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Authors

Wu, Linwei

Ning, Daliang

Zhang, Bing

et al.

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1 **Global Diversity and Biogeography of Bacterial Communities in Wastewater Treatment**
2 **Plants**

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4 Linwei Wu^{1,2†}, Daliang Ning^{2,3,1†}, Bing Zhang^{1,2†}, Yong Li⁴, Ping Zhang^{2,5}, Xiaoyu Shan¹,
5 Qiuting Zhang¹, Mathew Brown⁶, Zhenxin Li⁷, Joy D. Van Nostrand², Fangqiong Ling⁸, Naijia
6 Xiao^{2,3}, Ya Zhang², Julia Vierheilig^{9,10}, George F. Wells¹¹, Yunfeng Yang¹, Ye Deng^{12,13}, Qichao
7 Tu¹², Aijie Wang¹³, Global Water Microbiome Consortium[‡], Tong Zhang¹⁴, Zhili He^{15,16}, Jurg
8 Keller¹⁷, Per H. Nielsen¹⁸, Pedro J. J. Alvarez¹⁹, Craig S. Criddle²⁰, Michael Wagner⁹, James M.
9 Tiedje²¹, Qiang He^{22,23*}, Thomas P. Curtis^{6*}, David A. Stahl²⁴, Lisa Alvarez-Cohen^{25,26}, Bruce E.
10 Rittmann²⁷, Xianghua Wen^{1*}, and Jizhong Zhou^{2,1,26*}

11
12
13 ¹State Key Joint Laboratory of Environment Simulation and Pollution Control, School of
14 Environment, Tsinghua University, Beijing, China; ²Institute for Environmental Genomics,
15 Department of Microbiology and Plant Biology, and School of Civil Engineering and
16 Environmental Sciences, University of Oklahoma, Norman, OK, USA; ³Consolidated Core
17 Laboratory, University of Oklahoma, Norman, Oklahoma, USA; ⁴College of Resource &
18 Environment Southwest University, Chongqing, China; ⁵Alkek Center for Metagenomics and
19 Microbiome Research, Department of Molecular Virology and Microbiology, Baylor College of
20 Medicine, Houston, TX, USA; ⁶School of Engineering, Newcastle University, Newcastle upon
21 Tyne, UK; ⁷School of Environment, Northeastern Normal University, Changchun, China;
22 ⁸Department of Energy, Environmental and Chemical Engineering, Washington University in St.
23 Louis, MO, USA; ⁹Department of Microbiology and Ecosystem Science, Division of Microbial
24 Ecology, Research Network 'Chemistry meets Microbiology', University of Vienna, Vienna,
25 Austria; ¹⁰Karl Landsteiner University of Health Sciences, Division of Water Quality and Health,
26 Krems, Austria & Interuniversity Cooperation Centre for Water and Health; ¹¹Department of
27 Civil and Environmental Engineering, Northwestern University, Evanston, IL, USA; ¹²Institute
28 for Marine Science and Technology, Shandong University, Qingdao, China; ¹³Research Center
29 for Eco-Environmental Sciences, Chinese Academy of Sciences, Beijing, China;
30 ¹⁴Environmental Biotechnology Laboratory, The University of Hong Kong, Hong Kong, China;
31 ¹⁵Environmental Microbiomics Research Center, School of Environmental Science and
32 Engineering, Sun Yat-Sen University, Guangzhou, China; ¹⁶Guangdong Provincial Key
33 Laboratory of Environmental Pollution Control and Remediation Technology, Guangzhou,
34 China; ¹⁷Advanced Water Management Centre, The University of Queensland, Brisbane, QLD,
35 Australia; ¹⁸Department of Chemistry and Bioscience, Center for Microbial Communities,
36 Aalborg University, Aalborg, Denmark; ¹⁹Department of Civil and Environmental Engineering,
37 Rice University, Houston, TX, USA; ²⁰Department of Civil and Environmental Engineering,
38 Stanford University, Stanford, CA, USA; ²¹Center for Microbial Ecology, Michigan State
39 University, East Lansing, MI, USA; ²²Department of Civil and Environmental Engineering, The
40 University of Tennessee, Knoxville, TN, USA; ²³Institute for a Secure and Sustainable
41 Environment, The University of Tennessee, Knoxville, TN, USA; ²⁴Department of Civil and
42 Environmental Engineering, University of Washington, Seattle, WA, USA; ²⁵Department of
43 Civil and Environmental Engineering, College of Engineering, University of California,
44 Berkeley, CA, USA; ²⁶Earth and Environmental Sciences, Lawrence Berkeley National
45 Laboratory, Berkeley, CA, USA; ²⁷Biodesign Swette Center for Environmental Biotechnology,
46 Arizona State University, Tempe, AZ, USA.

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† These authors contributed equally to this work.

‡ Other Global Water Microbiome Consortium members are listed at the end of this paper.

* Corresponding authors. To whom correspondence should be addressed regarding analysis, synthesis, and reprints, E-mail: jzhou@ou.edu. To whom correspondence should be addressed regarding experimental design and sampling, E-mails: xhwen@tsinghua.edu.cn, tom.curtis@newcastle.ac.uk, qianghe@utk.edu.

56 **Microorganisms in wastewater treatment plants (WWTPs) are essential for water**
57 **purification to protect public and environmental health. However, their diversity and the**
58 **factors that control it are poorly understood. Using a systematic global-sampling effort, we**
59 **analyzed the 16S rRNA gene sequences from ~1,200 activated sludge samples taken from**
60 **269 WWTPs in 23 countries on 6 continents. Our analyses revealed that the global**
61 **activated sludge bacterial communities contain ~1 billion bacterial phylotypes with a**
62 **Poisson lognormal diversity distribution. Despite this high diversity, activated sludge has a**
63 **small global core bacterial community (n = 28 OTUs) that is strongly linked to activated**
64 **sludge performance. Meta-analyses with global datasets associate the activated sludge**
65 **microbiomes most closely to freshwater populations. In contrast to macroorganism**
66 **diversity, activated sludge bacterial communities show no latitudinal gradient.**
67 **Furthermore, their spatial turnover is scale-dependent and appears to be largely driven by**
68 **stochastic processes (dispersal, drift), although deterministic factors (temperature, organic**
69 **input) also are important. Our findings enhance mechanistic understanding of the global**
70 **diversity and biogeography of activated sludge bacterial communities within a theoretical**
71 **ecology framework and have important implications for microbial ecology and wastewater**
72 **treatment processes.**

73 **Introduction**

74

75 Microorganisms, the most diverse group of life on Earth¹, play crucial roles in the
76 biogeochemical cycling of carbon (C), nitrogen (N), sulfur (S), phosphorus (P), and various
77 metals. Unraveling the mechanisms generating and underlying microbial biodiversity is key to
78 predicting ecosystem responses to environmental changes² and improving bioprocesses, such as
79 wastewater treatment and soil remediation³. With recent advances in metagenomic technologies⁴,
80 microbial biodiversity and distribution are being intensively studied in a wide variety of
81 environments⁵⁻⁷, including the human gut, oceans, freshwater, air, and soil. However, we are just
82 beginning to understand the diversity and biogeography of microbial communities in wastewater
83 treatment plants (WWTPs)^{3,8}.

84

85 More than 300 km³ of wastewater is produced globally each year⁹. This volume equals one
86 seventh of the global river volume¹⁰. About 60% of this wastewater is treated prior to release,
87 and biological processes such as activated sludge are widely used in WWTPs⁹. Activated sludge
88 employs microbial flocs or granules to remove C, N, P, micropollutants (e.g., toxins, pesticides,
89 hormones, pharmaceuticals), and pathogens¹¹. Activated sludge relies on complex and
90 incompletely defined microbial communities. As the largest application of biotechnology in the
91 world¹², activated sludge is a vital infrastructure of modern urban societies¹³. Despite recent
92 advances in understanding the microbial ecology of activated sludge¹⁴⁻¹⁶, the global picture of
93 microbial diversity and distribution remains elusive. This information is essential to resolving
94 controversies concerning the relative importance of stochastic versus deterministic community
95 assembly in activated sludge³. Such information is also important for identifying key players in
96 the process and providing a basis for targeted manipulation of activated sludge microbiomes.

97
98 We created a Global Water Microbiome Consortium (GWMC) (<http://gwmc.ou.edu/>) and
99 conducted a global campaign for systematically collecting and analyzing activated sludge
100 microbiomes. We collected activated sludge samples from 269 WWTPs in 86 cities, 23
101 countries, and 6 continents (Fig. 1a, Supplementary Table 1). Deep sequencing and analysis of
102 16S rRNA genes were performed to address fundamental ecological questions, including: (i)
103 What is the extent of global diversity of activated sludge microbial communities? (ii) Does a
104 core microbiome exist in activated sludge processes across different continents? (iii) Do
105 activated sludge microbiomes show a latitudinal diversity gradient (LDG)? (iv) Is microbial
106 biodiversity important to function in activated sludge processes? and (v) What is the relative
107 importance of deterministic versus stochastic factors in regulating the composition, distribution,
108 and functions of activated sludge microbial communities?

109

110 **Species abundance distributions**

111

112 Species abundance distribution (SAD), a universal tool in ecology¹⁷ and central to biodiversity
113 theory, has not been rigorously tested in microbial ecology until recently¹⁸. Here, we tested
114 common SAD models, including Poisson lognormal, log-series, Broken-stick, and Zipf. The
115 Poisson lognormal model explained 99% of the variation of the activated sludge bacterial SADs,
116 compared with 72% for log-series, 94% for Zipf model, and 14% for Broken-stick (Fig. 1b;
117 Supplementary Table 2). Consistent with previous studies¹⁸, the Poisson lognormal model gave
118 the best fit to the observed SADs.

119

120 **Extent of global microbial diversity**

121
122 One grand challenge in biodiversity research is determining the number of species in an
123 ecological system¹⁹. We estimated the global richness of activated sludge bacterial communities
124 based on two parameters^{19,20}. One is the total number of individuals (N_T), which was estimated
125 as $4 - 6 \times 10^{23}$ bacteria in the global activated sludge community, based on published data⁹. The
126 other is the quantity of the most abundant taxa (N_{max}), which can be estimated based on either
127 our sequence data or the dominance-scaling law¹⁹. The lognormal model predicts $1.1 (\pm 0.07) \times$
128 10^9 species in activated sludge systems globally, with N_{max} at 1.2% of N_T based on our sequence
129 data. The number of species increases only slightly, to $2.0 (\pm 0.2) \times 10^9$ species, using $N_{max} =$
130 $0.4 \times N_T^{0.93}$ from the dominance-scaling law¹⁹ (Fig. 1c). The estimates of global activated sludge
131 bacterial richness are only about one order of magnitude lower than that of the global ocean
132 microbiome¹⁹ ($\sim 10^{10}$), even though the world's oceans represent an enormously larger
133 ecosystem, which could be attributed to the higher volumetric productivity, thus higher
134 concentration of bacterial cells, in activated sludge.

135

136 **Global core bacterial community**

137
138 Previous studies have reported the core community in WWTPs at regional scales. For example,
139 core genera existed in Danish¹⁴ and Asian¹⁵ WWTPs, but less than 10% of the genera overlapped.
140 Thus, a global core cannot be established from those regional studies.

141

142 At the global scale, occupancy-frequency and occupancy-abundance analyses revealed a hyper-
143 dominant pattern (Supplementary Fig. 1a) in which the 866 most abundant OTUs (1.39% of the
144 total OTU number) accounted for 50.06% of the total abundance. Similar hyper-dominance
145 patterns were observed in other macro-²¹ and microbial communities²².

146

147 A core bacterial community was determined based on abundance and occurrence frequency of
148 OTUs (see Methods for details). About 0.05% (28 OTUs) constituted a global core that
149 accounted for $12.4\% \pm 0.2\%$ (mean \pm SE) of the sequences in activated sludge samples (Fig. 2a;
150 Supplementary Table 3). Most (82%) of the core community members belonged to
151 *Proteobacteria*, with 15 OTUs classified as β -*Proteobacteria* (Fig. 2b). The most abundant OTU,
152 accounting for $1.14\% \pm 0.05\%$ of the sequence abundance in activated sludge samples and
153 occurring in 85% of all samples, was 99% similar to the γ -proteobacterium *Dokdonella*
154 *kunshanensis* DC-3²³. The second most abundant OTU ($0.89\% \pm 0.06\%$ in relative abundance
155 and occurring in 96% of all samples) belonged to *Zoogloea*, a dominant genus in activated
156 sludge communities¹⁵, with *Z. ramigera* known to enhance the flocculation of activated sludge²⁴.
157 A *Nitrospira* OTU (OTU_6) was also identified as a core taxon, reflecting its importance for
158 nitrite oxidation or complete ammonia oxidation in activated sludge^{25,26}. OTU_7 is closely
159 related to *Arcobacter* species, which are highly abundant in raw sewage²⁷ and include potential
160 pathogens, such as *A. cryaerophilus*, *A. butzleri*, and *A. skirrowii*²⁸. Furthermore, two putative
161 polyphosphate- accumulating organisms (PAOs), a “*Candidatus* Accumulimonas” OTU
162 (OTU_37) and a “*Candidatus* Accumulibacter” OTU (OTU_25), were identified as core taxa,
163 although only 149 out of the 269 sampled WWTPs operate as enhanced biological P removal

164 (EBPR) systems. Apparently, “*Candidatus Accumulimonas*” and “*Candidatus Accumulibacter*”
165 exhibit some metabolic versatility.

166
167 The global core community has some overlap with previous studies. For example, *Zoogloea*
168 species were proposed as core denitrifiers, and certain *Saprospiraceae* species play an important
169 role in hydrolysis in EBPR systems²⁹. However, some discrepancies also occurred. Saunders et al.
170 showed *Nitrotoga* rather than *Nitrospira* as primary nitrite-oxidizers in Danish WWTPs¹⁴.
171 Lawson et al. found low abundances of both *Nitrotoga* and *Nitrospira* in a pilot-scale EBPR
172 treatment plant, but *Nitrotoga* maintained high potential activities based on high SSU
173 rRNA:rDNA ratios³⁰. Regarding PAOs, we identified “*Candidatus Accumulimonas*” and
174 “*Candidatus Accumulibacter*” as global core taxa, while *Tetrasphaera* was the core PAO in
175 Danish WWTPs^{14,31}.

176
177 We similarly determined core communities for a variety of ecosystems at the global scale based
178 on the Earth Microbiome Project (EMP) datasets⁵. Soil, human feces, air, and freshwater
179 microbiomes had 9, 6, 2, and 1 bacterial OTUs identified as core taxa, respectively
180 (Supplementary Table 4). No core taxa were found for animal feces and the ocean, possibly due
181 to highly variable community compositions. Notably, the core community for activated sludge
182 had no overlap with the other habitats, suggesting that activated sludge selects for a unique core
183 community.

184

185 **Latitudinal diversity pattern**

186

187 Latitudinal diversity gradients (LDG), whereby species richness tends to decrease as latitude
188 increases³², are well documented in plant and animal ecology³³. Recently, several studies
189 examined LDG patterns in natural microbial communities, but found no clear trends^{6,7,34}. In
190 contrast, activated sludge operates under relatively stable and similar conditions everywhere.
191 Thus, one might not expect activated sludge microbial communities to exhibit LDG.

192

193 We examined the relationship between OTU richness and latitude. OTU richness peaked at
194 intermediate latitude, with a mean air temperature $\sim 15^{\circ}\text{C}$ (Fig. 1d). As taxonomic and
195 phylogenetic diversity were highly correlated ($R^2 = 0.92$), the trend was similar for phylogenetic
196 diversity (Supplementary Fig. 2a). These results suggest that a LDG does not occur in activated
197 sludge microbiomes; this parallels the global ocean microbiome⁷, but contrasts with some
198 ocean³⁴ and soil communities³⁵. In addition, the relationship between bacterial richness and
199 temperature (Supplementary Fig. 2b, c) did not fit predictions from the metabolic theory of
200 ecology³⁶. This theory cannot explain bacterial richness based on air temperature
201 (Supplementary Fig. 2b, $R^2 < 0.001$) and mixed liquid temperature (Supplementary Fig. 2c, $R^2 =$
202 0.03).

203

204 **Continental-level differences in bacterial community structure**

205

206 Variations in community composition (β -diversity) are key for understanding community
207 assembly mechanisms^{2,37} and ecosystem functioning³⁸. To understand how the bacterial
208 community composition of activated sludge varied across different spatial scales, we examined
209 taxonomic and phylogenetic diversity. First, diversity was highest in Asia and lowest in South

210 America (Supplementary Table 5). Second, considerable variations between activated sludge
211 samples were observed even at the phylum level (Supplementary Fig. 1b). Although the
212 taxonomic and phylogenetic community structures were not clearly separated at the OTU level in
213 two-dimensional ordinations (Supplementary Fig. 1c, d), PERMANOVA indicated that
214 taxonomic and phylogenetic composition were significantly different ($P < 0.001$) between any
215 two continents (Supplementary Table 6). Third, climate and activated sludge process type
216 exerted significant effects ($P = 0.001$) on microbial community structure, but these were
217 overwhelmed by continental geographical separation (Supplementary Table 7). For example,
218 bacterial communities of the same climate type in North America and Asia were distinguished by
219 their continental origins rather than being clustered together (Supplementary Fig. 1e, f). While
220 the activated sludge bacterial communities had higher similarity to those of freshwater and soil
221 than to other environments (Fig. 3a), they harbored a unique microbiome distinctly different
222 from all other habitats (Supplementary Table 8).

223

224 A Bayesian approach³⁹ was employed to identify potential sources of activated sludge bacterial
225 communities at the genus level. The most dominant potential source was freshwater, attributing
226 on average 46% of genera, followed by soil (17% on average) and ocean (12% on average) (Fig.
227 3b). Apparently, environmental characteristics are more similar between an activated sludge
228 bioreactor and freshwater than the others. Activated sludge and freshwater have potentially high
229 immigration events through connected water systems, such as wastewater discharge to rivers
230 after treatment.

231

232 **Scale-dependent distance-decay patterns**

233

234 Another fundamental pattern in ecology is the distance-decay relationship (DDR)^{17,40}, in which
235 community similarity decreases as geographic distance increases. Consistent with results in
236 other domains³⁷, we hypothesized that (i) the slope of the DDR curve would vary over local,
237 regional, and global scales, and (ii) the spatial turnover rates of activated sludge microbial
238 communities would be lower than those observed in natural habitats, especially for non-flowing
239 ecosystems, such as soils⁴¹.

240

241 Supporting our first hypothesis, significant negative DDRs ($P < 0.001$) were observed across all
242 scales based on taxonomic diversity (slope = -0.06 for Sorensen, -0.08 for Bray-Curtis, and -0.08
243 for Canberra distance) and phylogenetic diversity (slope = -0.04 for unweighted Unifrac, and -
244 0.02 for weighted Unifrac) (Fig. 4a, Supplementary Table 9). The slopes of DDRs depended
245 significantly on spatial scale. The DDR slopes across cities within a continent (-0.13 ~ -0.16 for
246 taxonomic similarity indices; -0.03 ~ -0.09 for phylogenetic similarity indices) were significantly
247 ($P = 0.001$) steeper (> 2 times) than the overall slopes for all similarity metrics (Supplementary
248 Table 9). Countering our second hypothesis, the overall spatial turnover rates of the activated
249 sludge communities were similar to those found in non-flowing natural habitats such as soils⁶
250 and sediments³⁷.

251

252 **Relationships between the community structure and activated sludge functions**

253

254 Understanding the relationships between biodiversity and ecosystem function is a critical topic in
255 ecology⁴². Despite decades of intensive studies, the biodiversity-function relationship is still
256 hotly debated, particularly in microbial ecology⁴³. A recent meta-analysis of the microbial
257 ecology literature found that less than one-half of all mechanistic claims were backed up by any
258 statistical tests⁴⁴. Since activated sludge is an engineered system, we hypothesized that there
259 would be a strong linkage between the activated sludge bacterial community structure and its
260 functions.

261
262 To assess functions, we calculated the removal rates of organic matter (biochemical oxygen
263 demand (BOD), chemical oxygen demand (COD)), total phosphorus, total nitrogen, and
264 ammonium nitrogen. Partial Mantel tests revealed that the distance-corrected changes of
265 activated sludge-community composition were significantly correlated with all measured
266 removal rates ($P < 0.032$), except for the ammonium-nitrogen removal rate ($P > 0.18$)
267 (Supplementary Table 10). Of the 28 global core OTUs, 27 were significantly correlated
268 (adjusted $P < 0.05$) with at least one of the five functions examined. Most of the correlations
269 (81%) were positive (Fig. 2c). Also, about 80% of the non-core OTUs showed significant
270 correlations (adjusted $P < 0.05$) with at least one function, and 40% of these correlations were
271 positive (Supplementary Fig. 3a). All of these results indicated that the structure of the activated
272 sludge bacterial communities, particularly the dominant populations, is critical to maintaining
273 activated sludge functions.

274
275 The global dataset also allows us to assess the importance of specific functional groups to
276 activated sludge functions. The nitrifying microbial community, including *Nitrospira* and

277 *Nitrosomonas* OTUs, showed a closer correlation with the ammonium- nitrogen removal rate
278 than did the whole community (Supplementary Table 10; P of Bray-Curtis distance =0.04).
279 Further analysis revealed significant positive correlations of *Nitrospira* (Spearman's $\rho = 0.40$,
280 adjusted $P < 0.001$) and *Nitrosomonas* (Spearman's $\rho = 0.21$, adjusted $P < 0.001$) abundance to
281 the percentages of ammonium-nitrogen removal (% of influent concentration), but not to the
282 ammonium-nitrogen removal rate (Supplementary Fig. 3b). *Nitrospira* was the top genus
283 correlating with the percentage of ammonium-nitrogen removal, corroborating its role in nitrite
284 oxidation in activated sludge. Regarding ammonium-oxidizing bacteria (AOB), an activated
285 sludge bioreactor harboured 15 *Nitrosomonas* OTUs on average, which made up $0.73\% \pm 0.06\%$
286 of the sequence abundance (Supplementary Table 11).

287

288 Consistent with our expectation, the activated sludge community composition was significantly
289 correlated with the TP removal rate for the samples from EBPR plants, but not for non-EBPR
290 plants (Supplementary Table 10), as P removal processes in non-EBPR plants are predominantly
291 chemical. The diversity of the three potential PAOs³¹ were significantly different ($P < 0.0001$,
292 two tailed paired-t test between any two organisms): 8.2 ± 0.2 “*Candidatus Accumulimonas*”
293 OTUs, 6.6 ± 0.2 “*Candidatus Accumulibacter*” OTUs, and 3.2 ± 0.1 *Tetrasphaera* OTUs within
294 a typical activated sludge bioreactor. While the relative abundance of “*Candidatus*
295 *Accumulimonas*” ($0.42\% \pm 0.06\%$) was not different from that of “*Candidatus Accumulibacter*”
296 ($0.42\% \pm 0.04\%$) (two tailed paired-t test, $P = 0.92$), both were more abundant than *Tetrasphaera*
297 (mean relative abundance $0.17\% \pm 0.02\%$) (two tailed paired-t test, $P < 0.0001$) (Supplementary
298 Table 12).

299

300 **Stochastic community assembly**

301

302 Since WWTPs are well-controlled engineered ecosystems, we hypothesized that the activated
303 sludge community assembly has a deterministic nature, and we calculated the null model-based
304 stochastic ratios⁴¹ with taxonomic and phylogenetic metrics. The average stochastic ratios based
305 on these four metrics all were higher than 0.75 (Fig. 4b), suggesting that stochastic factors were
306 more important than deterministic factors in influencing community composition, at least
307 partially contradicting our hypothesis.

308

309 To discern the relative importance of various factors contributing to spatial turnover of the
310 activated sludge bacterial communities, we performed multiple ‘regression on matrices’ (MRM)
311 analyses and a subsequent variance partition analysis (VPA) based on various taxonomic and
312 phylogenetic diversity metrics (Fig. 4c, Supplementary Fig. 4). Over all scales, the MRM model
313 explained considerable and significant portions of the community variations based on Bray-
314 Curtis similarity ($R^2 = 0.46$, $P = 0.001$) (Fig. 4c), with >50% variations unexplained. Among
315 these, 25%, 11%, and 10% of the variations were explained by geographical distance,
316 environmental variables, and their interactions, respectively (Fig. 4c). Similar trends were
317 observed across different scales, with environmental variables explaining < 30% of community
318 variations based on different similarity metrics (Supplementary Fig. 4). These results support
319 those inferred from the null-model-based stochastic ratio analysis.

320

321 **Environmental drivers of community composition**

322

323 Because both stochastic and deterministic factors are important in forming the activated sludge
324 community assembly, we attempted to discern the roles of individual deterministic factors in
325 shaping community structure. We correlated the geographic distance-corrected dissimilarities of
326 community composition with those of environmental variables by the partial Mantel test
327 (Supplementary Fig. 5a, Supplementary Table 13). Overall, the microbial community
328 composition had strong correlations with absolute latitude, mean annual temperature (MAT),
329 solids retention time (SRT, the average time which activated sludge solids are in the system), and
330 influent COD and BOD concentrations, representing organic matter ($r_m = 0.23-0.30$, $P = 0.001$).

331

332 More in-depth analysis by structural equation modeling (SEM) revealed direct and indirect
333 effects of the environmental drivers (Fig. 5a). Consistent with the Mantel test, temperature had
334 the strongest direct effects on PC1 representing the community structure (standardized path
335 coefficient, $\beta = 0.50$, $P < 0.001$). It also had weak negative impacts on species richness ($\beta = -$
336 0.14 , $P < 0.001$). This is consistent with previous observations at local^{45,46} and regional⁴⁷ scales
337 that highlighted temperature as a key factor influencing activated sludge community structure
338 and, in particular, abundance and diversity of slow-growing microorganisms such as AOB and
339 nitrite oxidizing bacteria (NOB).

340

341 Various biotic and abiotic factors (e.g., food-to-microorganisms ratio [F/M] (the ratio of organic
342 matter to microorganisms), dissolved oxygen concentration, and SRT) directly affected BOD-
343 removal rates (Fig. 5a). Influent BOD likely has an impact on bacterial composition through its

344 effect on the F/M ratio ($\beta = 0.31$, $P < 0.001$), which is inversely related to the SRT. Influent
345 BOD is the most influential environmental variable directly related to bacterial richness ($\beta = -$
346 0.28 , $P < 0.001$), and the abundance-weighted mean rRNA gene copy number significantly
347 increased with the influent BOD ($R^2 = 0.19$, $P < 0.0001$; Fig. 5b). All of these results are
348 consistent with the resource-competition theory⁴⁸, which predicts that high species diversity
349 occurs with low to intermediate supply of resources, but fast-growing r-strategists outcompete
350 efficient-scavenging K-strategists at high resource levels⁴⁹.

351
352 To independently test the strength of correlation for each of the three strongest parameters
353 (temperature, SRT, and influent BOD) with bacterial community structure, we performed
354 random-forest analysis, a machine learning-based method. Using species abundance as the input
355 data, the model predicted temperature, SRT, and influent BOD with an explained variance of
356 69%, 25%, and 18%, respectively (Fig. 5c, Supplementary Fig. 5b). When controlling for spatial
357 auto-correlation, models of temperature continued to have higher accuracy (Supplementary Fig.
358 5b). For example, the America-fitted model of temperature, i.e., a model trained solely by North
359 and South America samples, was able to capture variations in the temperatures of Asia samples
360 (cross-validated $R^2 = 0.47$) (Fig. 5c). The random-forest model also revealed the most important
361 OTUs for predicting temperature (Supplementary Fig. 5c). These results corroborate that
362 temperature is the major environmental variable shaping the activated sludge bacterial
363 compositions at the global scale, although it only has a weak effect on species richness (Fig. 5a).

364

365 **Conclusions and future perspectives**

366

367 Through well-coordinated international efforts, we systematically examined global diversity and
368 biogeography of activated sludge bacterial communities within the context of theoretical ecology
369 frameworks. Our findings enhance understanding of microbial ecology in activated sludge,
370 setting the stage for various future analyses of WWTP microbiomes, as well as other microbial
371 communities that span the globe.

372

373 Based on experimental and theoretical analyses, we estimate that activated sludge systems are
374 globally inhabited by $\sim 10^9$ different bacterial species. In contrast, only about 10^4 species have
375 been cultivated and studied in detail¹⁹. If we assume that all cultivated species are present in
376 activated sludge, potentially 99.999% of activated sludge microbial taxa remain uncultured.

377 Although more and more microorganisms have been genomically characterized, exploring
378 physiological attributes, which requires cultivation, represents a formidable task for future
379 microbiologists and process engineers⁵⁰. This finding also highlights how little we know of the
380 world's microbiome, even in one of the most common and well-controlled systems in the built
381 environment. Despite the very large diversity in activated sludge, a functionally important
382 global core community consists of fewer than 30 taxa. This core might serve as the “most
383 wanted” list for future experimental efforts to understand their genetic, biochemical,
384 physiological, and ecological traits.

385

386 Even though activated sludge is a managed ecosystem, its bacterial composition appears to be
387 driven most likely by stochastic processes, such as dispersal and drift, which apparently
388 contradicts conventional wisdom. However, deterministic factors (e.g., temperature, SRT, and
389 organic C inputs) play important roles in regulating the structure of the activated sludge

390 community. This could be important for developing operating strategies to maintain biodiversity
391 that promotes stable system performance. Perhaps one could overcome dispersal limitation by
392 establishing WWTPs, or repopulating failed WWTPs using an inoculum of activated sludge from
393 functioning WWTPs, which is a common practice in environmental engineering. Alternately,
394 one could alternate organic C loadings and/or operational conditions to manipulate the activated
395 sludge community's structure to select for the microorganisms having the desired functions.

396

397 Finally, apart from the practical implications of this study, it appears that the global bacterial
398 communities in activated sludge follow various macroecological patterns, such as SADs, DDRs,
399 resource theory, and community assembly mechanisms. Given that activated sludge can be
400 controlled and monitored, it could be an excellent system for testing how well different
401 macroecological theories apply to microbial ecology: e.g., the relationships among biodiversity,
402 food-web interactions, succession, stability, and ecosystem functioning.

403 **References**

- 404
- 405 1 Torsvik, V., Øvreås, L. & Thingstad, T. F. Prokaryotic diversity--magnitude, dynamics,
406 and controlling factors. *Science* **296**, 1064-1066 (2002).
- 407 2 Chase, J. M. & Myers, J. A. Disentangling the importance of ecological niches from
408 stochastic processes across scales. *Philos Trans R Soc Lond B Biol Sci* **366**, 2351-2363
409 (2011).
- 410 3 Ofitearu, I. D. *et al.* Combined niche and neutral effects in a microbial wastewater
411 treatment community. *Proc Natl Acad Sci USA* **107**, 15345-15350 (2010).
- 412 4 Zhou, J. *et al.* High-Throughput Metagenomic Technologies for Complex Microbial
413 Community Analysis: Open and Closed Formats. *mBio* **6**, e02288-02214 (2015).
- 414 5 Thompson, L. R. *et al.* A communal catalogue reveals Earth's multiscale microbial
415 diversity. *Nature* **551**, 457-463, doi:10.1038/nature24621 (2017).
- 416 6 Fierer, N. & Jackson, R. B. The diversity and biogeography of soil bacterial communities.
417 *Proc Natl Acad Sci USA* **103**, 626-631 (2006).
- 418 7 Sunagawa, S. *et al.* Structure and function of the global ocean microbiome. *Science* **348**,
419 1261359 (2015).
- 420 8 National Academies of Sciences, E. & Medicine. *Microbiomes of the built environment:
421 a research agenda for indoor microbiology, human health, and buildings.* (National
422 Academies Press, 2017).
- 423 9 Mateo-Sagasta, J., Raschid-Sally, L. & Thebo, A. in *Wastewater* 15-38 (Springer, 2015).
- 424 10 Gleick, P. H. Water resources. *Encyclopedia of climate and weather* **2**, 817-823 (1996).
- 425 11 van Loosdrecht, M. C. & Brdjanovic, D. Anticipating the next century of wastewater
426 treatment. *Science* **344**, 1452-1453 (2014).
- 427 12 Xia, S. *et al.* Bacterial community structure in geographically distributed biological
428 wastewater treatment reactors. *Environ Sci Technol* **44**, 7391-7396 (2010).
- 429 13 Grant, S. B. *et al.* Taking the “waste” out of “wastewater” for human water security and
430 ecosystem sustainability. *Science* **337**, 681-686 (2012).
- 431 14 Saunders, A. M., Albertsen, M., Vollertsen, J. & Nielsen, P. H. The activated sludge
432 ecosystem contains a core community of abundant organisms. *ISME J* **10**, 11 (2016).
- 433 15 Zhang, T., Shao, M.-F. & Ye, L. 454 Pyrosequencing reveals bacterial diversity of
434 activated sludge from 14 sewage treatment plants. *ISME J* **6**, 1137-1147 (2012).
- 435 16 Wagner, M. & Loy, A. Bacterial community composition and function in sewage
436 treatment systems. *Curr Opin Biotechnol* **13**, 218-227 (2002).
- 437 17 Morlon, H. *et al.* Spatial patterns of phylogenetic diversity. *Ecol Lett* **14**, 141-149 (2011).
- 438 18 Shoemaker, W. R., Locey, K. J. & Lennon, J. T. A macroecological theory of microbial
439 biodiversity. *Nat Ecol Evol* **1**, 107, doi:10.1038/s41559-017-0107 (2017).
- 440 19 Locey, K. J. & Lennon, J. T. Scaling laws predict global microbial diversity. *Proc Natl
441 Acad Sci USA* **113**, 5970-5975 (2016).
- 442 20 Curtis, T. P., Sloan, W. T. & Scannell, J. W. Estimating prokaryotic diversity and its
443 limits. *Proc Natl Acad Sci USA* **99**, 10494-10499 (2002).
- 444 21 Ter Steege, H. *et al.* Hyperdominance in the Amazonian tree flora. *Science* **342**, 1243092
445 (2013).
- 446 22 De Vargas, C. *et al.* Eukaryotic plankton diversity in the sunlit ocean. *Science* **348**,
447 1261605 (2015).

448 23 Li, Y. *et al.* Dokdonella kunshanensis sp. nov., isolated from activated sludge, and
449 emended description of the genus Dokdonella. *Int J Syst Evol Microbiol* **63**, 1519-1523
450 (2013).

451 24 Rosselló-Mora, R. A., Wagner, M., Amann, R. & Schleifer, K.-H. The abundance of
452 Zoogloea ramigera in sewage treatment plants. *Appl Environ Microbiol* **61**, 702-707
453 (1995).

454 25 Daims, H. *et al.* Complete nitrification by Nitrospira bacteria. *Nature* **528**, 504 (2015).

455 26 Daims, H., Nielsen, J. L., Nielsen, P. H., Schleifer, K.-H. & Wagner, M. In Situ
456 Characterization of Nitrospira-Like Nitrite-Oxidizing Bacteria Active in Wastewater
457 Treatment Plants. *Appl Environ Microbiol* **67**, 5273-5284 (2001).

458 27 Fisher, J. C., Levican, A., Figueras, M. J. & McLellan, S. L. Population dynamics and
459 ecology of Arcobacter in sewage. *Front Microbiol* **5** (2014).

460 28 Collado, L. & Figueras, M. J. Taxonomy, epidemiology, and clinical relevance of the
461 genus Arcobacter. *Clin Microbiol Rev* **24**, 174-192 (2011).

462 29 Nielsen, P. H., Saunders, A. M., Hansen, A. A., Larsen, P. & Nielsen, J. L. Microbial
463 communities involved in enhanced biological phosphorus removal from wastewater—a
464 model system in environmental biotechnology. *Curr Opin Biotechnol* **23**, 452-459 (2012).

465 30 Lawson, C. E. *et al.* Rare taxa have potential to make metabolic contributions in
466 enhanced biological phosphorus removal ecosystems. *Environ Microbiol* **17**, 4979-4993
467 (2015).

468 31 Stokholm-Bjerregaard, M. *et al.* A critical assessment of the microorganisms proposed to
469 be important to enhanced biological phosphorus removal in full-scale wastewater
470 treatment systems. *Front Microbiol* **8**, 718 (2017).

471 32 Hillebrand, H. On the generality of the latitudinal diversity gradient. *Am Nat* **163**, 192-
472 211 (2004).

473 33 Martiny, J. B. H. *et al.* Microbial biogeography: putting microorganisms on the map. *Nat*
474 *Rev Microbiol* **4**, 102-112 (2006).

475 34 Fuhrman, J. A. *et al.* A latitudinal diversity gradient in planktonic marine bacteria. *Proc*
476 *Natl Acad Sci USA* **105**, 7774-7778, doi:10.1073/pnas.0803070105 (2008).

477 35 Zhou, J. *et al.* Temperature mediates continental-scale diversity of microbes in forest
478 soils. *Nat Commun* **7**, 12083, doi:10.1038/ncomms12083 (2016).

479 36 Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M. & West, G. B. Toward a
480 metabolic theory of ecology. *Ecology* **85**, 1771-1789 (2004).

481 37 Martiny, J. B., Eisen, J. A., Penn, K., Allison, S. D. & Horner-Devine, M. C. Drivers of
482 bacterial beta-diversity depend on spatial scale. *Proc Natl Acad Sci USA* **108**, 7850-7854,
483 doi:10.1073/pnas.1016308108 (2011).

484 38 Zhou, J. *et al.* Stochastic assembly leads to alternative communities with distinct
485 functions in a bioreactor microbial community. *mBio* **4**, e00584-00512 (2013).

486 39 Knights, D. *et al.* Bayesian community-wide culture-independent microbial source
487 tracking. *Nat Methods* **8**, 761-763 (2011).

488 40 Zhou, J. & Ning, D. Stochastic Community Assembly: Does It Matter in Microbial
489 Ecology? *Microbiol Mol Biol Rev* **81**, e00002-00017 (2017).

490 41 Zhou, J. *et al.* Stochasticity, succession, and environmental perturbations in a fluidic
491 ecosystem. *Proc Natl Acad Sci USA* **111**, E836-E845 (2014).

492 42 Hooper, D. U. *et al.* A global synthesis reveals biodiversity loss as a major driver of
493 ecosystem change. *Nature* **486**, 105 (2012).

494 43 Krause, S. *et al.* Trait-based approaches for understanding microbial biodiversity and
495 ecosystem functioning. *Front Microbiol* **5**, 251 (2014).

496 44 Bier, R. L. *et al.* Linking microbial community structure and microbial processes: an
497 empirical and conceptual overview. *FEMS Microbiol Ecol* **91**, doi:10.1093/femsec/fiv113
498 (2015).

499 45 Wells, G.F. *et al.* Ammonia-oxidizing communities in a highly aerated full-scale
500 activated sludge bioreactor: betaproteobacterial dynamics and low relative abundance of
501 Crenarchaea. *Environ Microbiol* **11**, 2310-2328 (2009).

502 46 Karkman, A., Mattila, K., Tamminen, M. & Virta, M. Cold temperature decreases
503 bacterial species richness in nitrogen-removing bioreactors treating inorganic mine
504 waters. *Biotechnol Bioeng* **108**, 2876-2883 (2011).

505 47 Griffin, J.S. & Wells, G.F. Regional synchrony in full-scale activated sludge bioreactors
506 due to deterministic microbial community assembly. *ISME J* **11**, 500-511 (2017).

507 48 Tilman, D. *Resource competition and community structure*. (Princeton university press,
508 1982).

509 49 Wu, L. *et al.* Microbial functional trait of rRNA operon copy numbers increases with
510 organic levels in anaerobic digesters. *ISME J* **11**, 2874-2878, doi:10.1038/ismej.2017.135
511 (2017).

512 50 Pedrós-Alió, C. & Manrubia, S. The vast unknown microbial biosphere. *Proc Natl Acad*
513 *Sci USA* **113**, 6585-6587 (2016).

514

515

516 **Materials & Correspondence**

517

518 To whom correspondence should be addressed regarding analysis, synthesis and reprints, E-mail:
519 jzhou@ou.edu. To whom correspondence should be addressed regarding experimental design
520 and sampling, E-mails: xhwen@tsinghua.edu.cn, tom.curtis@newcastle.ac.uk, qianghe@utk.edu.

521

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523

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533

534 **Author contributions**

535 All authors contributed experimental assistance and intellectual input to this study. The original
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537 by JZ.Z., XH.W., T.P.C., Q.H., ZL.H., and DL.N.; Sample collections were coordinated by Q.H.,
538 DL.N., XH.W., T.P.C., B.Z., M.B., G.F.W., JZ.Z., and other GWMC members. J.D.V.N and
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540 J.V., A.N.R., D.D.C.V., C.E., L.C., J.C.A., C.D.L., L.C.M., A.C., Pa.B., D.A.) did DNA
541 extraction. P.Z. performed DNA sequencing with the help from LY.W.; LW.W., DL.N., JZ.Z.,
542 B.Z., XY.S., QT.Z., FQ.L., NJ.X., and RM.T. performed data analysis with help from Y.D.,
543 QC.T., T.Z., Ya.Z and AJ.W.; LW.W., JZ.Z., and DL.N. wrote the manuscript with the help from
544 B.E.R., L.A.-C., M.W., C.S.C., D.A.S., G.F.W., J.M.T., P.J.J.A., J.K., J.V., P.H.N., R.G.L.,
545 XH.W., ZL.H., and YF.Y..

546 **Global Water Microbiome Consortium**

547

548 Members besides the authors listed on the first page:

549 Dany Acevedo¹, Miriam Agullo-Barcelo², Gary L. Andersen^{3,4}, Juliana Calabria de Araujo⁵,
550 Kevin Boehnke⁶, Philip Bond², Charles B. Bott⁷, Patricia Bovio⁸, Rebecca K. Brewster⁶, Faizal
551 Bux⁹, Angela Cabezas⁸, Léa Cabrol^{10,11}, Si Chen¹², Ting Chen^{13,14}, Claudia Etchebehere⁸,
552 Amanda Ford⁷, Dominic Frigon¹⁵, Janeth Sanabria Gómez¹, James S. Griffin¹⁶, April Z. Gu¹⁷,
553 Moshe Habagil¹⁸, Lauren Hale¹⁹, Steven D. Hardeman²⁰, Marc Harmon²¹, Harald Horn²²,
554 Zhiqiang Hu²³, Shameem Jauffur^{15,24}, David R. Johnson²⁵, Alexander Keucken^{18,26}, Sheena
555 Kumari⁹, Cintia Dutra Leal⁵, Laura A. Lebrun²⁷, Jangho Lee²⁸, Minjoo Lee²⁸, Zarraz MP Lee²⁹,
556 Mengyan Li³⁰, Xu Li³¹, Yu Liu^{32,29}, Richard G. Luthy³³, Leda C. Mendonça-Hagler³⁴, Francisca
557 Gleire Rodriguez de Menezes³⁵, Arthur J. Meyers³⁶, Amin Mohebbi^{31,37}, Adrian Oehmen³⁸,
558 Andrew Palmer³⁹, Prathap Parameswaran⁴⁰, Joonhong Park²⁸, Deborah Patsch²⁵, Valeria
559 Reginatto⁴¹, Francis L. de los Reyes III⁴², Adalberto Noyola Robles⁴³, Simona Rossetti⁴⁴,

560 Jatinder Sidhu³⁹, William T. Sloan⁴⁵, Kylie Smith³⁹, Oscarina Viana de Sousa³⁵, Kyle Stephens⁴⁶,
561 Renmao Tian¹⁹, Nicholas B. Tooker¹⁷, Daniel De los Cobos Vasconcelos⁴³, Steve Wakelin⁴⁷, Bei
562 Wang⁴⁸, Joseph E. Weaver⁴², Stephanie West²², Paul Wilmes²⁷, Sung-Geun Woo³³, Jer-Horng
563 Wu⁴⁹, Liyou Wu¹⁹, Chuanwu Xi⁶, Meiyang Xu⁵⁰, Tao Yan⁵¹, Min Yang⁵², Michelle Young⁴⁰,
564 Haowei Yue⁵³, Qian Zhang⁵¹, Wen Zhang⁵⁴, Yu Zhang⁵², Hongde Zhou⁴⁸

565
566 **Steering Committee:** Jizhong Zhou^{19,53,55}, Xianghua Wen⁵³, Thomas P. Curtis⁵⁶, Qiang He^{12,57},
567 Zhili He^{58,59}; **Contact:** Jizhong Zhou (jzhou@ou.edu), Daliang Ning^{19,60,53} (ningdaliang@ou.edu)

568
569 ¹Environmental Microbiology and Biotechnology Laboratory, Engineering School of
570 Environmental & Natural Resources, Engineering Faculty, Universidad del Valle–Sede
571 Meléndez, Cali, Colombia; ²Advanced Water Management Centre, The University of
572 Queensland, Brisbane, QLD, Australia; ³Department of Environmental Science, Policy, and
573 Management, University of California, Berkeley, USA; ⁴Environmental Genomics and Systems
574 Biology Division, Lawrence Berkeley National Laboratory, Berkeley, USA; ⁵Universidade
575 Federal de Minas Gerais, Departamento de Engenharia Sanitária e Ambiental, Brazil;
576 ⁶Department of Environmental Health Sciences, The University of Michigan, Ann Arbor, MI,
577 USA; ⁷Hampton Roads Sanitation District (HRSD), Virginia Beach, VA, USA; ⁸Microbial
578 Ecology Laboratory, Microbial Biochemistry and Genomics Department, Biological Research
579 Institute “Clemente Estable”, Montevideo, Uruguay; ⁹Institute of Water and Wastewater
580 Technology, Durban University of Technology, South Africa; ¹⁰Aix-Marseille University CNRS
581 IRD, MIO UM110 Mediterranean Institute of Oceanography, Marseille, France; ¹¹Escuela de
582 Ingeniería Bioquímica, Pontificia Universidad Católica de Valparaíso, Valparaíso, Chile;
583 ¹²Department of Civil and Environmental Engineering, The University of Tennessee, Knoxville,
584 TN, USA; ¹³Department of Computer Science, Tsinghua University, Beijing, China;
585 ¹⁴Bioinformatics Division, Tsinghua National Laboratory of Information Science and
586 Technology, Tsinghua University, Beijing, China; ¹⁵Microbial Community Engineering
587 Laboratory, Department of Civil Engineering and Applied Mechanics, McGill University,
588 Montreal, Canada; ¹⁶Department of Chemical and Biological Engineering, Northwestern
589 University, Evanston, IL, USA; ¹⁷Department of Civil and Environmental Engineering,
590 Northeastern University, Boston, MA, USA; ¹⁸Vatten & Miljö i Väst AB (VIVAB), Falkenberg,
591 Sweden; ¹⁹Institute for Environmental Genomics, Department of Microbiology and Plant
592 Biology, and School of Civil Engineering and Environmental Sciences, University of Oklahoma,
593 Norman, OK, USA; ²⁰Norman Water Reclamation Facility, Norman, OK, USA; ²¹Golden Heart
594 Utilities, Fairbanks, AK, USA; ²²Karlsruhe Institute of Technology, Engler-Bunte-Institut, Water
595 Chemistry and Water Technology, Germany; ²³Department of Civil and Environmental
596 Engineering, University of Missouri, Columbia, MO, USA; ²⁴Department of Building, Civil and
597 Environmental Engineering, Concordia University, Montreal, Canada; ²⁵Department of
598 Environmental Microbiology, Eawag, Dübendorf, Switzerland; ²⁶Water Resources Engineering,
599 Faculty of Engineering, Lund University, Lund, Sweden; ²⁷Luxembourg Centre for Systems
600 Biomedicine, University of Luxembourg, Belvaux, Luxembourg; ²⁸Department of Civil and
601 Environmental Engineering, Yonsei University, Seoul, South Korea; ²⁹Advanced Environmental
602 Biotechnology Centre, Nanyang Environment & Water Research Institute, Nanyang
603 Technological University, Singapore; ³⁰Department of Civil and Environmental Engineering,
604 Rice University, Houston, TX, USA; ³¹Department of Civil Engineering, University of
605 Nebraska-Lincoln, NE, USA; ³²School of Civil and Environmental Engineering, Nanyang

606 Technological University, Singapore; ³³Department of Civil and Environmental Engineering,
607 Stanford University, Stanford, CA, USA; ³⁴Plant Biotechnology Program, Federal University of
608 Rio de Janeiro, UFRJ, Brazil; ³⁵Federal University of Ceará, UFC, Brazil; ³⁶University of
609 Tennessee, Center for Environmental Biotechnology, Knoxville, TN, USA; ³⁷Department of
610 Civil Engineering, Construction Management and Environmental Engineering, Northern Arizona
611 University, Flagstaff, AZ, USA; ³⁸UCIBIO, REQUIMTE, Departamento de Química, Faculdade
612 de Ciências e Tecnologia, Universidade Nova de Lisboa, Caparica, Portugal; ³⁹CSIRO Land and
613 Water, Ecosciences Precinct, QLD, Australia; ⁴⁰Biodesign Swette Center for Environmental
614 Biotechnology, Arizona State University, Tempe, AZ, USA; ⁴¹Departamento de Química,
615 Universidade de São Paulo, Faculdade de Filosofia Ciências e Letras de Ribeirão Preto –
616 FFCLRP, Ribeirão Preto, SP, Brazil; ⁴²Department of Civil, Construction, and Environmental
617 Engineering, North Carolina State University, NC, USA; ⁴³Grupo de Investigación en Procesos
618 Anaerobios, Instituto de Ingeniería, Universidad Nacional Autónoma de México, México, DF,
619 Mexico; ⁴⁴CNR-IRSA, National Research Council, Water Research Institute, Rome, Italy;
620 ⁴⁵Infrastructure and Environment Research Division, School of Engineering, University of
621 Glasgow, Glasgow, UK; ⁴⁶Tryon Creek & Columbia Blvd. Wastewater Treatment Plants, Bureau
622 of Environmental Services, City of Portland, OR, USA; ⁴⁷Scion Research, Christchurch, New
623 Zealand; ⁴⁸School of Engineering, University of Guelph, Guelph, Ontario, Canada; ⁴⁹Department
624 of Environmental Engineering, National Cheng Kung University, Tainan City, Taiwan, ROC;
625 ⁵⁰State Key Laboratory of Applied Microbiology Southern China, Guangdong Provincial Key
626 Laboratory of Microbial Culture Collection and Application, Guangdong Institute of
627 Microbiology, Guangzhou, Guangdong, China; ⁵¹Department of Civil and Environmental
628 Engineering, University of Hawaii at Manoa, Honolulu, HI, USA; ⁵²State Key Laboratory of
629 Environmental Aquatic Chemistry, Research Center for Eco-Environmental Sciences, Chinese
630 Academy of Sciences, Beijing, China; ⁵³State Key Joint Laboratory of Environment Simulation
631 and Pollution Control, School of Environment, Tsinghua University, Beijing, China;
632 ⁵⁴Department of Civil Engineering, University of Arkansas, Fayetteville, AR, USA; ⁵⁵Earth and
633 Environmental Sciences, Lawrence Berkeley National Laboratory, Berkeley, CA, USA; ⁵⁶School
634 of Engineering, Newcastle University, Newcastle upon Tyne, UK; ⁵⁷Institute for a Secure and
635 Sustainable Environment, The University of Tennessee, Knoxville, TN, USA; ⁵⁸Environmental
636 Microbiomics Research Center, School of Environmental Science and Engineering, Sun Yat-Sen
637 University, Guangzhou, China; ⁵⁹Guangdong Provincial Key Laboratory of Environmental
638 Pollution Control and Remediation Technology, Guangzhou, China; ⁶⁰Consolidated Core
639 Laboratory, University of Oklahoma, Norman, Oklahoma, USA.

640

641 **Competing interests**

642

643 The authors declare no competing financial interests.

644

645 **Methods**

646

647 **Global sampling and meta-data collection**

648

649 The Global Water Microbiome Consortium (GWMC) was initiated in May 2014 as a platform to
650 facilitate international collaboration and communication on research and education for global
651 water microbiome studies (<http://gwmc.ou.edu/>). The GWMC is a collaboration across more
652 than 70 research groups from 23 countries. As the first initiative of GWMC, we launched this
653 study with a global sampling campaign targeting municipal wastewater treatment plants
654 (WWTPs) by focusing on the activated sludge process. Unlike the Earth Microbiome Project
655 (EMP), which employed a bottom-up strategy to solicit microbial samples⁵, we used a top-down
656 approach to select WWTPs for sampling by considering their latitudes, climate zones, spatial
657 scales, activated sludge process type, and accessibility for sampling.

658

659 The main goal of this study was to provide system-level mechanistic understanding of global
660 diversity and distribution of municipal WWTP microbiomes. WWTPs were selected based on
661 the following criteria: **(i) Continental-level geographic locations.** Samples were obtained from
662 all continents except for Antarctica, but with special focus on North America, Asia, and Europe
663 (Fig. 1a). Because of the low accessibility, WWTPs in Africa and South America were under-
664 represented. **(ii) Latitude.** To address questions related to latitudinal diversity gradient (LDG),
665 WWTPs were intensively sampled in North America along the East and West Coasts, and
666 Highway 35, as well as Highway 40 (from East to West) (Fig. 1a), in Asia, Europe, and Australia.
667 The WWTPs sampled spanned latitudes from 43.6°S to 64.8 °N. **(iii) Climate zones.** Since
668 climate could have significant impacts on microbial communities, the samples covered 17

669 different climate types (Supplementary Fig. 6). To distinguish independent effects of continents
670 versus climate zones, we increased sampling efforts for climate zones that were present in
671 multiple continents, such as Humid Subtropical Climate. **(iv) Scales.** The samples were
672 collected from very broad spatial scales: global (across 6 continents), regional (e.g., individual
673 continents or climate zones), and local (e.g., individual cities). Within some cities, multiple
674 WWTPs and multiple samples per WWTP were collected; **(v) Wastewater treatment process**
675 **types.** To address the relationship of structure to function for activated sludge, we sampled the
676 aerobic zone of conventional plug flow, oxidation ditch, sequential batch reactors,
677 anaerobic/anoxic/oxic (A²O), and other activated sludge process types.

678

679 A unified protocol was used for sampling, sample preservation, metadata collection, DNA
680 extraction, sequencing, and sequence analysis, to minimize potential experimental variations^{4,51-}
681 ⁵³. Detailed sampling and metadata collection methods and protocols are available at the
682 GWMC web site (<http://gwmc.ou.edu/protocols/view/11>).

683

684 Sampling was carried out in June to November 2014 in the Northern Hemisphere and December
685 2014 to April 2015 in the Southern Hemisphere. The sampling time was generally between
686 10:00 am to 2:00 pm, when the WWTPs were relatively stable under normal conditions.

687 Although we tried to collect the global samples in the same season, seasonal temporal turnover in
688 activated sludge communities could have had some effect on the community variations we
689 observed to some degree. Based on limited published work^{54,55}, such temporal variations should
690 be much smaller than the spatial variations at the global/continental scales. For example, a
691 previous study on 5-year temporal dynamics of activated sludge community showed no

692 significant seasonal succession⁵⁴. It's also revealed that the activated sludge communities were
693 relatively stable across three months, with average Bray-Curtis distance 0.45 ± 0.10 (mean \pm SD)
694 between samples⁵⁵; this variation was smaller than our observed mean variations even at local
695 city level (0.54 ± 0.19) (Fig. 4a).

696

697 At local scale, we defined a city based on it having a large enough geographic scale, not on an
698 administrative division (see Supplementary Table 1 for defined cities). For each city, we usually
699 collected at least 12 samples, and had ≥ 12 samples/city in 77% cities, with < 3 samples/city in
700 only 1% of cities. We also sampled at least 2 WWTPs in 72% of the cities. In each plant, we
701 collected at least 3 mixed liquor samples, generally from 3 different positions (the front, middle,
702 and end part) of the aerobic zone in each aeration tank. In a few cases (3.3% plants), where only
703 one sampling position was applicable, 3 samples were taken in sequence with at least 30-min
704 interval. Altogether, we collected 1,186 activated sludge samples from 269 WWTPs across 23
705 countries from global scale (e.g., across 6 continents), regional scale (e.g., individual continents),
706 to local scale (e.g., geographic sites or individual cities) (Fig. 1a).

707

708 At each sampling position, approximately 1 liter mixed liquor was sampled and well mixed, and
709 40 mL was transferred into a sterile tube. The mixed liquor samples were kept on ice ($\leq 4^\circ\text{C}$),
710 transported to laboratory within 24 hours, divided into aliquots, and then centrifuged at 4°C ,
711 15,000 g for 10 min to collect pellets. Sludge pellets were transported (if necessary) with dry ice
712 to the designated laboratories within 48 hours and preserved at -80°C before DNA extraction.

713

714 Along with the sludge samples, associated metadata, conforming to the Genomic Standards
715 Consortium's MIxS and Environmental Ontology (ENVO) Standards^{56,57}, were provided by
716 plant managers and/or investigators (Supplementary Table 1; Supplementary Fig. 7). We
717 collected metadata (e.g., chemical properties, operation conditions, process type) from each plant
718 using a standard sampling data sheet, which ensured that the data from all plants was in the same
719 format. Raw metadata were processed as one metadata table (Supplementary Table 1) and
720 classified into three categories: geological variables, plant operation and monitoring variables,
721 and sample properties. The geological variables included latitude and longitude; ambient climate
722 variables such as climate type, mean annual temperature (MAT), and precipitation; and
723 population size and gross domestic product (GDP) for the city where the WWTP was located.

724

725 Climate type was determined by the Köppen-Geiger climate classification⁵⁸. GDP and
726 population data were derived from the Brookings analysis of Global Metro Monitor⁵⁹. Variables
727 related to plant design and operation include plant age, design capacity, actual flow rate, volume
728 of aeration tanks, hydraulic retention time (HRT) and solids retention time (SRT). The activated
729 sludge process type, aerator type, and coupling with N removal processes (nitrification and
730 denitrification) in the WWTP were also provided by the plant managers as possible. Plant
731 monitoring variables include influent and effluent biochemical oxygen demand (BOD) and
732 chemical oxygen demand (COD) representing organic carbon (C) level, total nitrogen and total
733 phosphorus representing nutrient level, ammonium N, as well as the food to microorganism (F/M)
734 ratio, indicating the average organic C loading to microorganisms. For sample properties, most
735 plant managers provided the yearly average value of mixed liquor suspended solids (MLSS),

736 indicating the concentration of biomass in the activated sludge, dissolved oxygen, pH, and mixed
737 liquid temperature; some provided the measured values when sampling.

738

739 Activated sludge performance was calculated as the specific removal rates (g per g biomass per
740 day) of organic C (BOD and COD), nutrients (total nitrogen and total phosphorus) and
741 ammonium nitrogen (NH₄-N):

742

$$\text{removal rate} = \frac{(\text{Influent}(X) - \text{Effluent}(X)) \times \text{flow rate}}{\text{MLSS} \times \text{aerobic tank volume}}$$

743

744 The WWTPs represent diverse geographies and a large range of climatic conditions, operation
745 parameters, and chemical conditions across and within continents (Supplementary Fig. 7). For
746 instance, the average influent BOD ranged from 30 to 1,000 mg/L. Such a broad range of
747 diverse parameters is critical to disentangling mechanisms of activated sludge microbial
748 community assembly.

749

750 **DNA Extraction**

751

752 To minimize the variations associated with sample processing, identical protocols were used in
753 DNA extraction and 16S rRNA gene sequencing. All samples from China and Japan were
754 shipped to Dr. Xianghua Wen's Laboratory at Tsinghua University for DNA extraction. All
755 other samples, including samples from Europe collected by Dr. Thomas Curtis at Newcastle
756 University, were shipped to Dr. Jizhong Zhou's Laboratory at University of Oklahoma (OU) for

757 DNA extraction. Due to the tight restriction of sample shipment in South Africa, Mexico, Chile,
758 Uruguay, and Brazil, the DNA was extracted by GWMC members in these countries. DNA was
759 extracted from sludge samples using MoBio PowerSoil DNA isolation kit. For each sample, a
760 pellet from 3 mL mixed liquor was used. In addition to the manufacture protocol, we always
761 placed exactly 12 bead tubes on the vortex evenly and vortex at maximum speed for 10 min to
762 minimize the lysis efficiency difference between samples. All DNA samples were processed at
763 OU for sequencing.

764

765 DNA quality for all samples was evaluated with a NanoDrop spectrophotometer (NanoDrop
766 Technologies Inc., Wilmington, DE, USA) at OU. Final DNA concentrations were quantified
767 using PicoGreen with a FLUO star Optima instrument (BMG Labtech, Jena, Germany). Purified
768 DNA was stored at -80 °C.

769

770 **16S rRNA gene sequencing and sequence processing**

771

772 The V4 region of the 16S rRNA gene was amplified and sequenced using standardized protocols
773 with the phasing amplicon sequencing (PAS) approach as described previously⁶⁰ and the primers
774 515F (GTGCCAGCMGCCGCGGTAA) and 806R (GGACTACHVGGGTWTCTAAT) of the
775 Earth Microbiome Project⁶¹. *In silico* primer coverage analysis using SILVA TestPrime 1.0⁶²
776 and SILVA dataset r123 showed that these primers cover 86.8% and 52.9% of all bacterial and
777 archaeal sequences with 0 mismatches, respectively.

778

779 To mitigate quantitative problems associated with amplicon sequencing⁵², the 16S rRNA gene
780 fragments were amplified from community DNAs (10 ng) with two-step PCR using lower
781 numbers of amplification cycles (10 and 20 cycles for the 1st and 2nd step, respectively). The
782 two-step PAS approach offers several advantages: lower amplification biases, better sequence-
783 read quality, higher effective sequence read numbers and length, and lower sequencing errors⁶⁰.
784 All samples were sequenced using the same MiSeq instrument at the Institute for Environmental
785 Genomics, OU. Generally, about 400 samples were combined for each round of MiSeq
786 sequencing. Since the numbers of sequence reads varied substantially from sample to sample,
787 most samples were sequenced more than once (e.g., 19% twice; 33%, three times; 43%, > 3
788 times) to meet the target number of about 30K sequencing reads per sample, as determined in our
789 previous analysis⁶³.

790
791 The numbers of sequences (reads) per sample ranged from 25,631 to 351,844 (Supplementary
792 Table 5), and a total of 96,148 OTUs were obtained. About 1.3% of these OTUs were from
793 archaea, which accounted for 0.13% of the total abundance. The choice of the PCR primer pair
794 506F/806R (that was also used in the EMP project) is very likely to have strongly influenced this
795 low archaeal abundance due to the much lower coverage of the primers of archaeal 16S rRNA
796 genes compared to the bacterial counterparts. Because of the low archaeal abundance, the term
797 “bacteria” is used for simplicity. Also, the terms microbiome and microbial (or bacterial)
798 community are used interchangeably.

799
800 Raw sequence data were processed as previously described³⁵, except for OTU generation by
801 UPARSE⁶⁴ at the 97% similarity threshold, resulting in 96,148 OTUs. We define operational

802 taxonomic units (OTUs) (based on 97% sequence similarity) for bacterial and archaeal
803 phylotypes. Although there is potential misconnection between OTUs and microbial species⁶⁵,
804 we use this popular definition for simplicity, and it also allows us to compare with previous
805 studies of other systems. The representative sequences were aligned using Clustal Omega
806 v1.2.2⁶⁶ for constructing the phylogenetic tree by FastTree2 v2.1.10⁶⁷. OTUs were
807 taxonomically annotated by RDP Classifier⁶⁸ with a confidence cutoff of 80%, using the MiDAS
808 database (version 2.1) which specifically provides a curated taxonomy for abundant and
809 functionally important microorganisms in activated sludge⁶⁹. After removal of the global
810 singletons⁶⁴, the sequence number in each sample was rarefied to the same depth (25,600
811 sequences per sample), resulting in 61,448 OTUs overall, which were used in subsequent
812 comparative analyses.

813

814 Although our sequencing depths were considerably higher than those in many similar studies⁷⁰,
815 rarefaction curves (Supplementary Fig. 2d, e) of activated sludge microbial communities
816 indicated that additional rare taxa were likely present in individual samples. Nevertheless,
817 pooling all sequences gave a sufficient number for estimating global- and continent-level
818 diversity of activated sludge microbial communities (Supplementary Fig. 2f, g). The global
819 OTU richness per sample was 2,309±559 (Supplementary Table 5). Besides richness, we also
820 calculated other alpha diversity indices on a global and regional scale (Supplementary Table 5).

821

822 The rRNA operon copy number for each OTU was estimated through the rrnDB database based
823 on its closest relatives with known rRNA operon copy number⁷¹. The abundance-weighted mean
824 rRNA operon copy number was then calculated for each sample as described previously⁴⁹.

825

826 **Sequence comparison against reference databases**

827

828 To compare the sequence diversity in this study to that in existing databases, the 96,148
829 representative sequences from the activated sludge samples were compared against the
830 representative set (97% similarity level) of full-length sequences from Greengenes 13.8⁷²
831 (released on August 2013) and the non-eukaryotic fraction of Silva 132 databases⁷³ (released on
832 December 2017). We used the open-source sequence search tool USEARCH10⁷⁴ in global
833 alignment search mode, and we required 97% similarity across the query sequence. Our
834 activated sludge sequences match to 38.6% of Greengenes and 37.2% of SILVA 16S rRNA gene
835 OTUs at 97% similarity. These matches accounted for 18.2% and 22.5% of the representative
836 sequences in our datasets, respectively, indicating that the majority of activated sludge microbial
837 species diversity is not yet captured in full-length sequence databases; this is similar to the
838 observations in the EMP⁵.

839

840 **Species abundance distribution (SAD) fitting**

841

842 We compared the SAD of each sample, based on the rank-abundance distribution, with
843 predictions from Poisson lognormal, log-series, Broken-stick, and Zipf models. Although
844 numerous SAD models are available, lognormal and log-series have been the most successful in
845 predicting SADs, and they are the standards for testing other models¹⁸. While the logseries
846 model is well supported by macroecological studies, the Poisson lognormal model is more
847 commonly observed with microorganisms¹⁸. By comparing (rank-for-rank) the observed and

848 predicted SADs using regression analysis, we could directly infer the percentages of variations in
 849 abundance among species explained by each model using the same code, developed by
 850 Shoemaker et al¹⁸.

851

852 **Estimation of global bacterial diversity of WWTPs**

853

854 We used the methods described in Curtis et al.²⁰ and Locey and Lennon¹⁹ to predict global
 855 bacterial richness (S_T) using the lognormal model. The lognormal prediction of S_T is based on
 856 the total abundance (N_T), the abundance of the most abundant species (N_{max}), and the assumption
 857 that the rarest species is a singleton, $N_{min} = 1$. In communities with N_T individuals, the richness
 858 can be estimated by:

$$859 \quad S_T = \frac{\sqrt{\pi}}{a} \exp \left\{ \left(a \log_2 \left(\sqrt{\frac{N_{max}}{N_{min}}} \right) \right)^2 \right\} \quad (i)$$

860

861 where a is an inverse measure of the width of the distribution, which can be numerically solved
 862 from:

$$863 \quad N_T = \frac{\sqrt{\pi N_{min} N_{max}}}{2a} \exp \left\{ \left(a \log_2 \left(\sqrt{\frac{N_{max}}{N_{min}}} \right) \right)^2 \right\} \exp \left\{ \left(\frac{\ln(2)}{2a} \right)^2 \right\} \left[\operatorname{erf} \left(a \log_2 \left(\sqrt{\frac{N_{max}}{N_{min}}} - \frac{\ln(2)}{2a} \right) \right) + \right. \\ 864 \quad \left. \operatorname{erf} \left(a \log_2 \left(\sqrt{\frac{N_{max}}{N_{min}}} + \frac{\ln(2)}{2a} \right) \right) \right] \quad (ii)$$

865

866 We used published data to estimate the total microbial abundance in WWTPs as follows.

867 Empirical records compiled from a variety of sources, for example, AQUASTAT⁷⁵ and Sato et al
 868 2013⁷⁶, suggest that about $330 \text{ km}^3 \text{ year}^{-1}$ of municipal wastewater are produced globally, of

869 which 60% is treated⁹. Assuming that they are all treated in WWTPs, then about 0.54 km³
870 municipal wastewater are treated by WWTPs globally per day. The total effective volume of
871 aerobic tanks of WWTPs can be estimated by:

$$872 \quad V = Q \times HRT \quad \text{(iii)}$$

873 where Q is the influent flow rate (m³ day⁻¹) and HRT is the hydraulic retention time (day) of the
874 aerobic tank. Our dataset indicates that the average HRT of aerobic tanks is 9.8 (± 0.3 s.e.) hours.
875 Thus, the total effective volume is estimated as 0.22 (± 0.007) km³. The total cells in activated
876 sludge are about 2.3 (± 0.4) × 10⁹ (ml⁻¹)⁷⁷; thus, N_T (global activated sludge bacterial abundance)
877 is about 4.0- 6.1 × 10²³.

878

879 We then estimated N_{max} based on the ratio of N_{max} to N_T of our sequencing data, i.e., the relative
880 abundance of the most abundant OTU, or using scaling law¹⁹. The knowledge of N_T, N_{max}, and
881 N_{min} allows equation (ii) to be solved numerically for the parameter *a* and, subsequently, for S_T
882 using equation (i).

883

884 Using the same method, we estimated the total bacterial richness of individual WWTPs, along
885 with WWTPs in the United States and China. The volume of aerobic tanks of a WWTP in
886 Beijing, China is 10,000 m³, making the total cells about 2.3 (± 0.4) × 10¹⁹. N_T of WWTPs in
887 US and China were estimated based on their published data of treating amount^{78,79}, with
888 activated sludge harboring similar numbers of species for the US (4.6 × 10⁸ to 1.1 × 10⁹) and
889 China (3.9 × 10⁸ to 1.0 × 10⁹). N_{max} was further estimated based on our 16S rRNA gene
890 sequencing data or using a scaling law¹⁹. The total bacterial richness estimates of individual
891 human gut, individual cow rumen, global ocean and Earth were taken from Locey and Lennon¹⁹.

892

893 **Core community determination**

894 A global-scale core microbial community was determined based on multiple reported measures.

895 First, “overall abundant OTUs” were filtered out according to mean relative abundance across all

896 samples (MRA)⁸⁰. Previous studies used different criteria (e.g., MRA > 1%^{30,81} or 0.1%^{82,83})

897 without any objective or standard rule. Thus, we selected all top 0.1% OTUs (62) as overall

898 abundant OTUs. Their MRA was higher than 0.2%, within the range of reported criteria.

899 Second, “ubiquitous OTUs” were defined as OTUs with occurrence frequency in more than 80%

900 of all samples⁸⁴. Finally, “frequently abundant OTUs” were selected based on their relative

901 abundances with a sample. In each sample, the OTUs were defined as abundant when they had a

902 higher relative abundance than other OTUs and made up the top 80% of the reads in the sample¹⁴.

903 A frequently abundant OTU was defined as abundant in at least half samples, which is stricter

904 than the reported criterion (10 in 26 samples¹⁴). Since the above three measures are

905 complementary to one another when defining core community, only OTUs fulfilling all three

906 criteria were defined as the global scale core bacterial community.

907

908 Following the same criteria as described above, the core community was identified for each

909 continent. That is, a core OTU for a specific continent should be one that was from the top 0.1%

910 OTUs of that continent; a core OTU also had to be detected in more than 80% of the samples and

911 dominant for more 50% of the samples of that continent.

912

913 **Comparison of bacterial community composition of WWTPs to natural habitats and source**

914 **tracking**

915
916 We downloaded the OTU table of 16S rRNA gene amplicon studies from the EMP
917 (ftp://ftp.microbio.me/emp/release1/otu_tables/closed_ref_greenegenes/emp_cr_gg_13_8.subset_5k.biom)⁵. This table was generated using closed reference against Greengenes 13.8 and
918 contained 5,000 global samples from multiple habitats. To compare community compositions at
919 the OTU level, our activated sludge OTUs were repicked using closed reference against
920 Greengenes 13.8, which picked 68.1% of the sequences. This OTU table was then merged with
921 the EMP OTU table. To give relatively equal representation of samples across environments, we
922 further collapsed our activated sludge samples at the plant level by summing the abundance of
923 each OTU across samples of the same plant, resulting in 269 activated sludge samples. Our
924 activated sludge samples and the EMP samples from freshwater (including that from freshwater
925 and freshwater biofilm), ocean (including that from sea water and biofilm), animal feces, human
926 feces, soil and air were selected from the merged OTU table. We then subsampled to 10,000
927 sequences per sample. To compare microbial community compositions across habitats, the
928 Nonmetric Multidimensional Scaling (NMS) analysis was performed using the Bray-Curtis
929 dissimilarity matrix.

930
931
932 The proportion of each activated sludge microbiota attributable to freshwater, soil, ocean, animal
933 and human feces, and air at the genus level were estimated using SourceTracker³⁹, which was
934 run through QIIME with default settings using activated sludge microbiota as the sink and those
935 in other habitats as sources. Genera detected in less than 1% of the samples were filtered out
936 before source-tracking modeling.

937

938 **Diversity analyses: α - and β -diversity and correlation with environment**

939

940 Richness and Faith's index were used to measure taxonomic and phylogenetic α -diversity,
941 respectively, and they were computed using the *Picante* R package⁸⁵. Other taxonomic α -
942 diversity indices, including Shannon index, Simpson index and Pielou's evenness, were
943 calculated using the *vegan* R package⁸⁶.

944

945 Bray-Curtis (abundance-based) and Sorensen (incidence-based) distances were calculated to
946 represent the taxonomic β -diversity using the *vegan* R package⁸⁶. Canberra's distance was also
947 calculated to give more weight to rare taxa, using the *vegan* R package⁸⁶. The weighted
948 (abundance-based) and unweighted UniFrac (incidence-based) distance⁸⁷ were calculated to
949 represent the phylogenetic β -diversity using the *GUniFrac* R package⁸⁸. For each environmental
950 variable, we performed a partial Mantel test to examine the correlation between environmental
951 variable and microbial community composition independent of geographical location (999
952 permutations) using the *vegan* R package⁸⁶.

953

954 PERMANOVA was applied to assess the difference of community composition among
955 continents, climate types, and activated sludge process types using the *vegan* R package⁸⁶. In
956 PERMANOVA, climate types were defined at main climate group level, which includes 5
957 groups: A (tropical), B (arid), C (temperate), D (cold), and E (polar)⁵⁸. The activated sludge
958 process types were classified into 9 general groups: complete mix, conventional plug flow,
959 sequential batch reactors (SBR), anaerobic/anoxic/oxic (A²O), anoxic/oxic (AO), oxidation ditch,
960 contact stabilization, pure oxygen and extended aeration.

961

962 **Distance Decay relationships**

963

964 The rate of the distance-decay relationship (DDR) was calculated as the slope of a linear least
965 squares regression on the relationship between ln-transformed geographic distance versus ln-
966 transformed bacterial community composition similarity. We used matrix permutation tests to
967 examine the statistical significance of the distance-decay slope³⁷. The samples were permuted
968 999 times, and the observed slope was compared with the distribution of values in the permuted
969 datasets. We also tested whether the slopes of the distance-decay curve at the three spatial scales
970 (0 to 100 km; 100 to 5,000 km; and 5,000 to 25,000 km) were significantly different from the
971 slope of the overall distance-decay curve, using matrix permutations to compare the observed
972 difference between slopes within the three spatial scales with the overall distance-decay slope to
973 that over 999 permutations.

974

975 **Estimating stochasticity of community assembly**

976

977 We assessed community-assembly stochasticity with a null-model-based index. The
978 Stochasticity ratio was described previously^{41,89}. Since null-model algorithms usually require a
979 high number of replicates, we selected 71 cities, each of which had more than 9 samples; we
980 randomly drew 9 samples from each city to make sampling even. We calculated stochasticity
981 ratio using taxonomic and phylogenetic metrics. Whether using the Bray-Curtis (abundance-
982 weighted) or Sorensen (unweighted) model, the stochasticity ratio was calculated based on
983 typical null-model algorithms for taxonomic metrics^{90,91}. When using weighted and unweighted

984 Unifrac, the stochasticity ratio was calculated based on typical null-model algorithms for
985 phylogenetic metrics^{91,92}. Samples within each city were considered sharing the same regional
986 species pool in null model algorithms.

987

988 **Partitioning the environment and distance effect**

989

990 To give a quantification of relative contribution of the environment effect versus the distance
991 effect on β -diversity, we performed a variation partition analysis (VPA) based on multiple
992 regression on matrices (MRM). We used a modified MRM approach as described previously³⁷.
993 Briefly, we first selected a non-redundant environmental variable set. The final set included
994 temperature, precipitation, design capacity, SRT, dissolved oxygen, pH, and influent BOD. The
995 highest correlation was between design capacity and SRT (Pearson' $r = -0.25$), and it indicated a
996 low level of collinearity among these variables. MRM was performed in different spatial scales.
997 Geographic distance and microbial community distance were ln-transformed. A Euclidean
998 distance matrix was calculated for each environmental variable. To reduce the effect of spurious
999 relationships between variables, we first ran the MRM test with all the variables in the non-
1000 redundant environmental variable set, removed the non-significant variables from this initial
1001 MRM test, and then reran the test³⁷. The significance of the partial regression was tested by
1002 matrix permutation for 999 times⁹³. In VPA, the R^2 of the selected environmental variables as
1003 independent matrices (R^2_E), geographical distance as independent matrix (R^2_G), and all matrices
1004 (R^2_T) were used to compute the four components of variations as described elsewhere⁹⁴: (i)
1005 pure environmental variation = $R^2_T - R^2_G$; (ii) pure geographical distance = $R^2_T - R^2_E$; (iii)

1006 spatially structured environmental variation = $R^2_G + R^2_E - R^2_T$; and (iv) unexplained variation
1007 = $1 - R^2_T$.

1008

1009 **Structural equation model (SEM)**

1010

1011 SEM was used to explore the direct and indirect relationships among environmental variables,
1012 bacterial communities, and activated sludge function. The community composition was
1013 represented by the first principal component (PC1) of Principal coordinate analysis (PCoA)
1014 based on Bray-Curtis distance. We first considered a full model that included all reasonable
1015 pathways, and then we sequentially eliminated non-significant pathways until we attained the
1016 final model whose pathways all were significant. To capture the quadratic correlation of SRT to
1017 diversity and BOD removal, we constructed a composite variable⁹⁴ of ‘SRT effect’ as a linear
1018 combination of SRT and the square of SRT (SRT.SQ). We used a χ^2 test and the root mean
1019 square error of approximation to evaluate the fit of model. The SEM-related analysis was
1020 performed using the *lavaan* R package⁹⁵.

1021

1022 **Random Forest models**

1023

1024 We applied a machine-learning model, random forest, to examine the strengths of the
1025 associations between environmental variable and compositional data, using the *randomForest* R
1026 package⁹⁶. We used OTUs as predictors and environmental variable as response data. To
1027 correct the potential spatial autocorrelation, we used OTU data at the plant level, by averaging
1028 the relative abundance of each OTU across samples of the same plant. OTUs which were

1029 detected in at least 20% of all the plants and in all continents were used for modelling. We
1030 allowed a baseline model to learn using the full data-set for training, and subsequently, we
1031 trained new random forests for each plant with customized training sets that excluded plants
1032 within a defined radius of the target plant. The size of this radius ranged from 0 to 5000 km. To
1033 delineate the model prediction strength, the cross-validated R^2 was calculated as $1 - \frac{\sum(y_i - \hat{y}_i)^2}{\sum(y_i - \bar{y}_i)^2}$
1034 where y_i is the value of the parameter for sample i , \hat{y}_i is the prediction for that same sample
1035 (obtained by held-out cross-validation), and \bar{y}_i is the overall mean (the summation runs over all
1036 the samples).

1037

1038 **Data availability**

1039

1040 The sample metadata are available in Supplementary Table 1. Sequences are available from the
1041 NCBI Sequence Read Archive with accession number PRJNA509305. OTU tables and
1042 representative sequences of the OTUs are available on the GWMC web site
1043 (<http://gwmc.ou.edu/data-disclose.html>).

1044

1045 **Code availability**

1046

1047 R codes on the statistical analyses are available at [https://github.com/Linwei-Wu/Global-](https://github.com/Linwei-Wu/Global-bacterial-diversity-in-WWTPs)
1048 [bacterial-diversity-in-WWTPs](https://github.com/Linwei-Wu/Global-bacterial-diversity-in-WWTPs).

1049

1050 **References of Methods**

1051

- 1052 51 Zhou, J. *et al.* Random Sampling Process Leads to Overestimation of β -Diversity of
1053 Microbial Communities. *mBio* **4** (2013).
1054 52 Zhou, J. *et al.* Reproducibility and quantitation of amplicon sequencing-based detection.
1055 *ISME J* **5**, 1303-1313 (2011).

1056 53 Sinha, R. *et al.* Assessment of variation in microbial community amplicon sequencing by
1057 the Microbiome Quality Control (MBQC) project consortium. *Nat Biotechnol* **35**, 1077
1058 (2017).

1059 54 Ju, F. & Zhang, T. Bacterial assembly and temporal dynamics in activated sludge of a
1060 full-scale municipal wastewater treatment plant. *ISME J* **9**, 683-695 (2015).

1061 55 Xia, Yu. *Diversity and temporal assembly patterns of microbial communities in*
1062 *municipal wastewater treatment systems*. PhD thesis, Univ. Tsinghua, Beijing, China
1063 (2016).

1064 56 Buttigieg, P. L., Morrison, N., Smith, B., Mungall, C. J. & Lewis, S. E. The environment
1065 ontology: contextualising biological and biomedical entities. *J Biomed Semantics* **4**, 43
1066 (2013).

1067 57 Yilmaz, P. *et al.* Minimum information about a marker gene sequence (MIMARKS) and
1068 minimum information about any (x) sequence (MIxS) specifications. *Nat Biotechnol* **29**,
1069 415-420 (2011).

1070 58 Peel, M. C., Finlayson, B. L. & McMahon, T. A. Updated world map of the Köppen-
1071 Geiger climate classification. *Hydrol Earth Syst Sci Discuss* **4**, 439-473 (2007).

1072 59 Alan Berube, J. L. T., Tao Ran, Joseph Parilla. *Global Metro Monitor*,
1073 <<https://www.brookings.edu/research/global-metro-monitor/>> (2015).

1074 60 Wu, L. *et al.* Phasing amplicon sequencing on Illumina Miseq for robust environmental
1075 microbial community analysis. *BMC Microbiol* **15**, 125 (2015).

1076 61 Caporaso, J. G. *et al.* Global patterns of 16S rRNA diversity at a depth of millions of
1077 sequences per sample. *Proc Natl Acad Sci USA* **108**, 4516-4522 (2011).

1078 62 Klindworth, A. *et al.* Evaluation of general 16S ribosomal RNA gene PCR primers for
1079 classical and next-generation sequencing-based diversity studies. *Nucleic Acids Res* **41**,
1080 e1-e1 (2013).

1081 63 Wen, C. *et al.* Evaluation of the reproducibility of amplicon sequencing with Illumina
1082 MiSeq platform. *PLoS One* **12**, e0176716 (2017).

1083 64 Edgar, R. C. UPARSE: highly accurate OTU sequences from microbial amplicon reads.
1084 *Nat Methods* **10**, 996-998 (2013).

1085 65 McLaren, M. R. & Callahan, B. J. In Nature, There Is Only Diversity. *mBio* **9**, e02149-
1086 02117 (2018).

1087 66 Sievers, F. *et al.* Fast, scalable generation of high-quality protein multiple sequence
1088 alignments using Clustal Omega. *Mol Syst Biol* **7**, doi:10.1038/msb.2011.75 (2011).

1089 67 Price, M. N., Dehal, P. S. & Arkin, A. P. FastTree 2 – Approximately maximum-
1090 likelihood trees for large alignments. *PLoS One* **5**, e9490,
1091 doi:10.1371/journal.pone.0009490 (2010).

1092 68 Wang, Q., Garrity, G. M., Tiedje, J. M. & Cole, J. R. Naive Bayesian classifier for rapid
1093 assignment of rRNA sequences into the new bacterial taxonomy. *Appl Environ Microbiol*
1094 **73**, 5261-5267 (2007).

1095 69 McIlroy, S. J. *et al.* MiDAS 2.0: an ecosystem-specific taxonomy and online database for
1096 the organisms of wastewater treatment systems expanded for anaerobic digester groups.
1097 *Database* **2017** (2017).

1098 70 Delgado-Baquerizo, M. *et al.* A global atlas of the dominant bacteria found in soil.
1099 *Science* **359**, 320-325 (2018).

1100 71 Stoddard, S. F., Smith, B. J., Hein, R., Roller, B. R. & Schmidt, T. M. rrnDB: improved
1101 tools for interpreting rRNA gene abundance in bacteria and archaea and a new foundation
1102 for future development. *Nucleic Acids Res*, gku1201 (2014).

1103 72 McDonald, D. *et al.* An improved Greengenes taxonomy with explicit ranks for
1104 ecological and evolutionary analyses of bacteria and archaea. *ISME J* **6**, 610 (2012).

1105 73 Quast, C. *et al.* The SILVA ribosomal RNA gene database project: improved data
1106 processing and web-based tools. *Nucleic Acids Res* **41**, D590-D596 (2012).

1107 74 Edgar, R. C. Search and clustering orders of magnitude faster than BLAST.
1108 *Bioinformatics* **26**, 2460-2461 (2010).

1109 75 AQUASTAT. *FAO global information system on water and agriculture. Wastewater*
1110 *section.*, <<http://www.fao.org/nr/water/aquastat/wastewater/index.stm>> (2014).

1111 76 Sato, T., Qadir, M., Yamamoto, S., Endo, T. & Zahoor, A. Global, regional, and country
1112 level need for data on wastewater generation, treatment, and use. *Agri Water Manag* **130**,
1113 1-13 (2013).

1114 77 Foladori, P., Bruni, L., Tamburini, S. & Ziglio, G. Direct quantification of bacterial
1115 biomass in influent, effluent and activated sludge of wastewater treatment plants by using
1116 flow cytometry. *Water Res* **44**, 3807-3818 (2010).

1117 78 Agency, U. S. E. P. *The Sources and Solutions: Wastewater*,
1118 <<https://www.epa.gov/nutrientpollution/sources-and-solutions-wastewater>> (2018).

1119 79 Chan, W. *Wastewater: Good To The Last Drop*,
1120 <<http://chinawaterrisk.org/resources/analysis-reviews/wastewater-good-to-the-last-drop/>>
1121 (2017).

1122 80 Hanski, I. Dynamics of regional distribution: the core and satellite species hypothesis.
1123 *Oikos*, 210-221 (1982).

1124 81 Galand, P. E., Casamayor, E. O., Kirchman, D. L. & Lovejoy, C. Ecology of the rare
1125 microbial biosphere of the Arctic Ocean. *Proc Natl Acad Sci USA* **106**, 22427-22432,
1126 doi:10.1073/pnas.0908284106 (2009).

1127 82 Székely, A. J. & Langenheder, S. The importance of species sorting differs between
1128 habitat generalists and specialists in bacterial communities. *FEMS Microbiol Ecol* **87**,
1129 102-112 (2014).

1130 83 Cheng, J. *et al.* Discordant temporal development of bacterial phyla and the emergence of
1131 core in the fecal microbiota of young children. *ISME J* **10**, 1002 (2016).

1132 84 Ju, F. & Zhang, T. Bacterial assembly and temporal dynamics in activated sludge of a
1133 full-scale municipal wastewater treatment plant. *ISME J*, doi:10.1038/ismej.2014.162
1134 (2014).

1135 85 Kembel, S. W. *et al.* Picante: R tools for integrating phylogenies and ecology.
1136 *Bioinformatics* **26**, 1463-1464 (2010).

1137 86 Oksanen, J. *et al.* Package ‘vegan’. *Community ecology package, version 2* (2013).

1138 87 Lozupone, C. & Knight, R. UniFrac: a new phylogenetic method for comparing microbial
1139 communities. *Appl Environ Microbiol* **71**, 8228-8235 (2005).

1140 88 Chen, J. GUniFrac: generalized UniFrac distances. *R package version 1*, 2012 (2012).

1141 89 Guo, X. *et al.* Climate warming leads to divergent succession of grassland microbial
1142 communities. *Nat Clim Change* **8**, 813 (2018).

1143 90 Chase, J. M., Kraft, N. J., Smith, K. G., Vellend, M. & Inouye, B. D. Using null models
1144 to disentangle variation in community dissimilarity from variation in α -diversity.
1145 *Ecosphere* **2**, art24 (2011).

1146 91 Stegen, J. C. *et al.* Quantifying community assembly processes and identifying features
1147 that impose them. *ISME J* **7**, 2069-2079 (2013).

1148 92 Kembel, S. W. Disentangling niche and neutral influences on community assembly:
1149 assessing the performance of community phylogenetic structure tests. *Ecol Lett* **12**, 949-
1150 960 (2009).

1151 93 Legendre, P., Lapointe, F. J. & Casgrain, P. Modeling brain evolution from behavior: a
1152 permutational regression approach. *Evolution* **48**, 1487-1499 (1994).

1153 94 Grace, J. B. & Bollen, K. A. Representing general theoretical concepts in structural
1154 equation models: the role of composite variables. *Environ Ecol Stat* **15**, 191-213 (2008).

1155 95 Rosseel, Y. Lavaan: An R package for structural equation modeling and more. Version
1156 0.5-12 (BETA). *Journal of statistical software* **48**, 1-36 (2012).

1157 96 Liaw, A. & Wiener, M. Classification and regression by randomForest. *R news* **2**, 18-22
1158 (2002).

1159 97 Faith, D. P. Conservation evaluation and phylogenetic diversity. *Biol Cons* **61**, 1-10
1160 (1992).

1161

1162 **Figures legends**

1163

1164 **Fig. 1. The Global Water Microbiome Consortium captures microbial diversity of globally**
1165 **distributed wastewater treatment plants (WWTPs).** (a) Geographical distribution of 269
1166 WWTPs where activated sludge samples and environmental data were collected. (b) Predicting
1167 species abundance distribution (SAD) of activated sludge bacterial communities. The grey line
1168 represents a SAD that was randomly chosen from our data. Each model was fit to the observed
1169 SAD (see Methods). Supplementary Fig.1a shows the variations of the SADs explained by each
1170 model across all 1186 activated sludge communities, indicating the best performance of the
1171 Poisson lognormal model. (c) Estimation of activated sludge microbial richness of WWTPs.
1172 Microbial species are defined as OTUs at 97% sequence similarity threshold. The microbial
1173 richness (S)-abundance (N) scaling relationship (dashed grey line with pink hull as 95%
1174 prediction interval), and the grey dots representing richness estimates from other systems were
1175 derived from Locey and Lennon¹⁹. Richness was predicted from the lognormal model using N_T
1176 estimated from published data, and N_{max} inferred from our sequencing data (filled circle) or N_{max}
1177 predicted from the dominance-scaling law¹⁹ (hollow circles). ‘WWTP’ indicates one WWTP, as
1178 do ‘Human gut’ and ‘Cow rumen’. (d) Latitudinal distribution of activated sludge bacterial
1179 diversity, plotting OTU richness against the absolute latitude of sampling locations shows the
1180 peak of richness at intermediate latitude (n = 1,186 biologically independent samples). The line
1181 shows the second order polynomial fit based on ordinary least squares regression. $P < 2 \times 10^{-16}$
1182 (two-sided) for both regression coefficients. The color gradient denotes the annual mean air
1183 temperature. Shapes of symbols denotes whether a sample originated from Northern (circle) or
1184 Southern Hemisphere (square).

1185

1186 **Fig. 2. Abundance, composition and functional importance of the global core operational**
1187 **taxonomic units (OTUs) in activated sludge.** (a) Percentage and relative abundance of the
1188 global core OTUs versus the remaining microbial OTUs. In total, 0.05% (28 out of 61,448
1189 OTUs) were identified as abundant and ubiquitous across wastewater treatment plants at global
1190 scale, which accounted for 12.4% of the 16S rRNA gene sequences in an activated sludge
1191 sample on average. (b) The taxonomic composition of the global core OTUs on phylum and
1192 class level. (c) Activated sludge functions were calculated as the removal rate of organic carbon
1193 (biochemical oxygen demand (BOD) removal, chemical oxygen demand (COD) removal),
1194 nutrients (total nitrogen (TN) and total phosphorus (TP) removal) and ammonia nitrogen (NH₄-N
1195 removal) (g chemical per g MLSS per day, where MLSS is mixed liquor suspended solids
1196 relating to microbial biomass). The color gradient on the right indicates Spearman’s rank
1197 correlation coefficients, with more positive values (dark blue) indicating stronger positive
1198 correlations, and more negative values (dark red) indicating stronger negative correlations. The
1199 asterisks denote the significance levels (two-sided) of the Spearman’s rank correlation
1200 coefficients (n = 1,186 biologically independent samples): *** $P < 0.001$, ** $P < 0.01$, * $P <$
1201 0.05 . In the correlation analysis, all OTUs detected in at least 20% of samples were included,
1202 and P values were adjusted for multiple testing using the Benjamini and Hochberg false

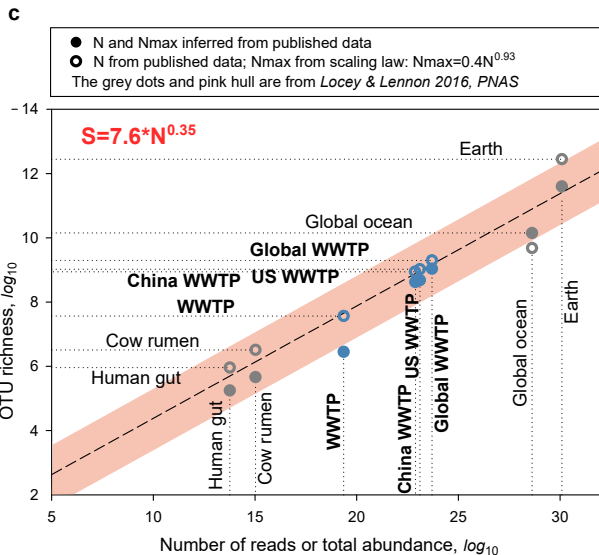
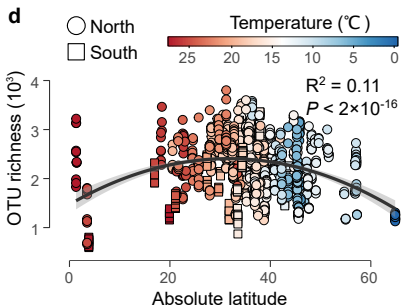
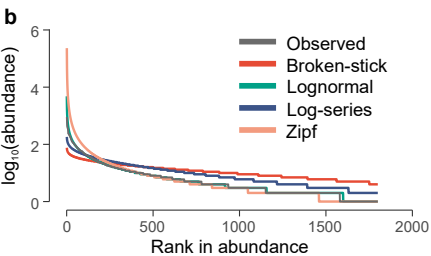
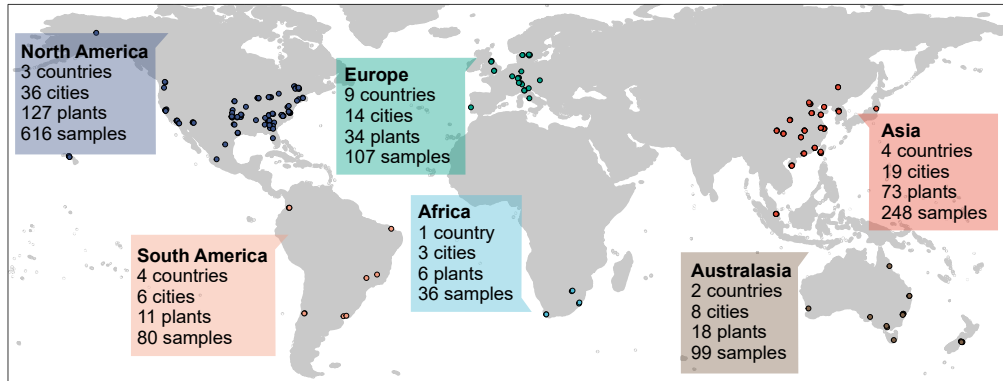
1203 discovery rate (FDR) controlling procedure (n = 14,235 pairwise cases). Only global core OTUs
1204 were shown, with their mean relative abundance indicated on the left of the heatmap.

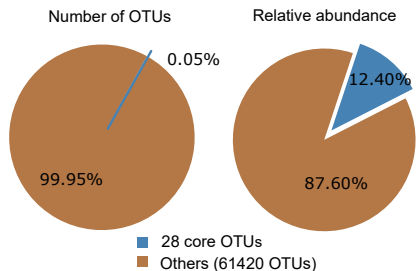
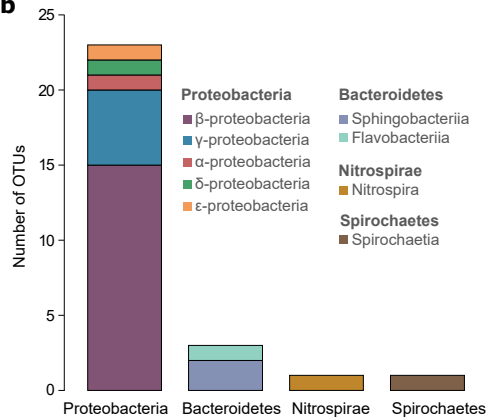
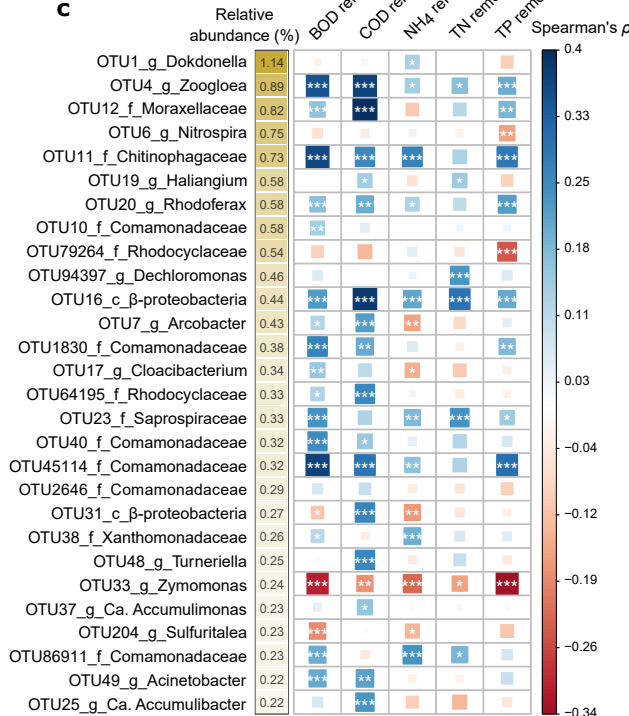
1205
1206 **Fig. 3. Comparing bacterial community compositions across continents and with other**
1207 **habitats.** (a) Nonmetric Multidimensional Scaling analysis (NMDS) showing that activated
1208 sludge of WWTPs harbored a unique microbiome as compared with other habitats. For
1209 comparison, we merged our OTU table (n = 269 WWTPs) with that released by EMP⁵, which
1210 contained thousands of bacterial communities from various habitats such as soil (n = 338
1211 samples), ocean (n = 969 samples), freshwater (n = 447 samples), air (n = 81 samples), human
1212 feces (n = 99 samples) and animal feces (n = 622 samples), but not activated sludge from
1213 WWTPs (see Methods for details). Bray-Curtis distance was calculated to represent the
1214 dissimilarity in bacterial community compositions. (b) Percentage of activated sludge bacterial
1215 genera attributable to air, animal and human feces, freshwater, ocean and soil, as determined by
1216 SourceTracker. In the boxplots, hinges show the 25, 50 and 75 percentiles. The upper whisker
1217 extends to the largest value no further than 1.5 * IQR from the upper hinge, where IQR is the
1218 inter-quartile range between the 25% and 75% quartiles; The lower whisker extends to the
1219 smallest value at most 1.5 * IQR from the lower hinge. Sample size: n = 6, 73, 18, 34, 127 and
1220 11 WWTPs for Africa, Asia, Australasia, Europe, North America and South America,
1221 respectively.

1222
1223
1224 **Fig. 4. Spatial turnover of the activated sludge bacterial communities.** (a) Distance-decay
1225 relationships (DDRs) based on Bray-Curtis similarity. Black line denotes the least-squares linear
1226 regression across all spatial scales (n= 702,705 pairwise distances). Colored lines denote
1227 separate regressions: within cities (n= 9,753 pairwise distances), within continents (n= 220,136
1228 pairwise distances), and intercontinental (n= 472,816 pairwise distances). *P* values (one-sided)
1229 for regression slopes were determined by matrix permutation tests. (b) Ecological stochasticity in
1230 bacterial community assembly estimated by stochasticity ratio, which is calculated for each pair
1231 of samples (n= 71 cities) based on taxonomic diversity (Taxo., Bray-Curtis/Sorensen) and
1232 phylogenetic diversity (Phyl., Unifrac) weighted with abundance (Wt) or not (Uw). Boxes and
1233 whiskers indicate quartiles and triangles indicate mean values. (c) Variance partition analysis
1234 showing relative contributions of geographic distance (Geo) and environmental variables (ENV)
1235 to the community variations based on Bray-Curtis distance.

1236
1237 **Fig. 5. Environmental drivers of the activated sludge community composition.** (a) A
1238 structural equation model (SEM) shows relationships among environmental variables,
1239 community composition, and WWTP functioning. The composite variable of ‘SRT effect’ was
1240 constructed as a linear combination of solids retention time (SRT) and the square of SRT
1241 (SRT.SQ). F/M is the food to microorganisms ratio. The community composition is represented
1242 by the first principal component score (PC1) from the Bray-Curtis distance-based principal
1243 coordinate analysis. Blue and red arrows represent significant (*P* < 0.05) positive and negative
1244 pathways, respectively. Numbers near the pathway arrow indicate the standard path coefficients

1245 (β). Arrow width is proportional to the strength of the relationship. R^2 represents the proportion
1246 of variance explained for every dependent variable. Model $\chi^2 = 13.92$, $df = 12$, $P = 0.31$, $n =$
1247 1,186 biologically independent samples; root mean square error of approximation (RMSEA) =
1248 0.012 with probability of a close fit = 1.00. **(b)** The average rRNA gene copy number of the
1249 community increased with the influent biochemical oxygen demand (BOD)/(1+recycle ratio)
1250 which approximates the influent BOD level of aerobic tank ($n = 641$ biologically independent
1251 samples). The P value (two-sided) denotes the significance of the slope of ordinary least squares
1252 regression. **(c)** The strength of association between taxonomic composition and temperature was
1253 tested by random forest ($n = 269$ WWTPs). The red diagonal shows the theoretical curve for
1254 perfect predictions. The inset shows a model trained on data from North and South America
1255 samples to predict the temperature in Asian samples ($n = 73$ WWTPs).
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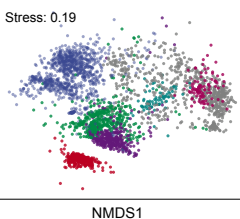


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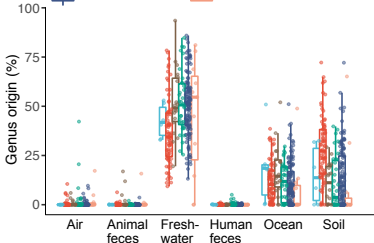
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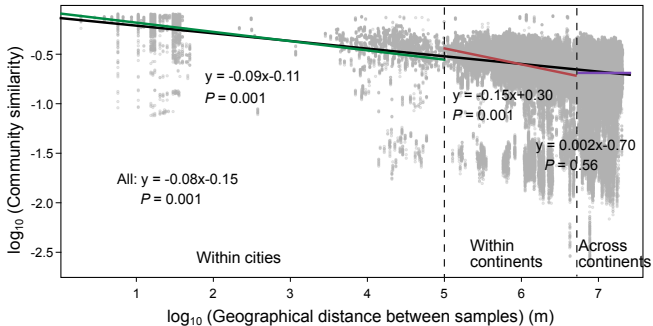
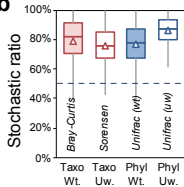
- WWTP ● Air ● Animal feces ● Soil
- Freshwater ● Human feces ● Ocean

NMDS2

**b**

- ▢ Africa ▢ Asia ▢ Australasia ▢ Europe
- ▢ North America ▢ South America



a**b****c**