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Targeted Rapid Synthesis of Redox-Active Dodecaborane Clusters

A thesis submitted in partial satisfaction
of the requirements for the degree Master of Science
in Chemistry

by

Alex Ian Wixtrom

2016

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ABSTRACT OF THE THESIS

Targeted Rapid Synthesis of Redox-Active Dodecaborane Clusters

by

Alex Ian Wixtrom

Master of Science in Chemistry

University of California, Los Angeles, 2016

Professor Alexander Michael Spokoyny, Chair

We have developed a fast, efficient route to obtain targeted perfunctionalized ether-linked alkyl and benzyl derivatives of the *closo*-[B₁₂(OH)₁₂]²⁻ icosahedral dodecaborate cluster via microwave-assisted synthesis. These icosahedral boron clusters exhibit unique properties including three-dimensional delocalization of the cage-bonding electrons, tunable photophysical properties, and a high degree of stability in air in both solid and solution states. Several B₁₂(OR)₁₂ clusters have been prepared with reaction times ranging from hours to several minutes using microwave-assisted synthesis. This represents a significant improvement upon previously reported protocols due to a dramatic reduction in required reaction times and the elimination of the need for inert atmosphere conditions. The robust nature of the new microwave-based method has been further demonstrated through the targeted synthesis of several new derivatives, which feature desired properties such as redox potentials up to 0.6 V more positive than previously known B₁₂(OR)₁₂ cluster compounds or substituents that allow further facile functionalization *via* thiol “click” S_NAr. We further demonstrate the ability to synthesize hybrid, vertex-differentiated cluster species B₁₂(OR)₁₁(OR')

via a single-step one-pot reaction that was formerly accessible only via multi-step reaction sequence.

The thesis of Alex Ian Wixtrom is approved.

Richard B. Kaner

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Dante A. Simonetti

Alexander Michael Spokoyny, Committee Chair

University of California, Los Angeles

2016

To my parents, for their encouragement and support,
and for always inspiring me to work hard in pursuit of my goals.

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Chapter 1: Introduction to Icosahedral Boron Cluster Chemistry

In 1912, the first borane compounds were discovered by Alfred Stock.¹⁻³ For decades these boron hydride compounds were considered exotic, with little practical value outside of academic curiosity, and their inherent pyrophoric properties made synthesis and characterization difficult. Borane chemistry was revitalized in the 1940's when certain borane compounds (pentaborane B₅H₉ and decaborane B₁₀H₁₄) were thought to be superior alternatives to hydrocarbons for rocket fuels.^{2,3} This notion was based on the fact that the energy contained in the B-H bond was greater than the C-H bond in traditional hydrocarbon fuels. Research efforts to produce high-energy fuels ultimately failed due to the discovery that borates formed from borane combustion clog jet engines, thus eliminating any potential efficiency gains from the higher energy,^{3,4} yet the interest in development of these fuels had successfully sparked the “renaissance era” of boron chemistry. A key discovery that elucidated a more accurate understanding of borane chemistry was Lipscomb's concept of three-center two-electron bonding.⁵ In his 1954 study, Lipscomb predicted the existence of an icosahedral boron hydride B₁₂H₁₂, which Longuet-Higgins and Roberts posited would only be stable as a dianion in a theoretical molecular orbital-based approach in 1955.^{5,6} In 1959 the possible formation of the predicted [B₁₂H₁₂]²⁻ was observed by Shapiro and Williams⁷, and a year later in 1960 the triethylammonium salt of [B₁₂H₁₂]²⁻ was isolated in a relatively low yield by Hawthorne and co-workers.⁸

The “renaissance era” in the field of boron cluster chemistry was led by the pioneering work of Hawthorne, Knoth, and Muetterties over the next few decades.^{2,9-22} Their investigations resulted in significantly improved synthetic methods to produce [B₁₂H₁₂]²⁻ on a much larger scale in high yields (>90%)² and the discovery that boron clusters can undergo facile functionalization chemistry in a similar fashion to some classical organic molecules such as benzene, unlocking

entirely new avenues of functionalized boron cluster development.^{3,23,24} As mentioned above, many boron hydride clusters were considered exotic and highly unstable species for many decades, undergoing rapid degradation if exposed to heat, acids, or bases. Even the widespread attempts to produce viable rocket fuels from these boranes was impeded by the unavoidable combustion products negatively impacting the jet engine performance.^{3,4} However, now that $[\text{B}_{12}\text{H}_{12}]^{2-}$ salts had been synthesized in greater yields, full characterization provided evidence of the surprising stability of the icosahedral cluster. $[\text{B}_{12}\text{H}_{12}]^{2-}$ salts were heated to 95 °C in strongly acidic (3 N HCl) and basic (3 N NaOH) media with little to no observed degradation, and heating the cluster to 810 °C under vacuum also resulted in no observable decomposition.¹⁶

The remarkable thermal stability exhibited by $[\text{B}_{12}\text{H}_{12}]^{2-}$ along with exceptional stability in acids and bases and the potential for functionalization of the cluster were very exciting discoveries, inspiring many groups to investigate possible novel boron cluster derivatives. The first persubstitution of the cluster where all 12 vertices of the icosahedron were functionalized was demonstrated by Knoth and co-workers. Knoth and his group were able to persubstitute $[\text{B}_{12}\text{H}_{12}]^{2-}$ with several halogens, successfully isolating $[\text{B}_{12}\text{F}_{12}]^{2-}$, $[\text{B}_{12}\text{Cl}_{12}]^{2-}$, $[\text{B}_{12}\text{Br}_{12}]^{2-}$ and $[\text{B}_{12}\text{I}_{12}]^{2-}$ derivatives.^{20,25} More recent studies have expanded upon these early persubstitution examples to produce additional derivatives of $[\text{B}_{12}\text{H}_{12}]^{2-}$ with improved synthetic methods and non-halogen functional groups, including $[\text{B}_{12}\text{Me}_{12}]^{2-}$ and $[\text{B}_{12}(\text{OH})_{12}]^{2-}$.^{26–30} The latter of these species, *closo*- $[\text{B}_{12}(\text{OH})_{12}]^{2-}$, is a particularly interesting derivative due to the possibility of further functionalization of the cluster *via* formation of ether, ester, carbonate, and carbamate linkages.^{29,31–33}

The exceptional stability of the parent dodecaborate cluster $[\text{B}_{12}\text{H}_{12}]^{2-}$ is one attractive feature of these boron cluster species, but another unique property is the three-dimensional

delocalization of two cage-bonding electrons across all of the boron atoms contained within. While Longuet-Higgins and Roberts concluded that the parent $[\text{B}_{12}\text{H}_{12}]^{2-}$ could only exist as a dianion⁶, the successful formation of various derivatives led others to investigate the possibility of oxidizing the cluster to produce radical 1- or neutral clusters. Controlled oxidation of the $[\text{B}_{12}\text{H}_{12}]^{2-}$ anion lead to irreversible degradation of the cluster due to the formation of a B–B linked dimer,²¹ though several of the synthesized perfunctionalized derivatives were observed to undergo reversible redox behavior.³⁴ Rupich and co-workers reported the single electron oxidation of $[\text{B}_{12}\text{X}_{12}]^{2-}$ to form a stable radical $[\text{B}_{12}\text{X}_{12}]^{1-}$,³⁵ and a recent study by Weber and co-workers confirmed the identity of the oxidized radical $[\text{B}_{12}\text{Cl}_{12}]^{1-}$ species.³⁶

The possible redox state of perfunctionalized clusters were further investigated by Hawthorne and co-workers *via* the characterization of $\text{B}_{12}(\text{OCH}_2\text{Ph})_{12}$ in three distinct redox states. These three redox states (*closo*- $[\text{B}_{12}(\text{OCH}_2\text{Ph})_{12}]^{2-}$, *hypocloso*- $[\text{B}_{12}(\text{OCH}_2\text{Ph})_{12}]^{1-}$, and *hypercloso*- $[\text{B}_{12}(\text{OCH}_2\text{Ph})_{12}]^0$) are accessible via two sequential and quasi-reversible one-electron oxidation reactions of the parent dianionic species.³⁷ Hawthorne's group later demonstrated the synthesis of several other benzyl and alkyl functionalized $\text{B}_{12}(\text{OR})_{12}$ clusters and characterized the redox behavior of these new derivatives, which exhibited similarly reversible behavior.³¹ The parent $[\text{B}_{12}(\text{OH})_{12}]^{2-}$ cluster these derivatives originated from was also shown to be capable of undergoing a one-electron oxidation by Schleid and co-workers, forming a stable radical $[\text{B}_{12}(\text{OH})_{12}]^{1-}$ species.³⁸ Perhaps the most intriguing feature of these cluster derivatives is that the redox potential of ether-linked $\text{B}_{12}(\text{OR})_{12}$ species can be rationally tuned as a function of the O-bound substituent, similar to metal-based redox-active inorganic complexes.^{31,34,39} This is a very interesting feature warranting further studies to probe the extent of this tunable nature, yet all reported synthetic routes to produce perfunctionalized $\text{B}_{12}(\text{OR})_{12}$ clusters to date require either extremely long reaction times (weeks) or highly specialized high-pressure equipment. All of these synthetic routes also

require strict inert atmospheric conditions, further adding to the difficulty of producing new derivatives.^{29,31,37} A new method to produce perfunctionalized $B_{12}(OR)_{12}$ clusters under less stringent inert atmosphere conditions in significantly shorter time would allow for much more rapid exploration of novel derivatives, allowing for targeted synthesis of cluster with specific properties for a plethora of applications.

Chapter 2: Rapid Synthesis of Redox-Active Dodecaborane B₁₂(OR)₁₂ Clusters Under Ambient Conditions

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We have developed a fast and efficient route to obtain perfunctionalized ether-linked alkyl and benzyl derivatives of the *closo*-[B₁₂(OH)₁₂]²⁻ icosahedral dodecaborate cluster via microwave-assisted synthesis. These icosahedral boron clusters exhibit three-dimensional delocalization of the cage-bonding electrons, tunable photophysical properties, and a high degree of stability in air in both solid and solution states. A series of *closo*-[B₁₂(OR)₁₂]²⁻, *hypocloso*-[B₁₂(OR)₁₂]¹⁻ and *hypercloso*-[B₁₂(OR)₁₂]⁰ clusters have been prepared with reaction times ranging from hours to several minutes. This method is superior to previously reported protocols since it dramatically decreases the reaction times required and eliminates the need for inert atmosphere conditions. The generality of the new microwave-based method has been further demonstrated through the synthesis of several new derivatives, which feature redox potentials up to 0.6 V more positive than previously known B₁₂(OR)₁₂ cluster compounds. We further show how this method can be applied to a one-pot synthesis of hybrid, vertex-differentiated species B₁₂(OR)₁₁(OR') that was formerly accessible only via multi-step reaction sequence.

Introduction

The existence of an icosahedral dodecaborate $[\text{B}_{12}\text{H}_{12}]$ cluster was first predicted by Lipscomb and co-workers in 1954.⁵ In a subsequent theoretical molecular orbital-based approach published in 1955⁶, Longuet-Higgins and Roberts predicted that such a cluster would only be stable as dianionic $[\text{B}_{12}\text{H}_{12}]^{2-}$. A 1959 study by Shapiro and Williams⁷ suggested the possible formation of a $[\text{B}_{12}\text{H}_{12}]^{2-}$ icosahedron, and in 1960 this cluster was first successfully isolated and characterized as a triethylammonium salt by Pitochelli and Hawthorne, albeit in a relatively low yield.⁸ Subsequent pioneering studies by Hawthorne, Knoth, Muetterties, and others initiated a new era in the field of boron cluster chemistry.^{2,9-22} Specifically, these groups have shown that $[\text{B}_{12}\text{H}_{12}]^{2-}$ can be prepared on a large scale in a high yield (>90%)² and undergo facile functionalization chemistry that parallels some properties of classical organic molecules (*e.g.* benzene).^{3,23,24} This was an exciting discovery, since previously many boron hydride clusters were perceived as highly unstable species prone to fast degradation by heat, acids, and bases. Conversely, $[\text{B}_{12}\text{H}_{12}]^{2-}$ salts were shown to be stable in acids and bases, and were thermally stable as high as 810 °C with no observable decomposition.¹⁶ Knoth and co-workers were the first to demonstrate the persubstitution of $[\text{B}_{12}\text{H}_{12}]^{2-}$, producing halogenated $[\text{B}_{12}\text{F}_{12}]^{2-}$, $[\text{B}_{12}\text{Cl}_{12}]^{2-}$, $[\text{B}_{12}\text{Br}_{12}]^{2-}$ and $[\text{B}_{12}\text{I}_{12}]^{2-}$ derivatives.^{20,25} In the past two decades, persubstitution of $[\text{B}_{12}\text{H}_{12}]^{2-}$ was improved with new synthetic methods and extended towards other functional groups including $[\text{B}_{12}\text{Me}_{12}]^{2-}$ and $[\text{B}_{12}(\text{OH})_{12}]^{2-}$.²⁶⁻³⁰ Among the perfunctionalized derivatives synthesized, *closo*- $[\text{B}_{12}(\text{OH})_{12}]^{2-}$ is particularly appealing, as it is capable of undergoing further functionalization by forming ether, ester, carbonate, and carbamate linkages.^{29,31-33}

While controlled oxidation of the parent $[\text{B}_{12}\text{H}_{12}]^{2-}$ anion leads to an irreversible cluster degradation forming a B–B linked dimer,²¹ several perfunctionalized variants have been

previously observed to undergo reversible redox behavior.³⁴ For example, Rupich reported that $[\text{B}_{12}\text{X}_{12}]^{2-}$ could undergo a single electron oxidation to form a stable radical $[\text{B}_{12}\text{X}_{12}]^{1-}$,³⁵ which was recently isolated and structurally confirmed as the oxidized radical $[\text{B}_{12}\text{Cl}_{12}]^{1-}$ species by Weber and co-workers.³⁶ Hawthorne and co-workers reported that the perfunctionalized $\text{B}_{12}(\text{OCH}_2\text{Ph})_{12}$ cluster can exist in three distinct redox states accessible via two sequential and quasi-reversible one-electron oxidation reactions of the parent dianionic *closo* species.³⁷ The same group later showed that other benzyl and alkyl functionalized $\text{B}_{12}(\text{OR})_{12}$ clusters can be synthesized, and all of these species exhibit similarly reversible redox behavior.³¹ Schleid and co-workers subsequently showed that the parent $[\text{B}_{12}(\text{OH})_{12}]^{2-}$ cluster can undergo a one-electron oxidation to form a stable radical $[\text{B}_{12}(\text{OH})_{12}]^{1-}$ species.³⁸ Interestingly, the redox potential of the ether-linked $\text{B}_{12}(\text{OR})_{12}$ species can be rationally tuned as a function of the O-bound substituent, reminiscent of many metal-based redox-active inorganic complexes.^{31,34,39} Unfortunately, all of the reported synthetic routes towards $\text{B}_{12}(\text{OR})_{12}$ clusters currently require either extremely long reaction times (weeks) or highly specialized high-pressure equipment. Furthermore, in all cases strict inert atmospheric conditions are also required for their synthesis.^{29,31,37}

Herein we report a rapid, scalable, and robust synthetic route to a wide range of perfunctionalized $\text{B}_{12}(\text{OR})_{12}$ cluster derivatives utilizing a bench-top microwave reactor. This technology has emerged over the past several decades and has been successfully employed in a large number of synthetic schemes⁴⁰ which necessitate the reaction heating above the boiling point of the solvent. Our work shows that the microwave-based method enables synthesis of perfunctionalized ether-linked boron clusters within minutes and does not require the use of inert atmosphere and rigorously dried solvents. We further show the synthetic utility of our method in the preparation of previously unknown $\text{B}_{12}(\text{OR})_{12}$ derivatives featuring highly oxidizing redox potentials as well as vertex-differentiated molecular architectures. Synthesized cluster species

were all isolated and characterized using solution-based NMR and IR spectroscopic tools and mass spectrometry. Electron delocalization of the radical state in these species was further evaluated by electron paramagnetic resonance (EPR) spectroscopy and elucidated by X-ray photoelectron spectroscopy (XPS).

Results and Discussion

The tetrabutylammonium (TBA) salt of *closo*-[B₁₂(OH)₁₂]²⁻ (TBA₂[**1**]) was chosen for use with our microwave synthesis due to its enhanced solubility in organic solvents compared to alkali metal salts of **1** (Figure 1).³¹ The synthesis of TBA₂[**1**] was adapted from previously reported protocols by Hawthorne and co-workers (see SI for details).^{31,41,42}

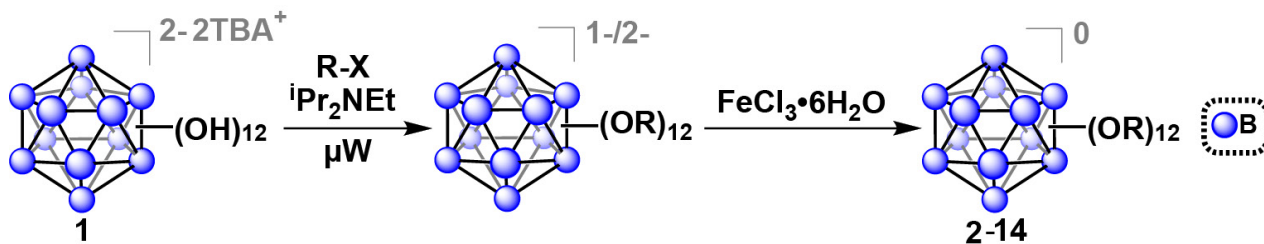


Figure 1. Synthetic route to produce functionalized ether-linked derivatives of the *hypercloso* boron clusters (**2-14**) via microwave-assisted synthesis. Precursor synthesis of [**1**]²⁻ was adapted from previously described methods (see SI for details).^{31,41,42}

Oxygen-free, anhydrous conditions (oven-dried glassware, dried and distilled solvents, nitrogen atmosphere) were initially employed for microwave-assisted syntheses of alkyl- and benzyl-functionalized *closo*-[B₁₂(OH)₁₂]²⁻ [**1**] ether-linked derivatives, due to previously described high-pressure and reflux-based methods necessitating stringent air and moisture free conditions. To our surprise, we discovered that microwave reactions utilizing previously reported conditions (benzyl bromide and TBA₂[**1**] in the presence of *N,N*-diisopropylethylamine (DIEA, Hünig's base)

in acetonitrile) are driven at a much higher rate, resulting in quantitative formation of a mixture of charged 1-/2- TBA salts of **2** within 15 minutes at 140 °C, as indicated by *in situ* ^{11}B NMR spectroscopy. Specifically, no parent ^{11}B NMR resonance at $\delta -18$ corresponding to the $[\text{B}_{12}(\text{OH})_{12}]^{2-}$ starting material is observed, and a singlet at $\delta -16$ can be seen instead. Concomitant presence of $[\mathbf{2}]^{1-}$ radical species in the product mixture can be deduced from the diagnostic pink color of the solution and its measured signature EPR signal (G-factor = 2.008121). Oxidation of the reaction mixture using $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ in 90/10 ethanol/acetonitrile followed by column chromatography on silica gel produces the pure neutral cluster $[\mathbf{2}]^0$ in 63% yield (Figure 2). Oxidation can be conveniently monitored by ^{11}B NMR, where the fully oxidized cluster $[\mathbf{2}]^0$ exhibits a downfield resonance shift at $\delta 41.8$. Overall, this represents a significant reduction in reaction duration from the originally reported 6 days and 4 hours required for reflux and high-pressure reactor methods, respectively, while retaining similar yield.³¹

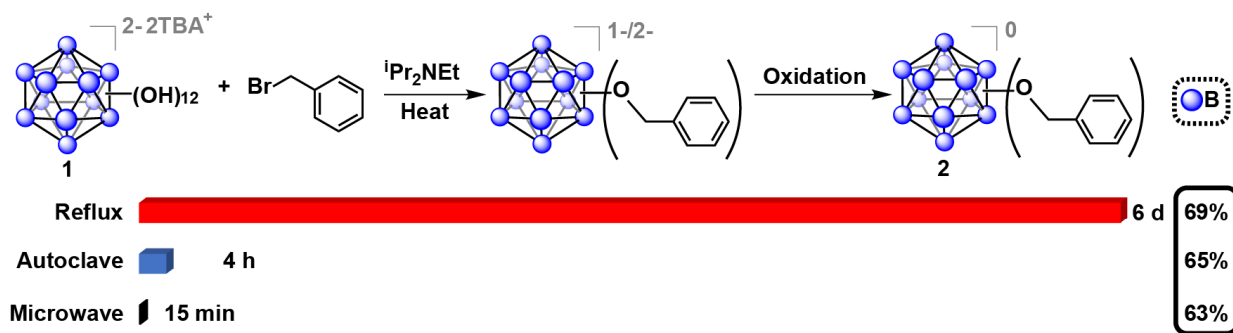


Figure 2. Synthesis of **2** from $\text{TBA}_2[\mathbf{1}]$ using microwave-based method along with two previously reported methods indicating dramatic reduction in reaction time without compromising the isolated yield of $[\mathbf{2}]^0$.³¹

Encouraged by these results, we then optimized the microwave reaction times with allyl bromide and bromoethane reagents independently, and in both cases observed that complete substitution can be accomplished within 15 - 30 minutes at 140 °C. These compounds were isolated

in their fully oxidized neutral form in a similar fashion to **[2]⁰**, (compounds **[3]⁰** and **[4]⁰**, respectively; Figure 3). Notably, previously reported high-pressure reactor synthesis of these species required 3 and 12 hours, respectively, for alkylation to occur at all twelve vertices, suggesting that the microwave-based method can be generally applied to several classes of ether-linked B₁₂(OR)₁₂ clusters and is potentially superior to the previously developed methods.³¹

The relatively short perfunctionalization reaction times made possible by this new microwave technique prompted us to investigate whether it would be possible to utilize ambient synthetic conditions. Specifically, we hypothesized that the fast rate of product formation would outcompete the rate of degradation stemming from the presence of adventitious air and moisture during the synthesis. We therefore tested the synthesis of **2** using as-received non-dried acetonitrile (see SI) with the reagents added to a reaction vessel open to air. The open-air synthesis of **2** proceeded with full conversion in 15 minutes as indicated by ¹¹B NMR spectroscopy on the crude mixture. Following the normal work-up procedure (see SI), **[2]⁰** was isolated in a 63% yield, suggesting that rigorous exclusion of air and moisture is not necessary for this transformation. This open-air synthesis method was successfully used for all subsequent ether-based cluster syntheses reported in this work. We decided to further explore the scope of this transformation by using longer-chain alkyl substituents. Hexyl chain substitution required increased reaction times compared to the shorter ethyl substituent, yet persubstitution was still achieved within two hours, as opposed to 8 hours when using a high-pressure reactor³¹ (isolated as **[5]⁰**, Figure 3). This increased reaction time likely stems from the increase in the length and size of the alkyl reagent affecting the kinetics of the reaction.^{43,44} To further probe the limits of the microwave-based method we tested hexene- and undecene-based electrophiles, and even with the requisite increase in reaction time to 7 and 8 hours respectively, persubstitution proceeded to full conversion. Neutral **[6]⁰** and **[7]⁰** were isolated in 43% and 28% yields, respectively, after oxidation and normal

purification procedures (Figure 3). These derivatives have not been synthesized prior to this report, and their preparation illustrates how one can dramatically increase the size of these ether-based dodecaborate clusters via a direct linkage of large substituents featuring terminal olefins onto $[\text{B}_{12}(\text{OH})_{12}]^{2-}$.

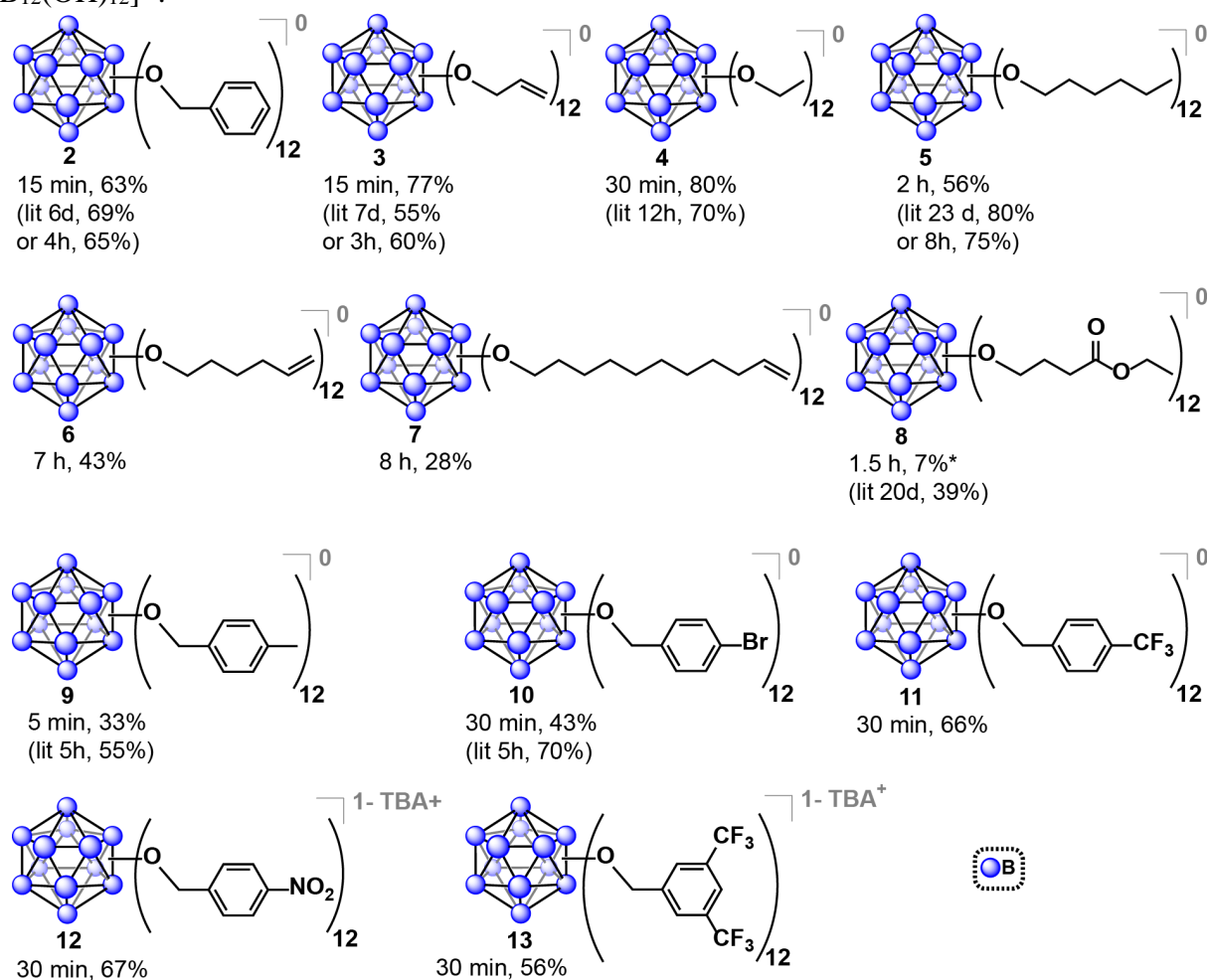


Figure 3. Synthesized $\text{B}_{12}(\text{OR})_{12}$ clusters via microwave-based method. Yields are reported for the species isolated in a designated oxidation state as an average of two independent trials. For previously synthesized species, reported yield is given for comparison.³¹ *Additional 8% of 1-/2-non-oxidized species collected.

Based on the success of the long-chain olefin-containing moieties, we investigated the compatibility of this system with less stable reagents by using ethyl 4-bromobutyrate with $\text{TBA}_2[1]$. Perfunctionalization with ethyl butyrate has been challenging using prior synthetic

methods, requiring multiple-step sequential additions of the alkyl halide and Hünig's base for 20 days while being handled under inert atmosphere conditions.⁴⁵ However, utilizing microwave-assisted synthesis, the same product (Figure 3, **[8]**⁰) was obtained via a single 1.5 hour reaction, followed by oxidation with FeCl₃·6H₂O overnight and purification with column chromatography on SephadexTM and silica gel.

Benzyl-substituted ether-based clusters can feature a high degree of electrochemical tunability as a function of the substituents attached to the aromatic ring.³⁹ Our method allows for the efficient synthesis of clusters containing both electron-rich (**9**) and electron-withdrawing (**10**) benzyl derivatives in yields matching previous methods but with significantly reduced reaction times (Figure 3). We were further intrigued by the possibility of extending the accessible electrochemical window for this class of compounds by utilizing benzyl halide precursors containing highly electron-withdrawing substituents. The perfunctionalized cluster featuring a trifluoromethyl (CF₃) group attached to the *para* position of the benzyl moiety was prepared using our method in 30 minutes, and following oxidation the isolated neutral compound **[11]**⁰ was obtained in 66% yield (Figure 3). The oxidation potential of **[11]**¹/**[11]**⁰ (E_{1/2} = 0.56 V vs Fc/Fc⁺) measured by cyclic voltammetry (CV) is particularly notable since it is higher than any reported B₁₂(OR)₁₂ cluster to date (previously 0.09 V vs Fc/Fc⁺).³⁹ Plotting the Hammett constants of various benzyl substituents⁴⁴ versus the redox potentials of [B₁₂(OR)₁₂] clusters perfunctionalized with these groups³⁹ (Figure 4) indicates the oxidation potential of these clusters can be rationally extended beyond the previously reported electrochemical window.

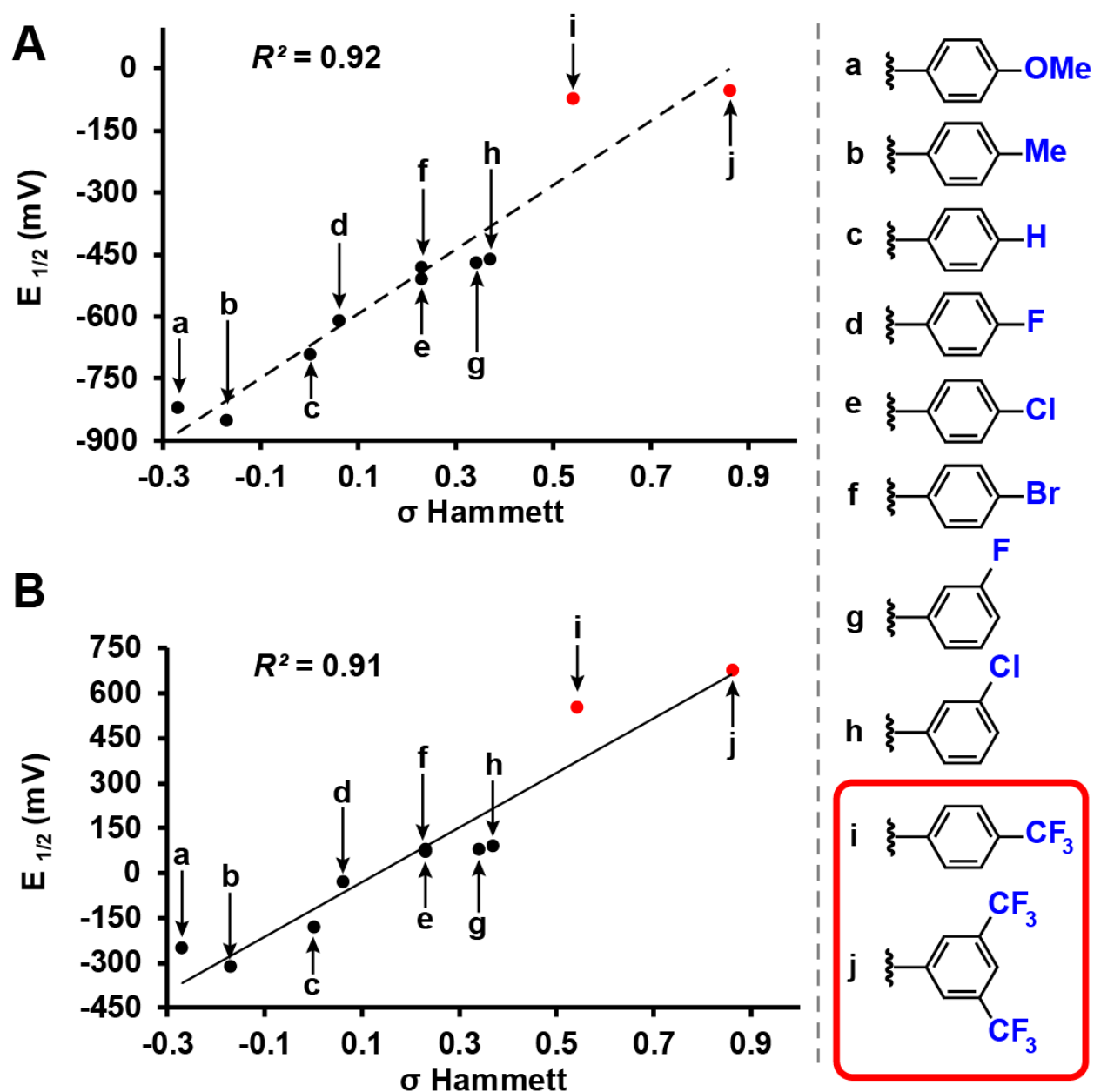


Figure 4. (a) Redox potential of $[B_{12}(OR)_{12}]^{2-/1-}$ and (b) $[B_{12}(OR)_{12}]^{1-/0}$ substituted with various benzyl substituents plotted vs Hammett constants.^{3,44} Previously characterized³⁹ (black) and new (red) $[B_{12}(OR)_{12}]^0$ clusters are shown.

For example, according to the trend suggested by this Hammett plot, a *para*-nitrobenzyl-substituted cluster should exhibit a higher 1-/0 oxidation potential than **11** (Figure 3 and Figure 4).⁴⁴ The perfunctionalized cluster **12** featuring *para*-nitro (NO_2) substituent attached to the benzyl

was successfully synthesized using our method in 30 minutes, however, oxidation of the reaction mixture containing $[\mathbf{12}]^{2-/1-}$ with $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ did not produce any *hypercloso*-neutral species $[\mathbf{12}]^0$. Instead, the radical cluster species $[\mathbf{12}]^{1-}$ was isolated as the only product in 67% yield (Figure 3). This is not surprising, given the predicted oxidation potential for the $[\mathbf{12}]^{1-}/[\mathbf{12}]^0$ redox couple is more positive than the oxidizing strength of $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$.⁴⁶ Attempts to use stronger chemical oxidants (e.g. ceric ammonium nitrate) resulted in cluster degradation. Furthermore, insufficient solubility of $[\mathbf{12}]^{1-}$ as a TBA salt precluded us from obtaining CV measurements for this derivative. Nevertheless, convinced we could expand the electrochemical window for $\text{B}_{12}(\text{OR})_{12}$ species featuring benzyl-based substituents, we turned our attention to a potentially more soluble compound containing a 3,5-*bis*(trifluoromethyl)benzyl group instead.

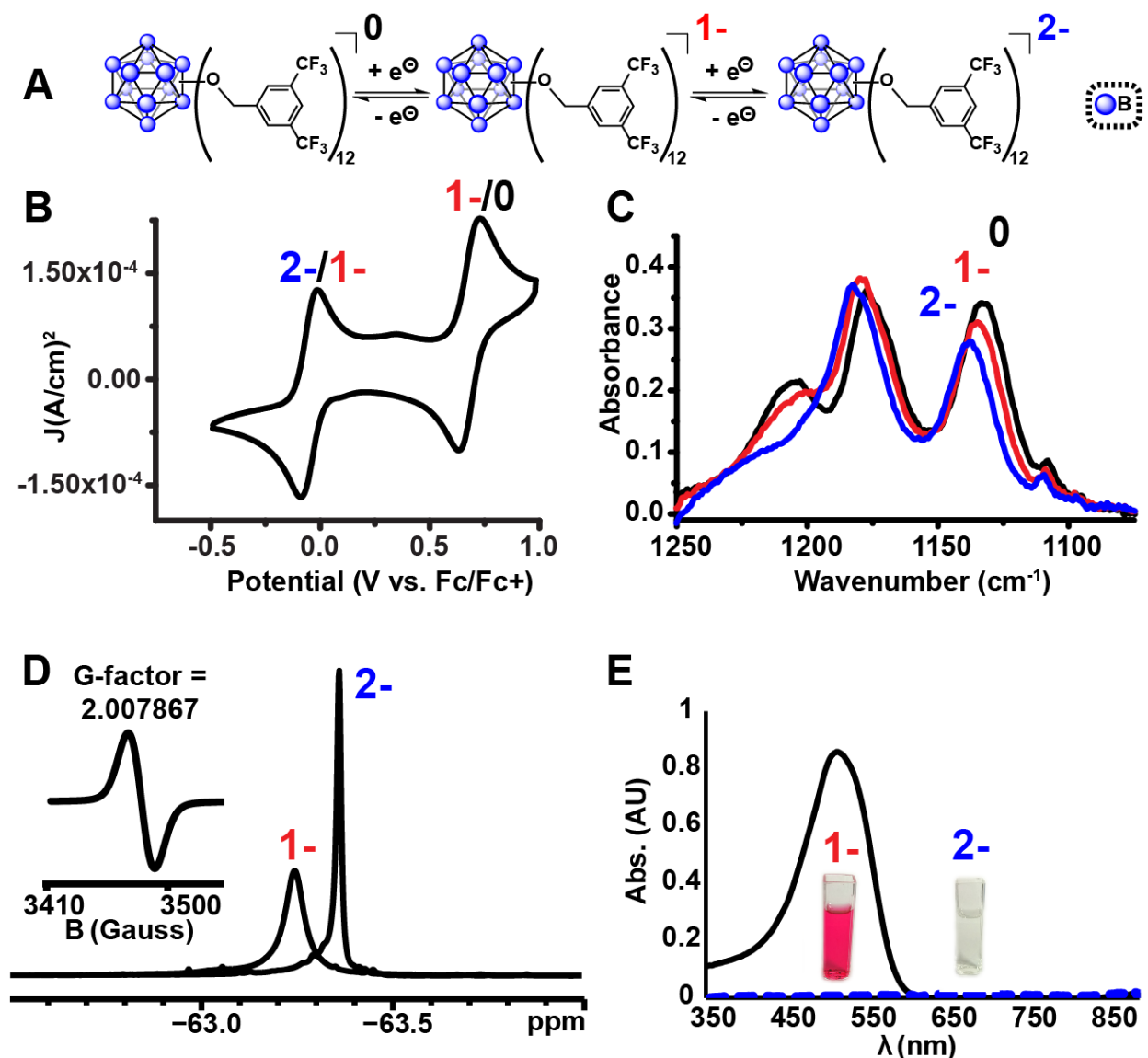


Figure 5. (a) Reversible redox activity of **13**; (b) Cyclic voltammogram (CV) demonstrating two independent, one-electron oxidation/reduction waves between 2⁻/1⁻, and 1⁻/0 states (1 mM **13** with 0.1 M TBAPF₆ in CH₂Cl₂; glassy carbon working electrode, Pt wire counter electrode, Ag/AgCl pseudoreference electrode behind a CoralPor tip; referenced to an internal ferrocene standard); (c) Infrared spectroelectrochemical (IRSEC)^{47,48} analysis of 0/1⁻/2⁻ states of **13**; (d) ¹⁹F NMR spectra of [13]¹⁻ and [13]²⁻ with EPR of [13]¹⁻ (inset); (e) UV-Vis spectra of [13]¹⁻ and [13]²⁻.

Using the microwave method described above, cluster **13** was synthesized in 30 minutes. Interestingly, unlike all of the synthesized clusters reported thus far, the post-microwave reaction mixture was colorless, which is a characteristic feature of the pure dianionic state [13]²⁻ for these

clusters. The lack of color persisted even after column chromatography purification on silica gel in air. The identity of the pure isolated TBA salt of **[13]**²⁻ (73% yield) was validated by full spectroscopic characterization and mass spectrometry (see SI). Oxidation of **[13]**²⁻ with FeCl₃·6H₂O did not produce the neutral cluster, rather the pure **[13]**¹⁻ radical species was isolated in 78% yield, resulting in 56% net yield for **[13]**¹⁻ species starting from TBA₂[**1**]. Use of stronger chemical oxidants such as ceric ammonium nitrate (CAN) degraded the boron cage, producing a diagnostic ¹¹B NMR resonance at δ 20 characteristic of borates.⁴⁹ Nevertheless, we were able to observe neutral cluster **[13]**⁰ electrochemically via infrared spectroelectrochemistry (IR-SEC) and cyclic voltammetry (CV) in CH₂Cl₂ (Figure 5, B, C). CVs of **[13]**¹⁻ as the TBA salt in CH₂Cl₂ showed two quasi-reversible redox features at -0.05 V and 0.68 V vs Fc/Fc⁺, corresponding to the 2-/1- and 1-/0 transitions, respectively. IR-SEC experiments where the applied potential was increased to more positive potentials incrementally on the TBA salt of **[13]**²⁻ showed subtle changes in the IR stretching modes for all three oxidation states of **[13]** around 1130-1140 cm⁻¹ and 1200-1220 cm⁻¹. The shift in these IR bands assigned to the B–O bond⁵⁰ to higher wavenumbers from **[13]**²⁻/**[13]**¹⁻/**[13]**⁰ is consistent with those observed in the same region for the analogous *para*-CF₃ compound **[11]**⁰ which can be isolated in its neutral form via direct synthesis (see SI). The high oxidation potential for the **[13]**¹⁻/**[13]**⁰ redox couple observed from cyclic voltammetry (E_{1/2} = 0.70 V vs Fc/Fc⁺) is notable since it is the highest observed 1-/0 oxidation potential for the B₁₂(OR)₁₂ class of clusters reported to date³⁹ and is ~130 mV higher than the *para*-CF₃ benzyl cluster **11**. ¹⁹F NMR spectroscopy provides another diagnostic handle on the oxidation state of this compound (Figure 5, D). ¹¹B NMR spectra for these clusters typically show a singlet around δ -14 to -16 for the 2- state, though the 1- state is silent due to the presence of the paramagnetic radical (confirmed by EPR, Figure 5D (inset)). However, with ¹⁹F NMR spectroscopy, a shift from a singlet resonance in **[13]**²⁻ at δ -63.36 to a broad singlet at δ -63.24 for

$[13]^{1-}$ was observed. This broadening is consistent with the paramagnetic nature of $[13]^{1-}$, where F atoms are located far enough from the unpaired electron-carrying B_{12} -based cluster core to be resolved by ^{19}F NMR spectroscopy.

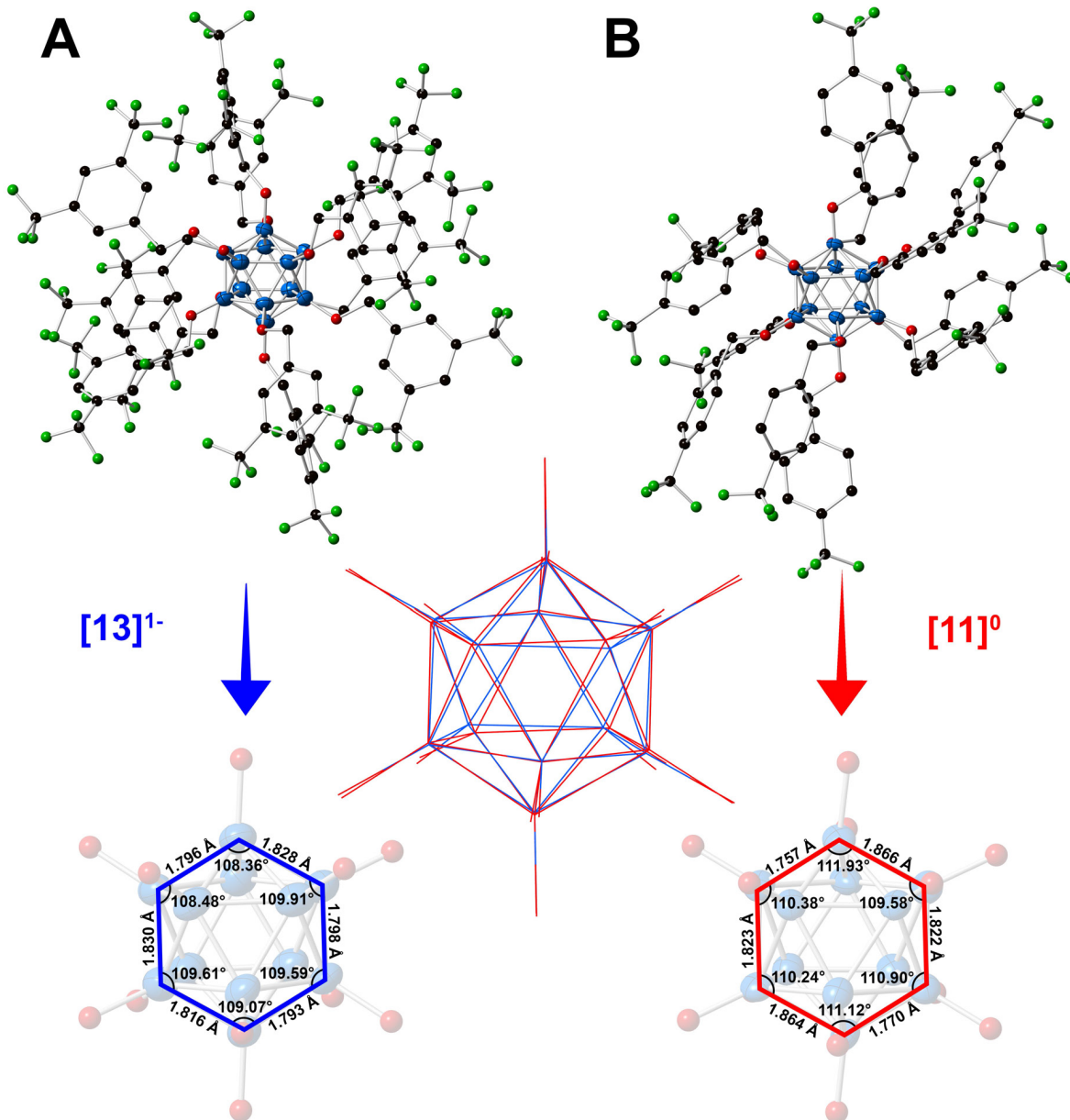


Figure 6. (A-B) Solid state X-ray structures for $[13]^{1-}$ and $[11]^{0}$ shown with 50% thermal ellipsoid probabilities for the boron atoms (hydrogen atoms omitted for clarity). B atoms are blue, O – red, C – black, and F – green. Selected bond lengths and angles of the boron cores (substituents omitted for clarity) for $[11]^{0}$ and $[13]^{1-}$ are shown on the bottom. Overlay of the two cores ($[13]^{1-}$ in blue, $[11]^{0}$ in red) is depicted in the middle.

The structural parameters of the boron clusters featuring persubstituted vertices can exhibit significant distortions in the solid state as a function of the substituent and the redox state as

determined by X-ray crystallographic studies. Specifically, $[2]^{2-}$ exhibits nearly identical B–B bond distances (1.781(4) – 1.824(4) Å) and angles (B–B–B $107.798^\circ < \alpha < 109.229^\circ$) as expected for a perfect icosahedron, yet as the cluster is oxidized to the electron-deficient $[2]^{1-}$ the structure expands and distorts slightly, with further distortion observed in the neutral state.³⁷ Additionally, the B–O bond lengths decrease as the cluster is oxidized from $[2]^{2-}$, which contains the longest average B–O distances, to the neutral $[2]^0$ state with the shortest B–O distances.³⁷ This observed trend of B–B bond lengthening, B–O bond contraction, and B–B–B angle distortion within the core as a function of cluster oxidation state is supported qualitatively by the crystal structures for neutral $[11]^0$ and radical $[13]^{1-}$ (Figure 6), which show comparable changes to those observed between $[2]^{1-}$ and $[2]^0$. Selected bond lengths and angles for $[11]^0$ and $[13]^{1-}$ are shown in Figure 6.

X-ray photoelectron spectroscopy (XPS) has been widely used to study oxidation states in inorganic compounds.^{51–54} We therefore decided to utilize this technique to further elucidate the oxidation state and effect of functionalization for these boron clusters. Boron XPS spectra for several representative clusters synthesized in our study (Figure 7) indicate a clear trend observed in the shift of B–B bond peak energies depending on the redox state of the functionalized cluster. The observed geometric distortion of these boron cluster icosahedra with oxidation from 2- to 1- and further to neutral species results in an increased B–B binding energy (Figure 7A). The nature of the substituents also produces a clear trend in these measurements, as compound **13** also exhibits an increase in B–B binding energy as the cluster increases in oxidation state from $[13]^{2-}$ to $[13]^{1-}$ (Figure 7B), yet both are higher in energy than that of neutral cluster $[11]^0$. However, despite the change in oxidation potential and binding energy observed from the XPS data from substituent effects, the nature of the electron radical delocalization throughout the boron-based core remains consistent. A single, broad symmetric EPR signal centered between 3450 and 3500 Gauss was

observed for all cluster species isolated in the radical form with g values ranging between 2.0079 and 2.0081 depending on the substituent (see SI). Due to the 3D delocalization of the single electron across the 12 boron nuclei comprising the cluster, there exist a large number of possible hyperfine couplings.³⁶ Overlap of these hyperfine couplings ultimately gives rise to the single broad line observed in the EPR spectra.

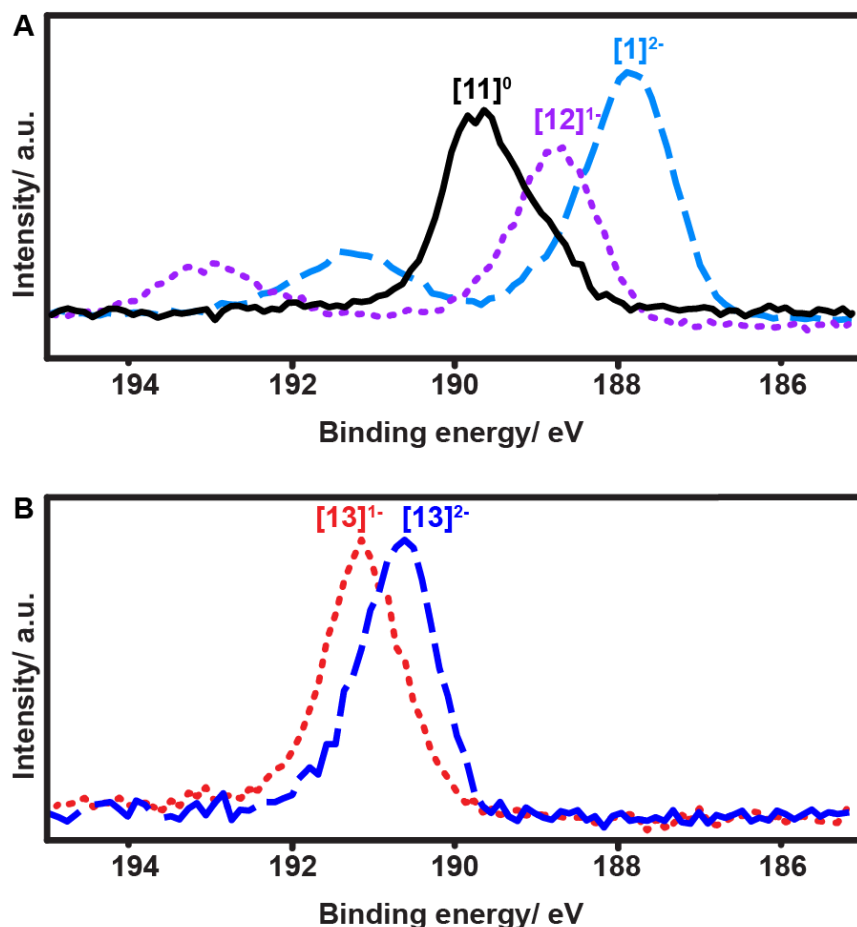


Figure 7. (a) Boron XPS spectra for $TBA_2[1]^{2-}$, $TBA[12]^{1-}$, and $[11]^0$ showing an increase in B–B bond energy with increasing oxidation state; (b) Boron XPS spectra for $TBA_2[13]^{2-}$ and $TBA[13]^{1-}$ indicating the higher B–B bond energies of the $[13]^{2-}$ and $[13]^{1-}$ anions compared to the other substituted clusters.

In addition to previously mentioned benefits such as shortened reaction durations and the lack of stringent requirements for inert reaction conditions, microwave-assisted synthesis allows for one-pot, single-step reactions that would otherwise require more elaborate protocols.

Monosubstitution of a benzyl ligand followed by persubstitution of the remaining eleven vertices has previously required a lengthy process involving multi-step syntheses,⁵⁵ whereas we demonstrate a one-pot approach enabled by our microwave-based method. For example, mixed-substituent $B_{12}(OEt)_{11}(OBn)$ cluster (**14**, Figure 8) can be formed in a single step simply by adding a stoichiometric amount of the desired reagents into a single reaction vessel.

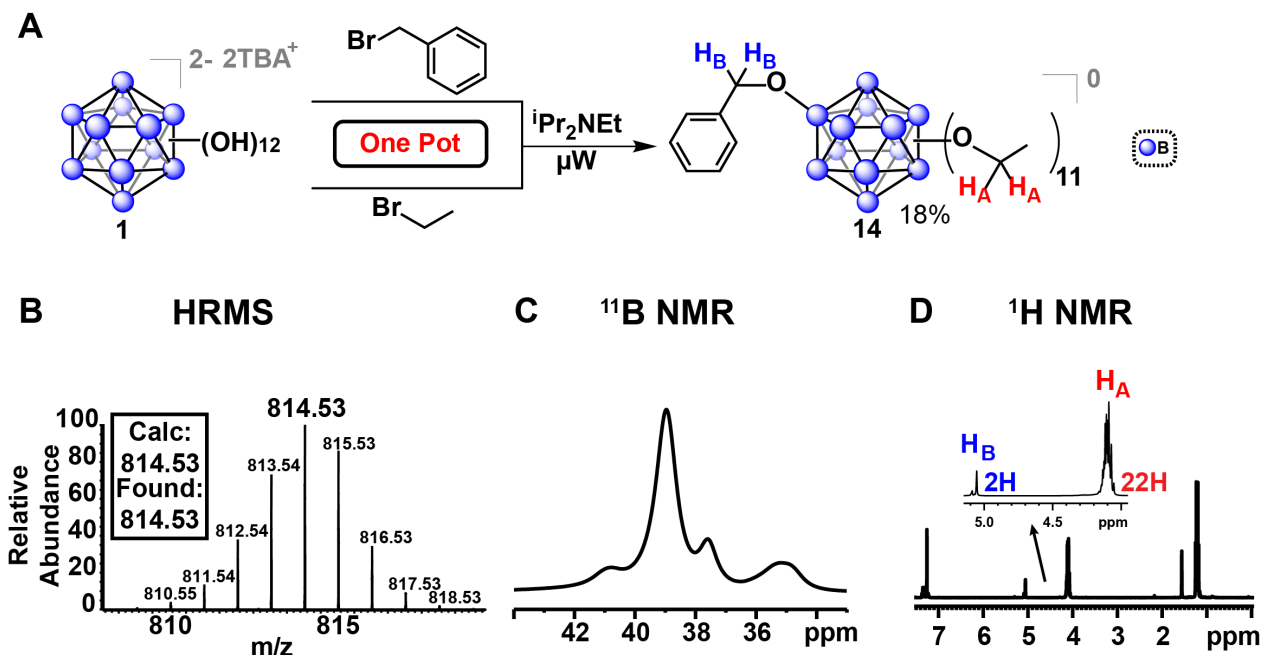


Figure 8. One-pot synthesis of vertex-differentiated *hypercloso* cluster [**14**]⁰ from a 60:1:1 molar mixture of benzyl bromide:bromoethane:TBA₂[**1**]. Isolated yield of [**14**]⁰ was 18% (compound [**4**]⁰ was also formed as an additional product of the reaction). ^{11}B NMR spectra indicates loss of the icosahedral symmetry due to vertex differentiation, 1H NMR integrations show 24H (CH₂) and 36H (CH₃) for the 11 ethyl groups and 2H (CH₂) and 5H (Ph) for the single benzyl moiety.

This reaction was completed in 30 minutes, producing three distinct species as a mixture which were oxidized with $FeCl_3 \cdot 6H_2O$ as previously described and subsequently isolated via column chromatography on silica: the perfunctionalized ethyl cluster [**4**]⁰, a small amount (<5%) of di-substituted [$B_{12}(OEt)_{10}(OBn)_2$]⁰ clusters, and the desired [**14**]⁰ in 18% yield. This method

represents a significantly faster route to produce this class of $[B_{12}(OR)_{11}(OR')]$ mixed-substituent clusters.

Conclusions

A rapid microwave-assisted synthetic route to perfunctionalized ether-linked $B_{12}(OR)_{12}$ clusters is disclosed and the robust nature of the technique demonstrated by the synthesis and characterization of multiple derivatives of **1**. For previously synthesized compounds, reaction duration was significantly reduced and prior requirements for oxygen-free and anhydrous reaction conditions were eliminated. Our method also allows for a unique one-pot synthesis of mixed-substituent clusters with good selectivity under the same open-air conditions. The cluster species described here maintain the attractive properties of earlier derivatives, behaving as redox-active cores which show delocalization of electrons throughout the entire 3D boron cage, while the new functional groups provide a significant expansion to the available tuneable redox potential window for this class of clusters.

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Experimental and Supporting Information

General considerations.

Initial microwave synthesis reactions were carried out under an inert atmosphere of nitrogen using standard glovebox techniques. All post-microwave work-up and characterization was performed under ambient conditions. All reactions designated as “open-air” were carried out and worked up under ambient conditions. The “ambient conditions” for this manuscript refer to room temperature (20 - 25 °C) and uncontrolled laboratory air.

Materials.

Deuterated solvents were purchased from Cambridge Isotope Laboratories and used as received. MilliQ water described in this manuscript refers to purified potable water with a resistivity at 25 °C of $\leq 18.2 \text{ M}\Omega\cdot\text{cm}$. $[\text{NEt}_3\text{H}]_2[\text{B}_{12}\text{H}_{12}]$ was purchased from Boron Specialties (USA). Ethanol (200 proof) was purchased from Decon Labs and used as received. $\text{FeCl}_3\cdot 6\text{H}_2\text{O}$ ($\geq 97\%$), $\text{CsOH}\cdot 1\text{H}_2\text{O}$ ($\geq 99.5\%$), H_2O_2 (30% in H_2O), $[\text{N}^n\text{Bu}_4]\text{OH}$ (40% in H_2O), bromoethane ($\geq 99\%$), 6-bromo-1-hexane (98%) allyl bromide (99%), 4-nitrobenzyl bromide (99%), acetonitrile ($\geq 99.9\%$), CH_2Cl_2 ($\geq 99.5\%$), ethyl acetate ($\geq 99.5\%$), hexanes ($\geq 98.5\%$), methanol ($\geq 99.8\%$), *N,N*-diisopropylethylamine ($\geq 99\%$), and tetrabutylammonium hexafluorophosphate ($\geq 99.0\%$, electrochemical grade and 98%, recrystallized from ethanol and dried under vacuum at 90 °C) were purchased from Sigma-Aldrich. Benzyl bromide (99%) and ethyl 4-bromobutyrate (98%) were purchased from Alfa Aesar, and 4-methylbenzyl bromide (98%) was purchased from Acros. 6-bromo-1-hexene (98%), undec-10-enyl bromide (95%), 4-trifluoromethylbenzyl bromide

(99%), and 3,5-bis(CF₃)₂-benzyl bromide (97%) were purchased from Oakwood. All reagents were used as received unless otherwise indicated.

Instruments.

Bruker AV400 and AV500 spectrometers were used to obtain ¹¹B, ¹³C{¹H}, ¹H, and ¹⁹F NMR spectra and Bruker Topspin software was used to process the NMR data. ¹³C{¹H} and ¹H NMR spectra were referenced to residual solvent resonances in deuterated solvents (due to high humidity H₂O resonances are often present). ¹¹B and ¹⁹F NMR spectra were referenced to BF₃·Et₂O and CFCl₃ standards, respectively. A Bruker EMX EPR spectrometer was used to acquire EPR spectra, with all spectra collected in CH₂Cl₂ at ambient temperature. Mass spectrometry data was acquired using a Thermo Scientific™ Q-Exactive™ Plus instrument with a quadrupole mass filter and Orbitrap mass analyzer. IR spectroscopy was acquired on solid samples using a PerkinElmer Spectrum Two FT-IR spectrometer equipped with a diamond universal ATR probe. X-ray photoelectron spectroscopy (XPS) data was acquired using an AXIS Ultra DLD instrument (Kratos Analytical Inc., Chestnut Ridge, NY, USA) with a monochromatic Al Kα X-ray source (10 mA for survey and high-resolution scans). A 300 x 700 nm oval spot size and ultrahigh vacuum (10⁻⁹ Torr) were used, with 160 eV pass energy for survey spectra and 20 eV for high-resolution spectra of B 1s using a 200 ms dwell time and 20 scans. All XPS peaks were externally referenced to the C 1s signal at 284.6 eV. The experimental setup and design of the infrared-spectroelectrochemistry (IR-SEC) cell has been published previously.^{47,48}

X-ray data collection and processing parameters.

For [11] and [13]¹⁻, a single crystal was mounted on a nylon loop using perfluoropolyether oil and cooled rapidly to 100 K with a stream of cold dinitrogen. Diffraction data were measured using a Bruker APEX-II CCD diffractometer using Mo- K_{α} radiation. The cell refinement and data reduction were carried out using Bruker SAINT and the structure was solved with SHELXS-97. All subsequent crystallographic calculations were performed using SHELXL-2013.

Cyclic voltammetry and IRSEC.

Cyclic voltammetry was performed on [11] and [14] using a CH Instruments CHI630D potentiostat with a glassy carbon disc working electrode, platinum wire counter electrode, and Ag/Ag⁺ wire pseudoreference. All experiments were conducted in 0.1M [NⁿBu₄]PF₆/CH₂Cl₂ with 0.5 mM analyte concentrations (11.2 mg in 10 mL for [11] and 3.7 mg in 10 mL for [14]). The CH₂Cl₂ was dried in house with a custom drying system running through two alumina columns prior to use. The solution was degassed by bubbling Ar, and the cyclic voltammetry was performed under Ar gas. For [11], a scan rate of 0.1 mV/s was used with Fc/Fc⁺ as an external standard. For [14], a scan rate of 0.5 mV/s was used with Fc/Fc⁺ as an internal standard.

IRSEC and cyclic voltammetry for [13] were performed using a Pine Instrument Company model AFCBP1 bipotentiostat and BAS Epsilon potentiostat, respectively. For IR-SEC, as the potential was scanned, thin-layer bulk electrolysis was monitored by Fourier-Transform Reflectance IR off the electrode surface. All experiments were conducted in 0.1 M [NⁿBu₄]PF₆/CH₂Cl₂ solutions with analyte concentrations of ~5 mM (13.9 mg in 1 mL) prepared under a nitrogen atmosphere. The IR-SEC cell used a glassy carbon working electrode, Pt wire

counter electrode, and Ag wire pseudoreference electrode. The anionic $[\text{N}^n\text{Bu}_4]_2[\mathbf{13}]^{2-}$ was used for IRSEC, starting at resting potential and increasing to more oxidizing potentials stepwise.

Microwave Synthesis.

Microwave reactions were performed using a CEM Discover SP microwave synthesis reactor. Except where noted otherwise, all reactions were performed in glass 10 mL microwave reactor vials purchased from CEM with silicone/PTFE caps. Flea micro PTFE-coated stir bars were used in the vials with magnetic stirring set to high and 15 seconds of premixing prior to the temperature ramping. All microwave reactions were carried out at 140 °C with the pressure release limit set to 250 psi (no reactions exceeded this limit to trigger venting) and the maximum wattage set to 250W (the power applied was dynamically controlled by the microwave instrument and did not exceed this limit for any reactions). Column chromatography was performed using 2.0 - 2.25 cm inner diameter glass fritted chromatography columns with 20-30 cm of slurry-packed silica gel to ensure full separation of reagents and products. Unfiltered pressurized air was used to assist column chromatography.

Dicesium Dodecahydroxy-*closo*-dodecaborate Cs₂[1].

CsOH·H₂O (14.00 g, 83.4 mmol) was dissolved in methanol (130 mL) in a 300 mL glass round bottom flask. $[\text{NEt}_3\text{H}]_2[\text{B}_{12}\text{H}_{12}]$ (13.3758 g, 38.9 mmol) was added along with a PTFE-coated stir bar, and the reaction was left to stir vigorously for 18 h at ambient temperature. The cloudy suspension was then filtered through a 60 mL fritted glass funnel and washed with methanol (3 x 20 mL). The resulting white solid was dried on the frit for 1.5 h then left under high vacuum for 12 h and complete conversion to Cs₂[*closo*-B₁₂H₁₂] was confirmed by the absence of amine

resonances in the ^1H NMR spectrum. Alternatively, commercially-obtained $\text{Cs}_2[\text{B}_{12}\text{H}_{12}]$ (98%, Strem) can be utilized for hydroxylation.

Note: The perhydroxylation procedure described herein should always be undertaken with caution and careful planning in order to ensure the $\text{Cs}_2[\text{B}_{12}\text{H}_{12}]$ reagent is pure and contains no organic contaminants. Blast shielding to contain any possible explosions should be utilized. Under no circumstances should the hydrogen peroxide used in the reaction come into contact with any organic material or solvents due to the possibility of an explosion. Synthesis of $\text{Cs}_2[\text{B}_{12}(\text{OH})_{12}]$ and the ion exchange to produce $[\text{N}^n\text{Bu}_4]_2[\text{B}_{12}(\text{OH})_{12}]$ have been described elsewhere,^{41,42} but will be reported here for convenience. $\text{Cs}_2[\text{B}_{12}\text{H}_{12}]$ (15.0 g, 36.8 mmol) was added to a glass three-necked round bottom flask with a water-cooled condensing coil in the top slot. The rear neck outlet contained a stopcock for venting pressure, and the front outlet was sealed with a glass stopper and secured with a plastic Keck clip. The apparatus was suspended in a silicone oil bath on a hot plate and secured, with a blast shield in front as a precaution against any potential explosion. The oil bath was heated to 95 °C, and a 50 mm oval PTFE stir bar was added to the flask, and the reaction was initiated with the addition of H_2O_2 (50 mL, 30% in H_2O). The flask was stoppered and the mixture allowed to stir at that temperature for 2 h. After 2 h, additional H_2O_2 (12 mL) was added, with the flask being vented, the glass stopper removed, and upon completion of addition, re-stoppering the flask. This addition of H_2O_2 was repeated every 2 h until a total volume of 60 mL was added to the reaction mixture. Upon completion of the addition, the oil bath temperature was increased to 105 °C, and additional H_2O_2 aliquots (10-15 mL) were added every 2 - 3 days, cooling the solution in the flask by raising it out of the oil bath and leaving it to cool for 20 – 30 minutes prior to each addition. After 14 days, the progress of the reaction was assessed *via* ^{11}B NMR, with reaction completion indicated by the appearance of a broad resonance at -18.0 ppm corresponding to $\text{Cs}_2[\text{B}_{12}(\text{OH})_{12}]$ and the disappearance of the resonance at -16 ppm corresponding to unreacted

Cs₂[B₁₂H₁₂]. Once the reaction is complete (as assessed by ¹¹B NMR), the mixture was cooled to 2 – 8 °C and cold MilliQ water was used to transfer the solution and solid product to a 150 mL glass fritted filter funnel. The crude product was washed with additional MilliQ water prior to drying on the filter frit under water-aspirator vacuum for 6 – 12 hours. Yield: 18.8 g (85 %).

From this point, NⁿBu₄ will be referred to as TBA. For the cation exchange, Dowex 50X8 (100-200 mesh, hydrogen form, Sigma-Aldrich) was washed with MilliQ water in a 500 mL glass beaker until neutral (decanting and discarding the wash fractions), and [TBA]OH (40% in H₂O) was added in 10 mL increments until basic with magnetic stirring using a 50 mm PTFE coated standard stir bar. After 30 minutes, the pH was assessed again, and if no longer basic, additional [TBA]OH was added to restore basicity and left to stir overnight covered with a watch glass. The resin was slurry-packed into a 5 x 30 cm column wrapped with heating tape (controller set to hold temperature at 50 °C), and washed with MilliQ water until neutral. Cs₂[B₁₂(OH)₁₂] (5.996 g, 10.0 mmol) was dissolved in boiling water (600 mL), cooled to 50 °C and slowly added to column. The product was washed with an additional 750 mL of 50 °C water, and the product was concentrated *in vacuo* and lyophilized to produce pure TBA₂[**1**]. Yield: 6.90 g (85 %). TBA₂[**1**] is a white solid. ¹H NMR (500 MHz, CDCl₃): δ 4.66 (s, 12H, OH), 3.08 (m, 8H, N-CH₂), 1.54 (m, 8H, N-CH₂CH₂), 1.25 (m, 8H, N-(CH₂)₂CH₂), 0.84 (m, 12H, N-(CH₂)₃CH₃). ¹¹B{¹H} NMR (128 MHz, D₂O): δ -17.9. *Note: TBA₂[1] is air-stable, but hygroscopic. Store under inert atmosphere or in a sealed desiccator to prevent excess absorption of water over extended periods of time under storage.*

General ether alkylation/benylation of TBA₂[1] to B₁₂(OR)₁₂ microwave procedure.

Reactions were performed using TBA₂[1] which was weighed and placed into a 10 mL glass microwave vial and transferred out of a nitrogen-filled glovebox, being opened to the air prior to synthesis. The acetonitrile solvent, base, and alkyl reagents were all used under ambient temperature and pressure conditions with no additional purification or drying. *Note: the initial inert-atmosphere trials mentioned in the main text for 2, 3, and 4 were prepared inside a nitrogen-filled glovebox using rigorously anhydrous solvent. Once the PTFE/silicone cap was placed on the microwave vial it was transferred out of the glovebox and the reactions were performed identically to the open-air reactions.* TBA₂[1] (50.0 mg, 0.061 mmol) was transferred to a 10 mL glass microwave vial containing a flea micro stir bar and dissolved in acetonitrile (1 mL). *N,N*-diisopropylethylamine (Hünig's base, 0.2 mL, 1.15 mmol) and alkyl halide (7.6 mmol) were added, and a PTFE/silicone cap was placed on the microwave vial. The mixture was heated to 140 °C with stirring in the microwave for 5 min to 8 hrs (depending upon the alkyl halide), with the progress of the reaction monitored *via* ¹¹B NMR spectroscopy. Multiple resonances between -14 and -16 ppm are first observed, indicating partial substitution of the 12 vertices. The reaction has reached completion when these resonances coalesce to a broad singlet resonance between -14 and -16 ppm corresponding to the fully substituted [B₁₂(OR)₁₂]²⁻ species. The color of the reaction mixture is typically a faint yellow initially, with the completed reaction mixture changing to a pink/purple, faint pink, or deep red/orange color indicative of the 1- species. Upon completion of the reaction, excess acetonitrile and base were removed *via* rotary evaporation. With the exception of **3**, **4**, **6**, and **14** the remaining reaction mixture containing product and unreacted alkyl halide were separated *via* column chromatography with silica gel. The unreacted alkyl halide (clear and colorless or slightly yellow/orange, UV active) was eluted first, followed by the elution of the remaining pink/purple product mixture consisting of 1-/2- species (note that the 2- species is

colorless). The excess solvent was removed *via* a rotary evaporator, and the remaining 1-/2- product mixture was dissolved in a 90/5/5 ethanol/acetonitrile/MilliQ H₂O or 90/10 ethanol/acetonitrile mixture and transferred to a 50 mL round bottom flask. FeCl₃·6H₂O (0.3 g, 1.11 mmol) was added to the dissolved mixture, and subsequently stirred vigorously for 12-24 h at ambient temperature. The solvent was removed from the resulting dark orange/brown mixture *via* rotary evaporation, and the neutral [*hypercloso*-B₁₂(OR)₁₂]⁰ or radical TBA[*hypocloso*-B₁₂(OR)₁₂]¹⁻ product was separated from the FeCl₃·6H₂O *via* column chromatography with silica gel. A dark orange, yellow-orange, or red-orange band consisting of neutral [*hypercloso*-B₁₂(OR)₁₂]⁰ was eluted with CH₂Cl₂, with a pink/purple band containing charged 1-/2- species eluting next if any product was not fully oxidized. The red or orange fraction containing the desired neutral closoomer was dried with rotary evaporation followed by high vacuum, and the above procedure for oxidation could be repeated on the remaining 1-/2- mixture to obtain additional neutral product if any non-fully oxidized product remains. *Note: if the final oxidized product appears to have any impurities (via ¹H NMR spectroscopy), eluting the product through an additional silica plug or short column with 1:1 CH₂Cl₂/hexanes should remove any contaminants.*

Dodeca(benzyloxy)-*hypercloso*-dodecaborane [2]

TBA₂[1] (50.0 mg, 0.061 mmol) was added to a 10 mL glass microwave vial and transferred out of a nitrogen filled glovebox, opened to the air, and dissolved in 1 mL acetonitrile. *N,N*-diisopropylethylamine (0.2 mL, 1.15 mmol) and benzyl bromide (1.74 mL, 14.7 mmol) were added along with a flea micro stir bar, the vial was sealed with a PTFE/silicone cap, and the mixture was heated at 140 °C with stirring in the microwave for 15 min. The volatiles were removed *via* rotary evaporation, and the excess reagent was eluted through a slurry-packed silica gel column with 65/35 hexanes/ethyl acetate, and the pink/purple product mixture was eluted with CH₂Cl₂.

The CH₂Cl₂ was removed *via* rotary evaporation, the remaining charged 1-/2- product mixture was dissolved in 5 mL 90/5/5 ethanol/acetonitrile/H₂O, FeCl₃·6H₂O (0.3 g, 1.11 mmol) was added and the mixture was left to stir for 12-24 h. Following oxidation, the solvent mixture was removed *via* rotary evaporation, and an orange band containing the neutral product was separated from the FeCl₃·6H₂O through a slurry-packed silica gel column with CH₂Cl₂. The CH₂Cl₂ was removed *via* rotary evaporation and the final neutral product **2** was dried under high vacuum to obtain an isolated yield of 54.3 mg (63%). Compound **2** is a dark orange solid. ¹H NMR (500 MHz, CDCl₃): δ 7.08 - 7.19 (m, 60H, C₆H₅), 5.25 (s, 24H, O-CH₂). ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 140.8, 128.4, 127.3, 73.4. ¹¹B{¹H} NMR (128 MHz, CDCl₃): δ 41.8. HRMS (Orbitrap): *m/z* calculated for C₈₄H₈₄B₁₂O₁₂ (M⁺), 1414.72 Da; found, 1414.72 Da.

Dodeca(allyloxy)-hypercloso-dodecaborane [3]

Note: this reaction should be performed with minimal exposure to light. TBA₂[**1**] (100.0 mg, 0.122 mmol) was added to a 10 mL glass microwave vial and transferred out of a nitrogen filled glovebox, opened to the air, and dissolved in 2 mL acetonitrile. *N,N*-diisopropylethylamine (0.4 mL, 2.30 mmol) and allyl bromide (1.28 mL, 14.68 mmol) were added along with a flea micro stir bar, the vial was sealed with a PTFE/silicone cap, and the mixture was heated at 140 °C with stirring in the microwave for 15 min. The volatiles and excess reagent were removed *via* rotary evaporation and the purple 1-/2- product mixture was dissolved in 5 mL 90/10 ethanol/acetonitrile, FeCl₃·6H₂O (0.3 g, 1.11 mmol) was added and the mixture was left to stir for 12-24 h (wrapped in foil to avoid excessive light exposure). Following oxidation, the solvent mixture was removed *via* rotary evaporation, and a yellow band containing the neutral product was separated from the FeCl₃·6H₂O through a slurry-packed silica gel column with CH₂Cl₂. The CH₂Cl₂ was removed *via* rotary evaporation and the final neutral product **3** was dried under high vacuum to obtain an

isolated yield of 76.8 mg (77%). Compound **3** is a dark yellow-orange viscous oil. ^1H NMR (500 MHz, CDCl_3): δ 5.91 – 5.99 (m, 12H, CH), 5.21 (dq, 12H, CH), 5.05 (dq, 12H, CH), 4.62 (m, 24H, O- $\underline{\text{CH}_2}$). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 136.9, 114.2, 71.6. $^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3): δ 41.1. HRMS (Orbitrap): m/z calculated for $\text{C}_{36}\text{H}_{60}\text{B}_{12}\text{O}_{12}$ (M^-), 814.53 Da; found, 814.53 Da. *Note: Compound 3 should be stored at -20 °C or used immediately.*

Dodeca(ethoxy)-hypercloso-dodecaborane [4]

TBA₂[**1**] (50.0 mg, 0.061 mmol) was added to a 10 mL glass microwave vial and transferred out of a nitrogen filled glovebox, opened to the air, and dissolved in 1 mL acetonitrile. *N,N*-diisopropylethylamine (0.2 mL, 1.15 mmol) and bromoethane (1.65 mL, 22.1 mmol) were added along with a flea micro stir bar, the vial was sealed with a PTFE/silicone cap, and the mixture was heated at 140 °C with stirring in the microwave for 30 min. The volatiles and excess reagent were removed *via* rotary evaporation, the remaining charged 1-/2- product mixture was dissolved in 4 mL 90/5/5 ethanol/acetonitrile/ H_2O , $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ (0.3 g, 1.11 mmol) was added and the mixture was left to stir for 12-24 h. Following oxidation, the solvent mixture was removed *via* rotary evaporation, and an orange band containing the neutral product was separated from the $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ through a slurry-packed silica gel column with CH_2Cl_2 . The CH_2Cl_2 was removed *via* rotary evaporation and the final neutral product **4** was dried under high vacuum to obtain an isolated yield of 32.7 mg (80%). Compound **4** is a dark orange solid. ^1H NMR (500 MHz, CDCl_3): δ 4.09 (q, 24H, O- $\underline{\text{CH}_2}$), 1.24 (t, 36H, CH_3). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 66.8, 17.8. $^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3): δ 37.7. HRMS (Orbitrap): m/z calculated for $\text{C}_{24}\text{H}_{60}\text{B}_{12}\text{O}_{12}$ (M^-), 670.53 Da; found, 670.53 Da.

Dodeca(hexoxy)-hypercloso-dodecaborane [5]

TBA₂[**1**] (99.0 mg, 0.121 mmol) was added to a 10 mL glass microwave vial and transferred out of a nitrogen filled glovebox, opened to the air, and dissolved in 2 mL acetonitrile. *N,N*-diisopropylethylamine (0.4 mL, 2.30 mmol) and 6-bromo-1-hexane (2.85 mL, 20.3 mmol) were added along with a stir bar, the vial was sealed with a PTFE/silicone cap, and the mixture was heated at 140 °C with stirring in the microwave for 2 h. The volatiles were removed *via* rotary evaporation at 65 °C, the remaining charged 1-/2- product mixture was dissolved in 5 mL 90/5/5 ethanol/acetonitrile/H₂O, FeCl₃·6H₂O (0.3 g, 1.11 mmol) was added and the mixture was left to stir for 12-24 h. Following oxidation, the solvent mixture was removed *via* rotary evaporation, and an orange band containing the neutral product was separated from the FeCl₃·6H₂O through a slurry-packed silica gel column with CH₂Cl₂. The CH₂Cl₂ was removed *via* rotary evaporation and the final neutral product **5** was dried under high vacuum to obtain an isolated yield of 91.4 mg (56%). Compound **5** is a dark yellow-orange oil. ¹H NMR (400 MHz, CDCl₃): δ 4.02 (t, 24H, O-CH₂), 1.54 (m, 24H, CH₂CH₂(CH₂)₃CH₃), 1.31, (m, 72H, CH₂CH₂(CH₂)₃CH₃), 0.89 (m, 36H, CH₃). ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 70.2, 32.2, 31.8, 25.9, 22.8, 14.1. ¹¹B{¹H} NMR (128 MHz, CDCl₃): δ 42.0. HRMS (Orbitrap): *m/z* calculated for C₇₂H₁₅₆B₁₂O₁₂ (M⁺), 1343.28 Da; found, 1343.28 Da.

Dodeca(6-hexeneoxy)-hypercloso-dodecaborane [6]

Note: this reaction should be performed with minimal exposure to light. TBA₂[**1**] 50.0 mg (0.061 mmol) was added to a 10 mL glass microwave vial and transferred out of a nitrogen filled glovebox, opened to the air, and dissolved in 1 mL acetonitrile. *N,N*-diisopropylethylamine (0.2

mL, 1.15 mmol) and 6-bromo-1-hexene (0.59 mL, 4.41 mmol) were added along with a stir bar, the vial was sealed with a PTFE/silicone cap, and the mixture was heated at 140 °C with stirring in the microwave for 7 h. The volatiles and excess reagent were removed *via* rotary evaporation, and the remaining charged 1-/2- product mixture was dissolved in 5 mL 90/10 ethanol/acetonitrile, FeCl₃·6H₂O (0.3 g, 1.11 mmol) was added and the mixture was left to stir for 12-24 h. Following oxidation, the solvent mixture was removed *via* rotary evaporation, and an orange band containing the neutral product was separated from the FeCl₃·6H₂O through a slurry-packed silica gel column with CH₂Cl₂. The CH₂Cl₂ was removed *via* rotary evaporation and the final neutral product **6** was dried under high vacuum to obtain an isolated yield of 35.0 mg (43%). Compound **6** is a dark yellow-orange oil. ¹H NMR (400 MHz, CDCl₃): δ 5.79 (m, 12H, CH), 4.95 (m, 24H, CH₂CH), 4.02 (t, 24H, O-CH₂), 2.05 (q, 24H, (CH₂)₃CH₂CH₂CH), 1.56 (m, 24H, CH₂CH₂(CH₂)₃CH), 1.43 (m, 24H, (CH₂)₂CH₂(CH₂)₂CH). ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 139.1, 114.2, 70.1, 33.6, 31.7, 25.5. ¹¹B{¹H} NMR (128 MHz, CDCl₃): δ 41.6. HRMS (Orbitrap): *m/z* calculated for C₇₂H₁₃₂B₁₂O₁₂ (M), 1319.09 Da; found, 1319.10 Da.

Dodeca(11-undeceneoxy)-hypercloso-dodecaborane [7]

Note: this reaction should be performed with minimal exposure to light. TBA₂[**1**] (50.0 mg, 0.061 mmol) was added to a 10 mL glass microwave vial and transferred out of a nitrogen filled glovebox, opened to the air, and dissolved in 1 mL acetonitrile. *N,N*-diisopropylethylamine (0.2 mL, 1.15 mmol) and undec-10-enyl bromide (1.54 mL, 7.33 mmol) were added along with a stir bar, the vial was sealed with a PTFE/silicone cap, and the mixture was reacted at 140 °C with stirring in the microwave for 8 h. The volatiles were removed *via* rotary evaporation, the excess reagent was eluted through a slurry-packed silica gel column with hexanes, and the product

mixture yellow-orange and red-orange fractions were eluted with CH₂Cl₂ followed by a pink fraction with ethyl acetate. The CH₂Cl₂/ethyl acetate was removed *via* rotary evaporation, the remaining charged 1-/2- product mixture was dissolved in 9 mL 49/49/2 ethanol/CH₂Cl₂/acetonitrile, FeCl₃·6H₂O (0.5 g, 1.85 mmol) was added and the mixture was left to stir for 12-24 h. Following oxidation, the solvent mixture was removed *via* rotary evaporation, and a dark brown/black band containing the neutral product was separated from the FeCl₃·6H₂O through a slurry-packed silica gel column with CH₂Cl₂. The CH₂Cl₂ was removed *via* rotary evaporation and the final neutral product **7** was dried under high vacuum to obtain an isolated yield of 37.1 mg (28%). Compound **7** is a dark, brown/black oil. ¹H NMR (400 MHz, CD₂Cl₂): δ 5.81 (m, 12H, CH), 4.98 (m, 12H, *trans*-CH₂CH), 4.91 (m, 12H, *cis*-CH₂CH), 4.01 (t, 24H, O-CH₂), 2.03 (m, 24H, O-CH₂CH₂), 1.54 (m, 24H, O-(CH₂)₂CH₂), 1.32 (m, 144H, O-(CH₂)₃(CH₂)₆CH₂CH. ¹³C{¹H} NMR (125 MHz, CD₂Cl₂): δ 139.3, 113.8, 70.4, 33.9, 32.3, 29.9, 29.7, 29.6, 29.3, 29.1, 26.2. ¹¹B{¹H} NMR (128 MHz, CD₂Cl₂): δ 41.5. *m/z* calculated for C₁₃₂H₂₅₂B₁₂O₁₂ (M⁺), 2161.03 Da; due to solubility issues, the molecule was incompatible with our M.S. instrument.

Dodeca(ethylbutyratoxy)-hypercloso-dodecaborane [8]

TBA₂[**1**] (100.0 mg, 0.122 mmol) was added to a 10 mL glass microwave vial and transferred out of a nitrogen filled glovebox, opened to the air, and dissolved in 2 mL acetonitrile. *N,N*-diisopropylethylamine (0.4 mL, 2.30 mmol) and ethyl 4-bromobutyrate (1.08 mL, 7.55 mmol) were added along with a stir bar, the vial was sealed with a PTFE/silicone cap, and the mixture was heated at 140 °C with stirring in the microwave for 1.5 h. The volatiles were removed *via* rotary evaporation, and the reaction mixture was eluted through Sephadex LH-20 size exclusion column with methanol, with the pink fraction containing the desired 1-/2- product collected. The methanol was removed *via* rotary evaporation, and the charged 1-/2- product mixture was oxidized

by eluting through a slurry-packed silica gel column with 90/10 CH₂Cl₂/ethanol, collecting the orange fraction. Following oxidation, the solvent mixture was removed *via* rotary evaporation, and the product was purified by eluting through a short (15 cm) silica gel column slurry-packed with 1:1 hexanes/ethyl ether, with ~50 mL 1:1 hexanes/ethyl ether followed by ~50 mL ethyl ether eluting an orange band. The volatiles were removed *via* rotary evaporation, and the final neutral product **8** was dried under high vacuum to obtain an isolated yield of 14.0 mg (7%). *Note: ~8% additional 1-/2- product was collected from the first silica column, and by repeating the oxidation step with the charged 1-/2- mixture additional 8 can be isolated.* Compound **8** is an orange solid. ¹H NMR (500 MHz, CDCl₃): δ 4.11 (q, 24H, O-CH₂), 4.03 (t, 24H, COOCH₂), 2.34 (t, 24H, CH₂COO), 1.87 (m, 24H, O-CH₂CH₂), 1.24 (t, 36H, CH₃). ¹³C{¹H} NMR (125 MHz, CDCl₃): δ, 173.3, 69.7, 60.3, 30.9, 27.4, 14.2. ¹¹B{¹H} NMR (128 MHz, CDCl₃): δ 41.9. HRMS (Orbitrap): *m/z* calculated for C₇₂H₁₃₂B₁₂O₃₆ (M⁻), 1702.97 Da; found, 1702.97 Da.

Dodeca(4-methylbenzyloxy)-hypercloso-dodecaborane [9]

TBA₂[**1**] (48.0 mg, 0.059 mmol) was added to a 10 mL glass microwave vial and transferred out of a nitrogen filled glovebox, opened to the air, and dissolved in 1 mL acetonitrile. *N,N*-diisopropylethylamine (0.2 mL, 1.15 mmol) and 4-methylbenzyl bromide (1.362 g, 7.36 mmol) were added along with a stir bar, the vial was sealed with a PTFE/silicone cap, and the mixture was heated at 140 °C with stirring in the microwave for 5 min. The volatiles were removed *via* rotary evaporation, and the excess reagent was eluted through a slurry-packed silica gel column with 90/10 hexanes/ethyl acetate, and the pink/purple product mixture was eluted with acetonitrile. The volatiles were removed *via* rotary evaporation, the remaining charged 1-/2- product mixture was dissolved in 5 mL 90/10 ethanol/acetonitrile, FeCl₃·6H₂O (0.3 g, 1.11 mmol) was added and the mixture was left to stir for 12-24 h. Following oxidation, the solvent mixture was removed *via*

rotary evaporation, and a dark orange band containing the neutral product was separated from the $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ through a slurry-packed silica gel column with CH_2Cl_2 . The CH_2Cl_2 was removed via rotary evaporation and the final neutral product **9** was dried under high vacuum to obtain an isolated yield of 30.9 mg (33%). Compound **9** is a brown/orange viscous oil. ^1H NMR (400 MHz, CDCl_3): δ 6.98 (m, 48H, C_6H_4), 5.17 (s, 24H, CH_2), 2.31 (s, 36H, CH_3). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 137.9, 136.4, 128.7, 127.3, 72.8, 21.2. $^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3): δ 41.6. HRMS (Orbitrap): m/z calculated for $\text{C}_{96}\text{H}_{108}\text{B}_{12}\text{O}_{12}$ (M⁺), 1582.90 Da; found, 1582.91 Da.

Dodeca(4-bromobenzyloxy)-hypercloso-dodecaborane [10]

TBA₂[**1**] (50.0 mg, 0.061 mmol) was added to a 10 mL glass microwave vial and transferred out of a nitrogen filled glovebox, opened to the air, and dissolved in 1 mL acetonitrile. *N,N*-diisopropylethylamine (0.2 mL, 1.15 mmol) and 4-bromobenzyl bromide (1.358 g, 7.36 mmol) were added along with a stir bar, the vial was sealed with a PTFE/silicone cap, and the mixture was heated at 140 °C with stirring in the microwave for 30 min. The volatiles were removed *via* rotary evaporation, and the excess reagent was eluted through a slurry-packed silica gel column with hexanes, and the red-pink product mixture was eluted with CH_2Cl_2 followed by ethyl acetate. The volatiles were removed *via* rotary evaporation, the remaining charged 1-/2-product mixture was dissolved in 5 mL 90/10 ethanol/acetonitrile, $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ (0.3 g, 1.11 mmol) was added and the mixture was left to stir for 12-24 h. Following oxidation, the solvent mixture was removed *via* rotary evaporation, and a dark orange band containing the neutral product was separated from the $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ through a slurry-packed silica gel column with CH_2Cl_2 . The CH_2Cl_2 was removed via rotary evaporation and the final neutral product **10** was dried under high

vacuum to obtain an isolated yield of 62.5 mg (43%). Compound **10** is a dark-orange solid. ^1H NMR (400 MHz, CDCl_3): δ 7.33 (d, 24H, *m*- C_6H_4), 6.86 (d, 24H, *o*- C_6H_4), 5.07 (s, 24H, O- CH_2). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ , 138.5, 131.6, 128.5, 121.6, 72.6. $^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3): δ 41.4. HRMS (Orbitrap): *m/z* calculated for $\text{C}_{84}\text{H}_{72}\text{B}_{12}\text{Br}_{12}\text{O}_{12}$ (M^-), 2361.63 Da; found, 2361.63 Da.

Dodeca(4-trifluoromethylbenzyloxy)-hypercloso-dodecaborane [11]

TBA₂[**1**] (50.0 mg, 0.061 mmol) was added to a 10 mL glass microwave vial and transferred out of a nitrogen filled glovebox, opened to the air, and dissolved in 1 mL acetonitrile. *N,N*-diisopropylethylamine (0.2 mL, 1.15 mmol) and 4-trifluoromethylbenzyl bromide (1.765 g, 7.5 mmol) were added along with a stir bar, the vial was sealed with a PTFE/silicone cap, and the mixture was reacted at 140 °C with stirring in the microwave for 30 min. The volatiles were removed *via* rotary evaporation, and the excess reagent was eluted through a slurry-packed silica gel column with hexanes, and the pink/purple product mixture was eluted with acetonitrile. The acetonitrile was removed *via* rotary evaporation, the remaining charged 1-/2- product mixture was dissolved in 5 mL of 90/10 ethanol/acetonitrile, $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ (0.4 g, 1.48 mmol) was added and the mixture was left to stir for 12-24 h. Following oxidation, the solvent mixture was removed *via* rotary evaporation, and an orange band containing the neutral product was separated from the $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ through a slurry-packed silica gel column with CH_2Cl_2 . The CH_2Cl_2 was removed *via* rotary evaporation and the final neutral product **11** was dried under high vacuum to obtain an isolated yield of 89.6 mg (66%). Compound **11** is a red-orange solid. ^1H NMR (500 MHz, CDCl_3): δ 7.38 - 7.48 (m, 24H, *m*- C_6H_4), 7.06 - 7.15 (m, 24H, *o*- C_6H_4), 5.27 (s, 24H, O- CH_2). $^{13}\text{C}\{^1\text{H}\}$

NMR (125 MHz, CDCl₃): δ 143.0, 130.6, 126.6, 125.7, 125.0, 122.8, 72.9. ¹¹B{¹H} NMR (128 MHz, CDCl₃): δ 41.8. ¹⁹F NMR (376 MHz, CDCl₃): δ -62.76 (s, 36F). HRMS (Orbitrap): *m/z* calculated for C₉H₇B₁₂F₃₆O₁₂ (M⁻), 2231.57 Da; found, 2231.56 Da. Crystallized from CDCl₃ and pentane at room temperature for 1 week to obtain a single crystal for X-ray diffraction analysis.

Dodeca(4-nitrobenzyloxy)-hypocloso-dodecaborane [12]¹⁻

TBA₂[1] (50.0 mg, 0.061 mmol) was added to a 10 mL glass microwave vial and transferred out of a nitrogen filled glovebox, opened to the air, and dissolved in 1 mL acetonitrile. *N,N*-diisopropylethylamine (0.2 mL, 1.15 mmol) and 4-nitrobenzyl bromide (1.585 g, 7.34 mmol) were added along with a stir bar, the vial was sealed with a PTFE/silicone cap, and the mixture was heated at 140 °C with stirring in the microwave for 30 min. The volatiles were removed *via* rotary evaporation, and the excess reagent was eluted through a slurry-packed silica gel column with 50/50 hexanes/CH₂Cl₂, and the orange product mixture was eluted with acetonitrile. The volatiles were removed *via* rotary evaporation, the remaining charged 1-/2- product mixture was dissolved in 5 mL 90/10 ethanol/acetonitrile, FeCl₃·6H₂O (0.3 g, 1.11 mmol) was added and the mixture was left to stir for 12-24 h. Following oxidation, the solvent mixture was removed *via* rotary evaporation, and the product mixture was washed with 150 – 200 mL ethanol in a glass fritted 30 mL filter funnel to remove the remaining FeCl₃·6H₂O. The remaining orange radical 1-product TBA[12]¹⁻ was removed from the funnel and dried under high vacuum to obtain an isolated yield of 89.0 mg (66%). Compound TBA[12]¹⁻ is an orange solid. ¹H NMR (400 MHz, CDCl₃): δ 8.49 - 7.41 (m, 48H, C₆H₄). *Note: The CH₂ signal is masked and all other peaks are quite broad due to the paramagnetic radical state of the molecule.* ¹³C{¹H} NMR (125 MHz, CDCl₃): δ 191.26, 147.47, 140.59, 130.55, 124.24. *Note: The CH₂ signal is masked and all other peaks are quite*

broad due to the paramagnetic radical state of the molecule. No resonances are visible by ^{11}B NMR due to paramagnetic broadening (a trace resonance at 20.4 ppm is indicative of borates, which result from decomposition). HRMS (Orbitrap): m/z calculated for $\text{C}_{84}\text{H}_{72}\text{B}_{12}\text{N}_{12}\text{O}_{36}$ (M^+), 1954.54 Da; found, 1954.54 Da.

Dodeca(3,5-bis(trifluoromethyl)benzyloxy)-hypocloso-dodecaborane [13] $^{1-}$

TBA $_2$ [1] (99.0 mg, 0.121 mmol) was added to a 10 mL glass microwave vial and transferred out of a nitrogen filled glovebox, opened to the air, and dissolved in 2 mL acetonitrile. *N,N*-diisopropylethylamine (0.4 mL, 2.30 mmol) and 3,5-bis(trifluoromethyl)benzyl bromide (2.68 mL, 14.6 mmol) were added along with a stir bar, the vial was sealed with a PTFE/silicone cap, and the mixture was heated at 140 °C with stirring in the microwave for 30 min. The volatiles were removed *via* rotary evaporation, and the excess reagent was eluted through a slurry-packed silica gel column with 65/35 hexanes/ethyl acetate, and the colorless/very light pink product mixture was eluted with CH_2Cl_2 . After removal of the CH_2Cl_2 *via* rotary evaporation, compound TBA $_2$ [13] $^{2-}$, a clear, colorless solid, was dried under high vacuum to obtain an isolated yield of 313.6 mg (73%). After spectroscopic characterization, the dianionic TBA $_2$ [13] $^{2-}$ was dissolved in 5 mL 90/10 ethanol/acetonitrile, 0.3 g (1.11 mmol) $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ was added and the mixture was left to stir for 12-24 h. Following oxidation, the solvent mixture was removed *via* rotary evaporation, and a red-purple band containing [13] $^{1-}$ was separated from the $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ through a slurry-packed silica gel column with CH_2Cl_2 . The CH_2Cl_2 was removed *via* rotary evaporation and the final isolated radical [13] $^{1-}$ was dried under high vacuum to obtain an isolated yield of 226.4 mg (56%). Compound [13] $^{1-}$ is a red-purple solid. ^1H NMR (500 MHz, CDCl_3): δ 7.40 – 8.74 (m, 36H, C_6H_3), 3.13 (m, 8H, N- CH_2), 1.65 (m, 8H, N- CH_2 - CH_2), 1.47 (m, 8H, N-(CH_2) $_2$ - CH_2), 1.05 (m, 12H, N-(CH_2) $_3$ - CH_3). *Note: The CH_2 signal for the cluster is masked and all other peaks*

are quite broad due to the paramagnetic radical state of the molecule. $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 189.0, 132.7, 132.5, 129.4, 124.6, 122.5, 121.0, 68.0, 59.2, 59.2, 59.1, 30.9, 25.6, 23.8, 19.7, 19.7, 13.4. No resonances are visible by ^{11}B NMR, due to paramagnetic broadening. ^{19}F NMR (376 MHz, CDCl_3): δ -63.22 (s, 72F). HRMS (Orbitrap): m/z calculated for $\text{C}_{108}\text{H}_{60}\text{B}_{12}\text{F}_{72}\text{O}_{12}$ (M^-), 3047.42 Da; found, 1523.71 ($z=2$) Da. Crystallized from CDCl_3 and pentane at room temperature for 1 week to obtain a single crystal for X-ray diffraction analysis.

Benzyloxy undeca(ethoxy)-hypercloso-dodecaborane [14]

TBA₂[1] (100.0 mg, 0.122 mmol) was added to a 10 mL glass microwave vial and transferred out of a nitrogen filled glovebox, opened to the air, and dissolved in 2 mL acetonitrile. *N,N*-diisopropylethylamine (0.4 mL, 2.30 mmol), benzyl bromide (0.0204 g, 0.119 mmol) and bromoethane (0.8055g, 7.39 mmol) were added along with a stir bar, the vial was sealed with a PTFE/silicone cap, and the mixture was heated at 140 °C with stirring in the microwave for 30 min. The volatiles and excess bromoethane were removed *via* rotary evaporation, the remaining charged 1-/2- product mixture was dissolved in 5 mL 90/10 ethanol/acetonitrile, $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ (0.3 g, 1.11 mmol) was added and the mixture was left to stir for 12-24 h. Following oxidation, the solvent mixture was removed *via* rotary evaporation, and the product mixture was separated from the $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ with CH_2Cl_2 . The CH_2Cl_2 was removed *via* rotary evaporation, and the product mixture was loaded onto a long (30 – 35 cm) silica gel column slurry-packed with 80/20 CH_2Cl_2 /hexanes, and the products were separated by eluting fractions with 80/20 CH_2Cl_2 /hexanes. The first orange band eluted contained randomly di-substituted benzyl₂ethyl₁₀ species, followed by an orange band with the neutral closomer **14**, with extra **4** in a third yellow-orange band eluting last. *Note: The fractions overlap, and due to the similar colors of the different products, thin layer chromatography (TLC) with 80/20 CH_2Cl_2 /hexanes was performed on the fractions near the band*

edges to determine which fractions contained a mixture of products. The fractions containing only a single species according to TLC that eluted after the di-substituted product and prior to the pure **4** bands were combined. The volatiles were removed *via* rotary evaporation, and the final neutral product **14** was dried under high vacuum to obtain an isolated yield of 15.7 mg (18%). Compound **14** is a yellow-orange solid. ^1H NMR (400 MHz, CDCl_3): δ 7.30 (m, 5H, C_6H_5), 5.07 (m, 2H, $\text{CH}_2\text{C}_6\text{H}_5$), 4.10 (m, 22H, CH_2CH_3), 1.22 (m, 33H, CH_2CH_3). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 141.2, 128.0, 126.6, 71.6, 66.8, 17.7. ^{11}B NMR (128 MHz, CDCl_3): δ 39.0, 35.2. HRMS (Orbitrap): m/z calculated for $\text{C}_{29}\text{H}_{62}\text{B}_{12}\text{O}_{12}$ (M^+), 732.54 Da; found, 732.54 Da.

Chapter 3: Additional B₁₂(OR)₁₂ Derivatives and Future Work

The B₁₂(OR)₁₂ clusters synthesized in this work have myriad future applications due to their unique properties including expanded redox potential windows for these classes of clusters and post-functionalization growth of the cluster. Targeted development of novel clusters with desired functional groups has already yielded promising initial results in two of these research directions in studies led by collaborators, with efforts on other projects ongoing. The principle focus of all immediate future work is to investigate the unique multivalent interactions possible with the perfunctionalized clusters, and the many potential applications these characteristics enable. Each of these avenues will require future studies to elucidate the full potential of the novel clusters synthesized in this study, yet early results have also been promising. The further study of these applications requiring unique, multivalent clusters with diverse functional groups is significantly accelerated due to the microwave-assisted synthetic methods developed in this work allowing for targeted synthesis of new clusters specifically designed for their intended usage scenarios. The first example of this directed synthesis of novel B₁₂(OR)₁₂ clusters for immediate application is demonstrated by the formation of two “scaffold” clusters for organomimetic atomically precise nanoparticles.

Two additional B₁₂(OR)₁₂ species were synthesized utilizing the general microwave-based methods to be used for multivalent organomimetic nanoparticle synthesis. After achieving successful perfunctionalization with highly electron-withdrawing substituents (as described in the previous chapter) for multiple compounds, the inclusion of perfluorinated groups was investigated. These groups would allow facile post-functionalization *via* established *para*-fluorine S_NAr “click” chemistry.⁵⁶ Utilizing identical conditions to the benzyl species [2], perfunctionalization of the cluster with 2,3,4,5,6-pentafluorobenzyl groups was accomplished by simply replacing the aryl

halide (benzyl bromide) with commercially available 2,3,4,5,6-pentafluorobenzyl bromide. This new species (Figure 9, **15**) was intended for use to serve as a well-defined rigid core molecular scaffold from which to construct atomically precise organomimetic nanoparticles *via* the aforementioned facile post-functionalization. Initial trials by Elaine A. Qian and co-workers (unpublished work) with small thiol-containing molecules yielded promising results, yet larger groups were slow to fully react with all 12 sites on the cluster, or failed to react entirely. The most likely cause of this decreased reactivity was steric hindrance due to the bulky perfluorinated benzyl rings preventing close approach of the larger thiol-containing molecules. To remedy this issue, yet another novel cluster (Figure 9, **16**) was synthesized featuring a phenyl “spacer” to alleviate the steric hindrance and enhance the speed of the S_NAr reaction.

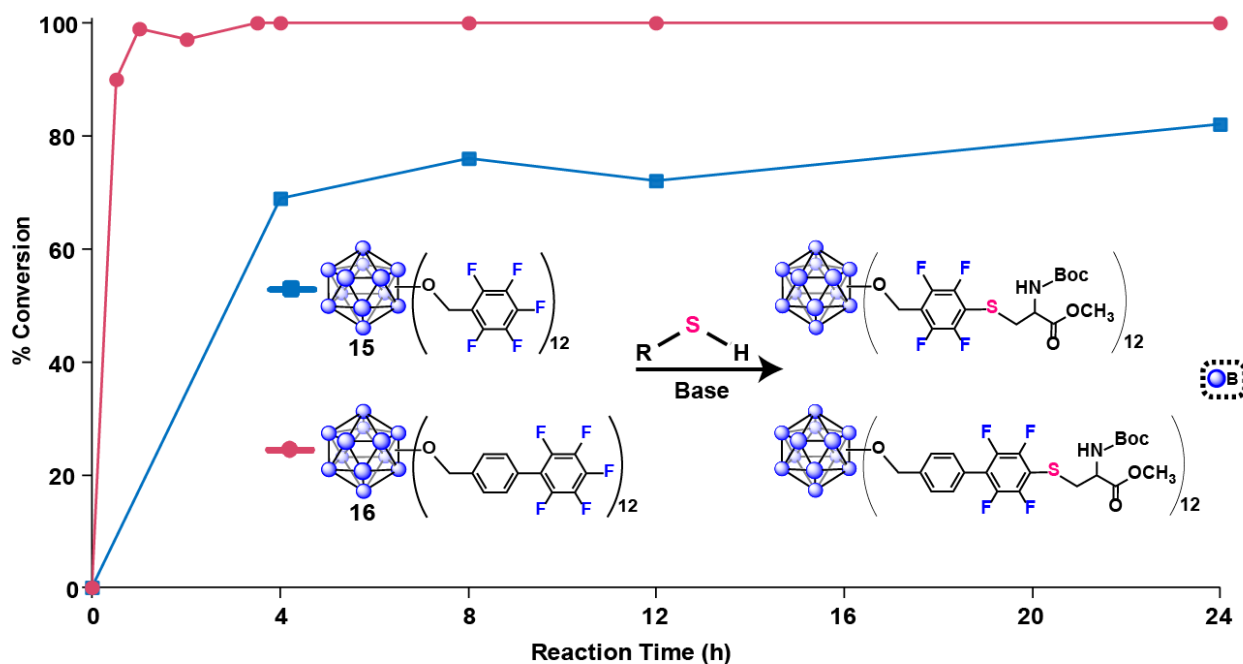


Figure 9. Compounds **15** and **16**; Reaction duration and conversion for *para*-fluorine S_NAr reactions of Boc-cysteine with compounds **15** and **16**.

To prepare compound **16**, the brominated precursor reagent $Br-CH_2C_6H_4C_6F_5$ was obtained *via* reduction of 4-(pentafluorophenyl)benzaldehyde with $NaBH_4$ in EtOH/THF (1:2) to form the

benzyl alcohol and subsequent treatment with PBr₃ to obtain the desired benzyl bromide (Figure 10).

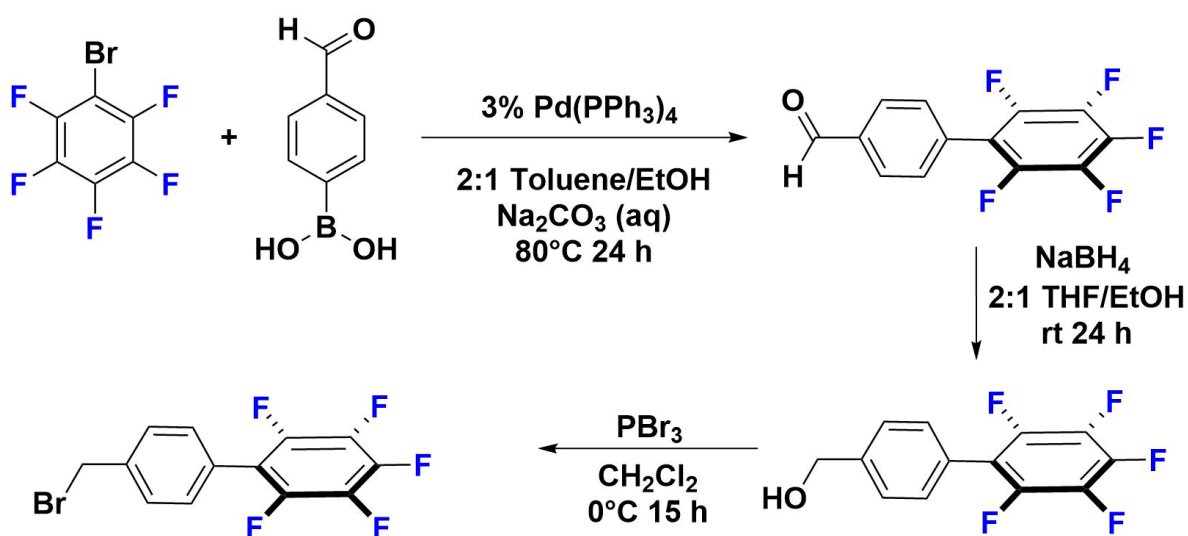


Figure 10. Synthesis of the Br-CH₂C₆H₄C₆F₅ reagent precursor for preparation of compound **16**.

Compound **16** was synthesized using the general microwave procedure with the developed Br-CH₂C₆H₄C₆F₅ reagent, once again only requiring an increase in reaction duration (to 1 h) and slight alteration of the column chromatography gradients for purification. Reactions of **16** with thiols were significantly more efficient than the first generation **15**, as demonstrated by the time required to fully substitute all 12 vertices with a bulky thiol-containing functional group (Figure 9). Compound **16** also allowed for more rapid attachment of larger and longer substituents, including sugars and polyethylene glycol (PEG). The synthesis of new perfunctionalized clusters with diverse substituents to enable specific applications (e.g. *para*-fluorine/thiol S_NAr click chemistry) is indicative of the robust nature of the microwave technique as a tool for targeted cluster design. Further studies investigating additional clusters with branched or further extended phenyl chains with terminal perfluorinated rings will probe the limits of this technique, and post-functionalization with thiol-containing groups will allow formation of myriad organomimetic nanoparticles with diverse multivalent applications based on the nature of the thiol groups decorating the rigid core.

Another application for these clusters with promising initial results is photo-initiated polymerization of styrenes. During the attempted synthesis of a $B_{12}(OR)_{12}$ derivative containing a polymerizable vinyl group on the substituent, *in situ* polymerization occurred in the microwave vial forming an insoluble solid, which was later determined to be a heavily crosslinked species. This led us to question if other clusters could be utilized to deliberately polymerize common monomers *via* photoexcitation, since the perfunctionalized clusters absorb in the visible region once oxidized to their neutral state. Initial trials by Marco S. Messina and co-workers (unpublished work) employing the benzyl (**2**) and pentafluorobenzyl (**15**) clusters to polymerize a variety of styrene derivatives were successful, resulting in polymer formation with high yields and low polydispersity (Figure 11). Another route being explored is the use of alkene-containing clusters such as compounds **3**, **6**, and **7** to construct heavily crosslinked polymers by incorporating styrene or other monomers. Further investigation of photopolymerization utilizing more oxidizing clusters, a broader substrate scope, and elucidation of the mechanism are ongoing.

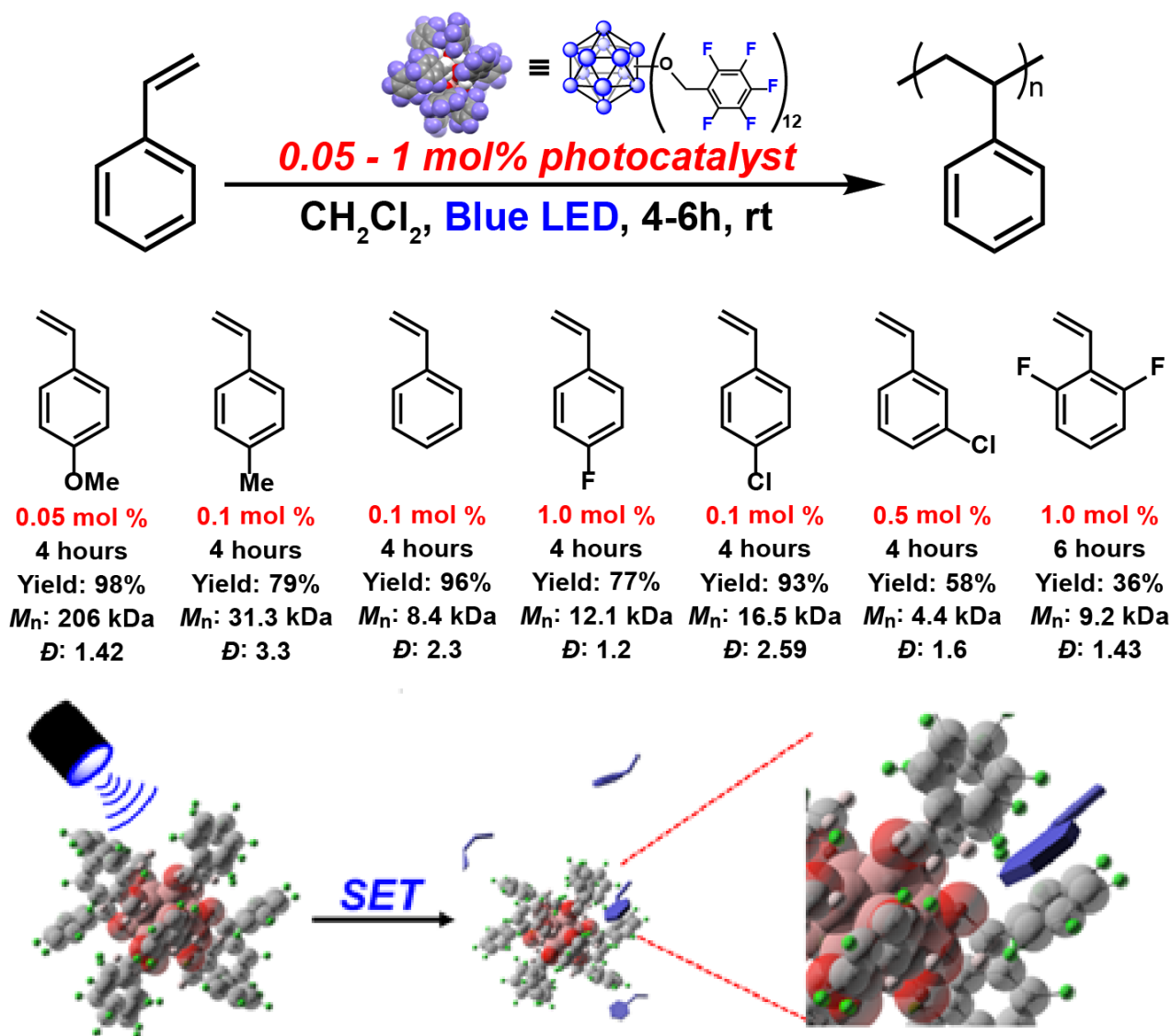


Figure 11. Photopolymerization of styrene derivatives utilizing compound **15**. Initial substrate scope with yields, average molecular weight, and polydispersity index.

These initial studies by collaborators have demonstrated the successful facile modifications to the cluster made possible by the developed microwave-assisted synthetic route yielding viable materials for the desired applications, and further studies for both applications are in progress. Immediate future work in new directions are based on exploiting the multivalent capabilities of the functionalized clusters for additional applications in several areas. One such application would be the formation of a tetrafluorido-containing cluster to probe the multivalent halogen bonding

interactions possible with such a cluster (Figure 12). This cluster would be synthesized using the general microwave synthetic procedure and utilizing a similar route to form the brominated reagent as shown in Figure 10. Recent studies have elucidated many possible applications incorporating XB halogen bonding, including the construction of molecular capsules⁵⁷, supramolecular rotors⁵⁸, and other molecular recognition scenarios⁵⁹. The unique multivalent interactions a cluster-based approach allow could lead to interesting self-assembly of three-dimensional complexes of the perfunctionalized tetrafluorido cluster and various XB acceptors.

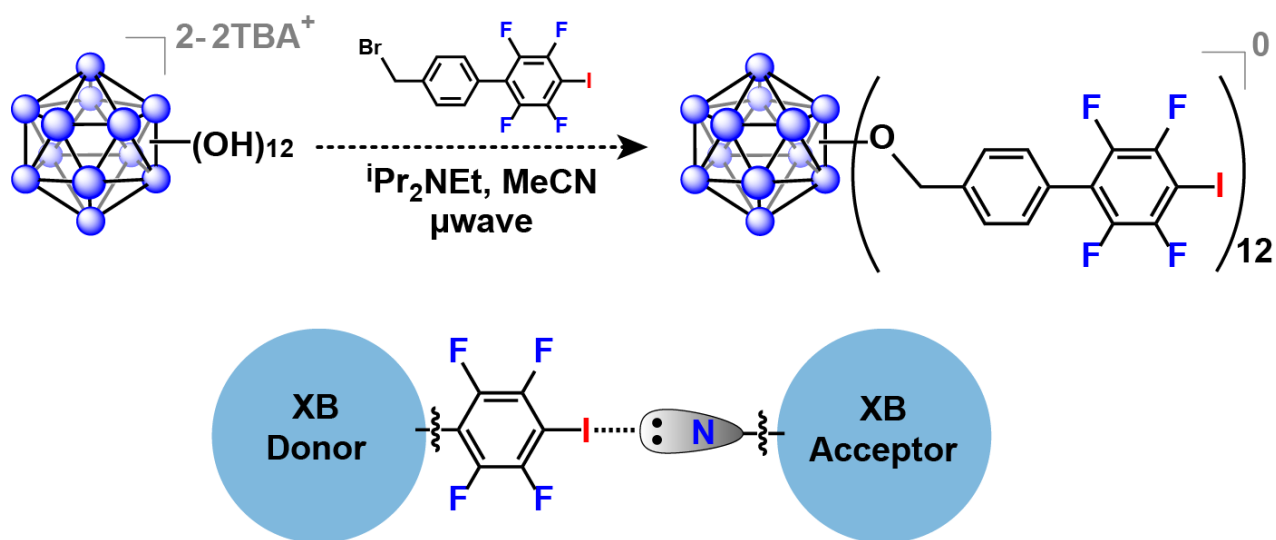


Figure 12. Synthesis of proposed tetrafluorido perfunctionalized cluster (top); scheme of XB donor-acceptor halogen bonding (bottom).

Formation of new classes of charge transfer complexes with various tetrathiafulvalene (TTF) compounds could be another use for the clusters synthesized in this work, as any phenyl ring-containing species such as **2**, **9-13**, and **15** could potentially form charge transfer complexes with TTF compounds, provided the redox potential of the neutral cluster is high enough to oxidize the TTF to TTF^+ . Initial NMR tube scale trials of this reaction with **15** have been promising, with

early NMR spectroscopic data indicating the formation of a $[\text{TTF}]^+[\mathbf{15}]^-$ ion pair. Full characterization of this reaction product and reactions with other TTF compounds is ongoing.

Energy storage is yet another application these novel boron clusters could be used for, in two possible battery architectures. Flow batteries have been considered for large-scale energy storage, due to their design accommodating the creation of very large batteries without the hazards or complexities associated with traditional batteries.⁶⁰ The design of a flow battery is fairly simple (Figure 13), yet given the right anodic and cathodic materials the potential energy density can be quite high, and recent studies have shown remarkable progress in advancing the performance of such devices.⁶⁰⁻⁶⁴ The molecules utilized in the anodic and cathodic chambers must be reversibly redox-active, stable compounds. The boron clusters synthesized in this work have exhibited redox potentials significantly higher than any previously reported $\text{B}_{12}(\text{OR})_{12}$ clusters, so by utilizing a pair of clusters with widely separated oxidation potentials a flow battery could be constructed. The performance of such a device could potentially be improved as more clusters are developed with even higher redox potentials.

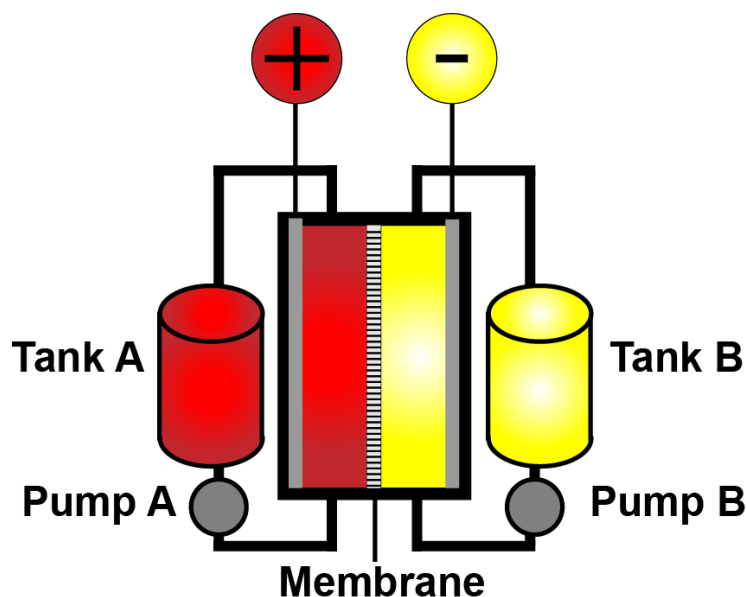


Figure 13. Flow battery general schematic, with anodic and cathodic chambers, pumps to circulate redox-active materials in solution past a membrane.

Flow batteries have the benefits of being easily scalable and simple to produce, but are not practical for use in smaller devices such as phones, tablets, and other personal electronics. Recent studies have demonstrated the successful formation of fully organic batteries utilizing nitroxyl radical polymers.⁶⁵ This class of battery could be extremely useful for many device applications where traditional rigid, heavy Li-ion or similar batteries are not suitable. Lightweight and flexible metal-free batteries could be invaluable for incorporation into many devices, though instability of the nitroxyl radical polymers utilized in current prototypes limits their immediate application. An alternative metal-free radical battery based on polymers containing redox-active stable radical icosahedral boron clusters (Figure 14) would potentially alleviate the stability issues due to the more robust nature of the boron clusters, while also allowing higher voltage performance due to the tunable wide redox potential window of the substituted $B_{12}(OR)_{12}$ clusters. A proposed first prototype based on this architecture would contain two different synthesized clusters with polymerizable functional groups. This would allow for photoinduced polymerization on the surface of indium tin oxide (ITO)-coated polyethylene terephthalate (PET) with a thin layer of electrolyte sandwiched in between the different cluster polymer layers (Figure 14). Two potential candidates among those synthesized so far would be the perfunctionalized allyl cluster (**3**) as one half of the cell, with a vinyl-containing derivative of the vertex-differentiated cluster **14** as the opposing half. This 4-vinylbenzyl cluster derivative (**17**, Figure 14) was successfully synthesized by employing the methods developed for **14** and exchanging the benzyl bromide for 4-vinylbenzyl bromide. Once again, this demonstrates the kind of facile modifications to form novel cluster derivatives with desired properties (in this case a single polymerizable terminal alkene) enabled by our microwave-assisted method. Both **3** and **17** are readily soluble in organic solvents, so *via* spin-coating or drop-casting a layer of each could be deposited onto PET substrates and

photopolymerized with UV light similarly to the preparation methods used in the aforementioned nitroxyl-based fully-organic metal-free battery.⁶⁵

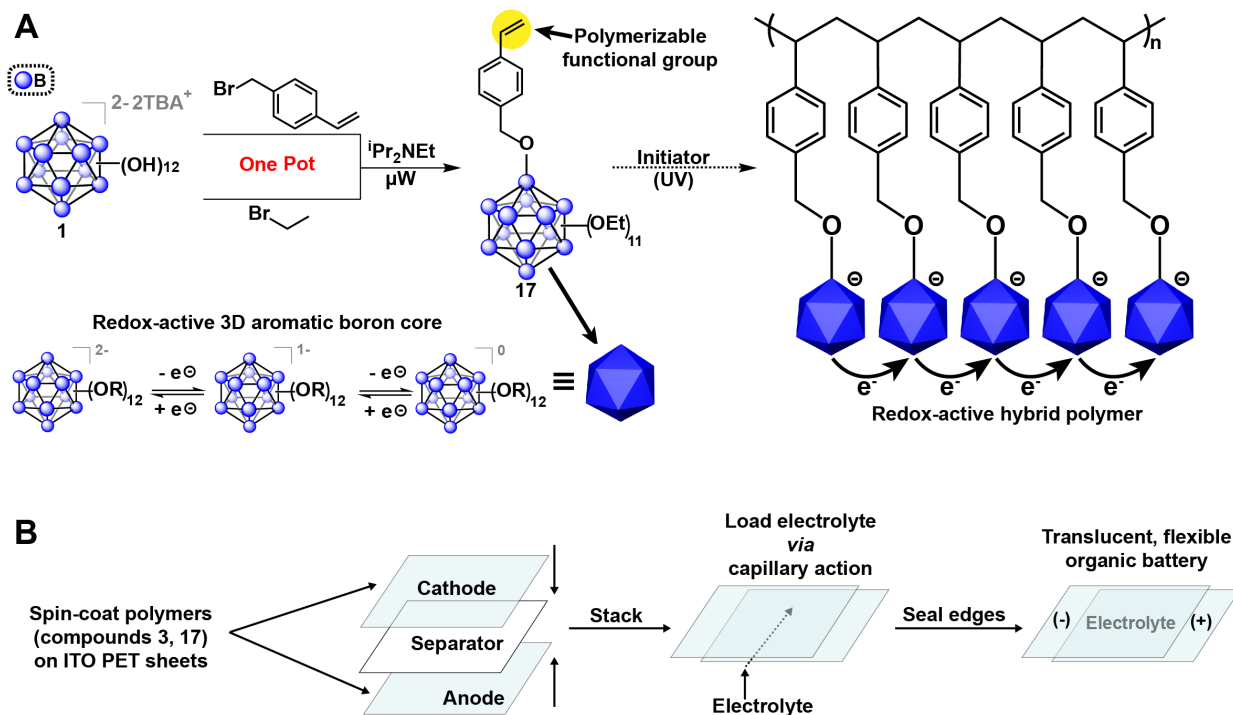


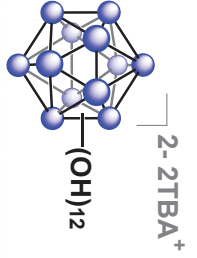
Figure 14. Scheme for synthesis of compound **17** (A) and proposed polymerization; schematic of a radical translucent, flexible organic battery from polymerized compounds **3** and **17**.

The boron cluster-containing polymers for the cathode and anode could both be rationally tuned for optimal performance, as the substituents attached to the boron core have been demonstrated to significantly modify the redox potential³⁹, and the backbone polymer could also be modified for enhanced performance.

Chapter 4: Conclusion

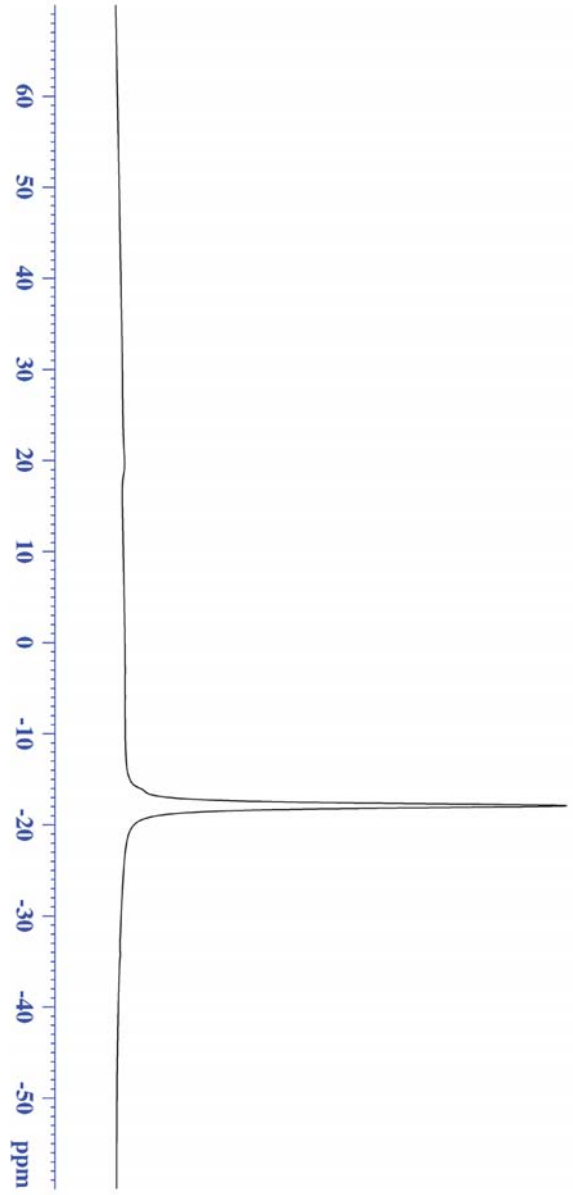
In conclusion, a new microwave-based method for synthesizing $B_{12}(OR)_{12}$ derivatives under ambient conditions was demonstrated. This method was successfully employed to synthesize several known derivatives with significantly reduced reaction times compared to previously reported protocols, while completely eliminating the need for handling in inert atmosphere. Additionally, the robust nature of the microwave-assisted synthesis method allowed the formation of novel $B_{12}(OR)_{12}$ species with diverse substituents for targeted applications. Hybrid vertex-differentiated $B_{12}(OR)_{11}(OR')$ clusters were also synthesized *via* a one-pot, single-step reaction, previously inaccessible except through lengthy multi-step protocols. There are many exciting applications for these newly developed $B_{12}(OR)_{12}$ clusters, with very promising initial results meriting further investigation. The targeted synthesis of clusters with specific functional groups for desired applications using the developed microwave method allows for significantly more rapid investigation of new research directions. The primary focus of the immediate future work utilizing this system will be exploring the possible multivalent interactions enabled by the three-dimensional nature of the perfunctionalized cluster.

Supplementary Spectra



¹¹B {¹H} NMR

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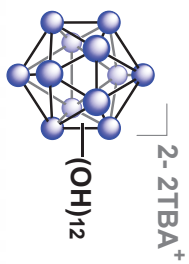
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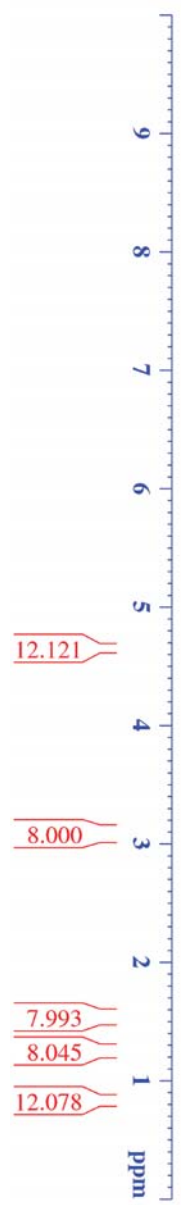
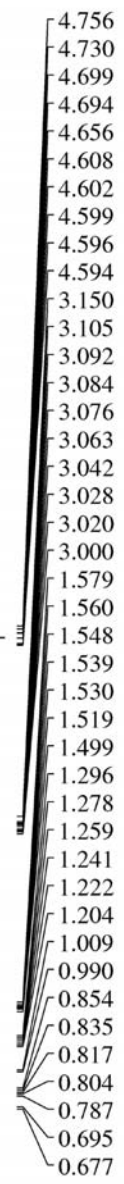
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¹H NMR



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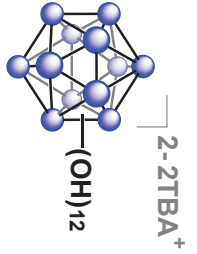
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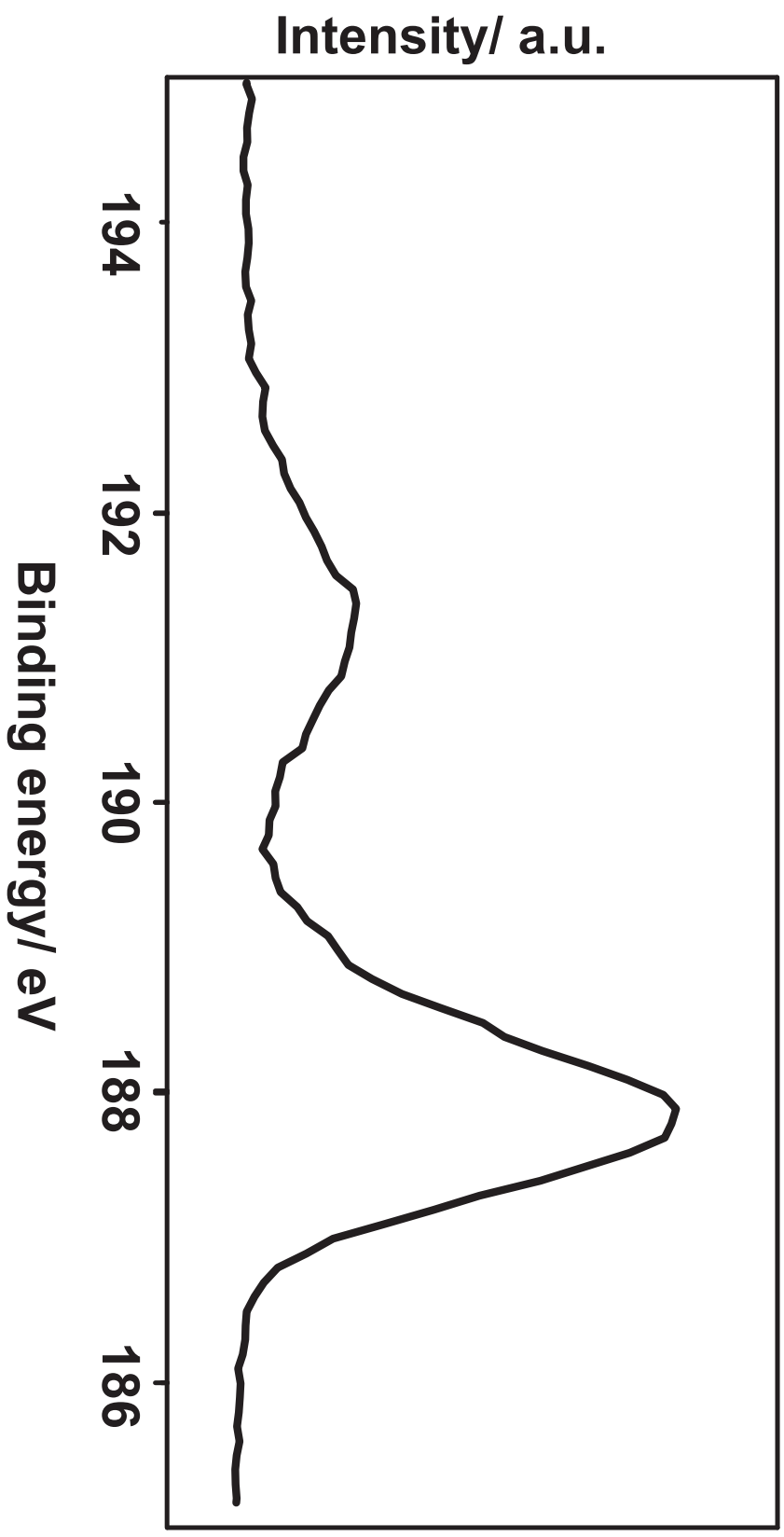
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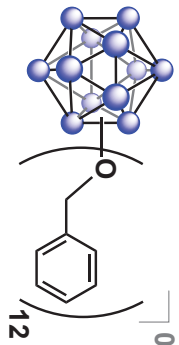




B 1s XPS



¹¹B {¹H} NMR



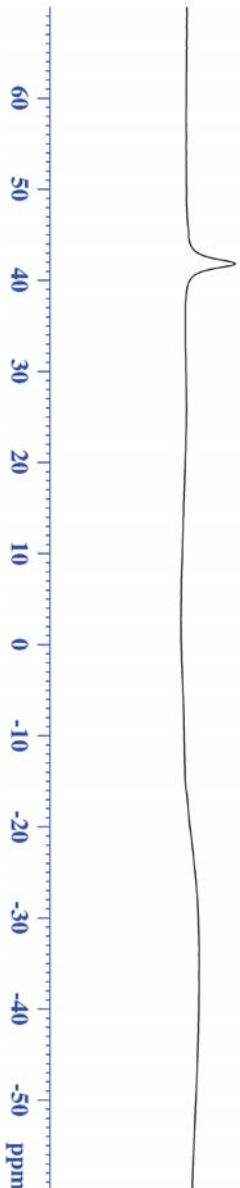
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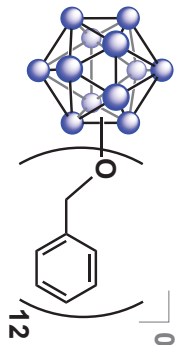
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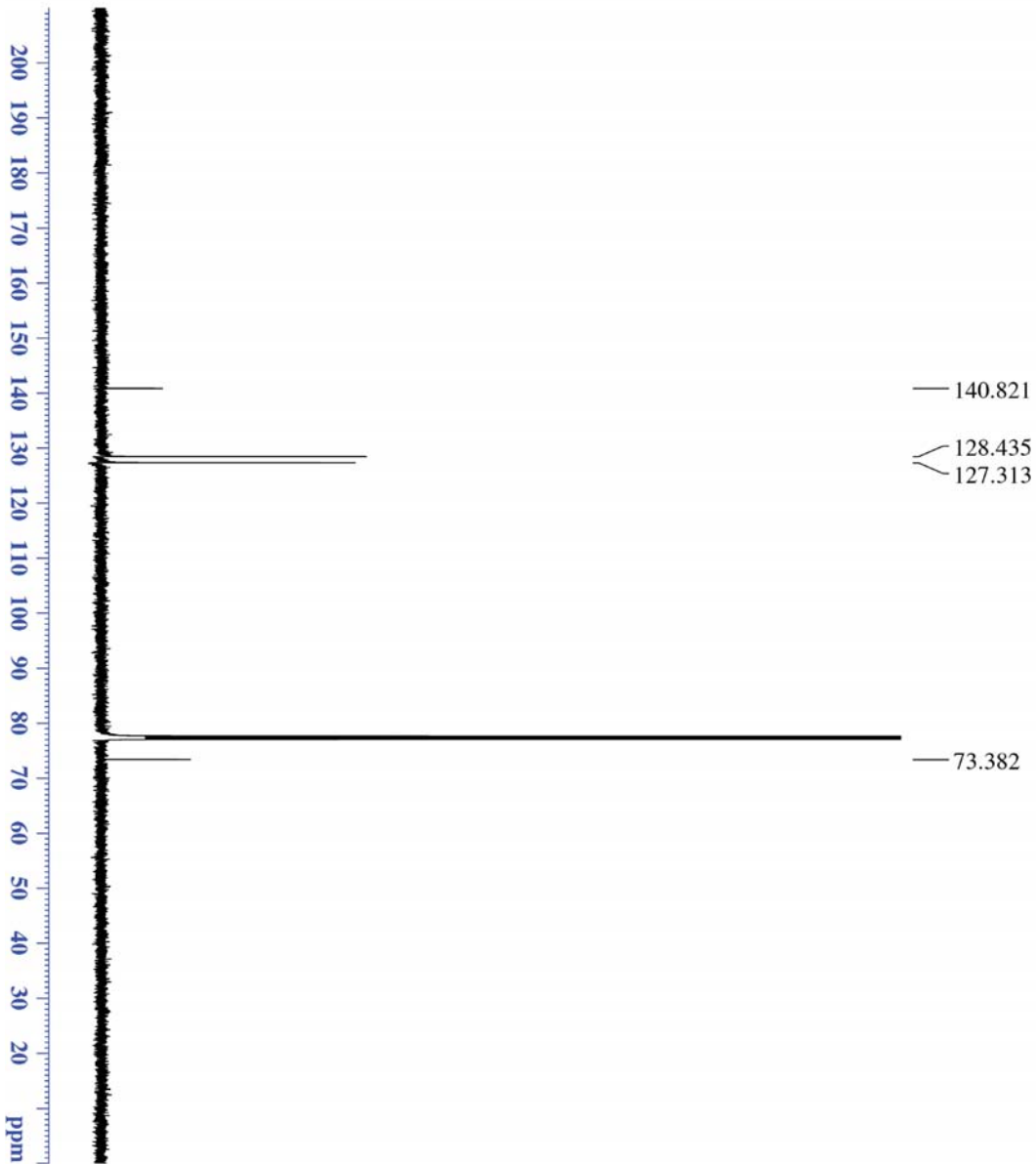
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¹³C NMR



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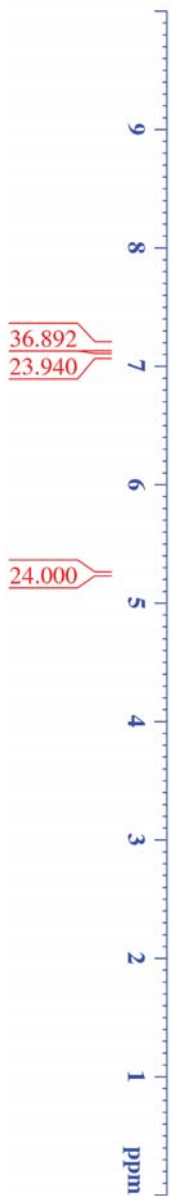
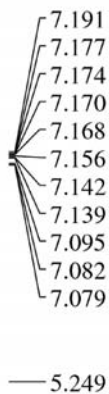
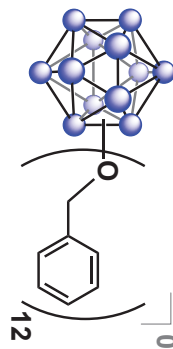
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¹H NMR

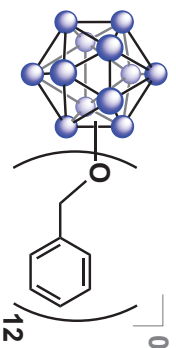


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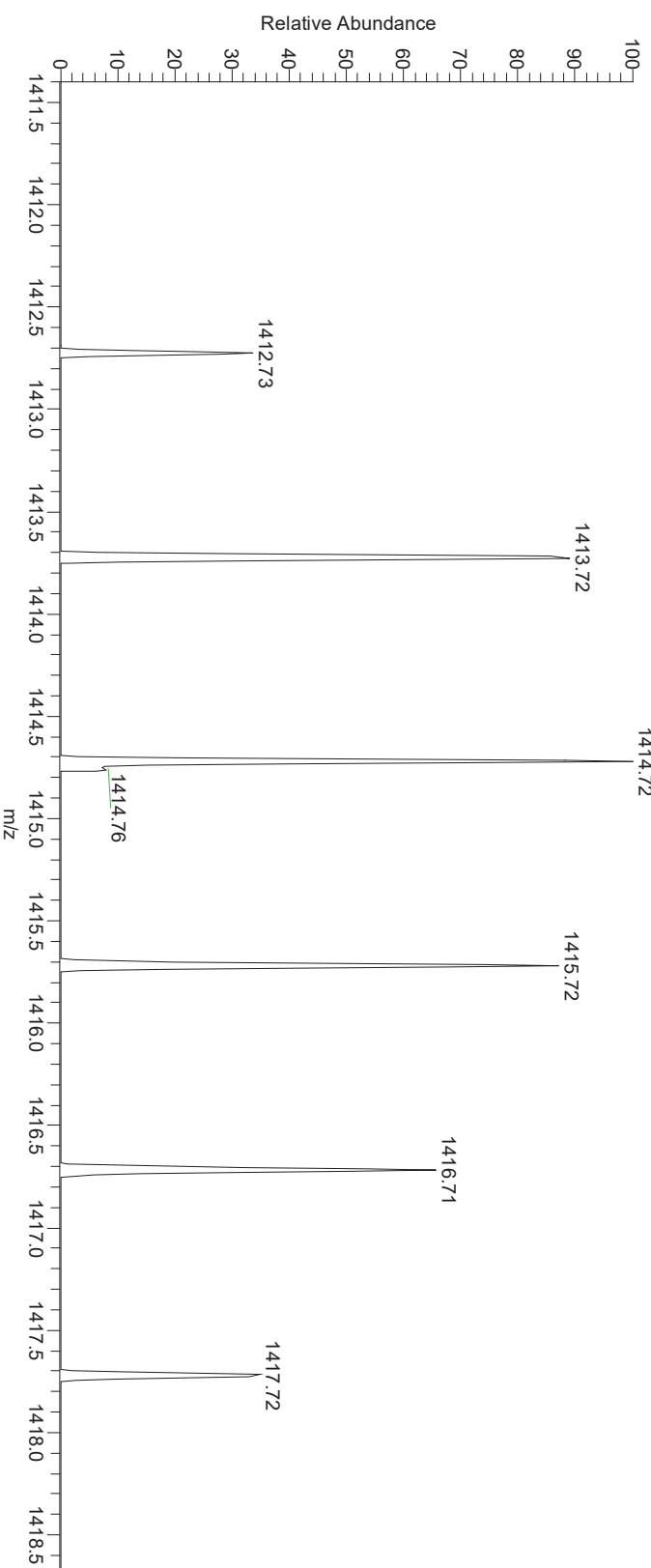
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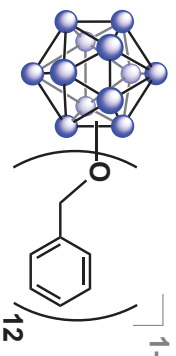
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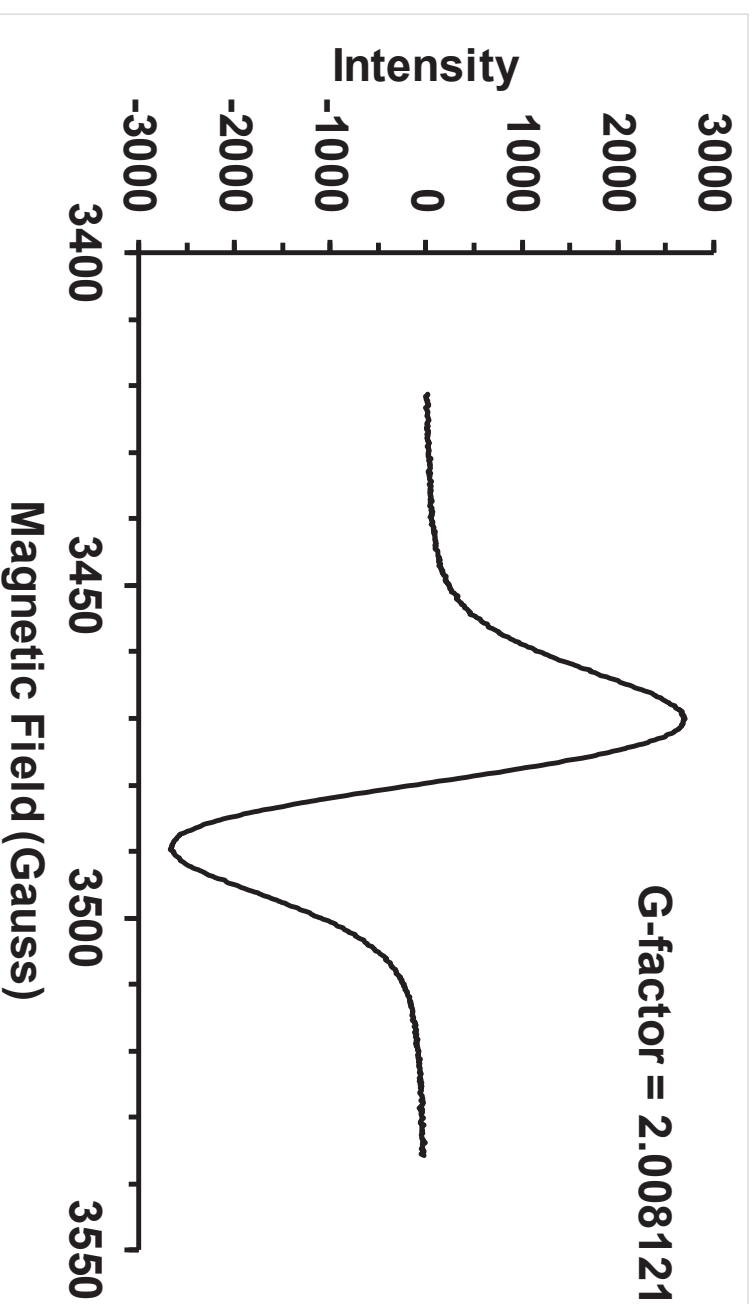
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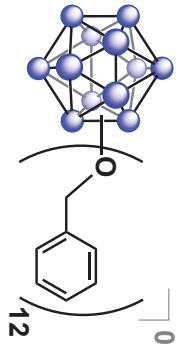
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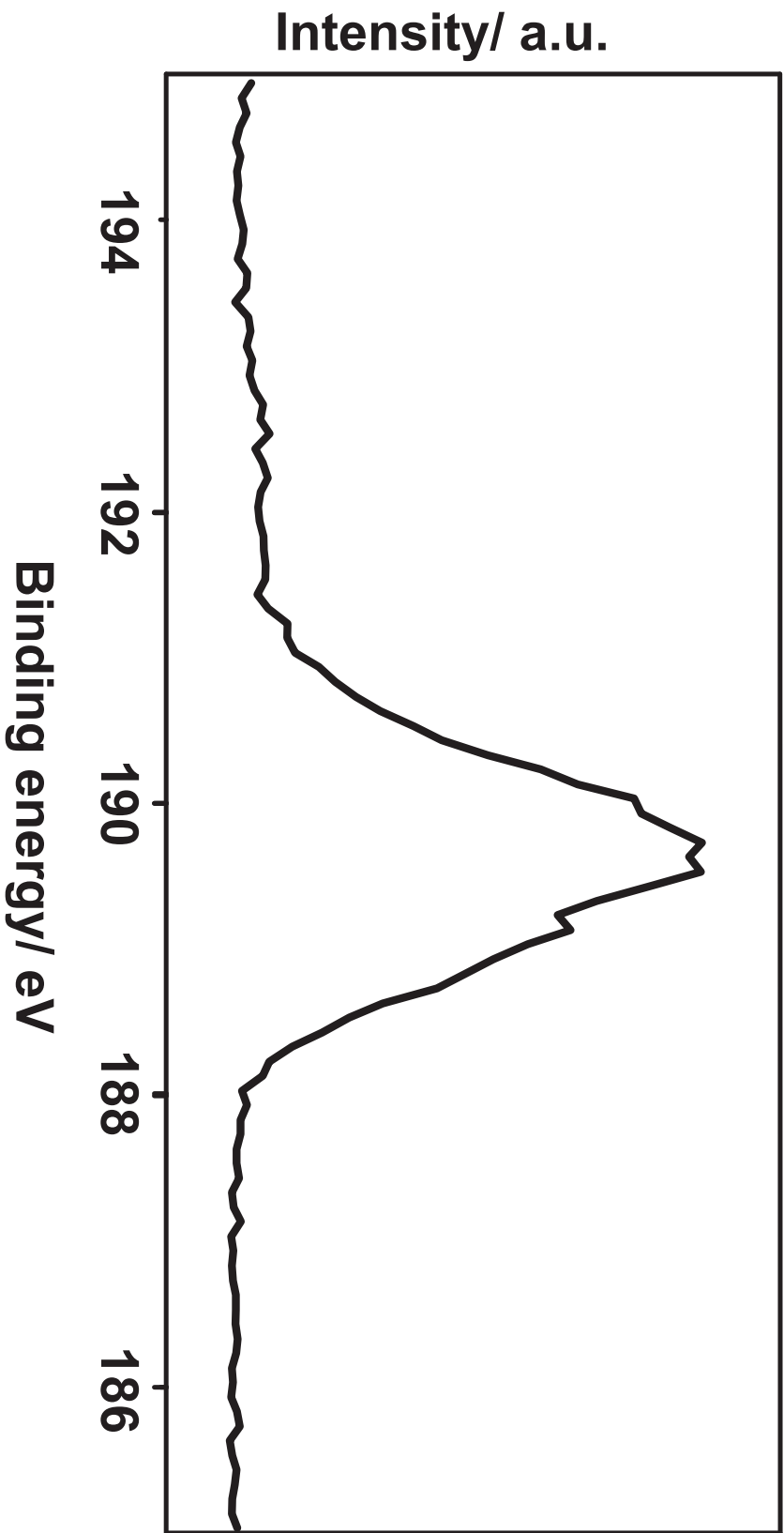
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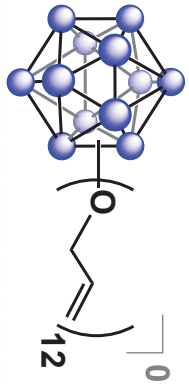
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B 1s XPS





¹¹B {¹H} NMR



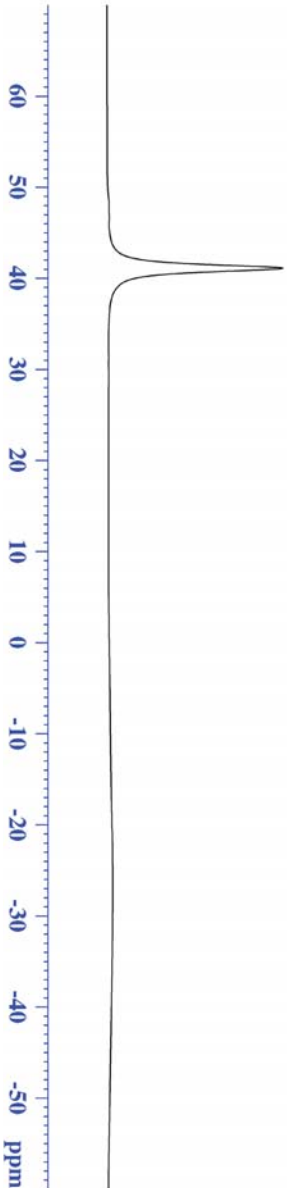
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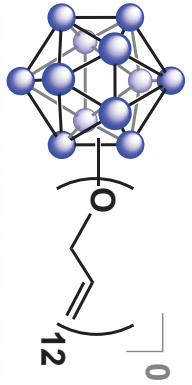
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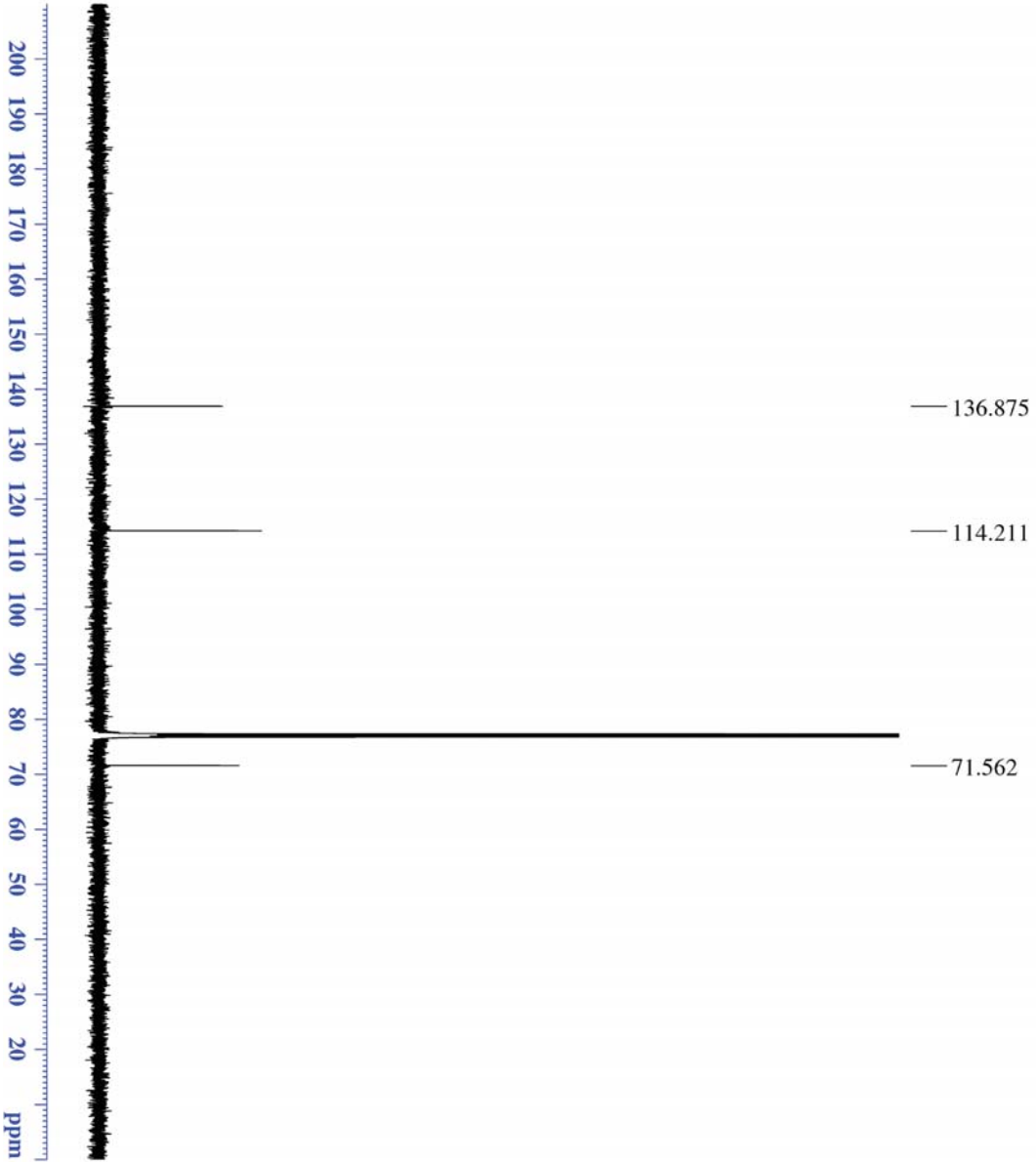
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¹³C NMR



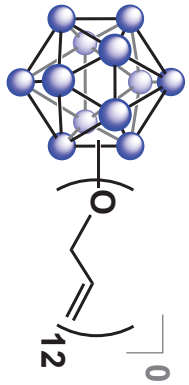
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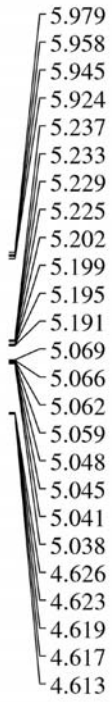
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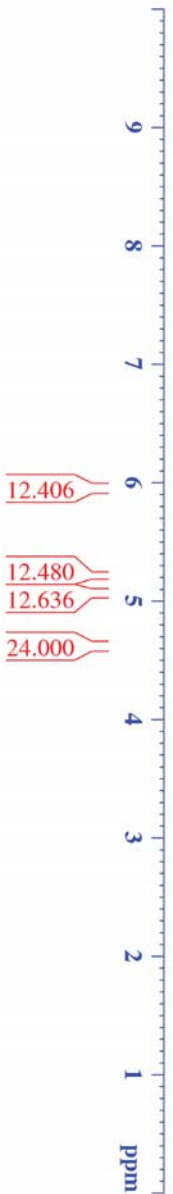
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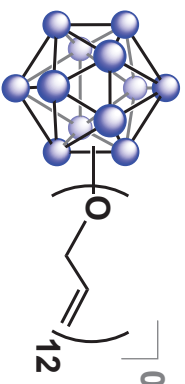


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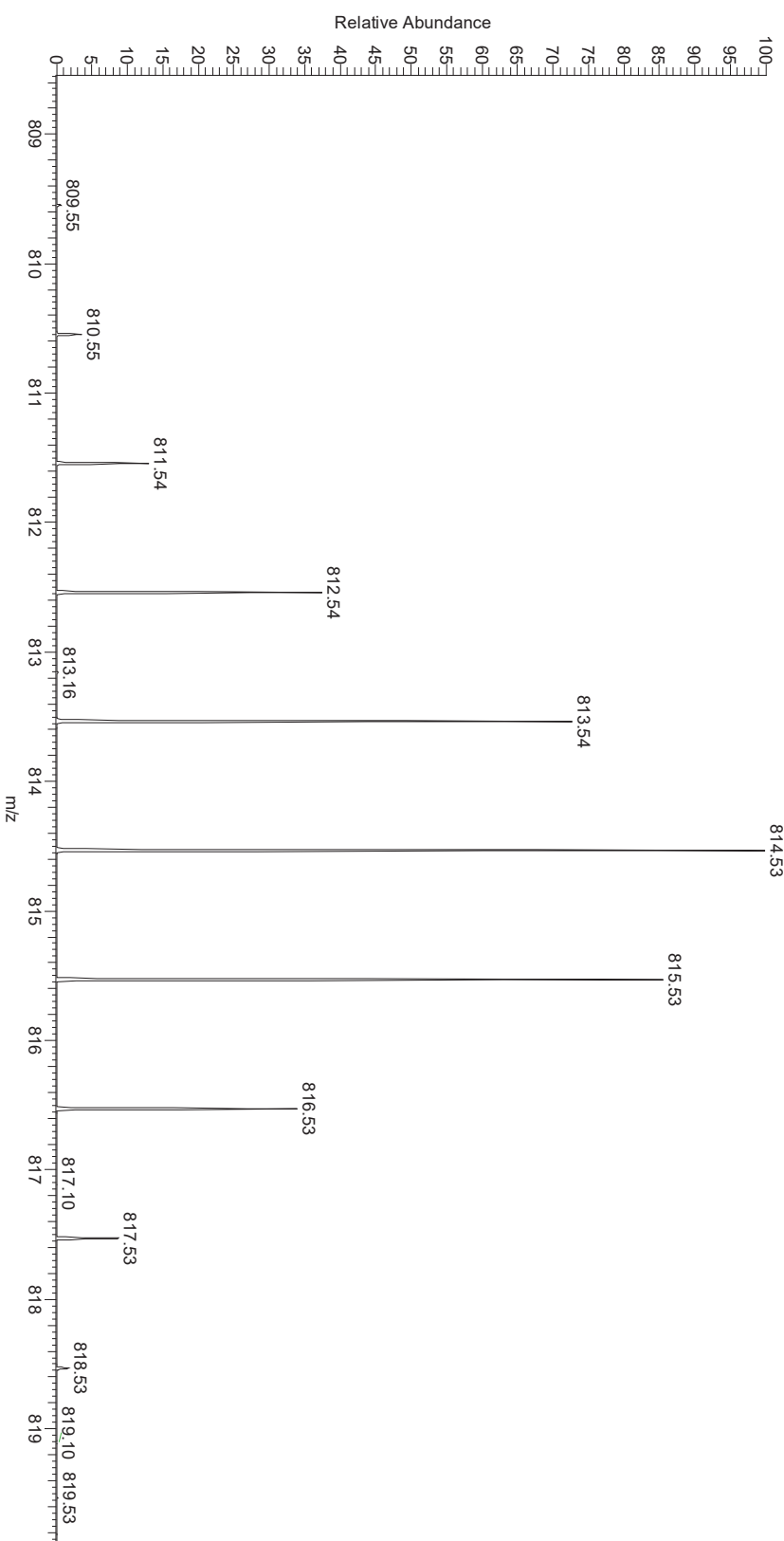
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 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

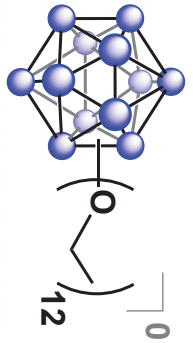




Q Exactive High-Res Mass Spec

Alex Alky_1 #1 RT: 0.01 AV: 1 NL: 5.39E7
T: FTMS - p ESI Full ms [500.00-5000.00]





¹¹B {¹H} NMR



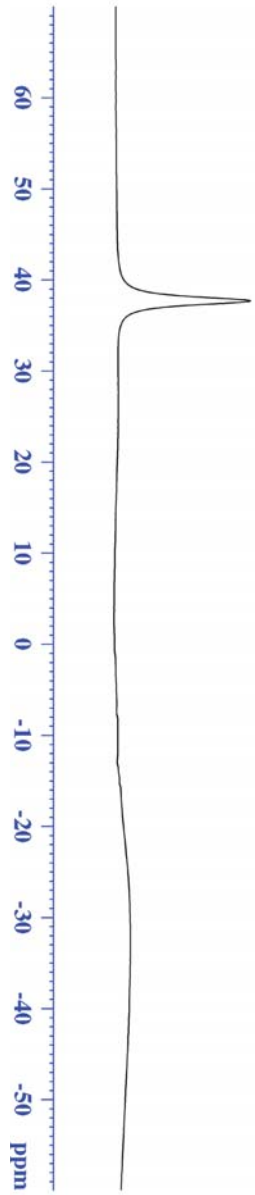
Current Data Parameters
 NAME B12(O.E0)12
 EXPNO 100
 PROCNO 1

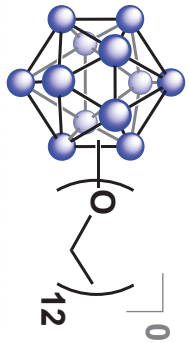
F2 - Acquisition Parameters
 Date_ 20150412
 Time 19.11
 INSTRUM av400
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 5096
 SOLVENT CDCl3
 NS 1024
 DS 0
 SWH 51020.406 Hz
 FIDRES 10.011854 Hz
 AQ 0.0449408 sec
 RG 189.85
 DW 9.800 usec
 DE 6.50 usec
 TE 299.1 K
 D1 0.00000400 sec
 D11 0.030000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 128.3776052 MHz
 NUC1 11B
 P1 10.00 usec
 PLW1 52.000000000 W

===== CHANNEL f2 =====
 SFO2 400.1324008 MHz
 NUC2 1H
 CPDPRG12 waltz16
 PCTPD2 90.00 usec
 PLW2 13.000000000 W
 PLW12 0.361110000 W

F2 - Processing parameters
 SI 32768
 SF 128.3776050 MHz
 WDW EM
 SSB 0
 LB 50.00 Hz
 GB 0
 PC 1.40

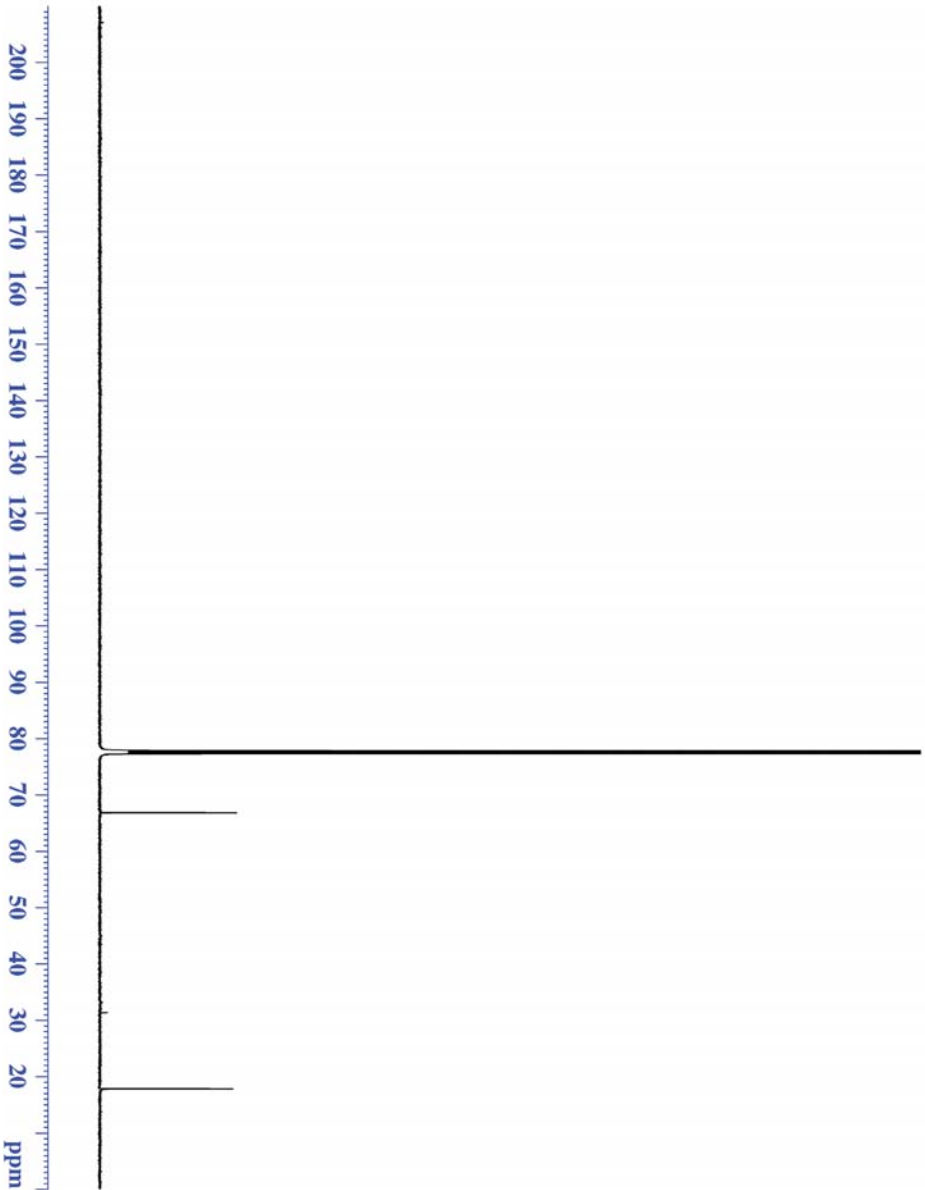




¹³C NMR



66.793
17.802



Current Data Parameters
 NAME B12(O-E0)12
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150420
 Time 21.23

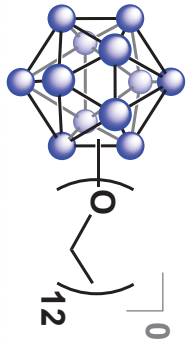
INSTRUM av500
 PROBHID 5 mm DCH 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 64
 DS 2

SWH 31250.000 Hz
 FIDRES 0.476837 Hz
 AQ 1.0485760 sec
 RG 204.54
 DW 16.000 usec
 DE 18.00 usec
 TE 298.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

==== CHANNEL f1 =====
 SFO1 125.772511 MHz
 NUC1 13C
 P1 9.63 usec
 PLW1 23.00000000 W

==== CHANNEL f2 =====
 SFO2 500.1330008 MHz
 NUC2 1H
 CPDPRGf2 waltz16
 PCPD2 80.00 usec
 PLW2 13.50000000 W
 PLW12 0.21094000 W
 PLW13 0.113500001 W

F2 - Processing parameters
 SF 131072
 SE 125.777890 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



¹H NMR

- 4.109
- 4.095
- 4.081
- 4.068
- 1.249
- 1.235
- 1.221

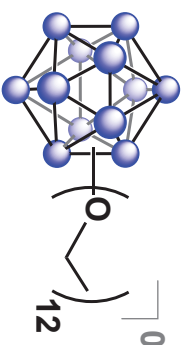


Current Data Parameters
 Name: B12(O-E0)12
 ExpNO: 11
 ProcNO: 1

F2 - Acquisition Parameters
 Date_: 20150420
 Time: 21.30
 INSTRUM: av500
 PROBHD: 5 mm DCH 13C-1
 PULPROG: zg30
 TD: 65536
 SOLVENT: CDCl3
 NS: 64
 DS: 0
 SWH: 10000.000 Hz
 FIDRES: 0.152588 Hz
 AQ: 3.2767999 sec
 RG: 52.41
 DW: 50.000 usec
 DE: 10.00 usec
 TE: 298.0 K
 D1: 2.00000000 sec
 TD0: 1

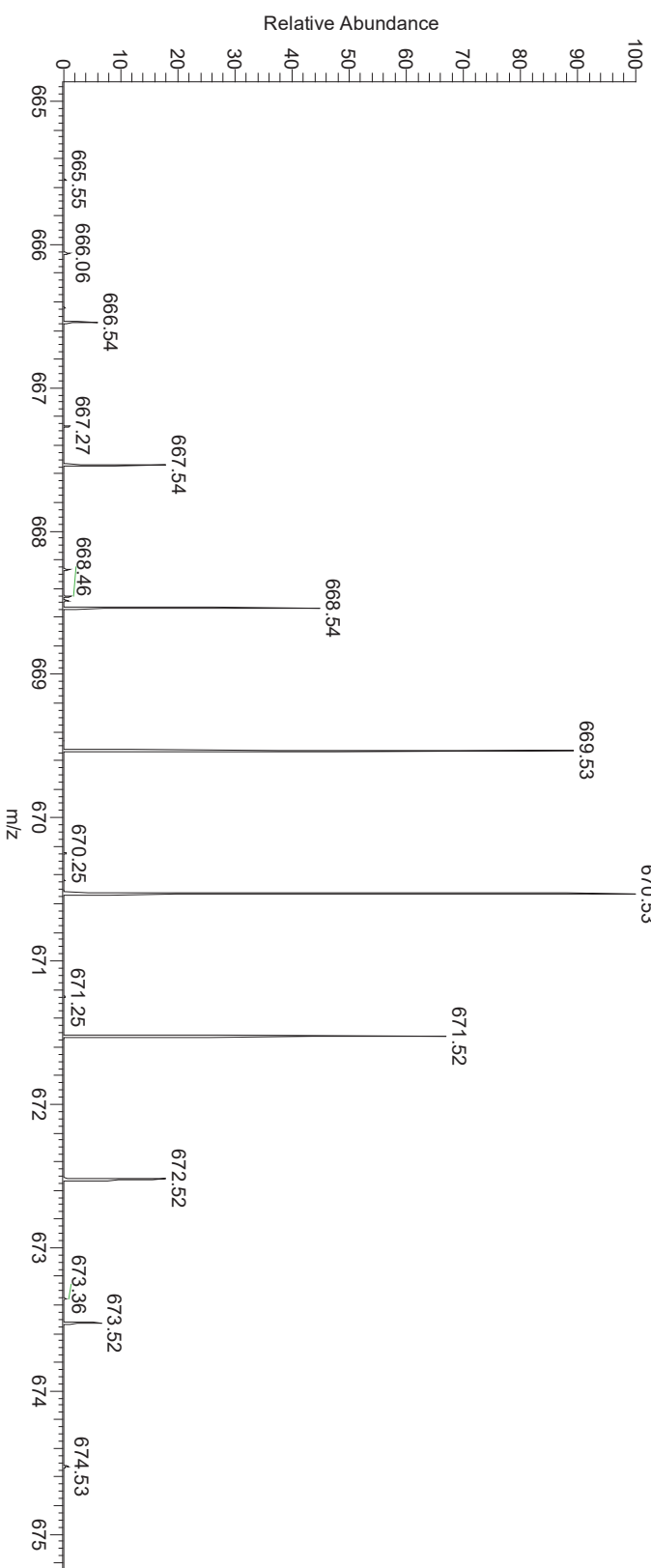
===== CHANNEL f1 =====
 SFO1 500.1330008 MHz
 NUC1 ¹H
 Pl 10.00 usec
 Plw1 13.50000000 W

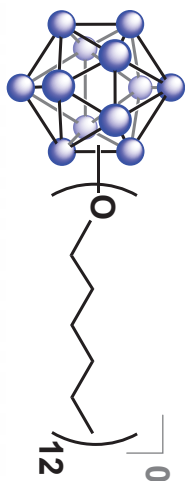
F2 - Processing parameters
 SI 65536
 SF 500.1300115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Q Exactive High-Res Mass Spec

Et#1 RT: 0.01 AV: 1 NL: 1.89E6
T: FTMS - p-ESI Fullms [300.00-1200.00]





41.983

¹¹B {¹H} NMR



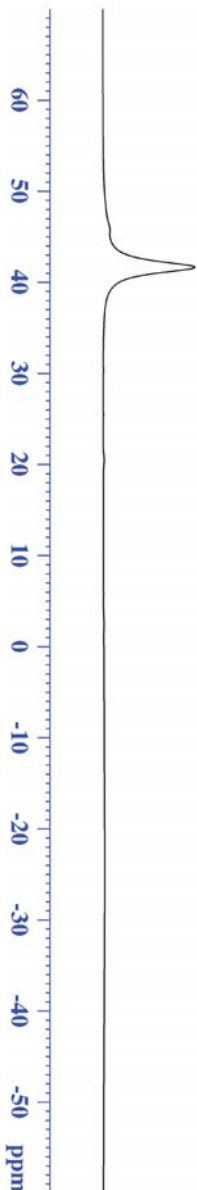
Current Data Parameters
 NAME B12(O.Hexyl)12
 EXPNO 20
 PROCNO 1

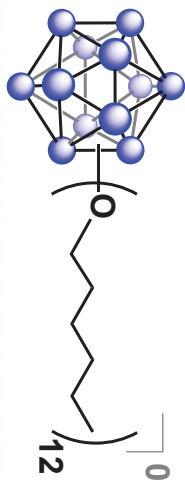
F2 - Acquisition Parameters
 Date_ 20150610
 Time 15:09
 INSTRUM av400
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 5096
 SOLVENT CDCl3
 NS 1024
 DS 0
 SWH 51020.406 Hz
 FIDRES 10.011854 Hz
 AQ 0.0449408 sec
 RG 189.85
 DW 9.800 usec
 DE 6.50 usec
 TE 299.2 K
 D1 0.00000400 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 128.3776052 MHz
 NUC1 11B
 P1 10.00 usec
 PLW1 52.000000000 W

===== CHANNEL f2 =====
 SFO2 400.1324008 MHz
 NUC2 1H
 CPDPRG12 waltz16
 PCTPD2 90.00 usec
 PLW2 13.000000000 W
 PLW12 0.36111000 W

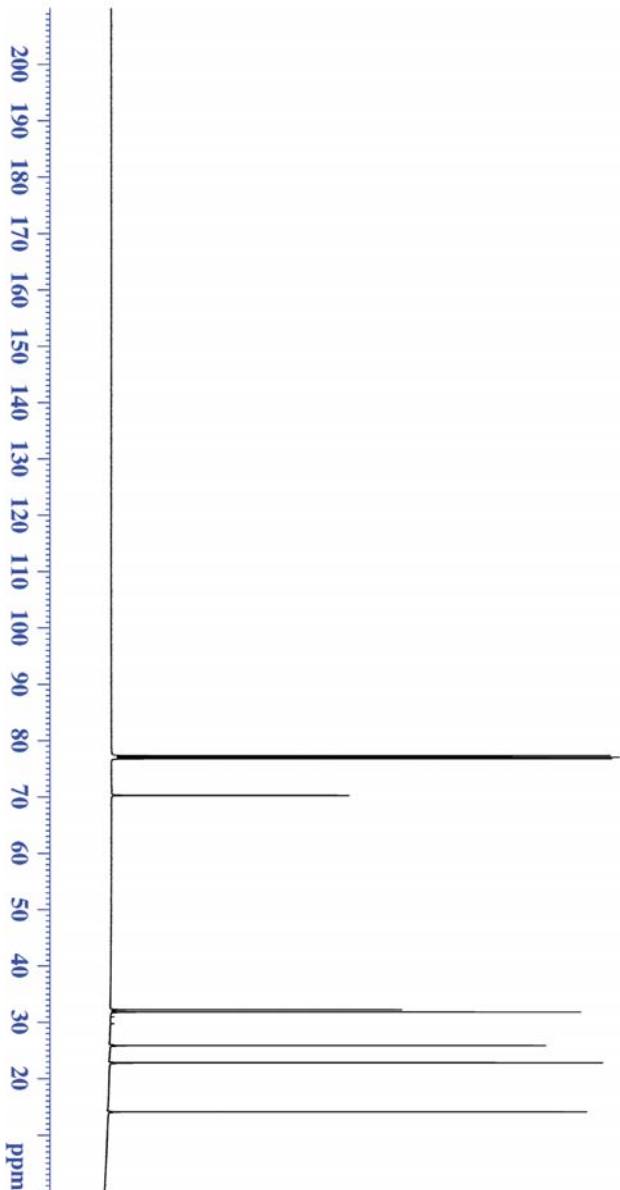
F2 - Processing parameters
 SI 32768
 SF 128.3776050 MHz
 WDW EM
 SSB 0
 LB 50.00 Hz
 GB 0
 PC 1.40





¹³C NMR

- 70.232
- 32.206
- 31.792
- 25.854
- 22.778
- 14.080



Current Data Parameters
 NAME B12(O-Hevy)12
 EXPNO 200
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150829
 Time 20.52

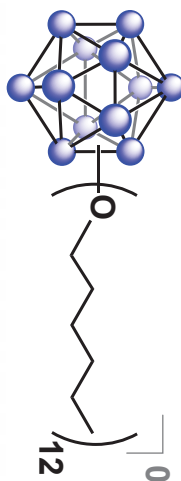
INSTRUM av500
 PROBHID 5 mm DGH 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 256
 DS 2

SWH 31250.000 Hz
 FIDRES 0.476837 Hz
 AQ 1.0485760 sec
 RG 204.54
 DW 16.000 usec
 DE 18.00 usec
 TE 298.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.772511 MHz
 NUC1 13C
 P1 9.63 usec
 PLW1 23.00000000 W

==== CHANNEL f2 =====
 SFO2 500.1330008 MHz
 NUC2 1H
 CPDPRGf2 waltz16
 PCPD2 80.00 usec
 PLW2 13.50000000 W
 PLW12 0.21094000 W
 PLW13 0.113500001 W

F2 - Processing parameters
 SF 131.072
 SE 125.777890 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



¹H NMR

- 4.037
- 4.021
- 4.005
- 1.571
- 1.555
- 1.539
- 1.520
- 1.504
- 1.374
- 1.353
- 1.334
- 1.327
- 1.316
- 1.312
- 1.290
- 1.283



Current Data Parameters
 NAME B12(O-Hexyl)12
 EXPNO 21
 PROCNO 1

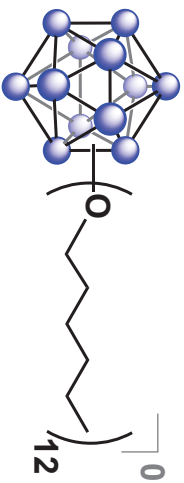
F2 - Acquisition Parameters
 Date_ 20150610
 Time 15.11

INSTRUM av400
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 52882
 SOLVENT CDCl3
 NS 8
 DS 0

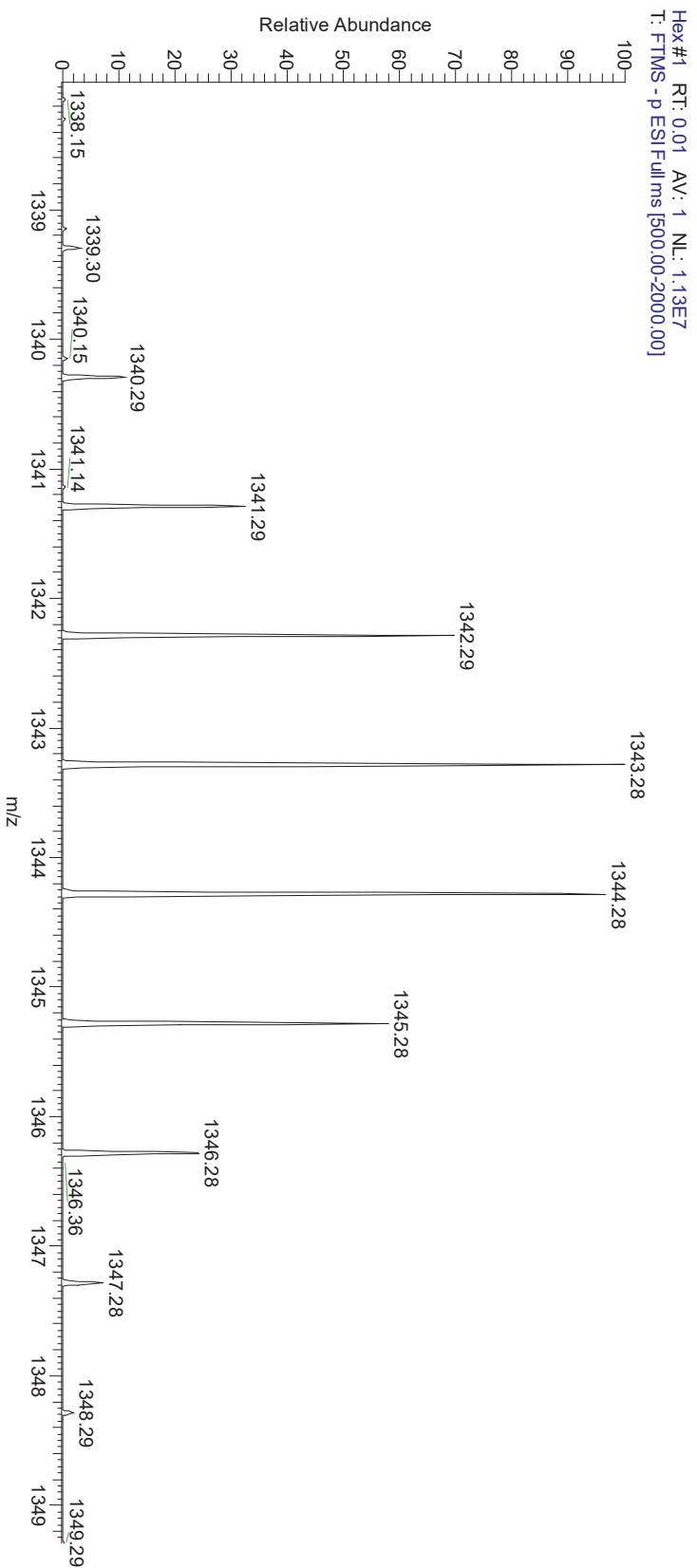
SWH 8012.820 Hz
 FIDRES 0.151523 Hz
 AQ 3.2998369 sec
 RG 28.22
 DW 62.400 usec
 DE 6.50 usec
 TE 299.0 K
 D1 2.00000000 sec
 TD0 1

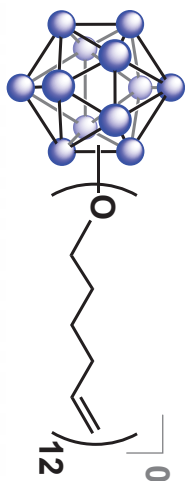
===== CHANNEL f1 =====
 SFO1 400.1324008 MHz
 NUC1 ¹H
 Pl 15.00 usec
 PLW1 13.000000000 W

F2 - Processing parameters
 SI 65536
 SF 400.1300184 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



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¹¹B {¹H} NMR



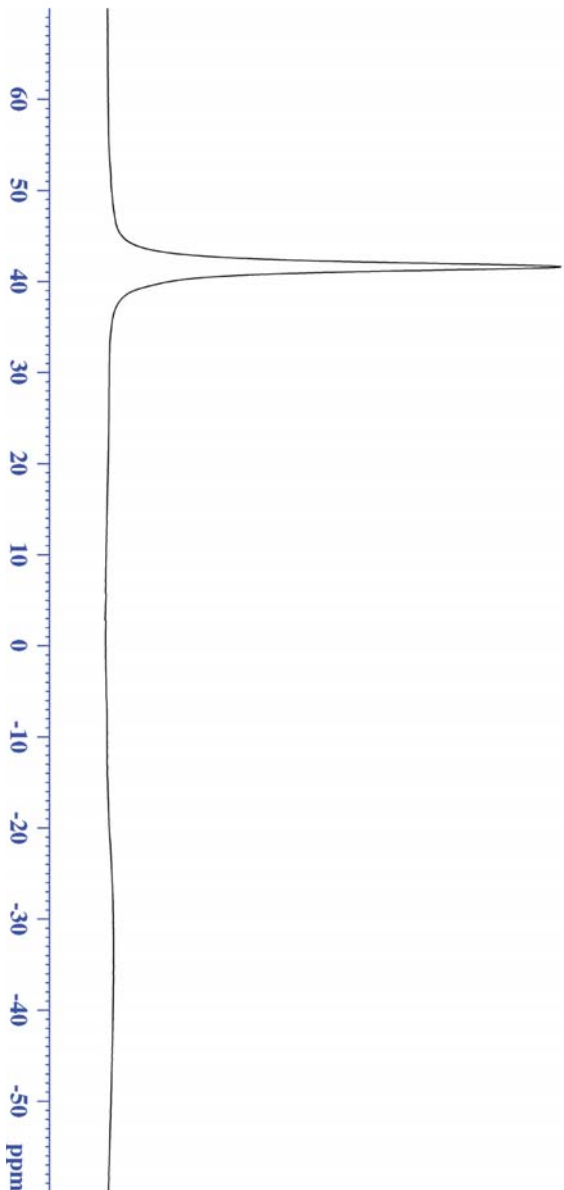
Current Data Parameters
 NAME Oct05_2015
 EXPNO 51
 PROCNO 1

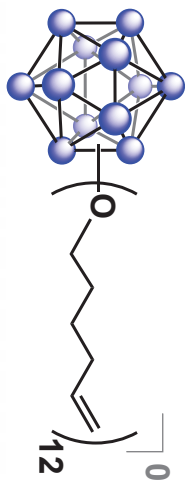
F2 - Acquisition Parameters
 Date_ 20151005
 Time 15.01
 INSTRUM av400
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 5096
 SOLVENT CDCl3
 NS 1024
 DS 0
 SWH 51020.406 Hz
 FIDRES 10.011854 Hz
 AQ 0.0449408 sec
 RG 189.85
 DW 9.800 usec
 DE 6.50 usec
 TE 299.1 K
 D1 0.05000000 sec
 D11 0.05000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 128.3776052 MHz
 NUC1 ¹¹B
 P1 10.00 usec
 PLW1 52.000000000 W

===== CHANNEL f2 =====
 SFO2 400.1324008 MHz
 NUC2 ¹H
 CPDPRG12 waltz16
 PCTPD2 90.00 usec
 PLW2 13.000000000 W
 PLW12 0.36111000 W

F2 - Processing parameters
 SI 32768
 SF 128.3776161 MHz
 WDW EM
 SSB 0
 LB 10.00 Hz
 GB 0
 PC 1.40





¹³C NMR



Current Data Parameters
 NAME B12(O-Hexene)12
 EXPNO 20
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20151021
 Time 17:44

INSTRUM av500
 PROBHID 5 mm DCH 13C-1
 PULPROG zgpg30

TD 65536
 SOLVENT CDCl3
 NS 256

DS 2
 SWH 31250.000 Hz
 FIDRES 0.476837 Hz

AQ 1.0485760 sec
 RG 204.54
 DW 16.000 usec

DE 18.00 usec
 TE 298.0 K
 D1 2.00000000 sec

D11 0.030000000 sec
 TD0 1

==== CHANNEL f1 =====
 SFO1 125.7722511 MHz
 NUC1 13C

P1 9.63 usec
 PLW1 23.000000000 W

==== CHANNEL f2 =====
 SFO2 500.1330008 MHz
 NUC2 1H

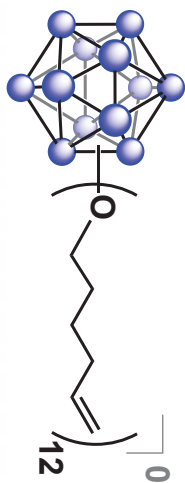
CPDPRGf2 waltz16
 PCPD2 80.00 usec
 PLW2 13.500000000 W

PLW12 0.21094000 W
 PLW13 0.113500001 W

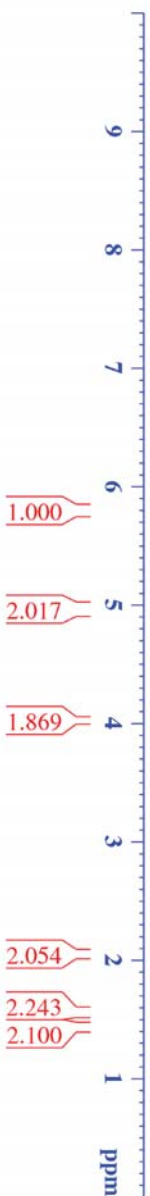
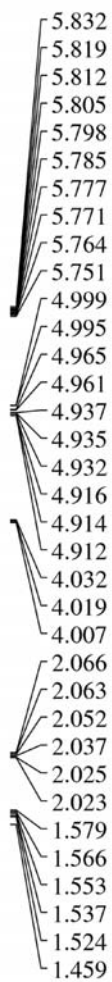
F2 - Processing parameters
 SF 131.072
 SE 125.7577890 MHz

WDW EM
 SSB 0
 LB 1.00 Hz

GB 0
 PC 1.40



¹H NMR

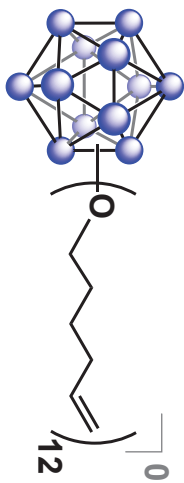


Current Data Parameters
 NAME B12(O-Hexene)12
 EXPNO 21
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20151021
 Time 17:48
 INSTRUM av500
 PROBHD 5 mm DCH 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 32
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 33.91
 DW 50.000 usec
 DE 10.00 usec
 TE 298.0 K
 D1 2.000000000 sec
 TD0 1

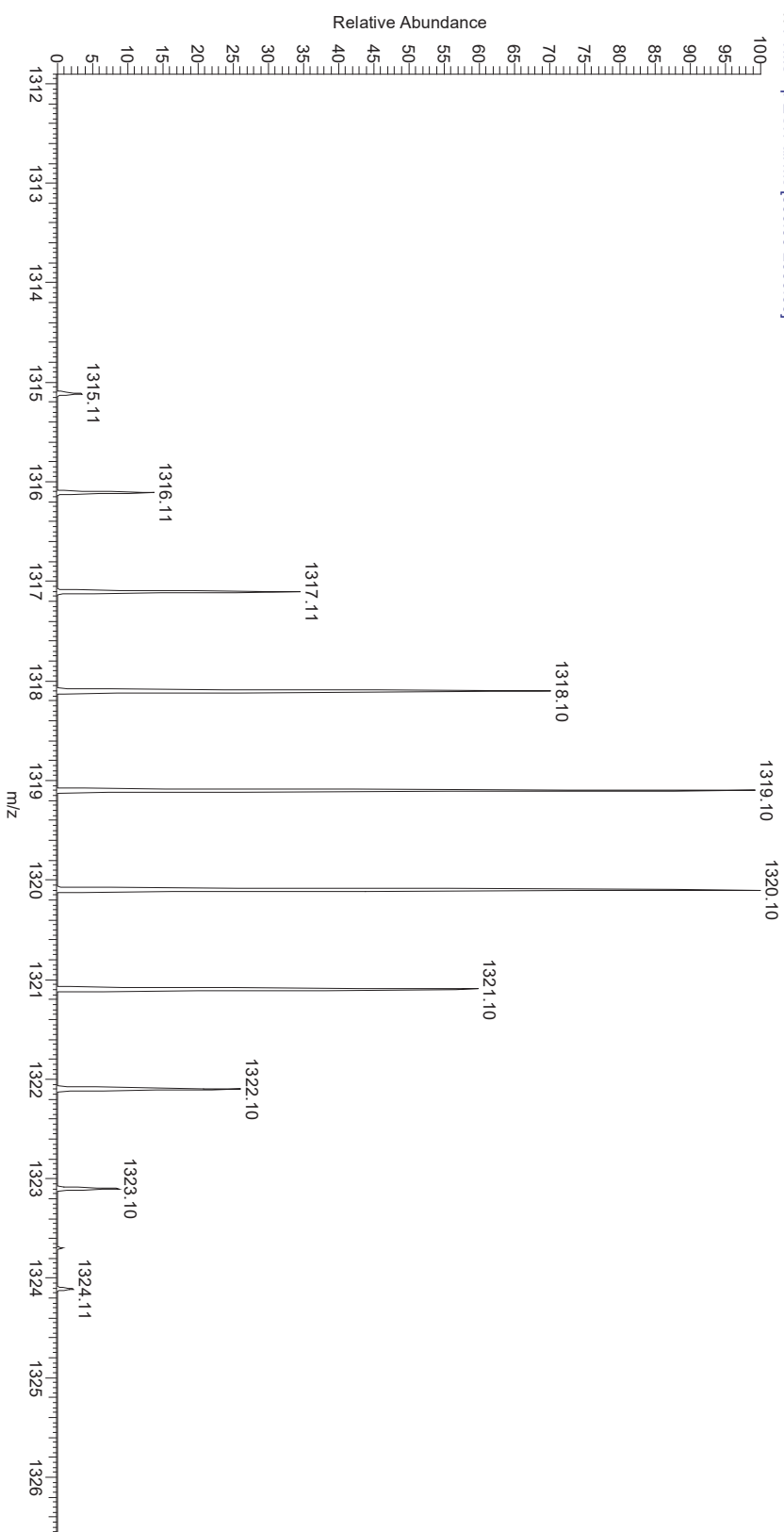
==== CHANNEL f1 ====
 SFO1 500.1330008 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 13.500000000 W
 F2 - Processing parameters
 SI 65536
 SF 500.1300146 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

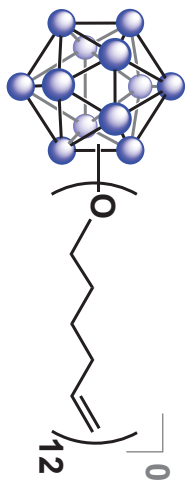




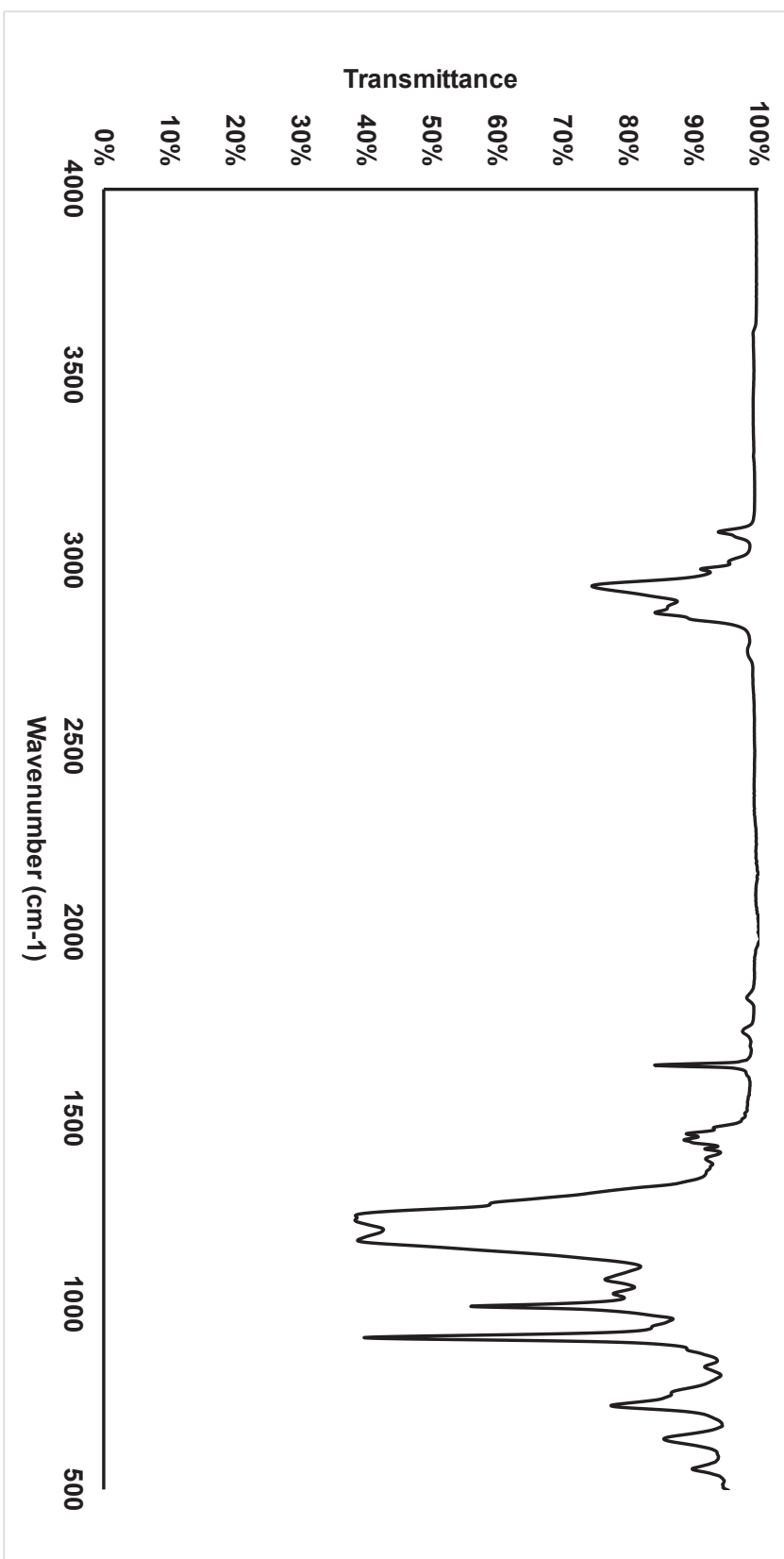
Q Exactive High-Res Mass Spec

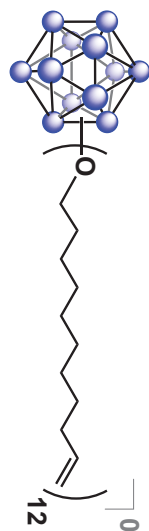
Hexene #1 RT: 0.01 AV: 1 NL: 6.90E5
T: FTMS - p ESI Full ms [500.00-2000.00]





IR

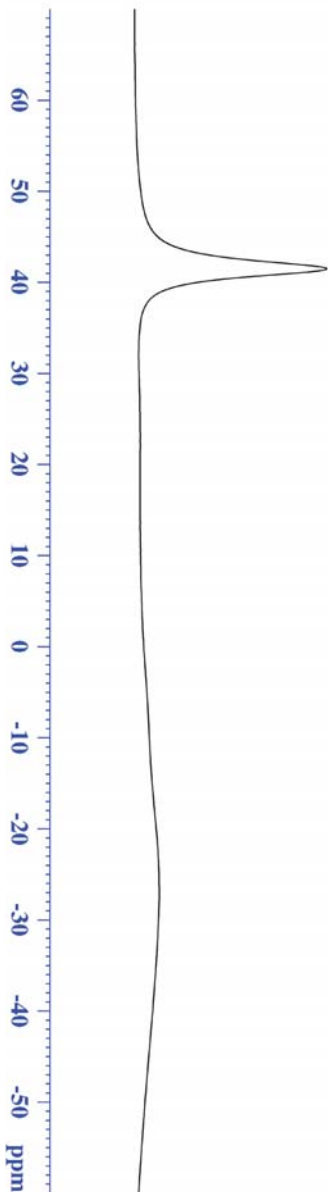




¹¹B {¹H} NMR



41.474



Current Data Parameters
 NAME B12(O-1-undecene)12
 EXPNO 20
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150626
 Time 12.16
 INSTRUM av400
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg3s
 TD 5096
 SOLVENT CD2Cl2
 NS 1024
 DS 0
 SWH 51020.406 Hz
 FIDRES 10.011854 Hz
 AQ 0.0449408 sec
 RG 189.85
 DW 9.800 usec
 DE 6.50 usec
 TE 299.1 K
 D1 0.00000400 sec
 D11 0.030000000 sec
 TD0 1

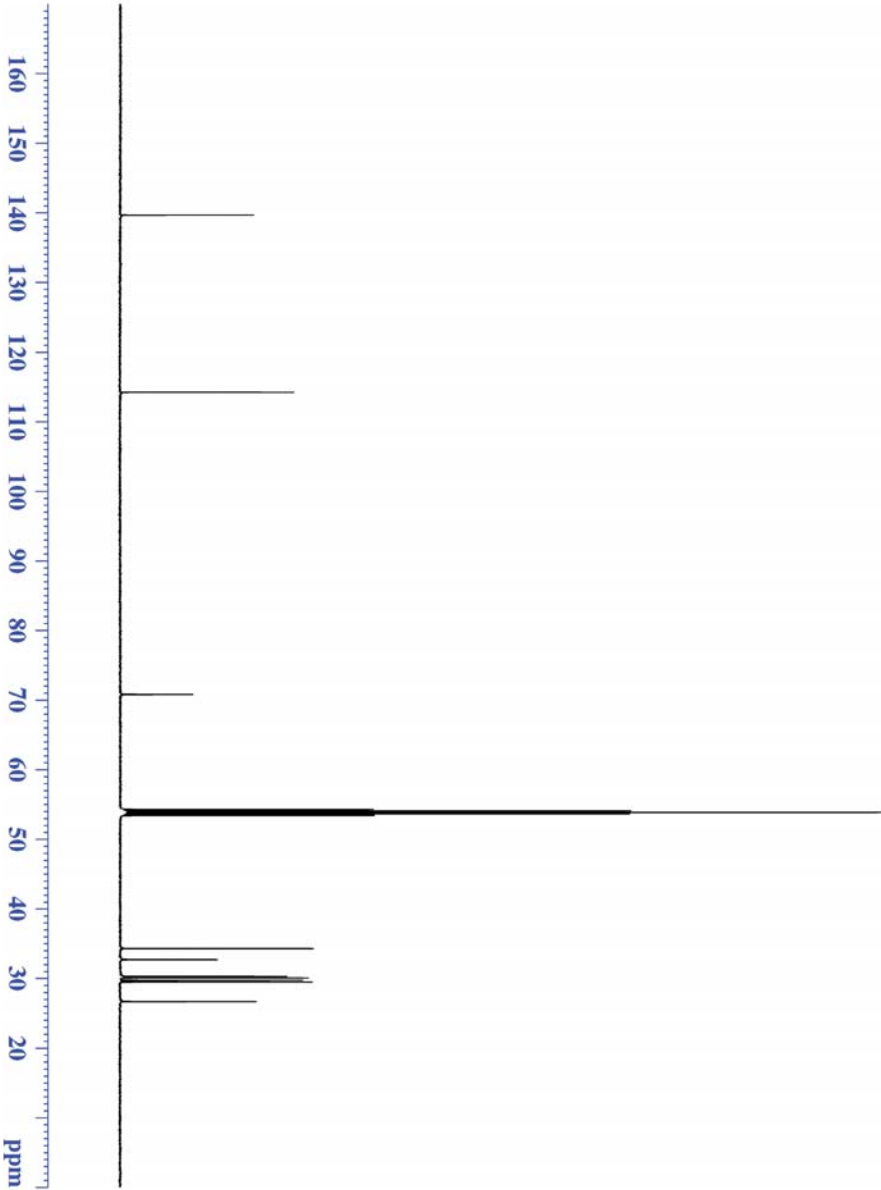
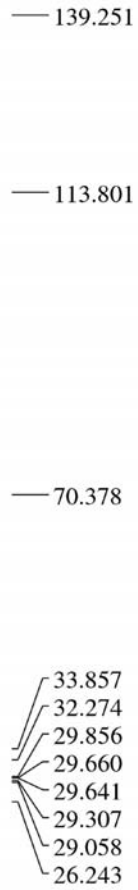
===== CHANNEL f1 =====
 SFO1 128.3776052 MHz
 NUC1 11B
 P1 10.00 usec
 PLW1 52.000000000 W

===== CHANNEL f2 =====
 SFO2 400.1324008 MHz
 NUC2 1H
 CPDPRG12 waltz16
 PCTPD2 90.00 usec
 PLW2 13.000000000 W
 PLW12 0.36111000 W

F2 - Processing parameters
 SI 32768
 SF 128.3776050 MHz
 WDW EM
 SSB 0
 LB 50.00 Hz
 GB 0
 PC 1.40



¹³C NMR



Current Data Parameters
 NAME Jun26-2015
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150626
 Time 14:05

INSTRUM av500
 PROBHID 5 mm DGH 13C-1
 PULPROG zgpg30

TD 65536
 SOLVENT CD2Cl2
 NS 64

DS 2
 SWH 31250.000 Hz
 FIDRES 0.476837 Hz

AQ 1.0485760 sec
 RG 204.54
 DW 16.000 usec

DE 18.00 usec
 TE 298.0 K
 D1 2.00000000 sec
 D11 0.030000000 sec
 TD0 1

==== CHANNEL f1 =====
 SFO1 125.772511 MHz
 NUC1 13C
 P1 9.63 usec
 PLW1 23.000000000 W

==== CHANNEL f2 =====
 SFO2 500.1330008 MHz
 NUC2 1H
 P2 13.000000000 W

==== CHANNEL f3 =====
 SFO3 125.772511 MHz
 NUC3 13C
 P3 9.63 usec
 PLW3 23.000000000 W

==== CHANNEL f4 =====
 SFO4 500.1330008 MHz
 NUC4 1H
 P4 13.000000000 W

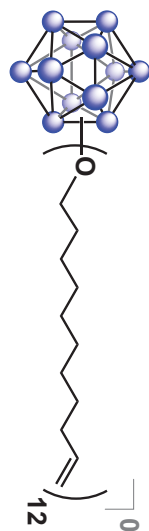
==== CHANNEL f5 =====
 SFO5 125.772511 MHz
 NUC5 13C
 P5 9.63 usec
 PLW5 23.000000000 W

==== CHANNEL f6 =====
 SFO6 500.1330008 MHz
 NUC6 1H
 P6 13.000000000 W

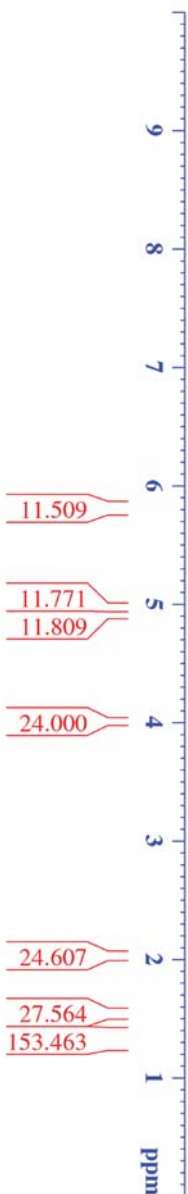
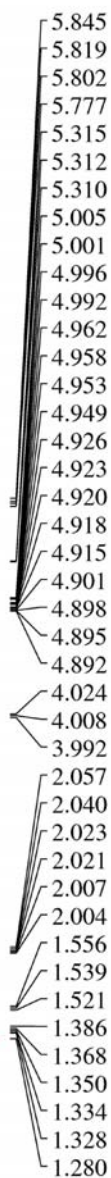
==== CHANNEL f7 =====
 SFO7 125.772511 MHz
 NUC7 13C
 P7 9.63 usec
 PLW7 23.000000000 W

==== CHANNEL f8 =====
 SFO8 500.1330008 MHz
 NUC8 1H
 P8 13.000000000 W

F2 - Processing parameters
 SI 131072
 SF 125.777890 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



¹H NMR



Current Data Parameters
 NAME B12(O-1-undecene)12
 EXPNO 21
 PROCNO 1

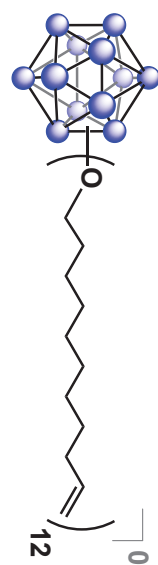
F2 - Acquisition Parameters
 Date_ 20150626
 Time 12.18
 INSTRUM av400
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 52882
 SOLVENT CD2Cl2
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.151523 Hz
 AQ 3.2998369 sec
 RG 83.63
 DW 62.400 usec
 DE 6.50 usec
 TE 299.0 K
 D1 2.00000000 sec
 TD0 1

==== CHANNEL f1 =====

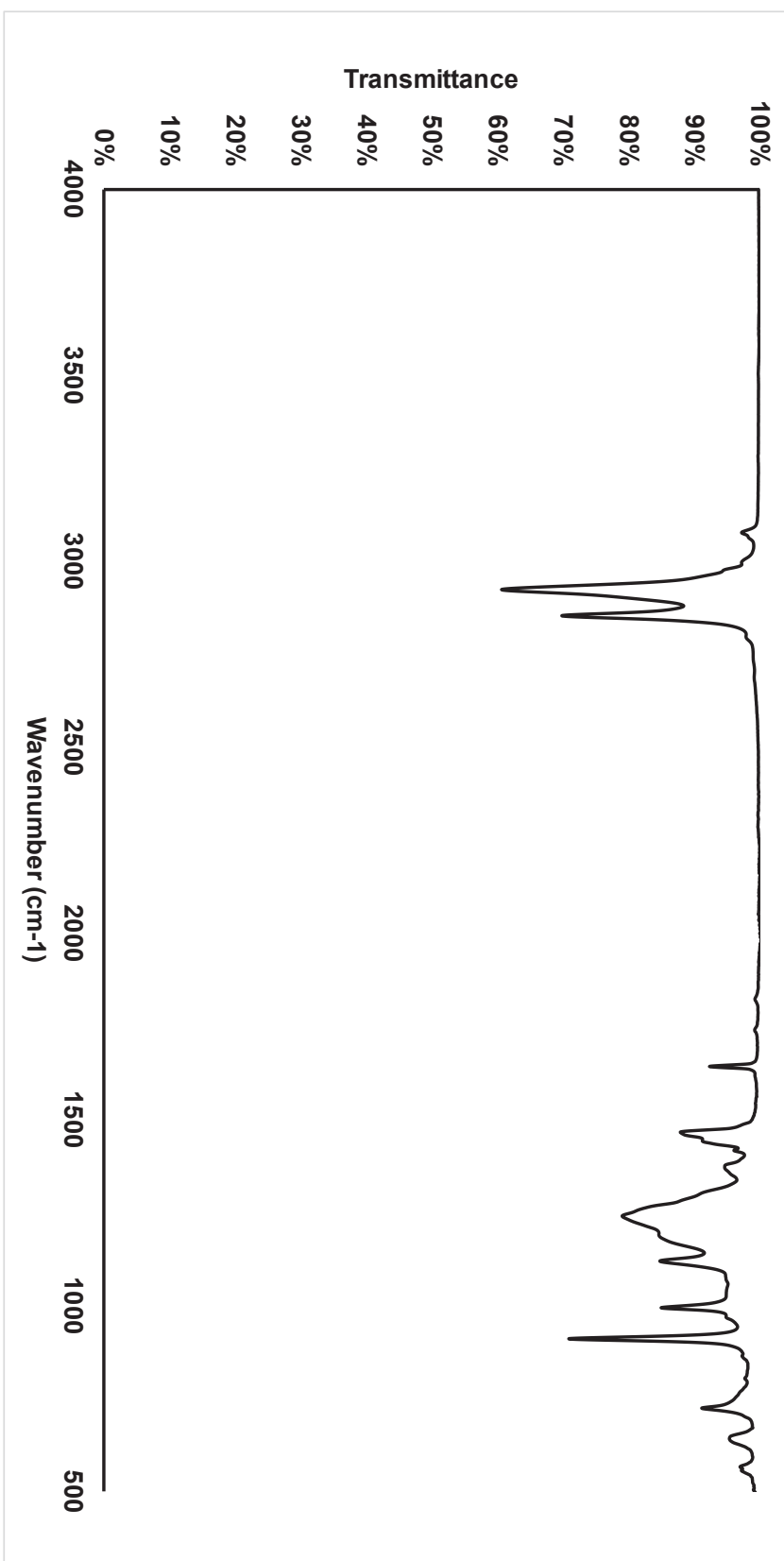
SFO1 400.1324008 MHz
 NUC1 1H
 P1 15.00 usec
 PLW1 13.00000000 W

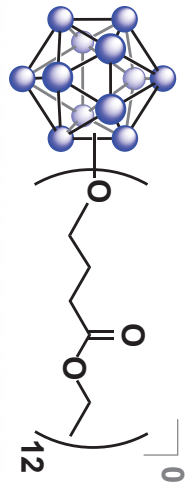
F2 - Processing parameters
 SI 65536
 SF 400.1300184 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





IR





$^{11}\text{B} \{^1\text{H}\}$ NMR



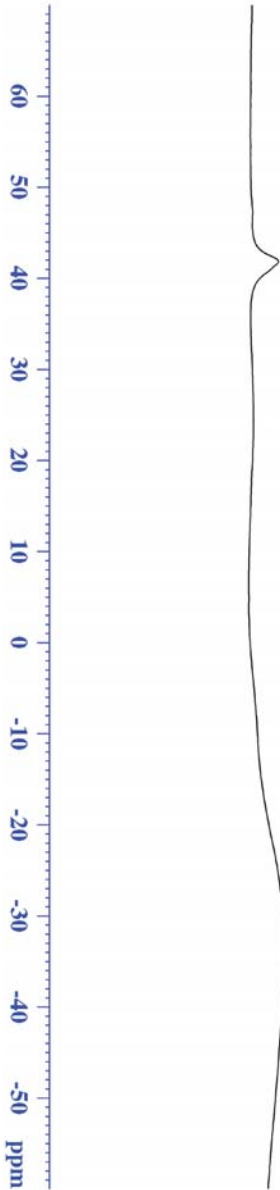
Current Data Parameters
 NAME B12(O:EBuBr)12
 EXPNO 110
 PROCNO 1

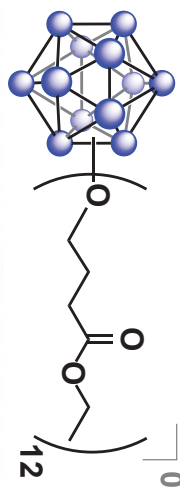
F2 - Acquisition Parameters
 Date_ 20150412
 Time 19.19
 INSTRUM av400
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 5096
 SOLVENT CDCl3
 NS 1024
 DS 0
 SWH 51020.406 Hz
 FIDRES 10.011854 Hz
 AQ 0.0449408 sec
 RG 189.85
 DW 9.800 usec
 DE 6.50 usec
 TE 299.1 K
 D1 0.00000400 sec
 D11 0.030000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 128.3776052 MHz
 NUC1 ^{11}B
 P1 10.00 usec
 PLW1 52.000000000 W

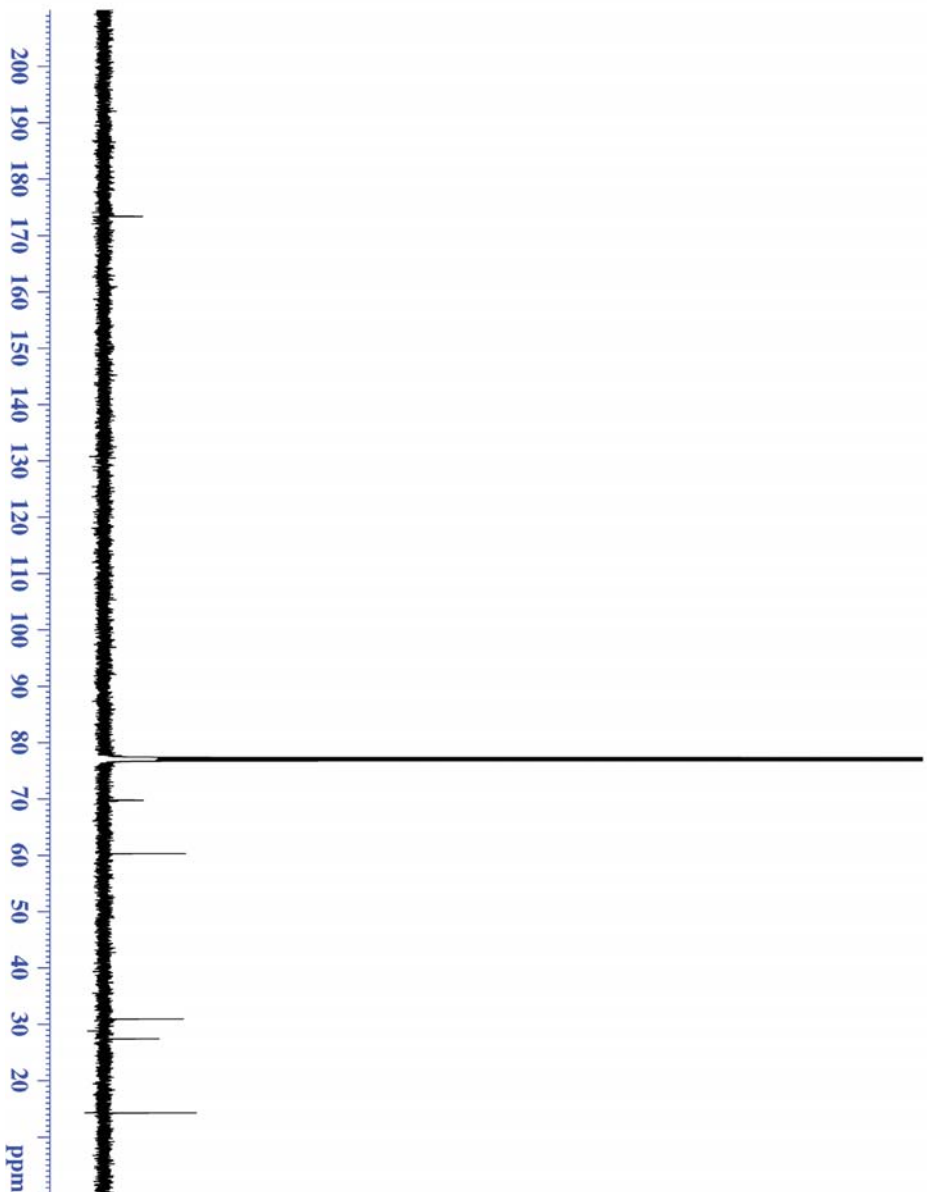
===== CHANNEL f2 =====
 SFO2 400.1324008 MHz
 NUC2 ^1H
 CPDPRG12 waltz16
 PCTPD2 90.00 usec
 PLW2 13.000000000 W
 PLW12 0.361110000 W

F2 - Processing parameters
 SI 32768
 SF 128.3776050 MHz
 WDW EM
 SSB 0
 LB 50.00 Hz
 GB 0
 PC 1.40





¹³C NMR



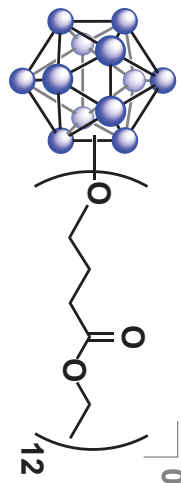
Current Data Parameters
 NAME B12(O-Etbutyr)12
 EXPNO 60
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150406
 Time 23:42
 INSTRUM av500
 PROBHID 5 mm DCH 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 64
 DS 2
 SWH 31250.000 Hz
 FIDRES 0.476837 Hz
 AQ 1.0485760 sec
 RG 204.54
 DW 16.000 usec
 DE 18.00 usec
 TE 298.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TDO 1

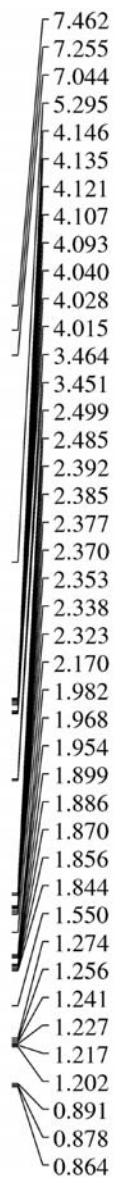
==== CHANNEL f1 =====
 SFO1 125.7722511 MHz
 NUC1 13C
 P1 9.63 usec
 PLW1 23.000000000 W

==== CHANNEL f2 =====
 SFO2 500.1330008 MHz
 NUC2 1H
 CPDPRGf2 waltz16
 PCPD2 80.00 usec
 PLW2 13.500000000 W
 PLW12 0.21094000 W
 PLW13 0.113500001 W

F2 - Processing parameters
 SF 131.072
 SE 125.757890 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



¹H NMR

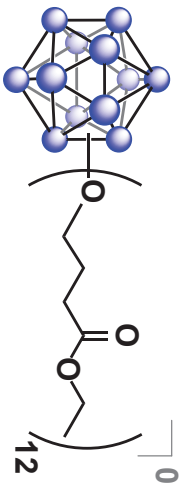


Current Data Parameters
 NAME B12(O-EtBu)12
 EXPNO 61
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150406
 Time 23.44
 INSTRUM av500
 PROBHD 5 mm DCH 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 59.34
 DW 50.000 usec
 DE 10.00 usec
 TE 298.0 K
 D1 2.00000000 sec
 TD0 1

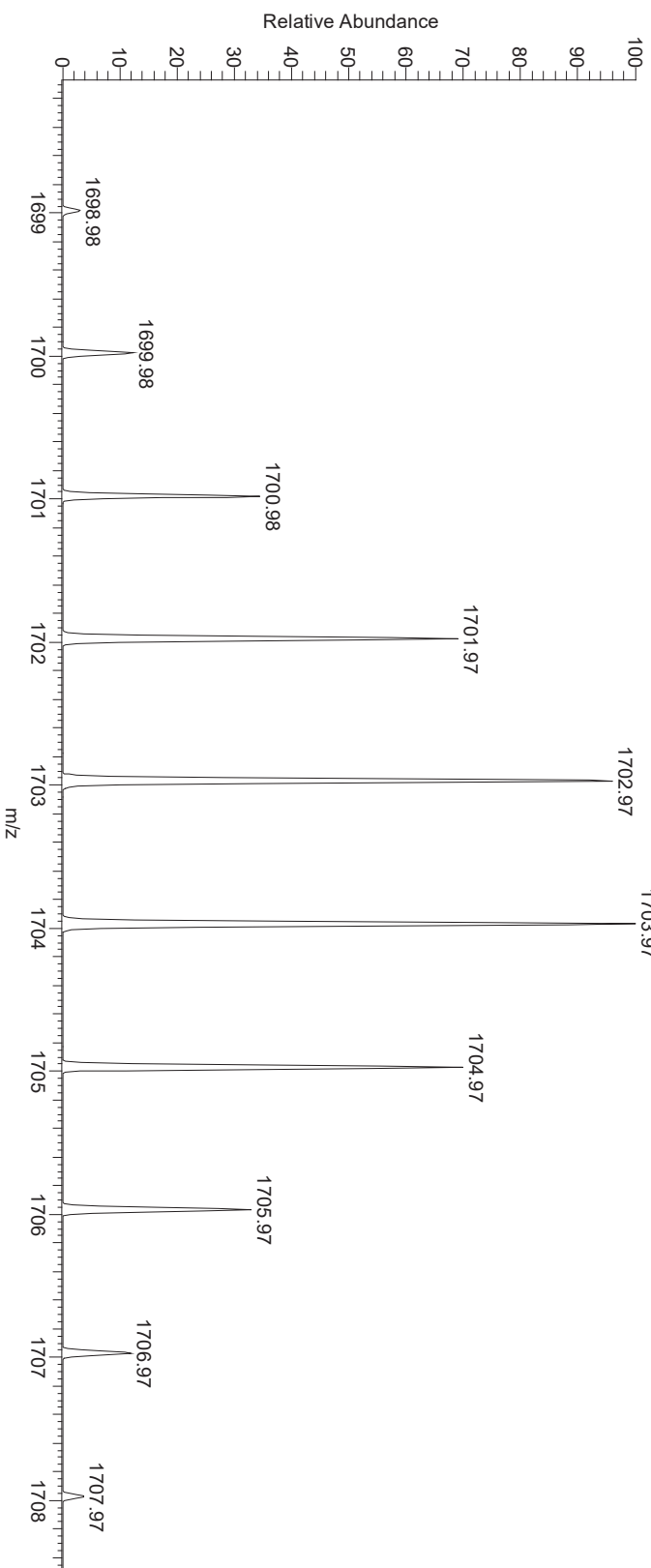
===== CHANNEL f1 =====
 SFO1 500.1330008 MHz
 NUC1 ¹H
 Pl 10.00 usec
 PLW1 13.50000000 W
 F2 - Processing parameters
 SI 65536
 SF 500.1300146 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





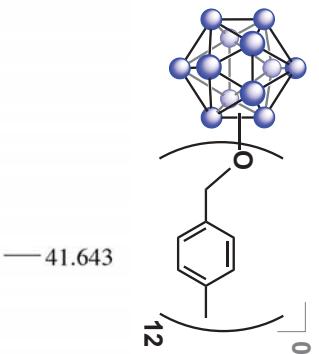
Q Exactive High-Res Mass Spec

EtBut#1 RT: 0.01 AV: 1 NL: 2.50E7
T: FTMS - p-ESI Full ms [800.00-2000.00]





^{11}B { ^1H } NMR



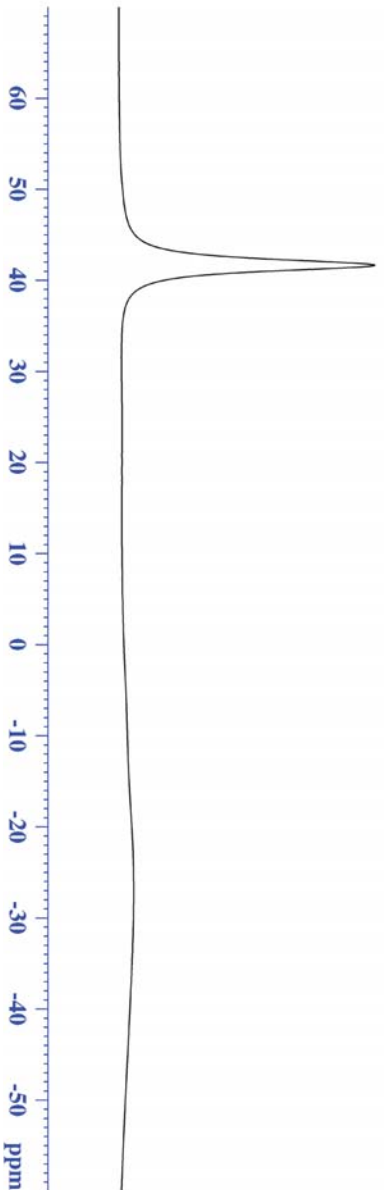
Current Data Parameters
NAME Aug29_2015
EXPNO 160
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150829
Time 21.02
INSTRUM av400
PROBHD 5 mm PABBO BB/
PULPROG zgpg3s
TD 5096
SOLVENT CDCl3
NS 1024
DS 0
SWH 51020.406 Hz
FIDRES 10.011854 Hz
AQ 0.0449408 sec
RG 189.85
DW 9.800 usec
DE 6.50 usec
TE 299.3 K
D1 0.00000400 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 128.3776052 MHz
NUC1 ^{11}B
P1 10.00 usec
PLW1 52.000000000 W

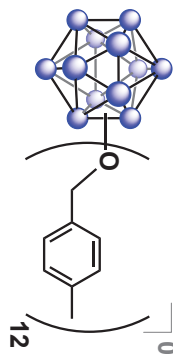
===== CHANNEL f2 =====
SFO2 400.1324008 MHz
NUC2 ^1H
CPDPRG12 waltz16
PCPD2 90.00 usec
PLW2 13.000000000 W
PLW12 0.36111000 W

F2 - Processing parameters
SI 32768
SF 128.3776050 MHz
WDW EM
SSB 0
LB 50.00 Hz
GB 0
PC 1.40





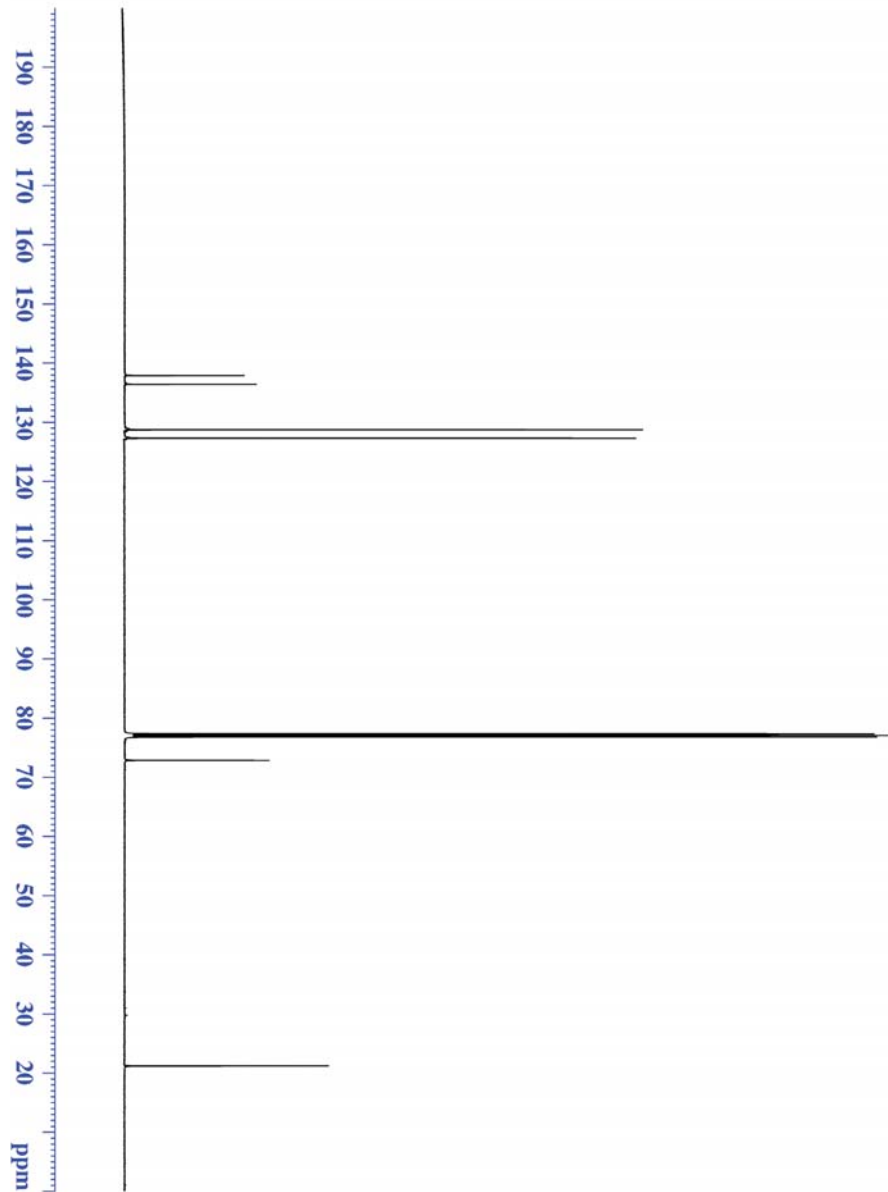
¹³C NMR



137.858
136.379
128.688
127.265

72.807

21.164



Current Data Parameters
NAME B12(O-4-MeBn)12
EXPNO 60
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150829
Time 22:04

INSTRUM av500
PROBHD 5 mm DCH 13C-1
PULPROG zgpg30

TD 65536
SOLVENT CDCl3

NS 256
DS 2

SWH 31250.000 Hz
FIDRES 0.476837 Hz

AQ 1.0485760 sec
RG 204.54

DW 16.000 usec
DE 18.00 usec

TE 298.0 K
D1 2.00000000 sec

D11 0.03000000 sec
TD0 1

==== CHANNEL f1 =====
SFO1 125.7722511 MHz
NUC1 13C
P1 9.63 usec
PLW1 23.000000000 W

==== CHANNEL f2 =====
SFO2 500.1330008 MHz
NUC2 1H
P2 13.000000000 W

==== CHANNEL f3 =====
SFO3 125.7722511 MHz
NUC3 13C
P3 9.63 usec
PLW3 23.000000000 W

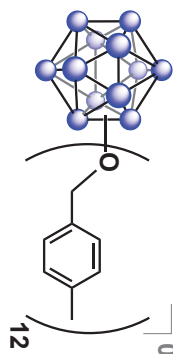
==== CHANNEL f4 =====
SFO4 500.1330008 MHz
NUC4 1H
P4 13.000000000 W

==== CHANNEL f5 =====
SFO5 125.7722511 MHz
NUC5 13C
P5 9.63 usec
PLW5 23.000000000 W

F2 - Processing parameters
SI 131072
SF 125.777890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



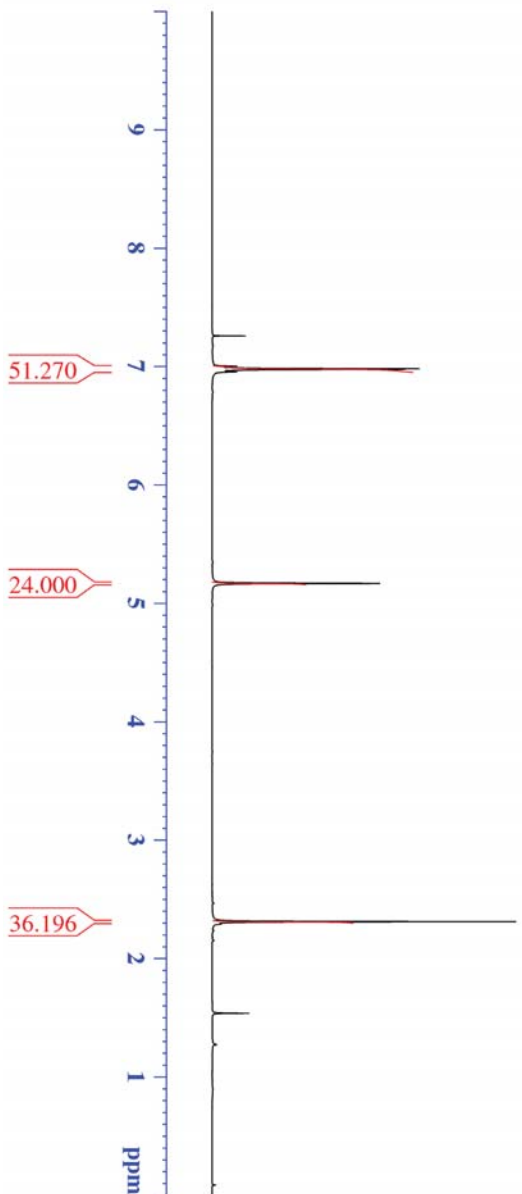
¹H NMR



7.002
6.981
6.975
6.954

5.167

2.310

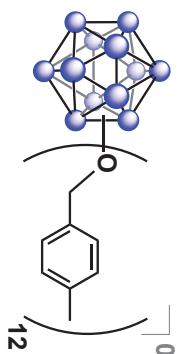


Current Data Parameters
NAME B12(O-4-Mebn)12
EXPNO 161
PROCNO 1

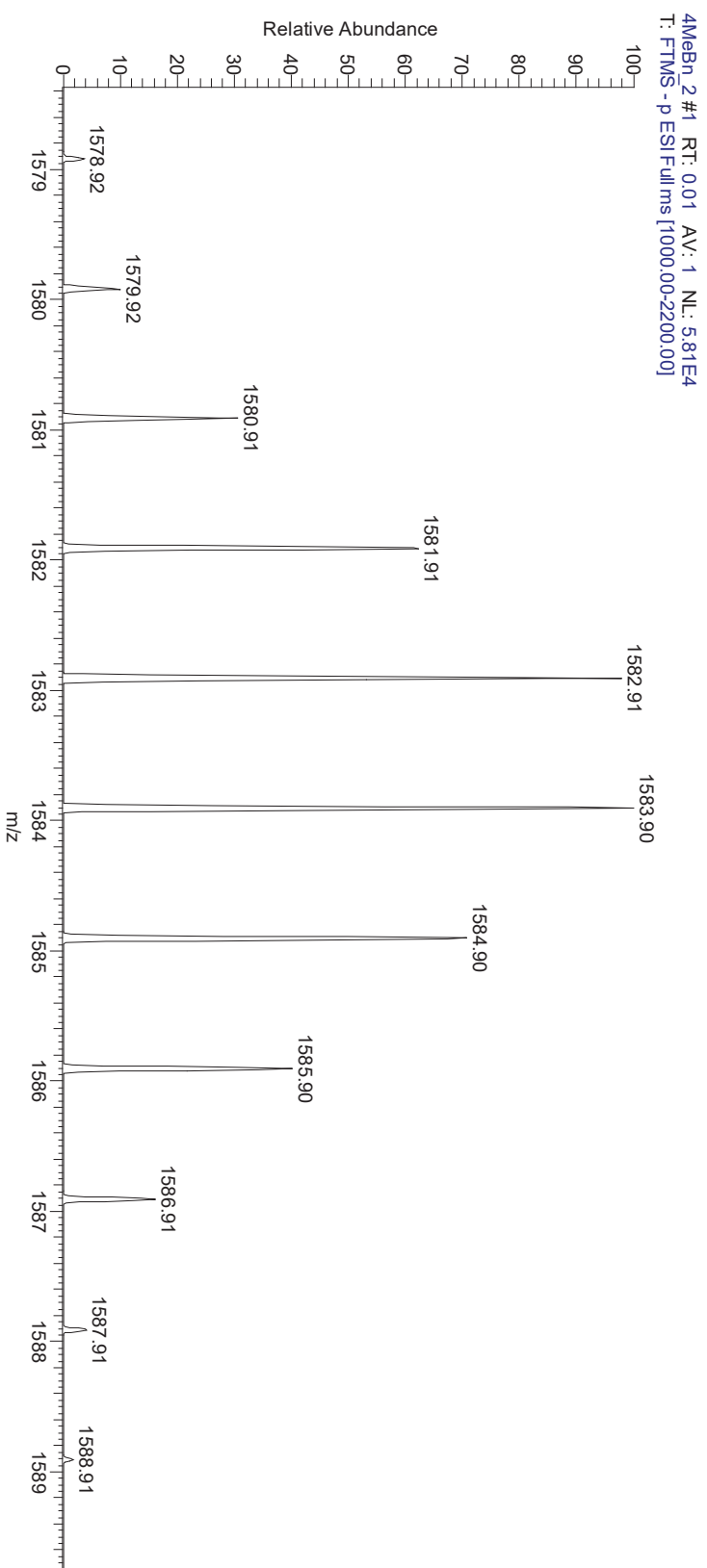
F2 - Acquisition Parameters
Date_ 20150829

Time 21.05
INSTRUM av400
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 52882
SOLVENT CDCl3
NS 32
DS 0
SWH 8012.820 Hz
FIDRES 0.151523 Hz
AQ 3.2998369 sec
RG 155.85
DW 62.400 usec
DE 6.50 usec
TE 299.0 K
D1 2.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1324008 MHz
NUC1 ¹H
Pl 15.00 usec
PLW1 13.000000000 W
F2 - Processing parameters
SI 65536
SF 400.1300184 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

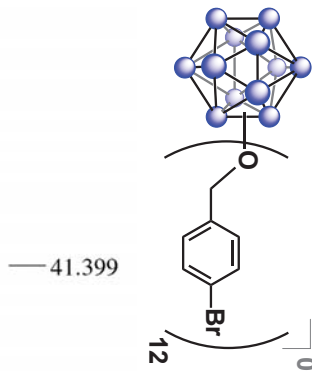


Q Exactive High-Res Mass Spec





^{11}B $\{^1\text{H}\}$ NMR



Current Data Parameters
NAME B12(O-4-BrBn)12
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150624
Time 10.50
INSTRUM av400
PROBHD 5 mm PABBO BB/
PULPROG zgpg3s
TD 5096
SOLVENT CDCl3
NS 1024
DS 0
SWH 51020.406 Hz
FIDRES 10.011854 Hz
AQ 0.0449408 sec
RG 189.85
DW 9.800 usec
DE 6.50 usec
TE 299.1 K
D1 0.00000400 sec
D11 0.03000000 sec
TD0 1

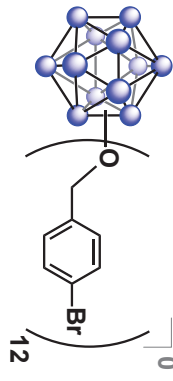
===== CHANNEL f1 =====
SFO1 128.3776052 MHz
NUC1 ^{11}B
P1 10.00 usec
PLW1 52.000000000 W

===== CHANNEL f2 =====
SFO2 400.1324008 MHz
NUC2 ^1H
CPDPRG12 waltz16
PCPD2 90.00 usec
PLW2 13.000000000 W
PLW12 0.36111000 W

F2 - Processing parameters
SI 32768
SF 128.3776050 MHz
WDW EM
SSB 0
LB 50.00 Hz
GB 0
PC 1.40



¹³C NMR



- 138.499
- 131.583
- 128.450
- 121.624
- 72.559
- 53.442



Current Data Parameters
NAME B12(O-4-BrBn)12
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151021
Time 17:14
INSTRUM av500
PROBHD 5 mm DCH 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 256
DS 2

SWH 31250.000 Hz
FIDRES 0.476837 Hz
AQ 1.0485760 sec
RG 204.54
DW 16.000 usec
DE 18.00 usec
TE 298.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

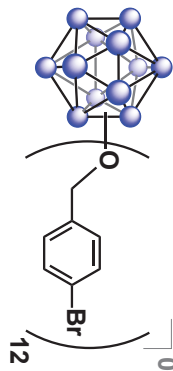
==== CHANNEL f1 =====
SFO1 125.7722511 MHz
NUC1 13C
P1 9.63 usec
PLW1 23.000000000 W

==== CHANNEL f2 =====
SFO2 500.1330008 MHz
NUC2 1H
CDDPRGf2 waltz16
PCPD2 80.00 usec
PLW2 13.500000000 W
PLW12 0.21094000 W
PLW13 0.113500001 W

F2 - Processing parameters
SF 131072
SE 125.777890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

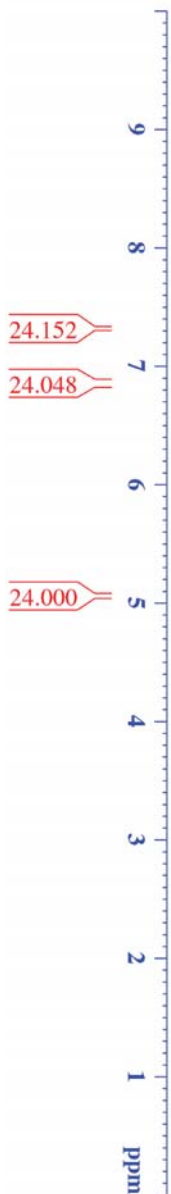


¹H NMR



7.328
7.311
6.857
6.841

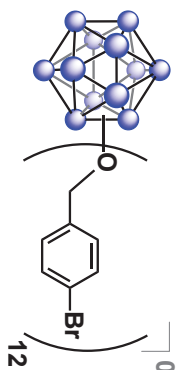
5.059



Current Data Parameters
NAME B12(O-4-BrBn)12
EXPNO 11
PROCNO 1

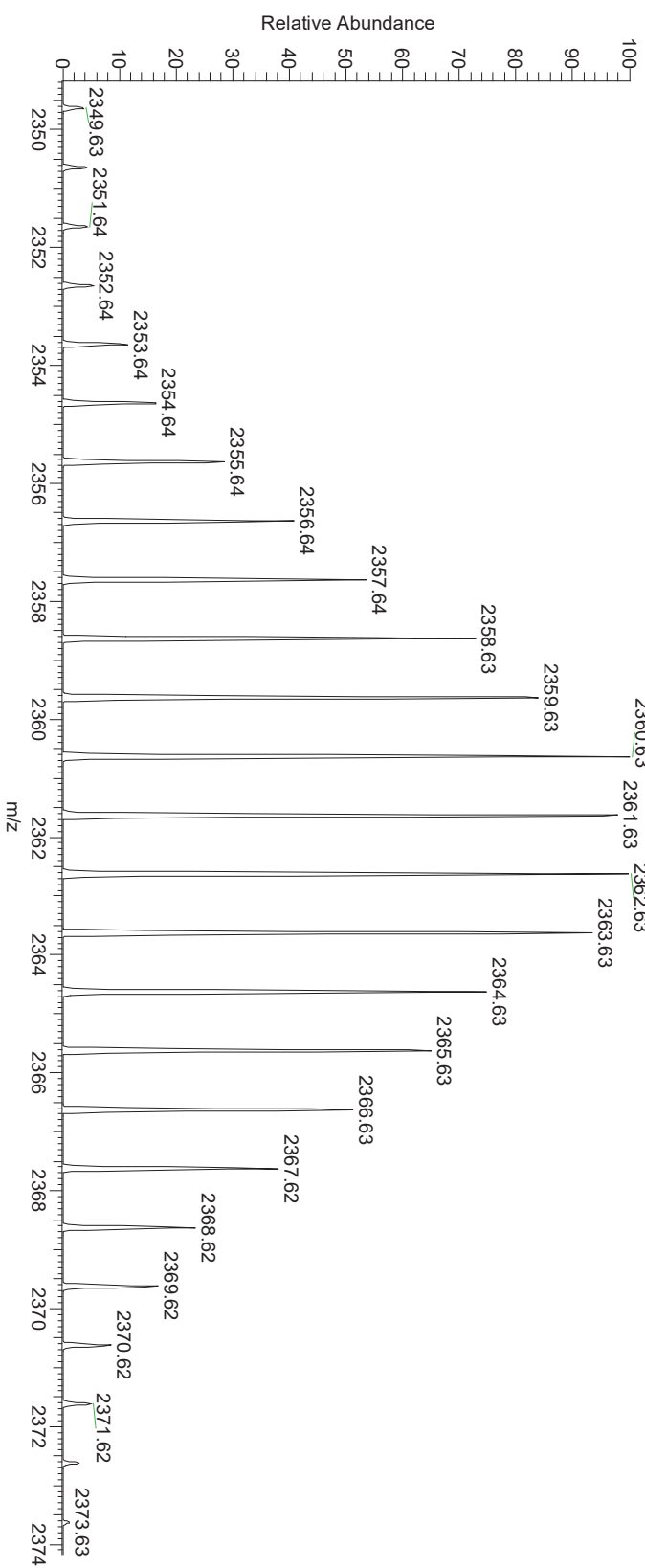
F2 - Acquisition Parameters
Date_ 20151021
Time 17:27
INSTRUM av500
PROBHD 5 mm DCH 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 32
DS 0
SWH 10000.000 Hz
FIDRES 0.152588 Hz
AQ 3.2767999 sec
RG 42.29
DW 50.000 usec
DE 10.00 usec
TE 298.0 K
D1 2.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 500.1330008 MHz
NUC1 1H
P1 10.00 usec
PLW1 13.50000000 W
F2 - Processing parameters
SI 65536
SF 500.1300146 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



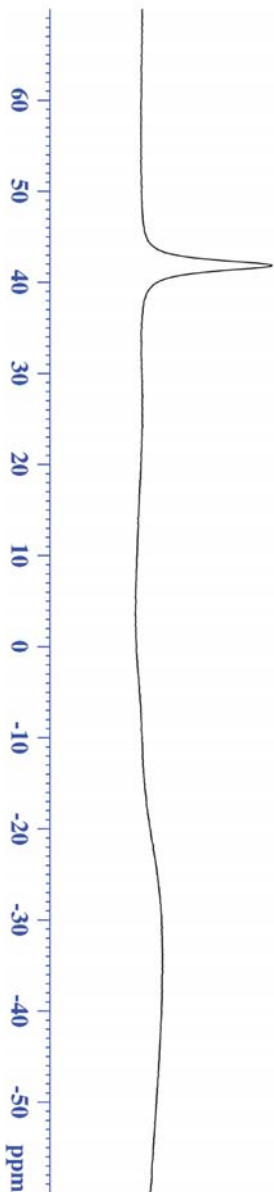
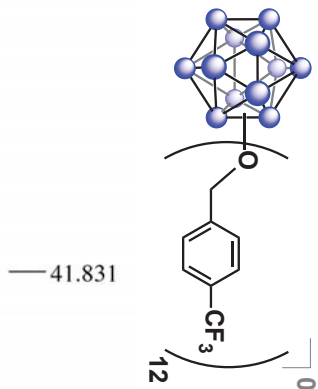
Q Exactive High-Res Mass Spec

4-BrBn#1 RT: 0.01 AV: 1 NL: 5.12E5
T: FTMS - p-ESI-Full.ms [1000.00-2800.00]





$^{11}\text{B} \{^1\text{H}\}$ NMR



Current Data Parameters
NAME B12(O-4-TFM(Bn))12
EXPNO 40
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150927
Time 14.58
INSTRUM av400
PROBHD 5 mm PABBO BB/
PULPROG zgpg3s
TD 5096
SOLVENT CDCl3
NS 1024
DS 0
SWH 51020.406 Hz
FIDRES 10.011854 Hz
AQ 0.0449408 sec
RG 189.85
DW 9.800 usec
DE 6.50 usec
TE 299.3 K
D1 0.05000000 sec
D11 0.05000000 sec
TDO 1

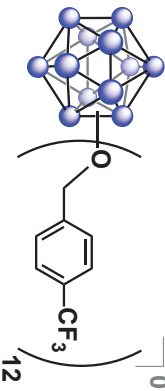
===== CHANNEL f1 =====
SFO1 128.3776052 MHz
NUC1 ^{11}B
P1 10.00 usec
PLW1 52.000000000 W

===== CHANNEL f2 =====
SFO2 400.1324008 MHz
NUC2 ^1H
CPDPRG12 waltz16
PCPD2 90.00 usec
PLW2 13.000000000 W
PLW12 0.36111000 W

F2 - Processing parameters
SI 32768
SF 128.3776161 MHz
WDW EM
SSB 0
LB 10.00 Hz
GB 0
PC 1.40

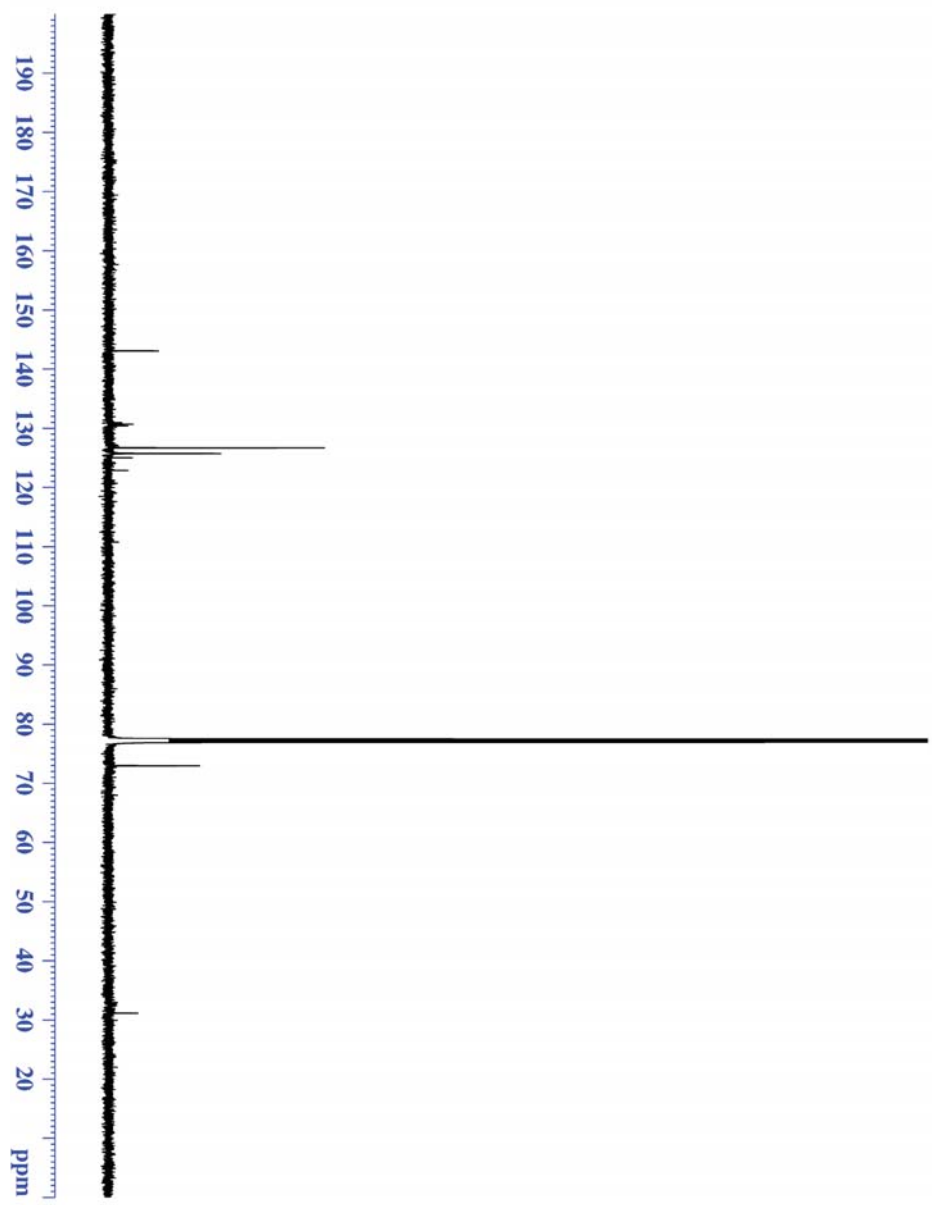


¹³C NMR



- 143.033
- 130.643
- 126.639
- 125.691
- 124.974
- 122.811

72.909



Current Data Parameters
NAME B12(O-4-TFMBr)12
EXPNO 20
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150928

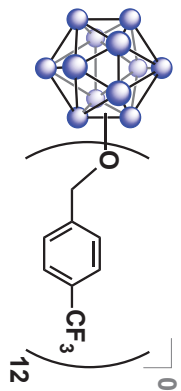
Time 13.45
INSTRUM av500
PROBHD 5 mm DGH 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 64
DS 2

SWH 31250.000 Hz
FIDRES 0.476837 Hz
AQ 1.0485760 sec
RG 204.54
DW 16.000 usec
DE 18.00 usec
TE 298.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

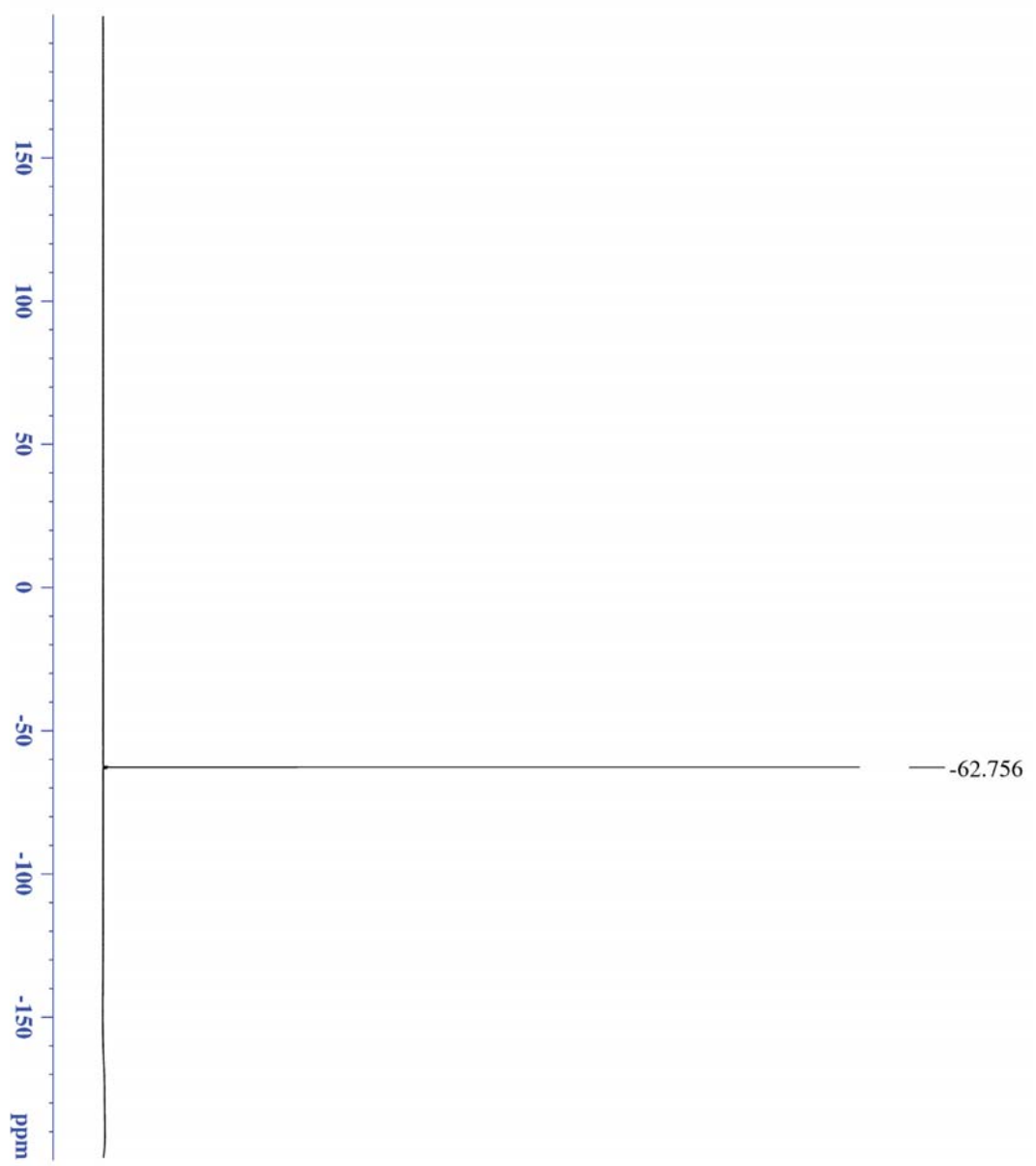
==== CHANNEL f1 =====
SFO1 125.772511 MHz
NUC1 13C
P1 9.63 usec
PLW1 23.00000000 W

==== CHANNEL f2 =====
SFO2 500.1330008 MHz
NUC2 1H
CDDPRGf2 waltz16
PCPD2 80.00 usec
PLW2 13.50000000 W
PLW12 0.21094000 W
PLW13 0.113500001 W

F2 - Processing parameters
SF 131072
SE 125.757722 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



¹⁹F NMR



Current Data Parameters
 NAME B12(O-4-TfMBn)12
 EXPNO 42
 PROCNO 1

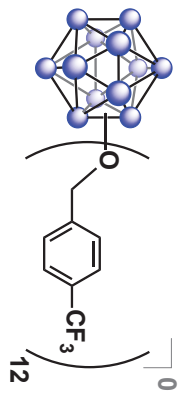
F2 - Acquisition Parameters
 Date_ 20150927
 Time 15:05
 INSTRUM av400
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 262144
 SOLVENT CDCl3
 NS 64
 DS 0
 SWH 150000.000 Hz
 FIDRES 0.572205 Hz
 AQ 0.8738133 sec
 RG 189.85
 DW 3.333 usec
 DE 6.50 usec
 TE 299.0 K
 D1 2.000000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 376.4983660 MHz
 NUC1 19F
 P1 14.50 usec
 PLW1 17.000000000 W

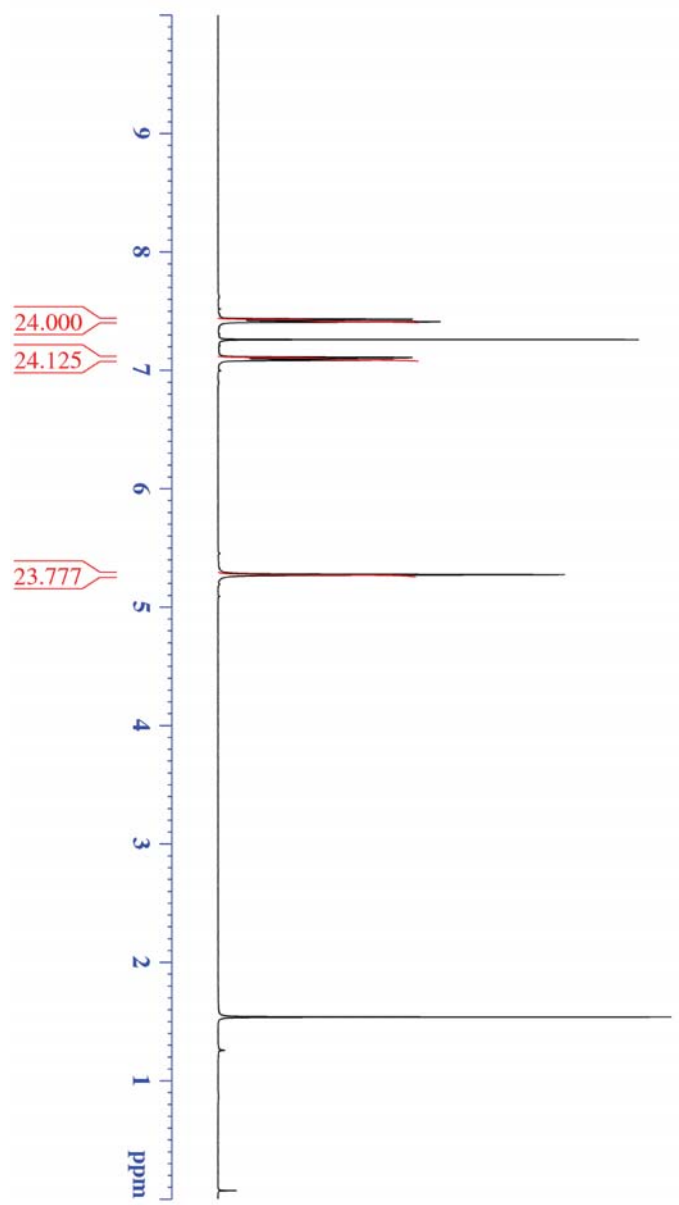
F2 - Processing parameters
 SI 262144
 SF 376.4983660 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.00



¹H NMR



7.430
7.409
7.107
7.087
5.272

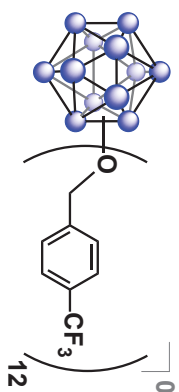


Current Data Parameters
NAME B12(O-4-TFMbn)12
EXPNO 41
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150927
Time 15.01
INSTRUM av400
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 52882
SOLVENT CDCl3
NS 32
DS 0
SWH 8012.820 Hz
FIDRES 0.151523 Hz
AQ 3.2998369 sec
RG 189.85
DW 62.400 usec
DE 6.50 usec
TE 299.0 K
D1 2.00000000 sec
TD0 1

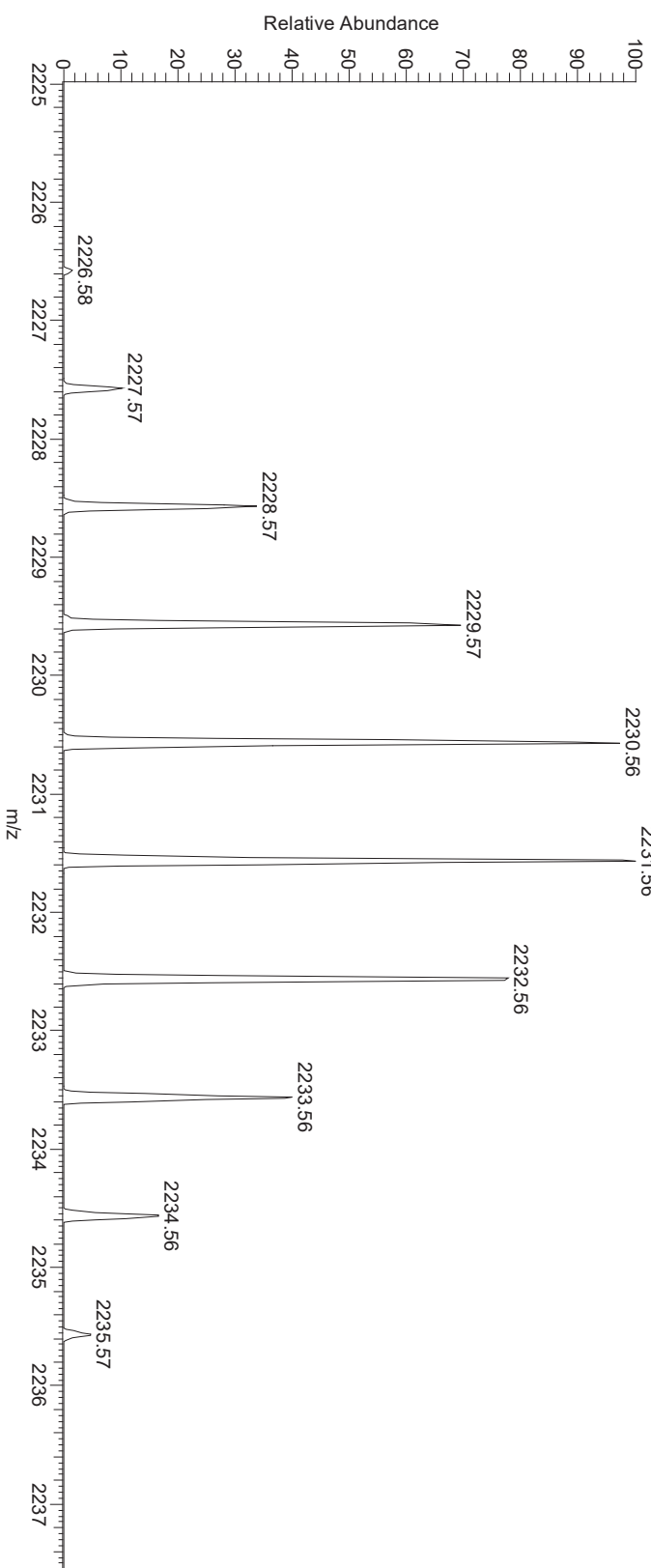
==== CHANNEL f1 =====
SFO1 400.1324008 MHz
NUC1 ¹H
Pl 15.00 usec
PLW1 13.000000000 W

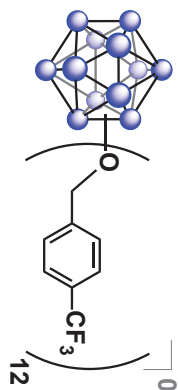
F2 - Processing parameters
SI 65536
SF 400.1300184 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



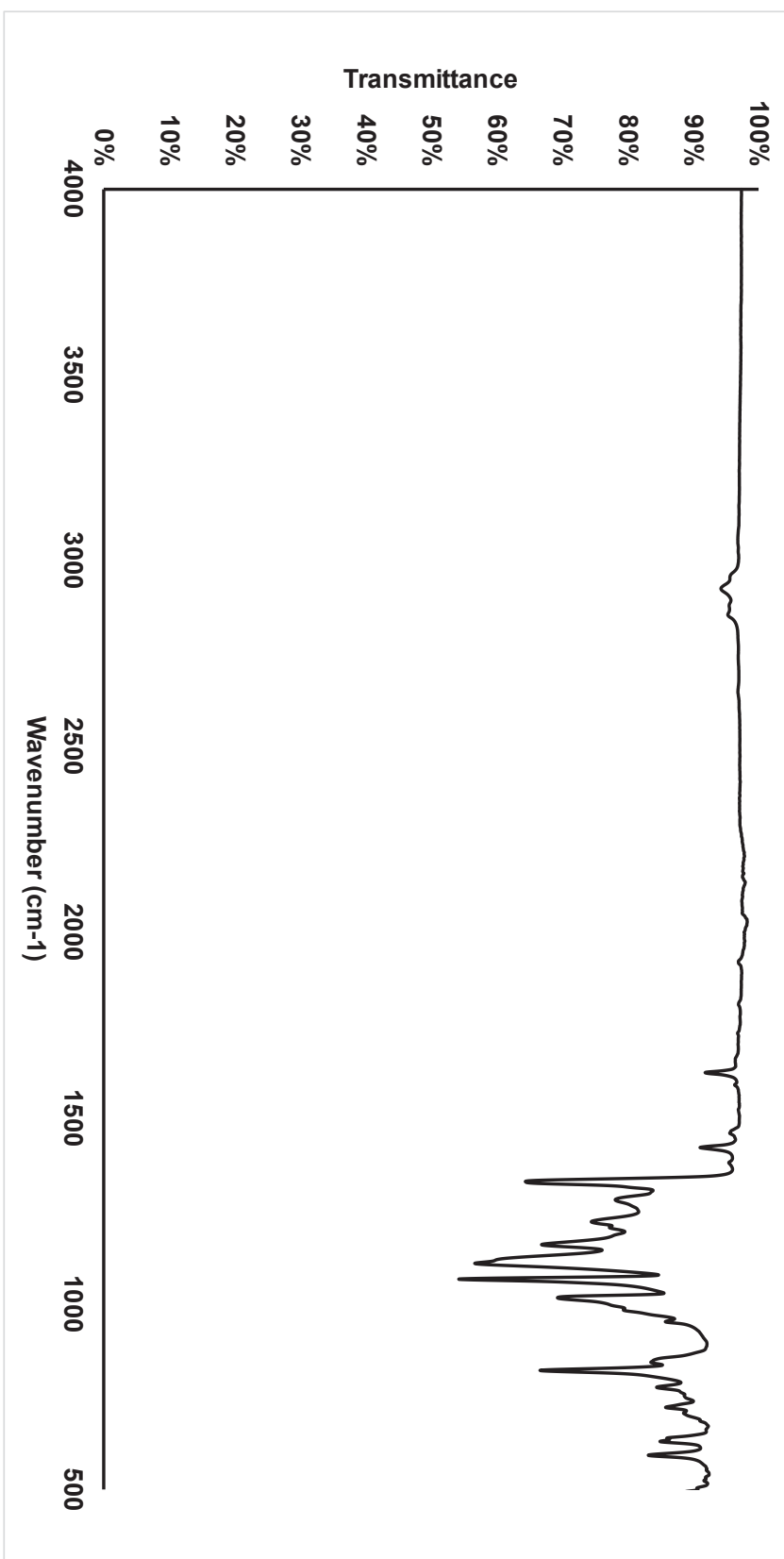
Q Exactive High-Res Mass Spec

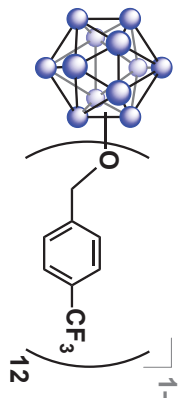
4-TFMBn#1 RT: 0.01 AV: 1 NL: 3.09E4
T: FTMS - p-ESI Fullms [800.00-2500.00]



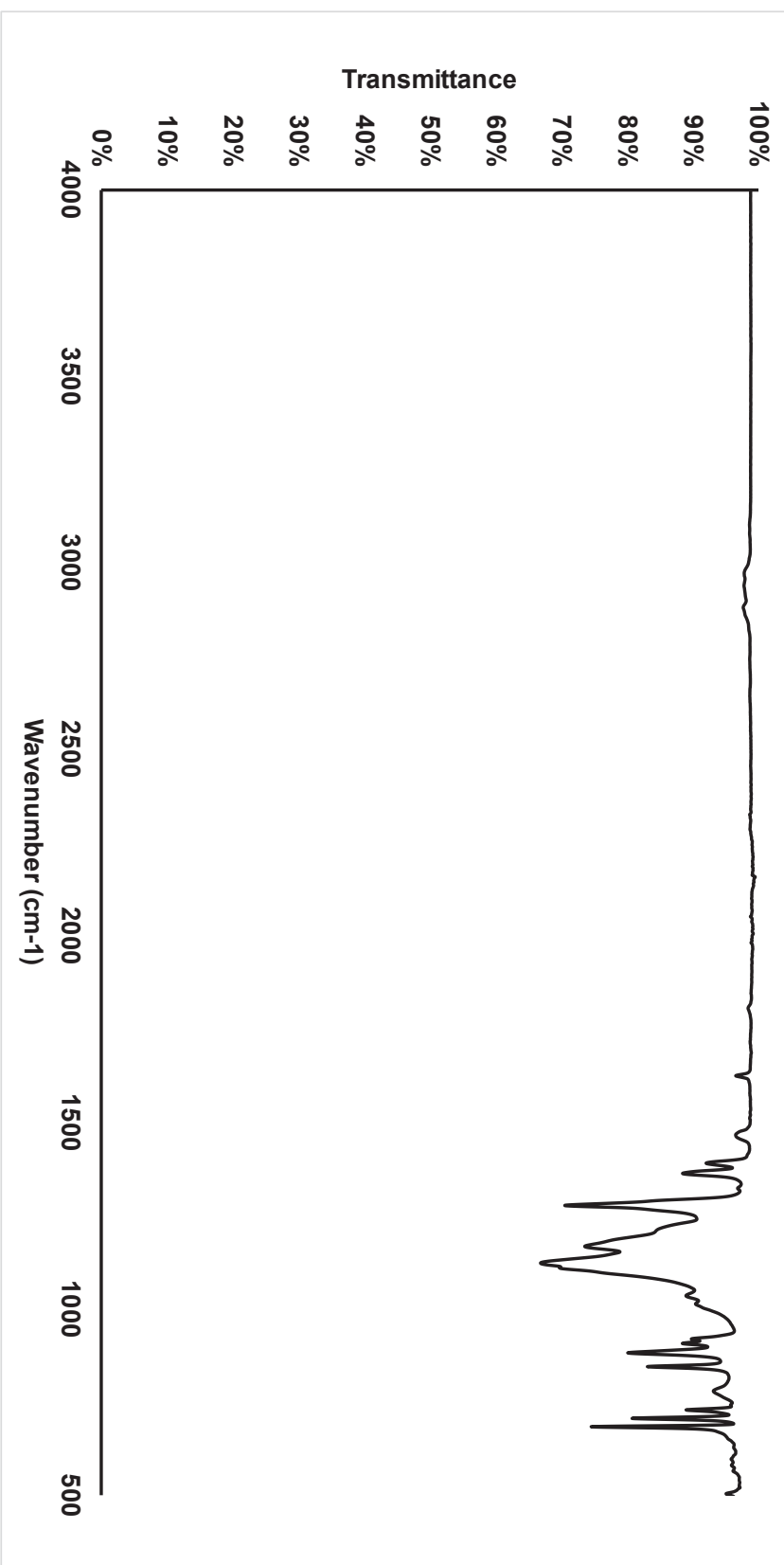


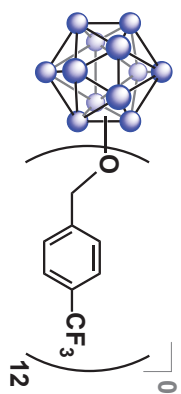
IR



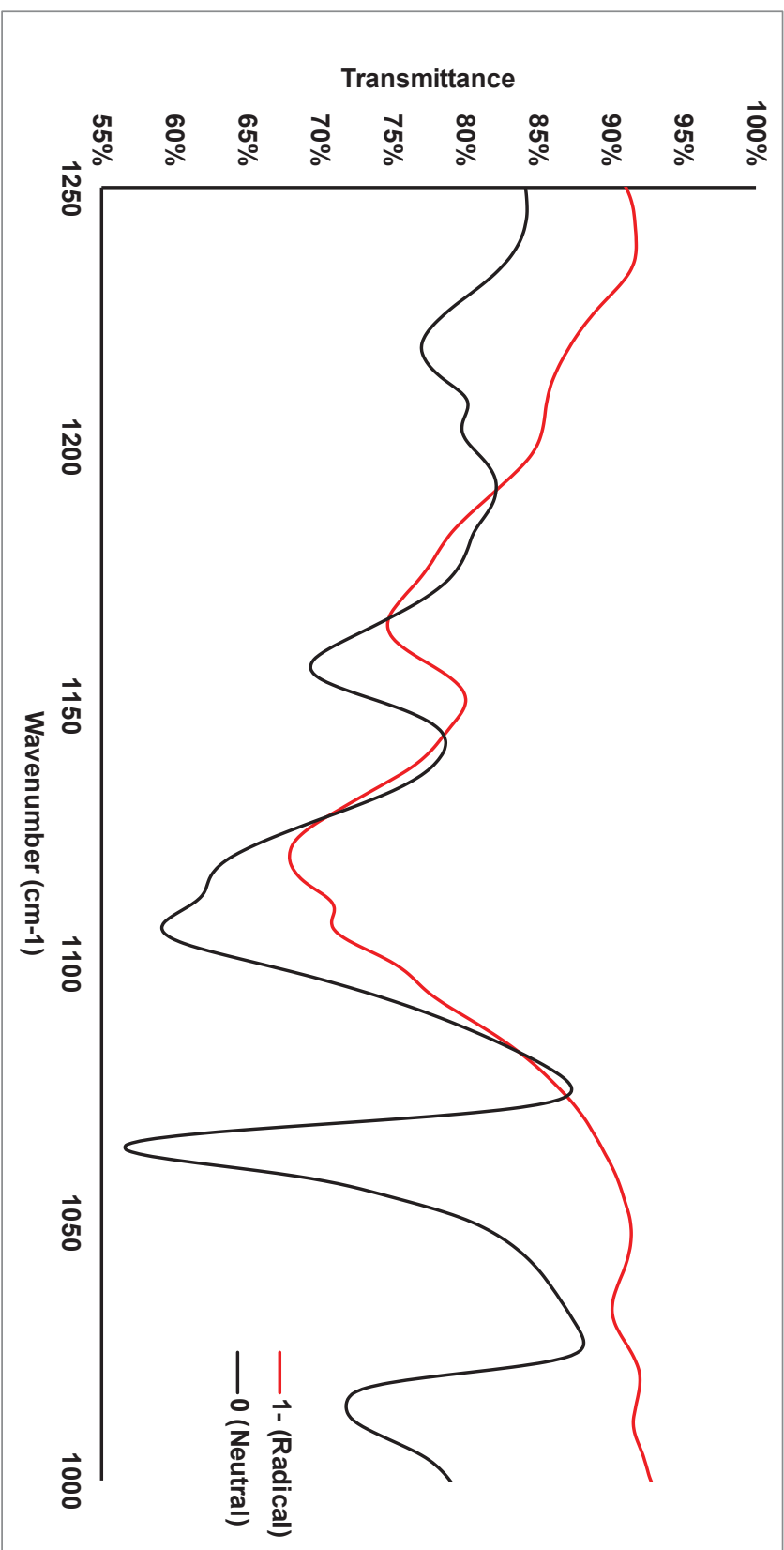


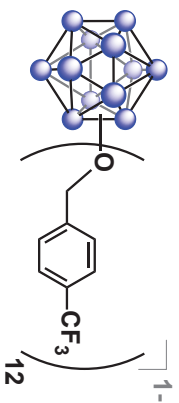
IR



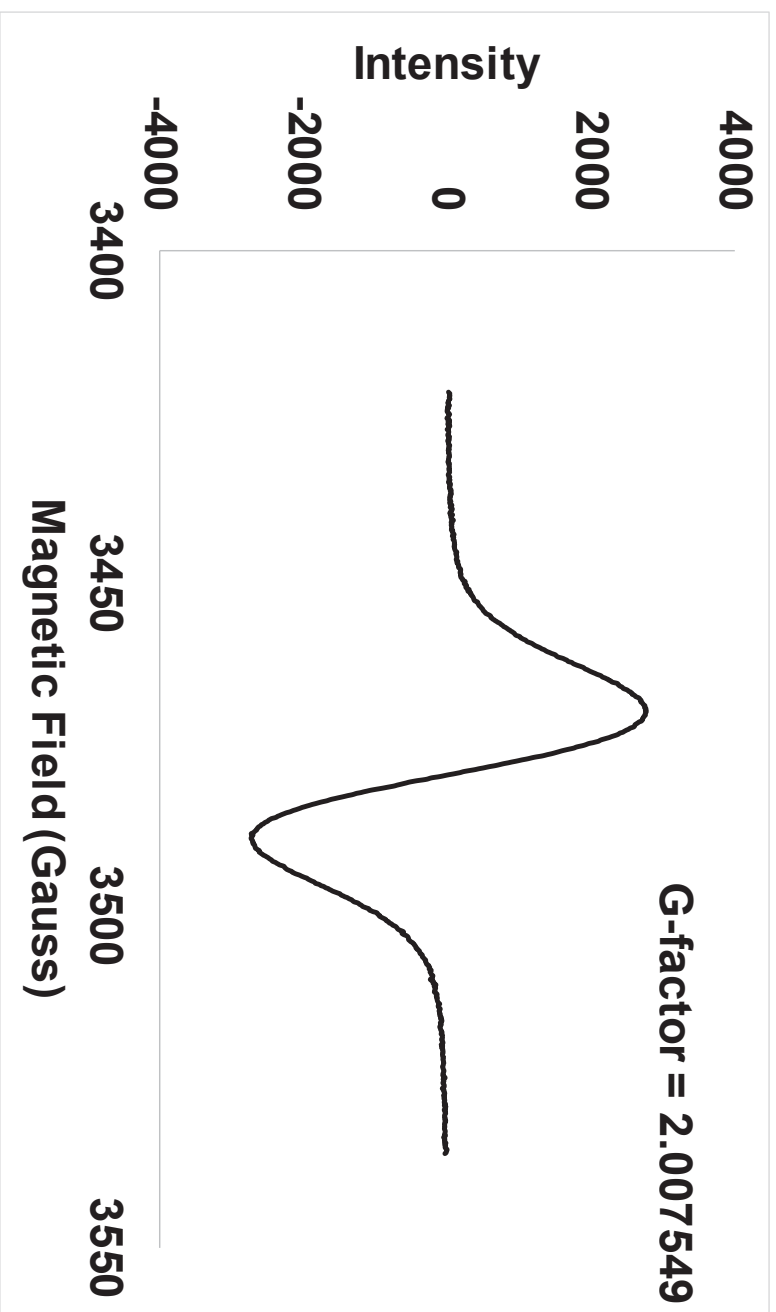


IR

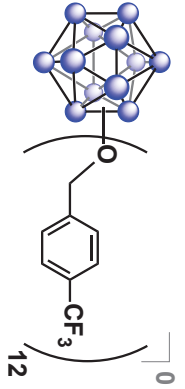




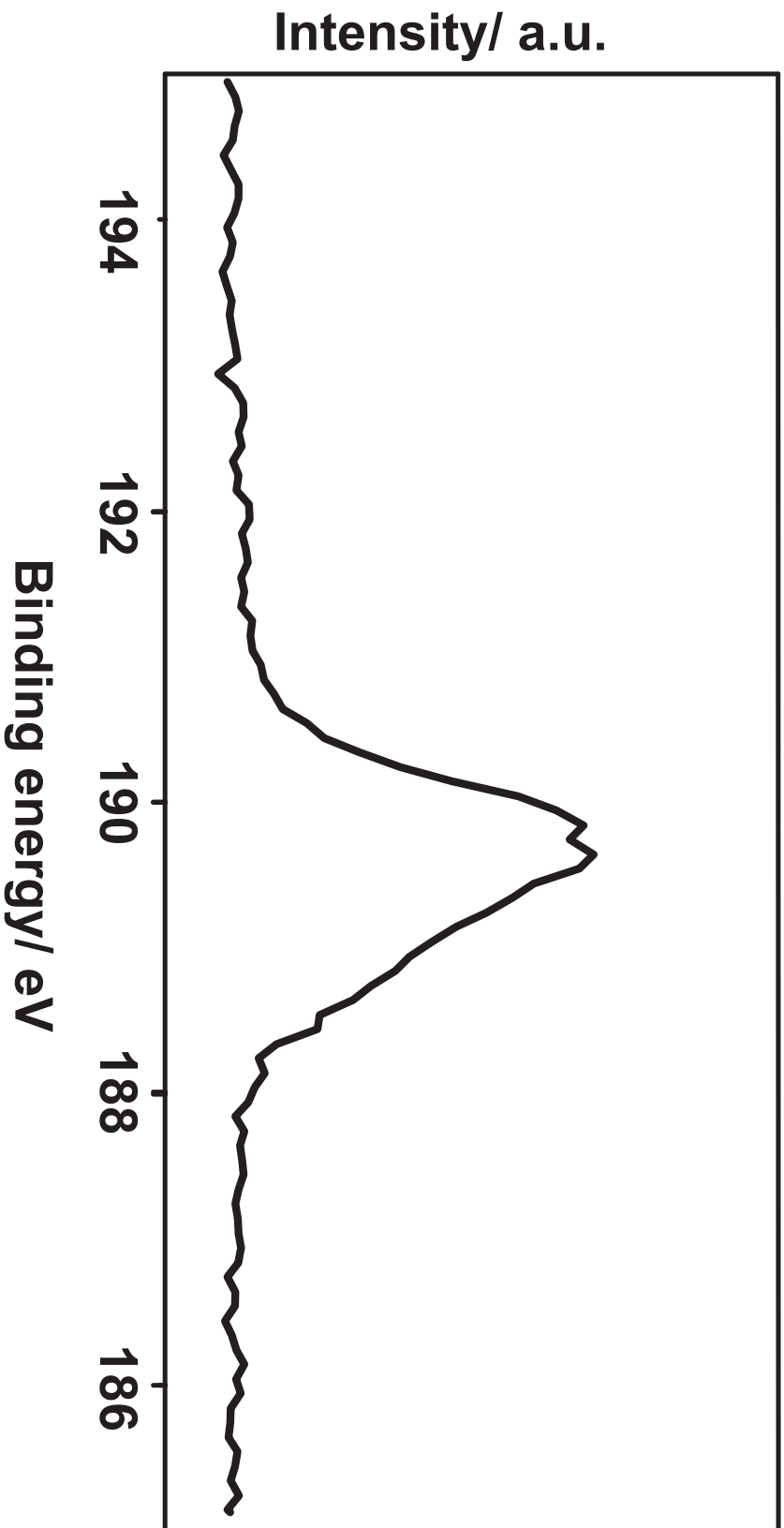
EPR

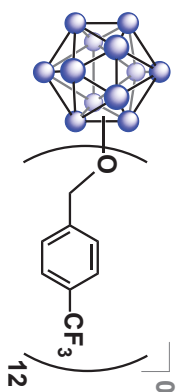


DOS Format
 ANZ 1024
 MIN -2733.186523
 MAX 2772.813477
 JSS 0
 GST 3421.274884
 GSI 114.543004
 JUN G
 JON Bruker BioSpin GmbH
 JDA 9/25/2015
 JTM 13:33
 JRE c:\programfiles\bruker-
 emx\syscal\st0103.cal
 JEX field-sweep
 JSD 1
 HCF 3478.546386
 HSW 114.543004
 EMF 0
 RCT 20.48
 RTC 20.48
 RRG 8.93E+03
 RMA 4
 MF 9.773981
 MP 6.38E-01
 MPD 25

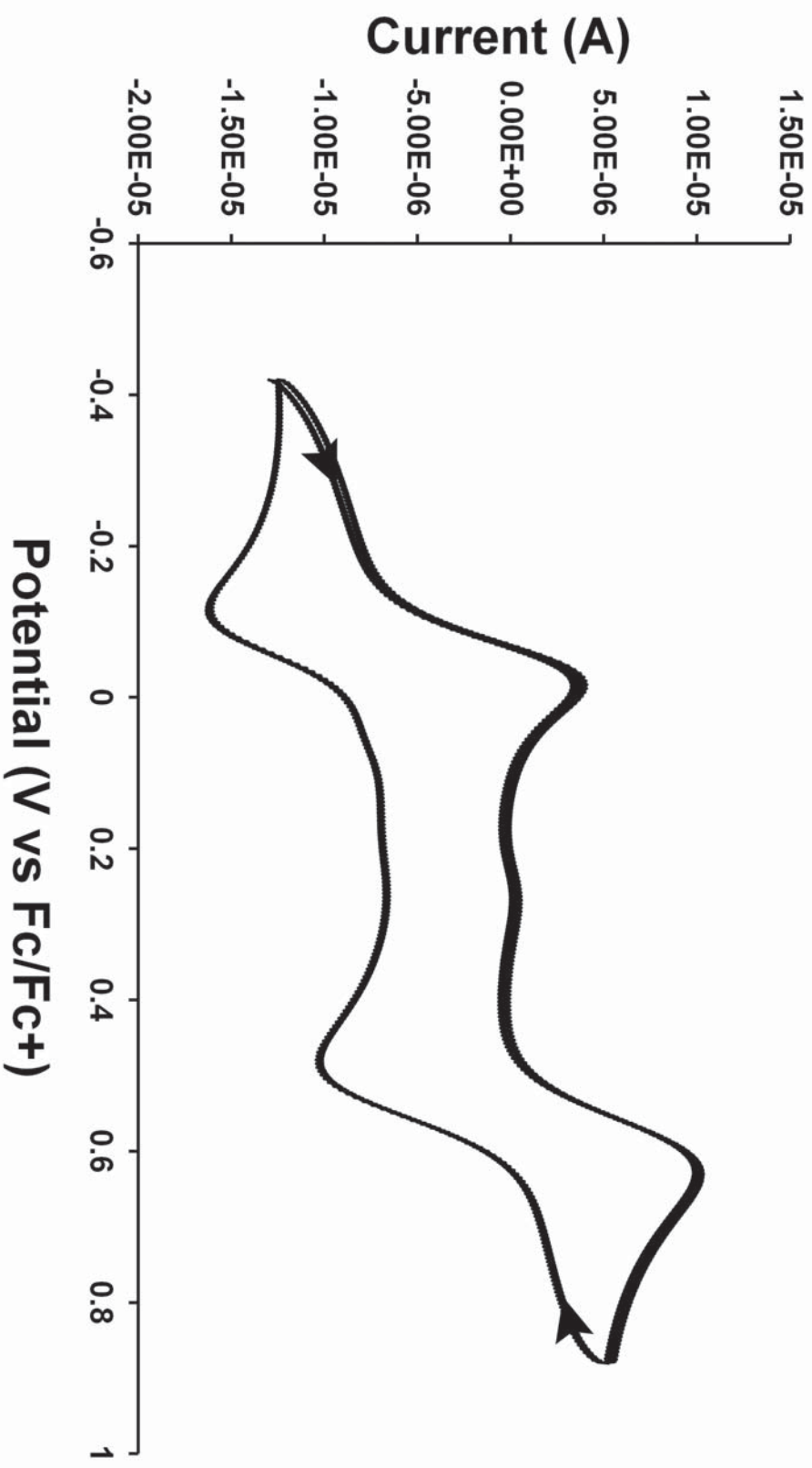


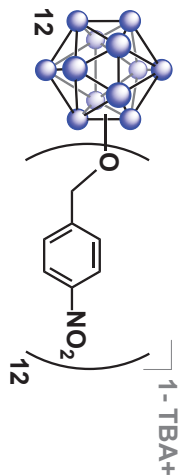
B 1s XPS





Cyclic voltammetry





¹¹B {¹H} NMR



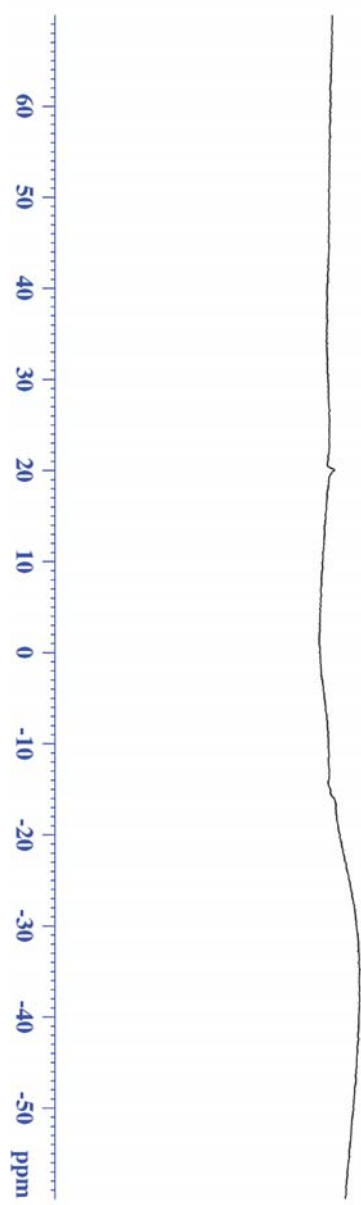
Current Data Parameters
 NAME B12(O-4-NO2Bn)12
 EXPNO 60
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20151101
 Time 19:40
 INSTRUM av400
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg3s
 TD 5096
 SOLVENT Acetone
 NS 1024
 DS 0
 SWH 51020.406 Hz
 FIDRES 10.011854 Hz
 AQ 0.0449408 sec
 RG 189.85
 DW 9.800 usec
 DE 6.50 usec
 TE 299.0 K
 D1 0.05000000 sec
 D11 0.05000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 128.3776052 MHz
 NUC1 11B
 P1 10.00 usec
 PLW1 52.000000000 W

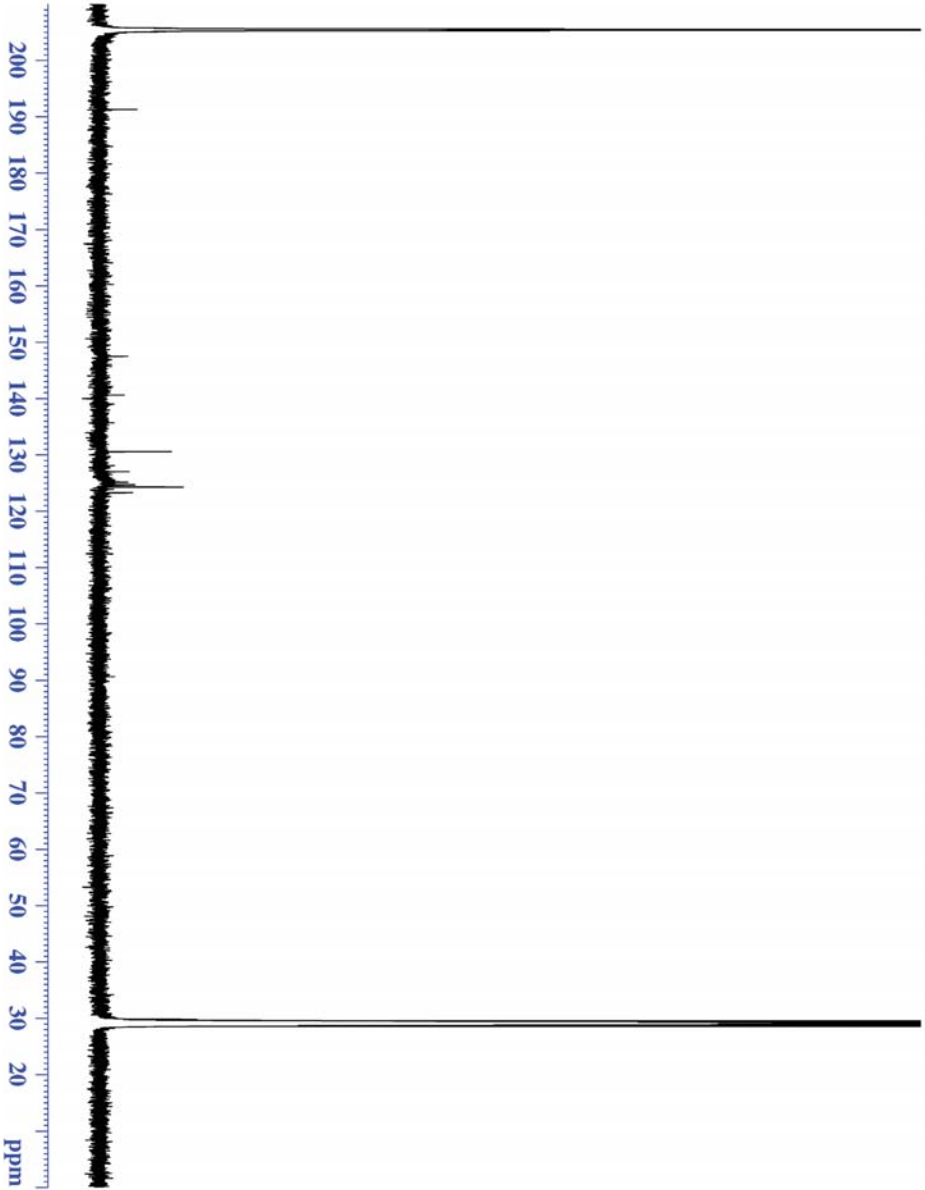
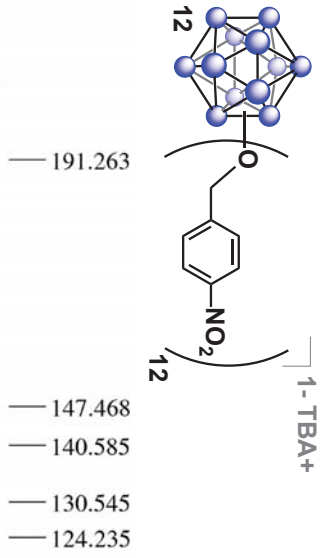
===== CHANNEL f2 =====
 SFO2 400.1324008 MHz
 NUC2 1H
 CPDPRG12 waltz16
 PCTPD2 90.00 usec
 PLW2 13.000000000 W
 PLW12 0.36111000 W

F2 - Processing parameters
 SI 32768
 SF 128.3776161 MHz
 WDW EM
 SSB 0
 LB 10.00 Hz
 GB 0
 PC 1.40





¹³C NMR



Current Data Parameters
NAME B12(O-4-NO2Bn)12
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151103
Time 11:12

INSTRUM av500
PROBHD 5 mm DGH 13C-1
PULPROG zgpg30
TD 65536
SOLVENT Acetone
NS 128
DS 2

SWH 31250.000 Hz
FIDRES 0.476837 Hz
AQ 1.0485760 sec
RG 204.54
DW 16.000 usec
DE 18.00 usec
TE 298.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

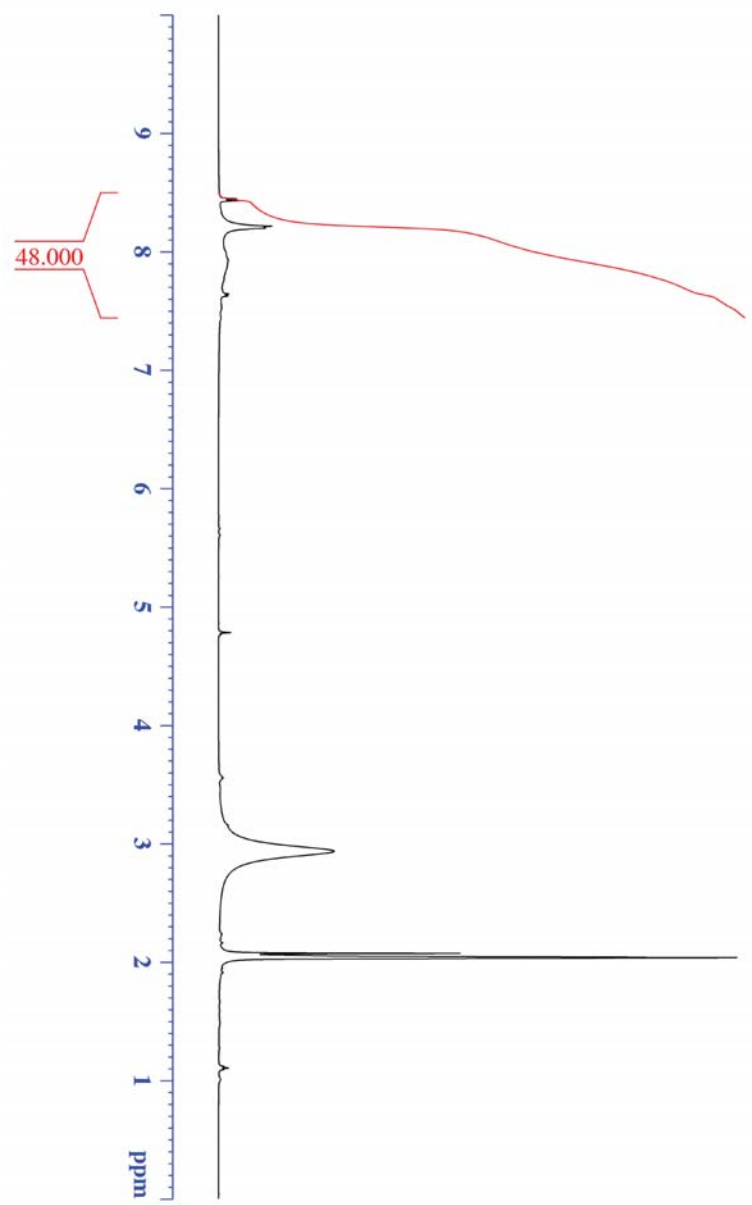
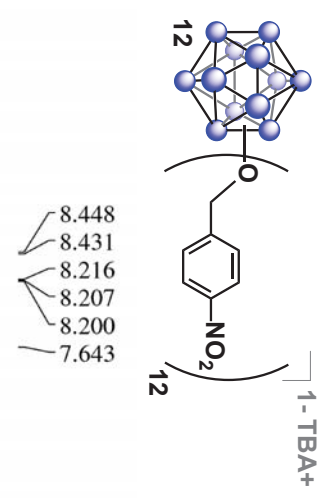
==== CHANNEL f1 =====
SFO1 125.7722511 MHz
NUC1 13C
P1 9.63 usec
PLW1 23.000000000 W

==== CHANNEL f2 =====
SFO2 500.1330008 MHz
NUC2 1H
CDDPRGf2 waltz16
PCPD2 80.00 usec
PLW2 13.500000000 W
PLW12 0.21094000 W
PLW13 0.113500001 W

F2 - Processing parameters
SF 131072
SE 125.7577890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



¹H NMR



Current Data Parameters
NAME B12(O-4-NO2Bn)12
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151103
Time 11.16

INSTRUM av500
PROBHD 5 mm DCH 13C-1
PULPROG zg30

TD 65536
SOLVENT Acetone
NS 32

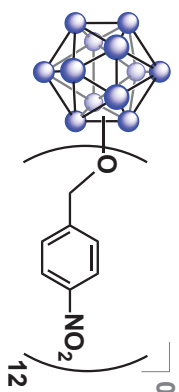
DS 0
SWH 10000.000 Hz
FIDRES 0.152588 Hz

AQ 3.2767999 sec
RG 52.41
DE 50.000 usec

TE 298.0 K
D1 2.000000000 sec
TD0 1

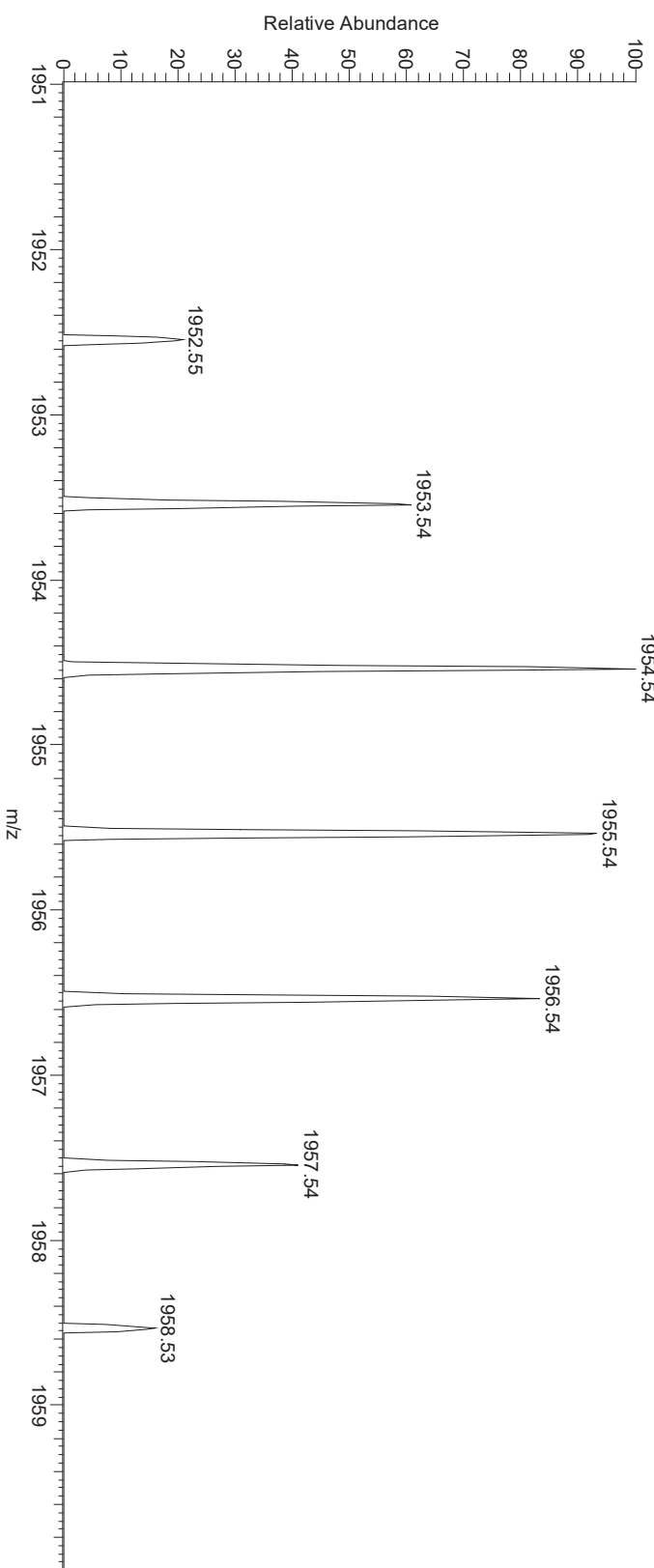
==== CHANNEL f1 =====
SFO1 500.1330008 MHz
NUC1 1H
P1 10.00 usec
PLW1 13.500000000 W

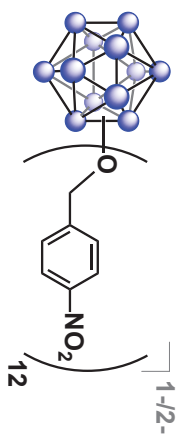
F2 - Processing parameters
SI 65536
SF 500.1300146 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



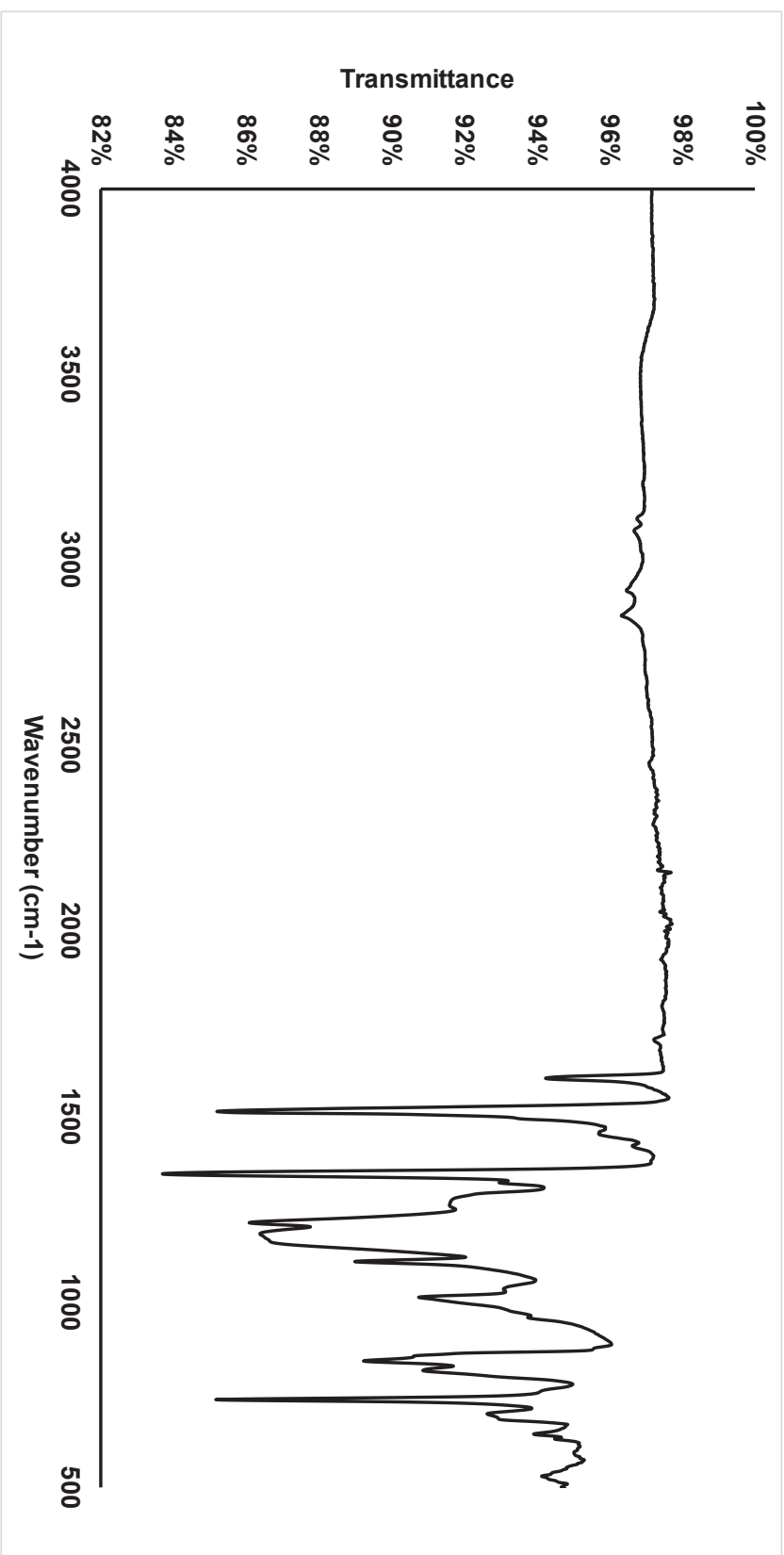
Q Exactive High-Res Mass Spec

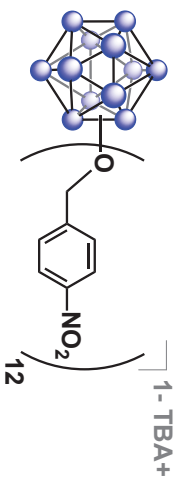
Nitro #1 RT: 0.01 AV: 1 NL: 3.75E5
T: FTMS - p-ESI-Full.ms [500.00-2500.00]



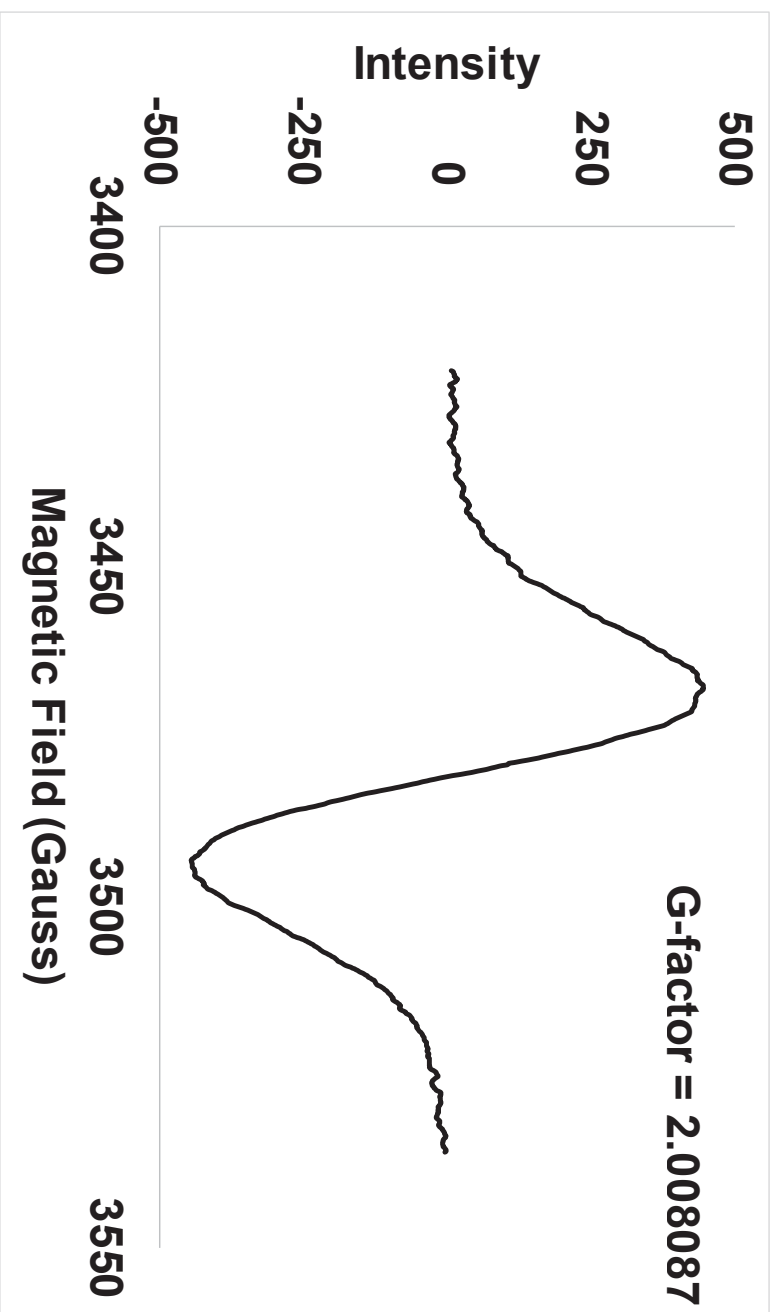


IR

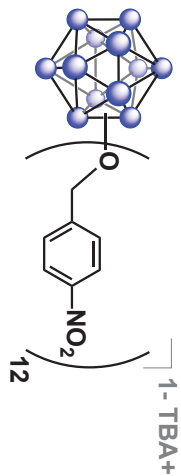




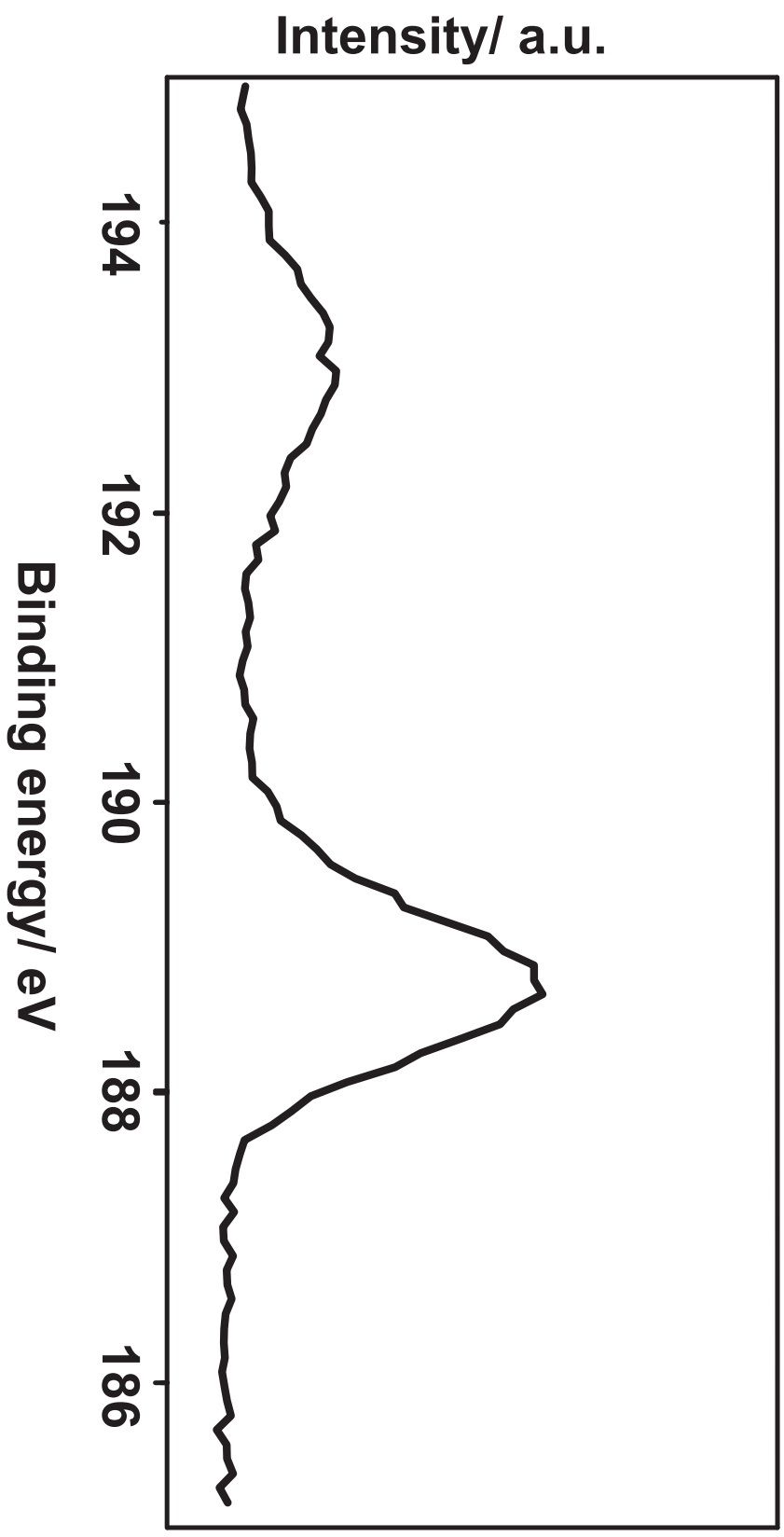
EPR

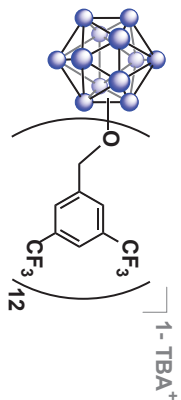


DOS Format
 ANZ 1024
 MIN -445.813477
 MAX 444.186523
 JSS 0
 GST 3421.28
 GSI 114.54
 JUN G
 JON Bruker Biospin GmbH
 JDA 9/29/2015
 JTM 16:26
 JRE c:\programfiles\bruker-
 emx\syscal\st0103.cal
 JEX field-sweep
 JSD 1
 HCF 3478.55
 HSW 114.54
 EMF 0
 RCT 20.48
 RTC 163.84
 RRG 8.93E+03
 RMA 4
 MF 9.776609
 MP 6.38E-01
 MPD 25



B 1s XPS





¹¹B {¹H} NMR



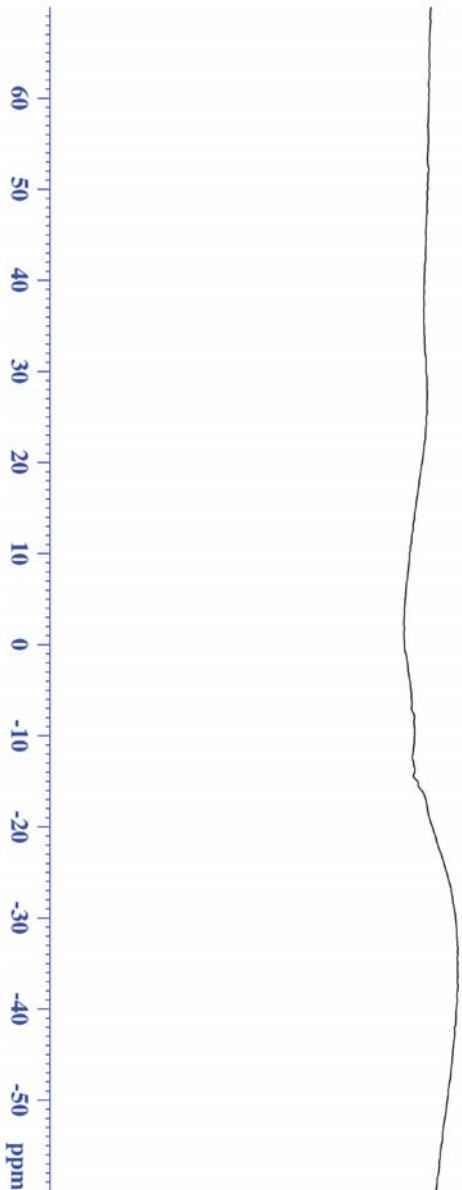
Current Data Parameters
 NAME B12(O-3,5-bis(TMBn))12
 EXPNO 210
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150824
 Time 20.51
 INSTRUM av400
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg3s
 TD 5096
 SOLVENT CDCl3
 NS 1024
 DS 0
 SWH 51020.406 Hz
 FIDRES 10.011854 Hz
 AQ 0.0449408 sec
 RG 189.85
 DW 9.800 usec
 DE 6.50 usec
 TE 299.2 K
 D1 0.00000400 sec
 D11 0.030000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 128.3776052 MHz
 NUC1 ¹¹B
 P1 10.00 usec
 PLW1 52.000000000 W

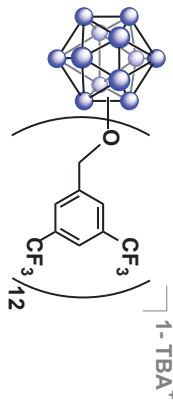
===== CHANNEL f2 =====
 SFO2 400.1324008 MHz
 NUC2 ¹H
 CPDPRG12 waltz16
 PCTPD2 90.00 usec
 PLW2 13.000000000 W
 PLW12 0.36111000 W

F2 - Processing parameters
 SI 32768
 SF 128.3776050 MHz
 WDW EM
 SSB 0
 LB 50.00 Hz
 GB 0
 PC 1.40





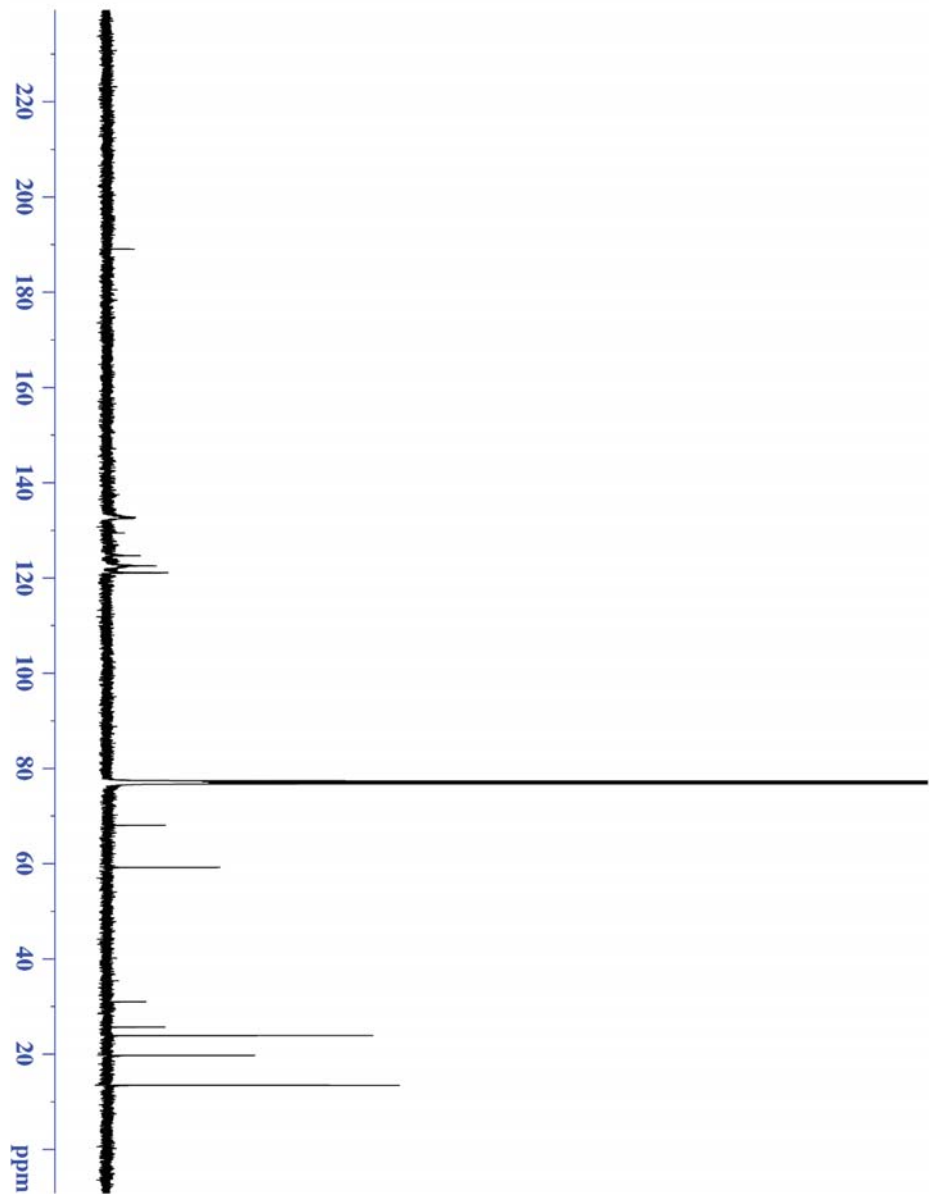
¹³C NMR



- 132.703
- 132.488
- 129.392
- 124.631
- 122.473
- 121.043

- 67.990
- 59.174
- 59.152
- 59.130

- 30.943
- 25.615
- 23.829
- 19.671
- 19.661
- 13.398



Current Data Parameters
 NAME B12(O-3,5-bis(TFMBn))12
 EXPNO 300
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150829
 Time 21.11

INSTRUM av500
 PROBHID 5 mm DCH 13C-1
 PULPROG zgpg30

TD 65536
 SOLVENT CDCl3
 NS 256

DS 2
 SWH 31250.000 Hz
 FIDRES 0.476837 Hz

AQ 1.0485760 sec
 RG 204.54
 DW 16.000 usec

DE 18.00 usec
 TE 298.0 K
 D1 2.00000000 sec
 D11 0.030000000 sec
 TD0 1

==== CHANNEL f1 =====
 SFO1 125.7722511 MHz
 NUC1 13C
 P1 9.63 usec
 PLW1 23.000000000 W

==== CHANNEL f2 =====
 SFO2 500.1330008 MHz
 NUC2 1H
 P2 13.000000000 W

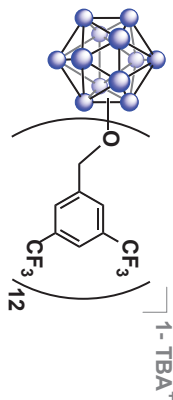
==== CHANNEL f3 =====
 SFO3 125.7722511 MHz
 NUC3 13C
 P3 9.63 usec
 PLW3 23.000000000 W

==== CHANNEL f4 =====
 SFO4 500.1330008 MHz
 NUC4 1H
 P4 13.000000000 W

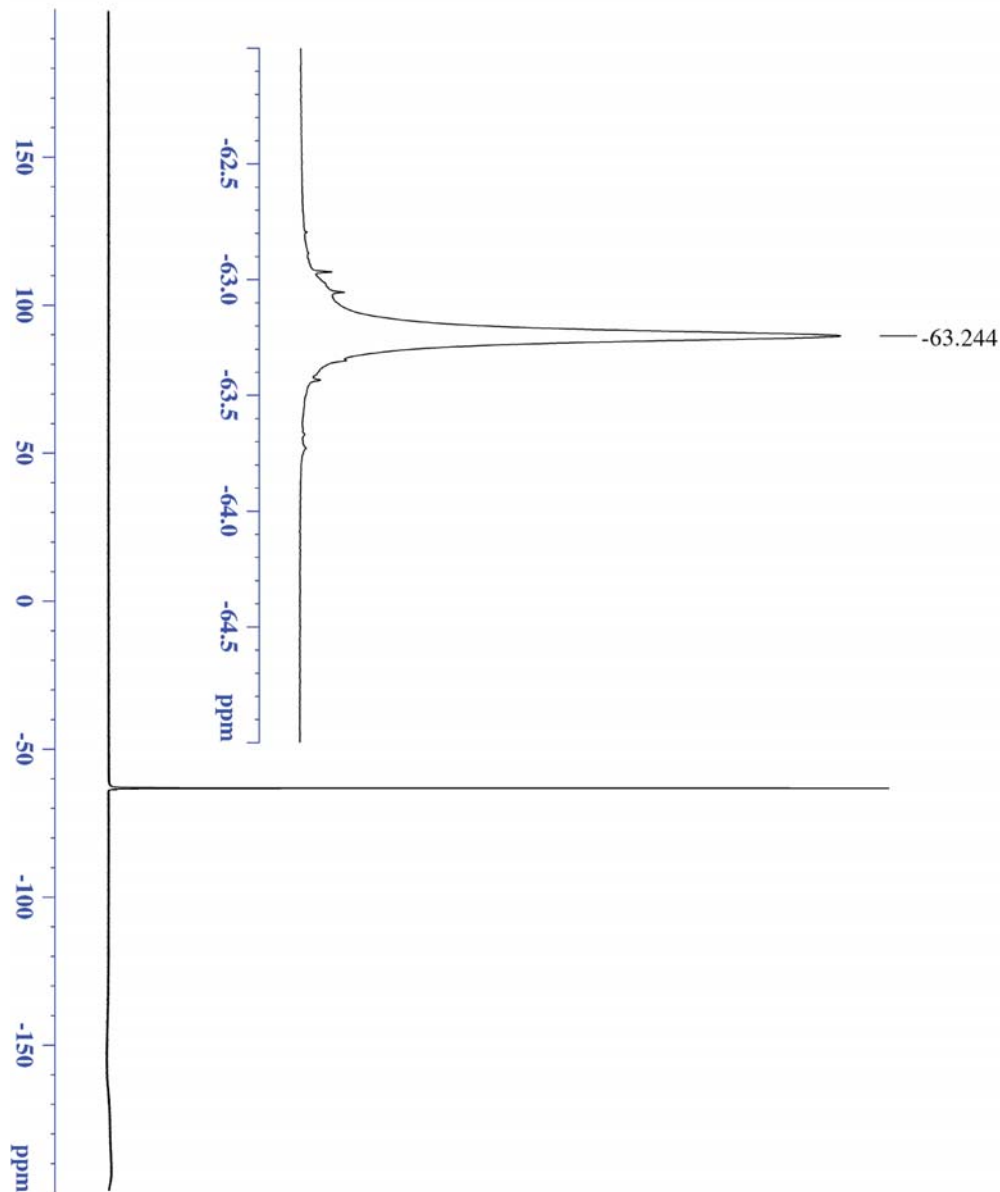
==== CHANNEL f5 =====
 SFO5 125.7722511 MHz
 NUC5 13C
 P5 9.63 usec
 PLW5 23.000000000 W

==== CHANNEL f6 =====
 SFO6 500.1330008 MHz
 NUC6 1H
 P6 13.000000000 W

F2 - Processing parameters
 SF 131.072
 SE 125.777890 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



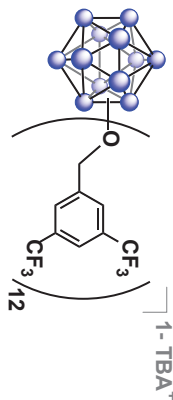
¹⁹F NMR



Current Data Parameters
 NAME B12(O-3,5-bisTFMBn)12
 EXPNO 212
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150824
 Time 21:00
 INSTRUM av400
 PROBHID 5 mm PABBO BB/
 PULPROG zgpg30
 TD 262144
 SOLVENT CDCl3
 NS 64
 DS 0
 SWH 150000.000 Hz
 FIDRES 0.572205 Hz
 AQ 0.8738133 sec
 RG 189.85
 DW 3.333 usec
 DE 6.50 usec
 TE 299.0 K
 D1 2.000000000 sec
 TD0 1

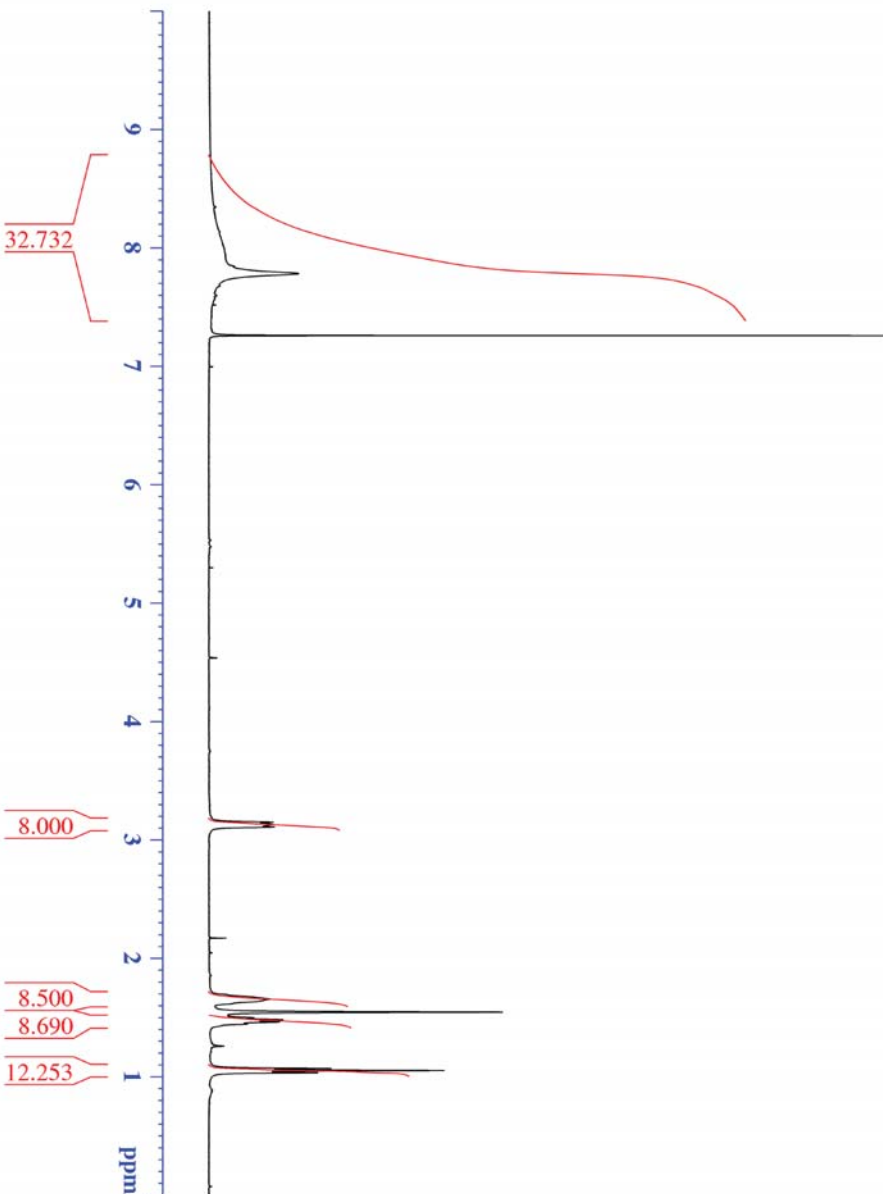
===== CHANNEL f1 =====
 SFO1 376.4983660 MHz
 NUC1 19F
 P1 14.50 usec
 PLW1 17.000000000 W
 F2 - Processing parameters
 SI 262144
 SF 376.4983660 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.00



¹H NMR

3.148
3.128
3.107

1.653
1.478
1.461
1.068
1.050
1.032



Current Data Parameters
NAME B12(O-3,5-bis(TFMBn))12
EXPNO 211
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150824

Time 20.54
INSTRUM av400
PROBHD 5 mm PABBO BB/
PULPROG zg30

TD 52882
SOLVENT CDCl3
NS 32

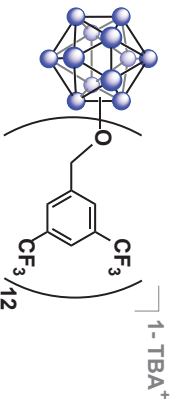
DS 0
SWH 8012.820 Hz
FIDRES 0.151523 Hz

AQ 3.2998369 sec
RG 189.85
DE 62.400 usec
TE 299.0 K
D1 2.000000000 sec
TD0 1

==== CHANNEL f1 ====

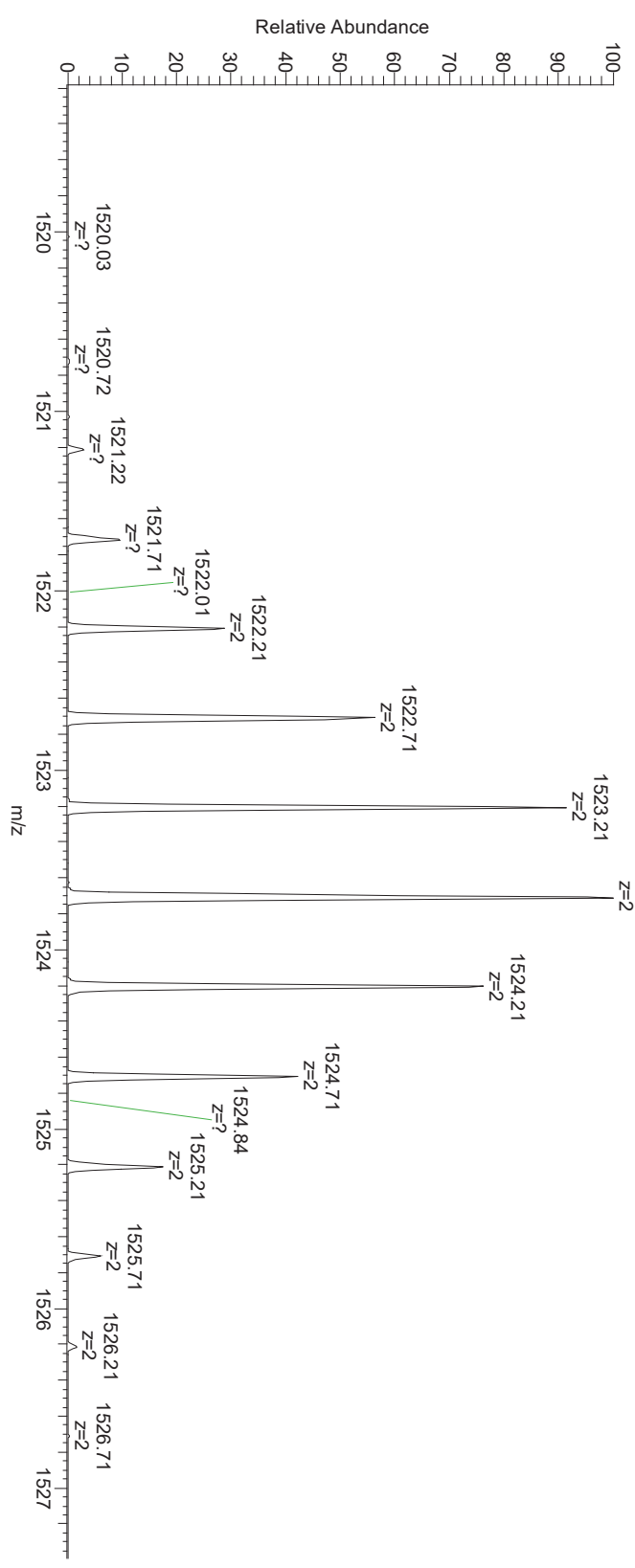
SFO1 400.1324008 MHz
NUC1 1H
P1 15.00 usec
PLW1 13.000000000 W

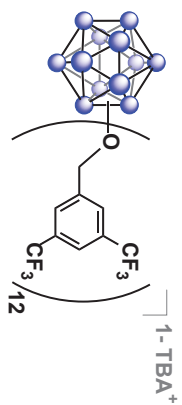
F2 - Processing parameters
SI 65536
SF 400.1300184 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



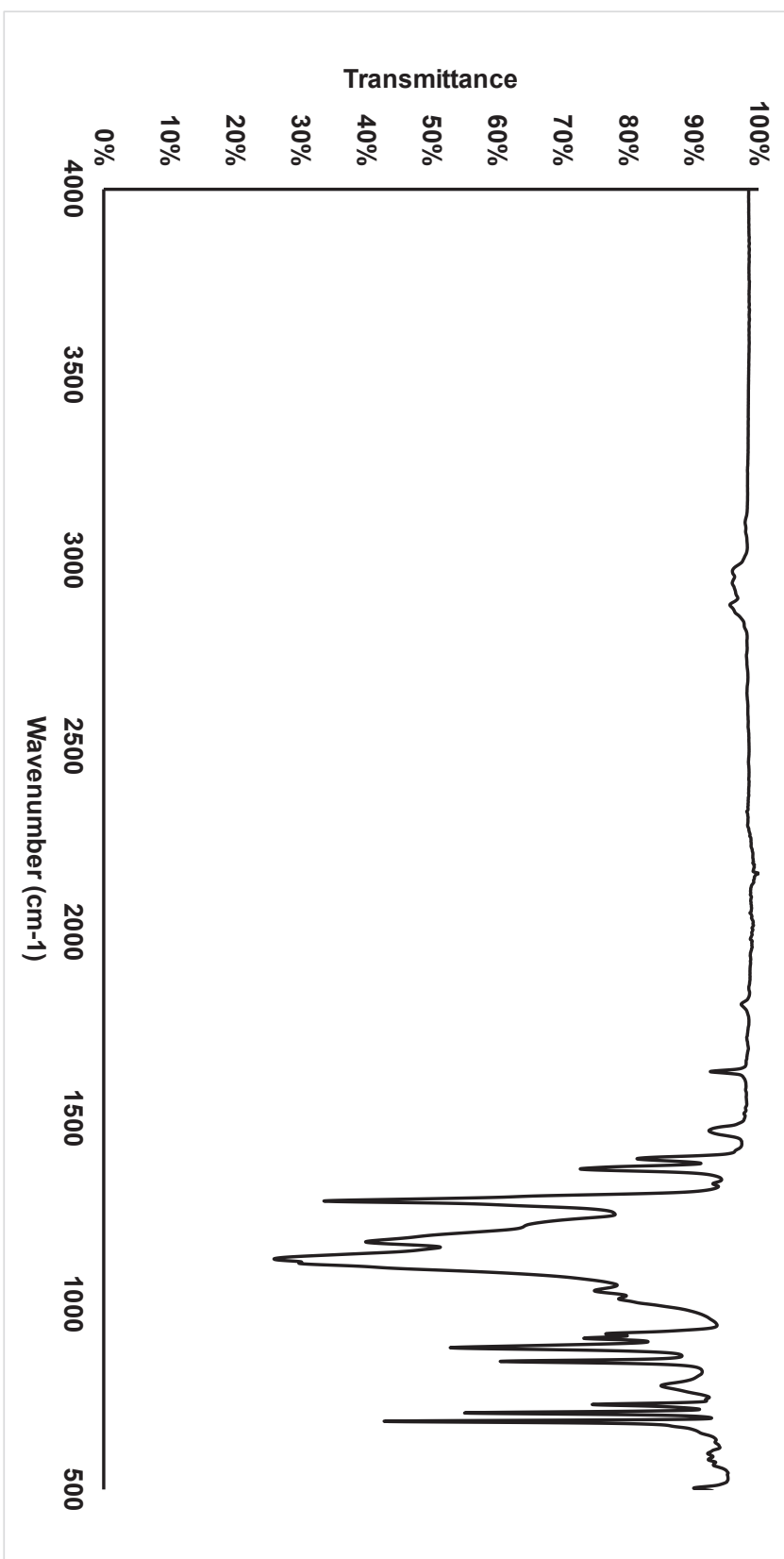
Q Exactive High-Res Mass Spec

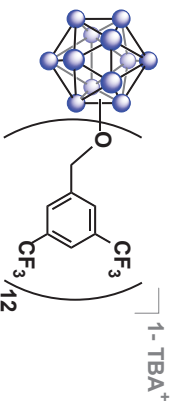
3-5-bis_2 #1 RT: 0.01 AV: 1 NL: 1.75E7
T: FTMS - p-ESI Fullms [500.00-4000.00]



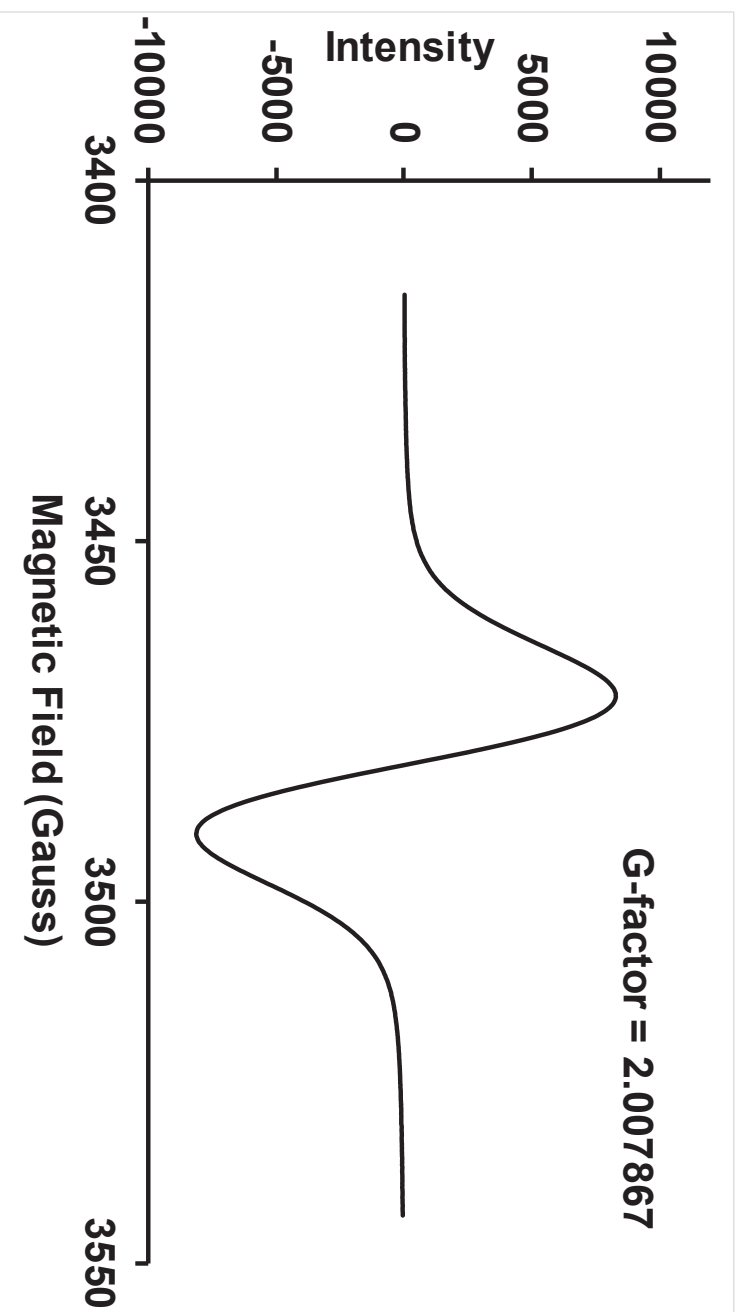


IR





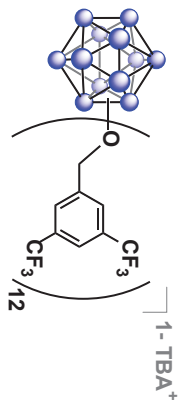
EPR



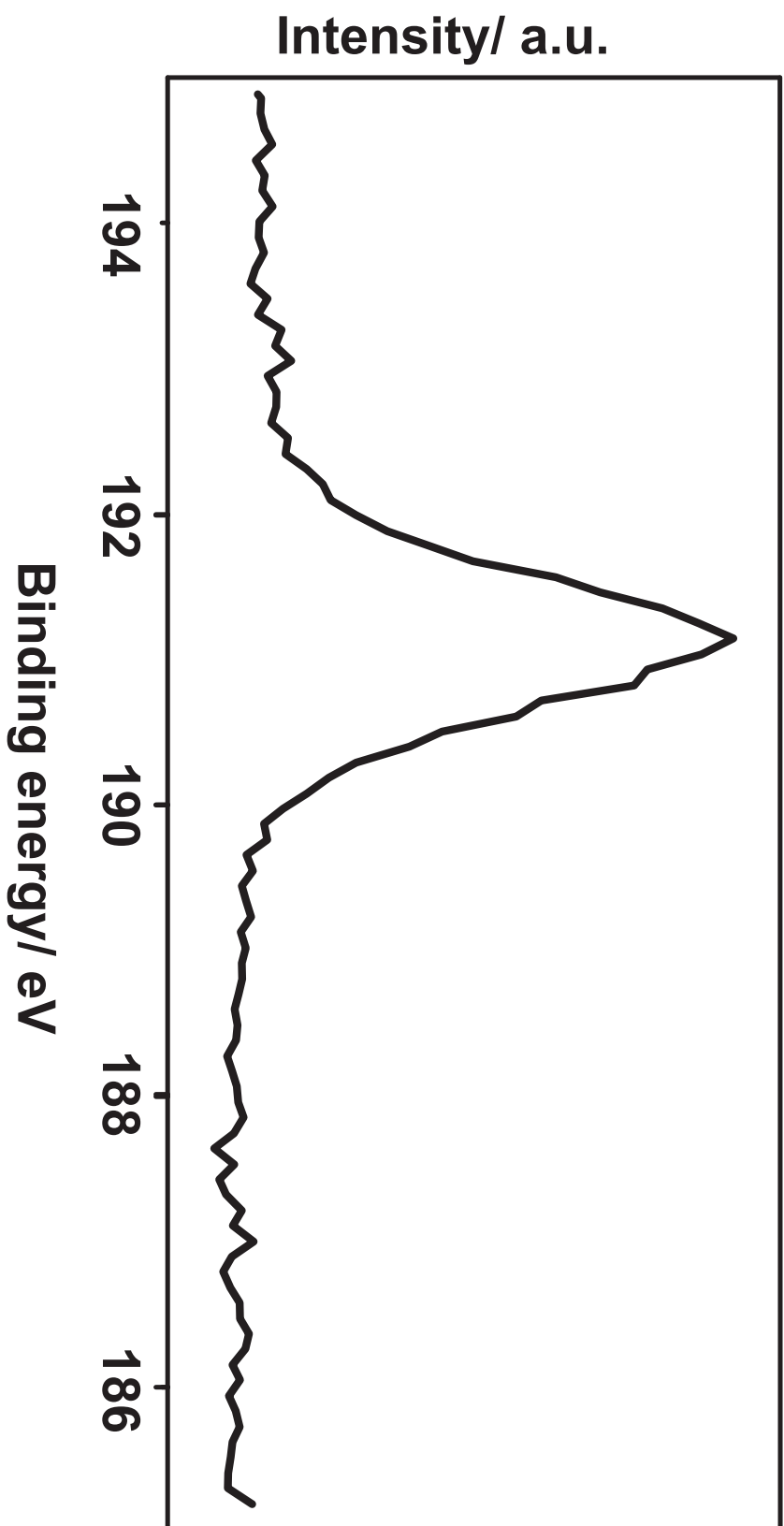
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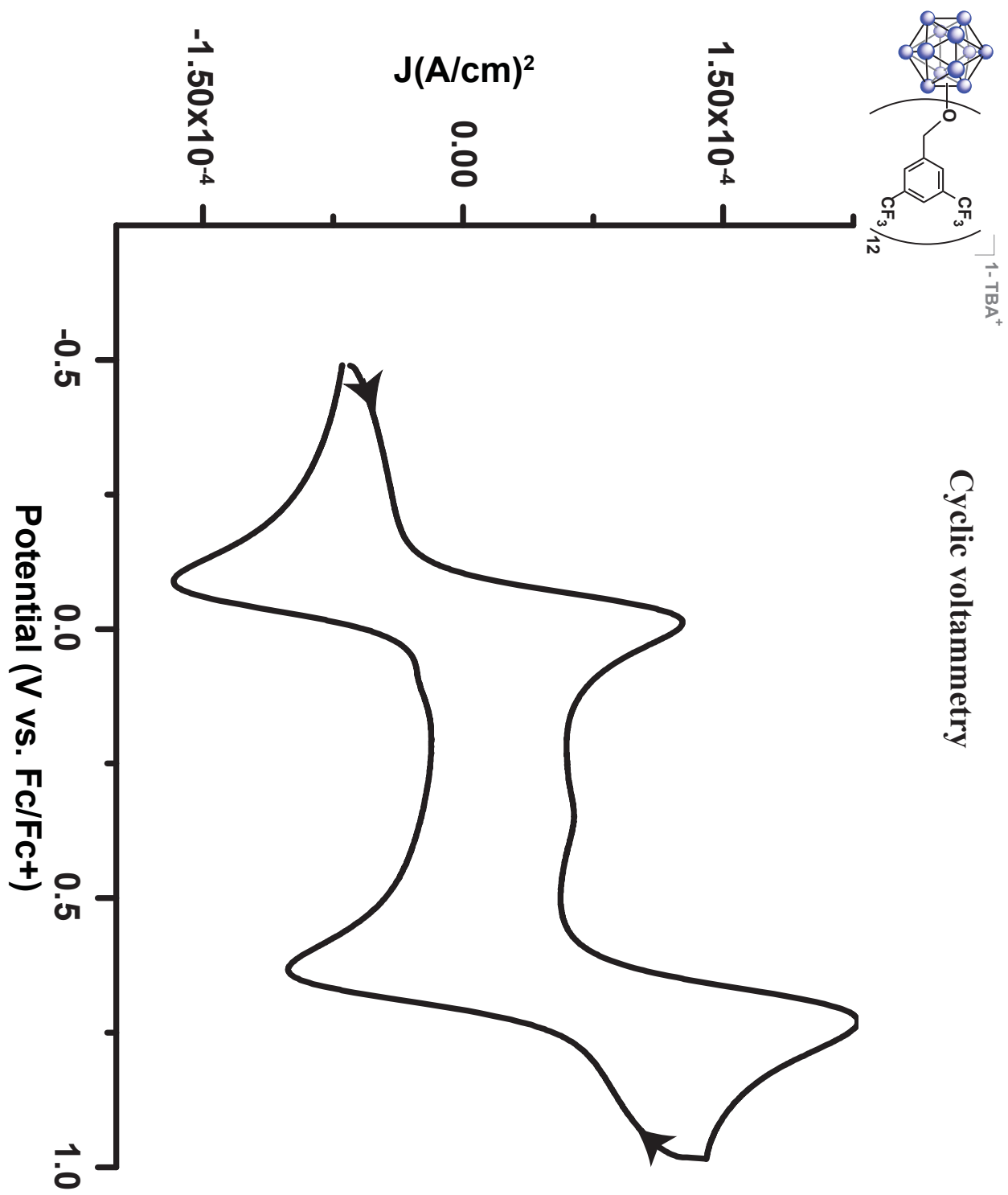
DOS Format
ANZ 1024
MIN -8134.161133
MAX 8283.838867
JSS 0
GST 3415.875
GSI 127.47
JUN G
JON Bruker Biospin GmbH
JDA 6/4/2015
JTM 16:02
JRE c:\programfiles\bruker-
emx\sycal\st0103.cal
JEX field-sweep
JSD 1
HCF 3479.61
HSW 127.47
EMF 0
RCT 20.48
RTC 20.48
RRG 2.83E+03
RMA 4
MF 9.778519
MP 6.38E-01

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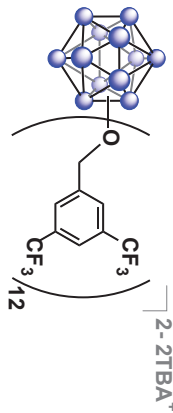
B 1s XPS



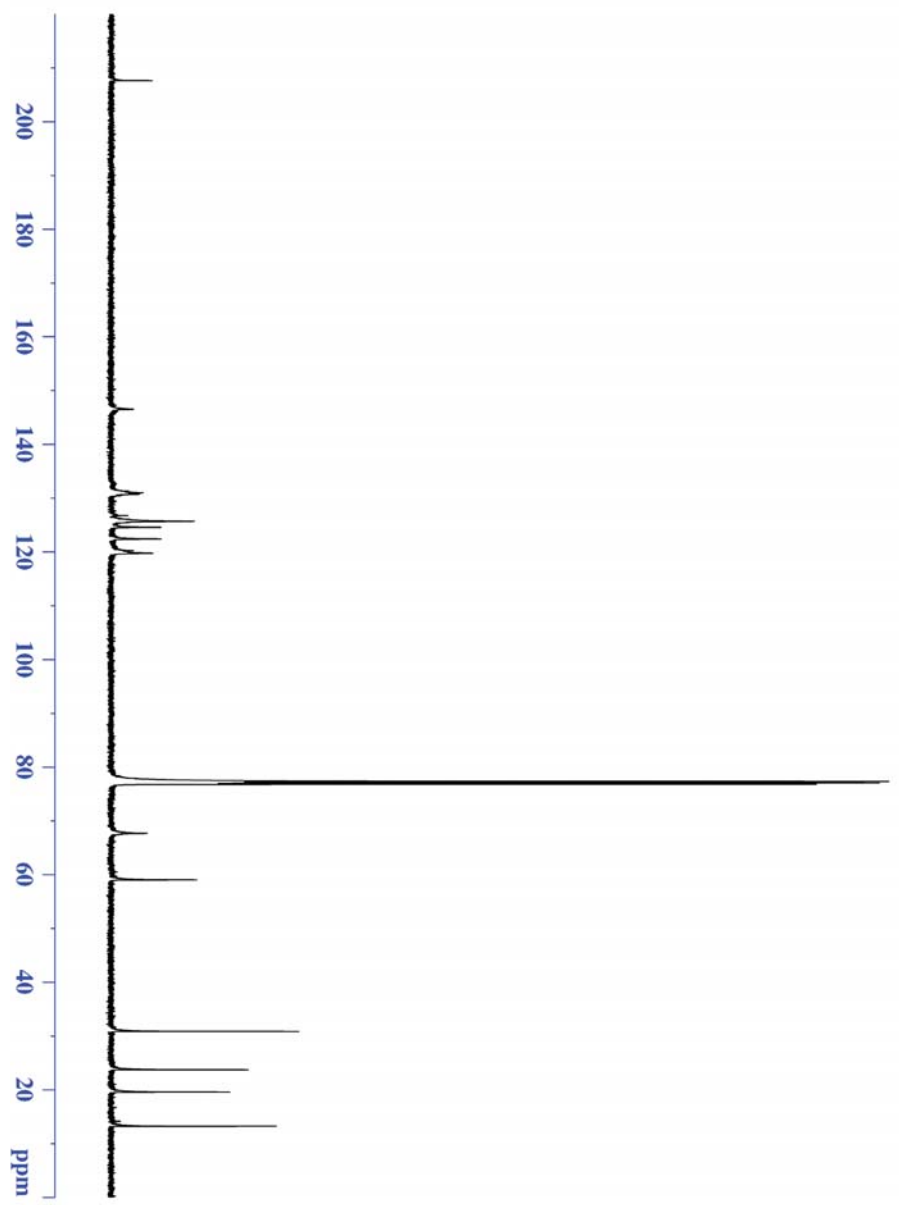




¹³C NMR



- 207.606
- 146.498
- 130.946
- 130.716
- 126.705
- 125.673
- 124.542
- 122.376
- 120.212
- 119.704
- 67.636
- 59.003
- 30.801
- 23.683
- 19.526
- 13.174



Current Data Parameters
 NAME B12(O-3,5-bis(TFMBn))12
 EXPNO 700
 PROCNO 1

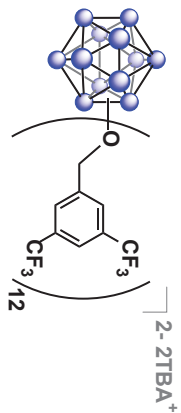
F2 - Acquisition Parameters
 Date_ 20150829
 Time 22.21
 INSTRUM av500
 PROBHID 5 mm DCH 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 256
 DS 2

SWH 31250.000 Hz
 FIDRES 0.476837 Hz
 AQ 1.0485760 sec
 RG 204.54
 DW 16.000 usec
 DE 18.00 usec
 TE 298.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TDO 1

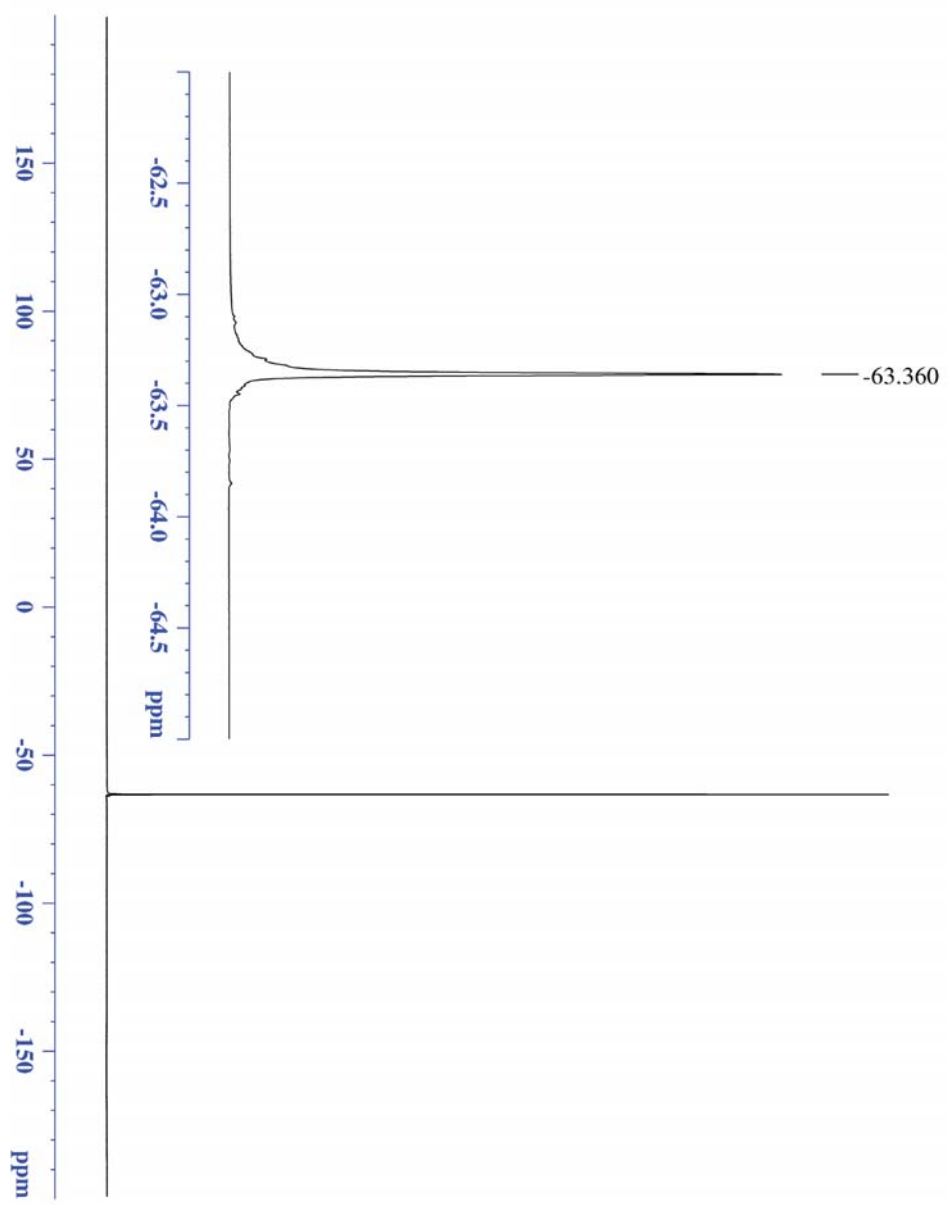
==== CHANNEL f1 =====
 SFO1 125.7722511 MHz
 NUC1 13C
 P1 9.63 usec
 PLW1 23.000000000 W

==== CHANNEL f2 =====
 SFO2 500.1330008 MHz
 NUC2 1H
 CPDPRGf2 waltz16
 PCPD2 80.00 usec
 PLW2 13.500000000 W
 PLW12 0.21094000 W
 PLW13 0.113500001 W

F2 - Processing parameters
 SF 131.072
 SE 125.777890 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



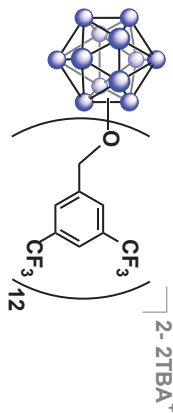
¹⁹F NMR



Current Data Parameters
 NAME B12(O-3,5-bisTFMBn)12
 EXPNO 121
 PROCNO 1

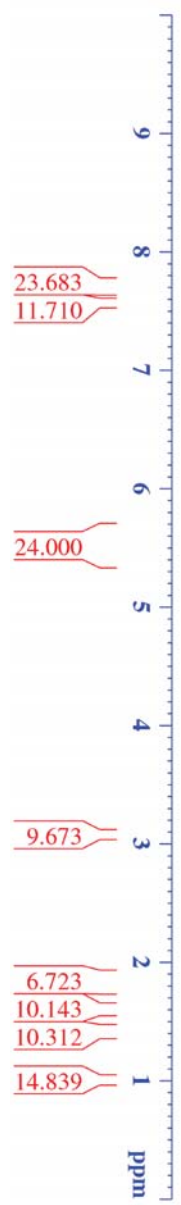
F2 - Acquisition Parameters
 Date_ 20150824
 Time 17:26
 INSTRUM av400
 PROBHID 5 mm PABBO BB/
 PULPROG zgpg30
 TD 262144
 SOLVENT CDCl3
 NS 32
 DS 0
 SWH 150000.000 Hz
 FIDRES 0.572205 Hz
 AQ 0.8738133 sec
 RG 189.85
 DW 3.333 usec
 DE 6.50 usec
 TE 299.0 K
 D1 2.000000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 376.4983660 MHz
 NUC1 19F
 P1 14.50 usec
 PLW1 17.000000000 W
 F2 - Processing parameters
 SI 262144
 SF 376.4983660 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.00



¹H NMR

- 7.702
- 7.561
- 5.528
- 3.098
- 3.077
- 3.056
- 1.622
- 1.604
- 1.586
- 1.441
- 1.423
- 1.405
- 1.387
- 1.017
- 0.999
- 0.981



Current Data Parameters
 NAME B12(O-3,5-bis(TFMBn))12
 EXPNO 122
 PROCNO 1

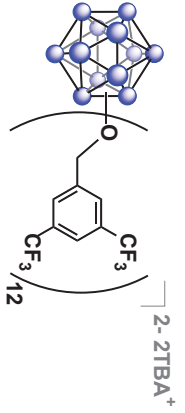
F2 - Acquisition Parameters
 Date_ 20150824
 Time 17.32

INSTRUM av400
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 52882
 SOLVENT CDCl3
 NS 64

DS 0
 SWH 8012.820 Hz
 FIDRES 0.151523 Hz
 AQ 3.2998369 sec
 RG 155.85
 DW 62.400 usec
 DE 6.50 usec
 TE 299.0 K
 D1 2.00000000 sec
 TD0 1

==== CHANNEL f1 =====
 SFO1 400.1324008 MHz
 NUC1 ¹H
 Pl 15.00 usec
 PLW1 13.000000000 W

F2 - Processing parameters
 SI 65536
 SF 400.1300184 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



¹¹B {¹H} NMR

-15.687



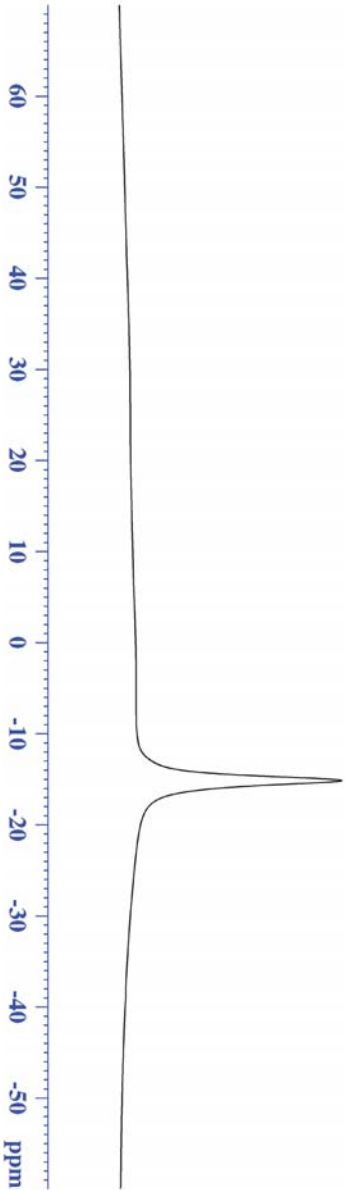
Current Data Parameters
 NAME B12(O-3,5-bis(TMbn))12
 EXPNO 120
 PROCNO 1

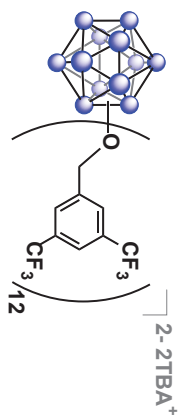
F2 - Acquisition Parameters
 Date_ 20150824
 Time 17.23
 INSTRUM av400
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 5096
 SOLVENT CDCl3
 NS 1024
 DS 0
 SWH 51020.406 Hz
 FIDRES 10.011854 Hz
 AQ 0.0449408 sec
 RG 189.85
 DW 9.800 usec
 DE 6.50 usec
 TE 299.2 K
 D1 0.00000400 sec
 D11 0.030000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 128.3776052 MHz
 NUC1 11B
 P1 10.00 usec
 PLW1 52.000000000 W

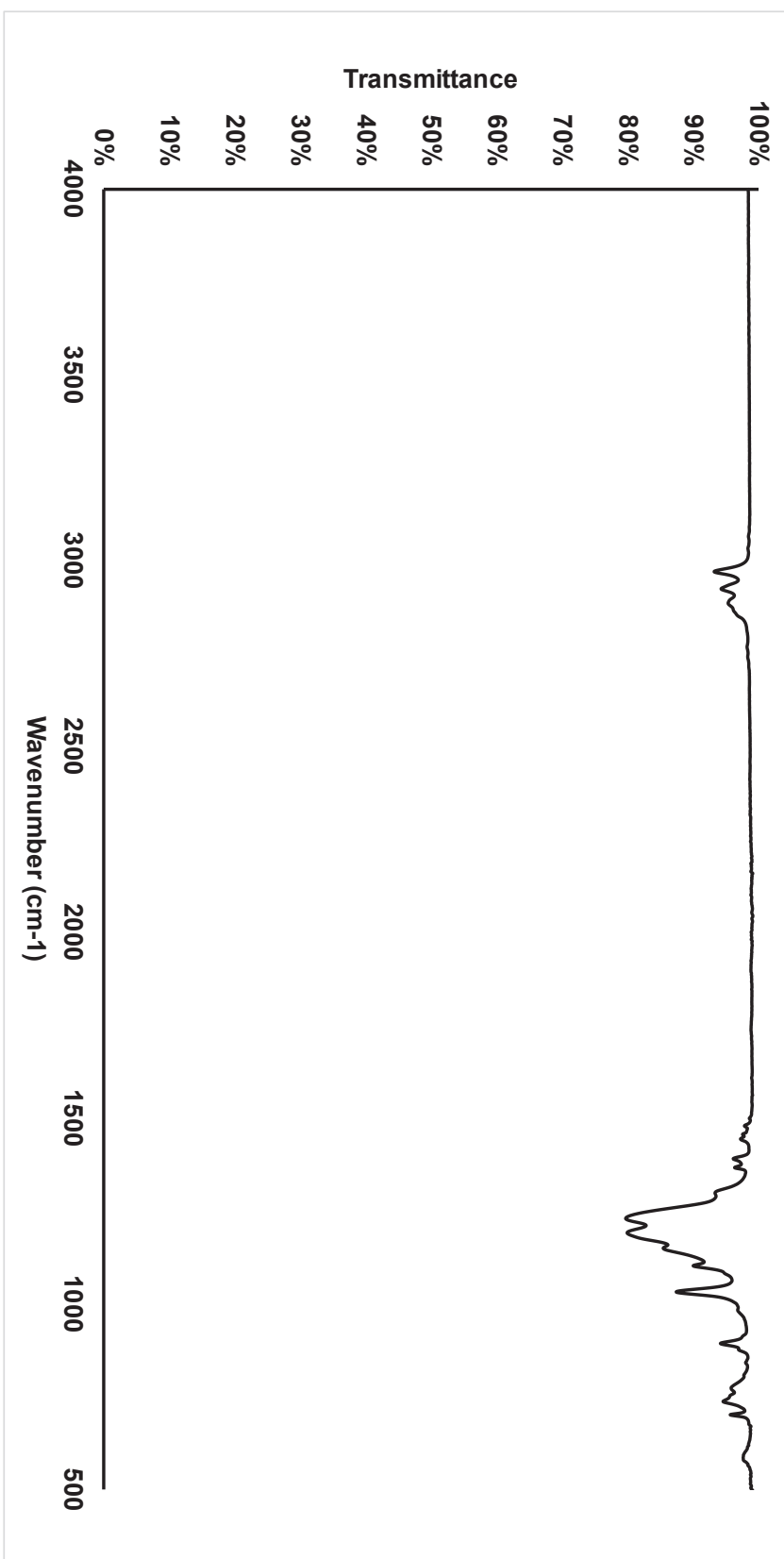
===== CHANNEL f2 =====
 SFO2 400.1324008 MHz
 NUC2 1H
 CPDPRG12 waltz16
 PCTPD2 90.00 usec
 PLW2 13.000000000 W
 PLW12 0.36111000 W

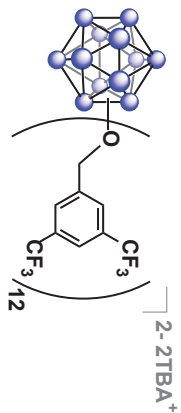
F2 - Processing parameters
 SI 32768
 SF 128.3776050 MHz
 WDW EM
 SSB 0
 LB 50.00 Hz
 GB 0
 PC 1.40



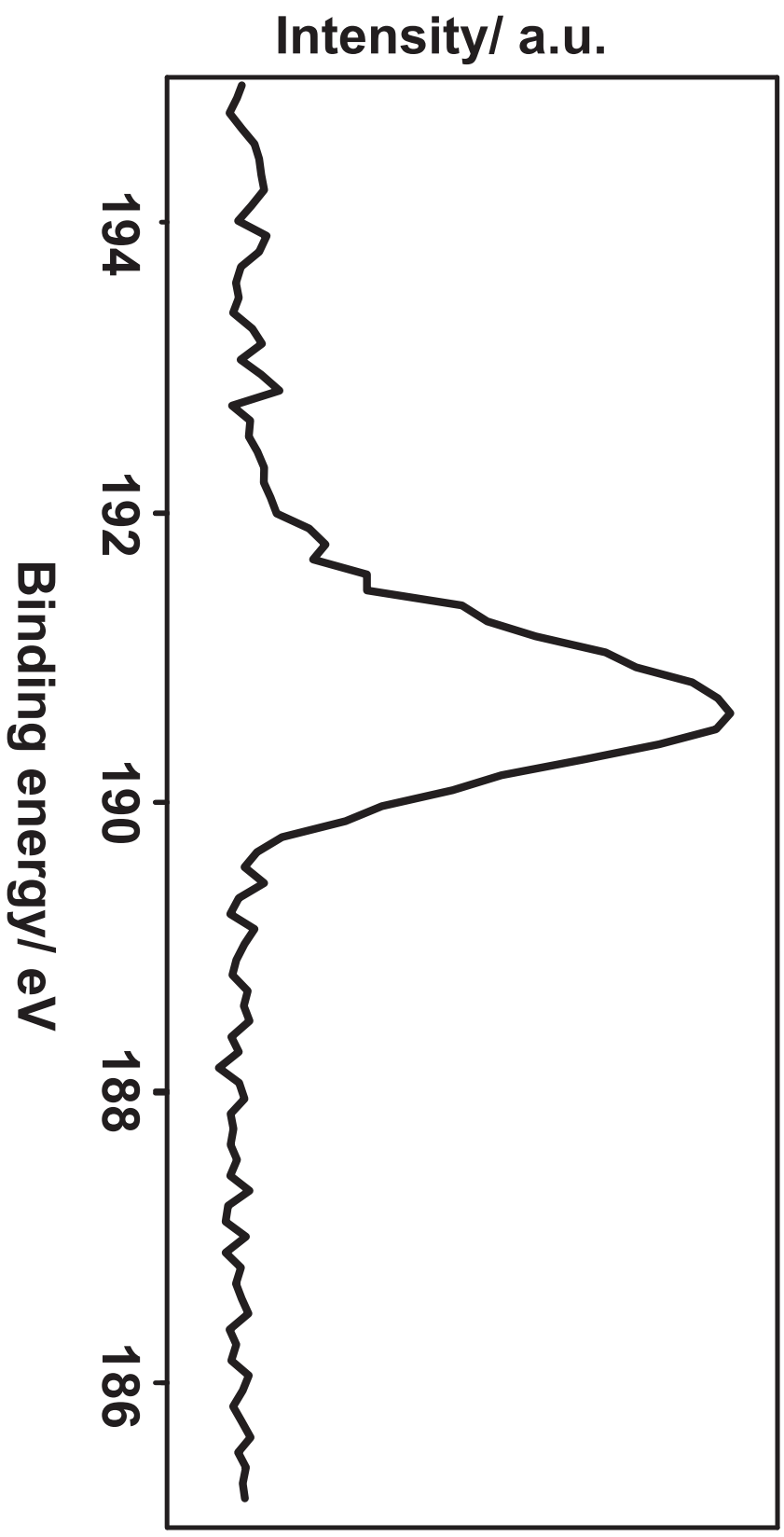


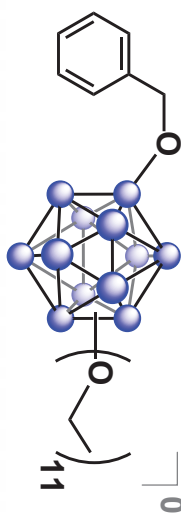
IR





B 1s XPS





— 39.064
— 35.228

¹¹B {¹H} NMR



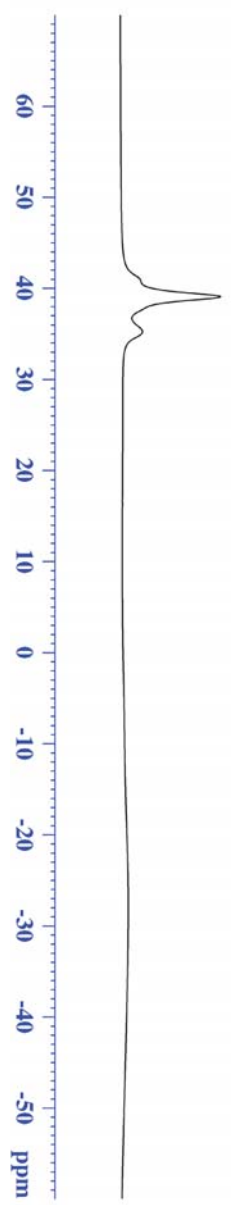
Current Data Parameters
 NAME B12(O-Bn)(O-Et)11
 EXPNO 30
 PROCNO 1

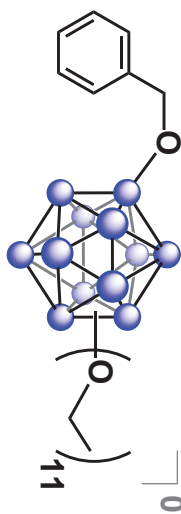
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 Time 15.01
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 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 5096
 SOLVENT CDCl3
 NS 1024
 DS 0
 SWH 51020.406 Hz
 FIDRES 10.011854 Hz
 AQ 0.0449408 sec
 RG 189.85
 DW 9.800 usec
 DE 6.50 usec
 TE 299.1 K
 D1 0.00000400 sec
 D11 0.030000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 128.3776052 MHz
 NUC1 ¹¹B
 P1 10.00 usec
 PLW1 52.000000000 W

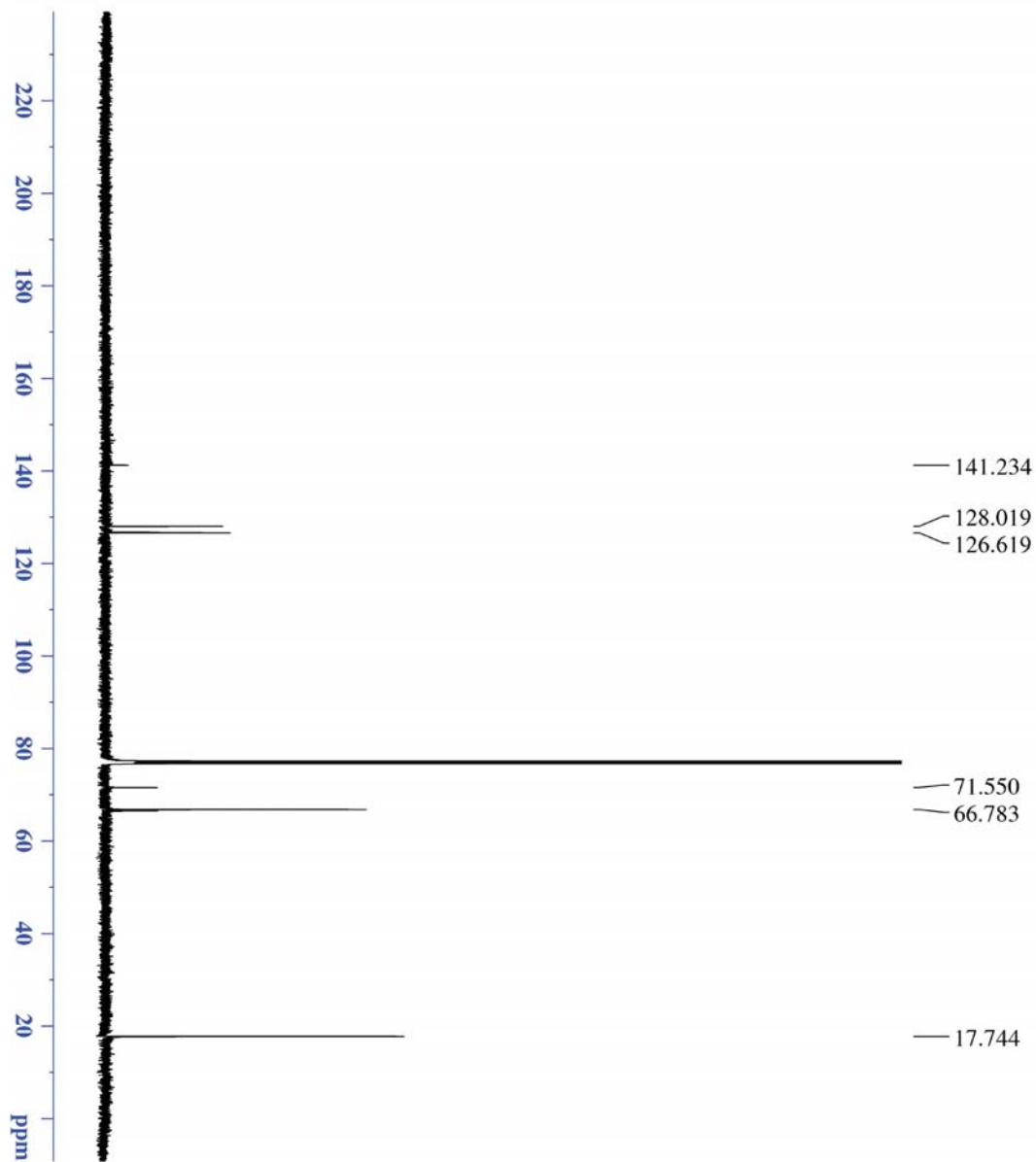
===== CHANNEL f2 =====
 SFO2 400.1324008 MHz
 NUC2 ¹H
 CPDPRG12 waltz16
 PCTPD2 90.00 usec
 PLW2 13.000000000 W
 PLW12 0.361110000 W

F2 - Processing parameters
 SI 32768
 SF 128.3776050 MHz
 WDW EM
 SSB 0
 LB 50.00 Hz
 GB 0
 PC 1.40





¹³C NMR



Current Data Parameters
 NAME B12(O-Bn)(O-Et)1
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150517
 Time 16:29

INSTRUM av500
 PROBHID 5 mm DCH 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3

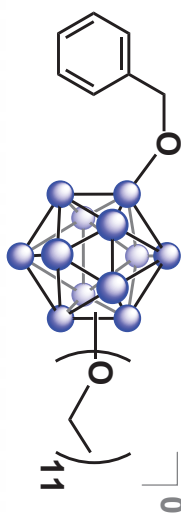
NS 2
 DS 4
 SWH 31250.000 Hz
 FIDRES 0.476837 Hz
 AQ 1.0485760 sec
 RG 13.13

DW 16.000 usec
 DE 18.00 usec
 TE 298.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

==== CHANNEL f1 =====
 SFO1 125.772511 MHz
 NUC1 13C
 P1 9.63 usec
 PLW1 23.00000000 W

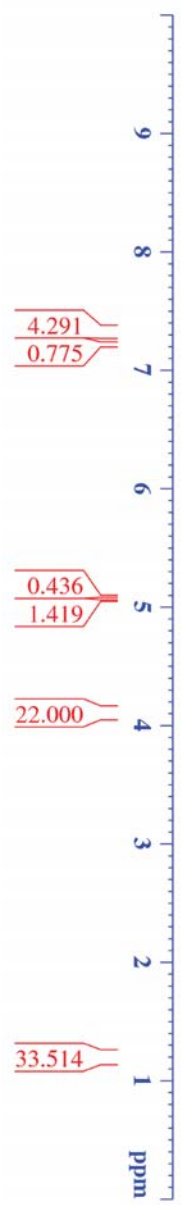
==== CHANNEL f2 =====
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 NUC2 1H
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 PCPD2 80.00 usec
 PLW2 13.50000000 W
 PLW12 0.21094000 W
 PLW13 0.113500001 W

F2 - Processing parameters
 SF 131.072
 SE 125.777892 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



¹H NMR

- 7.367
- 7.350
- 7.332
- 7.327
- 7.314
- 7.295
- 7.277
- 7.248
- 7.230
- 7.212
- 5.088
- 5.055
- 4.160
- 4.143
- 4.134
- 4.125
- 4.117
- 4.107
- 4.099
- 4.090
- 4.082
- 4.072
- 4.052
- 1.258
- 1.251
- 1.241
- 1.233
- 1.224
- 1.215
- 1.207
- 1.196
- 1.179
- 1.156



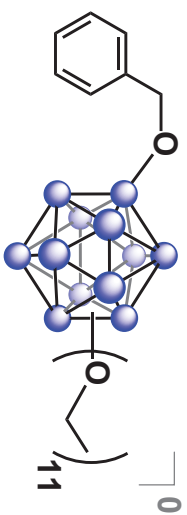
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 NAME B12(O-Bn)(O-E)11
 EXPNO 31
 PROCNO 1

F2 - Acquisition Parameters
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 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 52882
 SOLVENT CDCl3
 NS 32
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.151523 Hz
 AQC 3.2998369 sec
 RG 155.85
 DW 62.400 usec
 DE 6.50 usec
 TE 299.0 K
 D1 2.000000000 sec
 TD0 1

==== CHANNEL f1 ====
 SFO1 400.1324008 MHz
 NUC1 1H
 P1 15.00 usec
 PLW1 13.000000000 W

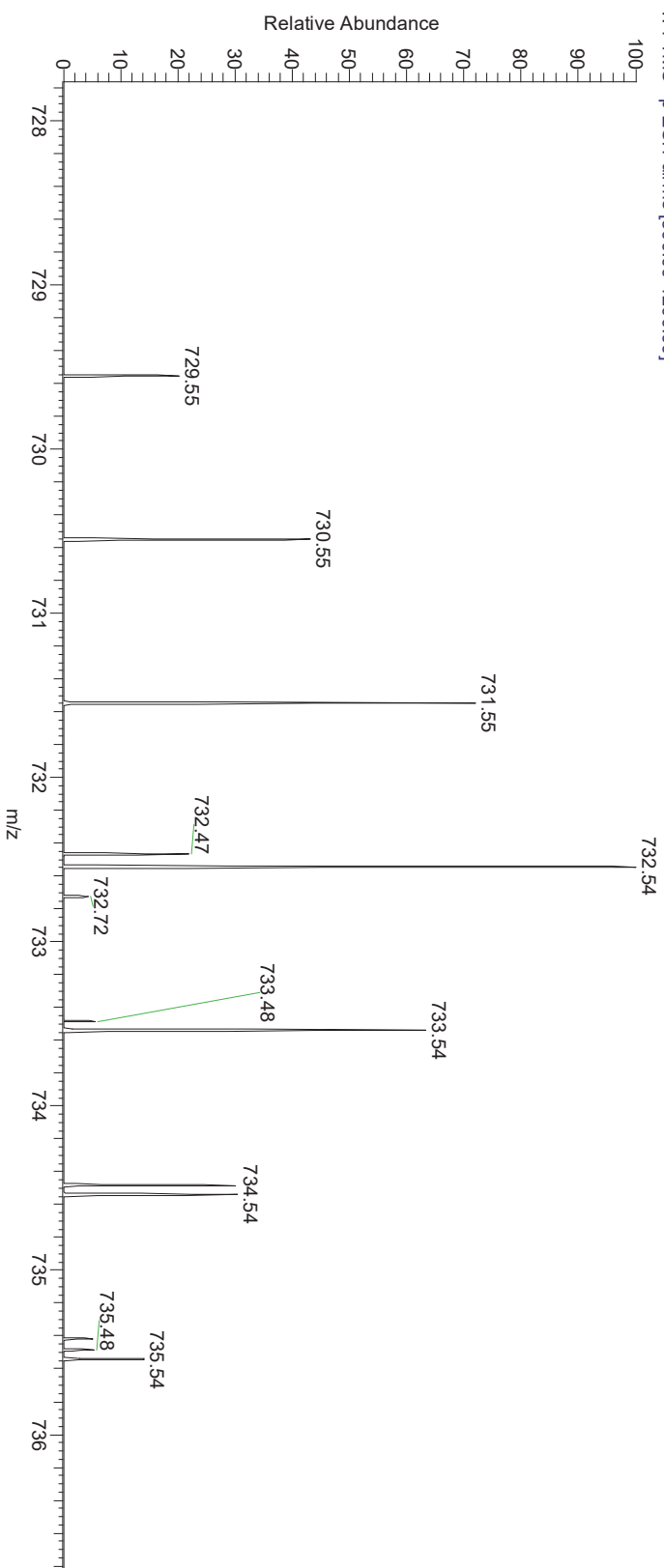
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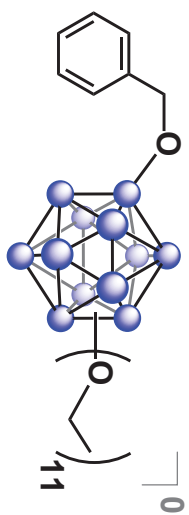




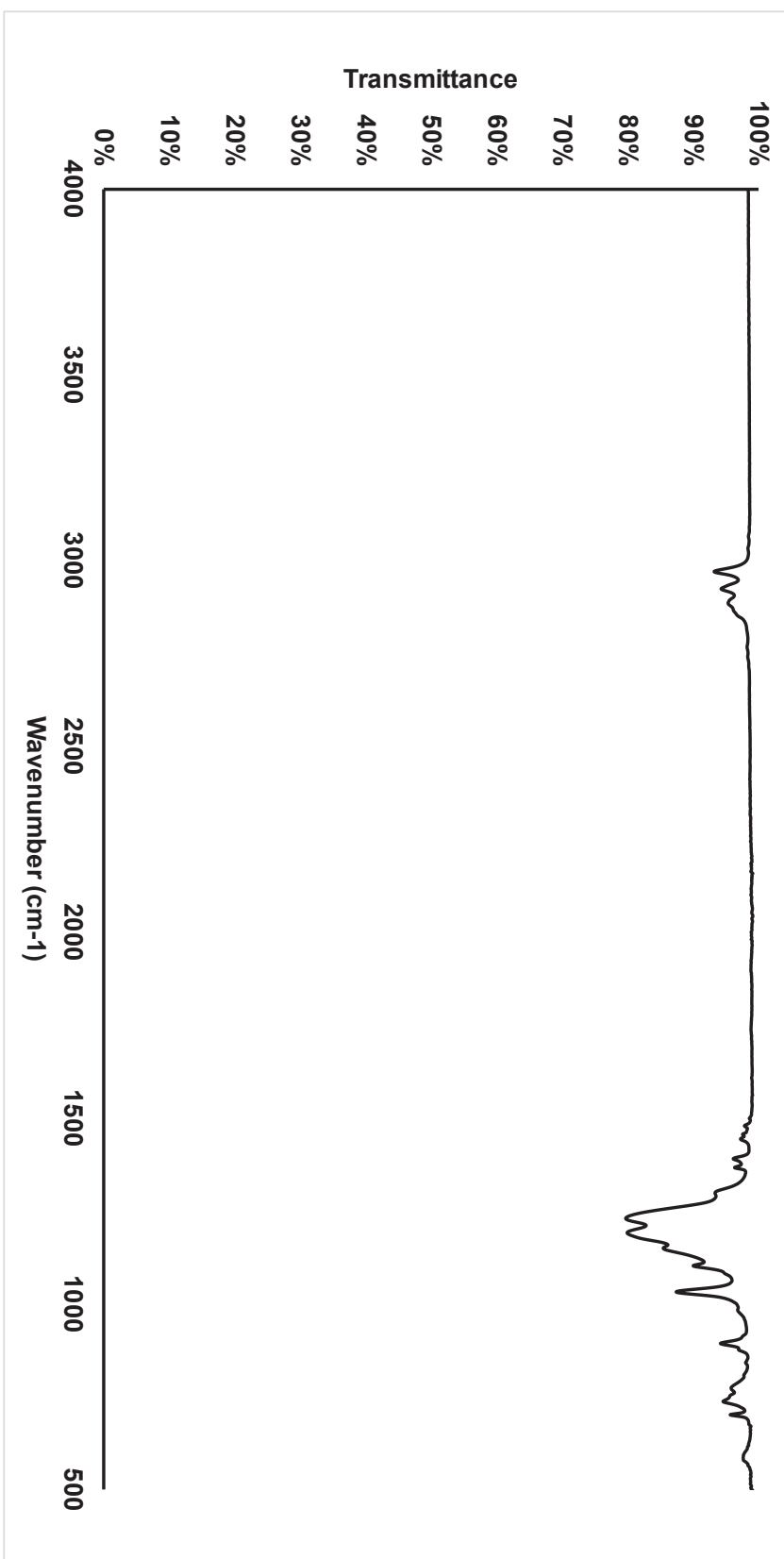
Q Exactive High-Res Mass Spec

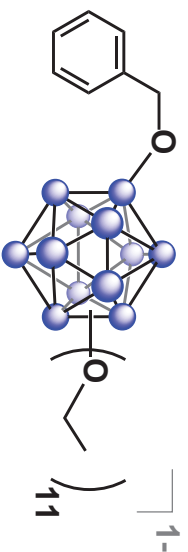
1Bn11Et#1 RT: 0.01 AV: 1 NL: 2.50E5
T: FTMS - p-ESI Fullms [500.00-1200.00]



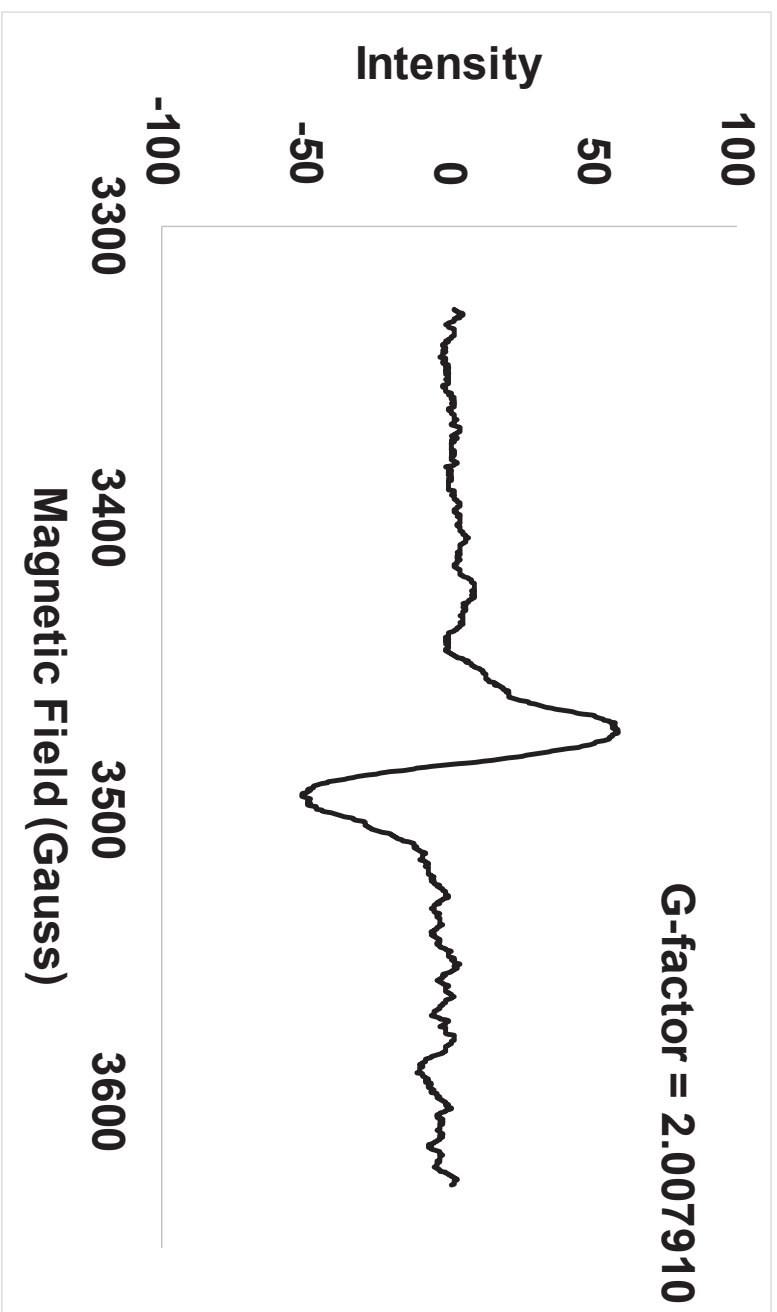


IR



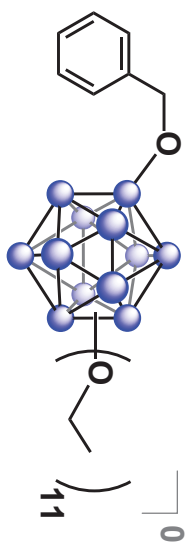


EPR

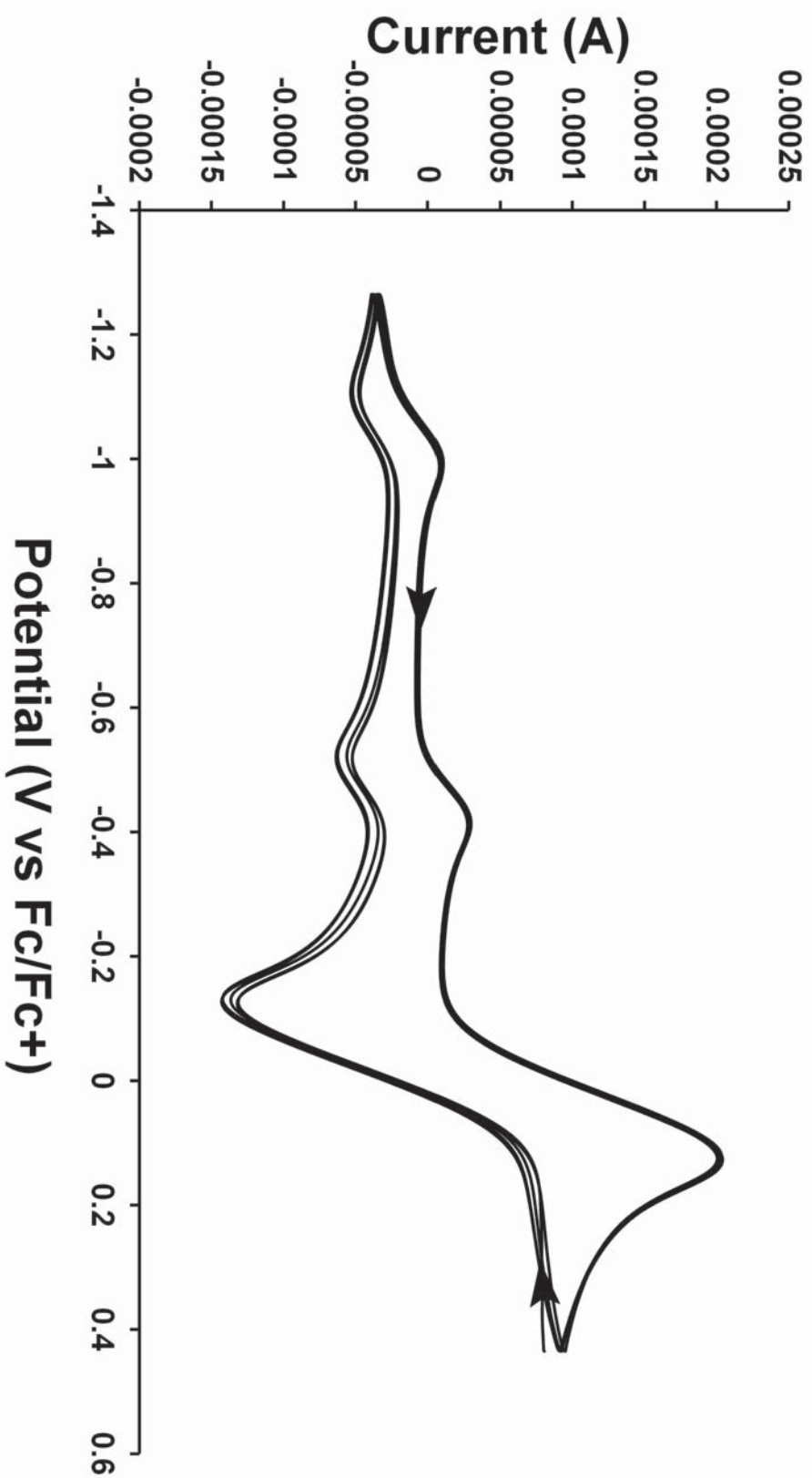


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DOS Format
ANZ 1024
MIN -51.404297
MAX 58.595703
JSS 0
GST 3328.55
GSI 300
JUN G
JON Bruker Biospin GmbH
JDA 11/2/2015
JTM 20:39
JRE c:\programfiles\bruker-
emx\sycal\st0103.cal
JEX field-sweep
JSD 1
HCF 3478.55
HSW 300
EMF 0
RCT 20.48
RTC 327.68
RRG 8.93E+03
RMA 4
MF 9.77575
MP 2.54E+00
MPD 19
  
```



Cyclic voltammetry



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