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Agonist-Specific Recruitment of Arrestin Isoforms Differentially Modify Delta Opioid Receptor Function

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Ligand-specific recruitment of arrestins facilitates functional selectivity of G-protein-coupled receptor signaling. Here, we describe agonist-selective recruitment of different arrestin isoforms to the delta opioid receptor in mice. A high-internalizing delta opioid receptor agonist (SNC80) preferentially recruited arrestin 2 and, in arrestin 2 knock-outs (KOs), we observed a significant increase in the potency of SNC80 to inhibit mechanical hyperalgesia and decreased acute tolerance. In contrast, the low-internalizing delta agonists (ARM390, JNJ20788560) preferentially recruited arrestin 3 with unaltered behavioral effects in arrestin 2 KOs. Surprisingly, arrestin 3 KO revealed an acute tolerance to these low-internalizing agonists, an effect never observed in wild-type animals. Furthermore, we examined delta opioid receptor–Ca²⁺ channel coupling in dorsal root ganglia desensitized by ARM390 and the rate of resensitization was correspondingly decreased in arrestin 3 KOs. Live-cell imaging in HEK293 cells revealed that delta opioid receptors are in pre-engaged complexes with arrestin 3 at the cell membrane and that ARM390 strengthens this membrane interaction. The disruption of these complexes in arrestin 3 KOs likely accounts for the altered responses to low-internalizing agonists. Together, our results show agonist-selective recruitment of arrestin isoforms and reveal a novel endogenous role of arrestin 3 as a facilitator of resensitization and an inhibitor of tolerance mechanisms.

Key words: arrestin; DRG; GPCR; pain; resensitization; tolerance

Significance Statement

Agonists that bind to the same receptor can produce highly distinct signaling events and arrestins are a major mediator of this ligand bias. Here, we demonstrate that delta opioid receptor agonists differentially recruit arrestin isoforms. We found that the high-internalizing agonist SNC80 preferentially recruits arrestin 2 and knock-out (KO) of this protein results in increased efficacy of SNC80. In contrast, low-internalizing agonists (ARM390 and JNJ20788560) preferentially recruit arrestin 3 and, surprisingly, KO of arrestin 3 produces acute tolerance and impaired receptor resensitization to these agonists. Arrestin 3 is in pre-engaged complexes with the delta opioid receptor at the cell membrane and low-internalizing agonists promote this interaction. This study reveals a novel role for arrestin 3 as a facilitator of receptor resensitization.

Introduction

G-protein-coupled receptors (GPCRs) regulate diverse biological functions and are the most abundant receptor class in the

human genome (Lagerström and Schiöth, 2008). These seven-transmembrane receptors exist in multiple conformations and ligands can stabilize different active states (Kenakin, 2003). Each receptor conformation can produce distinct receptor–effector

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complexes that initiate differing signaling and receptor trafficking events (Kenakin, 2012; Reiter et al., 2012). The existence of multiple activation states confer an added degree of complexity to GPCR signaling and allow for highly distinct responses among ligands that act at the same receptor. This concept, referred to as ligand-directed signaling, functional selectivity, or biased agonism (Galandrin et al., 2007; Luttrell, 2014), has important biological and pharmacological implications.

Agonist-selective recruitment of arrestins is a major source of functional selectivity between ligands that bind to the same receptor (Reiter et al., 2012). Arrestins are multifaceted proteins that play an important role in GPCR trafficking and signaling (Shenoy and Lefkowitz, 2011). Agonist binding can produce receptor phosphorylation by G-protein-receptor kinases (GRKs), which in turn results in the binding of arrestins, thus terminating the G-protein-dependent signaling at the membrane (Pierce et al., 2002). Arrestins are also an intracellular scaffold hosting a number of signaling networks, including MAP kinases, c-Src, and Akt-GSK3 (Rajagopal et al., 2010; Shenoy and Lefkowitz, 2011; Reiter et al., 2012). More recently, novel roles of arrestins have emerged as regulators of cytoskeletal proteins (Mittal et al., 2013), protein ubiquitination (Shenoy, 2014), and trafficking of ion channels (Shenoy and Lefkowitz, 2011). Although arrestins have been studied extensively, novel functional aspects continue to be uncovered.

Delta opioid receptor agonists are currently being developed for the treatment of chronic pain and psychiatric disorders (anxiety and depression; Pradhan et al., 2011; van Rijn et al., 2013; Gendron et al., 2015). Similar to other GPCRs, delta opioid receptors are also modulated by arrestins (Lowe et al., 2002; Zhang and Horvath, 2005; Qiu et al., 2007; Raehal and Bohn, 2011). In cellular models, both arrestin 2 (β -arrestin 1) and arrestin 3 (β -arrestin 2) can mediate delta opioid receptor desensitization and internalization after receptor phosphorylation (Qiu et al., 2007; Hong et al., 2009). However, evidence from a number of *in vitro* and *in vivo* studies has revealed biased agonism at the delta opioid receptor (for review, see Pradhan et al., 2012). Live-cell imaging studies using bioluminescence resonance energy transfer (BRET) revealed that, in response to different agonists, the delta opioid receptor can adopt distinct receptor conformations (Audet et al., 2008; Audet et al., 2012; Richard-Lalonde et al., 2013; Charfi et al., 2014; 2015). In addition, delta receptor trafficking is agonist dependent, which has important behavioral consequences (Law et al., 2000; Zhang et al., 2005; Bradbury et al., 2009; Pradhan et al., 2009; Tudashki et al., 2014). Previously, we compared two delta agonists with similar binding and analgesic properties, but high- (SNC80) or low- (ARM390) receptor internalization potencies. After repeated injection, SNC80, but not ARM390, produced acute behavioral tolerance. Upon examination of receptor trafficking, we showed that receptor internalization was directly correlated with receptor function *in vivo* (Pradhan et al., 2009). We also examined the long-term effects of treating with these different delta agonists and found that distinct forms of chronic tolerance were observed depending on the internalization property of the drug (Pradhan et al., 2010).

The aim of this study was to determine the behavioral significance of delta opioid receptor interactions with arrestin 2 and arrestin 3 and whether this regulation was ligand specific. We show that, *in vivo*, delta agonists recruit arrestins in a selective manner: binding of a high-internalizing agonist produces preferential interaction between the receptor and arrestin 2 and low-internalizing agonists preferentially promote arrestin 3-receptor interactions. We also show for the first time that increased arres-

tin 3 engagement with the delta opioid receptor can promote receptor resensitization, in marked contrast to its traditional role as a facilitator of desensitization.

Materials and Methods

Animals. In this study, arrestin 2 and arrestin 3 knock-outs (KOs), DOR-eGFP knock-in, and arrestin 3 KO \times DOR-eGFP KI mice were used. Arrestin 2 and 3 KOs were generously provided by R. Lefkowitz (Duke University). DOR-eGFP knock-in mice were described previously (Scherrer et al., 2006). Mice were obtained through heterozygous pairings and both males and females were used. Mice, aged 12 weeks on average, were housed in a temperature- and humidity-controlled animal colony on a 12 h light/dark cycle. Food and water were available *ad libitum*. All animal experiments were performed according to Association for Assessment and Accreditation of Laboratory Animal Care guidelines as administered by the University of California–Los Angeles Office of Animal Research Oversight and the University of Illinois–Chicago Animal Care Committee.

Inflammatory pain model. All experiments were performed between 8:00 and 16:00. In all cases, mice were habituated to the testing area for 20 min daily for 2 d before baseline testing. For mechanical responses, the threshold for responses to punctate mechanical stimuli (mechanical hyperalgesia) was tested according to the up-and-down method (Chaplan et al., 1994). In this case, the plantar surface of the hindpaw was stimulated with a series of 8 von Frey filaments (bending force ranging from 0.01 to 2 g). A response was defined as a lifting or shaking of the paw upon stimulation. Inflammatory pain was induced by injecting complete Freund's adjuvant (CFA) into the paw. Before the injection of CFA, baseline mechanical responses were determined. Inflammation was induced by injecting 13 μ l of CFA into the plantar surface of the paw and animals were subsequently tested 72 h after injection.

Drugs. All drugs were administered in a volume of 10 ml/kg. SNC80 was dissolved in 0.9% saline, pH 5.5, and injected intraperitoneally. ARM390 and JNJ20788560 were dissolved in dH₂O and administered by oral gavage. Animals were tested 45 min after treatment with SNC80 and ARM390 and 30 min after treatment with JNJ20788560.

DRG preparation and electrophysiological recording. Delta opioid receptor voltage-dependent Ca²⁺ channel (VDCC) inhibition was assessed by whole-cell patch-clamp recordings from acutely dissociated L4–L6 DRGs. Adult mice were treated with CFA and, after 72 h, they were killed and the ipsilateral DRGs collected in complete saline solution (CSS) containing the following (in mM): 137 NaCl, 5.3 KCl, 1 MgCl₂, 25 sorbitol, 10 HEPES, and 3 CaCl₂ and incubated in collagenase [1.25 U of Liberase TH (high thermolysin); Roche], 250 nm EDTA for 20 min at 32°C, transferred to fresh CSS containing collagenase [1.25 U of Liberase TM (medium thermolysin); Roche] with 250 nm EDTA and 0.25 U of papain (Roche), and incubated for 10 min at 32°C. After 2 washes and physical trituration through a series of graded Pasteur pipettes, the cells were spun (1000 rpm, 3 min) and plated in Neurobasal/B27/Glucose/Antibiotic/Antimycotic (Life Technologies) supplemented with 10 ng/ml NGF (Life Technologies). All recordings were performed within 6 h after plating. For each recording, culture medium was replaced by an external solution that contained the following (in mM): 130 TEA-Cl, 10 CaCl₂, 5 HEPES, 25 D-glucose, and 2.5 \times 10⁻⁴ tetrodotoxin, pH 7.2. Recording electrodes contained the following (in mM): 105 CsCl, 40 HEPES, 5 D-glucose, 2.5 MgCl₂, 10 EGTA, 2 Mg²⁺-ATP, and 0.5 Na⁺-GTP, pH 7.2. The potential difference between the open electrode and the bath ground was zeroed before establishing a \geq 1 G Ω resistance seal. No compensation was made for the cancellation of liquid junction potential. Ca²⁺ currents were activated by depolarizing neurons from -80 to 10 mV for 100 ms at 10 s intervals. Currents were low-pass filtered at 2 kHz and digitized (Digidata; Molecular Devices) at 10 kHz. Leak currents were nulled using the P/4 subtraction method. DRG neurons were rapidly and continuously superfused (5 ml/min) with external solution. SNC80 (1 μ M) was diluted into external solution on the day of the experiment and applied through the perfusion system. Experiments were performed at room temperature (22–24°C). Mean Ca²⁺ current amplitudes were measured (pCLAMP 9.0; Molecular Devices) between 5 and 10 ms

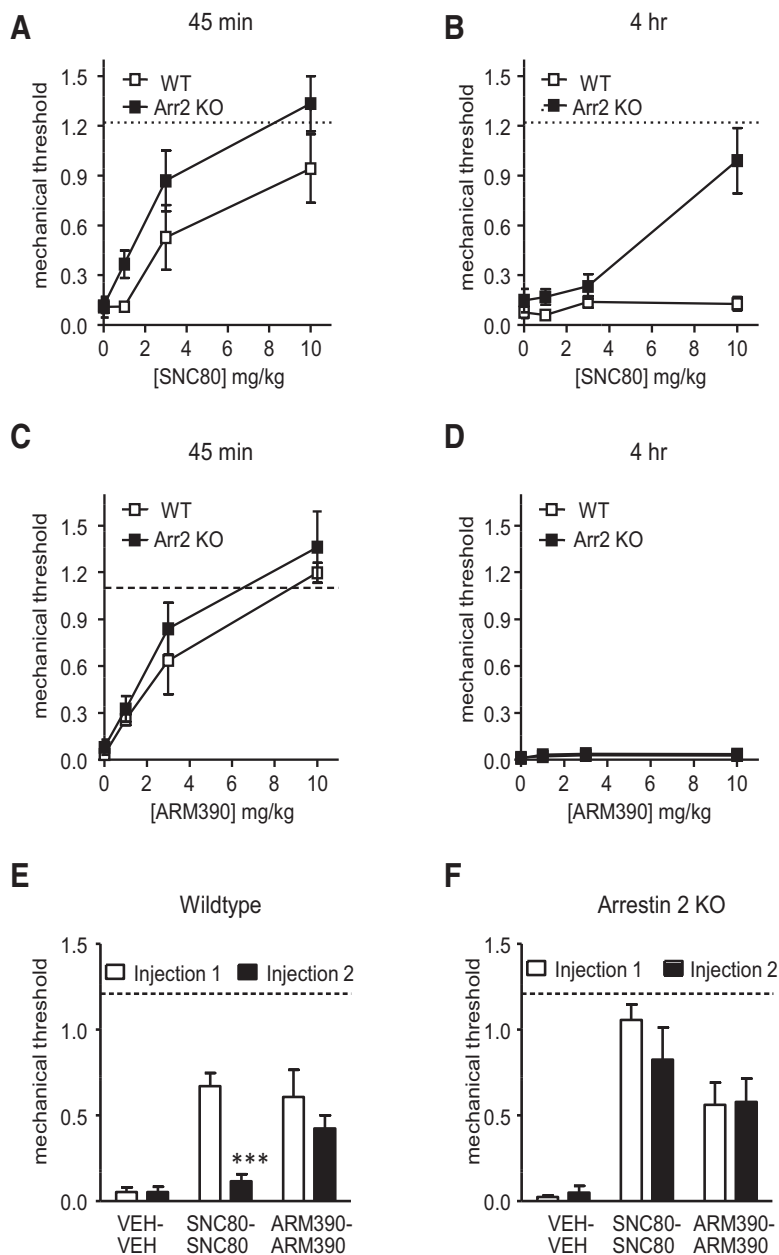


Figure 1. SNC80, but not ARM390, has increased potency and duration of action, and decreased acute tolerance in arrestin 2 KO mice. In a CFA model of inflammatory pain, we performed a dose and time response to SNC80 and ARM390. The anti-hyperalgesic effect of SNC80 (IP; **A**) was significantly enhanced in arrestin 2 KO mice ($p < 0.05$), and was longer lasting (**B**) ($p < 0.001$). **C, D**, ARM390 (PO) was equipotent in arrestin 2 WT and KO mice (**C**) and anti-hyperalgesic effects were lost 4 h after administration regardless of genotype (**D**). For acute tolerance experiments, Injection 1 represents mechanical responses in WT (**E**) and arrestin 2 KO (**F**) mice acutely treated with vehicle (control), SNC80 (3 mg/kg), or ARM390 (3 mg/kg). For Injection 2, animals were rechallenged with the same drugs and doses 4 h after Injection 1. Dashed lines represent baseline mechanical responses pre-CFA. For dose/time responses, two-way ANOVA was used ($n = 5-7$ mice/group). For acute tolerance, paired t tests with Bonferroni correction were used ($***p < 0.001$; $n = 7-10$ mice/group). SNC80 is more effective in arrestin 2 KO mice.

after initiating the depolarizing step. Mean current amplitudes were then plotted against time. Stable recordings were fitted by a linear function to compare, by extrapolation, control current amplitude with the current amplitude recorded in the presence of opioid receptor agonists.

Confocal microscopy. To determine the subcellular distribution of DOR-eGFP after agonist stimulation, mice were anesthetized with pentobarbital (somnosol, 150 mg/kg) and intracardially perfused with 10 ml of 0.1 M phosphate buffer (PB), followed by 30 ml of 4% paraformaldehyde in 0.1 M PB, pH 7.4. Dorsal root ganglia (DRG) were then postfixed for 2 h at 4°C in the fixative solution. The tissue was then cryoprotected at

4°C in a 30% sucrose, 0.1 M PB solution until the tissue sank. Sections were cut at 14 μ m in a cryostat and mounted on charged glass slides. Sections were immunostained for GFP. Briefly, tissue was blocked with 5% normal donkey serum/0.3% Triton X-100/0.1 M PB for 2 h. Sections were incubated overnight with rabbit anti-GFP antibody (1:1000; Life Sciences) at room temperature. Sections were washed and incubated with a donkey anti-rabbit secondary antibody conjugated to Alexa Fluor 488 (1:1000; Life Sciences) for 2 h at room temperature and then washed in PBS and coverslipped. All samples were observed under a Zeiss LSM 710 confocal microscope (63 \times objective and numerical aperture of 1.2) and Zen (Zeiss) software was used for image acquisition. Quantification of cell surface mean fluorescence intensity was determined using ImageJ software. Nuclear fluorescence defined the background level and was subtracted from the cell membrane fluorescence measures (for further details, see Scherrer et al., 2006). In total, three to five DRGs/mouse were analyzed and there were five to nine mice/group.

BRET analysis. For BRET in cell population, cells were counted by measuring protein concentration (Dc protein assay kit; Bio-Rad). For total fluorescence and luminescence measurements, cells were dispersed in 96 well plates with clear bottoms (Costar) at a density of 100,000 cells per well. Total fluorescence of the cells was measured in the Mithras LB 940 instrument (Berthold Technologies) using an excitation filter at 485 nm and an emission filter at 530 nm. The light emitted at 530 nm upon light excitation at 485 nm (Fluo) is indicative of the amount of YFP-tagged proteins. Cells were then incubated with Deep Blue C coelenterazine (5 mM) to measure the total luminescence using the Mithras. The light emitted at 400 nm in presence of Deep Blue C (Lumi) is indicative of the amount of Rluc-tagged proteins. For BRET measurements, cells coexpressing δ OR-Rluc and Arr2-YFP or Arr3-YFP constructs were distributed in white 96-well microplates (Corning) at a density of 100 000 cells per well. BRET was initiated by adding 5 mM coelenterazine H and the ratio of the light emitted by YFP (510–550 nm) to the light emitted by the Rluc (460–500 nm) was measured using a fluorimeter (Mithras; Berthold Technologies). Values were corrected by subtracting the background ratio detected when the Rluc construct was expressed alone.

Single-cell BRET imaging in HEK293 cells to study the subcellular localization of the interaction between arrestins and δ OR-Rluc8 were performed according to previous protocols (Coulon et al., 2008; Perroy, 2010). Briefly, images were obtained using a Plan-Apochromat 63 \times /1.40 oil M27 objective at room temperature. Transfected cells were first identified using a monochromatic light and appropriate filter to excite YFP (exciter HQ480/40, emitter HQ525/50; Chroma) or mOrange (exciter HQ540/40, emitter HQ600/50; Chroma). The light source was then switched off until the end of the experiment. Coelenterazine H (20 μ M) was applied for 5 min before acquisition with MetaMorph software (Molecular Devices). BRET images were collected every 45 s by sequential acquisitions from the 480 nm, 535 and 562 nm channels of 15 s each, using the evolve

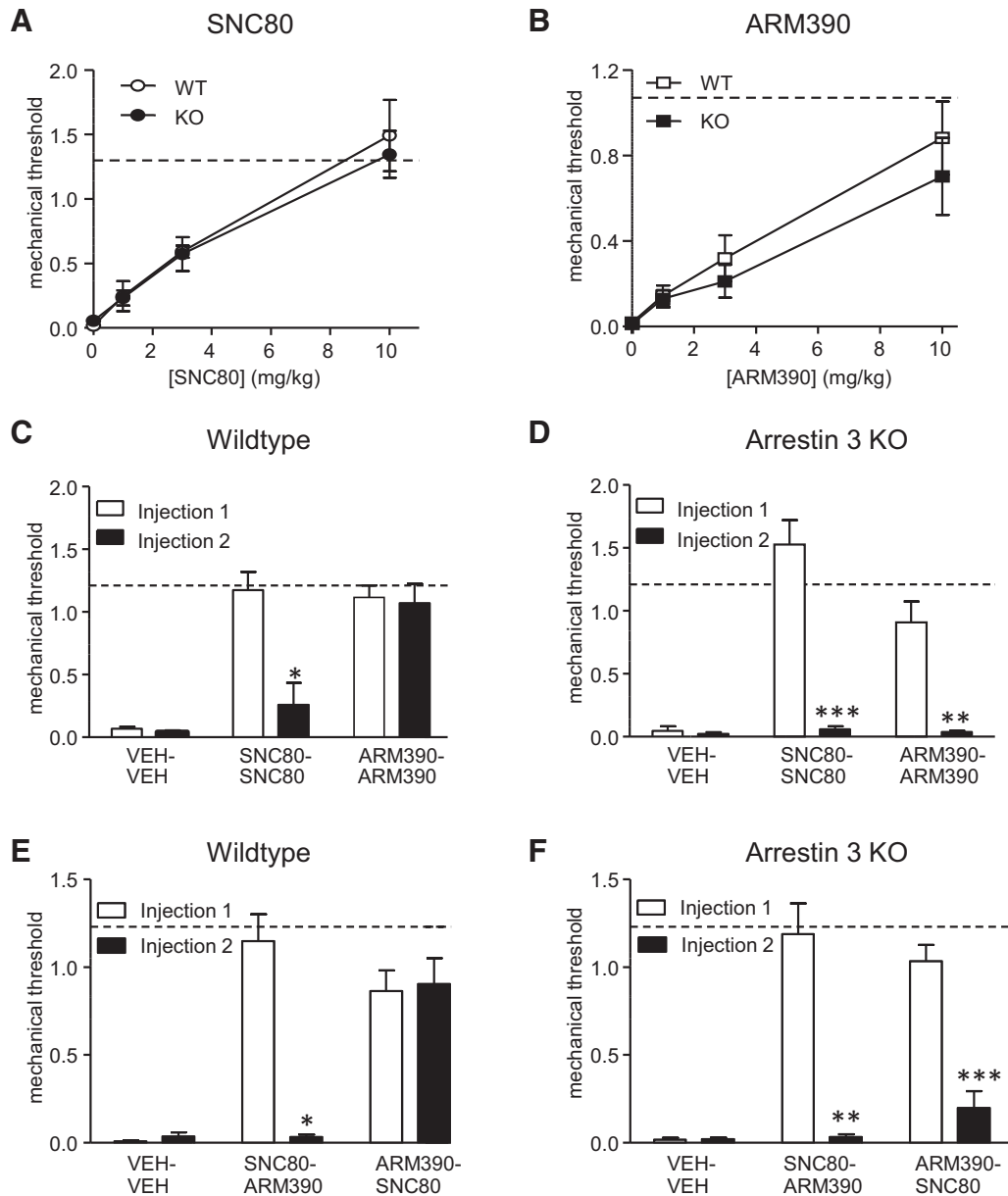


Figure 2. Acute tolerance and cross-tolerance to ARM390 is gained in arrestin 3 KO mice. Mechanical responses were determined in the CFA model of inflammatory pain. Arrestin 3 WT and KO animals were treated with differing doses of SNC80 (**A**) or ARM390 (**B**) and tested 45 min after drug administration ($n = 5-7$ /group). KO of arrestin 3 does not change potency of either delta agonist. For acute tolerance and cross-tolerance experiments (**C-F**), Injection 1 and Injection 2 were 4 h apart. WT and arrestin 3 KO mice were treated with vehicle (control), SNC80 (10 mg/kg), or ARM390 (10 mg/kg) and then rechallenged 4 h later with the same drug (**C, D**) or the other drug (**E, F**). Dashed lines represent baseline mechanical responses pre-CFA. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, paired t test with Bonferroni correction ($n = 4-10$ mice/group). In the absence of arrestin 3, ARM390 produces acute and cross-tolerance at the delta opioid receptor.

camera from Photometrics. Drugs were added 3 min after the first acquisition. Sequential acquisitions were performed at 5 MHz (Gain 3950, binning 1) with emission filters D480/60 nm, HQ535/50 nm, and HQ585/40 nm to select em480 (Rluc), em535 (YFP), and em562 (mOrange) wavelengths, respectively. An exclusive threshold was applied on the Em 480 image from 0 to 4000 counts to exclude these non-reliable weak values. The pixel-by-pixel 535 nm/480 nm ratios, BRET1 images were calculated by dividing the absolute blue or yellow intensities per pixel of images obtained at 535 nm over 480 nm. The pixel-by-pixel 562 nm/480 nm ratios, BRET3 images, were also obtained by dividing the absolute blue or orange intensities per pixel of images obtained at 562 nm over 480 nm. These numerical ratios (comprising between 0 and 1 for BRET1 and between 0 and 0.3 for BRET3) were translated and visualized with a continuous 256 pseudocolor look-up table as displayed in the figures. The average intensity of the 535 nm/480 and 562 nm/480 nm fluorescence ratios were determined in the square region of 20×20

pixels drawn on the subcellular compartment of interest using ImageJ software. The mean \pm SEM obtained from three to six cells and seven square regions per cell in the same subcellular compartment and in identical stimulating conditions were averaged.

Statistical analysis. Data are expressed as mean \pm SEM. All graphs were produced in GraphPad Prism version 4 and all statistical analysis was performed in SigmaStat software. For pain experiments, two-way repeated-measures ANOVAs were performed or, where cited, t tests with Bonferroni correction were used. For electrophysiological and BRET experiments, one-way ANOVA was performed. For all ANOVA analyses, a Holm-Sidak *post hoc* analysis was performed when appropriate.

Results

We compared delta opioid receptor specific agonists with similar binding and anti-hyperalgesic properties, but high (SNC80) or

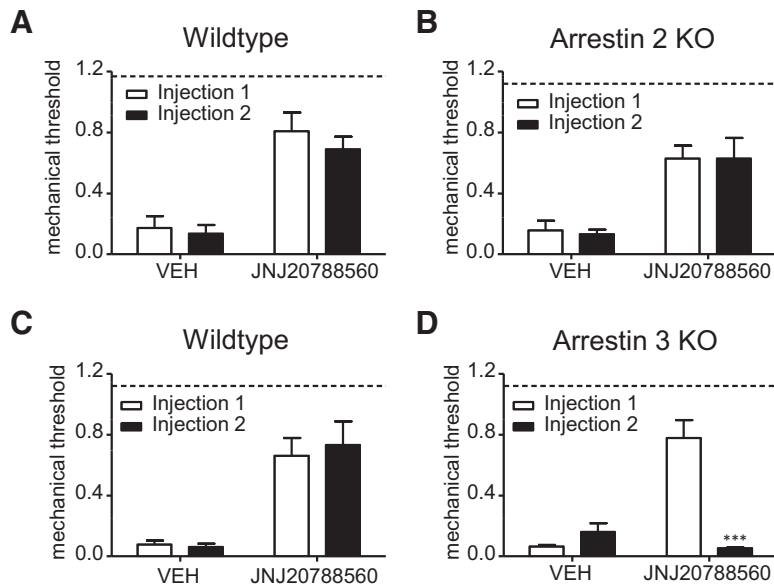


Figure 3. The low-internalizing agonist JNJ20788560 shows similar properties to ARM390 in arrestin 2 and 3 KO mice. Mechanical responses were determined in the CFA model of inflammatory pain. Arrestin 2 WT (**A**), arrestin 2 KO (**B**), arrestin 3 WT (**C**), and arrestin 3 KO (**D**) mice were tested with vehicle (VEH) or JNJ20788560 (100 mg/kg, p.o.) and then rechallenge with the same compound 4 h later. Dashed lines represent baseline mechanical responses pre-CFA. *** $p < 0.001$, two-way repeated-measures ANOVA ($n = 7$ –15 mice/group). KO of arrestin 3 produces acute behavioral tolerance to low-internalizing delta agonists.

low (ARM390 or JNJ20788560) internalization potencies (Pradhan et al., 2009; Pradhan et al., 2010). We characterized the ligand-specific effects in both arrestin 2 and arrestin 3 KO mice. The CFA mouse model of inflammatory pain was used throughout the *in vivo* experiments.

Acute behavioral tolerance to SNC80, but not ARM390, is attenuated in arrestin 2 KO mice

We initially tested the effect of arrestin 2 KO on the anti-hyperalgesic effect of delta opioid receptor agonists in the CFA inflammatory pain model. We performed a dose and time response to SNC80 and ARM390 in arrestin 2 wild-type (WT) and KO mice. CFA was injected into the hindpaw and, 72 h later, mice were injected with varying doses of SNC80 and ARM390 and tested 45 min (Fig. 1*A,C*) and 4 h (Fig. 1*B,D*) after injection. SNC80 was significantly more potent in arrestin 2 KO mice (Fig. 1*A*) and, at the highest dose, its pain-relieving effects were longer lasting (Fig. 1*B*). In contrast, ARM390 had similar potency (Fig. 1*C*) and duration of action in arrestin 2 WT and KO animals (Fig. 1*D*).

We showed previously that repeated injection of the high-internalizing agonist SNC80 produces acute behavioral tolerance, whereas the low-internalizing agonist ARM390 does not (Pradhan et al., 2009). Here, we tested arrestin 2 WT and KO within the same paradigm. To avoid the increased duration of action of the high dose of SNC80, we tested these animals with a submaximal 3 mg/kg dose. CFA was injected into the mouse paw and severe mechanical hypersensitivity was observed 72 h later (Fig. 1*E,F*, dashed line vs vehicle controls). In WT and arrestin 2 KO mice, an initial injection of SNC80 and ARM390 attenuated this hyperalgesia (Fig. 1*E*, Injection 1). As reported previously, a second injection of SNC80 was ineffective in WT animals (Fig. 1*E*, Injection 2); however, this acute behavioral tolerance was not observed in arrestin 2 KOs (Fig. 1*F*, Injection 2). A second injection of ARM390 was equally effective in both WT and KO mice. These results indicate that arrestin 2 disrupts acute tolerance to high-internalizing, but not low-internalizing, agonists.

Acute behavioral tolerance to ARM390 is enhanced in arrestin 3 KO mice

Arrestin 3 has been shown previously to regulate opioid receptor trafficking in a number of cellular model systems (Qiu et al., 2007; Hong et al., 2009). We examined the *in vivo* effect of arrestin 3 on the regulation of delta opioid receptors after agonist binding. An initial dose–response curve showed that there was no significant difference between arrestin 3 WT and KO mice to either SNC80 (Fig. 2*A*) or ARM390 (Fig. 2*B*). We performed an acute tolerance experiment and an initial injection of SNC80 and ARM390 was effective at reducing CFA-induced hyperalgesia (Fig. 2*C,D*, Injection 1). A second injection of SNC80 produced acute behavioral tolerance in both arrestin 3 WT and KO mice (Fig. 2*C,D*, Injection 2), which is consistent with the notion that SNC80-bound delta opioid receptors preferentially recruit arrestin 2. Surprisingly, arrestin 3 KOs acquired acute behavioral tolerance to the low-internalizing agonist ARM390 (Fig. 2*D*, Injection 2). Acute tolerance to ARM390 is

not observed in WT animals (Figs. 1*E*, 2*C*, Injection 2, and Pradhan et al., 2009; Pradhan et al., 2010) and the low-internalizing properties of ARM390 would suggest that arrestins would not be recruited after its binding. To confirm that this tolerance was due to changes at the delta opioid receptor, we did a cross-tolerance study in which the animals initially received SNC80 or ARM390 and were then rechallenged 4 h later with the other drug. An initial injection of SNC80 produced cross-tolerance to ARM390 in both WT and arrestin 3 KO mice (Fig. 2*E,F*). In contrast, injection of ARM390 only produced cross-tolerance in arrestin 3 KO mice, an effect that was not observed in WT controls (Fig. 2*E,F*, Injection 2). Therefore, KO of arrestin 3 results in a gain of acute tolerance to the low-internalizing agonist ARM390 and implicates arrestin 3 as being protective against delta opioid receptor tolerance mechanisms after binding to low-internalizing, but not high-internalizing, agonists.

We also determined the time to recover from the acute tolerance induced by ARM390 in arrestin 3 KOs. We found that a second injection 24 h after the first injection of ARM390 showed a pain-relieving effect of this agonist comparable to Injection 1 (Injection 1: 0.987 ± 0.097 vs Injection 2 24 h later: 1.27 ± 0.32). How this recovery occurs is currently unknown, but possibilities include *de novo* receptor synthesis, trafficking of receptor from intracellular stores, or an unknown recycling/resensitization process.

To confirm that ARM390 was not the only delta agonist that resulted in this unique interaction with arrestin 3, we tested another low-internalizing agonist, JNJ20788560 (Codd et al., 2009; Fig. 3). Similar to ARM390, KO of arrestin 2 did not affect acute tolerance (Fig. 3*A,B*); however, complete behavioral acute tolerance to JNJ20788560 was observed in arrestin 3 KO mice (Fig. 3*C,D*). The interaction of arrestin 3 to protect against acute tolerance at the delta opioid receptor is therefore not unique to ARM390 and also appears to be a property of other low-internalizing delta agonists.

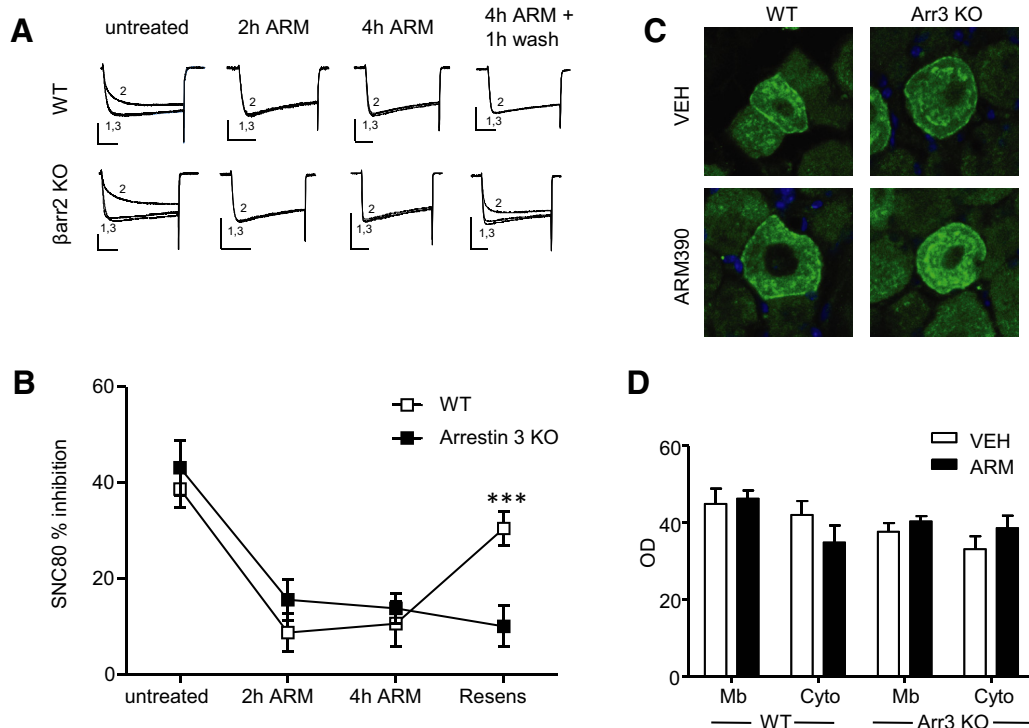


Figure 4. Arrestin 3 KO attenuates the rate of resensitization of delta opioid receptor function after ARM390, but does not alter receptor trafficking. **A**, Representative curves and mean SNC 80% inhibition from patch-clamp analysis of Ca^{2+} channel inhibition in DRGs from arrestin 3 WT and KO mice. Exemplar currents show VDCCs before (1), during (2), and after (3) SNC80 ($1 \mu M$) application. **B**, Desensitization, assessed by a $1 \mu M$ test dose of SNC80, occurred equally in both WT and KO DRGs after 2 and 4 h of ARM390 (100 nM) pre-incubation. Thereafter, SNC80 inhibition of VDCCs recovered after 1 h of washout in WT, but not in DRGs from arrestin 3 KO mice $***p < 0.001$, one-way ANOVA with Holm–Sidak *post hoc* analysis ($n = 7–12$ cells). In addition, DOR-eGFP reporter mice were crossed with arrestin 3 WT and KOs. Mice were treated with vehicle or ARM390 (10 mg/kg, p.o.) and DRGs were collected 4 h later. **C**, DRGs were analyzed by confocal microscopy and representative images are shown. **D**, Mean cell membrane (Mb) and cytosolic (Cyto) fluorescence was quantified in three to five neurons from each mouse, with five to nine mice/group. Delta opioid receptors in arrestin 3 KOs showed decreased resensitization after ARM390 that was not due to enhanced receptor internalization.

Arrestin 3 deletion impairs resensitization of delta opioid receptor function in DRG

We focused the next experiments on characterizing the acute behavioral tolerance to low-internalizing agonists observed in arrestin 3 KOs. We started by examining the modulation of Ca^{2+} channels by delta opioid receptors in DRG. DRGs are the first-order neurons in the pain transmission pathway and the pain-relieving effects of delta agonists within peripheral pain models are mediated by a subset of these neurons (Gavériaux-Ruff et al., 2011). We assessed delta opioid receptor inhibition of Ca^{2+} currents as a measure of receptor function (Walwyn et al., 2005). The conditions used in these experiments were chosen specifically to reflect our *in vivo* acute tolerance paradigm. Adult arrestin 3 WT and KO mice were injected with CFA and L4–L6 ipsilateral DRGs were collected 72 h later and acutely dissociated. We assessed both the rates of desensitization and resensitization of delta opioid receptor inhibition of VDCCs by a $1 \mu M$ SNC80 challenge. Desensitization was defined as loss of delta opioid receptor inhibition of VDCCs after 2 or 4 h application of ARM390; and resensitization as the recovery of this inhibition after a 1 h washout. We observed full VDCC desensitization, as defined by reduced SNC80–VDCC inhibition, by 4 h in both arrestin 3 WT and KO mice (Fig. 4A,B). Resensitization, as defined by a recovery of SNC80–VDCC inhibition after 1 h of washout, occurred in WT, but not arrestin 3 KO, DRGs (Fig. 4A,B). These results indicate that arrestin 3 endogenously facilitates resensitization of the delta opioid receptor after ARM390-induced desensitization.

We also determined whether ARM390 induced internalization in arrestin 3 KO mice, which might explain the acquired

acute tolerance in these animals. We crossed the reporter DOR-eGFP knock-in mice (Scherrer et al., 2006) with arrestin 3 KO mice. Mice were injected with CFA and, 72 h later, treated with ARM390; receptor localization was determined 4 h after drug administration. ARM390 did not appear to cause significant internalization of DOR-eGFP in DRGs from arrestin 3 KO mice (Fig. 4C,D). We conclude that it is unlikely that the behavioral desensitization induced by ARM390 in arrestin 3 KO is due to receptor sequestration.

Arrestin 3 and delta opioid receptors are pre-engaged at the cell surface and ARM390 binding increases this interaction

Our *in vivo* behavioral and electrophysiological results suggest that agonist bias at the delta opioid receptor can result from differential recruitment of arrestin isoforms. To examine the basal and agonist-induced interactions between delta opioid receptors and arrestin 2 or 3 directly, we used BRET imaging in transfected HEK293 cells. We used this assay to examine the protein–protein interactions that occur basally and soon after agonist activation of the receptor. Initially, we examined the basal interaction between delta opioid receptor and each arrestin alone in the absence of agonist. The C terminus of the delta opioid receptor was fused to the energy donor *Renilla luciferase* (δOR -Rluc8) and each arrestin was fused to the energy acceptor YFP (Arr2-YFP, Arr3-YFP). Under conditions in which δOR -Rluc8 expression levels were kept constant, BRET signal increased hyperbolically as a function of Arr3-YFP, but not Arr2-YFP, expression level (Fig. 5A). Saturation of the BRET signal when all of the donor was linked to the acceptor indicated a specific basal inter-

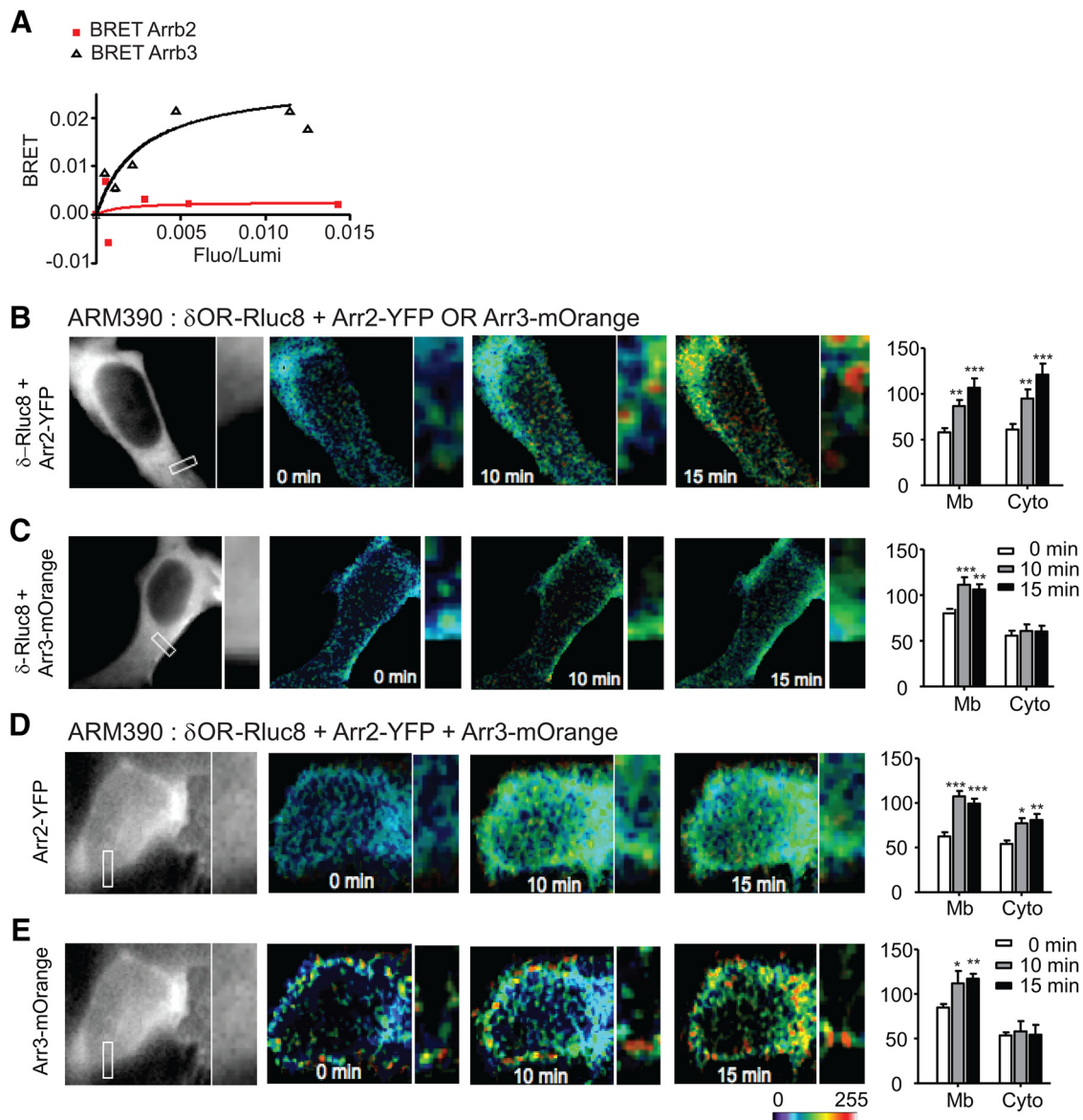


Figure 5. Arrestin 3 and delta opioid receptors are pre-engaged on the cell membrane and ARM390 increases this association. **A**, Basal BRET signal between δ OR-Rluc and Arr2-YFP or Arr3-YFP. HEK293 cells were cotransfected with a constant concentration of δ OR-Rluc and increasing concentrations of Arr2-YFP or Arr3-YFP plasmids. The data shown are representatives of five independent experiments. The curves were fitted using a nonlinear regression equation assuming a single binding site. BRET is expressed as a function of δ OR expression for a constant amount of arrestin and shows increased basal interaction at the cell membrane between δ OR-Rluc8 and Arr3-YFP, which is not observed between δ OR-Rluc8 and Arr2-YFP. **B–E**, To determine the effect of ARM390 on delta opioid receptor-arrestin interactions, HEK293 cells were transfected with δ OR-Rluc8 and Arr2-YFP (**B**), δ OR-Rluc8 and Arr3-mOrange (**C**), or δ OR-Rluc8, Arr2-YFP, and Arr3-mOrange (**D**, **E**). For the three partners-BRET experiment, **D** shows representative images from the Arr2-YFP channel and **E** those from the Arr3-mOrange channel. Cells were incubated with ARM390 (100 nM) and mean BRET intensity was quantified for the cell membrane (Mb) and cytosol (Cyto) at 0, 10, and 15 min after ARM390 administration. * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$, one-way ANOVA with Holm–Sidak *post hoc* analysis ($n = 3–6$ cells and 7 square regions per subcellular compartment). Delta opioid receptors are pre-engaged with Arr3 at the cell membrane and this interaction is strengthened after ARM390 binding.

action between δ OR-Rluc8 and Arr3-YFP proteins in absence of ligand, an effect not observed between δ OR-Rluc8 and Arr2-YFP.

We next examined the effect of delta agonists on delta opioid receptor-arrestin interactions. In this case, the C termini of arrestin 2 and arrestin 3 were fused to the energy acceptors YFP (Arr2-YFP) and mOrange (Arr3-mOrange), respectively. We initially performed BRET analysis by expressing δ OR-Rluc8 with either Arr2-YFP or Arr3-mOrange separately. To determine the subcellular interactions between these molecules, we quantified cell membrane and cytosolic interactions separately. We used a dose of ARM390 (0.1 μ M), which was not shown previously to induce internalization in primary cell cultures (Pradhan et al.,

2009). Application of ARM390 produced a time-dependent increase in BRET signal between δ OR-Rluc8 and Arr2-YFP at both the membrane and cytosol (Fig. 5B). These results indicate that arrestin 2 induces intracellular complexes with the delta opioid receptor after ARM390 binding in the absence of arrestin 3. In contrast, when we examined BRET between δ OR-Rluc8 and Arr3-mOrange, ARM390 produced a significant enhancement of BRET signal at the plasma membrane specifically (Fig. 5C, Mb), but did not appear to cause detectable intracellular interactions (Fig. 5C, Cyto).

We next coexpressed δ OR-Rluc8 with both Arr2-YFP and Arr3-mOrange. After the addition of ARM390, δ OR-Rluc8 and Arr2-YFP interactions were increased at the cell membrane, al-

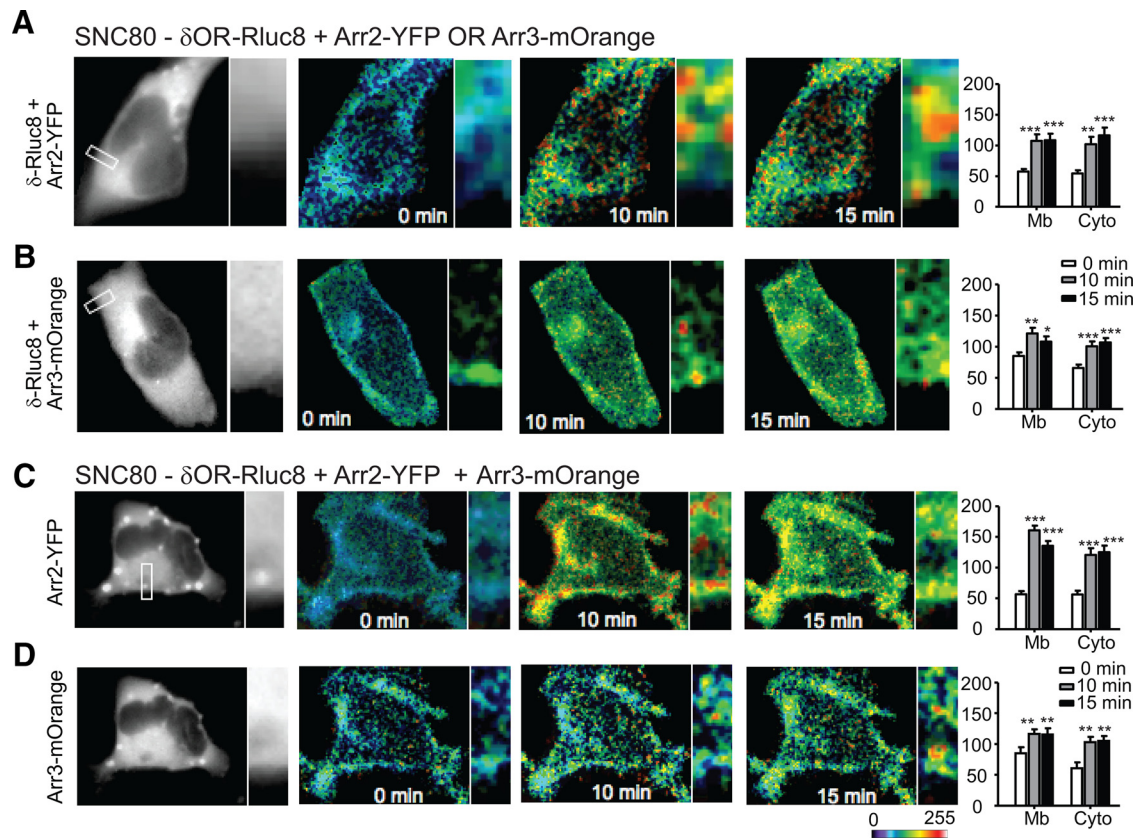


Figure 6. SNC80 causes receptor internalization with both arrestin 2 and 3. HEK293 cells transfected with δ OR-Rluc8 and Arr2-YFP (**A**), δ OR-Rluc8 and Arr3-mOrange (**B**), or δ OR-Rluc8, Arr2-YFP, and Arr3-mOrange (**C, D**). For the three partner-BRET experiment, **C** shows representative images from the Arr2-YFP channel and **D** those from the Arr3-mOrange channel. Cells were incubated with SNC80 (100 nM) and mean BRET intensity was quantified for the cell membrane (Mb) and the cytoplasm (Cyto) at time 0, 10, and 15 min after SNC80 administration. * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$, one-way ANOVA with Holm–Sidak *post hoc* analysis ($n = 3–6$ cells and 7 square regions per subcellular compartment). The high-internalizing agonist SNC80 induced robust internalization with either arrestin 2 or 3.

though the cytosolic interaction observed in the absence of arrestin 3 was attenuated (Fig. 5*B* vs *D*, Cyto). ARM390 continued to increase the interaction between δ OR-Rluc8 and Arr3-mOrange only at the cell membrane (Fig. 5*E*, Mb). Together, these data indicate that there is a preexisting basal interaction between the delta opioid receptor and arrestin 3 at the cell membrane, which is strengthened by the binding of ARM390. This enhanced membrane interaction appears to protect the delta opioid receptor from intracellular interactions with arrestin 2.

Finally, as a control, we analyzed BRET interactions among arrestin 2, arrestin 3, and delta opioid receptors after SNC80 administration (Fig. 6). The addition of SNC80 (0.1 μ M) significantly increased membrane and cytosolic interactions between δ OR-Rluc8 and both Arr2-YFP (Fig. 6*A*) and Arr3-mOrange (Fig. 6*B*). This was also true when all three proteins were coexpressed (Fig. 6*C, D*), results that are consistent with SNC80 being a high-internalizing agonist.

Discussion

In this study, we found that delta opioid receptor agonists preferentially recruit specific arrestins in a ligand-selective manner (Fig. 7). The high-internalizing agonist SNC80 preferentially recruited arrestin 2 and KO of arrestin 2 resulted in increased potency and duration of action and decreased acute tolerance to this agonist. In contrast, the low-internalizing agonists preferentially engaged arrestin 3. Surprisingly, KO of arrestin 3 resulted in gain of acute tolerance to the low-internalizing agonists ARM390 and JNJ20788560, an effect not observed in WT mice. Electrophysiolog-

ical recordings from adult DRGs showed that, in the absence of arrestin 3, desensitization rates were unaltered, but resensitization, the recovery from the desensitized state, was markedly delayed. BRET analysis revealed that, in contrast to arrestin 2, delta opioid receptors were pre-engaged with arrestin 3 at the cell membrane and the low-internalizing agonist ARM390 strengthened this association. Overall, these results show a ligand-selective recruitment of arrestins by different delta opioid receptor agonists and reveal a novel endogenous role of arrestin 3 as an inhibitor of acute tolerance and facilitator of receptor resensitization.

We found that the anti-allodynic effects of the high-internalizing agonist SNC80 were modulated by arrestin 2 and not arrestin 3. Our BRET data, along with another BRET-based study (Audet et al., 2012), demonstrate that SNC80 activation recruits both arrestin 2 and 3 isoforms. However, our *in vivo* results show that, in a chronic pain model, there is a predominance of arrestin 2-regulated behavioral effects by SNC80; and KO of arrestin 2 results in increased drug potency, duration of action, and decreased acute tolerance. In contrast, no change in the anti-hyperalgesic effect of SNC80 was observed in arrestin 3 KO. A previous study also showed that KO of arrestin 2 resulted in increased SNC80-induced hyperlocomotion and cellular functionality (Mittal et al., 2013). In that study, arrestin 2 regulated delta opioid receptor trafficking to the cell membrane and KO of arrestin 2 resulted in enhanced externalization of receptors in response to SNC80 via a deregulated ROCK/LIMK pathway. The enhancement of agonist-induced externalization, and potentially

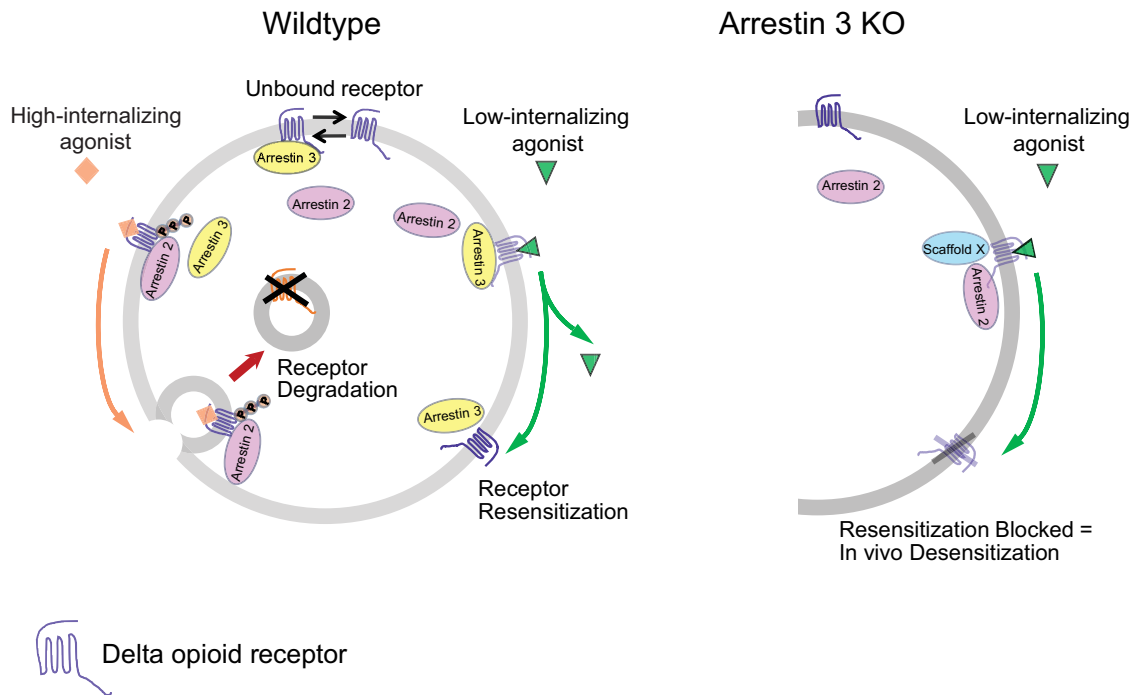


Figure 7. Arrestins are recruited in a ligand-specific manner to the delta opioid receptor. Under basal conditions (no agonist bound), delta opioid receptors are in a pre-engaged complex with arrestin 3 at the cell membrane. High-internalizing agonist such as SNC80 induce substantial receptor phosphorylation, preferentially recruit arrestin 2, and result in receptor internalization and degradation. In contrast, low-internalizing agonists such as ARM390 and JNJ20788560 induce a receptor conformation that strengthens the engagement between delta opioid receptor and arrestin 3 at the cell membrane and encourages receptor resensitization, thus preventing acute behavioral tolerance. In the absence of arrestin 3, low-internalizing agonists undergo desensitization similarly to WT animals, but we hypothesize that a different receptor complex is formed, perhaps with arrestin 2 or another GPCR modulator, which hinders resensitization. The mechanism underlying arrestin 3-mediated receptor resensitization remains to be elucidated.

altered desensitization/resensitization and trafficking, explain the increased efficacy and lack of tolerance to SNC80 in arrestin 2 KO.

We were surprised to find that the low-internalizing agonists ARM390 and JNJ20788560 recruited arrestins because neither compound induces robust delta opioid receptor internalization (Pradhan et al., 2009). Furthermore, arrestin 3 KO resulted in a gain of acute tolerance to both low-internalizing agonists, an effect not observed in WT mice (Pradhan et al., 2009; Pradhan et al., 2010). These results are in direct contrast to the findings for mu opioid receptors, in which arrestin 3 KO exhibits decreased mu opioid receptor desensitization (Mittal et al., 2013) and decreased anti-nociceptive tolerance (Bohn et al., 2000; Bohn et al., 2002) that have been attributed to facilitation of mu receptor resensitization (Dang and Christie, 2012; Williams et al., 2013; Connor et al., 2015). Together, these data suggest that endogenously arrestin 3 attenuates resensitization to mu, yet enhances delta opioid receptor resensitization. We speculate that this converse role of arrestin 3 is orchestrated via the highly divergent C-terminal domains of mu and delta, which are known to mediate interactions with many regulatory proteins, including arrestins.

Our BRET analysis provides insight on how low-internalizing agonists might result in acute tolerance in arrestin 3 KO. Live-cell imaging revealed that, in a naive state, there was a basal engagement between the delta opioid receptor and arrestin 3 at the cell membrane, an interaction not observed with arrestin 2. This arrestin 3 interaction was strengthened with ARM390 and this enhanced membrane interaction could promote recovery of the delta opioid receptor after binding to low-internalizing agonists. One other recent study also showed that ARM390 could recruit

arrestin 3 in a cell-based recruitment assay (Chiang et al., 2015). To better reflect our *in vivo* acute tolerance experiments, electrophysiological experiments on adult DRGs were performed using longer time points. These studies revealed that, although delta opioid receptors desensitized equally to ARM390 in both arrestin 3 WT and KO DRGs, subsequent resensitization was impaired in arrestin 3 KO. These results indicate that the presence of arrestin 3 is required to facilitate the return of delta opioid receptor functionality after acute exposure to a low-internalizing agonist.

One interpretation of our data is that, in the absence of arrestin 3, the delta opioid receptor is more susceptible to long-term desensitization by other receptor modulators, perhaps including arrestin 2. Results from our BRET experiments indicate that, in the absence of arrestin 3, ARM390 increases the intracellular complexes between the delta receptor and arrestin 2. Interestingly, in coexpression studies, the binding of ARM390 also recruited arrestin 2 to the membrane, but competition with arrestin 3 prevented further interaction. *In vivo*, we did not observe significant internalization of DOR-eGFP in arrestin 3 KO mice, but this does not exclude the possibility that arrestin 2 mediates a long-duration desensitization at or close to the cell membrane. For example, other modulators of desensitization such as GRKs or calmodulin could have an enhanced role in regulation of ARM390-bound delta opioid receptors in arrestin 3 KO.

An alternative explanation is that arrestin 3 facilitates recycling after delta opioid receptor binding to low-internalizing agonists. Although ARM390 has not been shown to produce profound receptor internalization, even in time-lapse cellular experiments (Pradhan et al., 2009), there is the possibility that low-internalizing agonists produce a rapid receptor recycling close to the membrane (Audet et al., 2012; Ong et al., 2015). In this cir-

cumstance, arrestin 3 could enable a quick replacement of desensitized receptors at the membrane with fully active recycled delta opioid receptors. *In vivo*, we did not observe significant receptor internalization to ARM390 in arrestin 3 KO-DOR-eGFP mice, but recycling close to the membrane would not be detected at this resolution.

The ligand-specific recruitment of arrestins observed in our studies is likely due to distinct conformational changes induced by high- and low-internalizing delta agonists. Ligand-initiated phosphorylation of the delta opioid receptor is an important step toward receptor desensitization and/or internalization. Phosphorylation of GPCRs at the C terminus can provide a substrate for high-affinity binding to arrestins. We have shown previously in brain samples that SNC80, but not ARM390, can produce phosphorylation at the serine 363 residue of the delta opioid receptor (Pradhan et al., 2009). Phosphorylation of this residue is a principal event for receptor internalization (Kouhen et al., 2000) and predicts internalization across numerous delta ligands (Bradbury et al., 2009). In addition, in coimmunoprecipitation studies of transfected HEK293 cells, arrestin 2 appears to have a higher affinity for phosphorylated delta opioid receptor compared with arrestin 3 (Zhang et al., 2008; but see Qiu et al., 2007). SNC80 produces receptor phosphorylation, which recruits arrestin 2 and facilitates receptor internalization, corresponding to the traditional role of arrestins as attenuators of GPCR signaling.

Although ARM390 does not phosphorylate the delta opioid receptor at the critical Ser363, it may phosphorylate the receptor at other residues, resulting in increased affinity between the receptor and arrestin 3 at the cell surface. Audet et al. (2012) showed that DPDPE, a high-internalizing delta agonist, induced a distinct conformational change that led to a transient interaction between the delta opioid receptor and arrestin 3 that encouraged rapid receptor recycling. High- and low-internalizing mu opioid receptor agonists have also been shown to encode distinct phosphorylation patterns in which internalization (but not interaction) with arrestins is only initiated after phosphorylation of critical residues (Lau et al., 2011; Doll et al., 2012; Just et al., 2013). Furthermore, phosphorylation-deficient mutants of the β_2 adrenergic receptor and angiotensin II receptor have also been shown to interact with arrestins in a transient manner (Shukla et al., 2008), also indicating that changes in receptor conformation may be sufficient to increase receptor–arrestin interactions in the absence of internalization. Our data indicate that the delta opioid receptor can adopt different conformational states that affect receptor–arrestin interactions.

Ligand-biased recruitment of different arrestins has been observed for other GPCRs *in vitro*. For example, the low-internalizing mu opioid receptor agonist morphine primarily recruits arrestin 3, whereas the high-internalizing agonist DAMGO can recruit either arrestin 2 or 3 (Groer et al., 2011). In addition, endogenous ligands for the C-C chemokine receptor 7 also show differential internalizing properties; and the high-internalizing ligand CCL19 preferentially recruits arrestin 3 over 2, whereas the low-internalizing ligand CCL21 engages neither (Byers et al., 2008). In addition, binding of ATP induces a greater interaction between P2Y2 receptor and arrestin 2, contrary to UTP, which nonselectively recruits both arrestins (Hoffmann et al., 2008). These studies support the notion that agonists for the same receptor can promote distinct receptor–arrestin complexes.

Our study reveals agonist-specific recruitment of arrestins at the delta opioid receptor. There is a clear distinction behaviorally for arrestin isoforms between the high-internalizing agonist SNC80 and

the low-internalizing agonists ARM390 and JNJ20788560. Our results show the behavioral implications of ligand-selective arrestin interactions at the delta opioid receptor and reveal a novel role for arrestin 3 as a facilitator of resensitization and inhibitor of tolerance mechanisms.

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