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## ALTERATION OF CEREBRAL NEUROTRANSMITTER RECEPTOR FUNCTION BY EXPOSURE OF RATS TO MANGANESE

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### SUMMARY

Adult male rats were treated on 15 successive days with i.p. injection of manganese chloride at 10 or 15 mg/kg body weight. 24 h after the last dose, the binding of the dopaminergic antagonist [<sup>3</sup>H]spiroperidol to striatal membranes was elevated significantly. At the lower dose, no other high-affinity binding site measured was altered. However, at the 15 mg/kg body weight dose, cerebellar GABA, frontal cortical serotonin, and striatal muscarinic binding were all depressed. Neither dose level altered striatal levels of enkephalin, substance P, dopamine, serotonin, or dihydroxyphenylacetic acid (DOPAC). Receptor binding measurements may be a sensitive means of detecting disturbances of specific neural circuits.

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### INTRODUCTION

Excessive exposure to manganese (Mn) during mining and processing operations [10, 21] and production of ferromanganese alloy [31, 32] may lead to chronic disabling disease. The clinical symptoms resemble Parkinsonism [11, 12, 21] although other behavioral disturbances also occur [24]. Decrease of dopamine levels in the brain of experimental animals [11, 22, 23] and improvement of some neurological symptoms in Mn poisoning cases following high doses of L-DOPA, the natural precursor of dopamine, suggests an effect of the metal on synaptic function of dopamine [8, 25]. Decreases in brain norepinephrine [22] and serotonin [19] indicates the involvement of other monoamines as well.

In the present study an attempt has been made to see whether Mn exerts its neurotoxic effects mainly by affecting the dopamine system. A survey of high-

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Abbreviations: DOPAC, 3,4-dihydroxyphenylacetic acid; 5-HIAA, 5-hydroxyindoleacetic acid; HVA, 3-methoxy-4-hydroxyphenylacetic acid; HPLC, high-performance liquid chromatography; 5-HT, 5-hydroxytryptamine; QNB, DL-[benzilic-4,4'-<sup>3</sup>H]quinuclidinyl benzilate.

affinity binding sites of various neurotransmitters was used as an objective means of detecting damage to a specific neuronal circuit [4]. Since the corpus striatum appears to be the chief site of injury, we have estimated the levels of dopamine and serotonin and their metabolites in this brain region. Levels of [met<sup>5</sup>]enkephalin and substance P, which may interact with the nigro-striatal dopamine system [15, 16] were also measured to see if Mn has any effect on neuroactive peptides.

## METHODS

### *(a) Manganese administration*

Male Fischer rats 6–7 weeks old were injected i.p. daily for 15 days with distilled water or manganese chloride (MnCl<sub>2</sub>) solution at 2 levels (10 and 15 mg/kg body weight). This dose range has previously been found to cause enzymic and histological changes in rodent brain [27, 30]. 24 h after the last injection rats were decapitated and brain regions dissected out [17] and frozen at -20°C.

### *(b) High-affinity binding assay*

A crude membrane fraction was prepared from brain regions by homogenization of tissue in 19 vols. of 0.32 M sucrose followed by centrifugation (50 000 × g, 10 min). The precipitate was homogenized in distilled water pH 7.4 and recentrifuged. The final pellet was suspended in 40 mM Tris-HCl pH 7.4 buffer at a concentration corresponding to 50 mg original tissue/ml.

Binding incubations were carried out in triplicate in 1 ml of medium containing 40 mM Tris-HCl pH 7.4 and appropriate labeled and unlabeled pharmacological agents. The incubation mixture used in the assay of serotonin included 10<sup>-5</sup>M pargyline, 4 · 10<sup>-3</sup>M CaCl<sub>2</sub> and 5.7 · 10<sup>-3</sup>M ascorbic acid. The amount of tissue used per tube corresponded to 5–10 mg original wet weight and contained 300–400 µg protein as determined by the method of Lowry et al. [20]. At the end of a 15 min incubation at 37°C samples were filtered on glass fiber discs (25 mm diameter, 0.3 µ pore size, Gelman Inc., Ann Arbor, MI) and washed twice rapidly with 5 ml Tris buffer. Filter discs were then dried and counted in 5 ml of a scintillation mixture using a scintillation counter at an efficiency of 38–43%. Control incubations were carried out simultaneously containing no unlabeled competing ligand in order to determine the extent of non-specific binding. The final concentration of unlabeled competing compounds in control incubations was 10<sup>-6</sup>M. The assay of the dopamine receptor was performed using 10<sup>-9</sup>M [1-phenyl-4 <sup>3</sup>H]spiroperidol (23 Ci/mmol) as the binding ligand and haloperidol as the competing compound in control tubes. In a parallel manner 10<sup>-9</sup>M QNB (29 Ci/mmol) was used to measure muscarinic sites with atropine as a competitor. Benzodiazepine sites were estimated with 0.7 · 10<sup>-9</sup>M [methyl-<sup>3</sup>H]diazepam (73 Ci/mmol) and the unlabeled compound

as a competitor.  $8.0 \cdot 10^{-9}$ M [methylene- $^3\text{H}(\text{N})$ ]muscimol (7.3 Ci/mmol) and unlabeled GABA were used in GABA-binding site assays.  $3.1 \cdot 10^{-9}$ M [1,2- $^3\text{H}(\text{N})$ ]serotonin (30 Ci/mmol) was used together with unlabeled serotonin in the assay of this receptor.  $6 \cdot 10^{-9}$ M [G- $^3\text{H}$ ]strychnine sulphate (13 Ci/mmol) and  $10^{-5}$ M unlabeled strychnine were used to assay the glycine receptor. The method used was essentially similar to other filtration binding methods. However, we felt it necessary to establish basic binding characteristics prior to studies on animals treated with manganese. These included delineation of saturability, specificity, reversibility, and regional distribution [4].

#### *(c) Assay of monoamine content*

The tissue concentrations of dopamine, DOPAC, HVA, 5-HT, and 5-HIAA were determined by HPLC with electrochemical detection [18]. Tissues were homogenized in 250  $\mu\text{l}$  of the mobile phase containing 20 ng of the internal standard (*N*-methyl-5-HT) and following centrifugation an aliquot of the resulting supernatant was injected into the LC. The striatal concentrations of the compounds were calculated from the ratio of their peak height relative to the internal standard using the slope and intercept of a standard curve.

#### *(d) Radioimmunoassay of neuropeptides*

Tissue was homogenized in 2N acetic acid, boiled for 5 min and centrifuged at  $25\,000 \times g$  for 20 min. The supernatant was lyophilized and the residue was reconstituted with  $\text{H}_2\text{O}$  and radioimmunoassayed for met-enkephalin and substance P using [tyrosyl-3,5- $^3\text{H}$ ]met-enkephalin (36 Ci/mmol) and  $^{125}\text{I}$ -substance P (original specific activity 65  $\mu\text{Ci}/\mu\text{g}$ ). Antisera were raised in rabbits using polylysine conjugates. Six injections were given at 2-week intervals before the animals were bled. The assay was carried out in polypropylene tubes. Nonlabeled peptide or brain extract was incubated with antiserum and [ $^3\text{H}$ ]met-enkephalin or  $^{125}\text{I}$ -substance P in 0.5 ml of 0.2 M Tris buffer, pH 7.4, containing 0.1% albumin and 0.06% dextran. The incubation was carried out at  $4^\circ\text{C}$  for 15–24 h. The [ $^3\text{H}$ ]met-enkephalin (or  $^{125}\text{I}$ -substance P) bound to antibody was separated from the free [ $^3\text{H}$ ]met-enkephalin (or  $^{125}\text{I}$ -substance P) by adding 0.2 ml of 1.5% charcoal slurry containing 0.15% dextran (suspended in 0.2 M Tris buffer, pH 7.4) and aliquot of supernatant fluid was counted in a liquid scintillation spectrometer. The validation and specificity of this method have been described in detail [14, 16].

#### *(e) Statistics*

Differences between groups were assessed using Fisher's Least Significant Difference Test after a one-way analysis of variance. The accepted level of

significance in all cases was  $P < 0.05$  using a two-tailed test. Each point represents values derived from 7–8 individual animals.

## RESULTS AND DISCUSSION

At the lower dose of Mn (10 mg/kg body weight, daily for 15 days), only one of the six labeled ligand-binding reactions was significantly different from that of the corresponding control value. Spiroperidol binding to striatal membranes was 61% higher in treated animals (Table I). At the higher dose of Mn (15 mg/kg body weight), striatal dopaminergic binding activity was still the only elevated parameter detected. However, striatal muscarinic binding was depressed below control values as was cerebellar GABA and cortical serotonin binding. The cerebral protein content of rats chronically exposed to manganese has been shown to remain unaffected [29].

Striatal levels of [met<sup>5</sup>]enkephalin and substance P were not significantly affected by the Mn treatment and were within 10% and 3% of control values respectively (Table II). The striatal content of dopamine, DOPAC, HVA, and serotonin were also unaltered by treatment. However, the level of 5-HIAA was significantly depressed to 75% of the control value in animals treated with 15 successive doses of Mn at 15 mg/kg (Table III).

The binding data suggest that assay of receptor–ligand interactions may be an especially sensitive means of detection of a derangement of a specific neurotransmitter system. A large increase of spiroperidol binding was seen in Mn-treated animals showing unaltered striatal levels of dopamine and its metabolic product, DOPAC and HVA. No changes in striatal neuropeptides thought to be associated with dopaminergic systems [15, 16] were found in rats exposed to Mn.

TABLE I

### EFFECT OF MANGANESE TREATMENT ON HIGH AFFINITY BINDING LEVELS IN SEVERAL BRAIN REGIONS

Data expressed as pmol ligand bound/g protein. Means derived from 7 individual animals per group, together with the standard error, are presented. Daily dosing for 15 days.

Region	Receptor	Manganese dose (daily, mg/kg)		
		0	10	15
Striatum	Dopamine	90 ± 5	145 ± 13 <sup>a</sup>	122 ± 11 <sup>a</sup>
	Muscarinic	1057 ± 65	1162 ± 39	811 ± 56 <sup>a</sup>
Frontal cortex	Benzodiazepine	23 ± 1	23 ± 3	26 ± 3
	Serotonin	73 ± 7	62 ± 4	48 ± 5
	Glycine	40 ± 4	46 ± 3	45 ± 2
Cerebellum	GABA	63 ± 6	52 ± 4	46 ± 2 <sup>a</sup>

<sup>a</sup>Differs from zero-dose ( $P < 0.05$ , Fisher's Least Significant Difference Test).

TABLE II

## STRIATAL LEVELS OF ENKEPHALIN AND SUBSTANCE P AFTER MANGANESE TREATMENT

Means derived from 8 individual animals per group are given together with the standard error. Daily dosing for 15 days.

Manganese dose (mg/kg, daily)	Peptide level (ng/mg protein)	
	[met <sup>5</sup> ]enkephalin	Substance P
0	19.0 ± 0.8	2.22 ± 0.05
10	20.6 ± 0.6	2.27 ± 0.06
15	20.6 ± 1.0	2.23 ± 0.10

TABLE III

## STRIATAL CONCENTRATIONS OF DOPAMINE, SEROTONIN, AND THEIR METABOLITES IN MANGANESE-TREATED RATS

Data are expressed as ng/mg protein; daily dosing for 15 days. Means and standard errors derived from data of 8 individual rats per group.

	Manganese dose (daily, mg/kg)		
	0	10	15
Dopamine	90 ± 2	99 ± 4	92 ± 3
DOPAC	16.8 ± 0.7	18.1 ± 0.9	17.1 ± 0.6
HVA	7.9 ± 0.3	8.6 ± 0.5	7.1 ± 0.3
Serotonin	3.4 ± 0.2	3.4 ± 0.2	3.2 ± 0.2
5-HIAA	5.5 ± 0.2	4.9 ± 0.4	4.1 <sup>a</sup> ± 0.2

<sup>a</sup>Differs from corresponding control ( $P < 0.05$ , Fisher's Least Significant Difference Test).

There is evidence that rats are relatively resistant to Mn, as evaluated by alterations in levels of dopamine, serotonin, and their metabolites [28]. Chronic exposure lasting many months may alter dopamine levels [8].

The depression of serotonin and GABA receptor activity seen at the higher (15 mg/kg) Mn dose may represent a less specific toxic effect of this metal. However, Mn has been shown to increase striatal GABA in rats after chronic administration [7] and cerebral serotonin levels are depressed in rats fed a diet containing 2 mg Mn chow [19]. The depressed level of the serotonin metabolite 5-HIAA in animals given the repeated 15 mg/kg Mn dose, also suggests serotonergic involvement. Since dopaminergic input to the cerebellum is minimal, the decreased GABA receptor binding activity observed in this region in animals treated at the higher Mn level, is unlikely to be a secondary response mediated directly by dopamine. However, in view of the complex interdependence of all neurotransmitter circuits, it may be that alteration in dopaminergic mechanisms underlies many other observed changes. The heightened level of striatal spiroperidol binding suggests a disuse supersensitivity-

like response although we have not been able to detect major changes in dopamine metabolism. Changes may be more readily detected in receptors, largely confined to the synapse, than in the concentrations of neurotransmitters or their metabolites which reflect the state of the entire neuron [4]. Mn treatment has been shown to modify the activity of tyrosine hydroxylase activity in striatum which may be related to the blockade of dopaminergic receptors [6]. The increase in the [<sup>3</sup>H]spiroperidol binding to striatum of Mn-exposed animals is consistent with this observation.

300 mg MnCl<sub>2</sub> given daily to Sprague–Dawley male rats for 30 days has been reported to increase striatal Mn to approx.  $0.5 \cdot 10^{-6}$ M [6]. Our dosing was for a shorter period at 1/10 the dose used in the above work. Since  $10^{-6}$ M MnCl<sub>2</sub> in vitro had no effect upon striatal spiroperidol binding (P.K. Seth, unpublished result), the effect of MnCl<sub>2</sub> dosing upon the striatal dopamine receptor in vivo, reported here is indirect.

No gross behavioral deficits, e.g. tremor, were observed in the treated rats. An earlier study involving  $10 \times$  the dose level of Mn for up to 8 months also failed to detect any sign of neurological disease [6]. The use of an appropriate pharmacological challenge such as altered responses to apomorphine might demonstrate abnormality of the dopamine system in Mn-treated animals before the onset of overt behavioral changes.

The effect of gestational and neonatal exposure of rat pups to Mn on neurotransmitter receptors is important since very early exposure to Mn has been reported to increase striatal catecholamine levels, an opposite effect to that found in the adult [9]. Pre- and postnatal effects on neurotransmitter-related phenomena are frequently opposite to those found in the adult. For example, prenatal exposure of rats to acrylamide depresses striatal dopaminergic receptors [2]; the opposite effect is found after acrylamide treatment of adult rats [1, 3, 5]. A parallel situation is found with chlordecone treatment except that neonatal exposure elevates striatal dopaminergic receptor binding which is depressed following adult exposure [26]. The striatal dopamine receptor may often be more sensitive to neurotoxicant-induced disturbances than are a variety of other transmitter receptor sites. This may account for the relatively common occurrence of tremor and other Parkinsonian symptoms found as a result of exposure to a variety of agents damaging to the CNS.

## REFERENCES

- 1 A.K. Agrawal, P.K. Seth, R.E. Squibb, H.A. Tilson, L.L. Uphouse and S.C. Bondy, Neurotransmitter receptors in brain regions of acrylamide-treated rats, I. Effects of a single exposure to acrylamide, *Pharmacol. Biochem. Behav.*, 14 (1981) 527–531.
- 2 A.K. Agrawal and R.E. Squibb, Effects of acrylamide given during gestation on dopamine receptor binding in pups, *Toxicol. Lett.*, 7 (1980) 233–238.
- 3 A.K. Agrawal, R.E. Squibb and S.C. Bondy, The effects of acrylamide upon the dopamine receptor, *Toxicol. Appl. Pharmacol.* 58 (1981) 89–99.

- 4 S.C. Bondy, Neurotransmitter binding interactions as a screen for neurotoxicity, in A. Vernadikis and K.N. Prasad (Eds.), *Mechanisms of Neurotoxic Substances*, Raven Press, New York, 1980, in press.
- 5 S.C. Bondy, H.A. Tilson and A.K. Agrawal, Neurotransmitter receptors in brain regions of acrylamide-treated rats, II. Effects of extended exposure to acrylamide, *Pharmacol. Biochem. Behav.*, 14 (1981) 533–537.
- 6 E. Bonilla, L-tyrosine hydroxylase activity in the rat brain after chronic oral administration of manganese chloride, *Neurobehav. Toxicol.*, 2 (1980) 37–41.
- 7 E. Bonilla, Increased GABA content in caudate nucleus of rats after chronic manganese administration, *J. Neurochem.*, 31 (1978) 551–552.
- 8 E. Bonilla and M. Diez-Ewald, Effect of L-DOPA on brain concentration of dopamine and homovanillic acid in rats after chronic manganese chloride administration, *J. Neurochem.*, 22 (1974) 297–299.
- 9 S.V. Chandra, G.S. Shukla and D.K. Saxena, Manganese-induced behavioral dysfunction and its neurochemical mechanism in growing mice, *J. Neurochem.*, 33 (1979) 1217–1221.
- 10 D.G. Cook, S. Fahn and K.A. Brait, Chronic manganese intoxication, *Arch. Neurol.*, 30 (1974) 59–64.
- 11 G.C. Cotzias, P.S. Papavasiliou, I. Mena, L.D. Tang and S.T. Miller, Manganese and catecholamines, *Adv. Neurol.*, 5 (1974) 235–243.
- 12 M. Goldman, Levo-dihydroxyphenylalanine, Parkinson's Disease and manganese poisoning, *Indust. Med.*, 41 (1972) 12–17.
- 13 F. Hefti, A simple sensitive method for measuring HVA and DOPAC in rat brain tissue using HPLC with electrochemical detection, *Life Sci.*, 25 (1979) 775–781.
- 14 J.S. Hong, E. Costa and H.Y.T. Yang, Effects of habenular lesions on the substance P content of various brain regions, *Brain Res.*, 118 (1976) 523–525.
- 15 J.S. Hong, H.Y.T. Yang and E. Costa, Substance P content of substantia nigra after chronic treatment with antischizophrenic drugs, *Neuropharmacology*, 17 (1978) 83–85.
- 16 J.S. Hong, H.Y.T. Yang, W. Fratta and E. Costa, Rat striatal methionine-enkephalin content after chronic treatment with cataleptogenic and non-cataleptogenic antischizophrenic drugs, *J. Pharmacol. Exp. Ther.*, 205 (1978) 141–147.
- 17 L.L. Iversen and J. Glowinski, Regional studies of catecholamines in the rat brain, I. The disposition of <sup>3</sup>H-norepinephrine, <sup>3</sup>H-dopamine, and <sup>3</sup>H-dopa in various regions of the brain, *J. Neurochem.*, 13 (1966) 655–669.
- 18 C.D. Kilts, R.B. Mailman and G.R. Breese, Simultaneous determination of dopamine, DOPAC, HVA, 5-HTP, 5-HT, and 5-HIAA by means of reverse phase HPLC with electrochemical detection, *J. Chromatogr.*, 1981 (submitted).
- 19 M. Kimura, N. Yagi and Y. Itokawa, Effect of subacute manganese feeding on serotonin metabolism in the rat, *J. Toxicol. Environ. Hlth.*, 4 (1978) 701–707.
- 20 O.H. Lowry, N.J. Rosebrough, A.L. Farr and R.J. Randall, Protein measurement with the Folin phenol reagent, *J. Biol. Chem.*, 193 (1951) 265–275.
- 21 I. Mena, O. Marin, S. Fuenzalida and G.C. Cotzias, Chronic manganese poisoning, clinical picture and manganese turnover, *Neurology*, 17 (1967) 128–136.
- 22 S.J. Mustafa and S.V. Chandra, Levels of 5-hydroxytryptamine, dopamine, and norepinephrine in whole brains of rabbits in chronic manganese toxicity, *J. Neurochem.*, 18 (1971) 931–933.
- 23 N.H. Neff, R.E. Barrett and E. Costa, Selective depletion of caudate nucleus dopamine and serotonin during chronic manganese dioxide administration to squirrel monkeys, *Experientia*, 25 (1969) 1140–1141.
- 24 B. Roussel and B. Renaud, Effect of chronic manganese intoxication on the sleep-wake cycle in the rat, *Neurosci. Lett.*, 4 (1977) 55–60.
- 25 W. Schunk, Problems associated with chronic occupational manganese poisoning, *Dtsch. Gesundheitsures*, 34 (1979) 1631–1633.



- 26 P.K. Seth, A.K. Agrawal and S.C. Bondy, Biochemical changes in the brain consequent to dietary exposure of developing and mature rats to chlordecone (kepone), *Toxicol. Appl. Pharmacol.* (1981) in press.
- 27 P.K. Seth, R. Husain, M. Mushtaq and S.V. Chandra, Effect of manganese on neonatal rat: manganese concentration and enzymic alterations in brain, *Acta Pharmacol. Toxicol.*, 40 (1977) 553–560.
- 28 G.S. Shukla and S.V. Chandra, Species variation in manganese induced changes in brain biogenic amines, *Toxicol. Lett.*, 3 (1979) 249–253.
- 29 G.S. Shukla, S.V. Chandra and P.K. Seth, Effect of manganese on the levels of DNA, RNA, DNase, and RNase in cerebellum and rest of brain regions of rat, *Acta Pharmacol. Toxicol.*, 39 (1976) 562–569.
- 30 J. Singh, R. Husain, S.K. Tandon, P.K. Seth and S.V. Chandra, Biochemical and histopathological alterations in early manganese toxicity in rats, *Environ. Physiol. Biochem.*, 4 (1974) 16–23.
- 31 L.T. Smyth, R.C. Ruhf, N.E. Whitman and T. Dugan, Clinical manganism and exposure to manganese in the production and processing of ferromanganese alloy, *J. Occup. Med.*, 15 (1973) 101–109.
- 32 S. Tanaka and J. Lieben, Manganese poisoning and exposure in Pennsylvania, *Arch. Environ. Hlth.*, (1969) 674–684.