* activation 386 energies ( $E_{aV}$  and  $E_{aJ}$ , respectively) and entropy terms ( $\Delta S_V$  and  $\Delta S_J$ , respectively), where data source was used as the random intercept. Mixed effects models were also used to investigate 388 relationships between  $A_{sat}$  and biochemical parameters ( $T_{opt}$ ,  $T_{opt}$ ,  $T_{opt}$ ,  $A_{25}$ ,  $V_{25}$ ,  $J_{25}$ ,  $\Omega$ ,  $E_{aV}$ , and *EaJ*) and individual climate variables. We found high collinearity between MAT and elevation (Fig. S3); therefore, elevation was removed from the individual bivariate regression models. We removed elevation as a continuous variable and grouped the data into four elevational groups (0- 500m, 501-1000m, 1001-2000m, and >2000m) to visually show the role that elevation played in our climate range for all bivariate regressions. Mixed effect models were also used to compare 394 leaf habit, successional type, and growth conditions, using an  $\alpha$  < 0.05. Due to available characterizations for our dataset, light environment (sun or shade) and leaf habit (deciduous or evergreen) were compared only for *Asat* parameters. Successional type (early or late) and growth environment (*in* or *ex situ*) were compared for both *Asat* and biochemical parameters (summary of samples used in each categorical analysis included in Table S2). Estimated *J<sup>25</sup>* and *V<sup>25</sup>* were compared between the default "plantecophys" package and our parameter estimates using a mixed effects model as described above.

 High variance inflation factors (VIF), a means of identifying potential collinearity, were assessed when we included both MAT and elevation in the same multivariate model, where full 403 models that included all four climate variables (MAT, AI,  $T_{range}$ , elevation) had at least one 404 variable VIF > 2 (VIF range 2.02-648.53). VIF on the full model was calculated using "vif" function in base R. Therefore, we used hierarchical partitioning to quantify which climate 406 variable had the highest explanatory power on parameter  $(T_{opt}, T_{opt}, T_{opt}, A_{25}, V_{25}, J_{25}, \Omega, E_{av}$ and *EaJ*) variance using the "rdacca.hp" package in R (Lai *et al.* 2022). Hierarchical partitioning

 is used in instances of high VIF because it estimates individual importance of predictors in all model subsets, where the subsets also include the full model (Lai *et al.* 2022). The individual effects were estimated via hierarchical partitioning and were calculated from the sum of the 411 calculated unique and shared contribution to the overall model's adjusted  $\mathbb{R}^2$ , where the model includes all individual variables of interest. The individual effect can be negative if the unique or shared contribution is negative due to high multicollinearity. In this calculation, the individual 414 effects were added to equal the total adjusted  $\mathbb{R}^2$ . 

**Results**

*Comparisons of biochemical estimations from global and tropical parameters*

 Global estimates of biochemical activation energies and entropy terms yielded higher biochemical parameter rates compared to tropical data derived estimates, suggesting that studies 420 in tropical systems would overestimate  $V_{25}$  and  $J_{25}$  if using global values.  $V_{25}$  and  $J_{25}$  estimated 421 from global datasets were both approximately 7% higher than those from tropical parameters (Table S3; Fig. S4A-B), resulting in no discernible difference in *JV* between parameter estimates (Fig. S4C).

*Primary climate variable influences on temperature parameters*

In bivariate regressions, the net photosynthetic and electron transport optimum

temperature increased with increasing temperature, while the maximum Rubisco carboxylation

- optimum temperature did not. *ToptA* was positively related with MAT, with MAT alone
- explaining 37% of *ToptA* variance (Fig. 1A; Table 2). *ToptA* did not strongly respond to *AI* or *Trange*
- (Figs. 2B, S5A). *ToptV* did not respond to any of the three climate variables (Figs. 1C-D, S5B;

431 Table 2). *ToptJ* increased with rising MAT, which explained 14% variation, and *ToptJ* did not 432 respond to *AI* or *Trange* (Figs. 1E-F, S5C).

433 While net photosynthetic rate did not show clear relationships with climate variables, the 434 rates of photosynthetic biochemical reactions decreased with a warmer climate. *A<sup>25</sup>* did not 435 respond to with MAT, *AI*, or *Trange* (Figs. 2A-B, S6A; Table 2). *V<sup>25</sup>* decreased as MAT rose 436 (marginal  $R^2 = 0.20$ ; Fig. 2C), did not respond to *AI* (Fig. 2D), and decreased with wider  $T_{range}$ 437 (marginal  $R^2 = 0.18$ ; Fig. S6B). Similarly,  $J_{25}$  decreased as MAT increased (marginal  $R^2 = 0.41$ ; 438 Fig. 2E), did not respond to AI (Fig. 2F), and increased as  $T_{range}$  increased (marginal  $R^2 = 0.28$ ; 439 Fig. S6C). The ratio between  $J_{max}$  and  $V_{cmax}$  at 25 °C (*J:V*) decreased with rising MAT (marginal 440  $R^2 = 0.28$ ; Fig. 3A), did not respond to *AI* (Fig. 3B; Table 2), and slightly increased with a wider 441 *T<sub>range</sub>* (marginal R<sup>2</sup> = 0.06; Fig. 3C). Neither net photosynthetic thermal niche (Ω) nor the 442 activation energy for *Vcmax* and *Jmax* responded to any climate variables (Fig. S7; Table 2).

443

### 444 *Growth environment influences on temperature response parameters*

445 Variables describing the rate of a photosynthetic process were higher in sun compared to 446 shade leaves, but temperature response parameters did not differ. Sun and shade leaf *ToptA* were 447 not significantly different from one another (Satterthwaite's method;  $p = 0.786$ ; Fig. 4A).  $A_{25}$  of 448 sun leaves was 1.5 times higher than shade leaves ( $p = 0.008$ ; Fig. 4B). Similar to  $T_{optA}$ , there 449 was no difference in  $\Omega$  between the two light environments ( $p = 0.210$ ; Fig. 4C). *V*<sub>25</sub> and *J*<sub>25</sub> of 450 sun leaves were 88% and 63% higher than the rate of shade leaves, respectively, (both  $p < 0.001$ ; 451 Fig. 4D-E), and *J:V* was slightly  $(\sim 10\%)$  higher in shade than sun leaves ( $p = 0.022$ ; Fig. 4F). 452 Plants grown *in situ* had higher biochemical response rates than *ex situ* grown plants, but 453 that did not lead to differences in *Asat* rates or parameters. There were no clear differences

454 between plants grown *in* or *ex situ* for  $A_{sat}$  parameters and rates  $T_{optA}$  ( $p = 0.085$ ),  $A_{25}$  ( $p = 0.096$ ), 455 or *Ω* (p = 0.313; Figs. S8A-C). *ToptV* (p = 0.974; Figs. S7D) and *EaV* (p = 0.102; Figs. S8F) did 456 not differ between *in* and *ex situ*, but plants grown *ex situ* had 40% higher  $V_{25}$  ( $p = 0.030$ ; Figs. 457 S8E). *ToptJ* did not differ between growth environments (p = 0.802; Fig. S8G), *J<sup>25</sup>* for plants 458 grown *ex situ* trended 48% higher than those grown *in situ* ( $p = 0.054$ ; Fig. S8H), and  $E_{aJ}$  was 459 around double in *in situ* than *ex situ* grown plants (p =0.002; Fig. S8I). Lastly, *J:V* also was not 460 different between the two growth environments ( $p = 0.696$ ; Fig. S8J).

461

#### 462 *Effects of plant functional type on temperature response parameters*

463 *ToptA* was higher in drought (semi-) deciduous, or raingreen, species compared to 464 broadleaf evergreen species, but other net photosynthetic temperature response rates and 465 variables did not differ between the two leaf habits.  $T_{optA}$  was  $\sim$  1 °C higher in drought (semi-) 466 deciduous compared to evergreen species ( $p = 0.009$ ; Fig. 5A). There were no differences 467 between evergreen and deciduous species for  $A_{25}$  (p = 0.347; Fig. 5B) or  $\Omega$  (p = 0.197; Fig. 5C). 468 Optimum temperatures of photosynthesis did not vary between successional types, but 469 rates of photosynthetic responses and the width of the photosynthetic responses were higher in 470 early compared to mid/late successional species. Early and mid/late successional species did not 471 differ in  $T_{optA}$  (p = 0.955; Fig 6A).  $A_{25}$  and  $\Omega$  (both p < 0.001; Fig. 6B-C) in early successional 472 species were ~83% and 32% higher than in mid/late successional species, respectively. *ToptV* did 473 not differ between successional types  $(p = 0.502; 6D)$  but, in terms of rates, mean early 474 successional *V<sup>25</sup>* was 61% higher than late successional species (p < 0.001; Fig. 6E). There were 475 no differences between successional types for *J:V* ( $p = 0.936$ ; Fig. 6F).  $T_{optJ}$  did not differ

476 between successional types ( $p = 0.644$ ; Fig 6G) but early successional species  $J_{25}$  was around 477 double that of late successional species ( $p < 0.001$ ; Fig. 6H).

478

#### 479 **Hierarchical Partitioning**

480 Except for *ToptA*, hierarchical partitioning revealed that no single climate or growth 481 environment variable explained a high amount of variation in our photosynthetic parameters. The 482 strongest predictor for  $T_{optA}$  variation was the elevation (individual adj  $R^2 = 0.159$ ; Fig 7A). With 483 a full model  $R^2 = 0.018$ , climate was not a strong predictor for  $A_{25}$ ; however, MAT (adj  $R^2 =$ 484 0.017) had a slightly stronger individual effect on  $A_{25}$  than other predictors (Fig. 7B).  $\Omega$  was 485 more strongly predicted by  $T_{range}$  (adj  $R^2 = 0.170$ ; Fig. 7C).  $T_{optV}$  was most strongly predicted by 486 *AI* (adj  $R^2 = 0.032$ ; Fig. 7D),  $V_{25}$  was slightly more predicted by MAT (adj  $R^2 = 0.053$ ; Fig. 7E), 487 and  $E_{aV}$  was most strongly predicted by MAT (adj  $R^2 = 0.128$ , Fig. 7F).  $T_{optJ}$  was not well 488 predicted by any climate variables; however, *Trange* explained slightly higher variation than other 489 variables (adj  $R^2$  = -0.040; Fig. 7G). Variance of  $J_{25}$  was more strongly explained by MAT (adj 490  $R^2 = 0.125$ ; Fig. 6H).  $E_{aJ}$  was more strongly driven by MAT (adj  $R^2 = 0.068$ ; Fig. 7I). *J*:V was 491 best explained by elevation (adj  $R^2 = 0.060$ ; Fig. 7J).

492

### 493 **Discussion**

### 494 *Climate drivers of the optimum temperature of photosynthesis*

 Globally (Kattge and Knorr 2007; Kumarathunge *et al.* 2019; Crous *et al.* 2022) and in tropical ecosystems (Tan *et al.* 2017), studies have found that the photosynthetic optimum temperature of net photosynthesis increases as growth temperatures increase. In partial support of our first hypothesis, the optimum temperatures of net photosynthesis (*ToptA*) and

 photosynthetic electron transport *(ToptJ*) rose with increasing mean annual temperature (MAT; Fig. 1A,E); however, the optimum temperature of Rubisco carboxylation *(ToptV*) did not (Fig 1C). 501 The slope of our tropical species responses to MAT ( $T_{optA}$  slope:  $0.59 \pm 0.15 \degree \text{C} \degree \text{C}^{-1}$ ; Table 3), is similar to and has overlapping standard error with a global analysis of *ToptA* response to growth 503 temperature ( $T_{optA}$  slope:  $0.62 \pm 0.1$  °C per increase in growth temperature; Kumarathunge *et al.*  2019), providing no evidence that different algorithms should be used to model tropical and global *ToptA* responses. *ToptJ* in our study also had a similar positive response as the global 506 analysis (current study:  $T_{optJ}$  slope:  $0.38 \pm 0.17$ MAT; Kumarathunge:  $T_{optJ}$  slope:  $0.63 \pm 0.17$  0.2*Tgrowth*; Kumarathunge *et al.* 2019). Our results for the optimum temperatures of *Vcmax* were not as consistent with Kumarathunge *et al.* (2019), where our *ToptV* did not respond to MAT (*ToptV* 509 slope:  $0.26 \pm 0.21$ MAT; Table 3), but the global analysis showed a positive relationship with 510 increasing growth temperature  $(T_{optV}$  slope:  $0.71 \pm 0.2T_{growth}$ : Kumarathunge *et al.* 2019). We 511 note, however, that our meta-analysis of tropical species' biochemical parameters (19.6 – 27.5) 512 °C) has narrower temperature range than the global meta-analysis ( $\sim$ 3.0 – 30.0 °C; Kumarathunge *et al.* 2019) which, along with the high variation in parameter values at each point along the MAT axis, might limit our ability to detect data trends. Additionally, the lower *ToptV*  MAT slope response provides some support for the common hypothesis that tropical species have adapted to more narrow climate envelopes and do not strongly respond to variations in growth temperature, potentially resulting in a reduced capability to acclimate to warmer temperatures (Janzen 1967; Cunningham and Read 2003; Dusenge *et al.* 2021). This idea is further supported by Kumarathunge *et al.* (2019) which found optimum temperature responses to growth temperature were more strongly driven by acclimation to growth temperature than adaptation to climate of origin. In a recent analysis across latitudes, Crous *et al*. (2022) found

 more negative photosynthetic responses to warmer temperatures in the tropics compared to cooler climates, suggesting constrained acclimation. Our *Jmax* and *Vcmax* temperature response 524 datasets cover MAT across a reduced range  $(19.6 - 27.5 \text{ °C})$  than our  $A_{sat}$  dataset  $(11.8 - 30.0 \text{ m})$  $\degree$ C). Additional studies investigating these biochemical parameters would enable the assessment of whether tropical forest species have systematically different temperature responses of these parameters than extra-tropical species.

 Contrary to our hypothesis, aridity index alone was not a strong predictor of photosynthetic temperature responses. None of our photosynthetic parameters or rates responded to aridity index (Figs. 1, 2, 3, S7). Compared with trees in temperate zones, fewer studies in the 531 tropics have investigated how rainfall affects  $T_{opt}$ .  $T_{optA}$  was found to increase as soils dry in a Puerto Rican tropical forest (Carter *et al.* 2020) and a savanna grassland ecosystem (Ma *et al.* 2017). However, Kumarathunge *et al.* (2020) found that the optimum temperature for tropical tree growth increases with water addition. Hierarchical partitioning showed aridity index as the most important measured climate component controlling *ToptV*; however, the individual aridity index effect on *ToptV* was very low (Fig. 7D). To date, the few studies that have investigated large-scale environmental controls on the biochemical components of photosynthesis focus solely on how temperature controls these important model parameters (Kattge and Knorr 2007; Tan *et al.* 2017; Kumarathunge *et al.* 2019; Crous *et al.* 2022). Even though these results suggest that aridity does not play a key role in controlling photosynthetic temperature responses, both temperature and rainfall play significant roles in modeled reductions in carbon gain in the Amazon rainforest (Galbraith *et al.* 2010). Future studies should investigate how other climate factors, such as aridity, influence photosynthetic optimum temperatures, as we know that a key

 constraint on photosynthetic optimization is the balance of carbon gain against water loss (Bloom *et al.* 1985; Wang *et al.* 2017).

- 
- 

### *Biochemical limitations at high temperatures*

 Limitations to the optimum temperature of net photosynthesis at moderate growth temperatures are often attributed to limitations of Rubisco carboxylation temperature response parameters (Lin *et al.* 2012; Yamaguchi *et al.* 2016), although not always (Wise *et al.* 2004; Cen and Sage 2005). When plants are grown at elevated temperatures, measured photosynthesis is increasingly limited by carboxylation as temperature rises, a trend that is driven both by stomatal 553 limitations on  $CO_2$  substrate and by the high temperature sensitivity of Rubisco carboxylation (Brooks and Farquhar 1985; Hikosaka *et al.* 2006). However, optimality theory of photosynthetic 555 capacity suggests that resources allocated to  $J_{max}$  and  $V_{cmax}$  at 25 °C are disproportionally reduced under warmer temperatures, resulting in reduced *J:V* (Smith and Keenan 2020; Wang *et al.* 2020). The limitation to *Jmax* is due to high temperatures reducing electron transport through PSII (Havaux 1996), and a greater investment in Rubisco carboxylation relative to electron transport to counteract the increased photorespiration at higher temperatures (Smith and Keenan 2020). This is supported by global meta-analyses showing declining *J:V* with increasing growth temperature (Kumarathunge *et al.* 2019; Crous *et al.* 2022). Our results support this, where both *V<sup>25</sup>* and *J<sup>25</sup>* decreased with increasing MAT but *J<sup>25</sup>* declined at a steeper rate (Fig. 2), resulting in a decreasing *J:V* with rising MAT (Fig. 3). Across our temperature range, our results are not consistent with those of previous global meta-analyses (Medlyn *et al.* 2002; Hikosaka *et al.* 2006; Kattge and Knorr 2007; Kumarathunge *et al.* 2019), where neither of our activation energy 566 terms of  $J_{max}$  ( $E_{aJ}$ ) or  $V_{cmax}$  ( $E_{aV}$ ) responded to temperature (Fig S8).  $E_{aV}$  activation energy is a



 *Photosynthetic differences between growth conditions, deciduousness, and successional types*  We found that the rate of photosynthesis was higher in sun leaves but there were no *Topt* differences between sun and shade leaves (Fig. 4), similar to the few studies that have investigated differences in *in situ* tropical photosynthetic responses to different canopy light conditions (Pearcy 1987; Slot *et al.* 2019; Hernández *et al.* 2020; but see Carter *et al.* 2021). Other biomes show similar results, and studies investigating differences in *ToptA* between upper canopy and understory leaves have found that *ToptA* either does not differ (Carter and Cavaleri 2018), or *ToptA* is higher in the upper canopy leaves (Jurik *et al.* 1988). Niinemets *et al.* (1999)

 showed that the optimum temperature of electron transport is higher in the upper canopy (higher incident radiation on average) compared to lower canopy leaves (lower spectral quality, lower average incident radiation), suggesting that the biochemical process of photosynthesis associated with light can adjust to different light conditions and higher temperatures. Within the tropics, Carter *et al.* (2021) found that *ToptA* decreased as canopy height and light increased, likely due to VPD induced stomatal limitations. Hernández *et al.* (2020) found trends toward higher *ToptV* in Panamanian sun leaves, yet *ToptJ* did not differ between light conditions. We did not have enough *Vcmax* or *Jmax* data classified as "shaded" and were unable to make a robust sun-shade comparison within our dataset. Even though we were able to make a comparison between *Asat* sun and shade leaves, we only had eight samples where shade leaves were measured (Table S2), suggesting we need much more temperature response measurements comparing sun and shade leaves in tropical forests. Even so, the growing evidence in tropical forests suggests that light conditions do not strongly control tropical *Topt*, and we may not need to distinguish between sun and shade leaves when modeling temperature responses in tropical forest canopies.

 Even though leaf habits, such as evergreen and deciduous species, often have different photosynthetic temperature responses (Yamori *et al.* 2014), global vegetation models usually do not implement separate temperature response parameters for different plant functional types due to insufficient data (Lombardozzi *et al.* 2015; Smith *et al.* 2016; Mercado *et al.* 2018). In the current study, *A<sup>25</sup>* did not differ but evergreen leaves had a slightly lower *ToptA* than semi-drought deciduous leaves (Fig. 5A,B). This suggests that global models should differentiate between "broadleaf evergreen tropical" and "semi-deciduous raingreen tropical" forests (Poulter *et al.* 2015), rather than considering all tropical regions as "broadleaf evergreen tropical. Although we did find a trend toward higher *ToptA* in semi-deciduous species, we note that all species labeled as

 'semi-deciduous' came from the same study (Slot and Winter 2017a), which had the highest 614 MAT (26.6 °C) of all the study sites included in the  $A_{sat}$  dataset. No species in our  $A - C_i$  dataset was characterized as either 'deciduous' or 'semi-deciduous' (Table S1), preventing any analysis on differences between leaf habit for *Jmax* and *Vcmax* data. Greater efforts should be made to better characterize differences between different plant functional types within the tropics and these data should be used to assess how vegetation models define tropical forest plant functional types.

 Generally, fast growing, early successional species have higher rates of photosynthesis (Wright *et al.* 2004). Our results agreed with this theory and, similar to Ziegler *et al.* (2020) and Mujawamariya *et al.* (2023), we found higher *A25*, *V25*, and *J<sup>25</sup>* in early successional species. Additionally, early successional species in a tropical dry forest were found to reside in higher temperature environments due to the higher light environment and more open forest structure in an early successional forest (Cao and Sanchez-Azofeifa 2017), suggesting that early successional seedlings and saplings might have higher optimum temperatures. However, our study that combined all species growth stages found no differences between successional types for *ToptA* (Fig. 6). Our results support a lack of clear differences between canopy species of different successional types in Slot and Winter (2017b) but differ from the results of Slot *et al.* (2016), which found higher optimum temperatures in early successional seedlings. Here, we highlight that Slot *et al.* (2016) was conducted on seedlings instead of canopy trees (Slot and Winter, 2017b). Future work should investigate differences in early successional seedling vs mature 632 canopy tree optimum temperatures. We did find that the net photosynthetic thermal niche  $(\Omega)$  was broader for early successional species than late successional species (Fig 6C). This is consistent with theory on 'fast' species with high rates of photosynthesis, as these species tend to invest in traits that allow productivity under a wide range of temperatures (Michaletz *et al.*

 2016). A wider thermal niche is likely beneficial to early successional forests that experience a wider, more dynamic range of temperatures (Holbo and Luvall 1989).

### *Opportunities for better parameterized functions*

 We present trends for the temperature parameters of net photosynthetic and biochemical processes of net photosynthesis in tropical regions. However, both stomatal conductance and daytime respiration can also play large roles in controlling photosynthetic temperature responses (Lin *et al.* 2012). Stomatal conductance, or vapor pressure deficit (VPD) which is the primary climate variable controlling stomatal conductance (Farquhar and Sharkey 1982), have been estimated to be the strongest predictors of photosynthetic decline with climate warming in the tropics (Lloyd and Farquhar 2008; Wu *et al.* 2017; Smith *et al.* 2020; Slot *et al.* 2024). This relationship between temperature, moisture, and stomatal conductance should also be investigated across tropical forests and is critical to understand photosynthetic responses to temperature as tropical forests become hotter and drier (Malhi *et al.* 2008). Further, our hierarchical partitioning could be further improved if we had included leaf functional traits. Most of our photosynthetic parameters were not well explained by any environmental factors. A meta- analysis by Atkin *et al.* (2015) found that plant functional types (broadleaf, conifer, grass type, shrubs) had the most explanatory power for predicting the rate of respiration globally. In addition, other plant trait factors, such as leaf nitrogen and leaf mass per area also improved their predictive models (Atkin *et al.* 2015). Including other factors, such as leaf habit or growth type (e.g., evergreen or deciduous; successional type) could provide valuable information for tropical biome photosynthesis modeling, and substantial efforts should be made to collect a larger variation of these data types, which were not available for many of the studies we analyzed. We

also note that this study presents results that under-represent African and Asian tropical forests.

Data from these regions could improve photosynthetic temperature response models.

*Conclusions*

 This study reports new predictive equations that describe photosynthetic temperature responses of tropical trees to different climate factors and describes pan-tropic differences related to plant growth conditions, growth habits, and successional strategies. Our novel analysis focusing on tropical woody species shows that *ToptA* and *ToptJ* responses to mean temperatures tended to align 667 with global meta-analyses; however, the optimum temperature of  $T_{optV}$  did not align with results found globally. A lower slope of photosynthetic biochemical parameter's *Topt* against MAT for tropical ecosystems suggests a lower capacity for these ecosystems to keep apace of climate change. While global carbon models should consider acclimation of the temperature response of photosynthetic parameters in order to allow for plant plasticity, the lower capacity for this response in tropical ecosystems should also be considered when making projections of ecosystem responses to climate change. Importantly, we did not find different temperature optimums between sun/shade leaves or successional types, but we did find differences between evergreen and semi-deciduous species' optimum temperatures. Vegetation models often define these systems solely as "broadleaf evergreen tropical", but functional types within tropical biomes have distinct temperature responses between "broadleaf evergreen tropical" and "semi- deciduous raingreen tropical" that should be considered to accurately represent tropical or global carbon dynamics.

#### **Acknowledgements**

Author contributions: KRC, MAC, AR, SPS, KSE, RJN, APW, PBR, SCR, TEW conceived of

- and designed the study, KRC and ECS organized and analyzed the data. KRC and MAC wrote
- the manuscript. KRC, AR, KSE, OA, NHAB, AWC, ZC, KYC, CED, MED, JRE, JFS, JWGK,
- ACM, BEM, PM, JR, MS, EST, JU, AV, KW, JW collected and contributed data to the study.
- All authors contributed to writing and editing the final manuscript.
- 

### **Funding Information**

 This work was supported by US Geological Survey John Wesley Powell Center Working Center for Analysis and Synthesis. Funding was also provided by U. S. Department of Energy Office of Science, Biological and Environmental Research Program awards [DE-SC-0012000, DE-SC- 0011806, DE-SC-0018942, 89243018S-SC-000014, and 89243018S-SC-000017]. Additional funding and support was provided by USDA Forest Service International Institute of Tropical Forestry (IITF). All research conducted at IITF is supported by the University of Puerto Rico. ORNL is managed by UT-Battelle, LLC, for the DOE under contract [DE-AC05-1008 00OR22725]. APW, SPS, AR, and KSE was supported by the Next Generation Ecosystem Experiments-Tropics (NGEE Tropics), funded by the U.S. Department of Energy, Office of Science, Office of Biological and Environmental Research and AR, KSE and SPS were also partially supported by the United States Department of Energy contract No. [DE-SC0012704] to Brookhaven National Laboratory, and AR and KSE by the U.S. Department of Energy Contract No. [DE-AC02-05CH11231] to Lawrence Berkeley National Laboratory. KYC gratefully acknowledges the Australian Research Council [DE160101484] supporting data collection on some Australian species. The contribution of PBR was supported by the U.S. NSF Biological Integration Institutes grant [DBI-2021898]. Any use of trade, firm, or product names is for



- **Brooks A, Farquhar GD**. **1985**. Effect of temperature on the CO2/O2 specificity of ribulose-
- 1,5-bisphosphate carboxylase/oxygenase and the rate of respiration in the light Estimates from
- gas-exchange measurements on spinach. *Planta* **165**: 397–406.
- **von Caemmerer S, Farquhar GD**. **1981**. Some relationships between the biochemistry of
- photosynthesis and the gas exchange of leaves. *Planta* **153**: 376–387.
- **Cao S, Sanchez-Azofeifa A**. **2017**. Modeling seasonal surface temperature variations in
- secondary tropical dry forests. *International Journal of Applied Earth Observation and*
- *Geoinformation* **62**: 122–134.
- **Carswell FE, Costa AL, Palheta M,** *et al.* **2002**. Seasonality in CO2 and H2O flux at an eastern
- Amazonian rain forest. *Journal of Geophysical Research: Atmospheres* **107**: LBA 43-1-LBA 43- 16.
- **Carter KR, Cavaleri MA**. **2018**. Within-canopy experimental leaf warming induces
- photosynthetic decline instead of acclimation in two Northern hardwood Species. *Frontiers in*
- *Forests and Global Change* **1**: doi: 10.3389/ffgc.2018.00011.
- **Carter KR, Wood TE, Reed SC,** *et al.* **2020**. Photosynthetic and respiratory acclimation of
- understory shrubs in response to in situ experimental warming of a wet tropical forest. *Frontiers*
- *in Forests and Global Change* **3**: 1–20.
- **Carter KR, Wood TE, Reed SC, Butts KM, Cavaleri MA**. **2021**. Experimental warming
- across a tropical forest canopy height gradient reveals minimal photosynthetic and respiratory
- acclimation. *Plant Cell and Environment* **44**: 2879–2897.
- **Cavaleri MA, Coble AP, Ryan MG, Bauerle WL, Loescher HW, Oberbauer SF**. **2017**.
- Tropical rainforest carbon sink declines during El Niño as a result of reduced photosynthesis and increased respiration rates. *New Phytologist*.
- **Cavaleri MA, Reed SC, Smith WK, Wood TE**. **2015**. Urgent need for warming experiments in
- tropical forests. *Global Change Biology* **21**: 2111–2121.
- **Cen YP, Sage RF**. **2005**. The regulation of Rubisco activity in response to variation in
- temperature and atmospheric CO2 partial pressure in sweet potato. *Plant Physiology* **139**: 979–
- **Choury Z, Wujeska-Klause A, Bourne A,** *et al.* **2022**. Tropical rainforest species have larger
- increases in temperature optima with warming than warm-temperate rainforest trees. *New Phytologist* **234**: 1220–1236.
- **Cox AJF, Hartley IP, Meir P,** *et al.* **2023**. Acclimation of photosynthetic capacity and foliar
- respiration in Andean tree species to temperature change. *New Phytologist* **238**: 2329–2344.
- **Crous KY, Cheesman AW, Middleby K,** *et al.* **2023**. Similar patterns of leaf temperatures and
- thermal acclimation to warming in temperate and tropical tree canopies. *Tree Physiology* **43**: 1383–1399.
- **Crous KY, Uddling J, De Kauwe MG**. **2022**. Temperature responses of photosynthesis and
- respiration in evergreen trees from boreal to tropical latitudes. *New Phytologist* **234**: 353–374.
- **Cunningham S, Read J**. **2002**. Comparison of temperate and tropical rainforest tree species:
- photosynthetic responses to growth temperature. *Oecologia* **133**: 112–119.
- **Cunningham SC, Read J**. **2003**. Do temperate rainforest trees have a greater ability to
- acclimate to changing temperatures than tropical rainforest trees? *New Phytologist* **157**: 55–64.
- **Curtis PS, Wang X**. **1998**. International Association for Ecology A Meta-Analysis of Elevated
- CO₂ Effects on Woody Plant Mass , Form , and Physiology. *Oecologia* **113**: 299–313.
- **Diffenbaugh NS, Scherer M**. **2011**. Observational and model evidence of global emergence of
- permanent, unprecedented heat in the 20th and 21st centuries. *Climatic Change* **107**: 615–624.
- **Dixon RK, Brown S, Houghton RA, Solomon AM, Trexler MC, Wisniewski J**. **1994**. Carbon
- pools and flux of global forest ecosystems. *Science* **263**: 185–190.
- **Doughty CE, Goulden ML**. **2008**. Are tropical forests near a high temperature threshold?
- *Journal of Geophysical Research* **113**: G00B07.
- **Doughty CE, Keany JM, Wiebe BC,** *et al.* **2023**. Tropical forests are approaching critical temperature thresholds. *Nature* **621**: 105–111.
- **Dusenge ME, Duarte AG, Way DA**. **2019**. Plant carbon metabolism and climate change:
- elevated CO2 and temperature impacts on photosynthesis, photorespiration and respiration. *New Phytologist* **221**: 32–49.
- **Dusenge ME, Wittemann M, Mujawamariya M,** *et al.* **2021**. Limited thermal acclimation of
- photosynthesis in tropical montane tree species. *Global Change Biology* **27**: 4860–4878.
- **Duursma RA**. **2015**. Plantecophys An R Package for Analysing and Modelling Leaf Gas Exchange Data. *PLoS ONE* **10**: e0143346.
- **Farquhar GD, Caemmerer S Von, Berry J a. 1980**. A biochemical model of photosynthesis
- CO2 fixation in leaves of C3 species. *Planta* **149**: 78–90.
- **Farquhar GD, Sharkey TD**. **1982**. Stomatal conductance and photosynthesis. *Annual Review of Plant Physiology* **33**: 317–345.
- **Fauset S, Freitas HC, Galbraith DR,** *et al.* **2018**. Differences in leaf thermoregulation and
- water-use strategies between three co-occurring Atlantic forest tree species. *Plant, Cell &*
- *Environment* **41**: 1618–1631.
- **Fauset S, Oliveira L, Buckeridge MS,** *et al.* **2019**. Contrasting responses of stomatal
- conductance and photosynthetic capacity to warming and elevated CO2 in the tropical tree
- species Alchornea glandulosa under heatwave conditions. *Environmental and Experimental Botany* **158**: 28–39.
- **Fick SE, Hijmans RJ**. **2017**. WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. *International Journal of Climatology* **37**: 4302–4312.
- **Fisher RA, Koven CD, Anderegg WRL,** *et al.* **2018**. Vegetation demographics in Earth System
- Models: A review of progress and priorities. *Global Change Biology* **24**: 35–54.
- **Friedlingstein P, Cox P, Betts R,** *et al.* **2006**. Climate-carbon cycle feedback analysis: Results
- from the C4MIP model intercomparison. *Journal of Climate* **19**: 3337–3353.
- **Galbraith D, Levy PE, Sitch S,** *et al.* **2010**. Multiple mechanisms of Amazonian forest biomass
- losses in three dynamic global vegetation models under climate change. *New Phytologist* **187**: 647–665.
- **Goulden ML, Miller SD, Da Rocha HR,** *et al.* **2004**. Diel and seasonal patterns of tropical
- forest CO2 exchange. *Ecological Applications* **14**: 42–54.
- **Green JK, Berry J, Ciais P, Zhang Y, Gentine P**. **2020**. Amazon rainforest photosynthesis
- increases in response to atmospheric dryness. *Science Advances* **6**: 1–10.
- **Greve P, Seneviratne SI**. **2015**. Assessment of future changes in water availability and aridity.
- *Geophysical Research Letters* **42**: 5493–5499.
- **Guan K, Pan M, Li H,** *et al.* **2015**. Photosynthetic seasonality of global tropical forests
- constrained by hydroclimate. *Nature Geoscience* **8**: 284–289.
- **Gurevitch J, Morrow LL, Wallace A, Walsh JS**. **1992**. A Meta-Analysis of Competition in
- Field Experiments Author ( s ): Jessica Gurevitch , Laura L . Morrow , Alison Wallace , Joseph S
- . Walsh Published by : The University of Chicago Press for The American Society of Naturalists
- Stable URL : http://www.jstor. *The American Naturalist* **140**: 539–572.
- **Havaux M**. **1996**. Short-tern response of Photosystem I to heat stress. *Photosynthesis Research*
- **47**: 85–97.
- **Hedges L V., Olkin I**. **1985**. *Statistical methods for meta-analysis*. New York, Academic.
- **Hernández GG, Winter K, Slot M**. **2020**. Similar temperature dependence of photosynthetic
- parameters in sun and shade leaves of three tropical tree species. *Tree physiology* **40**: 637–651.
- **Hikosaka K, Ishikawa K, Borjigidai A, Muller O, Onoda Y**. **2006**. Temperature acclimation
- of photosynthesis: Mechanisms involved in the changes in temperature dependence of
- photosynthetic rate. *Journal of Experimental Botany* **57**: 291–302.
- **Holbo HR, Luvall JC**. **1989**. Modeling surface temperature distributions in forest landscapes.
- *Remote Sensing of Environment* **27**: 11–24.
- **Janzen DH**. **1967**. Why mountain passes are higher in the tropics. *The American Naturalist* **101**: 233–249.
- **Jaramillo C, Ochoa D, Contreras L,** *et al.* **2010**. Effects of rapids global warming at the
- Paleocene-Eocene boundary on neotropical vegetation. *Science* **330**: 957–961.
- **June T, Evans JR, Farquhar GD**. **2004**. A simple new equation for the reversible temperature
- dependence of photosynthetic electron transport: a study on soybean leaf. *Functional Plant Biology* **31**: 275–283.
- **Jurik TW, Weber JA, Gates DM**. **1988**. Effects of Temperature and Light on Photosynthesis of
- Dominant Species of a Northern Hardwood Forest. *Botanical Review* **149**: 203–208.
- **Kattge J, Knorr W**. **2007**. Temperature acclimation in a biochemical model of photosynthesis:
- A reanalysis of data from 36 species. *Plant, Cell and Environment* **30**: 1176–1190.
- **Kattge J, Knorr W, Raddatz T, Wirth C**. **2009**. Quantifying photosynthetic capacity and its
- relationship to leaf nitrogen content for global-scale terrestrial biosphere models. *Global Change Biology* **15**: 976–991.
- **Kitajima K, Mulkey SS, Wright SJ**. **1997**. Seasonal leaf phenotypes in the canopy of a tropical
- dry forest: Photosynthetic characteristics and associated traits. *Oecologia* **109**: 490–498.
- **Kositsup B, Montpied P, Kasemsap P, Thaler P, Améglio T, Dreyer E**. **2009**. Photosynthetic
- capacity and temperature responses of photosynthesis of rubber trees (Hevea brasiliensis Müll.
- Arg.) acclimate to changes in ambient temperatures. *Trees - Structure and Function* **23**: 357–
- 365.
- **Kullberg AT, Slot M, Feeley KJ**. **2023**. Thermal optimum of photosynthesis is controlled by
- stomatal conductance and does not acclimate across an urban thermal gradient in six subtropical tree species. *Plant Cell and Environment*: 1–19.
- **Kumarathunge DP, Drake JE, Tjoelker MG,** *et al.* **2020**. The temperature optima for tree
- seedling photosynthesis and growth depend on water inputs. *Global Change Biology* **26**: 2544– 2560.
- **Kumarathunge DP, Medlyn BE, Drake JE,** *et al.* **2019**. Acclimation and adaptation
- components of the temperature dependence of plant photosynthesis at the global scale. *New*
- *Phytologist* **222**: 768–784.
- **Lai J, Zou Y, Zhang J, Peres-Neto PR**. **2022**. Generalizing hierarchical and variation
- partitioning in multiple regression and canonical analyses using the rdacca.hp R package.
- *Methods in Ecology and Evolution* **13**: 782–788.
- **Lin YS, Medlyn BE, Ellsworth DS**. **2012**. Temperature responses of leaf net photosynthesis:
- The role of component processes. *Tree Physiology* **32**: 219–231.
- **Lloyd J, Farquhar GD**. **2008**. Effects of rising temperatures and [CO2] on the physiology of
- tropical forest trees. *Philosophical transactions of the Royal Society of London. Series B,*
- *Biological sciences* **363**: 1811–7.
- **Lombardozzi DL, Bonan GB, Smith NG, Dukes JS, Fisher RA**. **2015**. Temperature
- acclimation of photosynthesis and respiration: A key uncertainty in the carbon cycle-climate
- feedback. *Geophysical Research Letters* **42**: 8624–8631.
- **Long SP**. **1991**. Modification of the response of photosynthetic productivity to rising
- temperature by atmospheric CO2 concentrations: Has its importance been underestimated?
- *Plant, Cell & Environment* **14**: 729–739.
- **Ma S, Osuna JL, Verfaillie J, Baldocchi DD**. **2017**. Photosynthetic responses to temperature
- across leaf–canopy–ecosystem scales: a 15-year study in a Californian oak-grass savanna. *Photosynthesis Research* **132**: 277–291.
- **Malhi Y, Roberts JT, Betts R a, Killeen TJ, Li W, Nobre C a**. **2008**. Climate Change,
- Deforestation, and the Fate of the Amazon. *Science* **319**: 169–172.
- **Marshall B, Biscoe P V. 1980**. A model for C3 leaves describing the dependence of net
- photosynthesis on irradiance II. *Journal of Experimental Botany* **31**: 41–48.
- **Matthews HD, Eby M, Ewen T, Friedlingstein P, Hawkins BJ**. **2007**. What determines the
- magnitude of carbon cycle-climate feedbacks? *Global Biogeochemical Cycles* **21**: 1–12.
- **Mau A, Reed S, Wood T, Cavaleri M**. **2018**. Temperate and tropical forest canopies are already
- functioning beyond their thermal thresholds for photosynthesis. *Forests* **9**: 47.
- **Medlyn B. E., Dreyer E, Ellsworth D,** *et al.* **2002**. Temperature response of parameters of a
- biochemically based model of photosynthesis. II. A review of experimental data. *Plant, Cell and*
- *Environment* **25**: 1167–1179.
- **Medlyn B E, Dreyer E, Ellsworth D**. **2002**. Temperature response of parameters of a
- biochemically-based model of photosynthesis. II. A review of experimental data. *Plant Cell Environ* **25**: 1167–1179.
- **Mercado LM, Medlyn BE, Huntingford C,** *et al.* **2018**. Large sensitivity in land carbon storage
- due to geographical and temporal variation in the thermal response of photosynthetic capacity.
- *New Phytologist* **218**: 1462–1477.
- **Michaletz ST, Weiser MD, McDowell NG,** *et al.* **2016**. The energetic and carbon economic origins of leaf thermoregulation. *Nature Plants* **2**: 1–8.
- 
- **Miller BD, Carter KR, Reed SC, Wood TE, Cavaleri MA**. **2021**. Only sun-lit leaves of the uppermost canopy exceed both air temperature and photosynthetic optima in a wet tropical
- forest. *Agricultural and Forest Meteorology* **301**–**302**: 108347.
- **Mora C, Frazier AG, Longman RJ,** *et al.* **2013**. The projected timing of climate departure from recent variability. *Nature* **502**: 183–7.
- **Mujawamariya M, Wittemann M, Dusenge ME,** *et al.* **2023**. Contrasting warming responses
- of photosynthesis in early- and late-successional tropical trees. *Tree Physiology* **43**: 1104–1117.
- **Niinemets Ü**. **2007**. Photosynthesis and resource distribution through plant canopies. *Plant, Cell*
- *and Environment* **30**: 1052–1071.
- **Niinemets Ü, Oja V, Kull O. 1999.** Shape of leaf photosynthetic electron transport versus
- temperature response curve is not constant along canopy light gradients in temperate deciduous
- trees. *Plant, Cell & Environment* **22**: 1497–1513.
- **Oliver RJ, Mercado LM, Clark DB,** *et al.* **2022**. Improved representation of plant physiology
- in the JULES-vn5.6 land surface model: photosynthesis, stomatal conductance and thermal
- acclimation. *Geoscientific Model Development* **15**: 5567–5592.
- **Pan Y, Birdsey RA, Phillips OL, Jackson RB**. **2013**. The structure, distribution, and biomass
- of the world's forests. *Annual Review of Ecology, Evolution, and Systematics* **44**: 593–622.
- **Pearcy R**. **1987**. Photosynthetic gas exchange responses of Australian tropical forest trees in
- canopy, gap and understorey micro-environments. *Functional Ecology* **1**: 169–178.
- **Poulter B, MacBean N, Hartley A,** *et al.* **2015**. Plant functional type classification for earth
- system models: Results from the European Space Agency's Land Cover Climate Change
- Initiative. *Geoscientific Model Development* **8**: 2315–2328.
- **De Pury DGG, Farquhar GD**. **1997**. Simple scaling of photosynthesis from leaves to canopies
- without the errors of big-leaf models. *Plant, Cell and Environment* **20**: 537–557.
- **R Core Team**. **2020**. *R: A language and environment for statistical computing. R Foundation*
- *for Statical Computing*.
- **Read J**. **1990**. Some Effects of Acclimation Temperature on Net Photosynthesis in Some
- Tropical and Extra- Tropical Australasian Nothofagus Species. *Journal of Ecology* **78**: 100–112.
- **Reef R, Slot M, Motro U,** *et al.* **2016**. The effects of CO2 and nutrient fertilisation on the
- growth and temperature response of the mangrove Avicennia germinans. *Photosynthesis*
- *Research* **129**: 159–170.
- **Reich PB, Bermudez R, Montgomery RA,** *et al.* **2022**. Even modest climate change may lead
- to major transitions in boreal forests. *Nature* **608**: 540–545.
- **Rey-Sánchez A, Slot M, Posada J, Kitajima K**. **2016**. Spatial and seasonal variation in leaf
- temperature within the canopy of a tropical forest. *Climate Research* **71**: 75–89.
- **Rogers A, Medlyn BE, Dukes JS,** *et al.* **2017**. A roadmap for improving the representation of photosynthesis in Earth system models. *New Phytologist* **213**: 22–42.
- **Ryu Y, Baldocchi DD, Kobayashi H,** *et al.* **2011**. Integration of MODIS land and atmosphere
- products with a coupled-process model to estimate gross primary productivity and
- evapotranspiration from 1 km to global scales. *Global Biogeochemical Cycles* **25**: 1–24.
- **Santos A, Carvalho W, Morel J,** *et al.* **2018**. Variations in precipitation and the equilibrium
- dynamics of a tropical forest tree community in south-eastern Brazil. *Journal of Tropical Forest Science* **30**: 597–605.
- **Scafaro AP, Posch BC, Evans JR, Farquhar GD, Atkin OK**. **2023**. Rubisco deactivation and
- chloroplast electron transport rates co limit photosynthesis above optimal leaf temperature in
- terrestrial plants. *Nature Communications* **14**: 1–9.
- **Scafaro AP, Xiang S, Long BM,** *et al.* **2017**. Strong thermal acclimation of photosynthesis in
- tropical and temperate wet-forest tree species: The importance of altered Rubisco content. *Global Change Biology* **23**: 2783–2800.
- **Van Schaik E, Killaars L, Smith NE,** *et al.* **2018**. Changes in surface hydrology, soil moisture
- and gross primary production in the Amazon during the 2015/2016 El Niño. *Philosophical*
- *Transactions of the Royal Society B: Biological Sciences* **373**.
- **Sheu BH, Lin CK**. **1999**. Photosynthetic response of seedlings of the sub-tropical tree Schima
- superba with exposure to elevated carbon dioxide and temperature. *Environmental and*
- *Experimental Botany* **41**: 57–65.
- **Šigut L, Holišová P, Klem K,** *et al.* **2015**. Does long-term cultivation of saplings under elevated
- CO2 concentration influence their photosynthetic response to temperature? *Annals of Botany* **116**: 929–939.
- **Sinclair TR, Murphy CE, Knoerr KR**. **1976**. Development and evaluation of simplified
- models for simulating canopy photosynthesis and transpiration. *Journal of Applied Ecology* **13**: 813–829.
- **Slot M, Garcia MA, Winter K**. **2016**. Temperature response of CO 2 exchange in three tropical
- tree species. *Functional Plant Biology* **43**: 468–478.
- **Slot M, Krause GH, Krause B, Hernández GG, Winter K**. **2019**. Photosynthetic heat
- tolerance of shade and sun leaves of three tropical tree species. *Photosynthesis Research* **141**: 119–130.
- **Slot M, Rey-Sánchez C, Gerber S, Lichstein JW, Winter K, Kitajima K**. **2014**. Thermal
- acclimation of leaf respiration of tropical trees and lianas: Response to experimental canopy
- warming, and consequences for tropical forest carbon balance. *Global Change Biology* **20**:
- 2915–2926.
- **Slot M, Rifai SW, Eze CE, Winter K**. **2024**. The stomatal response to vapor pressure deficit
- drives the apparent temperature response of photosynthesis in tropical forests. *New Phytologist*.
- **Slot M, Winter K**. **2017a**. In situ temperature response of photosynthesis of 42 tree and liana
- species in the canopy of two Panamanian lowland tropical forests with contrasting rainfall regimes. *New Phytologist*.
- **Slot M, Winter K**. **2017b**. Photosynthetic acclimation to warming in tropical forest tree seedlings. *Journal of Experimental Botany* **68**: 2275–2284.
- **Slot M, Winter K**. **2018**. High tolerance of tropical sapling growth and gas exchange to
- moderate warming. *Functional Ecology* **32**: 599–611.
- **Smith NG, Dukes JS**. **2013**. Plant respiration and photosynthesis in global-scale models:
- Incorporating acclimation to temperature and CO2. *Global Change Biology* **19**: 45–63.
- **Smith NG, Keenan TF**. **2020**. Mechanisms underlying leaf photosynthetic acclimation to
- warming and elevated CO2 as inferred from least-cost optimality theory. *Global Change Biology* **26**: 5202–5216.
- **Smith NG, Malyshev SL, Shevliakova E, Kattge J, Dukes JS**. **2016**. Foliar temperature
- acclimation reduces simulated carbon sensitivity to climate. *Nature Climate Change* **6**: 407–411.
- **Smith MN, Taylor TC, van Haren J,** *et al.* **2020**. Empirical evidence for resilience of tropical
- forest photosynthesis in a warmer world. *Nature Plants* **6**: 1225–1230.
- **Tagesson T, Schurgers G, Horion S,** *et al.* **2020**. Recent divergence in the contributions of
- tropical and boreal forests to the terrestrial carbon sink. *Nature Ecology and Evolution* **4**: 202– 993 209.<br>994 **Tan**
- **Tan ZH, Zeng J, Zhang YJ,** *et al.* **2017**. Optimum air temperature for tropical forest
- photosynthesis: Mechanisms involved and implications for climate warming. *Environmental Research Letters* **12**.
- **Vargas G G, Cordero S RA**. **2013**. Photosynthetic responses to temperature of two tropical
- rainforest tree species from Costa Rica. *Trees* **27**: 1261–1270.
- **Varhammar A, Mclean CM, Dusenge ME,** *et al.* **2015**. Photosynthetic temperature responses
- of tree species in Rwanda : evidence of pronounced negative effects of high temperature in montane rainforest climax species. *New Phytologist*.
- **Vårhammar A, Wallin G, Mclean CM,** *et al.* **2015**. Photosynthetic temperature responses of
- tree species in Rwanda: Evidence of pronounced negative effects of high temperature in montane rainforest climax species. *New Phytologist* **206**: 1000–1012.
- **Wang H, Atkin OK, Keenan TF,** *et al.* **2020**. Acclimation of leaf respiration consistent with optimal photosynthetic capacity. *Global Change Biology* **26**: 2573–2583.
- **Wang YP, Leuning R**. **1998**. A two-leaf model for canopy conductance, photosynthesis and
- partitioning of available energy I: Model description and comparison with a multi-layered model. *Agricultural and Forest Meteorology* **91**: 89–111.
- **Wang H, Prentice IC, Davis TW, Keenan TF, Wright IJ, Peng C**. **2017**. Photosynthetic
- responses to altitude: an explanation based on optimality principles. *New Phytologist* **213**: 976–
- 982.
- **Way DA, Oren R**. **2010**. Differential responses to changes in growth temperature between trees
- from different functional groups and biomes: a review and synthesis of data. *Tree Physiology* **30**: 669–688.
- **Way DA, Yamori W**. **2014**. Thermal acclimation of photosynthesis: On the importance of
- adjusting our definitions and accounting for thermal acclimation of respiration. *Photosynthesis*
- *Research* **119**: 89–100.
- **Williams JW, Jackson ST, Kutzbach JE**. **2007**. Projected distributions of novel and
- disappearing climates by 2100 AD. *Proceedings of the National Academy of Sciences* **104**: 5738–5742.
- **Wise RR, Olson AJ, Schrader SM, Sharkey TD**. **2004**. Electron transport is the functional
- limitation of photosynthesis in field-grown Pima cotton plants at high temperature. *Plant, Cell and Environment* **27**: 717–724.
- **Wittemann M, Andersson MX, Ntirugulirwa B, Tarvainen L, Wallin G, Uddling J**. **2022**.
- Temperature acclimation of net photosynthesis and its underlying component processes in four tropical tree species. *Tree Physiology* **42**: 1188–1202.
- **Wright IJ, Westoby M, Reich PB,** *et al.* **2004**. The worldwide leaf economics spectrum. *Nature* **428**: 821–827.
- **Wu J, Albert LP, Lopes AP,** *et al.* **2016**. Leaf development and demography explain
- photosynthetic seasonality in Amazon evergreen forests. *Science* **351**: 972–976.
- **Wu J, Guan K, Hayek M,** *et al.* **2017**. Partitioning controls on Amazon forest photosynthesis
- between environmental and biotic factors at hourly to interannual timescales. *Global Change*
- *Biology* **23**: 1240–1257.
- **Yamaguchi DP, Nakaji T, Hiura T, Hikosaka K**. **2016**. Effects of seasonal change and
- experimental warming on the temperature dependence of photosynthesis in the canopy leaves of Quercus serrata. *Tree Physiology* **36**: 1283–1295.
- **Yamori W, Hikosaka K, Way DA**. **2014**. Temperature response of photosynthesis in C3, C4,
- and CAM plants: Temperature acclimation and temperature adaptation. *Photosynthesis Research* : 101–117.
- **Yamori W, Noguchi K, Terashima I**. **2005**. Temperature acclimation of photosynthesis in
- spinach leaves: Analyses of photosynthetic components and temperature dependencies of photosynthetic partial reactions. *Plant, Cell and Environment* **28**: 536–547.
- **Yan J, Zhang Y, Yu G,** *et al.* **2013**. Seasonal and inter-annual variations in net ecosystem
- exchange of two old-growth forests in southern China. *Agricultural and Forest Meteorology*
- **182**–**183**: 257–265.
- **Zarakas CM, Swann ALS, Koven C, Marielle N, Taylor TC**. **2024**. Different model
- assumptions about plant hydraulics and photosynthetic temperature acclimation yield diverging
- implications for tropical forest resilience. *Global Change Biology* **30**: 1–16.
- **Ziegler C, Dusenge ME, Nyirambangutse B, Zibera E, Wallin G, Uddling J**. **2020**.
- Contrasting Dependencies of Photosynthetic Capacity on Leaf Nitrogen in Early- and Late-
- Successional Tropical Montane Tree Species. *Frontiers in Plant Science* **11**: 1–12.
- 
- 

 **Figure 1** The optimum temperature of net photosynthesis and biochemical responses to mean annual growth temperature and aridity index. *ToptA* response to A) MAT and B) aridity index. *ToptV* response to C) MAT and D) aridity index. *ToptJ* response to E) MAT and F) aridity index. Regression equations are weighted by number of observations that are used to calculate each temperature response mean. Size of data point depicts weight of each mean where larger data points carry a greater weight. Line represents linear regression fits (Table 2). Shaded area around line represents confidence intervals. Color represents altitude groupings of < 500m (blue-green), 500-999m (turquoise), 1000-2000m (beige), NA (gray).

**Figure 2** The rate of net and the biochemical components of photosynthesis at 25 °C responses to three primary climate variables. *A<sup>25</sup>* response to **A)** MAT, **B)** aridity index where higher aridity index indicates wetter conditions. *V<sup>25</sup>* response to **D)** MAT, **E)** aridity index. *J<sup>25</sup>* response to **G)** MAT, **H)** aridity index. Regression equations are weighted by number of observations used to calculate each temperature response mean. Size of data point depicts weight of each mean where larger data points carry a greater weight. Solid line represents significant linear regression fits (Table 2). Shaded area around line represents confidence intervals. Color represents altitude groupings of < 500m (blue-green), 500-999m (turquoise), 1000-2000m (beige), >2000m (black), NA (gray).

 **Figure 3** The ratio between rate of *Jmax* and *Vcmax* responses to three primary climate variables. 1076 The ratio between the rate of  $J_{max}$  at 25 °C and  $V_{cmax}$  at 25 °C (*JV*) responses to **A**) mean annual temperature (MAT), **B)** aridity index, and **C)** mean annual temperature range (*Trange*). Regression equations are weighted by number of observations that are used to calculate each temperature

 response mean. Size of data point depicts the sample size used to weight each mean where larger data points carry a greater weight. Shaded area around line represents confident intervals. Color represents altitude groupings of < 500m (blue green), 500-999m (turquoise), 1000-2000m (beige), >2000m (black), NA (gray).

 **Figure 4** Boxplots displaying the net photosynthetic and biochemistry at 25 °C parameter differences with leaf light environment. The distribution of shade and sun growth leaves for **A)**  $T_{optA}$ , **B**)  $A_{25}$ , **C**)  $\Omega$ , **D**)  $V_{25}$ , **E**)  $J_{25}$ , and F) the ratio of  $J_{max}$  to  $V_{cmax}$ .  $\Omega$  indicates the difference in  $T_{opt}$  and the temperature where the rate of photosynthesis is 37% of  $T_{opt}$ . The boxes display median and interquartile range. The whiskers represent 1.5 times the interquartile range. Data beyond the whiskers are outside of 1.5 times the interquartile range. Asterisks denotes significant 1090 differences between treatments based on a Satterthwaite test,  $* p < 0.05$ ,  $** p < 0.01$ ,  $*** p <$ 1091 0.001.  $A_{max}$ : sun n = 89 shade n = 6;  $k_{25}$ : sun n = 248, shade n = 23.

 **Figure 5** Boxplots displaying the net photosynthetic parameter differences between species of different leaf habit. The distribution of evergreen and semi-deciduous species for **A)** *ToptA*, **B)** *A*<sub>25</sub>, and **C**) Ω. The boxes display median and interquartile range. The whiskers represent 1.5 times the interquartile range. Data beyond the whiskers are outside of 1.5 times the interquartile range. Asterisks denotes significant differences between treatments based on a Satterthwaite test,  $* p < 0.01$ . evergreen n = 45, semideciduous n = 23.

 **Figure 6** Boxplots displaying the net photosynthetic parameter differences between successional 1101 stratus. The distribution of early and late successional species for **A**)  $T_{optA}$ , **B**)  $A_{25}$ , and **C**)  $\Omega$ , **D**)

- 1102  $T_{optV}$ , **E**)  $V_{25}$ , **F**) *JV*, **G**)  $T_{optJ}$ , **H**)  $J_{25}$ . The boxes display median and interquartile range. The
- 1103 whiskers represent 1.5 times the interquartile range. Data beyond the whiskers are outside of 1.5
- 1104 times the interquartile range. Asterisks denotes significant differences between treatments based
- 1105 on a Satterthwaite test, \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ .  $A_{max}$ : early  $n = 20$ , mid/late  $n =$
- 1106 22;  $k_{25}$ : early n = 14, shade n = 17;  $A C_i$ : early n = 8, shade n = 7.
- 1107
- 1108 **Figure 7** Hierarchical partitioning results for relative individual importance of individual climate
- 1109 variables on T<sub>optA</sub> (A), A<sub>25</sub> (B),  $\Omega$  (C), T<sub>optV</sub> (D), V<sub>25</sub> (E), H<sub>aV</sub> (F), T<sub>optJ</sub> (G), J<sub>25</sub> (H), H<sub>aJ</sub> (I), JV
- 1110 (J). Individual effects sums to the calculated total explained variation (adj  $R^2$ ).

## 1111 Tables

1112 **Table 1** Abbreviations and descriptions





1115 **Table 2** Regression equations for each photosynthetic parameter response to individual climate variables.



1119 respectively; °C), the rate of  $V_{cmax}$  ( $V_{25}$ ; µmol m<sup>-2</sup> s<sup>-1</sup>) and  $J_{max}$  ( $J_{25}$ ; µmol m<sup>-2</sup> s<sup>-1</sup>) at 25 °C, and the activation energy term for  $V_{cmax}$ 

1120  $(E_{aV}$ ; kJ mol<sup>-1</sup>) and  $J_{max}$  ( $E_{aJ}$ ; kJ mol<sup>-1</sup>). Climate variables are mean annual temperature (MAT; °C), aridity index, and the mean annual

1121 temperature range from the maximum temperature of the warmest month and the minimum temperature of the coldest month.

Intercepts and slopes are given as means  $\pm$  standard error. Bolded values indicate regression results with p-values < 0.05. Marginal  $r^2$ 1122

1123 provides the model variance of only the model fixed effect, whereas, conditional  $r^2$  provides variance of the model with both the fixed

1124 and random effects.















