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UNIVERSITY OF CALIFORNIA,  
IRVINE

Dual Catalytic Systems with Gold(I)

THESIS

submitted in partial satisfaction of the requirements  
for the degree of

MASTER OF SCIENCE

in Chemistry

by

Joel Johnson

Thesis Committee:  
Professor Chris D. Vanderwal, Chair  
Professor Suzanne A. Blum  
Professor Vy M. Dong

2015



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Prof. Martin Burke, my organic chemistry II professor. Learning to think by analogy and think in terms of molecular orbital interactions have helped me rationalize chemistry I have never seen.

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Prof. Suzanne Blum, my graduate research advisor. Thank you for giving me the opportunity to do the chemistry I was interested in doing.

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**Abstract Of Thesis**  
Dual Catalytic Systems with Gold(I)

By

Joel Johnson

Master of Science in Chemistry

University of California, Irvine, 2015

Professor Chris D. Vanderwal, Chair

**Chapter 1.** Described herein is a study on the functional group tolerance of the gold/palladium dual catalyzed synthesis of 4-allylisocoumarins. A number of motifs commonly found in compounds with biological activity and intermediates towards such compounds were prepared and tested. The effect of silver on the product distribution was determined.

**Chapter 2.** A kinetic study of the alkoxyboration reaction showed the reaction is second order overall: first order with respect to both IPrAuTFA and NaTFA. NaTFA was determined to be both a Lewis base, by enhancing the nucleophilicity the B–O  $\sigma$  bond, and as a reagent needed to decrease the amount of IPrAuTFA converted to the catalytically inactive IPrAuCl. Increasing the amount of trifluoroacetate eventually lead to a decrease in the reaction rate. This effect is attributed to over addition of the Lewis base into the borate ester that is formed after the cyclization step. The rate-determining step was determined to be the transmetalation step.

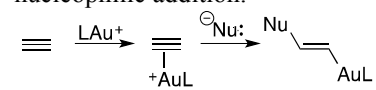
## Introduction

This introduction will present to those without an in depth knowledge of the chemical theory the information needed to understand the subsequent chapters. The subsequent chapters discuss a specific type of catalysis called *dual catalysis*, but first, a discussion of catalysis should occur. Catalysis is the field of study revolving around the development of reactions that use a reagent called a catalyst to increase the rate of the reaction. Catalysts work by either providing an alternative, lower energy pathway for a reaction to traverse or by lowering the activation energy of the rate-determining step of the reaction. The activation energy is inversely proportional to the rate of the reaction. Reactions with lower activation energies have faster rates. As the name suggests, dual catalysis involves a chemical transformation that is catalyzed by two catalysts. The advantage of a dual catalytic system is that otherwise impossible transformations and pathways become viable using this type of catalysis.

The dual catalytic system that is the main focus of research in the Blum group is one where gold(I) is one component of the dual catalytic system. Gold complexes are known to be carbophilic  $\pi$  acids, meaning

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Figure 0.1 Gold(I) catalyzed nucleophilic addition.



electron density can be withdrawn from a  $\pi$  system, enabling them to accept electron density from a nucleophile. The gold catalyst can be regenerated in a number of ways, including transmetalation with another metal or acting as a nucleophile towards an electrophile. We seek to discover and develop these types of reactions.

In addition to discovering new reactions, understanding the mechanistic underpinnings of how these reactions work is equally important. This is done by first forming a mechanistic hypothesis based on literature precedent and a thorough understanding of the principles that govern chemical reactions pathways. Determining both what reagents affect the rate and reaction intermediates are two ways in which one can aid in the determination of the true mechanism.

## Chapter 1

### Functional Group Tolerance and Silver Effect of Gold/Palladium Catalyzed Synthesis of 4-allylisocoumarins

**Abstract:** Described herein is a study on the functional group tolerance of the gold/palladium dual catalyzed synthesis of 4-allylisocoumarins. A number of motifs commonly found in compounds with biological activity and intermediates towards such compounds were prepared and tested. The effect of silver on the product distribution was determined.

#### INTRODUCTION

Dual metal catalyzed reactions can facilitate transformations that could not be reproduced otherwise.<sup>1</sup> The advantages of dual metal catalytic systems are usually counterbalanced by undesired redox chemistry, ligand exchange, and functional group incompatibility.<sup>2</sup> In 2009, Blum and coworkers reported the gold/palladium dual

catalyzed cyclization of allyl allenates and benzoates to form 4-allyl butenolides and isocoumarins respectively (Figure 1.1).<sup>3</sup> Figure 1.2 shows a plausible mechanism for the gold/palladium dual catalyzed cyclo-isomerization reaction.

Many of the intermediates in the putative mechanism can traverse alternative pathways, leading to the formation of side products. The organogold intermediates **B** and **C** can act as nucleophiles as opposed to participating in transmetalation with the palladium  $\pi$  allyl intermediate. This is the manner in which gold catalysts are typically regenerated, where the electrophile is a proton.<sup>4</sup> Intermediate **B** also contains an oxocarbenium fragment capable of acting as an electrophilic allylating agent as opposed to acting as the species capable of being oxidatively added into

by palladium(0). Palladium  $\pi$ -allyl intermediates have also been reported to act as electrophiles, such as in the

Figure 1.1 Work from Blum et al.

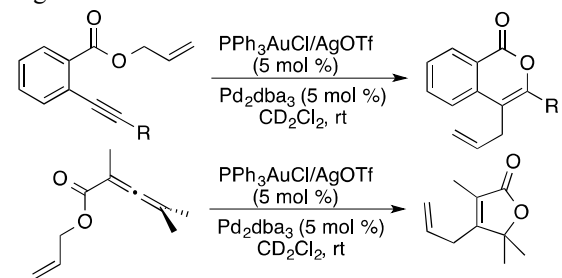
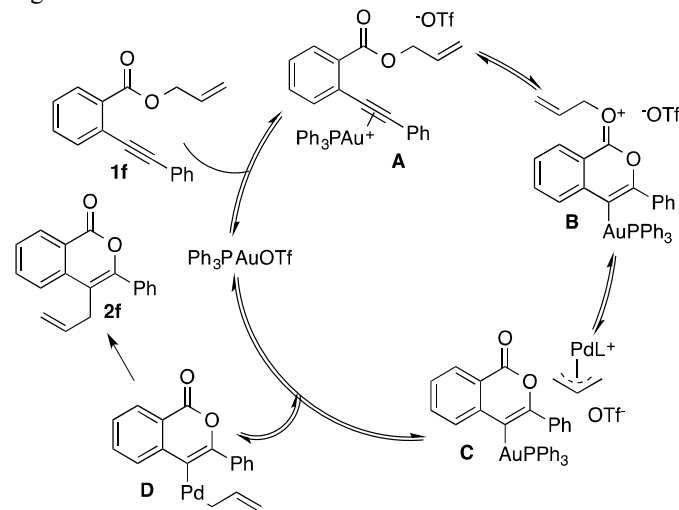


Figure 1.2 Plausible mechanism of the reaction.



Tsuji-Trost reaction.<sup>5</sup>

In the seminal publication, no appreciable functional group tolerance study was conducted, however, it was reported that aryl bromides were tolerated by the reaction conditions. The following report shows a subset of the functional groups tolerated, a downstream functionalization reaction, along with the effect silver(I) has on the product distribution.

## RESULTS AND DISCUSSION

Table 1.1 Gold and palladium dual-catalyzed synthesis of lactones 2a-2e<sup>a</sup>.

Entry	Starting Material	Product	Isolated Yield (%)	2:3 <sup>c</sup>
1 <sup>b</sup>			24	11:1
2 <sup>c</sup>			33	10:1
3 <sup>d</sup>			83	Only 2
4			71	12:1
5 <sup>b</sup>			50	16:1

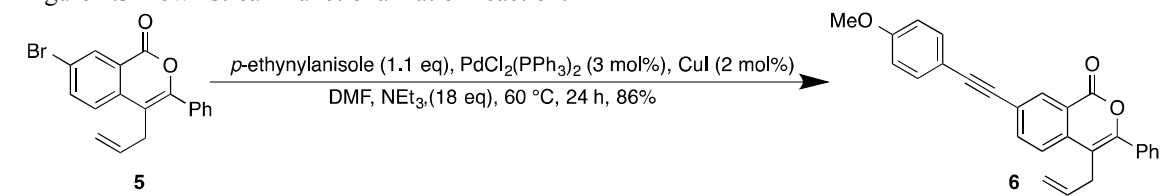
<sup>a</sup>Unless otherwise noted, reaction conditions: substrate **1** (1.0 equiv), PPh<sub>3</sub>AuCl (5 mol%), AgOTf (5 mol%), and Pd<sub>2</sub>dba<sub>3</sub> (5 mol%) in dry CD<sub>2</sub>Cl<sub>2</sub> (0.1 M substrate) were stirred at ambient temperature.

<sup>b</sup>Reaction was carried out at 60 °C in DCE-d<sub>4</sub>. <sup>c</sup>Reaction was carried out at 40 °C in CD<sub>2</sub>Cl<sub>2</sub>. <sup>d</sup>Reaction was carried out at 50 °C in DCE-d<sub>4</sub>. <sup>e</sup>The ratio was determined via analysis of the crude <sup>1</sup>H NMR spectrum.

The results from the functional group tolerance study are shown in Table 1.1. As stated in the previous section, each substrate has a functional group capable of undergoing an undesired chemical reaction with one of the catalysts or putative intermediates in the catalytic cycle (Figure 1.2). Substrates **1a**, **1c**, and **1e** all contain Lewis basic atoms capable of ligating to the cationic gold catalyst, which could lead to rate retardation or complete inhibition of product formation under nominal conditions. Notably, in order for the reaction to proceed at a viable rate using these substrates, higher temperatures were necessary. Also note that substrate **1a** can act as a proton source, which could lead to the protodeauration of intermediates **B** or **C**. The protodeaurated product was isolated in 10% yield. No allyl ethers were detected, suggesting that the phenol may not have acted as a nucleophile towards one of the electrophilic allyl species. The reaction with **1d**, which contains a propargylic cyclopropyl group capable of rearrangement under acidic conditions, proceeded at ambient temperature, affording the product in 71% yield.<sup>6</sup> Substrate **1b** contains an electrophilic cross coupling partner capable of undergoing oxidative addition with palladium(0). The reaction temperature using this substrate needed to be increased to 40 °C. The temperature increase may have been necessitated by the reversible oxidative addition removing the active palladium catalyst from the reaction mixture or due to the formation of palladium black. One potential selling point of this substrate is that it is an electrophilic cross coupling partner, which creates the opportunity for downstream functionalization.

As proof of concept, downstream functionalization reactions were conducted. Starting with the compound **5**, a Sonogashira Coupling with *p*-ethynylanisole produced the product, **6**, in 86% yield (Figure 1.3).

Figure 1.3 Down stream functionalization reaction.

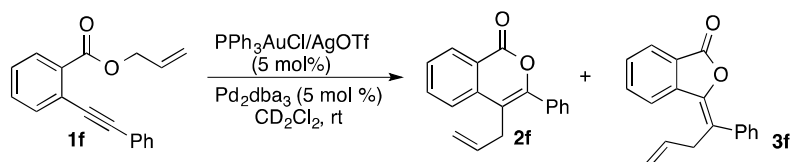


A paper published by Shi and coworkers detailed the role silver(I) can play in gold catalysis.<sup>7</sup> The effect of silver on the reaction was determined (Table 1.2). It seems the product distribution is dependent on the silver content in the reaction medium.

In summary, a number of motifs are tolerated by the reaction conditions, including: Lewis basic atoms, electrophilic cross coupling partners, and motifs known to rearrange under acidic conditions, albeit, a small



Table 1.2 Effect of silver on the reaction.



Entry	Conditions <sup>a</sup>	<sup>1</sup> H NMR yield (%) <sup>b</sup>	Product ratio 2:3
1	Ag(I) filtered <sup>c</sup>	96	8:1–16:1
2	Ag(I) unfiltered	98	8:1→20:1
3	AgOTf only <sup>d</sup>	20	>20:1

<sup>a</sup>PPh<sub>3</sub>AuCl (5 mol %), AgOTf (5 mol %), and Pd<sub>2</sub>dba<sub>3</sub> (5 mol %) in CD<sub>2</sub>Cl<sub>2</sub> at rt. <sup>b</sup><sup>1</sup>H NMR spectroscopy yield was determined after 24 h based on the ratio of product to mesitylene (the internal standard). <sup>c</sup>AgCl and AgOTf were removed via filtration through Celite or glass fiber paper. <sup>d</sup>In the absence of PPh<sub>3</sub>AuCl.

modification in temperature was needed in order for the reaction to proceed at a viable rate in a number of cases.

The tolerance of some functional groups can allow for down stream functionalization. Silver(I) also seems to have an effect on the product distribution of the reaction.

## REFERENCES

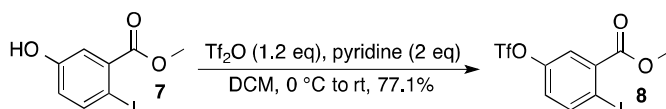
- 1.) Hansmann, M.M.; Hashmi, A.S.K.; Lautens, M. *Org. Lett.* **2013**, *15*, 3226 – 3229.
- 2.) Weber, D.; Gagné, M. *Chem. Commun.* **2011**, *47*, 5172 – 5174.
- 3.) Shi, Y.; Roth, K. E.; Ramgren, S. D.; Blum, S. A. *J. Am. Chem. Soc.* **2009**, *131*, 18022 – 18023.
- 4.) Hashmi, A.S.K. *Chem. Rev.* **2007**, *107*, 3180 – 3211.
- 5.) I. Ibrahim; A. Córdova *Angew. Chem. Int. Ed.* **2006**, *45*, 1952 – 1956.
- 6.) Olah, G. A.; Reddy, V. P.; Prakash, G. K. S. *Chem. Rev.* **1992**, *92*, 69 – 95.
- 7.) Wang, D.; Cai, R.; Sharma, S.; Jirak, J.; Thummanapeli, S. K.; Akhmedov, N. G.; Zhang, H.; Liu, X.; Petersen, J. L.; Shi, X. *J. Am. Chem. Soc.* **2012**, *134*, 9012 – 9019.

## EXPERIMENTAL SECTION

General Procedure for transesterification:

To a flame dried round bottom flask equipped with a stir bar and activated four angstrom molecular sieves, allyl alcohol (10 mmol) was dissolved in dry THF (10 mL). The reaction flask was cooled to –78 °C. *n*-butyllithium (1.6 M, 5 mmol) was added, the mixture was allowed to warm to room temperature. The methyl ester (1 mmol) was then transferred to the reaction flask with minimal amounts of THF. The reaction was allowed to stir over night at room temperature under nitrogen. The reaction was added to water (10 mL) and extracted with dichloromethane (10 mL × 3). The combined organic layers were dried with MgSO<sub>4</sub> and concentrated. The compound was purified via chromatography (20% EtOAc in hexanes), unless stated otherwise. Like fractions were combined, concentrated, and dried under high vacuum.

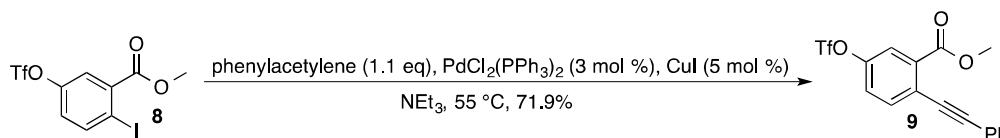
Synthesis of **8**:



Made according to literature precedent.<sup>1</sup> To a flame dried round bottom flask equipped with a stir bar, **7** (0.9653 g, 3.472 mmol, 1.000 equiv) and pyridine (0.56 mL, 6.9 mmol, 2.0 equiv) was dissolved in dry DCM (12 mL) under

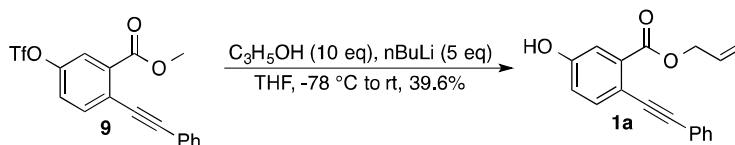
nitrogen. Triflic anhydride (1.0 mL, 4.2 mmol, 1.2 equiv) was added at 0 °C. The mixture was allowed to warm to room temperature and stir for six hours. The reaction mixture was quantitatively transferred to a separatory funnel and washed with saturated NH<sub>4</sub>Cl (aq.). The organic layer was dried with MgSO<sub>4</sub>, which was removed via gravity filtration. The solution was concentrated. The compound was purified by column chromatography (0 to 20% EtOAc in hexanes). The product, **8** (1.0974 g, 77.1%), was isolated in analytically pure form. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 8.09 (d, J = 8.7, 1H), 7.74 (d, J = 3.0 Hz, 1H), 7.10 (dd, J = 8.7 Hz, 3.0 Hz, 1H), 3.97 (s, 3H); <sup>13</sup>C (CDCl<sub>3</sub>, 125 MHz) δ 165.00, 149.17, 143.34, 136.84, 125.55, 124.13, 118.69 (q, J = 319.1 Hz, 1C), 93.55, 53.11; <sup>19</sup>F (CDCl<sub>3</sub>, 376 MHz) δ -72.6; HRMS *m/z* calcd for C<sub>9</sub>H<sub>10</sub>O<sub>5</sub>F<sub>3</sub>ISN (M + NH<sub>4</sub>)<sup>+</sup> 427.9276, found 427.9269.

#### Synthesis of **9**:



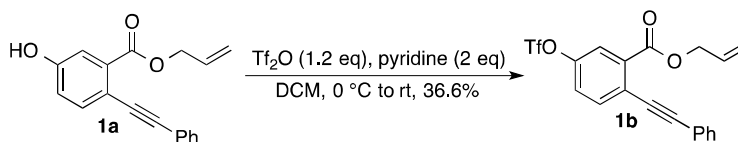
To a 25 mL round bottom flask equipped with a stir bar, **8** (1.0974 g, 2.6759 mmol, 1.0000 equiv), phenylacetylene (0.33 mL, 2.9 mmol, 1.1 equiv), PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub> (56.3 mg, 80.3 μmol, 0.03 equiv), CuI (25.4 mg, 80.3 μmol, 0.05 equiv) was dissolved in degassed triethylamine (8.9 mL). The mixture was allowed to stir under nitrogen overnight at 55 °C. The reaction mixture was concentrated under nitrogen and purified by column chromatography (0 to 20% EtOAc in hexanes). The product, **9** (0.7388 g, 71.9%), was isolated. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.90 (d, J = 2.6 Hz, 1H), 7.73 (d, J = 8.6 Hz, 1H), 7.58 (m, 2H), 7.42 (dd, J = 8.6 Hz, 2.7 Hz, 1H), 7.38 (m, 3H), 3.99 (s, 3H); <sup>13</sup>C NMR (CD<sub>2</sub>Cl<sub>2</sub>, 125 MHz) δ 165.32, 148.85, 136.47, 134.62, 132.40, 129.77, 129.19, 125.34, 124.86, 124.12, 123.28, 120.64, 119.27 (t, J = 318.4 Hz), 96.86, 87.14; <sup>19</sup>F NMR (CDCl<sub>3</sub>, 376 MHz) δ -72.7; HRMS *m/z* calcd for C<sub>17</sub>H<sub>11</sub>O<sub>5</sub>F<sub>3</sub>SNa (M + Na)<sup>+</sup> 407.0177, found 407.0172.

#### Synthesis of **1a**:



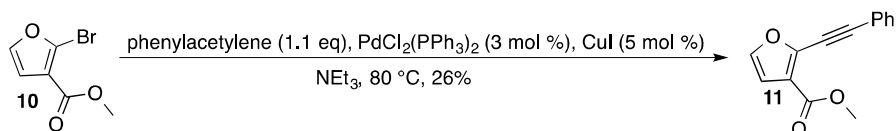
General procedure for transesterification followed. **9** (0.7388 g, 1.922 mmol, 1.000 equiv), allylic alcohol (1.31 mL, 19.2 mmol, 10.0 equiv), *n*-butyllithium (6.0 mL, 9.6 mmol, 5.0 equiv). The product, **1a** (0.2634 g, 39.6%), was isolated in analytically pure form. <sup>1</sup>H (CDCl<sub>3</sub>, 600 MHz) δ 7.59 (m, 3H), 7.52 (d, J = 2.7 Hz, 1H), 7.39 (m, 3H), 7.05 (dd, J = 8.5 Hz, 2.7 Hz, 1H), 6.10 (ddt, J = 17.2 Hz, 10.4 Hz, 5.6 Hz, 1H), 5.49 (app-dq, J = 17.2, 1.4 Hz, 1H), 5.39 (s, 1H), 5.3230 (app-dq, J = 10.4 Hz, 1.4 Hz, 1H), 4.92 (app-dt, J = 5.6 Hz, 1.4 Hz, 2H); <sup>13</sup>C (CDCl<sub>3</sub>, 125 MHz) δ 167.4, 155.8, 135.8, 132.9, 131.6, 128.4, 128.3, 123.6, 119.7, 117.5, 115.8, 92.7, 88.1, 52.6; HRMS *m/z* calcd for C<sub>18</sub>H<sub>14</sub>O<sub>3</sub>Na (M + Na)<sup>+</sup> 301.0841, found 301.0846.

#### Synthesis of **1b**:



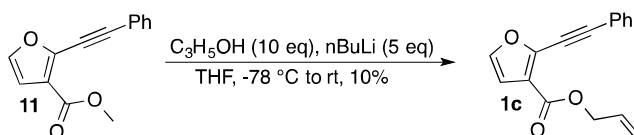
Same procedure used to make **8**. **1a** (124.1 mg, 0.4459 mmol, 1.000 equiv), pyridine (70. μL, 0.89 mmol, 2.0 equiv), triflic anhydride (0.11 mL, 0.67 mmol, 1.2 equiv), dry DCM (1.7 mL). The product, **1b** (67.0 mg, 36.6%), was isolated in analytically pure form. <sup>1</sup>H (CDCl<sub>3</sub>, 600 MHz) δ 7.91 (d, J = 2.7 Hz, 1H), 7.73 (d, J = 8.6 Hz, 1H), 7.57 (m, 2H), 7.42 (dd, J = 8.6 Hz, 2.7 Hz, 1H), 7.38 (m, 3H), 6.05 (ddt, J = 17.1 Hz, 10.4 Hz, 5.8 Hz, 1H), 5.45 (app-dd, J = 17.1 Hz, 1.4 Hz, 1H), 5.30 (app-dd, J = 10.4 Hz, 1.4 Hz, 1H), 4.89 (dt, J = 5.8 Hz, 1.4 Hz, 2H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 125 MHz) δ 164.2, 148.3, 136.1, 134.0, 132.0, 131.8, 129.3, 128.6, 124.9, 124.6, 123.8, 122.8, 119.3, 118.9 (t, J = 319.0 Hz), 96.8, 86.7, 66.7; <sup>19</sup>F NMR (CDCl<sub>3</sub>, 376 MHz) δ -72.7; HRMS *m/z* calcd for C<sub>19</sub>H<sub>13</sub>F<sub>3</sub>O<sub>5</sub>SNa (M + Na)<sup>+</sup> 433.0334, found 433.0326.

### Synthesis of **11**:



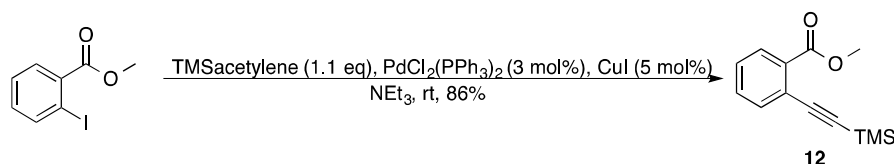
To a 5 mL one neck round bottom flask equipped a reflux condenser and stir bar, **10** (0.100 g, 0.487 mmol, 1.00 equiv), phenylacetylene (0.06 mL, 0.5 mmol, 1.1 equiv), PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub> (17.7 mg, 24.4 μmol, 0.050 equiv), and CuI (4.6 mg, 24 μmol, 0.050 equiv) was dissolved in NEt<sub>3</sub> (0.98 mL). The reaction was stirred under nitrogen at 80 °C. The reaction was allowed to cool to room temperature and then concentrated. The compound was isolated by column chromatography (0 to 10% EtOAc/hexanes). Compound **11** was purified via Prep TLC (5% EtOAc in hexanes). The product, **11** (29.0 mg, 26%), was isolated in analytically pure form. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.60 (m, 2H), 7.39 (m, 4H), 6.80 (d, J = 1.9 Hz, 1H) 3.91 (s, 3H); <sup>13</sup>C (CDCl<sub>3</sub>, 125 MHz) δ 163.1, 143.5, 140.8, 132.0, 129.6, 128.7, 121.9, 121.3, 111.7, 98.1, 78.8, 52.0; HRMS *m/z* calcd for C<sub>15</sub>H<sub>14</sub>O<sub>2</sub>Na (M + Na)<sup>+</sup> 249.0892, found 249.0894.

### Synthesis of **1c**:



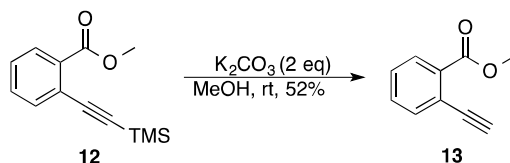
General procedure for transesterification followed. **11** (29.0 mg, 0.128 mmol, 1.00 equiv), allyl alcohol (90. μL, 1.3 mmol, 10. equiv), *n*-BuLi (1.6 M, 0.40 mL, 0.64 mmol, 5.0 equiv). The compound was purified via Prep TLC (2% EtOAc in hexanes). The product, **1c** (3.1 mg, 10 %) was isolated in analytically pure form. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.59 (m, 2H), 7.38 (m, 4H), 6.82 (d, J = 1.9 Hz, 1H) 6.03 (ddt, J = 17.2 Hz, 10.5 Hz, 5.6 Hz, 1H), 5.46 (app-dd, J = 17.2, 1.4 Hz, 1H), 5.28 (app-dd, J = 10.5 Hz, 1.4 Hz, 1H), 4.82 (app-d, J = 5.6 Hz, 2H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 125 MHz) δ 162.2, 143.4, 140.6, 132.1, 131.9, 129.4, 128.5, 121.7, 118.3, 111.6, 98.0, 78.8, 77.3, 65.4; HRMS *m/z* calcd for C<sub>16</sub>H<sub>12</sub>O<sub>3</sub>Na (M + Na)<sup>+</sup> 275.0684, found 275.0693.

### Synthesis of **12**:



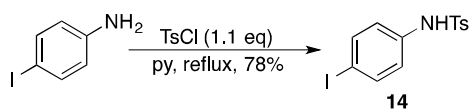
To a round bottom flask, methyl 2-iodobenzoate (0.79 mL, 5.4 mmol, 1.0 eq), PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub> (113 mg, 0.161 mmol, 0.030 equiv), and CuI (30.7 mg, 0.161 mmol, 0.050 equiv) were added. The flask was capped and purged with nitrogen. Degassed triethylamine (5.4 mL) was added to dissolve the solids. Trimethylsilylacetylene (0.83 mL, 5.9 mmol, 1.1 equiv) was added slowly. The reaction mixture was allowed to stir at room temperature. The reaction was monitored by TLC until completion. The solution was concentrated under N<sub>2</sub>. The compound was purified and isolated via column chromatography (0 to 20% over 10 column volumes). The product **12** (1.0790 g, 86.32%) was obtained in analytically pure form. <sup>1</sup>H NMR (CD<sub>2</sub>Cl<sub>2</sub>, 500 MHz) δ 7.87 (ddd, J = 7.9 Hz, 1.4 Hz, 0.5 Hz, 1H), 7.57 (ddd, J = 7.7 Hz, 1.3 Hz, 0.5 Hz) 7.47 (td, J = 7.5 Hz, 1.4 Hz) 7.39 (td, J = 7.8 Hz, 1.4 Hz, 1H), 3.89 (s, 3H), 0.26 (s, 9H); <sup>13</sup>C NMR (CD<sub>2</sub>Cl<sub>2</sub>, 125 MHz) δ 167.0353, 134.7105, 133.1035, 131.8509, 130.4719, 128.6152, 123.4034, 103.5347, 99.8476, 52.2508, -0.1093; HRMS *m/z* calcd for C<sub>13</sub>H<sub>16</sub>O<sub>2</sub>SiNa (M + Na)<sup>+</sup> 255.0817, found 255.0821. In agreement with literature precedent.<sup>2</sup>

### Synthesis of **13**:



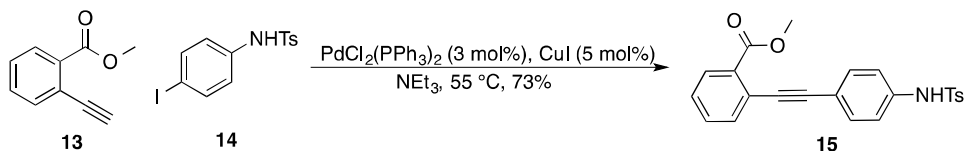
Prepare according to literature precedent.<sup>3</sup> To a 5 mL round bottom flask **12** (1.08 g, 4.64 mmol, 1.00 equiv) and  $K_2CO_3$  (1.29 g, 9.29 mmol, 2.00 equiv) were added. MeOH (2 mL) was added. The reaction was monitored by TLC until completion. The reaction was added to water (10 mL) and extracted with DCM (10 mL  $\times$  3). The combined organic layers were dried with  $MgSO_4$  and concentrated. The compound was purified via column chromatography (0% to 20% EtOAc in hexanes). The product, **13** (390.1 mg, 52%), was isolated in analytically pure form.  $^1H$  NMR ( $CD_2Cl_2$ , 500 MHz)  $\delta$  7.8952 (dd,  $J = 7.9$  Hz, 1.6 Hz, 1H), 7.60 (dd,  $J = 7.7$  Hz, 1.1 Hz, 1H), 7.48 (app-td,  $J = 7.6$  Hz, 1.4 Hz, 1H), 7.41 (app-td,  $J = 7.7$  Hz, 1.3 Hz, 1H), 3.88 (s, 3H), 3.39 (s, 1H);  $^{13}C$  NMR ( $CD_2Cl_2$ , 125 MHz)  $\delta$  166.7, 135.3, 133.3, 132.1, 130.6, 129.0, 82.4, 82.3, 52.4.

Synthesis of **14**:



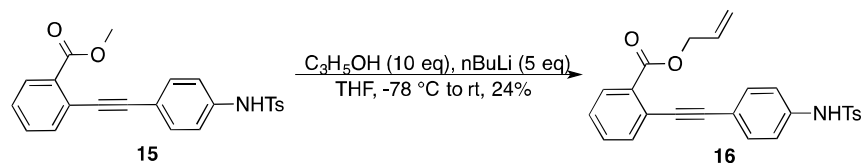
Prepared according to a literature source.<sup>4</sup> To a flame dried round bottom flask, 4-iodoaniline (200. mg, 0.914 mmol, 1.00 equiv) and tosyl chloride (191 mg, 1.00 mmol, 1.10 equiv) were added. Pyridine (1.1 mL) was added to dissolve the solids. The reaction was stirred under nitrogen at reflux overnight. Upon cooling to room temperature the reaction mixture was added to EtOAc (5 mL) and washed with saturated  $NH_4Cl$  (aq) (3  $\times$  5mL). The organic layer was dried with  $MgSO_4$ , which was removed via gravity filtration. The organic solution was concentrated. The compound was purified via column chromatography (0 to 30% EtOAc in hexanes over 15 column volumes). The product, **14** (266.5 mg, 78.27%), was isolated in analytically pure form.  $^1H$  NMR ( $CDCl_3$ , 600 MHz)  $\delta$  7.70 (d,  $J = 8.3$ , 2H), 7.59 (s, 1H), 7.49 (d,  $J = 8.8$  Hz, 2H), 7.22 (d,  $J = 8.2$  Hz, 2H), 6.87 (d,  $J = 8.8$  Hz, 2H) 2.37 (s, 2H); HRMS  $m/z$  calcd for  $C_{13}H_{16}O_2SiN_2$  ( $M + NH_4$ )<sup>+</sup> 390.9977, found 390.9974.

Synthesis of **15**:



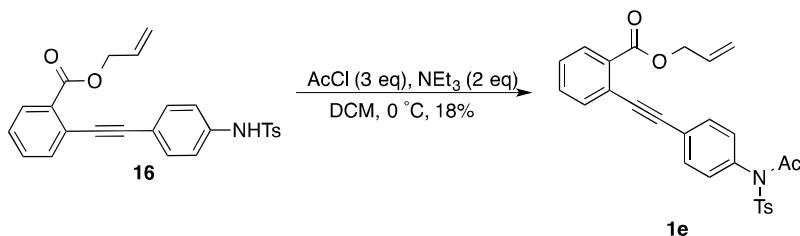
Compounds **14** (265.5 mg, 0.7117 mmol, 1.000 equiv), **13** (125.4 mg, 0.7829 mmol, 1.100 equiv),  $PdCl_2(PPh_3)_2$  (15.0 mg, 23.4  $\mu$ mol, 0.030 equiv), and  $CuI$  (6.8 mg, 36  $\mu$ mol, 0.050 equiv) were added to a 5 mL round bottom flask equipped with a stir bar. The flask was allowed to sit at room temperature while under dynamic nitrogen for 10 min. Degassed triethylamine (1.1 mL) was added. The reaction was heated to 55  $^\circ C$  and allowed to stir under nitrogen overnight. The reaction mixture was concentrated. The compound was purified and isolated via column chromatography (0 to 20% EtOAc in hexanes). The product, **15** (211.4 mg, 73.25%), was isolated in analytically pure form.  $^1H$  NMR ( $CDCl_3$ , 600 MHz)  $\delta$  7.97 (dd,  $J = 7.9$  Hz, 1.0 Hz, 1H), 7.68 (d,  $J = 8.3$  Hz, 2H), 7.5987 (dd,  $J = 7.8$  Hz, 0.9 Hz, 1H), 7.48 (app-td,  $J = 7.5$  Hz, 1.3 Hz, 1H), 7.42 (d,  $J = 8.6$  Hz, 2H), 7.37 (app-td,  $J = 7.7$  Hz, 1.3 Hz, 1H), 7.23 (d,  $J = 8.0$  Hz, 2H), 7.07 (d,  $J = 8.6$  Hz, 2H), 6.97 (s, 1H), 3.9405 (s, 3H), 2.38 (s, 3H);  $^{13}C$  NMR ( $CDCl_3$ , 125 MHz)  $\delta$  166.7, 144.2, 136.8, 135.8, 134.0, 132.9, 131.8, 131.7, 130.6, 129.8, 128.0, 127.3, 123.7, 120.8, 120.0, 93.7, 88.6, 52.3, 21.6; HRMS  $m/z$  calcd for  $C_{23}H_{18}NO_4S$  ( $M - H$ )<sup>-</sup> 404.0956, found 404.0948.

Synthesis of **16**:



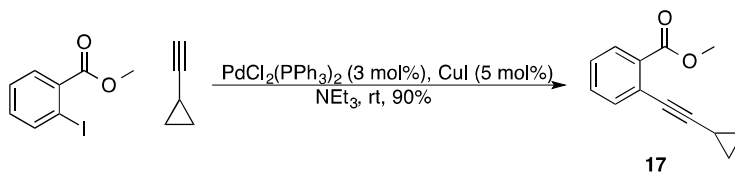
General procedure for transesterification used. **13** (211 mg, 0.520 mmol, 1.00 equiv), allyl alcohol (0.35 mL, 5.2 mmol, 10. equiv), *n*-BuLi (1.6 M, 1.60 mL, 2.60 mmol, 5.00 equiv). The product **16** (53.7 mg, 24%) was isolated in analytically pure form. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.99 (dd, *J* = 7.9 Hz, 1.0 Hz, 1H), 7.71 (d, *J* = 8.3 Hz, 2H), 7.60 (m, 2H), 7.47 (app-td, *J* = 7.6 Hz, 1.3 Hz, 1H), 7.40 (d, *J* = 8.6 Hz, 2H), 7.36 (app-td, *J* = 7.6 Hz, 1.0 Hz, 1H), 7.22 (d, *J* = 8.3 Hz, 2H), 7.10 (d, *J* = 8.6 Hz, 1H), 6.03 (ddt, *J* = 17.2 Hz, 10.4 Hz, 5.6 Hz, 1H), 5.41 (dd, *J* = 17.2 Hz, 1.4 Hz, 1H), 5.24 (dd, *J* = 10.4 Hz, 1.4 Hz, 1H), 4.85 (app-dt, *J* = 5.6 Hz, 1.3 Hz, 2H), 2.36 (s, 3H); HRMS *m/z* calcd for C<sub>25</sub>H<sub>21</sub>NO<sub>4</sub>SNa (M + Na)<sup>+</sup> 454.1089, found 454.1087.

#### Synthesis of **1e**:



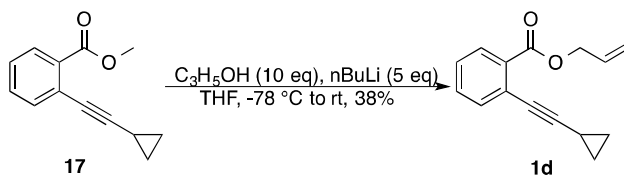
To a flame dried 5 mL round bottom flask, **16** (19.4 mg, 44.9 μmol, 1.00 equiv) and triethylamine (12.5 μL, 89.9 μmol, 2.00 equiv) was dissolved in dry dichloromethane (0.45 mL). The reaction vessel was cooled to 0 °C. Acetyl chloride (14.4 μL, 0.20 mmol, 4.5 equiv) was added via a syringe. The reaction mixture was allowed to stir under N<sub>2</sub> over night. The reaction was quenched with saturated NH<sub>4</sub>Cl (0.50 mL) and transferred to a 4 mL vial. The product was extracted with dichloromethane (3 × 5 mL). The organic layers were combined, dried with MgSO<sub>4</sub>, and concentrated in vacuo. The compound was purified via Prep TLC (25% EtOAc in hexanes). The product, **1e** (3.8 mg, 18%), was obtained in analytically pure form. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 8.04 (dd, *J* = 7.9 Hz, 1.0 Hz, 1H), 7.91 (d, *J* = 8.0 Hz, 2H), 7.67 (app-d, *J* = 8.3 Hz, 3 H), 7.53 (app-td, *J* = 7.6 Hz, 1.3 Hz, 1H), 7.43 (app-td, *J* = 7.8 Hz, 1.3 Hz, 1H), 7.35 (d, *J* = 8.1 Hz, 2H), 7.24 (d, *J* = 8.4 Hz, 2H), 6.06 (ddt, *J* = 17.1 Hz, 10.3 Hz, 5.6 Hz, 1H), 5.45 (app-dq, *J* = 17.1 Hz, 1.4 Hz, 1H), 5.29 (app-dd, *J* = 10.4 Hz, 1.3 Hz, 1H), 4.88 (app-dt, *J* = 5.6 Hz, 1.3 Hz, 2H), 2.46 (s, 3H), 1.90 (s, 3H); HRMS *m/z* calcd for C<sub>27</sub>H<sub>23</sub>NO<sub>5</sub>SNa (M + Na)<sup>+</sup> 496.1195, found 496.1197.

#### Synthesis of **17**:



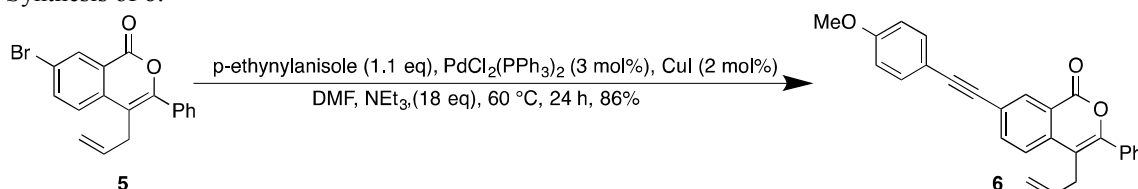
To a 5 mL round bottom flask equipped with a stir bar, methyl 2-iodobenzoate (0.37 mL, 2.5 mmol, 1.0 equiv), PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub> (17.5 mg, 25.0 μmol, 0.010 equiv), and CuI (9.5 mg, 50. μmol, 0.020 equiv) were added and allowed to sit under N<sub>2</sub> for 10 minutes. Degassed triethylamine (2.5 mL) was added to the reaction vessel. Cyclopropylacetylene (0.23 mL, 2.7 mmol, 1.1 equiv) was added slowly. The reaction mixture was allowed to stir at room temperature, under N<sub>2</sub>, over night. The reaction was concentrated under N<sub>2</sub>. The compound was purified by column chromatography (0% to 20% EtOAc in hexanes). The product, **17** (488.7 mg, 90%), was isolated in analytically pure form. <sup>1</sup>H NMR (CD<sub>2</sub>Cl<sub>2</sub>, 500 MHz) 7.83 (d, *J* = 7.9 Hz, 1H), 7.45 (d, *J* = 7.5 Hz, 1H), 7.41 (t, *J* = 7.5 Hz, 1H), 7.30 (t, *J* = 7.7 Hz, 1H), 3.86 (s, 3H), 1.48 (m, 1H), 0.90 (m, 2H), 0.81 (m, 2H); <sup>13</sup>C NMR (CD<sub>2</sub>Cl<sub>2</sub>, 125 MHz), δ 167.1, 134.3, 132.6, 131.9, 130.5, 127.5, 124.7, 89.4, 74.7, 52.3, 9.1, 0.8; HRMS *m/z* calcd for C<sub>13</sub>H<sub>12</sub>O<sub>2</sub>Na (M + Na)<sup>+</sup> 223.0735, found 223.0736.

#### Synthesis of **1d**:



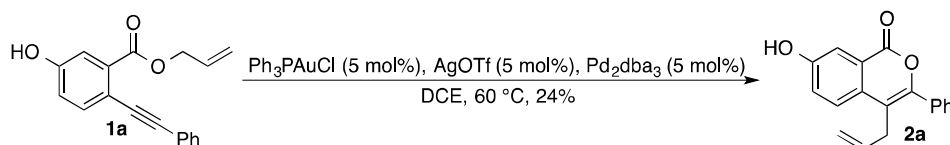
General procedure for transesterification followed. **17** (448.7 mg, 2.240 mmol, 1.00 equiv), allyl alcohol (1.52 mL, 22.4 mmol, 10. equiv), *n*-BuLi (1.6 M, 7.0 mL, 11 mmol, 5.0 equiv). The product, **1d** (193.5 mg, 38%), was isolated in analytically pure form.  $^1\text{H NMR}$  ( $\text{CD}_2\text{Cl}_2$ , 500 MHz)  $\delta$  7.87 (d,  $J = 7.9$  Hz, 1H), 7.47 (d,  $J = 7.7$  Hz, 1H), 7.43 (app-t,  $J = 7.6$  Hz, 1H), 7.32 (app-t,  $J = 7.4$  Hz, 1H), 6.06 (ddt,  $J = 17.2$  Hz, 10.4 Hz, 5.6 Hz, 1H), 5.43 (app-d,  $J = 17.2$  Hz, 1H), 5.29 (app-d,  $J = 10.4$  Hz, 1H), 4.80 (app-d,  $J = 5.6$  Hz, 2H), 1.48 (m, 1H), 0.89 (m, 2H), 0.82 (m, 2H);  $^{13}\text{C NMR}$  ( $\text{CD}_2\text{Cl}_2$ , 125 MHz)  $\delta$  166.4, 134.4, 132.8, 132.5, 131.9, 130.5, 127.5, 124.7, 118.3, 99.4, 74.8, 66.0, 9.0, 0.8; HRMS  $m/z$  calcd for  $\text{C}_{15}\text{H}_{14}\text{O}_2\text{Na}$  ( $\text{M} + \text{Na}$ ) $^+$  249.0892, found 249.0884.

#### Synthesis of **6**:



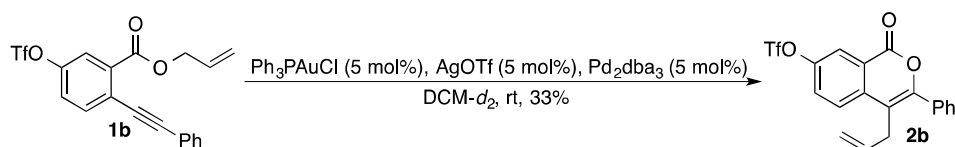
To a 5 mL round bottom flask equipped with a stir bar, **5** (45.3 mg, 0.133 mmol, 1.00 equiv),  $\text{PdCl}_2(\text{PPh}_3)_2$  (2.8 mg, 40.  $\mu\text{mol}$ , 0.030 equiv), CuI (0.5 mg, 30  $\mu\text{mol}$ , 0.020 equiv) were added. Degassed DMF (1 mL) was added via syringe. *p*-ethynylanisole (19  $\mu\text{L}$ , 0.15 mmol, 1.1 equiv) and  $\text{NEt}_3$  (0.33 mL) were added sequentially. The reaction was allowed to stir under nitrogen at 60 °C for 24 h. The reaction mixture was added to brine (5 mL) and extracted into DCM ( $3 \times 5$  mL). The combined organic layers were washed with brine, dried with  $\text{MgSO}_4$ , and concentrated. The product was crystallized with a mixture of hexanes and DCM, then isolated. The product, **6** (42 mg, 86%), was isolated and fully characterized  $^1\text{H NMR}$  ( $\text{CD}_2\text{Cl}_2$ , 600 MHz)  $\delta$  8.44 (d,  $J = 1.7$  Hz, 1H), 7.84 (dd,  $J = 8.3$  Hz, 1.8 Hz, 1H), 7.64 (m, 2H), 7.60 (d,  $J = 8.4$  Hz, 1H), 7.51 (d,  $J = 8.8$  Hz, 2H), 7.48 (m, 3H), 6.92 (d,  $J = 8.9$  Hz, 2H), 6.1238 (ddt,  $J = 17.2$  Hz, 10.4 Hz, 5.6 Hz, 1H), 5.22 (app-dq,  $J = 10.4$  Hz, 1.3 Hz, 1H), 5.08 (app-dq,  $J = 17.2$  Hz, 1.3 Hz, 1H), 3.83 (s, 3H), 3.46 (app-dt,  $J = 5.6$  Hz, 1.3 Hz, 2H);  $^{13}\text{C NMR}$  ( $\text{CD}_2\text{Cl}_2$ , 125 MHz)  $\delta$  161.3, 160.1, 152.9, 137.1, 136.8, 135.6, 133.1, 133.0, 132.0, 129.7, 128.6, 128.3, 124.6, 123.4, 121.2, 116.9, 114.6, 114.1, 110.7, 91.5, 86.8, 55.4, 31.2; HRMS  $m/z$  calcd for  $\text{C}_{27}\text{H}_{20}\text{O}_3\text{Na}$  ( $\text{M} + \text{Na}$ ) $^+$  415.1310, found 415.1302.

#### Synthesis of **2a**:



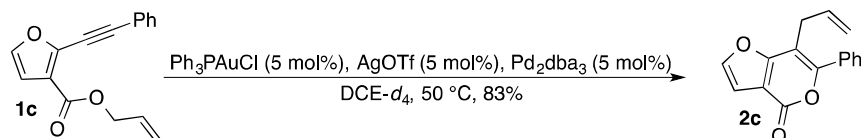
In a one dram vial, the active gold catalyst was generated with  $\text{Ph}_3\text{PAuCl}$  (1.9 mg, 3.8  $\mu\text{mol}$ , 0.050 equiv) and AgOTf (1.0 mg, 3.8  $\mu\text{mol}$ , 0.050 equiv) in DCE (0.20 mL). The active gold catalysis solution was transferred to a one dram vial containing **1a** (21 mg, 76  $\mu\text{mol}$ , 1.0 equiv) and mixed thoroughly. The solution containing the active gold catalyst and **1a** was transferred to a one dram vial containing  $\text{Pd}_2\text{dba}_3$  (3.5 mg, 3.8  $\mu\text{mol}$ , 0.050 equiv). The mixture was diluted to a total of 0.76 mL DCE. The reaction temperature was increased to 60 °C and the reaction was allowed to stir under an inert atmosphere for 8 h. The reaction mixture was concentrated in vacuo. The product was isolated and purified via silica gel column chromatography (0% to 40% EtOAc/hexanes). The product **2a** (5 mg, 24%) was fully characterized.  $^1\text{H NMR}$  ( $\text{CDCl}_3$ , 600 MHz)  $\delta$  7.98 (d,  $J = 2.6$  Hz, 1H), 7.62 (m, 2H), 7.54 (d,  $J = 8.9$  Hz, 1H), 7.44 (m, 3H), 7.35 (dd,  $J = 8.9$  Hz, 2.2 Hz, 1H), 6.71 (s, br, 1H), 6.10 (ddt,  $J = 17.8$  Hz, 10.1 Hz, 3.5 Hz, 1H), 5.22 (d,  $J = 10.1$  Hz, 1H), 5.09 (d,  $J = 17.8$  Hz, 1H), 3.45 (d,  $J = 3.5$ , 2H);  $^{13}\text{C NMR}$  ( $\text{CDCl}_3$ , 125 MHz)  $\delta$  163.3, 156.4, 150.5, 136.0, 133.2, 131.5, 129.7, 128.9, 128.5, 126.6, 124.3, 122.3, 117.4, 114.3, 111.3, 31.6; HRMS  $m/z$  calcd for  $\text{C}_{18}\text{H}_{14}\text{O}_3\text{Na}$  ( $\text{M} + \text{Na}$ ) $^+$  301.0841, found 301.0840.

### Synthesis of **2b**:



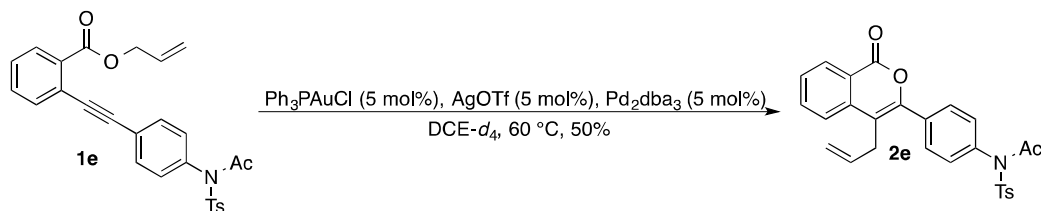
$\text{Ph}_3\text{PAuCl}$  (1.4 mg, 2.8  $\mu\text{mol}$ , 0.050 equiv) was dissolved in  $\text{CD}_2\text{Cl}_2$  (0.10 mL) and thoroughly mixed with  $\text{AgOTf}$  (0.7 mg, 2.8  $\mu\text{mol}$ , 0.050 equiv) to generate the active gold catalyst. The active gold catalyst was added to the **1b** (23 mg, 55  $\mu\text{mol}$ , 1.0 equiv) and thoroughly mixed. The mixture was then added to  $\text{Pd}_2\text{dba}_3$  (2.3 mg, 2.8  $\mu\text{mol}$ , 0.050 equiv). The reaction was allowed to stir at room temperature for 24 h. Afterwards, the reaction temperature was raised to 40  $^\circ\text{C}$ , the reaction was allowed to stir for 8 h. An excess of dimethylaminoethanethiol and triethylamine was added to the reaction mixture. The reaction mixture was allowed to stir for an additional 24 h at room temperature. The product was isolated and purified via silica gel column chromatography. The product **2b** (7.6 mg, 33%) was fully characterized.  $^1\text{H NMR}$  ( $\text{CDCl}_3$ , 600 MHz)  $\delta$  8.26 (d,  $J = 2.5$  Hz, 1H), 7.71 (d,  $J = 8.9$  Hz, 1H), 7.63 (m, 3H), 7.47 (m, 3H), 6.11 (ddt,  $J = 17.5$  Hz, 10.1 Hz, 5 Hz, 1H), 5.27 (d,  $J = 10.1$ , 1H), 5.10 (d,  $J = 17.5$  Hz, 1H), 3.47 (d,  $J = 5$  Hz, 2H);  $^{13}\text{C NMR}$  ( $\text{CDCl}_3$ , 125 MHz)  $\delta$  160.7, 153.8, 148.2, 138.0, 135.0, 132.2, 130.1, 128.6, 128.3, 127.9, 126.9, 122.6, 122.0, 118.6 (t,  $J = 318$  Hz, 1C), 117.7, 109.8, 31.4;  $^{19}\text{F NMR}$  ( $\text{CDCl}_3$ , 376 MHz)  $\delta$  -72.6; HRMS  $m/z$  calcd for  $\text{C}_{19}\text{H}_{13}\text{F}_3\text{O}_5\text{SNa}$  ( $M + \text{Na}$ ) $^+$  433.0334, found 433.0339.

### Synthesis of **2c**:



$\text{Ph}_3\text{PAuCl}$  (1.6 mg, 3  $\mu\text{mol}$ , 0.05 equiv) was dissolved in  $\text{C}_2\text{D}_4\text{Cl}_2$  (0.10 mL) and thoroughly mixed with  $\text{AgOTf}$  (0.9 mg, 3  $\mu\text{mol}$ , 0.05 equiv) to generate the active gold catalyst. The active gold catalyst was added to the **1c** (16.7 mg, 66  $\mu\text{mol}$ , 1.0 equiv) and thoroughly mixed. The mixture was then added to  $\text{Pd}_2\text{dba}_3$  (3.0 mg, 3  $\mu\text{mol}$ , 0.05 equiv). The reaction mixture was diluted to 0.66 mL. The reaction temperature was increased to 50  $^\circ\text{C}$ . The reaction was allowed to stir for 6 h. The reaction was cooled to room temperature. An excess of dimethylaminoethanethiol and triethylamine was added and the reaction mixture was allowed to stir at room temperature over night. The product was isolated and purified via column chromatography (gradient from 0% to 10% EtOAc/hexanes). The product **2c** (13.9 mg, 83%) was fully characterized.  $^1\text{H NMR}$  ( $\text{C}_2\text{D}_4\text{Cl}_2$ , 600 MHz)  $\delta$  7.61 (m, 3H), 7.47 (m, 3H), 6.91 (d,  $J = 1.8$  Hz, 1H), 6.07 (ddt,  $J = 17.3$  Hz, 11.0, 5.7 Hz, 1H), 5.16 (d,  $J = 11.0$ , 1H), 5.09 (d,  $J = 17.3$  Hz, 1H), 3.42 (d,  $J = 5.7$  Hz, 2H);  $^{13}\text{C NMR}$  ( $\text{CDCl}_3$ , 125 MHz)  $\delta$  162.3, 158.7, 156.1, 144.8, 135.0, 132.3, 130.1, 129.2, 128.7, 125.6, 116.6, 109.8, 107.8, 106.5, 29.5; HRMS  $m/z$  calcd for  $\text{C}_{16}\text{H}_{12}\text{O}_3\text{Na}$  ( $M + \text{Na}$ ) $^+$  275.0684, found 275.0693.

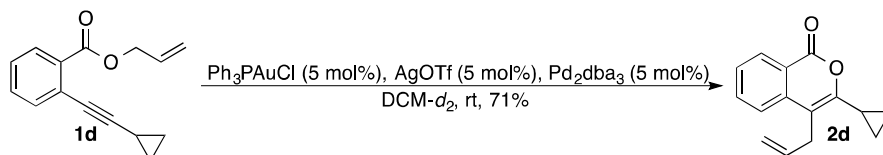
### Synthesis of **2e**:



$\text{Ph}_3\text{PAuCl}$  (1.5 mg, 3  $\mu\text{mol}$ , 0.05 equiv) was dissolved in  $\text{C}_2\text{D}_4\text{Cl}_2$  (0.10 mL) and thoroughly mixed with  $\text{AgOTf}$  (0.9 mg, 3  $\mu\text{mol}$ , 0.05 equiv) to generate the active gold catalyst. The active gold catalyst was added to **1e** (30 mg, 64  $\mu\text{mol}$ , 1 equiv) and thoroughly mixed. The mixture was then added to  $\text{Pd}_2\text{dba}_3$  (3.0 mg, 3  $\mu\text{mol}$ , 0.05 equiv). The reaction mixture was diluted to 0.64 mL. The reaction temperature was increased to 60  $^\circ\text{C}$ . The reaction was allowed to stir for 9 h. The reaction was cooled to room temperature. The product was isolated and purified via column chromatography (0% to 30% EtOAc/hexanes). The product **2e** (15 mg, 50%) was fully characterized.  $^1\text{H NMR}$  ( $\text{CDCl}_3$ , 600 MHz)  $\delta$  8.40 (d,  $J = 7.9$  Hz, 1H), 7.93 (d,  $J = 8.1$  Hz, 2H), 7.79 (m, 3H), 7.64 (d,  $J = 8.1$  Hz, 1H), 7.58 (app-t,  $J = 7.4$  Hz, 1H), 7.36 (app-t,  $J = 7.2$ , 4H), 6.17 (ddt,  $J = 17.5$  Hz, 10.1 Hz, 2.4 Hz, 1H), 5.28 (d,  $J = 10.1$  Hz, 1H), 5.10 (d,  $J = 17.5$  Hz, 1H), 3.53 (d,  $J = 2.4$  Hz, 2H), 2.47 (s, 3H), 1.92 (s, 3H);  $^{13}\text{C NMR}$  ( $\text{CDCl}_3$ , 125 MHz)

$\delta$  169.7, 162.0, 150.7, 145.2, 137.7, 137.5, 135.8, 135.3, 134.8, 134.5, 130.1, 129.9, 129.8, 129.5, 129.4, 129.2, 128.4, 124.4, 121.0, 117.5, 111.6, 31.3, 25.1, 21.7; HRMS  $m/z$  calcd for  $C_{27}H_{23}NO_5SNa$  ( $M + Na$ )<sup>+</sup> 496.1195, found 496.1197.

Synthesis of **2d**:



$Ph_3PAuCl$  (2.4 mg, 5  $\mu$ mol, 0.05 equiv) was dissolved in  $CD_2Cl_2$  (1 mL) and thoroughly mixed with  $AgOTf$  (1.3 mg, 5  $\mu$ mol, 0.05 equiv) to generate the active gold catalyst. The active gold catalyst was added to **1d** (23 mg, 0.10 mmol, 1.0 equiv) and thoroughly mixed. The mixture was then added to  $Pd_2dba_3$  (4.6 mg, 5  $\mu$ mol, 0.05 eq). The reaction was allowed to stir at room temperature for 24 h. The products were purified by silica gel chromatography (0% to 10% EtOAc/hexanes). The product **2d** (16 mg, 71%) was fully characterized  $^1H$  NMR ( $CDCl_3$ , 600 MHz)  $\delta$  8.28 (dd,  $J = 7.9$  Hz, 1.1 Hz, 1H), 7.70 (app-td,  $J = 7.4$  Hz, 1.4 Hz, 1H), 7.48 (d,  $J = 8.2$  Hz, 1H), 7.42 (app-td,  $J = 7.2$  Hz, 1.3 Hz, 1H), 5.99 (ddt,  $J = 17.1$  Hz, 10.4 Hz, 5.7 Hz, 1H), 5.12 (app-dq,  $J = 10.5$  Hz, 1.3 Hz, 1H), 5.10 (app-dq,  $J = 17.2$ , 1.4 Hz, 1H), 3.53 (app -dt,  $J = 5.6$  Hz, 1.3 Hz, 2H), 1.97 (m, 1H), 1.18 (m, 2H), 0.94 (m, 2H);  $^{13}C$  NMR ( $CDCl_3$ , 125 MHz)  $\delta$  162.6, 154.5, 138.3, 134.9, 134.8, 130.0, 126.9, 122.6, 116.5, 109.2, 29.8, 11.1, 7.3; HRMS  $m/z$  calcd for  $C_{15}H_{14}O_2H$  ( $M + H$ )<sup>+</sup> 227.1072, found 227.1076.

## REFERENCES FOR EXPERIMENTAL SECTION

- 1.) Huang, Z.; Liu, Z.; Zhou, S. *J. Am. Chem. Soc.* **2011**, *133*, 15882 – 15885.
- 2.) Spivey, A. C.; McKendrick, J.; Srikanan, R. *J. Org. Chem.* **2003**, *68*, 1843 – 1851.
- 3.) Marchal, E.; Uriac, P.; Legouin, B.; Toupet, L.; Weghe, P. *Tetrahedron*, **2007**, *63*, 9979 – 9990.
- 4.) Gioiello, A.; Rosatelli, E.; Teofrati, M.; Filipponi, P.; Pellicciari, R. *ACS. Comb. Sci.* **2013**, *15*, 235 – 239.



## Chapter 2

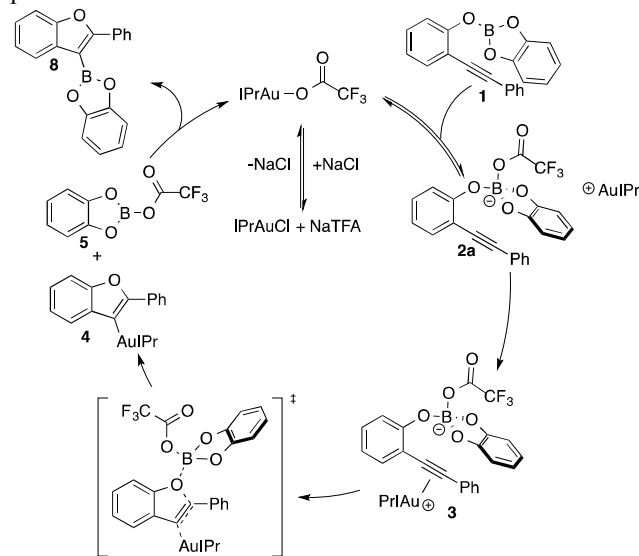
### Mechanistic Studies of The Alkoxyboration Reaction

**Abstract:** A kinetic study of the alkoxyboration reaction showed the reaction is second order overall: first order with respect to both IPrAuTFA and NaTFA. NaTFA was determined to be both a Lewis base, by enhancing the nucleophilicity the B–O  $\sigma$  bond, and as a reagent needed to decrease the amount of IPrAuTFA converted to the catalytically inactive IPrAuCl. Increasing the amount of trifluoroacetate eventually lead to a decrease in the reaction rate. This effect is attributed to over addition of the Lewis base into the borate ester that is formed after the cyclization step. The rate-determining step was determined to be the transmetalation step.

### INTRODUCTION

In 2014, the Blum group reported a gold catalyzed alkoxyboration reaction, allowing for the synthesis of 3-borylbenzofurans.<sup>1</sup> In the seminal publication, the authors proposed that the reaction was catalyzed by a bifunctional catalyst capable of activating both the B–O  $\sigma$  bond and C–C  $\pi$  bond. The proposed mechanism is shown in Figure 2.1. In this mechanism, the trifluoroacetate ion is transferred to the boric ester to form a gold borate ion pair, **2a**. Upon binding of the cationic gold complex to the

Figure 2.1 The proposed mechanism from the seminal publication.



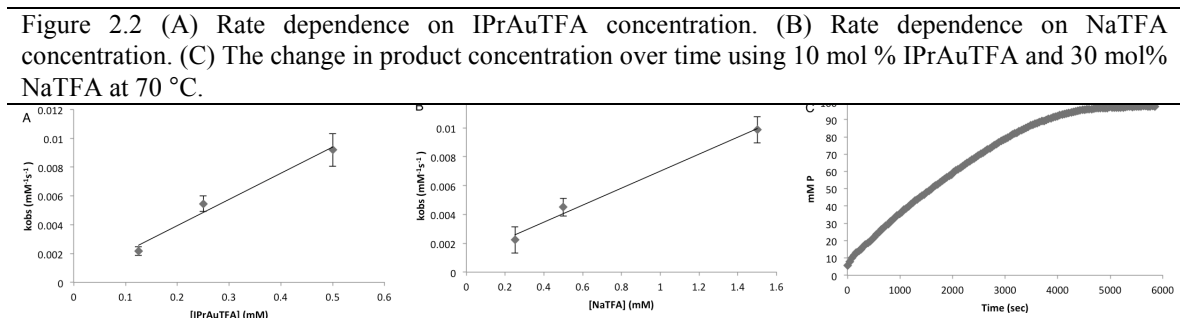
alkyne, **3**, cyclization yields a vinyl organogold intermediate, **4**, and a borate ester, **5**. Transmetalation yields the desired product, **8**. The proposed mechanism is dissimilar to other reactions that are formal 1,3-transpositions due to the migration occurring in an intermolecular fashion.<sup>2,3</sup> The role of sodium trifluoroacetate (NaTFA) along with the manner in which transmetalation occurred remained unclear. The purpose of this study was to determine the mechanism of the alkoxyboration reaction.

### RESULTS AND DISCUSSION

Initial studies focused on determining the order with respect to IPrAuTFA, NaTFA, and **1**. The kinetic data (Figure 2.2) obtained is consistent with the rate equation shown in eq 1. Note, increasing the amount of NaTFA from 15 mol % (1.5 mM) to 30 mol % (3.0 mM) showed no statistically significant increase in rate.

$$\text{Rate} = k[\text{IPrAuTFA}][\text{NaTFA}] \quad (1)$$

Due to the enormous amount of literature precedent surrounding gold(I) complexes acting as carbophilic  $\pi$  acids, IPrAuTFA is thought to act as such.<sup>4</sup> However, the manner in which NaTFA affects the rate of the reaction is not as obvious.



In order to deconvolute the role of NaTFA, mass spectrometry and NMR spectroscopy were used. Saturation of a 100 mM solution of **1** in toluene with NaTFA followed by equilibration at 50 °C showed one small peak around the -75 ppm region in the <sup>19</sup>F NMR, a range consistent with compounds containing the TFA fragment.<sup>5</sup> A saturated solution of NaTFA in toluene produces no peaks consistent with NaTFA at 50 °C, suggesting NaTFA alone is not soluble. Mass spectrometry data suggests that NaTFA can be solubilized via complexation with the boric ester. Borate **6** (Figure 2.5) was observed via ESI-MS. Since the only source of the catecholborane fragment is the boric ester, the TFA ion must react with **1** in order to produce the observed species. A m/z value of 425.1 with a isotopic distribution indicative of a boron containing compound was observed in the MS spectrum, suggesting that there may be **2** in solution, but due to the low intensity nothing can be said definitively. It is also notable that tetrakis(2,2,2-trifluoroacetoxy)borate, B(TFA)<sub>4</sub><sup>-</sup>, was also observed, suggesting that if the TFA concentration is too high over-addition can occur, which would lead to a decrease in yield and rate.

To test this hypothesis, tetrabutylammonium trifluoroacetate (TTFA) was synthesized and used as a substitute for NaTFA. The rate decreased dramatically in the case where TTFA was used (Table 2.1). This result is consistent with the hypothesis that there exists an ideal concentration of TFA, such that solutions with higher concentrations of TFA may open an alternative pathway where the catecholborane fragment can be removed from

the reaction medium and solutions with lower concentrations may suffer from both catalyst quenching by exogenous chloride and a lack of borate in solution. Ion pairs suspended in solution may also have a detrimental effect on the rate of the reaction. The active gold catalyst (presumably IPrAu<sup>+</sup>) can be quenched by NaCl to generate IPrAuCl (observed by ESI-MS). The concentration of chloride can be reduced via the common ion effect. Also, a metathesis reaction can regenerate the precatalyst from

IPrAuCl. Based on this reasoning, it is reasonable to believe that Na<sup>+</sup> may have an effect on the rate.

In order to determine the rate dependence of Na<sup>+</sup> on the reaction, the reaction without any

Table 2.1 Observed rate constants shown for comparison.	
Conditions	k <sub>obs</sub> (mM <sup>-1</sup> s <sup>-1</sup> × 10 <sup>-3</sup> )
5 mol % IPrAuTFA, 15 mol % NaTFA	9.9 ± 0.9
5 mol % IPrAuTFA, 15 mol % TTFA	0.096 ± 0.111
5 mol % IPrAuTFA, 15 mol % SHFA	0.72 ± 0.62
5 mol % IPrAuTFA	0.015 ± 0.059
5 mol % IPrAuTFA (No NaCl)	55 ± 7

additional salt (outside the NaCl generated by the reaction) was used as the control experiment for comparison. Replacing NaTFA with sodium hexafluoroantimonate (SHFA) led to a major decrease in yield relative to the case where NaTFA was present (Table 2.1). However, comparison of the control and experimental rate constants suggest that Na<sup>+</sup> may have a favorable effect on the rate. Due to the low precision of the calculated rate constants, there is ambiguity that lies in whether or not the increase in rate is statistically significant. Using the T.TEST function in Microsoft Excel, the probability that the difference in the rate constants is insignificant was determined to be 0.09. To rephrase for clarity, there is a 91% chance that the differences in the rate constants are statistically significant and that Na<sup>+</sup> concentration influences the rate of the reaction.

The common ion effect and a metathesis may be occurring simultaneously. The common ion effect seems to play a minor role in reducing the amount of catalyst quenching (assuming NaTFA and SHFA have similar solubility products in toluene). A metathesis reaction may account for the low rate when SHFA is used as a co-catalyst.

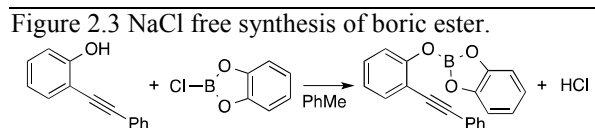


A metathesis reaction to produce IPrAuSbF<sub>6</sub> would not produce a bifunctional catalyst capable of enhancing both the electrophilicity and nucleophilicity of the substrate (eq 2).

The nature of the transmetalation step may also be discerned by analysis of the rate constants shown in Table 2.1. The experimentally determined rate constant for the reaction with 5 mol % IPrAuTFA in the presence of NaCl is 660 times slower than the reaction with 5 mol % IPrAuTFA and 15 mol % NaTFA. If the transmetalation occurred via a four centered transition state, the relative rate would not be as large because every turnover would

result in the formation of the precatalyst. A metathesis reaction with NaCl may lead to a decrease in rate due to formation of IPrAuCl, but the likelihood of a reaction with NaCl relative to a reaction with the catalyst is low due to the low solubility of NaCl in toluene. An alternative explanation is that putative organogold intermediate, **4**, may act as a nucleophile towards the borate ester, **5**, resulting in a gold borate ion pair, **7**. Dissociation of TFA would produce the desired product. This hypothesis is consistent with the data obtained from a recent study from the Blum group.<sup>6</sup>

The rate retardation attributed to NaCl can easily be discerned via its elimination from the reaction medium. The reaction of

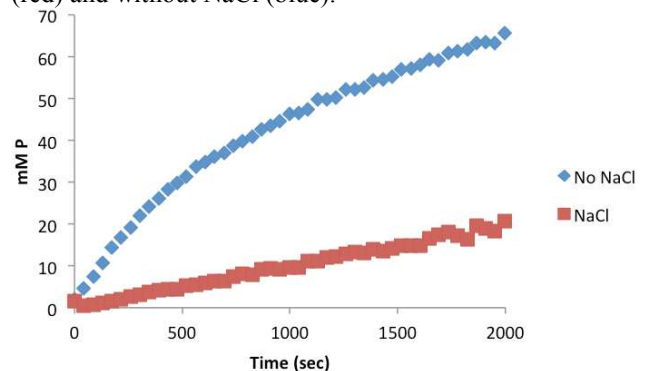


chlorocatecholborane and 2-(phenylethynyl)phenol generated the desired boric ester and hydrogen chloride gas (Figure 2.3). The container was evacuated to remove hydrogen chloride.

The kinetic studies using the boric ester synthesized in this fashion show a different kinetic profile than what is observed under nominal conditions. Using 5 mol % IPrAuTFA, the initial rate (Table 2.1, Figure 2.4) is about 5.5 times higher than the initial rate using 5 mol % IPrAuTFA and 15 mol % NaTFA in the presence of NaCl (Table 2.1, Figure 2.4). The rate quickly decreases about 40% of the initial rate after about 20% conversion to product (figure 2.4).

The rate decrease may be due to the manner in which NaCl affects the electronic nature of the solvent. When NaCl is present, ions are more stable. After the formation of complex **2**, the species is persistent enough to allow the cationic gold catalyst to form the  $\pi$  complex, **3**. When NaCl is absent, ionic species are not as stable. Upon formation of **2**, disproportionation occurs to form **5**

Figure 2.4 Change in product concentration over time with (red) and without NaCl (blue).



and a phenolate. The phenolate can then bind to the cationic gold complex, resulting a lower amount of gold catalyst to induce cyclization, which would result in a lower, steady rate after equilibration.

Notably, in absence of NaCl, NaTFA retards the rate. The active catalyst and precatalyst could be in equilibrium. Introduction of more TFA in solution would lead to a shift in the equilibrium resulting in less active

catalyst in solution. A greater amount of the TFA in the reaction medium could result in increased amounts of borate **6** in solution, which may not be a reactive partner in gold to boron transmetalation. It appears that doubling the concentration of TFA in the reaction medium cuts the rate in half. The current data suggest that the reaction is inverse first order with respect to TFA when NaCl is absent.

Table 2.2. The activation parameters from the Eyring analysis.

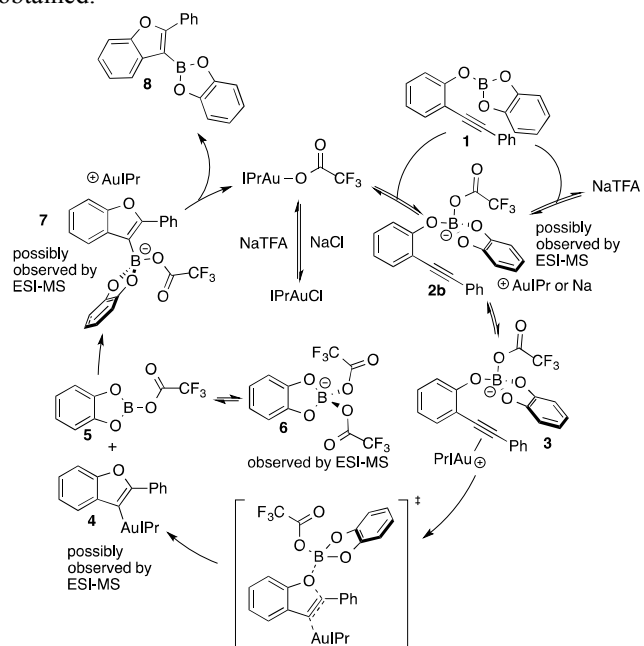
$\Delta G^\ddagger$ (kcal mol <sup>-1</sup> )	$\Delta H^\ddagger$ (kcal mol <sup>-1</sup> )	$\Delta S^\ddagger$ (cal K <sup>-1</sup> mol <sup>-1</sup> )
26.34 ± 1.47	13.44 ± 1.47	- 39.96 ± 4.58

An Eyring analysis was carried out to discern the rate-determining step (RDS) of the reaction. The thermodynamic values are listed in

Table 2.2. The entropy of activation is consistent with a bimolecular RDS.<sup>7</sup> There are two bimolecular steps in the proposed mechanism, substrate binding and transmetalation. The positive enthalpy of activation indicates that energy is required for the transformation to occur. In the case of substrate binding, the enthalpy of activation should be negative, also if substrate binding was the RDS, the reaction would not be zero order with respect to substrate. The data from the Eyring analysis suggests the transmetalation is the RDS of the reaction.

The data obtained in this study is consistent with the mechanism shown in Figure 2.5, which takes into account the ability of NaTFA to act as a source of TFA for **1** and **5**. The intermediate after the transmetalation step is also included. In conclusion, the alkoxyboration reaction is second order overall, first order with respect to both IPrAuTFA and NaTFA. NaTFA acts as both a species that reduces the degree of catalyst quenching and acts as a source of TFA that can produce the borate necessary for cyclization to occur. The rate of the reaction seems to be sensitive to the amount of TFA present in solution.

Figure 2.5 The Proposal mechanism based on the data obtained.



Solutions with higher concentrations of TFA had lower observed rate constants. The data obtained from the Eyring analysis suggests the RDS is the transmetalation.

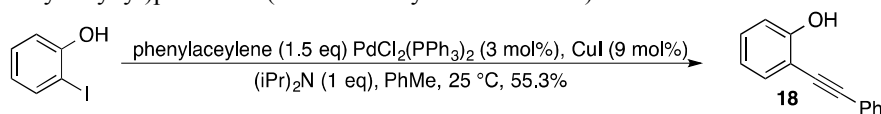
## REFERENCES

- Hirner, J. J.; Faizi, D. J.; Blum, S. A. *J. Am. Chem. Soc.* **2014**, *136*, 4740 – 4745.
- Nakamura, I.; Yamagishi U.; Song, D.; Konta, S.; Yamamoto Y. *Angew. Chem. Int. Ed.* **2007**, *46*, 2284 – 2287.
- Tokimizu, Y.; Oishi, S.; Fujii, N.; Ohno, H. *Angew. Chem. Int. Ed.* **2015**, *56*, 7862 – 7866.
- Li, Z.; Brouwer, C.; He, C. *Chem. Rev.* **2008**, *108*, 3239 – 3265.
- Wang, F.; Chen, D.; Deng, H.; Chen, Q.; Liu, X.; Jian, X. *Tetrahedron* **2014**, *70*, 2582 – 2590.
- Hirner, J. J.; Blum, S.A. *Tetrahedron* **2015**, *71*, 4445 – 4449.
- Denmark S. E.; Bui, T. *J. Org. Chem.* **2005**, *70*, 10393 – 10399.

## EXPERIMENTAL

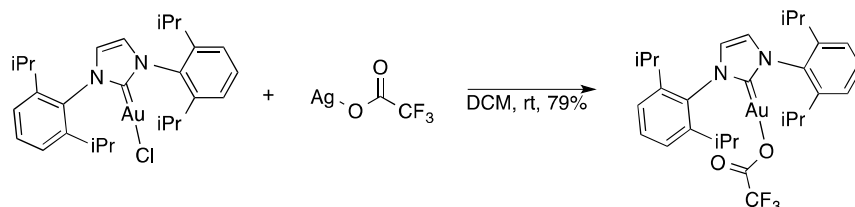
### I. Synthetic Procedures

Synthesis of 2-(phenylethynyl)phenol **18** (Conducted by Darius J. Faizi)



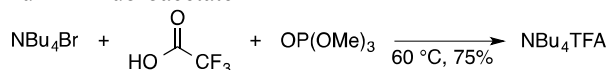
A flame dried 1-L round bottomed flask equipped with a stir bar was charged with 2-iodophenol (14.0 g, 3.60 mmol, 1.00 equiv), copper iodide (1.09 g, 5.71 mmol, 0.08980 equiv), and bis(triphenylphosphine)palladium dichloride (1.33 g, 1.89 mmol, 0.0297 equiv). Toluene (320 mL), diisopropylamine (8.90 mL, 63.6, 1.00 equiv), and phenylacetylene (10.5 mL, 95.4 mmol, 1.50 equiv) were added sequentially. The reaction mixture was stirred for 8.5 h at 25 °C. Ethyl acetate (100 mL) was added and the mixture was washed with saturated ammonium chloride (50 mL × 1) then brine (50 mL × 1). The organic layer was isolated and dried with sodium sulfate. Upon concentration of the solution, a dark brown oil was obtained. The material was purified via column chromatography (12% EtOAc in hexanes). Like fractions were collected, concentrated and dried under high vacuum. The resultant oil was recrystallized from hexanes to afford a brown crystalline material (6.82 g, 55.3%). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz): δ 7.57–7.55 (m, 2H), 7.44 (dd, *J* = 7.6, 1.6 Hz, 1H), 7.40–7.38 (m, 3H), 7.30–7.27 (m, 1H), 7.03 (dd, *J* = 8.3, 0.8 Hz, 1H), 6.96 (td, *J* = 7.5, 1.0 Hz, 1H), 5.90 (s, 1H). Spectral Information is in agreement with literature precedent.<sup>1</sup>

Synthesis of IPrAuTFA



IPrAuCl (124. mg, 0.200 mmol, 1.00 equiv) and AgTFA (44.2 mg, 0.200 mmol, 1.00 equiv) were weighed in separate vials. IPrAuCl was dissolved in DCM (1 mL). The solution was transferred to the vial with AgTFA. DCM (1 mL) was used to rinse the IPrAuCl vial, washings were transferred to the vial with AgTFA. The reaction mixture was then capped with a teflon coated cap and covered in aluminum foil and allowed to stir for 30 h in a glovebox. The reaction mixture was then filtered through glass fiber filter paper. The resultant solution was concentrated inside the glovebox. No further purification was performed, a colorless solid was isolated (110.4 mg, 79%). <sup>1</sup>H (PhMe-*d*<sub>8</sub>, 400 MHz) δ 7.13 (t, *J* = 8 Hz, 2H), 6.30 (s, 2H), 2.46 (app-sep, *J* = 7 Hz, 4H), 1.35 (d, *J* = 7 Hz, 12H), 1.02 (d, *J* = 7 Hz, 12H). Spectral Information is in agreement with literature precedent.<sup>2</sup>

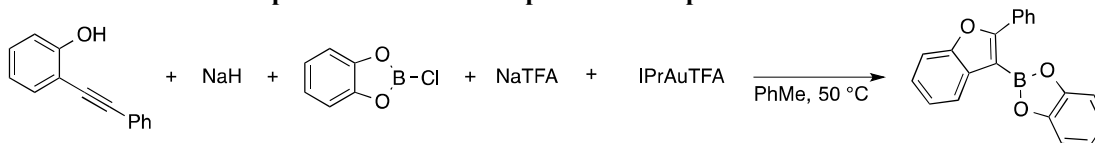
Synthesis of Tetrabutylammonium Trifluoroacetate



The product was prepared according to literature precedent.<sup>3</sup> Trifluoroacetic acid (0.04 mL, 0.5 mmol, 1 equiv) was added to a 10 mL round bottom flask with trimethyl phosphate (0.7 mL, 6.0 mmol, 12 equiv) dropwise. While stirring tetrabutylammonium bromide (161 mg, 0.500 mmol, 1.00 equiv) was added. The reaction was allowed to stir at 60 °C overnight. Excess trimethylphosphate was removed via vacuum distillation at 85 °C. The resultant residue was dissolved in DCM (2 mL), 25 drops of a 10% (w/v) NH<sub>4</sub>OH aqueous solution was added. The pH of the

solution was checked while stirring. Water (2 mL) was added when pH = 7. The layers were separated. The aqueous layer was extracted with DCM (2 mL × 2). The organic fractions were combined, concentrated, and dried under high vacuum at 100 °C overnight. No further purification was performed, the product was isolated as a beige solid (133.3, 75%). <sup>1</sup>H (CDCl<sub>3</sub>, 600 MHz) δ 3.30 (t, J = 8 Hz, 8H), 1.65 (app-quin, J = 8 Hz, 8H), 1.44 (app-sex, J = 8 Hz, 8H), 1.01 (t, J = 8 Hz, 12H); <sup>19</sup>F (CDCl<sub>3</sub>, 376 MHz) δ 75.10.

## II. General Procedure for Preparation of Kinetic Experiment Samples



2-(phenylethynyl)phenol (34.0 mg, 0.175 mmol, 1.00 equiv), 89% w/w NaH (4.7 mg, 0.18 mmol, 1.0 equiv), B-chlorocatecholborane (27.0 mg, 0.175 mmol, 1.00 equiv), Sodium trifluoroacetate (7.2 mg, 0.053 mmol, 0.30 equiv), IPrAuTFA (12.3 mg, 0.018 mmol, 0.10 equiv) were weighed in separate 4 mL vials. 2-(phenylethynyl)phenol was dissolved in toluene-*d*<sub>8</sub> (250 μL) and transferred to the vial with NaH. The mixture was stirred by hand briefly and allowed to sit for 15 min. The mixture was then transferred to the vial with B-chlorocatecholborane. Toluene-*d*<sub>8</sub> (250 μL × 2) was used to rinse through the first two vials. The mixture was allowed to stir for 30 min with occasional stirring by hand. Toluene-*d*<sub>8</sub> was added to NaTFA to create a suspension, which was then transferred to the reaction mixture. This process was repeated until the total volume of solvent added to the reaction container was 1400 μL. The IPrAuTFA was dissolved in toluene-*d*<sub>8</sub> (350 μL). From the boric ester stock solution, 400 μL was transferred to a J-Young tube and capped with a rubber septum. This was repeated a total of three times.

## III. General Procedure for Kinetic Experiments

One J-Young tube and one 250 μL capped (with a rubber stopper) gas tight syringe with 100 μL of the IPrAuTFA solution were removed from the glovebox. The rubber spectrum on the J-Young tube was wrapped in parafilm. The sample in the J-Young tube was immediately transported to the NMR spectrometer, which underwent temperature calibration before sample injection. The contents of the syringe were injected into the J-Young tube. The J-Young was shaken and inserted into the NMR spectrometer. Acquisition of spectra immediately followed. The acquisition time was set to 6 s. A 90° pulse was used. The line broadening was set to 1 Hz. The number of scans per experiment = 1. The number of dummy scans = 0. Fifty experiments were conducted sequentially with 34 s delays between experiments.

## IV. Raw Kinetic Data

Catalyst Loadings	average rate (mM/s × 10 <sup>-3</sup> )	St. Dev. (mM/s × 10 <sup>-3</sup> )
7.5 mol % IPrAuTFA, 30 mol % NaTFA	11	0.1
5 mol % IPrAuTFA, 30 mol % NaTFA	9.2	1.1
5 mol % IPrAuTFA, 15 mol % NaTFA	9.9	0.9
5 mol % IPrAuTFA, 5 mol % NaTFA	4.5	0.6
2.5 mol % IPrAuTFA, 30 mol % NaTFA	5.5	0.6
5 mol % IPrAuTFA, 15 mol % NaBArF	0	0
5 mol % IPrAuTFA, No salt	0.015	0.059
5 mol % IPrAuTFA, 15 mol % NaSbF <sub>6</sub>	0.72	0.6
5 mol % IPrAuTFA, 15 mol % NBu <sub>4</sub> TFA	0.096	0.111
1.25 mol % IPrAuTFA, 30 mol % NaTFA	2.2	0.3
5 mol % IPrAuTFA, 2.5 mol % NaTFA	2.2	0.9
5 mol % IPrAuTFA, No NaTFA/NaCl	55	6
5 mol % IPrAuTFA, 5 mol % NaTFA, no NaCl	20	1

5 mol % IPrAuTFA, 10 mol % NaTFA, no NaCl

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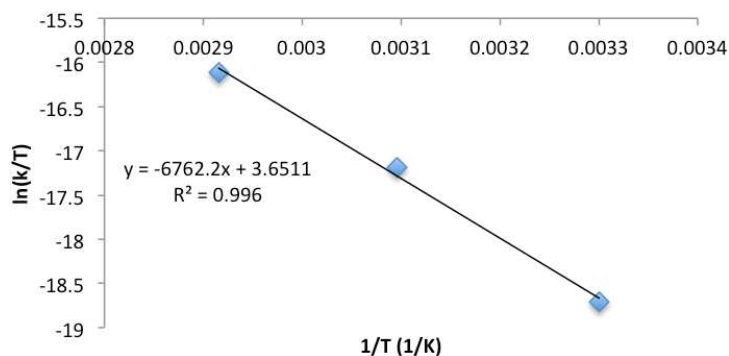
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## V. EYRING ANALYSIS DATA

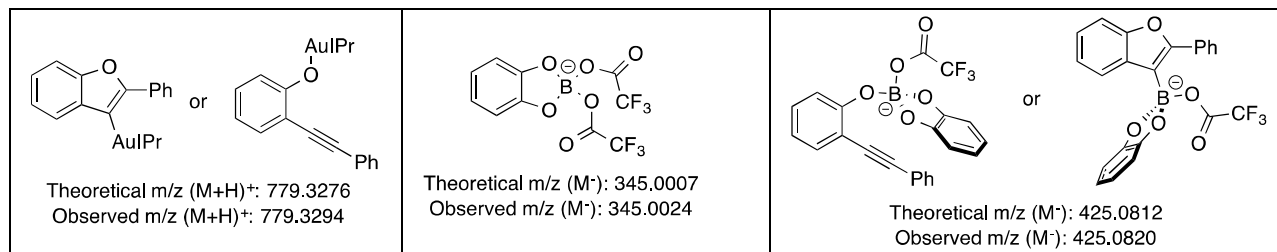
Temperature (°C)	Average rate (M/s × 10 <sup>-5</sup> )	St. Dev. (M/s × 10 <sup>-5</sup> )
70	3.4	0.2
50	1.1	0.1
30	0.23	0.04

Raw Eyring Analysis Data		Activation Parameters		
1/T (K <sup>-1</sup> )	ln(k/T)	Parameter	Value	St. Dev.
0.0029	-16.11	$\Delta G^\ddagger$ (kcal mol <sup>-1</sup> )	26.3	1.5
0.0031	-17.19	$\Delta S^\ddagger$ (cal K <sup>-1</sup> mol <sup>-1</sup> )	-40.0	4.6
0.0033	-18.71	$\Delta H^\ddagger$ (kcal mol <sup>-1</sup> )	13.4	1.5

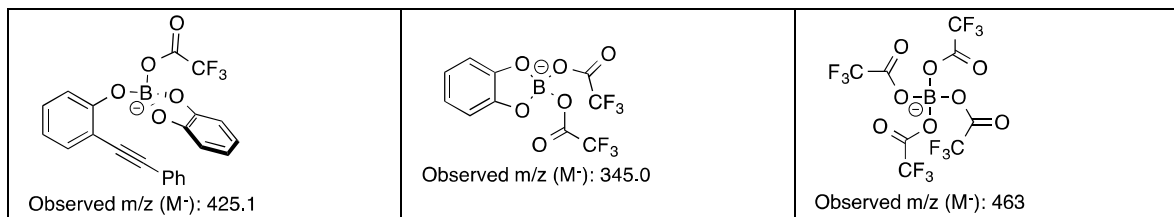
### Eyring Analysis



## VI. Reaction Intermediates Found By High Resolution Mass Spectrometry



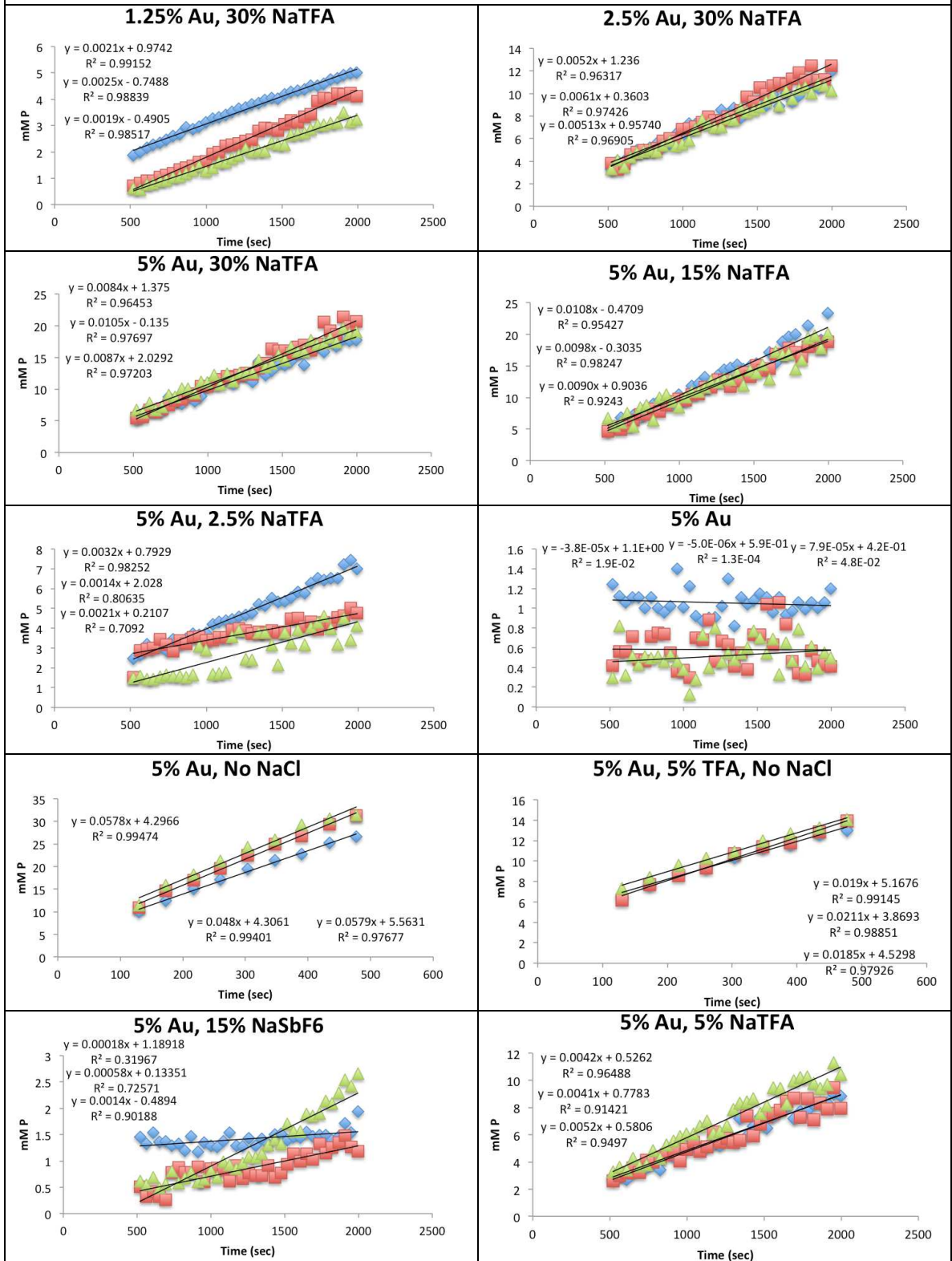
## VII. Species Found in NaTFA Solubility Study by Low Resolution Mass Spectrometry

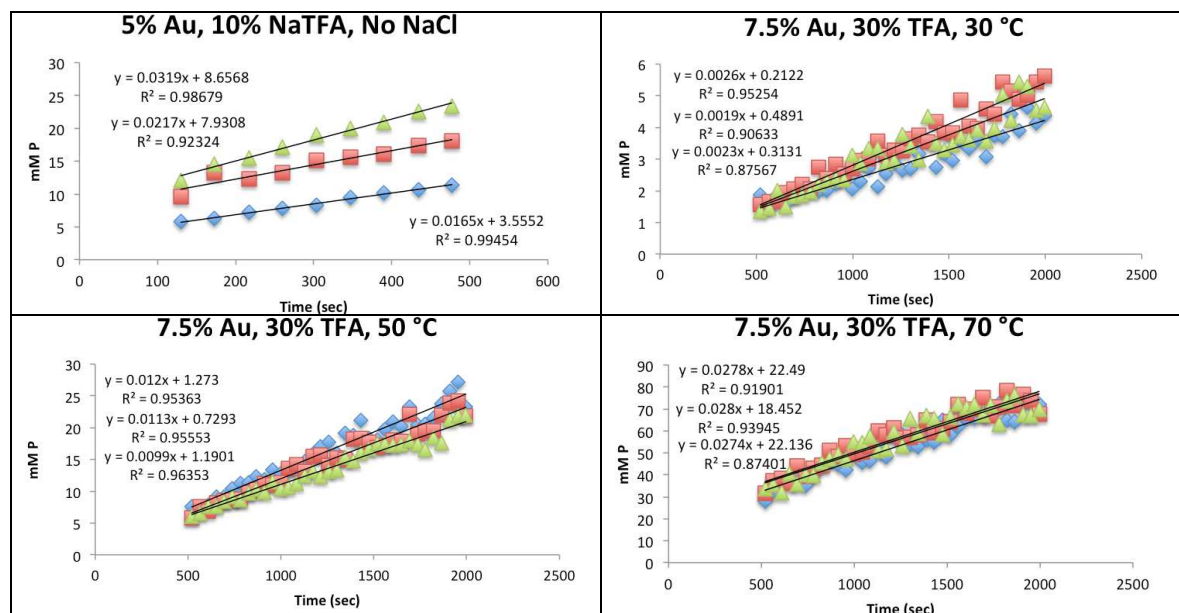


## IX. Kinetic Plots



Note: 1.25% Au = 1.25 mol % IPrAuTfA



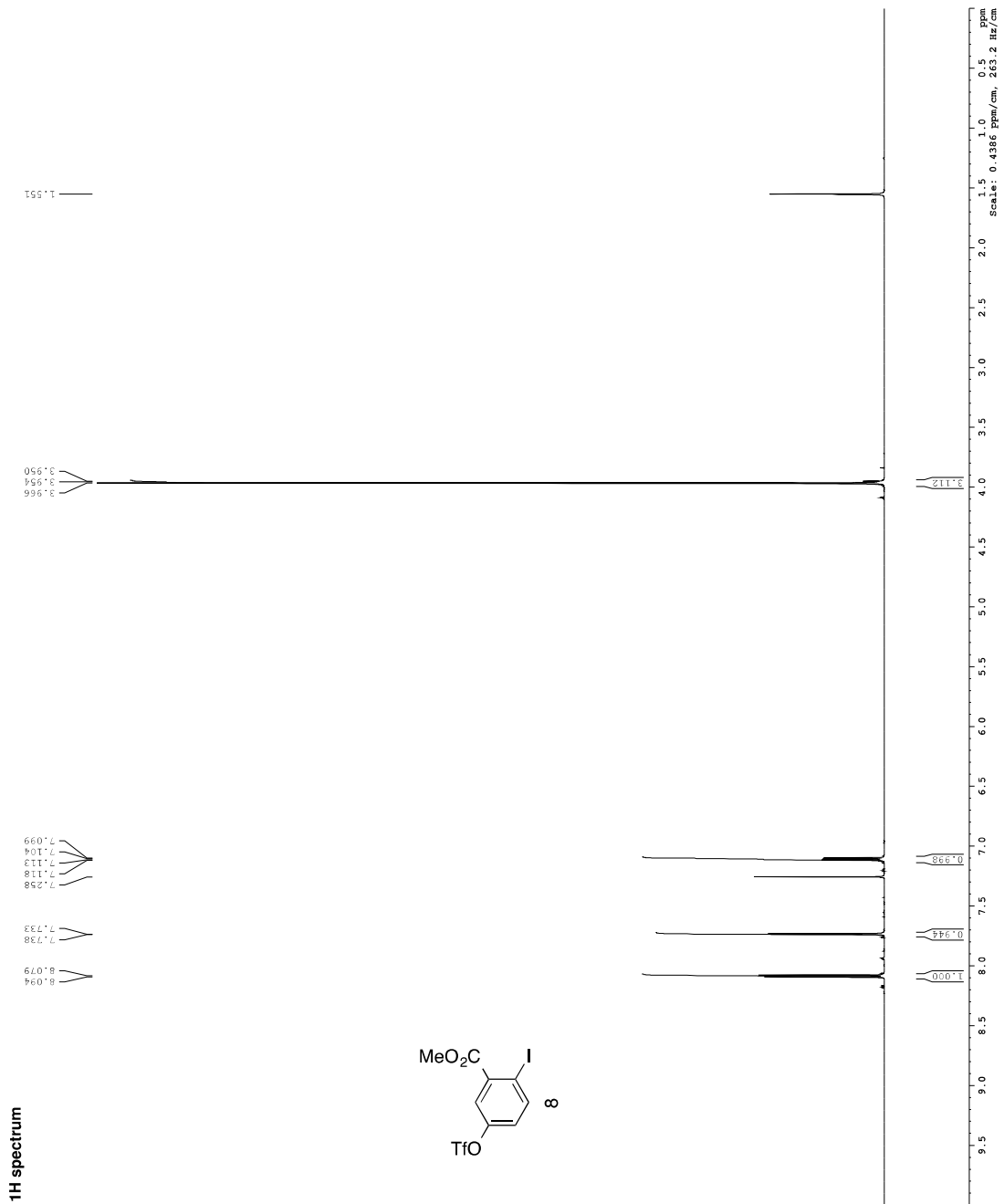


## REFERENCES FOR EXPERIMENTAL SECTION

- 1.) Hirner, J. J.; Faizi, D. J.; Blum, S. A. *J. Am. Chem. Soc.* **2014**, *136*, 4740 – 4745.
- 2.) Chong, E.; Blum, S. A. *J. Am. Chem. Soc.* **2015**, *137*, 10144 – 10147.
- 3.) Jeon, J. Y.; Varghese, J. K.; Park, J. H.; Lee, S.; Lee, B. Y. *Eur. J. Org. Chem.* **2012**, *2012*, 3566 – 3569.

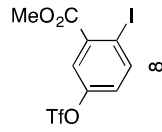
# Appendix A: NMR Spectra

Current Data Parameters  
 Name: 1302017  
 PROCNO: 1  
 F2 - Acquisition Parameters  
 Date\_ Time: 2011.06.11.06  
 INSTRUM: spect  
 PULPROG: zgpg30  
 SCLPRG: zgpg30  
 SOLVENT: CDCl3  
 US: 2  
 SWH: 916.385 Hz  
 FWHM: 5.4998878 Hz  
 AQ: 57.000 usec  
 RG: 248.0 usec  
 SF: 600.130070 MHz  
 EQ: 248.0 usec  
 ZF0: 0.10000000 usec  
 ===== CHANNEL f1 =====  
 NUC1: 1H  
 P1: 12.00 usec  
 PL1: 0.00 dB  
 ZF1: 22.0444556 MHz  
 SF01: 600.130070 MHz  
 F2 - Processing parameters  
 SI: 32768  
 SF: 600.130070 MHz  
 EQ2: 248.0 usec  
 SFO2: 600.130070 MHz  
 LB: 0.200 Hz  
 GB: 0.000 Hz  
 PC: 1.000

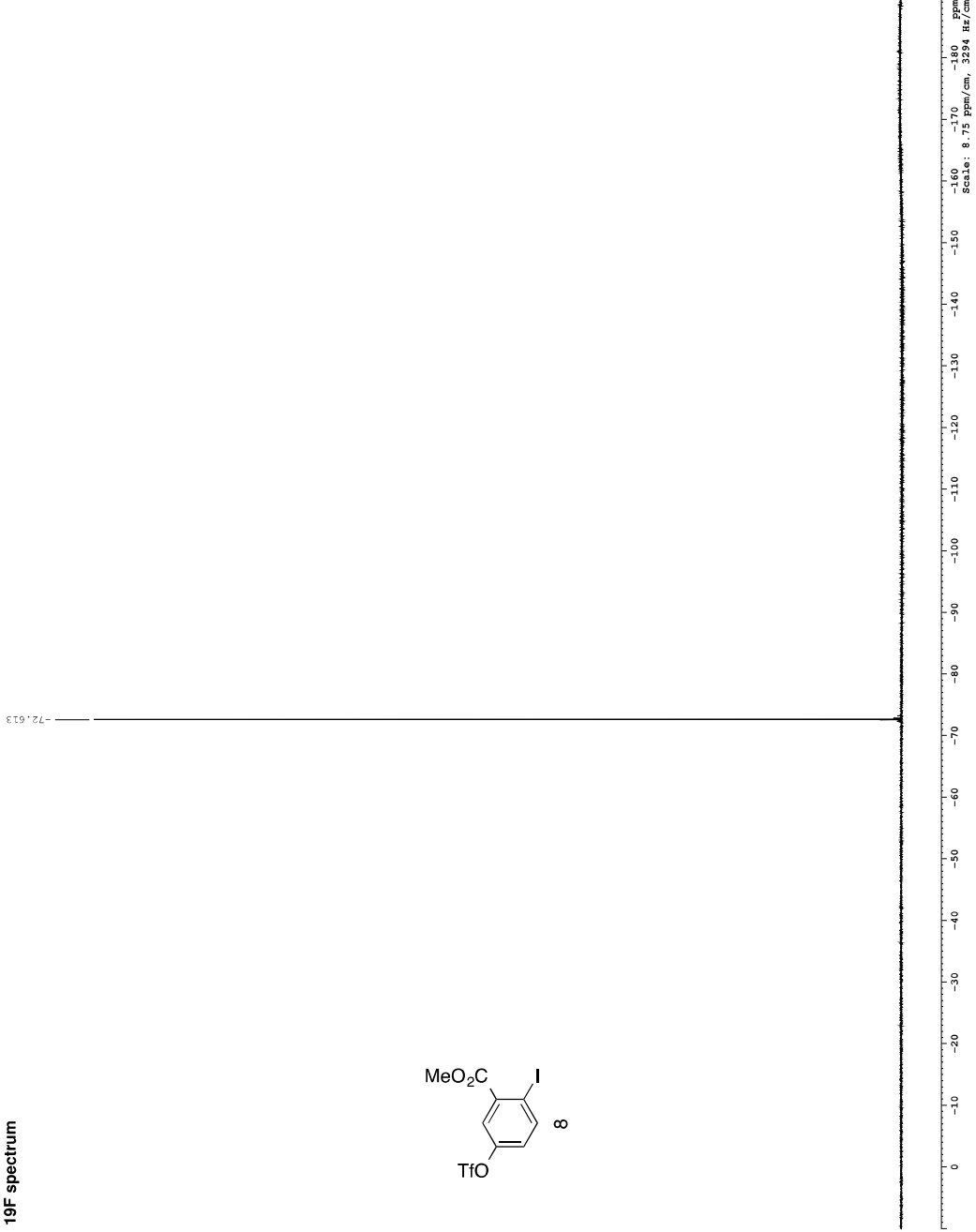




19F spectrum

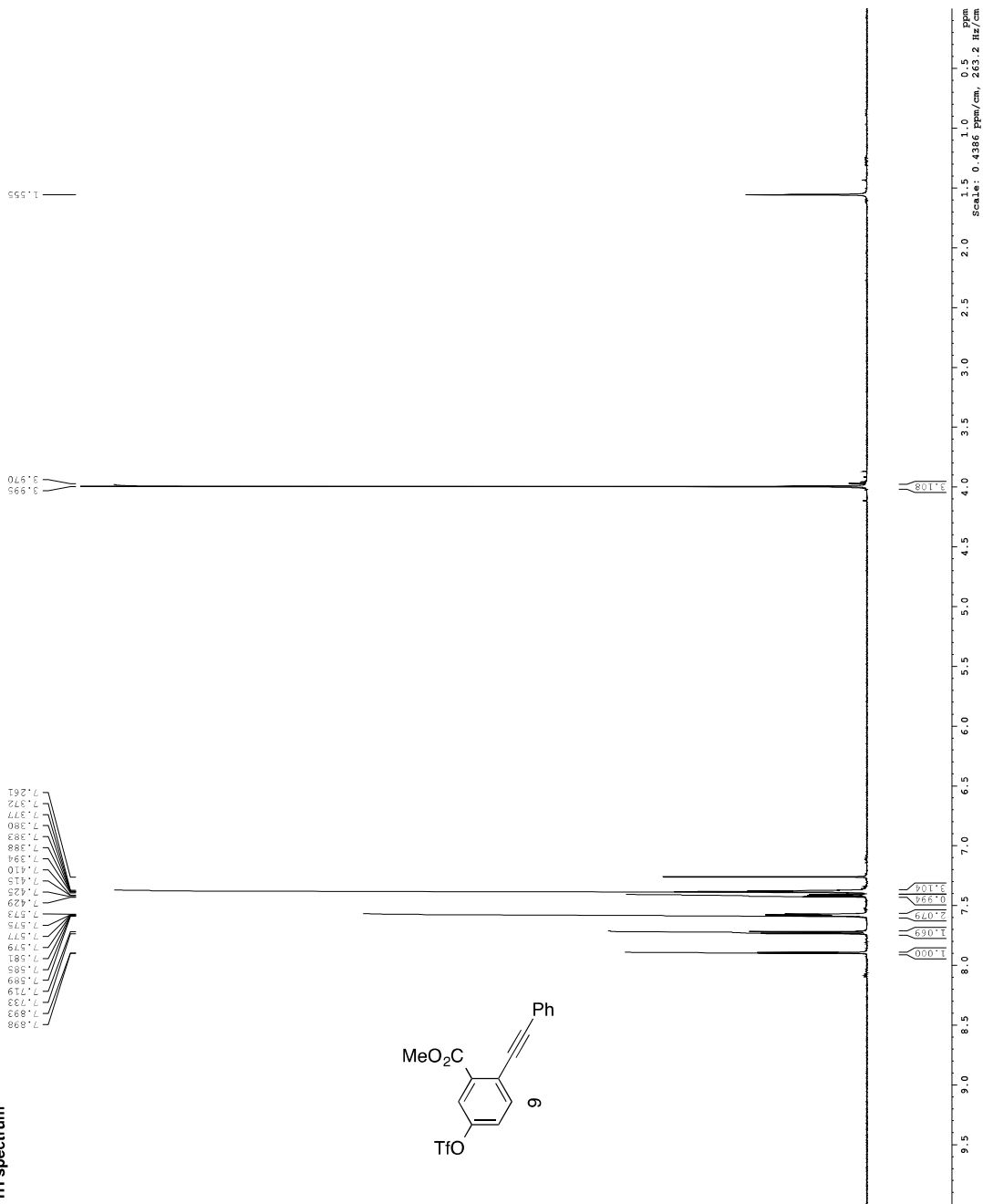


Current Data Parameters  
NAME 550201\_3  
PROCNO 1  
F2 - Acquisition Parameters  
Time 20 11 22  
INSTRUM spect  
PULPROG zgpg30  
SOLVENT CDCl3  
NS 2  
DS 2  
SWH 7130.269 Hz  
AQ 0.433544 sec  
RG 6.66 usec  
TE 300.0 K  
DE 1.0000000 sec  
===== CHANNEL f1 =====  
NUC1 19F  
P1 22.00 usec  
PL 0.00 dB  
RFQ 376.4651401 MHz  
F2 - Processing parameters  
SI 376.4651401 MHz  
WDW EM  
SSB 0  
GB 0  
PC 1.00



1H spectrum

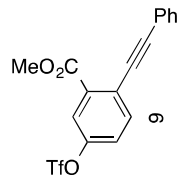
Current Data Parameters  
 Date\_ 20060503  
 Time\_ 11:29  
 F2 - Acquisition Parameters  
 INSTRUM spect  
 PULPROG zgpg30  
 SOLVENT dms-d6  
 NS 1280  
 DS 4  
 SWH 615.265 Hz  
 FWHM 0.850 Hz  
 AQ 0.0850 sec  
 RG 512.000 usec  
 TE 300.2 K  
 DE 1.00000000  
 L1 0.10000000 sec  
 L2  
 L3  
 ===== CHANNEL f1 =====  
 NUC1 1H  
 P1 12.00 usec  
 PL1 0.00 dB  
 SFO1 400.130000 MHz  
 F2 - Processing parameters  
 SI 32768  
 SF 400.130000 MHz  
 EQ 2  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



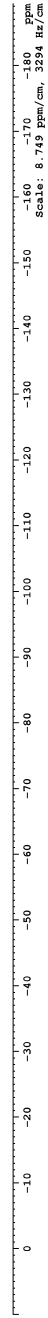


19F spectrum

Current Data Parameters  
NAME 55020\_3  
PROCNO 1  
F2 - Acquisition Parameters  
Time 2012.01  
INSTRUM spect  
PULPROG zgpg30  
SOLVENT CDCl3  
NS 1  
DS 2  
SWH 7530.209 Hz  
AQ 0.4335644 sec  
RG 6.676 usec  
TE 300.0 K sec  
DE 1.0000000 sec  
===== CHANNEL f1 =====  
NUC1 19F  
P1 22.50 usec  
PL 0.00 dB  
RFQ1 376.4651401 MHz  
F2 - Processing parameters  
SI 376.4651401 MHz  
WDW EM  
SSB 0  
GB 0  
PC 1.00



-72.654





1H spectrum

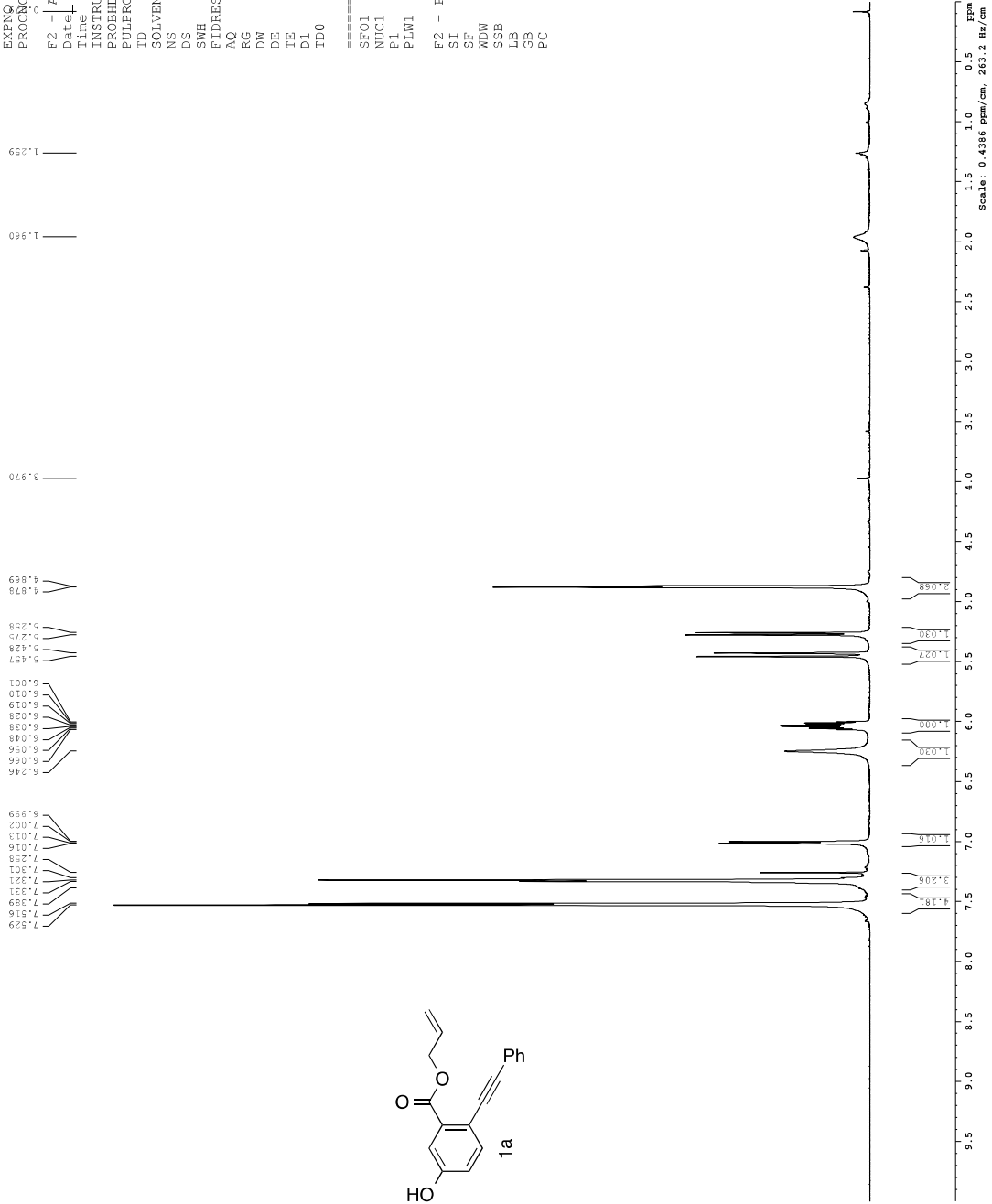
```

Current Data Parameters
NAME      JJ02136
EXPNO    1
PROCNO   1

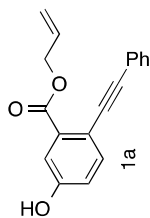
F2 - Acquisition Parameters
Date_    20140522
Time     11.07
INSTRUM  av600
PROBHD   5 mm TBI 1H/13
PULPROG  zg30
TD        98074
SOLVENT  CDCl3
NS        8
DS        2
SWH       9615.385 Hz
FIDRES    0.098042 Hz
AQ         5.0398478 sec
RG         406
DW         52.000 use
DE         14.54 use
TE         298.1 K
D1         0.1000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1     600.1342009 MHz
NUC1      1H
P1         8.00 use
PLW1     23.01441956 W

F2 - Processing Parameters
SI         65536
SF         600.1300355 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
  
```



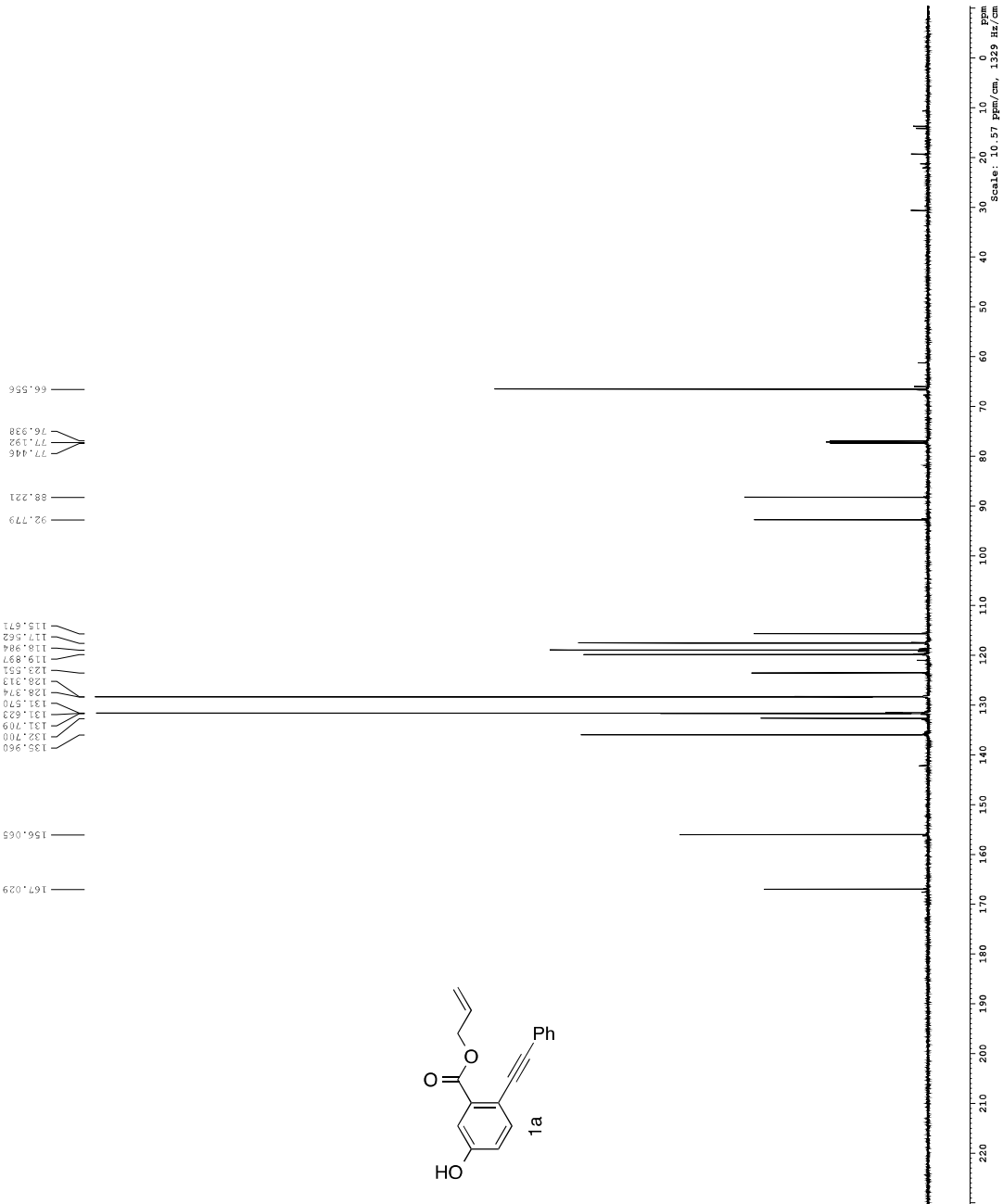
Z-restored spin-echo 13C spectrum with 1H decoupling



```

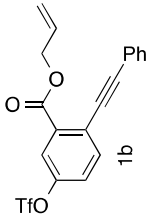
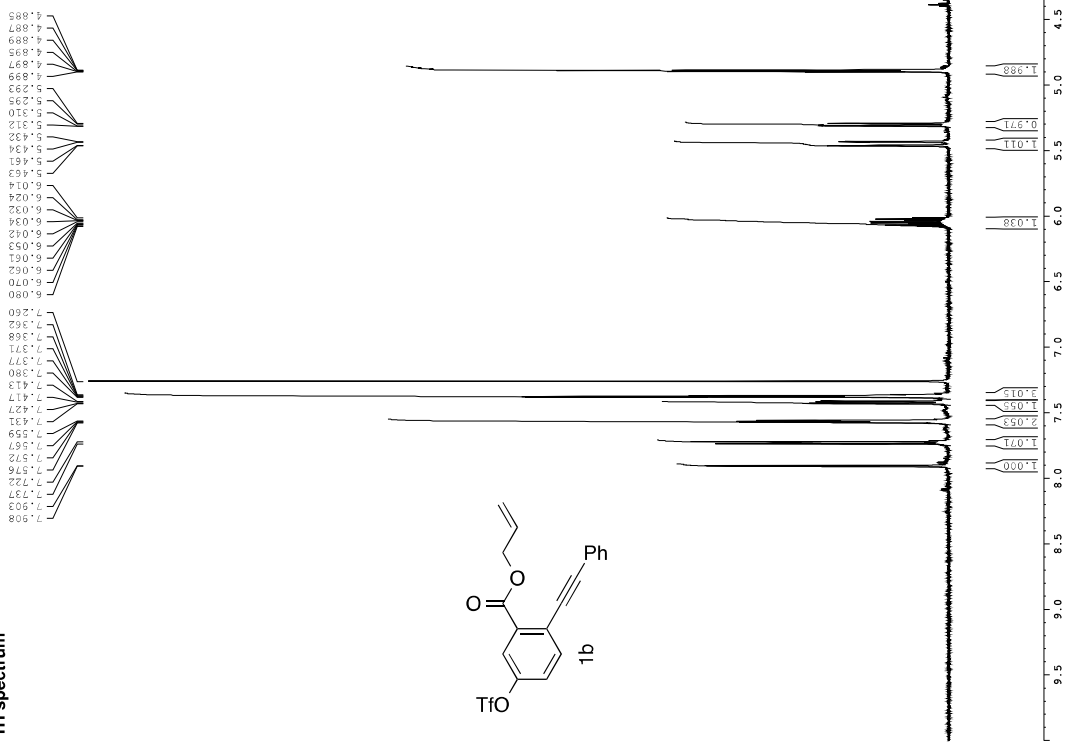
Current Data Parameters
NAME: j300c2
PROCNO: 1
F2 - Acquisition Parameters
Date_ 201203
Time_ 10:55
INSTRUM: spect
PROBHD: 5 mm CHX-10
PULPROG: zgpg30
SOLVENT: SpinelDMSO-d6
ACQNT: 600
AQ: 0.29000000 sec
RG: 3130
SFO: 125.760124 MHz
NUC1: 13C
NUC2: 1H
F1: 500.130498 MHz
F2: 125.760124 MHz
AQ: 0.29000000 sec
RG: 3130
SFO: 125.760124 MHz
SC: 1
SI: 32768
SR: 0.00012500 sec
RECEIVED: 0
PRT: 0.00012500 sec
DECODE: 1
CHUNK: 0
SI: 3130
===== CHANNEL f1 =====
NUC1: 13C
F1: 500.130498 MHz
P1: 12.00 dB
PL1: 0.00 dB
PL2: 0.00 dB
PL3: 0.00 dB
PL4: 0.00 dB
PL5: 0.00 dB
RF1: 125.760124 MHz
SC: 1
SFO: 125.760124 MHz
SSAMPL: 32768
SSAMPL: 65536
SWMODE: 2
SFRF: 0 Hz
SFRF2: 0 Hz
===== CHANNEL f2 =====
NAME: zgpg30
PROCNO: 1
F2: 125.760124 MHz
P2: 12.00 dB
PL2: 0.00 dB
PL3: 0.00 dB
PL4: 0.00 dB
PL5: 0.00 dB
RF2: 125.760124 MHz
SC: 1
SFO: 125.760124 MHz
SSAMPL: 65536
SWMODE: 2
SFRF: 0 Hz
SFRF2: 0 Hz
===== Processing parameters =====
SI: 32768
RG: 125.760124 MHz
AQ: 0.29000000 sec
SFO: 125.760124 MHz
SFRF: 0 Hz
SFRF2: 0 Hz

```



### <sup>1</sup>H spectrum

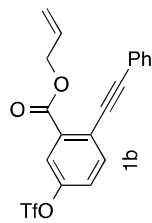
```
Current Data Parameters
=====
PROCNO      1
PROBHD      1
F2 - Acquisition Parameters
=====
Time_      2016_03_17_10:23
INSTRUM     spect
PROBHD      spect
PULPROG     zgpg30
SOLVENT     CDCl3
DS          2
SFO1        400.132009 MHz
AQ          0.10000000 sec
RG          327.5
TE          298.2 K
FIDRES      0.10000000 sec
===== CHANNEL f1 =====
NUC1        13C
P1          2.00000000 sec
PC          1.00
===== CHANNEL f2 =====
NUC2        13C
PC2         2.00000000 sec
===== CHANNEL f3 =====
NUC3        13C
PC3         2.00000000 sec
===== CHANNEL f4 =====
NUC4        13C
PC4         2.00000000 sec
F2 - Processing parameters
=====
SI          32768
SF          400.132009 MHz
WDW         EM
SSB         0
LB          0.30 Hz
GB          0
PC          1.00
```



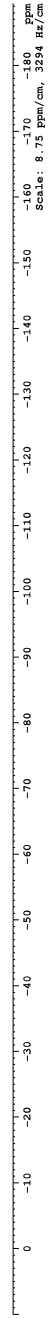


19F spectrum

Current Data Parameters  
NAME 550203  
PROCNO 1  
F2 - Acquisition Parameters  
Time 20 16.51  
INSTRUM spect  
PULPROG zgpg30  
SOLVENT CDCl3  
NS 1  
DS 2  
SWH 7530.269 Hz  
AQ 0.433564 sec  
RG 6.520 usec  
TE 300.0 K sec  
D1 2.0000000 sec  
===== CHANNEL f1 =====  
NUC1 19F  
P1 22.50 usec  
PL 0.00 dB  
RFQ 376.4651401 MHz  
F2 - Processing parameters  
SI 376.4651401 MHz  
WDW EM  
SSB 0  
GB 0  
PC 1.00

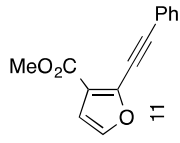
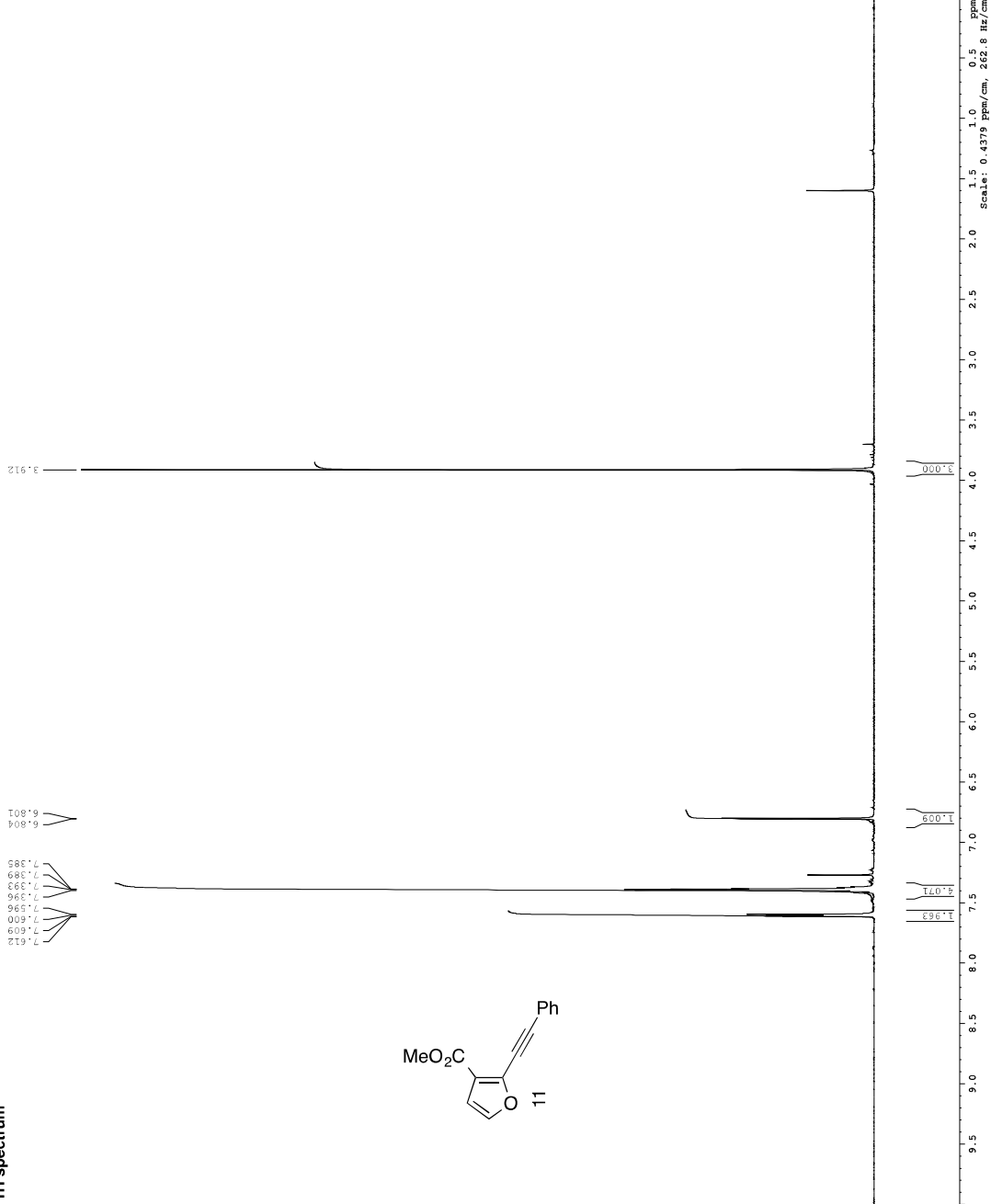


-72.655



1H spectrum

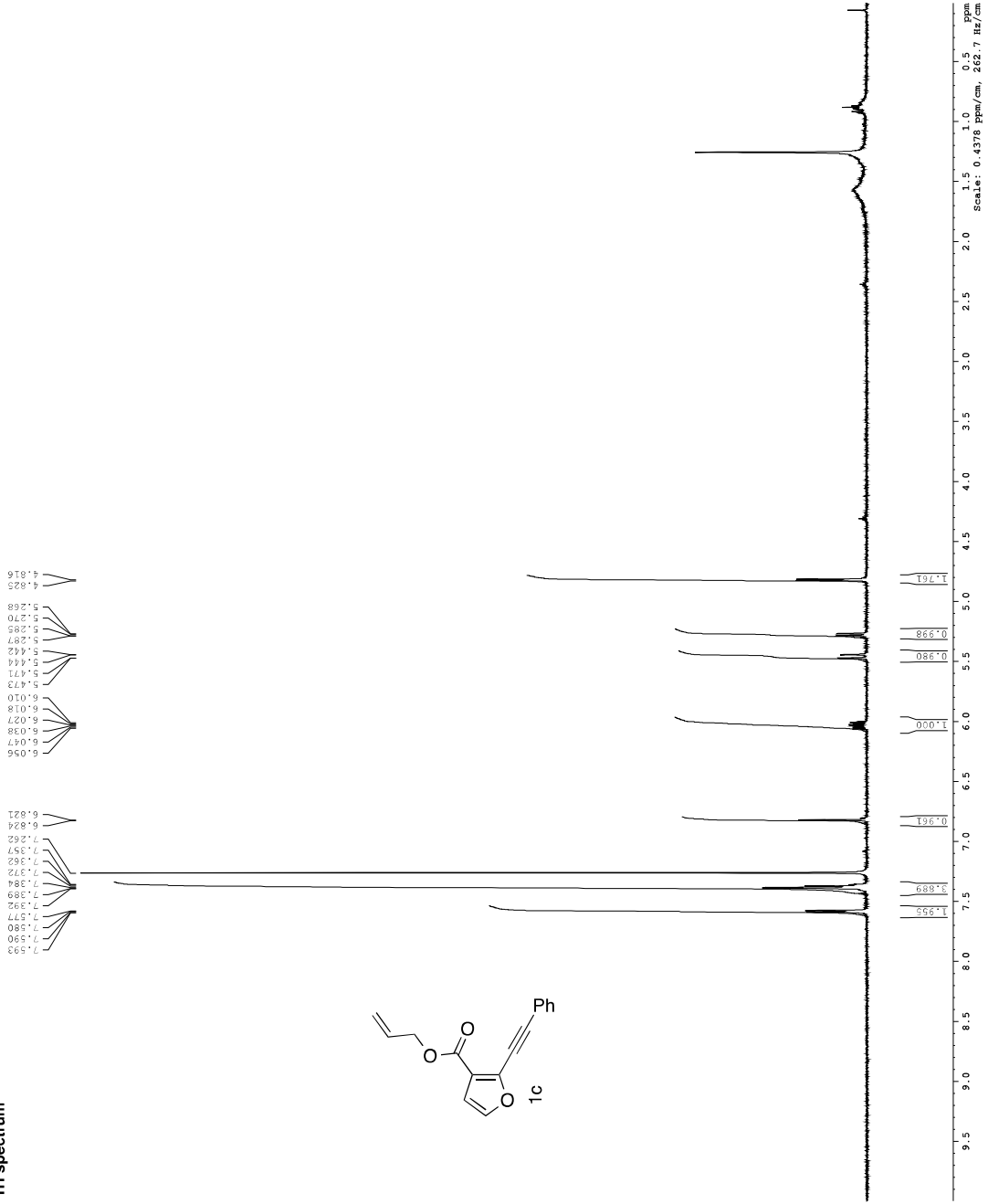
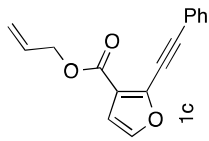
Current Data Parameters  
Date\_ 20060505  
Time\_ 11:28  
F2 - Acquisition Parameters  
Time 20.1428  
INSTRUM spect  
PROBHD 5 mm TBI BBO  
PULPROG zgpg30  
SOLVENT CDCl3  
NS 2  
DS 2  
SWH 615.265 Hz  
AQ 0.0850 sec  
RG 512.000 Hz  
FID 52.000 usec  
TE 300.2 K  
TE 298.1 usec  
L1 0.10000000 sec  
===== CHANNEL f1 =====  
NUC1 1H  
P1 12.00 usec  
PL1 0.00 dB  
ZG4 23.014126 usec  
SFO1 600.1310209 MHz  
F2 - Processing parameters  
SI 32768  
SF 600.1310209 MHz  
WDW EM  
SSB 2  
LB 0.30 Hz  
GB 0  
PC 1.00





1H spectrum

Current Data Parameters  
Date\_ 20060505  
Time\_ 11:02:00  
PROCNO 1  
F2 - Acquisition Parameters  
Time\_ 2012.28  
INSTRUM spect  
PROBHD 5 mm TBI 5MM  
PULPROG zgpg30  
SOLVENT CDCl3  
DS 4  
FID 320  
SFO1 601.330209 MHz  
AQ 5.0988279 sec  
RG 327.5  
TE 300.2 K  
D1 0.10000000 sec  
===== CHANNEL f1 =====  
NUC1 1H  
P1 12.00 nsec  
PL1 0.00 dB  
SFO1 601.330209 MHz  
F2 - Processing parameters  
SI 32768  
SF 600.1300346 MHz  
WDW EM  
SSB 2  
LB 0.30 Hz  
GB 0  
PC 1.00

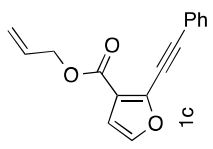
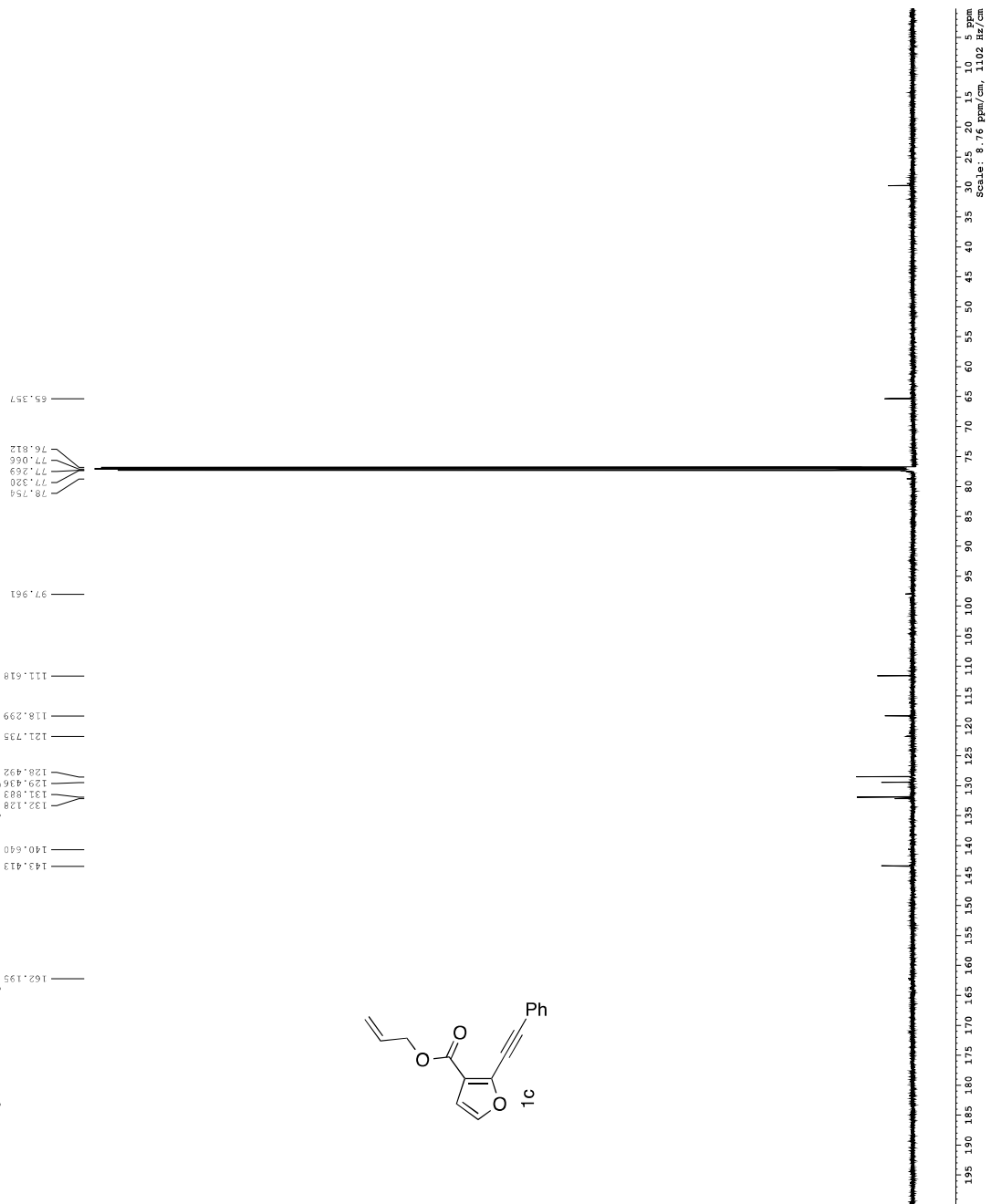




Z-restored spin-echo 13C spectrum with 1H decoupling

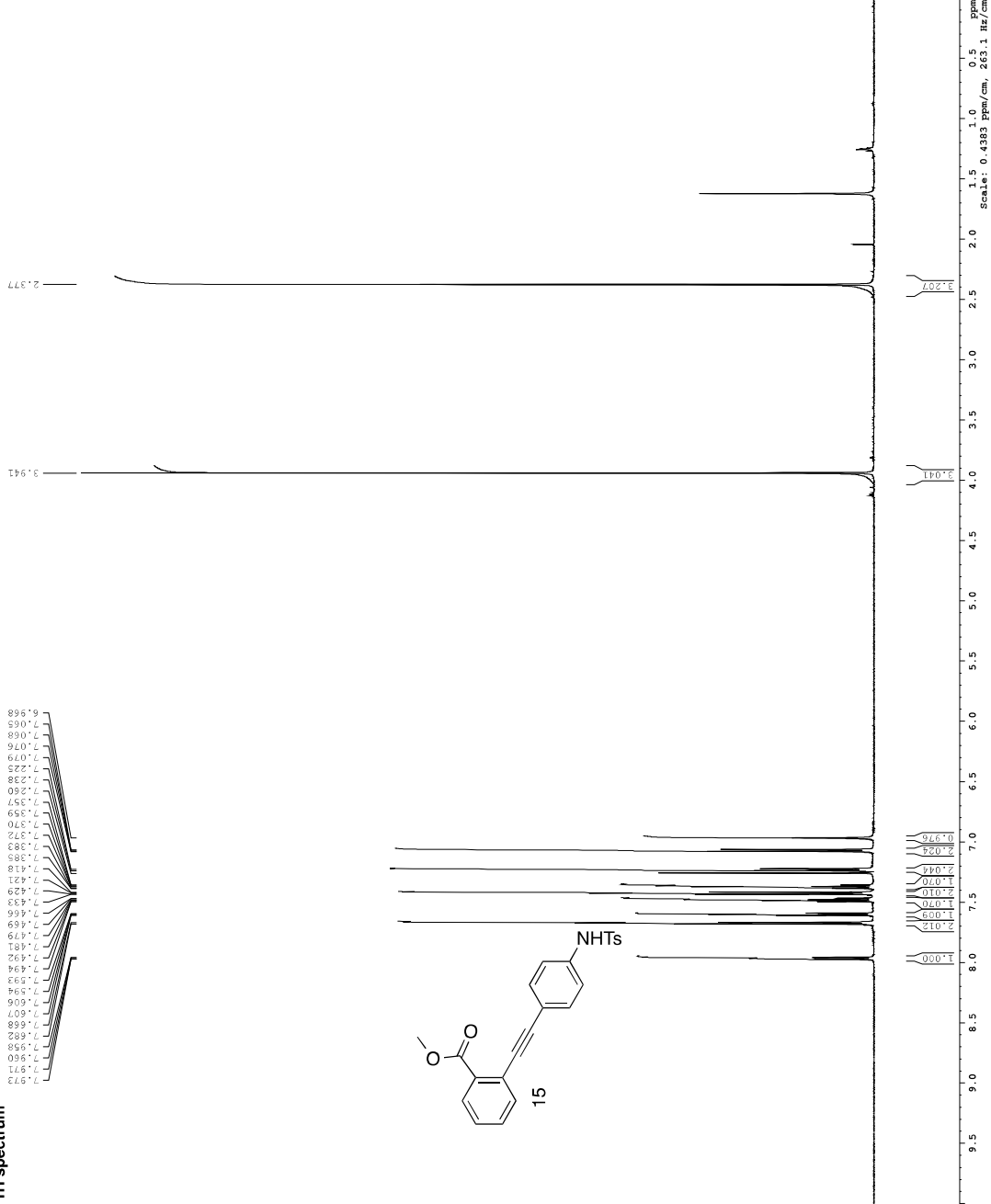
```

Current Data Parameters
=====
PROCNO 1
F2 - Acquisition Parameters
=====
Date_ 20110326
Time 0.23
INSTRUM spect
PROBHD 5 mm C13100
PULPROG zgpg30
SOLVENT CDCl3
NS 1987
DS 4
SWH 30396.031 Hz
AQ 0.0888 sec
RG 655.5
FIDRES 1.6813340 e-6
AQRES 0.00010000000000000000
TE 300.2 K
DE 2.00000000000000000000
L1 0.00000000000000000000
L2 0.00000000000000000000
L3 0.00000000000000000000
RGRES 3 Hz
SFO 101.25361 MHz
===== CHANNEL f1 =====
NUC1 13C
P1 1.32 usec
PL1 0 dB
PL12 0 dB
PL13 0 dB
PL14 0 dB
PL15 0 dB
PL16 0 dB
PL17 0 dB
PL18 0 dB
PL19 0 dB
PL20 0 dB
PL21 0 dB
PL22 0 dB
PL23 0 dB
PL24 0 dB
PL25 0 dB
PL26 0 dB
PL27 0 dB
PL28 0 dB
PL29 0 dB
PL30 0 dB
PL31 0 dB
PL32 0 dB
PL33 0 dB
PL34 0 dB
PL35 0 dB
PL36 0 dB
PL37 0 dB
PL38 0 dB
PL39 0 dB
PL40 0 dB
PL41 0 dB
PL42 0 dB
PL43 0 dB
PL44 0 dB
PL45 0 dB
PL46 0 dB
PL47 0 dB
PL48 0 dB
PL49 0 dB
PL50 0 dB
PL51 0 dB
PL52 0 dB
PL53 0 dB
PL54 0 dB
PL55 0 dB
PL56 0 dB
PL57 0 dB
PL58 0 dB
PL59 0 dB
PL60 0 dB
PL61 0 dB
PL62 0 dB
PL63 0 dB
PL64 0 dB
PL65 0 dB
PL66 0 dB
PL67 0 dB
PL68 0 dB
PL69 0 dB
PL70 0 dB
PL71 0 dB
PL72 0 dB
PL73 0 dB
PL74 0 dB
PL75 0 dB
PL76 0 dB
PL77 0 dB
PL78 0 dB
PL79 0 dB
PL80 0 dB
PL81 0 dB
PL82 0 dB
PL83 0 dB
PL84 0 dB
PL85 0 dB
PL86 0 dB
PL87 0 dB
PL88 0 dB
PL89 0 dB
PL90 0 dB
PL91 0 dB
PL92 0 dB
PL93 0 dB
PL94 0 dB
PL95 0 dB
PL96 0 dB
PL97 0 dB
PL98 0 dB
PL99 0 dB
PL100 0 dB
===== CHANNEL f2 =====
NUC2 13C
P2 1.32 usec
PL2 0 dB
PL23 0 dB
PL24 0 dB
PL25 0 dB
PL26 0 dB
PL27 0 dB
PL28 0 dB
PL29 0 dB
PL30 0 dB
PL31 0 dB
PL32 0 dB
PL33 0 dB
PL34 0 dB
PL35 0 dB
PL36 0 dB
PL37 0 dB
PL38 0 dB
PL39 0 dB
PL40 0 dB
PL41 0 dB
PL42 0 dB
PL43 0 dB
PL44 0 dB
PL45 0 dB
PL46 0 dB
PL47 0 dB
PL48 0 dB
PL49 0 dB
PL50 0 dB
PL51 0 dB
PL52 0 dB
PL53 0 dB
PL54 0 dB
PL55 0 dB
PL56 0 dB
PL57 0 dB
PL58 0 dB
PL59 0 dB
PL60 0 dB
PL61 0 dB
PL62 0 dB
PL63 0 dB
PL64 0 dB
PL65 0 dB
PL66 0 dB
PL67 0 dB
PL68 0 dB
PL69 0 dB
PL70 0 dB
PL71 0 dB
PL72 0 dB
PL73 0 dB
PL74 0 dB
PL75 0 dB
PL76 0 dB
PL77 0 dB
PL78 0 dB
PL79 0 dB
PL80 0 dB
PL81 0 dB
PL82 0 dB
PL83 0 dB
PL84 0 dB
PL85 0 dB
PL86 0 dB
PL87 0 dB
PL88 0 dB
PL89 0 dB
PL90 0 dB
PL91 0 dB
PL92 0 dB
PL93 0 dB
PL94 0 dB
PL95 0 dB
PL96 0 dB
PL97 0 dB
PL98 0 dB
PL99 0 dB
PL100 0 dB
===== GRAB =====
GRAB 0
===== GRADIENT =====
G1 0.00 usec
G2 0.00 usec
G3 0.00 usec
G4 0.00 usec
G5 0.00 usec
G6 0.00 usec
G7 0.00 usec
G8 0.00 usec
G9 0.00 usec
G10 0.00 usec
G11 0.00 usec
G12 0.00 usec
G13 0.00 usec
G14 0.00 usec
G15 0.00 usec
G16 0.00 usec
G17 0.00 usec
G18 0.00 usec
G19 0.00 usec
G20 0.00 usec
G21 0.00 usec
G22 0.00 usec
G23 0.00 usec
G24 0.00 usec
G25 0.00 usec
G26 0.00 usec
G27 0.00 usec
G28 0.00 usec
G29 0.00 usec
G30 0.00 usec
G31 0.00 usec
G32 0.00 usec
G33 0.00 usec
G34 0.00 usec
G35 0.00 usec
G36 0.00 usec
G37 0.00 usec
G38 0.00 usec
G39 0.00 usec
G40 0.00 usec
G41 0.00 usec
G42 0.00 usec
G43 0.00 usec
G44 0.00 usec
G45 0.00 usec
G46 0.00 usec
G47 0.00 usec
G48 0.00 usec
G49 0.00 usec
G50 0.00 usec
G51 0.00 usec
G52 0.00 usec
G53 0.00 usec
G54 0.00 usec
G55 0.00 usec
G56 0.00 usec
G57 0.00 usec
G58 0.00 usec
G59 0.00 usec
G60 0.00 usec
G61 0.00 usec
G62 0.00 usec
G63 0.00 usec
G64 0.00 usec
G65 0.00 usec
G66 0.00 usec
G67 0.00 usec
G68 0.00 usec
G69 0.00 usec
G70 0.00 usec
G71 0.00 usec
G72 0.00 usec
G73 0.00 usec
G74 0.00 usec
G75 0.00 usec
G76 0.00 usec
G77 0.00 usec
G78 0.00 usec
G79 0.00 usec
G80 0.00 usec
G81 0.00 usec
G82 0.00 usec
G83 0.00 usec
G84 0.00 usec
G85 0.00 usec
G86 0.00 usec
G87 0.00 usec
G88 0.00 usec
G89 0.00 usec
G90 0.00 usec
G91 0.00 usec
G92 0.00 usec
G93 0.00 usec
G94 0.00 usec
G95 0.00 usec
G96 0.00 usec
G97 0.00 usec
G98 0.00 usec
G99 0.00 usec
G100 0.00 usec
===== Processing Parameters =====
SI 32768
SF 125.7601152 MHz
WDW EM
SSB 0
GB 0
PC 1.00 Hz
=====
  
```



**1H spectrum**

Current Data Parameters  
 Name: 550202  
 PROCNO: 1  
 F2 - Acquisition Parameters  
 Date\_ Time: 2011.07.11.47  
 INSTRUM: spect  
 PULPROG: zgpg30  
 SOLVENT: CDCl3  
 NS: 128  
 DS: 4  
 SWH: 615.265 Hz  
 AQ: 0.18859 sec  
 FIDRES: 5.098879 e-6  
 RG: 52.000 usec  
 TE: 300.2 K  
 DE: 1.00000000  
 L1: 0.10000000 sec  
 ===== CHANNEL f1 =====  
 NUC1: 1H  
 P1: 12.00 usec  
 PL1: 0.00 dB  
 ZG4: 23.014126 usec  
 SFO1: 600.1310209 MHz  
 F2 - Processing parameters  
 SI: 32768  
 SF: 600.1310209 MHz  
 EQ2: EX  
 AS: 2  
 LB: 0.30 Hz  
 GB: 0  
 SC: 1.00



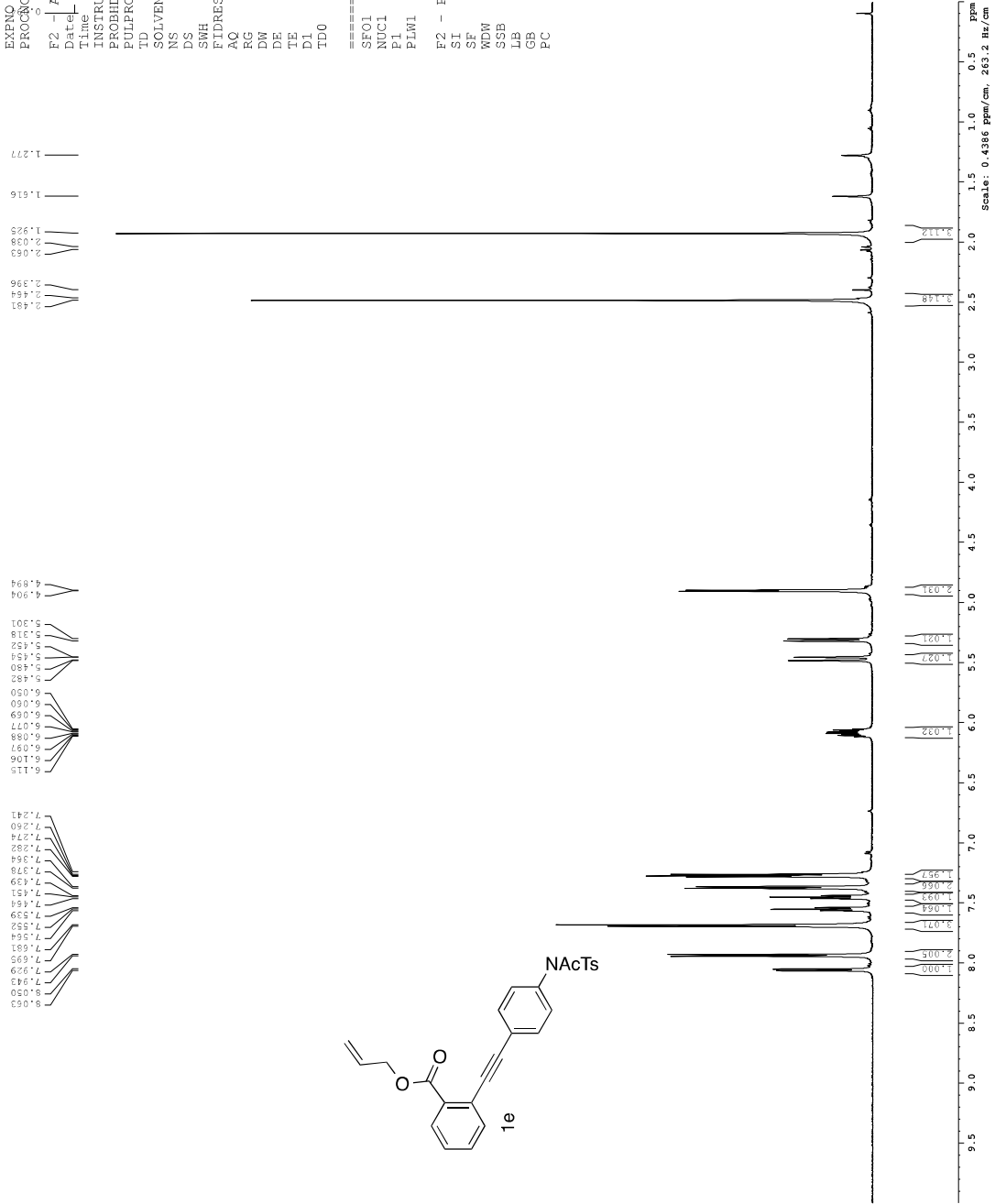






1H spectrum

Current Data Parameters  
 NAME JJ02145  
 EXPNO 3  
 PROCNO 1  
 F2 - Acquisition Parameters  
 Date\_ 20140617  
 Time 7.27  
 INSTRUM av600  
 PROBHD 5 mm TBI 1H/13  
 PULPROG zg30  
 TD 98074  
 SOLVENT CDCl3  
 NS 8  
 DS 2  
 SWH 9615.385 Hz  
 FIDRES 0.098042 Hz  
 AQ 5.0398478 sec  
 RG 724  
 DW 52.000 use  
 DE 14.54 use  
 TE 298.0 K  
 D1 0.1000000 sec  
 TD0 1  
 ===== CHANNEL f1 =====  
 SF01 600.1342009 MHz  
 NUC1 1H  
 P1 8.00 use  
 PLW1 23.01441956 W  
 F2 - Processing Parameters  
 SI 65536  
 SF 600.1300211 MHz  
 WDW EM  
 SSB 0  
 LB 0  
 GB 0  
 PC 1.00





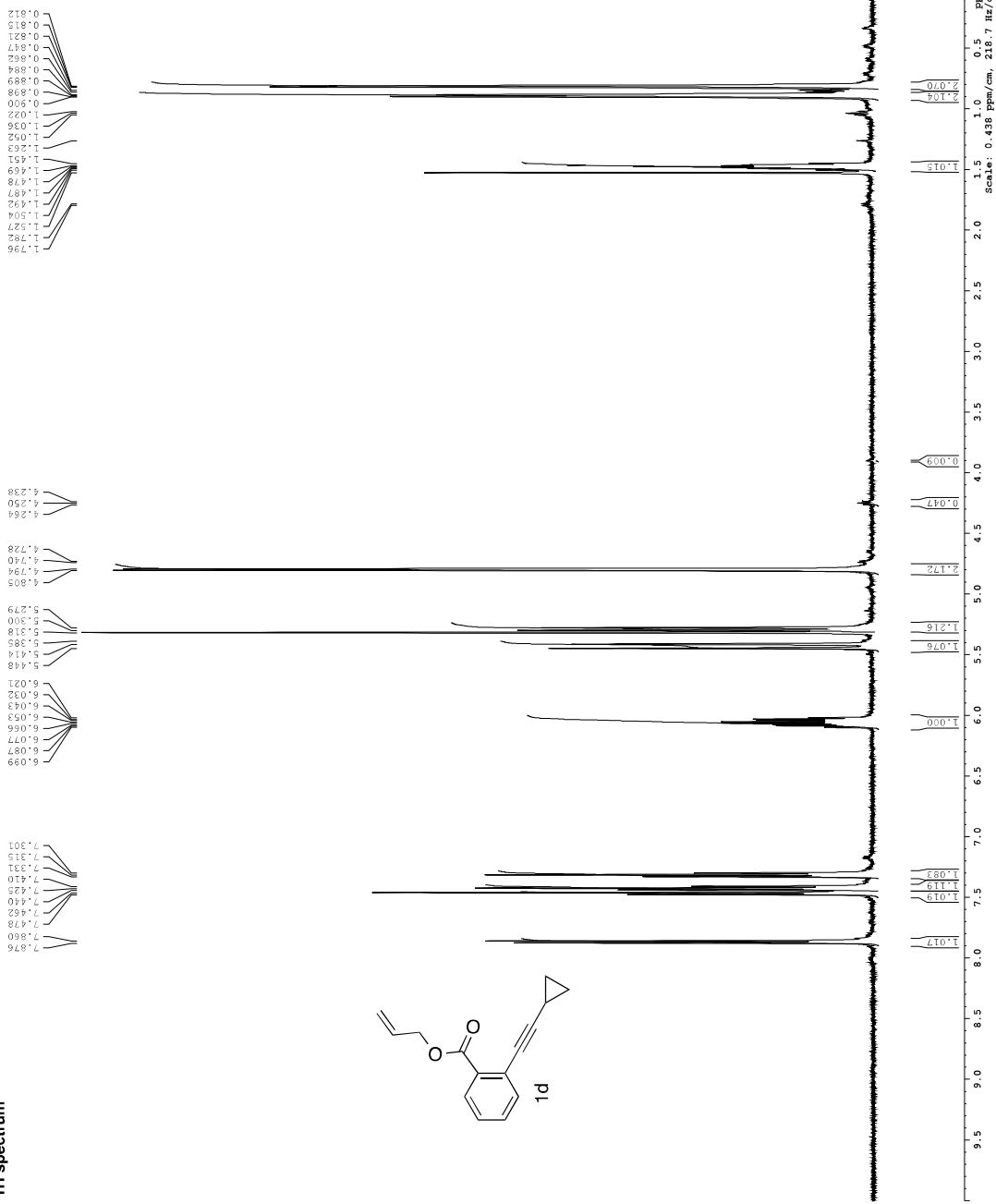






**<sup>1</sup>H spectrum**

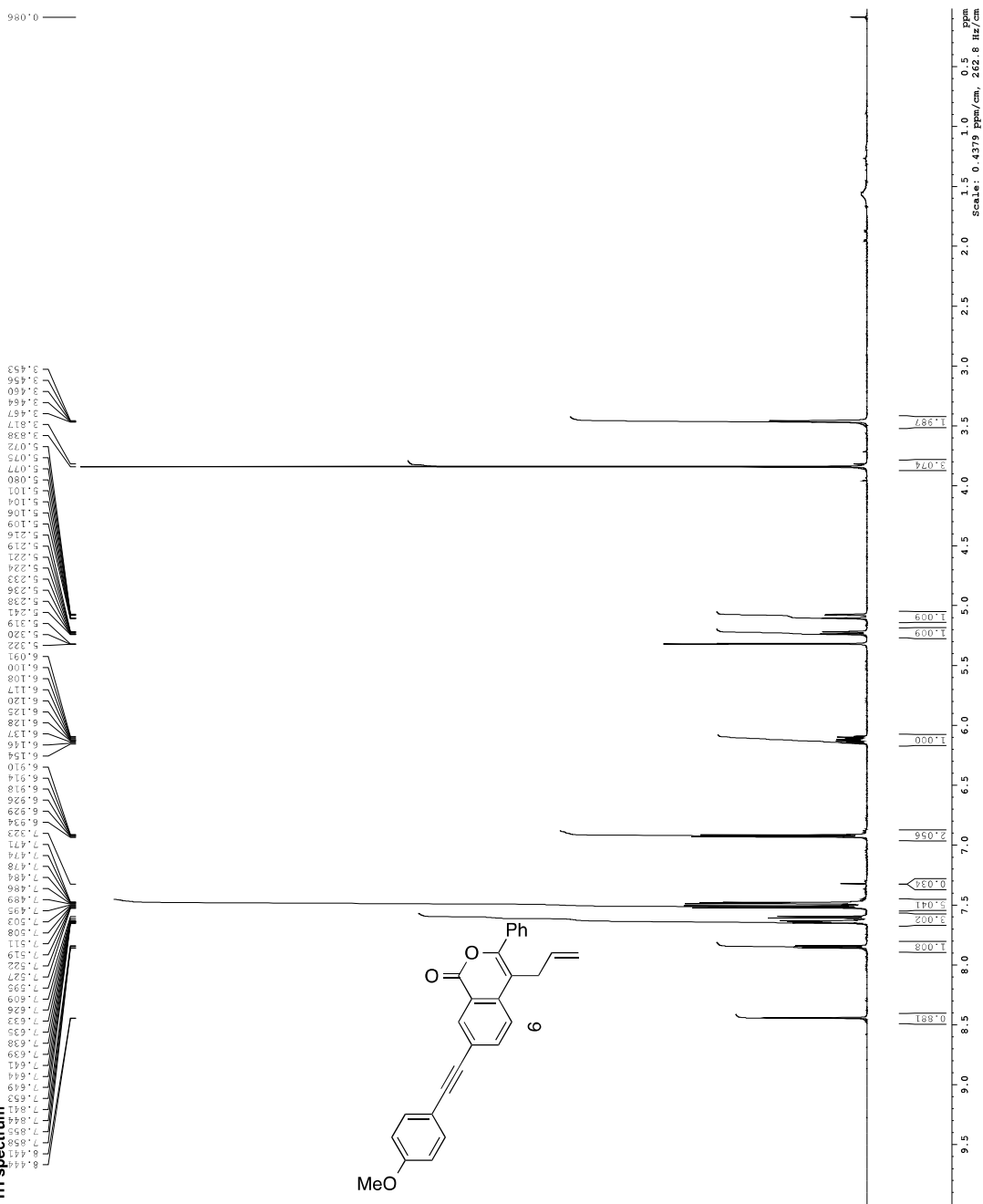
Current Data Parameters  
 Date\_ 13/12  
 Time\_ 1  
 F2 - Acquisition Parameters  
 Date\_ 2013-09-19  
 Time\_ 9:59  
 INSTRUM spect  
 PULPROG zgpg30  
 FIDRES 0.000000  
 AQ 4.230  
 SOLVENT CDCl3  
 NS 2  
 DS 2  
 SWH 6013.820 Hz  
 FWHM 0.1500 Hz  
 AQ 0.000000 sec  
 SFO 5.098374 sec  
 RG 655.400 usec  
 TD 65536  
 TE 298.0 K usec  
 DE 0.000000 sec  
 ACQUIS 0 sec  
 MD 0.01300000 sec  
 ===== CHANNEL f1 =====  
 NU1 12.00 usec  
 PL1 0.00 dB  
 PR1 489.294450 MHz  
 =====  
 SI - Processing Parameters  
 SF 499.822361 MHz  
 RG 655.400 usec  
 DE 0.000000 sec  
 AS 0.000000 sec  
 GB 0.00 Hz  
 GC 0.00 Hz  
 SC 1.00





Current Data Parameters  
 20000 1312  
 F2 - Acquisition Parameters  
 Time 20.145  
 INSTRUM spect  
 PULPROG zgpg30  
 SOLVENT CDCl3  
 NS 2  
 DS 2  
 SWH 615.265 Hz  
 AQ 5.098879 sec  
 RG 52.000 usec  
 TE 300.0 K  
 DE 1.000 usec  
 LL 0.10000000 sec  
 Channel f1 1H  
 NUC1 1H  
 ZG4 23.044156 usec  
 SFO1 600.1320209 MHz  
 F2 - Processing parameters  
 SF 600.132025 MHz  
 WW 6000  
 EQ  
 SFO 600.132025 MHz  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

1H spectrum



Current Data Parameters  
 Job: 0114  
 PROCNO: 1

==== Acquisition Parameters  
 Date\_ Time: 2014.14.10  
 INSTRUM: spect  
 F1FREQ: 125.7603384 MHz  
 PULPROG: zgpg30  
 SOLVENT: CDCl3  
 DS: 4  
 SWH: 30399.931 Hz  
 AQ: 0.4081836 sec  
 F2 - Acquisition Parameters  
 Date\_ Time: 2014.14.10  
 INSTRUM: spect  
 F2FREQ: 500.136099 MHz  
 PULPROG: zgpg30  
 SOLVENT: CDCl3  
 DS: 4  
 SWH: 30399.931 Hz  
 AQ: 0.4081836 sec  
 F2 - Acquisition Parameters  
 Date\_ Time: 2014.14.10  
 INSTRUM: spect  
 F2FREQ: 500.136099 MHz  
 PULPROG: zgpg30  
 SOLVENT: CDCl3  
 DS: 4  
 SWH: 30399.931 Hz  
 AQ: 0.4081836 sec  
 F2 - Acquisition Parameters  
 Date\_ Time: 2014.14.10  
 INSTRUM: spect  
 F2FREQ: 500.136099 MHz  
 PULPROG: zgpg30  
 SOLVENT: CDCl3  
 DS: 4  
 SWH: 30399.931 Hz  
 AQ: 0.4081836 sec

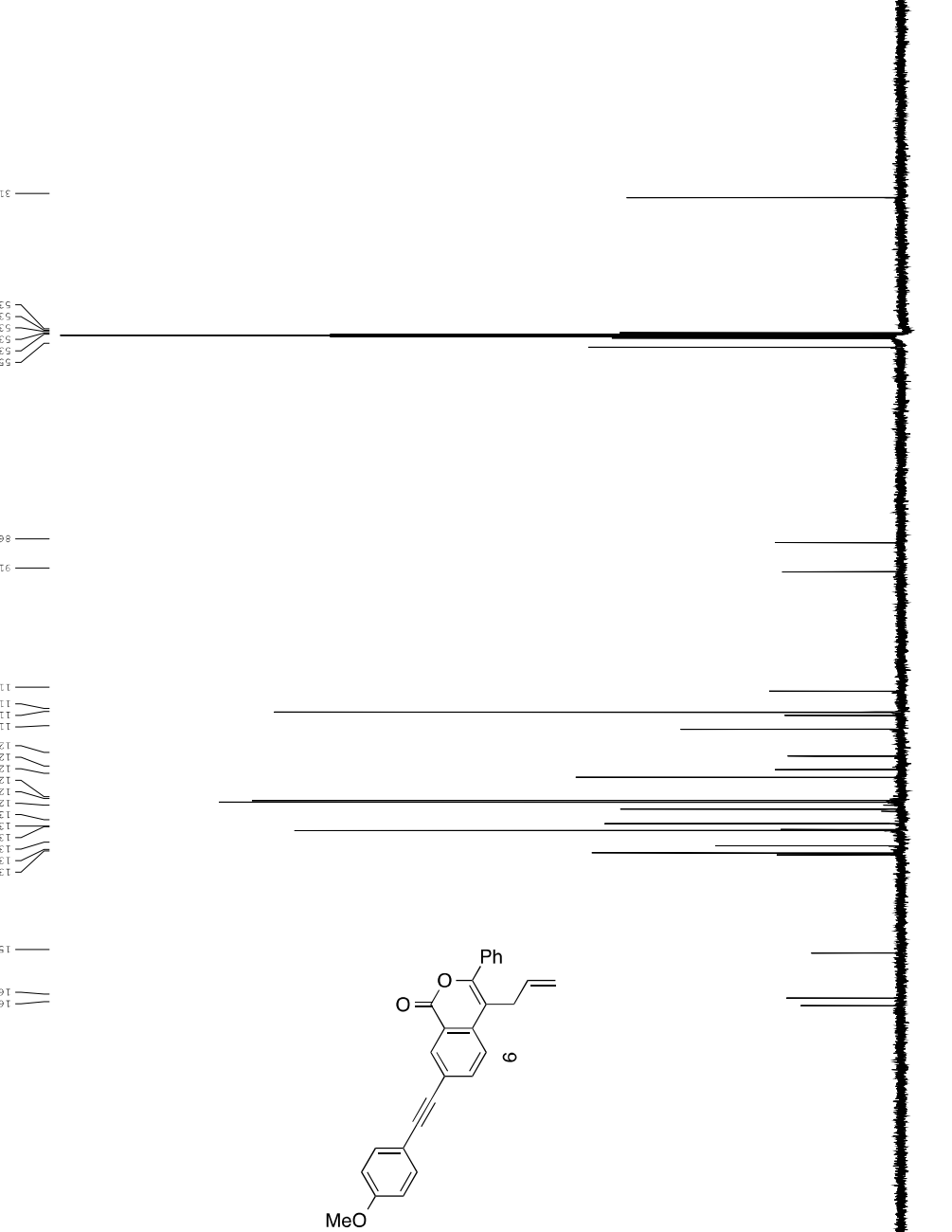
==== CHANNEL f1 =====  
 NUCL1: <sup>13</sup>C  
 P1: 13.00 usec  
 PL1: 0.00 dB  
 FWHM: 320.000 Hz  
 SFO1: 125.7603384 MHz  
 SF01A: 500.136099 MHz  
 SF01B: 500.136099 MHz  
 SF01C: 500.136099 MHz  
 SF01D: 500.136099 MHz  
 SF01E: 500.136099 MHz  
 SF01F: 500.136099 MHz  
 SF01G: 500.136099 MHz  
 SF01H: 500.136099 MHz  
 SF01I: 500.136099 MHz  
 SF01J: 500.136099 MHz  
 SF01K: 500.136099 MHz  
 SF01L: 500.136099 MHz  
 SF01M: 500.136099 MHz  
 SF01N: 500.136099 MHz  
 SF01O: 500.136099 MHz  
 SF01P: 500.136099 MHz  
 SF01Q: 500.136099 MHz  
 SF01R: 500.136099 MHz  
 SF01S: 500.136099 MHz  
 SF01T: 500.136099 MHz  
 SF01U: 500.136099 MHz  
 SF01V: 500.136099 MHz  
 SF01W: 500.136099 MHz  
 SF01X: 500.136099 MHz  
 SF01Y: 500.136099 MHz  
 SF01Z: 500.136099 MHz  
 =====

==== CHANNEL f2 =====  
 NUCL2: <sup>13</sup>C  
 P2: 13.00 usec  
 PL2: 0.00 dB  
 FWHM: 320.000 Hz  
 SFO2: 125.7603384 MHz  
 SF02A: 500.136099 MHz  
 SF02B: 500.136099 MHz  
 SF02C: 500.136099 MHz  
 SF02D: 500.136099 MHz  
 SF02E: 500.136099 MHz  
 SF02F: 500.136099 MHz  
 SF02G: 500.136099 MHz  
 SF02H: 500.136099 MHz  
 SF02I: 500.136099 MHz  
 SF02J: 500.136099 MHz  
 SF02K: 500.136099 MHz  
 SF02L: 500.136099 MHz  
 SF02M: 500.136099 MHz  
 SF02N: 500.136099 MHz  
 SF02O: 500.136099 MHz  
 SF02P: 500.136099 MHz  
 SF02Q: 500.136099 MHz  
 SF02R: 500.136099 MHz  
 SF02S: 500.136099 MHz  
 SF02T: 500.136099 MHz  
 SF02U: 500.136099 MHz  
 SF02V: 500.136099 MHz  
 SF02W: 500.136099 MHz  
 SF02X: 500.136099 MHz  
 SF02Y: 500.136099 MHz  
 SF02Z: 500.136099 MHz  
 =====

==== CHANNEL f3 =====  
 NUCL3: <sup>13</sup>C  
 P3: 13.00 usec  
 PL3: 0.00 dB  
 FWHM: 320.000 Hz  
 SFO3: 125.7603384 MHz  
 SF03A: 500.136099 MHz  
 SF03B: 500.136099 MHz  
 SF03C: 500.136099 MHz  
 SF03D: 500.136099 MHz  
 SF03E: 500.136099 MHz  
 SF03F: 500.136099 MHz  
 SF03G: 500.136099 MHz  
 SF03H: 500.136099 MHz  
 SF03I: 500.136099 MHz  
 SF03J: 500.136099 MHz  
 SF03K: 500.136099 MHz  
 SF03L: 500.136099 MHz  
 SF03M: 500.136099 MHz  
 SF03N: 500.136099 MHz  
 SF03O: 500.136099 MHz  
 SF03P: 500.136099 MHz  
 SF03Q: 500.136099 MHz  
 SF03R: 500.136099 MHz  
 SF03S: 500.136099 MHz  
 SF03T: 500.136099 MHz  
 SF03U: 500.136099 MHz  
 SF03V: 500.136099 MHz  
 SF03W: 500.136099 MHz  
 SF03X: 500.136099 MHz  
 SF03Y: 500.136099 MHz  
 SF03Z: 500.136099 MHz  
 =====

==== CHANNEL f4 =====  
 NUCL4: <sup>13</sup>C  
 P4: 13.00 usec  
 PL4: 0.00 dB  
 FWHM: 320.000 Hz  
 SFO4: 125.7603384 MHz  
 SF04A: 500.136099 MHz  
 SF04B: 500.136099 MHz  
 SF04C: 500.136099 MHz  
 SF04D: 500.136099 MHz  
 SF04E: 500.136099 MHz  
 SF04F: 500.136099 MHz  
 SF04G: 500.136099 MHz  
 SF04H: 500.136099 MHz  
 SF04I: 500.136099 MHz  
 SF04J: 500.136099 MHz  
 SF04K: 500.136099 MHz  
 SF04L: 500.136099 MHz  
 SF04M: 500.136099 MHz  
 SF04N: 500.136099 MHz  
 SF04O: 500.136099 MHz  
 SF04P: 500.136099 MHz  
 SF04Q: 500.136099 MHz  
 SF04R: 500.136099 MHz  
 SF04S: 500.136099 MHz  
 SF04T: 500.136099 MHz  
 SF04U: 500.136099 MHz  
 SF04V: 500.136099 MHz  
 SF04W: 500.136099 MHz  
 SF04X: 500.136099 MHz  
 SF04Y: 500.136099 MHz  
 SF04Z: 500.136099 MHz  
 =====

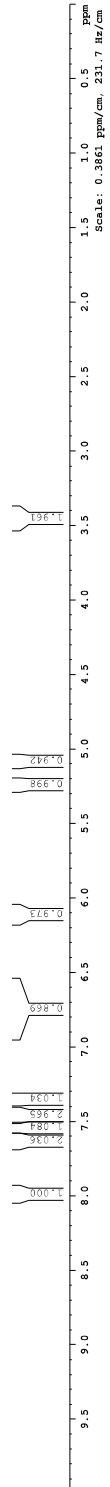
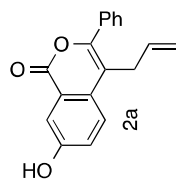
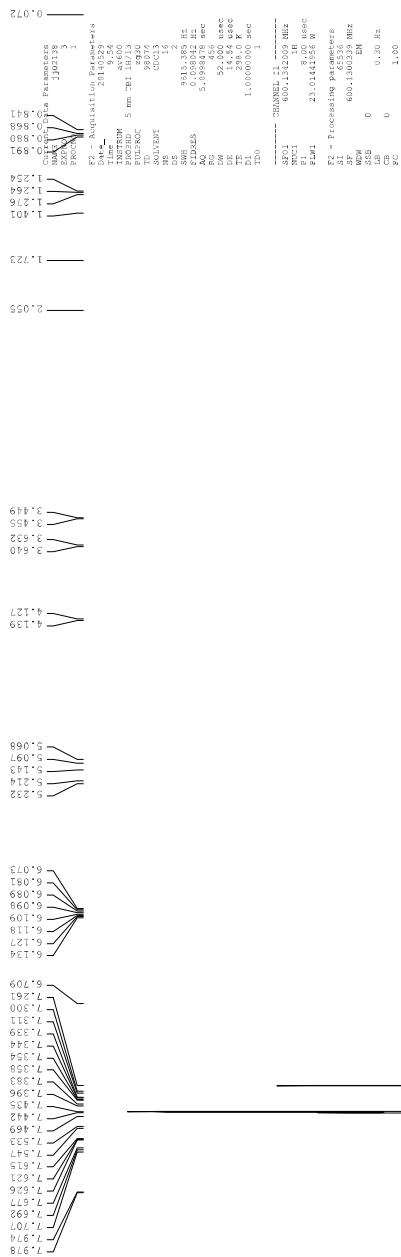
==== CHANNEL f5 =====  
 NUCL5: <sup>13</sup>C  
 P5: 13.00 usec  
 PL5: 0.00 dB  
 FWHM: 320.000 Hz  
 SFO5: 125.7603384 MHz  
 SF05A: 500.136099 MHz  
 SF05B: 500.136099 MHz  
 SF05C: 500.136099 MHz  
 SF05D: 500.136099 MHz  
 SF05E: 500.136099 MHz  
 SF05F: 500.136099 MHz  
 SF05G: 500.136099 MHz  
 SF05H: 500.136099 MHz  
 SF05I: 500.136099 MHz  
 SF05J: 500.136099 MHz  
 SF05K: 500.136099 MHz  
 SF05L: 500.136099 MHz  
 SF05M: 500.136099 MHz  
 SF05N: 500.136099 MHz  
 SF05O: 500.136099 MHz  
 SF05P: 500.136099 MHz  
 SF05Q: 500.136099 MHz  
 SF05R: 500.136099 MHz  
 SF05S: 500.136099 MHz  
 SF05T: 500.136099 MHz  
 SF05U: 500.136099 MHz  
 SF05V: 500.136099 MHz  
 SF05W: 500.136099 MHz  
 SF05X: 500.136099 MHz  
 SF05Y: 500.136099 MHz  
 SF05Z: 500.136099 MHz  
 =====



137.110, 136.893, 135.879, 133.490, 133.056, 132.090, 129.746, 128.658, 128.362, 124.606, 121.229, 118.919, 114.626, 114.138, 110.747, 91.527, 86.817, 55.255, 53.693, 53.677, 53.461, 52.242, 51.028, 31.246

195 190 185 180 175 170 165 160 155 150 145 140 135 130 125 120 115 110 105 100 95 90 85 80 75 70 65 60 55 50 45 40 35 30 25 20 15 10 5 ppm  
 Scale: 8.777 PPM/cm, 1104 Hz/cm

1H spectrum







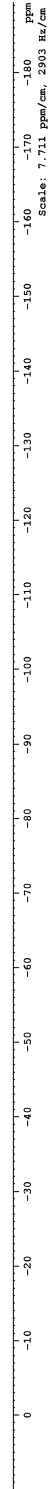
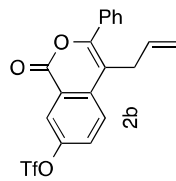




19F spectrum

-72.585

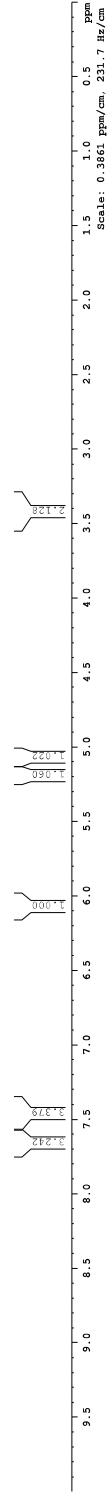
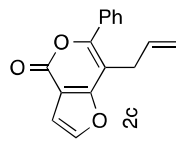
Current Data Parameters  
NAME J06212  
PROCNO 1  
F2 - Acquisition Parameters  
Time 20.11  
Time 11.54  
PULPROG zgpg30  
PROCNO 5  
QUP 0.0000  
F1-NUC 19F  
F2-NUC 19F  
SOLVENT CDCl3  
DS 2  
SBS 3187.500 Hz  
AQ 0.465  
RG 655.814 sec  
SR 6.165 usec  
TE 29.2 K  
D1 2.0000000 sec  
WDW1 HANNING  
SI 22.50 usec  
SF 376.486100 MHz  
F2 376.486100 MHz  
SI 655.814  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.00



Current Data Parameters  
NAME: j06114  
EXPNO: 1  
PROCNO: 1

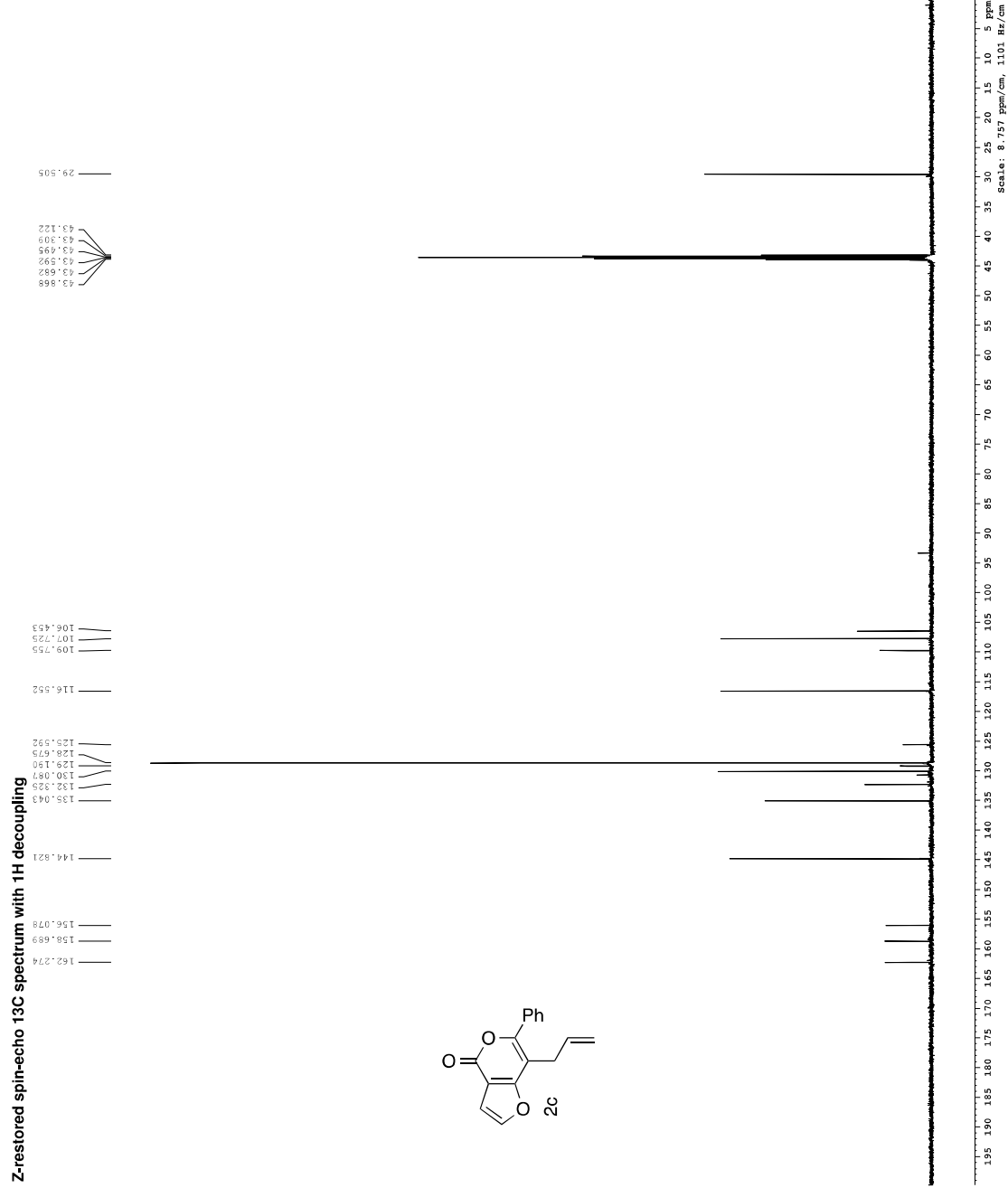
F2 - Acquisition Parameters  
Date\_ Acq: 20110124  
Time: 12.12  
INSTRUM: spect  
PROBHD: 5 mm QNP 1H/13  
PULPROG: zgpg30  
AQ: 0.08476 sec  
SOLVENT: CDCl3  
DS: 2  
SWH: 7910.388 Hz  
AQ: 0.08476 sec  
NUC1: 13C  
NUC2: 13C  
TE: 300.2 K  
D0: 0.11000000 sec  
SFO1: 125.761 MHz  
SFO2: 125.761 MHz  
SFO3: 600.1314209 MHz  
SFO4: 600.1314209 MHz  
RG: 300  
RG2: 300  
RG3: 300  
RG4: 23.0144155 g  
P2 - Processing parameters  
SI: 32768  
SF: 600.1308662 MHz  
WDW: EM  
SSB: 0  
LB: 0.30 Hz  
GB: 0  
RC: 1.00

7.651  
7.658  
7.668  
7.688  
7.587  
7.501  
7.413  
7.468  
7.464  
7.407  
7.123  
6.933  
6.910  
6.099  
6.090  
6.072  
6.082  
6.072  
6.053  
6.045  
6.036  
5.163  
5.146  
5.101  
5.072  
3.719  
3.719  
3.418  
3.418



Z-restored spin-echo 13C spectrum with 1H decoupling

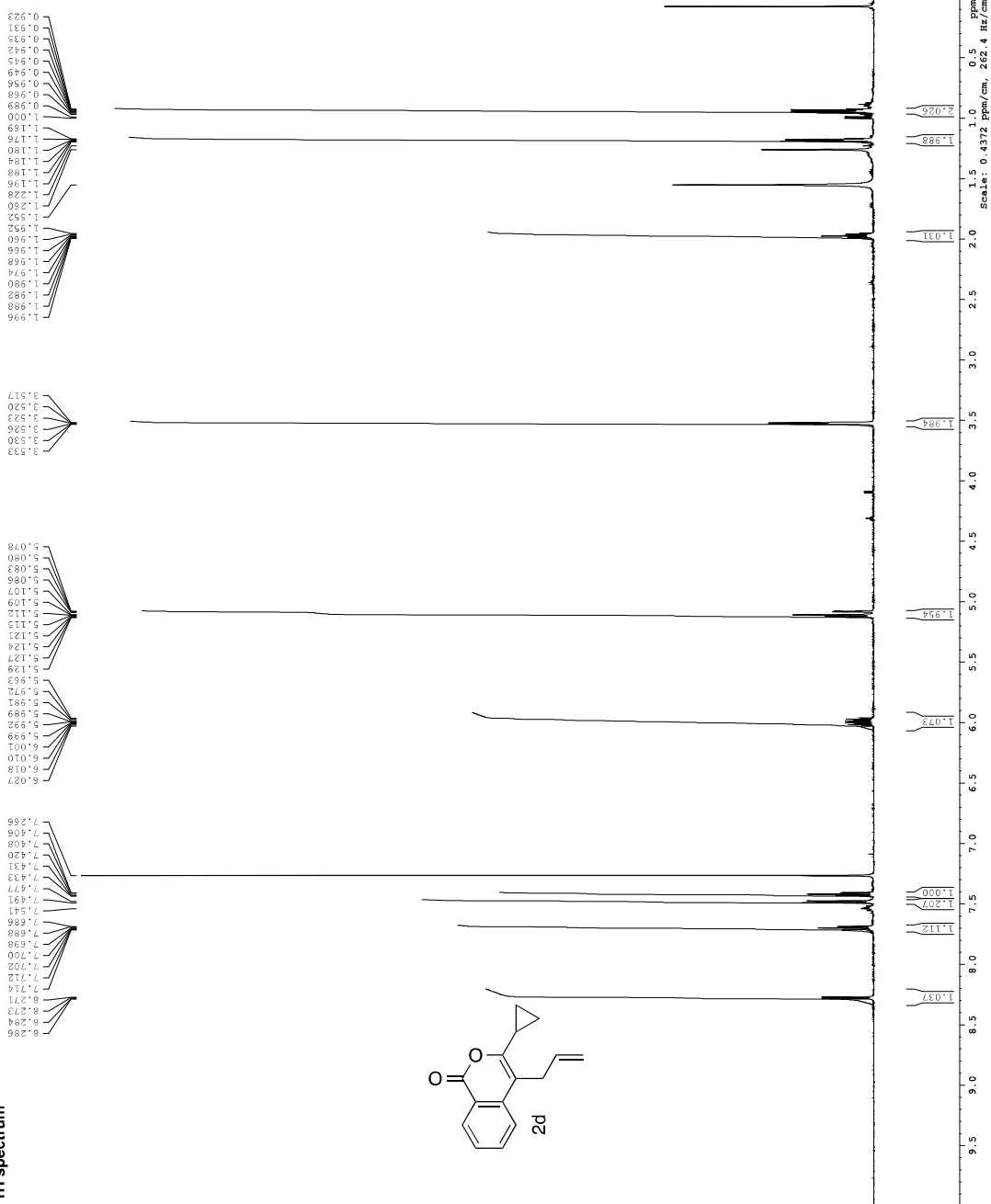
Current Data Parameters  
NAME: JMW136  
PROBHD: 5 mm QNP130  
PULPROG: zgpg30  
PROCNO: 1  
RG: 1  
AQ: 0.013446 sec  
RG: 20.000000 sec  
TE: 300.2 K  
DELTA: 0.25000000 sec  
DECOUPL1: 13C  
DECOUPL2: 13C  
DECOUPL3: 13C  
DECOUPL4: 13C  
DECOUPL5: 13C  
DECOUPL6: 13C  
DECOUPL7: 13C  
DECOUPL8: 13C  
DECOUPL9: 13C  
DECOUPL10: 13C  
DECOUPL11: 13C  
DECOUPL12: 13C  
DECOUPL13: 13C  
DECOUPL14: 13C  
DECOUPL15: 13C  
DECOUPL16: 13C  
DECOUPL17: 13C  
DECOUPL18: 13C  
DECOUPL19: 13C  
DECOUPL20: 13C  
DECOUPL21: 13C  
DECOUPL22: 13C  
DECOUPL23: 13C  
DECOUPL24: 13C  
DECOUPL25: 13C  
DECOUPL26: 13C  
DECOUPL27: 13C  
DECOUPL28: 13C  
DECOUPL29: 13C  
DECOUPL30: 13C  
DECOUPL31: 13C  
DECOUPL32: 13C  
DECOUPL33: 13C  
DECOUPL34: 13C  
DECOUPL35: 13C  
DECOUPL36: 13C  
DECOUPL37: 13C  
DECOUPL38: 13C  
DECOUPL39: 13C  
DECOUPL40: 13C  
DECOUPL41: 13C  
DECOUPL42: 13C  
DECOUPL43: 13C  
DECOUPL44: 13C  
DECOUPL45: 13C  
DECOUPL46: 13C  
DECOUPL47: 13C  
DECOUPL48: 13C  
DECOUPL49: 13C  
DECOUPL50: 13C  
DECOUPL51: 13C  
DECOUPL52: 13C  
DECOUPL53: 13C  
DECOUPL54: 13C  
DECOUPL55: 13C  
DECOUPL56: 13C  
DECOUPL57: 13C  
DECOUPL58: 13C  
DECOUPL59: 13C  
DECOUPL60: 13C  
DECOUPL61: 13C  
DECOUPL62: 13C  
DECOUPL63: 13C  
DECOUPL64: 13C  
DECOUPL65: 13C  
DECOUPL66: 13C  
DECOUPL67: 13C  
DECOUPL68: 13C  
DECOUPL69: 13C  
DECOUPL70: 13C  
DECOUPL71: 13C  
DECOUPL72: 13C  
DECOUPL73: 13C  
DECOUPL74: 13C  
DECOUPL75: 13C  
DECOUPL76: 13C  
DECOUPL77: 13C  
DECOUPL78: 13C  
DECOUPL79: 13C  
DECOUPL80: 13C  
DECOUPL81: 13C  
DECOUPL82: 13C  
DECOUPL83: 13C  
DECOUPL84: 13C  
DECOUPL85: 13C  
DECOUPL86: 13C  
DECOUPL87: 13C  
DECOUPL88: 13C  
DECOUPL89: 13C  
DECOUPL90: 13C  
DECOUPL91: 13C  
DECOUPL92: 13C  
DECOUPL93: 13C  
DECOUPL94: 13C  
DECOUPL95: 13C  
DECOUPL96: 13C  
DECOUPL97: 13C  
DECOUPL98: 13C  
DECOUPL99: 13C  
DECOUPL100: 13C



195 190 185 180 175 170 165 160 155 150 145 140 135 130 125 120 115 110 105 100 95 90 85 80 75 70 65 60 55 50 45 40 35 30 25 20 15 10 5 ppm  
Scale: 8.757 ppm/cm, 1101 Hz/cm

**1H spectrum**

Current Data Parameters  
 20000 1316 4  
 F2 - Acquisition Parameters  
 Time 20.44  
 INSTRUM 1H  
 PULPROG zgpg30  
 SOLVENT CDCl3  
 NS 2  
 DS 2  
 SWH 615.265 Hz  
 FWHM 5.098879 sec  
 AQ 52.000 usec  
 RG 328.0 usec  
 TE 298.0 usec  
 DE 1.0000000 sec  
 L1 0.10000000 sec  
 D1 0.10000000 sec  
 CHANNEL f1 1H  
 NUC1 1H  
 ZG4 23.044126 usec  
 SFO1 600.1310209 MHz  
 F2 - Processing parameters  
 SF 600.131021 MHz  
 EQ 2  
 EX 2  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



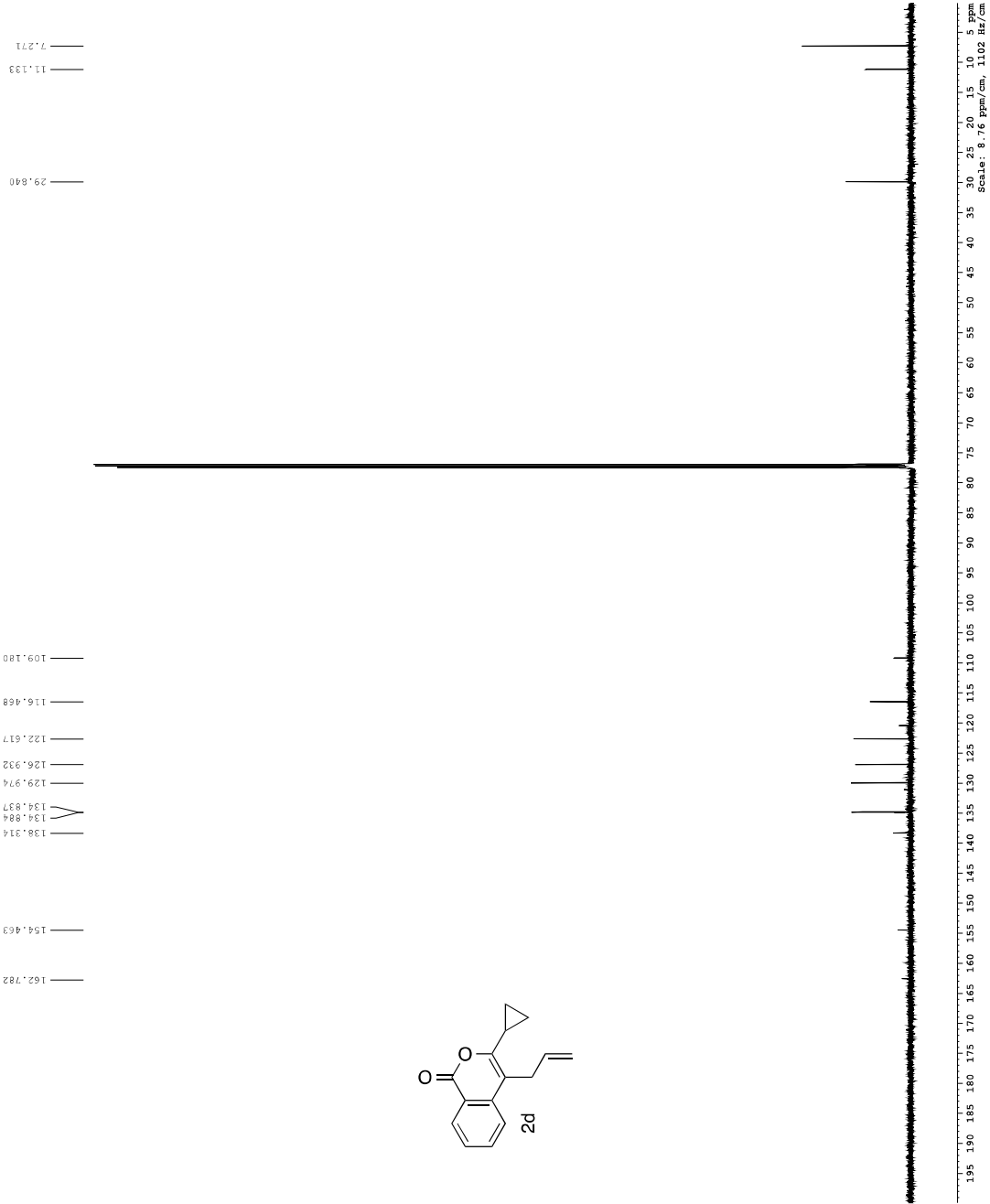
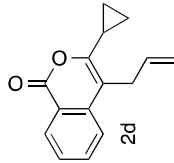
Z-restored spin-echo 13C spectrum with 1H decoupling

Current Data Parameters  
Date\_ 3/16/00  
Time\_ 11:33

F2 - Acquisition Parameters  
Time 20:22:53  
INSTRUM 8 mm CPY130  
PULPROG zgpg30  
SOLVENT CDCl3  
NS 512  
DS 4  
SWH 3000.031 Hz  
FIDRES 1.081350 epc  
AQ 0.050000 sec  
RG 327.500  
FQ 16.500 MHz  
TE 300.2 K  
D1 0.2500000 sec  
d11 0.0300000 sec  
d12 0.0300000 sec  
d15 0.0000000 sec  
RGREST 3 Hz  
SFOFFS 0.0150000 MHz  
SFOFF 0.0150000 MHz

===== CHANNEL f1 =====  
NUC1 13C  
P1 1.32 usec  
PL1 0.00 dB  
PL12 0.00 dB  
PL13 0.00 dB  
PL14 0.00 dB  
PL15 0.00 dB  
PL16 0.00 dB  
PL17 0.00 dB  
PL18 0.00 dB  
PL19 0.00 dB  
PL20 0.00 dB  
PL21 135.99100 dB  
PL22 0.0000000 dB  
PL23 0.0000000 dB  
PL24 0.0000000 dB  
PL25 0.0000000 dB  
PL26 0.0000000 dB  
PL27 0.0000000 dB  
PL28 0.0000000 dB  
PL29 0.0000000 dB  
PL30 0.0000000 dB  
PL31 0.0000000 dB  
PL32 0.0000000 dB  
PL33 0.0000000 dB  
PL34 0.0000000 dB  
PL35 0.0000000 dB  
PL36 0.0000000 dB  
PL37 0.0000000 dB  
PL38 0.0000000 dB  
PL39 0.0000000 dB  
PL40 0.0000000 dB  
PL41 0.0000000 dB  
PL42 0.0000000 dB  
PL43 0.0000000 dB  
PL44 0.0000000 dB  
PL45 0.0000000 dB  
PL46 0.0000000 dB  
PL47 0.0000000 dB  
PL48 0.0000000 dB  
PL49 0.0000000 dB  
PL50 0.0000000 dB  
PL51 0.0000000 dB  
PL52 0.0000000 dB  
PL53 0.0000000 dB  
PL54 0.0000000 dB  
PL55 0.0000000 dB  
PL56 0.0000000 dB  
PL57 0.0000000 dB  
PL58 0.0000000 dB  
PL59 0.0000000 dB  
PL60 0.0000000 dB  
PL61 0.0000000 dB  
PL62 0.0000000 dB  
PL63 0.0000000 dB  
PL64 0.0000000 dB  
PL65 0.0000000 dB  
PL66 0.0000000 dB  
PL67 0.0000000 dB  
PL68 0.0000000 dB  
PL69 0.0000000 dB  
PL70 0.0000000 dB  
PL71 0.0000000 dB  
PL72 0.0000000 dB  
PL73 0.0000000 dB  
PL74 0.0000000 dB  
PL75 0.0000000 dB  
PL76 0.0000000 dB  
PL77 0.0000000 dB  
PL78 0.0000000 dB  
PL79 0.0000000 dB  
PL80 0.0000000 dB  
PL81 0.0000000 dB  
PL82 0.0000000 dB  
PL83 0.0000000 dB  
PL84 0.0000000 dB  
PL85 0.0000000 dB  
PL86 0.0000000 dB  
PL87 0.0000000 dB  
PL88 0.0000000 dB  
PL89 0.0000000 dB  
PL90 0.0000000 dB  
PL91 0.0000000 dB  
PL92 0.0000000 dB  
PL93 0.0000000 dB  
PL94 0.0000000 dB  
PL95 0.0000000 dB  
PL96 0.0000000 dB  
PL97 0.0000000 dB  
PL98 0.0000000 dB  
PL99 0.0000000 dB  
PL100 0.0000000 dB

===== CHANNEL f2 =====  
NUC2 130  
P2 100.00 usec  
PL2 0.00 dB  
PL21 0.00 dB  
PL22 0.00 dB  
PL23 0.00 dB  
PL24 0.00 dB  
PL25 0.00 dB  
PL26 0.00 dB  
PL27 0.00 dB  
PL28 0.00 dB  
PL29 0.00 dB  
PL30 0.00 dB  
PL31 0.00 dB  
PL32 0.00 dB  
PL33 0.00 dB  
PL34 0.00 dB  
PL35 0.00 dB  
PL36 0.00 dB  
PL37 0.00 dB  
PL38 0.00 dB  
PL39 0.00 dB  
PL40 0.00 dB  
PL41 0.00 dB  
PL42 0.00 dB  
PL43 0.00 dB  
PL44 0.00 dB  
PL45 0.00 dB  
PL46 0.00 dB  
PL47 0.00 dB  
PL48 0.00 dB  
PL49 0.00 dB  
PL50 0.00 dB  
PL51 0.00 dB  
PL52 0.00 dB  
PL53 0.00 dB  
PL54 0.00 dB  
PL55 0.00 dB  
PL56 0.00 dB  
PL57 0.00 dB  
PL58 0.00 dB  
PL59 0.00 dB  
PL60 0.00 dB  
PL61 0.00 dB  
PL62 0.00 dB  
PL63 0.00 dB  
PL64 0.00 dB  
PL65 0.00 dB  
PL66 0.00 dB  
PL67 0.00 dB  
PL68 0.00 dB  
PL69 0.00 dB  
PL70 0.00 dB  
PL71 0.00 dB  
PL72 0.00 dB  
PL73 0.00 dB  
PL74 0.00 dB  
PL75 0.00 dB  
PL76 0.00 dB  
PL77 0.00 dB  
PL78 0.00 dB  
PL79 0.00 dB  
PL80 0.00 dB  
PL81 0.00 dB  
PL82 0.00 dB  
PL83 0.00 dB  
PL84 0.00 dB  
PL85 0.00 dB  
PL86 0.00 dB  
PL87 0.00 dB  
PL88 0.00 dB  
PL89 0.00 dB  
PL90 0.00 dB  
PL91 0.00 dB  
PL92 0.00 dB  
PL93 0.00 dB  
PL94 0.00 dB  
PL95 0.00 dB  
PL96 0.00 dB  
PL97 0.00 dB  
PL98 0.00 dB  
PL99 0.00 dB  
PL100 0.00 dB



Scale: 8.76 PPM/cm, 1102 Hz/cm







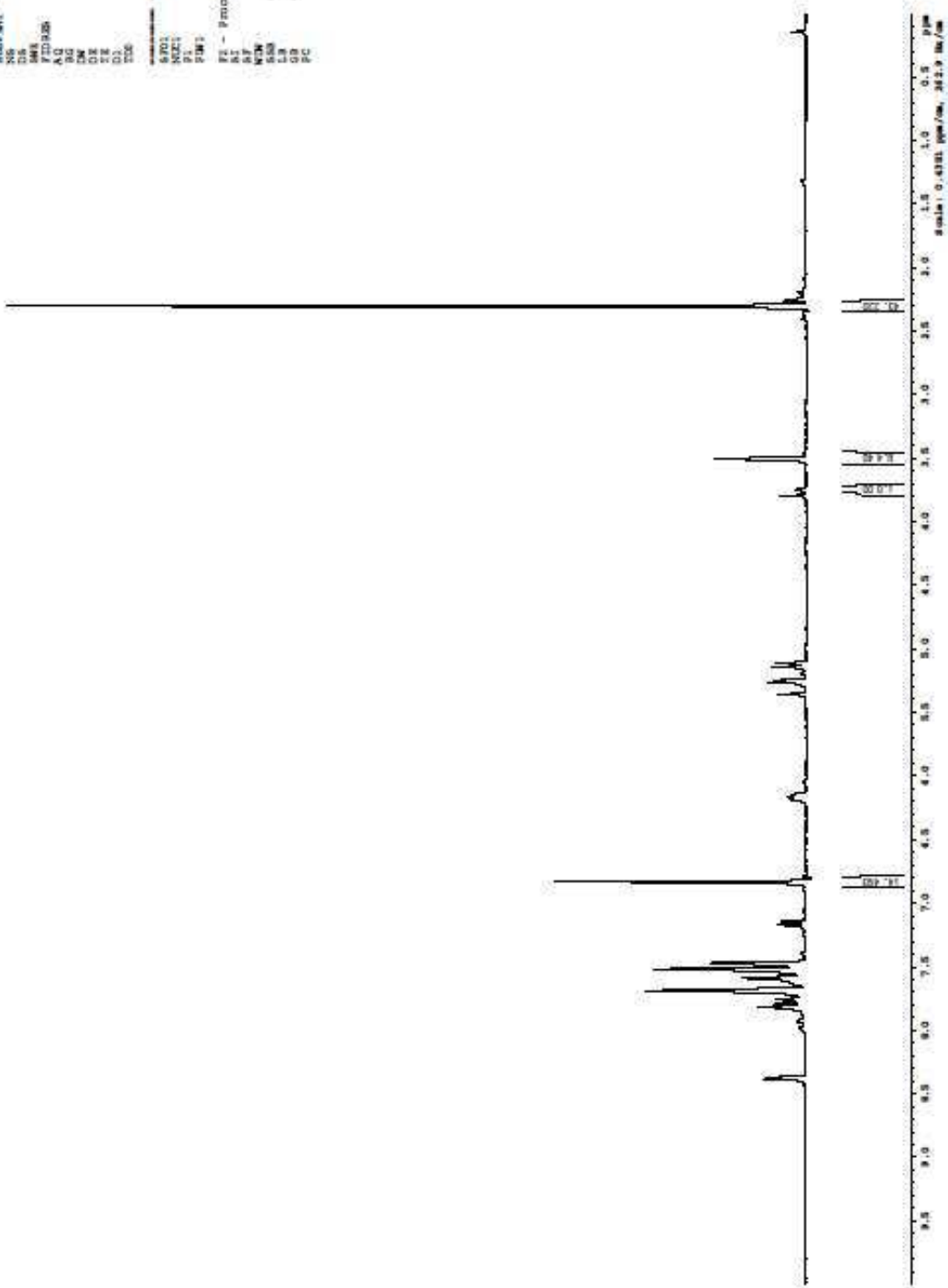
1H spectrum

Current Data Parameters  
NAME: 2152149  
EXPNO: 1  
PROCNO: 1

F2 - Acquisition Parameters  
Date\_ 20140111  
Time 19.46  
INSTRUM spect  
PROBHD 5 mm QNP 1H/13  
PULPROG zgpg30  
TD 65536  
FIDRES 0.4074  
AQUANT 1  
RG 327.5  
AQ 0.0145 HZ  
F2 - Processing parameters  
F2FREQ 125.7612 MHz  
AQ 0.0145 HZ  
RG 327.5  
WDW EM  
SSB 0  
GB 0  
PC 1.00

===== CHANNEL f1 =====  
NUC1 13C  
P1 1.00  
F1 101.253175 MHz  
F2 - Processing parameters  
F2FREQ 400.130000 MHz  
WDW EM  
SSB 0  
GB 0  
PC 1.00

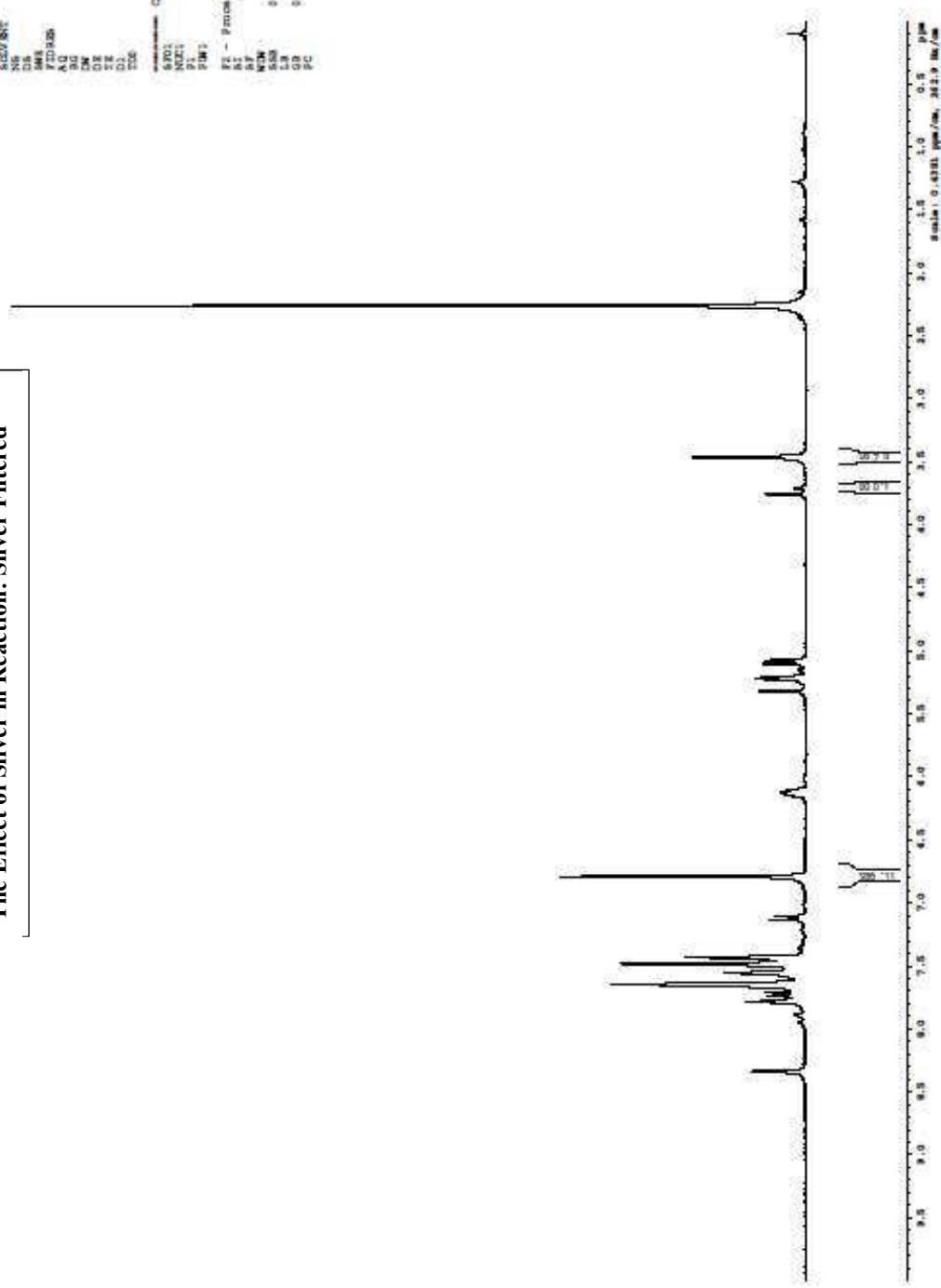
The Effect of Silver in Reaction: Silver Unfiltered



1H spectrum

The Effect of Silver in Reaction: Silver Filtered

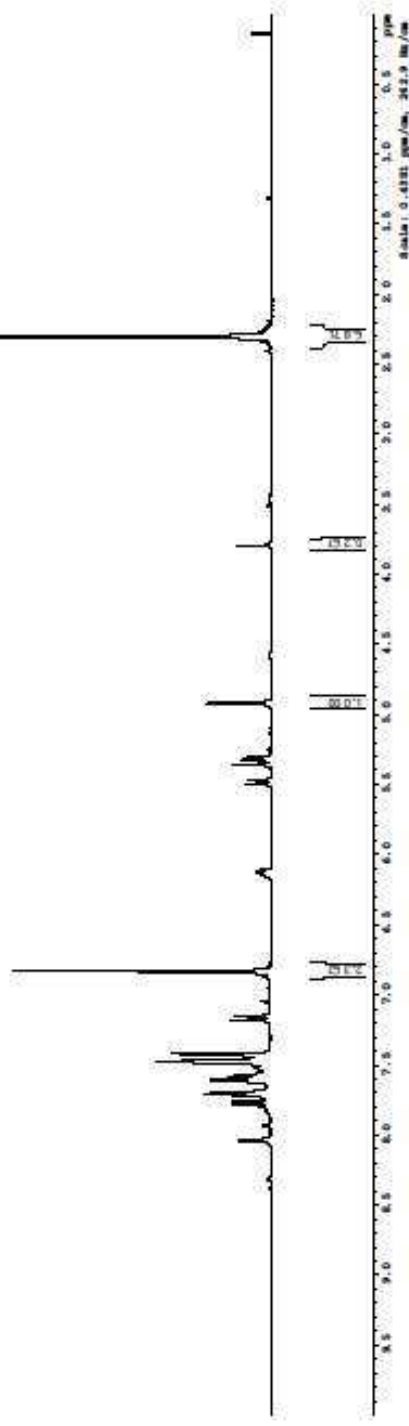
Current Data Parameters  
NAME: 374213  
PROCNO: 1  
F2 - Acquisition Parameters  
Date\_ Time: 2011-07-11 22:47  
INSTRUM: spect  
PROBHD: 5 mm QNP 1H/13  
PULPROG: zgpg30  
TD: 65536  
SOLVENT: CDCl3  
NS: 1  
DS: 4  
AQ: 0.131 s  
RG: 327.680  
FIDRES: 0.0462 Hz  
AQ: 5.094471 sec  
RG: 404  
AQ: 31.000 sec  
RG: 256.0  
AQ: 2.500 sec  
RG: 0.1  
AQ: 40.000000 sec  
TD: 1  
===== CHANNEL f1 =====  
NUC1: 13C  
P1: 12.00 usec  
PL1: 0.00 dB  
SFO1: 125.7611538 MHz  
F2 - Processing parameters  
SI: 32768  
SF: 125.7611538 MHz  
WDW: EM  
SSB: 0  
LB: 0.30 Hz  
GB: 0  
PC: 1.00

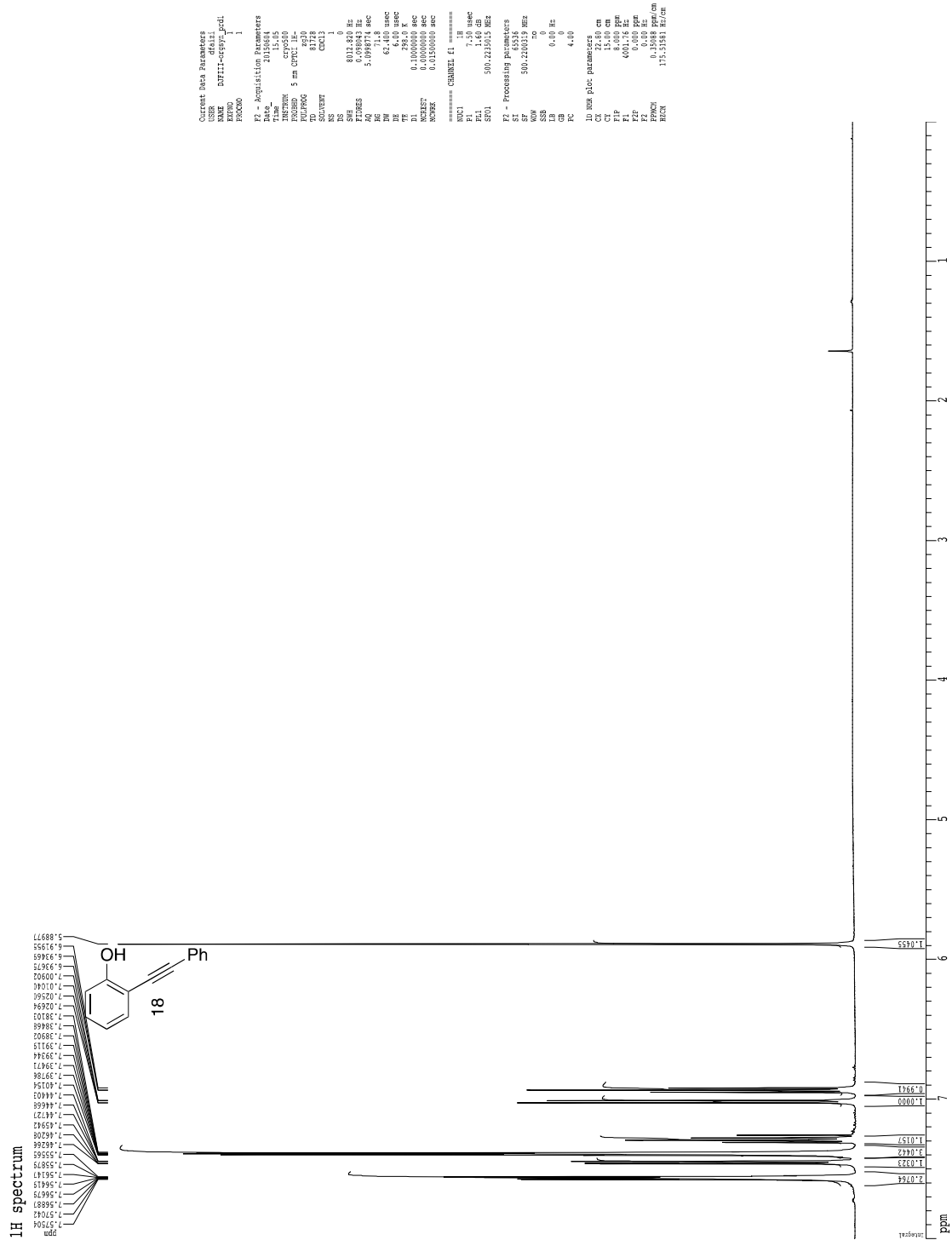


1H spectrum

CUSTOMER DATA PARAMS FOR  
NAME: 370213  
PROJECT: 1  
PRG: 1  
F2 - Acquisition Parameters  
Date\_: 20140111  
Time: 14:34  
INSTRUM: spect  
PROBHD: 5 mm QNP 1H/13  
PULPROG: zgpg30  
TD: 65536  
AQ: 0.4674  
RG: 327.5  
SOLVENT: CDCl3  
NS: 1  
DS: 4  
SWH: 9425.345 Hz  
FIDRES: 0.094642 Hz  
AQ: 5.098478 sec  
RG: 327.5  
SOLVENT: CDCl3  
D1: 2.00  
D2: 0.00000000 sec  
D3: 1  
D5: 1  
===== CHANNEL f1 =====  
NUC1: 13C  
P1: 1.00  
PL1: 0.00000000 sec  
===== CHANNEL f2 =====  
NUC2: 1H  
P2: 0.00000000 sec  
PL2: 0.00000000 sec  
===== END =====

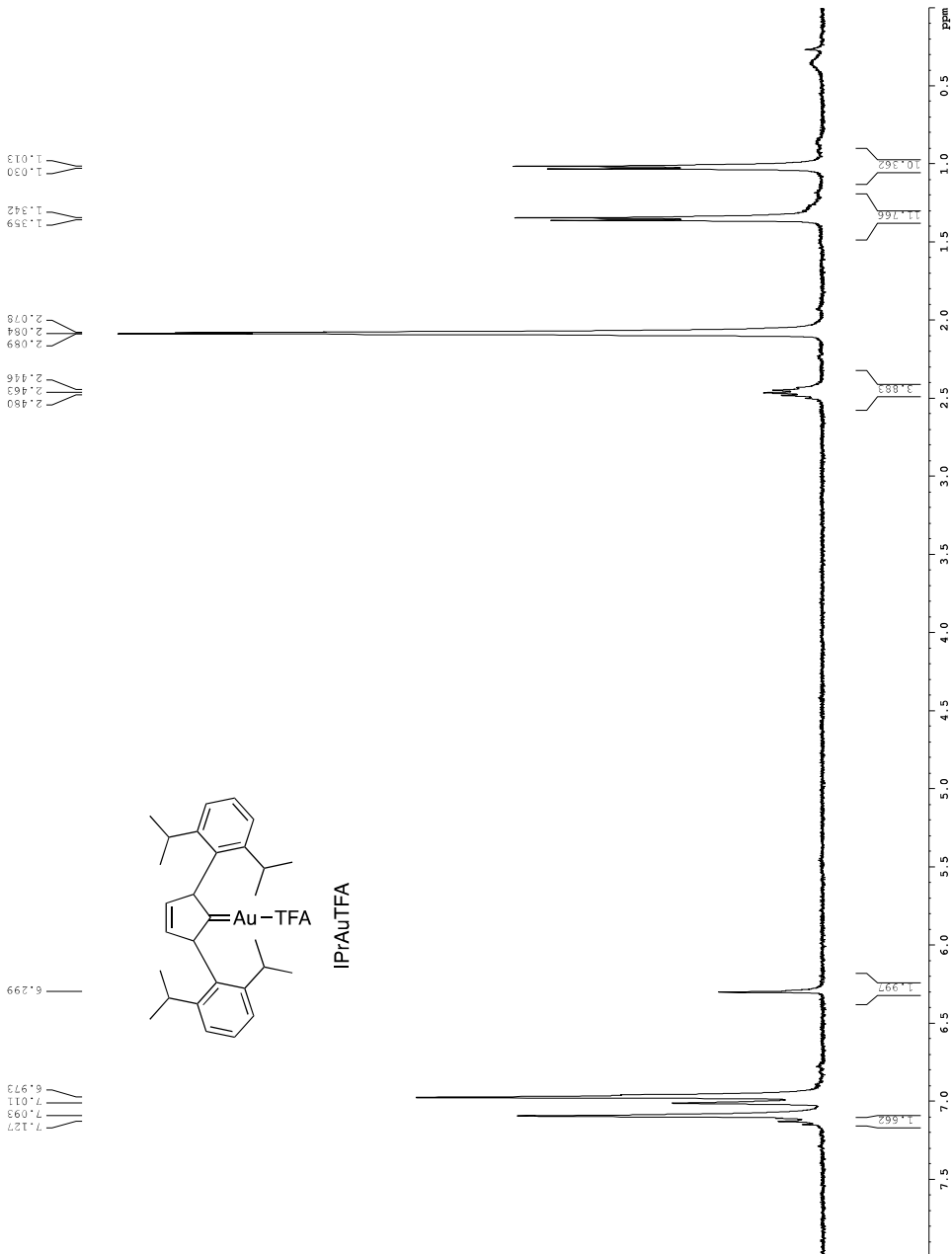
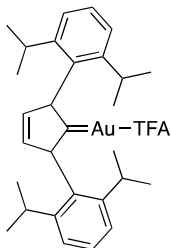
The Effect of Silver in Reaction: Silver Only





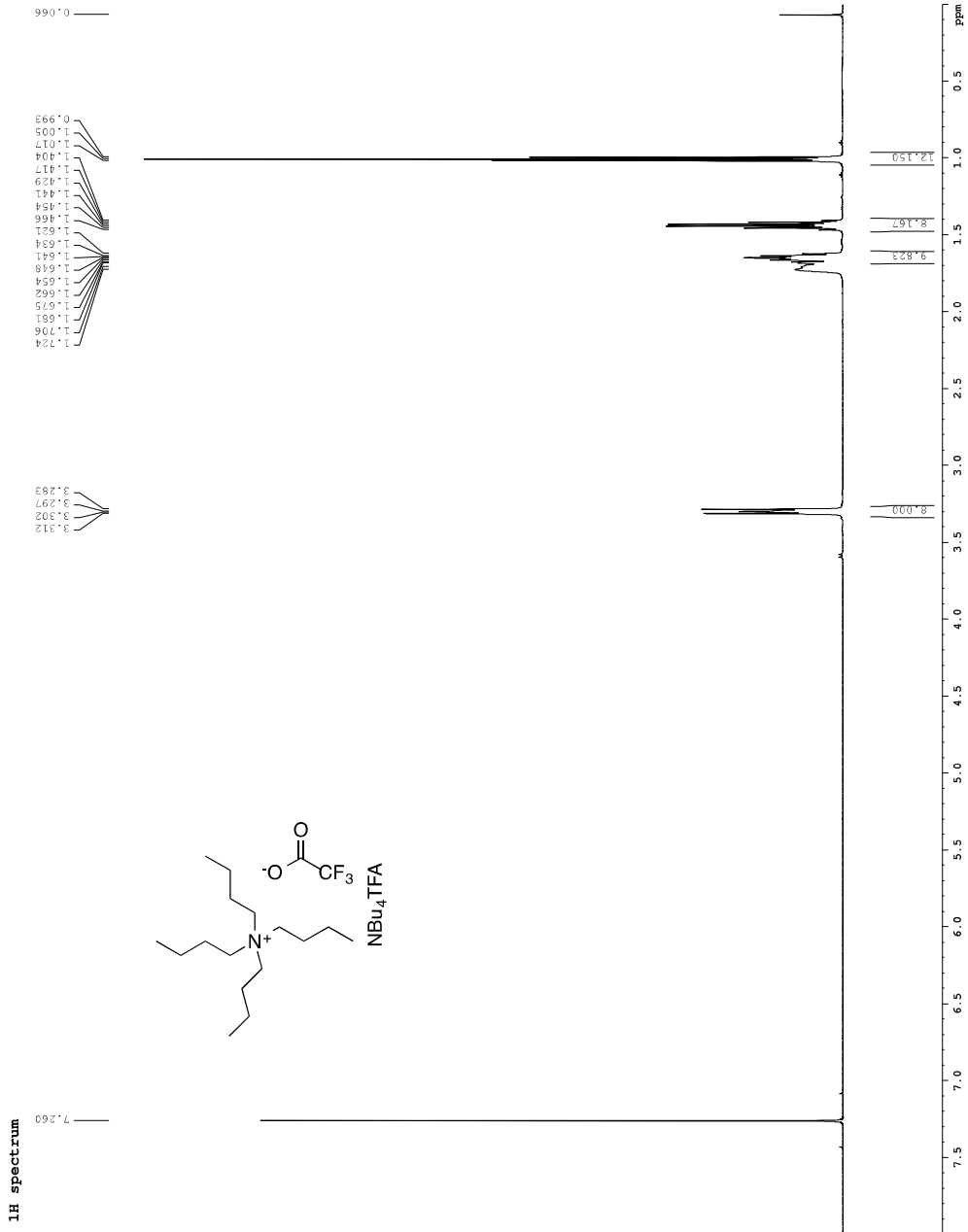
**<sup>1</sup>H spectrum**

Current Data Parameters  
 NAME J101152  
 PROCNO 1  
 F2 - Acquisition Parameters  
 Date\_ 20141119  
 Time 19:13  
 INSTRUM spect400  
 PROBHD 5 mm QNP 1H/13  
 PULPROG zgpg30  
 ID 65536  
 SOLVENT DMS  
 NS 18  
 DS 2  
 SWH 6410.62  
 FIDRES 0.097613 Hz  
 AQ 5.1118678 sec  
 DD 78.100 usec  
 DE 4.50 usec  
 TE 298.0 K  
 MCHST 0 sec  
 MCPRK 0.0100000 sec  
 CHANNEL f1  
 NUC1 <sup>1</sup>H  
 P1 12.00 usec  
 PL 0 dB  
 SFO1 400.1328605 MHz  
 F2 - Processing parameters  
 SI 65536  
 SF 400.1301134 MHz  
 DS 4  
 MSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 2.00



```

Current: Data Parameters
NAME      JJ04015
PROBHD    1
=====
F2 - Acquisition Parameters
Date_      2016.11
Time       16.11
INSTRUM    SPC00
PULPROG    zgpg30
TD         65536
SOLVENT    CDCl3
NS         8
DS         2
SWH         6615.2 Hz
AQ         0.098476 sec
RG         512
DE         52.000 usec
TE         296.2 K
TD0        1
=====
CHANNEL f1
SFO1       600.132409 MHz
NUC1       1H
P1         12.00 nsec
PL1        24.50770093 dB
=====
F2 - Processing parameters
SI         65535
SF         600.1300954 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
  
```



**<sup>1</sup>H spectrum**  
**Sample Spectrum at T = 130 s**

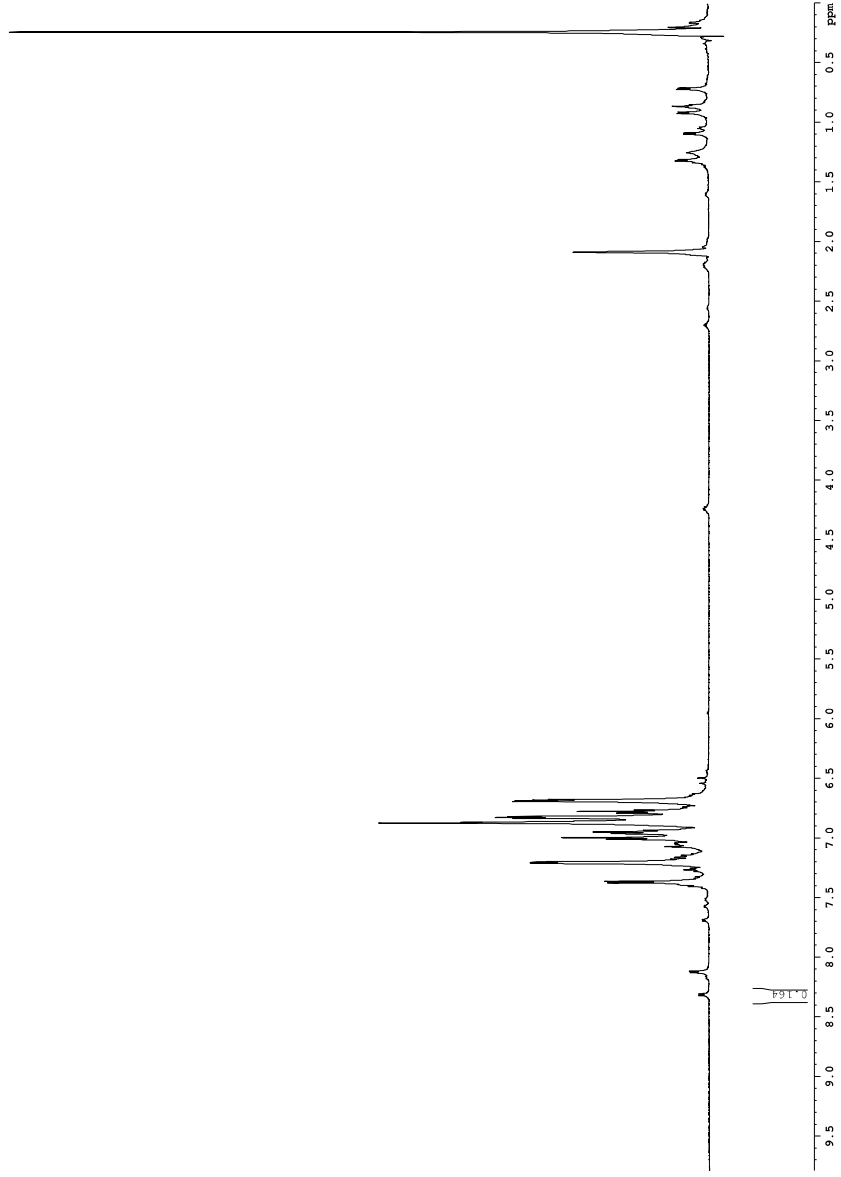
```

Current Data Parameters
Name      J104009
EXPNO    4
PROCNO   1

F2 - Acquisition Parameters
Date_    20130308
Time     21.33
INSTRUM  S
PROBHD   5 mm BBO BB-1H
PULPROG  zgpg30
TD        115382
SOLVENT  CDCl3
NS        1
DS        1
SWH       9615.385 Hz
FIDRES    0.083335 Hz
AQ        5.9598641 sec
RG        327.5
DM        52.000 usec
DE        10.50 usec
TE        340.6 K
DQ        0.10000000 sec
TD0       1

===== CHANNEL f1 =====
SFO1     500.1342019 MHz
NUC1     13C
P1        9.00 usec
PL1       0.00 dB
PLW1     60.25500052 W

F2 - Processing parameters
SI        65536
SF        500.1331355 MHz
WDW       EM
SSB       0
GB        0
PC        1.00 Hz
FC        1.00
  
```



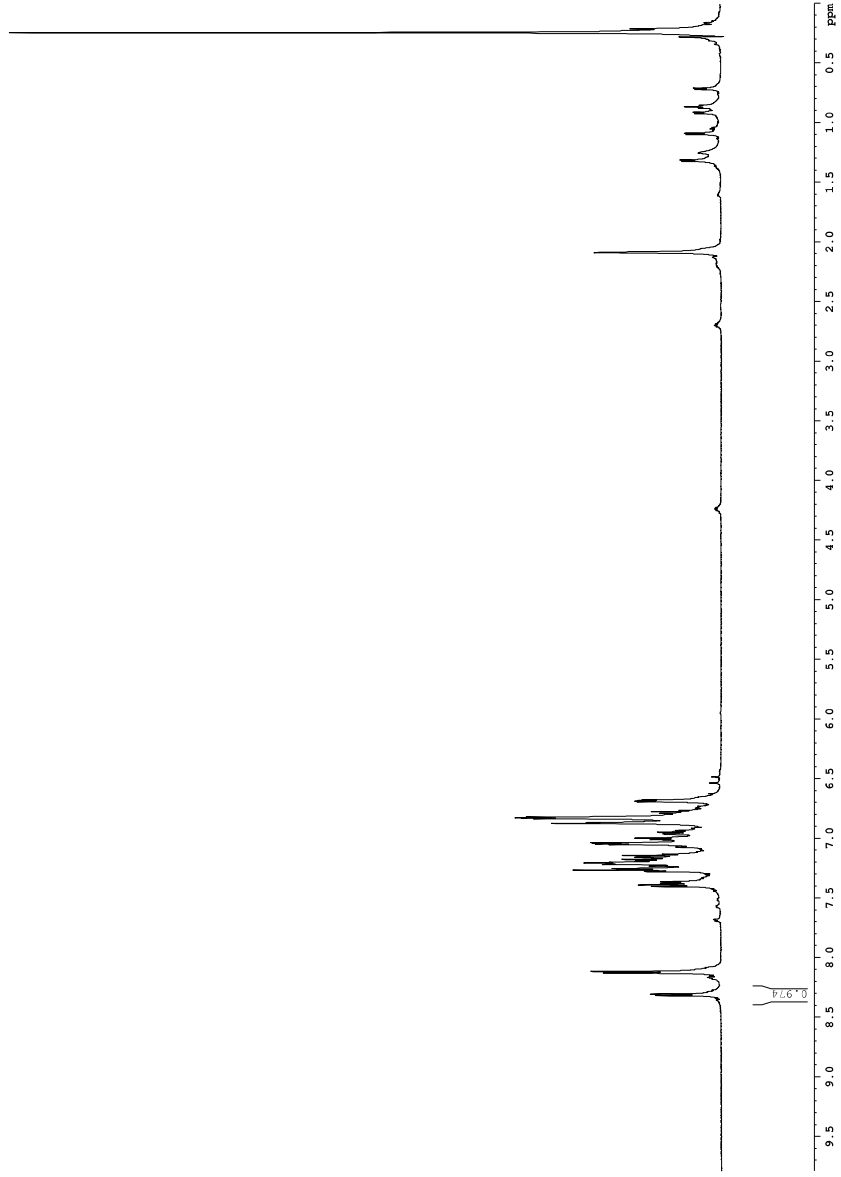
**<sup>1</sup>H spectrum**  
**Sample Spectrum at T = 954 s**

```

Current Data Parameters
Name      J104823
EXPNO    23
PROCNO   1

F2 - Acquisition Parameters
Date_    20150509
Time     21.47
INSTRUM  spect
PROBHD   5 mm BBO BB-1H
PULPROG  zgpg30
TD        115382
SOLVENT  CDCl3
NS        1
DS        1
SWH       9615.385 Hz
FIDRES    0.083335 Hz
AQ         5.9598641 sec
RG         512
WDW        EM
SSB        0
LB         10.50 Hz
GB         0
PC         341.1 K
TD0        0.10000000 sec

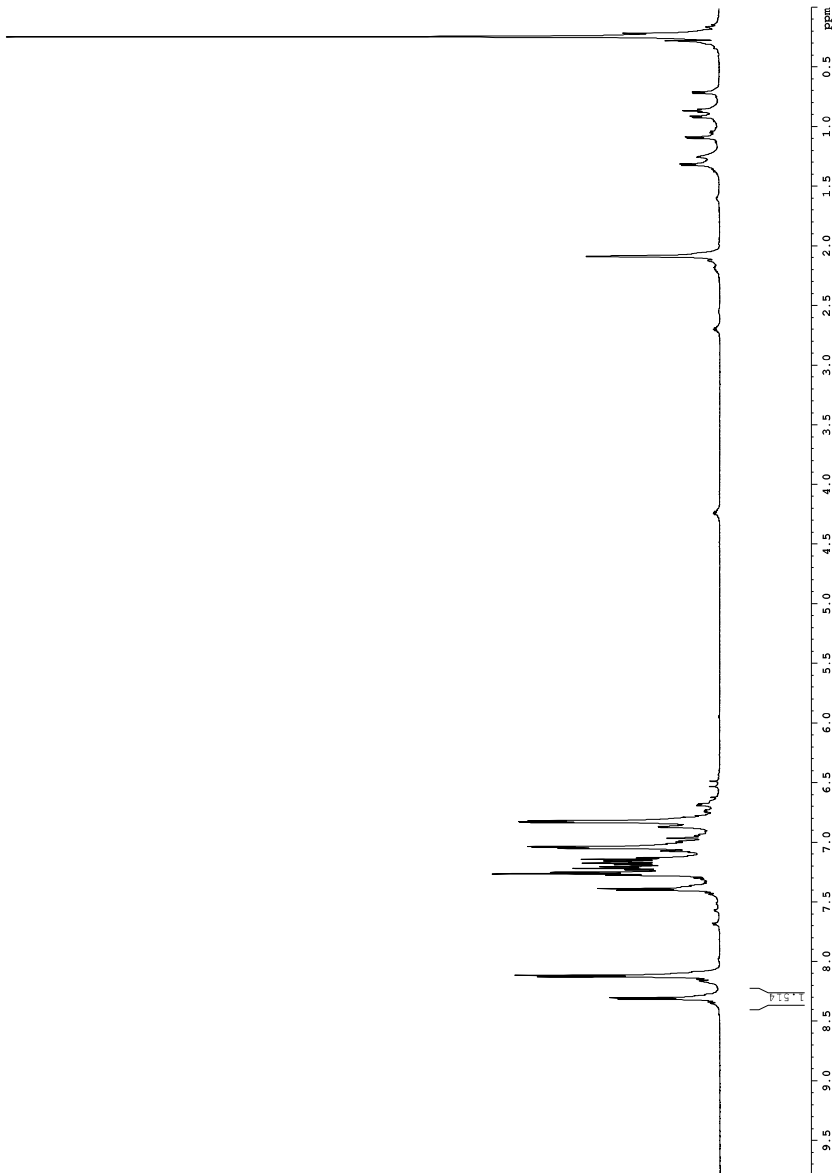
===== CHANNEL f1 =====
SFO1     500.1342019 MHz
NUC1      13C
P1        9.00 usec
PL1       0.00 dB
PLWL      50.25500052 W
=====
F2 - Processing parameters
SI         65536
SF         500.1331349 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.00
  
```





1H spectrum  
Sample Spectrum at T = 1995 s

Current Data Parameters  
Name J104097  
EXPNO 1  
PROCNO 1  
F2 - Acquisition Parameters  
Date\_ 20120303  
Time 22.06  
INSTRUM S min\_BBO BB-1H  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 115382  
SOLVENT CDCl3  
NS 1  
DS 1  
SWH 9615.385 Hz  
FIDRES 0.083335 Hz  
AQ 5.9598641 sec  
RG 655  
DM 52.000 usec  
DE 10.50 usec  
TE 339.8 K  
D0 0.10000000 sec  
TD0 1  
===== CHANNEL f1 =====  
SFO1 500.134209 MHz  
NUC1 13C  
P1 9.00 usec  
PL1 0.00 dB  
PLWL 50.25500052 W  
F2 - Processing parameters  
SI 65536  
SF 500.134355 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
FC 1.00



**<sup>1</sup>H spectrum**  
**Mesitylene ERETIC Reference Spectrum**

```

Current Data Parameters
Name_ mes eretic 50 degree tbi
EXPNO 1
PROCNO 1
F2 - Acquisition Parameters
Date_ 20120201
Time_ 12.14
INSTRUM spect
PROBHD 5 mm TBI LH/13
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 1
DS 1
SWH 9615.385 Hz
FIDRES 0.083335 Hz
AQ 5.9598641 sec
RG 655.36
DM 52.000 usec
DE 11.13 usec
TE 323.5 K
D0 34.00000000 sec
TDO 1
===== CHANNEL F1 =====
SFO1 500.1342019 MHz
P1 8.00 usec
ELW1 24.54700089 W
F2 - Processing parameters
SI 65536
SF 500.1300010 MHz
WDW EM
SSB 0
GB 0
PC 1.00
FC 1.00
  
```

