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A specific amino acid motif of *HLA-DRB1* mediates risk and interacts with smoking history in Parkinson's disease

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Parkinson's disease (PD) is a neurodegenerative disease in which genetic risk has been mapped to *HLA*, but precise allelic associations have been difficult to infer due to limitations in genotyping methodology. Mapping PD risk at highest possible resolution, we performed sequencing of 11 *HLA* genes in 1,597 PD cases and 1,606 controls. We found that susceptibility to PD can be explained by a specific combination of amino acids at positions 70–74 on the HLA-DRB1 molecule. Previously identified as the primary risk factor in rheumatoid arthritis and referred to as the "shared epitope" (SE), the residues Q/R-K/R-R-A-A at positions 70–74 in combination with valine at position 11 (11-V) is highly protective in PD, while risk is attributable to the identical epitope in the absence of 11-V. Notably, these effects are modified by history of cigarette smoking, with a strong protective effect mediated by a positive history of smoking in combination with the SE and 11-V ($P = 10^{-4}$; odds ratio, 0.51; 95% confidence interval, 0.36–0.72) and risk attributable to never smoking in combination with the SE without 11-V ($P = 0.01$; odds ratio, 1.51; 95% confidence interval, 1.08–2.12). The association of specific combinations of amino acids that participate in critical peptide-binding pockets of the HLA class II molecule implicates antigen presentation in PD pathogenesis and provides further support for genetic control of neuroinflammation in disease. The interaction of *HLA-DRB1* with smoking history in disease predisposition, along with predicted patterns of peptide binding to HLA, provide a molecular model that explains the unique epidemiology of smoking in PD.

Parkinson's disease | HLA | smoking | shared epitope

Parkinson's disease (PD; MIM: 168600) is a chronic, progressive neurodegenerative disease characterized by bradykinesia, tremor, rigidity, and postural instability plus a constellation of associated features including autonomic, sensory, cognitive, and psychological changes. With both familial and sporadic forms of the disease and a clear genetic component, numerous genome-wide association studies (GWAS), fine-mapping, and candidate gene studies over the past two decades have revealed dozens of risk-associated genetic variants. Consistent with underlying immunoregulatory dysfunction and inflammatory processes operating in PD, immune system genetic factors are consistently represented among replicated PD loci. Most notably, a clear association peak has been shown to map to the major histocompatibility complex (*MHC*) on chromosome 6 (6p21.3), which houses the *HLA* genes.

HLA class II gene products are expressed on the surface of professional antigen-presenting cells, which generally present peptide fragments of extracellular origin to helper (CD4^{+}) T cells (1). HLA class I molecules are expressed on all nucleated cells and present antigens originated from intracellular sources to cytotoxic (CD8^{+}) T cells for killing tumorous or infected cells (2). While an association of HLA class I with PD was first observed more than four decades ago (3), more contemporaneous single nucleotide polymorphism (SNP) associations point to the class II region of the locus (4, 5). In an early GWAS, the association peak in the MHC was found with rs3129882 within *HLA-DRA* (MIM: 142860) (6), although attempts to replicate this observation have generated varied and conflicting results (7–13).

Recent large studies have used SNP-based GWAS to fine-map the *MHC* association signal and to impute functional alleles of the *HLA* system (4, 5, 12, 14–17), but the complexity of the region presents major challenges for studies aimed at defining the causative variants. Extremely high levels of polymorphism,

Significance

Parkinson's disease (PD) is a chronic, progressive neurodegenerative disease with both familial and sporadic forms and a clear genetic component. In addition, underlying immunoregulatory dysfunction and inflammatory processes have been implicated in PD pathogenesis. In this study, deep sequencing of *HLA* genes, which encode highly variable cell surface immune receptors, reveals specific variants conferring either risk or protection in PD. Because a history of cigarette smoking is known to be protective in PD, we analyzed the interaction of these genetic variants with smoking history in PD patients and healthy controls and found that the genetic effects are modified by history of cigarette smoking. These results provide a molecular model that explains the unique epidemiology of smoking in PD.

GENETICS

structural variation, an extensive history of gene duplication, conversion and recombination events, and unusually high levels of linkage disequilibrium (LD) confound attempts to infer precise allelic associations from SNP data (18). The few studies that have directly queried *HLA* variation through traditional genotyping methods have produced mixed results and have been hampered by a relatively small sample size, low-resolution *HLA* genotyping methodology, or both (19–22).

To unravel the immunogenetic underpinnings of the disease, we undertook a large-scale deep-sequencing study of the *HLA* genes in PD. We analyzed *HLA* variation at high resolution using a well-validated next-generation sequencing (NGS) method in 1,941 PD cases and 1,612 controls from the United States with European ancestry. By sequencing comprehensively across 11 full-length *HLA* genes— *HLA-DRB1* (MIM: 142857), *HLA-DRB3* (MIM: 612735), *HLA-DRB4*, *HLA-DRB5* (MIM: 604776), *HLA-DQA1* (MIM: 146880), *HLA-DQB1* (MIM: 604305), *HLA-DPA1* (MIM: 142880), *HLA-DPB1* (MIM: 142858), *HLA-A* (MIM:142800), *HLA-B* (MIM: 142830), and *HLA-C* (MIM: 142840)—we were able to examine associations at the highest possible resolution, yielding the most complete representation to date of the complex immunogenetics underlying PD. We found that protection in PD can be explained by a specific epitope at positions 70–74 on the HLA-DRB1 molecule in combination with valine at position 11 (11-V), while risk is attributable to the identical epitope in the absence of 11-V, and that these effects are modified by a history of cigarette smoking. In identifying the source of association signals in the disease and interaction with a key environmental factor, we provide a framework for understanding the nature of the immune component in PD pathogenesis.

Results

High-Resolution Genotyping Demonstrates That Risk and Protection in PD Are Mediated by *HLA-DRB1*.

All 11 of the highly-polymorphic *HLA* class I and II genes were sequenced from 1,597 PD cases and 1,606 controls (*SI Appendix*, Table S1). We first analyzed data for the entire molecule at polypeptide sequence resolution, described in the HLA nomenclature by the first two fields of the allele name (23), to examine the entire functional unit of interest. In this analysis, we considered results sufficient to warrant follow-up when significant at a false discovery rate (FDR) (24) of 10%, corresponding to a cutoff of $P = 0.02$. We found strong protective effects for *HLA-DRB1*04:01* [$P = 0.007$; odds ratio (OR), 0.78; 95% confidence interval (CI), 0.65–0.93] and *HLA-DQB1*03:02* ($P = 0.006$; OR, 0.78; 95% CI, 0.66–0.93) (Fig. 1). These alleles are in strong LD, forming part of the *HLA-DQA1*03:01~HLA-DQB1*03:02~HLA-DRB1*04:01* class II haplotype, which is also significantly associated with disease ($P = 0.008$; OR, 0.66; 95% CI, 0.54–0.84). Confirming *HLA-DRB1* as the source of this signal, another *HLA-DRB1*04:01*–containing haplotype, *HLA-DQA1*03:03~HLA-DQB1*03:01~HLA-DRB1*04:01*, was observed at nearly equal frequency (Table 1) and is also significantly protective against developing PD ($P = 0.005$; OR, 0.66; 95% CI, 0.54–0.85). While both haplotypes are in LD with *HLA-DRB4*01:03*, this allele may be excluded as causative because it is also observed in some *HLA-DRB1*07:01* haplotypes not associated with disease. Furthermore, no association remained for *HLA-DQB1*03:02* on conditioning on *HLA-DRB1*04:01*.

Complementing this finding, we observed borderline significance for PD risk mediated by *HLA-DRB1*01:01* ($P = 0.012$; OR, 1.26; 95% CI, 1.05–1.50). This result remained significant after conditioning for *HLA-DRB1*04:01* ($P = 0.02$; OR, 1.23;
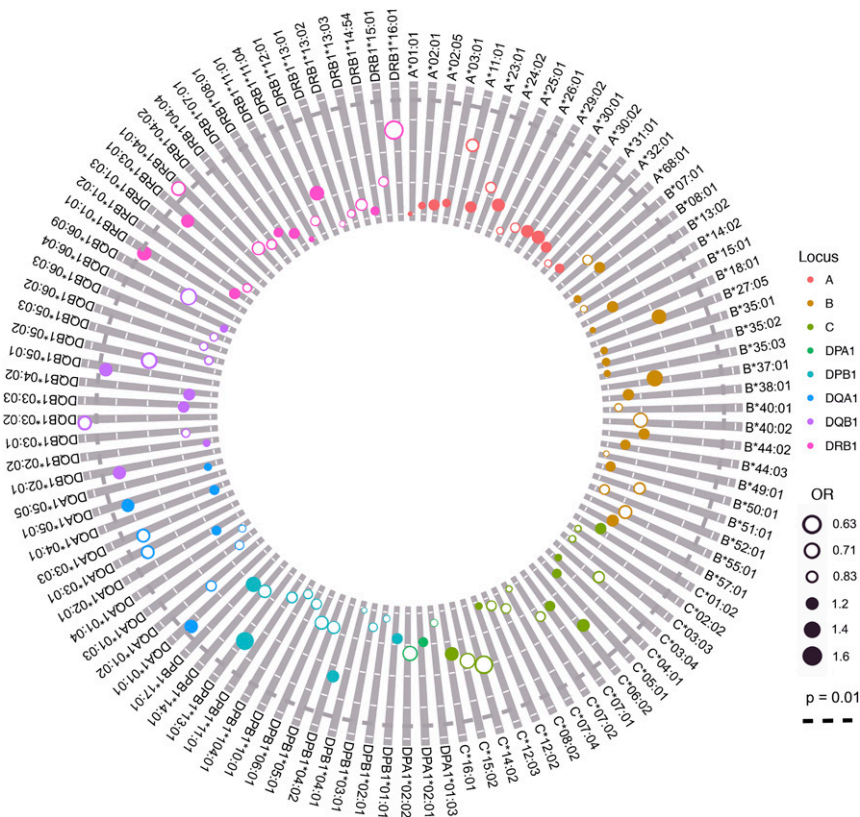
Fig. 1. Association analysis results for all *HLA* class I and class II alleles. *HLA* alleles that were detected in the study population are shown around the perimeter. Decreasing *P* values are indicated by the distance of points from the center, with *P* = 0.01 indicated by the black dotted concentric line. The magnitude of ORs is indicated by the diameter of points, with filled points indicating OR >1 and open points indicating OR <1. The magnitude of global linkage disequilibrium between loci is indicated by the width of interior lines.

Table 1. HLA-DQA1~DQB1~DRB1 haplotype frequencies in PD cases and controls

DQA1~DQB1~DRB1	Control frequency	Case frequency
01:02~06:02~15:01	0.124	0.122
05:01~02:01~03:01	0.114	0.137
02:01~02:02~07:01	0.091	0.092
01:01~05:01~01:01	0.079	0.099
01:03~06:03~13:01	0.055	0.053
05:05~03:01~11:01	0.051	0.051
03:03~03:01~04:01	0.047	0.039
03:01~03:02~04:01	0.037	0.031
05:05~03:01~11:04	0.037	0.031
01:02~06:04~13:02	0.034	0.034
02:01~03:03~07:01	0.033	0.038
03:01~03:02~04:04	0.032	0.027
01:04~05:03~14:54	0.02	0.02
01:02~05:02~16:01	0.019	0.011
04:01~04:02~08:01	0.018	0.02
05:05~03:01~12:01	0.017	0.02
03:01~03:02~04:02	0.015	0.012
01:01~05:01~01:02	0.014	0.016
01:02~06:09~13:02	0.013	0.01
01:03~06:01~15:02	0.011	0.008
01:01~05:01~01:03	0.011	0.011
05:05~03:01~13:03	0.01	0.01
03:03~03:01~04:07	0.009	0.01

95% CI, 1.03–1.49), confirming it as an independent association. *HLA-DRB1*01:01* is observed in near-complete LD with *HLA-DQA1*01:01* and *HLA-DQB1*05:01* as a haplotype in populations with European ancestry (25, 26). Although we also found similar associations and significance levels for the *HLA-DQA1* and *HLA-DQB1* alleles of this haplotype (Fig. 1), their effect sizes were smaller, and on conditioning for *HLA-DRB1*01:01*, no effect remained for these alleles. This finding suggests that any observed *HLA-DQA1/-DQB1* association is attributable to LD with the primary predisposing allele, *HLA-DRB1*01:01*. Taken together, our results strongly implicate *HLA-DRB1* as the operative locus in disease.

Our genotyping methodology provides information for virtually all coding and noncoding variations at the *HLA* loci; however, we did not observe any appreciable differences between cases and controls with respect to noncoding variation at any locus. Likewise, there were no significant differences between cases and controls for alleles of any class I locus (*HLA-A, -B,* and *-C*) or class II loci *HLA-DPA1, HLA-DPB1, HLA-DRB3, HLA-DRB4,* and *HLA-DRB5* after conditioning on the *HLA-DRB1* associations. Similarly, tests for interaction between *HLA* alleles and between *HLA* and known PD risk SNPs, when available in our dataset, did not yield any significant results.

Specific Amino Acid Residues of HLA-DRB1 Mediate Risk and Protection in PD. To decipher the precise structural elements mediating the suggested *HLA-DRB1* associations with PD, we examined the contribution of individual amino acids of the *HLA-DRB1* molecule to disease risk. Omnibus testing revealed four positions to be significantly associated with PD: 11 ($P = 0.002$; $p_{corr} = 0.05$), 13 ($P = 0.001$; $p_{corr} = 0.04$), 26 ($P = 0.001$; $p_{corr} = 0.03$), and 33 ($P = 0.001$; $p_{corr} = 0.04$). Of these, positions 11, 13, and 26 are highly polymorphic and occupy key positions in the peptide-binding groove of the class II molecule. Results of analysis of amino acid variation for each position are provided in *SI Appendix*, Table S2. The effect at position 26 is driven by phenylalanine, mediating protection ($P = 3 \times 10^{-5}$; OR, 0.79; 95% CI, 0.71–0.89). Position 11 is extremely polymorphic, with

six different residues observed across this dataset; of these, significant associations are observed for valine, which mediates protection ($P = 0.006$; OR, 0.83; 95% CI, 0.73–0.95), and lysine, which mediates risk ($P = 0.006$; OR, 1.24; 95% CI, 1.06–1.44). Position 13 is also extremely polymorphic and is in very strong yet incomplete LD with position 11; however, the association at this position appears to be driven mainly by a protective effect of histidine ($P = 0.005$; OR, 0.82; 95% CI, 0.72–0.95). Examination of these amino acid associations in a replication cohort of an additional 462 controls and 536 PD patients confirms the findings for 11-V ($P = 0.02$; OR, 0.7; 95% CI, 0.51–0.97) and 13-H ($P = 0.03$; OR, 0.7; 95% CI, 0.5–0.97).

The protective effects observed for 11-V, 13-H, and 26-F can be attributed to association of these variants with alleles of the *HLA-DRB1*04* lineage, aligning with our initial observation of protection mediated by *HLA-DRB1*04:01*. Likewise, 11-L is specific to alleles of the *HLA-DRB1*01* lineage, in line with our observation of risk mediated by *HLA-DRB1*01:01*. Thus, while the extreme levels of polymorphism at *HLA-DRB1* hampers our power to confirm high-resolution allelic associations beyond 10% FDR or to fully account for the burden of multiple comparisons, the totality and context of variation at individual amino acids support our initial observations in high-resolution alleles. Indeed, combining alleles of the *HLA-DRB1*04* lineage, we find significant protection at this resolution ($P = 0.001$, $p_{corr} = 0.01$; OR, 0.79; 95% CI, 0.69–0.91). Confirming this observation, *HLA-DRB1*04* is also significantly protective in our replication cohort, with a nearly identical effect size ($P = 0.04$; OR, 0.77; 95% CI, 0.60–0.99).

The "Shared Epitope" and Position 11 Explain *HLA* Associations in PD. Notably, along with *HLA-DRB1*01:01*, which we find to be predisposing in PD, *HLA-DRB1*04:01* and *HLA-DRB1*10:01* share a common set of amino acids at positions 70–74 (70-Q/R, 71-R/K, 72-R, 73-A, and 74-A), referred to as the "shared epitope" (SE) and originally described with respect to predisposition to rheumatoid arthritis (RA; MIM: 180300) (27). In our dataset, seven *HLA-DRB1* alleles have the SE, including *HLA-DRB1*10:01* and additional alleles of the *HLA-DRB1*04* and *HLA-DRB1*01* lineages (Table 2). Looking more closely at the lower-frequency *HLA-DRB1*04* alleles in our dataset, we observed that only *HLA-DRB1*04* alleles with the SE are increased in controls compared with PD cases (Table 2), while those without the SE are not.

Given these findings, we examined the association of *HLA-DRB1* with PD with respect to the SE in combination with position 11-V. We found that the risk for PD is mediated by the SE in the absence of 11-V ($P = 0.01$; OR, 1.25; 95% CI, 1.05–1.50). While not statistically significant, a trend toward a risk for the SE without 11-V was seen in our replication cohort. Most striking, we found that protection in PD can be explained by the SE in combination with 11-V ($P = 0.001$; OR, 0.76; 95% CI, 0.64–0.89), and that this result is clearly reproducible in our replication cohort ($P = 0.01$; OR, 0.61; 95% CI, 0.42–0.90), confirming a role for this specific set of amino acids in disease.

Smoking History and the *HLA* SE Interact in Risk and Protection in PD. A protective effect of cigarette smoking in PD is well established (28). Moreover, recent work suggests an interaction between smoking history and variation in *HLA-DRB1* in disease (17), but that analysis examined only a single SNP in the locus. Having identified the SE as the structural variant of *HLA-DRB1* associated with disease, we sought to examine in greater detail the role of smoking history in combination with HLA variation. We analyzed the interaction of smoking history with the *HLA-DRB1* SE in 837 PD cases and 918 controls for whom data on cigarette smoking history were available. These individuals had self-reported as either a former or current smoker or having never smoked; for

Table 2. Amino acid residues for HLA-DRB1 allotype SE and position 11 and frequencies in PD cases and controls

HLA-DRB1	11	70	71	72	73	74	Controls	Cases
01:01	L	Q	R	R	A	A	0.083	0.099
01:02	L	Q	R	R	A	A	0.014	0.016
04:01	V	Q	K	R	A	A	0.085	0.076
04:04	V	Q	R	R	A	A	0.034	0.027
04:05	V	Q	R	R	A	A	0.006	0.004
04:08	V	Q	R	R	A	A	0.006	0.004
10:01	V	R	R	R	A	A	0.009	0.008

The HLA-DRB1 SE is defined as residues Q/R-K/R-R-A-A at position 70–74.

our analysis, we dichotomized these groups into those with any history of smoking and those with no history of smoking. *HLA-DRB1* alleles were coded according to the presence of the SE with and without 11-V (SE$^+$V11$^+$ and SE$^+$V11$^-$), and the impact on disease occurrence was determined by multiple logistic regression with gender, principal component (PC) 1, PC2, PC3, and PC4 as covariates. No significant differences with respect to SE frequencies were observed between individuals for whom smoking data were available and those for whom it was not available (*SI Appendix*, Table S3).

To study whether the effect of the SE genotype on disease was modified by smoking history, an interaction term (SE × smoking history) was included in the model. The importance of each possible risk factor under consideration was assessed by the Wald test and confirmed by the likelihood ratio test on the deviance of the model. We found that the interaction between positive smoking history and homozygosity for SE+V11$^+$ alleles was significant ($P = 0.02$), suggesting that protection mediated by SE$^+$V11$^+$ *HLA-DRB1* alleles was further enhanced by a history of smoking. The data show that the combined effect of a history of smoking with the presence of one or two SE$^+$V11$^+$ alleles was highly protective in PD ($P = 10^{-4}$; OR, 0.51; 95% CI, 0.36–0.72) (*SI Appendix*, Table S4), with an effect size greater than that when considering *HLA-DRB1* alone in the same individuals (OR, 0.65; 95% CI, 0.52–0.82). Likewise, the interaction term for a negative history of smoking with homozygosity for SE$^+$V11$^-$ alleles also showed a strong trend ($P = 0.06$), and we found that a history of having never smoked in combination with SE$^+$V11$^-$ alleles are predisposing to PD ($P = 0.01$; OR, 1.51; 95% CI, 1.08–2.12), with a greater effect size than for *HLA* alone (OR, 1.29; 95% CI, 1.01–1.66). Thus, it appears that the protective effect in PD of a positive history of smoking can be explained in part by interaction with *HLA* variation, suggesting that molecular changes mediated by cigarette smoke may be involved in alteration of *HLA* antigen presentation.

Citrullination of α-Synuclein Peptides Is Predicted to Alter Binding Affinity to HLA SE Alleles. Because α-synuclein aggregates in the PD brain, and T-cell recognition of α-synuclein peptides by PD patients has been recently reported and proposed as an autoantigen in disease (29), we explored the binding affinity of peptides derived from α-synuclein to the HLA SE allotypes using a bioinformatic prediction method. We used the NetMHCII3.2 server (30) to predict binding of peptides from α-synuclein to SE alleles associated with PD risk (V11$^-$) or protection (V11$^+$). We input the entire protein sequence of α-synuclein and predicted the probable peptides that bind to *HLA-DRB1*01:01* and *01:02* (SE$^+$V11$^-$) and to *HLA-DRB1*04:01, *04:04*, and *04:05* (SE$^+$V11$^+$). Overall, we found that peptides derived from α-synuclein were predicted to bind with higher affinity to SE$^+$V11$^-$ (risk) alleles compared with SE$^+$V11$^+$ (protective) alleles ($P = 0.004$); in fact, among the 126 peptides tested, none reached

the threshold for predicted binding (<500 nM) to the SE$^+$V11$^+$ alleles (*SI Appendix*, Table S5).

Given our finding that a history of smoking in our cohort interacts with *HLA-DRB1* in PD, and that smoking significantly increases posttranslational modifications such as citrullination and homocitrullination (31), which in turn may alter peptide-binding affinity for HLA class II (32), we performed replicate peptide-binding predictions with all lysine modified to homocitrulline residues (*SI Appendix*, Table S6). For SE$^+$V11$^-$ alleles, peptide modification is predicted to reduce the binding affinity of α-synuclein ($P = 2.2 \times 10^{-16}$). Conversely, peptide modification is predicted to increase the overall binding affinity to the SE$^+$V11$^+$ alleles ($P = 0.0001$). Importantly, the predicted increase in affinity for the SE$^+$V11$^+$ alleles resulted in a number of α-synuclein–derived peptides meeting the affinity threshold for binding. These results suggest that posttranslational modification of peptides mediated by cigarette smoking may simultaneously increase peptide affinity for protective *HLA-DRB1* alleles such as *HLA-DRB1*04:01* and decrease peptide affinity for risk *HLA-DRB1* alleles such as *HLA-DRB1*01:01*.

Discussion

Despite clear evidence for the role of *HLA* in genetic predisposition to PD, the complex and highly polymorphic character of the region has been a barrier to defining the precise nature of its contribution to disease. Here we have performed deep sequencing across entire genes of all polymorphic classical *HLA* loci. In doing so, we were able to identify the specific amino acids and elucidate structural features mediating both risk and protection in PD. Our data establish that the entirety of *HLA* risk and protection in PD can be explained by the SE of HLA-DRB1 at positions 70–74 in the β-chain α-helix defining the sides of the peptide-binding groove, in combination with position 11 in the β-pleated sheet on the floor of the groove.

While our findings are in agreement with numerous previous studies that have identified *HLA-DRB1*04* alleles as protective in PD (12, 14, 17, 20), by examining the *HLA* genes at sequence level resolution we refine these observations to clarify that only alleles of this lineage bearing the SE are protective. Extending the understanding of the role of HLA in PD, we find that another SE allele, *HLA-DRB1*01:01*, is predisposing in disease. While *HLA-DRB1*01:01* has not been previously associated with PD, a recent meta-analysis of genome-wide studies in PD identified *HLA-DRB6*, a pseudogene on the same haplotype, as a risk allele in PD (33).

Originally defined in rheumatoid arthritis, the "SE hypothesis" sought to explain the *HLA-DRB1* predisposition to disease across several allelic lineages (27). Among Europeans, the most common SE alleles belong to the *HLA-DRB1*01* and *HLA-DRB1*04* lineages and are predisposing to RA. The crystal structures for the SE alleles *HLA-DRB1*01:01* and *HLA-DRB1*04:01* reveal many mutual features with respect to the antigen-binding groove and have been shown to present several peptides in common; for example, both alleles have been crystallized with influenza virus protein hemagglutinin residues 306–318 (34). In contrast to RA, our observation in PD is of opposing effects for these alleles, with the direction of effect mediated by the presence or absence of 11-V; however, it is within reason that both risk and protective alleles present the same peptide.

In support of that model, a landmark study examining HLA function in Goodpasture disease demonstrated that risk vs. protective *HLA* alleles presenting the same immunodominant autoreactive peptide can produce markedly different T-cell responses (35), with the protective allele producing a more regulatory phenotype. We propose that HLA peptide specificity in PD is determined by the SE, residues of which participate in the P4, P7, and P9 pockets of the peptide-binding groove (36), while the shift

to a regulatory phenotype is mediated by 11-V, which defines the specificity for the P6 pocket (Fig. 2).

Our observation that the protective effect of cigarette smoking interacts with the *HLA-DRB1* SE draws another intriguing parallel to RA. In contrast to the protection afforded in PD, smoking is a well-established risk factor in RA (37). A common feature in RA is the presence of antibodies to citrullinated protein antigens (38), the generation of which has been associated with smoking (39). Further evidence suggests that citrullination enhances the binding affinity of some arithrogenic peptides to SE alleles (40), suggesting a pathway for interaction between smoking and *HLA* in RA. Our finding that citrullination increases the predicted affinity for *HLA-DRB1* alleles protective in PD while decreasing the predicted affinity for *HLA-DRB1* alleles predisposing to PD supports the notion that the epidemiologic association of smoking in both RA and PD is mediated by its posttranslational modification of antigens presented by *HLA-DRB1*. Finally, and in support of these observations, epidemiologic studies have shown that individuals with RA are at reduced risk of developing PD (41).

These results also lend credence to the notion that PD is, at least in part, autoimmune in nature. Further underscoring this notion, a recent study demonstrating T-cell autoreactivity to HLA class II presented peptides derived from α-synuclein, which accumulates in the PD brain (29), suggested that neuronal loss may be mediated by autoimmune mechanisms.

In summary, we find a clear and replicable role for *HLA* class II in predisposition and protection in PD and provide an explanation for the association of smoking with protection from disease. The association of specific combinations of amino acids that participate in critical peptide-binding pockets of the HLA class II molecule implies antigen presentation as a key factor in disease and strongly supports a role for immunogenetic variation in mediating the immunopathology of PD.

Methods

The study was approved by the University of California San Francisco (UCSF) Institutional Review Board.

Datasets.
Discovery cohort. The discovery cohort consisted of the National Institutes of Health (NIH)-GWAS and NIH-NeuroX datasets derived from the National Institute of Neurological Disorders and Stroke-funded Neurogenetics Repository
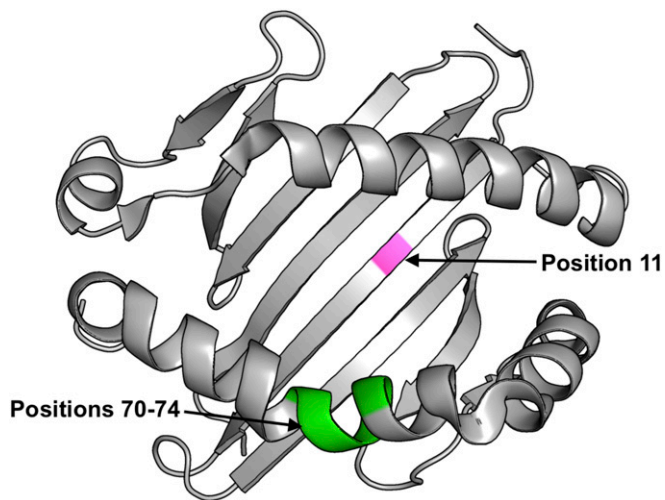

Fig. 2. HLA-DRB1 amino acid positions mediating risk and protection in PD. Position 11 (pink) is located in the β-pleated sheet on the floor of the peptide-binding groove of the HLA-DR molecule. The SE at positions 70–74 (green) is located on the second α-helix, bordering the peptide-binding groove of HLA-DR.

at the Coriell Institute for Medical Research, as well as the UCSF-GeneMSA dataset. After initial quality controls performed on *HLA* sequencing data, 3,647 samples with calling at all loci were received for analysis. Patients were removed from the discovery dataset if (*i*) they did not have available SNP data, (*ii*) they self-identified as nonwhite or non-Caucasian, (*iii*) they had an SNP genotyping rate <90% (see below), or (*iv*) the genetic population stratification analysis identified them as outliers (see below). The number of patients removed at each step of sample quality control is presented in *SI Appendix*, Table S7. This process resulted in a discovery dataset of 1,606 controls and 1,597 PD patients. ***Replication cohort.*** The replication cohort consisted of two Michael J. Fox Foundation for Parkinson's Research datasets (BioFIND and DATATOP) and UCSF controls (UCSF1). Patients were removed from the replication dataset if (*i*) they did not have an allele call at all loci considered for replication (*HLA-DQA1, HLA-DQB1,* and *HLA-DRB1*) or (*ii*) they self-identified as nonwhite or non-Caucasian or of Hispanic origin (*SI Appendix*, Table S8). Because no SNP data were available for these datasets, genetic population stratification was not performed. This process resulted in a replication dataset of 462 controls and 536 PD patients.

Demographic data for the discovery and replication cohorts are presented in *SI Appendix*, Table S9. All patients were non-Hispanic whites from the United States with idiopathic PD. All controls are reported to be unrelated non-Hispanic white and free from neurologic disorders. Informed consent was obtained for each participant under locally approved protocols.

SNP Data. Samples from the NIH dataset were genotyped on one of two Illumina arrays, the Immunochip or the NeuroX chip, which we refer to as the NIH-GWAS and NIH-NeuroX datasets, respectively. Some of the UCSF controls were genotyped on the HumanHap 550 BeadChip and are referred to as UCSF-GeneMSA dataset. For the UCSF-GeneMSA dataset, SNP positions were updated to the HG19 assembly using the LiftOver tool (https://genome.ucsc.edu/cgi-bin/hgLiftOver). Quality controls on samples included the genotyping rate per individuals and genetic outlier assessment, defined as individuals >6 SDs outside the mean for the first 10 PCs. Quality controls on SNPs included duplicated variant removal, Hardy–Weinberg equilibrium departure ($P < 0.001$), and low SNP call rate (<95%) (*SI Appendix*, Table S10). Quality control checks were performed at the dataset level, using R (42) and Plink v1.07 (43) software.

Control for Population Stratification. Population stratification analysis was performed using a core set of 8,006 SNPs. These 8,006 SNPs remained after excluding those that (*i*) did not pass quality control checks as described above; (*ii*) were not genotyped in the NIH-GWAS, NIH-NeuroX, and UCSF-GeneMSA datasets; (*iii*) were located on nonautosomal chromosomes; (*iv*) were from the extended MHC region (all markers on chromosome 6 from 26 to 36 Mb); (*v*) were from other regions of extended LD (all markers on chromosome 8 from 6 to 16 Mb and on chromosome 17 from 40 to 45 Mb); (*vi*) had a minor allele frequency <1%; or (*vii*) were from LD pruning so that no marker had a pairwise $r^2 > 0.1$ with any marker within a 100-SNP window. SNP data management was done using R (42) and Plink v1.07 (43). PCs were computed using the SNPRelate R package (44) on the final set of SNPs. Samples at >6 SDs from the mean for any of the first 10 PCs were considered population-specific outliers and removed (*SI Appendix*, Fig. S1). Using a final set of 7,918 SNPs, population stratification was also performed in regard to the 1000 Genomes white samples (*SI Appendix*, Fig. S2). A total of 12 samples were removed using these criteria.

HLA Next-Generation Sequencing and Genotyping. DNA samples were typed for 11 *HLA* loci—*HLA-A, -B, -C, -DPA1, -DPB1, -DQA1, -DQB1, -DRB1, -DRB3, -DRB4,* and *-DRB5*—using the MIA FORA NGS high-throughput typing kit and analysis software (Immucor) and performed according to the manufacturer's semiautomated protocol. Details are provided in *SI Appendix, Methods*.

Statistical Analysis. All analyses were performed in R (42). Associations of *HLA* alleles with disease were tested at all levels of resolution using logistic regression. Conditioned analyses were performed by adding specific alleles/variants of interest as covariates. Only *HLA* alleles with a frequency >1% were analyzed. Frequencies of *HLA* alleles and an omnibus test for association with specific amino acids were computed using the BIGDAWG R package (45). Logistic regression was also used to test for interactions between *HLA* alleles and between *HLA* alleles and known PD risk SNPs. Haplotype estimations and association analyses with disease were performed using the "haplo.glm" function in the haplo.stats R package (46), with gender and the first four PCs as covariates. All analyses were adjusted for gender and the first four PCs to correct for population stratification (*SI Appendix*, Fig. S3).

To test for interaction between smoking history and *HLA*, the model parameters of logistic regression were estimated using the maximum likelihood method. The Wald test was used to test the mean difference between the estimated coefficient and the null parameter; typically the null hypothesis assumed it equal to 0. The significance of the interaction term was further confirmed by the likelihood ratio test on the deviance between the full model (with interaction term) and the reduced model (without interaction term). Power was assessed through simulation to obtain empirical distributions for *P* values. Data were simulated 100 times based on the distribution of the predictors. The effect (β) of each predictor was used to determine the outcome (0 or 1). A binary logistic regression was fit on the simulated dataset, and the *P* value of the interaction term was extracted. The power to detect significant interactions with a single copy of SE^+V11^+ was 0.96, and the power to detect interactions with two copies of SE^+V11^+ was 0.36. Likewise, the power to detect significant interactions with a single copy of SE^+V11^- was 0.82, and the power to detect interactions with two copies of SE^+V11^- was 0.70.

The FDR was calculated as described previously (47) and was set to 10%, giving a significance threshold of $P < 0.02$ for the primary association analysis. Correction for multiple testing was performed using the Bonferroni method.

The data supporting the findings of this study are available from the corresponding author on request.

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