

UC Irvine

UC Irvine Previously Published Works

Title

Evolving Interactions and Emergent Functions in Microbial Consortia.

Permalink

<https://escholarship.org/uc/item/0wm8q3tz>

Journal

mSystems, 6(4)

ISSN

2379-5077

Author

Rodríguez-Verdugo, Alejandra

Publication Date

2021-08-31

DOI

10.1128/mSystems.00774-21

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <https://creativecommons.org/licenses/by/4.0/>

Peer reviewed



Evolving Interactions and Emergent Functions in Microbial Consortia

 Alejandra Rodríguez-Verdugo^a

^aDepartment of Ecology and Evolutionary Biology, University of California, Irvine, California, USA

ABSTRACT Microbial communities are constantly challenged with environmental stressors, such as antimicrobials, pollutants, and global warming. How do they respond to these changes? Answering this question is crucial given that microbial communities perform essential functions for life on Earth. Our research aims to understand and predict communities' responses to change by addressing the following questions. (i) How do eco-evolutionary feedbacks influence microbial community dynamics? (ii) How do multiple interacting species in a microbial community alter evolutionary processes? (iii) To what extent do microbial communities respond to change by ecological versus evolutionary processes? To answer these questions, we use microbial communities of reduced complexity coupled with experimental evolution, genome sequencing, and mathematical modeling. The overall expectation from this integrative research approach is to generate general concepts that extend beyond specific bacterial species and provide fundamental insights into the consequences of evolution on the functioning of whole microbial communities.

KEYWORDS evolution, microbial ecology, environmental change, species interactions, microbial communities, ecosystem functions, collective metabolism

Microbial communities perform pivotal functions, from the cycling of elements through Earth's ecosystems to the digestion of complex foods that shape human health and disease. Many of these functions emerge from the interaction of multiple species working as a collective (1). For example, the degradation of complex polymers in nature often requires a division of labor among species for their breakdown. A key question is how these emergent functions could persist, in the long-term, in face of environmental stressors, such as antimicrobials, pollutants, and global warming. To answer this question, we first need to understand how species and their interactions evolve in response to change.

Achieving such understanding is extraordinarily difficult given that microbial communities are incredibly diverse in terms of numbers of different microbes, genes, and interactions. Progress on this problem requires both top-down studies that deal with the complexity of natural communities and bottom-up research that uses simpler experimental setups (2). My research group works at the intersection between top-down and bottom-up research and strives to understand how microbial communities respond to environmental change and the consequences of these changes on collective functions. By studying simplified communities under both laboratory and field conditions and by addressing fundamental questions, we aim to make conceptual advances in the emerging field of microbial evolutionary ecology.

HOW DO ECO-EVOLUTIONARY FEEDBACKS INFLUENCE MICROBIAL COMMUNITY DYNAMICS?

Rapid evolution can impact short-term ecological dynamics, and these altered ecological dynamics can feedback to affect subsequent evolutionary change (3). Eco-

Citation Rodríguez-Verdugo A. 2021. Evolving interactions and emergent functions in microbial consortia. *mSystems* 6:e00774-21. <https://doi.org/10.1128/mSystems.00774-21>.

Copyright © 2021 Rodríguez-Verdugo. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to alejandr1@uci.edu.

Conflict of Interest Disclosures: A.R.-V. has nothing to disclose.

The views expressed in this article do not necessarily reflect the views of the journal or of ASM.

This article is part of a special series sponsored by Floré.

Published 24 August 2021

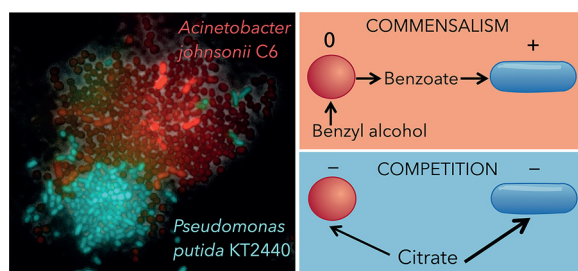


FIG 1 The interaction between *A. johnsonii* and *P. putida* changes according to the resource provided.

evolutionary feedbacks have been well documented for pairs of microbes with reciprocal negative interactions, such as predator-prey and host-parasite interactions (4–6). Less is known about how eco-evolutionary feedbacks alter other types of ecological interactions, such as commensalism, in which one species unilaterally benefits another species. This knowledge is critical given that commensal interactions, especially those based on metabolic exchanges (the waste of one species being the substrate for another), underlay important functions, such the breakdown of indigestible polymers in animals (7). One important question is how do commensal interactions based on metabolic exchanges evolve through time and drive microbial community dynamics?

My research has contributed to answering this question by studying the long-term stability of a commensal interaction based on metabolic exchanges. We have been studying the interaction between *Acinetobacter johnsonii* and *Pseudomonas putida*, two bacterial species that were originally isolated from a polluted aquifer in Denmark (8). This consortium has been a model system to study commensal interactions for more than 20 years, in part, because species collectively degrade toxic and recalcitrant compounds and therefore can be used for bioremediation (9, 10). Another advantage of working with such a consortium is that species and their interactions are easy to manipulate by controlling the chemical composition of the culture media. Using a simple mathematical model, we showed that interactions are based solely on the use of resources from the external environment (11). We reached this conclusion by building a mathematical model informed by single-species behaviors and by accurately predicting ecological dynamics in a two-species consortium (11). When grown in an environment limited by citrate, species compete for it (i.e., exploitation competition). Instead, when grown in an environment limited by benzyl alcohol, *P. putida* cross-feeds on the benzoate excreted by *A. johnsonii* (i.e., commensalism) (Fig. 1). Thus, by manipulating the resources, we can manipulate species interactions and address questions related to the long-term stability of different types of species interactions, including interactions that change over time in response to external fluctuations in resources.

To assess the stability of a cross-feeding commensal interaction over evolutionary timescales, we conducted a laboratory evolution experiment over 200 generations using cocultures of *A. johnsonii* and *P. putida* (12). We found that species rapidly adapted to their culture conditions through *de novo* mutations. Importantly, rapid evolution had different consequences on species coexistence according to the environment. In a constant environment, species stably coexisted, while in a fluctuating environment with daily switches between commensalism and competition, rapid evolution sometimes led to the extinction of *A. johnsonii*. We concluded that eco-evolutionary feedbacks are important drivers of community dynamics in a fluctuating environment but are relatively negligible in a constant environment in which commensalism is the sole type of interaction. Thus, by finding the conditions in which a commensal interaction is stable over evolutionary timescales, we have provided the field of synthetic ecology with a successful example of evolutionary stable consortia

(13). The question remains on how cross-feeding commensal interactions could evolve to give rise to new emergent functions (e.g., enhanced biodegradation efficiency). How do species that utilize the waste products of other species ameliorate the collective metabolism of a multispecies system? These questions can be tackled by evolving the *Acinetobacter-Pseudomonas* consortium with high replication under selective pressures promoting metabolic exchanges. By investigating the fundamental principles governing the evolution of collective metabolism of multispecies systems, we will contribute to explaining the widespread occurrence of cross-feeding interactions (e.g., metabolic dependencies) in natural communities (14, 15).

DO MULTIPLE INTERACTING SPECIES IN A MICROBIAL COMMUNITY ALTER EVOLUTIONARY PROCESSES?

Although two-species consortia exist in nature and play important roles for ecosystem functioning (16), most microbial communities are composed of more than two species (often containing hundreds or thousands of microbial species). Thus, a major question in microbial evolutionary ecology is whether a focal species responds to selective pressures from one or multiple interacting species (17). In pairwise evolution (coevolution), a population's evolution is mainly driven by another species: e.g., evolution of antipredator defense. Instead, in diffuse evolution, a population's evolution is driven by multiple species (18). Determining if evolution is pairwise or diffuse is important for predicting evolutionary trajectories in complex microbial communities. To address this important issue, we are currently assembling simple multispecies communities and plan to assess to what extent the community context (the presence of more than two species) determines the direction and rate of evolution of community members.

TO WHAT EXTENT DO MICROBIAL COMMUNITIES RESPOND TO ENVIRONMENTAL CHANGE BY ECOLOGICAL VERSUS EVOLUTIONARY PROCESSES?

Microbial communities respond to environmental change through shifts in the identity and frequency of different species (ecological responses) and different genotypes (evolutionary responses). What is the relative contribution of ecological versus evolutionary processes to mediating responses to change? We previously discussed how adaptive evolution mediated by *de novo* mutations can influence ecological dynamics in laboratory settings (12). What remains largely unknown is the contribution of adaptive evolution in communities under natural conditions (19, 20). Filling this knowledge gap is important because evolutionary adaptation may influence key ecosystem functions, such the rates of carbon and nutrient cycling. For example, environmental stressors may lead to the decline or extinction of taxa that are key for carbon cycling. In contrast to these demographic responses, evolution may be able to restore lost functions and even prevent species extinction (e.g., evolutionary rescue). On the other hand, evolutionary adaptation may entail tradeoffs between stress tolerance and other functional traits (21). This example illustrates the importance of identifying mechanisms mediating responses to environmental change to predict consequences for ecosystem functions.

One important ecosystem function is plant litter decomposition, which is performed by microbial communities (i.e., leaf litter microbiomes). I have recently joined a collaboration, The Loma Ridge Climate Change Experiment, aiming to understand how the plant litter microbiome responds to drought in Southern California (22, 23). We are assembling multispecies consortia representative of the leaf litter microbiome from the Loma Ridge grassland site. Our consortia are simple enough that we can easily manipulate them and conduct dozens of evolution experiments in parallel, but are complex enough so that we can capture some of the features of the leaf litter microbiome: e.g., consortia are grown in leaf litter extract in environments with spatial and temporal structures. Importantly, these consortia can be used to conduct manipulative field evolution experiments. In these experiments, consortia are

enclosed with litter substrates in microbial cages made of nylon membranes with microscopic pores. This allows the consortia to experience the abiotic conditions from the field while preventing migration from local microbial species (20). These closed systems allow us to easily track changes in species and genotypes frequencies through time (20). Taken together, we expect this approach will bring us one step closer to understanding how communities respond to change in nature and how these responses influence important ecosystems functions.

FUTURE DIRECTIONS

Using an integrative research approach based on evolution experiments of simple microbial consortia, whole-genome sequencing, and mathematical modeling, we have gained general insights into the mechanisms by which microbial communities respond to environmental change. We are currently working to understand how these responses affect community-level functions important for ecosystem functioning. The next exciting research directions will be (i) to study whether natural selection can act on community-level properties (24) and (ii) to quantify evolutionary rates and mechanisms mediating responses to environmental change under field conditions. By addressing these questions, our research will have lasting impacts on both our understanding of eco-evolutionary dynamics in communities and our current views of the capabilities of organisms in changing environments.

ACKNOWLEDGMENTS

I thank Katrine Whiteson for inviting me to write this commentary and Jack Gilbert for providing valuable feedback on an earlier version of the manuscript. I thank current and past members of my lab, especially two talented undergraduate students, Melissa Martens and Parmis Abdoli, supported by UC Irvine's Summer Undergraduate Research Fellowship (SURF) to work on the Loma Ridge project funded by the U.S. Department of Energy (DE-SC0020382).

REFERENCES

- Gilbert JA, Henry C. 2015. Predicting ecosystem emergent properties at multiple scales. *Environ Microbiol Rep* 7:20–22. <https://doi.org/10.1111/1758-2229.12258>.
- De Roy K, Marzorati M, Van den Abbeele P, Van de Wiele T, Boon N. 2014. Synthetic microbial ecosystems: an exciting tool to understand and apply microbial communities. *Environ Microbiol* 16:1472–1481. <https://doi.org/10.1111/1462-2920.12343>.
- Rodríguez-Verdugo A, Buckley J, Stapley J. 2017. The genomic basis of eco-evolutionary dynamics. *Mol Ecol* 26:1456–1464. <https://doi.org/10.1111/mec.14045>.
- Hall AR, Scanlan PD, Morgan AD, Buckling A. 2011. Host-parasite coevolutionary arms races give way to fluctuating selection. *Ecol Lett* 14:635–642. <https://doi.org/10.1111/j.1461-0248.2011.01624.x>.
- Hiltunen T, Becks L. 2014. Consumer co-evolution as an important component of the eco-evolutionary feedback. *Nat Commun* 5:5226. <https://doi.org/10.1038/ncomms6226>.
- Retel C, Kowallik V, Huang W, Werner B, Künzel S, Becks L, Feulner PGD. 2019. The feedback between selection and demography shapes genomic diversity during coevolution. *Sci Adv* 5:eaax0530. <https://doi.org/10.1126/sciadv.aax0530>.
- Solden LM, Naas AE, Roux S, Daly RA, Collins WB, Nicora CD, Purvine SO, Hoyt DW, Schückel J, Jørgensen B, Willats W, Spalinger DE, Firkins JL, Lipton MS, Sullivan MB, Pope PB, Wrighton KC. 2018. Interspecies cross-feeding orchestrates carbon degradation in the rumen ecosystem. *Nat Microbiol* 3:1274–1284. <https://doi.org/10.1038/s41564-018-0225-4>.
- Møller S, Sternberg C, Andersen JB, Christensen BB, Ramos JL, Givskov M, Molin S. 1998. In situ gene expression in mixed-culture biofilms: evidence of metabolic interactions between community members. *Appl Environ Microbiol* 64:721–732. <https://doi.org/10.1128/AEM.64.2.721-732.1998>.
- Hansen SK, Rainey PB, Haagensen JAJ, Molin S. 2007. Evolution of species interactions in a biofilm community. *Nature* 445:533–536. <https://doi.org/10.1038/nature05514>.
- Haagensen JA, Hansen SK, Christensen BB, Pamp SJ, Molin S. 2015. Development of spatial distribution patterns by biofilm cells. *Appl Environ Microbiol* 81:6120–6128. <https://doi.org/10.1128/AEM.01614-15>.
- Rodríguez-Verdugo A, Vulin C, Ackermann M. 2019. The rate of environmental fluctuations shapes ecological dynamics in a two-species microbial system. *Ecol Lett* 22:838–846. <https://doi.org/10.1111/ele.13241>.
- Rodríguez-Verdugo A, Ackermann M. 2021. Rapid evolution destabilizes species interactions in a fluctuating environment. *ISME J* 15:450–460. <https://doi.org/10.1038/s41396-020-00787-9>.
- Escalante AE, Rebolledo-Gómez M, Benítez M, Travisano M. 2015. Ecological perspectives on synthetic biology: insights from microbial population biology. *Front Microbiol* 6:143. <https://doi.org/10.3389/fmicb.2015.00143>.
- Ponomarova O, Patil KR. 2015. Metabolic interactions in microbial communities: untangling the Gordian knot. *Curr Opin Microbiol* 27:37–44. <https://doi.org/10.1016/j.mib.2015.06.014>.
- D'Souza G, Shitut S, Preussger D, Yousif G, Waschina S, Kost C. 2018. Ecology and evolution of metabolic cross-feeding interactions in bacteria. *Nat Prod Rep* 35:455–488. <https://doi.org/10.1039/c8np00009c>.
- Dekas AE, Poretsky RS, Orphan VJ. 2009. Deep-sea archaea fix and share nitrogen in methane-consuming microbial consortia. *Science* 326:422–426. <https://doi.org/10.1126/science.1178223>.
- Scheuerl T, Hopkins M, Nowell RW, Rivett DW, Barraclough TG, Bell T. 2020. Bacterial adaptation is constrained in complex communities. *Nat Commun* 11:754. <https://doi.org/10.1038/s41467-020-14570-z>.
- terHorst CP, Zee PC, Heath KD, Miller TE, Pastore AI, Patel S, Schreiber SJ, Wade MJ, Walsh MR. 2018. Evolution in a community context: trait responses to multiple species interactions. *Am Nat* 191:368–380. <https://doi.org/10.1086/695835>.
- Denef VJ, Banfield JF. 2012. In situ evolutionary rate measurements show ecological success of recently emerged bacterial hybrids. *Science* 336:462–466. <https://doi.org/10.1126/science.1218389>.

20. Chase AB, Weihe C, Martiny JBH. 2021. Adaptive differentiation and rapid evolution of a soil bacterium along a climate gradient. *Proc Natl Acad Sci U S A* 118:e2101254118. <https://doi.org/10.1073/pnas.2101254118>.
21. Malik AA, Martiny JBH, Brodie EL, Martiny AC, Treseder KK, Allison SD. 2020. Defining trait-based microbial strategies with consequences for soil carbon cycling under climate change. *ISME J* 14:1–9. <https://doi.org/10.1038/s41396-019-0510-0>.
22. Allison SD, Lu Y, Weihe C, Goulden ML, Martiny AC, Treseder KK, Martiny JB. 2013. Microbial abundance and composition influence litter decomposition response to environmental change. *Ecology* 94:714–725. <https://doi.org/10.1890/12-1243.1>.
23. Glassman SI, Weihe C, Li J, Albright MBN, Looby CI, Martiny AC, Treseder KK, Allison SD, Martiny JBH. 2018. Decomposition responses to climate depend on microbial community composition. *Proc Natl Acad Sci U S A* 115:11994–11999. <https://doi.org/10.1073/pnas.1811269115>.
24. Day MD, Beck D, Foster JA. 2011. Microbial communities as experimental units. *Bioscience* 61:398–406. <https://doi.org/10.1525/bio.2011.61.5.9>.